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## **Carbon Nanotubes Modified for Cellular** Membrane Integration.

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#### Abstract

In this examination, we create functionalized type of carbon a nanotube with detergent molecules designed to integrate with cellular membranes via attractive an electrical force between our molecular construct and the membrane itself.

examination, Upon the functionality of our construct shows bond between the carbon a detergent nanotube the and molecules. Hopefully opening the door to more advanced studies of cellular interactions.

#### Introduction

medicine has been Modern exploring advanced methods of biological integration of circuitry and physiological constructs for some time. With goals of modifying the behavior of many different types of biological structures from individual proteins to entire cells via the introduction of foreign bodies.

However the integration of biological and non-biological systems has been extremely difficult. Non-biological systems either lack the capacity to properly interface with cellular systems, or they are damaging to the cellular systems with which they are integrated. The goal of this examination is the creation of a biologically compatible, and conductive macromolecular highly construct. In theory, we will modify, and functionalize carbon nanotubes such that they are no longer biologically incomparable through the addition of other molecules. Modifying the properties of these carbon nanotubes will create the possibility of performing functions with them in the biological sciences. Generally, a carbon nanotube (CNT) is hydrophobic, which prevents both waterdispersion in solution and is part of what prevents a CNT from interacting with a cell. Our method around this is the use of two detergent molecules, SDS, and TWEEN, [fig. 1] and [fig. 2]. By attaching these molecules, the hydrophobic hydrophillic nature of the molecule will cause the hydrophobic end to face the CNT while the hydrophillic end will face away from the CNT. After fictionalization in acids, the CNT will have been partially dissolved and there will be available binding sites on the surface. Utilizing these binding sites, it is possible to bind the hydrophobic end of the detergent molecule to the wall of the CNT.

#### **Examination of Structure**

Via observing the absorption spectra of the Tween 80 solution, it is possible to correct for all substances in their non-bound state. Any further spectra appearance are the result of molecular interaction and covalent bonding.

The spectra show an absorption peak at 300nm which is unaccounted for from other sources.



#### **Molecular Properties**

Tween and SDS are molecules that have a charged 'head' and a long uncharged 'tail'. Just like the phospholipids which make up the cellular membrane of animal cells. However, these molecular heads are positively charged as opposed to phospholipid bilayers which are negatively charged at the head.

This makes them 'sticky' with respect to a cellular bilayer. However the substrate of a CNT prevent them from tearing the phopholipid bilayer of the cellular membrane apart.



## Discussion

The new spectral peak at 300nm indicates that there exists a change in material properties. The most likely cause of this occurrence is a covalent bond at the reactive binding sites which have been generated on the surface of the nanotube.

Using further analysis, it is now necessary to test the properties of the constructed nanotube as well as place it in an interactive media. A proper characterization of this structure should lead to potential uses in terms of analyzing cellular media and possible use in medical development.

This this could be useful in the creation of a connective molecular structure on the nanoscale such that individual cells, or parts of individual cells can be monitored for activity



## **Functional result:**



Figure 4. Functionalized Multiwall Carbon Nanotube with SDS.

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#### Citations;

Ye, J., Cui, H., Wen, Y., Ottova, A., Tien, H., Xu, G. Sheu, F., Self-assembly of bilayer lipid membrane at multiwalled carbon nanotubes towards the development of photo-switched functional device, Electrochemistry Communications 7 (2005) 81-86.

Wallace, E., Sansom, M., Carbon nanotube self-assembly with lipids and detergent: a molecular dynamics study, Nanotechnology 20 (2009)

Sanka, M. S. L. R., Role f Functionalized Carbon nanotubes in Stabilization of RNA, University of Bridgeport Biomedical Engineering Thesis.

http://www.chm.bris.ac.uk/ As an image source http://nanogloss.com/ As an image source