



Highly sensitive detection of estradiol by a SERS sensor based on TiO₂ covered with gold nanoparticles

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Abstract

We propose the use of gold nanoparticles grown on the surface of nanoporous TiO₂ films as surface-enhanced Raman scattering (SERS) sensors for the detection of 17 β -estradiol. Gold deposition on top of a TiO₂ surface leads to the formation of nanoparticles the plasmonic properties of which fulfil the requirements of a SERS sensor. The morphological and optical properties of the surface were investigated. Specifically, we demonstrate that the TiO₂ background pressure during pulsed laser deposition and the annealing conditions offer control over the formation of Au nanoparticles with different sizes, shapes and distributions, yielding a versatile sensor. We have exploited the surface for the detection of 17 β -estradiol, an emerging contaminant in environmental waters. We have found a limit of detection of 1 nM with a sensitivity allowing for a dynamic range of five orders of magnitude (up to 100 μ M).

Introduction

Surface-enhanced Raman scattering (SERS) as a sensing tool requires the optimization of a surface and its functionalization. The surface should provide a good enhancement over a large

range of wavelengths, to detect molecules with various fingerprints, while it should also be easy to fabricate at reduced cost. In addition, the surface functionalization needs to guarantee the

selection, detection and quantification of a target molecule, e.g., a biomarker [1-3] or a pollutant [4,5] dissolved in complex media such as blood, plasma, urine, or river and sea water.

SERS is mainly based on an electromagnetic effect that originates from the excitation of plasmon resonances, in particular of localized surface plasmons (LSPs) in metallic nanoparticles (NPs). Other effects may contribute to the enhancement such as the formation of hot spots or lightning rod effects [6-8]. Many surfaces were proposed for SERS including rough metallic surfaces [9,10], colloidal solutions [11], and structures with controlled size, distance and shape obtained via lithography techniques [2,5,12]. However, these techniques can be time-consuming and expensive. Recently, the use of composite systems of dielectrics (TiO₂, ZnO) and metallic NPs has gathered increasing attention regarding SERS applications, because the plasmonic enhancement provided by metallic NPs can be combined with the optical properties of the semiconductor such as light trapping, scattering, and antireflection abilities [13-15]. These composite microstructures have also shown to maximize the path of the Raman excitation laser beam within the substrate, leading to signals with higher intensity.

Samransuksamer et al. [16] used TiO₂ nanorods decorated with Au NPs, deposited via precipitation by soaking in HAuCl₄ solution, as composite SERS substrates for the detection of methylene blue. They reported a successful SERS enhancement, compared to bare Si substrates, with an enhancement factor of ca. 10⁶ and a lower detection limit of 100 nM. Li et al. [15] studied Au NP-coated TiO₂ nanotube arrays as SERS substrate for the detection of rhodamine 6G and other organic molecules. They obtained stable and reproducible results with a detection limit down to 10 μM, while also showing high recyclability through cleaning via UV irradiation. However, a main drawback of these methodologies is the use of aggressive solvents, which can induce damage especially in delicate applications involving polymeric substrates. Also, the control over the size and shape of AuNPs and, thus, over their plasmonic behavior is often limited.

Here, we propose the use of a nanostructured hierarchically organized TiO₂ film as a template for the growth of Au NPs (in the following the samples will be referred as TiO₂/Au). Both TiO₂ film and Au NPs were synthesized by vapor phase deposition techniques (involving pulsed laser deposition and thermal evaporation) avoiding the use of solvents, while accurately tuning the morphology and the plasmonic properties. Specifically, TiO₂ films with different porosities have been deposited, with different Au NP sizes and coverages. Then, the growth parameters of TiO₂ and of the AuNPs were selected in order to obtain the maximum SERS enhancement. In a second step, the

Au NPs were functionalized with aptamers (a biorecognition element), specific to the natural estrogen 17β-estradiol (E2) [17] for SERS detection. The all-solid configuration of the TiO₂/Au surface makes it a good candidate for in situ detection.

Surface functionalization with aptamers is gathering interest because they possess many of the important qualities required for the functionalization of SERS sensors [18-21]. Aptamers are single-stranded DNA molecules that are specifically selected to bind to a target molecule. They are relatively cheap and their chemistry is easy to tune so that they can attach to a metallic surface. Also, they can be selected to be short enough to guarantee that the targeted molecule is in the enhancement volume of the plasmonic nanoparticle (the effect of SERS decreases exponentially with the distance from the surface and is negligible beyond 5 nm). Another interesting feature of aptamers is that their Raman fingerprint is easily recognizable, as DNA is an extensively studied molecule.

In this study, we focus on the detection of E2 with an aptamer-functionalized sensor. E2 is the main female hormone responsible for growth, reproduction, breast development, maturation, bone formation, and childbearing in mammals. It is the most potent estrogen [22]. Estrogens found in the environment originate from human and animal excretions and are released into surface waters from agricultural activities, non-treated waste or wastewater treatment effluents [23]. High concentrations of E2 have been found in surface and groundwater in urban areas, leading to rising concerns in the EU. Studies have revealed that in some fish species, exposure to E2 has led to the feminization of males [24,25]. Routine instrumental methods for the detection of E2 in environmental waters are well established [26]. They are very specific and very sensitive, however, they are also time-consuming and expensive [27]. SERS sensors are therefore investigated as an alternative as they present the potential for in situ near-real-time analysis.

In the following we will present the possibilities of TiO₂ porous surfaces decorated with Au NPs regarding the use as SERS sensors. The tuning of TiO₂ growth and Au deposition gives access to a variety of surfaces with specific optical properties. In a second part, we show that it is possible to detect as low as 1 nM of E2 using these surfaces.

Experimental

Growth of the TiO₂/Au nanostructured surfaces

TiO₂/Au substrates were synthesized using a two-step deposition. First, a nanostructured TiO₂ film was synthesized by pulsed laser deposition (PLD). Then, a Au NP layer was

deposited on top by thermal evaporation of Au followed by solid-state dewetting to induce the formation of NPs. A TiO₂ (99.9%) target was ablated using a Nd:YAG laser ($\lambda = 532$ nm) with a pulse duration of 5–7 ns and 10 Hz repetition rate. The laser fluence on the target is 3.5 J·cm⁻² and the pulse energy is 200 mJ. Film synthesis was carried out at room temperature in oxygen atmosphere, using both Si(100) and soda-lime glass substrates, which were mounted on a sample holder at a fixed target-to-substrate distance of 5 cm. Changing the background pressure within the deposition chamber allowed for a tuning of the film morphology (i.e., a higher pressure resulted in a higher film porosity) [28,29]. Samples were therefore deposited at a pressure of 8 or 12 Pa to obtain different film porosities.

A thin layer of Au (99.9%) was then evaporated using an Edwards E306A resistively heated thermal evaporator. The equivalent (i.e. nominal) thickness of the evaporated layers was monitored with a quartz microbalance sensor. Three different values of Au thickness, namely 3, 6, and 15 nm, were selected enabling the formation of different sizes of AuNPs through subsequent annealing. Selected samples underwent annealing at 500 °C for 2 h in air, in a Lenton muffle furnace with 4 °C/min heating ramp. The thermal treatment was carried out to induce both crystallization of the as-deposited amorphous TiO₂ into the anatase phase (as discussed in [28,29]) and the formation of AuNPs exploiting dewetting of the Au films.

A field-emission scanning electron microscope (FEG-SEM, Zeiss Supra 40) was used to perform morphological characterizations of the films deposited on Si(100) substrates. The average size distribution of Au NPs was estimated through statistical analysis of top-view SEM images with the open source software ImageJ[®]. Since the shape of the Au NPs was not always perfectly circular, their area was measured with ImageJ[®] in order to calculate the equivalent diameter, which was used to define the NP size.

Chemicals and reagents

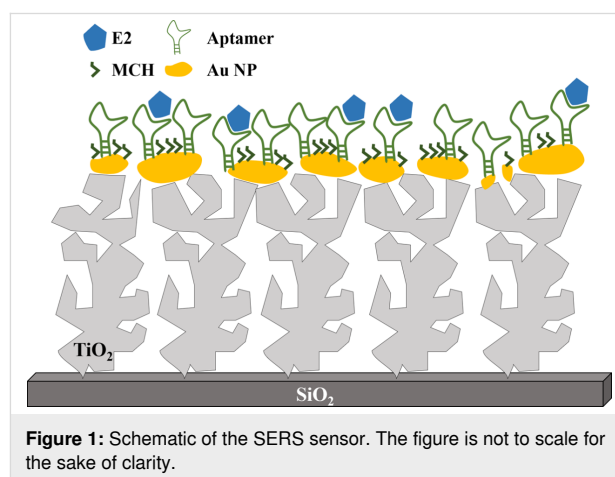
Mercaptobenzoic acid (MBA), 6-mercapto-1-hexanol (MCH), 17 β -estradiol (E2), and ethanol were purchased from Sigma-Aldrich.

A 17 β -estradiol binding aptamer was previously isolated by the SELEX process by Kim and co-workers [17]. It was purchased from Eurogentec with the following 76-mer-long sequence SH-C6-5'-GCT-TCC-AGC-TTA-TTG-AAT-TAC-ACG-CAG-AGG-GTA-GCG-GCT-CTG-CGC-ATT-CAA-TTG-CTG-CGC-GCT-GAA-GCG-CGG-AAG-C-3'. The thiol group was added to the 5' end of the aptamer for functionalization via a Au–S bond. The aptamer was diluted and aliquoted upon arrival and stored at –18 °C.

Sample functionalization

MBA was used as a common Raman reporter to find out the best deposition parameters for Raman scattering enhancement. The thiol group is known to have a strong affinity to Au but none to TiO₂. Moreover, the Raman spectrum of MBA is well known and the molecule demonstrates a large scattering cross section [30]. For the functionalization of TiO₂/Au surfaces, MBA was diluted in ethanol at a concentration of 2.9 mM. The TiO₂/Au samples were then soaked in the solution overnight. After that, they were thoroughly rinsed with ethanol before drying with nitrogen.

For E2 detection, the TiO₂/Au surfaces were first functionalized with the aptamer. For this a fresh 3 μ M solution of aptamer was prepared in Tris-HCl buffer solution (20 mM Tris-HCl contained 1 M of HCl, 0.1 M of NaCl and 5 mM of MgCl₂, pH 7.5). The samples were left for 2 h in this solution before rinsing with Tris-HCl buffer solution and Milli-Q water. In order to prevent unspecific interactions between E2 and the gold surface, the sample was then left for 2 h in a solution of the blocking agent MCH. MCH occupies gold sites that are not functionalized with the aptamer (Figure 1). This prevents the deposition of unwanted molecules from the sample, which could blur the signal. It also prevents the amine groups of DNA to form weak bonds with the gold and it helps the aptamer to have a homogeneous orientation on the surface [31].



The MCH solution was prepared in phosphate-buffered saline (PBS) solution mixed with 10 mM MgCl₂. The concentration of MCH was constant at 14.6 μ M. The samples were rinsed three times with PBS and reverse osmosis water (RO water) to remove any unbound or excess molecules after the deposition.

Samples with different concentrations of E2 were prepared in order to test the sensitivity of the sensor. E2 was first diluted in ethanol at saturation (36 mM). This solution was further diluted

in RO water to obtain E2 solutions with concentrations of 1 nM, 10 nM, 100 nM, 1 μ M, 10 μ M, 100 μ M, and 1 mM. Samples were left in the E2 solutions for 1 h before being rinsed with RO water and blown dry. Figure 1 gives a schematic of the final system.

Optical and SERS measurements

Plasmon resonance was evaluated via optical spectroscopy. For this purpose, transmission spectra were acquired using a UV–vis–NIR spectrophotometer (PerkinElmer Lambda 1050) with a 150 mm diameter integrating sphere in the range of 250–2000 nm, illuminating the sample from the glass substrate side. All acquired spectra were normalized with respect to the contribution of the glass substrate.

The SERS spectra were recorded with a micro-Raman spectrophotometer (Jobin-Yvon Labram 300), using a 100 \times magnification objective (NA = 0.90) in back-scattering geometry, with a spectral resolution of 3 cm^{-1} and a spatial resolution of about 1 μm . The employed excitation wavelength was 633 nm, with a power of 1 mW for an acquisition time of 300 s. This trade-off between power and duration of the acquisition has been chosen after a series of tests in which the power was gradually decreased from 10 mW to 100 μ W. The typical peak of silicon at 521 cm^{-1} was used as an internal reference to normalize the intensities of all the spectra. The spectra presented here are the average of four spectra taken at different locations of each sample.

Results and Discussion

Sample growth and structural characterization

Morphology

Nanostructured TiO_2 films with hierarchical micrometer/nanometer-scale morphology and tuned porosity were deposited by pulsed laser deposition (PLD) as already reported in [28]. By increasing the background O_2 pressure during deposition it is possible to deposit films that are more porous. Samples were therefore synthesized at background pressures of 8 or 12 Pa. Au layers were then evaporated on top of the TiO_2 films. Three

nominal thickness values of 3, 6, and 15 nm were chosen, in order to obtain NPs with different diameters (Table 1). After deposition of Au, samples underwent an annealing treatment in a furnace at 500 $^\circ\text{C}$ for 2 h, which leads to the crystallization of TiO_2 to the anatase phase (as demonstrated by Raman spectra, not shown) and caused dewetting in the Au layer with the subsequent formation of NPs [32]. Evaporation and dewetting of Au on TiO_2 layers with different porosity was carried out to exploit the effect of the surface morphology on the formation of Au NPs, yielding different size distributions and densities.

Figure 2 shows the two boundary cases of 3 and 15 nm of evaporated Au on a TiO_2 film deposited at 12 Pa. The observations were similar for the 8 Pa films. The effects of heat treatment are clearly visible. In the case of 15 nm of Au (Figure 2c,d), a continuous layer is formed on the underlying TiO_2 surface, which upon thermal treatment forms isolated, large Au nanoislands. Evaporation of 3 nm of Au resulted instead in a non-continuous nanostructured layer, for which thermal treatment led to the growth of well separated AuNPs (Figure 2a,b).

Statistical analyses of the top-view SEM images after annealing (some of which are reported in Figure 3) proved that by increasing the amount of evaporated Au, it was possible to increase the NP size. The average equivalent NP diameters varied from 12 up to 115 nm (Table 1). Moreover, the morphology of the TiO_2 film also played a role in determining the final Au NP diameter, with smaller equivalent diameters obtained in the case of more porous films deposited at 12 Pa. Finally, the Au coverage increased with the amount of Au deposited on the TiO_2 surface up to almost 30%, while it was almost independent of the TiO_2 porosity.

Optical properties

Figure 4 reports the optical transmittance of the TiO_2/Au samples. Our previous studies involving TiO_2/Au systems have shown that the reflectance has low values of about 5% and can thus be considered negligible [28]. A general decrease of the optical transmittance was observed for large quantities of Au and the transmittance was influenced by the growth conditions and the thermal treatment. For example, in the case of 6 nm of

Table 1: Average equivalent diameter with standard deviation of Au NPs on TiO_2 films after annealing.

	8 Pa			12 Pa		
	Au 3 nm	Au 6 nm	Au 15 nm	Au 3 nm	Au 6 nm	Au 15 nm
average diameter [nm]	12	23	115	13	19	64
standard deviation [nm]	4	12	88	6	13	42
Au surface coverage [%]	9.5	21.6	35.1	9.4	22.2	35.7

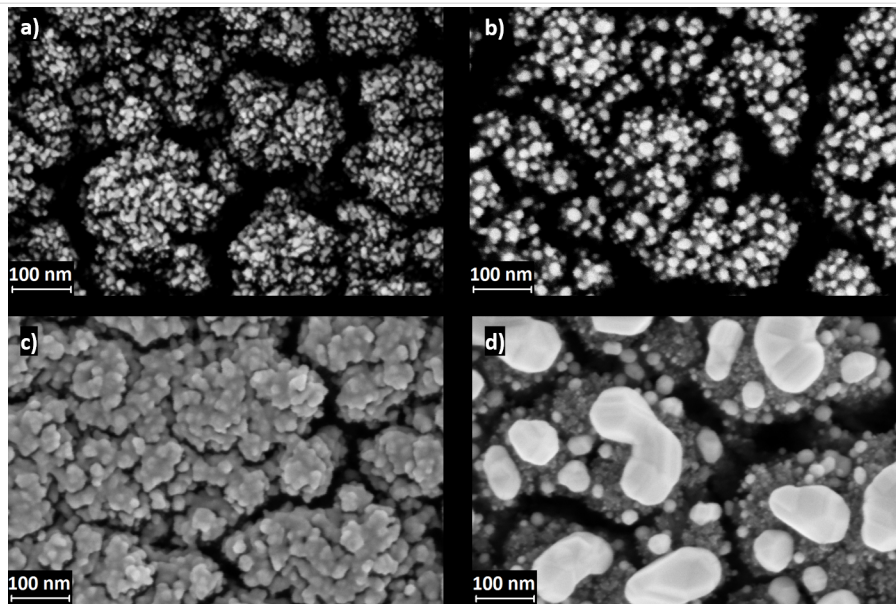


Figure 2: SEM top-view images showing 12 Pa TiO₂ samples with 3 nm of evaporated Au before (a) and after (b) annealing. Panels (c) and (d) show 12 Pa TiO₂ samples with 15 nm of evaporated Au, respectively, before (Au continuous layer) and after annealing (completely formed NPs).

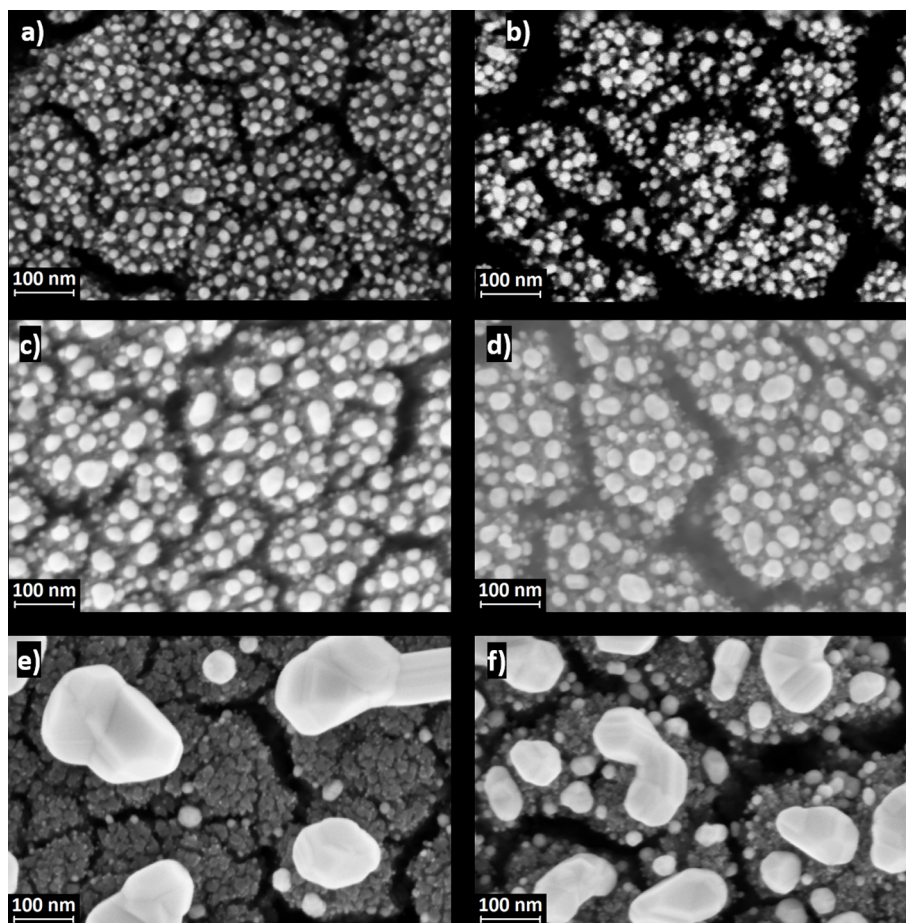


Figure 3: SEM top-view images of TiO₂ 8 Pa with 3 nm (a), 6 nm (c), and 15 nm (e) of Au, and of TiO₂ 12 Pa with 3 nm (b), 6 nm (d), and 15 nm (f) of Au. All micrographs were acquired after annealing.

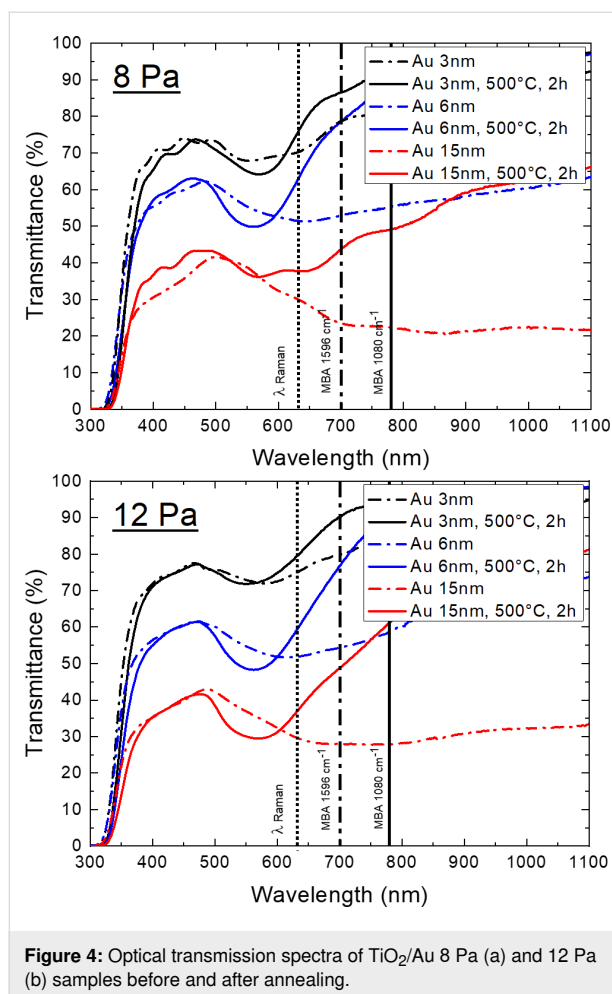


Figure 4: Optical transmission spectra of TiO_2/Au 8 Pa (a) and 12 Pa (b) samples before and after annealing.

Au the wavelength at which maximum absorption occurs changed after annealing from about 620 to 560 nm (Table 2). These trends were outlined also in works by Dorono-Mor et al. [33] and Karakouz and co-workers [34]. More specifically, the as-deposited TiO_2/Au samples displayed a very broad absorption, especially for greater amounts of evaporated Au, when an almost continuous Au layer was formed. However, after annealing the absorption peak became sharper and blue-shifted,

which can be attributed to the formation of Au NPs. In the annealed samples the LSPR resonance (LSPR) red-shifted as a function of the Au NP size, i.e., the wavelength for maximum absorption varied from 549 nm (for the 12 Pa sample with 3 nm of Au) up to a maximum value of 575 nm (for the 8 Pa sample with 15 nm of Au) [35]. Moreover, the full width at half maximum (FWHM) increased as a function of the AuNP size, due to the fact that NPs exhibited a broader dispersion, as reported also by Gaspar and co-workers [36]. In other words, the optical properties and the LSPR changed due to the morphology change of the deposited Au, from an almost continuous layer to various shape/aspect ratios, size distributions, and average distances between the Au NPs, during annealing (Figure 3).

Selection of the TiO_2/Au surfaces for E2 detection

To test the TiO_2/Au surfaces as SERS substrates, MBA was used as it is a well-known Raman reporter, showing two intense characteristic peaks at 1080 and 1590 cm^{-1} from aromatic ring vibrations [30]. The structure of MBA and the grafting process (thiol–gold interaction) guarantee that the molecule will preferentially attach to gold [37,38]. The purpose here was to select TiO_2 growth and Au deposition parameters that would allow for the best enhancement possible over the largest wavelength range.

MBA was grafted on samples produced under the different growth conditions presented in the previous section. The detailed Raman spectra of MBA are displayed in Figure 5. MBA is barely detectable on the TiO_2/Au 3 nm samples. The peaks start to be visible when the TiO_2 surface is decorated with more than 6 nm of Au. A rule of thumb is that the highest enhancement is achieved when the wavelength of the LSPR is between the excitation wavelength (here 632 nm) and the Raman wavelength of the peak in consideration [5,39]. As shown in Figure 4 the plasmon resonances for all the samples were outside of this interval. The plasmonic properties of the NPs probably contribute to the enhancement of the Raman scat-

Table 2: LSPR wavelength (minima of transmission spectra) and corresponding FWHM.

	8 Pa				
	3 nm		6 nm		15 nm
	as deposited	500 °C, 2 h	as deposited	500 °C, 2 h	500 °C, 2 h
plasmon wavelength [nm]	553	569	637	560	575
FWHM [nm]	118	91	313	94	—
	12 Pa				
plasmon wavelength [nm]	579	549	615	562	569
FWHM [nm]	110	87	216	102	114

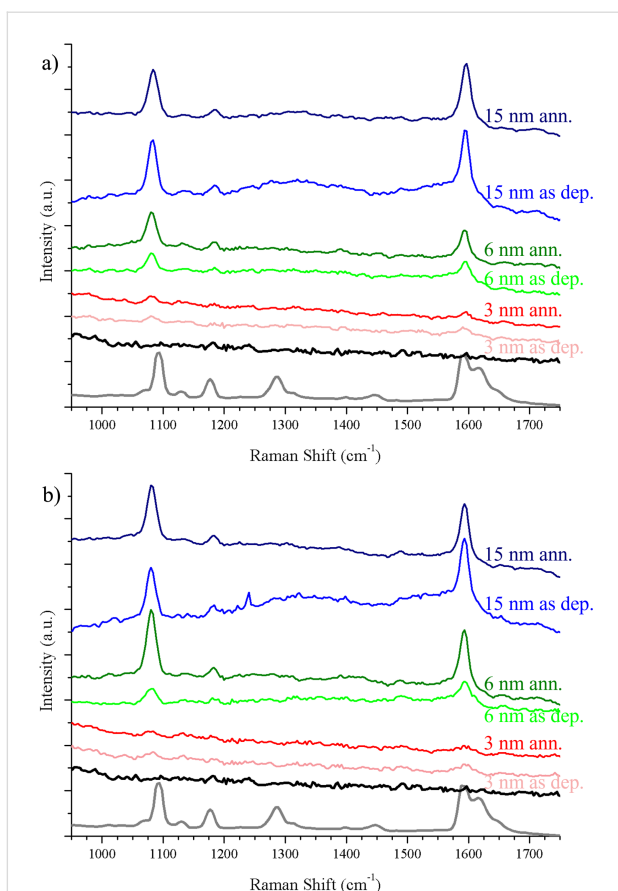


Figure 5: SERS measurements. The red, green and blue lines show SERS spectra of MBA deposited on, respectively, 3, 6, and 15 nm of Au deposited on TiO₂ surfaces at 8 Pa (a) and 12 Pa (b). The corresponding brighter color means that there was no annealing of the TiO₂. Grey line: Raman spectrum of MBA powder (the intensity has been scaled down to be comparable with the SERS spectra). Black line: spectrum of MBA deposited on a TiO₂ surface without Au, yielding no SERS signal.

tering. However, it also originates from the proximity of the NPs, which is higher for higher Au coverages (Table 1). The electric field between two nanoparticles is extraordinarily enhanced when the NPs are close to each other [7,40] and form so-called hot spots.

To compare the enhancement capacity of the TiO₂/Au samples, the enhancement factor (EF) was calculated as follows [40]:

$$EF = \frac{I_{\text{SERS}}}{I_{\text{Raman}}} \frac{N_{\text{Raman}}}{N_{\text{SERS}}} \quad (1)$$

I_{SERS} and I_{Raman} are the intensities of the analyzed peak of MBA acquired in SERS configuration and from MBA powder in standard Raman configuration. N_{Raman} is the number of molecules within the excited laser volume, which can be calculated from the density and the molar mass of MBA ($\rho_{\text{MBA}} = 1.5 \text{ g}\cdot\text{cm}^{-3}$, $M_{\text{MBA}} = 154.19 \text{ g}\cdot\text{mol}^{-1}$), the laser spot area (A_{spot}), and the penetration depth of the focused laser beam (which was assumed to be $h = 12 \mu\text{m}$), as:

$$M_{\text{Raman}} = A_{\text{spot}} h \frac{\rho_{\text{MBA}}}{M_{\text{MBA}}} N_{\text{A}}, \quad (2)$$

where N_{A} is the Avogadro constant. N_{SERS} corresponds to the number of molecules adsorbed on the AuNP surface within the laser spot, which we assume to be a single monolayer of MBA fully covering the surface [38,41]. From the SERS and Raman spectra of Figure 5 the EF values were calculated (Table 3). The sample that gave the most homogeneous EF through over the investigated wavelength range was TiO₂/Au 6 nm deposited at 12 Pa and annealed at 500 °C for 2 h. Hence, it was chosen for the subsequent examination of the of E2.

Table 3: Enhancement factor values for MBA, calculated for the peaks at 1080 and 1590 cm⁻¹.

	Au 3 nm		8 Pa		Au 15 nm	
	as deposited	500 °C, 2 h	Au 6 nm 500 °C, 2 h		as deposited	500 °C, 2 h
1080 cm ⁻¹	1.9·10 ⁵	2.9·10 ⁵	5.2·10 ⁵		8.5·10 ⁴	2.5·10 ⁵
1590 cm ⁻¹	9.3·10 ⁴	8.2·10 ⁴	2.9·10 ⁵		5.0·10 ⁴	2.3·10 ⁵
12 Pa						
1080 cm ⁻¹	1.5·10 ⁵	1.1·10 ⁵	3.7·10 ⁵		8.9·10 ⁴	3.4·10 ⁵
1590 cm ⁻¹	8.0·10 ⁴	3.9·10 ⁴	3.3·10 ⁵		6.0·10 ⁴	2.4·10 ⁵
Si(100)						
1080 cm ⁻¹	—	—	2.7·10 ⁴		—	—
1590 cm ⁻¹	—	—	1.5·10 ⁴		—	—

Even though the SERS enhancement might be mainly attributed to an electromagnetic effect arising from the Au NPs, the presence of TiO₂ could also be beneficial for different reasons. In addition to the abovementioned influence of the TiO₂ surface on the size of the Au NPs (and consequently their plasmonic properties), a first hint can be found in the optical properties of semiconductor nanostructured materials. Their light-scattering, light-trapping and antireflection abilities, have already been reported to improve SERS enhancement [14–16]. In addition, the nanostructured morphology contributes to provide a larger available surface for Au NP growth, but also for analyte molecules to be adsorbed. The positive influence of the nanostructured TiO₂ film was also confirmed by comparing SERS enhancements of the composite TiO₂/Au 6 nm sensor with that of a bare Si(100) substrate on which the same equivalent thickness of Au (6 nm) was evaporated and annealed at 500 °C for 2 h. Figure 6 shows a clearly larger enhancement for both MBA peaks on the TiO₂/Au sensor. The resulting EF is about one order of magnitude higher than that of the bare Si(100) substrate (Table 3). Therefore, the presence of the nanostructured TiO₂ film benefits the overall SERS effect.

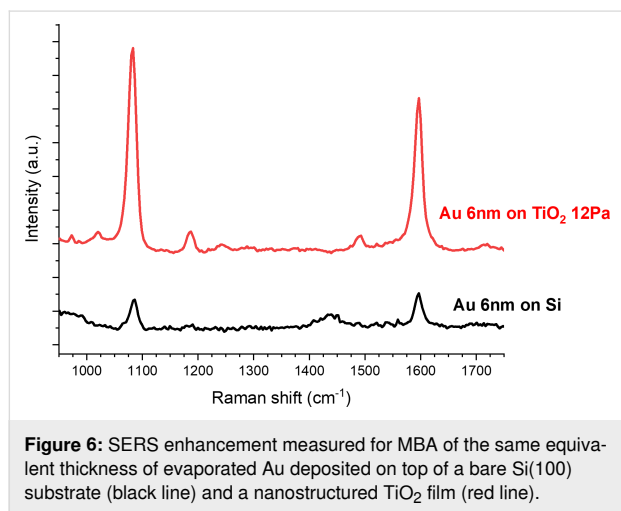


Figure 6: SERS enhancement measured for MBA of the same equivalent thickness of evaporated Au deposited on top of a bare Si(100) substrate (black line) and a nanostructured TiO₂ film (red line).

Application to E2 detection

Figure 7a presents the Raman spectrum of E2 powder and the SERS spectra of E2 measured with the selected sensor. The signal of the empty sensor is designated as “Apt+MCH”. It mostly reflects the signal of the aptamer as MCH is known to have a very low Raman cross section and is thus not expected to yield a significant signal.

From the comparison between the spectra of the empty sensor (Apt+MCH) and of the hormone (E2 powder), it is possible to distinguish the zones where the E2 fingerprint can be found in the SERS spectra. These curves were acquired at concentrations ranging from 1 nM to 1 mM of E2. The fingerprint of E2

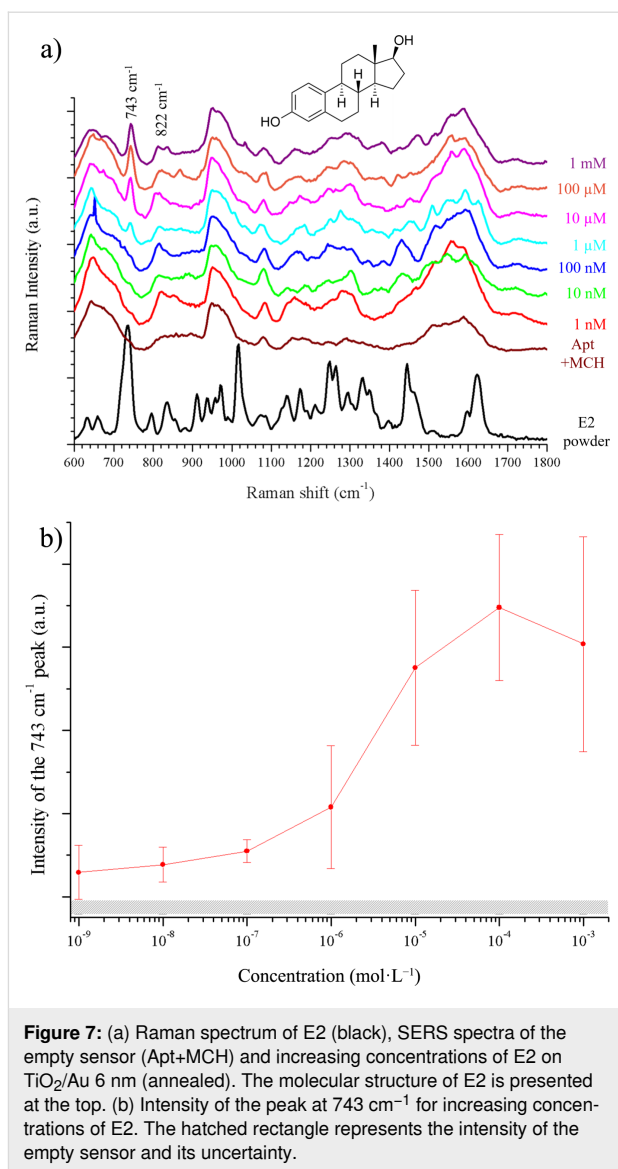


Figure 7: (a) Raman spectrum of E2 (black), SERS spectra of the empty sensor (Apt+MCH) and increasing concentrations of E2 on TiO₂/Au 6 nm (annealed). The molecular structure of E2 is presented at the top. (b) Intensity of the peak at 743 cm⁻¹ for increasing concentrations of E2. The hatched rectangle represents the intensity of the empty sensor and its uncertainty.

is clearly emerging from the aptamer spectra as the concentration increases. Shifts from the Raman spectra of the powder can be observed, which are due to the fact that the molecule is bound to the aptamer, affecting the vibration frequencies. For instance, the peak at 743 cm⁻¹ corresponds to the peak at 730 cm⁻¹ in the E2 Raman spectra, while the peak at 822 cm⁻¹ corresponds to the peak at 830 cm⁻¹. These two peaks are, respectively, attributed to bending of CCH and bending of CH [42]. They become clearer as the E2 concentration increases. Another feature can be seen in the region between 1200 and 1400 cm⁻¹ corresponding to aromatic ring deformation modes, in-plane OH bending modes, and aliphatic/aromatic CH bending modes [43]. In order to study the evolution of the spectra with E2 concentration, we have plotted the total intensity of the peak at 743 cm⁻¹ as a function of the E2 concentration (Figure 7b). The error bars reflect the homogeneity of the sam-

ple surface. The signal increases with the concentration and reaches saturation at 100 μM . The hatched rectangle is the intensity of the same zone but for the empty sensor (without E2). Even at the lowest concentration (1 nM) the peak intensity is above this reference signal. The TiO_2/Au 6 nm sensor was tested from 1 nM to 1 mM. The quantification is possible between 1 nM and 10 μM , i.e., the sensor has a dynamic range of at least four orders of magnitude.

Conclusion

TiO_2 nanoporous surfaces covered with Au NPs were tested as SERS surfaces for the detection of 17 β -estradiol. Different conditions of Au deposition were considered as they lead to different shapes, sizes and distributions of the Au NPs. The TiO_2/Au 6 nm deposited at 12 Pa and annealed for 2 h at 500 $^\circ\text{C}$ gives an enhancement factor (EF) of $3.7 \cdot 10^5$ and $3.4 \cdot 10^5$ at, respectively, 1080 and 1590 cm^{-1} . These high EF values for two distant wavelengths has been exploited to test the detection of E2 in water. For the detection, the surfaces were functionalized with aptamers in order to guarantee a good specificity [17]. We thus have produced a sensor that is specific (with the use of aptamer), can detect low concentrations (1 nM, compatible with environmentally relevant concentration) and has a wide dynamic range (up to 100 μM). These results combined with the fact that the sensor is all solid makes the nanoporous TiO_2/Au systems interesting for environmental detection applications.

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Preprint

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References

- Cottat, M.; D'Andrea, C.; Yasukuni, R.; Malashikhina, N.; Grinyte, R.; Lidgi-Guigui, N.; Fazio, B.; Sutton, A.; Oudar, O.; Charneau, N.; Pavlov, V.; Toma, A.; Di Fabrizio, E.; Gucciardi, P. G.; Lamy de la Chapelle, M. *J. Phys. Chem. C* **2015**, *119*, 15532–15540. doi:10.1021/acs.jpcc.5b03681
- Cottat, M.; Lidgi-Guigui, N.; Tijunelyte, I.; Barbillon, G.; Hamouda, F.; Gogol, P.; Aassime, A.; Lourtioz, J.-M.; Bartenlian, B.; de la Chapelle, M. L. *Nanoscale Res. Lett.* **2014**, *9*, 623. doi:10.1186/1556-276x-9-623
- Galarreta, B. C.; Norton, P. R.; Lagugne-Labarhet, F. *Langmuir* **2011**, *27*, 1494–1498. doi:10.1021/la1047497
- Tijunelyte, I.; Dupont, N.; Milosevic, I.; Barbey, C.; Rinnert, E.; Lidgi-Guigui, N.; Guenin, E.; de la Chapelle, M. L. *Environ. Sci. Pollut. Res.* **2017**, *24*, 27077–27089. doi:10.1007/s11356-015-4361-6
- Guillot, N.; Shen, H.; Frémaux, B.; Péron, O.; Rinnert, E.; Toury, T.; Lamy de la Chapelle, M. *Appl. Phys. Lett.* **2010**, *97*, 023113. doi:10.1063/1.3462068
- Moskovits, M. *J. Raman Spectrosc.* **2005**, *36*, 485–496. doi:10.1002/jrs.1362
- Radziuk, D.; Moehwald, H. *Phys. Chem. Chem. Phys.* **2015**, *17*, 21072–21093. doi:10.1039/c4cp04946b
- Campion, A.; Kambhampati, P. *Chem. Soc. Rev.* **1998**, *27*, 241–250. doi:10.1039/a827241z
- Jeanmaire, D. L.; Van Duyne, R. P. *J. Electroanal. Chem. Interfacial Electrochem.* **1977**, *84*, 1–20. doi:10.1016/s0022-0728(77)80224-6
- Albrecht, M. G.; Creighton, J. A. *J. Am. Chem. Soc.* **1977**, *99*, 5215–5217. doi:10.1021/ja00457a071
- Kneipp, K.; Wang, Y.; Kneipp, H.; Perelman, L. T.; Itzkan, I.; Dasari, R. R.; Feld, M. S. *Phys. Rev. Lett.* **1997**, *78*, 1667–1670. doi:10.1103/physrevlett.78.1667
- Hulteen, J. C.; Treichel, D. A.; Smith, M. T.; Duval, M. L.; Jensen, T. R.; Van Duyne, R. P. *J. Phys. Chem. B* **1999**, *103*, 3854–3863. doi:10.1021/jp9904771
- Ben-Jaber, S.; Peveler, W. J.; Quesada-Cabrera, R.; Cortés, E.; Sotelo-Vazquez, C.; Abdul-Karim, N.; Maier, S. A.; Parkin, I. P. *Nat. Commun.* **2016**, *7*, 12189. doi:10.1038/ncomms12189
- Alessandri, I.; Lombardi, J. R. *Chem. Rev.* **2016**, *116*, 14921–14981. doi:10.1021/acs.chemrev.6b00365
- Li, X.; Chen, G.; Yang, L.; Jin, Z.; Liu, J. *Adv. Funct. Mater.* **2010**, *20*, 2815–2824. doi:10.1002/adfm.201000792
- Samransuksamer, B.; Horprathum, M.; Jutarosaga, T.; Kopwithaya, A.; Limwichean, S.; Nuntawong, N.; Chananonawathorn, C.; Patthanasettakul, V.; Muthitamongkol, P.; Treetong, A.; Klamchuen, A.; Leelapojanaporn, A.; Thanachayanont, C.; Eiamchai, P. *Sens. Actuators, B* **2018**, *277*, 102–113. doi:10.1016/j.snb.2018.08.136
- Kim, Y. S.; Jung, H. S.; Matsuura, T.; Lee, H. Y.; Kawai, T.; Gu, M. B. *Biosens. Bioelectron.* **2007**, *22*, 2525–2531. doi:10.1016/j.bios.2006.10.004
- Cottat, M.; Lidgi-Guigui, N.; Hamouda, F.; Bartenlian, B.; Venkataraman, D.; Marks, R. S.; Steele, T. W. J.; de la Chapelle, M. L. *J. Opt. (Bristol, U. K.)* **2015**, *17*, 114019. doi:10.1088/2040-8978/17/11/114019
- Pavlov, V.; Willner, I.; Dishon, A.; Kotler, M. *Biosens. Bioelectron.* **2004**, *20*, 1011–1021. doi:10.1016/j.bios.2004.06.020
- Song, S.; Wang, L.; Li, J.; Fan, C.; Zhao, J. *TrAC, Trends Anal. Chem.* **2008**, *27*, 108–117. doi:10.1016/j.trac.2007.12.004
- Terracciano, M.; Rea, I.; Borbone, N.; Moretta, R.; Oliviero, G.; Piccialli, G.; De Stefano, L. *Molecules* **2019**, *24*, 2216. doi:10.3390/molecules24122216
- Hamid, H.; Eskicioglu, C. *Water Res.* **2012**, *46*, 5813–5833. doi:10.1016/j.watres.2012.08.002
- Liu, S.; Cheng, R.; Chen, Y.; Shi, H.; Zhao, G. *Sens. Actuators, B* **2018**, *254*, 1157–1164. doi:10.1016/j.snb.2017.08.003

24. Wise, A.; O'Brien, K.; Woodruff, T. *Environ. Sci. Technol.* **2011**, *45*, 51–60. doi:10.1021/es1014482
25. Vajda, A. M.; Barber, L. B.; Gray, J. L.; Lopez, E. M.; Woodling, J. D.; Norris, D. O. *Environ. Sci. Technol.* **2008**, *42*, 3407–3414. doi:10.1021/es0720661
26. Snyder, S. A.; Keith, T. L.; Verbrugge, D. A.; Snyder, E. M.; Gross, T. S.; Kannan, K.; Giesy, J. P. *Environ. Sci. Technol.* **1999**, *33*, 2814–2820. doi:10.1021/es981294f
27. Long, F.; Zhu, A.; Shi, H. *Sensors* **2013**, *13*, 13928–13948. doi:10.3390/s131013928
28. Bricchi, B. R.; Ghidelli, M.; Mascaretti, L.; Zapelli, A.; Russo, V.; Casari, C. S.; Terraneo, G.; Alessandri, I.; Ducati, C.; Li Bassi, A. *Mater. Des.* **2018**, *156*, 311–319. doi:10.1016/j.matdes.2018.06.051
29. Di Fonzo, F.; Casari, C. S.; Russo, V.; Brunella, M. F.; Li Bassi, A.; Bottani, C. E. *Nanotechnology* **2009**, *20*, 015604. doi:10.1088/0957-4484/20/1/015604
30. Michota, A.; Bukowska, J. *J. Raman Spectrosc.* **2003**, *34*, 21–25. doi:10.1002/jrs.928
31. Garai-Ibabe, G.; Grinyte, R.; Golub, E. I.; Canaan, A.; de la Chapelle, M. L.; Marks, R. S.; Pavlov, V. *Biosens. Bioelectron.* **2011**, *30*, 272–275. doi:10.1016/j.bios.2011.09.025
32. Ghidelli, M.; Mascaretti, L.; Bricchi, B. R.; Zapelli, A.; Russo, V.; Casari, C. S.; Li Bassi, A. *Appl. Surf. Sci.* **2018**, *434*, 1064–1073. doi:10.1016/j.apsusc.2017.11.025
33. Doron-Mor, I.; Barkay, Z.; Filip-Granit, N.; Vaskevich, A.; Rubinstein, I. *Chem. Mater.* **2004**, *16*, 3476–3483. doi:10.1021/cm049605a
34. Karakouz, T.; Holder, D.; Goomanovsky, M.; Vaskevich, A.; Rubinstein, I. *Chem. Mater.* **2009**, *21*, 5875–5885. doi:10.1021/cm902676d
35. Tesler, A. B.; Chuntunov, L.; Karakouz, T.; Bendikov, T. A.; Haran, G.; Vaskevich, A.; Rubinstein, I. *J. Phys. Chem. C* **2011**, *115*, 24642–24652. doi:10.1021/jp209114j
36. Gaspar, D.; Pimentel, A. C.; Mateus, T.; Leitão, J. P.; Soares, J.; Falcão, B. P.; Araújo, A.; Vicente, A.; Filonovich, S. A.; Águas, H.; Martins, R.; Ferreira, I. *Sci. Rep.* **2013**, *3*, 1469. doi:10.1038/srep01469
37. Gucciardi, P. G.; de la Chapelle, M. L.; Lidgi-Guigui, N., Eds. *Handbook of Enhanced Spectroscopy*; Jenny Stanford Publishing: 2015. doi:10.1201/b19175
38. Love, J. C.; Estroff, L. A.; Kriebel, J. K.; Nuzzo, R. G.; Whitesides, G. M. *Chem. Rev.* **2005**, *105*, 1103–1170. doi:10.1021/cr0300789
39. Félidj, N.; Aubard, J.; Lévi, G.; Krenn, J. R.; Hohenau, A.; Schider, G.; Leitner, A.; Aussenegg, F. R. *Appl. Phys. Lett.* **2003**, *82*, 3095–3097. doi:10.1063/1.1571979
40. Le Ru, E. C.; Etchegoin, P. G. *MRS Bull.* **2013**, *38*, 631–640. doi:10.1557/mrs.2013.158
41. Ferreira, Q.; Brotas, G.; Alcácer, L.; Morgado, J. Characterization of self-assembled monolayers of thiols on gold using scanning tunneling microscopy. *Proc. Conf. Telecommun. - ConfTele*; 2009; Vol. 1, pp 149–152. <https://www.it.pt/Publications/PaperConference/7432>
42. Minaeva, V. A.; Minaev, B. F.; Hovorun, D. M. *Ukr. Biokhim. Zh.* **2008**, *80*, 82–95. <http://ubj.biochemistry.org.ua/index.php/en/journal-archive/2008/n-4-july-august-34155/480-vibrational-spectra-of-the-steroid-hormones-estradiol-and-estriol-calculated-by-density-functional-theory-the-role-of-low-frequency-vibrations-minaeva-v-a-minaev-b-f-hovorun-d-m>
43. Vedad, J.; Mojica, E.-R. E.; Desamero, R. Z. B. *Vib. Spectrosc.* **2018**, *96*, 93–100. doi:10.1016/j.vibspec.2018.02.011

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