

ACKNOWLEDGMENT

I would like to express my deep gratitude to Dr. K.F. Brown for his encouragement and continued interest in my work. In particular I want to direct my thanks to him for his suggestions and comments in the development of the papers published based on this work.

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My thanks are due to the members of the academic staff, technical staff and fellow postgraduate students, for the pleasant time I have enjoyed

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I am indebted to Mr. K. Murray and M.J. Taylor for their help in construction of some items of equipment used for this work and to Mrs. Anita Auzins, Mrs. Anna Marie Pokorny and Ann Broughton for their help in typing my work.

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Historically pharmacy has been primarily concerned with the forms of administration. Most effort has been directed to making formulations stable and acceptable to the patient with regard to appearance, colour, taste and smell and form of administration. It has become apparent, however, that other factors are important in drug formulation as techniques in pharmacology, medicine and analysis of drugs in the body improved. Emphasis has been directed toward studying the interactions between drug formulations and the organism and this resulted in the establishment of biopharmaceutics as a distinct field within the pharmaceutical sciences.

One of the major biopharmaceutical problems has been absorption in relation to drug formulation. Many very potent drugs have high lipid/water partition coefficients which facilitates their penetration through biological membranes. While this property lowers the biological barrier and favours absorption, it also paradoxically creates formulation problems because such drugs are frequently only very slightly soluble in water.

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It is generally accepted that limited absorption after oral intake is frequently the result of slow dissolution. This kind of absorption problem occurs so frequently that bioavailability testing has become an important part of modern drug and dosage form design and production control. Many examples of differences in bioavailability resulting from differences in dissolution behaviour have been given in the literature (1, 2). Research into dissolution kinetics of drugs is therefore of great importance.

CHAPTER 1

INTRODUCTION

Historically pharmacy has been primarily concerned with the forms in which drugs are administered. Most effort has been directed to making formulations of drugs physically and chemically stable and acceptable to the patient with regard to appearance, colour, taste and smell and form of administration. It has become apparent, however, that other factors are important in drug formulation as techniques in pharmacology, medicine and analysis of drugs in the body improved. Emphasis has been directed toward studying the interactions between drug formulations and the organism and this resulted in the establishment of biopharmaceutics as a distinct field within the pharmaceutical sciences.

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The numerous reviews published confirm the importance of the field (1-5).

The main purpose of most dissolution tests is to *describe* or *quantitate* the dissolution of a compound or dosage form. The dissolution process is most frequently described in the form of a dissolution profile. Although this is the most exact representation it is useful in practice to *summarize and quantitate* the dissolution kinetics in terms of one or more parameters e.g. the so-called dissolution rate constant. In order to do so, it is necessary to decide on some kind of mathematical description.

If the purpose is just to *summarize* dissolution data then a great number of arbitrary *approximating functions* can be used, e.g. polynomials and multiexponential expressions. Although the original data may be regenerated closely from parameters obtained in this way it is generally not reliable to make predictions or extrapolate on this basis. Furthermore, in the absence of mechanistic understanding and because of the substantial number of parameters involved (as in the case of polynomials), it is not possible to characterize the dissolution in a meaningful way. The use of arbitrary approximating functions is therefore of limited use. It is thus more meaningful to use a *mathematical model* which has some mechanistic significance at least conceptually and which adequately describes the dissolution behaviour. The extent to which the *true* mechanism of the system should be approximated by the mathematical model is a somewhat philosophic question, which depends on the purpose of the investigation. For practicability, the model chosen must involve a compromise between its correctness and its simplicity.

Dissolution is a heterogeneous process which is virtually impossible to describe rigorously because of the complexity of the mass transport phenomenon in an agitated heterogeneous system. It is apparent

from Chapter 4 that even when the dissolution mechanism of a free falling single spherical particle is analysed, using several simplifying assumptions, a "rigorous" mathematical model becomes too complex to be of practical significance. The description of the kinetics must therefore rely on a simpler model. As shown in Chapter 4 several such models can be derived on the basis of various assumptions about the mass-transport mechanism in the interfacial region.

The model for a *multiparticulate* system must be based on a model for the single particle dissolution behaviour together with a model for the particle size distribution.

Several *multiparticulate* dissolution models may be postulated. A decision about the model which best characterises the dissolution behaviour must be a compromise between the extent of experimental verification, its simplicity, and whether it is mechanistically meaningful. It may be possible to postulate very simple models which fit a particular set of dissolution data very well. However, oversimplification of the dissolution kinetics may lead to a lack of generality or flexibility. For example it may be possible to derive simple models for the dissolution of monodisperse systems. However, these will usually not fit dissolution data from polydisperse systems adequately; whereas more complex models for polydisperse systems may describe monodisperse systems as a special case.

The evaluation of how well a model fits dissolution data must rely on regression analysis. If the model is linear or can be transformed to linear form then the curve fitting procedure is generally simple. However, the *multiparticulate* dissolution models for polydisperse systems derived in this thesis are of a nonlinear form which can only be fitted and analysed

properly using a nonlinear regression computer program. As described in Chapter 9 several such programs are available. However, none of these programs are written for interactive time sharing and dedicated to mathematical modeling. The program FUNFIT described in Chapter 8 has been developed with these features. It is a general nonlinear regression program which can be used in any branch of science.

The various apparatus used for dissolution testing have been reviewed by Hersey (6), who classified them according to the type of agitation. Methods for Studying Dissolution Behaviour *in vitro*.

The large number of dissolution rate measurement procedures and apparatus which have been described indicates that a universally acceptable method has not yet been developed. The many different techniques have been well reviewed by Hersey (6), Wagner (5) and Braun & Walker (7) and more recently by Hersey and Marty (8) and Groves (9).

The objectives of various tests are often very different. One worker may be interested in the fundamental dissolution behaviour, another in the effect of agitation and type of vehicle flow, or perhaps in the thermodynamics of the dissolution process, thus many kinds of dissolution apparatus are adapted to the study of fundamental principles of the process as for example the rotating-disc method, where the experimental conditions are better defined in terms of surface area of drug and agitation. Others are aimed at investigating dissolution behaviour of dosage forms *in vitro* to provide an estimate of their behaviour *in vivo*. The goal in these cases is to establish an *in vitro* - *in vivo* correlation such that a particular drug can be screened *in vitro* to provide an index of its expected behaviour *in vivo*. This requires that the composition of the drug formulation, the materials used and the production techniques are constant. Prediction of *in vivo* performance from *in vitro* data cannot be guaranteed but is only

intended to enable screening of dosage forms more readily and economically than *in vivo* procedures. Other reasons for performing dissolution tests include development purposes to guide the pharmaceutical formulator in the preparation of optimum dosage forms of drugs for clinical trial or for control purposes to ensure that a given pharmaceutical product is essentially uniform from lot to lot.

The various apparatus used for dissolution testing have been reviewed by Hersey (6), who classified them according to the type of agitation (free and forced convection) and the existence of sink or non-sink conditions.

In the past, the term sink conditions has most frequently referred to the situation where the concentration of dissolved drug is kept small, of the order of about 10% of the solubility, such that re-deposition of dissolved material onto the dissolving solid is considered negligible. This is somewhat vague and imprecise. In Chapter 4 of this work the sink condition is defined in more precise mathematical terms.

The following section of this chapter is not intended to be a comprehensive review of apparatus used in dissolution testing, this has already been done by several authors (5-9), but is an attempt to outline briefly the basic principles of their design and discuss their advantages and limitations.

#### Apparatus with non-sink conditions and constant vehicle volume.

Most apparatus described in the literature belongs to this group because of the simplicity of design and operation. They are based on a constant volume and differ only in the shape of the dissolution container, the way agitation is supplied or in the position of the

dissolving sample. One of the most widely used techniques is the "beaker method" often credited to Levy & Hayes (11) although it was used earlier by Parrott & coworkers (12). It has been used widely for fundamental dissolution rate studies (13, 14) and in modified form for dissolution studies by Shefter & Higuchi (15) and other workers.

The rotating disc apparatus consists of a disc holder to which a tablet or compressed disc of the pure drug is fixed and rotated at suitable constant speed in a round bottomed flask. The method, proposed by Levy & Sahli (19), has been used chiefly for studies of intrinsic dissolution rate and studies concerned with heterogeneous reaction kinetics, diffusion layer theory, effect of agitation on dissolution rate and other fundamental problems and has therefore found wide use (20-22). In interpreting the results of this apparatus it is important to consider that the enormous pressures (for example 50,000 psi on a 0.5 inch diameter disc) often used to make these discs from pure drug powder may introduce changes in crystal form and consequently alter the physical properties of the drug. Clevely & Williams (23) reported that grinding of crystalline barbituric acid derivatives may produce changes in polymorphic form.

The static disc technique employs forced convection in the dissolution liquid and thus has less defined agitation than the rotating disc. Other disc methods include the solvometer technique (24, 25) and the hanging pellet method (13).

There are several disadvantages of the methods mentioned above. They will be discussed under following headings: (1) non-sink conditions, (2) agitation, (3) sampling, (4) introduction of drug sample.

### Non-sink conditions

If the object of the dissolution test is to establish an *in vitro* - *in vivo* correlation or to provide a rough guide to the drug release rate of a dosage form *in vivo* then the methods above are of limited value according to Gibaldi & Feldman (26). These authors stated that unless sink conditions are maintained, *in vitro* results will bear little relationship to *in vivo* observations, for drugs that show dissolution rate limited absorption.

Sampling. Dissolution testing is obviously most relevant for those drugs which represent the greatest dissolution problems. In general such drugs are the least soluble, which means it will frequently be necessary to use exceedingly large volumes of solvent to follow dissolution behaviour for more than just a small fraction of the drug sample used. For example, to follow the dissolution of a tablet containing 5 mg glibenclamide to completion in 0.1 M HCl, it is theoretically necessary to use 500 ml solvent (solubility of glibenclamide is 0.5 mg/100 ml). Dissolution will be very slow, however, because the process slow continuously as saturation is approached. In practice it is usually not convenient to exceed about 20% of saturation. Thus in the example above 2.5 l of solvent would be necessary. This is a rather inconvenient and unwieldy volume for handling and maintaining proper agitation and temperature control.

### Agitation

The large volume of solvent required for apparatus operating under non-sink conditions causes problems in the maintenance of suitable agitation. If the drug is in the form of a powder or disintegrating dosage form a high degree of agitation will be required to suspend the drug



particles in the solvent so that they are all exposed to similar conditions of agitation. On the basis of *in vivo* data Hamlin *et.al.* (27) indicated that only low agitation rates could adequately differentiate rates of release from solid dosage forms. These authors studied the rates of dissolution of two polymorphic forms of methylprednisolone and found that sensitivity in distinguishing between the rates decreased at higher agitation intensities. This confirms the importance of using a dissolution apparatus where agitation is well defined and can be varied over a wide range.

### Sampling.

Several problems are encountered with respect to sampling in the types of apparatus mentioned above. It is impossible to sample without disturbing the dissolution process to some extent. Sampling affects solvent volume, which in turn may affect agitation conditions. Replacement of removed solvent is not an entirely satisfactory solution to the problem because it results in a discontinuity or small drop in the concentration of dissolved drug. The agitation conditions may change during additions of replacement liquid and affect the homogeneity of the system by creating "pockets" of fresh solvent which mix only slowly with the bulk. The latter is particularly important under conditions of low agitation. The most serious problem in sampling these systems seems to be the fact that the most precise and detailed characterisation of the dissolution process requires the greatest frequency of sampling, which compounds the errors due to the sampling procedure. The time taken for sampling is difficult to define precisely. In most cases it is considered to be instantaneous rather than a time interval. Such errors may be quite substantial at the initial stages of the experiment, where the concentration is changing very rapidly.

### Introduction of drug sample

It has been suggested that only a small amount of solid sample should be used for a complete dissolution test if the solubility is low. Furthermore, slightly soluble drugs are often ground finely or micronised which increases their surface-free energy so that particles may adhere and be difficult to disperse evenly in the solvent. They frequently adhere to the side of the dissolution container and a significant fraction of the powder may be washed up the side of the container where it is no longer exposed to the vehicle. The powder may also aggregate and float on the surface.

It can thus be seen that the most widely used type of dissolution apparatus (non-sink, constant, vehicle volume) has several serious design disadvantages which limits its usefulness in fundamental dissolution kinetics studies.

### Dissolution apparatus with non-sink conditions, constant vehicle volume and automatic recording

Schroeter & Wagner (28) appear to be the first to describe an automatic recording dissolution apparatus. They combined a beaker type apparatus with a peristaltic pump. Filtered solvent was pumped to an automatic recording spectrophotometer and returned to the beaker. This procedure has several advantages. It provides unlimited data points, there is no discontinuity in the system due to sampling and the number of sources of error are reduced. Furthermore, it is fast and convenient.

Among the disadvantages are the facts that particles may be trapped in the filter system, the concentration range is limited by the spectrophotometer; however variable path length absorption cells can extend

the range. The system is only useful where there is little background absorption from solvent or excipients. The problems associated with the beaker method including introduction of sample, agitation, solvent volume, non-sink conditions are still present.

The technique described by Baum & Walker (7) represents a considerable improvement in the method used by Schroeter & Wagner. Agitation is performed by the solvent as it flows through a column type dissolution chamber bounded at both ends by mesh screens. Solvent is recycled continuously through the column via a beaker or a flask. Sampling can be done from a collection reservoir or better by passing the filtered solvent through an automatic recording spectrophotometer. This system has several advantages compared to that of Schroeter & Wagner. Firstly, the solvent flow or agitation experienced by the particles is better defined and more easily adjustable. Entrapment of particles by the screen or filter should have little effect since they will still be exposed to almost the same solvent flow. Wetting and dispersion problems are reduced. In addition much larger solvent volumes can be handled readily so that sink conditions can be approached without increasing agitation rate. This system could be improved by altering to a non-recycled open system.

Dissolution apparatus with sink conditions: non-recycled open systems.

These types of apparatus include those most recently developed and represent the most suitable apparatus available for studies of dissolution kinetics. They consist essentially of a dissolution vessel with continuous input of fresh solvent and output of filtered solvent containing dissolved drug. The concentration of drug can be continuously monitored by some automated analytical procedure or solution sample may be

collected in fractions and assayed separately. The various methods usually differ only in the design of the dissolution vessel and the mode of agitation, which may be produced either by the solvent flow or other means. The most common flow-through dissolution cells are cylindrical, with filters at both ends to enclose the sample and use the solvent flow as the only source of agitation (29-32).

Lapidus & Lordi (33) described a flow-cell which included a holder for compressed drug discs or tablets, enabling drug release measurements under sink conditions from a constant surface area. Their method represents a valuable alternative to the rotating disc technique for studying dissolution kinetics of pure drugs.

There are many advantages to using the continuous-flow column type apparatus. It is a flexible system that permits changes to be made readily, even within a test run, of important factors such as temperature, flow rate and vehicle composition including pH, viscosity, drug concentration and surfactant concentration.

Data obtained using a continuous recording technique with this system are in differential form. Thus a direct recording is made of the variation of release rate with time. This is superior to integral data (cumulative amounts dissolved with time) which tends to obscure small changes in dissolution rate particularly if only relatively few fractions are taken. A continuous recording yields unlimited data points which enable very precise characterisation of the dissolution behaviour. For this reason, it is also suitable for automatic data processing (34). Unlike most other systems, in particular the beaker method, sampling does not influence the dissolution process.

Normally the problem of wetting is minimal. However, when necessary

wetting can be accomplished by introduction of a surfactant solution for a short period initially and allowing it to be washed away immediately by fresh solvent (30).

#### EXPERIMENTAL

The dissolution process can be followed to completion provided sufficient solvent is available and that the analytical method is sufficiently sensitive. All particles of a powder experience essentially the same intensity of solvent flow, including those particles which collect on the filters or screen at the ends of the column, thus agitation conditions are related to the solvent flow rate in a meaningful way and the latter is easy to define and control. Complete sink condition can be approximated very well. Thus results are more reproducible and instantaneous initial drug release measurements can be made, in contrast to most other methods. Furthermore, the continuous flow method is fast, convenient and very suitable for routine dissolution tests.

It is apparent that the continuous flow dissolution cell apparatus is a powerful and versatile tool in studies of dissolution kinetics.

The high precision dissolution apparatus described in Chapter 2 belongs to this category.

Flexible polyethylene tubing (i.d., 0.35 cm) was used throughout with the exceptions of that used in the pump, which was a silicon tubing (silastic, Dow Corning, i.d., 0.335 cm and o.d., 0.465 cm). It is an open tube type meter that monitors the pressure governing the flow rate of liquid.

#### The Dissolution Cell

Figure 2.2 shows a detailed diagram of the dissolution cell constructed for this work. Powder to be investigated is spread in a thin evenly distributed layer,  $c$ , in a sandwich-like arrangement between two

CHAPTER 2

EXPERIMENTAL

Dissolution Apparatus

A diagram of the continuous-flow recording dissolution apparatus is shown in Fig. 2.1.  $R_1$  and  $R_2$  are 20 litre reservoirs containing the dissolution media. P is a peristaltic pump (MHRE Watson-Marlow Ltd., England) transporting the liquid from  $R_1$  or  $R_2$  through a heat exchanger E, which adjusts the liquid to the required temperature before it reaches the dissolution cell D. This is immersed in a water filled, jacketed beaker, B, maintained at the same temperature as the dissolution liquid leaving E. Liquid from the dissolution cell, D, passes through a flow-cell, F, in the spectrophotometer, S, (Perkin-Elmer 124) which was fitted with chartrecorder, R, and finally accumulates in the container, C.  $V_1$  and  $V_2$  are two-way valves which enable by-pass of the dissolution cell, D, for zero line adjustment of the spectrophotometer with blank liquid from the reservoir.  $V_3$  is a similar valve by means of which liquid can be drawn from either reservoir  $R_1$  or  $R_2$ . Flexible polyethylene tubing (i.d., 0.35 cm) was used throughout with the exceptions of that used in the pump, which was a silicon tubing (silastic, Dow Corning, i.d., 0.335 cm and o.d., 0.465 cm). L is an open tube type meter that monitors the pressure governing the flow rate of liquid.

The Dissolution Cell

Figure 2.2 shows a detailed diagram of the dissolution cell constructed for this work. Powder to be investigated is spread in a thin evenly distributed layer, c, in a sandwich-like arrangement between two



Figure 2.1 Diagram of continuous-flow dissolution apparatus:  $R_1$  and  $R_2$  are reservoirs containing the dissolution liquid; P, peristaltic pump; E, head exchanger (Fig. 2.3); D dissolution cell (Fig. 2.2); B, water filled jacketed beaker; F, spectrophotometer flow-cell (Fig. 2.5); S, spectrophotometer with reorder R.  $V_1$ ,  $V_2$  and  $V_3$  are two way valves; C, collection vessel; T, thermostat; L, pressure meter.

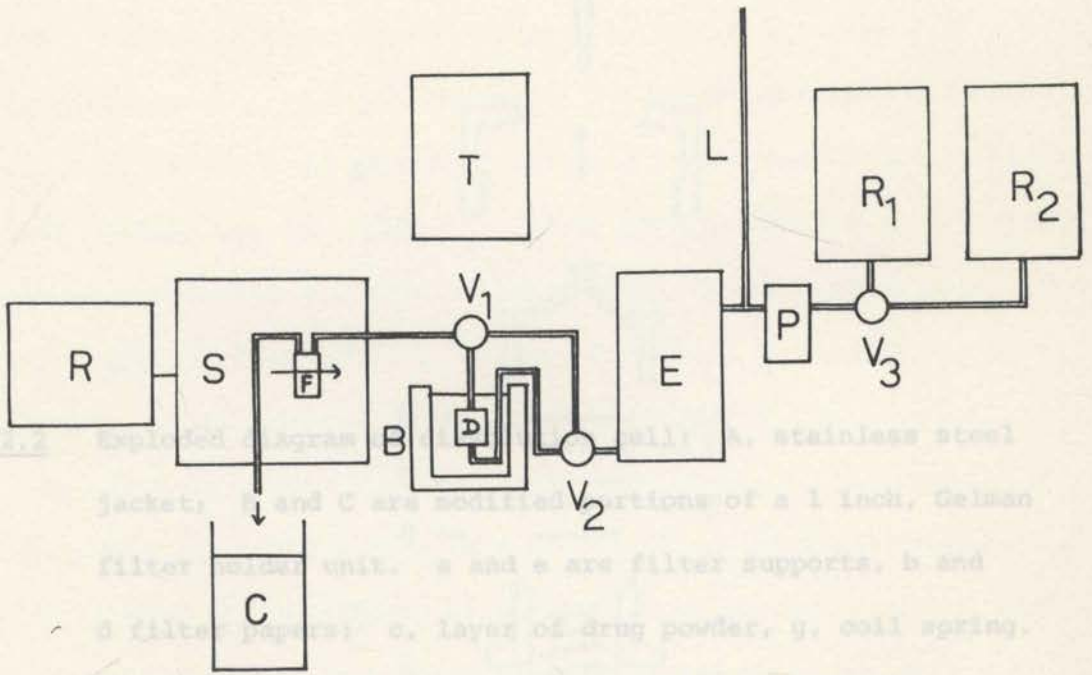


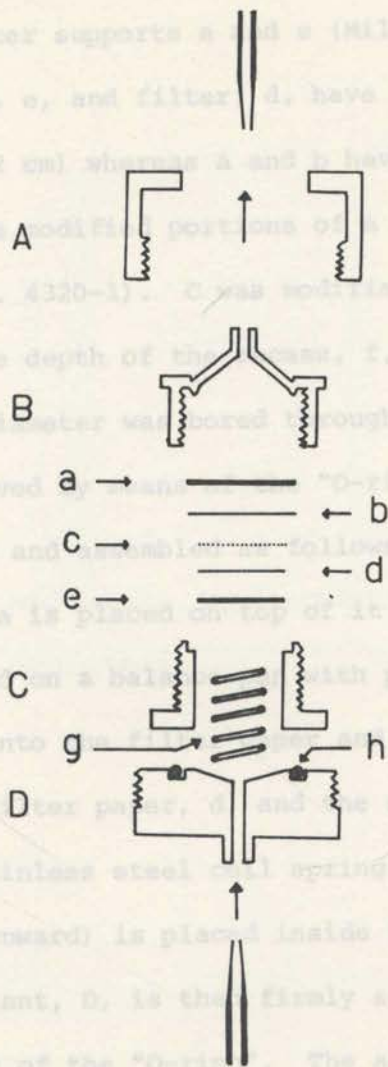
Figure 3.2 Exploded diagram of a pharmaceutical filtration system. A, stainless steel jacket; B and C are modified portions of a 1 inch, Gelman filter unit. s and e are filter supports, b and d filter papers; c, layer of drug powder, y, coil spring, D, reflex fitting which screws into A; h, "o-ring".





Figure 2.2 Exploded diagram of dissolution cell: A, stainless steel jacket; B and C are modified portions of a 1 inch, Gelman filter holder unit. a and e are filter supports, b and d filter papers; c, layer of drug powder, g, coil spring. D, teflon fitting which screws into A; h, "o-ring".

paper filters b and c (Whatman quantitative filter paper) supported on both sides by stainless steel filter supports a and d (Millipore cat. no. XX 3002503). Filter support, e, and filter d, have a diameter equal to the inside diameter of C (2.2 cm) whereas a and b have the same diameter as the recess f. B and C are modified portions of a 1 inch Gelsman, filter holder, unit (Gelsman cat. no. 4320-1). C was modified from the commercial filter unit by increasing the depth of the recess, f, to contain both a and b. In addition a hole, 2.2 cm diameter was bored through the centre. A water tight pressure seal is achieved by means of an "O-ring", h.



The cell is loaded and assembled as follows: Filter paper b is positioned in the recess f, a is placed on top of it and B and C screwed together. The unit is placed on a balance with portion B as the base. Powder is weighed directly onto the filter paper b distributed as a thin, uniform layer. The second filter paper, c and the support, e, are placed on top together with the stainless steel filter support, d. The complete B - C unit (with B still downward) is placed inside the stainless steel jacket, A. The Teflon component, D, is then firmly screwed to A to form a tight seal with C by means of the "O-ring", h. The assembled unit is placed upright in a tripod stand with the hoses connected at both ends and the whole is immersed in the jacketed beaker (B on Fig. 2.1) for thermal equilibration 10 minutes before any run.

Heat Exchange

The equipment for temperature regulation was also designed specially for this work, to overcome the problems of maintaining large volumes of dissolution fluid at constant temperature. A diagram is shown in Fig. 2.1. It consists of a cylindrical PVC container 40 cm in depth with an

paper filters b and d (Whatman quantitative filter paper) supported on both sides by stainless steel filter supports a and e (Millipore cat. no. XX 3002503). Filter support, e, and filter, d, have a diameter equal to the inside diameter of C (2.2 cm) whereas a and b have the same diameter as the recess f. B and C are modified portions of a 1 inch Gelman, filter holder, unit (Gelman cat. no. 4320-1). C was modified from the commercial filter unit by increasing the depth of the recess, f, to contain both a and b. In addition a hole, 2.2 cm diameter was bored through the centre. A water tight pressure seal is achieved by means of the "O-ring", h.

The cell is loaded and assembled as follows: Filter paper b is positioned in the recess f, a is placed on top of it and B and C screwed together. The unit is placed on a balance pan with portion B as the base. Powder is weighed directly onto the filter paper and distributed as a thin, uniform layer. The second filter paper, d, and the support, e, are placed on top together with the stainless steel coil spring, g. The complete B - C unit (with B still downward) is placed inside the stainless steel jacket, A. The Teflon component, D, is then firmly screwed to A to form a tight seal with C by means of the "O-ring". The assembled unit is placed upright in a tripod stand with the hoses connected at both ends and the whole is immersed in the jacketed beaker (B on Fig. 2.1) for thermal equilibration 10 minutes before any run.

#### Heat Exchanger

The equipment for temperature regulation was also designed specially for this work, to overcome the problems of maintaining large volumes of dissolution fluid at constant temperature. A diagram is shown in Fig. 2.3. It consists of a cylindrical PVC container 40 cm in depth with an

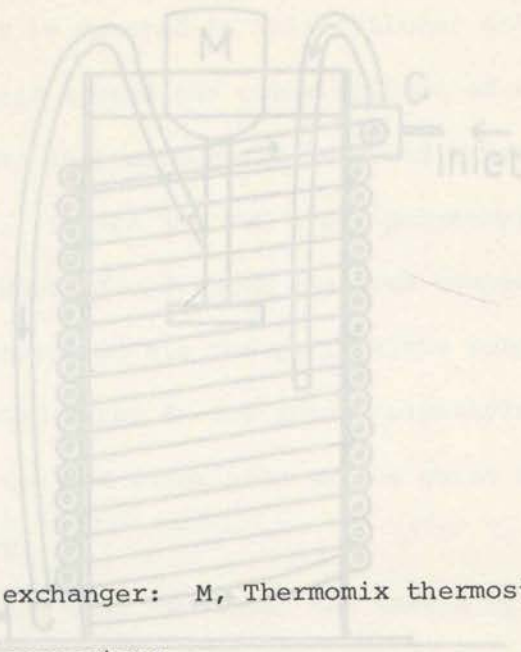
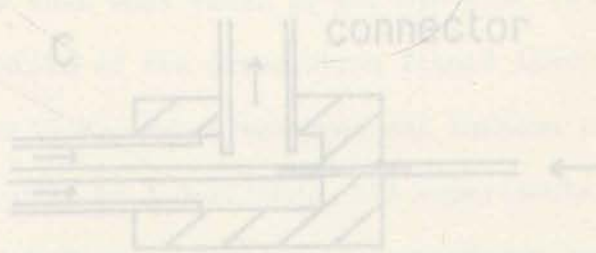
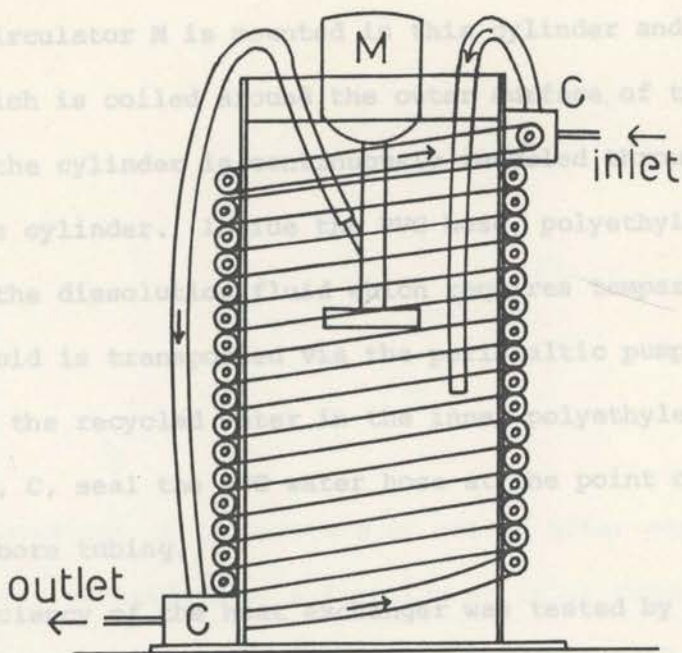


Figure 2.3 Diagram of heat exchanger: M, Thermomix thermostat water circulator; C, connectors.

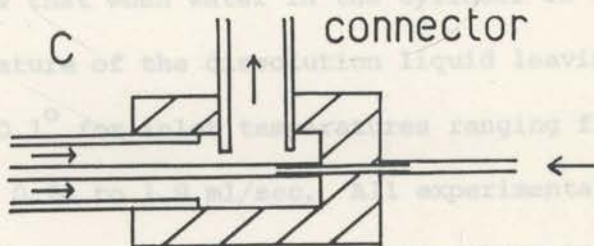


inside diameter of 1.5 cm filled with distilled water. A Thermo-  
 thermostat water circulator M is connected to the cylinder and connected to  
 a soft PVC hose which is coiled around the cylinder. Heated water from the cylinder  
 and returned to the cylinder. polyethylenes tubing is threaded carrying the dissolution  
 The dissolution fluid is transferred by a peristaltic pump and flows  
 counter-current to the recycled water through the polyethylene hose.  
 Special connectors, C, seal the point of entry and exit of the small bore tubing



The efficiency of the heat exchanger was tested by passing water  
 of varying initial temperature through the apparatus at two flow rates and  
 determining the temperature of the outlet. The results are graphed in

Fig. 2.4. These data show that when water in the cylinder is maintained  
 at  $37.7 \pm 0.1^\circ$  the temperature of the dissolution liquid leaving the  
 heat exchanger is  $37.5 \pm 0.1^\circ$  for flow rates ranging from between  
 $5^\circ$  and  $30^\circ$  and flow rates



carried out within these boundary conditions of temperature and flow rate.  
 Equilibration of temperature was achieved within 10 minutes irrespective  
 of starting temperature.

The heat exchanger is a very convenient piece of apparatus  
 since it is highly efficient and eliminates the practical difficulties  
 associated with maintaining large volumes of dissolution medium at  
 constant temperature.

An inexpensive flow-cell for the spectrophotometer was also  
 designed and is shown in schematic form in Fig. 2.5. It consists of a  
 standard rectangular quartz cell of 1 cm path length to which is fitted

inside diameter of 15.5 cm filled with distilled water. A Thermomix thermostat water circulator M is mounted in this cylinder and connected to a soft PVC hose which is coiled around the outer surface of the cylinder. Heated water from the cylinder is continuously recycled through the hose and returned to the cylinder. Inside the PVC hose, polyethylene tubing is threaded carrying the dissolution fluid which requires temperature regulation. The dissolution fluid is transported via the peristaltic pump and flows counter-current to the recycled water in the inner polyethylene hose. Special connectors, C, seal the PVC water hose at the point of entry and exit of the small bore tubing.

The efficiency of the heat exchanger was tested by passing water of varying initial temperature through the apparatus at two flow rates and determining the temperature of the outlet. The results are graphed in Fig. 2.4. These data show that when water in the cylinder is maintained at  $37.7 \pm 0.1^{\circ}$  the temperature of the dissolution liquid leaving the heat exchanger is  $37.5 \pm 0.1^{\circ}$  for inlet temperatures ranging from between  $5^{\circ}$  and  $30^{\circ}$  and flow rates 0.24 to 1.9 ml/sec. All experimental work was carried out within these boundary conditions of temperature and flow rate. Equilibration of temperature was achieved within 10 minutes irrespective of starting temperature.

The heat exchanger is a very convenient piece of apparatus since it is highly efficient and eliminates the practical difficulties associated with maintaining large volumes of dissolution medium of constant temperature.

An inexpensive flow-cell for the spectrophotometer was also designed and is shown in schematic form in Fig. 2.5. It consists of a standard rectangular quartz cell of 1 cm path length to which is fitted

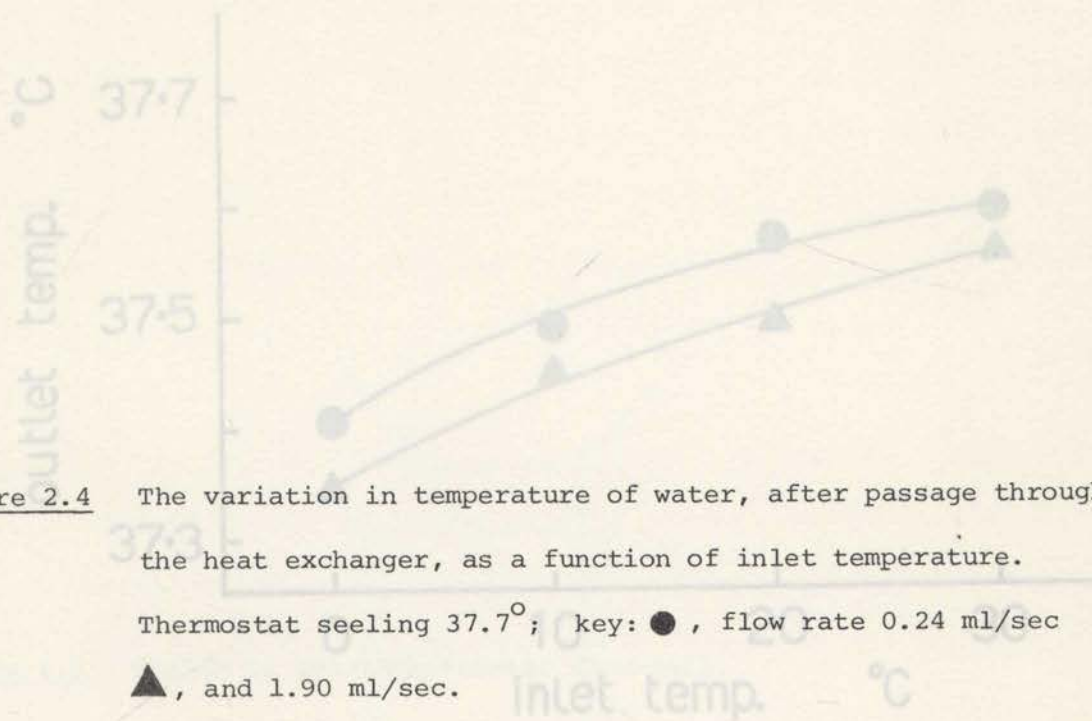


Figure 2.4 The variation in temperature of water, after passage through the heat exchanger, as a function of inlet temperature.

Thermostat setting  $37.7^{\circ}$ ; key: ●, flow rate 0.24 ml/sec ▲, and 1.90 ml/sec.

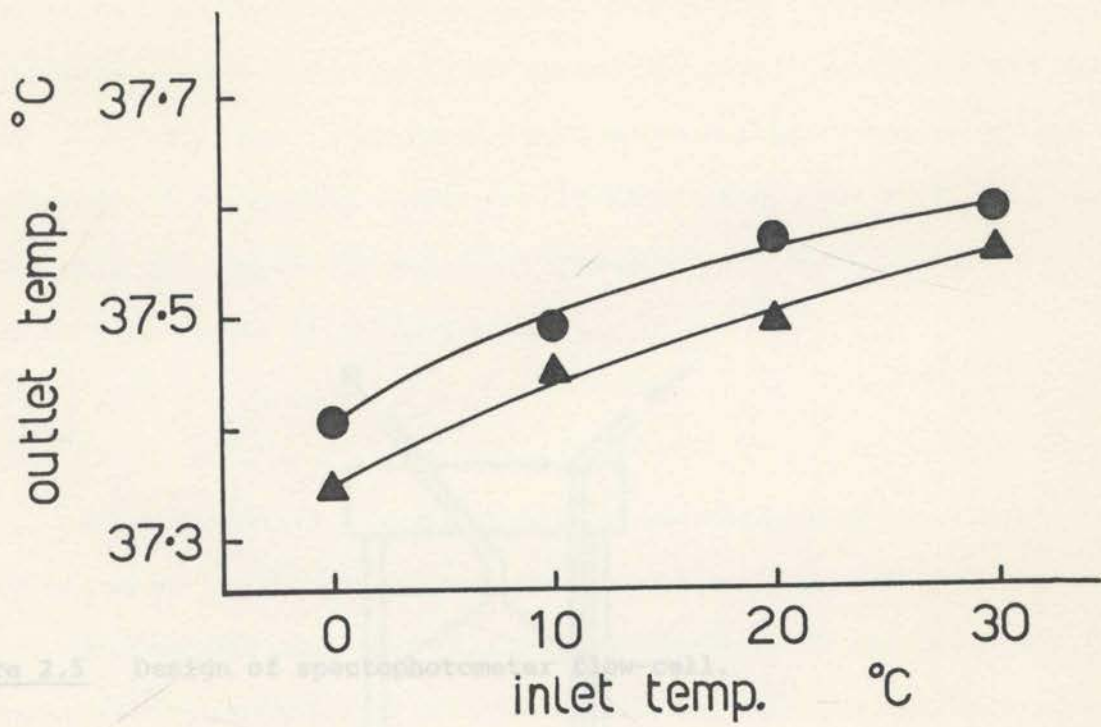


Figure 2.5 Design of spectrophotometer



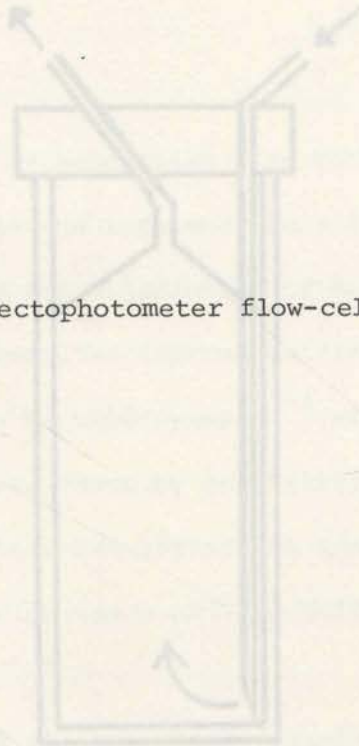
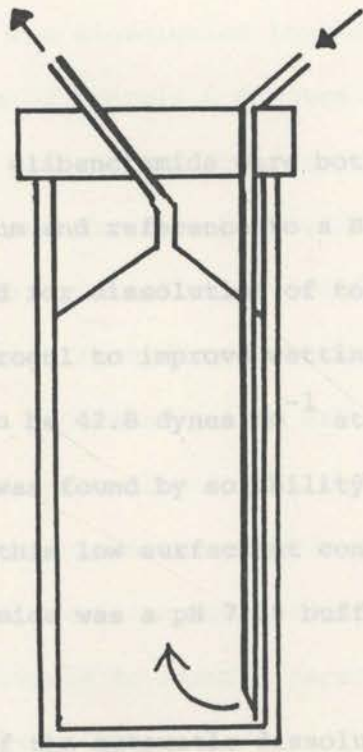


Figure 2.5 Design of spectrophotometer flow-cell.

a specially designed rectangular teflon stopper with silicon rubber seal. A narrow bore stainless steel tube passes through the stopper and extends from one side of the cell. This tube is adjustable in length and does not obstruct the light path. Dissolution medium containing dissolved solute is introduced to the bottom of the cell by this means. The solution flows upward across the light path and exits through a cone shaped outlet which assists in the removal of gas bubbles.

Analytical

Tolbutamide and glibenclamide were both determined by ultraviolet spectrophotometry at 230 m $\mu$  and reference to a Beer-law curve. The vehicle used for dissolution of tolbutamide was 0.1 N HCL containing  $10^{-5}$  M cetomacrogol to improve wetting. The surface tension of this solution was found to be 42.8 dynes/cm<sup>2</sup> at 37<sup>o</sup> (mean of 10 measurements with a.d. 3%). It was found by solubility measurements, that no solubilization occurs at this low surface concentration. The dissolution liquid used for glibenclamide was a pH 7 buffer, I = 0.05, containing  $10^{-5}$  M cetomacrogol.



The operation of the dissolution apparatus (Fig. 2.1) is self-explanatory from the previous description. Values of absorbance were read from the chart recording at 2.5 minute intervals from the beginning of the experiment. These values together with the volumetric flow rate, the Beer-law constant, the initial amount used, the amount of undissolved drug remaining in the dissolution cell at the end of the experiment and a code for the particular data treatment desired were fed into a digital computer and processed according to a FORTRAN program. The amount of undissolved drug remaining was found by disconnecting the

a specially designed rectangular teflon stopper with silicon rubber seal. A narrow bore stainless steel tube passes through the stopper and extends from one side of the cell. This tube is adjustable in length and does not obstruct the light path. Dissolution medium containing dissolved solute is introduced to the bottom of the cell by this means. The solution flows upward across the light path and exits through a cone shaped outlet which assists in the removal of gas bubbles.

Analytical

Tolbutamide and glibenclamide were both determined by ultraviolet spectrophotometry at 220 nm and reference to a Beer-Law curve.

2.6. The vehicle used for dissolution of tolbutamide was 0.1 M HCL containing  $10^{-5}$  M cetomacrogol to improve wetting. The surface tension of this solution was found to be  $42.8 \text{ dynes cm}^{-1}$  at  $37^{\circ}$  (mean of 10 measurements with s.d. 5%). It was found by solubility measurements, that no solubilization occurs at this low surfactant concentration. The dissolution liquid used for glibenclamide was a pH 7.25 buffer,  $I = 0.05$ , containing  $10^{-5}$  M cetomacrogol.

The operation of the automatic dissolution apparatus (Fig. 2.1) is self-explanatory from the previous description. Values of absorbance were read from the chart recording at 2.5 minutes intervals from the beginning of the experiment. These values together with the volumetric flow rate, the Beer-law constant, the initial amount used, the amount of undissolved drug remaining in the dissolution cell at the end of the experiment and a code for the particular data treatment desired were fed into a digital computer and processed according to a FORTRAN program. The amount of undissolved drug remaining was found by disconnecting the

dissolution cell and transferring the filter paper/drug/filter paper "sandwich" to a volumetric flask. The drug was then dissolved in 95% ethanol and the solution assayed. This procedure increases the accuracy of the dissolution curves generated by the computer because it enables corrections to be made on the principle of mass balance.

A test of this accuracy was made using 12.5 mg 60/85 mesh fraction of tolbutamide, a flow rate of 0.149 cm/sec and 0.1 M HCL +  $10^{-5}$  M cetomacrogol as solvent. The dissolution liquid containing dissolved drug was collected at intervals of exactly 5 minutes as it left the spectrophotometer cell (F in Fig 2.1) and assayed for drug. The cube root of the calculated amount undissolved was then plotted versus time as seen in Fig. 2.6. On the same figure are plotted values generated by the computer from the absorbance readings taken from the chart recording. It appears from the plot that the computer generated points are almost coincidental with the points obtained from direct analysis. A chi-square test showed no significant difference between the two methods ( $P > 0.99$ ).

The accuracy and precision obtained using this experimental technique have made it possible to examine aspects of the dissolution kinetics which would be impossible with most other methods.

### Dynamic Dialysis

The data used for the evaluation of the new method of obtaining drug-macromolecule binding parameters described in Chapter 11 was kindly supplied by Dr. M.J. Crooks (35). The dynamic dialysis technique used was that described by Meyer and Guttman (36). Chlorpropamide was dialysed from 1% bovine serum albumin in 0.067 M phosphate buffer pH 7.4 at 37° in the presence of a fixed pre-concentration of warfarin ( $1.6 \times 10^{-5}$  M).

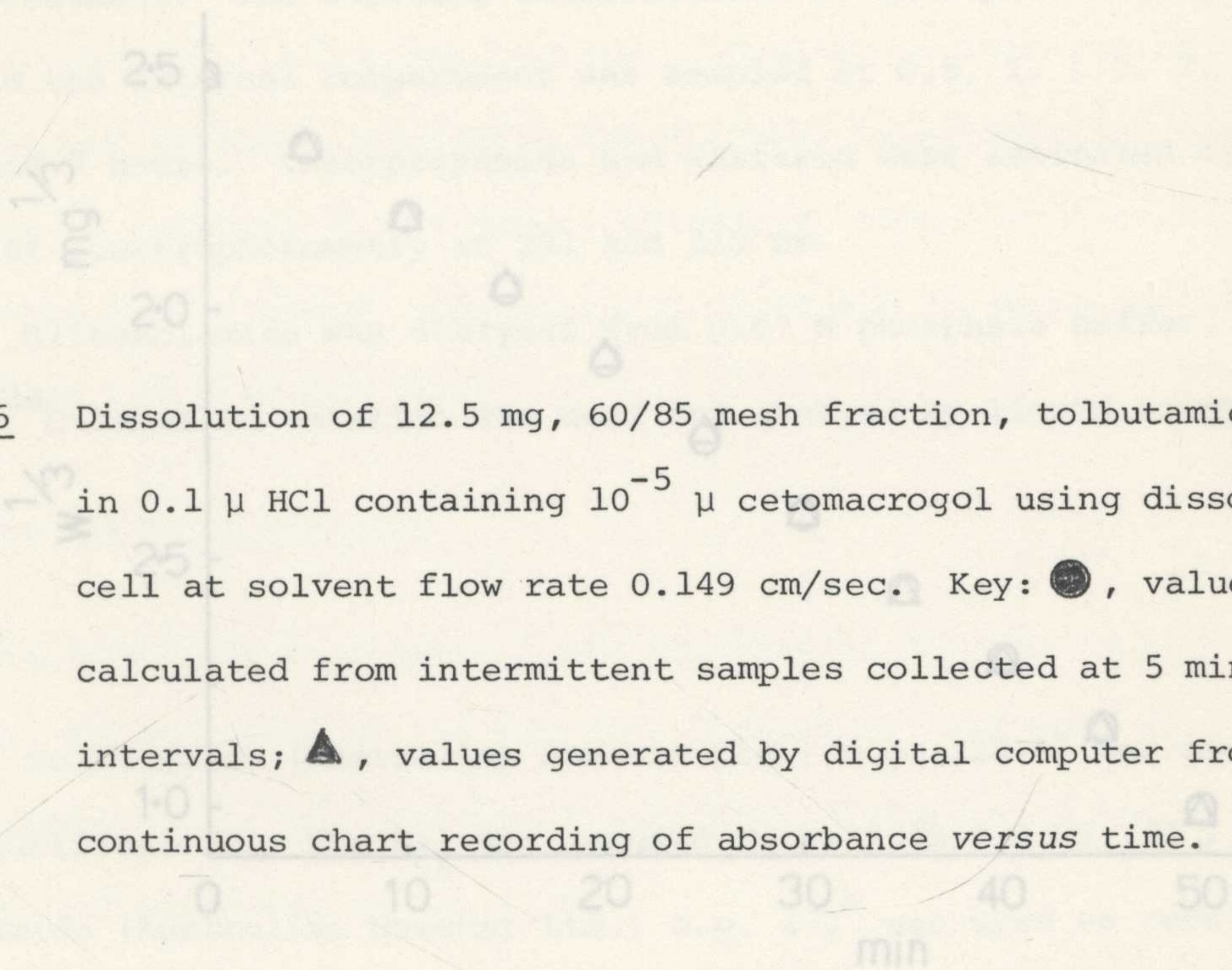
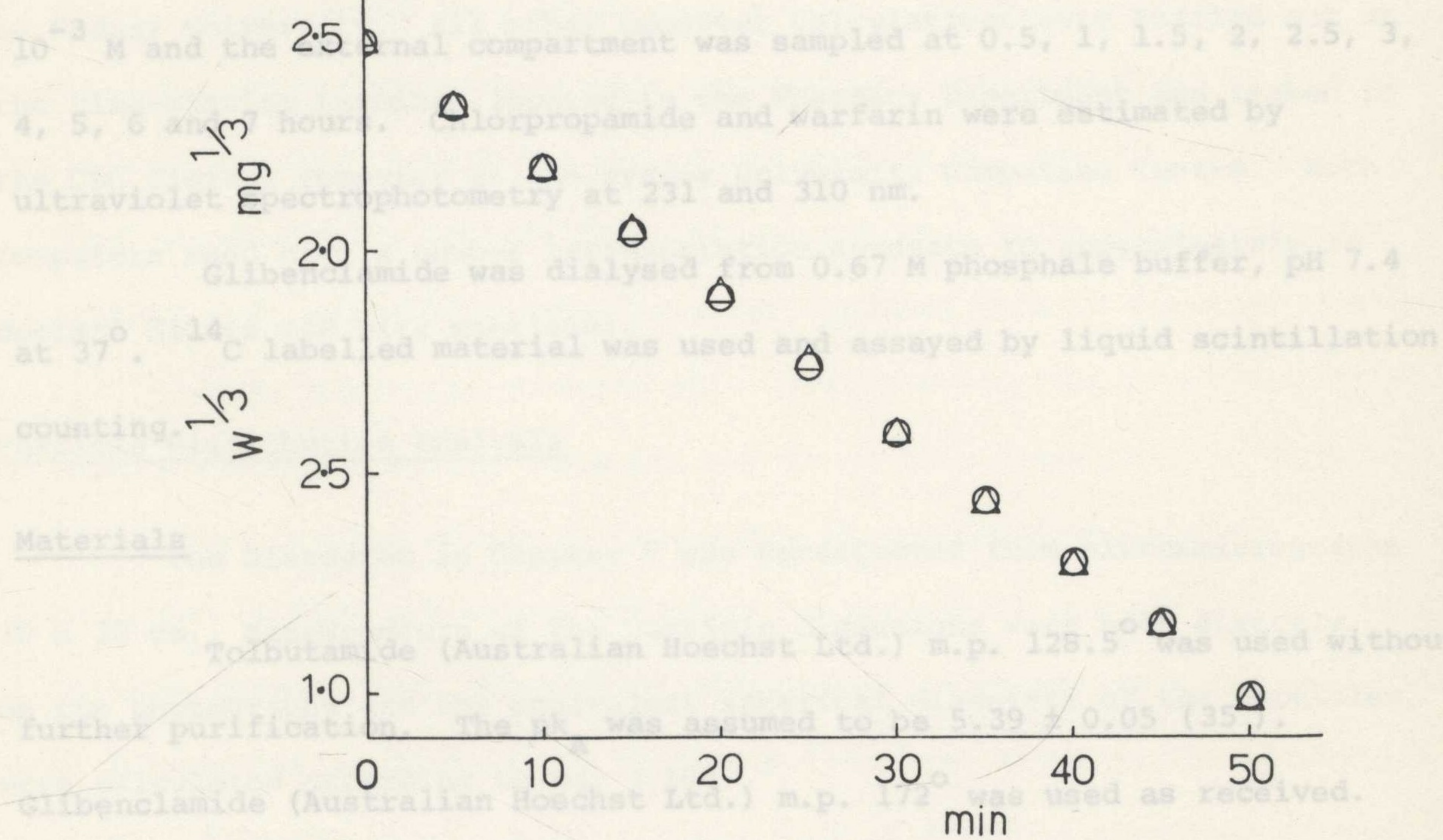


Figure 2.6 Dissolution of 12.5 mg, 60/85 mesh fraction, tolbutamide in 0.1  $\mu$  HCl containing  $10^{-5}$   $\mu$  cetomacrogol using dissolution cell at solvent flow rate 0.149 cm/sec. Key: ●, values calculated from intermittent samples collected at 5 min's intervals; ▲, values generated by digital computer from continuous chart recording of absorbance versus time.

Although warfarin displaces some chlorpropamide, the latter still appears to bind to two classes of sites (37). This system was selected so that such a model of two classes of sites could be appraised by the new method of data treatment. The starting concentration of chlorpromide was  $3.62 \times 10^{-3}$  M and the



Materials

Tolbutamide (Australian Hoechst Ltd.) m.p. 125.5° was used without further purification. The  $pK_a$  was assumed to be  $5.39 \pm 0.05$  (35). Glibenclamide (Australian Hoechst Ltd.) m.p. 172° was used as received. The  $pK_a$  was estimated as  $6.50 \pm 0.05$  (35). Cetomacrogol was obtained from Glovers Chemicals Ltd. The concentration used the surfactant had negligible absorbance at 220 nm.

Data Treatment

The dissolution rate data ( $dw/dt$  vs. time) used for the accuracy test of the dissolution apparatus (Fig. 2.6) were integrated by means of a FORTRAN program (38). The dissolution rate data presented in Chapter 10 and the dialysis rate data in Chapter 11 were analysed according to the respective mathematical models by nonlinear least squares regression techniques, using the FORTRAN program FUNFIT listed in the appendix. The subroutine, MODEL, defining the mathematical models are listed in the

Although warfarin displaces some chlorpromamide, the latter still appears to bind to two classes of sites (37). This system was selected so that such a model of two classes of sites could be appraised by the new method of data treatment. The starting concentration of chlorpromamide was  $3.62 \times 10^{-3}$  M and the external compartment was sampled at 0.5, 1, 1.5, 2, 2.5, 3, 4, 5, 6 and 7 hours. Chlorpromamide and warfarin were estimated by ultraviolet spectrophotometry at 231 and 310 nm.

Glibenclamide was dialysed from 0.67 M phosphate buffer, pH 7.4 at 37°.  $^{14}$ C labelled material was used and assayed by liquid scintillation counting.

### Materials

Tolbutamide (Australian Hoechst Ltd.) m.p. 128.5° was used without further purification. The  $pK_A$  was assumed to be  $5.39 \pm 0.05$  (35). Glibenclamide (Australian Hoechst Ltd.) m.p. 172° was used as received. The  $pK_A$  was estimated as  $6.50 \pm 0.05$  (35). Cetomacrogol was obtained from Glovers Chemicals Ltd. The concentration used the surfactant had negligible absorbance at 220 nm.

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respective chapters.

The calculations and drawings of the theoretical dissolution curves in chapter 6 was done using a CDC digital computer equipped with a calcomp plotter installed at the National Standards Laboratory, C.S.I.R.O., at Sydney University. All other computer calculations were carried out at the time-sharing terminal located in the Pharmacy Department and linked to the CDC digital computer at the Sydney University Computing Centre. Both computers used have a number representation accurate to approximately 14 decimal digits (48 bits mantissa).

#### Particle Distribution Analysis

The histogram in Chapter 7 was constructed from electronmicrographs 30 x 30 cm. Measurements of the particle dimensions were made directly on the photographs and the equivalent spherical diameters of the particles were calculated according to Eq. 4.104.

#### Pharmacokinetic Data

The data used for the comparison of NONLIN and FUNFIT in Chapter 9 was kindly supplied by Mr. A. Somogyi. Although artificially generated data containing random noise just as well could have been used in such a comparison it was felt that the biological data would give a more realistic basis for comparison.



CHAPTER 3

BIOLOGICAL ASPECTS AND METHODS OF ENHANCING DISSOLUTION

Potent drugs very often have a high lipid/water partition coefficient which facilitates their penetration through biological membranes. While this property lowers the biological barrier and favours absorption it also paradoxically creates formulation problems, since these drugs are frequently only slightly soluble in water, causing them to dissolve slowly.

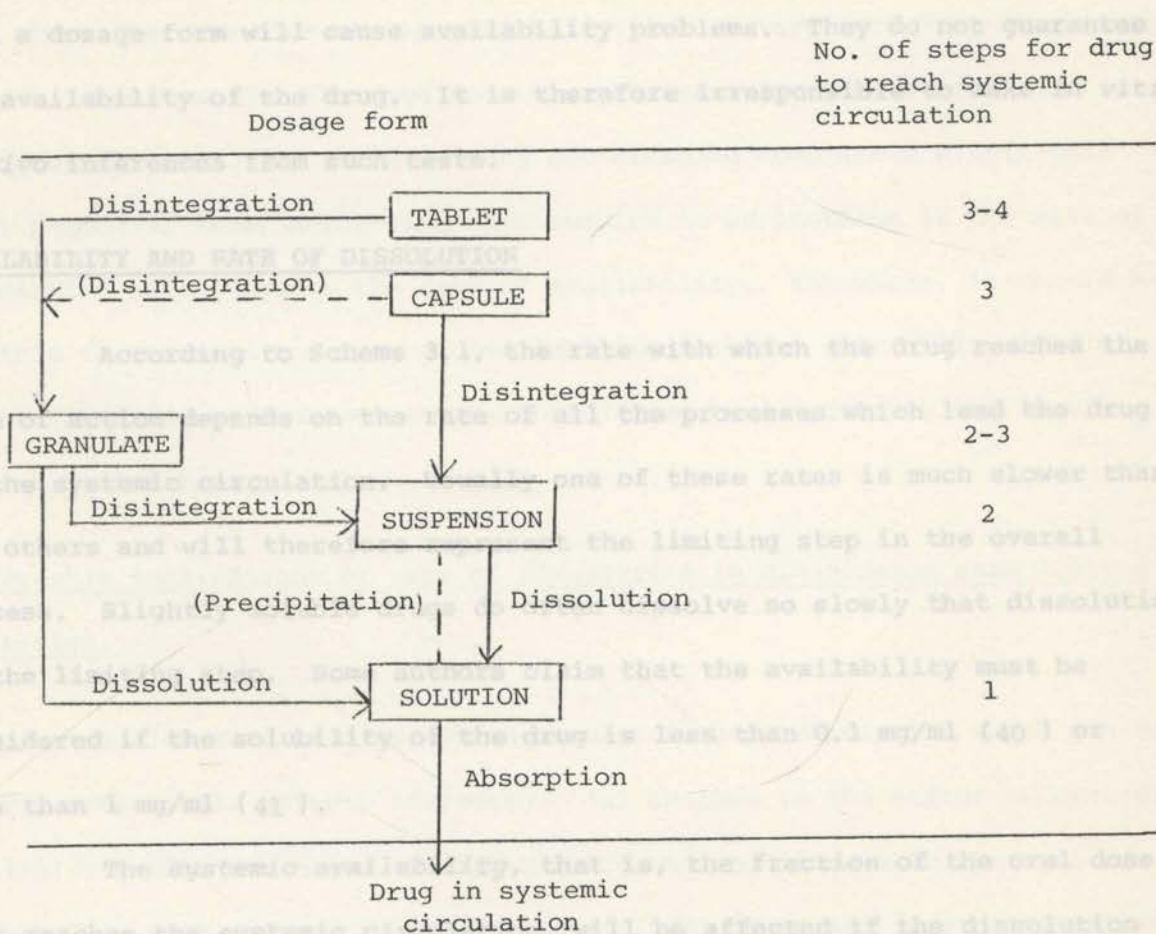
It is generally accepted that limited absorption after oral intake of poorly soluble drugs is often the result of slow dissolution. This kind of absorption problem occurs so frequently that bioavailability and dissolution testing has become important in evaluation and control of slowly dissolving drugs. Many examples of differences in bioavailability resulting from differences in dissolution behaviour have been given in the literature (1, 39). Research into dissolution kinetics of drugs is therefore of great importance and a better understanding of the influence of the formulation on the dissolution process is necessary to overcome these problems. The numerous reviews published recently confirm the importance of the field (2,3-5).

Dissolution as a step in the pathway to systemic circulation.

When a drug is given orally it has to go through a number of steps before it reaches the systemic circulation. Scheme 3.1 illustrates this for various oral dosage forms. It is expected from the number of steps, that the availability of the dosage forms should be of the order solution > suspension > granulate > capsule > tablet.

Scheme 3.1

Pathways to systemic circulation for oral dosage forms



Solid dosage forms are often tested *in vitro* for their disintegration or dissolution behaviour. Such tests essentially provide only information about these processes in relation to the particular experimental conditions. They are primarily useful in evaluating how the disintegration or dissolution is influenced by the drug formulation and by factors such as agitation, pH, vehicle composition etc. and are mainly of value in the development of better formulations.

The aim has often been to use the tests as a measure or indication of how the dosage form will perform *in vivo*. Most conclusions of this nature will however be unreliable because these *in vitro* tests in their very simple

forms cannot properly reflect the rather complex *in vivo* system. The *in vitro* tests are only able to tell in some but not all cases when a dosage form will cause availability problems. They do not guarantee the availability of the drug. It is therefore irresponsible to make *in vitro* - *in vivo* inferences from such tests.

#### AVAILABILITY AND RATE OF DISSOLUTION

According to Scheme 3.1, the rate with which the drug reaches the site of action depends on the rate of all the processes which lead the drug to the systemic circulation. Usually one of these rates is much slower than the others and will therefore represent the limiting step in the overall process. Slightly soluble drugs do often dissolve so slowly that dissolution is the limiting step. Some authors claim that the availability must be considered if the solubility of the drug is less than 0.1 mg/ml (40) or less than 1 mg/ml (41).

The *systemic availability*, that is, the fraction of the oral dose that reaches the systemic circulation, will be affected if the dissolution rate is too slow. Much more easily affected, however, is the *rate of availability*, that is, the rate with which the drug enters the systemic circulation. This availability for drugs absorbed by passive diffusion will theoretically be affected in any case where the drug is not completely dissolved when it reaches the main absorption site. This implies that the dissolution rate of even relatively rapidly dissolving drugs will affect the rate of availability.

The stomach is the first place where absorption of a swallowed dosage form takes place. The absorption rate at this site is rather poor compared to the intestine due to the relatively small mucosal surface area,

the limited agitation and the longer diffusion pathway. The absorption rate first becomes substantial when the drug enters the intestine. Studies have shown that the stomach emptying half-life can vary from 7 to 22 minutes (42,43). The time for the drug to enter the intestine should be significantly less. If the drug does not dissolve completely within this short period of time, any action that results in an increase in the rate of dissolution will increase the rate of availability. Therefore, it should be possible to increase the rate of availability for even relatively rapidly dissolving drugs by a change in the formulation that increases the *in vivo* rate of dissolution.

Therapeutic implications of rate of dissolution in dissolution rate limited absorption.

A slow rate of dissolution results in poor absorption which can cause several undesirable effects including: (a) changes in the extent of systemic availability with decreased therapeutic effect, (b) changes in the rate of availability with delayed onset of activity, (c) increased inter- and intra-subject variability and unpredictability of response, (d) increased residency of drug in gastrointestinal tract with increased damage to mucosal tissue (Aspirin, KCl, steroids) (39). It is therefore nearly always advantageous to formulate both slowly and readily dissolving drugs so they dissolve as fast as possible in the organism. The only exceptions are drugs with a short biological half-life where a slow release dosage form is desirable to avoid the inconvenience of too frequent drug intake. The effect of dissolution rate on pharmacological activity has been discussed by Levy (44).

Factors affecting drug dissolution from solid dosage forms.

The dissolution of a drug from a solid dosage form is influenced by several factors that can be summarised as shown in Scheme 3.2.

Scheme 3.2

Factors affecting drug dissolution from solid dosage forms

I Disintegration

II Environmental factors

- (1) agitation
- (2) Dissolution media
  - (a) drug concentration and gradient
  - (b) pH
  - (c) viscosity
  - (d) interfacial tension
  - (e) complexation
  - (f) solubilization

III Factors related to the drug itself

- (a) solubility
- (b) polymorphism
- (c) solvation
- (d) salt form
- (e) particle size
- (f) particle size distribution

It is possible by proper formulation techniques to influence the majority of these factors such that a higher *in vivo* dissolution rate can be achieved.

Disintegration

A rapid disintegration of the dosage form (tablet, capsule) is necessary in order to achieve fast dissolution. The factors influencing

disintegration have been reviewed by Wagner (45). The aim of disintegration is to make the drug particles available for dissolution as *quickly and efficiently* as possible. Therefore, a disintegration test should include a test for both disintegration time and disintegration efficiency. This efficiency is measured in terms of the availability of disintegrated particles for dissolution. A conventional disintegration (time) test is therefore actually only useful when it is combined with a dissolution test.

The factors influencing dissolution (Scheme 6.2) can be divided into environmental factors and intrinsic factors, the latter being factors related to the drug itself. The environmental factors can be influenced either by additives in the dosage form or by the administration of the drug e.g. whether it is given before, or after a meal or with or without liquid etc. The intrinsic factors can be manipulated by physical or chemical means only e.g. by change in particle size or by making a salt-form of the drug.

#### Environmental factors

##### Agitation

It is well known that dissolution rate increases with increasing agitation (46-48). There does not seem to be any way the *in vivo* agitation can be influenced by drug formulation. It is possibly too drastic to include other drugs that increase gastric emptying rate and gastrointestinal motility. It would be more worthwhile to consider the dependence of agitation on the composition of the food. It has been shown that meals of low viscosity are emptied more rapidly than meals of high viscosity (49) and that fats and fatty acids inhibit gastric motility (50). The physical activity of the patient also plays a role. Walking generally produces a higher gastrointestinal motility than lying (1).

pH is not seen to be any way by which the formulation of a solid drug

(tablet, capsule) can influence significantly the viscosity of the gastrointestinal contents. Noyes and Whitney have shown that the rate of dissolution of a drug is proportional to its solubility (51). Most drugs are weak acids or bases and their solubility is dependent on the pH of the vehicle. By use of the Henderson-Hasselbalch equation it can be shown that the

solubility,  $C$ , of a weak base having solubility,  $C_o$ , in unionised form is dependent on the hydrogen ion concentration as follows:

$$C = C_o (1 + K_b (H^+) / K_w) \quad (3.1)$$

where  $K_w$  is the ion product of water and  $K_b$  is the dissociation constant of the base. The corresponding expression for a weak acid is:

$$C = C_o (1 + K_a / (H^+)) \quad (3.2)$$

The addition of small amounts of alkaline buffer substances to formulations of weak acids results in a higher pH in the vicinity of the drug particles which can enhance the dissolution of the drug (52). Buffers can in this way help to increase the dissolution rate, but their effect on drug absorption should also be considered. It is unfortunate that the effect of pH on the dissolution rates of acids and bases is opposite to the effect of surfactants on the dissolution rate will therefore probably result in overly optimistic expectations about the *in vivo* effect. The absorption of a weak acid from solution is optimal at low pH where the rate of dissolution of the weak acid is lowest. On the other hand, the rapid

dissolution of a weak base in the stomach acid content is not so important because little absorption occurs in this part of the gastrointestinal tract. The effect of complexation on the dissolution rate of drugs has been extensively investigated (58-60). Most soluble macromolecules such as

Viscosity ular weight polyols, gums and cellulose derivatives that are able to form complexes with drugs, can increase the water solubility of the drugs.

It has been shown by several investigators (53-55) that the dissolution rate decreases with increasing viscosity of the vehicle. There

does not seem to be any way by which the formulation of a solid drug (tablet, capsule) can influence significantly the viscosity of the gastrointestinal contents. The implication of viscosity and viscosity-enhancing agents used in suspension on drug absorption has been discussed by Gibaldi (56).

### Interfacial tension

It is well recognised that the dissolution rate is proportional to the effective surface area of the drug, i.e. the surface area available for dissolution. The effective surface area for hydrophobic drugs is usually considerably less than the real surface area because the interfacial tension between the solid and the liquid does not allow complete wetting. The effective surface area can however be increased by the addition of a surface active agent that facilitates wetting and hence results in an increase in the dissolution rate. It was shown by Finholdt and Solvang (57) that the gastric juice in most humans contains surface active agents and has a surface tension that is considerably less than the 0.1M HCl that is often used in *in vitro* experiments. *In vitro* investigations of the effect of surfactants on the dissolution rate will therefore probably result in overly optimistic expectations about the *in vivo* effect.

### Complexation

The effect of complexation on the dissolution rate of drugs has been extensively investigated (58-60). Most soluble macromolecules such as high molecular weight polyols, gums and cellulose derivatives that are able to form complexes with drugs, can increase the water solubility of the drugs. There will not, however, be a proportional increase in the dissolution rate



since the drugs have to dissolve before complex formation can take place. One investigator explains that the increased dissolution rate observed in the presence of certain complex forming agents is possibly caused by a lowering of the energy change for transferring drug molecules from crystal to solution (58). The viscosity increasing properties of macromolecular complex forming agents will, however, reduce the diffusion rate of drugs and often counteract the former effect, causing a possible decrease in the dissolution rate. Complexing agents have primarily been used in dosage forms in order to increase the solubility or stability of a drug rather than to influence its dissolution behaviour. It is important to consider the implications of complex formation on drug absorption before any such agents are used (56,61).

#### Solubilization

The effect of solubilizing agents on the dissolution process has been investigated by several authors (62-64). The increase in the solubility due to micellar solubilization will usually not result in a proportional increase in the dissolution rate. The significant increase in the rate sometimes found is probably caused by the wetting effect rather than the solubilizing effect. It is therefore apparent considering Finholdt and Solvangs' investigations (57) that these agents are limited in their ability to increase the *in vivo* dissolution rate. They are mainly used for technical reasons in production but are also occasionally used for improving absorption, although there still seems to be controversy as to whether they actually enhance or retard drug absorption (56).

FACTORS RELATED TO THE DRUG ITSELF

Solubility

It was early recognised that the rate of dissolution in a diffusion-controlled dissolution process is proportional to the solubility (51). Several papers have discussed dissolution rate in relation to drug solubility (65-67). It has been pointed out that the solubility of very small particles increases with decreasing particle size because of an increase in vapor pressure of the solid (66).

The solubility of a drug is determined by the interaction between the solid drug and the solvent. There are therefore two ways in which the solubility and hence the dissolution rate can be manipulated: by changing the vehicle composition, for example by use of buffering agents to alter pH in the vicinity of the drug particles, as discussed previously; or by changing the crystal form.

Polymorphism

Most drugs can exist in at least two crystal forms. In certain classes of compounds the incidence of polymorphism is even greater. The metastable crystal forms have a lower melting point and a higher solubility than the stable forms. The increased solubility generally results in an increased dissolution rate. The amorphous form represents the highest energy level of the molecules in solid form. A pronounced difference in therapeutic activity between the amorphous and crystalline forms of drugs has been observed in several cases (67,68). This difference can only be due to a difference in the *in vivo* dissolution rate of the two forms, because a drug has to dissolve before it can be absorbed and the properties of the dissolved

drug do not depend on its original crystal structure. The above observations therefore strongly confirm the role of the dissolution rate in the absorption

process. Several drugs can exist in the form of salts and it is well documented that these usually have faster dissolution and absorption rates of enhancing the dissolution rate. Polymorphism of drugs has for this reason than their parent compounds (73-75). Sodium salts of weak acids dissolve much more rapidly than the corresponding acid forms, regardless of the initial problem in using the most soluble crystal form of a drug to achieve faster pH. Their fast dissolution in the low pH of the stomach can be explained in terms of the ability of these salts to increase the pH at the drug-liquid interface, causing a fast release and high concentration in the diffusion layer. Some drugs may subsequently re-precipitate in the bulk fluid, but it can be ensured that they will not change to less therapeutically active crystal forms.

usually in the form of very fine particles that readily dissolve through further dilution or absorption in the gastrointestinal tract (44). Chemical

Solvation can in certain cases preclude the use of the drug in salt form.

The sodium salt of aspirin is very unstable in solution and even the solid form is rather unstable. The sodium or potassium salts may react with atmospheric carbon dioxide and water to precipitate out poorly soluble parent compounds. This occurs on the surface and thereby retards the dissolution of caffeine, theophylline and glutethimide have been observed (75). The and absorption rates (76,77). The alkalinity of some salts may furthermore anhydrous forms dissolved faster in these three cases. The n-pentanol cause epigastric distress after oral intake of the drug in a solid form. and ethylacetate solvates of fluorocortisone dissolved faster, however, than the non-solvated forms (72). The use of solvated or non-solvated forms of

a drug to increase the dissolution rate can be troublesome, since conversion

between forms sometimes occurs during or subsequent to the formulation of a method of increasing the dissolution rate. The surface area to weight ratio of a particle of any shape varies inversely with its diameter. Therefore humidity-dependent as well. Therefore a complete study of the forms should be done under different temperature and humidity conditions before they powder will increase substantially by particle size reduction, in particular are used in pharmaceutical preparations.

for fine particles. The greater effective surface area of the drug in contact with the gastrointestinal fluid will result in more rapid dissolution and

### Salt form

Several drugs can exist in the form of salts and it is well documented that these usually have faster dissolution and absorption rates than their parent compounds (73-75 ). Sodium salts of weak acids dissolve much more rapidly than the corresponding acid forms, regardless of the initial pH. Their fast dissolution in the low pH of the stomach can be explained in terms of the ability of these salts to increase the pH at the drug-liquid interface, causing a fast release and high concentration in the diffusion layer. Some drugs may subsequently re-precipitate in the bulk fluid, but usually in the form of very fine particles that readily dissolve through further dilution or absorption in the gastrointestinal tract (44). Chemical stability can in certain cases preclude the use of the drug in salt form. The sodium salt of aspirin is very unstable in solution and even the solid form is rather unstable. The sodium or potassium salts may react with atmospheric carbon dioxide and water to precipitate out poorly soluble parent compounds. This occurs on the surface and thereby retards the dissolution and absorption rates (76,77). The alkalinity of some salts may furthermore cause epigastric distress after oral intake of the drug in a solid form.

### Particle size

Particle size reduction is undoubtedly the most used and important method of increasing the dissolution rate. The surface area to weight ratio of a particle of any shape varies inversely with its diameter. Therefore the total surface area and hence also the effective surface area of a drug powder will increase substantially by particle size reduction, in particular for fine particles. The greater effective surface area of the drug in contact with the gastrointestinal fluid will result in more rapid dissolution and

absorption. This has led most drug firms to micronize (particle size  $< 25 \mu$ ) very slightly soluble drugs for use in oral dosage forms. There are numerous reports of the better dissolution and availability of micronised drug compared to crystalline (78-82). Particle size reduction of a drug does not always influence its *systemic* availability. It was found for example that 50, 200, 400 and 800  $\mu$  powders of chloramphenicol were absorbed to the same extent. The 50  $\mu$  and 200  $\mu$  powders showed essentially the same absorption rate or rate of availability with a peak blood level at one hour, while the 400  $\mu$  and 800  $\mu$  powders were different with peak blood levels at two and three hours respectively (83). These findings indicate that the *in vivo* dissolution rate of the 50  $\mu$  and 200  $\mu$  powders has been fast enough to enable both powders to dissolve before they reach the main absorption site. However, this was not the case for the 400  $\mu$  and 800  $\mu$  powders that had different rates of availability. Therefore, it can be concluded that nothing has been gained in therapeutic efficacy by reducing the particle size to less than 200  $\mu$ . Such findings are of value in situations where it is better to use coarse rather than fine particles because of production or stability reasons. A particle size reduction to enhance dissolution should seriously be considered when the absorption site is in the stomach or upper region of the intestine. If, on the other hand, the site is in the last section of the intestine, drug absorption may be nearly independent of particle size. This results when dissolution occurs before this section is reached, and depends on the stomach emptying rate and the peristaltic activity (2).

Relatively large particle sizes are often needed to give local action in the terminal part of the gastrointestinal tract (84,85). A weak organic basic drug will often rapidly dissolve in the form of an hydrochloride in the acidic content of the stomach. However, when it passes into the slightly basic content of the intestine, it can precipitate out

as the un-ionized compound. The precipitation will be in the form of very fine, rapidly dissolving particles, so the absorption rate of the compound will be independent of the initial particle size (52). The absorption of a slightly soluble weak acid drug should, on the other hand, be much more dependent on particle size. A reduction in particle size will largely facilitate its dissolution in the acidic gastric juice, where it is only slightly soluble. It then forms the more soluble salt when it passes into the duodenum and small intestine. However, the salt is highly ionised and its absorption rate is decreased, since only a small fraction of the drug molecules is in the undissociated state.

Such considerations regarding the effect of particle size reduction in relation to the acidic or basic character of the drug are based on the pH-partition theory for absorption and should therefore be subject to discussion and further investigation. Particle size reduction can in certain cases reduce the therapeutic efficacy of a drug if it is unstable in gastric juice, as for example penicillin G or erythromycin, fast dissolution will enhance degradation in the stomach. The effect of particle size in relationship to absorption and activity has been reviewed by Fincher (2).

Particle size reduction can be performed in several ways: (a) trituration, (b) ball milling, (c) fluid energy micronization or (d) controlled precipitation including spray drying (86). There are, however, limitations as to how much the particles can be reduced in size and how suitable an extreme particle size reduction would be with respect to formulation and dissolution. Fine particles very often show a strong tendency to aggregate and agglomerate due to their increased surface energy and the stronger van der Waals attraction between non-polar molecules. Furthermore, electrostatic charges in fine hydrophobic powders can cause severe technical

difficulties in production. The problems associated with the wettability of fine powders have already been discussed. Drugs with plastic properties are difficult to subdivide by mechanical means (a-c), since they tend to stick together, even if produced by controlled precipitation (d).

Lin et al. (87) found that the *in vitro* dissolution rates of micronized griseofulvin and glutethimide were slower than those of their coarser particles. The opposite finding was reported by Chiou and Riegelman for griseofulvin (88). The results of Lin et al. can be explained by the strong agglomeration and reduced wettability of the micronized powder. The previously mentioned investigations of Finholdt and Solvang (57) indicate that Chiou and Riegelman's results better reflect what is expected *in vivo*. Several reports about the better availability of drugs in micronized form strongly support this (78-80).

Drugs can also be introduced to the gastrointestinal fluids in the form of very fine particles formed by precipitation *in vivo*. This can be done in several ways (52). The drug can be dissolved in a non-aqueous water-miscible solvent from which it precipitates out by dilution in the gastrointestinal tract. Solutions of sodium or potassium salts of an acidic drug will in the same way precipitate out at the low pH existing in the stomach. Formulations resulting in *in vivo* drug precipitation should have better availability than formulations of drugs in micronized form, because agglomeration and wettability problems are avoided and the particle size of the precipitate usually will be smaller than that which can be produced by micronization. However, the many disadvantages of liquid drug formulations compared to solid with respect to stability and convenience do not seem to counteract the advantage of the somewhat better availability.

formulation form is the problem of aging. Solid dispersions often contain the drug in a high energy, metastable form which, under prolonged storage,

Solid Dispersions

Drug formulations based on solid dispersions of slightly soluble drugs have received considerable attention in recent years. The pharmaceutical application of these systems has been extensively reviewed by Chiou and Riegelman (89). Solid dispersions are micro-crystalline or molecular dispersions of a poorly soluble drug in a solid water-soluble matrix. The drug either precipitates out in a fine particle form which is readily available for dissolution or becomes solubilized when the matrix rapidly dissolves in the gastrointestinal juice. Solid dispersions may be classified as: (a) simple-eutectic mixtures, (b) solid solutions, (c) glass solutions and glass suspensions, (d) amorphous dispersion in crystalline carriers and various combinations of these systems. Several reports indicate that the availability and dissolution rate of these formulations are superior to formulations containing micronized drugs (90,91). Solid dispersions appear to be as available as the liquid formulations from which the drug precipitates out *in vivo*, but they do not have the disadvantages of the latter with respect to their convenience. The stability also seems to be much better although very little investigation has been done in this field (89).

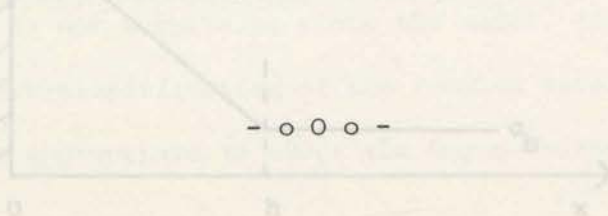
Solid dispersions appear, therefore, to be a very promising new approach in the formulation of slightly soluble, slowly dissolving drugs. The major problem in designing this formulation form lies in finding a suitable production method and in finding a matrix material with the right properties such as low toxicity, high water solubility, high solubilizing capacity for the drug, thermal and chemical stability and suitability for processing. Possibly the main hindrance to the introduction of this formulation form is the problem of aging. Solid dispersions often contain the drug in a high energy, metastable form which, under prolonged storage,



can be transformed into a stable but less soluble and more slowly dissolving form. Solid solutions (molecular solid dispersions) often contain the drug in a supersaturated form which, through the slow diffusion process occurring in solids, can precipitate out during storage, particularly at elevated temperatures (90). The aging of solid dispersions should be an important research subject for the pharmaceutical scientist, hopefully resulting in the commercial acceptance of this unique dosage form.

### Particle size distribution

NERNST FILM In recent years there has been increasing interest in the effect of particle size distribution on the dissolution behaviour of drugs (92-96). The reduced systemic availability observed for certain drug formulations can possibly be explained in terms of a "size distribution effect". In most drug powders there is a considerable difference between the sizes of the larger and smaller particles, which produces an important difference in their time for complete dissolution. This time is for a particle, dissolving according to Hixson-Crowell's cube root law (97), proportional to its initial diameter. The larger particles can because of their manifold longer dissolution time in such cases reach the distal part of the intestine and only be partly dissolved. The further dissolution in this section of the intestine with its content of increased viscosity, solid matter and low water content will be rather slow and the drug will possibly get eliminated from the body by defaecation before completion of its dissolution. Therefore in order to avoid such availability problems it would be good practice to have standards for the particle sizes of slightly soluble drugs.



CHAPTER 4

SINGLE PARTICLE DISSOLUTION

From geometrical considerations (fig. 4.1):

There are three dominant models describing the interfacial mass transport in a dissolving dynamic (agitated) system. These are:

1. Nernst stagnant film theory (98).
2. Danckwerts surface renewal theory (99).
3. Interfacial solvation rate limited dissolution theory (100,101).

NERNST FILM THEORY

The theory of Nernst assumes a stagnant layer of solvent at the solid-liquid interface. The mass transport of solute through the layer is accomplished by simple molecular diffusion in a steady state fashion following Fick's law of diffusion. Once passed the stagnant layer the solute is then mixed quickly by convection and diffusion in the bulk of the liquid.

The Nernst model predicts a concentration profile from a plane surface in two dimensions as shown schematically in Fig. 4.1, where  $h$  refers to the thickness of the so-called diffusion layer,  $c_i$  is the interfacial concentration and  $c_b$  the bulk concentration. The concentration gradient in the stagnant layer,  $0 \leq x \leq h$ , is then constant because the diffusion flux,

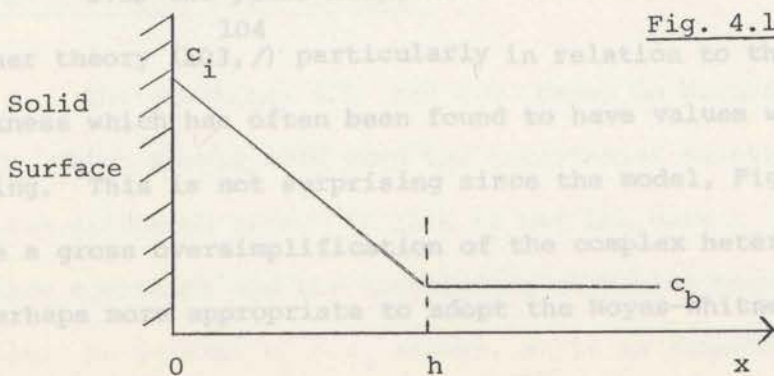


Fig. 4.1

$J_D$ , is constant during steady state:

$$J_D = -Ddc/dx = \text{constant} \tag{4.1}$$

From geometrical considerations (fig. 4.1):

$$dc/dx = -(c_i - c_b)/h \tag{4.2}$$

so that  $J_D = (D/h) (c_i - c_b)$  (4.3)

If it is assumed that the interfacial reaction rate is large compared to the rate of diffusion then  $c_i$  can be considered close to the solubility concentration  $c_o$  so 4.3 can be approximated by:

$$J_D \sim (D/h) (c_s - c_b) \tag{4.4}$$

The dissolution rate from a plane surface of area, A, will then be:

$$dw/dt = -(DA/h) (c_s - c_b) \tag{4.5}$$

If in 4.5  $D/h$  is assumed constant, the equation reduces to the Noyes-Whitney equation:

$$dw/dt = -kA (c_s - c_b) \tag{4.6}$$

after the workers who appear to be the first to have verified this equation experimentally (51). They did not attach any mechanistic significance to the quantity k. Nernst and Brunner (102) extended the Noyes-Whitney equation to include the concept of diffusion layer thickness and diffusion coefficient as presented in 4.5.

Over the years there have been several critics of the Nernst-Brunner theory (103,/) particularly in relation to the diffusion layer thickness which has often been found to have values without any physical meaning. This is not surprising since the model, Fig. 4.1, would appear to be a gross oversimplification of the complex heterogeneous process. It is perhaps more appropriate to adopt the Noyes-Whitney equation recognising

that the quantity  $k$  depends in some way on  $D$ , the hydrodynamic conditions and the geometry of the dissolving object.

It has been shown that  $k$  in Eq. 4.6 under fixed experimental conditions can often be considered constant and the equation has been successfully applied in several cases.

#### DANCKWERTS THEORY

Danckwerts (99) rejected the idea of a stagnant film and proposed a model based on the assumption that liquid motion is turbulent and extends to the surface of the dissolving crystal. The physical interpretation is that pockets of fresh solvent reach the interface by turbulence and renew parts of the surface, while 'old' pockets containing solute simultaneously leave the surfaces. The mathematical description of this system includes a parameter,  $\gamma$ , known as the mean rate of surface renewal:

$$\frac{dw}{dt} = A \sqrt{\gamma D} (c_s - c_b) \quad (4.7)$$

For the dissolution of small particles Goyan (105) proposed the following model which combines 4.5 and 4.7 :

$$\frac{dw}{dt} = -A \left( \frac{D}{r} + \sqrt{\gamma D} \right) (c_s - c_b) \quad (4.8)$$

When the radius of the particle,  $r$ , diminishes as  $\gamma \rightarrow 0$  this equation reduces to 4.5 (in the case  $r = h$ ).

#### LIMITED SOLVATION RATE THEORIES

The equations 4.5 and 4.6 based on Nernst film theory assume that  $c_i \sim c_s$  which should hold when the interfacial reaction rate is large compared with the diffusion rate. If this is not the case  $c_s$  must be replaced by  $c_i$  in these equations and the dissolution mechanism becomes significantly more complex. In general  $c_i < c_s$  always, so it is somewhat difficult to decide

between diffusion and reaction rate controlled dissolution.

Wurster and Taylor (104) proposed a method based on the temperature dependence of  $k$ . They claim that the process is diffusion-controlled if the temperature coefficient is approximately 1.3 and interfacial reaction rate controlled if it is close to 2.0. Higuchi (4) derived the following equation for an interfacial reaction rate and diffusion rate controlled process:

$$dw/dt = - \frac{AD}{h + D/k_i} (c_s - c_b) \quad (4.9)$$

where  $k_i$  was termed the effective interfacial transport rate constant. This equation was presented in slightly modified form in a later publication (60). It is seen that when the effective interfacial transport rate constant is large i.e. when  $k_i \gg D/h$  the equation reduces to 4.5 and the process is diffusion rate limited. If  $k_i \ll D/h$  equation 4.9 reduces to:

$$dw/dt = - A k_i (c_s - c_b) \quad (4.10)$$

and the process is truly reaction rate controlled.

### SPHERICAL DISSOLUTION

Nernst theory was derived for a plane interface. It is of interest to apply similar assumptions to a spherical particle, i.e. assume (Fig. 4.2):

1. Spherical symmetry in dissolution
2.  $c = c_i \sim c_s$  as  $r = r_t$
3.  $c = c_b$  at  $r = r_t + h$
4. Solute transported only by Ficks diffusion ( $J_D = -D\partial c/\partial r$ ) in stagnant layer,  $r_t \leq r \leq r_t + h$
5. quasi steady state conditions in the sense that the mass transport rate through a spherical surface in the diffusion

layer is constant at a fixed time, i.e.

$$(4\pi r_t^2 D \frac{dc}{dr})_t = \text{constant} \quad r_t \leq r \leq r_t + h \quad (4.11)$$

or: 
$$\left[ \frac{d(r^2 \frac{dc}{dr})}{dr} \right]_t = 0 \quad (4.12)$$

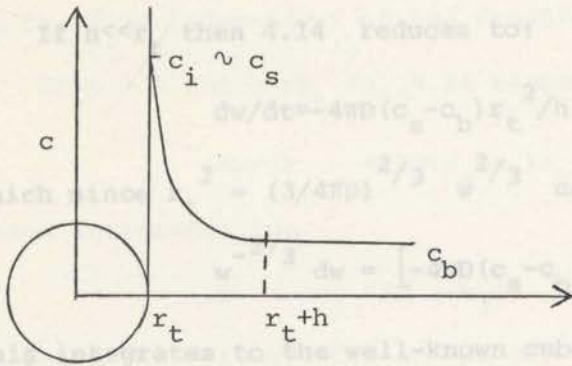


Fig. 4.2

Solving 4.12 under boundary conditions 2 and 3 above leads to:

$$\left( \frac{dc}{dr} \right)_t = - (c_s - c_b) \frac{r_t (r_t + h)}{hr^2} \quad r_t \leq r \leq r_t + h \quad (4.13)$$

From this expression, it is seen that the concentration gradient in the diffusion layer is not constant as it was in the case for the plane body (Fig. 4.1). The dissolution rate of the spherical particle according to assumption 4 is thus given by:

$$\frac{dw}{dt} = 4\pi r_t^2 D \left( \frac{dc}{dr} \right)_{r=r_t} = - 4\pi D (c_s - c_b) (r_t + h) r_t / h \quad (4.14)$$

The diffusion layer thickness,  $h$ , is related to the intensity of the solvent flow or agitation in the vicinity of the spherical interface. This agitation will depend in some way on the size,  $r_t$ , of the particle. Therefore, some functional relationship must exist between the thickness of the diffusion layer and the radius of the dissolving particle.

Several functional relationships between  $h$  and  $r_t$  can be assumed. On this basis, integration of 4.14 will then lead to different models for single particle dissolution as demonstrated in the following section.

to simple relationships based on the following power model:

Constant Diffusion Layer Thickness.

Firstly, it would be of interest to investigate the case where h is constant independent of particle size, since this assumption has often been made in the literature. Two cases will be considered:

1. If  $h \ll r_t$  then 4.14 reduces to:

$$dw/dt \approx -4\pi D(c_s - c_b) r_t^2 / h \quad (4.15)$$

which since  $r_t^2 = (3/4\pi\rho)^{2/3} w^{2/3}$  can be written:

$$w^{-2/3} dw = [-4\pi D(c_s - c_b) (3/(4\pi\rho))^{2/3} / h] dt \quad (4.16)$$

this integrates to the well-known cube root law (97)

$$w^{1/3} = w_o^{1/3} - k_1 t \quad (4.17)$$

with:  $k_1 = (4\pi/3)^{1/3} D(c_s - c_b) \rho^{-2/3} / h \quad (4.18)$

2. If  $h \gg r_t$  then 4.14 reduces to:

$$dw/dt \approx -4\pi D(c_s - c_b) r_t \quad (4.19)$$

which similarly to above integrates to:

$$w^{2/3} = w_o^{2/3} - k_2 t \quad (4.20)$$

where  $k_2 = 2(4\pi/3)^{2/3} D \rho^{-1/3} (c_s - c_b) \quad (4.21)$

It is interesting to note that in this case the diffusion layer thickness, h, is not a part of the rate parameter  $k_2$ .

Variable Diffusion Layer Thickness.

It is expected from hydrodynamic considerations that h decreases with increasing r. Any monotonically decreasing functional relationship between h and r could be considered. This investigation will be restricted to simple relationships based on the following power model:

$$h = k_p r_t^{-p} \quad k_p, p > 0 \quad (4.22)$$

where  $k_p$  and  $p$  are some positive constants. Several cases can be considered depending upon the value of  $p$  and the size of  $h$  relative to  $r_t$ . It should first be noted that in the cases where  $h \gg r_t$  Eq. 4.14 still leads to Eq. 4.20 independent of the functional relationship between  $h$  and  $r_t$ .

1. If  $p = 1$  and  $h \ll r_t$  Eq. 4.14 becomes:

$$dw/dt = -4\pi D(c_s - c_b) r_t^3 / k_p \quad (4.23)$$

which integrates to:

$$w = w_o \text{EXP}(-k_3 t) \quad (4.24)$$

$$\text{where } k_3 = 3D(c_s - c_b) / k_p \rho \quad (4.25)$$

2. If  $p = 1$  and  $h$  is not very different in magnitude from  $r_t$  Eq. 4.14 leads to the following expression:

$$w^{2/3} = (w_o^{2/3} + k_4) \text{EXP}(-k_3 t) - k_4 \quad (4.26)$$

$$\text{where } k_4 = k_p (4\pi\rho/3)^{2/3} \quad (4.27)$$

and  $k_3$  is given by 4.25.

3. If  $p = 2$  and  $h \ll r_t$  the following equation is obtained:

$$w^{1/3} = (w_o^{-1/3} + k_5 t)^{-1} \quad (4.28)$$

$$\text{where } k_5 = 4\pi D(c_s - c_b) (4\pi\rho/3)^{4/3} / 3k_p \quad (4.27)$$

#### APPROACHES NOT BASED ON DIFFUSION LAYER ASSUMPTIONS

The Noyes-Whitney model applied to a spherical particle takes the form:

$$dw/dt = -kA(c_s - c_b) \quad (4.28)$$

where  $k$  may or may not be constant or depend on the particle radius.

1. If  $k = \text{constant}$  4.28 integrates to yield the model proposed by



Hixson and Crowell (97)<sup>1</sup>:

$$w^{1/3} = w_o^{1/3} - k_6 t \quad (4.29)$$

where<sup>1</sup>  $k_6 = k(c_s - c_b) (4\pi/3) \rho^{-2/3}$  (4.30)

2. If  $k$  is inversely proportional to the particle radius, say,  $k = k_7/r_t$  then 4.28 integrates to:

$$w^{2/3} = w_o^{2/3} - k_8 t \quad (4.31)$$

where  $k_8 = 2k_7(c_s - c_b) (4\pi/3)^{2/3} \rho^{-1/3}$  (4.32)

3. If  $k$  is inversely proportional to the square root of the particle radius, i.e.  $k = k_9 r_t^{-1/2}$ , Eq. 4.28 integrates to yield:

$$w^{1/2} = w_o^{1/2} - k_{10} t \quad (4.33)$$

$$k_{10} = k_9 (2\pi)^{1/2} (2\rho/3)^{-1/3} (c_s - c_b) \quad (4.34)$$

Sink Conditions.

The models for single particle dissolution presented above are derived for dissolution under sink condition. This condition can best be explained in a mathematical context to be a condition under which the change in  $c_b$  is so small that the errors, introduced by considering  $(c_s - c_b)$  or  $(c_i - c_b)$  to be constant, are acceptable. This definition is related to the concept of ignoring any time dependence of the concentration terms in the integration of the dissolution rate equation.

Comparing Models.

It is interesting to note that several of the dissolution models above could be derived both on the basis of the diffusion layer theory and

1. These workers did not specify the composition of  $k_6$  but assumed it did not vary with the progress of the dissolution.

on the more simple model 4.28. In both approaches certain assumptions were made about the relationship between either the diffusion layer thickness,  $h$ , and the particle size or the rate parameter  $k$  (4.28) and the particle size.

In spite of oversimplification of the dissolution mechanism in the diffusion layer approach, it is appealing since it has greater conceptual value than the other approach. The flexibility of models based on a diffusion layer seems also to be greater.

Of the models considered above the cube root model (4.17, 4.29), the square root model (4.33) and the 2/3-root model (4.20, 4.31) have all been proposed in the literature (97,106,107).

There still seems to be some controversy as to which of these models best describes dissolution of the single particle. This is mainly because there appears to be no accurate experimental information directly concerning the dissolution of single particles.

The inferences made have been based on multiparticle dissolution from which it may be difficult to deduce single particle dissolution behaviour, particularly when the particle size distribution is not completely monodisperse (see Chapter 6).

In the experimental evaluation of the 1/3, 1/2 and 2/3 root models in Chapter 7, the cube root model appears to be the best although further investigation is necessary to confirm this finding. It may well be that a more flexible model applies which is close to the 1/3 root model in the beginning and approaches the 1/2 and 2/3 root model as the dissolution process progresses. Such behaviour is expected if the dissolution follows Eq. 4.14. This equation reduces to the 1/3 model if  $h \ll r_t$ , which is likely to be true in the initial stages of dissolution if the particle size then is relatively large. Later when the particle is reduced in size the hydrodynamic activity in the vicinity of the interface will be considerably

smaller resulting in an increased  $h$  value so that for  $h \gg r_t$  the dissolution reaction approaches the  $2/3$  root model. The intermediate state will result in a "square-root dissolution". This condition should occur when  $(r_t+h)r_t/h \sim r_t^{3/2}$  i.e. when  $h \sim r_t^2/(r_t^{3/2} - r_t)$ .

STATIC DISSOLUTION MODELS

Study of single particle dissolution in which there is no agitation far from the centre of the particle, which is the same as the initial concentration. In most cases therefore  $c_\infty = 0$ . which the solute mass transport in most cases is completely diffusion-controlled.

If a spherical particle is considered the system is mathematically well defined. Substantial theoretical studies have been performed in this area in chemical engineering (108-113). The many different mathematical approaches presented have been based on different assumptions. For example, the assumption that the process is wholly diffusion controlled (109,/) or partly diffusion controlled (113). Most of the mathematical models have been described in terms of partial differential equations which for given boundary conditions can only be solved numerically using a digital computer.

It is interesting to note that the radius vs. time relationship computed in this way by Cable and Evans (109) and Ready and Cooper (112) is approximately linear except for an initial transient period. This means that the dissolution of a spherical particle under static conditions can be approximated (disregarding the initial phase) by the cube root law. (The  $1/3$  model implies a linear decrease in radius with time.)

Of the equations presented for static spherical dissolution, the following equations given by Rosner (113) are appealing because of their simplicity:

B.C.1.  $c = c_s \quad r = r_t \quad t > 0$  (4.38)

B.C.2.  $c = 0 \quad r > r_0 \quad t = 0$  (4.39)

B.C.3.  $c = 0 \quad r = \infty$  (4.40)

$$dr/dt = (D\alpha/r) \ln ((1-c_\infty)/(1-c_i)) \quad (4.35)$$

$$k (c_s - c_i)^p = (D/r) \ln ((1-c_\infty)/(1-c_i)) \quad (4.36)$$

Where  $\alpha$  is the ratio of solvent density/solute density,  $k$  is the rate constant governing the interface kinetics,  $p$  is the order of the solvation reaction. It was assumed that this reaction was of the ordinary Berthroud form, (100), i.e., that  $p = 1$ . The term  $c_\infty$  is the concentration infinitely far from the centre of the particle, which is the same as the initial concentration. In most cases therefore  $c_\infty = 0$ .

The above two equations define, in a computationally simple way, the variation of particle radius with time for given values of  $D$ ,  $\alpha$ ,  $c_s$  and  $r_0$ . For example 4.36 can be solved numerically for  $c_i$  and this value substituted into 4.35 which by numerical integration provides the  $r_t$ -value for the chosen value of  $t$ . Such calculations were performed on a computer by Ridgway and Peacock (114) for various simple organic compounds.

The validity of the above two equations depends primarily on the following extended quasi steady-state assumption: "The instantaneous concentration field surrounding the dissolving sphere is approximated by the steady-state concentration field surrounding a hypothetical spherical solid of the same size, through which solute is being artificially forced at a mass rate equal to the instantaneous rate of dissolution".

If the interfacial concentration  $c_i$  is constant and close to the solubility concentration  $c_b$  the dissolution can be obtained directly by solving Fick's law:

$$\partial c / \partial t = D \nabla^2 c \quad (4.37)$$

under the following boundary conditions:

$$\text{B.C.1.} \quad c = c_s \quad r = r_t \quad t > 0 \quad (4.38)$$

$$\text{B.C.2.} \quad c = 0 \quad r > r_0 \quad t = 0 \quad (4.39)$$

$$\text{B.C.3.} \quad c = 0 \quad r = \infty \quad (4.40)$$

This yields the following expression for the concentration profile in the liquid (115):

$$c(r,t) = c_s \frac{r_t}{r} \operatorname{erfc} \frac{r-r_t}{2\sqrt{Dt}} \quad (4.41)$$

so that the mass flux per unit surface area at the interface is:

$$J_i = -D(\partial c/\partial r)_{r=r_t} = Dc_s \left[ (\pi Dt)^{-1/2} + 1/r_t \right] \quad (4.42)$$

Furthermore:

$$dw/dt = -4\pi r_t^2 J_i \quad (4.43)$$

Now since  $dw/dt = d(4\pi r_t^3 \rho/3)/dt = 4\pi \rho r_t^2 dr_t/dt$ , 4.42 becomes:

$$dr_t/dt = -J_i/\rho \quad (4.44)$$

i.e. 
$$dr_t/dt = -\frac{Dc_s}{\rho} \left[ (\pi Dt)^{-1/2} + 1/r_t \right] \quad (4.45)$$

This equation can readily be integrated numerically to yield the time dependence of  $r_t$  and hence a numerical solution for the single particle dissolution for given values of  $\rho$ ,  $D$  and  $c_s$ .

As dissolution proceeds the term  $(\pi Dt)^{-1/2}$  in the bracket in 4.45 decreases while in the same time the term  $1/r_t$  increases. Thus after the initial transition period  $((\pi Dt)^{-1/2} + 1/r_t) \sim 1/r_t$  and 4.45 integrates to yield the 2/3-root law:

$$w^{2/3} = w_0^{2/3} - k_{11} t \quad (4.46)$$

where 
$$k_{11} = 2(4\pi/3)^{2/3} D \rho^{-1/3} c_s \quad (4.47)$$

It is interesting and satisfactory to note that this model was also derived for the dynamic system (4.20), considering the diffusion layer theory for the case  $h \gg r_t$  i.e. for a system with little agitation. In fact the rate parameters  $k_{11}$  and  $k_2$  in 4.47 and 4.21 are identical for  $c_b = 0$ .

A DYNAMIC MODEL FOR A FREE FALLING SPHERICAL PARTICLE

It is very difficult to define, qualitatively or quantitatively, the agitation in the close vicinity of a particle in a mechanically stirred liquid. A better defined experimental system would be the examination of a spherical particle which falls freely through an unstirred liquid. Analysis of such a system should be particularly valuable in the study of single particle dissolution.

In the following section an attempt is made to describe this system in a mathematically rigorous way. In order to do so the following assumptions are made and will be referred to where necessary in the derivation:

- A1. The particle remains spherical during dissolution.
- A2. The interfacial concentration,  $c_i$ , is constant and very close to the saturation concentration,  $c_s$ .
- A3. The temperature and densities remain constant.
- A4. There is no  $c_s/r$  dependence.
- A5. The effect of Brownian motions is negligible.
- A6. The particle falls without producing turbulence, i.e. the Reynold's number,  $Re$ , is less than 0.1:

$$Re = 2r_t v_\infty \rho_1 / \mu < 0.1 \quad (4.48)$$

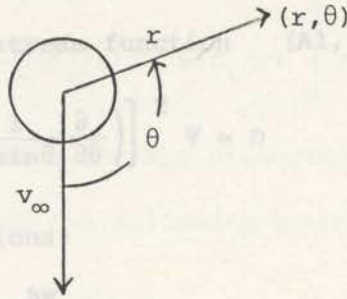
(The velocity of the particle in the direction of gravity is denoted  $v_\infty$ , the viscosity of the liquid  $\mu$  and its density  $\rho_1$ .)

- A7. The fluid is incompressible.
- A8. The fluid flow behaves in a Newtonian way.
- A9. Fick's laws of diffusion are obeyed with  $D$ =constant.

Spherical symmetry makes it convenient to choose a spherical coordinate system with the particle centre as origin and the  $\theta$  coordinate

starting counter-clockwise from the direction of the gravity (Fig. 4.3) of the particle and the velocity  $v_\infty$  with which the particle is falling through the liquid. This velocity can be related through Stokes' equation to the particle radius  $r_c$ .

Fig. 4.3



The  $r$  and  $\theta$  components of  $\underline{v}$  can be found by solving the following differential equation for the stream function  $\psi$  (A1, A3, A4-48):

$$\left[ \frac{\partial^2}{\partial r^2} + \frac{\sin\theta}{r} \frac{\partial}{\partial \theta} - \frac{\partial}{\partial \theta} \left( \frac{\partial}{\partial \theta} \right) \right] \psi = 0 \quad (4.53)$$

subject to the boundary conditions

B.C.1  $v_r = -\frac{1}{r} \frac{\partial \psi}{\partial \theta} = 0$  at  $r=r_c$  (4.54)

B.C.2  $v_\theta = \frac{1}{r \sin\theta} \frac{\partial \psi}{\partial r} = 0$  at  $r=r_c$  (4.55)

First it is necessary to derive an extension of Fick's law valid for incompressible liquids (A7.) in motion: A shell mass balance on an arbitrary volume element,  $p$ , in the bulk of the liquid yields (A9.):

$$\iiint_p \partial c / \partial t \, dp = \iint_s (-\underline{v} \cdot \underline{n} c + D \nabla c \cdot \underline{n}) \, ds \quad (4.49)$$

where  $s$  is the surface of the element and  $\underline{n}$  is the normal to the surface directed outward and  $\underline{v}$  is the fluid velocity vector. The divergence theorem applied on r.h.s. of 4.49 yields:

$$\iiint_p \partial c / \partial t \, dp = \iiint_p (-\nabla \cdot \underline{v} c + D \nabla^2 c) \, dp \quad (4.50)$$

which also may be written:

$$\iiint_p \partial c / \partial t \, dp = \iiint_p (-\underline{v} \cdot \nabla c - c \nabla \cdot \underline{v} + D \nabla^2 c) \, dp \quad (4.51)$$

Equating left and right integrand in 4.51, noting  $\nabla \cdot \underline{v} = 0$  (A7.) it follows that:

$$\partial c / \partial t = D \nabla^2 c - \underline{v} \cdot \nabla c \quad (4.52)$$

This is the extended form of Fick's second law which takes into account that the liquid in which diffusion takes place is in motion.

where  $\underline{v}$  The fluid velocity vector function  $\underline{v}$  above depends on the geometry of the particle and the velocity  $v_\infty$  with which the particle is falling through the liquid. This velocity can be related through Stokes' equation to the particle radius  $r_t$ .

The  $r$  and  $\theta$  components of  $\underline{v}$  can be found by solving the following differential equation for the stream function (A1, A3, A5-A8):

$$\left[ \frac{\partial^2}{\partial r^2} + \frac{\sin\theta}{r^2} \frac{\partial}{\partial \theta} \left( \frac{1}{\sin\theta} \frac{\partial}{\partial \theta} \right) \right]^2 \Psi = 0 \quad (4.53)$$

subject to the boundary conditions:

$$\text{B.C.1. } v_r = -\frac{1}{r^2 \sin\theta} \frac{\partial \Psi}{\partial \theta} = 0 \text{ at } r=r_t \quad (4.54)$$

$$\text{B.C.2. } v_\infty = \frac{1}{r \sin\theta} \frac{\partial \Psi}{\partial r} = 0 \text{ at } r=r_t \quad (4.55)$$

$$\text{B.C.3. } \Psi \rightarrow -\frac{1}{2} v_\infty r^2 \sin^2 \theta \text{ for } r \rightarrow \infty \quad (4.56)$$

The velocity components are then obtained

$$v_r = -v_\infty \left[ 1 - \frac{3}{2} \left( \frac{r_t}{r} \right) + \frac{1}{2} \left( \frac{r_t}{r} \right)^3 \right] \cos\theta \quad 0 \leq \theta \leq \pi \quad (4.57)$$

$$v_\theta = v_\infty \left[ 1 - \frac{3}{4} \left( \frac{r_t}{r} \right) - \frac{1}{4} \left( \frac{r_t}{r} \right)^3 \right] \sin\theta, \quad 0 \leq \theta \leq \pi \quad (4.58)$$

The  $r$  and  $\theta$  components of  $\underline{\nabla}c$  in 4.52 is  $\partial c/\partial r$  and  $r^{-1} \partial c/\partial \theta$  respectively thus:

$$\underline{v} \cdot \underline{\nabla}c = -v_\infty \left[ 1 - \frac{3}{2} \left( \frac{r_t}{r} \right) + \frac{1}{2} \left( \frac{r_t}{r} \right)^3 \right] \frac{\partial c}{\partial r} \cos\theta \quad (4.59)$$

$$+ v_\infty \left[ 1 - \frac{3}{4} \left( \frac{r_t}{r} \right) - \frac{1}{4} \left( \frac{r_t}{r} \right)^3 \right] r^{-1} \frac{\partial c}{\partial \theta} \sin\theta \quad (4.60)$$

When there is no  $\phi$ -dependence the  $\nabla^2$  operator in spherical coordinates is:

$$\nabla^2 = \frac{1}{r^2} \frac{\partial}{\partial r} \left( r^2 \frac{\partial}{\partial r} \right) + \frac{1}{r^2 \sin\theta} \frac{\partial}{\partial \theta} \left( \sin\theta \frac{\partial}{\partial \theta} \right) \quad (4.60)$$

The velocity  $v_\infty$  is related to  $r_t$  by Stokes' law:

$$v_\infty = (2\Delta\rho g/9\mu) r_t^2 \quad (4.61)$$



where  $g$  is the acceleration of gravity and  $\Delta\rho$  the solid-liquid density difference. Equations 4.52 and 4.59-61 can then be summarized to give:

$$\frac{\partial c}{\partial t} = \frac{D}{r^2} \frac{\partial}{\partial r} \left( r^2 \frac{\partial c}{\partial r} \right) + \frac{D}{r^2 \sin\theta} \frac{\partial}{\partial \theta} \left( \sin\theta \frac{\partial c}{\partial \theta} \right) \quad (4.71)$$

$$- (2\Delta\rho g/9\mu) \left[ 1 - \frac{3}{2}(r_t/r) + \frac{1}{2}(r_t/r)^3 \right] r_t^2 \frac{\partial c}{\partial r} \cos\theta \quad (4.62)$$

$$+ (2\Delta\rho g/9\mu) \left[ 1 - \frac{3}{4}(r_t/r) - \frac{1}{4}(r_t/r)^3 \right] r_t^2 \frac{\partial c}{\partial \theta} \sin\theta \quad (4.73)$$

which is the differential equation for the dissolution of spherical particle.

The equation is to be solved for the following boundary conditions:

$$\text{B.C.1. } c(r_t, \theta, 0) = c_s \quad (4.63)$$

$$\text{B.C.2. } c(\infty, \theta, t) = 0 \quad (4.64)$$

$$\text{B.C.3. } c(r, \theta, 0) = 0 \quad r > r_t \quad (4.65)$$

Because 4.62 is a nonlinear partial differential equation, it seems impossible to solve analytically using conventional techniques. The equation can, however, be solved numerically using a digital computer. It would then be convenient first to transform the variables involved to dimensionless quantities such as:

$$c^* = c/c_s \quad 0 \leq c^* < 1 \quad (4.66)$$

$$t^* = Dt/r_o^2 \quad 0 \leq t^* < \infty \quad (4.67)$$

$$r^* = r/r_o \quad 1 \leq r^* < \infty \quad (4.68)$$

$$u = \cos\theta \quad -1 \leq u \leq 1 \quad (4.69)$$

It can then be shown that 4.62 becomes:

$$\frac{\partial c^*}{\partial t^*} = \frac{\partial}{\partial r^*} \left( r^* \frac{\partial c^*}{\partial r^*} \right) + \frac{\partial}{\partial u} \left( (1-u^2) \frac{\partial c^*}{\partial u} \right) \quad (4.70)$$

$$- \frac{1}{2} Sc Re^0 \left[ 1 - \frac{3}{2}(r_t^*/r^*) + \frac{1}{2}(r_t^*/r^*)^3 \right] r_t^{*2} u \frac{\partial c^*}{\partial r^*}$$

$$+ \frac{1}{2} Sc Re^0 \left[ 1 - \frac{3}{4}(r_t^*/r^*) - \frac{1}{4}(r_t^*/r^*)^3 \right] r_t^{*2} (1-u^2) \frac{\partial c^*}{\partial u}$$

where  $Sc$  is the Schmidt number ( $Sc = \mu/\Delta\rho D$ ) and  $Re^{\circ}$  is the initial Reynold number ( $Re^{\circ} = 2 r_{\infty} v_{\infty}^{\circ} \rho_1/\mu$ ). The transformed equation is then to be solved under the following conditions:

$$\text{B.C.1. } c^*(1, u, 0) = 1 \quad (4.71)$$

$$\text{B.C.2. } c^*(\infty, u, t^*) = 0 \quad (4.72)$$

$$\text{B.C.3. } c^*(r^*, u, 0) = 0 \quad r^* > 1 \quad (4.73)$$

The numerical solution obtained by solving 4.70 for various values of  $ScRe^{\circ}$  will then be general in the sense that it encompasses an infinite number of combinations of values for the parameters  $\mu$ ,  $\rho_1$ ,  $\Delta\rho$ ,  $D$  and  $c_s$ .

#### NONSPHERICAL ISOTROPIC DISSOLUTION

The models for single particle dissolution considered previously in this chapter are all derived for spherical particles. Application of such models to real particle systems is complicated by the fact that pure drug particles are not spherical. The usual approach has been to treat the real particles as if they were spherical having the same surface area or volume. Such approximations may introduce substantial errors.

The influence of shape factors on dissolution kinetics of drugs has been discussed for tablets and controlled release tablets (116,117), but little attention has been given to single drug particles (118,119).

This section presents exact isotropic single-particle dissolution equations for several nonspherical forms and formulas enabling calculation of the diameters of hypothetical spherical particles which closely approximate the dissolution of these forms.

Assume that dissolution takes place isotropically, that is, that the rate of dissolution per unit surface area,  $J$ , is constant so that the following equation can be written:

$$dw/dt = -JA \quad (4.74.)$$

where  $w$  is the amount undissolved and  $A$  is the surface area. This equation implies that the boundary of a plane interface retreats with constant speed during dissolution such that:

$$ds/dt = -J/\rho \quad (4.75.)$$

where  $\rho$  is the solid density, and  $s$  is the distance perpendicular to the interface from some fixed reference point in the dissolving solid.

Equation 4.75. integrates to:

$$s = s_0 - Jt/\rho \quad (4.76.)$$

where  $s_0$  is the initial ( $t=0$ ) distance to the reference point. When this equation is applied to the isotropic dissolution of a spherical particle, the following equation arises:

$$w/w_0 = (1 - Jt/r_0\rho)^3 \quad (4.77.)$$

$$\text{or} \quad (w/w_0)^{1/3} = 1 - Jt/r_0\rho \quad (4.78.)$$

where  $r_0$  is the initial particle radius.

Therefore, when a spherical particle dissolves isotropically ( $J=\text{constant}$ ), it obeys the Hixson and Crowell (97) cube root law, that is, a plot of  $(w/w_0)^{1/3}$  vs.  $t$  is linear.

#### Dissolution of Prismatic Particles.

Structures I-VI (Fig. 4.4.) are 10 simple forms of the six crystal systems and illustrate the dimensional quantities  $b_0$ ,  $c_0$ ,  $l_0$ ,  $h_0$  and  $\alpha$  used in the following derivations. It is assumed, without loss of generality, that  $b_0 < c_0 < l_0$ .

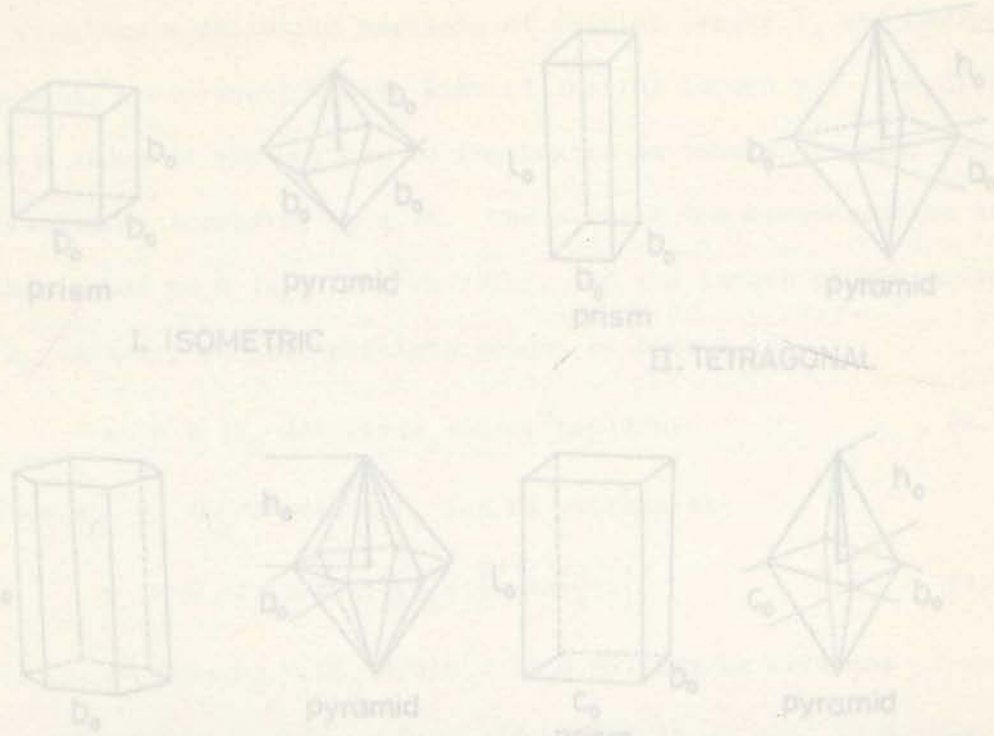
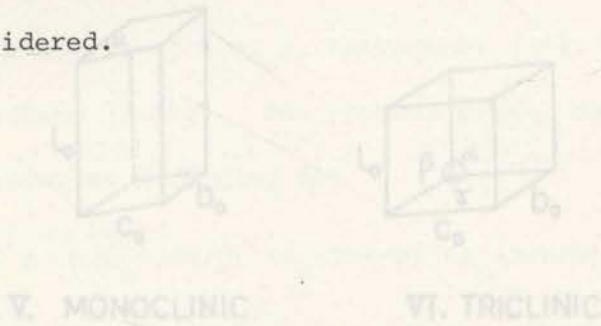
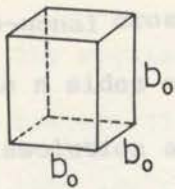
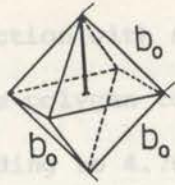


Figure 4.4 Illustration of the six basic crystal forms for which spherical approximations to the theoretical isotropic dissolution are considered.

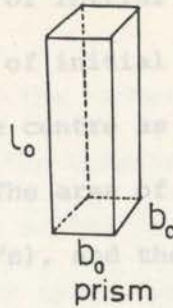




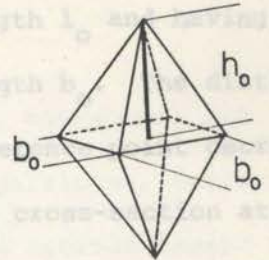
prism



pyramid



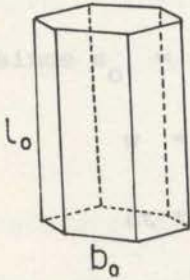
prism



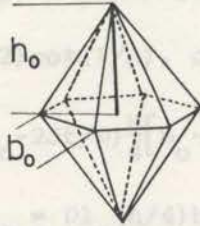
pyramid

I. ISOMETRIC

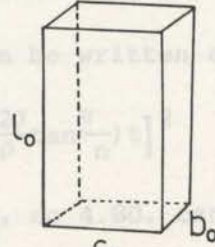
II. TETRAGONAL



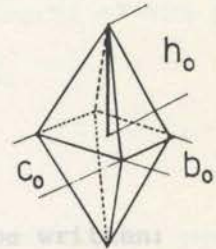
prism



pyramid



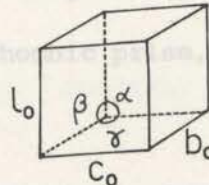
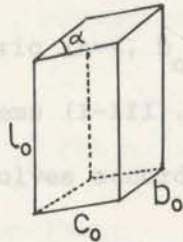
prism



pyramid

III. HEXAGONAL

IV. RHOMBIC



V. MONOCLINIC

VI. TRICLINIC

Equation 4.81...  
 Form of the isometric, tetragonal, and hexagonal  
 (n=6), crystal system...  
 cross-section, dissolves according to:  
 because of the isotropic retreat of all surfaces. Similarly to 4.81., this  
 equation can be written:

$$w/w_0 = (1 - 2\lambda t/h_0 \rho) (1 - 2\lambda t/c_0 \rho) (1 - 2\lambda t/l_0 \rho) \quad (4.83.)$$

The monoclinic prismatic particles, having a parallelogram cross-section with an acute angle  $\alpha$ , at any time has a cross-sectional area equal to  $bc \sin \alpha$ , where  $b = b_0 - [(2\lambda/\rho) \sin \alpha] t$  and  $c = c_0 - [(2\lambda/\rho) \sin \alpha] t$ , so it dissolves according to:

$$w = \rho \left[ 1 - \frac{2\lambda t}{\rho} \right] \left[ b_0 - \left( \frac{2\lambda}{\rho} \sin \alpha \right) t \right] \left[ c_0 - \left( \frac{2\lambda}{\rho} \sin \alpha \right) t \right] \sin \alpha \quad (4.84.)$$

$$\text{or: } w/w_0 = \left( 1 - \frac{2\lambda t}{l_0 \rho} \right) \left[ 1 - \left( \frac{2\lambda}{c_0 \rho} \sin \alpha \right) t \right] \left[ 1 - \left( \frac{2\lambda}{b_0 \rho} \sin \alpha \right) t \right] \quad (4.85.)$$

Consider a prismatic particle of initial length  $l_0$  and having a regular  $n$ -gonal cross-section with side of initial length  $b_0$ . The distance,  $s$  from the  $n$  sides of the polygon to the centre as reference point decreases during dissolution according to 4.76. The area of the cross-section at any time is then equal to  $n (s_0 - Jt/\rho)^2 \tan(\pi/n)$ , and the length of the prism is equal to  $l_0 - (2Jt/\rho)$ , so that particle weight at time  $t$  is:

$$w = \rho (l_0 - 2Jt/\rho) n (s_0 - Jt/\rho)^2 \tan(\pi/n) \quad (4.79.)$$

which, since  $s_0 = (b_0/2) \cot(\pi/n)$ , can be written as:

$$w = \rho (l_0 - 2Jt/\rho) \frac{n}{4} \left[ b_0 - \left( \frac{2J}{\rho} \tan \frac{\pi}{n} \right) t \right]^2 \quad (4.80.)$$

at  $t=0$ ,  $w_0 = \rho l_0 (n/4) b_0^2$ , so 4.80. can be written:

$$w/w_0 = \left( 1 - \frac{2J}{l_0 \rho} t \right) \left[ 1 - \left( \frac{2J}{b_0 \rho} \tan \frac{\pi}{n} \right) t \right]^2 \quad (4.81.)$$

Equation 4.81. comprises the exact dissolution equation for the prismatic forms of the isometric ( $n=4$ ,  $b_0=l_0$ ), tetragonal ( $n=4$ ,  $b_0=l_0$ ), and hexagonal ( $n=6$ ), crystal systems (I-III). The rhombic prism, having a rectangular cross-section, dissolves according to:

$$w = \rho (b_0 - 2Jt/\rho) (c_0 - 2Jt/\rho) (l_0 - 2Jt/\rho) \quad (4.82.)$$

because of the isotropic retreat of all surfaces. Similarly to 4.81., this equation can be written:

$$w/w_0 = (1 - 2Jt/b_0 \rho) (1 - 2Jt/c_0 \rho) (1 - 2Jt/l_0 \rho) \quad (4.83.)$$

The monoclinic prismatic particle, having a parallelogram cross-section with an acute angle  $\alpha$ , at any time has a cross-sectional area equal to  $bc \sin \alpha$ , where  $b=b_0 - [(2J/\rho) \sin \alpha] t$  and  $c=c_0 - [(2J/\rho) \sin \alpha] t$ , so it dissolves according to:

$$w = \rho (l_0 - 2Jt/\rho) \left[ b_0 - \left( \frac{2J}{\rho} \sin \alpha \right) t \right] \left[ c_0 - \left( \frac{2J}{\rho} \sin \alpha \right) t \right] \sin \alpha \quad (4.84.)$$

$$\text{or: } w/w_0 = \left( 1 - \frac{2Jt}{l_0 \rho} \right) \left[ 1 - \left( \frac{2J}{b_0 \rho} \sin \alpha \right) t \right] \left[ 1 - \left( \frac{2J}{c_0 \rho} \sin \alpha \right) t \right] \quad (4.85.)$$

Dissolution of Pyramidal Particles.

The regular pyramidal forms of the isometric, tetragonal and hexagonal systems (I-III) all dissolve like spherical particles, following the "cube root law": all plane surfaces of the pyramid retreat toward its centre of symmetry with the same constant speed during isotropic dissolution. Therefore, the shape of the pyramid remains the same while its size diminishes. For example, it can be shown geometrically that all lengths of the prism decrease by a factor of  $(1-Jt/r_0\rho)$ , where  $r_0$ , given by:

$$r_0 = \frac{1}{2} h_0 b_0 (h_0^2 + b_0^2/4)^{-1/2} \quad (4.86.)$$

is the radius of the largest sphere that can be contained in the pyramid initially. The weight of the regular n-gonal prism at time t is equal to  $\frac{1}{6} \rho n h_t b_t^2 \cot(\pi/n)$ , where the height,  $h_t$ , and side,  $b_t$ , are  $h_t = h_0 (1-Jt/r_0\rho)$  and  $b_t = b_0 (1-Jt/r_0\rho)$ , respectively, according to the above theory. Thus, its weight is:

$$w = \frac{1}{6} \rho n h_0 b_0^2 (1-Jt/r_0\rho)^3 \cot(\pi/n) \quad (4.87.)$$

from which it follows that:

$$w/w_0 = (1-Jt/r_0\rho)^3 \quad (4.88.)$$

This equation is identical to 4.77. Therefore, a regular pyramidal crystal form dissolves in identical manner to the largest (hypothetical) spherical particle that can be contained within its boundaries initially. This is also approximately true for an irregular pyramidal form such as the rhombic pyramid when the irregularity is not too extreme. It can be shown, using a double integration approach, that this crystal form dissolves according to:

$$w/w_0 = (1-Jt/pr_1)^2 \left[ 1 - \left( \frac{1}{4r_1} + \frac{3}{4r_2} \right) \frac{J}{\rho} t \right] \quad (4.89.)$$

where

$$r_1 = \frac{1}{2} h_o b_o (h_o^2 + b_o^2/4)^{-1/2} \quad (4.90.)$$

$$r_2 = \frac{1}{2} h_o c_o (h_o^2 + c_o^2/4)^{-1/2} \quad (4.91.)$$

The deviation from spherical particle dissolution (4.77.) arises from the fact  $b_o \neq c_o$ . If  $b_o = c_o$ , then 4.89. reduces to the special cases 4.88. as expected.

To evaluate these single-particle dissolution equations, it is convenient to present them in a transformed simplified form which better illustrates their *intrinsic dissolution profile* (96). For example 4.81. can be transformed to:

$$w/w_o = (1-Ft^*) (1-t^*)^2 \quad (4.92.)$$

or  $(w/w_o)^{1/3} = (1-Ft^*)^{1/3} (1-t^*)^{2/3} \quad (4.93.)$

where:

$$t^* = \left( \frac{2J}{b_o \rho} \tan \frac{\pi}{n} \right) t \quad (4.94.)$$

is denoted *time length* and:

$$F = \frac{b_o}{l_o} \cot (\pi/n) \quad (4.95.)$$

is denoted the *shape ratio*. This form of the equation clearly shows that the intrinsic dissolution profile depends only on the value of the dimensionless shape ratio,  $F$ , which defines the particle shape. Furthermore, the transformation makes it more convenient to evaluate the extent to which dissolution of the prismatic particle deviates from spherical particle dissolution (i.e. from the cube root law). For  $F=1$  i.e. when  $(w/w_o)^{1/3} = 1-t^*$ , there is no such deviation; however, as  $F$  decreases, the deviation becomes more significant, i.e., when the length of the particle relative to its side length or "diameter" becomes more extreme.



It is seen (Fig. 4.5.) that as F decreases, the deviation from the cube root law becomes larger. Dissolution then approaches "the square root law", that is, a linear relationship between  $(w/w_0)^{1/2}$  and time length (or time). This is in agreement with the fact that, for small F values, 4.92. approximates  $(w/w_0)^{1/2} = 1-t^*$ .

The cube root law and the square root law were each postulated previously as a model for the dissolution of spherical particles under sink conditions (97,106). Pure drug particles are not spherical, however, but are often prismatic in shape. Therefore, the particle shape effect should be considered in any experimental evaluation of such models.

The dissolution equation for a rhombic pyramidal particle, 4.89., can also be transformed to 4.92. where:

then: 
$$t^* = \frac{2J}{\rho} (h_0^2 + b_0^2/4)^{-1/2} t \quad (4.96.)$$

and: 
$$F = \frac{1}{4} + \frac{3}{4} \left( \frac{h_0^2 + c_0^2/4}{h_0^2 + b_0^2/4} \right)^{1/2} \quad (4.97.)$$

It is seen (Fig. 4.6.) that the shape ratio, F, for this pyramidal particle form does not deviate much from 1 for most shapes, indicating that in most cases dissolution closely approximates that of a spherical particle.

Dissolution equations for either a rhombic (4.83.) or a monoclinic (4.85.) particle can similarly be written in a common transformed form as:

$$w/w_0 = (1-F_1 t^*) (1-F_2 t^*) (1-t^*) \quad (4.98.)$$

where  $F_1 = b_0/c_0$ ,  $F_2 = b_0/l_0$ , and  $t^* = 2Jt/b_0\rho$  for a rhombic particle and  $F_1 = b_0/c_0$ ,  $F_2 = (b_0/l_0)\sin\alpha$ , and  $t^* = (2J/b_0\rho)\sin\alpha t$  for a monoclinic particle.

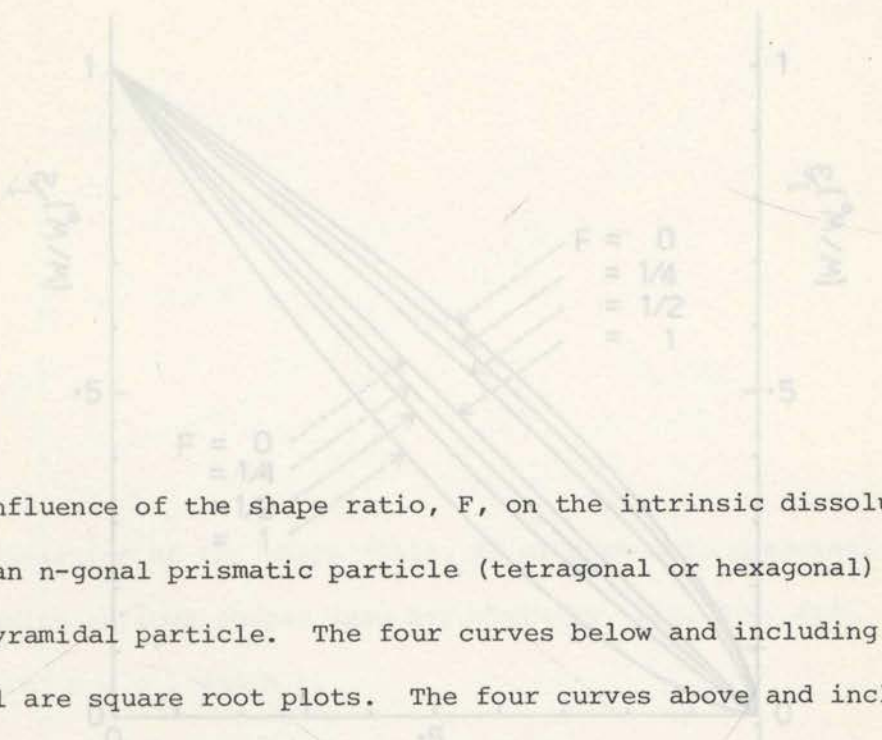


Figure 4.5

Influence of the shape ratio,  $F$ , on the intrinsic dissolution profile of an  $n$ -gonal prismatic particle (tetragonal or hexagonal) or a rhombic pyramidal particle. The four curves below and including the diagonal are square root plots. The four curves above and including the diagonal are cube root plots. The dissolution deviates increasingly from the  $w/w_0^{1/3}$  versus  $t^*$  linear relationship (the cube root law) as the shape ratio becomes less than 1 and approaches a linear  $(w/w_0)^{1/2}$  versus  $t^*$  relationship (the square root law).

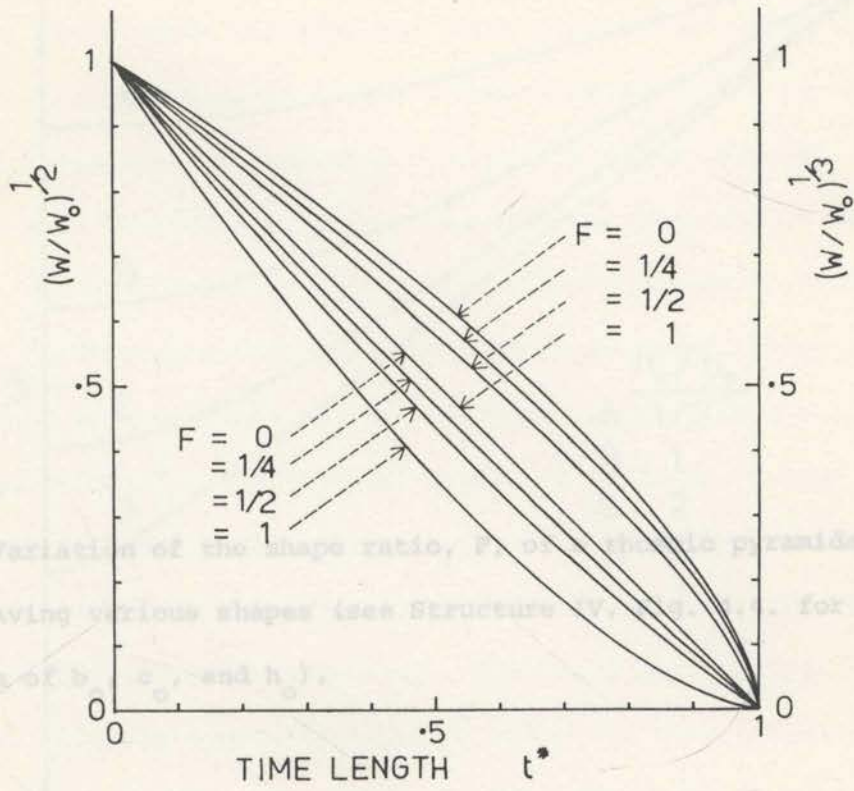


Figure 4.5.

Variation of the shape ratio,  $F$ , of a spherical pyramidal particle having various shapes (see Structure 4.1.1.1. for definitions of  $b$ ,  $c$ , and  $h$ ).

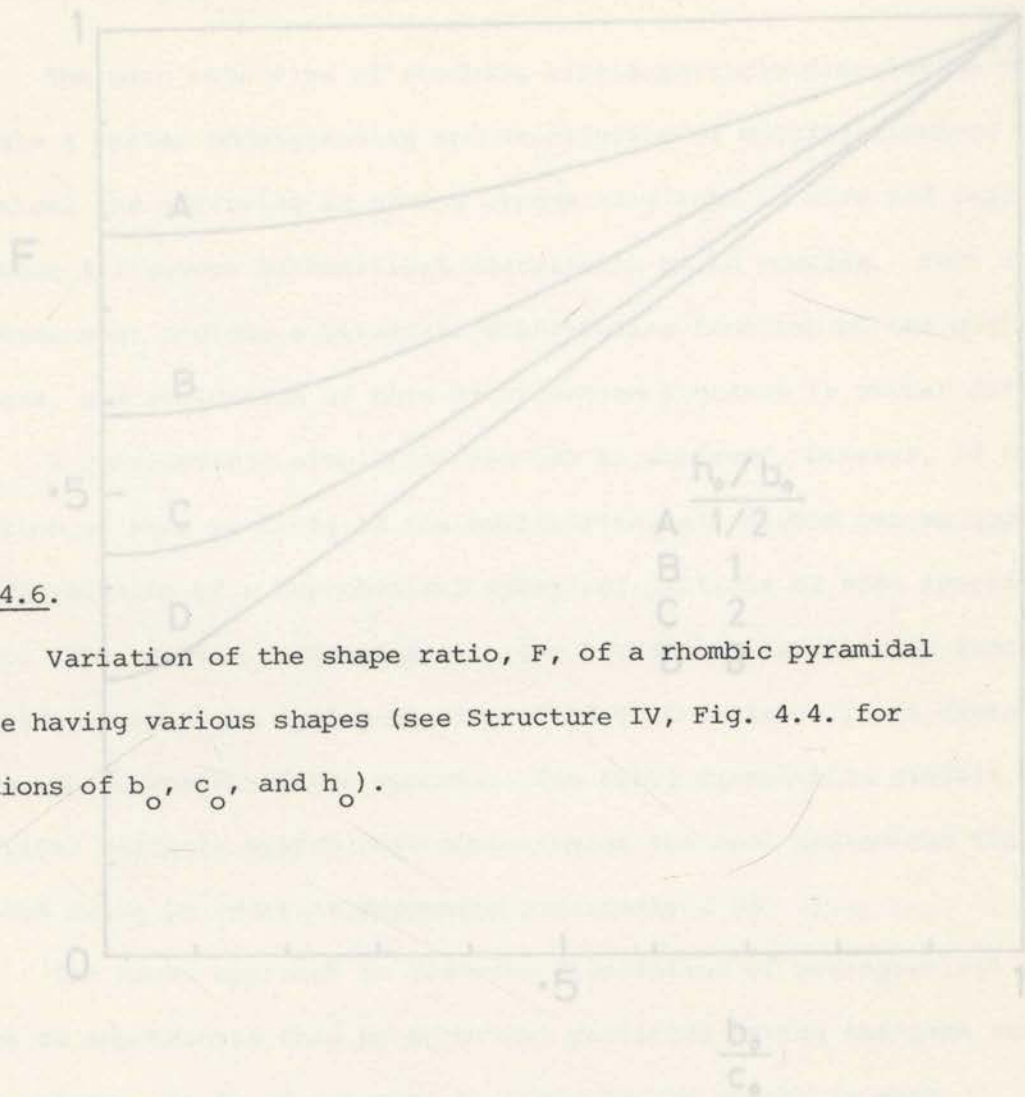
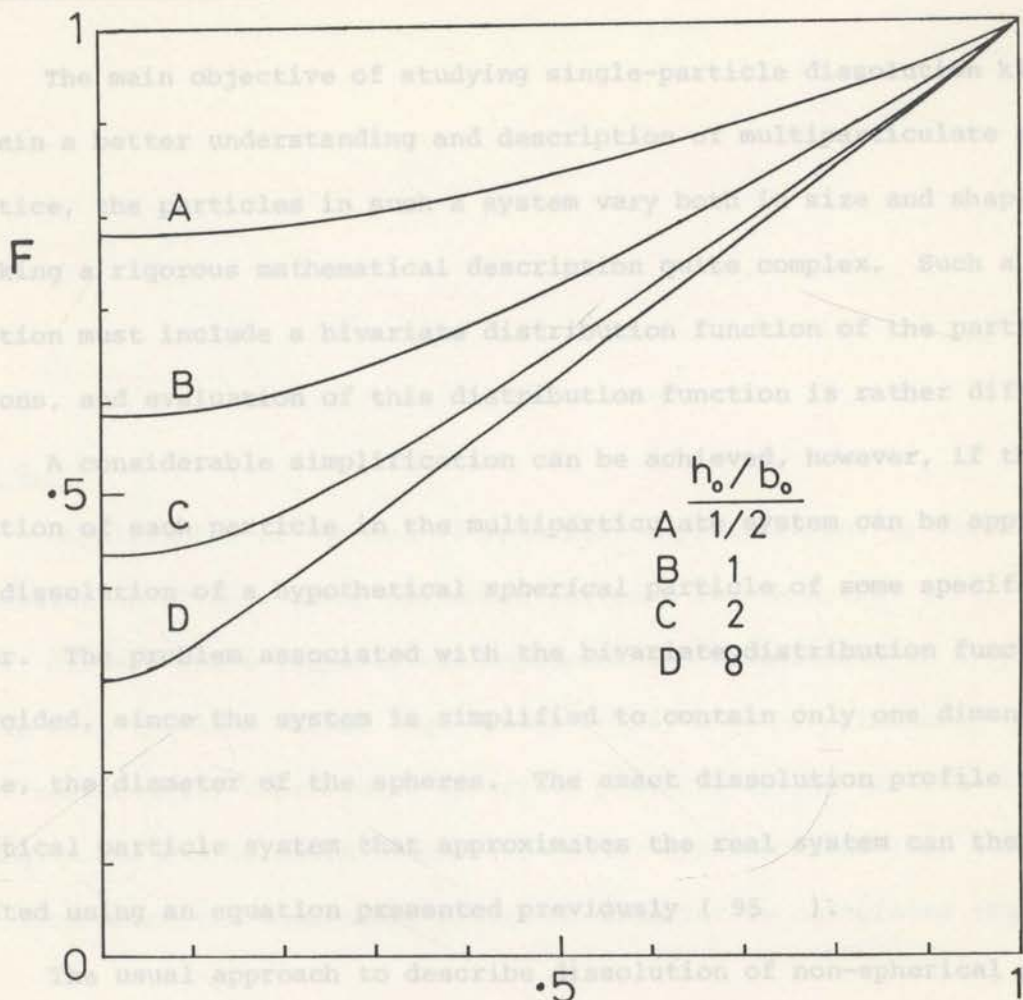


Figure 4.6.

Variation of the shape ratio,  $F$ , of a rhombic pyramidal particle having various shapes (see Structure IV, Fig. 4.4. for definitions of  $b_0$ ,  $c_0$ , and  $h_0$ ).

Spherical Approximations.



The main objective of studying single-particle dissolution kinetics is to gain a better understanding and description of multicomponent systems. In practice, the particles in such a system vary both in size and shape, thus making a rigorous mathematical description quite complex. Such a description must include a bivariate distribution function of the particle dimensions, and a measure of this distribution function is rather difficult. A considerable simplification can be achieved, however, if the dissolution of each particle in the multicomponent system can be approximated by the dissolution of a hypothetical spherical particle of some specified diameter. The problem associated with the bivariate distribution function is then avoided, since the system is simplified to contain only one dimensionless variable, the diameter of the spheres. The exact dissolution profile of the hypothetical particle system that approximates the real system can then be calculated using an equation presented previously (95).

A usual approach to describe dissolution of non-spherical particles has been to approximate them by spherical particles having the same surface area or volume. It is of interest to evaluate the errors in such approximations. A spherical particle, having the same surface area as an n-gonal prismatic particle with shape ratio F, dissolves according to:

$$(w/w_0)^{1/3} = 1 - \left( \frac{2\pi c_0 \cot \frac{\pi}{n}}{n(1+2/F)} \right)^{1/3} t^* \quad (4.99.)$$

Or if it has the same volume, it dissolves according to:

$$(w/w_0)^{1/3} = 1 - \left( \frac{2\pi c_0 \cot \frac{\pi}{n}}{3n} \right)^{1/3} t^* \quad (4.100.)$$

where  $c^*$  and F are defined by 4.94. and 4.95. Figure 4.7. shows the substantial errors introduced by such approximations based on equal surface area or volume. This is not only the case for  $F = 1/4$  but for all other values of the shape

Spherical Approximations.

The main objective of studying single-particle dissolution kinetics is to gain a better understanding and description of multiparticulate systems. In practice, the particles in such a system vary both in size and shape, thus making a rigorous mathematical description quite complex. Such a description must include a bivariate distribution function of the particle dimensions, and evaluation of this distribution function is rather difficult.

A considerable simplification can be achieved, however, if the dissolution of each particle in the multiparticulate system can be approximated by the dissolution of a hypothetical *spherical* particle of some specified diameter. The problem associated with the bivariate distribution function is then avoided, since the system is simplified to contain only one dimensionless variable, the diameter of the spheres. The exact dissolution profile of the hypothetical particle system that approximates the real system can then be calculated using an equation presented previously ( 95 ).

The usual approach to describe dissolution of non-spherical particles has been to approximate them by spherical particles having the same surface area or volume. It is of interest to evaluate the errors in such approximations. A spherical particle, having the same surface area as an n-gonal prismatic particle with shape ratio F, dissolves according to:

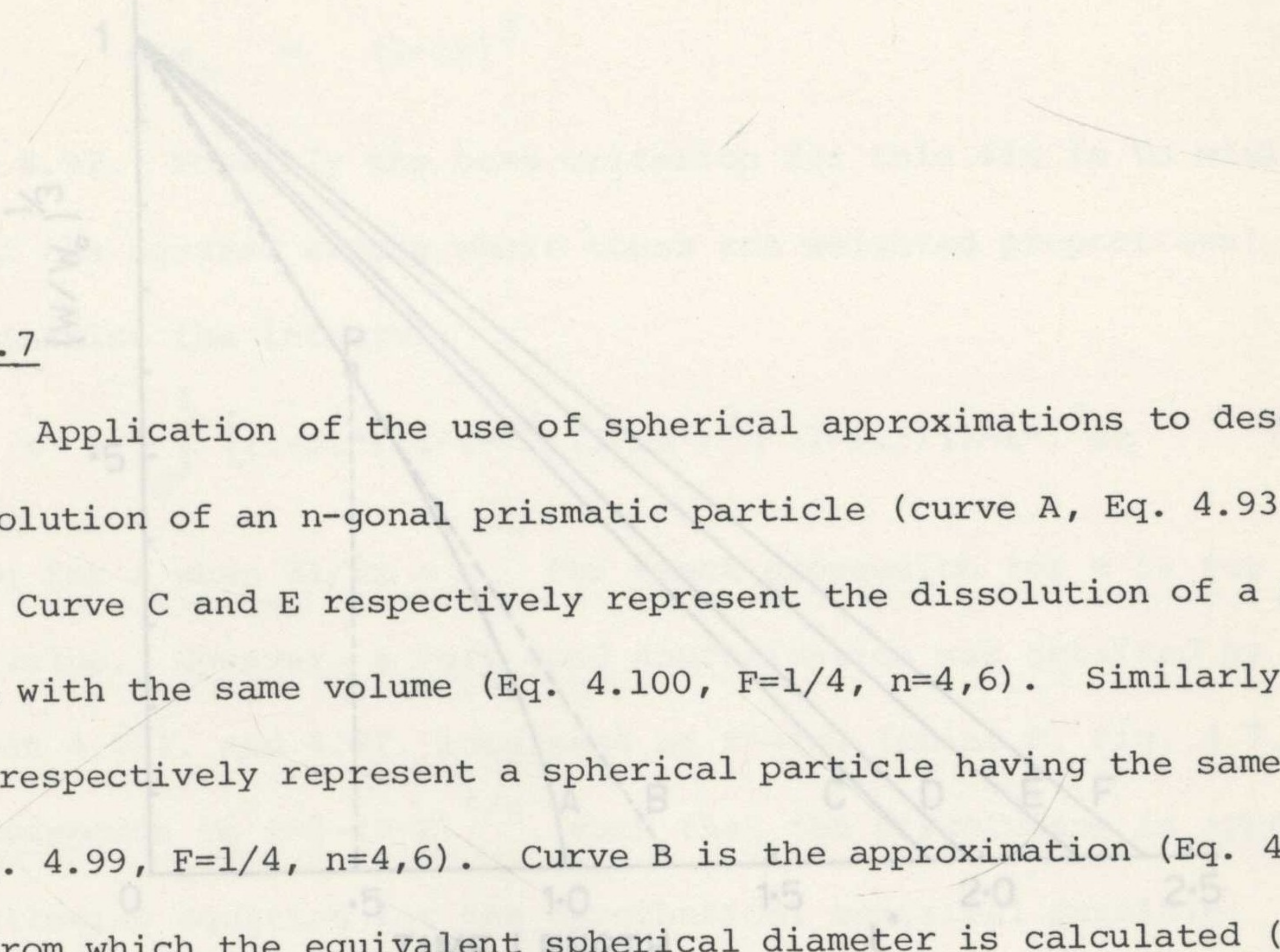
$$(w/w_0)^{1/3} = 1 - \left( \frac{2\pi \cot \frac{\pi}{n}}{n(1+2/F)} \right)^{1/2} t^* \quad (4.99.)$$

Or if it has the same volume, it dissolves according to:

$$(w/w_0)^{1/3} = 1 - \left( \frac{2\pi F \cot \frac{\pi}{n}}{3n} \right)^{1/3} t^* \quad (4.100.)$$

where  $t^*$  and  $F$  are defined by 4.94. and 4.95. Figure 4.7. shows the substantial errors introduced by such approximations based on equal surface area or volume. This is not only the case for  $F = 1/4$  but for all other values of the shape

Figure 4.7



Application of the use of spherical approximations to describe the dissolution of an n-gonal prismatic particle (curve A, Eq. 4.93,  $F=1/4$ ). Curve C and E respectively represent the dissolution of a spherical particle with the same volume (Eq. 4.100,  $F=1/4$ ,  $n=4,6$ ). Similarly curve D and F respectively represent a spherical particle having the same surface area (Eq. 4.99,  $F=1/4$ ,  $n=4,6$ ). Curve B is the approximation (Eq. 4.103,  $F=1/4$ ) from which the equivalent spherical diameter is calculated (Eq. 4.104).

ratio less than 1.

The problem of finding the diameter of the spherical particle that best approximates the dissolution of an n-gonal prismatic particle is mathematically the same as finding a quantity, x, such that:

$$(w/w_0)^{1/3} = (1-xt)^3 \tag{4.101.}$$

best fits 4.92. Possibly the best criterion for this fit is to minimize the sum of the squared errors where these are weighted proportional to  $w/w_0$ , i.e. to minimize the integral

$$\int_0^1 [(1-pt^*)^3 - (1-t^*)^3]^2 (1-pt^*) (1-t^*)^2 dt \tag{4.102.}$$

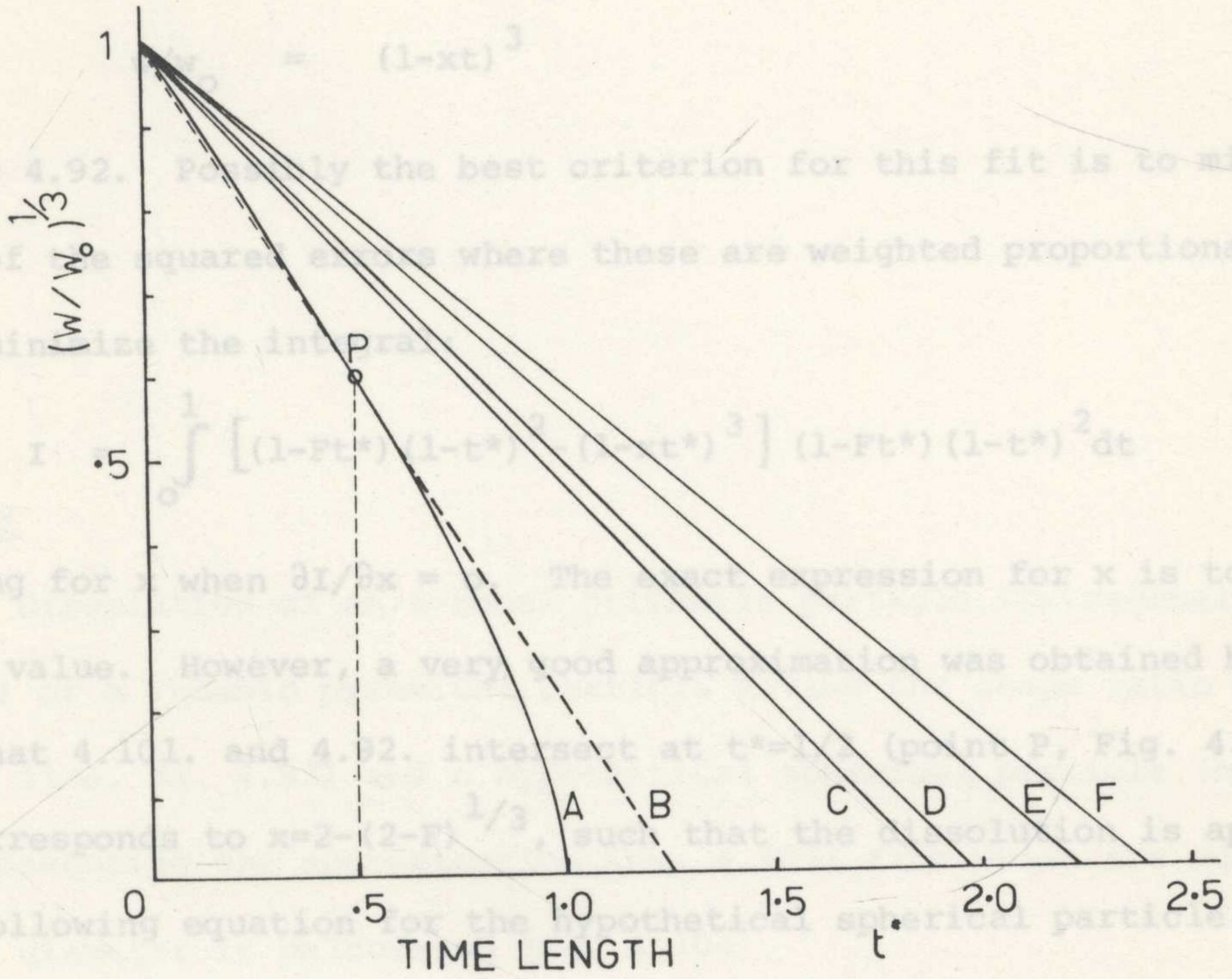
by solving for x when  $\partial I/\partial x = 0$ . The exact expression for x is too complex to be of value. However, a very good approximation was obtained by choosing x such that 4.101. and 4.92. intersect at  $t^*=0.5$  (point P, Fig. 4.7.), which corresponds to  $x=2-(2-F)^{1/3}$ , such that the dissolution is approximated

$$(w/w_0)^{1/3} = 1 - [2-(2-F)^{1/3}] t^* \tag{4.103.}$$

The equivalent spherical diameter, a, i.e., the diameter of this spherical particle, can then be obtained by equaling the right-hand sides of 4.103. and 4.78. from which it follows that:

$$a = \frac{2\sqrt{t}}{[2-(2-F)^{1/3}] \rho t^*} \tag{4.104.}$$

Although the approximation given by 4.103. (curve B, Fig. 4.7.) does not seem to be a particularly good fit to the exact dissolution curve calculated for the n-gonal particle (curve A), when the same two curves are plotted as  $w/w_0$  vs. time length (Fig. 4.8.) instead of as  $(w/w_0)^{1/3}$ , it is clear that this is because of the nonlinear scaling in the cube root plot. The stippled curve (Fig. 4.8.) representing the spherical approximation shows





ratio less than 1.

The problem of finding the diameter of the spherical particle that best approximates the dissolution of an n-gonal prismatic particle is mathematically the same as finding a quantity,  $x$ , such that:

$$w/w_0 = (1-xt)^3 \quad (4.101.)$$

best fits 4.92. Possibly the best criterion for this fit is to minimize the sum of the squared errors where these are weighted proportional to  $w/w_0$ , i.e. to minimize the integral:

$$I = \int_0^1 [(1-Ft^*)(1-t^*)^2 - (1-xt^*)^3]^2 (1-Ft^*)(1-t^*)^2 dt \quad (4.102.)$$

Figure 4.8

by solving for  $x$  when  $\partial I/\partial x = 0$ . The exact expression for  $x$  is too complex to be of value. However, a very good approximation was obtained by choosing  $x$  such that 4.101. and 4.92. intersect at  $t^*=1/2$  (point P, Fig. 4.7.), which corresponds to  $x=2-(2-F)^{1/3}$ , such that the dissolution is approximated by the following equation for the hypothetical spherical particle:

$$(w/w_0)^{1/3} = 1 - [2-(2-F)^{1/3}] t^* \quad (4.103.)$$

The equivalent spherical diameter,  $a$ , i.e., the diameter of this spherical particle, can then be obtained by equaling the right-hand sides of 4.103. and 4.78. from which it follows that:

$$a = \frac{2Jt}{[2-(2-F)^{1/3}] \rho t^*} \quad (4.104.)$$

Although the approximation given by 4.103. (curve B, Fig. 4.7.) does not seem to be a particularly good fit to the exact dissolution curve calculated for the n-gonal particle (curve A), when the same two curves are plotted as  $w/w_0$  vs. time length (Fig. 4.8.) instead of as  $(w/w_0)^{1/3}$ , it is clear that this is because of the nonlinear scaling in the cube root plot. The stippled curve (Fig. 4.8.) representing the spherical approximation shows

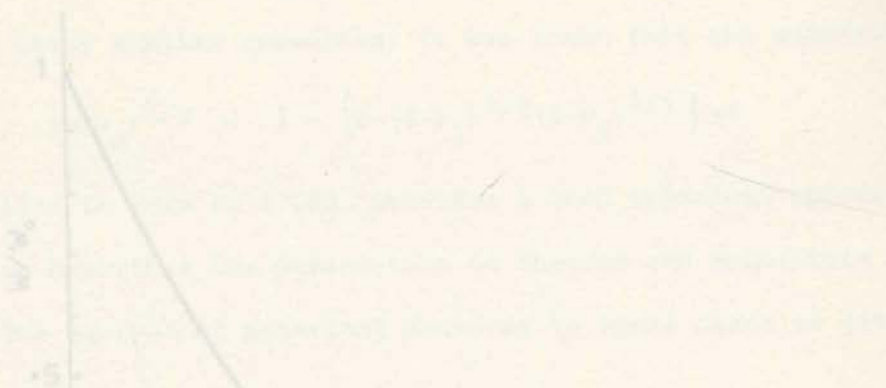


Figure 4.8

Dissolution of an n-gonal prismatic particle (tetragonal or hexagonal) or a rhombic pyramidal particle having the shape ratio  $F=1/4$  (unbroken line, Eq. 4.93) and a hypothetical spherical particle (broken line) representing the approximation (Eq. 4.103) from which the equivalent spherical diameter is calculated (Eq. 4.104).

excellent fit to the true dissolution. The weighted errors of the spherical approximation were calculated for various values of the shape ratio  $F$  and showed (Fig. 4.9.) that this choice for the approximation was satisfactory.

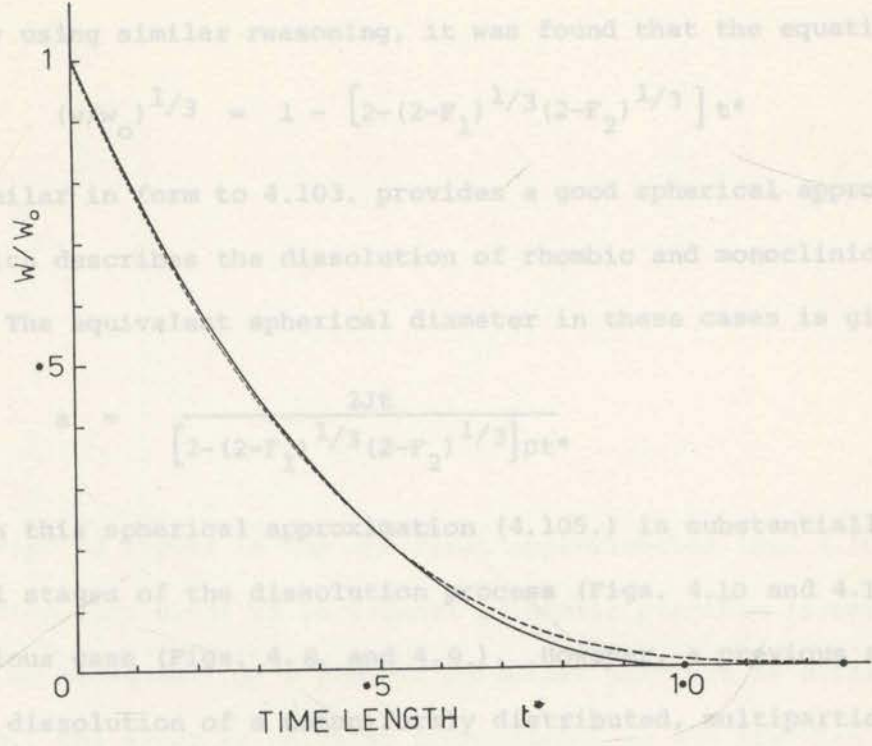
By using similar reasoning, it was found that the equation:

$$\left(\frac{W}{W_0}\right)^{1/3} = 1 - \left[2 - (2-F_1)^{1/3}(2-F_2)^{1/3}\right] t^* \quad (4.103.)$$

which is similar in form to 4.103, provides a good spherical approximation to 4.98, which describes the dissolution of rhombic and monoclinic prismatic particles. The equivalent spherical diameter in these cases is given by:

$$d^* = \frac{2FE}{\left[2 - (2-F_1)^{1/3}(2-F_2)^{1/3}\right] \rho t^*} \quad (4.105.)$$

The error in this spherical approximation (4.105.) is substantially greater in the final stages of the dissolution process (Figs. 4.10 and 4.11.) than in the previous work (Figs. 4.8 and 4.9.)



showed that dissolution of a distributed, multiparticulate system is only slightly affected by the dissolution behavior of the smallest particles. Substantial truncation at the lower end of the particle-size distribution had very little effect on the dissolution profile calculated (95). Thus, approximation error in the later stage of the single-particle dissolution does not introduce the same degree of error when applied to a nonuniformly distributed, multiparticulate system. The approximation (4.105.) should, therefore, yield considerably better results when applied to a multiparticulate system than might be judged from Fig. 4.10. This explains the choice of the particular weighting of the errors in the approximation procedure.

Table 4.1 summarizes the dissolution of the particle forms shown in Structures I-VI and gives formulas for the calculation of the equivalent spherical diameter in each case. The dissolution of these 10 crystal shapes can be described by three basic transformed equations of the form in the

excellent fit to the true dissolution. The weighted errors of the spherical approximation were calculated for various values of the shape ratio F and showed (Fig. 4.9.) that this choice for the approximation was satisfactory.

By using similar reasoning, it was found that the equation:

$$(w/w_0)^{1/3} = 1 - [2 - (2-F_1)^{1/3} (2-F_2)^{1/3}] t^* \quad (4.105.)$$

which is similar in form to 4.103. provides a good spherical approximation to 4.98, which describes the dissolution of rhombic and monoclinic prismatic particles. The equivalent spherical diameter in these cases is given by:

$$a = \frac{2Jt}{[2 - (2-F_1)^{1/3} (2-F_2)^{1/3}] \rho t^*} \quad (4.106.)$$

Figure 4.9

The error in this spherical approximation (4.105.) is substantially greater in the final stages of the dissolution process (Figs. 4.10 and 4.11.) than in the previous case (Figs. 4.8. and 4.9.). However, a previous study (96) showed that dissolution of a nonuniformly distributed, multiparticulate system is only slightly affected by the dissolution behaviour of the smallest particles. Substantial truncation at the lower end of the particle-size distribution had very little effect on the dissolution profile calculated (95). Thus, approximation error in the later stage of the single-particle dissolution does not introduce the same degree of error when applied to a nonuniformly distributed, multiparticulate system. The approximation (4.105.) should, therefore, yield considerably better results when applied to a multiparticulate system than might be judged from Fig. 4.10. This explains the choice of the particular weighting of the errors in the approximation procedure.

Table 4.1 summarizes the dissolution of the particle forms shown in Structures I-VI and gives formulas for the calculation of the equivalent spherical diameter in each case. The dissolution of these 10 crystal shapes can be described by three basic transformed equations of cubic form in time-

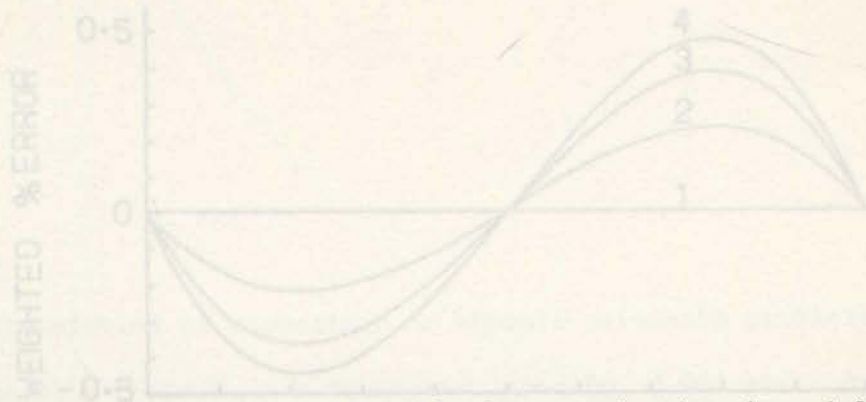


Figure 4.9

Graph of errors in the spherical approximation (Eq. 4.103) of the dissolution (Eq. 4.93) of an n-gonal prismatic particle (isometric, tetragonal, or hexagonal) or a rhombic pyramidal particle of different shape ratios. The error is weighted proportional to the fraction undissolved ( $w/w_0$ ). Key: 1,  $F = 1$ ; 2,  $F = \frac{1}{2}$ ; 3,  $F = \frac{1}{3}$ , and 4,  $F = \frac{1}{4}$ .

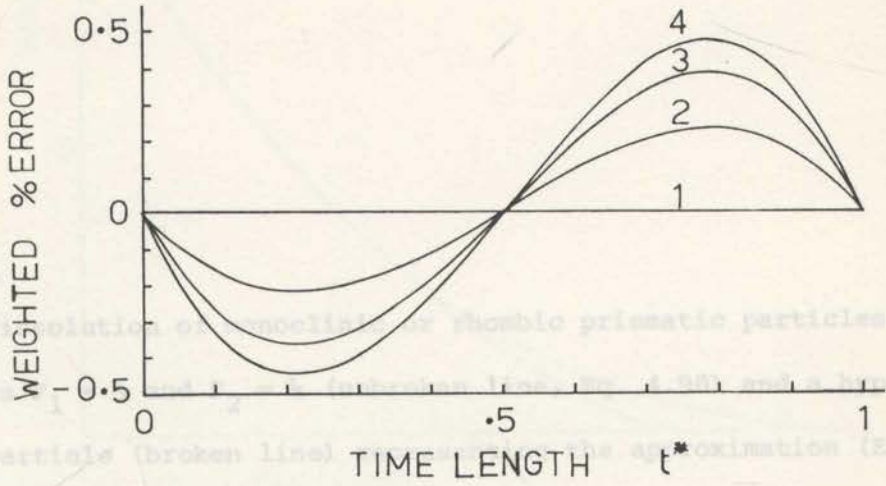


Figure 4.10

Dispersion of spherical particles with shape ratios 1, 2, 3, and 4. The approximation (Eq. 4.103) from which the equivalent spherical diameter is calculated (Eq. 4.104).

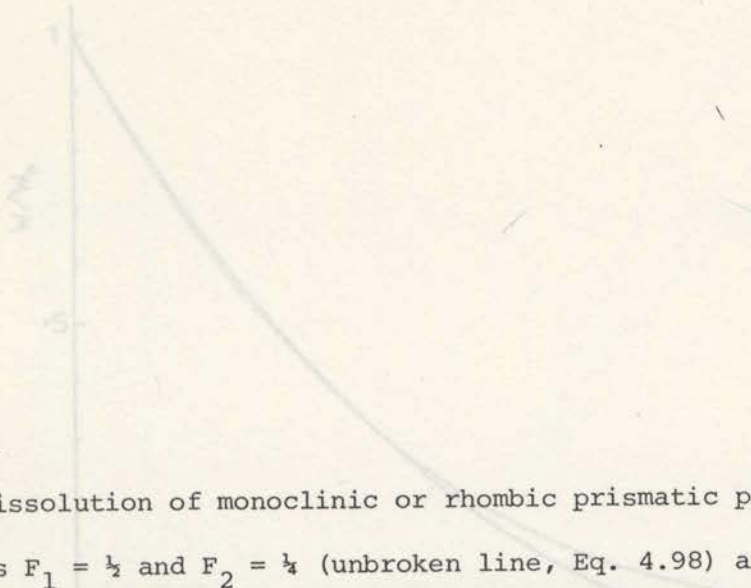


Figure 4.10

Dissolution of monoclinic or rhombic prismatic particles with shape ratios  $F_1 = \frac{1}{2}$  and  $F_2 = \frac{1}{4}$  (unbroken line, Eq. 4.98) and a hypothetical spherical particle (broken line) representing the approximation (Eq. 4.103) from which the equivalent spherical diameter is calculated (Eq. 4.104).

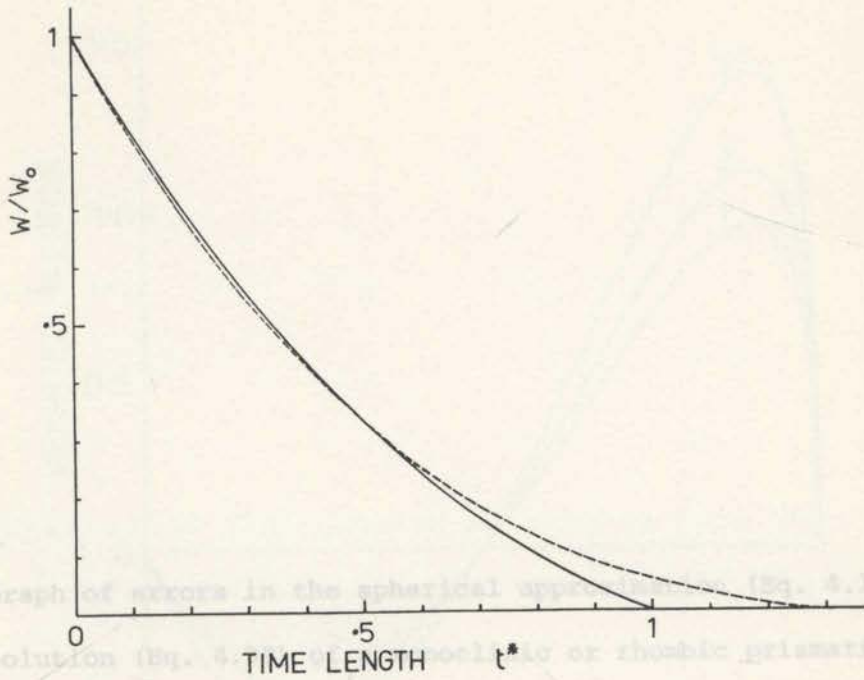


Figure 4.11

Graph of errors in the spherical approximation (Eq. 4.103) to the dissolution (Eq. 4.101) of a spherical or rhombic prismatic particle of different shape ratios. The error is weighted proportional to the fraction undissolved ( $w/w_0$ ). Keys 1,  $r_2 = \frac{1}{4}$ ,  $r_1 = \frac{1}{2}$ ; and 2,  $r_2 = \frac{1}{3}$ ,  $r_1 = \frac{1}{2}$ .





Figure 4.11

Graph of errors in the spherical approximation (Eq. 4.103) to the dissolution (Eq. 4.98) of a monoclinic or rhombic prismatic particle of different shape ratios. The error is weighted proportional to the fraction undissolved ( $w/w_0$ ). Key: 1,  $F_2 = \frac{1}{8}$ ; 2,  $F_2 = \frac{1}{4}$ ; and 3,  $F_2 = \frac{1}{3}$ .  $F_1 = \frac{1}{2}$ .

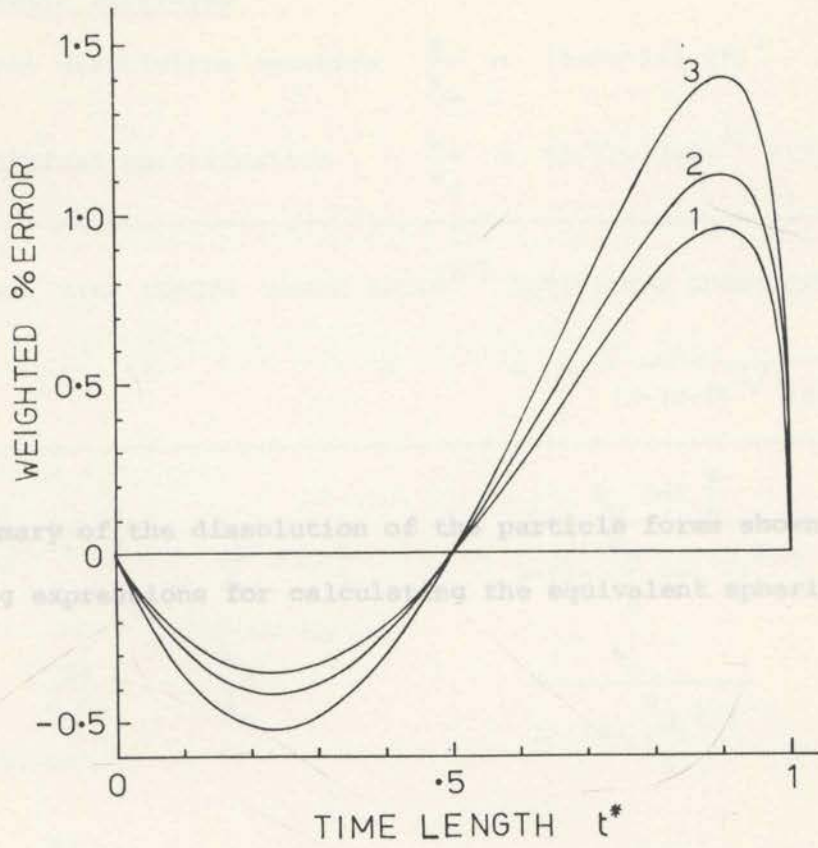


Table 4.1

Summary of the dissolution of the particle forms shown in Fig. 4.7, including expressions for calculating the equivalent spherical diameter.

DISSOLUTION OF PRISMATIC PARTICLES <sup>(a)</sup>

n-Gonal Prismatic Particles

Exact dissolution equation  $\frac{V}{V_0} = (1 - P t^*) (1 - t^*)^2$

Spherical approximation  $\frac{V}{V_0} = (1 - (2 - (2 - P)^{1/3}) t^*)^3$

CRYSTAL SYSTEM	TIME LENGTH	SHAPE RATIO <sup>(b)</sup>	EQUIVALENT SPHERICAL DIAMETER
	$t^*$	$P$	$a = \frac{2t^*}{(2 - (2 - P)^{1/3}) t^*}$

Table 4.1

Summary of the dissolution of the particle forms shown in Fig. 4.7. including expressions for calculating the equivalent spherical diameter.

Tetragonal (n=4)	$\frac{2b_0}{b_0} t^*$	$\frac{b_0}{1_0}$	$2 - (2 - \frac{b_0}{1_0})^{1/3}$
Isometric (n=4, $b_0=1_0$ )	$\frac{2b_0}{b_0} t^*$	1	$b_0$
Hexagonal (n=6)	$\frac{2\sqrt{3}b_0}{3b_0} t^*$	$\frac{b_0}{\sqrt{3}1_0}$	$2 - (2 - \sqrt{3} \frac{b_0}{1_0})^{1/3}$

Rhombic and Monoclinic Particles

Exact dissolution equations  $\frac{V}{V_0} = (1 - P_1 t^*) (1 - P_2 t^*) (1 - t^*)$

Spherical approximation  $\frac{V}{V_0} = (1 - (2 - (2 - P_1)^{1/3} (2 - P_2)^{1/3}) t^*)^3$

CRYSTAL SYSTEM	TIME LENGTH	SHAPE RATIOS	EQUIVALENT SPHERICAL DIAMETER
	$t^*$	$P_1 \quad P_2$	$a = \frac{2t^*}{(2 - (2 - P_1)^{1/3} (2 - P_2)^{1/3}) t^*}$

DISSOLUTION OF PRISMATIC PARTICLES (a)

n-Gonal Prismatic Particles

Exact dissolution equation  $\frac{w}{w_0} = (1-Ft^*)(1-t^*)^2$

Spherical approximation  $\frac{w}{w_0} = (1-(2-(2-F)^{1/3})t^*)^3$

CRYSTAL SYSTEM TIME LENGTH SHAPE RATIO<sup>(b)</sup> EQUIVALENT SPHERICAL DIAMETER

DISSOLUTION OF PYRAMIDAL PARTICLES

$$a = \frac{2Jt}{(2-(2-F)^{1/3})\rho t^*}$$

n-Gonal Pyramidal Particles

n-Gonal  $(\frac{2J}{b_0 \rho} \tan \frac{\pi}{n}) t$   $\frac{b_0}{l_0} \cot \frac{\pi}{n}$   $\frac{b_0 \cot \frac{\pi}{n}}{2-(2-\frac{b_0}{l_0} \cot \frac{\pi}{n})^{1/3}}$

Tetragonal (n=4)  $\frac{2J}{b_0 \rho} t$   $\frac{b_0}{l_0}$   $\frac{b_0}{2-(2-\frac{b_0}{l_0})^{1/3}}$

Isometric (n=4,  $b_0=l_0$ )  $\frac{2J}{b_0 \rho} t$  1  $b_0$

Hexagonal (n=6)  $\frac{2\sqrt{3}J}{3b_0 \rho} t$   $\sqrt{3} \frac{b_0}{l_0}$   $\frac{\sqrt{3} b_0}{2-(2-\sqrt{3} \frac{b_0}{l_0})^{1/3}}$

Rhombic and Monoclinic Particles

Exact dissolution equations  $\frac{w}{w_0} = (1-F_1 t^*)(1-F_2 t^*)(1-t^*)$

Spherical approximation  $\frac{w}{w_0} = (1-(2-(2-F_1)^{1/3}(2-F_2)^{1/3})t^*)^3$

CRYSTAL SYSTEM TIME LENGTH SHAPE RATIOS EQUIVALENT SPHERICAL DIAMETER

$$a = \frac{2Jt}{(2-(2-F_1)^{1/3}(2-F_2)^{1/3})\rho t^*}$$

Rhombic	$\frac{2J}{b_o}$	$\frac{b_o}{c_o}$	$\frac{b_o}{l_o}$	$\frac{b_o}{2 - (2 - \frac{b_o}{c_o})^{1/3} (2 - \frac{b_o}{l_o})^{1/3}}$
Monoclinic	$\frac{2J}{b_o \rho \sin \alpha} t$	$\frac{b_o}{c_o}$	$\frac{b_o}{l_o} \sin \alpha$	$\frac{b_o \sin \alpha}{2 - (2 - \frac{b_o}{c_o})^{1/3} (2 - \frac{b_o}{l_o} \sin \alpha)^{1/3}}$

DISSOLUTION OF PYRAMIDAL PARTICLES

n-Gonal Pyramidal Particles

(Isometric, Tetragonal, Hexagonal)

Exact dissolution equation	$\frac{w}{w_o} = (1-t^*)^3$
Time length	$t^* = \frac{2J \sqrt{h_o^2 + b_o^2/4}}{\rho} t$
Equivalent spherical diameter <sup>(c)</sup>	$a = \frac{h_o b_o}{\sqrt{h_o^2 + b_o^2/4}}$

Rhombic Pyramidal Particle

Exact dissolution equation	$\frac{w}{w_o} = (1-Ft^*) (1-t^*)^2$
Spherical approximation	$\frac{w}{w_o} = (1 - (2 - (2-F)^{1/3}) t^*)^3$
Time length	$t^* = \frac{2J \sqrt{h_o^2 + b_o^2/4}}{\rho} t$
Shape ratio	$F = \frac{1}{4} + \frac{3}{4} \sqrt{\frac{h_o^2 + c_o^2/4}{h_o^2 + b_o^2/4}}$

length of ...  
Equivalent spherical diameter  $a = \frac{1}{(2-(2-F))^{1/3} \sqrt{h_o^2 + b_o^2/4}}$   
polynomial in time in all cases.

The basic assumptions behind these derivations is that the rate of

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dissolution per unit surface area,  $J$ , remains constant during dissolution

(a) Figure 1 defines the quantities  $b_o$ ,  $c_o$ ,  $l_o$  and  $\alpha$  used.

(b) When  $F=1$ , i.e.  $\frac{b_o}{l_o} = \tan \frac{\pi}{n}$  then the equivalent spherical diameter is equal to the biggest sphere that can be contained in the prismatic body. The spherical approximation of the dissolution will then become exact. Later stages dissolution is slower than calculated. However, this

(c) This diameter is equal to the biggest sphere the pyramid can contain.

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expression given for isotropic conditions. Thus, the approximating curve (stippled line, Fig. 4.8. and 4.10) is above the calculated dissolution curve in the later stages. The true dissolution curve, because of the rounding effect, is above the calculated curve and hence closer to the approximation.

Excellent agreement between experimental and calculated results was obtained for the dissolution of a multiparticulate system of particles, approximately tetragonal prismatic in shape, when the respective spherical approximations were applied (130).

length or time. Dissolution can thus be described exactly by a third degree polynomial in time in all cases.

The basic assumptions behind these derivations is that the rate of dissolution per unit surface area,  $J$ , remains constant during dissolution and is the same everywhere at the interface of the dissolving crystal. This assumption can only be approximately true in practice under complete sink conditions. The higher activity at the crystal edges results in a larger  $J$  value in these areas and, therefore, a "rounding off" of the shape, so that during the later stages dissolution is slower than calculated. However, this should result in an improvement in the fit of the spherical approximation and could result in a closer fit to the real dissolution than the exact expression given for isotropic conditions. Thus, the approximating curve (stippled line, Fig. 4.8. and 4.10) is above the calculated dissolution curve in the later stages. The true dissolution curve, because of the rounding effect, is above the calculated curve and hence closer to the approximation.

Excellent agreement between experimental and calculated results was obtained for the dissolution of a multiparticulate system of particles, approximately tetragonal prismatic in shape, when the respective spherical approximations were applied (120).

Narrow-disperse powders.

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If the initial particle size distribution can be considered to be infinitely narrow, i.e. if all the particles are of the same initial size,  $r_0$ , then the multiparticulate model becomes identical to the simple particle model when considered on a "fraction undissolved versus time basis".

For example if the single particle dissolution follows the cube root model:

CHAPTER 5

MULTIPARTICULATE DISSOLUTION

The dissolution of a multiparticulate system is considerably more complex to characterize than a single particle system. There are two main reasons for this. One reason is that it may be difficult to account for the particle size distribution effect (see Chapter 6) which is present when the powder is not monodisperse. Another reason is that there may be significant interaction between the dissolving particles; the dissolution of a particle in the powder will, in general, influence the dissolution of other particles and often to varying degrees. In evaluating the intrinsic dissolution properties of a powder it is therefore necessary to establish experimental conditions which minimize such interactions. This is made possible by the "single layer-flow through" principle employed in the dissolution cell used in these experiments (see Chapter 7 and 10).

Under such sink conditions it is possible to establish a multiparticulate dissolution model based on a single particle dissolution model and the initial particle size distribution. Such a model should describe the intrinsic dissolution properties of a powder.

Monodisperse powders.

If the initial particle size distribution can be considered to be infinitely narrow, i.e. if all the particles are of the same initial size,  $r_0$ , then the multiparticulate model becomes identical to the simple particle model when considered on a "fraction undissolved versus time basis".

For example if the *single* particle dissolution follows the cube root model:



Derivation.  $w^{1/3} = w_0^{1/3} - kt$  (5.1.)

Then the multiparticulate model becomes:

$$W^{1/3} = W_0^{1/3} - n^{1/3} kt$$
 (5.2.)

where  $n = W_0/w_0$  (5.3.)

is the number of particles in the powder. Eqs. 5.1. and 5.2. are thus identical in the sense that:

$$W/W_0 = w/w_0 = \left(1 - \frac{k}{1/3}t\right)^3$$
 (5.4.)

Polydisperse powders.

-94

Several investigators (107,92) have considered the problem of exactly describing the dissolution profile of powders in relation to their particle size distribution. These authors have been concerned with powders initially consisting of particles with log-normal size distribution. Earlier attempts made use of approximations (107) or computer simulations (92). More recently, Brooke (93,94) developed an equation that permits calculation of the dissolution profile of such powders without the aid of a computer. This equation was later presented in a form to account for trimcated log-normal distributions.

A general equation that exactly describes the entire dissolution profile of powders under sink conditions is derived in the following section. The equation is valid for particles having any initial size distribution and dissolving according to any explicit single particle model. The equation is then applied to obtain an exact expression for a log-normal powder considering the cube root model.

$$f(w) = \frac{h(w)}{\int_0^w h(x) dx} \text{ for } w > 0$$
 (5.5.)

Derivation, General Case.

Consider a powder consisting of particles which initially (t=0) have a weight density (probability distribution)  $f_0(w_0)$ . Let the particles dissolve independently of each other according to:

$$w = g(w_0, t, A) \tag{5.5.}$$

where  $w$  and  $w_0$  are the particle weights at time  $t$  and  $t=0$ , respectively and  $A$  collectively represents dissolution parameters such as solubility, particle density and particle shape factors. The inverse dissolution function is defined as:

$$w_0 = g^{-1}(w, t, A) \tag{5.6.}$$

Using the rules of transformation of independent variables (121) the particle weight density function at time  $t$  becomes:

$$h(w) = f_0 [g^{-1}(w, t, A)] \frac{d}{dw} g^{-1}(w, t, A) \tag{5.7.}$$

For 5.7. to hold, the following conditions for  $g$  must be satisfied: (a)  $g^{-1}$  must be a strictly increasing function of  $w$  for all  $t$  values, (b)  $g$  must decrease strictly with time until equal to zero, (c)  $g$  must remain equal to zero beyond that time. The latter two conditions ensure that the dissolution function reflects the actual physical conditions of the dissolution process. The first condition will rarely be violated because in application,  $g$  is nearly always a strictly increasing function of  $w_0$  for all  $t$  values. It is obvious that the second condition must be met by any dissolution equation.

The third condition is not satisfied for most equations in the literature (97,106,/). To overcome this problem, it is necessary to redefine the particle weight density function such that it is generally applicable:

$$f(w) = \frac{h(w)}{\int_0^\infty h(w) dw} \quad \text{for } w > 0 \tag{5.8.}$$

These values are not to be absolute but rather represent limits giving the best fit when the actual particle distribution is approximated by any particular function. Therefore, they also represent truncation limit of the function.

$$f(w) = 0 \quad \text{for } w \leq 0 \quad (5.9.)$$

where division by the integral is necessary to satisfy the condition that the total integral (from  $-\infty$  to  $+\infty$ ) of  $f(w)$  must be equal to 1.

The weight of undissolved powder,  $W$ , at any time,  $t$ , is equal to the product of the number of particles remaining,  $N_t$ , and the mean particle weight, which for a large number of particles is the same as the expected value of  $w$ ,  $E_t(w)$ . Therefore, the following general equation can be written:

$$W = N_t E_t(w) \quad (5.10.)$$

The number of particles remaining at time  $t$  is:

$$N_t = N_0 \int_0^{\infty} h(w) dw \quad (5.11.)$$

where the initial numbers of particles  $N_0$  is equal to the initial powder weight,  $W_0$ , divided by the initial mean particle weight:

$$N_0 = \frac{W_0}{\int_0^{\infty} w f_0(w) dw} \quad (5.12.)$$

The mean particle weight as time  $t$  is given by:

$$E_t(w) = \frac{\int_{-\infty}^{\infty} wf(w) dw}{\int_0^{\infty} h(w) dw} \quad (5.13.)$$

which according to 5.8. can be written:

$$E_t(w) = \frac{\int_0^{\infty} wh(w) dw}{\int_0^{\infty} h(w) dw} \quad (5.14.)$$

substituting Eqs. 5.11., 5.12. and 5.14. into Eq. 5.10. yields:

$$\frac{W}{W_0} = \frac{\int_0^{\infty} wh(w) dw}{\int_0^{\infty} w f_0(w) dw} \quad (5.15.)$$

This equation relates to unbounded particle weight distributions. In practice, the distribution is always bounded, so the limits of the integration must be changed accordingly.

Let  $m_0$  and  $M_0$  denote the initial weights of the smallest and largest particle, respectively.<sup>1</sup> These values then represent the lower and

1. These values are not intended to be absolute but rather represent limits giving the best fit when the actual particle distribution is approximated by any particular function. Therefore, they also represent truncation limits of the function.





in the numerator of Eqs. 5.17. and 5.22. reduces to the constant integral in the denominator of zero time, thus making the ratio  $W/W_0$  equal to 1 as expected.

The general mathematical models expressed by 5.17. and 5.22. require the use of a computer for numerical evaluation because they are in integral form. However, if the initial particle weight or particle-size distribution can be approximated by some simple function, then the model can often be solved in terms of an expression suitable for evaluating without the use of a computer.

Special Case: Log-Normal Powders and the Cube Root Model.

Figure 5.1. Carstensen and Musa (92 ) have pointed out in their review of the literature that procedures such as milling, grinding and precipitation, which are based on random processes, produce particles having skewed distribution functions which often can be approximated by a log-normal distribution. It is therefore of interest to examine this special case.

Consider a powder consisting of spherical particles of initial diameters  $a_0$ , distributed such that  $\ln a_0$  approximates a normal distribution with mean  $\mu$  and standard deviation  $\sigma$ , truncated at  $\ln d_0 = \mu - i\sigma$  and  $\ln D_0 = \mu + j\sigma$ , (Fig. 5.1.) where  $i$  and  $j$  are trimcation parameters. The density function of  $\ln a_0$  is then given by:

$$u(\ln a_0) = \frac{N(\ln a_0, \mu, \sigma)}{\int_{\ln a_0 = \mu - i\sigma}^{\ln a_0 = \mu + j\sigma} N(\ln a_0, \mu, \sigma) d \ln a_0} \quad (5.27.)$$

$$\mu - i\sigma \leq \ln a_0 \leq \mu + j\sigma$$

where the normal density function  $N$  is defined as:

$$N(x, \mu, \sigma) = \frac{1}{\sigma\sqrt{2\pi}} \text{EXP} \left[ -(x-\mu)^2 / 2\sigma^2 \right] \quad (5.28.)$$

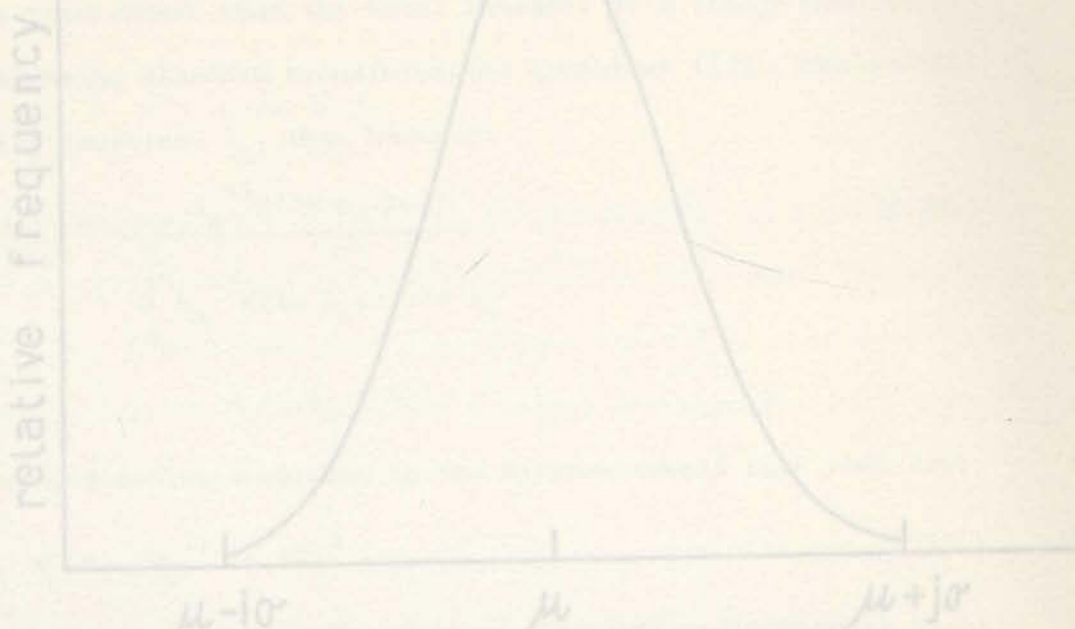
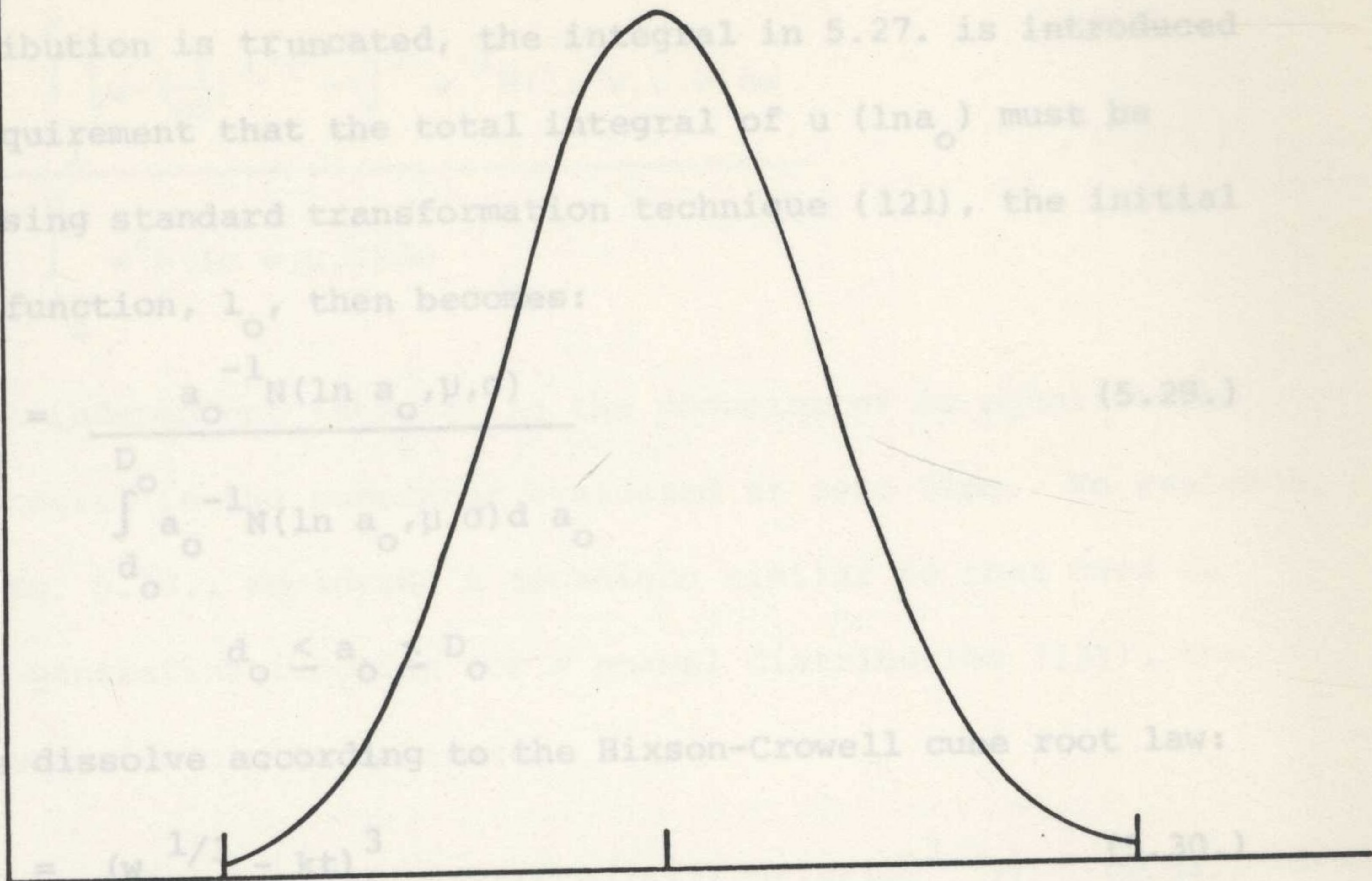


Figure 5.1. Illustration of the parameters in a truncated log-normal distribution.

relative frequency



ln(particle diameter)

Because the distribution is truncated, the integral in 5.27. is introduced to satisfy the requirement that the total integral of  $u(\ln a_0)$  must be equal to 1. By using standard transformation technique (121), the initial particle density function,  $l_0$ , then becomes:

$$l_0(a_0) = \frac{a_0^{-1} N(\ln a_0, \mu, \sigma)}{\int_{d_0}^D a_0^{-1} N(\ln a_0, \mu, \sigma) da_0} \quad (5.29.)$$

Let the particles dissolve according to the Hixson-Crowell cube root law:

$$w = (w_0^{1/3} - kt)^3 \quad (5.30.)$$

where  $k$  is a positive constant. The inverse dissolution function is then:

$$w_0 = (w^{1/3} + kt)^3 \quad (5.31.)$$

After the initial particle-size distribution (5.29.) and the particle dissolution equation (5.30. and its inverse 5.31.) are defined, the relationship giving the dissolution profile can then be derived by means of 5.22. in the following way:

After inserting Eqs. 5.29.-5.31. into Eq. 5.22., the integral in the numerator,  $I_n$  of 5.22. becomes:

$$I_n = \frac{\int_{R_1}^{R_2} \left[ \left( \frac{D\pi}{6} \right)^{1/3} w - kt \right]^3 w^{-1} N(\ln w, \mu, \sigma) dw}{\int_{d_0}^D w^{-1} N(\ln w, \mu, \sigma) dw} \quad (5.32.)$$

It was indicated previously that  $W/W_0 = I_n / (I_n)_{t=0}$ . By using this fact,  $W/W_0$  can be written:



Because the distribution is truncated, the integral in 5.27. is introduced to satisfy the requirement that the total integral of  $u(\ln a_0)$  must be equal to 1. By using standard transformation technique (121), the initial particle density function,  $l_0$ , then becomes:

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$d_0 \leq a_0 \leq D_0$

Let the particles dissolve according to the Hixson-Crowell cube root law:

$$w = (w_0^{1/3} - kt)^3 \quad (5.30.)$$

where  $k$  is a positive constant. The inverse dissolution function is then:

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After inserting Eqs. 5.29.-5.31. into Eq. 5.22., the integral in the numerator,  $I_n$  of 5.22. becomes:

$$I_n = \frac{\int_{R_1}^{R_2} \left[ \left( \frac{\rho\pi}{6} \right)^{1/3} w - kt \right]^3 w^{-1} N(\ln w, \mu, \sigma) dw}{\int_{d_0}^D w^{-1} N(\ln w, \mu, \sigma) dw} \quad (5.32.)$$

It was indicated previously that  $W/W_0 = I_n / (I_n)_{t=0}$ . By using this fact,

$W/W_0$  can be written:

$$\frac{W}{W_0} = \frac{\int_{R_1}^{R_2} \left[ w - \left( \frac{6}{\rho\pi} \right)^{1/3} kt \right]^3 w^{-1} N(\ln w, \mu, \sigma) dw}{\int_{D_0}^D w^2 N(\ln w, \mu, \sigma) dw} \quad (5.33.)$$

In 5.33., the time-independent integral in the denominator is equal to the time-dependent integral in the numerator evaluated at zero time. To evaluate the numerator of Eq. 5.33., employing a technique similar to that used to derive the moment-generating function for a normal distribution (121), the following useful equation can be obtained:

$$\int_q^r x^s N(\ln x, \mu, \sigma) dx = (F(A) - F(B)) \text{EXP} \left[ (s+1) (\mu + (s+1)\sigma^2/2) \right] \quad (5.34.)$$

where  $A = (\ln r - \mu)/\sigma - (s+1)\sigma \quad (5.35.)$

$B = (\ln q - \mu)/\sigma - (s+1)\sigma \quad (5.36.)$

and the function  $F(\ )$  is the area under the standard normal curve function given by:

$$F(x) = \frac{1}{\sqrt{2\pi}} \int_{-\infty}^x e^{-x^2/2} dx \quad (5.37.)$$

Thus, to evaluate the numerator,  $I$ , in 5.33., the term  $(w-Kt)^3 w^{-1}$  is expanded as  $w^2 - 3(Kt)w + 3(Kt)^2 w^{-1}$  (letting  $K = (6/\rho\pi)^{1/3} k$  for simplicity). The above formula is then applied term by term by putting  $s = 2, 1, 0,$  and  $-1$ ;  $r = R_2$  and  $q = R_1$  and by treating  $t$  as a constant for the purpose of integration. The following equation is then derived using  $W/W_0 = I_n / (I_n)_{t=0}$  and noting that  $\ln D_0 = \mu - i\sigma$  and  $\ln D_0 = \mu + j\sigma$ :

is similar to the equation  $\dots$  used previously (92-94) in which  $\gamma$  denotes the time length. Hooke (94) derived an equation similar to 5.33 for log-normal powders. Directions were given for the changes required in the equation at the critical time. However, in his equation the first term is constant, equal

$$\begin{aligned}
 W/W_0 = & \left[ \frac{F\left(\frac{T_2 - \mu}{\sigma} - 3\sigma\right) - F\left(\frac{T_1 - \mu}{\sigma} - 3\sigma\right)}{F(j-3\sigma) - F(-i-3\sigma)} \right] \\
 -3(Kt) & \left[ \frac{F\left(\frac{T_2 - \mu}{\sigma} - 2\sigma\right) - F\left(\frac{T_1 - \mu}{\sigma} - 2\sigma\right)}{F(j-3\sigma) - F(-i-3\sigma)} \right] \text{EXP}\left[-\mu - 5\sigma^2/2\right] \quad (5.38.) \\
 +3(Kt)^2 & \left[ \frac{F\left(\frac{T_2 - \mu}{\sigma} - \sigma\right) - F\left(\frac{T_1 - \mu}{\sigma} - \sigma\right)}{F(j-3\sigma) - F(-i-3\sigma)} \right] \text{EXP}\left[-2\mu - 4\sigma^2\right] \\
 -(Kt)^3 & \left[ \frac{F\left(\frac{T_2 - \mu}{\sigma}\right) - F\left(\frac{T_1 - \mu}{\sigma}\right)}{F(j-3\sigma) - F(-i-3\sigma)} \right] \text{EXP}\left[-3\mu - 9\sigma^2/2\right]
 \end{aligned}$$

where:

$$T_1 = \mu - i\sigma \text{ for } \ln Kt \leq \mu - i\sigma \quad (5.39.)$$

$$T_1 = \ln Kt \text{ for } \ln Kt > \mu - i\sigma \quad (5.40.)$$

$$T_2 = \mu + j\sigma \text{ for } \ln Kt \leq \mu + j\sigma \quad (5.41.)$$

$$T_2 = \ln Kt \text{ for } \ln Kt > \mu + j\sigma \quad (5.42.)$$

$$\text{and } K = (6/\rho\pi)^{1/3} k \quad (5.43.)$$

The change in T, at timelength  $Kt = \text{EXP}(\mu - i\sigma)$  (critical time) corresponds to the time when the smallest particles, initially having a diameter  $d_0 = \text{EXP}(\mu - i\sigma)$ , begin to disappear. The change at  $Kt = \text{EXP}(\mu + j\sigma) = D_0$  signifies the end of the dissolution process, so  $W/W_0$  becomes zero after that timelength.

Equation 5.38. describes the complete dissolution profile of log-normal powders and any sieve fraction of such powders. It assumes that the particles dissolve according to the cube root law (5.30.) which also can be written in the form  $a = a_0 - Kt$  for spherical particles, where  $a_0$  and  $a$  are the particle diameters at time zero and t, respectively. This relationship is similar to the equation  $a = a_0 - \tau$  used previously (92-94) in which  $\tau$  denotes the timelength.

Brooke (94) derived an equation similar to 5.38 for log-normal powders. Directions were given for the changes required in the equation at the critical time. However, in his equation the first term is constant, equal

to 1. Therefore, his equation is incorrect if applied to dissolution after the critical time. The error so introduced becomes quite substantial for large timelengths and large values of  $\sigma$ .

Brooke calculated values of  $W/W_0$  for various values of  $\tau/\text{EXP}(\mu)$  (the latter terminology corresponds to  $\text{EXP}(-\mu)Kt$  used here). This procedure represents an ingenious method of "scaling time" (by the factor  $\text{EXP}(-\mu)K$ ) so the dissolution profile becomes independent of the parameters  $\mu$  and  $k$  (Eq. 5.30.) and only depends on  $\sigma$ , enabling the effect of  $\sigma$  alone to be assessed. In the present work, however, cube root type plots of  $(W/W_0)^{1/3}$  versus  $\text{EXP}(-\mu)Kt$  are used for better comparison with the fundamental particle dissolution equation (5.30) which obeys the cube root law.

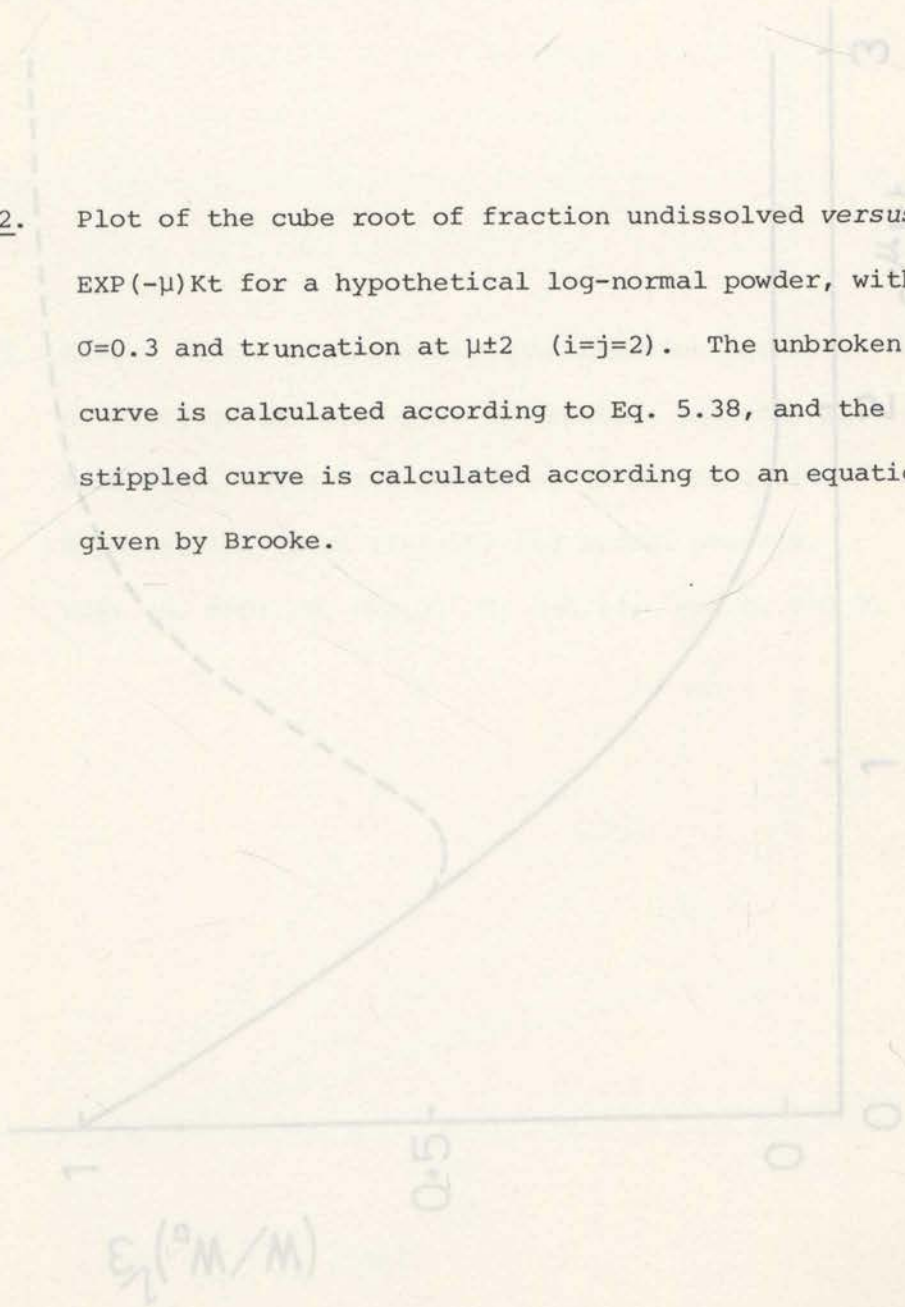
Figure 5.2. shows such a plot for powders truncated at  $\mu \pm 2\sigma$  ( $i=j=2$ ) and having  $\sigma=0.3$ . The curvature of the unbroken line, calculated according to Eq. 5.38. is logically expected. The stippled line represents the dissolution profile calculated according to Brooke's equation (Eq. 4 of Ref. 94 ). The two profiles are, as expected, identical until critical time ( $\text{EXP}(-\mu)Kt=0.5488$ ), but the later part clearly demonstrates the limitation of his equation.

Figure 5.3. demonstrates the effect of  $\sigma$  on the dissolution profile for a powder initially having an ideal distribution ( $i=j=10$ ). Powders of uniform particle size, i.e.,  $\sigma=0$ , give linear cube root plots as expected, while the deviation from linearity is significant for larger  $\sigma$  values. An increase in  $\sigma$  results in a decrease in the initial slopes of the curves, which is consistent with calculations made by Brooke (94 ). Among powders having the same logarithmic mean diameter,  $\mu$ , those with broadest distribution will have the slowest initial release rate.

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3. A Cyber 76 digital computer equipped with Calcomp platter was used for calculations and plots. Numerical evaluations were tested to six digits.

Figure 5.2. Plot of the cube root of fraction undissolved versus  $\text{EXP}(-\mu)Kt$  for a hypothetical log-normal powder, with  $\sigma=0.3$  and truncation at  $\mu \pm 2$  ( $i=j=2$ ). The unbroken curve is calculated according to Eq. 5.38, and the stippled curve is calculated according to an equation given by Brooke.



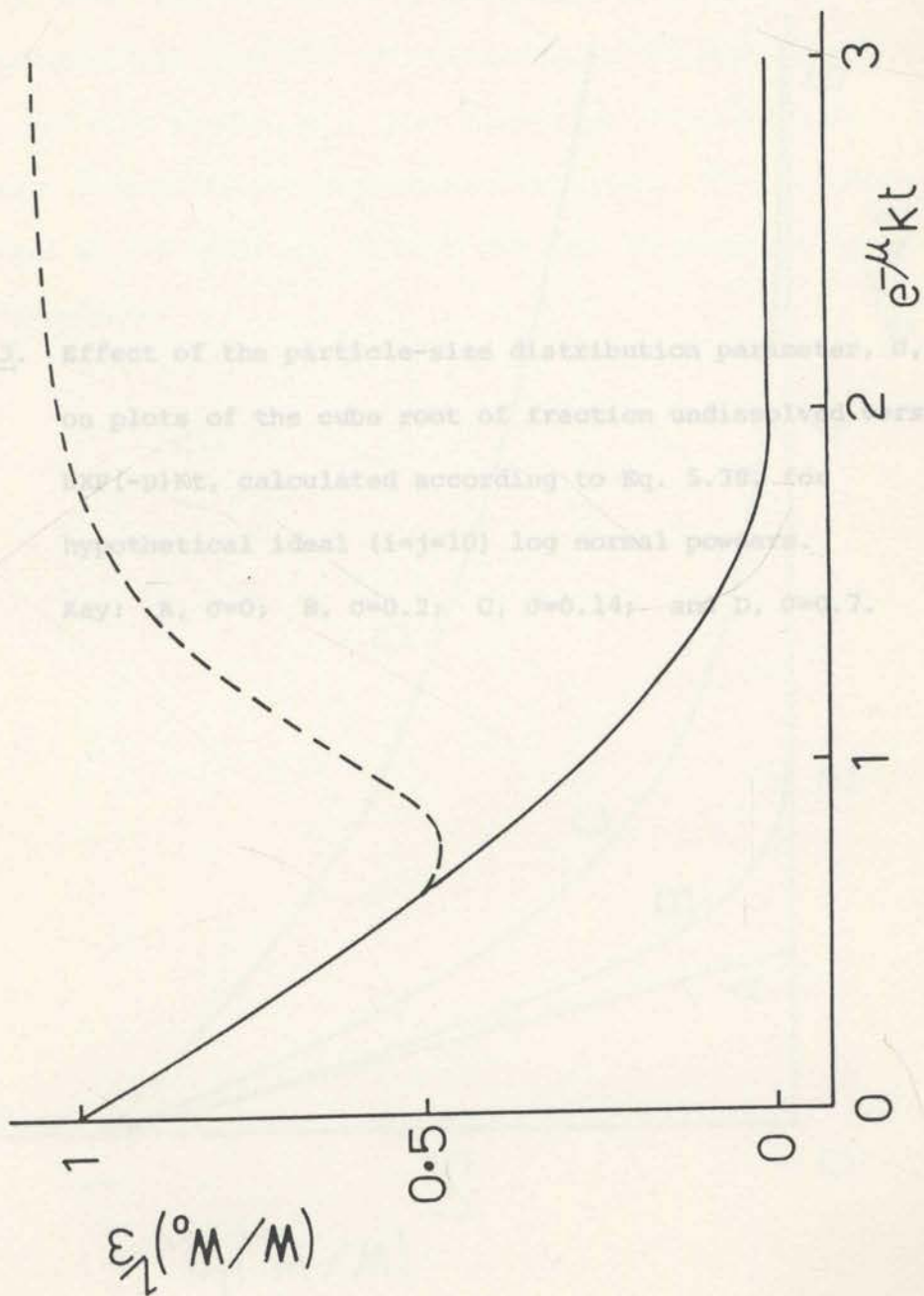


Figure 5.3. Effect of the particle-size distribution parameter,  $\sigma$ , on plots of the cube root of fraction undissolved versus  $e^{-\kappa kt}$ , calculated according to Eq. 5.18, for hypothetical ideal ( $i=j=10$ ) log normal powders. Key: A,  $\sigma=0$ ; B,  $\sigma=0.2$ ; C,  $\sigma=0.14$ ; and D,  $\sigma=0.7$ .

Figure 5.3. Effect of the particle-size distribution parameter,  $\sigma$ , on plots of the cube root of fraction undissolved versus  $\text{EXP}(-\mu)Kt$ , calculated according to Eq. 5.38. for hypothetical ideal ( $i=j=10$ ) log normal powders.  
Key: A,  $\sigma=0$ ; B,  $\sigma=0.2$ ; C,  $\sigma=0.14$ ; and D,  $\sigma=0.7$ .

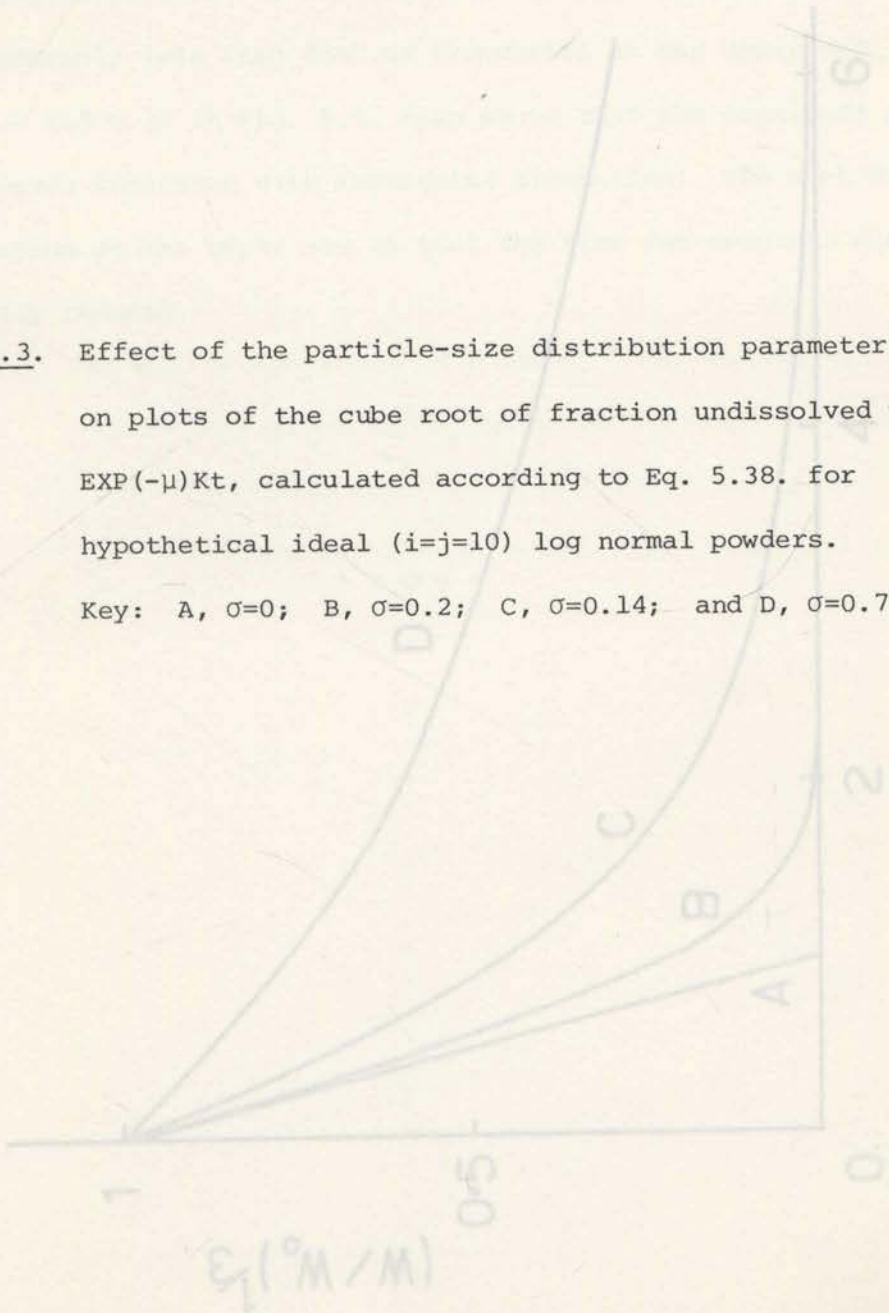


Figure 5.4. and 5.5. show the effect of truncation on the dissolution profile when the initial  $\ln$  (particle diameter) - distribution approximates a normal distribution ( $\sigma=0.5$ ) with various degrees of upper (Fig. 5.4.) and lower (Fig. 5.5.) end truncation. Comparison of the two figures indicates that the effect of truncation at the low end is very small and considerably less than that of truncation at the upper end. The curves for  $\gamma=10, 1, 0.3$  and  $0.25$  in Fig. 5.4. also shows that the magnitude of the slopes of the curves increases with increasing truncation. The most marked effect of truncation at the upper end is that the time for complete dissolution is drastically reduced.

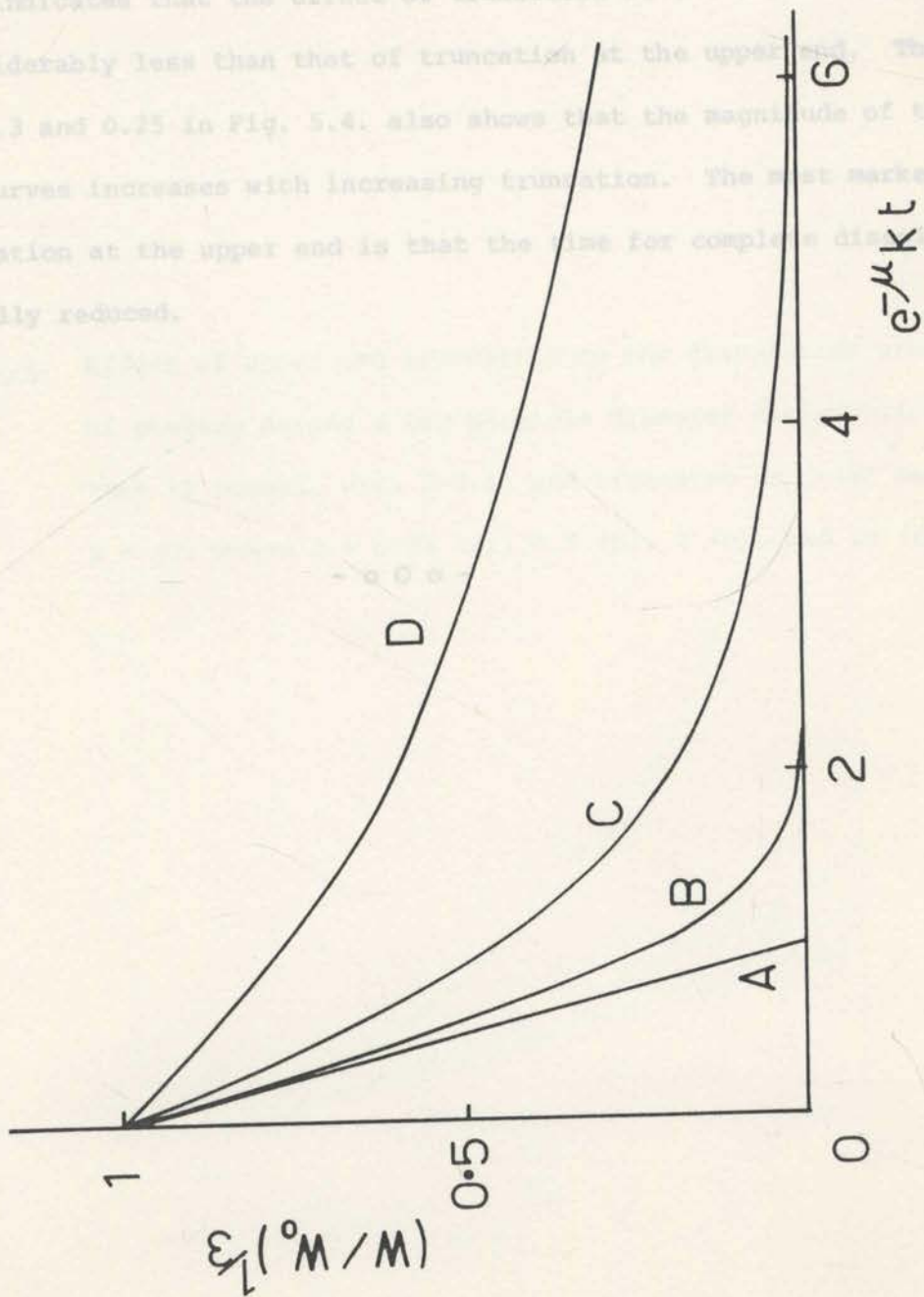
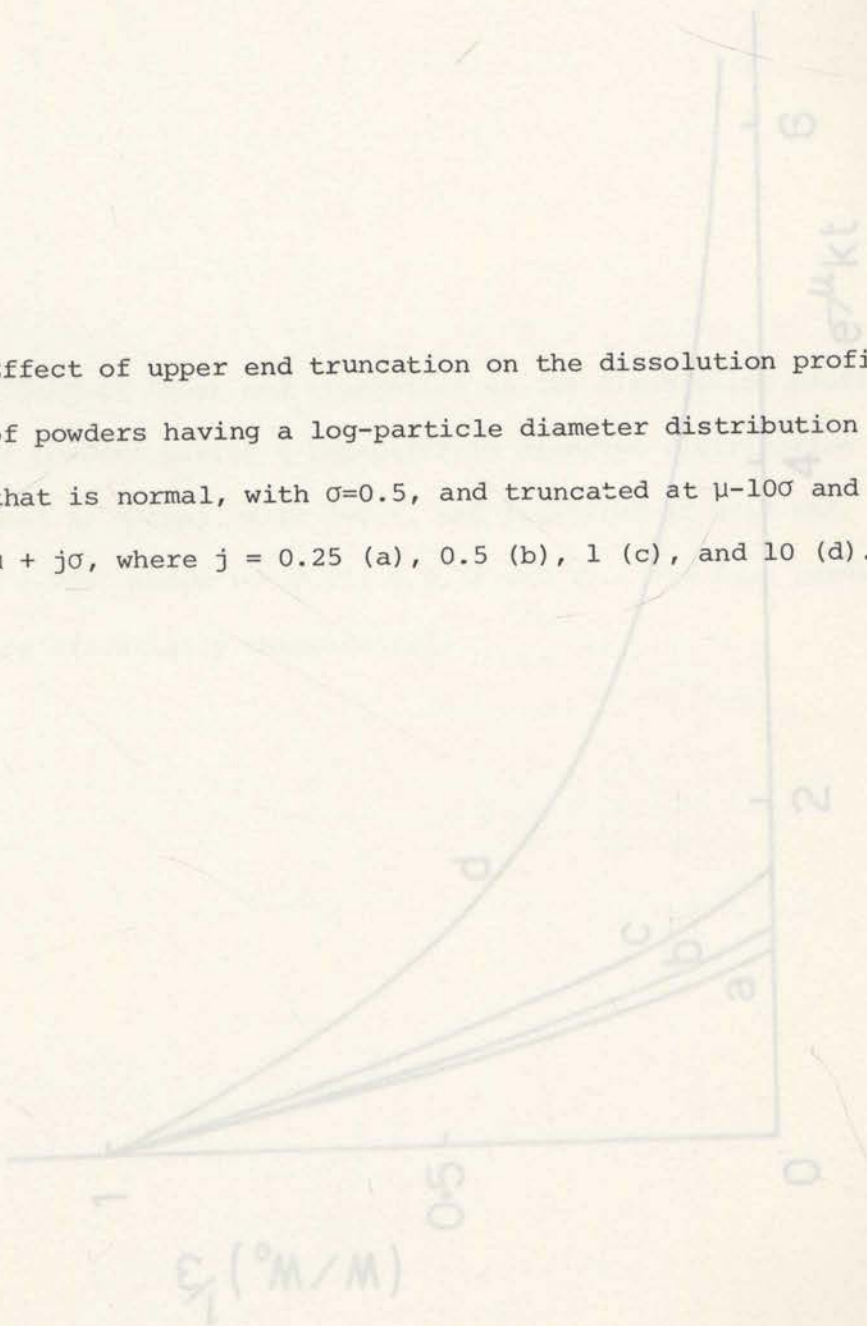




Figure 5.4. and 5.5. show the effect of truncation on the dissolution profile when the initial  $\ln$  (particle diameter) - distribution approximates a normal distribution ( $\sigma=0.5$ ) with various degrees of upper (Fig. 5.4.) and lower (Fig. 5.5.) end truncation. Comparison of the two figures indicates that the effect of truncation at the low end is very small and considerably less than that of truncation at the upper end. The curves for  $j=10,1,0.3$  and  $0.25$  in Fig. 5.4. also shows that the magnitude of the slopes of the curves increases with increasing truncation. The most marked effect of truncation at the upper end is that the time for complete dissolution is drastically reduced.

Figure 5.4. Effect of upper end truncation on the dissolution profile of powders having a log-particle diameter distribution that is normal, with  $\sigma=0.5$ , and truncated at  $\mu-10j$  and  $\mu + j\sigma$ , where  $j = 0.25$  (a),  $0.5$  (b),  $1$  (c), and  $10$  (d).  
- o o o -

Figure 5.4. Effect of upper end truncation on the dissolution profile of powders having a log-particle diameter distribution that is normal, with  $\sigma=0.5$ , and truncated at  $\mu-10\sigma$  and  $\mu + j\sigma$ , where  $j = 0.25$  (a),  $0.5$  (b),  $1$  (c), and  $10$  (d).



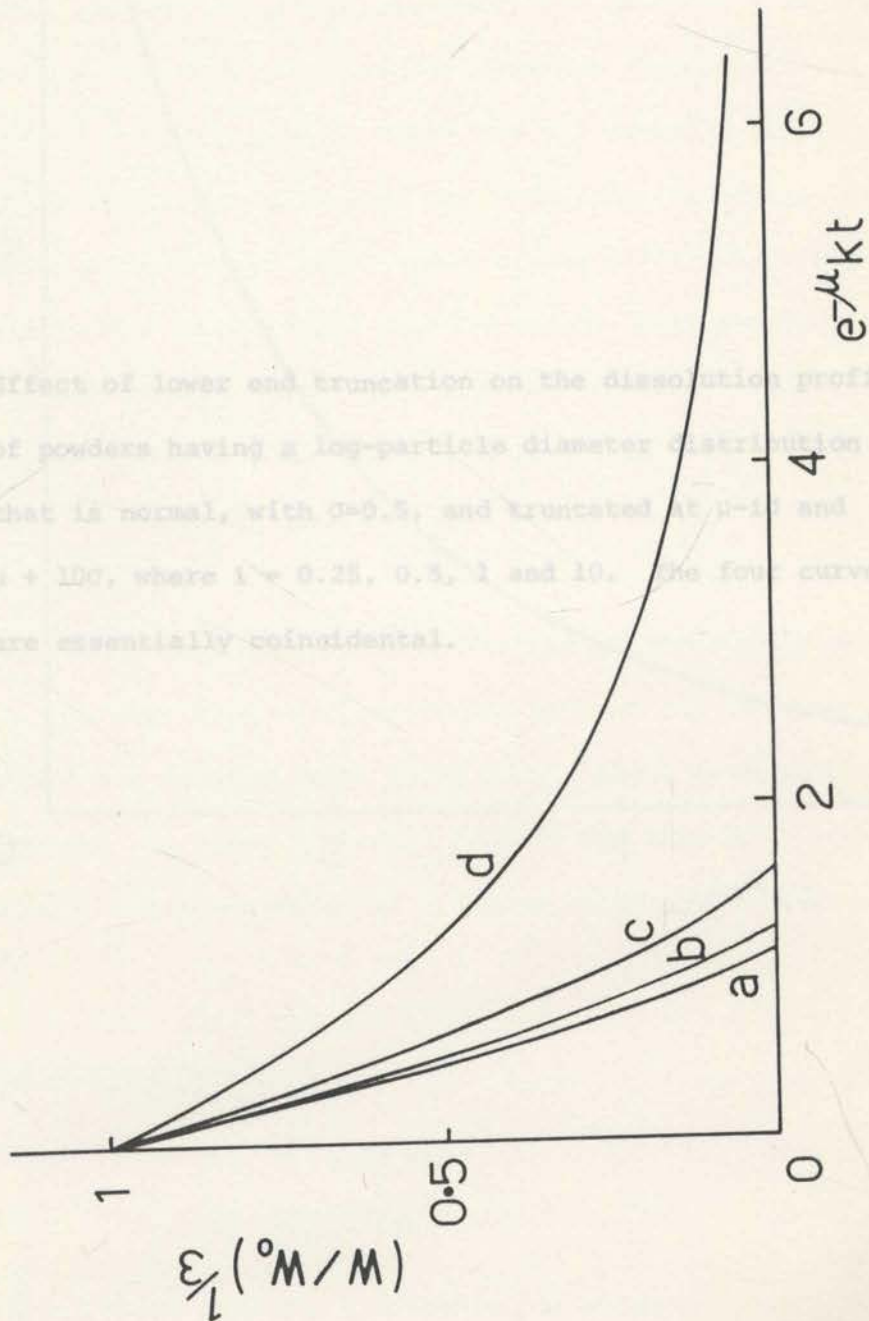


Figure 5.5. Effect of lower and truncation on the dissolution profile of powders having a log-particle diameter distribution that is normal, with  $\sigma=0.5$ , and truncated at  $\mu-1\sigma$  and  $\mu+1\sigma$ , where  $l = 0.25, 0.5, 1$  and  $10$ . The four curves are essentially coincidental.

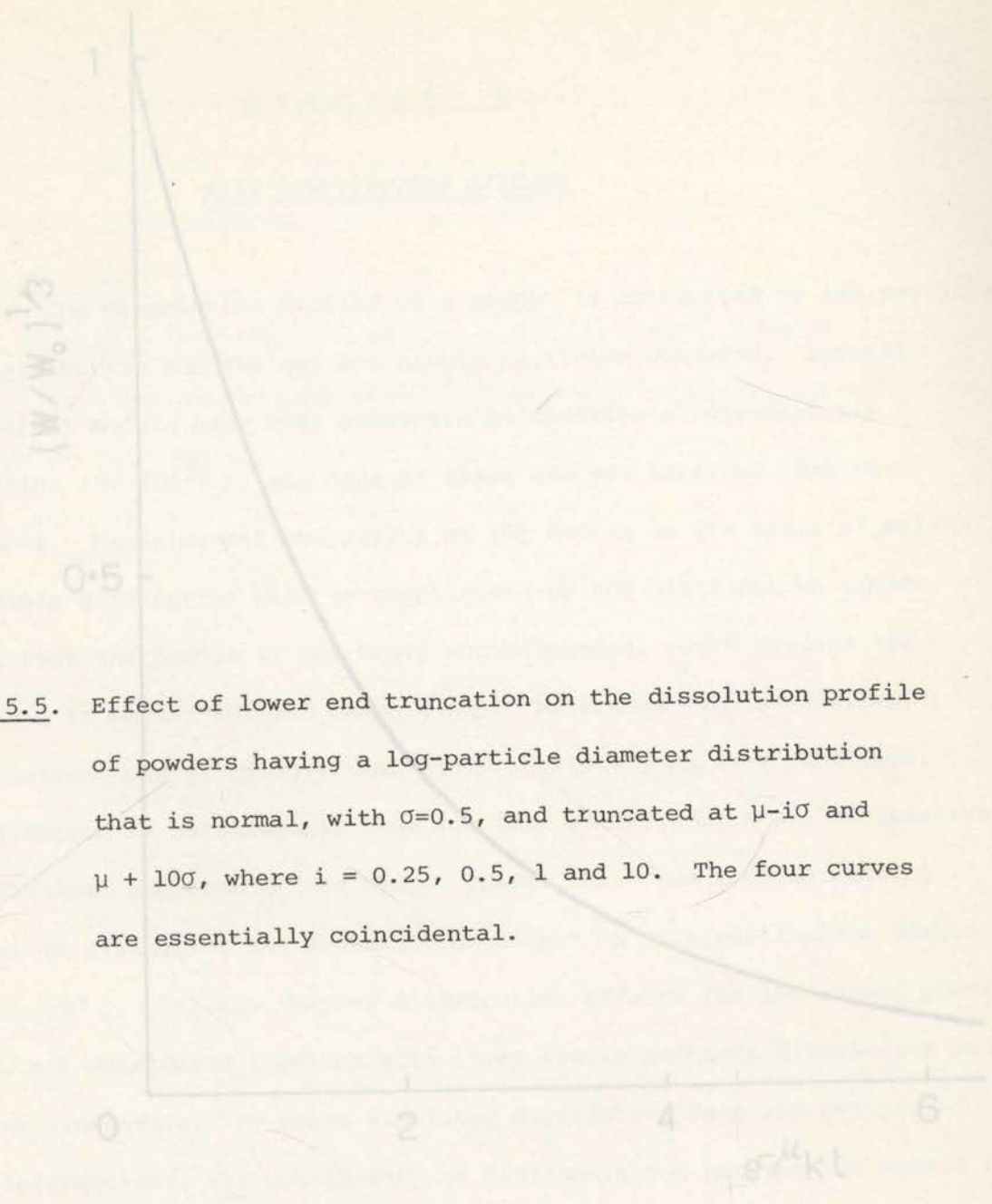


Figure 5.5. Effect of lower end truncation on the dissolution profile of powders having a log-particle diameter distribution that is normal, with  $\sigma=0.5$ , and truncated at  $\mu-i\sigma$  and  $\mu + 10\sigma$ , where  $i = 0.25, 0.5, 1$  and  $10$ . The four curves are essentially coincidental.

CHAPTER 6

SIZE DISTRIBUTION EFFECTS

$(w/w_0)^{1/3}$

0.5

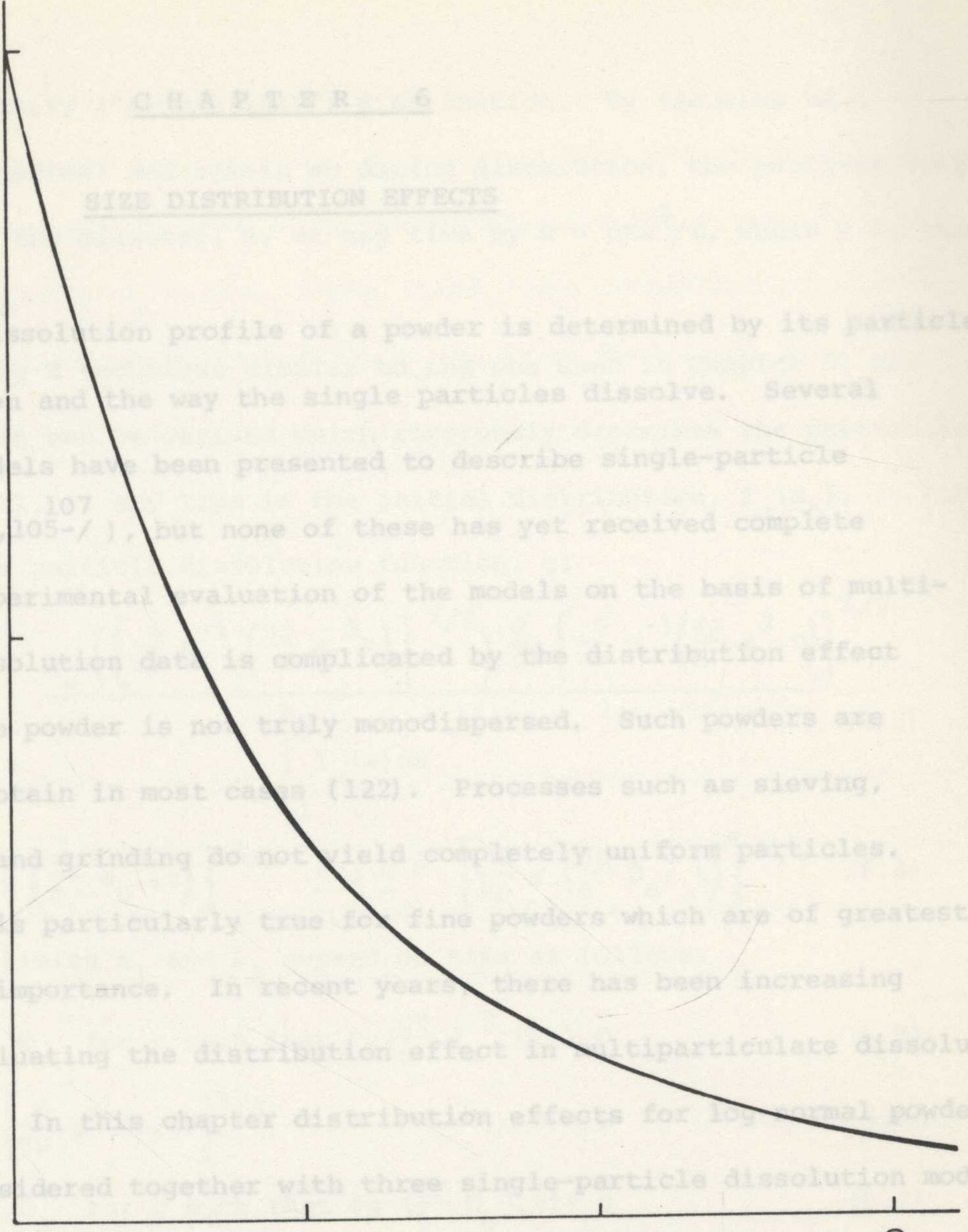
0

2

4

6

$e^{-\mu kt}$



Theoretical Considerations

Let:  $w = g(w_0, t)$  (6.1)

and  $w_0 = g^{-1}(w, t)$  (6.2)

describe the dissolution equation and inverse dissolution equation, respectively, for a single particle, where  $w$  and  $w_0$  are the particle weights at time  $t$  and 0, respectively. Further, let  $l_0$  ( $a_0$ ) denote the initial ( $t=0$ )

CHAPTER 6

SIZE DISTRIBUTION EFFECTS

The dissolution profile of a powder is determined by its particle-size distribution and the way the single particles dissolve. Several mathematical models have been presented to describe single-particle dissolution (97,105-107), but none of these has yet received complete acceptance. Experimental evaluation of the models on the basis of multiparticulate dissolution data is complicated by the distribution effect present when the powder is not truly monodispersed. Such powders are impossible to obtain in most cases (122). Processes such as sieving, precipitation, and grinding do not yield completely uniform particles. This situation is particularly true for fine powders which are of greatest pharmaceutical importance. In recent years, there has been increasing interest in evaluating the distribution effect in multiparticulate dissolution (92-96, 107). In this chapter distribution effects for log-normal powder systems are considered together with three single-particle dissolution models from the literature. By using simulated dissolution data and particle-size distributions, the possibility of distinguishing between the models is investigated.

Theoretical Considerations

Let:  $w = g(w_0, t)$  (6.1)

and  $w_0 = g^{-1}(w, t)$  (6.2)

describe the dissolution equation and inverse dissolution equation, respectively, for a single particle, where  $w$  and  $w_0$  are the particle weights at time  $t$  and  $0$ , respectively. Further, let  $l_0$  ( $a_0$ ) denote the initial ( $t=0$ )

particle-size density ("probability") distribution. By assuming that particles are spherical and remain so during dissolution, the particle weight,  $w$ , is related to the diameter,  $a$ , at any time by  $w = \rho\pi a^3/6$ , where  $\rho$  is the particle density.

By using a technique similar to the one used in Chapter 5, the following equation can be derived which rigorously describes the particle-size distribution,  $l(a)$ , at any time if the initial distribution,  $l_0(a_0)$ , is known together with the particle dissolution function,  $g$ :

$$l(a) = \frac{l_0 \left( \left[ \frac{6}{\pi\rho} g^{-1} \left( \frac{\pi\rho}{6} a^3, t \right) \right]^{1/3} \right) \frac{d}{da} \left[ \frac{6}{\pi\rho} g^{-1} \left( \frac{\pi\rho}{6} a^3, t \right) \right]^{1/3}}{\int_{L_1}^{L_2} l_0(a) da} \quad (6.3)$$

For 
$$P \left[ \frac{6}{\pi\rho} g \left( \frac{\pi\rho}{6} d_0^3, t \right) \right]^{1/3} \leq a \leq P \left[ \frac{6}{\pi\rho} g \left( \frac{\pi\rho}{6} D_0^3, t \right) \right]^{1/3} \quad (6.4)$$

The integration limits  $L_1$  and  $L_2$  depend on time as follows:

$$L_1 = d_0 \text{ for } t \text{ such that } P g \left( \frac{\rho\pi}{6} d_0^3, t \right) > 0 \quad (6.5)$$

$$L_1 = \left[ \frac{6}{\pi\rho} g^{-1}(0, t) \right]^{1/3} \text{ " " " " " " } = 0 \quad (6.6)$$

$$L_2 = D_0 \text{ for } t \text{ such that } P g \left( \frac{\rho\pi}{6} D_0^3, t \right) > 0 \quad (6.7)$$

$$L_2 = \left[ \frac{6}{\pi\rho} g^{-1}(0, t) \right]^{1/3} \text{ " " " " " " } = 0 \quad (6.8)$$

where  $d_0$  and  $D_0$  denote the initial diameters of the smallest and largest particles, respectively. The operator  $P$  has been introduced to make the expression generally applicable (95). It is defined to be equal to one in the time period before the operand becomes zero and is equal to zero beyond that time. The lower integration limit  $L_1$  changes value at  $g \left[ \left( \frac{\rho\pi}{6} \right) d_0^3, t \right] = 0$ , that is the critical time when the particles start to disappear. The time at which  $g \left[ \left( \frac{\rho\pi}{6} \right) D_0^3, t \right] = 0$  corresponds to the disappearance of the last particle and marks the completion of the dissolution process.

Many powders have size distributions that are approximately log-normal (122). Consider such a powder distributed such that  $\ln a_o$  approximates a normal distribution with mean  $\mu$  and standard deviation  $\sigma$  truncated at  $\ln d_o = \mu - i\sigma$  and at  $\ln D_o = \mu + j\sigma$ , where  $i$  and  $j$  are truncation parameters. The initial particle-size distribution,  $l_o(a_o)$ , is then given by (96):

$$l_o(a_o) = \frac{a_o^{-1} N(\ln a_o, \mu, \sigma)}{\int_{d_o}^{D_o} a_o^{-1} N(\ln a_o, \mu, \sigma) da_o} \quad (6.9)$$

$$d_o \leq a_o \leq D_o$$

where  $N(\ )$  is the normal distribution with  $\ln a_o$  as the variable.

The change in the particle-size distribution during dissolution depends on the way the individual particles dissolve. Three widely known models for single-particle dissolution are considered. When written in the same form as 6.1, the cube root law (97) can be expressed as:

$$w = (w_o^{1/3} - k_3 t)^3 \quad (6.10)$$

In a similar way, the equation presented by Niebergal et al. (106) can be written simply:

$$w = (w_o^{1/2} - k_w t)^2 \quad (6.11)$$

and the model proposed by Higuchi and Hiestand (107) can be written:

$$w = (w_o^{2/3} - k_1 t)^{3/2} \quad (6.12)$$

For simplicity and because their evaluation is not important to the theoretical discussion, the constants  $k_1$ ,  $k_2$  and  $k_3$  are used in place of the original time coefficients which included parameters such as the shape factor, particle density, and diffusion coefficient. In the following section, Eqs. 6.12, 6.11 and 6.10 will be referred to as models 1, 2 and 3, respectively. By having defined the initial size distribution  $l_o(a_o)$  (Eq. 6.9),



and the dissolution function (Eqs. 6.10-12) the size distribution at time t,  $l(a)$ , can be expressed applying 6.3:

$$l(a) = \frac{a^{\left(\frac{3}{m} - 1\right)} \left(\frac{3}{a^m + Kt}\right)^{-1} N\left[\ln\left(\frac{3}{a^m + Kt}\right)^{\frac{m}{3}}, \mu, \sigma\right]}{F\left(\frac{T_2 - \mu}{\sigma}\right) - F\left(\frac{T_1 - \mu}{\sigma}\right)} \quad (6.13)$$

For  $P(\text{EXP}[3(\mu - i\sigma)/m] - Kt)^{\frac{m}{3}} \leq a \leq P(\text{EXP}[3(\mu + j\sigma)/m] - Kt)^{\frac{m}{3}}$

and  $l(a) = 0$  elsewhere. Also:

$$T_1 = \mu - i\sigma \quad \text{for } \frac{m}{3} \ln(Kt) < \mu - i \quad (6.14)$$

$$T_1 = \frac{m}{3} \ln(Kt) \quad " \quad " \quad " \quad \geq \mu - i \quad (6.15)$$

$$T_2 = \mu + j\sigma \quad \text{for } \frac{m}{3} \ln(Kt) < \mu + j \quad (6.16)$$

$$T_2 = \frac{m}{3} \ln(Kt) \quad " \quad " \quad " \quad \geq \mu + j \quad (6.17)$$

$$\text{and } K = (6/\rho\pi)^{\frac{1}{m}} k \quad (6.18)$$

Equation 6.13 describes the size distribution for all three models. For Model 1,  $m = 3/2$ ; for Model 2,  $m = 2$ ; and for Model 3,  $m = 3$ . The constants  $k$  and  $K$  for each model should be  $k_1, k_2, k_3$  and  $K_1, K_2, K_3$ , respectively. The function  $F(\ )$  is the commonly tabulated area under the standard normal curve function defined earlier (5.37).

The main particle size (diameter),  $\bar{a}$  can be obtained by applying the usual integration approach used in mathematical expectation:

$$\bar{a} = \frac{\int_R a^{\frac{3}{m}} \left(\frac{3}{a^m + Kt}\right)^{-1} N\left[\ln\left(\frac{3}{a^m + Kt}\right)^{\frac{m}{3}}, \mu, \sigma\right] da}{F\left(\frac{T_2 - \mu}{\sigma}\right) - F\left(\frac{T_1 - \mu}{\sigma}\right)} \quad (6.19)$$

The integration interval  $R$  in 6.19 is the same as the interval for  $a$  defined in 6.13. Equation 6.19 considers Model 1 and 2 ( $m = 3/2$  and  $m = 2$ , respectively). The mean particle size for the third model ( $m = 3$ ) simplifies further to:

$$\bar{a} = \frac{F\left(\frac{T_2 - \mu}{\sigma} - \sigma\right) - F\left(\frac{T_1 - \mu}{\sigma} - \sigma\right)}{F\left(\frac{T_2 - \mu}{\sigma}\right) - F\left(\frac{T_1 - \mu}{\sigma}\right)} \text{EXP}(\mu + \sigma^2/2) - K_3 t \quad (6.20)$$

Where  $T_1$  and  $T_2$  are still defined as in Eqs. 6.14-17. The exact dissolution profile of a log-normal powder with single particles dissolving according to each of the three models can be derived using 5.22 presented earlier (96):

$$\frac{W}{W_0} = \sum_{n=0}^m \binom{m}{n} (-Kt)^{(m-n)} \frac{F(A) - F(B)}{F(j-3\sigma) - F(-i-3\sigma)} \text{EXP}(C) \quad (6.21)$$

For which

$$A = (T_2 - \mu)/\sigma - 3n\sigma/m \quad (6.22)$$

$$B = (T_1 - \mu)/\sigma - 3n\sigma/m \quad (6.23)$$

and 
$$C = \frac{3}{m} (n-m) (\mu + \frac{3}{m} (n+m)\sigma^2/2) \quad (6.24)$$

Where  $W$  and  $W_0$  are the amounts of undissolved powder at time  $t$  and  $0$ , respectively. Equation 6.21 with  $m = 3$ , although presented in a more compact form, is identical to 5.38 derived earlier. Equation 6.21 is not defined for Model 1 ( $m = 3/2$ ) which must be considered separately:

$$\frac{W}{W_0} = \frac{\int_{R_1}^{R_2} (w^2 - K_1 t)^{3/2} w^{-1} N(\ln w, \mu, \sigma) dw}{F(j-3\sigma) - F(-i-3\sigma)} \text{EXP}(-3\mu - 9\sigma^2/2) \quad (6.25)$$

where

$$R_1 = \text{EXP}(\mu - i\sigma) \quad \text{for } (K_1 t)^{1/2} < \text{EXP}(\mu - i\sigma) \quad (6.26)$$

$$R_1 = (K_1 t)^{1/2} \quad \text{" " } \geq \text{EXP}(\mu - i\sigma) \quad (6.27)$$

$$R_2 = \text{EXP}(\mu + j\sigma) \quad \text{for } (K_1 t)^{1/2} < \text{EXP}(\mu + j\sigma) \quad (6.28)$$

$$R_2 = (K_1 t)^{1/2} \quad \text{" " } \geq \text{EXP}(\mu + j\sigma) \quad (6.29)$$

The derivation of these equations are based on two assumptions:

- (a) that the particles in the multiparticulate system dissolve independently of each other, which will be approximated well under sink conditions; and
- (b) that they dissolve according to the same single-particle dissolution

model having fixed parameters (for these cases,  $k_1$ ,  $k_2$  and  $k_3$  are the same for all particles and do not vary during dissolution). If these conditions exist, then it is possible to propose some general rules concerning the dissolution process. These rules are explained in relation to what will be termed "the intrinsic dissolution profile", which can be defined in the following way: Dissolution curves have the same intrinsic dissolution profile if, by a suitable scaling of time, they can be brought into each other in a  $W/W_0$  versus time plot (Fig. 6.1).

It should be clear from observation of Eqs. 5.17 and 5.22 that the coefficient of time in an expression correctly defining the multiparticulate dissolution profile originates directly from the coefficient of time in the single-particle dissolution equation. Thus a different value of the rate parameter, that is, a different coefficient of time in the single-particle dissolution equation, has the same effect as a different scaling of time. Therefore, the intrinsic dissolution profile will still be the same. The following rule can thus be stated:

1. *The intrinsic dissolution profile is independent of the value of the rate parameter, that is, the coefficient of time in the single-particle dissolution equation.*

According to this rule, the rate parameter  $k_1$ ,  $k_2$  and  $k_3$  (6.10-12) have no influence on the intrinsic dissolution profile. Furthermore, there will always be a proportional relationship between the coefficient of time in the multiparticulate dissolution equation and the rate parameter. The following rule can therefore be stated:

2. *In two systems having identical particle-size distributions, the time-scaling factor that brings one dissolution curve into another is equal to the factor with which the rate parameters are proportionally related in the two systems. (Fig. 6.1)*

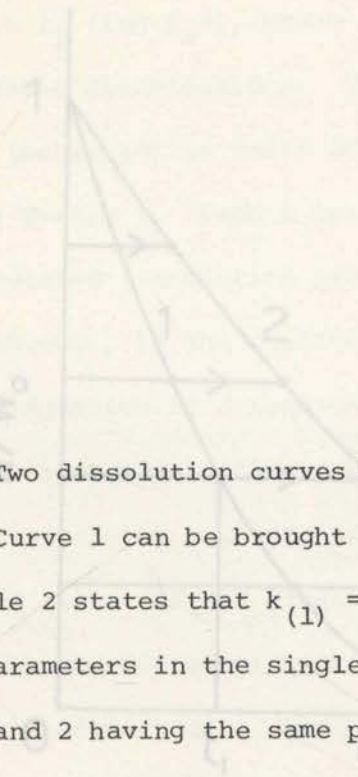
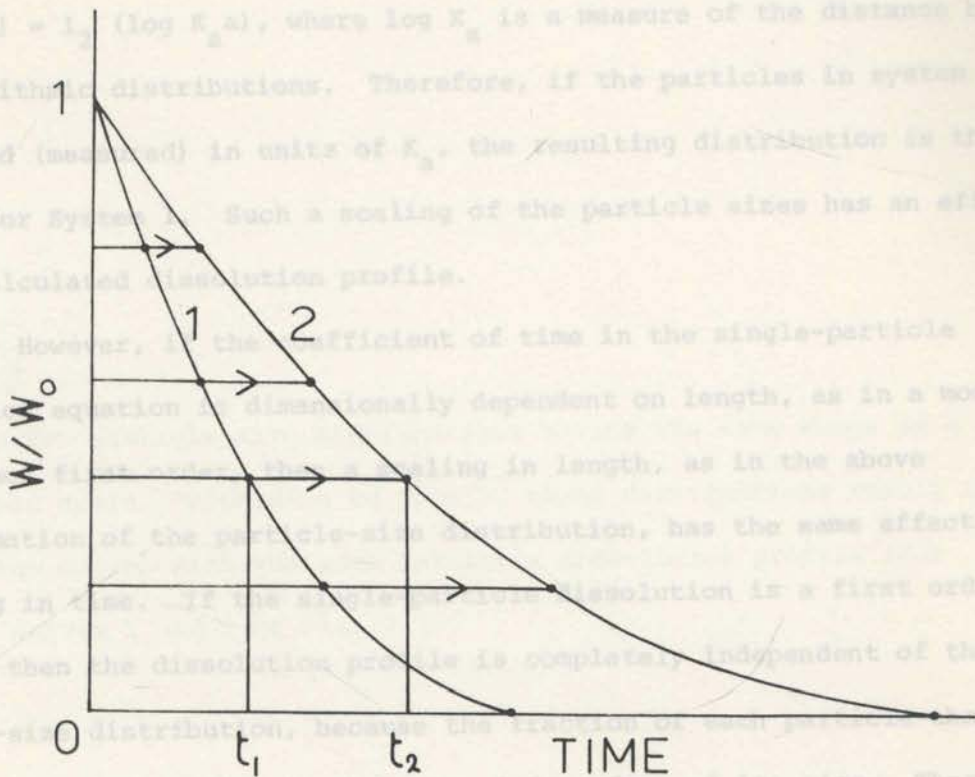


Figure 6.1

Two dissolution curves having the same intrinsic dissolution profile. Curve 1 can be brought into curve 2 by a time-scaling factor,  $t_2/t_1$ . Rule 2 states that  $k_{(1)} = (t_2/t_1)k_{(2)}$ , where  $k_{(1)}$  and  $k_{(2)}$  are the rate parameters in the single-particle dissolution equation for Systems 1 and 2 having the same particle-size distribution. These statements also include plots where  $W/W_0$  is raised to any other exponent.

Consider two particle-size distributions that are distributed on a logarithmic scale  $l_1(\log a)$  and  $l_2(\log a)$  having the same shape. Then for any diameter,  $A$  (Fig. 5.2)  $l_1(a) = l_2(\log a + \log K_0)$ ; i.e.,  $l_1(\log a) = l_2(\log K_0 a)$ , where  $\log K_0$  is a measure of the distance between the logarithmic distributions. Therefore, if the particles in system 2 are scaled (measured) in units of  $K_0$ , the resulting distribution is the same as that for system 1. Such a scaling of the particle sizes has an effect on the calculated dissolution profile.



However, if the coefficient of time in the single-particle dissolution equation is dimensionally dependent on length, as in a model that is first order in length, as in the above transformation of the particle-size distribution, has the same effect as a scaling in time. If the single-particle dissolution is a first order process, then the dissolution profile is completely independent of the particle-size distribution, because the time for each particle to dissolve in a given time is the same, independent of its size. Therefore, it can be concluded that systems 1 and 2 have the same intrinsic dissolution profile, and the following rule can be given:

3. Two powders dissolving according to the same single-particle dissolution model have the same intrinsic dissolution profile if their particle-size distributions are of the same shape on a logarithmic scale, (Fig. 5.2)

It follows from this rule that it is not the "position" of the distribution, that is, not the actual size of the particles, but the shape of the distribution that affects the intrinsic profile. Thus it can be stated that:

4. The intrinsic dissolution profile does not depend on the actual size of the particles but on the shape of their distribution.

Consider two particle-size distributions that are distributed on a logarithmic scale  $l_1(\log a)$  and  $l_2(\log a)$  having the same shape. Then for any diameter,  $a$  (Fig. 6.2)  $l_1(a) = l_2(\log a + \log K_a)$ ; i.e.,  $l_1(\log a) = l_2(\log K_a a)$ , where  $\log K_a$  is a measure of the distance between the logarithmic distributions. Therefore, if the particles in system 2 are scaled (measured) in units of  $K_a$ , the resulting distribution is the same as that for System 1. Such a scaling of the particle sizes has an effect on the calculated dissolution profile.

However, if the coefficient of time in the single-particle dissolution equation is dimensionally dependent on length, as in a model that is not first order, then a scaling in length, as in the above transformation of the particle-size distribution, has the same effect as a scaling in time. If the single-particle dissolution is a first order process, then the dissolution profile is completely independent of the particle-size distribution, because the fraction of each particle that dissolves in a given time is the same, independent of its size. Therefore, it can be concluded that system 1 and 2 have the same intrinsic dissolution profile, and the following rule can be given:

3. *Two powders dissolving according to the same single-particle dissolution model have the same intrinsic dissolution profile if their particle-size distributions are of the same shape on a logarithmic scale.*

(Fig. 6.2)

It follows from this rule that it is not the "position" of the distribution, that is, not the actual size of the particles, but the shape of the distribution that affects the intrinsic profile. Thus it can be stated that:

4. *The intrinsic dissolution profile does not depend on the actual size of the particles but on the shape of their distribution.*

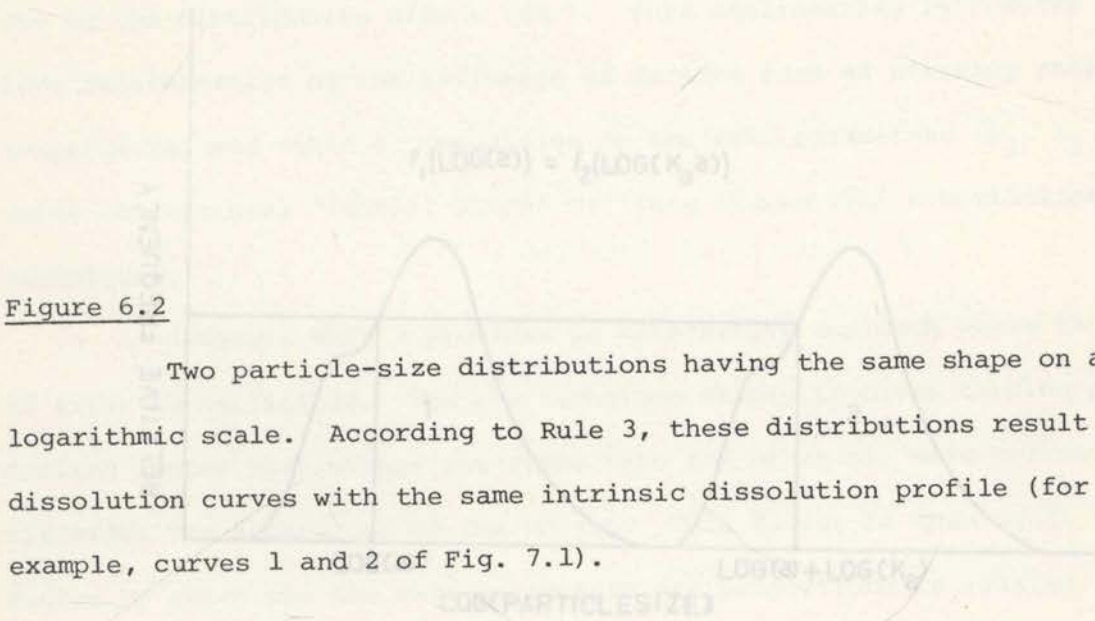
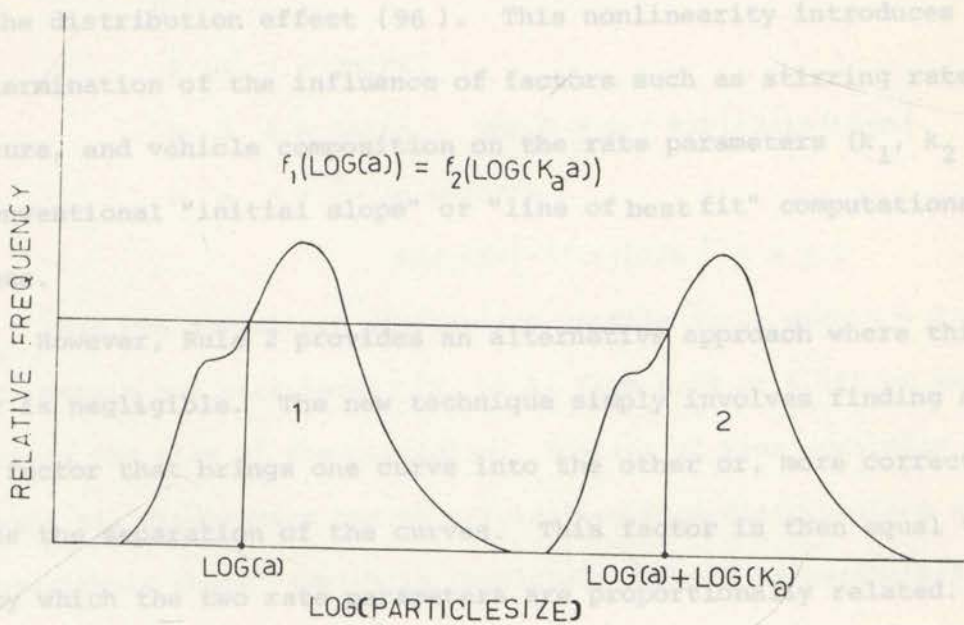


Figure 6.2

Two particle-size distributions having the same shape on a logarithmic scale. According to Rule 3, these distributions result in dissolution curves with the same intrinsic dissolution profile (for example, curves 1 and 2 of Fig. 7.1).

According to Rule 3, the concept of time scaling in dissolution studies should have some practical application. It has already been stated that it is very difficult to prepare completely monodispersed powders; therefore, some nonlinearity is always present in the dissolution profile due to the distribution effect (96). This nonlinearity introduces errors into determination of the influence of factors such as stirring rate, temperature, and vehicle composition on the rate parameters ( $k_1$ ,  $k_2$  or  $k_3$ ) using conventional "initial slope" or "line of best fit" computational techniques.



However, Rule 2 provides an alternative approach where this type of error is negligible. The new technique simply involves finding a time-scaling factor that brings one curve into the other or, more correctly, minimizes the variance of the curves. This factor is then equal to the factor by which the two rate parameters are proportionally related. Perhaps the best criteria is to minimize the squared horizontal differences between the curves.

According to Rules 1 and 3, it should be possible to normalize the calculated dissolution profiles for log-normal powders by appropriate scaling of time to a form that does not depend on either the rate parameter ( $k_1$ ,  $k_2$  or  $k_3$ ) or the actual sizes of the particles. One approach is to scale time in the  $W/W_0$  versus time plot as the time fraction  $\psi$ , defined as the fraction of the time necessary for complete dissolution. The expression defining the resulting normalized dissolution profile can be obtained in the following way, using Models 2 and 3 as examples. The time for complete dissolution,  $t_0$ , is given by

$(m/3) \ln(Kt_0) = \psi + \sigma$ ; then since  $\psi = t/t_0$ , it follows that:

$$\psi = K \exp[-3(\psi + \sigma)/m] \quad (6.30)$$



According to Rule 2, the concept of time scaling in dissolution studies should have some practical application. It has already been stated that it is very difficult to prepare completely monodispersed powders; therefore, some nonlinearity is always present in the dissolution profile due to the distribution effect (96). This nonlinearity introduces errors into determination of the influence of factors such as stirring rate, temperature, and vehicle composition on the rate parameters ( $k_1$ ,  $k_2$  or  $k_3$ ) using conventional "initial slope" or "line of best fit" computational techniques.

However, Rule 2 provides an alternative approach where this type of error is negligible. The new technique simply involves finding a time-scaling factor that brings one curve into the other or, more correctly, minimizes the separation of the curves. This factor is then equal to the factor by which the two rate parameters are proportionally related. Perhaps the best criteria is to minimize the squared horizontal differences between the curves.

According to Rules 1 and 3, it should be possible to normalize the calculated dissolution profiles for log-normal powders by appropriate scaling of time to a form that does not depend on either the rate parameter ( $k_1$ ,  $k_2$  or  $k_3$ ) or the actual sizes of the particles. One approach is to scale time in the  $W/W_0$  versus time plot as the time fraction  $\psi$ , defined as the fraction of the time necessary for complete dissolution. The expression defining the resulting normalized dissolution profile can be obtained in the following way, using Models 2 and 3 as examples. The time for complete dissolution,  $t_0$ , is given by

$(m/3) \ln(Kt_0) = \mu + j\sigma$ ; then since  $\psi = t/t_0$  it follows that:

$$\psi = Kt \text{EXP} \left[ -3(\mu + j\sigma)/m \right] \quad (6.30)$$

Accordingly, the  $Kt$  terms in 6.21 can be substituted by  $\psi^{3(\mu+j\sigma)/m}$ , which causes the  $\mu$  term to cancel out. After rearrangement, 6.21 can be written:

$$\frac{W}{W_0} = \sum_{n=0}^m \binom{m}{n} (-\psi)^{(m-n)} \frac{F(A) - F(B)}{F(j-3\sigma) - F(-i-3\sigma)} \text{EXP}(C) \quad (6.31)$$

where:

$$A = j - 3n\sigma/m$$

$$B = i - 3n\sigma/m \quad \text{for } 0 \leq \psi \leq \text{EXP}[-3(i+j)\sigma/m]$$

$$B = j + \frac{m}{3} \ln \psi - 3n\sigma/m$$

$$\text{for } \text{EXP}[-3(i+j)\sigma/m] \leq \psi \leq 1$$

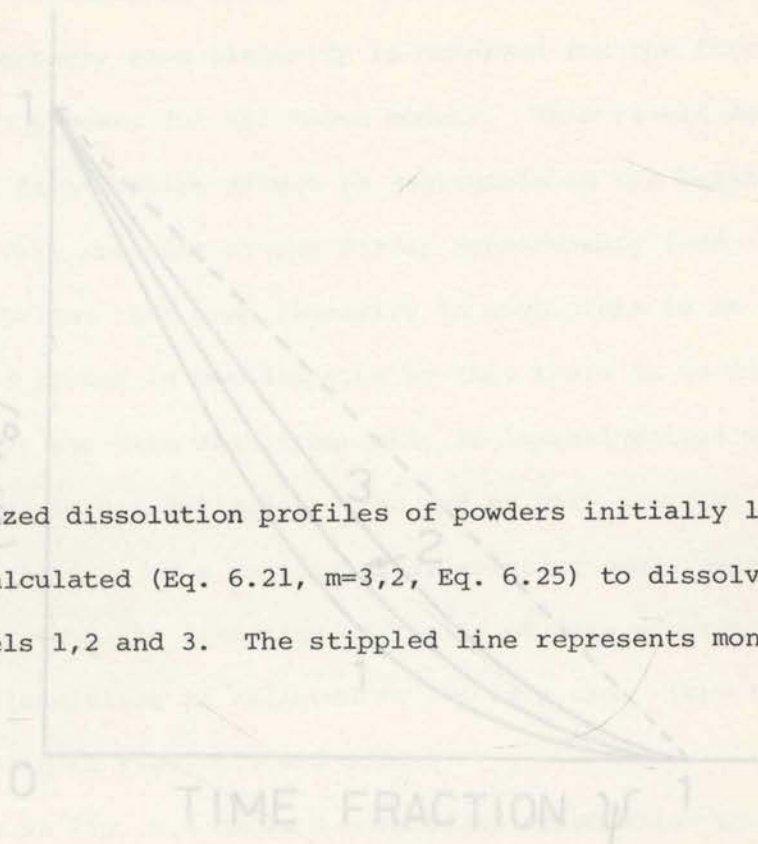
Thus the normalized dissolution profile (6.31 with  $m = 2$  or  $m = 3$ ) does not contain any rate terms ( $k_3$  or  $k_2$  from 6.10 or 6.11) or any term ( $\mu$ ) representing the size of the particles. Scaling of time according to 6.30 has brought all dissolution curves originating from distributions with the same "logarithmic shape" (which is completely defined by parameters  $\sigma$ ,  $i$  and  $j$ ) into one single curve (6.31) which does not depend on the size of the particles or the rate parameter from the single-particle equation. This confirms Rule 1 and 3.

The transformation has essentially normalized all possible systems having the same intrinsic dissolution profile into one single curve. This curve is unique in that it makes it possible to evaluate the isolated distribution effect. This evaluation is best done by plotting in a way that linearizes the underlying single-particle dissolution equation (6.10-12), by using  $(W/W_0)^{1/m}$  instead of  $W/W_0$  in the plot. Such a plot will be linear with slope = -1 for a true monodisperse system. Any deviation from this linearity and slope will be due solely to the distribution effect.

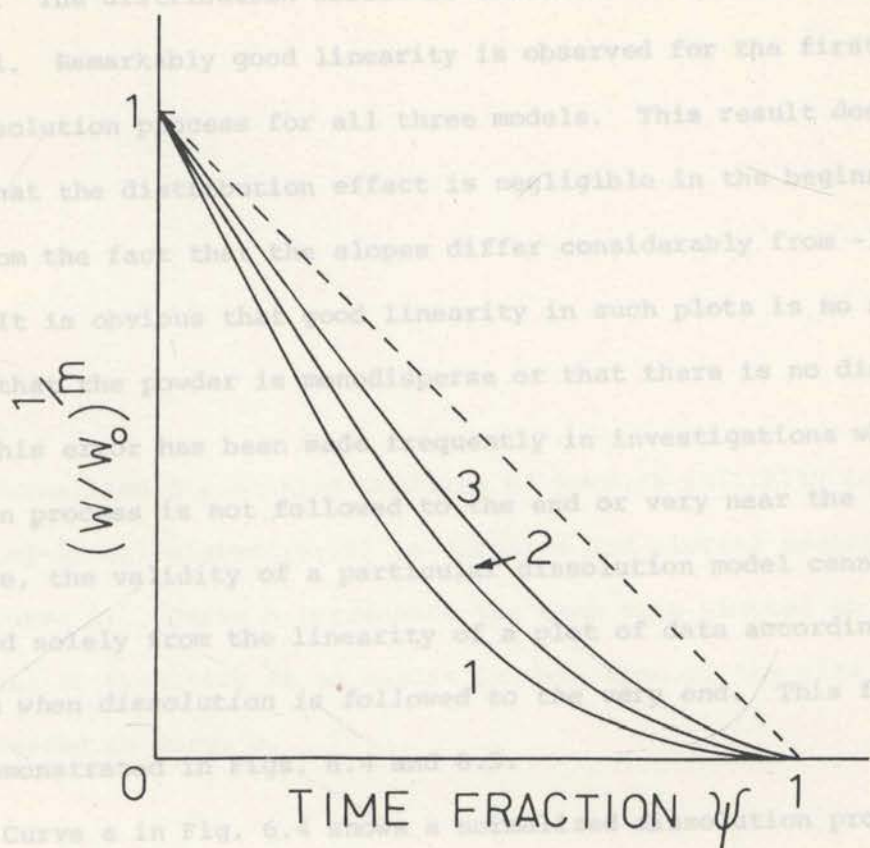
Figure 6.3 shows such normalized dissolution profiles, calculated according to 6.21 and 6.31, for powders initially log-normal, having distribution (shape) parameters  $\sigma = 0.2$  and  $i = j = 2$ , for particles dissolving

Figure 6.3

Normalized dissolution profiles of powders initially log-normal ( $\sigma=0.2$ ,  $i=j=2$ ) calculated (Eq. 6.21,  $m=3,2$ , Eq. 6.25) to dissolve and plotted according to models 1,2 and 3. The stippled line represents monodisperse powders ( $\sigma=0$ ).



according to each of the three models (6.10-12). The fraction of undissolved powder,  $W/W_0$ , is raised to the powers of 1/3, 1/2 and 2/3 for the reason just given. The distribution effect is smallest for Model 3 and greatest for Model 1. Remarkably good linearity is observed for the first part of the dissolution process for all three models. This result does not mean, however, that the distribution effect is negligible in the beginning, as is seen from the fact that the slopes differ considerably from -1.



It is obvious that good linearity in such plots is no necessary criterion that the powder is well dispersed or that there is no distribution effect. This error has been made frequently in investigations where the dissolution process is not followed to the end or very near the end. Furthermore, the validity of a particular dissolution model cannot always be assumed solely from the linearity of a part of data according to that model even when dissolution is followed to the end. This fact is clearly demonstrated in Fig. 6.3.

Curve c in Fig. 6.3 shows a dissolution profile of log-normal powders ( $\sigma=0.14$ ,  $l=j=2$ ) calculated (6.25) to dissolve and plotted ( $W/W_0$  to power of 2/3) according to Model 1. The size distribution effect is clearly reflected in the nonlinearity of the curve. By plotting the same data according to an incorrect model, Model 3 ( $W/W_0$  to power of 1/3), the size distribution effect is almost entirely cancelled and surprisingly good linearity is obtained that extends to the very end of the dissolution process (curve b).

Figure 6.5 shows the same phenomenon for powders ( $\sigma = 0.14$ ,  $l = j = 2$ ) where the particles are calculated to dissolve according to Model 2 (6.31,  $m = 2$ ).

A judgment based solely on the linearity of such plots will often lead to false conclusions about the validity of the model, even where

according to each of the three models (6.10-12). The fraction of undissolved powder,  $W/W_0$ , is raised to the powers of  $1/3$ ,  $1/2$  and  $2/3$  for the reason just given. The distribution effect is smallest for Model 3 and greatest for Model 1. Remarkably good linearity is observed for the first part of the dissolution process for all three models. This result does not mean, however, that the distribution effect is negligible in the beginning, as is seen from the fact that the slopes differ considerably from  $-1$ .

It is obvious that good linearity in such plots is no necessary criterion that the powder is monodisperse or that there is no distribution effect. This error has been made frequently in investigations where the dissolution process is not followed to the end or very near the end. Furthermore, the validity of a particular dissolution model cannot always be assessed solely from the linearity of a plot of data according to that model *even when dissolution is followed to the very end*. This fact is clearly demonstrated in Figs. 6.4 and 6.5.

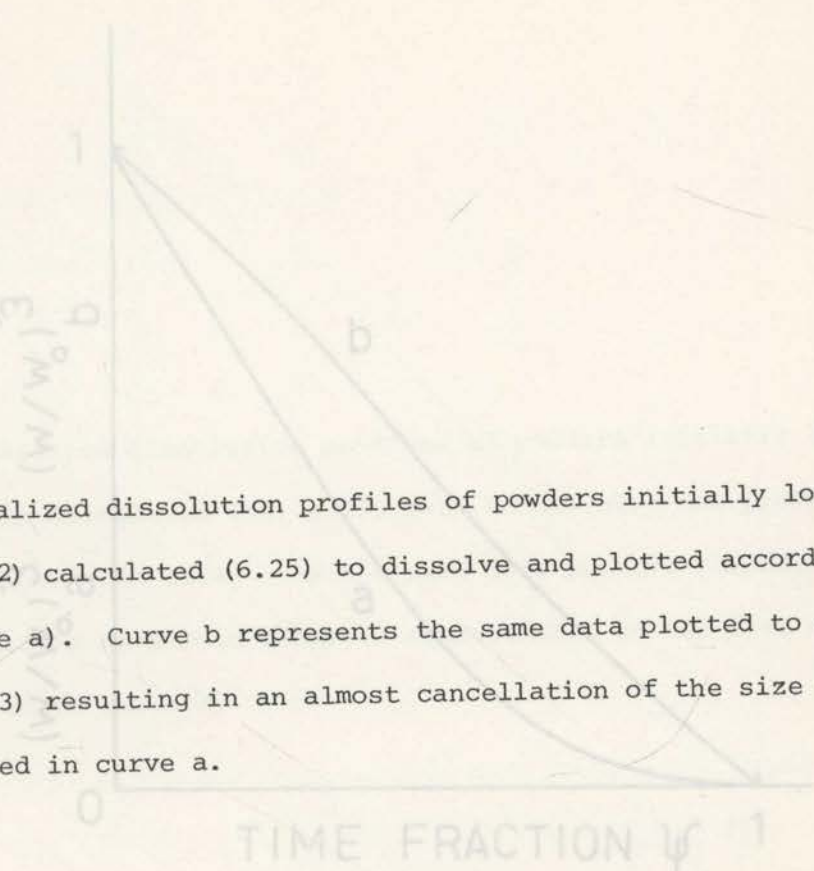
Curve a in Fig. 6.4 shows a normalized dissolution profile of log-normal powders ( $\sigma=0.14$ ,  $i=j=2$ ) calculated (6.25) to dissolve and plotted ( $W/W_0$  to power of  $2/3$ ) according to Model 1. The size distribution effect is clearly reflect in the nonlinearity of the curve. By plotting the same data according to an incorrect model, Model 3 ( $W/W_0$  to power of  $1/3$ ), the size distribution effect is almost entirely cancelled and surprisingly good linearity is obtained *that extends to the very end of the dissolution process* (curve b).

Figure 6.5 shows the same phenomenon for powders ( $\sigma = 0.12$ ,  $i = j = 2$ ) where the particles are calculated to dissolve according to Model 2 (6.31,  $m = 2$ ).

A judgment based solely on the linearity of such plots will often lead to false conclusions about the validity of the model, even where

Figure 6.4

Normalized dissolution profiles of powders initially log-normal ( $\sigma=0.14$ ,  $i=j=2$ ) calculated (6.25) to dissolve and plotted according to model 1 (curve a). Curve b represents the same data plotted to an incorrect model (model 3) resulting in an almost cancellation of the size distribution effect observed in curve a.



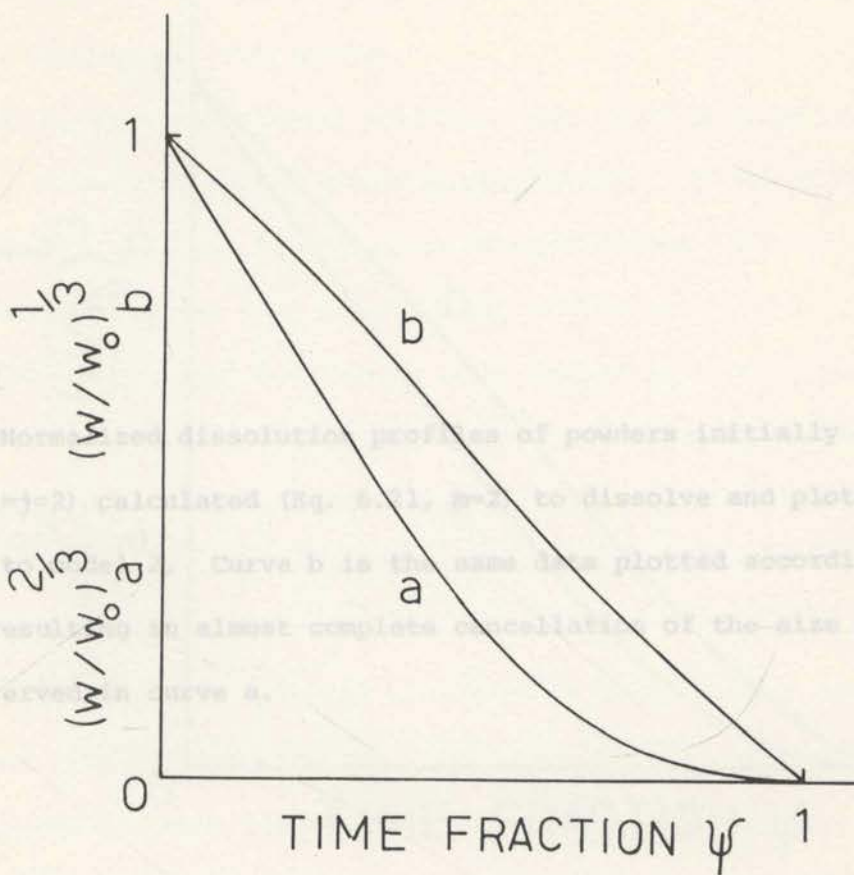


Figure 6.5

Normalized dissolution profiles of powders initially log-normal ( $q=0.12$ ,  $l=2$ ) calculated (Eq. 6.21,  $n=2$ ) to dissolve and plotted according to (6.22). Curve 'a' is the same as plotted according to model 1, resulting in almost complete dissolution of the size distribution effect observed in curve 'a'.

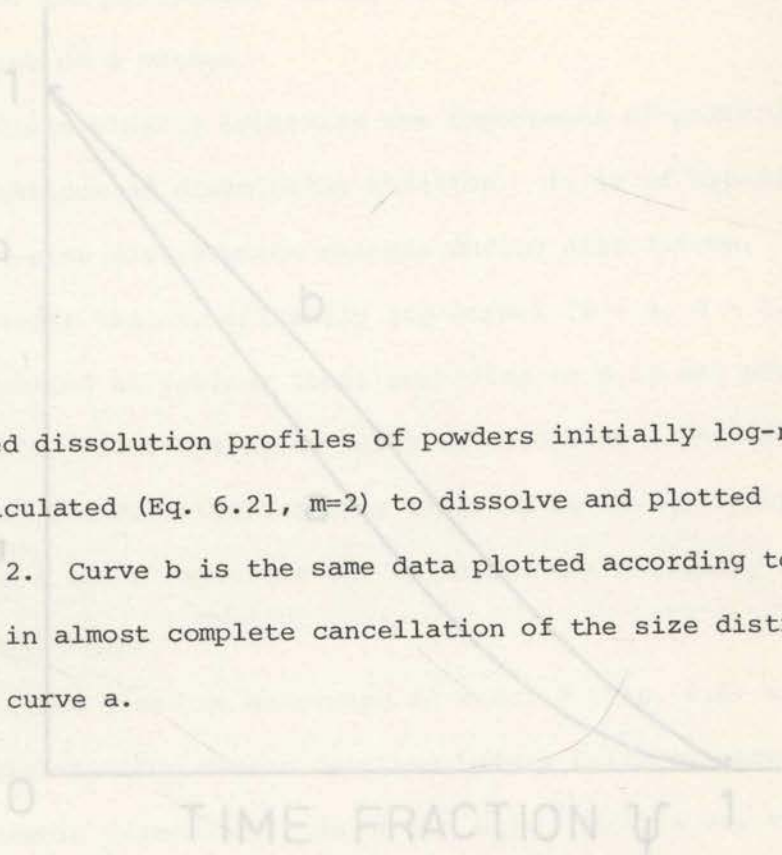


Figure 6.5

Normalized dissolution profiles of powders initially log-normal ( $\sigma=0.12$ ,  $i=j=2$ ) calculated (Eq. 6.21,  $m=2$ ) to dissolve and plotted according to model 2. Curve b is the same data plotted according to model 3, resulting in almost complete cancellation of the size distribution effect observed in curve a.



dissolution is followed to completion, unless an analysis of particle size distribution is made. The phenomenon demonstrated in Figs. 6.4 and 6.6 are not special cases for the particular values of  $\sigma$  chosen but were observed to apply for a wide range of  $\sigma$  values.

These findings clearly emphasize the importance of particle-size analysis in investigations of dissolution kinetics. It is of interest to see how the particle size distribution changes during dissolution. The distribution of powder that is initially log-normal ( $\mu = 5, \sigma = 0.2, 1 = j = 2$ ) was calculated at various times according to 6.13 and plotted in Figs. 6.6, 6.7 and 6.8, illustrating dissolution according to Models 3, 2 and 1, respectively. The distributions are labelled in chronological order from A to B. Curves A and B represent the distributions initially and at critical time, respectively.

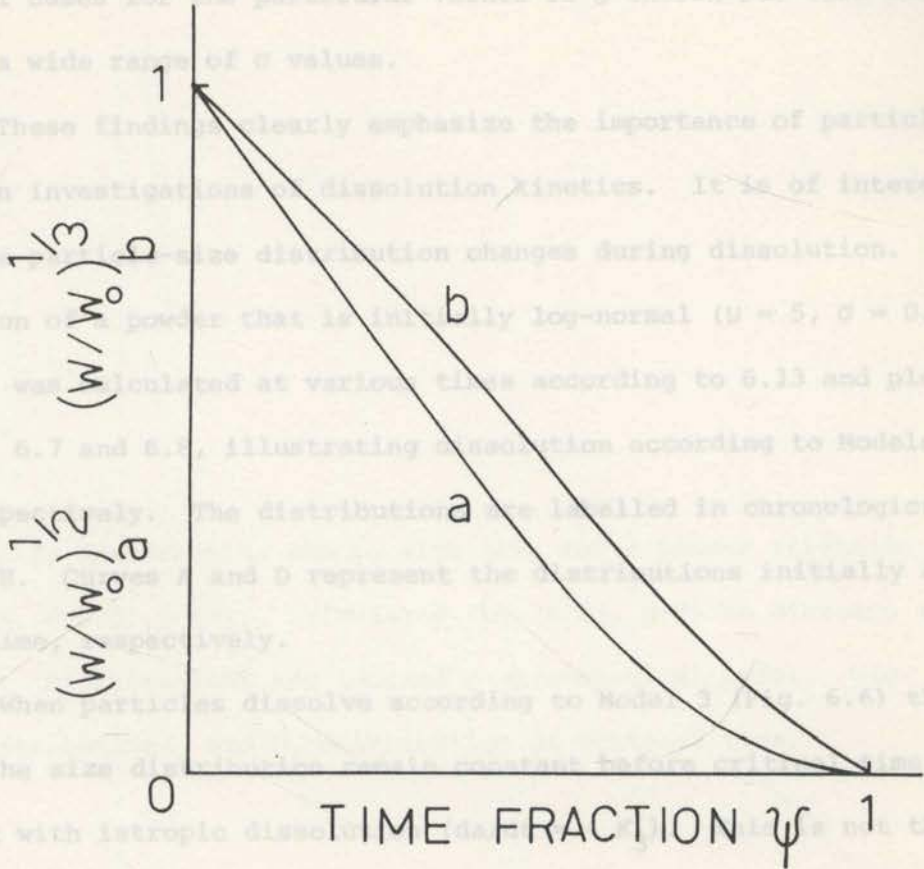
When particles dissolve according to Model 3 (Fig. 6.6) then the shape of the size distribution remains constant before critical time, consistent with isotropic dissolution. However, for Models 1 and 2, the case for dissolution according to Models 1 and 2 where the absolute rate of change in size of the particles,  $da/dt$ , increases with time. For Model 1:

$$da/dt = -\frac{k_1}{2} (a_0^2 - k_1 t)^{-1/2} \quad (6.32)$$

and for Model 2:

$$da/dt = -\frac{2}{3} k_2 (a^{3/2} - k_2 t)^{-1/3} \quad (6.33)$$

As a result, the distribution broadens before critical time (Figs. 6.7 and 6.8) and is particularly affected at the small particles end as zero is approached where  $da/dt$  takes extreme values. Because of the latter effect, near the end of the dissolution process the relative frequency of the very small particles increases with increasing size for Models 1 and 2 (Fig. 6.8 and 6.7) while it decreases for Model 3 (Fig. 6.6). This information



dissolution is followed to competition, unless an analysis of particle size distribution is made. The phenomenon demonstrated in Figs. 6.4 and 6.5 are not special cases for the particular values of  $\sigma$  chosen but were observed to apply for a wide range of  $\sigma$  values.

These findings clearly emphasize the importance of particle-size analysis in investigations of dissolution kinetics. It is of interest to see how the particle-size distribution changes during dissolution. The distribution of a powder that is initially log-normal ( $\mu = 5, \sigma = 0.2, i = j = 2$ ) was calculated at various times according to 6.13 and plotted in Figs. 6.6, 6.7 and 6.8, illustrating dissolution according to Models 3, 2 and 1, respectively. The distributions are labelled in chronological order from A to H. Curves A and D represent the distributions initially and at critical time, respectively.

When particles dissolve according to Model 3 (Fig. 6.6) then the shape of the size distribution remain constant before critical time, consistent with isotropic dissolution ( $da/dt = -K_3$ ). This is not the case for dissolution according to Models 1 and 2 where the absolute rate of change in size of the particles,  $da/dt$ , increases with time. For Model 1:

$$da/dt = -\frac{K_1}{2} (a_0^2 - K_1 t)^{-1/2} \quad (6.32)$$

and for Model 2:

$$da/dt = -\frac{2}{3} K_2 (a^{3/2} - K_2 t)^{-1/3} \quad (6.33)$$

As a result, the distribution broadens before critical time (Figs. 6.7 and 6.8) and is particularly affected at the small particle end as zero is approached where  $da/dt$  takes extreme values. Because of the latter effect, near the end of the dissolution process the relative frequency of the very small particles increases with increasing size for Models 1 and 2 (Fig. 6.8 and 6.7) while it decreases for Model 3 (Fig. 6.6). This information

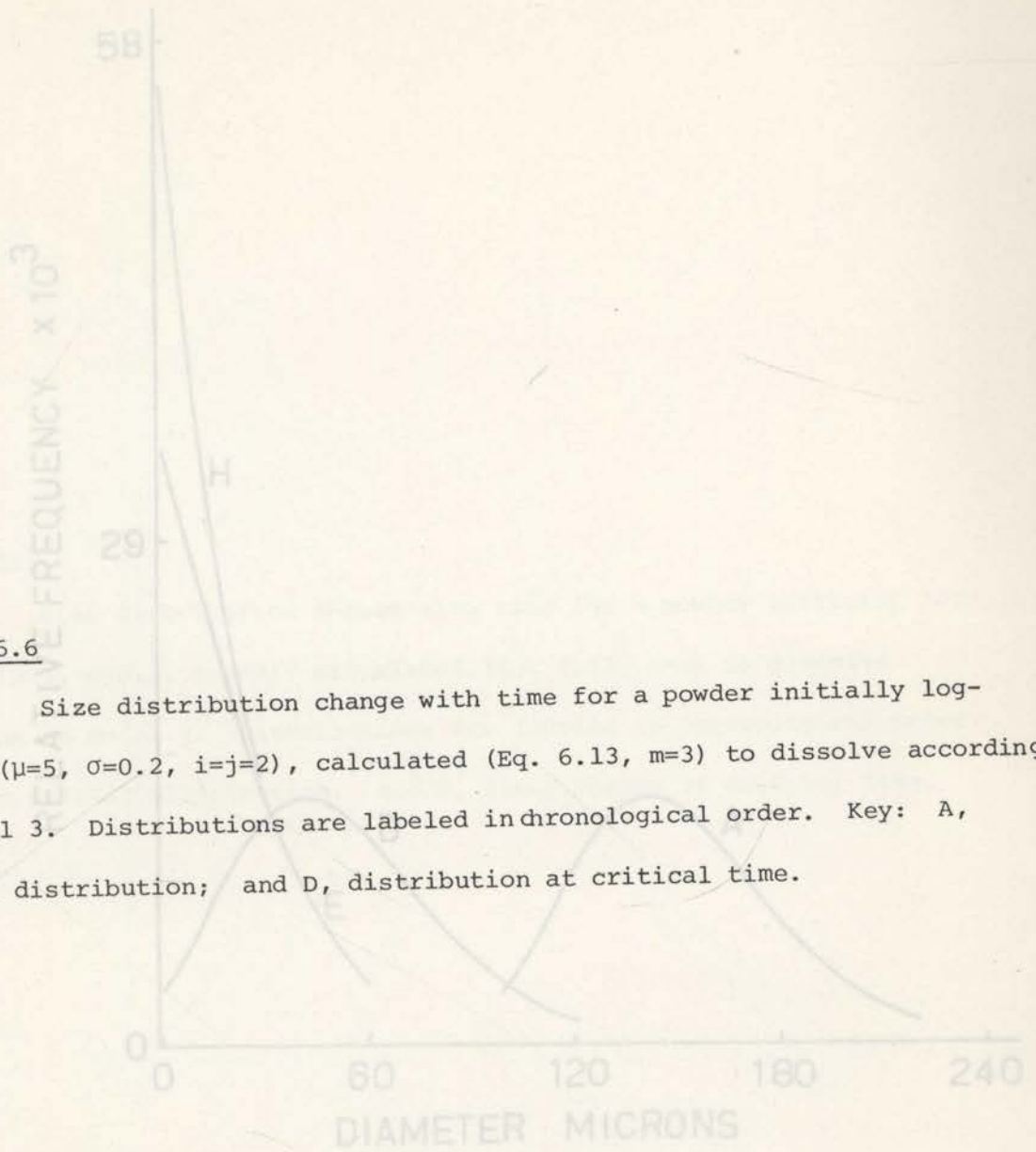


Figure 6.6

Size distribution change with time for a powder initially log-normal ( $\mu=5, \sigma=0.2, i=j=2$ ), calculated (Eq. 6.13,  $m=3$ ) to dissolve according to Model 3. Distributions are labeled in chronological order. Key: A, initial distribution; and D, distribution at critical time.

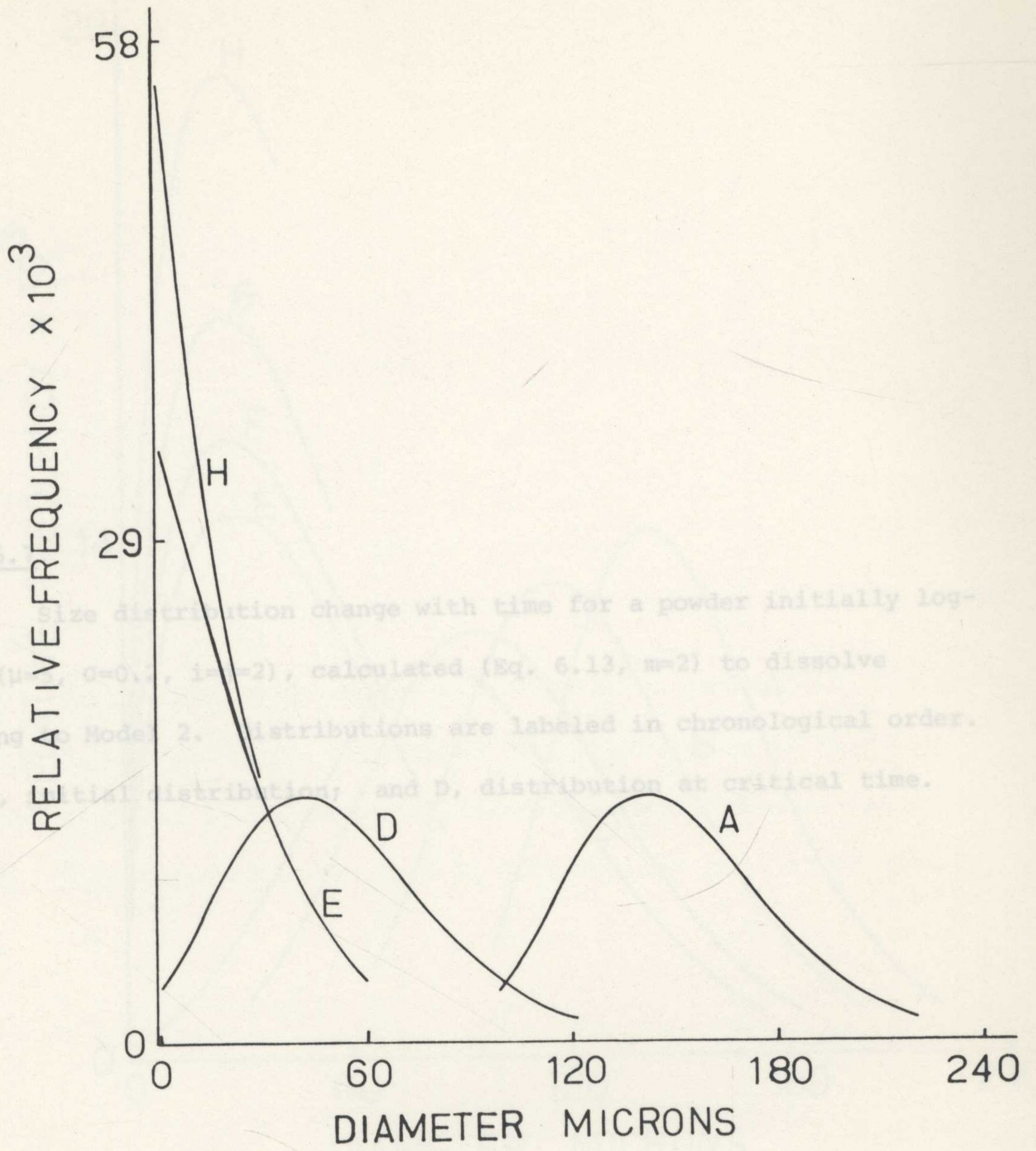


Figure 6.

size distribution change with time for a powder initially log-normal ( $\mu = 0.1$ ,  $\sigma = 2$ ), calculated (Eq. 6.13,  $m=2$ ) to dissolve according to Model 2. Distributions are labeled in chronological order. Key: A, initial distribution; and D, distribution at critical time.

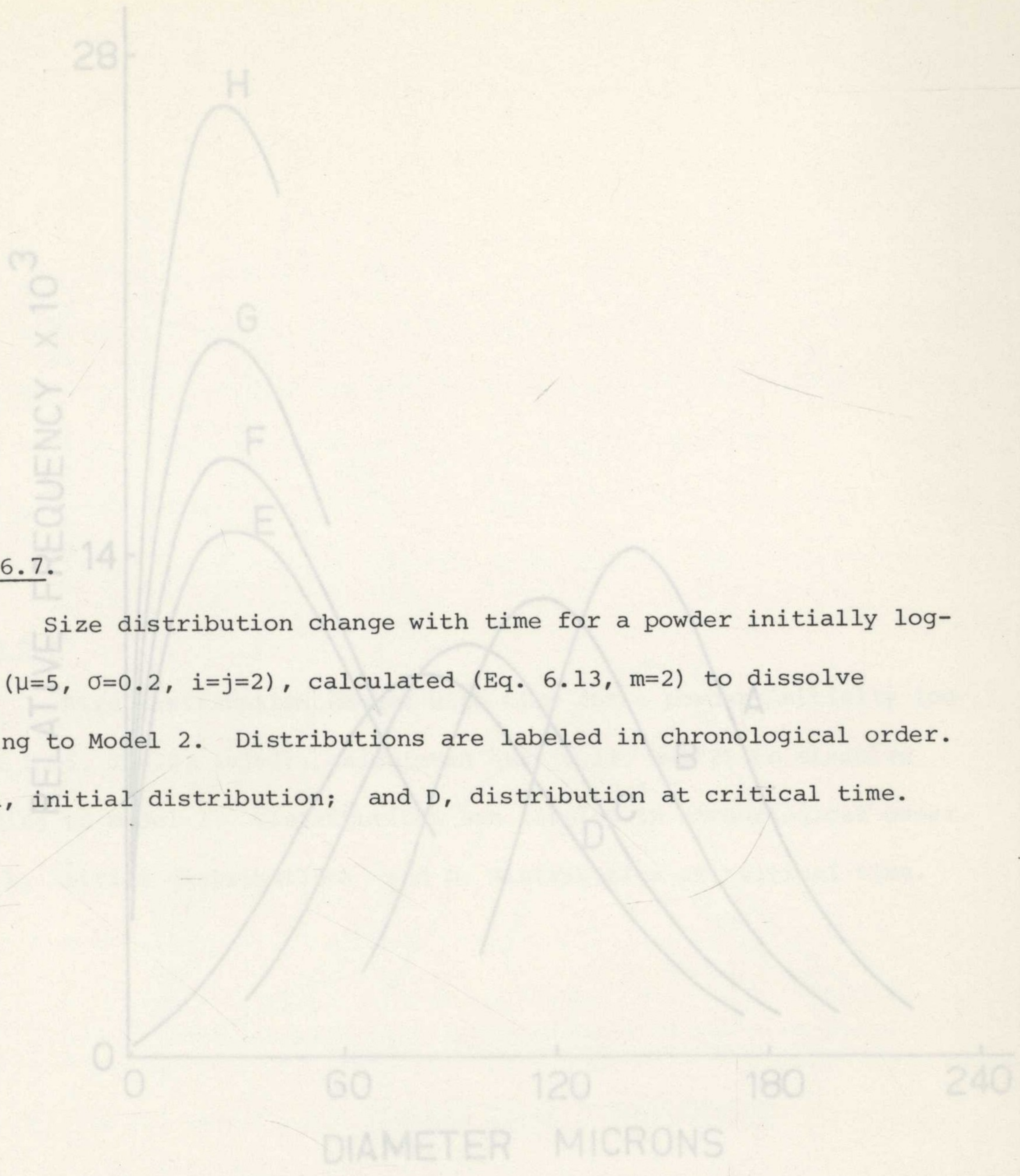
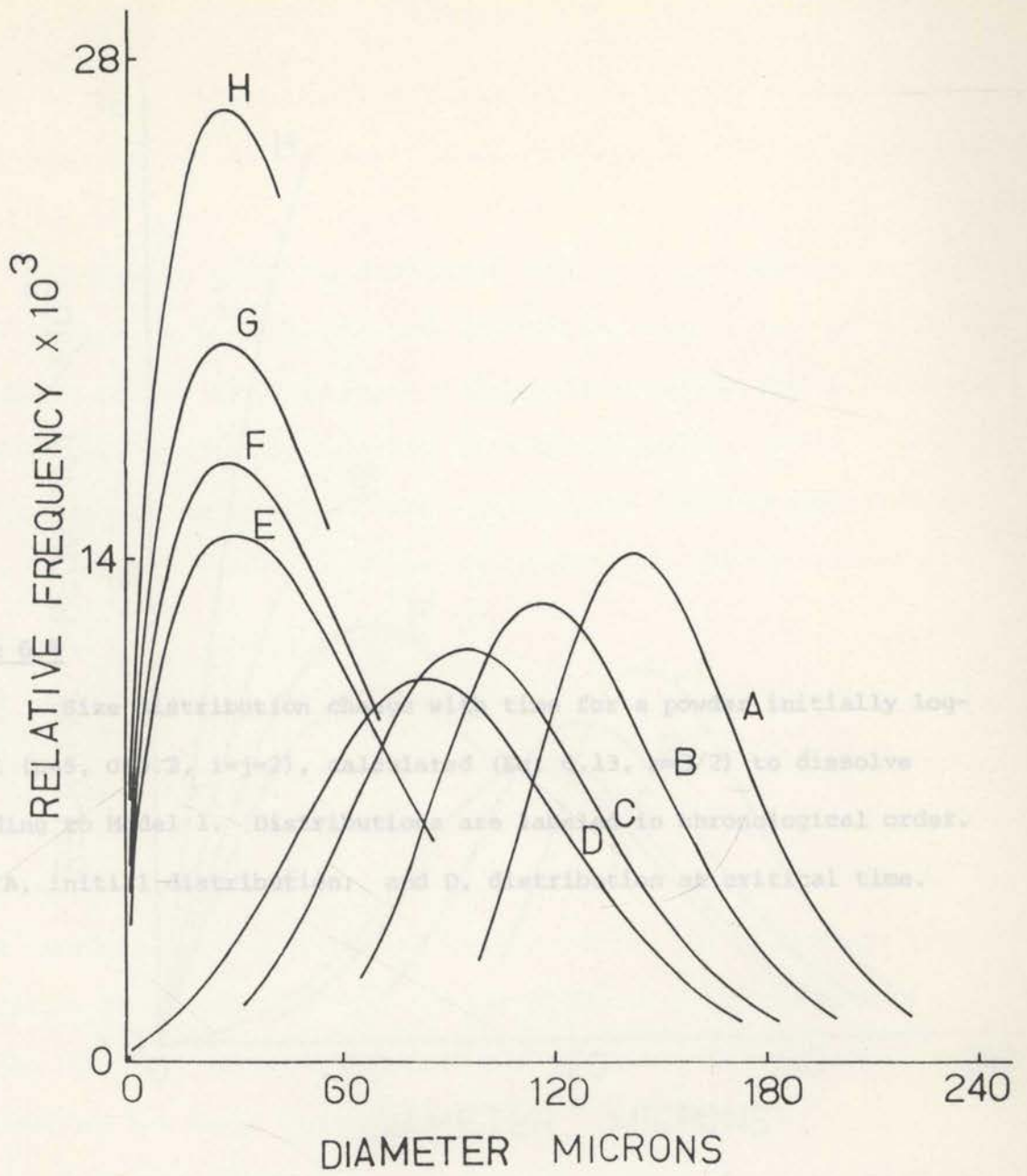


Figure 6.7.

Size distribution change with time for a powder initially log-normal ( $\mu=5, \sigma=0.2, i=j=2$ ), calculated (Eq. 6.13,  $m=2$ ) to dissolve according to Model 2. Distributions are labeled in chronological order. Key: A, initial distribution; and D, distribution at critical time.



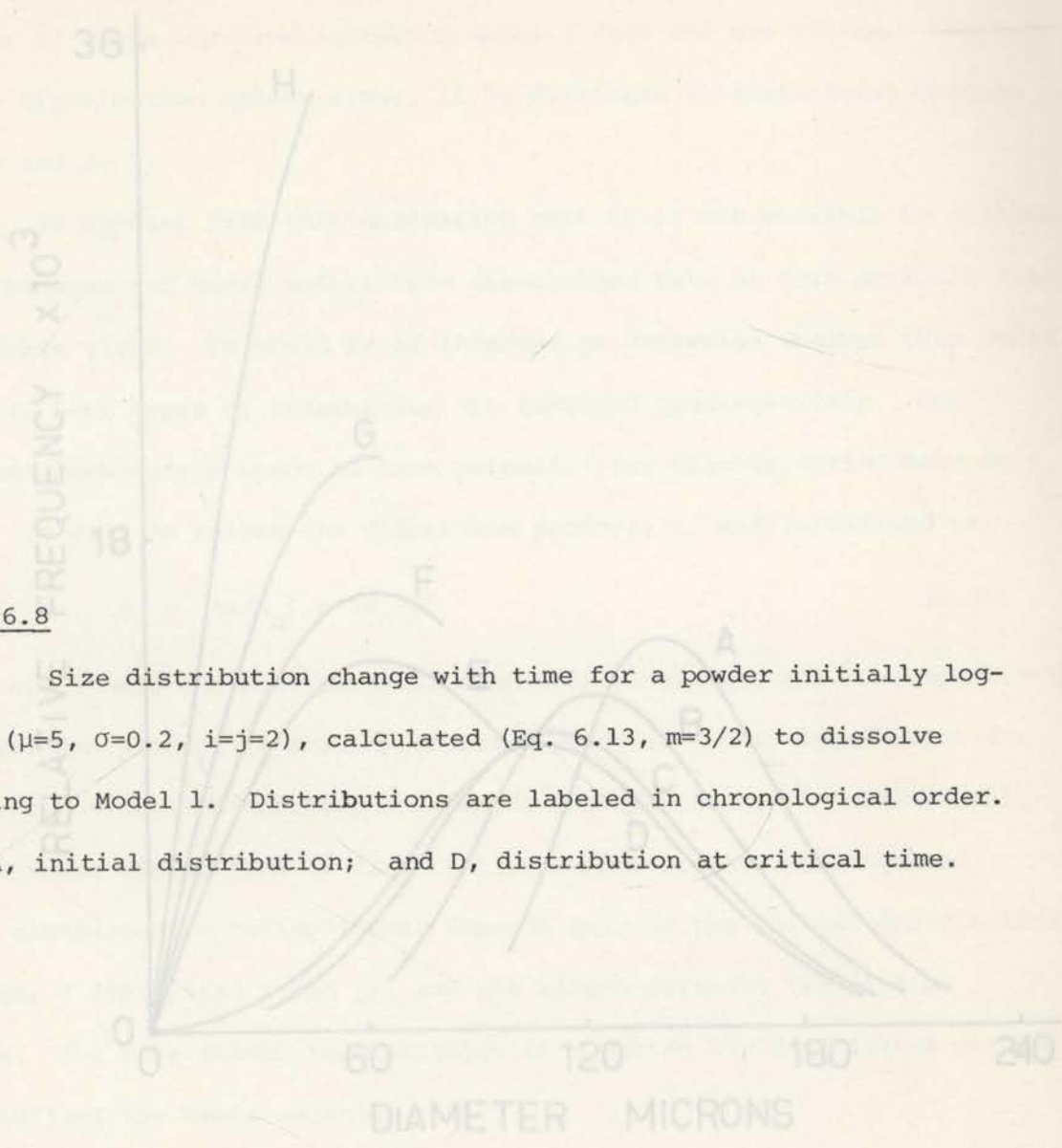


Figure 6.8

Size distribution change with time for a powder initially log-normal ( $\mu=5$ ,  $\sigma=0.2$ ,  $i=j=2$ ), calculated (Eq. 6.13,  $m=3/2$ ) to dissolve according to Model 1. Distributions are labeled in chronological order. Key: A, initial distribution; and D, distribution at critical time.

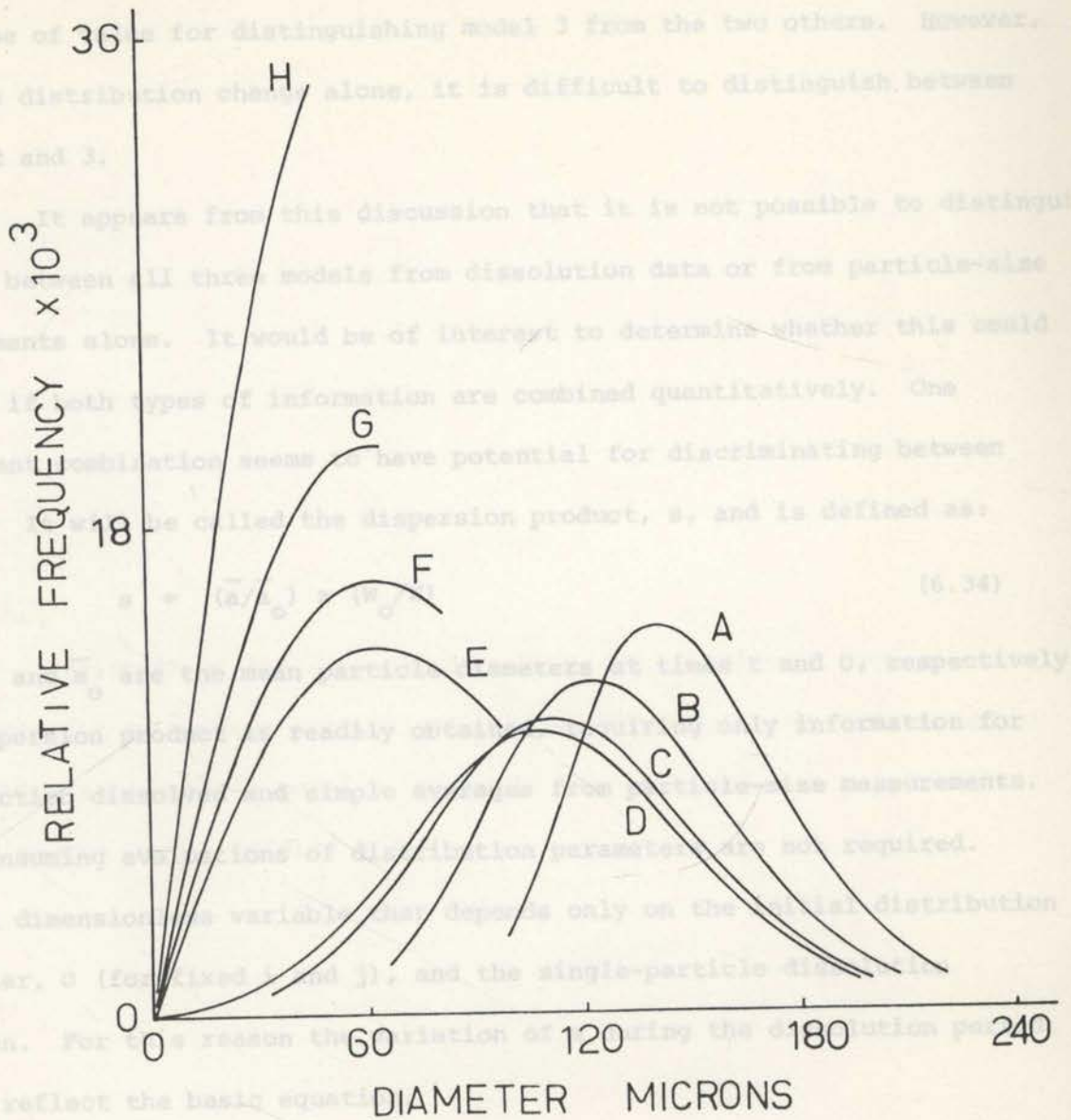


Figure 6.9 shows this variation for powders initially log-normally distributed ( $\sigma = 0.2$ ,  $i = j = z$ ). The curves representing the three dissolution models are significantly different. The basic shape of the curve remains the same for varying values of  $\sigma$ , although the minima shift to the right and to higher values for very narrow distributions. All three curves approach  $\alpha = 1$  (stippled line) when  $\sigma$  approaches zero as expected for a completely monodisperse powder. The values of the three minima remain approximately constant. When  $\sigma$  ranges from somewhat less than 0.1 to at least 0.2 (Fig. 6.10), which encompasses most fine powder distributions



should be of value for distinguishing model 3 from the two others. However, from the distribution change alone, it is difficult to distinguish between Models 2 and 3.

It appears from this discussion that it is not possible to distinguish clearly between all three models from dissolution data or from particle-size measurements alone. It would be of interest to determine whether this could be done if both types of information are combined quantitatively. One convenient combination seems to have potential for discriminating between models. It will be called the dispersion product,  $s$ , and is defined as:

Figure 6.9 
$$s = (\bar{a}/\bar{a}_0) \times (W_0/W) \quad (6.34)$$

where  $\bar{a}$  and  $\bar{a}_0$  are the mean particle diameters at times  $t$  and  $0$ , respectively. The dispersion product is readily obtained, requiring only information for the fraction dissolved and simple averages from particle-size measurements. Time-consuming evaluations of distribution parameters are not required. It is a dimensionless variable that depends only on the initial distribution parameter,  $\sigma$  (for fixed  $i$  and  $j$ ), and the single-particle dissolution equation. For this reason the variation of  $s$  during the dissolution period should reflect the basic equation.

Figure 6.9 shows this variation for powders initially log-normally distributed ( $\sigma = 0.2$ ,  $i = j = z$ ). The curves representing the three dissolution models are significantly different. The basic shape of the curve remains the same for varying values of  $\sigma$ , although the minima shift to the right and to higher values for very narrow distributions. All three curves approach  $s = 1$  (stippled line) when  $\sigma$  approaches zero as expected for a completely monodisperse powder. The values of the three minima remain approximately constant. When  $\sigma$  ranges from somewhat less than 0.1 to at least 0.2 (Fig. 6.10), which encompasses most fine powder distributions

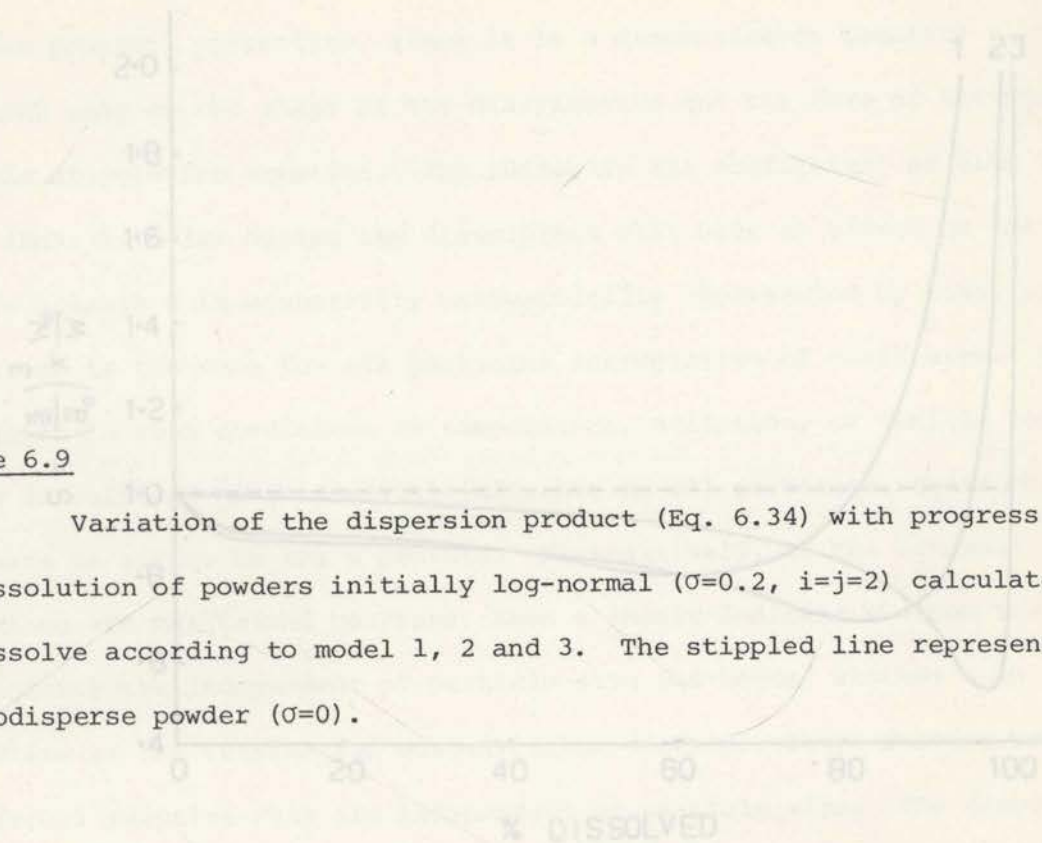
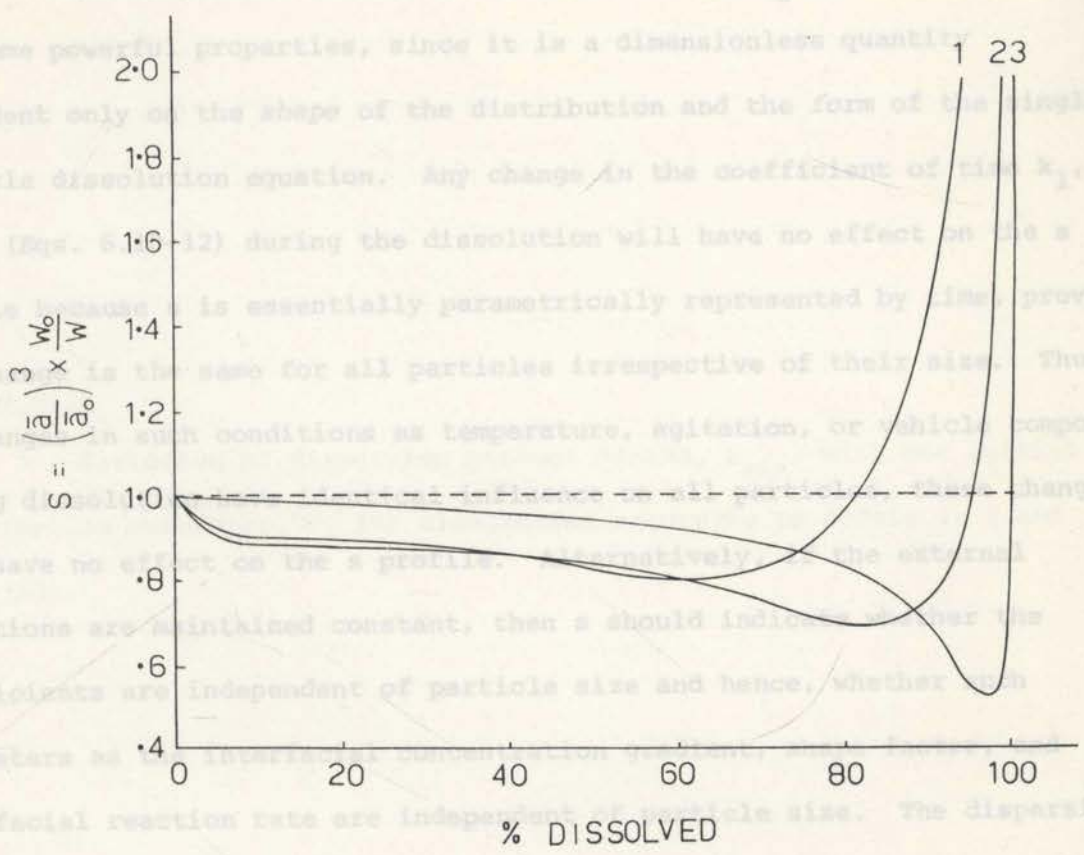


Figure 6.9

Variation of the dispersion product (Eq. 6.34) with progress of dissolution of powders initially log-normal ( $\sigma=0.2, i=j=2$ ) calculated to dissolve according to model 1, 2 and 3. The stippled line represents a monodisperse powder ( $\sigma=0$ ).

encountered in practice. Theoretically, the considerable difference between the  $s_{min}$  values should make it possible to distinguish between the three models.

The dispersion product profile is versus  $W/W_0$  or percent dissolved has some powerful properties, since it is a dimensionless quantity dependent only of the shape of the distribution and the form of the single-particle dissolution equation. Any change in the coefficient of time  $k_1$ ,  $k_2$  or  $k_3$  (Eqs. 5, 6, 12) during the dissolution will have no effect on the  $s$  profile because  $s$  is essentially parametrically represented by  $W/W_0$  provided the  $W/W_0$  is the same for all particles irrespective of their size. Thus, if changes in such conditions as temperature, agitation, or vehicle composition during dissolution will have no effect on the  $s$  profile. Alternatively, if the external conditions are maintained constant, then  $s$  should indicate whether the coefficients are independent of particle size and hence, whether such parameters as interfacial reaction rate are independent of particle size. The dispersion product should, therefore, be a valuable tool in dissolution kinetic studies.



The extent to which these mathematical models can be applied to describe the dissolution of a "real" powder depends on three assumptions.

1. As mentioned earlier, it was assumed that the particles dissolve independently of each other. This should be approximated well under sink conditions.

2. It was assumed that the dissolution of each particle in the powder can be described by an equation having the same parameter values for all the particles. In practice, this assumption is rarely valid because of differences in individual particle shapes, crystal structure, and interaction with the vehicle. However, these types of effects probably can be averaged to produce a parameter value for the single-particle dissolution

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The extent to which these mathematical models can be applied to describe the dissolution of a "real" powder depends on three assumptions.

1. As mentioned earlier, it was assumed that the particles dissolve independently of each other. This should be approximated well under sink conditions.

2. It was assumed that the dissolution of each particle in the powder can be described by an equation having the same parameter value ( $k$ ) for all the particles. In practice, this assumption is rarely valid because of differences in individual particle shapes, crystal structure, and interaction with the vehicle. However, these types of effects probably can be averaged to produce a parameter value for the single-particle dissolution

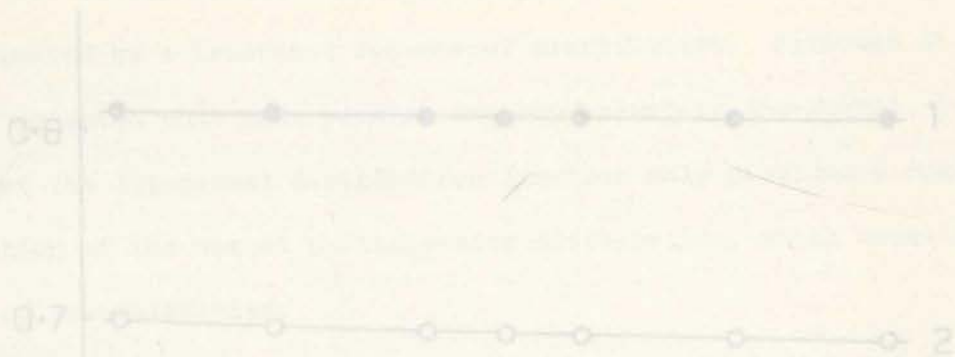


Figure 6.10

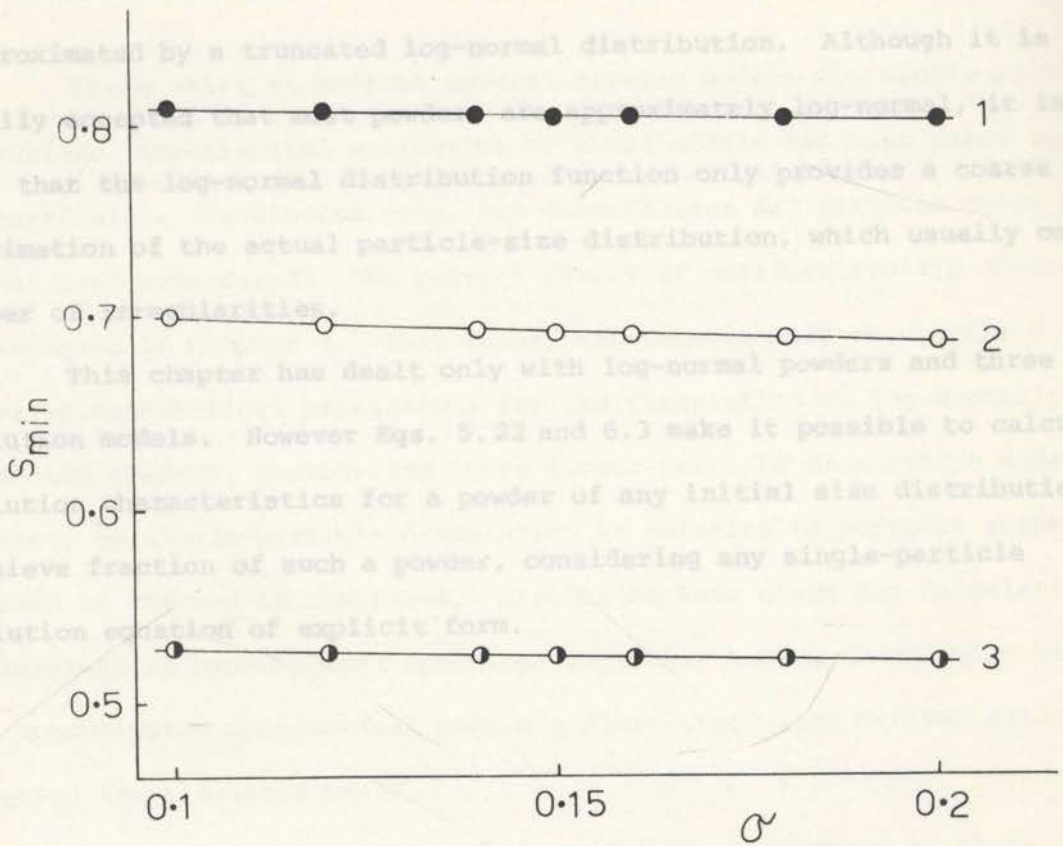
Variation of dispersion product minima,  $S_{min}$ , with the initial distribution parameter,  $\sigma$ , for dissolution according to Models 1, 2 and 3 ( $i=j=2$ ).



model which, when used in the multiparticulate dissolution equation, results in a good approximation of the actual multiparticulate dissolution behavior.

3. It was assumed that the initial particle-size distribution can be approximated by a truncated log-normal distribution. Although it is generally likely that the log-normal distribution function only provides a coarse approximation of the actual particle-size distribution, which usually contains a number of particles.

This chapter has dealt only with log-normal powders and three dissolution models. However Eqs. 5.22 and 6.3 make it possible to calculate dissolution characteristics for a powder of any initial size distribution, or a sieve fraction of such a powder, considering any single-particle dissolution equation of explicit form.



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EXPERIMENTAL EVALUATION OF THREE DISSOLUTION MODELS

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generally accepted that most powders are approximately log-normal, it is likely that the log-normal distribution function only provides a coarse approximation of the actual particle-size distribution, which usually contains a number of irregularities.

This chapter has dealt only with log-normal powders and three dissolution models. However Eqs. 5.22 and 6.3 make it possible to calculate dissolution characteristics for a powder of any initial size distribution, or a sieve fraction of such a powder, considering any single-particle dissolution equation of explicit form.

The theory of single-particle dissolution in relation to particle shape was discussed at the end of Chapter 4. Directions were given for calculating the dissizers of hypothetical spherical particles having dissolution behavior which approximates nonspherical particle dissolution with minimum error on a weighted least squares basis.

This chapter demonstrates the combined application of these theories to explain the dissolution kinetics of 60/85-mesh fraction of tobutamide such that both size distribution effects and particle shape effects are considered. By using a time-scaling approach, the three single-particle dissolution kinetic models are evaluated.

Theoretical Considerations

The three equations for dissolution of spherical particles under sink conditions considered can be written in common form as:

$$w = \left( w_{\infty} \frac{1}{n} - kt \right)^m \quad m = 3, 2, \frac{3}{2} \quad (7.1)$$
$$k = k_1, k_2, k_3$$

CHAPTER 7

EXPERIMENTAL EVALUATION OF THREE DISSOLUTION MODELS

There exist at present several kinetic models for single particle dissolution. Experimental evaluation of these models has been based on multiparticulate dissolution data, but distribution and particle shape effects have not been considered. The general theory of multiparticulate dissolution was discussed in Chapter 5. This theory was subsequently in Chapter 6 used to develop mathematical expressions for the dissolution of log-normally distributed powders, considering three single-particle dissolution models. The theory of single-particle dissolution in relation to particle shape was discussed at the end of Chapter 4. Directions were given for calculating the diameters of hypothetical spherical particles having dissolution behaviour which approximates nonspherical particle dissolution with minimum error on a weighted least squares basis.

This chapter demonstrates the combined application of these theories to explain the dissolution kinetics of 60/85-mesh fraction of tolbutamide such that both size distribution effects and particle shape effects are considered. By using a time-scaling approach, the three single-particle dissolution kinetic models are evaluated.

Theoretical Considerations

The three equations for dissolution of spherical particles under sink conditions considered can be written in common form as:

$$w = (w_0^{\frac{1}{m}} - kt)^m \quad m = 3, 2, \frac{3}{2} \quad (7.1)$$

$$k = k_1, k_2, k_3$$



$$\int_{R_2} (w^2 - kt)^{3/2} w^{-1} N(\ln w, \mu, \sigma) dw$$

representing the cube root, the square root and the 3/2-root models, respectively (97,106,107). The positive constants  $k_1$ ,  $k_2$  and  $k_3$  replace the original coefficients of time. These contained quantities such as density, diffusion coefficient and shape factor. This simplification is made because the aim is not to evaluate the theoretical basis of the three equations but solely to assess them as models for describing the dissolution kinetic data.  $\ln w$  as variable, defined by:

These equations (7.1) do not strictly describe the dissolution correctly in their present form since  $w$  does not vanish for  $t \rightarrow \infty$ . A more correct formulation would therefore be:

$$w = (w_0^{\frac{1}{m}} - kt)^m \text{ for } t \leq w_0^{\frac{1}{m}} / k \quad (7.2)$$

$$w = 0 \text{ for } t > w_0^{\frac{1}{m}} / k$$

The general multiparticulate dissolution equation presented earlier (5.22) however was developed such that it accepts the single particle dissolution function in the forms of both 7.1 and 7.2. Equation 7.2 will be used for simplicity.

In Chapter 6 the following equations were presented to describe the dissolution profile of a multiparticulate log-normal system where the spherical particles dissolve according to each of the three models (7.1):

$$\frac{W}{W_0} = \sum_{n=0}^m \binom{m}{n} (-Kt)^{(m-n)} \frac{F(A) - F(B)}{F(j-3\sigma) - F(-i-3\sigma)} \text{EXP}(C) \quad (7.3)$$

where

$$A = (T_2 - \mu) / \sigma - 3n\sigma / m$$

$$B = (T_1 - \mu) / \sigma - 3n\sigma / m$$

$$C = \text{EXP} \left[ \frac{3}{m} (n-m) \left( \mu + \frac{3}{m} (n+m) \sigma^2 / 2 \right) \right]$$

$$T_1 = \max \left[ \frac{m}{3} \ln(Kt), \mu - i\sigma \right]$$

$$T_2 = \max \left[ \frac{m}{3} \ln(Kt), \mu + j\sigma \right]$$

$$\frac{W}{W_0} = \frac{\int_{R_1}^{R_2} (w^2 - Kt)^{3/2} w^{-1} N(\ln w, \mu, \sigma) dw}{F(j-3\sigma) - F(-i-3\sigma)} \text{EXP}(-3\mu - 9\sigma^2/2) \quad (7.4)$$

where  $R_1 = \max \left[ (Kt)^{1/2}, \text{EXP}(\mu - i\sigma) \right]$   
 $R_2 = \max \left[ (Kt)^{1/2}, \text{EXP}(\mu + j\sigma) \right]$

The function  $F(\ )$  above is the area under standard normal curve function defined earlier (5.37). The function  $N(\ln w, \mu, \sigma)$  is the normal distribution function with  $\ln w$  as variable, defined by:

$$N(\ln w, \mu, \sigma) = \frac{1}{\sigma\sqrt{2\pi}} \text{EXP} \left[ -\frac{1}{2} \left( \frac{\ln w - \mu}{\sigma} \right)^2 \right]$$

The constant  $K$  is related to  $k_1, k_2$  and  $k_3$  by:

$$K = (6/\rho\pi)^m k_i \quad i = 1, 2, 3 \quad (7.6)$$

where  $\rho$  is the particle density.

It is assumed that the particle diameter ( $a$ ) distribution is "log-normal", that is,  $\ln a$  can be approximated by a normal distribution (mean =  $\mu$ , standard deviation =  $\sigma$ ) truncated at the lower end at  $\mu - i\sigma$  and at the upper end at  $\mu + j\sigma$ , where  $i$  and  $j$  are truncation parameters (Fig. 5.1).

The multiparticulate dissolution equations above consider spherical drug particles. Such particles are only encountered when the drug exists in liquid form as an emulsion. In solid form the particles are not spherical. The drug used for the dissolution tests was a 60/85 mesh fraction of Tolbutamide consisting of particles approximately tetragonal prismatic in shape. It was shown in Chapter 4 that the dissolution of such particles can be approximated well by the dissolution of hypothetical spherical particles. The equivalent spherical diameter,  $a$ , is the diameter of the spherical particle that best approximates the dissolution of the non-spherical particle, and is given by:

$$a = \frac{b_0}{2 - (2 - \frac{b_0}{l_0})^{1/3}} \quad (7.7)$$

where  $l_0$  and  $b_0$  are the length and side respectively of the tetragonal particle. In this way dissolution of the non-spherical particle system can be suitably described by the dissolution of a hypothetical spherical particle system that can be rigorously treated using the equations above.

It is evident from these that  $k$ ,  $\mu$ ,  $\sigma$ ,  $i$ ,  $j$  and  $\rho$  must be known to calculate the dissolution profile. The distribution parameters  $\mu$ ,  $\sigma$ ,  $i$  and  $j$  can be obtained from micrographs and  $\rho$  by a standard method, but the single particle rate parameter  $k$  is unknown. It is possible however to calculate the exact intrinsic dissolution profile with much less information. According to the rules given previously (Chapter 6) for multiparticulate dissolution only the *shape* of the initial distribution, that is, of the above six parameters only  $\sigma$ ,  $i$  and  $j$  are required to calculate the intrinsic dissolution profile when the single particle dissolution model is known. The concept of time scaling was discussed in Chapter 6. By such an approach it is possible to evaluate quantitatively the difference between the actual dissolution data and the calculated intrinsic dissolution profile.

Figure 7.1 illustrates this application of time scaling. Curve B through the experimental data points represents the dissolution curve  $((w/w_0)^m \text{ versus } t)$  and curve A the corresponding calculated normalised intrinsic dissolution profile  $((w/w_0)^m \text{ versus } \psi)$ .

Let,  $N$ , be the number of data points and,  $f$ , the time scaling factor that brings curve B "into" curve A such that the sum of the squared deviations between the curves given by:

$$ss = \sum_{i=1}^N (ft_i - \psi_i)^2 \quad (7.8)$$

is a minimum. This means  $f$  is obtained from  $\partial ss / \partial f = 0$  which gives:

$$f = \frac{\sum_{i=1}^N \psi_i t_i}{\sum_{i=1}^N t_i^2} \quad (7.9)$$

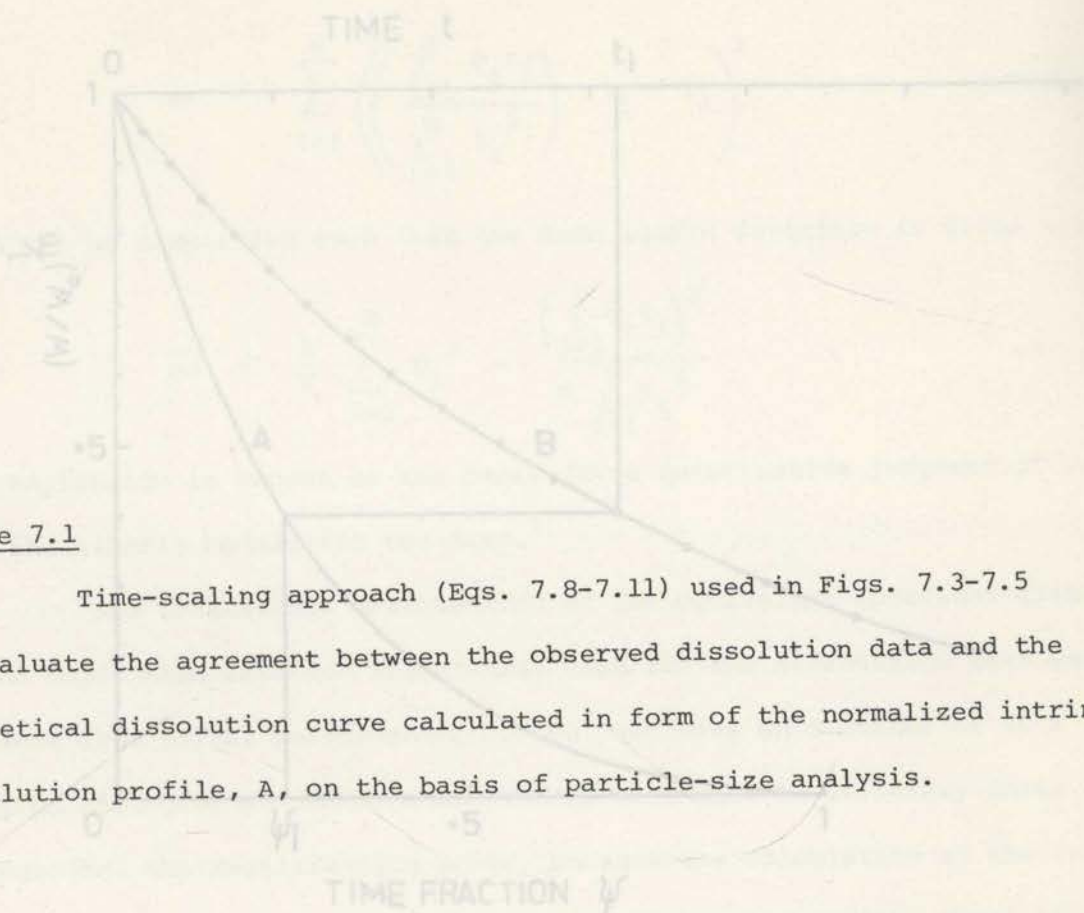
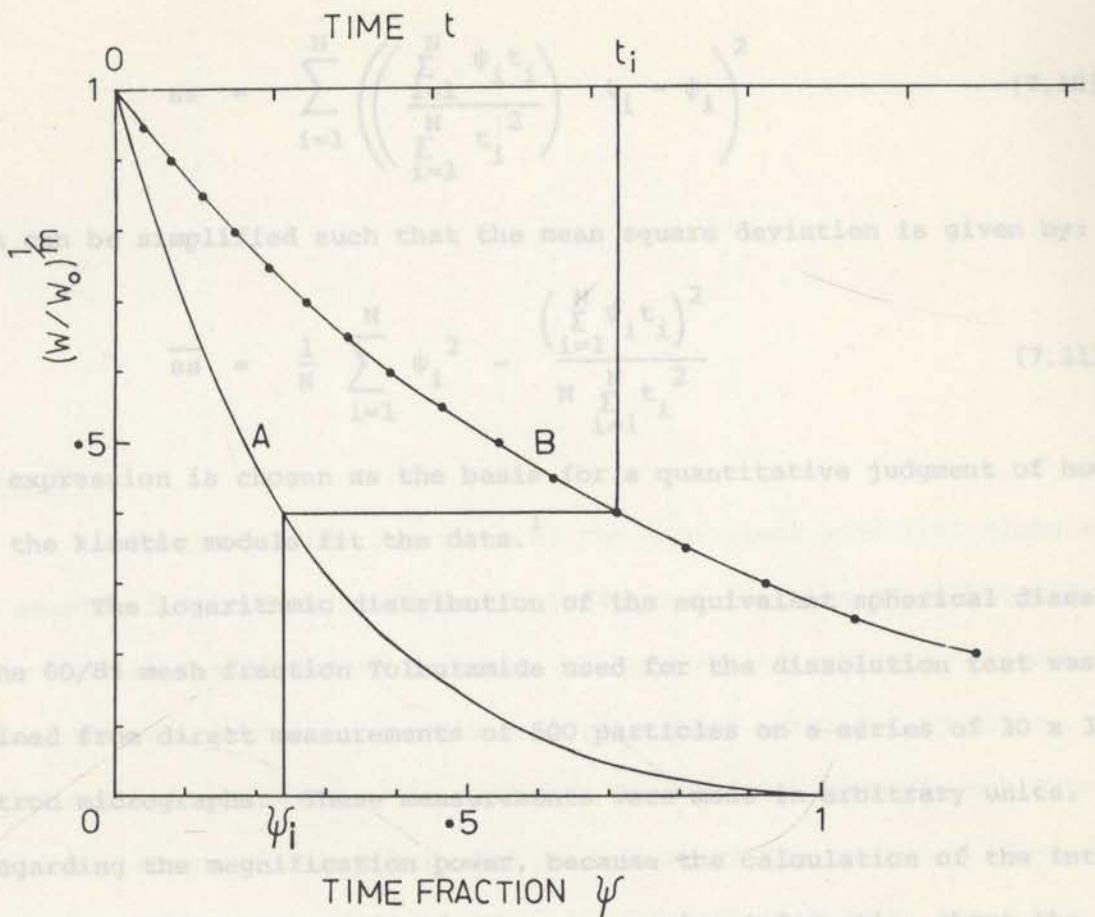


Figure 7.1

Time-scaling approach (Eqs. 7.8-7.11) used in Figs. 7.3-7.5 to evaluate the agreement between the observed dissolution data and the theoretical dissolution curve calculated in form of the normalized intrinsic dissolution profile, A, on the basis of particle-size analysis.

The minimum of the sum of squares is therefore:



which is simplified such that the mean square deviation is given by:

$$ss = \frac{1}{N} \sum_{i=1}^N \psi_i^2 - \left( \frac{\sum_{i=1}^N \psi_i}{N} \right)^2 \quad (7.11)$$

This expression is chosen as the basis for quantitative judgment of how well the kinetic models fit the data.

The logarithmic distribution of the equivalent spherical diameters of the 60/80 mesh fraction tubastide used for the dissolution test was obtained from direct measurements of 500 particles on a series of 10 x 10 cm electrostatic precipitator plates.

regarding the magnification power, because the calculation of the intrinsic dissolution profile as mentioned does not require information about the actual sizes of the particles but the shape of their distribution (that is,  $\sigma$ ,  $\mu$  and  $\lambda$  for a log-normal powder). Each particle was approximated by the tetragonal prismatic body which fitted best, and its equivalent spherical diameter was calculated using Eq. 7.7.

The histogram of the logarithm of these diameters (Fig. 7.2) shows a good fit to a normal distribution with standard deviation  $\sigma = 0.395$  and mean

1. The alternative approach, to bring curve A "into" curve B, that has the character of a curve fitting to the data points would yield the same result in a comparison of the models. The above time scaling of the data is used for the convenience of plotting and to better illustrate the predicted time for complete dissolution.

The minimum of the sum of squares is therefore:

$$ss = \sum_{i=1}^N \left( \left( \frac{\sum_{i=1}^N \psi_i t_i}{\sum_{i=1}^N t_i^2} \right) t_i - \psi_i \right)^2 \quad (7.10)$$

which can be simplified such that the mean square deviation is given by:

$$\overline{ss} = \frac{1}{N} \sum_{i=1}^N \psi_i^2 - \frac{\left( \frac{\sum_{i=1}^N \psi_i t_i}{\sum_{i=1}^N t_i^2} \right)^2}{N} \quad (7.11)$$

This expression is chosen as the basis for a quantitative judgment of how well the kinetic models fit the data.<sup>1</sup>

The logarithmic distribution of the equivalent spherical diameters of the 60/85 mesh fraction Tolbutamide used for the dissolution test was obtained from direct measurements of 500 particles on a series of 30 x 30 cm electron micrographs. These measurements were made in arbitrary units, disregarding the magnification power, because the calculation of the intrinsic dissolution profile as mentioned does not require information about the actual sizes of the particles but the shape of their distribution (that is,  $\sigma$ ,  $i$  and  $j$  for a log-normal powder). Each particle was approximated by the tetragonal prismatic body which fitted best, and its equivalent spherical diameter was calculated using Eq. 7.7.

The histogram of the logarithm of these diameters (Fig. 7.2) shows a good fit to a normal distribution with standard deviation  $\sigma = 0.395$  and mean

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1. The alternative approach, to bring curve A "into" curve B, that has the character of a curve fitting to the data points would yield the same result in a comparison of the models. The above time scaling of the data is used for the convenience of plotting and to better illustrate the predicted time for complete dissolution.

relative frequency

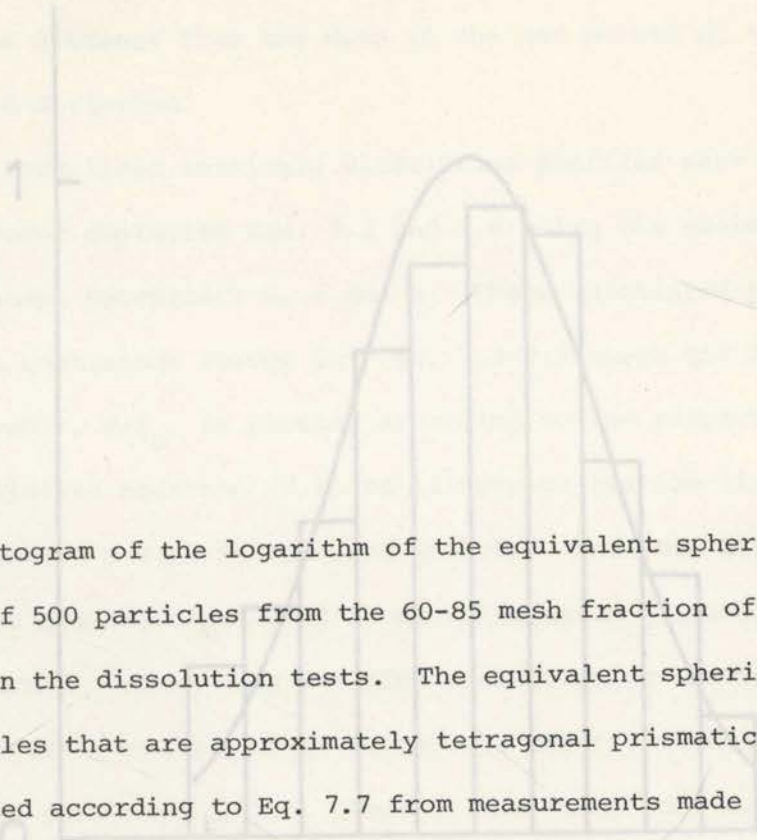
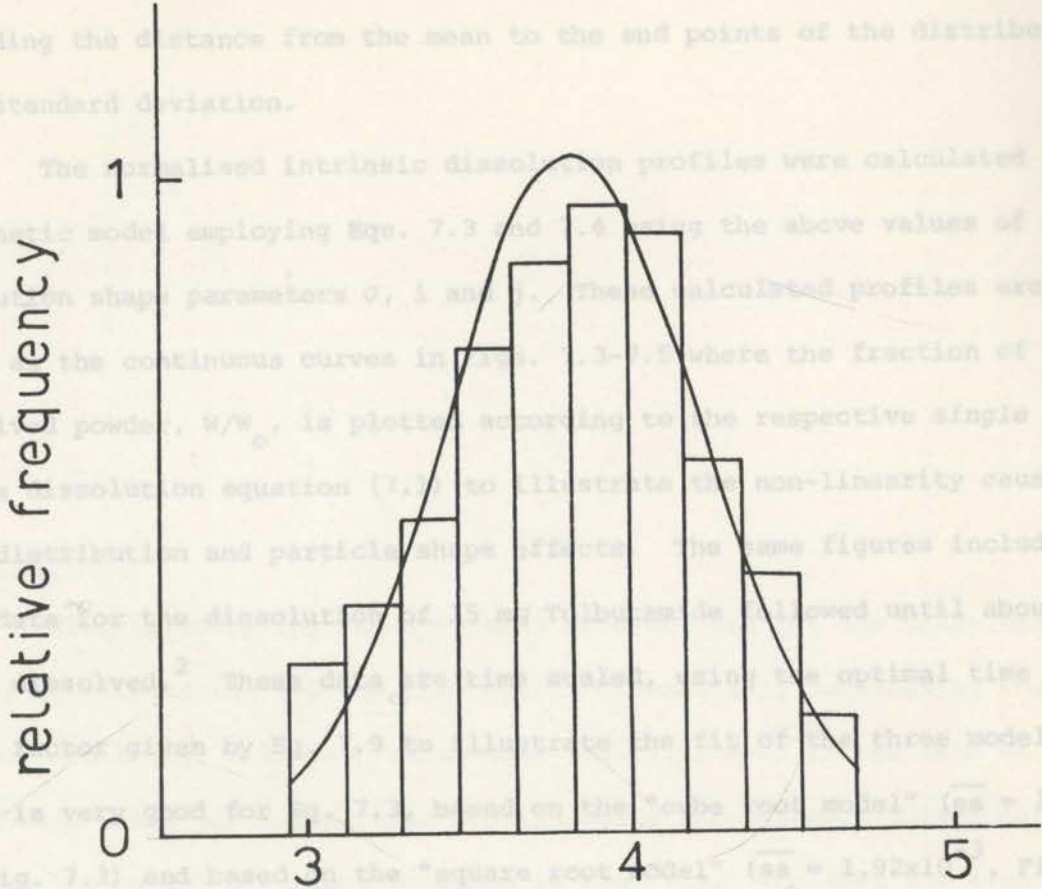


Figure 7.2

Histogram of the logarithm of the equivalent spherical diameters of a sample of 500 particles from the 60-85 mesh fraction of tolbutamide powder used in the dissolution tests. The equivalent spherical diameters of the particles that are approximately tetragonal prismatic in shape were calculated according to Eq. 7.7 from measurements made (in arbitrary units) on electron micrographs. The parameters of the truncated log-normal distribution which best fits this diameter distribution are:  $\sigma = 0.395$ ,  $\mu = 3.82$ ,  $i = 2.25$ , and  $j = 2.20$ .

$\mu = 3.62$ . The truncation parameters,  $i = 2.25$ , and  $j = 2.20$ , were obtained by dividing the distance from the mean to the end points of the distribution by the standard deviation.

The calculated intrinsic dissolution profiles were calculated for each kinetic model employing Eqs. 7.3 and 7.4 using the above values of the distribution shape parameters  $\sigma$ ,  $i$  and  $j$ . These calculated profiles are graphed as the continuous curves in Figs. 7.3-7.5 where the fraction of undissolved powder,  $W/W_0$ , is plotted according to the respective single particle dissolution equation (7.1) to illustrate the non-linearity caused by the distribution and particle shape effects. The same figures include a set of data for the dissolution of 15 mg Tolbutamide followed until about 90% had dissolved. These data are time scaled, using the optimal time scaling factor given by Eq. 7.3 to illustrate the fit of the three models. The fit is very good for Fig. 7.3 based on the "cube root model" ( $\bar{x}_s = 1.10 \times 10^{-3}$ , Fig. 7.3) and based on the "square root model" ( $\bar{x}_s = 1.92 \times 10^{-3}$ , Fig. 7.4)



ln (equivalent diameter)  
arbitrary units

but not so good for the "square root model" ( $\bar{x}_s = 1.92 \times 10^{-3}$ , Fig. 7.4) and not so good for the "2/3-root model" ( $\bar{x}_s = 1.10 \times 10^{-3}$ , Fig. 7.5). The dissolution data were analyzed using different amounts of Tolbutamide. The  $\bar{x}_s$  values (Table 7.1) indicate (F-test,  $P < .05$ ) that the "cube root model" describes the single particle dissolution best with the "square root model" almost as good, however, the "2/3-root model" is relatively poor.

In order to make the above evaluations of the three models by comparing dissolution data with theoretical calculations it is necessary that the experimental conditions are consistent with the assumptions behind these calculations. The three assumptions on which Eqs. 7.3 and 7.4 are based are:

1. The absorbance after 90% dissolution was so small that substantial error would have been introduced if the process were followed much further.



$\mu = 3.82$ . The truncation parameters,  $i = 2.25$ , and  $j = 2.20$ , were obtained by dividing the distance from the mean to the end points of the distribution by the standard deviation.

The normalised intrinsic dissolution profiles were calculated for each kinetic model employing Eqs. 7.3 and 7.4 using the above values of the distribution shape parameters  $\sigma$ ,  $i$  and  $j$ . These calculated profiles are graphed as the continuous curves in Figs. 7.3-7.5 where the fraction of undissolved powder,  $W/W_0$ , is plotted according to the respective single particle dissolution equation (7.1) to illustrate the non-linearity caused by the distribution and particle shape effects. The same figures include a set of data for the dissolution of 15 mg Tolbutamide followed until about 90% had dissolved.<sup>2</sup> These data are time scaled, using the optimal time scaling factor given by Eq. 7.9 to illustrate the fit of the three models. The fit is very good for Eq. 7.3, based on the "cube root model" ( $\overline{ss} = 1.10 \times 10^{-3}$ , Fig. 7.3) and based on the "square root model" ( $\overline{ss} = 1.92 \times 10^{-3}$ , Fig. 7.4) but not so good for the "2/3-root model" ( $\overline{ss} = 6.70 \times 10^{-3}$ , Fig. 7.5).

The dissolution test was done several times using different amounts of Tolbutamide. The  $\overline{ss}$  - values (Table 7.1) indicate (F-test,  $P < .05$ ) that the "cube root model" describes the single particle dissolution best with the "square root model" almost as good, however, the "2/3-root model" is relatively poor.

In order to make the above evaluations of the three models by comparing dissolution data with theoretical calculations it is necessary that the experimental conditions are consistent with the assumptions behind these calculations. The three assumptions on which Eqs. 7.3 and 7.4 are based are:

---

2. The absorbance after 90% dissolution was so small that substantial error would have been introduced if the process were followed much further.



Figure 7.3

Plot illustrating the agreement between data for the dissolution of 15 mg of tolbutamide and the theoretical dissolution, considering the cube root model (Eq. 7.1,  $m=3$ ) calculated (Eq. 7.3,  $m=3$ ) in the form of a normalized intrinsic dissolution profile using the parameters from the truncated log-normal distribution shown in Fig. 7.2. The data are time scaled using the scaling factor given by Eq. 7.9. The mean square deviation  $\overline{ss} = 1.10 \times 10^{-3}$ .

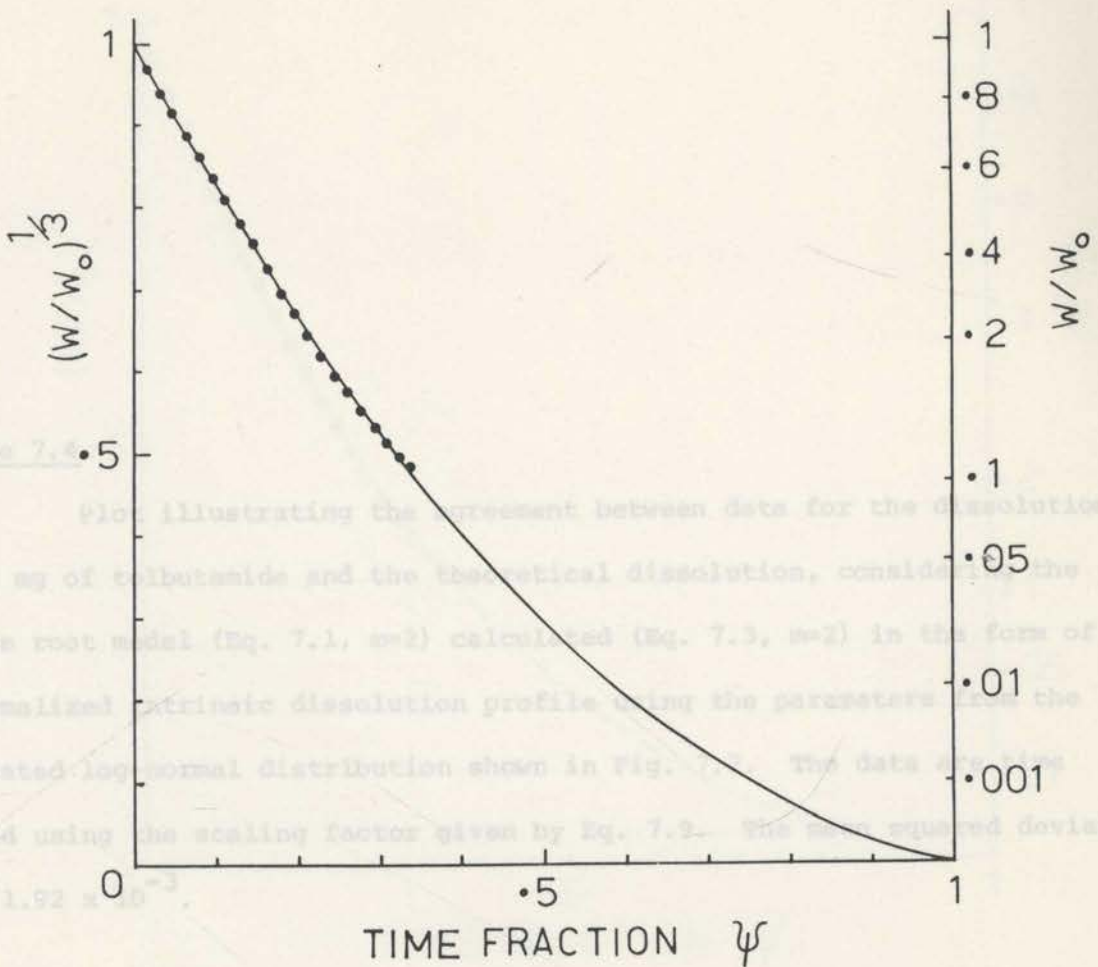


Figure 7.4. Plot illustrating the agreement between data for the dissolution of 15 mg of salbutamol and the theoretical dissolution, considered by the square root model (Eq. 7.1,  $n=2$ ) calculated (Eq. 7.3,  $n=2$ ) in the form of a normalized extrinsic dissolution profile using the parameters from the truncated lognormal distribution shown in Fig. 7.3. The data are scaled using the scaling factor given by Eq. 7.2. The mean squared deviation  $\overline{d^2} = 1.92 \times 10^{-3}$ .

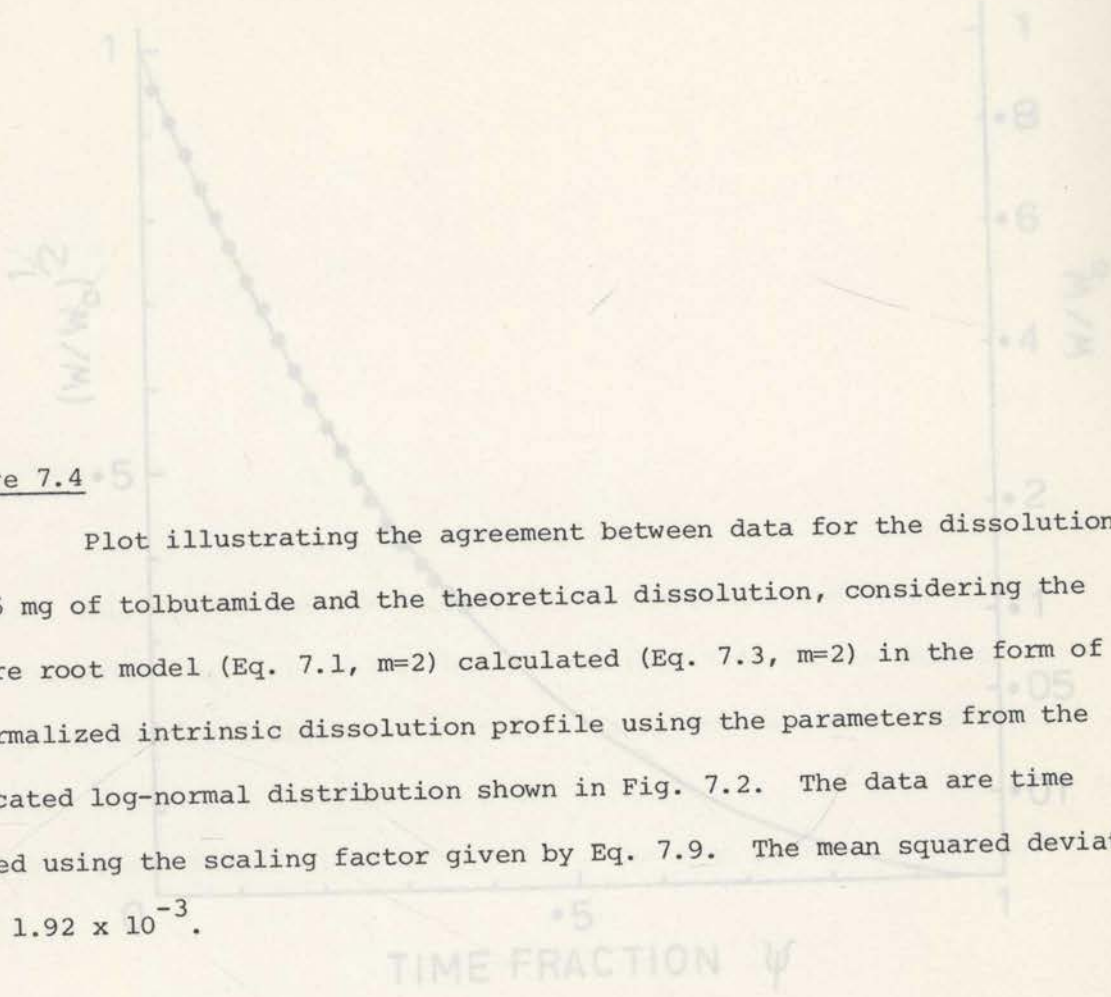


Figure 7.4

Plot illustrating the agreement between data for the dissolution of 15 mg of tolbutamide and the theoretical dissolution, considering the square root model (Eq. 7.1,  $m=2$ ) calculated (Eq. 7.3,  $m=2$ ) in the form of a normalized intrinsic dissolution profile using the parameters from the truncated log-normal distribution shown in Fig. 7.2. The data are time scaled using the scaling factor given by Eq. 7.9. The mean squared deviation  $\overline{ss} = 1.92 \times 10^{-3}$ .

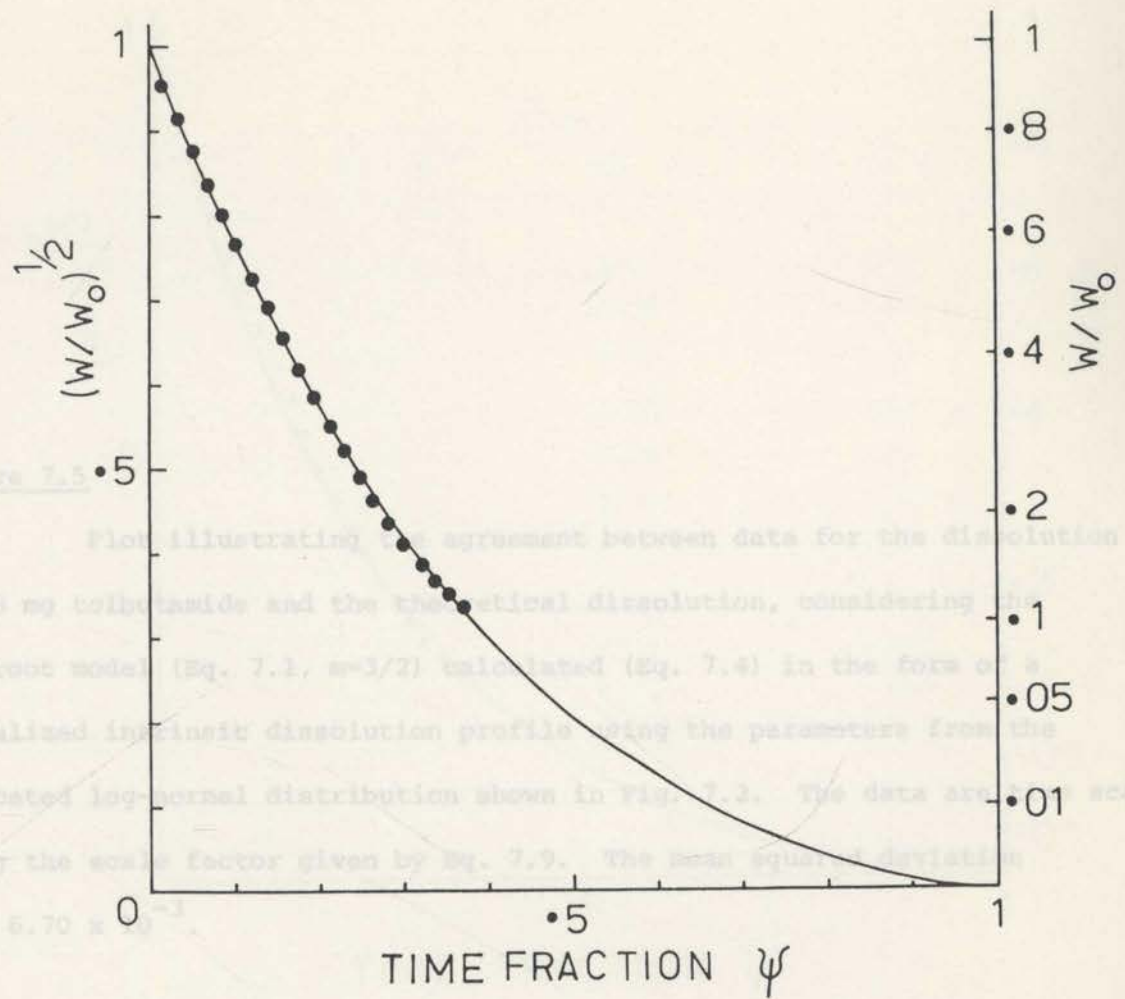


Figure 7.5 Plot illustrating the agreement between data for the dissolution of 15 mg valbutamide and the theoretical dissolution, considering a 2/3-root model (Eq. 7.1,  $n=3/2$ ) calculated (Eq. 7.4) in the form of a normalized infinite dissolution profile using the parameters from the truncated log-normal distribution shown in Fig. 7.2. The data are scaled using the scale factor given by Eq. 7.9. The mean squared deviation  $ss = 6.70 \times 10^{-3}$

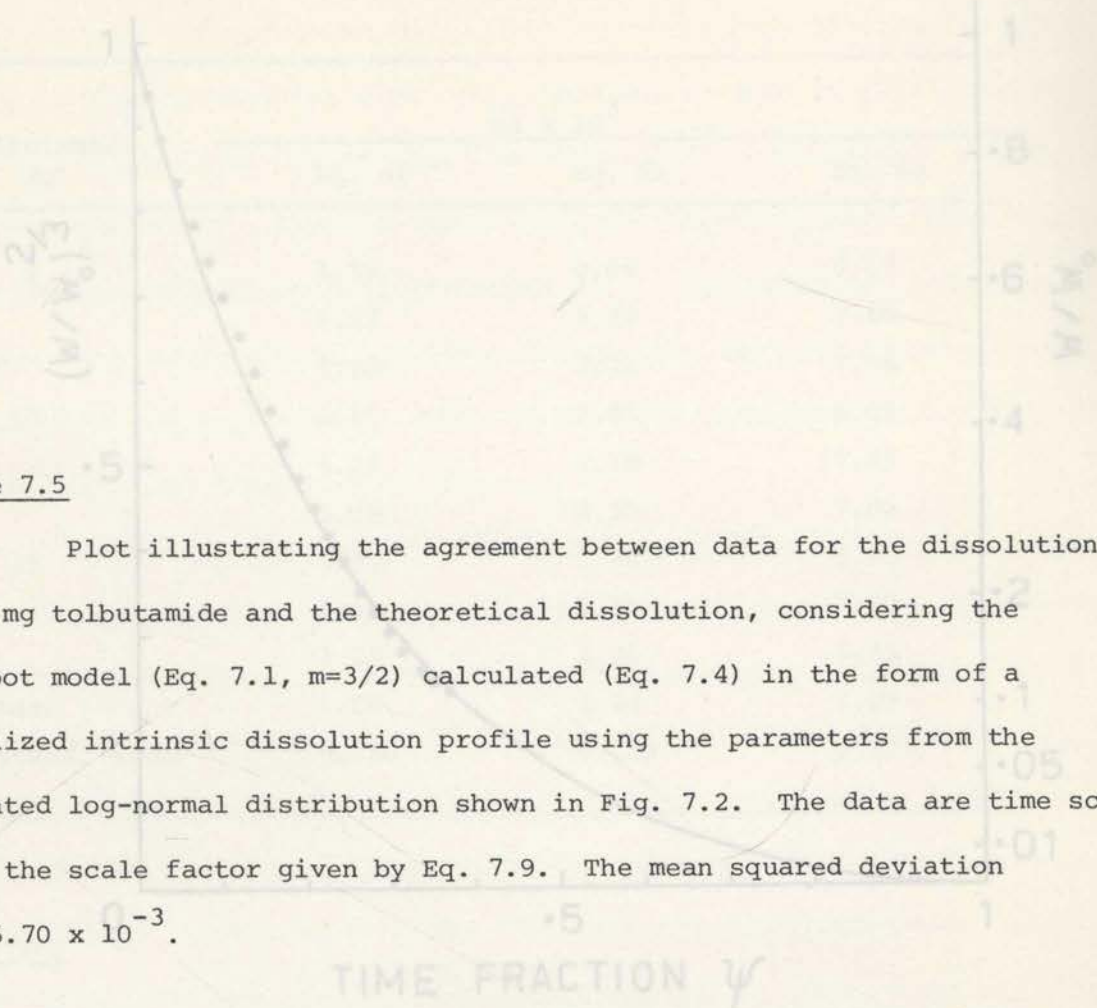


Figure 7.5

Plot illustrating the agreement between data for the dissolution of 15 mg tolbutamide and the theoretical dissolution, considering the 2/3-root model (Eq. 7.1,  $m=3/2$ ) calculated (Eq. 7.4) in the form of a normalized intrinsic dissolution profile using the parameters from the truncated log-normal distribution shown in Fig. 7.2. The data are time scaled using the scale factor given by Eq. 7.9. The mean squared deviation  $\overline{ss} = 6.70 \times 10^{-3}$ .

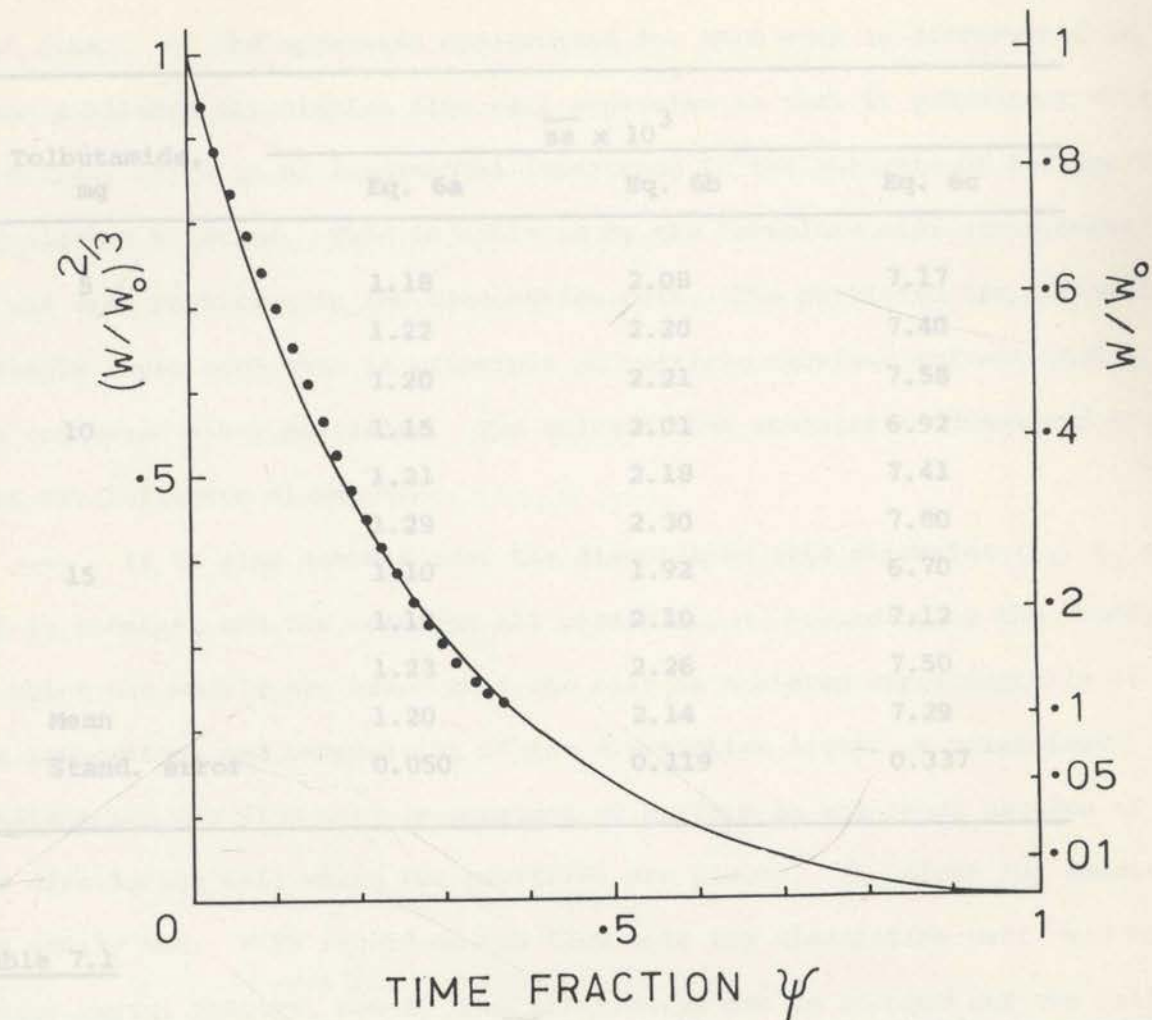


Table 7.1

Mean square deviation,  $ss$  (Eq. 7.11) as a quantitative comparison of the fit of the three multiparticulate dissolution equations. Eqs. 7.3 ( $m=2,3$ ) and Eq. 7.4 (based on the single-particle dissolution models of Eq. 7.1 ( $m=3,2, 3/2$ )) to the data from the dissolution of various amounts of 60/85 mesh fraction of tolbutamide.

1. — It is assumed that the particles dissolve independently of each other. — The apparatus constructed for this work is different from

Tolbutamide, mg	$\overline{ss} \times 10^3$		
	Eq. 6a	Eq. 6b	Eq. 6c
5	1.18	2.08	7.17
	1.22	2.20	7.40
	1.20	2.21	7.58
10	1.15	2.01	6.92
	1.21	2.18	7.41
	1.29	2.30	7.80
15	1.10	1.92	6.70
	1.19	2.10	7.12
	1.23	2.26	7.50
Mean	1.20	2.14	7.29
Stand. error	0.050	0.119	0.337

Table 7.1

Mean square deviation,  $\overline{ss}$  (Eq. 7.11) as a quantitative comparison of the fit of the three multiparticulate dissolution equations. Eqs. 7.3 (m=2,3) and Eq. 7.4 (based on the single-particle dissolution models of Eq. 7.1 (m=3,2, 3/2) to the data from the dissolution of various amounts of 60/85 mesh fraction of tolbutamide.

3. — It is further assumed that the initial particle size distribution can be approximated by a truncated log-normal distribution function. — Figure 7.2 shows that this is a good approximation for the 300 particles measured, however, it does not guarantee the correctness of the



1. — It is assumed that the particles dissolve independently of each other. - The apparatus constructed for this work is different from other published dissolution flow cell apparatus in that it guarantees this condition, which is of fundamental importance in the analysis of multiparticulate dissolution kinetics. This is achieved by the "absolute sink arrangement" of the drug particles in the dissolution cell. The particles are placed in a single layer such that in principle no particle receives solvent that has contacted other particles. The solvent thus contains no dissolved drug that may influence dissolution. -

2. — It is also assumed that the dissolution rate parameter ( $k_1$ ,  $k_2$  or  $k_3$ ) is constant and the same for all particles. - According to the theory on which the models are based this can only be achieved experimentally if the temperature and composition of the dissolution liquid is maintained constant and the flow rate is constant or uniform in the cross section of the dissolution cell where the particles are placed. The first two conditions are easily met. With regard to the flow rate the dissolution cell used has a very useful feature, namely that the process can be stopped and the cell rapidly disconnected allowing the particles to be inspected at any stage of the dissolution process. Such inspections showed (after microscopic measurements) uniform dissolution over the whole particle layer indicating a uniform flow rate. The fact that the particles in the dissolution cell can be inspected in this way makes it possible for the dissolution data to be combined with particle size measurements.

3. — It is further assumed that the initial particle size (diameter) distribution can be approximated by a truncated log-normal distribution function. - Figure 7.2 shows that this is a good approximation for the 500 particles measured, however, it does not guarantee the correctness of the

results in a mathematical expression which is much more complex than Eqs.

assumption that this small sample represents the particle size distribution in the samples used for the dissolution tests, although the uniformity of the powder supported this assumption.

These investigations indicate that it is possible to describe mathematically, the dissolution of a multiparticulate system with a high degree of accuracy by considering both the particle size distribution effect and the particle shape effect discussed earlier. It is evident from the dissolution data obtained, that among the three models investigated, the cube root model describes the kinetics best.

It is possible that more complex and flexible models for single particle dissolution could describe the dissolution more adequately. The fact that the  $\overline{ss}$  values for the cube root and the square root model are almost the same suggests a model with properties between these two. The Danckwerts model as discussed by Goyan (105) is given by:

$$-dw/dt = A((Dp)^{\frac{1}{2}} + D/a)C_a \quad (7.12)$$

- where
- w = weight undissolved
  - A = the surface area
  - D = the diffusion coefficient
  - p = a quantity related to stirring
  - a = the radius of the particle
  - C<sub>a</sub> = the steady state concentration.

This model is very flexible. When  $(Dp)^{\frac{1}{2}}$  predominates, the apparent model would be the cube root model. As the quantity D/a becomes more important, then the square root model will become the apparent model. Finally, as the quantity D/a predominates the "squared cube root model" will become the apparent model. However, the Danckwerts model, when applied to the log-normal case, results in a mathematical expression which is much more complex than Eqs.

7.3 and 7.4.

CHAPTER 8

The fit of the dissolution data to the cube root model is excellent. Therefore, if an application of the Danckwerts model results in an even better fit, this improvement will likely be statistically insignificant considering the magnitude of the experimental errors. In such a case the Hixson-Crowell model should be preferred because of its simplicity.

of obtaining estimates of the parameters in a particular model. If the mathematical model is nonlinear in its parameters and cannot be transformed to a linear form<sup>1</sup> it is necessary to use a computer to cope with the complexity of computations. Several computer programs are available for the treatment of nonlinear regression (123-132). Considerable time and effort is often spent in applying the more powerful and versatile of these programs and their complexity frequently inhibits the less experienced. Even for the experienced user their inflexible and strict input structure may often lead to errors that can be difficult to find because of the lack of specific error messages. The time spent on correcting such errors is of considerable inconvenience, particularly when the program used is not written for interactive time-sharing.

This chapter describes a powerful and versatile FORTRAN computer program, FUNFIT, for nonlinear regression and curve fitting that does not have the disadvantages described above. It is written for the maximum convenience of the user and utilizes the many advantages of interactive

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1. The expression  $y = ae^{bx}$  is linear in the parameter  $a$  and nonlinear in  $b$ . By transforming to  $\ln y = \ln a + bx$  it is linear in  $(\ln a)$  and  $b$ . The expression  $y = ae^{bx} + ce^{dx}$  is nonlinear in  $b$  and  $d$  and cannot be transformed to a linear form by standard means.

CHAPTER 8

- FUNFIT - A TIME SHARING PROGRAM FOR GENERAL  
NONLINEAR REGRESSION AND CURVEFITTING

Many investigators must deal with the problems of evaluating how well one or more mathematical models describe a certain physical system or of obtaining estimates of the parameters in a particular model. If the mathematical model is nonlinear in its parameters and cannot be transformed to a linear form<sup>1</sup> it is necessary to use a computer to cope with the complexity of computations. Several computer programs are available for the treatment of nonlinear regression (123-132). Considerable time and effort is often spent in applying the more powerful and versatile of these programs and their complexity frequently inhibits the less experienced. Even for the experienced user their inflexible and strict input structure may often lead to errors that can be difficult to find because of the lack of specific error messages. The time spent on correcting such errors is of considerable inconvenience, particularly when the program used is not written for interactive time-sharing.

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$$S = \sum_{i=1}^n y_i^2 = \sum_{i=1}^n [y_i - f(\theta, x_i)]^2 \quad (8.2)$$

time-sharing. The input occurs essentially as a communication with the computer. Questions are asked about which particular data treatment is desired and instructions are given how to enter data. Every input is extensively checked for numerical, logical and typing errors, so these can be corrected immediately. A special command 'BACK' makes it easy to edit previous inputs so multiple runs under various conditions can be made quickly. The program offers an extensive analysis of residuals that most other programs neglect, and a lattice search to obtain suitable initial parameter estimates. A lattice search combined with contour maps makes it possible to investigate whether a better solution of the nonlinear regression problem may exist.

The above example represents a simple case with only 1 independent variable. The function to be fitted could, in fact, have 2 independent variables.

### THEORY

The Central Limit Theorem of probability theory justifies the assumption that random errors in a set of observations are normally distributed. If the mathematical model is correct, the independent variable exact, and the errors are independent and normally distributed with zero mean and the same variance, then the method of least squares is the best choice for the estimation of the parameters because the estimates obtained will be maximum likelihood estimates.

For example the equation to be fitted to some observations might be:

$$y = p_1 e^{p_2 x} + p_3 e^{p_4 x} = f(\underline{p}, x) \quad (8.1)$$

where  $\underline{p} = [p_1, p_2, p_3, p_4]^T$  is the parameter vector and  $x$  the independent variable. The problem is then to minimize the sum of the squared residuals, that is:

$$SS = \sum_{i=1}^{NOBS} e_i^2 = \sum_{i=1}^{NOBS} [y_i - f(\underline{p}, x_i)]^2 \quad (8.2)$$

where  $[(y_i, x_i), i = 1, 2, \dots, \text{NOBS}]$  are the observations. There may also be reasons to weight the observations in a certain way. The weighted residual sum of squares is then defined as<sup>2</sup>

$$SS = \sum_{i=1}^{\text{NOBS}} e_{w,i}^2 = \sum_{i=1}^{\text{NOBS}} w_i [y_i - f(\underline{P}, x_i)]^2 \quad (8.3)$$

where  $w_i$  is the weight of the  $i$ -th observation. Unweighted data are in fact data having the same weight i.e.,  $w_i = 1$ . The relationship between weighted and unweighted residuals is

$$e_{w,i} = \sqrt{w_i} e_i \quad (8.4)$$

The above example represents a simple case with only 1 independent variable,  $x$ . The function to be fitted could, in fact, have 2 independent variables i.e.,  $\underline{x} = [x_1, x_2]^T$ . The observations are then  $[(y_i, x_{1,i}, x_{2,i}), i = 1, 2, \dots, \text{NOBS}]$  (e.g. concentration, time temperature) and are represented in 3 dimensions. The problem is then no longer a least squares curve-fitting but a least squares surface-fitting. 'FUNFIT' can fit 'hypersurfaces' with up to 9 independent variables.

SIMULANTEOUS FITTING OF SEVERAL RESPONSE SYSTEMS

The system under investigation can sometimes be measured for more than one response or dependent variable,  $y$ . For example blood levels and urinary excretion of a drug could be measured in the same pharmacokinetic experiment or, in an experiment in chemical reaction kinetics more than one reaction product could be followed. In general, if there is a functional

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2. In FUNFIT the weights are normalized i.e. scaled so that their sum is equal to the number of observations, NOBS. This is done to achieve a better comparison of weighted and unweighted sum-of-squares values.

expression for each kind of measured response and these expressions contain one or more *common* parameters it is likely that a simultaneous least squares fit of all the functions would provide more reliable parameter estimates than would be obtained by fitting each function individually. If, for example, 2 different kinds of response are measured in a system, the simultaneous least squares fitting problem is to minimize 'the sum of the sum of squared residuals' given by:

$$SS = \sum_{i=1}^{NOBS_1} w_{1,i} [y_{1,i} - f_1(\underline{P}_1, X_{1,i})]^2 + \sum_{i=1}^{NOBS_2} w_{2,i} [y_{2,i} - f_2(\underline{P}_2, X_{2,i})]^2 \quad (8.5)$$

where the parameter vectors  $\underline{P}_1$  and  $\underline{P}_2$  contain one or more common parameters. The symbols are used in analogy with equation 8.3 and subscripts 1 and 2 denote response systems 1 and 2. Hence, functions  $f_1$  and  $f_2$  are to be fitted simultaneously to  $NOBS_1$  observations from response system 1 and  $NOBS_2$  observations from response system 2 respectively. It is possible using 'FUNFIT' to fit simultaneously up to 10 functions (response systems) each containing up to 20 parameters and 9 independent variables, with or without weighting of the single observations in each response system. The sum-of-squares function can therefore be summarized by a general extension of

Eq. 8.5:

$$SS = \sum_{j=1}^{NFUNC} \sum_{i=1}^{NOBS_j} w_{j,i} [y_{j,i} - f_j(\underline{P}_j, X_{j,i})]^2 \quad (8.6)$$

The total number of observations is

$$NOBS = \sum_{j=1}^{NFUNC} NOBS_j \quad (8.7)$$

where  $NOBS_j$  is the number of observations for the  $j$ -th function.

WEIGHTING OF RESPONSE SYSTEMS

The problem of weighting is of particular importance in simultaneous fitting for two reasons: 1. There may be a large difference in the orders of magnitude of the values of the dependent variables in each response system. 2. The variances of the errors may differ considerably between the systems.

The effect of condition 1. can be reduced by the proper choice of units for the dependent variables. Equation 8.6 assumes that the residual variance between each individual response system is the same. However this is seldom the case (i.e. condition 2. applies) and each response system must be given its own weight. It is generally accepted that, in a statistical sense, the best weighting scheme is to make the weight of each observation inversely proportional to the variance of the error (as estimated by the residual variance). Therefore the weight of each response system should be made proportional to the reciprocal of its residual variance which is obtained when the system is fitted individually by least squares (i.e. not simultaneously with other systems). Therefore the weight for the  $j$ -th response system should be:

$$W_j = \frac{NOBS_j - NPAR_j}{\left[ \sum_{i=1}^{NOBS_j} w_{j,i} [y_{n,i} - f_j(p_j, X_{j,i})]^2 \right]_{\min}} \quad (8.8)$$

where  $NPAR_j$  is the number of parameters in the function  $f_j$  and  $(NOBS_j - NPAR_j)$  is the residual degrees of freedom. Equation 8.6 can therefore be written in an improved form:

It should be noted that for a given set of observations the residual sum of squares,  $SS$ , depends only on the parameters because in Eq's



$$SS = \sum_{j=1}^{NFUNC} W_j \sum_{i=1}^{NOBS_j} w_{j,i} [y_{j,i} - f_j(\underline{P}_j, \underline{X}_{j,i})]^2 \quad (8.9)$$

where  $W_j$  signifies the weights of the  $j$ -th response system and  $w_{j,i}$  the weights of the observations within that system. For example, if it is desired in a pharmacokinetic experiment to fit blood and urine data simultaneously, an individual fitting of each of the two systems should be done first. The weights to be used for each system in the simultaneous fitting are calculated as the residual degrees of freedom divided by the (weighted or unweighted) residual sum-of-squares obtained from the individual fittings.

Fitting implicit functions

It is necessary to apply a special approach if a function to be fitted is of implicit form; i.e., if the dependent variable,  $y$ , cannot be expressed explicitly as a function of the independent variable(s),  $\underline{X}$  and the parameters,  $\underline{P}$ .

The function may be described by:

$$g(y, \underline{X}, \underline{P}) = 0 \quad (8.10)$$

(e.g.  $y^2 + X^{p1} + (X + y)^{p2} = 0$ ) from which  $y$  can be found by an iterative procedure only when numerical values of  $\underline{X}$  and  $\underline{P}$  are given. The calculated values of  $y$  are found by including in the user-supplied subroutine 'MODEL' a suitable algorithm for finding the root of  $g$ . This algorithm will serve the purpose of solving Eq. 8.10 for  $y$  so the subroutine indirectly defines the functional relationship between  $y$  and  $\underline{X}, \underline{P}$ .

MINIMIZATION

It should be noted that for a given set of observations the residual sum of squares,  $SS$ , depends only on the parameters because in Eq's

8.2, 8.6 and 8.9, the quantities  $y$ ,  $\underline{X}$ ,  $w$ ,  $W$ ,  $NFUNC$  and  $NOBS$  are all numbers that are given. These equations can therefore be written in a shorter form as

$$SS = SS(\underline{P}) \quad (8.11)$$

indicating that  $SS$  is a function of the parameters only. The least squares fitting and parameter estimation is then simply reduced to a function minimization problem, namely to find the particular values of the elements of the parameter vector,  $\underline{P}$ , that minimize  $SS$ .

If the sum-of-squares function is strictly convex in a specified convex parameter space then it will only have one minimum (133). This condition is guaranteed when fitting linear but not nonlinear functions for which there may exist more than one minimum. There is, in general, in the latter case no guarantee that the minimum found by any nonlinear regression program is the smallest, giving the best possible fit and parameter estimates. This problem can be illustrated in the case of two parameters where Eq. 8.11 describes a surface in 3 dimensions: Computer programs find the minimum by some iterative procedure and therefore require initial estimates of the parameters  $p_1$  and  $p_2$  together with their initial step sizes  $\Delta p_1$  and  $\Delta p_2$ .<sup>3</sup> For example if the  $SS$  surface has two troughs and the initial starting point  $(p_1, p_2)$  lies on the slope of one of these and the same is the case for the points  $(p_1 + \Delta p_1, p_2)$ ,  $(p_1, p_2 + \Delta p_2)$  or  $(p_1 + \Delta p_1, p_2 + \Delta p_2)$  then the strategy of the minimization algorithms employed is to 'run downhill' so that the parameter estimates in the next iteration step will usually be closer to the bottom of that trough while the other trough which may contain a smaller minimum, is overlooked. Mathematically this problem exists also for sum-of-

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3. If the initial step sizes are not included in the input they are chosen in relation to the upper and lower limits given for the parameters in  $NFUNC$ . For example a lattice

squares surfaces in more than 3 dimensions (hyper surfaces) but is then impossible to visualize.

The importance of good (i.e. close) initial parameter estimates is obvious. The choice of small initial parameter step sizes will, as seen above, result in an increased probability of overlooking a smaller minimum. Large initial step sizes require more iterations and computation time but reduce this probability because a larger part of the parameter space is evaluated.

Most minimization algorithms in nonlinear regression programs are based on the gradient method of Gauss-Newton (134). These are particularly efficient in finding a minimum for a sum-of-squares function but are more likely to overlook a better minimum than non-gradient methods that usually take longer to converge. The simplex method of Nelder and Mead (135) used in 'FUNFIT' appears to be a particularly suitable non-gradient method. It has statistical application, is rapidly convergent and, on the way to the minimum, covers a large section of the parameter space, reducing the probability of missing a global (smallest) minimum. It has proven to be a very robust method that will always find a minimum and further has the advantage over gradient methods that nonlinear parameter constraints can more easily be employed. The economic disadvantage of the somewhat slower minimization procedure is insignificant in most cases.

#### LATTICE SEARCH

The most reliable way to search for a minimum smaller than that found by some minimization procedure is to evaluate  $SS(P)$  at each point in a grid or lattice that spans over the whole parameter space. The computation time for such a procedure increases exponentially and very rapidly with the number of parameters and divisions per parameter. For example a lattice

consisting of just 6 parameters with 10 divisions requires  $10^6$  SS-evaluations each of which requires NOBS function evaluations. If SS ( $\underline{P}$ ) is a surface in 3 dimensions (2 parameter case) its approximate shape can be visualized from a contour plot constructed from the SS-values of the points in a grid of the two parameters. If there are more than two parameters a composite picture of the sum-of-squares surface can be built up by "slicing" the parameter space, that is by fixing all except two parameters at a time. Contour plots help greatly in finding a global minimum and considerably reduce the number of function evaluations that otherwise would be required in such a search.

the parameters is related to the 'variability' of the SS-surface in the region of the minimum because a flat surface allows the parameters to have a wide range of values resulting in only a small variation

CONVERGENCE CRITERION

The accuracy with which the exact position of the minimum of SS ( $\underline{P}$ ) can be determined is always limited. After a certain number of iteration steps the SS-values of the points used in calculating the position of the next  $\underline{P}$  estimate differ so little from each other that further iterations will introduce round-off errors. It is because of this and for reasons of economy, necessary to specify a stopping criterion for the minimization process. A necessary (but not sufficient) condition for  $\underline{P}$  to be close to a minimum is that the SS-surface in the proximity of  $\underline{P}$  is "sufficiently flat". The minimization process is often considered to have converged when the SS ( $\underline{P}$ ) value does not change more than a certain amount from one iteration step to the next. This guarantees a certain degree of 'flatness' as long as there is a significant change in the parameter vector  $\underline{P}$  between the iterations. However such a stopping criterion evaluates the 'flatness' in a somewhat imprecise way and since SS ( $\underline{P}$ ) is not usually dimensionless, depends on the units used for the experimental data. The stopping criterion

used in 'FUNFIT' checks the 'flatness' in terms of the coefficient of variation (percent) of the SS-values of some points closely surrounding  $\underline{P}$ .

For many applications it is important to get statistical estimates of the variability of the parameters obtained. As indicated above, this flatness is in this way defined more strictly and does not depend on the units used. Results can therefore be compared on a more consistent basis information is found in the shape of the SS-surface near the minimum. The complexity of the SS( $\underline{P}$ ) expression in the nonlinear case does not allow than in the former case.

The stopping criterion determines how 'accurately' the parameters are determined. Its value should be chosen in relation to the variability of the parameters. There is no reason to choose a very small stopping criterion to get very 'accurate' parameters if their values are statistically rather uncertain. The SS ( $\underline{P}$ ) surface is approximated in the region of the final parameter estimates,  $\underline{P}$ , by a quadratic surface from a Taylor series of the SS-surface in the region of the minimum because a flat surface allows expansion where third- and higher order terms are neglected; i.e., the parameters to have a wide range of values resulting in only a small variation of the SS ( $\underline{P}$ ) value. The stopping criterion used in 'FUNFIT' therefore has an advantage over other commonly used criteria because it is based on the 'variability' of the surface and hence determines the 'accuracy' of the parameters in relation to their variability. To avoid termination on a flat plateau of the SS-surface or near a 'saddle point' it must further be ensured that the surface around the point found at convergence is convex. In gradient methods this is often done by checking whether the matrix of the second partial derivatives (the Hessian matrix) is positive definite as is the case if all its eigenvalues are positive (133). In 'FUNFIT' the minimization process is allowed to continue a certain number of iteration steps after the 'flatness criterion' has been satisfied and the process is considered to have converged if the new parameter values and SS-values are sufficiently close to the previous values.

It can be seen from this equation that in order to decrease the variances of the parameters, the number of observations, NOBS, should be relatively

STATISTICAL ESTIMATION

For many applications it is important to get statistical estimates of the variability of the parameters obtained. As indicated above, this information is found in the shape of the SS-surface near the minimum. The complexity of the SS(P) expression in the nonlinear case does not allow exact statistical estimates to be obtained. These are obtained instead by approximating the surface in the region of the minimum with a more simple surface for which exact, i.e. unbiased, statistical estimates can be calculated. The SS (P) surface is approximated in the region of the final parameter estimates,  $\hat{\underline{P}}$ , by a quadratic surface from a Taylor series expansion where third- and higher order terms are neglected; i.e.,

$$SS(\underline{P}) \approx SS(\hat{\underline{P}}) + \nabla^T SS(\hat{\underline{P}}) (\underline{P} - \hat{\underline{P}}) + \frac{1}{2} (\underline{P} - \hat{\underline{P}})^T \nabla^2 SS(\hat{\underline{P}}) (\underline{P} - \hat{\underline{P}}) \quad (8.12)$$

where  $\nabla^T = (\partial/\partial p_1 + \partial/\partial p_2 + \dots + \partial/\partial p_n)$  and n is the number of parameters. The statistical information is found in the term  $\nabla^2 SS(\hat{\underline{P}})$ , that is the square matrix of the second partial derivatives of SS(P) evaluated at  $\hat{\underline{P}}$ , the so-called Hessian matrix:

$$H(\hat{\underline{P}}) = \nabla^2 SS(\hat{\underline{P}}) = \begin{bmatrix} \frac{\partial^2 SS(\hat{\underline{P}})}{\partial p_1^2} & \dots & \frac{\partial^2 SS(\hat{\underline{P}})}{\partial p_1 \partial p_n} \\ \dots & \dots & \dots \\ \frac{\partial^2 SS(\hat{\underline{P}})}{\partial p_1 \partial p_n} & \dots & \frac{\partial^2 SS(\hat{\underline{P}})}{\partial p_n^2} \end{bmatrix} \quad (8.13)$$

The sample variance-covariance matrix of the parameters, V, is the inverse of the Hessian matrix multiplied by the residual variance estimate:

$$V = \frac{SS(\hat{\underline{P}})}{NOBS-n} H^{-1}(\hat{\underline{P}}) \quad (8.14)$$

It can be seen from this equation that in order to decrease the variances of the parameters, the number of observations, NOBS, should be relatively

large compared to the number of parameters,  $n$ . It is recommended that NOBS should not be less than about  $3n$  in statistical estimations. The covariance between the  $i$ -th and the  $j$ -th parameter is the  $(i,j)$ -th element of the variance-covariance matrix:

$$\text{COV}(\hat{p}_i, \hat{p}_j) = v_{i,j} \quad (8.15)$$

and the variance of the  $i$ -th parameter is the  $i$ -th diagonal element:

$$\text{var}(\hat{p}_i) = \text{COV}(\hat{p}_i, \hat{p}_i) = v_{i,i} \quad (8.16)$$

The elements of the correlation matrix,  $\rho_{i,j}$ , are calculated from the variance-covariance matrix by:

$$\rho_{i,j} = \frac{\text{COV}(\hat{p}_i, \hat{p}_j)}{\sqrt{\text{Var}(\hat{p}_i) \text{Var}(\hat{p}_j)}} \quad (8.17)$$

The joint probability distribution of the estimated parameters is multivariate normal and is given by (136).

$$g(\underline{P}) = \frac{1}{(2\pi)^{n/2} \sqrt{|V|}} \exp \left[ - \frac{(\underline{P} - \hat{\underline{P}})^T V^{-1} (\underline{P} - \hat{\underline{P}})}{2} \right] \quad (8.18)$$

### ASSUMPTIONS MADE IN THE STATISTICAL EVALUATIONS

The reliability of the statistical estimates above depends on the following assumptions:

1. The independent variable(s) is without error.
2. The errors are independent, have zero mean and the same variance,

$\sigma^2$ , i.e.:

$$\text{COV}(\epsilon_i, \epsilon_j) = 0 \quad (8.19)$$

$i \neq j$

$$E(\epsilon_i) = 0 \quad (8.20)$$

$$\text{Var}(\epsilon_i) = \sigma^2 \quad (8.21)$$

For the F- test, t- test and confidence limits to be valid the errors must furthermore be normally distributed with zero mean and the same variance:

$$\epsilon_i \sim N(0, \sigma^2) \tag{8.22}$$

3. The mathematical model is correct.

4. Equation 8.12 is exact.

When conditions 1., 2. and 3. are satisfied the least squares estimate of  $\underline{P}$  is also the maximum likelihood estimate of  $\underline{P}$  because the likelihood function,  $L$ , can be written (137):

$$L(\underline{P}, \sigma^2) = (2\pi\sigma^2)^{-\frac{n}{2}} \exp[-SS(\underline{P})/2\sigma^2] \tag{8.23}$$

which is maximized when  $SS(\underline{P})$  is minimized ( $\sigma^2 = \text{constant}$ ). Therefore the least squares parameter estimates are unbiased under these 3 conditions.

Assumption 4. is only true when linear but not nonlinear functions are fitted. The most crucial point in nonlinear regression appears to be how good an approximation Eq. 8.12 is, because the statistical estimates (variances, covariances) are based on formulae and analysis from linear regression theory. The estimates are therefore biased and often called 'asymptotic estimates' because of the asymptotic property of the Taylor series expansion (Eq. 8.12). It has been found in simulation studies that the standard deviations of parameters in nonlinear regression can be two- to three-fold different from their true values (138). Parameter estimates are usually less biased because they are less sensitive to violation of the above conditions.

WEIGHTING

Where assumption 2. is not true because the error variance,  $\sigma^2$ , is not constant, it is recommended that weights inversely proportional to



the individual variances be used, according to the following weighting scheme:

$$w_i = \frac{1}{\sigma_i^2}, \quad i = 1, 2, \dots, \text{NOBS} \quad (8.24)$$

Thus if,  $\epsilon_i \sim N(0, \sigma_i^2)$

$$\epsilon_i \sim N(0, \sigma_i^2) \quad (8.25)$$

then the weighted errors will fulfil condition 2. because

$$\epsilon_{w,i} = w \epsilon_i = \frac{\epsilon_i}{\sigma_i} \sim N(0, 1) \quad (8.26)$$

so that this weighting will provide improved estimates. The error variances,  $\sigma_i^2$ , can only be estimated by repeated experiments. However in some weighting situations a certain functional relationship is assumed between the error and the dependent or independent variable. For example in pharmacokinetics it is often assumed that the standard deviation of the errors is proportional to the plasma concentration, so the data are assigned weights inversely proportional to the square of the concentration. It is not valid statistically to use weights just to get a better fit. There must be a sound basis for the weighting scheme used.

CONFIDENCE REGIONS AND CONFIDENCE LIMITS

When assumptions 1. to 4. are true it can be shown that  $SS(\hat{P})$  is distributed as chi-square with (NOBS-n) degrees of freedom:

$$SS(\hat{P}) \sim \sigma^2 \chi_{\text{NOBS-n}}^2 \quad (8.27)$$

and  $SS(P) - SS(\hat{P}) \sim \sigma^2 \chi_n^2 \quad (8.28)$

Both  $SS(P) - SS(\hat{P})$  and  $SS(\hat{P})$  follow a  $\chi^2$  distribution and are independently distributed. The ratio

$$\frac{[SS(\underline{P}) - SS(\hat{\underline{P}})]/n}{SS(\hat{\underline{P}})\lambda\text{NOBS}-n} \sim F(n, \text{NOBS}-n) \quad (8.29)$$

is therefore distributed as an F-distribution with n and (NOBS-n) degrees of freedom respectively (136). The term SS(P) can be isolated from this equation so that the critical points of the F-distribution provide the exact (1-α) 100% confidence region of the parameters:

$$SS(\underline{P}) = SS(\hat{\underline{P}}) \left[ 1 + \frac{n}{\text{NOBS}-n} F_{\alpha, n, \text{NOBS}-n} \right] \quad (8.30)$$

The set of parameter values, P, for which this equation is satisfied forms a closed contour line (two parameter case) or a contour surface inside which the probability of *simultaneously* finding the true (population) parameter values is (1-α) 100%. In the nonlinear case where assumption 4. is violated, the confidence contours are still exactly defined by Eq. 8.30 but the (1-α) 100% confidence level is only approximated.

When linear equations are fitted the confidence region is ellipsoidal in shape and given by:

$$(\underline{P}-\hat{\underline{P}})^T H^{-1} (\underline{P}-\hat{\underline{P}}) \leq \frac{n}{\text{NOBS}-n} SS(\hat{\underline{P}}) F_{\alpha, n, \text{NOBS}-n} \quad (8.31)$$

Because the parameters are correlated it is not possible to define confidence limits for each individual parameter. Limits are however often defined in terms of tangent points of 'support planes' i.e. planes parallel to the parameter coordinate axis and tangent to the ellipsoidal region defined by Eq. 8.31. The approximate (1-α) 100% 'support plane' confidence limits of the parameters are given by (123)

$$\hat{p}_i \pm SD_i \sqrt{F_{\alpha, n, \text{NOBS}-n} \times n} \quad (8.32)$$

where  $SD_i$  is the standard deviation of the i-th parameter.

Assuming no correlation among the parameters the 'uni-plane' confidence limits are given by (123)

$$\hat{p}_i \pm t_{\alpha, \text{NOBS-n}} \times \text{SD}_i \quad (8.33)$$

where  $t_{\alpha, \text{NOBS-n}}$  is the t-statistic with probability level  $\alpha$  and (NOBS-n) degrees of freedom. Confidence limits are sometimes desired for the ratio of two parameters, for example  $p_1/p_2$ . This can be calculated using the following expression (139):

$$\frac{\hat{p}_1 \hat{p}_2 - t_{\alpha, \text{NOBS-n}}^2 \text{COV}(\hat{p}_1, \hat{p}_2) \pm t_{\alpha, \text{NOBS-n}} A}{\hat{p}_2 - t_{\alpha, \text{NOBS-n}}^2 \text{Var}(\hat{p}_2)} \quad (8.34)$$

where

$$A = \hat{p}_1^2 \text{Var}(\hat{p}_2) + \hat{p}_2^2 \text{Var}(\hat{p}_1) - 2\hat{p}_1 \hat{p}_2 \text{COV}(\hat{p}_1, \hat{p}_2) \quad (8.35)$$

$$- t_{\alpha, \text{NOBS-n}}^2 \left[ \text{Var}(\hat{p}_1) - \frac{\text{COV}(\hat{p}_1, \hat{p}_2)^2}{\text{Var}(\hat{p}_2)} \right] \quad (8.39)$$

STANDARD DEVIATION OF A FUNCTION OF THE PARAMETERS

Use of variance-covariance matrix

The variance-covariance matrix Eq. 8.14 that is printed out in most computer programs allows the standard deviation to be calculated for any quantity that is expressed as a function of one or more of the parameters. Let such a quantity be denoted

$$g = g(\hat{p}_1, \hat{p}_2, \dots, \hat{p}_k) \quad (8.36)$$

then its standard deviation is given by the following formula which is based on a Taylor series expansion (140)

$$(SD_g)^2 \approx \sum_{i,j=1}^k \frac{\partial g}{\partial p_i} \frac{\partial g}{\partial p_j} \text{COV}(\hat{p}_i, \hat{p}_j) \quad (8.37)$$

Equation 8.37 often leads to rather complex expressions if  $g$  is a nonlinear function of more than a few parameters. However, this problem can be avoided in most cases and the standard deviation of  $g$  obtained directly without the truncation errors that may be introduced when Eq. 8.37 is used. This can be done if, as is usually the case, Eq. 8.36 can be solved explicitly or numerically for one of its parameters, so that this parameter,  $p_i$ , can be expressed as a function of  $g$  and the remaining parameters, i.e.

$$SD_g \approx \left| \frac{dg}{dp_i} \right| SD_i \quad (8.38)$$

In particular if  $g$  is linear and of the form  $a\hat{p} + b$  ( $a$  and  $b$  are constants) then  $SD_g = aSD_p$  exactly. 2. If  $g$  is a linear function of the form:

$$g = \sum_{i=1}^k a_i \hat{p}_i \quad (8.39)$$

then Eq. 8.37 becomes

$$(SD_g)^2 = \sum_{i=1}^k a_i^2 \text{Var}(\hat{p}_i) + \sum_{i \neq j} a_i a_j \text{COV}(\hat{p}_i, \hat{p}_j) \quad (8.40)$$

3. If  $g$  is a ration of two parameters, for example  $g = \hat{p}_1 / \hat{p}_2$ , Eq. 8.37 becomes

$$(SD_g)^2 \approx \left( \frac{\partial g}{\partial p_1} \right)^2 \text{Var}(\hat{p}_1) + \left( \frac{\partial g}{\partial p_2} \right)^2 \text{Var}(\hat{p}_2) + 2 \frac{\partial g}{\partial p_1} \frac{\partial g}{\partial p_2} \text{COV}(\hat{p}_1, \hat{p}_2) \quad (8.41)$$

which in this case gives

$$(SD_g)^2 \approx \frac{\text{Var}(\hat{p}_1)}{\hat{p}_2^2} + \frac{\hat{p}_1^2 \text{Var}(\hat{p}_2)}{\hat{p}_2^4} - \frac{2\hat{p}_1 \text{COV}(\hat{p}_1, \hat{p}_2)}{\hat{p}_2^3} \quad (8.42)$$

Use of the transformation technique

Equation 8.37 often leads to rather complex expressions if  $g$  is a nonlinear function of more than a few parameters. However, this problem can be avoided in most cases and the standard deviation of  $g$  obtained directly without the truncation errors that may be introduced when Eq. 8.37 is used. This can be done if, as is usually the case, Eq. 8.36 can be solved explicitly or numerically for one of its parameters, so that this parameter,  $\hat{p}_i$ , can be expressed as a function of  $g$  and the remaining parameters, i.e.

$$\hat{p}_i = h(g, \hat{p}_1, \hat{p}_2 \dots, \hat{p}_{k-1}) \tag{8.43}$$

The function to be fitted to the data points can then be transformed so that it contains  $g$  as a parameter essentially replacing  $p_i$ . To evaluate the function,  $p_i$  is then simply calculated using Eq. 8.43 (where  $g$  and the  $(k-1)$  parameters are among the  $k$  input parameters). Initial estimates must be obtained for  $g$  since it acts as an input parameter. This is done by calculating it from Eq. 8.36 using the initial estimates of the  $k$  parameters or by using the final least squares estimates obtained from curve-fitting based on the original untransformed function.

This transformation technique is of considerable importance and highly recommended. For example, it allows the standard deviation to be calculated for essentially any pharmacokinetic quantity that is expressed in terms of the macro- or microparameters in the model.

EXAMINATION OF RESIDUALS

It is of fundamental importance to analyse the pattern of the residuals because this is essentially the only way of examining whether the

basic assumptions behind the nonlinear estimation are violated. Most general nonlinear programs seem to ignore this. Perhaps the best way to examine the residuals is to plot them against the independent and dependent variables (141). Significant systematic deviations can be visualized in this way. The assessment is somewhat complicated by the fact that there will always be a correlation between the residuals because NOBS residuals are only associated with (NOBS-n) degrees of freedom. 'FUNFIT' includes the Durbin-Watson statistic, given by (142):

$$d = \frac{\sum_{i=2}^{\text{NOBS}} (e_i - e_{i-1})^2}{\sum_{i=1}^{\text{NOBS}} e_i^2} \quad (8.44)$$

to test for excessive serial correlation (systematic deviation) among the residuals. The statistic is compared with tabulated critical points at a given significance level.

The fundamental assumption of random errors is also tested using two nonparametric tests that will be called the 'group' statistic and the 'number' statistic. The 'group' statistic is based on an analysis of groups (runs) of residuals of equal sign. For example the sequence of residuals (+++) (--) (+) (--) (+) forms  $r = 5$  groups. The least number of residuals with the same sign is  $L = 4$ . If NOBS residuals, with equal probability of being negative and positive, form a sequence with  $r$  groups, then the probability of getting  $\leq r$  groups is (143)

$$P(\leq r) = \binom{\text{NOBS}}{L}^{-1} \sum_{i=2}^r f_r \quad (8.45)$$

where  $f_r = 2 \binom{L-1}{\frac{i}{2}-1} \binom{\text{NOBS}-L-1}{\frac{i}{2}-1}$  when  $i$  is even (8.46)

and 
$$f_r = \binom{L-1}{\frac{i-1}{2}} \binom{NOBS-L-1}{\frac{i-3}{2}} + \binom{L-1}{\frac{i-3}{2}} \binom{NOBS-L-1}{\frac{i-1}{2}} \quad (8.47)$$

when  $i$  is odd. The 'number' statistic is defined as the probability of getting  $L$  or less residuals of same sign and is given by

$$P(\leq L) = 2^{-NOBS} \sum_{i=1}^{L+1} \binom{NOBS}{i-1} \quad (8.48)$$

where the large brackets denote binomial coefficients i.e.  $\binom{x}{y} = x! / (y!(x-y)!)$ .

If  $P(\leq r) < 0.05$  or  $P(\leq L) < 0.05$  then the hypothesis that the residuals are random should be rejected (with  $\alpha$ -error  $< 0.05$ ). This can be used to reject the basic assumption of normal errors provided the model is correct. However, it can also mean that the mathematical model is wrong if it is assumed that the errors are in fact random with zero mean.

OUTLIERS

Several repeated experiments will be required to examine whether the errors are all normally distributed. When only one experiment is available a test must rely on the fact that if all the errors are deviates from the same normal distribution, they will then collectively be normally distributed. If the mathematical model is correct and it is assumed that  $E(\epsilon_i) = 0$  and  $Var(\epsilon_i) = \sigma^2$  then the residuals are unbiased estimates of the errors, i.e.  $E(e_i) = \epsilon_i$  and the residual variance given by

$$s^2 = \frac{\sum_{i=1}^{NOBS} (e_i - \bar{e})^2}{NOBS-n} \approx \frac{\sum_{i=1}^{NOBS} e_i^2}{NOBS-n} = \frac{SS(\hat{P})}{NOBS-n} \quad (8.49)$$

estimates  $\sigma^2$ . Further, if it is assumed that  $\epsilon_i \sim N(0, \sigma^2)$  it follows that  $\epsilon_i / \sigma \sim N(0, 1)$ . Now, if the 4 basic assumptions mentioned previously are true then  $e_i / s$ , the unit normal deviate form of the residuals, estimates  $\epsilon_i / \sigma$  and is  $N(0, 1)$ .

The normal deviate form of the residuals can be used to test for outliers, that is, data-points which, in a statistical sense, are not typical of the rest of the data. The  $i$ -th observation is an outlier if the normal deviate  $e_i/s$  falls in the critical region of the  $N(0,1)$  distribution, that is if  $|e_i/s| > 1.96$  ( $\alpha < 0.05$ ). If  $\text{NOBS} - n$  is smaller than 30 the normal deviate should be compared with the critical values of the  $t$ -distribution with  $(\text{NOBS} - n)$  degrees of freedom.

Outliers should be submitted to particularly careful examination since they may provide information of vital interest. They should only be rejected if they are caused by errors in the recording or the experimental technique (144). Outliers may also occur if the mathematical model is incorrect.

If there are many observations it may be useful to construct a half-normal plot of the residuals. The linearity of such a plot gives information about outliers and how normally the residuals are distributed (141). The above analysis extends also to weighted residuals.

COMPARISON OF PARAMETERS AND MODELS

It can be of interest to investigate whether there is a significant difference in a certain parameter,  $\hat{p}_i$ , between two experiments (1 and 2).

If it is assumed the parameter is normally distributed a  $t$ -test can then be applied.

$$t = \frac{\hat{p}_{i,1} - \hat{p}_{i,2}}{\sqrt{\text{Var}(\hat{p}_{i,1}) + \text{Var}(\hat{p}_{i,2})}} \quad (8.50)$$

If  $t$  falls in the critical region  $|t| > t_{\alpha/2}$  of a  $t$ -distribution with  $(\text{NOBS}_1 + \text{NOBS}_2 - 2n)$  degrees of freedom, the null hypothesis that  $\hat{p}_{i,1}$  and  $\hat{p}_{i,2}$  are the same should be rejected at significance level  $\alpha$ .



SCHEME 1 USER - COMPUTER INTERACTION DURING EXECUTION

For any particular system there may be more than one mathematical model which could be appropriate. It is generally accepted that if two models give approximately "equal" fits, the simpler model should be chosen unless the more complex model can be justified on other criteria. At present there does not seem to be any simple, rigorously based statistical test which makes it possible to distinguish between alternative models. The following test, based on a comparison of residual variances, is no exception. It can however be used as a guide in the absence of other methods.

Let  $SS(\hat{P}_{(1)}) / (NOBS_1 - n_1) > SS(\hat{P}_{(2)}) / (NOBS_2 - n_2)$  be the residual variances of models 1 and 2 respectively. If it is assumed  $SS(\hat{P}_{(1)})$  and  $SS(\hat{P}_{(2)})$  are independent random variables having chi-square distributions with  $(NOBS_1 - n_1)$  and  $(NOBS_2 - n_2)$  degrees of freedom, the variance ratio given by:

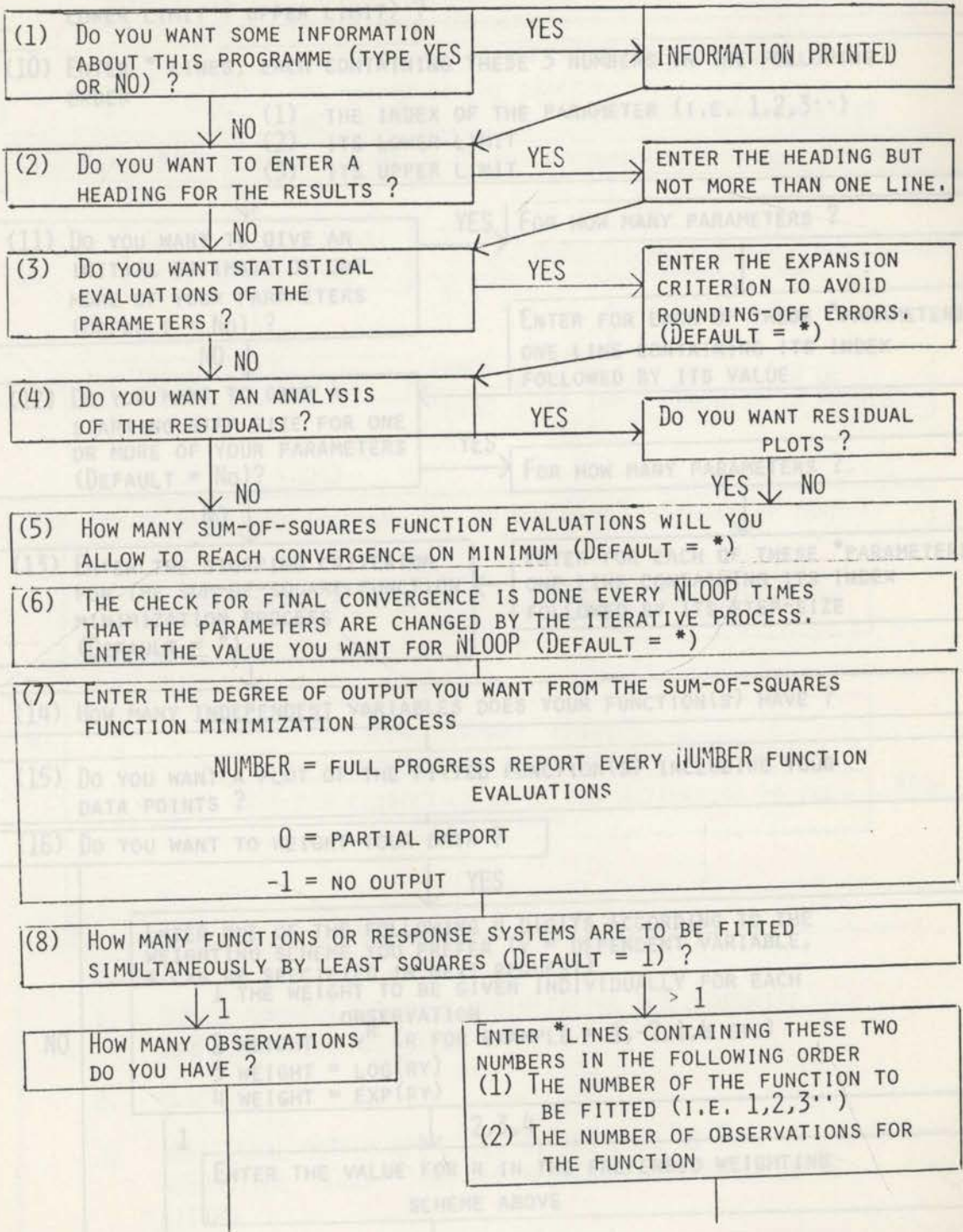
$$F = \frac{SS(\hat{P}_{(1)})}{SS(\hat{P}_{(2)})} \frac{NOBS_2 - n_2}{NOBS_1 - n_1} \quad (8.51)$$

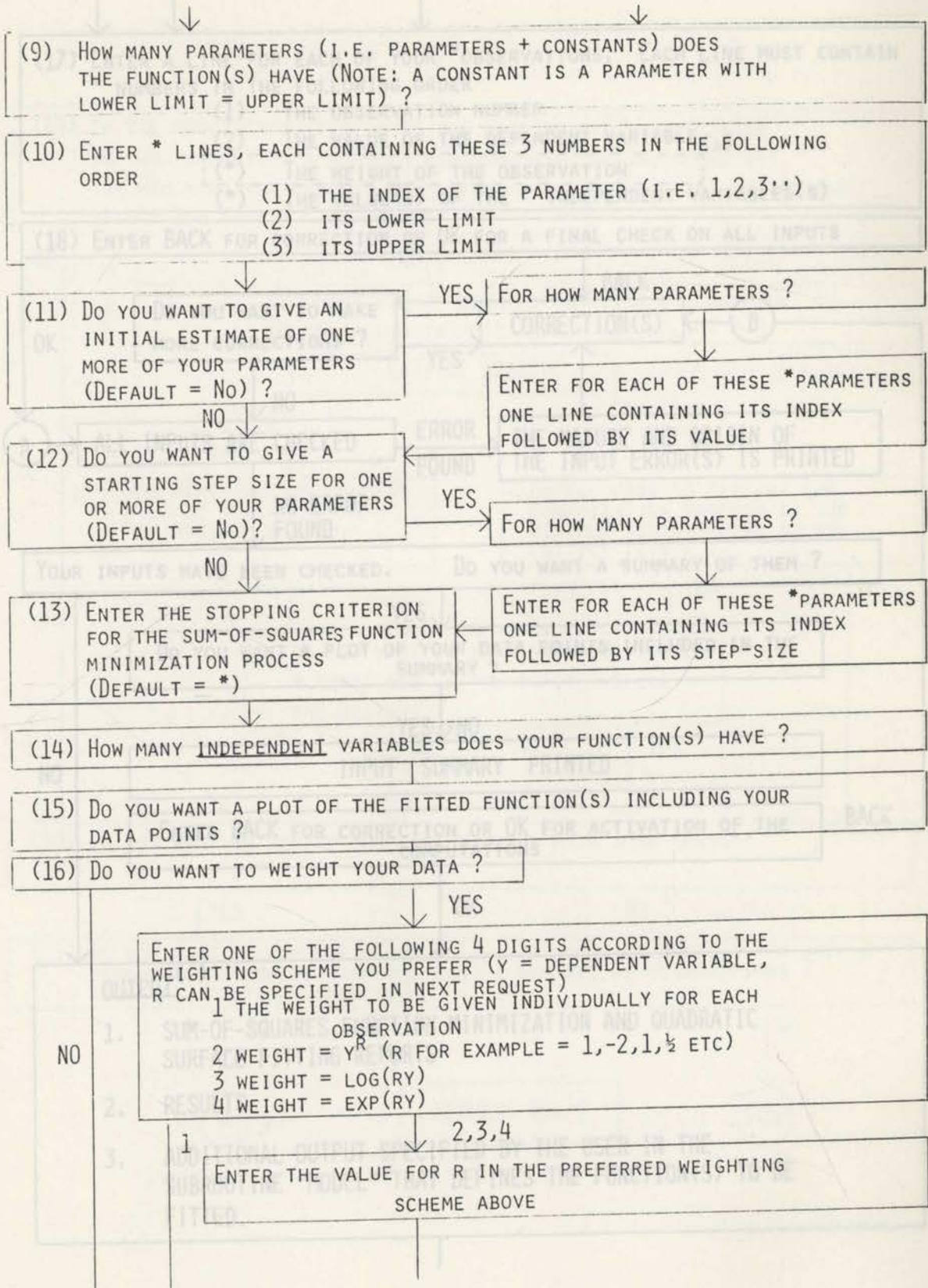
follows an F- distribution with  $(NOBS_1 - n_1)$  and  $(NOBS_2 - n_2)$  degrees of freedom. Therefore, if  $F > F_{\alpha, NOBS_1 - n_1, NOBS_2 - n_2}$ , the null hypothesis that the residual variances are identical should be rejected. The hypothesis that the residual variance of model 2 is significantly smaller than that of model 1, (i.e. that model 2 fits the experimental data better than model 1) can therefore be accepted. If F does not exceed the critical value, further investigations are required or the simpler of the two models should be chosen.

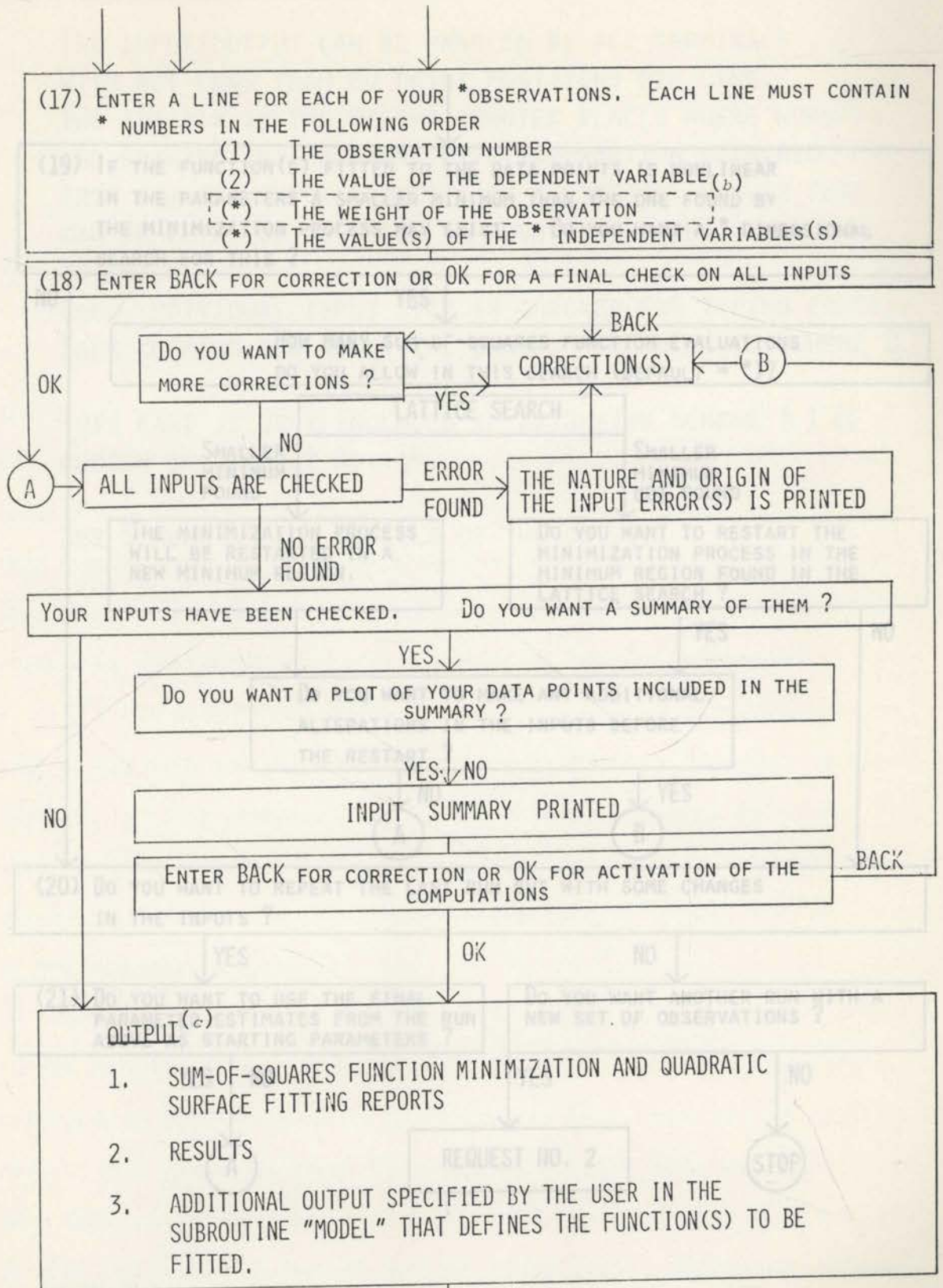
INPUT TO FUNFIT

The input is provided in a communication between the user and the computer (scheme 8.1). Questions are asked about which data treatment is wanted and depending on the answers, the next questions are given in relation

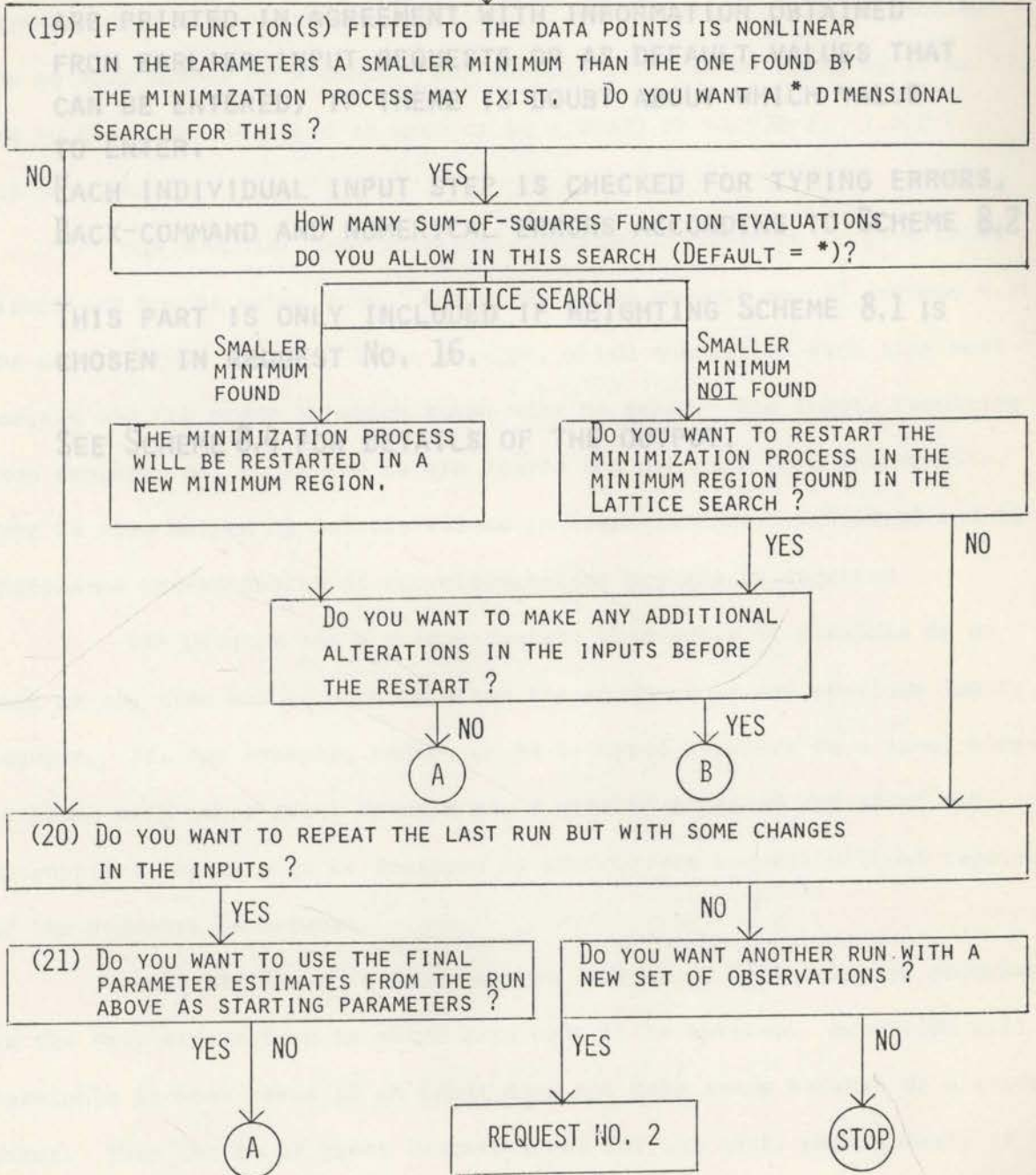
SCHEME I USER - COMPUTER INTERACTION DURING EXECUTION OF THE TIME-SHARING PROGRAMME "FUNFIT" FOR GENERAL NONLINEAR REGRESSION AND CURVE FITTING<sup>(a)</sup>







THE INPUT/OUTPUT CAN BE HANDLED BY ALL TERMINALS WITH NOT LESS THAN 80 PRINT POSITIONS PER LINE. THE ASTERIX IN THE SCHEME DENOTES PLACES WHERE NUMBERS



(a) THE INPUT/OUTPUT CAN BE HANDLED BY ALL TERMINALS WITH NOT LESS THAN 80 PRINT POSITIONS PER LINE. THE ASTERIX IN THE SCHEME DENOTES PLACES WHERE NUMBERS ARE PRINTED IN AGREEMENT WITH INFORMATION OBTAINED FROM EARLIER INPUT REQUESTS OR AS DEFAULT VALUES THAT CAN BE ENTERED, IF THERE IS DOUBT ABOUT WHICH VALUE TO ENTER. EACH INDIVIDUAL INPUT STEP IS CHECKED FOR TYPING ERRORS, BACK-COMMAND AND NUMERICAL ERRORS ACCORDING TO SCHEME 8.2

(b) THIS PART IS ONLY INCLUDED IF WEIGHTING SCHEME 8.1 IS CHOSEN IN REQUEST No. 16.

(c) SEE SCHEME 8.4 FOR DETAILS OF THE OUTPUT.

The program has a unique feature that makes it possible to go back at any time and correct or alter the input under any previous input request. If, for example, BACK4 or B4 is typed anywhere on a line, alone or along with other data, request no. 4 will be repeated and after the appropriate input, will be followed by the current request without repetition of the requests in between.

Possibly the most inconvenient feature of most computer programs is the very strict form in which data have to be entered. Execution will terminate in most cases if an input does not make sense because of a typing error. This can be of great inconvenience for the user, particularly if many data have to be entered. 'PUNFIT' does not have this disadvantage. It will not terminate for any kind of typing error. Instead every input is checked in several ways (scheme 8.2). Typing errors, logical errors and numerical errors are all detected. It is, for example, not uncommon to

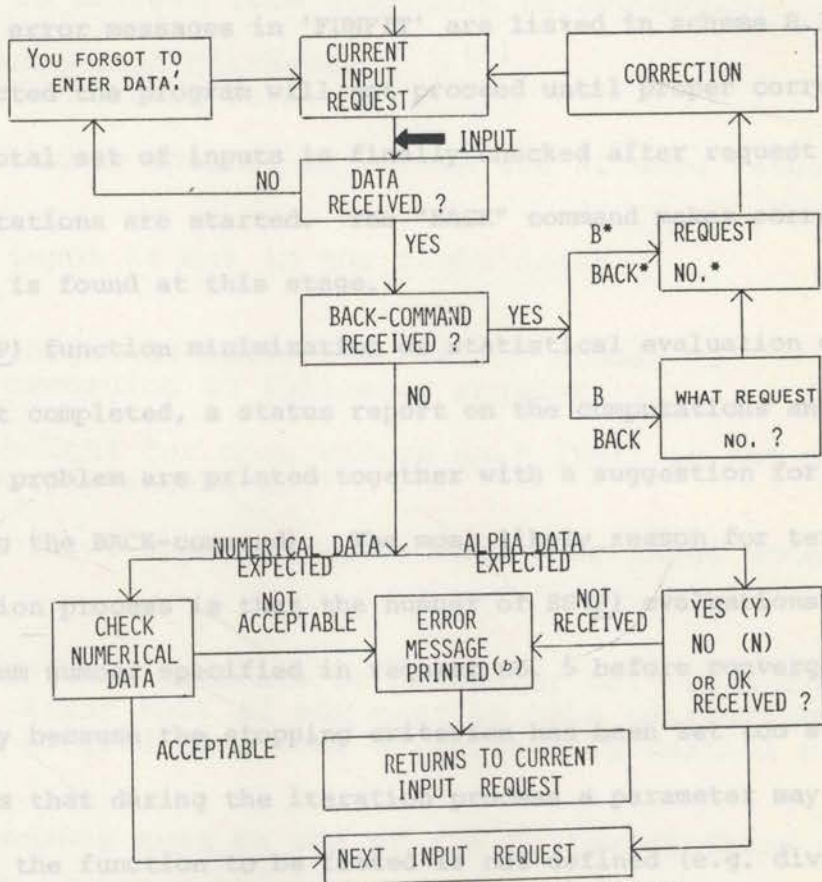
to the particular treatment chosen. The answers can be YES or NO or, in short form, Y or N. Where numerical data are requested, these can be typed anywhere on a line in integer, decimal or exponential form separated by one or more blanks or a comma. For example 17.0 can be typed as 17 or 17. and -0.00153 can be typed as such or as -.00153 or -1.53E-3, -1.53E-03, -153E-5, etc.

The computer uses previously entered information to give exact directions how to enter data. For example, in request no. 17 (scheme 8.1), the user is told how many lines to type, which quantities each line must contain and the order in which these must be typed. The inputs resulting from requests no. 8, 14 and 16 are stored and used for this instruction. The user is also helped by default values in requests no. 3, 5, 6, 8, 12, 13 and 19 where some understanding of the minimization process is required.

The program has a unique feature that makes it possible to go back at any time and correct or alter the input under any previous input request. If, for example, BACK4 or B4 is typed anywhere on a line, alone or along with other data, request no. 4 will be repeated and after the appropriate input, will be followed by the current request without repetition of the requests in between.

Possibly the most inconvenient feature of most computer programs is the very strict form in which data have to be entered. Execution will terminate in most cases if an input does not make sense because of a typing error. This can be of great inconvenience for the user, particularly if many data have to be entered. 'FUNFIT' does not have this disadvantage. *It will not terminate for any kind of typing error.* Instead every input is checked in several ways (scheme 8.2). Typing errors, logical errors and numerical errors are all detected. It is, for example, not uncommon to

SCHEME 8.2 STRUCTURE OF THE DATA CHECK PERFORMED AT EACH INDIVIDUAL INPUT STEP IN SCHEME 8.1 (a)



(a) THE INPUT DATA ARE CHECKED FOR NUMERICAL, LOGICAL AND TYPING ERRORS. IF AN ERROR IS MADE IT IS ESSENTIALLY IMPOSSIBLE TO PROCEED BEFORE IT HAS BEEN PROPERLY CORRECTED. ALL INPUT DATA ARE FURTHERMORE CHECKED COLLECTIVELY AFTER REQUEST NO.18 IN SCHEME 8.1 SUCCESSFUL RUNS SHOULD IN THIS WAY BE GUARANTEED IN MOST CASES.

(b) SCHEME 8.3 GIVES EXAMPLES OF SOME OF THE MANY INPUT ERROR MESSAGES IN FUNFIT.



type a comma in place of a decimal point or an "0" instead of a zero;

such errors are clearly indicated by the program and the last input line is repeated for immediate correction. An error message is also printed

so corrections can be made if an input does not agree with an earlier input.

Some of the many error messages in 'FUNFIT' are listed in scheme 8.3. If an error is detected the program will not proceed until proper corrections

are made. The total set of inputs is finally checked after request no. 18 before the computations are started. The "BACK" command makes corrections

easy if an error is found at this stage.

If SS(P) function minimization or statistical evaluation of the parameters is not completed, a status report on the computations and a

diagnosis of the problem are printed together with a suggestion for a suitable correction (using the BACK-command). The most likely reason for termination

of the minimization process is that the number of SS(P) evaluations may exceed the maximum number specified in request no. 5 before convergence is

reached, probably because the stopping criterion has been set too strictly. Another reason is that during the iteration process a parameter may reach

a value at which the function to be fitted is not defined (e.g. division by zero, taking the logarithm of a non-positive number, etc.). This can be

avoided by a proper formulation of the user-supplied subroutine 'MODEL' and by the right choice of parameter limits.

At convergence, the point in n-dimensional parameter space defined by  $\hat{P}$  is surrounded by other points which form a polygon with  $\hat{P}$  as centroid. The size of this polygon depends on the accuracy with which the SS(P) minimum is determined and diminishes for small values of the stopping criterion.

The SS-values at the vertices, the centroid and mid-points of the sides of the polygon provide the surface points used in the quadratic surface fitting for estimating the variability of the parameters (135). Because of the small

SCHEME 8.3 SOME INPUT ERROR MESSAGES IN FUNFIT<sup>(a)</sup>

---

Input error, too many numbers on line above.\* are expected.  
Reenter last line in correct form according to request.

Input error, too few numbers on line above.\* are expected.  
Reenter last line in correct form according to request.

Input error. Unrecognized character ( )<sup>(b)</sup>.  
Reenter last line in correct form.

Input error. The number entered is not in the allowed range.  
- Try again -

Input error. The input is not in the right order.  
- Try again -

Input error. You forgot to enter observation number \*.  
Reenter your data according to following request:

Input error. The weight for observation no.\* (Y=\*) is not defined  
by the chosen weighting scheme no.\*

Input error. Your input does not agree with your input under  
request no.\*  
- Try again -

Input error. The initial estimate of parameter no.\* is not within  
the limits given. Enter BACK if you want to make correction or  
OK if you want the program to choose an acceptable initial param-  
eter estimate.

Input error, your answer must be one of the following three only  
YES NO BACK (Y,N,B)

- Try again -

Input error. Your input under request no.\* does not agree with  
the input under request no.\*. What request number do you want to  
go back to for making corrections?

Input error. You forgot to enter data.  
- Try again -

Input error. The weight of the dependent variable is negative.  
Reenter last line in correct form.

---

(a) The appropriate numbers are printed where \* appears in the text.

(b) The mistyped symbol is printed in the bracket. This feature  
prevents termination of the program when for example common  
errors such as typing ',' instead of '.' or the letter O in-  
stead of zero are committed.

size of the polygon at convergence these points will be so close together that numerical rounding-off errors can be significant in the fitting procedure. To prevent this, the polygon is expanded around the centroid before the quadratic surface is fitted until the SS-values at the vertices exceed that at the centroid by more than a given value of the expansion criterion (request no. 3). Two tests are then done to ensure that the statistical evaluation of the parameters is not significantly subject to numerical errors: The minimum of the fitted surface is compared with the minimum  $SS(\hat{P})$  found by the minimization process to check for round-off errors; also, the surface points used in the fitting procedure must lie in a convex region of the  $SS(P)$  surface in order to be close to the true minimum and this is checked by evaluating whether the Hessian matrix (Eq. 8.13) is positive definite.

It is however most unlikely that there will be any computational difficulties either in the minimization process or in the statistical evaluation of the parameters if the recommended (default) values are chosen in requests no. 3,5 and 13.

Lower and upper parameter limits are specified in request no. 10 to prevent convergence on an  $SS(P)$  minimum that may give unrealistic parameter estimates. If an initial estimate is not given for a parameter in request no. 11 the mid-point of the range defined by its lower and upper limits is chosen. Default values for the initial step sizes (request no. 12) are chosen in relation to the initial parameter estimates and their lower and upper limits so that a large part of the parameter space is searched in the  $SS(P)$  minimization process and the chance of overlooking a smaller minimum is reduced.

Requests no. 11 and 12 also provide the option of choosing one or more initial parameter estimates and their respective step sizes independently

of their lower and upper limits. This option can be useful if it is desirable to increase the chance of finding a minimum that gives final estimates close to the initial estimates instead of finding another even smaller minimum that may exist but would give less realistic parameter estimates.

By entering 1 in request no. 5 a lattice search can be made before the start of a run. The coordinates of the point in the lattice which has the smallest  $SS(\underline{P})$  value can then be chosen as the initial estimates of the parameters, and the run can be started after input no. 5 has been modified (using a B5 command). This 'pre-search' for the best starting values increases the chance of finding the global minimum.

#### APPLICATIONS OF THE 'BACK' COMMAND

The B\*\* command is not only useful for correcting input errors but has wider application. For example, if the analysis of the residuals suggests a particular weighting scheme, a new run with weighted data can easily be made. By typing B16, request no. 16 is repeated and the desired weighting scheme can be chosen (e.g.  $R = -2$  will give weights proportional to  $y^{-2}$ ). The 'communication' then continues from the current request and a new run, this time with weighted data, will be started by typing B18 followed by OK and N in succession. A new run in which one or more of the parameters is kept constant can, in a similar way, be made by repeating request no. 10 and fixing the parameters in question by setting upper limit=lower limit=the fixed value.

The lattice search that can be chosen in request no. 19 includes, in the case of two parameters (2-dimensional search), a contour map of the  $SS(\underline{P})$  surface, together with matrices of the  $SS(\underline{P})$  values and the sign of

the partial derivatives of  $SS(P)$  with respect to each of the two parameters at the points in the search grid. In the case of only one parameter (all others fixed) a plot will be printed of  $SS(P)$  versus the parameter.

The B\*\* command is particularly useful in connection with a lattice search involving more than 2 parameters if a composite picture of the  $SS(P)$  surface is desired. This is done as follows: The command B10 is typed in response to request no. 19 (or after a lattice search) and all but two parameters are fixed. Then after the following 2-dimensional search, B19 is typed, followed again by B10. This procedure can be repeated as many times as desired, each time 'slicing' the parameter space in a different way.

If two or more response systems are fitted simultaneously the data for these are stacked and numbered in succession (in request no. 17). Therefore, a new run where only the first response system is fitted individually can easily be made by typing B8 followed by 1, B18 and N in succession. If the first run was a simultaneous fitting of a pharmacokinetic model using both blood and urine data the effect of including or excluding the urine response system can quickly be evaluated.

Every time the B\*\* command is used the computer stores the current request position and returns to this immediately after correction of a previous input. It is however, possible to erase this memory and continue without returning from a previous request by typing the same B\*\* command twice in succession. This procedure is useful if, for example, the first 7 inputs for a new run with different observations are identical to those of a previous run. These inputs are then taken to be the same and need not be repeated for the new run if B8 is typed twice in succession after completion of the previous run.

In fitting pharmacokinetic models it is often necessary to calculate from the final parameter estimates, quantities such as half-lives, clearances and volumes of distribution or to make a plot of the amount of drug in a peripheral compartment versus time. It will be demonstrated in the following section how the subroutine 'MODEL' which describes the equation(s) to be

SCHEME 8.4 SUMMARY OF OUTPUT FROM 'FUNFIT' (4)

OUTPUT

Scheme 8.4 summarizes the maximum output possible from 'FUNFIT' for a single run. *The sections printed in italics denote the standard (minimum) output that is always printed (or displayed on a terminal screen).* The degree of output will range from this to the maximum output according to the user's specifications.

The optional input summary is useful as a check for numerical errors before activation of the computations or may serve as an extra copy of the experimental data. A minimization report should be chosen if the user-supplied subroutine 'MODEL' specifies parameter constraints that may cause convergence problems. The report is also of value because it provides a table of SS(P) values and parameter values that gives some information about the sum-of-squares surface and how large a section of the parameter space has been searched on the way to the minimum.

The quadratic surface fitting report is only printed in conjunction with the minimization report when statistical evaluation of the parameters has been requested (no. 3). The variance-covariance and correlation matrices of the parameters printed in this section will also appear in the general result section. The plots printed in the result section are line plots and are therefore of low accuracy but provide the essential information. The exact coordinates of the points in the plots are tabulated so precise plots can be produced manually if desired.

In fitting pharmacokinetic models it is often necessary to calculate from the final parameter estimates, quantities such as half-lives, clearances and volumes of distribution or to make a plot of the amount of drug in a peripheral compartment versus time. It will be demonstrated in the following section how the subroutine 'MODEL' which describes the equation(s) to be

SCHEME 8.4 SUMMARY OF OUTPUT FROM 'FUNFIT' <sup>(a)</sup>

INPUT SUMMARY <sup>(b)</sup>

- Heading for problem. - Number of: variable parameters, constant parameters, observations and independent variables for each response system. Table of parameters with lower and upper limits, initial estimates and step sizes. - Table of observations with dependent variable(s), independent variable(s), weight (if any) and normalized weight. - Plot of dependent variable(s) versus independent variable(s). - Weighting scheme used, expansion and stopping criterion. -

SUM-OF-SQUARES FUNCTION MINIMIZATION REPORT <sup>(c)</sup>

- Table of evaluation no., sum-of-squares values and parameters. - sum-of-square value and parameter estimates at convergence and number of evaluations used to reach convergence. -

FITTING OF QUADRATIC SURFACE IN REGION OF MINIMUM <sup>(c)</sup>

- Minimum of quadratic surface and parameter values at minimum. - Generalised inverse of information matrix ( $H^{-1}$ ). - Information matrix (H). - Correlation matrix (Eq.8.17). - Number of evaluations used in the fitting. -

RESULTS

- Heading for problem. - Table of parameters with their lower and upper limits, initial and final estimates. - Table of standard deviation, coefficient of variation and 95% confidence limits of the parameters. - Graphical illustration of the relative position of

the calculated parameters in their specified range.- Residual sum of squares, regression sum of squares, sum of squared response, mean of response, residual mean square, regression mean square, mean of residuals and correlation coefficient.- Weighted residual sum of squares, weighted residual mean squares, mean of weighted residuals.- Table of dependent variable(s), independent variable(s), observed and calculated responses, difference in response, differences expressed as percentages and as normal deviates.- Table of weights, normalized weights, weighted residuals and normal deviate form of weighted residuals.- Plot of calculated (fitted) curve including experimental data points. Table of coordinates in plot.- Analysis of variance table.- Durbin-Watson statistics for serial correlation of residuals. 'Run test' and 'number test' for randomness of residuals.- Residual plots: residuals versus dependent and independent variables. Weighted residuals versus dependent and independent variables.- Variance-covariance matrix.- Correlation matrix.- OUTPUT SPECIFIED IN THE USER-SUPPLIED SUBROUTINE 'MODEL'.

#### LATTICE SEARCH FOR GLOBAL MINIMUM<sup>(d)</sup>

The output in this section depends on the number of variable parameters as follows: (1 PARAMETER) - Plot of sum-of-squares function values versus the parameter in its specified range.- Table of coordinates of points in the plot.- Sum-of-squares and parameter values at minimum found in the unidimensional search.- (2 PARAMETERS)- Residual sum-of-squares matrix.

- Matrices of the sign of the partial derivatives of the sum-of-squares function with respect to each parameter.
- Sum-of-squares contour map.- Parameter values at the grid point



that has the smallest sum-of-squares value.- (> 2 PARAMETERS).-

Total number of lattice points evaluated. Number of divisions of each parameter interval.- Parameter values of the grid point with the smallest sum-of-squares value.

'PLOT', which enables the user to make special plots in this section.

- (a) The scheme summarizes the maximum degree of output possible from FUNFIT for a single run. The sections printed in italics denote the standard, minimum output that is always printed. The degree of output will vary in this range according to the user's specifications.
- (b) The input summary can be chosen to check all the inputs before the activation of the computations or may serve as an extra copy of the experimental data.
- (c) It is useful to choose a minimization report if special parameter constraints have been specified in the user supplied subroutine 'MODEL' that specifies the function(s) to be fitted.
- (d) The lattice search can be used to evaluate the contour of the sum-of-squares surface when there are only two parameters. A composite picture of the surface can be built up if there are more than two by fixing all but two parameters at a time (see text).

The structure can best be illustrated by some examples.

1. A single Equation, one Dependent and Independent Variable:

To fit the two-exponential equation:

$$C = P_1 e^{-P_2 t} + P_3 e^{-P_4 t} \tag{8.52}$$

the subroutine can be written simply as:

```

SUBROUTINE MODEL (C,T,P,IPRINT)
  DIMENSION P(4)
  E = P(1)*EXP(-P(2)*T) + P(3)*EXP(-P(4)*T)
  RETURN
END

```

fitted, can include a special section containing such additional calculations and plots. This 'user-supplied output' will then be printed as the last part of the results section. The program includes an easily applicable subroutine, 'PLOT', which enables the user to make special plots in this section.

DEFINING THE EQUATION(S) TO BE FITTED

In order to use FUNFIT the function(s) to be fitted must first be defined by the user in a special subroutine called MODEL. The structure of this subroutine is:

```
SUBROUTINE MODEL (Y,X,P,IPRINT)
```

where Y denotes the dependent variable(s); X the independent variable(s); P the parameter vector and IPRINT is an integer variable which is controlled by FUNFIT and is used if additional output is desired. The subroutine must define Y as a function of X and P.

The structure can best be illustrated by some examples,

1. A single Equation, one Dependent and Independent Variable:

To fit the two-exponential equation:

$$c = p_1 e^{-p_2 t} + p_3 e^{-p_4 t} \tag{8.52}$$

the subroutine can be written simply as:

```
SUBROUTINE MODEL (C,T,P,IPRINT)
```

```
DIMENSION P(4)
```

```
C = P(1)*EXP(-P(2)*T) + P(3)*EXP(-P(4)*T)
```

```
RETURN
```

```
END
```

1. It is recommended to use NPOINTS=75 and NLines=50.

Special Output

If it is desired to calculate a quantity, say:

$$A = p_1/p_2 + p_3/p_4 \tag{8.53}$$

from the final least squares parameter estimates this can be done by the computer by including the following special output section in the subroutine:

```
DIMENSION P(4),F2(75),TT(75)
IF (IPRINT.EQ.Ø) RETURN
COMMON XMIN, XMAX
A = P(1)/P(2) + P(3)/P(4)
WRITE(6,1)A
1 FORMAT(" A=", E12.6)
```

just before the RETURN statement. It is seen that the IPRINT parameter controls the execution of the special output section. This section is printed once just after the general output section described in scheme 8.4.

An easily applicable plotting routine is available for special plots the user may wish to make in the special output section. Its structure is

```
SUBROUTINE PLOT (XARRAY,YARRAY,NPOINTS,NLINES)
```

where XARRAY and YARRAY are two arrays of dimension NPOINTS which contain the coordinates to the points to be plotted using NLINES of the output device.<sup>3</sup>

In calculating the points for such plots it is frequently convenient to know the interval over which the observations are taken, i.e. the smallest and largest X value. This information can be made available by including the common statement:

```
COMMON XMIN, XMAX
```

---

3. It is recommended to use NPOINTS=75 and NLINES=50.

The following subroutine illustrates the use of PLOT in terms of a plot (75 points) of the function:

$$f_2 = \left| e^{-P_4 t} - e^{-P_2 t} \right| \quad (8.54)$$

over the range of the t values, i.e. from XMIN to XMAX:

```
SUBROUTINE MODEL (C,T,P,IPRINT)
```

```
DIMENSION P(4),F2(75),TT(75)
```

```
COMMON XMIN, XMAX
```

```
C=P(1)*EXP(-P(2)*T) + P(3)*EXP(-P(4)*T)
```

```
IF (IPRINT.EQ.0) RETURN
```

```
DO 1 I = 1,75
```

```
TT(I) = XMIN + (I-1)*XMAX/74.
```

```
1 F2(I) = ABS(EXP(-P(4)**TT(I)) - EXP(-P(2)*TT(I)))
```

```
CALL PLOT (TT,F2,75,50)
```

```
RETURN
```

```
END
```

### Fitting Several Functions Simultaneously

Example  
The general structure of the user supplied subroutine MODEL when N response systems or equations are to be fitted simultaneously can be illustrated schematically as follows:

```
SUBROUTINE MODEL (Y,X,P,IPRINT)
```

```
DIMENSION P( )
```

```
COMMON/FUNNUM/ITHFUN
```

```
GO TO (1,2,...,N) ITHFUN
```

```
1 Y = (1st function)
```

```
RETURN
```

2 Y = (2nd function)

RETURN

$$y_2 = \frac{\sigma^2}{P_2 - P_1} (\exp(-P_1 t) - \exp(-P_2 t))$$

.

N Y = (Nth function)

RETURN

END

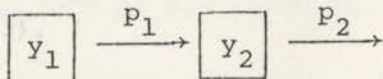
It is desired to estimate  $p_1$  and  $p_2$  from the set of observations of  $(y_1, t)$  and  $(y_2, t)$ . This can be done by fitting simultaneously 8.57 and 8.58 to the observations.

These equations can conveniently be defined as follows:

The basis of this structure can be explained in the following way: During the input procedure each observation point is numbered consecutively and automatically assigned a label ITHFUN (i-th function) indicating which response system it belongs to. The user supplied subroutine MODEL is called by FUNFIT for each observation point. The parameter ITHFUN that is introduced by the COMMON block named FUNNUM (function number) transfer control to the i-th function which is then calculated. Control then returns to FUNFIT where the squared residual at that particular point is calculated, and added to the previous squared residual in the process of calculating the sum of squared residuals.

Example

Consider the following linear compartmental system:



where  $y_1$  and  $y_2$ , which are measured at various times  $t$ , are given by:

$$dy_1/dt = -P_1 y_1 \tag{8.55}$$

$$dy_2/dt = P_1 y_1 - P_2 y_2 \tag{8.56}$$

which (for  $y_1=y_0$  and  $y_2=0$  at  $t=0$ ) integrates to:

various temperatures  $T$  (K). A simplified model for this reaction may be written as:

$$y_1 = y_0 \text{EXP}(-p_1 t) \tag{8.57}$$

$$y_2 = \frac{y_0 p_1}{p_2 - p_1} (\text{EXP}(-p_1 t) - \text{EXP}(-p_2 t)) \tag{8.58}$$

It is desired to estimate  $p_1$  and  $p_2$  from the set of observations of  $(y_1, t)$  and  $(y_2, t)$ . This can be done by fitting simultaneously 8.57 and 8.58 to the observations.

These equations can conveniently be defined as follows:

```
SUBROUTINE MODEL (Y,T,P,IPRINT)
```

```
  DIMENSION P(3)
```

```
  COMMON/FUNNUM/J
```

```
  GO TO (1,2) J
```

```
  1  Y = P(3)*EXP(-P(1)*T)
```

```
  RETURN
```

```
  2  Y = P(3)*P(1)/(P(2)-P(1))*(EXP(-P(1)*T)-EXP(-P(2)*T))
```

```
  RETURN
```

```
  END
```

The quantity  $y_0$  can either be defined as a constant by setting its upper and lower limits equal to the same (constant) value in the input, or it can be defined as a parameter that is to be estimated by assigning appropriate bounds for  $P(3)$ .

### Fitting an Equation with 2 Independent Variables, a Simple Example

Consider a first-order reaction:



where the fraction remaining,  $f_A$ , of A is measured at various times,  $t$ , at various temperatures  $T$  (°K). A simplified model for this reaction may be written as:

$$f_A = \text{EXP}(-p_1 t \text{EXP}(-p_2/T)) \quad (8.59)$$

where  $p_1$  and  $p_2$  are parameters to be determined from a set of  $(f_A, t, T)$  data points. This model can be defined in the following way for FUNFIT:

```

SUBROUTINE MODEL (FA,X,P,IPRINT)
DIMENSION P(2),X(2)
FA = EXP(-P(1)*X(1)*EXP(-P(2)/X(2)))
RETURN
END
    
```

where  $X(1)$  acts as the time,  $t$ , and  $X(2)$  as the absolute temperature ( $^{\circ}\text{K}$ ).

Fitting of Implicit Equations, A Simple Example

It is, in the following equation:

$$p_1 x = \ln y + p_2 y \quad x, y, p_1, p_2 > 0 \quad (8.60)$$

not possible to express  $y$  explicitly as a function of the independent variable  $x$  so a special technique must be used to express and fit the function:

$$y = f(x, p_1, p_2) \quad (8.61)$$

The numerical problem of defining 8.60 in the form of 8.61 is the same as finding the root of the equation:

$$p_1 x - \ln y - p_2 y = 0 \quad (8.62)$$

for given values of  $x$ ,  $p_1$  and  $p_2$ ; where  $p_{1,2}$  are the parameters to be determined from a set of  $(x, y)$  observations. Let the left-hand side of 8.62 be denoted  $g(y)$  then  $y$  can be found by the Newton-Raphson iteration:

$$y_{i+1} = y_i - \frac{g(y_i)}{g'(y_i)} \quad (8.63)$$

where 
$$g'(y) = \frac{\partial g(y)}{\partial y} = -\frac{1}{y} - p_2 \quad (8.64)$$

i.e. 8.63 becomes: 
$$y_{i+1} = y_i + \frac{p_1 x - \ln y_i - p_2 y_i}{\frac{1}{y_i} + p_2} \quad (8.65)$$

Since  $y$  and  $p_2 > 0$  then  $g'(y) < 0$  so the function  $g(y)$  will be monotonously decreasing:

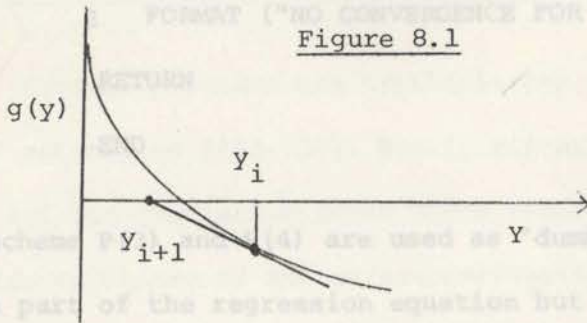


Figure 8.1

In the iteration 8.65  $y_{i+1}$  may become non-positive for which  $g(y)$  will not be defined in the next iteration. To prevent this to occur it is convenient to define:

$$y_{i+1} = y_i/2 \quad \text{if} \quad y_{i+1} \leq 0 \quad (8.66)$$

The iteration 8.65 may be considered to have converged when the relative change in  $y$  between iterations is less than 0.0001% i.e. when

$$|(y_{i+1} - y_i)/y_i| < 10^{-6}.$$

The subroutine to define 8.61 (i.e. 8.60) may thus be defined in the following way:

```

SUBROUTINE MODEL(Y,X,P,IPRINT)
DIMENSION P(4)
C     P(3) = CONVERGENCE CRITERION
C     P(4) = MAX.NO.OF ITERATIONS
MAX = IFIX (P(4))
DO 1 I = 1, MAX
YSAVE = Y
Y = Y + (P(1)*X-ALOG(Y)-P(2)*Y)/(1/Y+P(2))
    
```



IF (Y.LE.Ø.) Y = YSAVE/2.

IF (ABS ((Y-YSAVE)/YSAVE) .LT. P (3)) RETURN

CURVE FITTING AND MODELLING IN PHARMACOKINETICS

1 CONTINUE

WRITE (6,2) X,Y,P

A Comparison of FURFIT and NONLIN computer programs.

2 FORMAT ("NO CONVERGENCE FOR X,Y,P =", 6E12.6)

RETURN

END

Several programs are available for nonlinear least squares parameter estimation (123-128). Nearly all are based on the Gauss-Newton or other related gradient methods since these are usually rapidly convergent. In this scheme P(3) and P(4) are used as "dummy" constant parameters which provide estimates of the variance-covariance matrix. However, such are not a part of the regression equation but included of computational gradient methods may fail when the residuals are large (145, 146), as is often the case in fitting equations to biological data, and they may converge on a non-stationary point (147) if great care is not taken by the user in choosing a suitable value for the step size used in the finite difference approximation of derivatives. The default value specified for this step size in a program may unfortunately apply successfully only in a limited number of cases. Such practical experiences are illustrated in this paper in the application of the program NONLIN (123) which is based on Hartley's modification of the Gauss-Newton algorithm (148).<sup>1</sup>

Such problems are eliminated in FURFIT. This program has implemented the adaptive simplex method of Nelder and Mead (135,150) which is a non-gradient method that does not require evaluation of derivatives. This method is less efficient than Gauss-Newton based methods but considerably more robust and reliable. It will never fail even under extreme conditions where the gradient methods may be unstable due to near singularity and ill-conditioning of the matrices used in the iterative procedure.

1. Numerical techniques in nonlinear parameter estimation have been reviewed by Chambers (149) and an excellent discussion has been given by Bercia (146).

CHAPTER 9

CURVE FITTING AND MODELLING IN PHARMACOKINETICS

A Comparison of FUNFIT and NONLIN computer programs.

Several programs are available for nonlinear least squares parameter estimation (123-128). Nearly all are based on the Gauss-Newton or other related gradient methods since these are usually rapidly convergent and provide estimates of the variance-covariance matrix. However, such gradient methods may fail when the residuals are large (145, 146), as is often the case in fitting equations to biological data, and they may converge on a non-stationary point (147) if great care is not taken by the user in choosing a suitable value for the step size used in the finite difference approximation of derivatives. The default value specified for this step size in a program may unfortunately apply successfully only in a limited number of cases. Such practical experiences are illustrated in this paper in the application of the program NONLIN (123) which is based on Hartley's modification of the Gauss-Newton algorithm (148).<sup>1</sup>

Such problems are eliminated in FUNFIT. This program has implemented the adaptive simplex method of Nelder and Mead (135, 150) which is a nongradient method that does not require evaluation of derivatives. This method is less efficient than Gauss-Newton based methods but considerably more robust and reliable. It will never fail even under extreme conditions where the gradient methods may be unstable due to near singularity and ill-conditioning of the matrices used in the iteration procedure.

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1. Numerical techniques in nonlinear parameter estimation have been reviewed by Chambers (149) and an excellent discussion has been given by Dennis (146).

The possibility of multiple solutions (multiple sums of squares minima) undoubtedly represents the greatest problem in nonlinear estimation. The problem is expected to be particularly pronounced in pharmacokinetic studies, because these often involve the fitting of multiexponential equations to rather variable biological data, and because the ratio of number of data points to number of parameters is often quite small. Parameter estimation under such conditions may produce spurious results and discrimination between pharmacokinetic models may be very difficult and unreliable. The problem can be reduced, but usually not eliminated, if a graphical or numerical method is available which provides good initial parameter estimates for the iteration procedure. This is seldom the case for models describing nonlinear pharmacokinetics. The best approach should therefore be to use an algorithm which is effective in finding the (statistically) best solution in terms of the smallest residual sum-of-squares value.

It is generally accepted that the nongradient search methods perform better than the gradient methods in this respect. In particular, the adaptive simplex method used in FUNFIT appears to be very suitable because of its unique minimization method.

Regardless of the choice of algorithm the question of which starting values the parameters should be given still seems to be the greatest practical problem the user faces in nonlinear estimation. Frequently, when no preliminary estimation technique is available the initial values are simply guesses which all too often produce unacceptable results in the first run. However, by studying these results, corrections can often be made so that acceptable results can be obtained in subsequent runs.

plans,  $\sigma$ , is given implicitly by:

Interactive programs are most convenient in such cases. In particular, FUNFIT has been designed so that it allows a highly interactive and flexible editing of input at any stage.

### Pharmacokinetic applications of FUNFIT

1. The classical linear, compartmental models are still the most often used models in pharmacokinetic studies. The evaluation of such models is well documented and has become a routine procedure in many investigations.
2. There has, however, been an increasing awareness that linear models cannot adequately describe certain drug disposition phenomena (151,152) and various nonlinear models have been postulated. These mathematical models are often of a form which requires a special technique for least squares fitting.
3. Often several possible models are investigated to explain a pharmacokinetic phenomenon. There has been increasing interest in discriminating between such models (152,153).

It is appropriate to discuss points 2. and 3. above:

#### Fitting of Implicit Functions: a simple example

Several of the models describing nonlinear pharmacokinetic phenomena can be expressed in an implicit form which can be fitted by defining the functional relationship between the variables explicitly by an iterative procedure.

Consider, for example, a simple one-compartmental model with intravenous injection in which the drug is eliminated partly by conversion to a single metabolite according to Michaelis-Menten kinetics and partly by excretion unchanged in the urine (154). The concentration of drug in plasma,  $c$ , is given implicitly by:

$$\ln \frac{c}{c_0} = \frac{V_m}{k_{lu} K_m} \ln \left[ \frac{k_{lu} K_m + V_m + k_{lu} c}{k_{lu} K_m + V_m} \right] - \left[ \frac{k_{lu} M + V_m}{K_m} \right] t \quad (9.1)$$

where  $V_m$  and  $K_m$  are the Michaelis-Menten parameters,  $C_0 = \text{Dose}/V_1$  and  $k_{lu}$  is the urinary elimination constant. This equation can be written more simply:

$$\ln c + A_1 \ln (A_2 + A_3 c) + A_4 = 0 \quad (9.2)$$

where  $A_1 = \frac{V_m}{k_{lu} K_m}$  (9.3)

$$A_2 = \frac{k_{lu} K_m + V_m}{k_{lu} K_m + V_m + k_{lu} c_0} \quad (9.4)$$

$$A_3 = \frac{k_{lu}}{k_{lu} K_m + V_m + k_{lu} c_0} \quad (9.5)$$

$$A_4 = \frac{k_{lu} K_m + V_m}{K_m} t - \ln c_0 \quad (9.6)$$

Discrimination between models

The dependent variable,  $c$ , cannot be isolated from 9.1 but must be found by an iterative procedure by solving 9.2 for  $c$ .<sup>2</sup> The Newton-Raphson algorithm provides a simple and rapidly convergent method. If the expression in 9.2 is denoted  $g(c)$  then  $c$  can be determined by the following iteration:

$$c_{i+1} = c_i - \frac{g(c_i)}{g'(c_i)} \quad (9.7)$$

where  $g'(c) = \frac{\partial g}{\partial c} = \frac{1}{c} + \frac{A_1 A_2}{A_2 + A_3 c}$  (9.8)

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2. It would be incorrect, as is sometimes done for equations of similar type, to fit directly the equation where  $t$  is expressed as a function of the dependent variable. The result would be unreliable because the dependent variable which accounts for nearly all of the errors in the data, is treated as an independent variable without error.

It is important that  $c_{i+1}$  in this iteration does not take a nonpositive value since this will terminate the execution of the program because  $g(c)$  ( $\ln c$ ) is not defined for  $c \leq 0$ . To prevent this it is most convenient to define

$$c_{i+1} = c_i/2 \quad \text{if} \quad c_{i+1} \leq 0 \quad (9.9)$$

This is an acceptable approach because  $g(c)$  is strictly increasing for  $c > 0$  since  $A_1, A_2, A_3, A_4 > 0$  and therefore,  $g'(c) > 0$ . The term  $\ln(A_2 + A_3 c)$  in  $g(c)$  will also be defined under these conditions.

The above procedure can be used for most implicit mathematical models in nonlinear pharmacokinetics. However special care must be taken to prevent the parameters wandering into a parameter space where the function(s) is not defined (e.g. logarithm of a nonpositive number, division by zero etc.).

Discrimination between models

The best criterion to use in discriminating between alternate pharmacokinetic models depends on the aim of the investigation and the application of the results (138,155-). If the main aim is to discriminate between models, the experiment should be designed so that the hypothesized models are placed in as much jeopardy as possible.

The problem is nevertheless considerably complicated by the substantial variation and low reproducibility of measurements in a biological system and the limited number of sample points available. Discrimination on a statistical basis requires information about the variability of the observations which can only be estimated by repeated experiments. A likely outcome of such experiments would often be that the system is "ill-conditioned" i.e. the variability of the data is too

large to allow a discrimination on a significant probability level.

Since the macroparametric representations of linear, compartmental models are all of the multi-exponential form:

$$c = \sum_{i=1}^n A_i e^{-\alpha_i t} \quad (A_i, \alpha_i > 0) \quad (9.10)$$

it appears appealing in routine investigations of raw pharmacokinetic data to apply a "multiple regression approach", similar to that used for linear systems, to determine the order,  $n$ , of the system.

This seems to be unreliable, however, for several reasons.

1. There exists no computer program which will inevitably find the "best" solution (smallest residual sum of squares) in a nonlinear least squares estimation which may have several minima.
2. Measurements in biological systems often produce substantial residual values which may give rise to multiple minima.
3. The number of minima will increase very rapidly as the number of exponential terms,  $n$ , to be fitted increases.

The problem of multiple minima can be reduced but not eliminated by a suitable procedure which gives good initial estimates, or by multiple runs with initial parameter values randomly taken from the parameter space, or by performing a lattice search. The interactive structure of FUNFIT makes it particularly suitable for performing multiple runs and lattice searches.

So-called "back-projection" or "stripping" is the technique most frequently used to obtain initial parameter estimates for models of the form described by 9.10. The stripping is either done graphically or automatically by the computer, in some cases employing a spline function representation of the data (152).

However, it is important to realise that this particular technique assumes that one or more exponential terms vanish in certain regions of the total drug level-time curve. In other words the method tends to disregard cases where two or more exponential terms dominate fairly equally throughout the whole time space investigated. Hence, the method may produce biased results. Discrimination between alternative models (9.10) on this basis must therefore be considered unreliable.

It would be appropriate in this connection to refer to a different method which does not introduce such a bias (158). This method is based on a linear shift operator technique which appears not to have been used previously in pharmacokinetic studies. Currently it seems to be the most suitable to use in obtaining initial parameter estimates in linear compartmental models.

In evaluating how well a model describes some data, three points must be considered:

1. How well do the calculated values agree with those observed, i.e. what is the sum of squared residuals or the correlation coefficient?
2. Does the fit agree with the basic assumptions made about the errors?
3. How predictive is the model?

A comparison of fits entirely in terms of sum of squared residuals (such as an F-test) must be considered insufficient.

### Analysis of Residuals

The importance of an analysis of residuals (137) seems to have been completely ignored in most computer programs. The basic assumptions in nonlinear least squares are: 1. The independent variable(s) is without error. 2. The errors,  $\epsilon_i$  in the dependent variable are independent



$(\text{COV}_{i \neq j}(\epsilon_i, \epsilon_j) = 0)$  and normally distributed with zero mean and the same variance ( $\epsilon_i \sim N(0, \sigma^2)$ ).

The theory of least squares predicts that if the model is correct then the *residuals* should reflect the above properties of the *errors*, in an unbiased way, provided the errors possess these properties<sup>3</sup> (140). The assessment of a model on the basis of a residual analysis must rely on the converse principle: If the *residuals* appear to be from the same normal distribution then, under assumption 2, the model should not be rejected.

To investigate whether the residuals are in fact equally normally distributed requires repeated experiments. When only one experiment is available a test must rely on the fact that if the residuals deviate from the same normal distribution, then collectively they will be normally distributed. The converse, however, is not true in general (140). Therefore, in the absence of repeated experiments it is necessary to make the additional assumption that if the residuals are collectively normally distributed then they are individually normally distributed also.

Possibly the best way to examine the residuals is to plot them against the independent and dependent variables (137,138). Significant systematic deviations can be visualized in this way. FUNFIT includes such plots and the following statistics which may be helpful in the assessment and comparison of models.

The Kolmogorov-Smirnov statistic (159) is used to test for normality of the residuals. The procedure is as follows: Given N residuals, the program calculates

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3. This is only strictly true in the linear case but approximately true for the nonlinear case.

$$D = \max_x |F^*(X) - S_N(X)| \quad (9.11)$$

where  $S_N(X)$  is the cumulative distribution of the residuals and  $F^*(X)$  is the cumulative normal distribution function with the same mean and variance as the residual sample. The calculated value of  $D$  is compared with the critical value obtained from a Monte Carlo calculation at a given significance level.

The fundamental assumption of random errors is also tested in FUNFIT using the "run test" and the "number test" as discussed in Chapter 8. The Durbin-Watson statistic to test for serial correlation among the residuals was also presented in that chapter together with the test for "outliers".

If the residual analysis reveals that the residuals do not appear to be significantly random or normally distributed then *this does not necessarily mean that the model is incorrect*. More exactly it means one is faced with the problem of either rejecting the hypothesis that the model is "correct", rejecting the assumption made about the errors, or rejecting the assumption that the computer program has found the "best" solution in the case of multiple sum of squares minima. In the last two cases the model cannot be verified. This clearly emphasizes the need for a computer program which is efficient in finding a global minimum, the need for accurate data to reduce or eliminate multiple minima, and the need for carefully designed experiments which do not introduce systematic or cumulative errors.

#### The predictive power of the model

The ultimate goal in mathematical modelling in pharmacokinetics is to establish models with significant predictive power. A similar goal

exists in modelling of economic systems. The voluminous literature in this area can undoubtedly give inspiration to future approaches in pharmacokinetics.

A very useful test of the predictive capabilities of a model is to test the hypothesis that the parameters do not depend on the model variables. This can be done readily if sufficient data are available.

The total set of data is first partitioned into 2 or more subsets. The parameters are then estimated separately for each subset and the parameter subsets are tested for any trend or for a functional relationship with the independent variable(s) by suitable correlation analysis. A test to establish whether the parameter subsets are significantly different from the parameter set obtained for the whole sample can also be employed (160,161). The highly interactive structure of FUNFIT readily facilitates such partitioning of the data enabling the above tests to be made.

A Comparison of FUNFIT and NONLIN

FUNFIT was applied to obtain parameter estimates of the following simplified 2- and 3-compartment models:

$$c = p_1 e^{-p_2 t} + p_3 e^{-p_4 t} \quad p_i > 0 \quad (9.12)$$

$$c = p_1 e^{-p_2 t} + p_3 e^{-p_4 t} + p_5 e^{-p_6 t} \quad p_i > 0 \quad (9.13)$$

which were used to describe the plasma profile of pancuronium after I.V. bolus injection in 4 human subjects. The data to which 9.12 and 9.13 were fitted are shown in table 9.1.

Identical parameter limits and initial estimates were chosen to those used in applying the 1969 version of NONLIN which appears to be the

most commonly used nonlinear regression program in pharmacokinetic investigations. The stopping criterion for FUNFIT was 0.1 (per cent) which gives approximately the same relative change in the SS value as

**Table 9.1** Pancuronium Bloodlevel Data to which Eqs. 9.12 and 9.13 are fitted using NONLIN and FUNFIT.

No.	I.A.		B.A.		M.C.		J.C.	
	TIME	CONC.	TIME	CONC.	TIME	CONC.	TIME	CONC.
	min	µg/ml	min	µg/ml	min	µg/ml	min	µg/ml
1	7.5	1.120	5.0	.600	5.0	1.033	5.0	1.440
2	10.0	.775	10.0	.556	10.0	.830	10.0	1.000
3	20.0	.545	15.0	.550	15.0	.800	15.0	.945
4	30.0	.510	20.0	.480	20.0	.680	20.0	.805
5	60.0	.395	30.0	.370	30.0	.555	30.0	.620
6	95.0	.416	60.0	.200	60.0	.255	60.0	.463
7	120.0	.166	90.0	.160	91.0	.235	90.0	.365
8	152.0	.200	120.0	.150	120.0	.220	120.0	.355
9	180.0	.200	255.0	.090	240.0	.143	143.0	.270
10	190.0	.168	361.0	.100	413.0	.095	257.0	.160
11	250.0	.130					408.0	.083
12	400.0	.067						

for the two programmes FUNFIT was started with initial parameter values identical to NONLIN's final estimates and with initial parameter step sizes = 0.1% of these parameter values. At the first iteration FUNFIT gave exactly the same SS-value as NONLIN's value at convergence but it did not accept this solution as a stationary point and converged to a significantly different solution (Table 9.2).

most commonly used nonlinear regression program in pharmacokinetic investigations. The stopping criterion for FUNFIT was 0.1 (per cent) which gives approximately the same relative change in the SS value at convergence as NONLIN with its differently defined stopping criterion set at TEST=0.0001. The step size used in NONLIN to approximate derivatives was DEL=0.001; the same value as that used in the test problems given in the NONLIN user's manual. The experimental plasma levels were recorded to 3 significant digits after the decimal point so the precision factor in NONLIN was chosen as IDIG=-9 to avoid significant truncation of calculated values. This NONLIN and FUNFIT should give identical SS values for identical parameter values. This was verified in all runs.

FUNFIT found a different solution than NONLIN in every case where a 3-compartment model (9.13) was fitted, and in half of the cases where a 2-compartment model (9.12) was fitted (Table 9.2). The residual sum of squares values obtained using FUNFIT were substantially lower than those obtained using NONLIN in all cases where there was a difference. The average percentage difference was -55% and -29% when fitting Eqs. 9.13 and 9.12 respectively. The differences were also reflected in the parameter values. Furthermore, the run test indicates that the residuals are more randomly distributed overall in the FUNFIT results.

To test NONLIN's results in the cases where results differed for the two programmes FUNFIT was started with initial parameter values identical to NONLIN's *final* estimates and with initial parameter step sizes = 0.1% of these parameter values. At the first iteration FUNFIT gave exactly the same SS-value as NONLIN's value at convergence but it did not accept this solution as a stationary point and converged to a significantly different solution (Table 9.2).

Table 9.2: Least squares fitting of Eq's 9.12 and 9.13 using NONLIN (N) and FUNFIT (F).

PATIENT /POINTS		SS	$\Delta SS\%$ <sup>(a)</sup>	RUNS <sup>(b)</sup>	P <sub>1</sub>	P <sub>2</sub>	P <sub>3</sub>	P <sub>4</sub>	P <sub>5</sub>	P <sub>6</sub>
IA/12	N	.7131E-1		6	.1452E+1	.2871	.4553	.3022E-1	.3986	.4165E-1
	F	.2410E-1	-66.2	9	.4924E+1	.3026	.4081E-1	.1686	.6013	.6632E-1
	F <sup>(c)</sup>	.2869E-1	-59.8	9	.2940E+1	.2370	.4855E-3	.3568E-1	.6025	.6712E-1
JC/11	N	.1825E-1		8	.1038E+2	.9543	.1139E+1	.7307E-1	.6039	.5048E-1
	F	.6631E-2	-63.7	9	.4649E+1	.5826	.8223	.5242E-1	.5597	.4721E-1
	F <sup>(c)</sup>	.6074E-2	-66.7	9	.1303E+2	.7920	.8369	.5411E-1	.5647	.4744E-1
BA/10	N	.8973E-2		6	.2614	.4341E-1	.3327	.3473E-1	.1697	.1873E-1
	F	.4653E-2	-48.1	6	.5603E-1	.5668	.5911	.2526E-1	.1041	.2758E-1
	F <sup>(c)</sup>	.4447E-2	-50.4	6	.7253E-1	.2467E-1	.5253	.2478E-1	.9702E-1	.3492E-1
MC/10	N	.1382E-1		4	.8224	.4488E-1	.7388E-1	.6326E-1	.3148	.2521E-1
	F	.7844E-2	-43.2	6	.1658E+1	.9200	.9374	.3623E-1	.2283	.1936E-1
	F <sup>(c)</sup>	.8346E-2	-39.6	6	.8740	.3600E-1	.818E-1	.6357E-1	.2336	.2035E-1
IA/12	N	.5102E-1		5	.1983E+1	.2367	.6586	.7449E-2		
	F	.2199E-1	-55.9	8	.1066E+2	.3999	.6202	.6858E-2		
	F <sup>(c)</sup>	.2199E-1	-55.9	9	.1049E+2	.3978	.6198	.6853E-2		
JC/11	N	.1621E-1		7	.1240E+1	.9424E-1	.6496	.5616E-2		
	F	.1621E-1	0	7	.1240E+1	.9425E-1	.6497	.5616E-2		
BA/10	N	.4528E-2		6	.5998	.2569E-1	.1012	.2100E-3		
	F	.4446E-2	- 1.8	6	.5975	.2470E-1	.9677E-1	.5866E-5		
	F <sup>(c)</sup>	.4449E-2	- 1.8	6	.5962	.2484	.9835E-1	.5402E-4		
MC/10	N	.8194E-2		6	.9464	.3760E-1	.2373	.2093E-2		
	F	.8194E-2	0	6	.9464	.3761E-1	.2374	.2095E-2		

(a)  $\Delta SS\% = 100(SS_{\text{FUNFIT}} - SS_{\text{NONLIN}}) / SS_{\text{NONLIN}}$

(b) See text for definition of runs.

(c) FUNFIT was in these cases started using NONLINS final parameter estimate as initial estimates and with initial parameter step sizes = 0.1% of these parameter values.

$$|\delta_i| \approx \left| \frac{\epsilon}{p_{i-1} u_{ii}} \right|$$

where  $\epsilon$  is the relative error in the computed  $f$ -value and  $u_{ii}$  is the  $i$ -th diagonal element of the Hessian matrix. This formula shows that the optimal

The detailed minimization report chosen in the investigation of this phenomenon, in fact, showed that the SS-function in NONLIN's convergence region had a significant gradient value indicating that NONLIN's solutions in these cases were not sufficiently close to the true sum of squares minimum.

The most likely reason for the failure of NONLIN to find a satisfactory solution in these cases appears to result from substantial errors in the approximation of derivatives.

The derivative with respect to the  $i$ -th parameter,  $p_i$ , of the function  $f(\underline{X}, \underline{P})$  to be fitted in NONLIN is approximated by a one-sided difference formula:

$$\frac{\partial f}{\partial p_i} \approx \frac{f(\underline{X}, p_1, p_2, \dots, p_i + \delta p_i, \dots, p_n) - f(\underline{X}, \underline{P})}{\delta p_i} \quad (9.14)$$

where the step size  $\delta$ , (DEL) is chosen by the user. The value of this quantity is critical to the accuracy of the derivative. In choosing a proper value for  $\delta$ , one has to steer between two hazards: 1. If the value chosen for  $\delta$  is too *small* the derivative will be substantially inaccurate because of the *rounding error* which arises when the two  $f$ -values in (9.14) are too close. 2. If  $\delta$  is set too *large* the derivative approximation will be too inaccurate because of the *truncation error* (9.14 is only accurate in the limit as  $\delta \rightarrow 0$ ). This indicates that there must be an optimal value for  $\delta$ . It can be shown that this value, for the  $i$ -th parameter, is approximately given by:

$$|\delta_i| \approx \left| \frac{4 \epsilon f}{P_i H_{ii}} \right| \quad (9.15)$$

where  $\epsilon$  is the relative error in the computed  $f$ -value and  $H_{ii}$  is the  $i$ -th diagonal element of the Hessian matrix. This formula shows that the optimal

$\delta$ -value differs from parameter to parameter. It is not uncommon to find a very large value for the ratio,  $\max(p_{i,ii})/\min(p_{i,ii})$ , indicating that the choice of a *single* common  $\delta$ -value as is done in NONLIN may not be adequate for all derivative evaluations and may cause convergence to a non-stationary point (147).

The above problem can be overcome in several ways:

1. By abolishing difference approximations and using exact analytical derivatives. This, however, may be of considerable inconvenience for the user who must define the analytical derivatives. It also limits the use of the program to equations for which analytical derivatives can be obtained.
2. By modifying the initial choice of  $\delta$  according to Eq. 19 or by other means (162). However, even if alterations are made according to 1. or 2. the Gauss-Newton methods may still converge in some cases to a point at which the gradient does not vanish (145).
3. By abolishing the linearization approach in the Gauss-Newton methods and using a general function minimization approach (163-165 ).
4. By the use of an algorithm which is not based on derivatives or derivative approximations as is done in FUNFIT. The disadvantage of the last approach is that more function evaluations are required to reach convergence. For most pharmacokinetic applications this disadvantage is not significant. However, in cases where many parameters (> about 12) are to be estimated or where the equation(s) to be fitted is very time-consuming to evaluate (for example in the fitting of a functional relationship described by a system of differential equations) the disadvantage may become significant.

The presence of multiple SS-minima in fitting the 3-compartment model is evident from the fact that in the cases where FUNFIT was started



with NONLIN's final estimate it converged on a different solution (Table 9.2). The difference between SS-values found by FUNFIT in consecutive runs was much smaller than the difference between NONLIN and FUNFIT's values.

To test for multiple minima in fitting the 2-compartment model to IA's data, FUNFIT was started randomly 10 times in the chosen parameter space. In 9 of these cases it found the same solution (Table 9.2) but in one case it converged to:

$$SS = .2209E-1 \text{ and } p_{1-4} = .8780E+1, .3744, .6162, .6797E-2.$$

It is encouraging that this minimum is larger than that found in the 9+2 other cases. The more frequent occurrence of different solutions in the 3-compartment fitting confirms that the problem of multiple minima increases with an increasing number of parameters.

In only about half the cases investigated did FUNFIT and NONLIN find a smaller SS value for the 3-compartment model than for the 2-compartment model. This clearly emphasises the problems in discriminating between nonlinear mathematical models as discussed.

If, in fitting linear compartmental models, 9.10, the lower limits for the coefficient parameters  $A_1$  are set to zero then, in theory, the fit in terms of SS of a higher order model (e.g.  $n=3$  vs  $n=2$ ) should always be better or at least as good as the fit of a lower order model.<sup>4</sup> Therefore, if under such conditions it is found that the higher order model does not improve the fit (SS) then there is reason to believe a better minimum exists. In such cases, the higher order model should be refitted with initial estimates of the common parameters equal to the final parameter estimates of the lower order models. If this procedure also fails to give a lower SS-value, then a third run should be made where the common parameters

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4. Provided of course that the parameter space for the lower order model is a subset of that of the higher order model.

are restricted within narrow limits around the optimal values for the lower order model, and where the parameters in the higher order term (e.g.  $p_5$  and  $p_6$  above) are much less restricted. The above approach will be successful in most cases and its use is recommended not only for fitting to sums of exponentials but also for any other "order system" where the terms are allowed to vanish. It should reduce significantly the problems associated with multiple minima and the problems of finding suitable initial estimates.

### Truncation

In the NONLIN program it is possible by using the IDIG parameter to specify various degrees of truncation of the calculated values for the dependent variable. For example, if IDIG is set at -3 then all calculated values of the dependent variable will be truncated to 3 significant digits after the decimal point. The philosophy behind the use of this parameter is that there is no reason to calculate the predicted values to any higher precision than the observed values. In adopting such a philosophy it must be realised that the results so obtained will be specific to the NONLIN program and in general cannot be compared with results obtained using other nonlinear regression programs. The difference between results obtained specifying virtually no truncation (IDIG= -9) and specifying truncation to the precision of the observations (IDIG= -3) was found to be very pronounced (Table 9.3). The substantial difference was reflected not only in the SS-values and parameter values but also in the randomness of the residuals.

Truncation (IDIG= -3), furthermore, strongly affects the errors in the derivative approximations since the value of  $\epsilon$  and hence,  $\delta$  in Eq. 9.15 will be affected. This may explain why NONLIN in one case (Table 9.3)

Table 9.3: Least squares fitting of Eq's 9.12 and 9.13 using NONLIN with precision factor -3 and -9.

PATIENT /POINTS	PRECISION- FACTOR	SS	$\Delta$ SS%  (a)	RUNS (b)	P <sub>1</sub>	P <sub>2</sub>	P <sub>3</sub>	P <sub>4</sub>	P <sub>5</sub>	P <sub>6</sub>
IA/12	-9	.7131E-1	48.7	6	.1452E+1	.2871	.4553	.3022E-1	.3986	.4165E-2
	-3	.3658E-1		8	.1793E+1	.1807	.3155	.1097E-1	.2949	.3697E-2
JC/11	-9	.1825E-1	35.2	8	.1028E+2	.9543	.1139E+1	.7307E-1	.6039	.5048E-2
	-3	.1183E-1		6	.112E+1	.1813	.5571	.3310E-1	.5060	.4237E-2
BA/10	-9	.8973E-2	68.8	6	.2614	.4341E-1	.3327	.3473E-1	.1697	.1873E-2
	-3	.1515E-1		4	.1396	.5172E-1	.4649	.4265E-1	.1734	.1953E-2
MC/10	-9	.1382E-1	582 (c)	4	.8224	.4488E-1	.7388E-1	.6326E-1	.3148	.2521E-2
	-3	.9427E-1		3	.8000	.6000	.3000	.1000	.4000E-1	.2000E-2
IA/12	-9	.5102E-1	74.9	5	.1983E+1	.2367	.6586	.7449E-2		
	-3	.8924E-1		4	.8795	.4713E-1	.3312	.2706E-2		
JC/11	-9	.1621E-1	165	7	.1240E+1	.9424E-1	.6496	.5616E-2		
	-3	.4306E-1		4	.1114E+1	.4835E-1	.4251	.2649E-2		
BH/10	-9	.4528E-2	17.1	6	.5998	.2569E-1	.1012	.2100E-3		
	-3	.5303E-2		6	.5547	.3019E-1	.1610	.2121E-2		
MC/10	-9	.8194E-2	2.78	6	.9464	.3760E-1	.2373	.2093E-2		
	-3	.8422		5	.9226	.4107E-1	.2760	.2779E-2		

(a)  $|\Delta$ SS%| =  $100 \times |SS_{(-9)} - SS_{(-3)}| / SS_{(-9)}$

(b) See text for definition of 'runs'.

(c) NONLIN failed to find a proper solution. At convergence after 5 iterations the final parameter estimates were identical to the initial estimates.

failed to converge properly.

The relative precision with which the parameter can be calculated will also be affected by truncation since it will not be possible to improve their estimates further when the computer has reduced the residual sum of squares to a value, SS, for which

$$\frac{SS}{SS(\text{true})} < 1 + \epsilon \quad (9.16)$$

where the error,  $\epsilon$ , depends on the degree of truncation chosen by IDIG (149). Furthermore the truncation procedure cannot be justified on a statistical basis since the theory of least squares assumes no errors in the calculated values. In fact great effort is often made to program the function to be fitted so that it can be evaluated with minimum errors to avoid biased results.

The user of the program NONLIN is, therefore, advised to use a value for the precision factor IDIG such that minimum truncation takes place.

The many nonlinear regression programs available have provided the scientist with a powerful tool useful for a great variety of problems. However, the results obtained have too often been accepted and used without an awareness of the limitations and possible unreliability of the program used, ignoring the numerical problems involved.

The complex structure of the program used has often resulted in an authoritative attitude which may go so far as using the program as a substitute for rational thought. There is a definite need for programs which are more reliable and which allow greater interaction between the user and the program, with the user in a more dominant role.

Estimation CHAPTER 10 provides a means of describing

dissolution characteristics of drug powders apparently more accurately than

previous approaches. The method should be of interest in the quality

control of drugs likely to cause bioavailability problems because of

dissolution rate limited absorption.

Chapter 7 demonstrated good agreement between the theoretical dissolution profile and experimental data for tolbutamide. The intrinsic dissolution profile was calculated, using the rigorous mathematical approach present in Chapter 5 and 6, from optical analysis of the size distribution of the 60/85 mesh fraction powder. Such an analysis is complicated to perform for micronized powders because of the highly irregular particle shapes and the degree of aggregation often found in such powders.

However it should be possible to determine by nonlinear regression analysis whether the dissolution of such a powder can be adequately described by one of the multiparticulate dissolution models presented earlier.

In this chapter various such models, based on a log-normal particle size distribution, are fitted by nonlinear least squares regression to data from the dissolution of micronized glibenclamide using the FUNFIT program. Estimates of parameters describing the *effective* initial particle size distribution are obtained together with estimates of a quantity defined as the *specific dissolution rate parameter*. A dissolution equation based on an ideal, untruncated log-normal distribution with the single particles dissolving according to the cube root law best describes the dissolution kinetics. Dissolution behaviour of glibenclamide can be well described by this model in terms of the *specific dissolution rate parameter* and one other parameter which accounts for the distribution effect termed the *dispersion parameter*.

Estimation of these two parameters provides a means of describing dissolution characteristics of drug powders apparently more correctly than previous approaches. The method should be of interest in the quality control of drugs likely to cause bioavailability problems because of dissolution rate limited absorption.

Theoretical

The dissolution equation for a log-normal powder, considering the cube root and the square root model, was given earlier by Eq. 6.21. This equation appears to contain 5 parameters, namely  $\sigma$ ,  $i$ ,  $j$ ,  $\mu$  and  $K_m$  which define the dissolution profile,  $W/W_0$  versus time. However, an attempt to obtain least squares estimates of all these 5 parameters from  $W/W_0$  vs. time data may fail because  $\mu$  and  $K_m$  can be fused into a single parameter:

$$K_m^* = e^{-\frac{3}{m} \mu} K_m \tag{10.1}$$

which will be called the *specific dissolution rate parameter*.

The uniqueness of these 4 parameters,  $\sigma$ ,  $i$ ,  $j$  and  $K_m^*$ , in defining the dissolution profile can be seen by substituting  $K_m = e^{\frac{3}{m} \mu} K_m^*$  into Eq. 10.1 resulting in total cancellation of  $\mu$ , in full agreement with the theory discussed previously:

$$\frac{W}{W_0} = \sum_{n=0}^m \binom{m}{n} (-K_m^* t)^{(m-n)} \frac{F(j - \frac{3n\sigma}{m}) - A}{F(j - 3\sigma) - F(-i - 3\sigma)} e^{\frac{9}{2} \left[ \frac{n}{m} \right]^2 \sigma^2} \tag{10.2}$$

where  $A = F(-i - \frac{3n\sigma}{m})$  for  $t < \frac{1}{K_m^*} e^{-\frac{3i\sigma}{m}}$  (10.3)

and  $A = F(\frac{m}{3\sigma} \ln(K_m^* t) - \frac{3n\sigma}{m})$  for  $\frac{1}{K_m^*} e^{\frac{3j\sigma}{m}} > t > \frac{1}{K_m^*} e^{-\frac{3i\sigma}{m}}$  (10.4)

and 
$$\frac{W}{W_0} = 0 \quad \text{for} \quad t \geq \frac{1}{K_m^*} e^{\frac{3j\sigma}{m}} \quad (10.5)$$

The continuous flow, recording apparatus used provides dissolution rate data. The fraction undissolved,  $W/W_0$ , versus time can be obtained by integrating. The parameters  $\sigma$ ,  $i$ ,  $j$  and  $K_m^*$  can then be estimated by nonlinear least squares regression analysis using Eq. 10.2. However the integrated data will contain integration errors. The integration also tends to "smooth" the original data so estimates of the variability of the parameters will be less reliable than if the original rate data were used. It is therefore useful to derive an expression for the release rate in order to estimate the 4 parameters directly from the original (rate) data.

By applying Eq. 5.33 of Chapter 5 the following equation is obtained:

$$\frac{W}{W_0} = \frac{\int_{R_1}^{R_2} [x^{\frac{3}{m}} - K_m^* t]^m x^{-1} N(\ln x, \mu, \sigma) dx}{\int_{d_0}^D x^2 N(\ln x, \mu, \sigma) dx} \quad (10.6)$$

where  $d_0 = e^{\mu - i\sigma} \quad (10.7)$

$D_0 = e^{\mu + j\sigma} \quad (10.8)$

$R_1 = (\max [K_m^* t, d_0^{\frac{3}{m}}])^{\frac{m}{3}} \quad (10.9)$

$R_2 = (\max [K_m^* t, D_0^{\frac{3}{m}}])^{\frac{m}{3}} \quad (10.10)$

An expression for the dissolution rate of drug,  $Q = -dw/dt$  can now be found by differentiating Eq. 10.6 with respect to time (using Leibnitz's rule). For abbreviation, let B denote the (constant) denominator of Eq. 10.6; differentiation then gives:

$$\frac{B}{W_0} Q = mK_m \int_{R_1}^{R_2} (x^{\frac{3}{m}-K_m t})^{(m-1)} x^{-1} N(\ln x, \mu, \sigma) dx \quad (10.11)$$

$$- (R_2^{\frac{3}{m}-K_m t})^m R_2^{-1} N(\ln R_2, \mu, \sigma) \frac{dR_2}{dt}$$

$$+ (R_1^{\frac{3}{m}-K_m t})^m R_1^{-1} N(\ln R_1, \mu, \sigma) \frac{dR_1}{dt} \quad (10.12)$$

Before critical time<sup>1</sup>  $R_1 = d_0$ ,  $R_2 = D_0$  so  $dR_1/dt = dR_2/dt = 0$ ,

and the last two terms of the right-hand side of Eq. 10.11 vanish. After the critical time,  $dR_2/dt$  is still zero but  $R_1 = (K_m t)^{\frac{m}{3}}$  (according to 10.9), thus the last term also vanishes because then:

$$R_1^{\frac{3}{m}-K_m t} = ((K_m t)^{\frac{m}{3}})^{\frac{3}{m}-K_m t} = 0$$

Equation 10.11 can therefore be simplified to:

$$Q = mK_m W_0 \frac{\int_{R_1}^{R_2} (x^{\frac{3}{m}-K_m t})^{(m-1)} x^{-1} N(\ln x, \mu, \sigma) dx}{\int_{d_0}^{D_0} x^2 N(\ln x, \mu, \sigma) dx} \quad (10.12)$$

The term  $(x^{\frac{3}{m}-K_m t})^{(m-1)} x^{-1}$  under the integral sign can be expanded to:

$$x - 2K_3 t + (K_3 t)^2 x^{-1} \quad (\text{for } m=3) \quad (10.13)$$

$$x^{\frac{1}{2}} - K_2 t x^{-1} \quad (\text{for } m=2)$$

and the formula given previously (5.34) can be applied to express the integrals in 10.11 in terms of the function F. This leads to the following

1. The critical time is the time when the first particles begin to disappear in the dissolution process, i.e.

$$t = \frac{1}{K_m} e^{\frac{3}{m}(\mu-i\sigma)}$$



expression for  $Q$  that can be evaluated more readily and exactly than 10.12:

$$Q = m K_m W_o \sum_{n=0}^{(m-1)} \binom{m-1}{n} (-K_m t)^{(m-n-1)} \frac{F\left(\frac{T_2 - \mu}{\sigma} - \frac{3n\sigma}{m}\right) - F\left(\frac{T_1 - \mu}{\sigma} - \frac{3n\sigma}{m}\right)}{F(j-3\sigma) - F(-i-3\sigma)} \times$$

$$e^{\frac{3}{m(n-m)} \left(\mu + \frac{3}{2m} (n+m)\sigma^2\right)} \quad (10.14)$$

where  $T_1$  and  $T_2$  are defined:

$$T_1 = \max\left[\frac{m}{3} \ln(Kt), \mu - i\sigma\right] \quad (10.15)$$

$$T_2 = \max\left[\frac{m}{3} \ln(Kt), \mu + j\sigma\right] \quad (10.16)$$

This expression for the dissolution rate  $Q$  contains 5 parameters ( $\sigma, i, j, \mu$  and  $K_m$ ). However, as before only 4 parameters ( $\sigma, i, j$  and  $K_m^*$ ) are needed to define uniquely the dissolution rate profile:

$$Q = m K_m^* W_o \sum_{n=0}^{(m-1)} \binom{m-1}{n} (-K_m^* t)^{(m-n-1)} \frac{F\left(j - \frac{3n\sigma}{m}\right) - A}{F(j-3\sigma) - F(-i-3\sigma)} \times$$

$$e^{\frac{9}{2} \left[\left(\frac{n}{m}\right)^2 - 1\right] \sigma^2} \quad (10.17)$$

This equation is the differential form of Eq. 10.2. The quantity  $A$  is defined as previously (10.3 and 10.4) and

$$Q = 0 \text{ for } t \geq \frac{1}{K_m^*} e^{\frac{3j\sigma}{m}}$$

The above expression is of considerable value since it allows the effective initial particle size distribution parameters  $\sigma, i$  and  $j$  to be determined together with the specific dissolution rate parameter,  $K_m^*$ , by nonlinear regression analysis of the dissolution rate profile ( $dW/dt$  versus time).

It is also of interest to investigate how well the dissolution behaviour can be described if the particle size distribution is considered

ideal, i.e. if  $i=j=\infty$ .

Noting that  $F(\infty) = 1$  and  $F(-\infty) = 0$ , Eq. 10.17 becomes, for an ideal distribution:

$$Q = m K_m^* W_o \sum_{n=0}^{(m-1)} \binom{m-1}{n} (-K_m^* t)^{(m-n-1)} \left[ 1 - F\left(\frac{m}{3\sigma} \ln(K_m^* t) - \frac{3n\sigma}{m}\right) \right] X \tag{10.18}$$

which,  $\frac{9}{2} \left[ \left(\frac{n}{m}\right)^2 - 1 \right] \sigma^2$

If the distribution is considered ideal at the lower end ( $i=\infty$ ) but truncated at the higher end, the expression becomes:

$$Q = m K_m^* W_o \sum_{n=0}^{(m-1)} \binom{m-1}{n} (-K_m^* t)^{(m-n-1)} \frac{F\left(j - \frac{3n\sigma}{m}\right) - F\left(\frac{m}{3\sigma} \ln(K_m^* t) - \frac{3n\sigma}{m}\right)}{F(j - 3\sigma)} X \tag{10.19}$$

$\frac{9}{2} \left[ \left(\frac{n}{m}\right)^2 - 1 \right] \sigma^2$

and if it is considered truncated at the lower end but not at the higher end ( $j=\infty$ ):

$$Q = m K_m^* W_o \sum_{n=0}^{(m-1)} \binom{m-1}{n} (-K_m^* t)^{(m-n-1)} \frac{1-A}{1-F(-i-3\sigma)} e^{\frac{9}{2} \left[ \left(\frac{n}{m}\right)^2 - 1 \right] \sigma^2} \tag{10.20}$$

Monodisperse powder

It is of interest to look at the limiting case where the particle size distribution is infinitely narrow, i.e.  $\sigma=0$ , since this case allows a better understanding of the specific dissolution rate parameter,  $K_m^*$ . It also provides a method of obtaining a suitable initial estimate of this parameter to use in nonlinear curve fitting.

When  $\sigma=i=j=0$ , Eq. 10.17 becomes:

A comparison of 10.27 and 10.23 shows that for a monodisperse powder the specific dissolution rate parameter,  $K_m^*$ , is common to both multiparticle

$$Q = m K_m^* W_o \sum_{n=0}^{(m-1)} \binom{m-1}{n} (-K_m^* t)^{(m-n-1)} \quad (10.21)$$

Equation 10.22 can be written which can be written more simply as:

$$Q = - \frac{dW}{dt} = m K_m^* W_o (1 - K_m^* t)^{(m-1)} \quad (10.22)$$

This equation can be used to obtain an initial estimate of  $K_m^*$  from the linear regression of  $Q$  on  $t$ , using dissolution rate data. The intercept value, which, after integration, can be written:

$$\left(\frac{W}{W_o}\right)^{\frac{1}{m}} = 1 - K_m^* t \quad (10.23)$$

As expected, since there is no size distribution effect this equation predicts that the powder will dissolve strictly according to the cube root or square root model.

When the powder is monodisperse  $\mu = \ln a_o$  and Eq. 10.1 becomes:

$$K_m^* = e^{-\frac{3}{m} \ln a_o} K_m = a_o^{-\frac{3}{m}} K_m \quad (10.24)$$

For spherical particles  $K_m = (6/\rho\pi)^{\frac{1}{m}} k_m$  where  $k_m$  is the rate parameter in the single particle dissolution model:

$$\frac{1}{w_m} = \frac{1}{w_o} - k_m t \quad (10.25)$$

thus 10.24 becomes:

$$K_m^* = a_o^{-\frac{3}{m}} (6/\rho\pi)^{\frac{1}{m}} k_m = w_o^{-\frac{1}{m}} k_m \quad (10.26)$$

When  $k_m = w_o^{\frac{1}{m}} K_m^*$ , from this expression is inserted in 10.25 the single particle dissolution model can be written:

$$\left(\frac{w}{w_o}\right)^{\frac{1}{m}} = 1 - K_m^* t \quad (10.27)$$

A comparison of 10.27 and 10.23 shows that for a monodisperse powder the specific dissolution rate parameter,  $K_m^*$ , is common to both multiparticulate

(10.23) and single particle dissolution (10.27). For both  $m=2$  and 3 it has the dimension of time<sup>-1</sup>.

Equation 10.22 can be written:

$$\frac{1}{Q^{m-1}} = (mW_o K_m^*)^{\frac{1}{m-1}} - (mW_o)^{\frac{1}{m-1}} (K_m^*)^{\frac{m}{m-1}} t \quad (10.28)$$

This equation can be used to obtain an initial estimate of  $K_m^*$  from the linear regression of  $\frac{1}{Q^{m-1}}$  on  $t$ , using dissolution rate data. The intercept value,  $(mW_o K_m^*)^{\frac{1}{m-1}}$ , divided by  $(mW_o)^{\frac{1}{m-1}}$  gives  $K_m^{*\frac{1}{m-1}}$ . (Fig. 10.1)

For a monodisperse powder 10.28 predicts a linear relationship between  $\sqrt{Q}$  and  $t$  when the single particles dissolve according to the cube root model ( $m=3$ ) and a linear relationship between  $Q$  and  $t$  when they dissolve according to the square root model ( $m=2$ ).

Significant deviations from linearity were observed when dissolution rate data for micronized glibenclamide were plotted in either of these ways (Figs. 10.1-10.5). Such deviations can arise if the powder is not monodisperse or if the single particles do not dissolve according to the single particle model given by 10.25.

Under an electron microscope the micronized glibenclamide used appears to be quite polydisperse. The observed deviation from linearity in the rate plots can thus be explained as a particle size distribution effect, assuming 10.25 to be valid. However, the rate data can also be explained by other single particle dissolution models in combination with a size distribution effect. Conclusions about the validity of a single particle model can only be made when the size distribution effect can be taken into account.

Chapter 7 dealing with the dissolution of 60/85, mesh fraction tolbutamide in relation to its particle size distribution indicated that

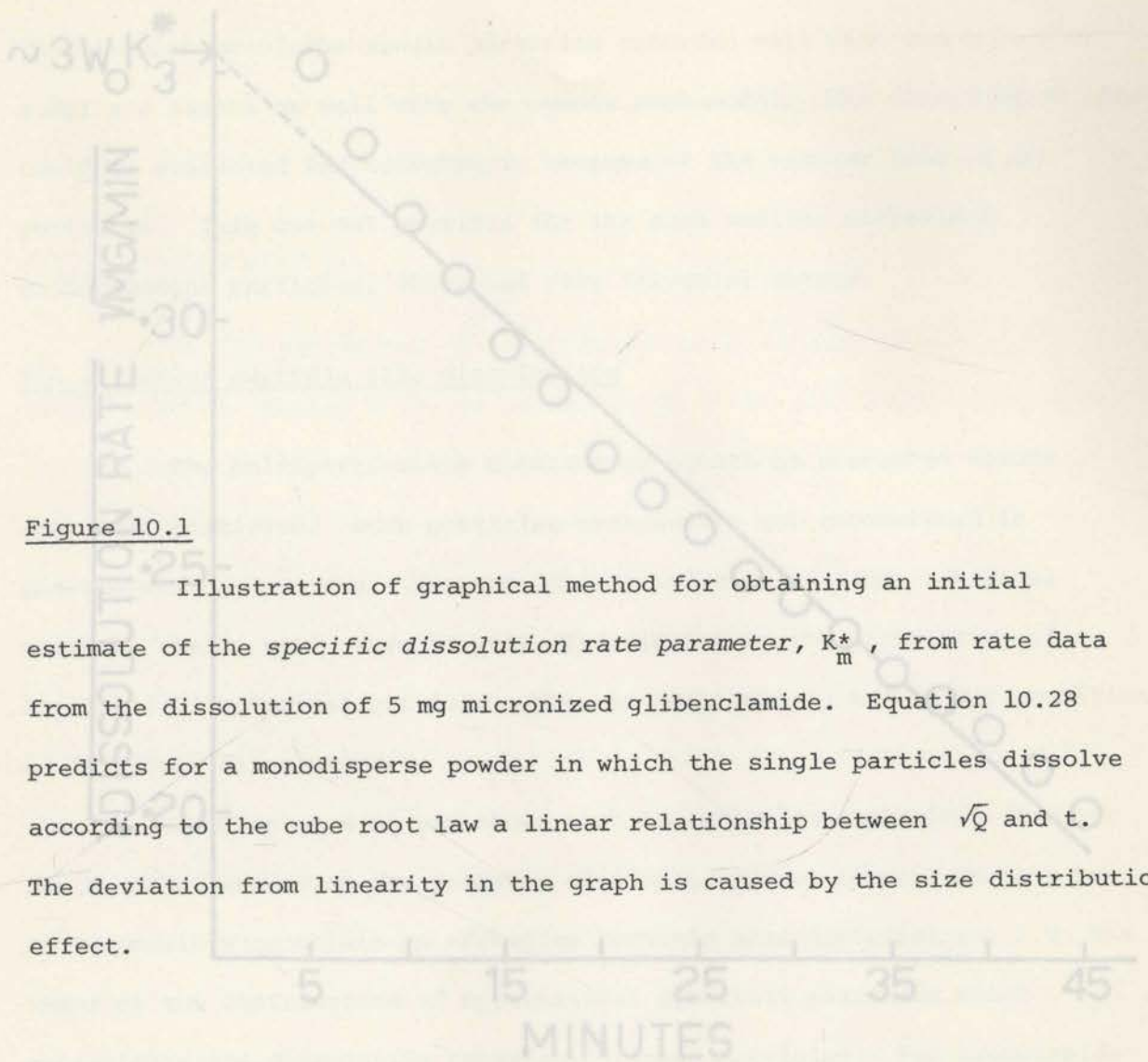
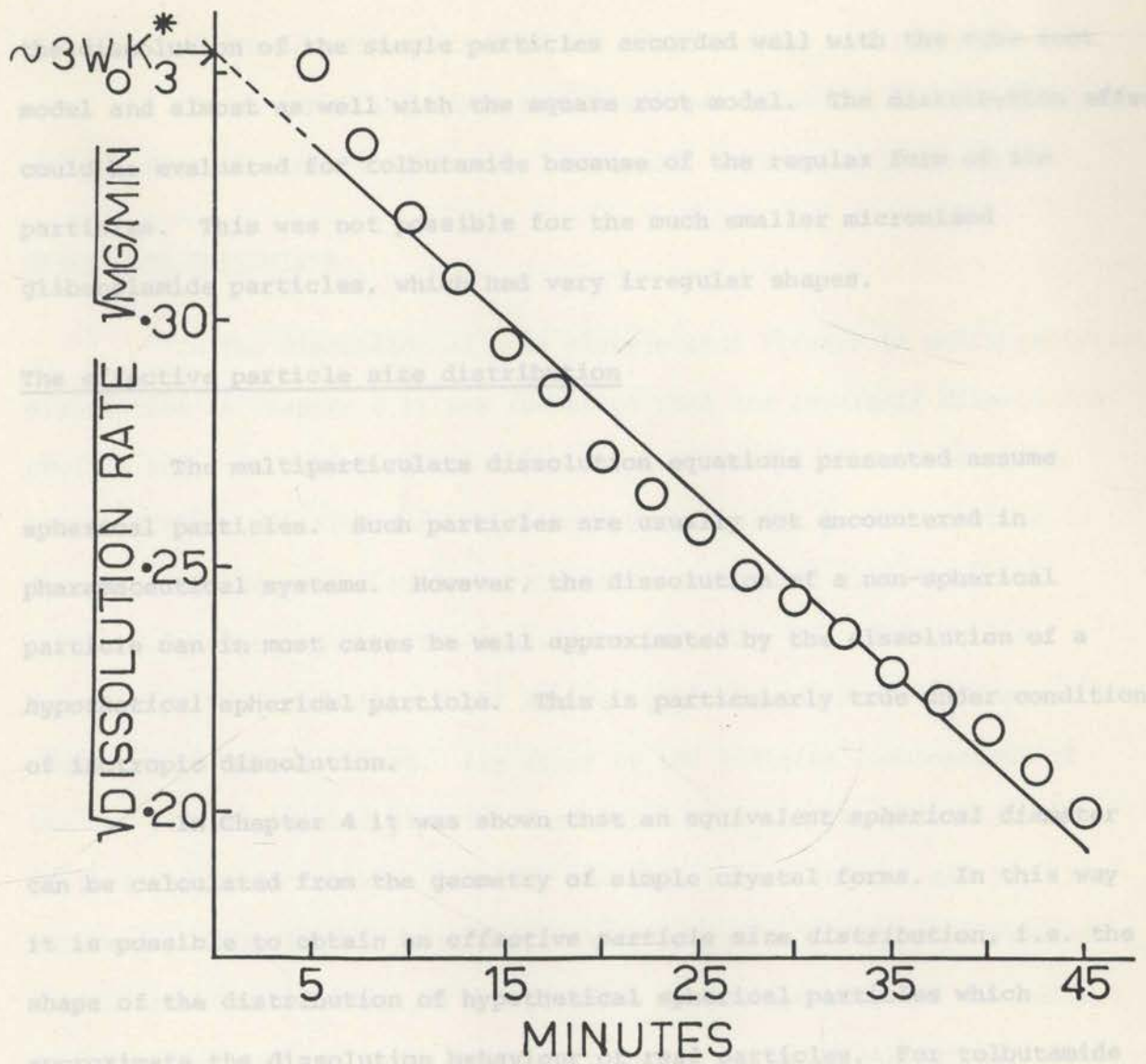


Figure 10.1

Illustration of graphical method for obtaining an initial estimate of the *specific dissolution rate parameter*,  $K_m^*$ , from rate data from the dissolution of 5 mg micronized glibenclamide. Equation 10.28 predicts for a monodisperse powder in which the single particles dissolve according to the cube root law a linear relationship between  $\sqrt{t}$  and  $t$ . The deviation from linearity in the graph is caused by the size distribution effect.



the dissolution of the single particles accorded well with the cube root model and almost as well with the square root model. The distribution effect could be evaluated for tolbutamide because of the regular form of its particles. This was not possible for the much smaller micronized glibenclamide particles, which had very irregular shapes.

#### The effective particle size distribution

The multiparticulate dissolution equations presented assume spherical particles. Such particles are usually not encountered in pharmaceutical systems. However, the dissolution of a non-spherical particle can in most cases be well approximated by the dissolution of a hypothetical spherical particle. This is particularly true under conditions of isotropic dissolution.

In Chapter 4 it was shown that an equivalent spherical diameter can be calculated from the geometry of simple crystal forms. In this way it is possible to obtain an effective particle size distribution, i.e. the shape of the distribution of hypothetical spherical particles which approximate the dissolution behaviour of real particles. For tolbutamide it was found that the effective distribution was approximately log-normal. Because of the irregular particle shapes it was not possible to make a *a priori* conclusions about the effective particle size distribution for glibenclamide. In these investigations the distribution is assumed to be log-normal, consistent with the results in Chapter 7 and the fact that powders are often found to have a log-normal particle size distribution (122,166). The distribution in Fig.5.1 therefore illustrates a log-normal approximation to the effective particle size distribution. The initial particle diameters are the diameters of the hypothetical spherical particles

that approximate the dissolution of the real nonspherical particles.

If the lower and upper truncation parameters,  $i$  or  $j$ , are finite the log-distribution is said to be truncated; otherwise it is ideal.

#### Regression parameters

In the discussion of size distribution effects in multiparticulate dissolution in Chapter 6 it was indicated that the *intrinsic dissolution profile* does not depend on the actual size of the particles but on the shape of their distribution. For the same reason it is not possible to determine by regression analysis the scale parameter,  $\mu$ , as might be erroneously expected from the appearance of 6.21. If, in fact, this equation is used to obtain least squares estimates of  $\mu$  and  $K_m^*$ , these would not be unique values. Any other of the infinite combinations of the two parameters that give the same value of  $K_m^*$  (10.1) will according to 10.2 result in the same fit (for the same values of  $\sigma$ ,  $i$  and  $j$ ). The regression analysis can thus only provide estimates of the *dimensionless* distribution parameters  $\sigma$ ,  $i$  and  $j$ , which define the *shape* of the initial distribution, and  $K_m^*$ . The scale or position of the distribution, given by  $\mu$ , is hidden in the specific dissolution rate parameter,  $K_m^*$ .

#### Curvefitting

Equation 10.17 was fitted by least squares to dissolution rate data from dissolution of 5 and 10 mg micronized glibenclamide using FUNFIT. The kinetic models given by 10.17-10.19 are defined in schemes 10.1-10.3

are present in FUNFIT as function routines named NRC and SUP respectively. The "model parameter",  $m$ , can be set at 3 or 2, depending on whether a model based on the cube- or square root model is to be fitted. The use of  $m$  is of great convenience since a fitting to both models can quickly be done with a single input of dissolution data.



respectively.<sup>2</sup>

The estimates obtained for the truncation parameters,  $i$  and  $j$  (Table 10.1, Eq. 10.17), were all larger than 2 and in most cases exceeded 4, indicating that the effective initial particle size distribution was close to ideal. It was shown in Chapter 5 that the effect on the dissolution profile of the lower truncation parameter,  $i$ , is negligible. Simulation studies also show that the influence of an increase in the upper truncation parameter,  $j$ , becomes insignificant when  $j$  is larger than 2. It is therefore expected from the values of  $i$  and  $j$  obtained using 10.17 that the simpler model, 10.18, which assumes an ideal distribution ( $i=j=\infty$ ) should fit the same data nearly as well. The values in table 10.1 and the curves fitted in Figs. 10.2-10.5 confirm this expectation. There does not seem to be any significant difference in either the  $K_m^*$ , the  $\sigma$  or the  $r$ -values for the two models. The residual plots in Figs. 10.2, 10.3 and Figs. 10.4, 10.5 also seem to be very similar. Equation 10.19 (upper truncation) and 10.20 (lower truncation) also gave similar results (and therefore, these have not been included).

#### Choice of the mathematical model

In agreement with the general principles of mathematical modelling, Eq. 10.17, should be considered as the model that best describes the dissolution of the micronized glibenclamide. This is because it is the

- 
2. The user of FUNFIT does not need to specify the binomial coefficients or the cumulative standard normal distribution function,  $F$ , since these are present in FUNFIT as function routines named NBC and SDF respectively. The "model parameter",  $m$ , can be set at 3 or 2, depending on whether a model based on the cube- or square root model is to be fitted. The use of  $m$  is of great convenience since a fitting to both models can quickly be done with a single input of dissolution data.

TABLE 10.1 Least squares estimates of rate- and distribution parameters obtained from nonlinear regression analysis of data from the dissolution of micronized glibenclamide, considering various models for multiparticulate dissolution kinetics.<sup>(a)</sup>

$W_0$ (mg)	SQUARE ROOT MODEL (m=2)			CUBE ROOT MODEL (m=3)			
	Eq.10.18 <sup>(b)</sup> Scheme 10.2	Eq.10.19 <sup>(c)</sup> Scheme 10.3	Eq.10.17 <sup>(d)</sup> Scheme 10.1	Eq.10.18 <sup>(b)</sup> Scheme 10.2	Eq.10.19 <sup>(c)</sup> Scheme 10.3	Eq.10.17 <sup>(d)</sup> Scheme 10.1	
5	$K_m^*$ (min <sup>-1</sup> )	0.02858 (6.39)	0.02858 (5.12)	0.02859 (6.51)	0.02367 (4.55)	0.02367 (4.44)	0.02449 (3.96)
	$\sigma$	0.4759 (2.93)	0.4760 (2.70)	0.4760 (3.28)	0.6184 (2.47)	0.6184 (2.11)	0.6315 (1.71)
	i	$\infty$	$\infty$	5.136	$\infty$	$\infty$	2.364
	j	$\infty$	5.299	5.655	$\infty$	9.807	4.172
	$r^{(e)}$	0.9935	0.9935	0.9935	0.9952	0.9952	0.9954
10	$K_m^*$ (min <sup>-1</sup> )	0.02633 (7.18)	0.02633 (10.6)	0.02633 (9.35)	0.02147 (9.45)	0.02147 (4.74)	0.02176 (4.03)
	$\sigma$	0.4955 (3.20)	0.4956 (7.24)	0.4955 (5.14)	0.6374 (3.29)	0.6374 (2.03)	0.6446 (1.89)
	i	$\infty$	$\infty$	8.912	$\infty$	$\infty$	4.577
	j	$\infty$	5.299	6.732	$\infty$	7.060	4.200
	$r^{(e)}$	0.9938	0.9938	0.9938	0.9957	0.9957	0.9958

- (a) The values in brackets are relative standard deviations (percent).  
 (b) The initial size distribution is considered ideal (i=j= $\infty$ , Fig.1).  
 (c) The initial size distribution is truncated at the upper end (i= $\infty$ , j< $\infty$ , Fig.1).  
 (d) The initial size distribution is truncated at both ends (Fig.1).  
 (e) Correlation coefficient.

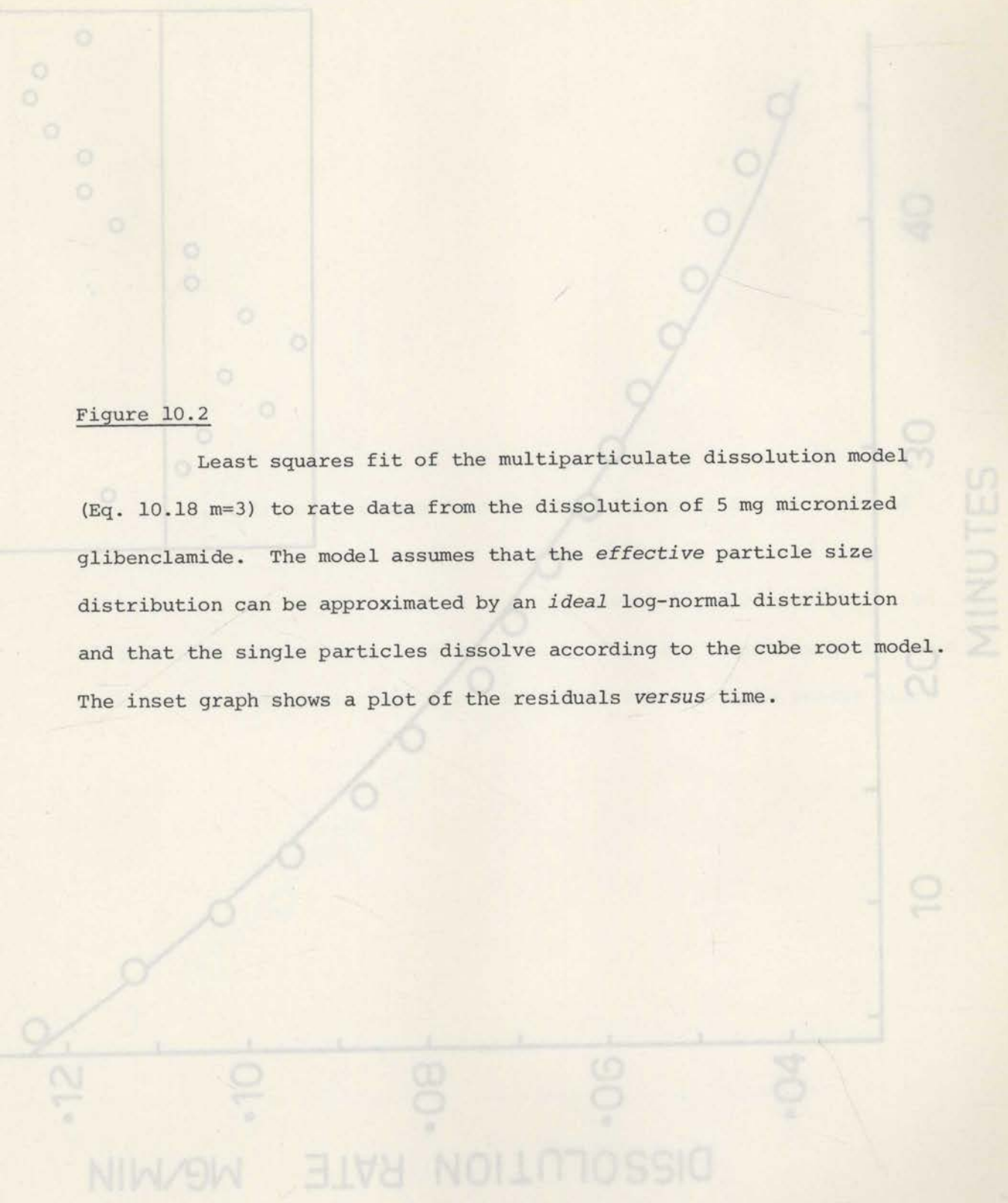


Figure 10.2

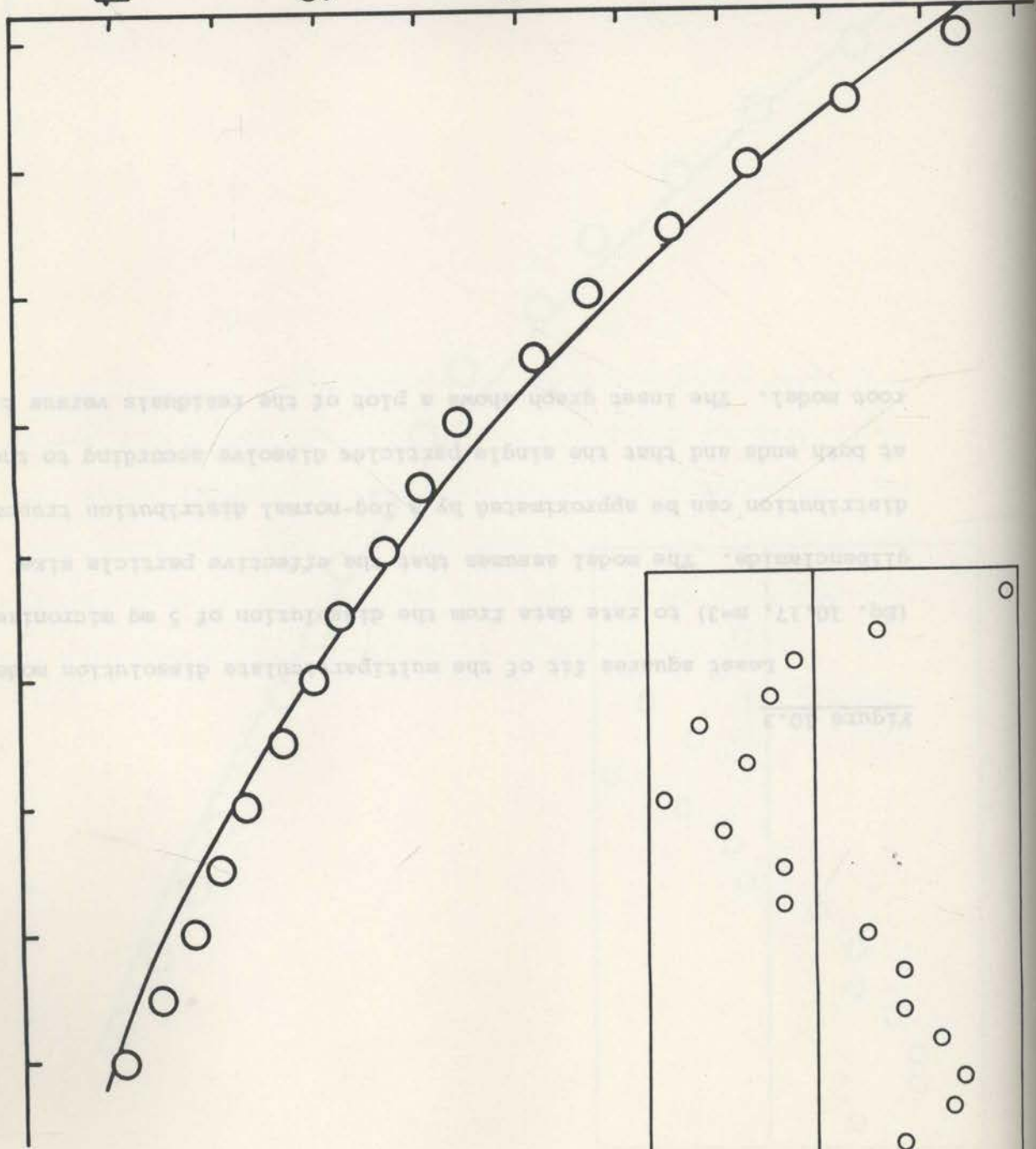
Least squares fit of the multiparticulate dissolution model (Eq. 10.18  $m=3$ ) to rate data from the dissolution of 5 mg micronized glibenclamide. The model assumes that the effective particle size distribution can be approximated by an ideal log-normal distribution and that the single particles dissolve according to the cube root model. The inset graph shows a plot of the residuals versus time.

DISSOLUTION RATE MG/MIN

.04 .06 .08 .10 .12

MINUTES

10 20 30 40



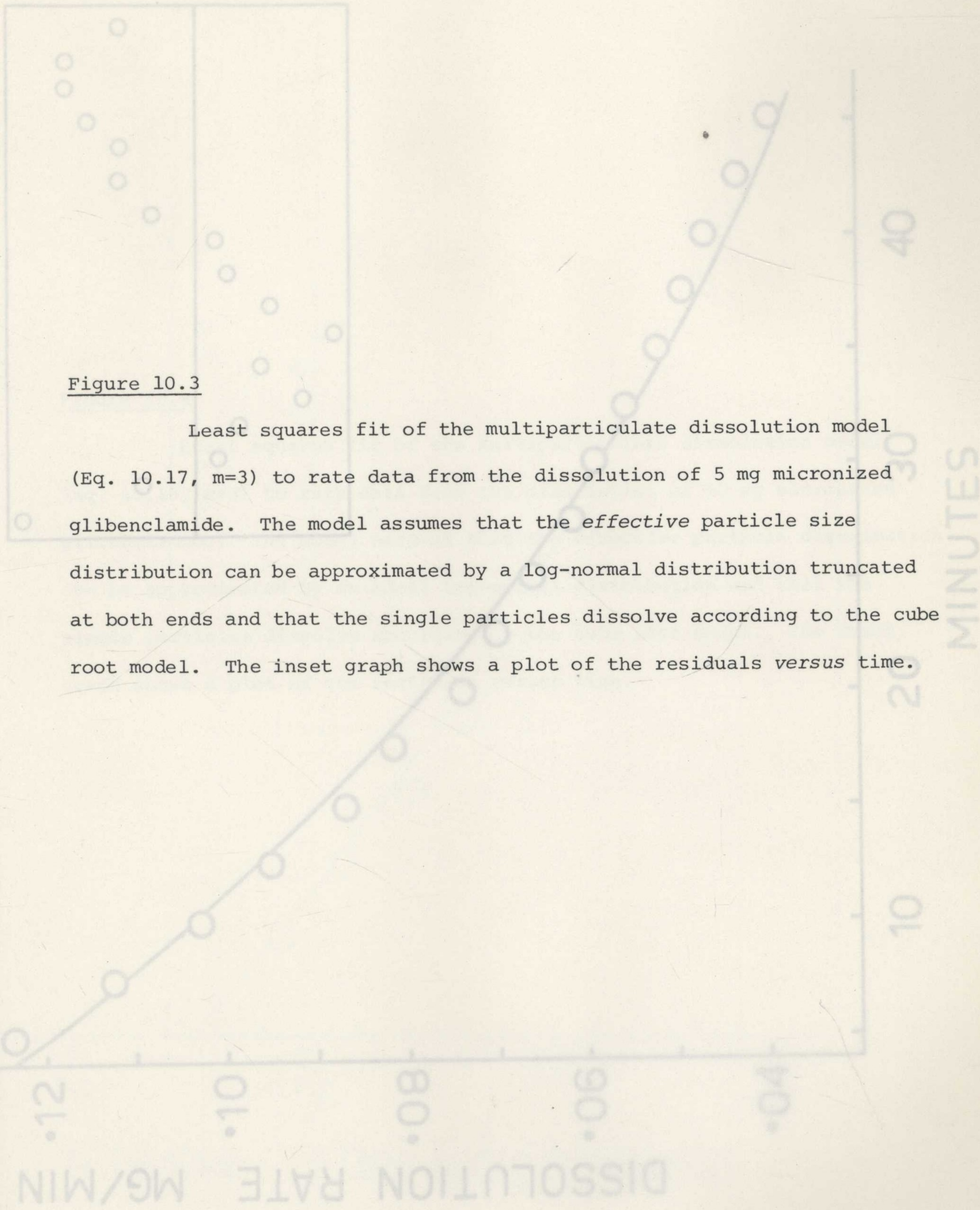


Figure 10.3

Least squares fit of the multiparticulate dissolution model (Eq. 10.17,  $m=3$ ) to rate data from the dissolution of 5 mg micronized glibenclamide. The model assumes that the *effective* particle size distribution can be approximated by a log-normal distribution truncated at both ends and that the single particles dissolve according to the cube root model. The inset graph shows a plot of the residuals versus time.

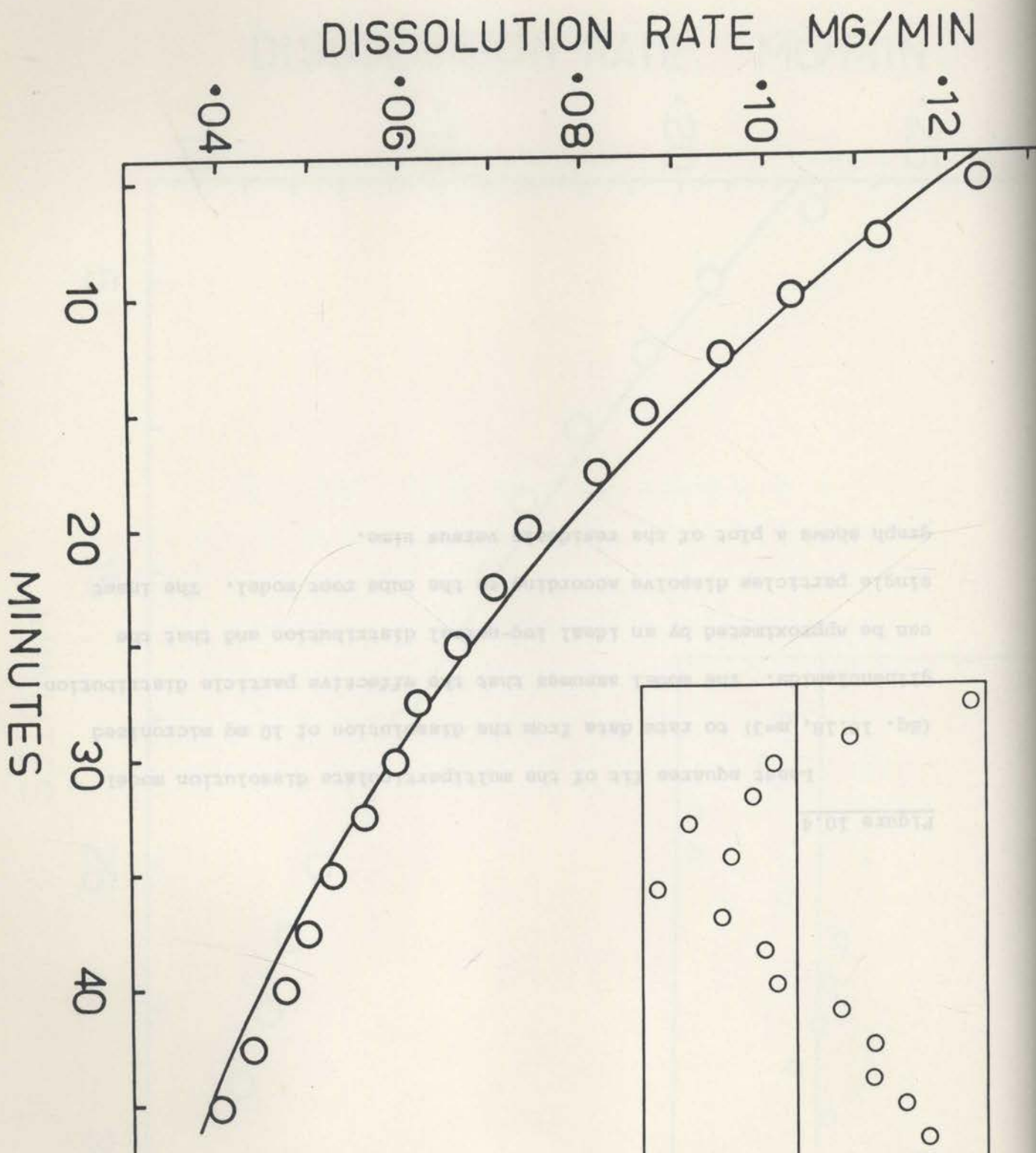
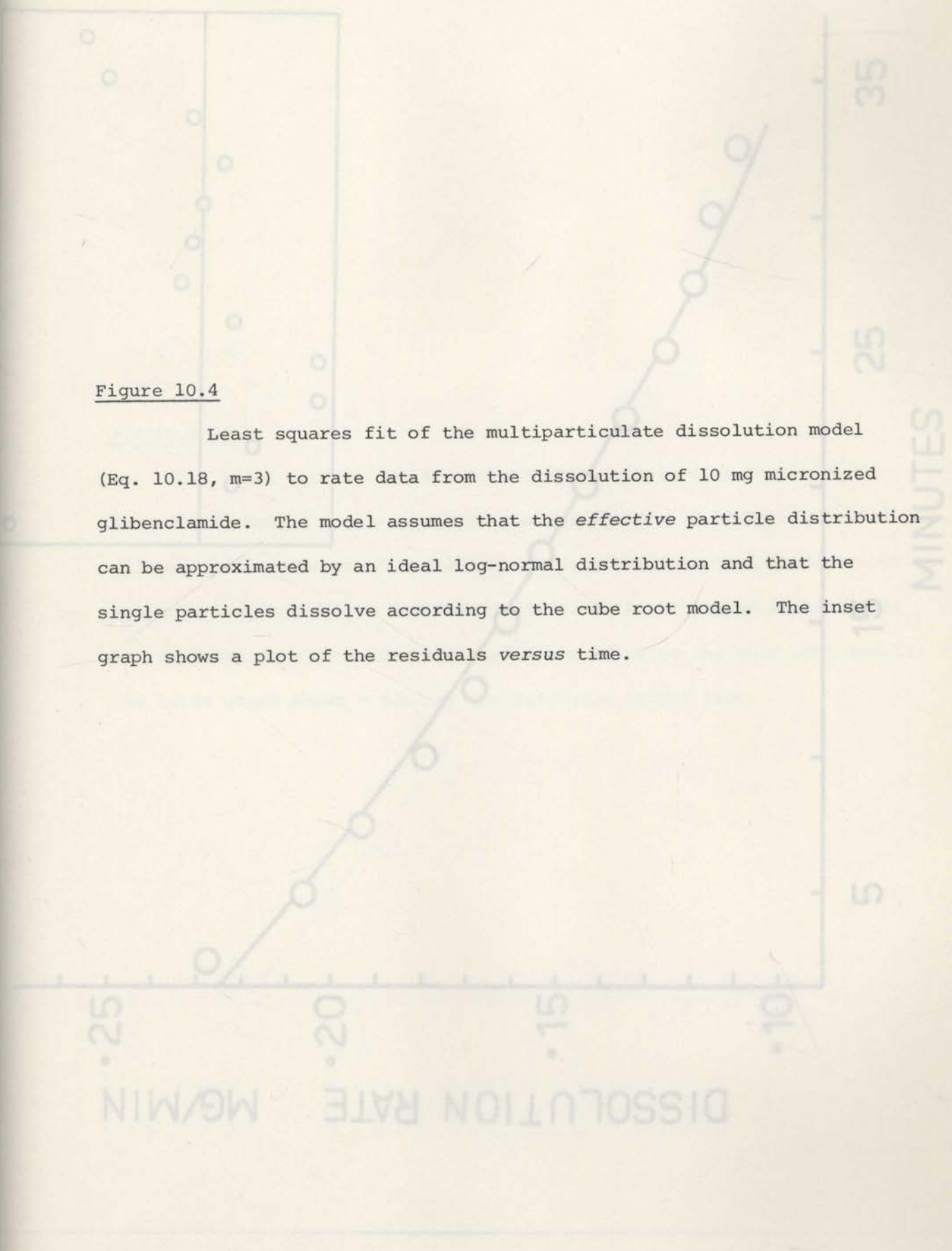


Figure 10.4

Least squares fit of the multiparticulate dissolution model (Eq. 10.18,  $m=3$ ) to rate data from the dissolution of 10 mg micronized glibenclamide. The model assumes that the *effective* particle distribution can be approximated by an ideal log-normal distribution and that the single particles dissolve according to the cube root model. The inset graph shows a plot of the residuals versus time.



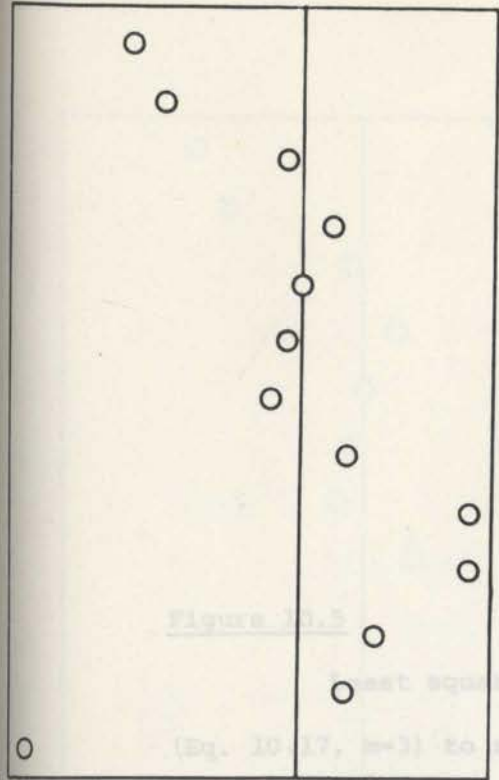
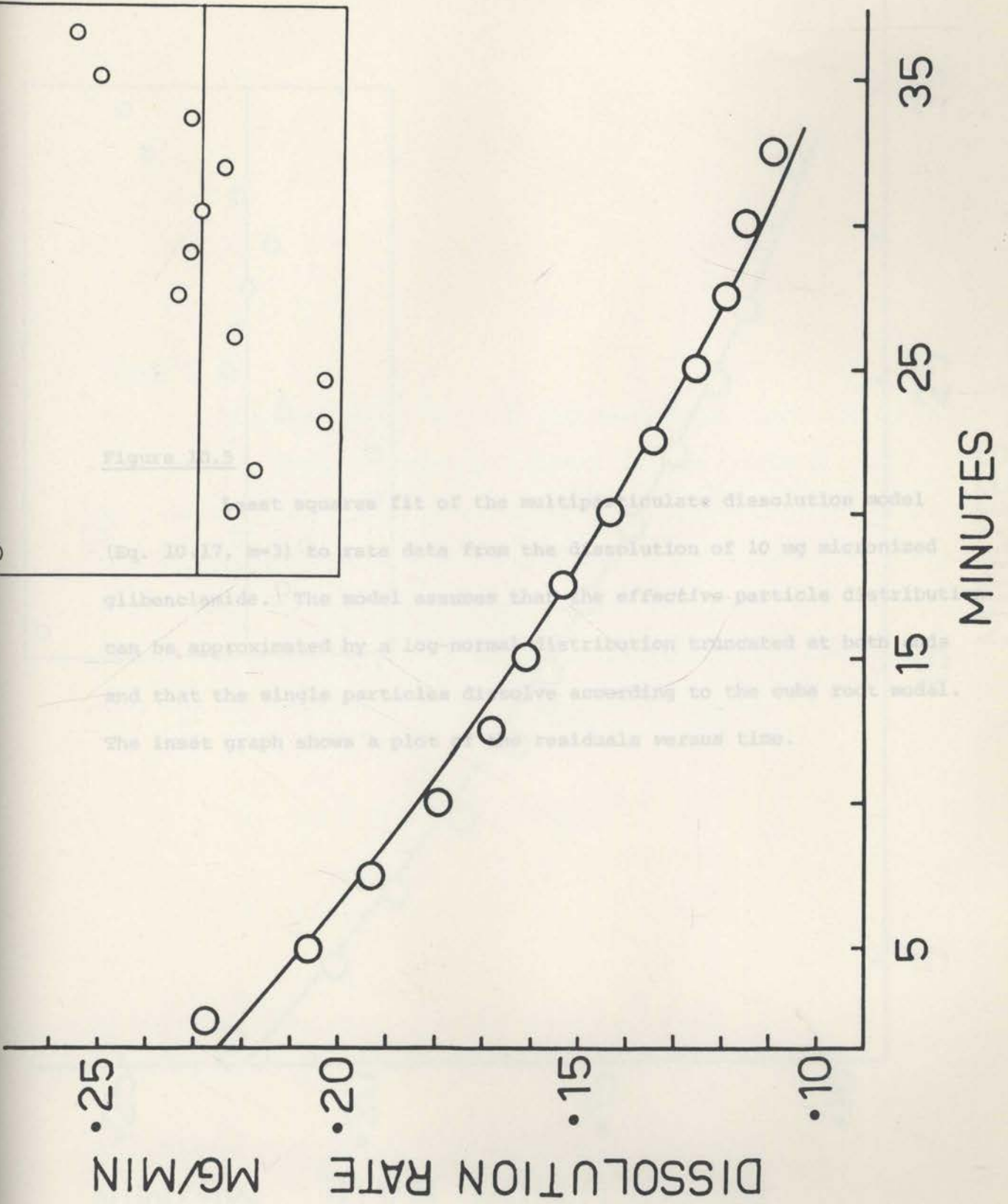


Figure 10.5

least squares fit of the multiparticle dissolution model (Eq. 10.17,  $n=3$ ) to rate data from the dissolution of 10 mg micronized glibenclamide. The model assumes that the effective particle distribution can be approximated by a log-normal distribution truncated at both ends and that the single particles dissolve according to the cube root model. The inset graph shows a plot of residuals versus time.





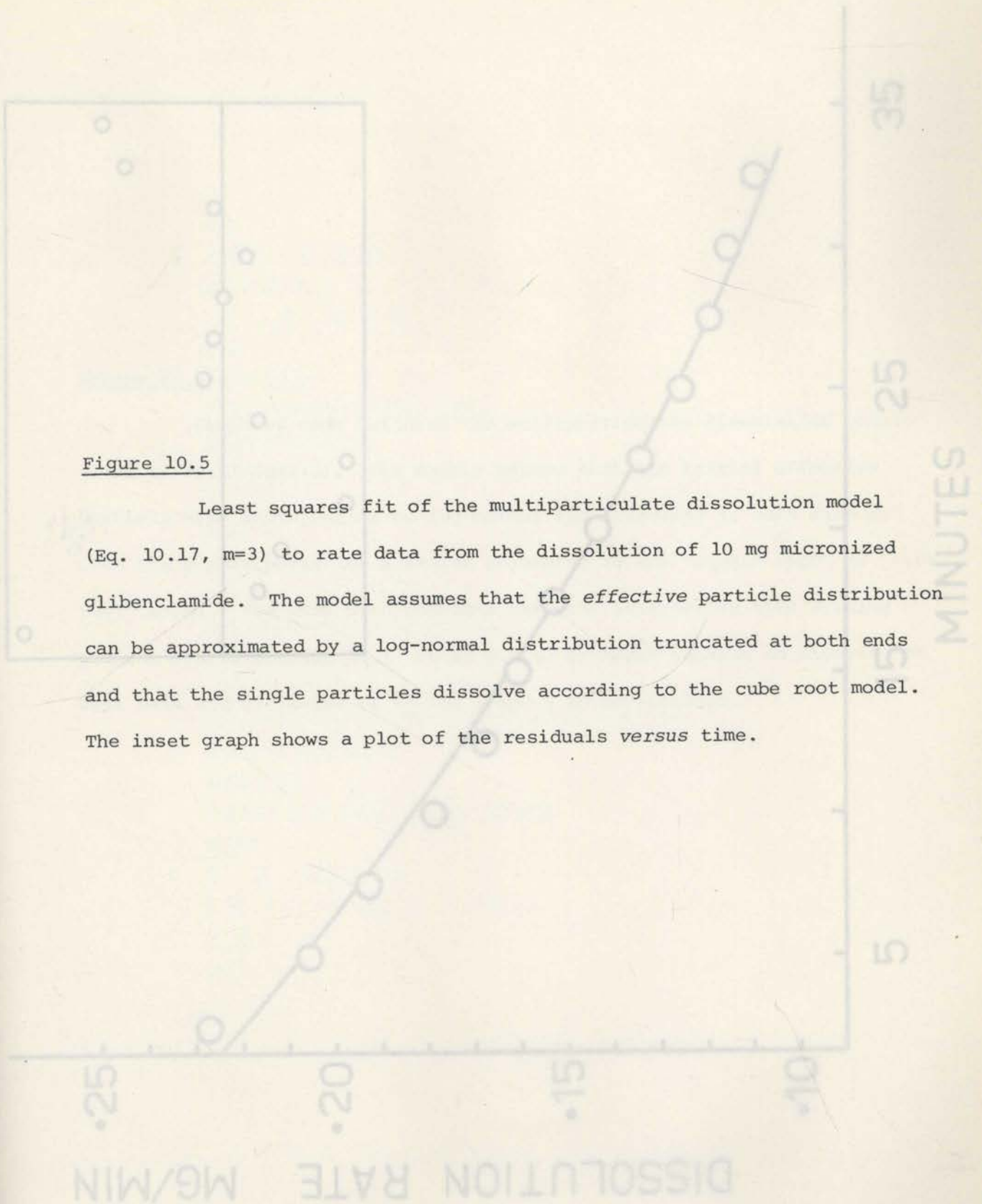


Figure 10.5

Least squares fit of the multiparticulate dissolution model (Eq. 10.17,  $m=3$ ) to rate data from the dissolution of 10 mg micronized glibenclamide. The model assumes that the *effective* particle distribution can be approximated by a log-normal distribution truncated at both ends and that the single particles dissolve according to the cube root model. The inset graph shows a plot of the residuals versus time.

DISSOLUTION RATE MG/MIN

•25  
•20  
•15  
•10

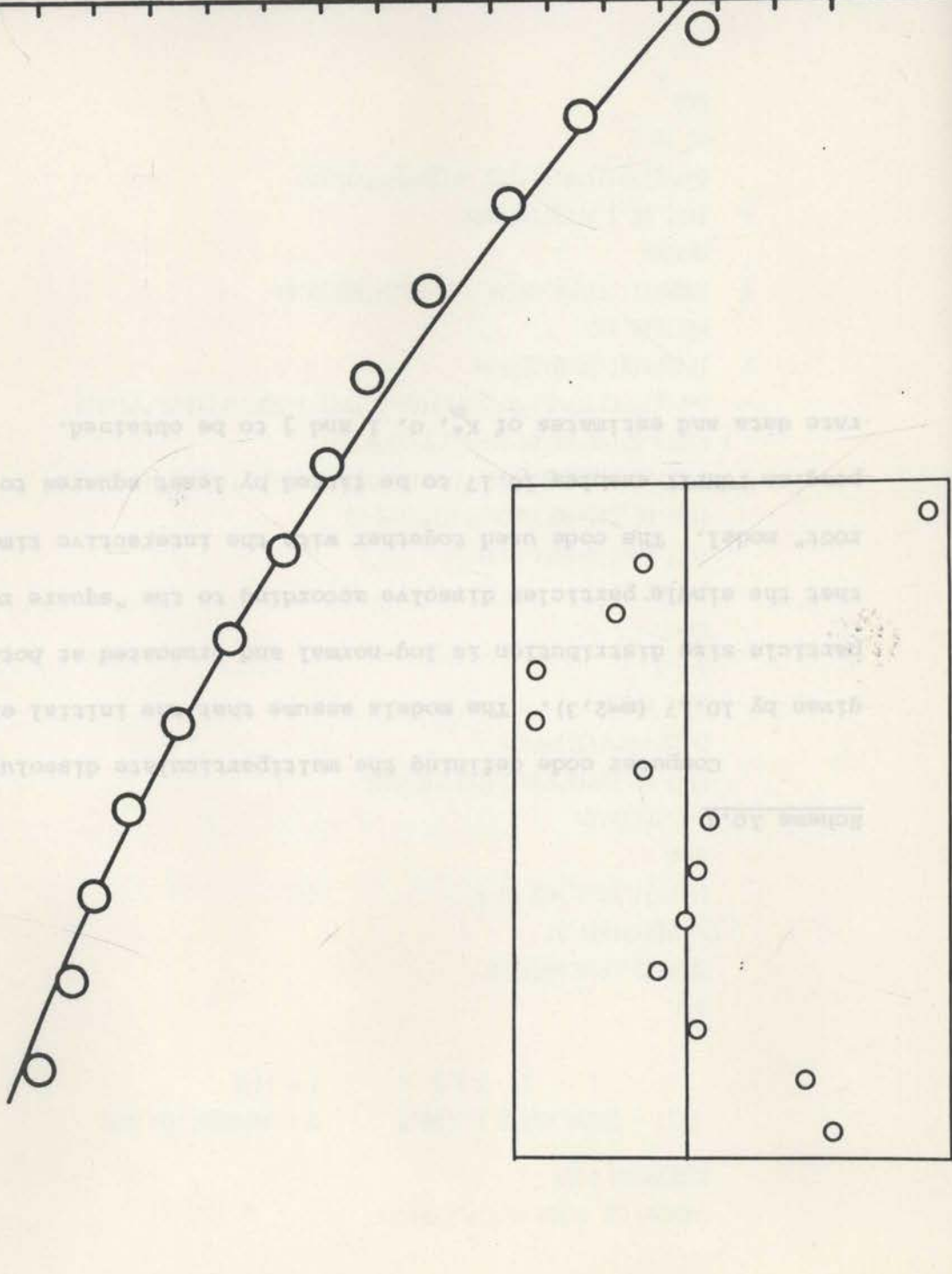
5

15

25

35

MINUTES



```

SUBROUTINE INDEL(O,T,P,IPRINT)
DIMENSION P(6)

P(1) = SIGMA,KSTAR,I,J,NO,N      Q = DISSOLUTION RATE
      1   2   3 4 5 6          T = TIME

```

```

Q=0.
IF(P(2).LE.0.)RETURN
N=IFIX(P(6)+.1)
IF(P(1).EQ.0.)GO TO 4
N=N

```

Scheme 10.1

Computer code defining the multiparticulate dissolution models given by 10.17 ( $m=2,3$ ). The models assume that the initial *effective* particle size distribution is log-normal and *truncated at both ends* and that the single particles dissolve according to the "square root" or "cube root" model. The code used together with the interactive time sharing program FUNFIT enables 10.17 to be fitted by least squares to dissolution rate data and estimates of  $K_m^*$ ,  $\sigma$ ,  $i$  and  $j$  to be obtained.

```

2 IF(IPRINT.EQ.0)RETURN
WRITE(6,5)P
3 FORMAT(' SIGMA,KSTAR,I,J,NO,N= ',/E12,4)
RETURN
4 IF(T.GE.1./P(2))RETURN
Q=P(6)*P(2)*P(5)*(1.-P(2)*T)**(M-1)
GO TO 2
END

```

SUBROUTINE MODEL(Q,T,P,IPRINT)

DIMENSION P(6)

P(1) = SIGMA,KSTAR,I,J,WØ,M  
1 2 3 4 5 6

Q = DISSOLUTION RATE

T = TIME

Q=Ø.

IF(P(2).LE.Ø.)RETURN

M=IFIX(P(6)+.1)

IF(P(1).EQ.Ø.)GO TO 4

AM=M

B=3.\*P(1)/AM

IF(T.GE.EXP(B\*P(4))/P(2))RETURN

C=EXP(-B\*P(3))/P(2)

DO 1 L=1,M

N=L-1

AN=N

D=B\*AN

IF(T.LT.C)A=SDF(-P(3)-D)

IF(T.GE.C)A=SDF(ALOG(P(2)\*T)/B-D)

1 Q=Q+FLOAT(NBC(M-1,N))\*((-P(2)\*T)\*\*(M-N-1))\*(SDF(P(4)-D)-A)\*

+ EXP(4.5\*(AN\*AN/AM/AM-1.))\*P(1)\*P(1)

Q=P(6)\*P(2)\*P(5)\*Q/(SDF(P(4)-3.\*P(1))-SDF(-P(3)-3.\*P(1)))

2 IF(IPRINT.EQ.Ø)RETURN

WRITE(6,3)P

3 FORMAT(' SIGMA,KSTAR,I,J,WØ,M=' /6E12.4)

RETURN

4 IF(T.GE.1./P(2))RETURN

Q=P(6)\*P(2)\*P(5)\*((1.-P(2)\*T)\*\*(M-1))

GO TO 2

END

```

SUBROUTINE MODEL(Q,T,P,IPRINT)
DIMENSION P(4)
P(1) = SIGMA,KSTAR,K0,Q Q = DISSOLUTION RATE

```

Scheme 10.2

Computer code defining the multiparticulate dissolution models given by 10.18 (m=2,3). The models assume that the initial *effective* particle size distribution is *ideal* log-normal and that the single particles dissolve according to the "square root" or "cube root" model. This code used together with the interactive time sharing program FUNFIT enables 10.18 to be fitted by least squares to dissolution rate data and estimates of  $K_m^*$  and  $\sigma$  to be obtained.

```

A=AN/CS.*P(1)
A=SQF(A*ALOG(P(2)*T)-AN/A)
1 Q=Q+FLOAT(NBC(0)-L,N)*((1-P(2)*T)**(M-N-1))*Q.-A)*
*EXP(4.5*(AN*AN/NV*AN-1.)*P(1)*P(1))
Q=P(4)*P(2)*P(3)*Q
2 IF(IPRINT.EQ.0)RETURN
WRITE(6,3)P
3 FORMAT(' SIGMA,KSTAR,K0,T='*NELL.4)
RETURN
4 IF(T.GE.1./P(2))RETURN
Q=P(4)*P(2)*P(3)*(Q.-P(2)*T)**(M-1)
GO TO 2
END

```

```

SUBROUTINE MODEL(Q,T,P,IPRINT)
DIMENSION P(4)
C
C
C
P(1) = SIGMA, KSTAR, W0, M      Q = DISSOLUTION RATE
      1      2      3  4      T = TIME

Q=0.
IF(P(2).LE.0.) RETURN
I=IFIX(P(4)+.1)
IF(P(1).EQ.0.) GO TO 4
AM=M
A=0.
DO 1 L=1,M
N=L-1
AN=I
IF(T.EQ.0.) GO TO 1
A=AM/(3.*P(1))
A=SDF(A*ALOG(P(2)*T)-AN/A)
1  Q=Q+FLOAT(IIBC(M-1,N))*((-P(2)*T)**(M-N-1))*(1.-A)*
+EXP(4.5*(AN*AN/AM/AM-1.))*P(1)*P(1)
Q=P(4)*P(2)*P(3)*Q
2  IF(IPRINT.EQ.0) RETURN
WRITE(6,3)P
3  FORMAT(' SIGMA, KSTAR, W0, M='4E11.4)
RETURN
4  IF(T.GE.1./P(2)) RETURN
Q=P(4)*P(2)*P(3)*((1.-P(2)*T)**(M-1))
GO TO 2
END

```

```

SUBROUTINE MODEL(Q, T, P, IPRINT)
DIMENSION P(5)
P(1) = SIGMA, KSTAR, J, M, Q = DISSOLUTION RATE
Q=0.

```

Scheme 10.3 F(P(2), LE, Q, IPRINT)

Computer code defining the multiparticulate dissolution models given by 10.19 ( $m=2,3$ ). The models assume that the initial *effective* particle size distribution is log-normal and *truncated at the upper end* ( $i=\infty, j<\infty$ ) and that the single particles dissolve according to the "square root" or "cube root" model. The code used together with the interactive time sharing program FUNFIT enables 10.19 to be fitted by least squares to dissolution rate data and estimates of  $K_m^*$ ,  $\sigma$  and  $j$  to be obtained.

```

A=SQRT(ALOG(P(2)*T)/S-B*AN)
1. Q=Q+FLOAT(NBC(M-1,N))*(1-P(2)*T)**(M-B-1)*(SQF(P(3)-D*ND-D)*
*EXP(4.5*(AN*AN/AR/AR-1.)*P(1)*P(1))
Q=P(5)*P(2)*P(4)*Q/SQF(P(3)-S.*P(1))
2. IF(IPRINT.EQ.0)RETURN
WRITE(6,3)P
3. FORMAT(' SIGMA, KSTAR, J, M, Q = 'SE1B,4)
RETURN
4. IF(T.GE.1./P(2))RETURN
Q=P(5)*P(2)*P(4)*(1.-P(2)*T)**(M-1)
GO TO 2
END

```

simplest of the models, containing only the two parameters  $K_1$  and  $\sigma$ , and fits the dissolution data just as well as the other models that contain more parameters. (Table 10.1) that the multiple solution model based on the cube root model (n=3) agrees best with the data. Thus, it can be concluded that the distribution using either 5 or 10 mg of powder (Table 10.1), of the 8 multiple solutions investigated it can therefore be concluded that the distribution parameter,  $\sigma$ , is not significantly different from 1.0. The distribution parameter,  $\sigma$ , which can be determined in integrated form as:

$$\frac{dW}{dt} = -K_1 \left( \frac{W}{W_0} \right)^{3-n} \left[ 1 - \frac{1}{\sigma} \ln \left( \frac{W}{W_0} \right) - n \right] e^{\frac{\sigma^2}{3} [n^2 - 9]} \quad (10.29)$$

$$\frac{W}{W_0} = \sum_{i=1}^3 \left( \frac{W_i}{W_0} \right)^{3-n} \left[ 1 - \frac{1}{\sigma} \ln \left( \frac{W_i}{W_0} \right) - n \right] e^{\frac{\sigma^2}{3} [n^2 - 9]} \quad (10.30)$$

best described by the dissolution behaviour in terms of  $\sigma$ , and the distribution parameter,  $\sigma$ . From the residual plots in Figs. 10.2-10.5 that there is a serial correlation between the residuals. The Durbin-Watson statistic indicates that this correlation is significant (142). Systematic deviation can be caused by non-random experimental errors. It can also be caused by a significant departure from the assumed single particle dissolution model 10.23 or by a deviation from homogeneity. The residual values are however so small in relation to the accuracy of the experimental technique that the correlation seems of little importance. The specific dissolution rate parameter,  $K_1$ , should theoretically

```

SUBROUTINE MODEL(Q,T,P,IPRINT)
DIMENSION P(5)
      P(1) = SIGMA, KSTAR, J, W0, M   Q = DISSOLUTION RATE
      T = TIME
Q=0.
IF(P(2).LE.0.)RETURN
M=IFIX(P(5)+.1)
IF(P(1).EQ.0.)GO TO 4
AM=M
B=3.*P(1)/AM
IF(T.GE.EXP(B*P(3))/P(2))RETURN
A=0.
DO 1 L=1,M
N=L-1
AIN=i
IF(T.EQ.0.)GO TO 1
A=SDF(ALOG(P(2)*T)/B-B*AN)
1  Q=Q+FLOAT(NBC(M-1,N))*((-P(2)*T)**(M-N-1))*(SDF(P(3)-B*AN)-A)*
+EXP(4.5*(AN*AN/AM/AM-1.)*P(1)*P(1))
Q=P(5)*P(2)*P(4)*Q/SDF(P(3)-3.*P(1))
2  IF(IPRINT.EQ.0)RETURN
WRITE(6,3)P
3  FORMAT(' SIGMA,KSTAR,J,W0,M= '5E10.4)
RETURN
4  IF(T.GE.1./P(2))RETURN
Q=P(5)*P(2)*P(4)*((1.-P(2)*T)**(M-1))
GO TO 2
END
    
```



simplest of the models, containing only the two parameters  $K_m^*$  and  $\sigma$ , and fits the dissolution data just as well as the other models that contain more parameters (i and j). Furthermore, it is clear (Table 10.1) that the multiparticulate dissolution model based on the cube root model ( $m=3$ ) agrees best with the dissolution data. This is true for either a truncated or ideal distribution using either 5 or 10 mg of powder (Table 10.1). Of the 8 multiparticulate models investigated it can therefore be concluded that the following equation:

$$\frac{dW}{dt} = -3K_3^* W_0 \sum_{n=0}^2 \binom{2}{n} (-K_3^* t)^{(2-n)} \left[ 1 - F\left(\frac{1}{\sigma} \ln(K_3^* t) - n\sigma\right) \right] e^{\frac{\sigma^2}{2} [n^2 - 9]} \quad (10.29)$$

which can be written in integrated form as:

$$\frac{W}{W_0} = \sum_{n=0}^3 \binom{3}{n} (-K_3^* t)^{(3-n)} \left[ 1 - F\left(\frac{1}{\sigma} \ln(K_3^* t) - n\sigma\right) \right] e^{\frac{\sigma^2}{2} [n^2 - 9]} \quad (10.30)$$

best describes the dissolution kinetics of the micronized glibenclamide.

These two equations uniquely characterise the dissolution behaviour in terms of the rate parameter,  $K_3^*$ , and the distribution parameter,  $\sigma$ .

It appears from the residual plots in Figs. 10.2-10.5 that there is a serial correlation between the residuals. The Durbin-Watson

statistic indicates ( $\alpha < 0.05$ ) that this correlation is significant (142).

Systematic deviation can be caused by non-random experimental errors.

It can also be caused by a significant departure from the assumed single particle dissolution model 10.25 or by a deviation from log-normality.

The residual values are however so small in relation to the accuracy of the experimental technique that the correlation seems of little importance.

The specific dissolution rate parameter,  $K_3^*$ , should theoretically

3. For example if it takes x min for a powder to dissolve, say 100 for a given  $K_3^*$  value, then it will take the powder x/2 minutes to dissolve to the same extent if the value of  $K_3^*$  is doubled (for illustration see Fig. 6.1).

be independent of the initial amount used,  $W_0$ . This is only found to be approximately true. Values of the rate parameter,  $K_m^*$ , obtained for  $W_0=10$  mg are consistently lower than for  $W_0=5$  mg (Table 10.1). The dissolution models consider dissolution under *complete sink conditions* i.e. conditions where there is no interaction between the dissolving particles. Using 10 mg of the very fine powder it has not been possible to load the dissolution cell with a single "layer" of particles in such a way that dissolved drug from any particle does not pass over other particles. The slightly larger  $K_m^*$  values observed when less powder is used in the cell agree with an expected smaller particle interaction.

Characterisation and quality control of drug powders

Previous approaches to characterizing the dissolution properties of drug powders have been based on equations describing monodisperse systems. In many cases, the so-called "dissolution rate constant" evaluated using such equations will not be sufficient to characterize the dissolution behaviour because the size distribution effect is not accounted for. This is particularly true for pharmaceutical systems which frequently involve highly polydisperse fine powders.

The use of nonlinear regression analysis to evaluate the specific dissolution rate parameter,  $K_m^*$ , and the distribution parameter,  $\sigma$ , represents a more exact and meaningful approach.

The properties of  $K_m^*$  make its interpretation particularly meaningful. These properties are best understood in relation to the concepts of *time scaling* and the *intrinsic dissolution profile* from which the following conclusion can be made:<sup>3</sup>

---

3. For example if it takes  $x$  min for a powder to dissolve, say 30% for a given  $K_m^*$  value, then it will take the powder  $x/2$  minutes to dissolve to the same extent if the value of  $K_m^*$  is doubled (for illustration see Fig. 6.1 ).

If  $K_m^*$  is changed by a factor  $\alpha$  then the time for complete dissolution, or the time for any particular fraction to dissolve, is changed by a factor of  $1/\alpha$ . This simple property makes  $K_m^*$  a particularly useful parameter.

The distribution parameter,  $\sigma$ , which appropriately should be called the dispersion parameter, is a single measure of how polydisperse a powder is, or more exactly, how much the dissolution behaviour deviates from that expected if the powder were completely monodisperse. A value close to zero characterizes a nearly monodisperse powder, while higher values indicate increasing "degrees of dispersion". Probably the most important property of  $\sigma$  is that it is a measure of how long it takes the last fraction of a polydisperse powder to dissolve. For example, it is seen from 10.5 that the time for complete dissolution increases exponentially with  $\sigma$ . For this reason it is most likely that very slightly soluble drugs, which exhibit dissolution rate limited absorption, will show a significant correlation between  $\sigma$  and systemic availability. Research in this area should be of considerable pharmaceutical interest.

Although the multiparticulate dissolution model defining  $K_m^*$  and  $\sigma$  may seem complex, the interpretation of these parameters is simple and they can be readily obtained. The experimental technique used requires a high precision, flow-through dissolution apparatus which is easy to standardize in combination with a nonlinear regression program.

The method could well become established as a routine procedure in quality control and further investigation could eventually result in improved standards for drug dissolution.

C H A P T E R 1 1

A METHOD OF OBTAINING DRUG-MACROMOLECULE BINDING

PARAMETERS DIRECTLY FROM DYNAMIC DIALYSIS DATA

The dynamic dialysis method for characterizing interactions of small molecules with macromolecules is well established (167-181). It has a number of advantages compared to equilibrium dialysis and ultrafiltration. A complete binding profile can be obtained rapidly in one experiment and the method utilizes only a small sample of macromolecule. As the method is based on a dynamic process, an equilibrium state does not need to be defined and compared to ultrafiltration there is no change in concentration of macromolecule.

Meyer and Guttman (169) designed a dynamic dialysis method to characterize drug-protein interactions. However this method has a number of limitations. The experimental data must be differentiated to evaluate binding parameters. It is recognised in numerical analysis that differentiation of discrete data may introduce substantial errors particularly if the number of data points is limited. An empirical equation was used to fit dialysis data to obtain instantaneous rates. A recent publication (179) has shown that the various empirical equations used can yield substantially different binding parameters.

The technique of Meyer and Guttman (169) requires that the rate constant for dialysis be determined in a separate experiment in the absence of macromolecules. It is assumed that the same rate constant will apply in the presence of macromolecules. This may be an unreasonable assumption as the rate constant depends on several factors such as the physico-chemical

state of the dialysis membrane (182, / ) which may change between runs.

A number of compounds are significantly bound to the membrane material (171,177). Using previous methods it has not been possible to determine the dialysis rate constant and account for the membrane binding in the determination of macromolecule binding parameters of such compounds.

In this chapter a new approach is presented which rigorously describes the total kinetics of the system in a form that enables binding parameters to be estimated accurately, directly from dialysis data. It eliminates the need to determine an accurate dialysis rate constant in a separate experiment. The method does not rely on differentiation of experimental data and should be applicable to compounds that are membrane bound.

THEORY

Consider an interaction between small molecules and macromolecules which can be described by the general binding expression:

$$\bar{v} = \sum_{i=1}^j \frac{n_i K_i D_f}{1 + K_i D_f} \tag{11.1}$$

where  $\bar{v}$  is the number of moles of small molecules bound per mole macro molecule,  $n_i$  is the number of binding sites in the  $i$ -th class of sites,  $K_i$  is the association constant for the interaction and  $D_f$  is the molar concentration of unbound small molecules. If a model with two classes ( $j=2$ ) is assumed then the total concentration of drug,  $D_t$ , in the protein compartment is given by (171):

$$D_t = D_f + P_t D_f \left[ \frac{n_1 K_1}{1 + K_1 D_f} + \frac{n_2 K_2}{1 + K_2 D_f} \right] \tag{11.2}$$

If sink conditions prevail the small molecules will leave the

protein compartment by a first order process:  
 now described by Eq's 11.5 and 11.9 in parametric form where the variable

$$x \text{ is the parameter } \frac{dD_t}{dt} = -K_e D_f \quad (11.3)$$

where  $K_e$  is the dialysis rate constant.

It is convenient to introduce a variable,  $s$ , defined as:

$$s = - \frac{dD_t}{dt} \quad (11.4)$$

so that  $D_f = s/K_e$  and Eq. 11.6 can be written:

$$D_t = \frac{s}{K_e} + P_t s \left[ \frac{n_1 K_1}{K_e + K_1 s} + \frac{n_2 K_2}{K_e + K_2 s} \right] \quad (11.5)$$

Taking the differential of this equation, noting  $dD_t = -s dt$ , it becomes

$$dt = - \left[ \frac{1}{K_e s} + P_t K_e \left( \frac{n_1 K_1}{s(K_e + K_1 s)^2} + \frac{n_2 K_2}{s(K_e + K_2 s)^2} \right) \right] ds \quad (11.6)$$

which integrated from  $t=0$  to  $t$  corresponding to  $s = s_0$  to  $s$  yields:

$$t = \left[ \frac{1}{K_e} \ln s + P_t n_1 K_1 \left[ \frac{1}{K_e + K_1 s} + \frac{1}{K_e} \ln \left( \frac{s}{K_e + K_1 s} \right) \right] \right] \quad (11.7)$$

$$+ P_t n_2 K_2 \left[ \frac{1}{K_e + K_2 s} + \frac{1}{K_e} \ln \left( \frac{s}{K_e + K_2 s} \right) \right] \Bigg|_{s_0}^s \quad (11.10)$$

then Eq. 11.9 can be solved by the Newton-Raphson method using the following

If the following function is defined:

$$f(x) = \frac{1}{K_e} \ln x + P_t n_1 K_1 \left[ \frac{1}{K_e + K_1 x} + \frac{1}{K_e} \ln \left( \frac{x}{K_e + K_1 x} \right) \right] \quad (11.8)$$

where the functions  $\phi$  and  $\phi'$  are defined by:

$$+ P_t n_2 K_2 \left[ \frac{1}{K_e + K_2 x} + \frac{1}{K_e} \ln \left( \frac{x}{K_e + K_2 x} \right) \right]$$

then Eq. 11.7 can be written more simply as:

$$t = f(s_0) - f(s) \quad (11.9)$$

$\phi$  is a monotone increasing function of  $s$  because  $\phi' = \phi' > 0$  ( $s > 0$ ).

The exact functional relationship describing the change of  $D_t$  with  $t$  is now described by Eq's 11.5 and 11.9 in *parametric form* where the variable  $s$  is the parameter. Each value of  $s$  defines by these equations a *unique* pair of  $D_t$  and  $t$  values.

The quantity  $s_0$  is the initial ( $t=0$ ) value of  $-dD_t/dt$  (Eq. 11.4), which would normally be determined by extrapolation. To avoid the errors and problems of such an extrapolation *it is convenient to define  $t=0$  at the first sampling time.* In this way  $s_0$  is  $-dD_t/dt$  at first sampling.

In order to determine the binding parameters by nonlinear regression it is necessary to define the exact functional relationship between  $D_t$  and  $t$ , for any values of  $n_1, K_1, n_2, K_2, s_0$  and  $K_e$  which are changing during the nonlinear fitting procedure. This can be done by determining the particular values of  $s$  which satisfy Eq. 11.9. These values are then used to determine the corresponding values of  $D_t$  by Eq. 11.5.

However Eq. 11.9 cannot be expressed explicitly in terms of  $s$  so some iterative procedure is needed to solve for  $s$ . The Newton-Raphson algorithm is particularly suitable because it is computationally compact and exhibits quadratic convergence.

If 
$$\phi(s) = t - f(s_0) + f(s) \tag{11.10}$$

then Eq. 11.9 can be solved by the Newton-Raphson method using the following iteration<sup>1</sup>:

$$s_{i+1} = s_i - \frac{\phi(s_i)}{\phi'(s_i)} \tag{11.11}$$

where the functions  $\phi$  and  $\phi'$  are defined by:

---

1. The function  $\phi$  is not defined for  $s \leq 0$  so it is necessary during the iteration procedure to prevent  $s$  from taking a nonpositive value. This is conveniently done by defining  $s_{i+1} = s_i/2$  if  $s_{i+1} \leq 0$  since  $\phi$  is a monotone increasing function of  $s$  because  $\phi' = f' > 0$  ( $s > 0$ ).

$$\phi'(s) = f'(s) = \frac{1}{K_e s} + \frac{P_t K_e}{s} \left[ \frac{n_1 K_1}{(K_e + K_1 s)^2} + \frac{n_2 K_2}{(K_e + K_2 s)^2} \right] \quad (11.12)$$

$$\phi(s) = t + \frac{1}{K_e} \ln \frac{s}{s_0} + P_t n_1 K_1 \left[ \frac{K_1 (s_0 - s)}{(K_e + K_1 s)(K_e + K_1 s_0)} + \frac{1}{K_e} \ln \frac{s}{s_0} \frac{(K_e + K_1 s_0)}{(K_e + K_1 s)} \right] \quad (11.13)$$

$$+ P_t n_2 K_2 \left[ \frac{K_2 (s_0 - s)}{(K_e + K_2 s)(K_e + K_2 s_0)} + \frac{1}{K_e} \ln \frac{s}{s_0} \frac{(K_e + K_2 s_0)}{(K_e + K_2 s)} \right]$$

The treatment outlined above is based on an assumption that binding occurs only to protein. However, some drugs, particularly those strongly protein bound can become significantly bound to the dialysis membrane. This means that  $K_e$  cannot be estimated from plots of  $\ln D_t$  vs.  $t$  in the absence of protein and previous techniques because of curvature estimated from the observed values of  $D_t$  vs.  $t$ . Any simple technique for slope estimation can be used since the actual accuracy of the estimates is of no importance for the final result.

The above derivations for 2 binding classes can easily be extended to any number ( $j$ ) of classes for which then:

$$D_t = \frac{s}{K_e} + P_t s \sum_{i=1}^j \frac{n_i K_i}{K_e + K_i s} \quad (11.14)$$

$$t = \left. \frac{1}{K_e} \ln s + P_t \sum_{i=1}^j n_i K_i \left[ \frac{1}{K_e + K_i s} + \frac{1}{K_e} \ln \left( \frac{s}{K_e + K_i s} \right) \right] \right|_s^{s_0} \quad (11.15)$$

and the iteration, 11.11, can still be used with

$$\phi'(s) = \frac{1}{K_e s} + \frac{P_t K_e}{s} \sum_{i=1}^j \frac{n_i K_i}{(K_e + K_i s)^2} \quad (11.16)$$

and  $K^*$  is the amount of small molecules bound per amount of available membrane material and  $K^*$  is the association constant for membrane binding.



$$\phi(s) = t + \frac{1}{K_e} \ln \frac{s}{s_0} \quad (11.17)$$

$$+ P_t \sum_{i=1}^j n_i K_i \left[ \frac{K_i (s_0 - s)}{(K_e + K_i s) (K_e + K_i s_0)} + \frac{1}{K_e} \ln \frac{s (K_e + K_i s_0)}{s_0 (K_e + K_i s)} \right]$$

Binding of Small Molecules by the Dialysis Membrane

The treatment outlined above is based on an assumption that binding occurs only to protein. However some drugs, particularly those strongly protein bound can become significantly bound to the dialysis membrane. This means that  $K_e$  cannot be estimated from plots of  $\ln D_t$  vs.  $t$  in the absence of protein using previous techniques because of curvature (Fig. 11.1).

A special technique is therefore required to estimate the dialysis rate constant and the membrane binding must be taken into account in the treatment of the dialysis behaviour of the small molecule-macromolecule system.

Determination of  $K_e$  and membrane binding parameters in absence of macromolecule.

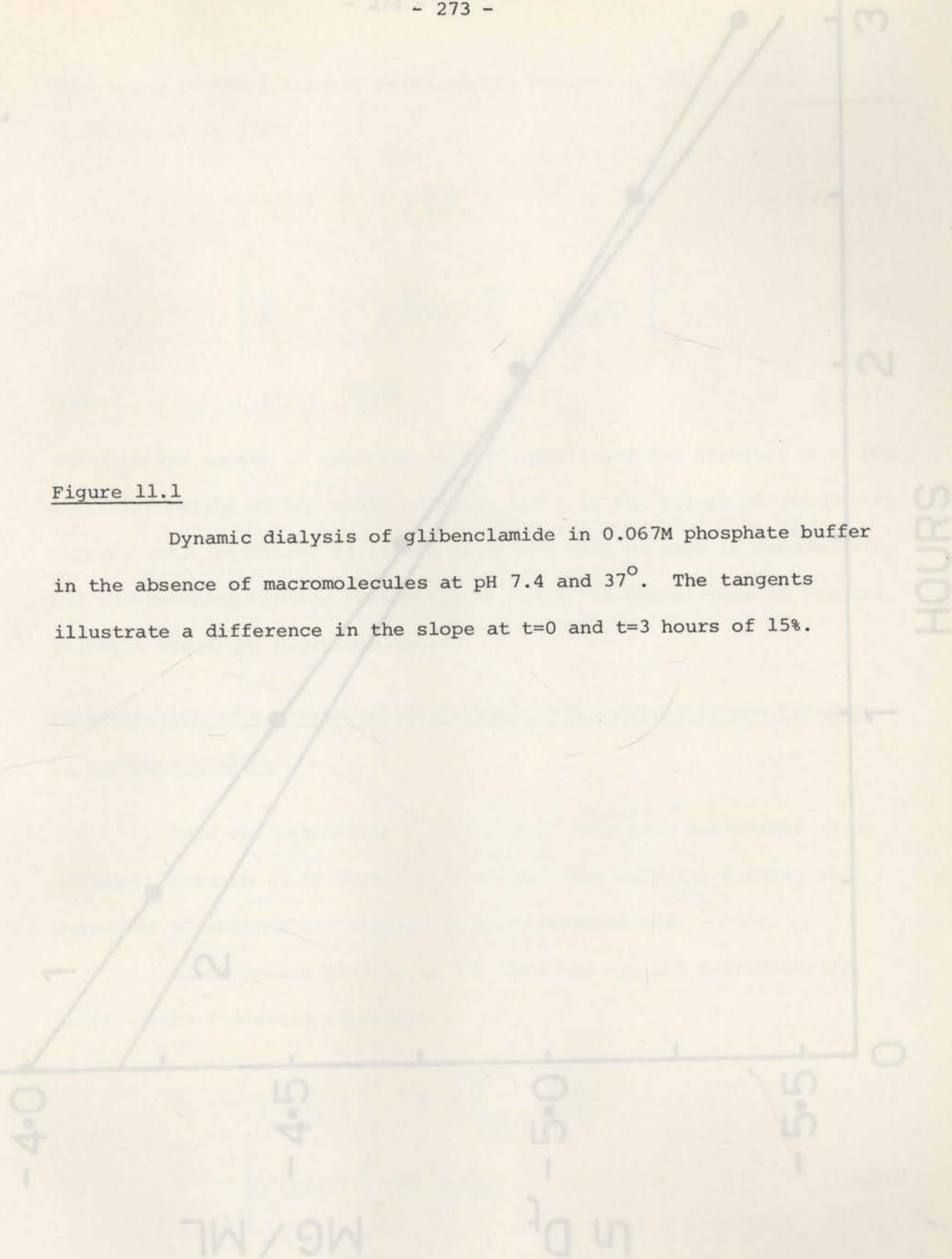
The binding of small molecules to the membrane can often be considered as a Langmuir type absorption phenomenon (171), which is mathematically analogous to binding to a single class of sites, and can be described by the equation:

$$\bar{v}^* = \frac{n^* K^* D_f}{1 + K^* D_f} \quad (11.18)$$

where  $\bar{v}^*$  is the amount of small molecules bound per amount of available membrane material and  $K^*$  is the association constant for membrane binding.

Figure 11.1

Dynamic dialysis of glibenclamide in 0.067M phosphate buffer in the absence of macromolecules at pH 7.4 and 37°. The tangents illustrate a difference in the slope at t=0 and t=3 hours of 15%.



This leads to the following relationship between  $D_t$  and  $D_s$  (Miller 11.14 and 11.15 ( $j=1$ )):

$$D_t = \frac{K}{K_0} + \frac{K^2}{K_0 + K^2}$$

$$\ln \frac{D_t}{D_s} = \frac{K}{K_0} \ln \frac{K_0 + K^2}{K_0} + \frac{K^2}{K_0 + K^2} \ln \frac{K_0 + K^2}{K_0}$$

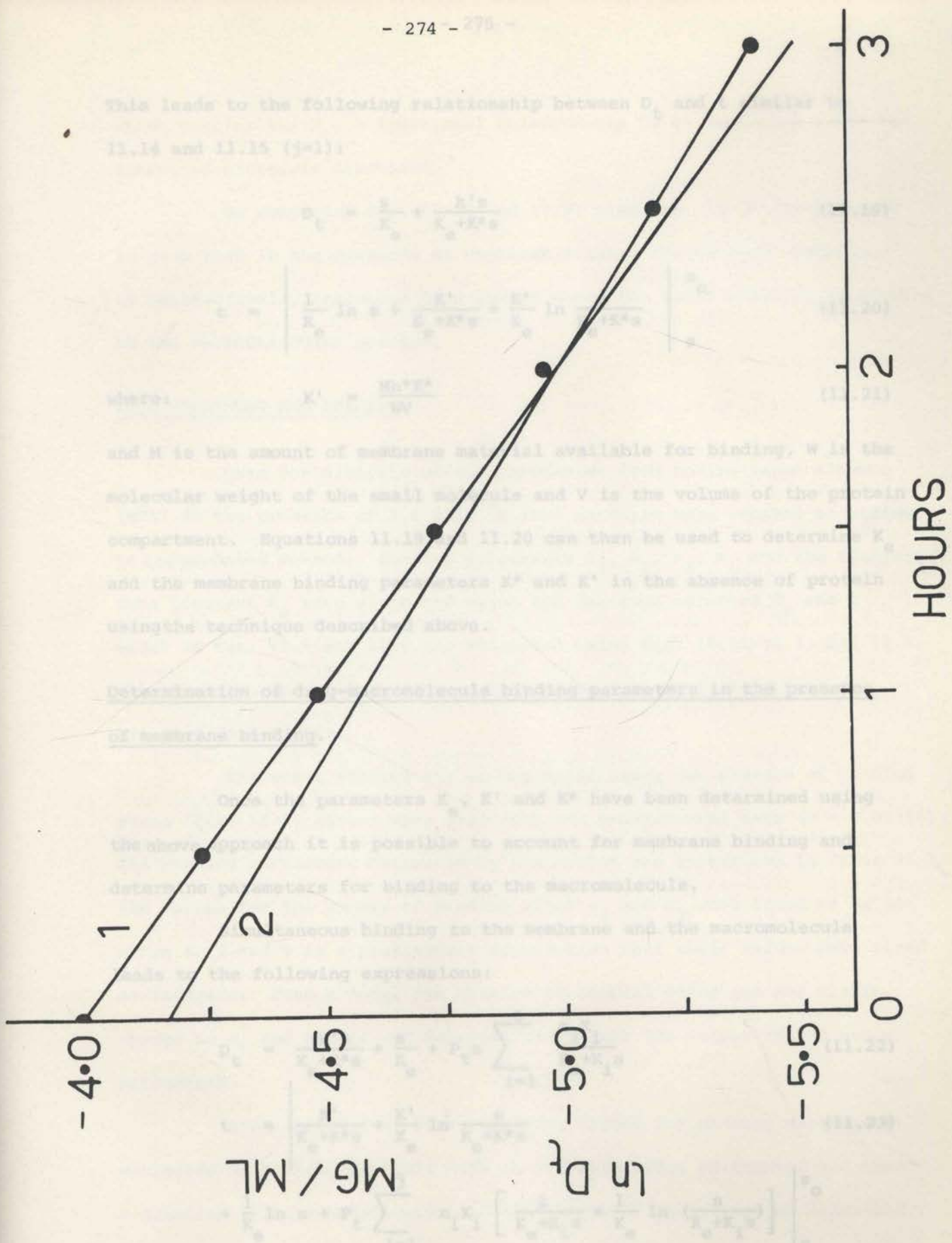
where:  $K = \frac{M_0 \cdot P^0}{W}$

and  $M$  is the amount of membrane material available for binding,  $W$  is the molecular weight of the small molecule and  $V$  is the volume of the protein compartment. Equations 11.14 and 11.20 can then be used to determine  $K_0$  and the membrane binding parameters  $K^0$  and  $K^1$  in the absence of protein using the technique described above.

Determination of  $K^0$ ,  $K^1$  and  $K^2$  membrane binding parameters in the presence of macromolecules

Once the parameters  $K_0$ ,  $K^1$  and  $K^2$  have been determined using the above approach it is possible to account for membrane binding and determine parameters for binding to the macromolecule,

simultaneous binding to the membrane and the macromolecule and the following expressions:



This leads to the following relationship between  $D_t$  and  $t$  similar to which enables the  $D_t$ ,  $t$  functional relationship to be evaluated using the 11.14 and 11.15 ( $j=1$ ):

$$D_t = \frac{s}{K_e} + \frac{k's}{K_e + K^*s} \quad (11.19)$$

$$t = \left[ \frac{1}{K_e} \ln s + \frac{K'}{K_e + K^*s} + \frac{K'}{K_e} \ln \frac{s}{K_e + K^*s} \right] \Bigg|_{s_0}^s \quad (11.20)$$

where:  $K' = \frac{Mn \cdot K^*}{WV} \quad (11.21)$

and  $M$  is the amount of membrane material available for binding,  $W$  is the molecular weight of the small molecule and  $V$  is the volume of the protein compartment. Equations 11.19 and 11.20 can then be used to determine  $K_e$  and the membrane binding parameters  $K^*$  and  $K'$  in the absence of protein using the technique described above.

Determination of drug-macromolecule binding parameters in the presence of membrane binding.

Once the parameters  $K_e$ ,  $K'$  and  $K^*$  have been determined using the above approach it is possible to account for membrane binding and determine parameters for binding to the macromolecule.

Simultaneous binding to the membrane and the macromolecule leads to the following expressions:

$$D_t = \frac{K's}{K_e + K^*s} + \frac{s}{K_e} + P_t s \sum_{i=1}^i \frac{n_i K_i}{K_e + K_i s} \quad (11.22)$$

$$t = \left[ \frac{K'}{K_e + K^*s} + \frac{K'}{K_e} \ln \frac{s}{K_e + K^*s} \right] \quad (11.23)$$

$$+ \frac{1}{K_e} \ln s + P_t \sum_{i=1}^j n_i K_i \left[ \frac{1}{K_e + K_i s} + \frac{1}{K_e} \ln \left( \frac{s}{K_e + K_i s} \right) \right] \Bigg|_{s_0}^s$$

which enables the  $D_t$ ,  $t$  functional relationship to be evaluated using the iterative procedure discussed.

By comparing Eqs. 11.22 and 11.23 with Eqs. 11.14 and 11.15 it is seen that in the presence of membrane binding the dialysis behaviour is mathematically analogous to a system where the small molecule is binding to two macromolecular species.

#### DATA TREATMENT AND RESULTS

Data for dialysis of chlorpropamide from bovine serum albumin (BSA) in the presence of  $1.6 \times 10^{-5}$  M free warfarin were treated according to the proposed method. Binding parameters  $n_1$ ,  $K_1$ ,  $n_2$ ,  $K_2$  and the dialysis rate constant  $K_e$  were estimated using the function relating  $D_t$  and  $t$  given in Eqs. 11.5 and 11.9 and evaluated using Eqs. 11.11-11.13 and 11.5. This function was programmed in a subroutine (scheme 11.1) which was executed with FUNFIT.

The least squares fit of the model using two classes of binding sites (Fig. 11.2) agreed very well with the experimental data ( $r = 0.99986$ ). The binding parameters estimated by the method are summarised in Table 11.1. The values for the number of binding sites  $n_1$  and  $n_2$  were found to be so close to 2 and 9 in a preliminary computation that their values were fixed as integers. Such a model has greater conceptual value and the slight change in  $n_1$  and  $n_2$  did not substantially alter the values of the other parameters.

Also included in Table 11.1 are values for binding parameters estimated by fitting dialysis data to a fourth order polynomial and then evaluation using a modification of the method of Hart (184) as described elsewhere (175). This method will be denoted derivative method I. A

SUBROUTINE MODEL (DT,X,P,IPRINT)  
DIMENSION X(2),P(8)

Scheme 11.1

C P=N1,K1,N2,K2,S0,KE,PT,MAX DT = TOTAL CONC. OF SMALL MOLECULES  
C 1 2 3 4 5 6 7 8 X(1) = TIME

The subroutine MODEL defines the functional relationship between  $D_t$  and  $t$  in a dynamic dialysis process. This function, given in parametric form by Eqs. 11.5 and 11.9, is calculated according to Eqs. 11.11-11.13 and 11.5. The functions PHI and PHIDER used in MODEL defines  $\phi(s)$  and  $\phi'(s)$  as given by Eqs. 11.13 and 11.12. The subroutine which is written specifically for FUNFIT enables least squares estimates of the binding parameters  $n_1, K_1, n_2, K_2$  (Eq. 11.1) and the dialysis rate constant  $K_e$  (Eq. 11.3) to be determined directly from dynamic dialysis data (Fig. 11.2). The subroutine specifies  $s$  as a second "independent variable",  $x(2)$ , which is used as an initial value in the iteration, Eq. 11.11, that in less than MAX cycles should determine  $s$  to an accuracy corresponding to an accuracy of  $t$  better than  $10^{-6}\%$ . The last two parameters ( $P(7)$  = total macromolecule concentration and  $P(8)$  = MAX) of the eight formal parameters are fixed during the least squares fitting procedure. A value of 50 for MAX should be sufficient to reach convergence (Eq. 11.11) even for pure starting values of  $P(1)$  to  $P(6)$ . However if convergence is not reached IPRINT is made equal to 1, the current values of the parameters are printed and the control is returned to the main program so that appropriate action can be taken.

DIMENSION P(8)  
A=ALOG(S/P(5))/P(6)  
B=P(5)-S  
C=P(6)+P(2)\*P(5)  
D=P(6)+P(2)\*S  
E=P(6)+P(4)\*P(5)  
F=P(6)+P(4)\*S  
PHI=T+A\*P(1)\*P(2)\*P(3)\*P(4)\*P(5)\*P(6)+  
P(1)\*P(2)\*P(4)\*P(5)\*P(6)/P(7)  
PHIDER=PHI  
RETURN  
END

SUBROUTINE MODEL (DT,X,P,IPRINT)  
 DIMENSION X(2),P(8)

C  
 C P=N1,K1,N2,K2,S0,KE,PT,MAX DT = TOTAL CONC. OF SMALL MOLECULES  
 C 1 2 3 4 5 6 7 8 X(1) = TIME  
 C X(2) = SLOPE ESTIMATE  
 C

S=X(2)  
 MAX=IFIX(P(8))  
 DO 1 I=1,MAX  
 SSAVE=S  
 PHIVAL=PHI(S,P,X(1))  
 S=S-PHIVAL/PHIDER(S,P)  
 IF(S.LE.0.)S=SSAVE/2.  
 TIME=X(1)  
 IF(TIME.EQ.0.)TIME=1.  
 IF(ABS(PHIVAL)/TIME.LT.1E-8) GO TO 2  
 1 CONTINUE  
 IPRINT=1  
 2 DT=S/P(6) + P(7)\*S\*(P(1)\*P(2)/(P(6)+P(2)\*S)+  
 P(3)\*P(4)/(P(6)+P(4)\*S))  
 IF(IPRINT.EQ.0)RETURN  
 WRITE(6,3)P  
 3 FORMAT(' N1,K1,N2,K2,S0,KE,PT,MAX =', (4E11.5))  
 RETURN  
 END

C  
 C  
 C FUNCTION PHIDER(S,P)  
 DIMENSION P(8)  
 PHIDER=1./(P(6)\*S) + (P(7)\*P(6)/S)\*(P(1)\*P(2)/((P(6)+P(2)\*S)\*\*2)+  
 P(3)\*P(4)/((P(6)+P(4)\*S)\*\*2))  
 +  
 RETURN  
 END

C  
 C  
 C FUNCTION PHI(S,P,T)  
 DIMENSION P(8)  
 A=ALOG(S/P(5))/P(6)  
 B=P(5)-S  
 C=P(6)+P(2)\*P(5)  
 D=P(6)+P(2)\*S  
 E=P(6)+P(4)\*P(5)  
 F=P(6)+P(4)\*S  
 PHI=T+A+P(7)\*P(1)\*P(2)\*(P(2)\*B/(C\*D)+A+ALOG(C/D)/P(6))+  
 + P(7)\*P(3)\*P(4)\*(P(4)\*B/(E\*F)+A+ALOG(E/F)/P(6))  
 RETURN  
 END

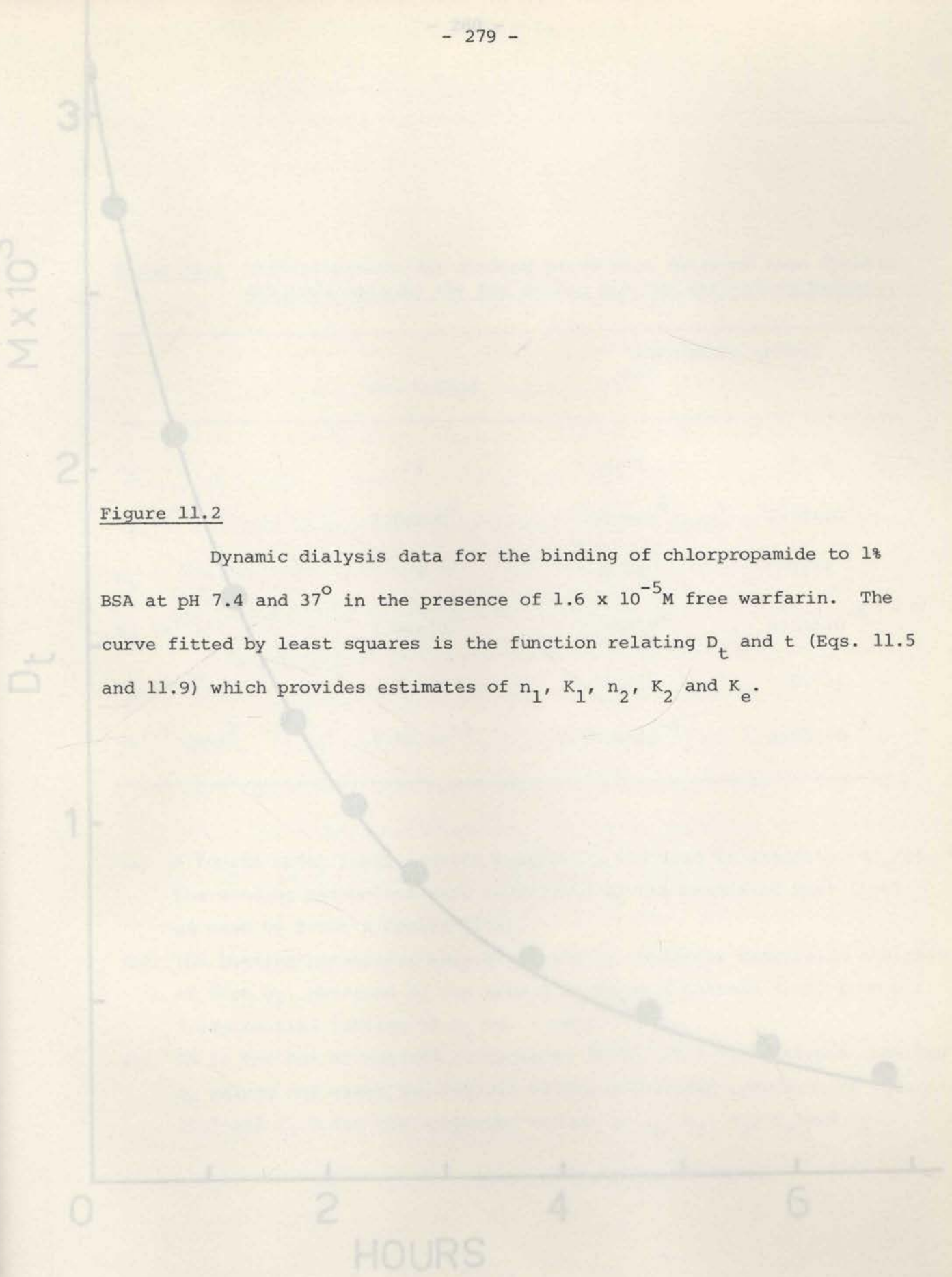


Figure 11.2

Dynamic dialysis data for the binding of chlorpropamide to 1% BSA at pH 7.4 and  $37^\circ$  in the presence of  $1.6 \times 10^{-5}$  M free warfarin. The curve fitted by least squares is the function relating  $D_t$  and  $t$  (Eqs. 11.5 and 11.9) which provides estimates of  $n_1$ ,  $K_1$ ,  $n_2$ ,  $K_2$  and  $K_e$ .



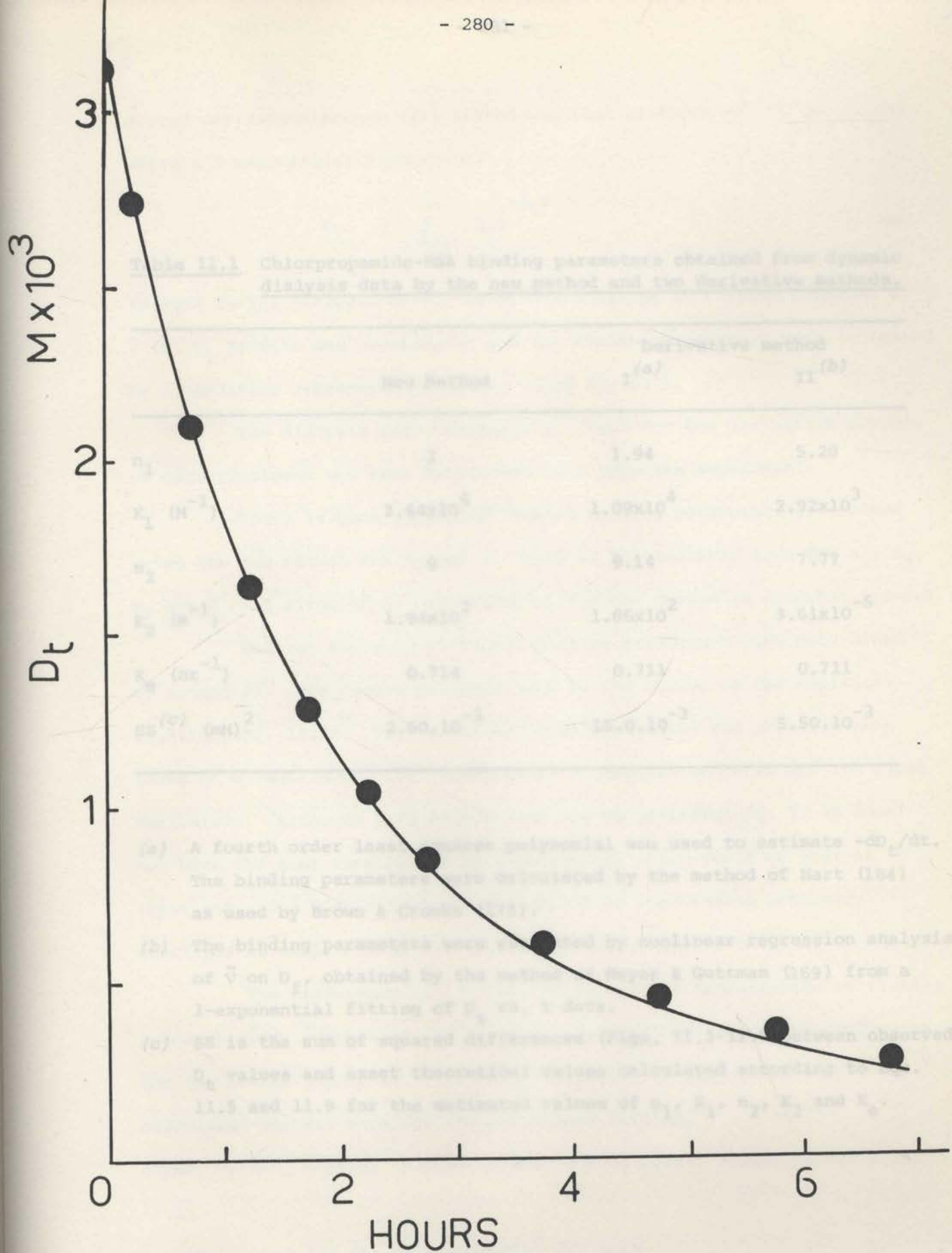


Table 11.1 Chlorpromazine-BSA binding parameters obtained from dynamic dialysis data by the new method and the derivative method.

Parameter	New Method	Derivative method	
		I (a)	II (b)
$n_1$	2	1.94	5.20
$K_1$ ( $M^{-1}$ )	$2.44 \times 10^5$	$1.09 \times 10^4$	$2.92 \times 10^3$
$n_2$	4	9.14	7.77
$K_2$ ( $M^{-1}$ )	$1.94 \times 10^2$	$1.06 \times 10^2$	$3.61 \times 10^{-5}$
$K_3$ ( $sec^{-1}$ )	0.711	0.711	0.711
$SS^{(c)}$ ( $M$ ) <sup>2</sup>	$2.50 \times 10^{-2}$	$15.0 \times 10^{-2}$	$5.50 \times 10^{-3}$

(a) A fourth order least squares polynomial was used to estimate  $-dD_t/dt$ . The binding parameters were calculated by the method of Hart (1961) as used by Brown & Crook (1971).

(b) The binding parameters were calculated by nonlinear regression analysis of  $\bar{v}$  on  $D_{EF}$  obtained by the method of Taylor & Gotsman (1969) from a 1-exponential fitting of  $D_t$  vs.  $t$  data.

(c)  $SS$  is the sum of squared differences (Fig. 11.1) between observed  $D_t$  values and exact theoretical values calculated according to eqs. 11.5 and 11.9 for the estimated values of  $n_1$ ,  $K_1$ ,  $n_2$ ,  $K_2$  and  $K_3$ .

second derivative method (11) tested was that of Meyer and Guttman (169) where a 3-exponential expression:

$$D_t = \sum_{i=1}^3 A_i e^{-\lambda_i t}$$

Table 11.1 Chlorpropamide-BSA binding parameters obtained from dynamic dialysis data by the new method and two derivative methods.

	New Method	Derivative method	
		I (a)	II (b)
$n_1$	2	1.94	5.28
$K_1$ ( $M^{-1}$ )	$2.64 \times 10^4$	$1.09 \times 10^4$	$2.92 \times 10^3$
$n_2$	9	9.14	7.77
$K_2$ ( $M^{-1}$ )	$1.94 \times 10^2$	$1.86 \times 10^2$	$3.61 \times 10^{-5}$
$K_e$ ( $Hr^{-1}$ )	0.714	0.711	0.711
SS (c) ( $mM$ ) <sup>2</sup>	$2.60 \cdot 10^{-3}$	$15.0 \cdot 10^{-3}$	$5.50 \cdot 10^{-3}$

(a) A fourth order least squares polynomial was used to estimate  $-dD_t/dt$ . The binding parameters were calculated by the method of Hart (184) as used by Brown & Crooks (175).

(b) The binding parameters were estimated by nonlinear regression analysis of  $\bar{V}$  on  $D_f$ , obtained by the method of Meyer & Guttman (169) from a 3-exponential fitting of  $D_t$  vs.  $t$  data.

(c) SS is the sum of squared differences (Figs. 11.3-11.5) between observed  $D_t$  values and exact theoretical values calculated according to Eqs. 11.5 and 11.9 for the estimated values of  $n_1$ ,  $K_1$ ,  $n_2$ ,  $K_2$  and  $K_e$ .

0.286, -0.317, -0.519, -0.0209, 8.46, 0.77, 0.381, -2.49, -1.42, 3.14,

-1.13

and the values for the triexponential fit were:

1.73, 1.91, 1.46, 1.36, -1.13, -1.05, -1.31, -1.55, -2.19, 1.10, 13.7,

second derivative method (II) tested was that of Meyer and Guttman (169) where a 3-exponential expression:

$$D_t = \sum_{i=1}^3 A_i e^{-\alpha_i t} \quad (11.24)$$

is used to fit dialysis data. By differentiation at various  $t$  values a  $\bar{V}$  vs.  $D_f$  profile was constructed and the binding parameters were estimated by a nonlinear regression technique using Eq. 11.1.

The dialysis rate constant,  $K_e$ , used for the derivative methods of data treatment was that determined in a separate experiment.

There is good agreement between binding parameters determined using the new method and method I. This is particularly true for  $n_1$ ,  $n_2$ ,  $K_2$  and  $K_e$  and although  $K_1$  determined by the new method is somewhat greater.

However entirely different binding parameters were determined by method II. The reason probably lies in the choice of the empirical equation, Eq. 11.24. The selection of this equation has probably been based on a requirement for "smoothness" of the fitted curve and its first derivative. Although this requirement may be satisfied, Eq. 11.24 does not have the same flexibility as a polynomial which could be just as important. The flexibility of polynomials to approximate arbitrary functions is explained by the well-known Taylor series theorem.

The difference in flexibility is clearly demonstrated by fitting a fourth order polynomial and 11.24 to exact dialysis data obtained using the new method (Table 11.1). The  $D_t$  residuals (expressed as % of calculated values) were for the polynomial fitting:

0.286, -0.217, -0.519, -0.0609, 0.466, 0.779, 0.381, -1.49, -1.42, 3.14, -1.12

and the values for the triexponential fit were:

1.73, 1.91, 1.46, 3.38, -1.19, -2.65, -3.91, -4.55, -2.19, 3.10, 10.7.

This shows that the polynomial is considerably more flexible. The triexponential fitting resulted in significant systematic deviation in residuals leading to bias in the slope values and  $D_t$  values and therefore a bias in the final results. The residual sum of squares ( $\text{mM}^2$ ) were  $5.85 \cdot 10^{-4}$  and  $1.04 \cdot 10^{-2}$  for the polynomial and triexponential fitting respectively.

A further disadvantage of using 11.24 is that multiple solutions are possible because this equation is nonlinear (in  $\alpha$ ) and therefore may result in several sum-of-squares minima. This is not the case with a polynomial which has a *unique* least squares solution.

The fitting of 11.24 to the generated, exact  $D_t$ ,  $t$  data was repeated several times with different initial estimates for  $A_i$  and  $\alpha_i$  but the same solution was obtained each time suggesting that the above fit is the best possible using the triexponential equation.

The use of a least squares polynomial to represent dialysis data, is on the other hand, expected to be less suitable than the triexponential when the experimental errors are large or where there are significant 'gaps' between observation points. This is due to the fact that the ordinary least squares polynomial fitting completely disregards derivative values. The derivative values are commonly found to be in large error at first and last observation points and just before or after 'gaps' in the data. This disadvantage of polynomials can be reduced considerably by imposing constraints on the derivative values by using least squares spline polynomials.

Such polynomials will compete favourably with 11.24 on data with large errors and 'gaps' particularly considering the fact that the problem of multiple minima, using 11.24, is much larger for large residual problems.

However, no matter which empirical equation is used the results obtained will theoretically never be as exact as obtained using the true equation as in fact done in the proposed method.

To investigate the bias introduced by using method I and II the exact  $D_t, t$  profile was calculated using the parameter values (Table 11.1) obtained using the two methods. The differences between the observed  $D_t$  values and the calculated values (Figs. 11.4 and 11.5) shows that the residuals are significantly larger in those methods than in the new method (Fig. 11.3). The residuals from method I are particularly biased in a positive direction although their pattern resembles that from the new method (Fig. 11.3).

On the other hand the residuals from method II (Fig. 11.5) show an entirely different pattern consistent with the fact that the binding parameters obtained using method II represent an entirely different solution.

A final check on the bias introduced by method I and II was made by applying them to exact  $D_t, t$  data generated from the parameter values (Table 11.1) obtained using the new method.

Method I found binding parameters relatively close to the true values although  $K_1$  seems to be somewhat different (Table 11.2). Method II found however an entirely different solution that is similar to the solution obtained using the real experimental data.

The value for the dialysis rate constant,  $K_e$ , estimated by the new method agrees very well with the value determined experimentally (Table 11.1). As stated previously this may not always be true as the permeability of the membrane may change between experiments.

Chlorpropamide does not appear to be significantly membrane bound.

Figure 11.3

Differences between observed  $D_t$  values and exact theoretical values calculated according to Eqs. 11.5 and 11.9 for the values of  $n_1$ ,  $K_1$ ,  $n_2$ ,  $K_2$  and  $K_e$  (Table 11.1) estimated by the proposed method.

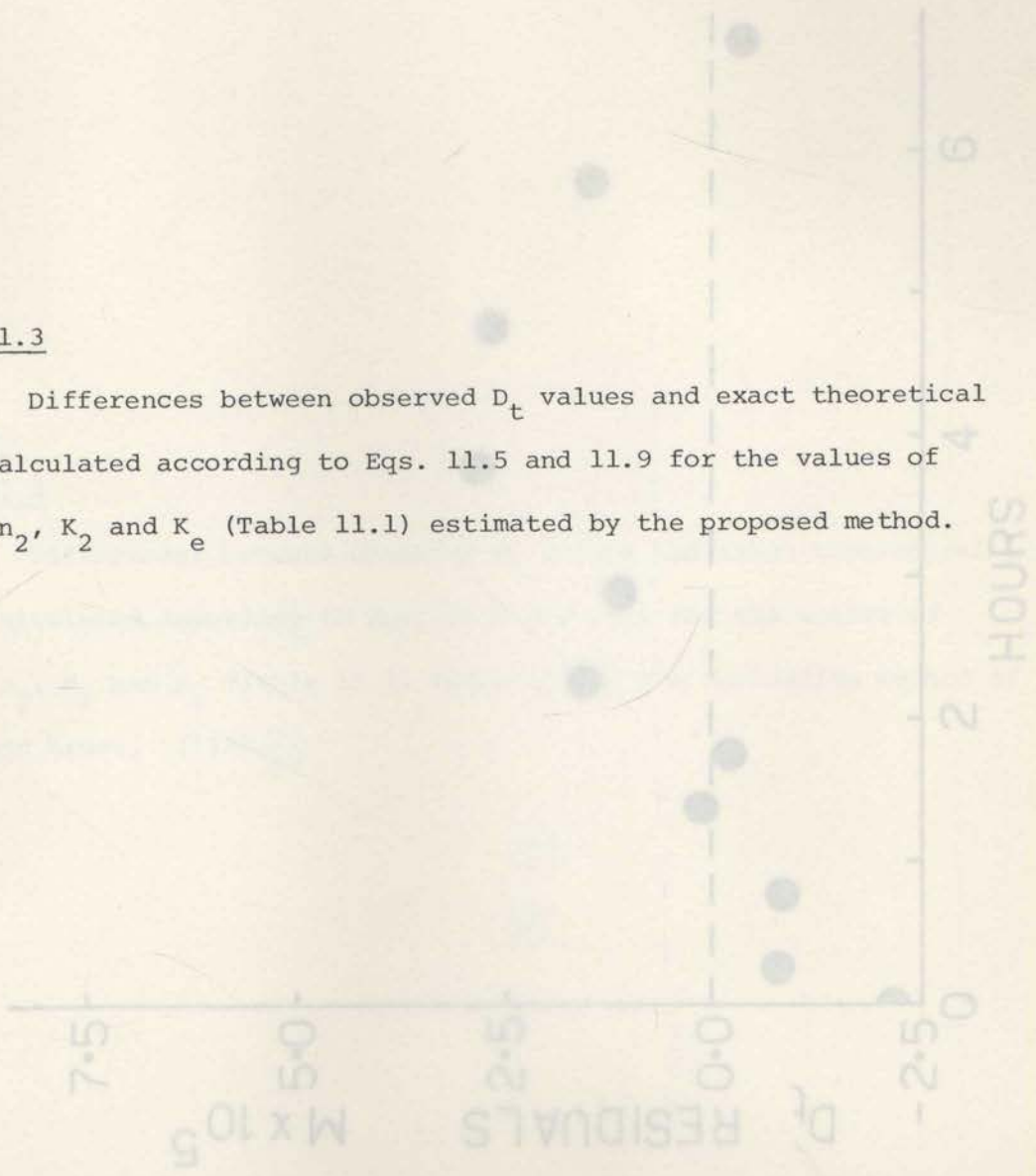


Figure 11.4

Differences between observed  $n_t$  values and exact theoretical values calculated according to Eqs. 11.7 and 11.9 for the values of  $n_1, K_1, n_2, K_2$  and  $K_0$  (Table 11.3) calculated by the derivative method of Crooks and Brown. (175)

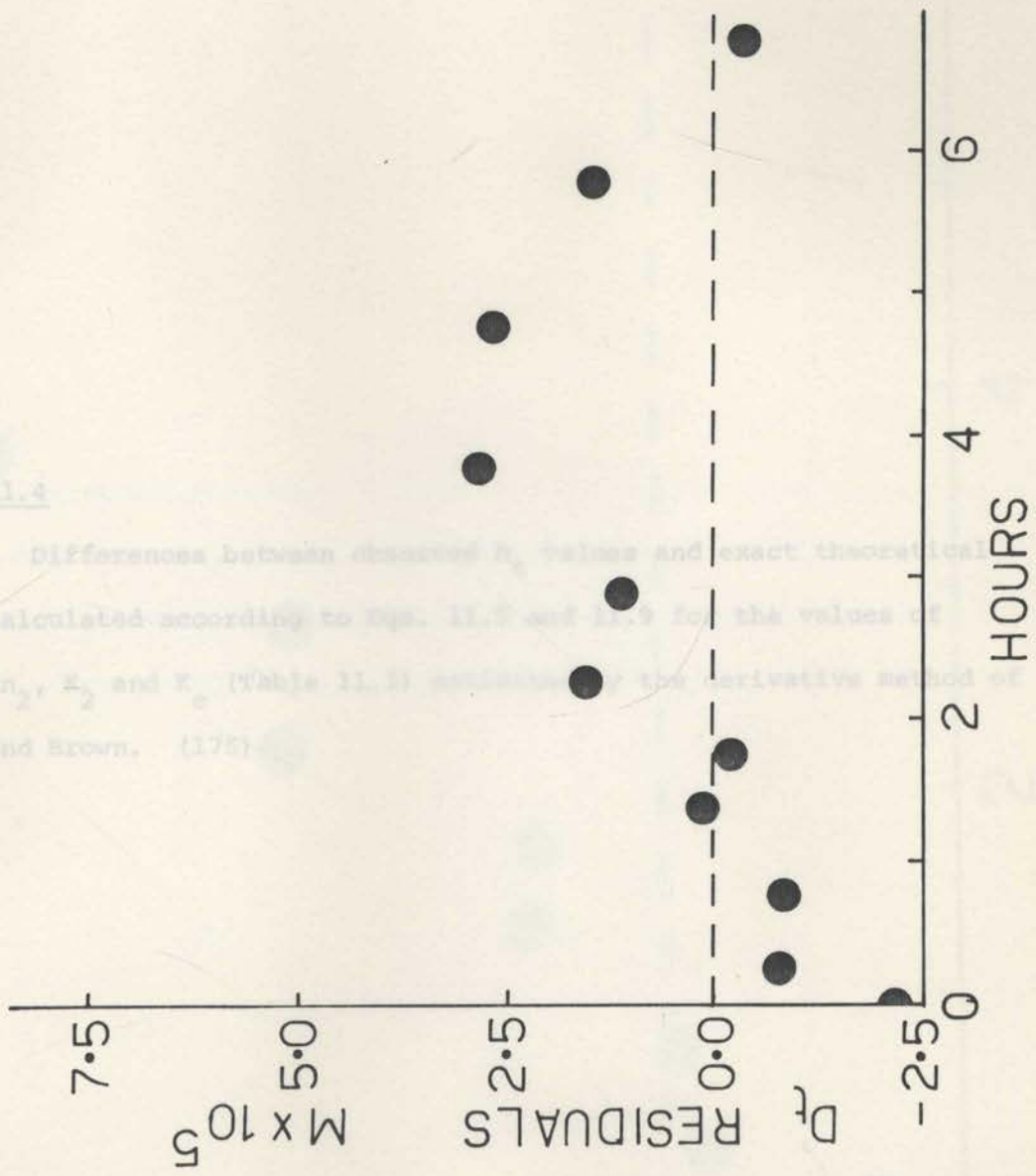


Figure 11.4

Differences between observed  $D_t$  values and exact theoretical values calculated according to Eqs. 11.5 and 11.9 for the values of  $n_1$ ,  $K_1$ ,  $n_2$ ,  $K_2$  and  $K_e$  (Table 11.1) estimated by the derivative method of Crooks and Brown. (175)

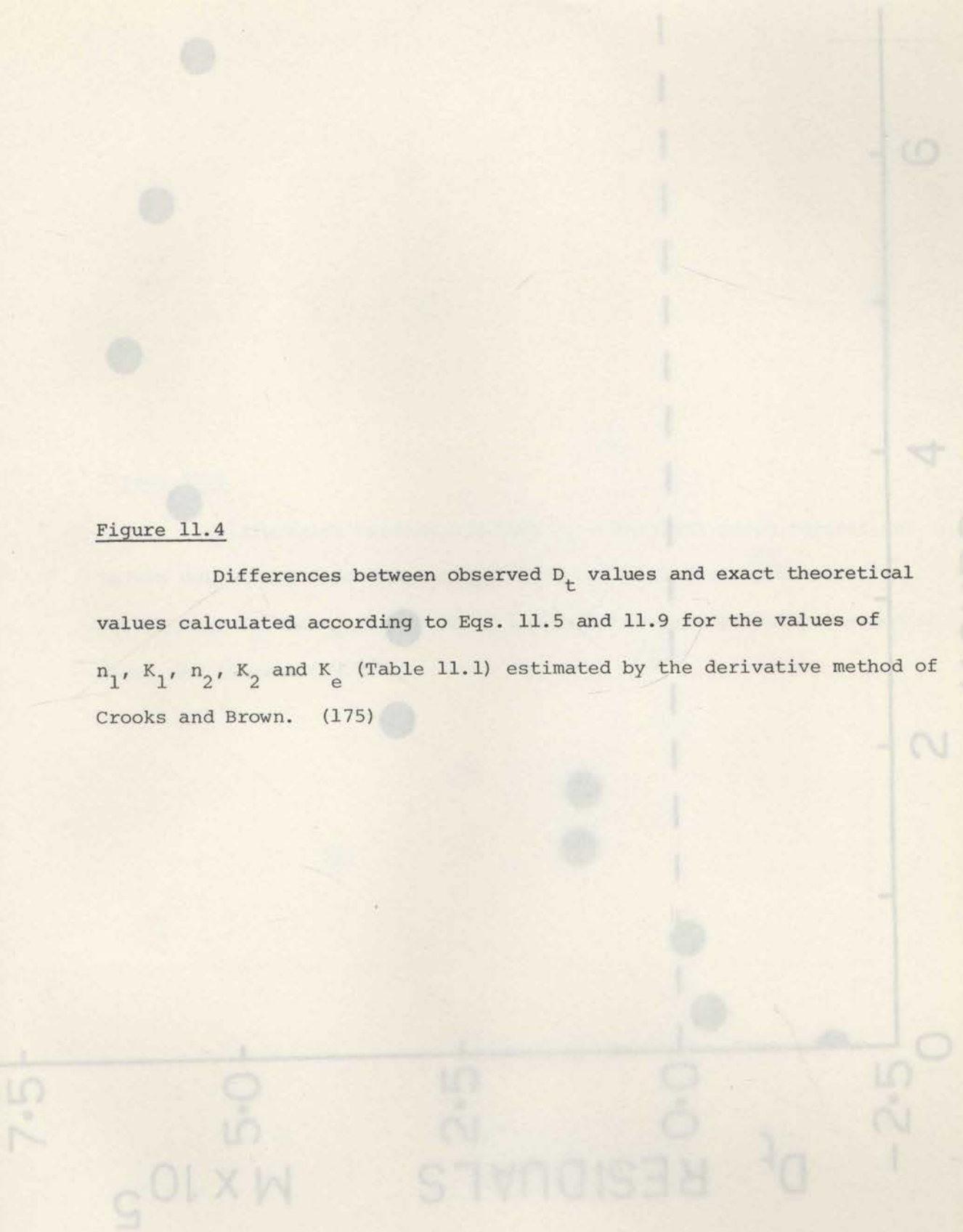




Figure 11.3

Differences between observed  $D_t$  values and exact theoretical values calculated according to Eqs. 11.1 and 11.2 for the values of  $n_1, K_1, n_2, K_2$  and  $K_0$  (Table 11.1) estimated by the derivative method of Meyer and Gattman. (16)

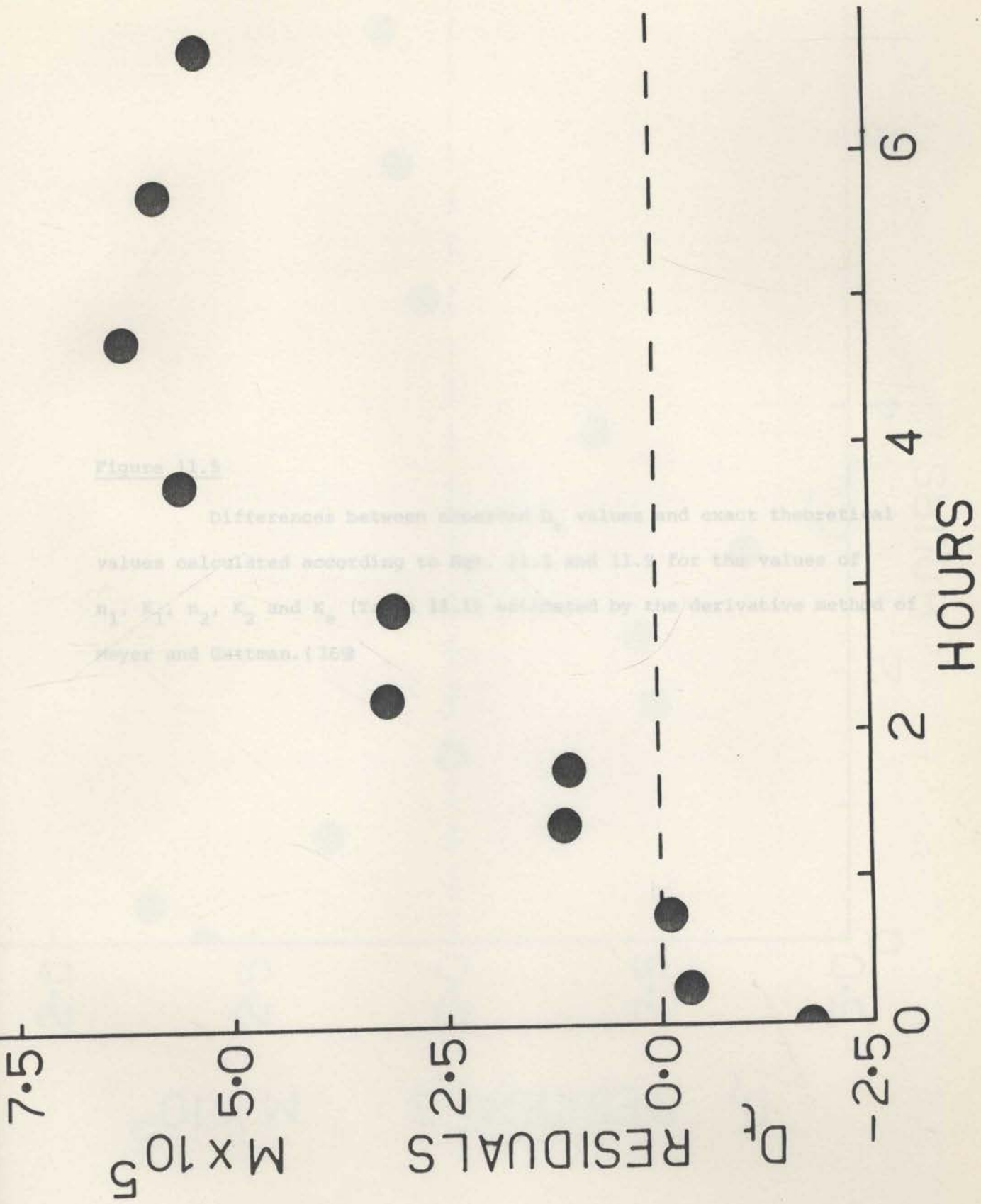
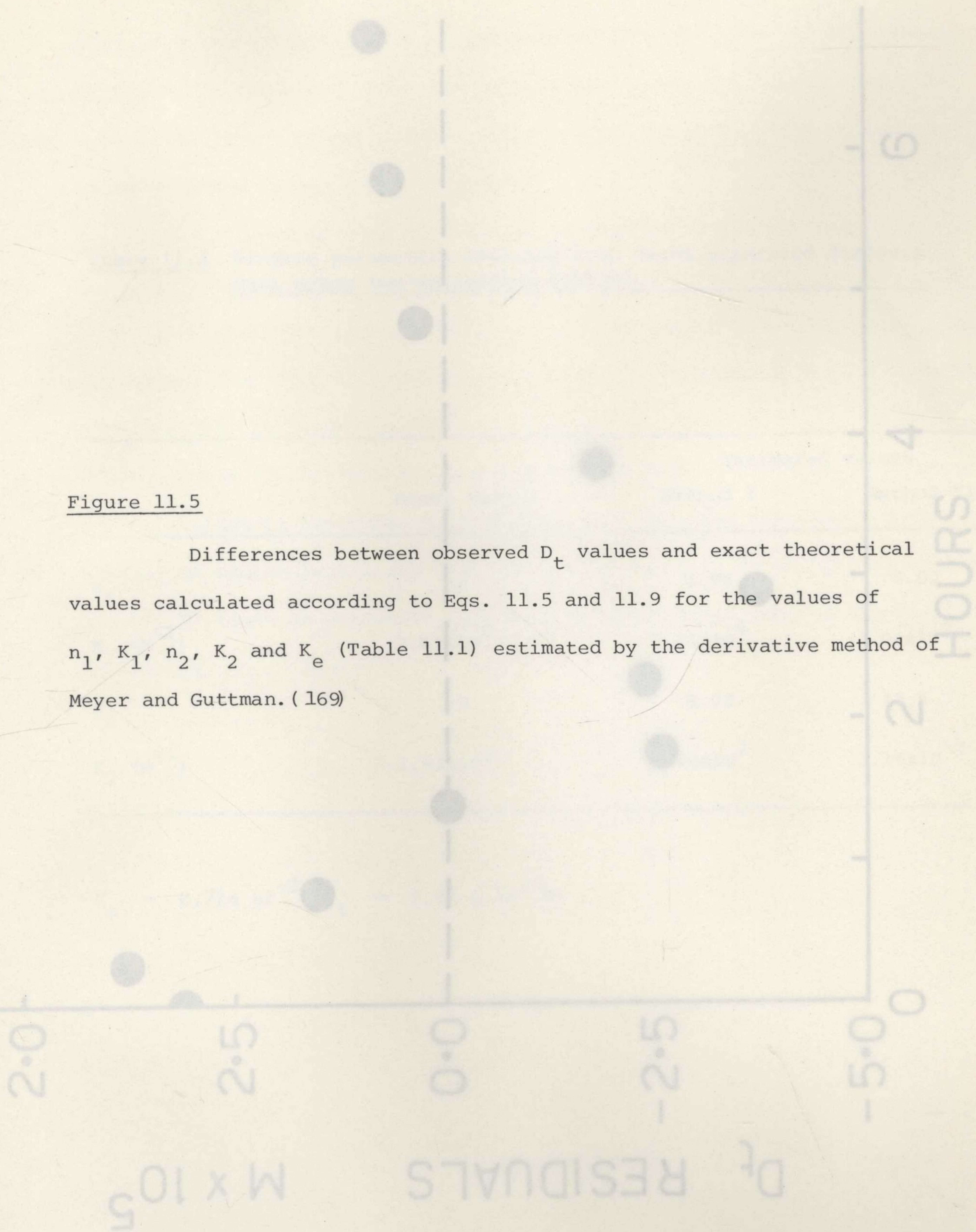


Figure 11.5

Differences between observed  $D_t$  values and exact theoretical values calculated according to Eqs. 11.5 and 11.9 for the values of  $n_1$ ,  $K_1$ ,  $n_2$ ,  $K_2$  and  $K_e$  (Table 11.1) estimated by the derivative method of Meyer and Guttman. (169)



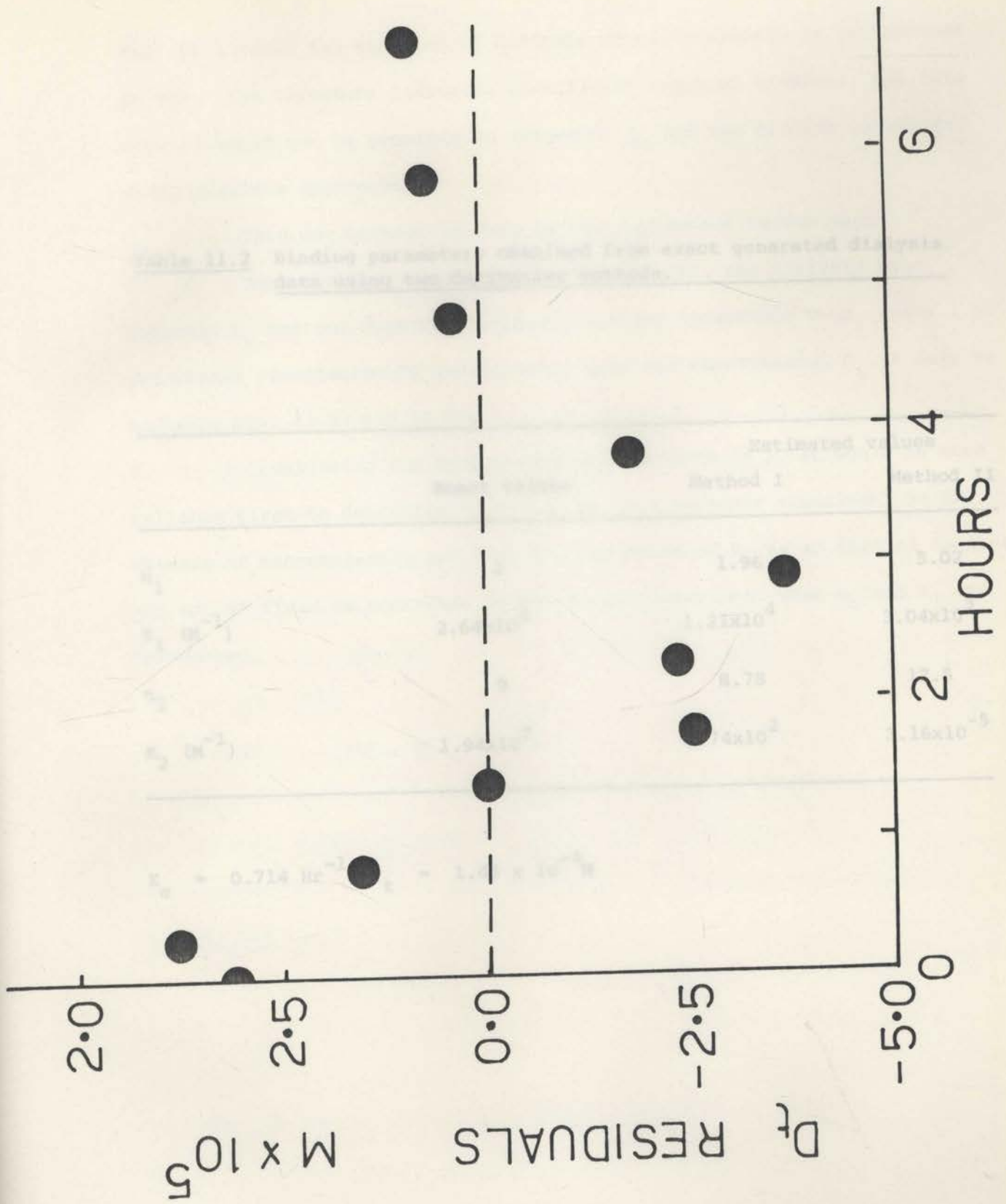


Fig. 11.1 shows the kinetics of dialysis of glibenclamide in the absence of BSA. The curvature indicates significant membrane binding. For this drug it would not be possible to determine  $K_1$  and the binding parameters using previous approaches.

Table 11.2 Binding parameters obtained from exact generated dialysis data using two derivative methods.

1. constant  $K_1$  and the drug-macromolecule binding parameters  $n_1, K_1$  can all be determined simultaneously and directly from the experimental  $Q_t, V$  data by applying Eqs. 11.22 and 11.23.

2. Considering the many parameters involved in the model, it is more reliable first to determine  $K_1, K^*$  and  $K^*$  in a separate experiment in the

absence of macromolecules and then use the value of  $K_1$  as an initial estimate

and  $K^*, K^*$  fixed as constants in the second experiment when  $n_1$  and  $K_1$  are determined.

	Exact values	Estimated values	
		Method I	Method II
$n_1$	2	1.96	5.02
$K_1 (M^{-1})$	$2.64 \times 10^4$	$1.21 \times 10^4$	$3.04 \times 10^3$
$n_2$	9	8.78	12.5
$K_2 (M^{-1})$	$1.94 \times 10^2$	$1.74 \times 10^2$	$3.16 \times 10^{-5}$

$$K_e = 0.714 \text{ Hr}^{-1}, P_t = 1.45 \times 10^{-4} \text{ M}$$

Fig. 11.1 shows the kinetics of dialysis of glibenclamide in the absence of BSA. The curvature indicates significant membrane binding. For this drug it would not be possible to determine  $K_e$  and the binding parameters using previous approaches. 57, 1825 (1968)

1. ~~Worst~~ This can however be done by the new method in two ways:

1. ~~Worse~~ The membrane binding parameters,  $K'$ ,  $K^*$ , the dialysis rate constant  $K_e$  and the drug-macromolecule binding parameters  $n_i, K_i$  can all be determined simultaneously and directly from the experimental  $D_t, t$  data by applying Eqs. 11.22 and 11.23. J. Pharm. Sci., 58, 611 (1969).

2. ~~Worse~~ Considering the many parameters involved in 1. it would be more reliable first to determine  $K_e, K'$  and  $K^*$  in a separate experiment in the absence of macromolecule and then use the value of  $K_e$  as an initial estimate and  $K', K^*$  fixed as constants in the second experiment when  $n_i$  and  $K_i$  are determined. E.L., Wurster, D.E. and Higuchi, T., J. Pharm. Sci., 44,

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APPENDIX

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$s_i$  i-th residual value.

$E(\cdot)$  expected value.

$E_s(w)$  mean particle weight of size  $s$ .

- o o o -

$\bar{w}$  =  $\frac{1}{N} \sum_{i=1}^N w_i$  = mean particle weight.

$\bar{w}_s$  =  $\frac{1}{N} \sum_{i=1}^N w_i^s$  = mean particle weight of size  $s$ .

$\bar{w}_s$  =  $\frac{1}{N} \sum_{i=1}^N w_i^s$  = mean particle weight of size  $s$ .

$\bar{w}_s$  =  $\frac{1}{N} \sum_{i=1}^N w_i^s$  = mean particle weight of size  $s$ .

$\bar{w}_s$  =  $\frac{1}{N} \sum_{i=1}^N w_i^s$  = mean particle weight of size  $s$ .

A P P E N D I X

Table of Symbols

$g$	acceleration of gravity.
$a$	diameter or equivalent spherical diameter.
$\bar{a}$	mean diameter.
$A$	surface area.
$\alpha$	acute angle of crystal (p. 69).
$b_0$	side length of crystal ( $t=0$ ).
$c_0$	solubility of compound in unionised form.
$c_b$	solute bulk concentration.
$c_i$	interfacial concentration of solute.
$c^*$	dimensionless concentration.
$COV( )$	covariance.
$d_0$	initial diameter of the smallest particle in a powder.
$D_0$	initial diameter of the largest particle in a powder.
$D$	diffusion coefficient.
$D_t$	total concentration of small molecule.
$D_f$	free concentration of small molecule.
$\nabla^2$	Laplacian operator.
$e_i$	$i$ -th residual value.
$E( )$	expected value.
$E_t(w)$	mean particle weight of time $t$ .
$erfc(x) = 1 - erf(x)$	complementary error function.
$erf(x) = \frac{2}{\sqrt{\pi}} \int_0^x e^{-u^2} du$	error function.
$f( )$	particle weight density function of time $t$ .
$F$	shape ratio.
$F(x) = \frac{1}{\sqrt{2\pi}} \int_{-\infty}^x e^{-u^2/2} du$	cumulative normal distribution function.
$F_{\alpha, n, p}$	critical point of $F$ -distribution.

$\psi$	stream function, time function.
$g$	acceleration of gravity.
$g(\ )$	single particle dissolution function (p. 97).
$g^{-1}(\ )$	inverse single particle dissolution function (p. 97).
$\gamma$	mean rate of surface renewal (p. 51).
$h$	diffusion layer thickness.
$H$	Hessian matrix.
$H^+$	$H_3O^+$ concentration.
$H_{ii}$	$i$ -th diagonal element of Hessian matrix.
$i$	lower truncation parameter (Fig. 5.1)
$j$	upper truncation parameter (Fig. 5.1)
$J$	interfacial mass flux (p. 66).
$J_D$	diffusion flux.
$k_i$	effective interfacial transport rate constant (p. 52).
$k_m$	rate parameter for model $m$ .
$K_a$	dissolution constant of acid.
$K_b$	dissociation constant of base.
$K_e$	dialysis rate constant (Chapter 11).
$K_i$	association constant for $i$ -th class (Chapter 11)
$K_m$	Michaelis Menten parameter (Chapter 11), rate parameter for model $m$ .
$K_w$	ion product of water.
$K^*$	specific dissolution rate parameter, association constant for membrane binding.
$l_o$	sidelength of crystal (Fig. 4.4).
$l_o(\ )$	initial particle size distribution.
$L(\ )$	likelihood function.



$m$	model parameter.
$m_o$	initial weight of the smallest particle.
$M_o$	initial weight of the largest particle.
$\mu$	logarithmic mean (Fig. 5.1). viscosity (Chapter 4).
$n_i$	number of sites in i-th class (Chapter 11)
$N( )$	normal distribution function (p. 101).
$N_t$	number of particles at time t.
$\bar{V}$	number of moles of small molecules bound per mol macromolecule.
$\bar{V}^*$	amount of small molecule bound per amount of available membrane material.
$\underline{P}$	parameter vector.
$P$	operator (defined p. 99).
$P_t$	total concentration of macromolecule (Chapter 11).
$\pi =$	3.1415...
$r_t$	radius of particle at time t.
$r^*$	dimensionless radius.
$\rho$	density
$\rho_{i,j}$	correlation coefficient between i-th and j-th element.
$R_e$	Reynolds number.
$s$	distance to fixed point (Chapter 4) dispersion product (Chapter 6) variable defined p. 269.
$Sc$	Schmidt number.
$SD_i$	standard deviation of i-th element.
$SS$	sum of squared residuals.
$S_N( )$	cumulative distribution of residuals.
$\sigma$	standard deviation of normal distribution (Fig. 5.1.)

- t time.
- $t^*$  dimensionless time (Chapter 4).
- $t_{\alpha,n}$  critical value of t-distribution.
- $V_{\infty}$  vertical velocity of free falling spherical particle.
- V variance-covariance matrix.
- $V_m$  Michaelis Menten parameter.
- Var( ) variance.
- w weight of single particle.
- W weight of undissolved powder.
- $\theta$  spherical coordinate (Chapter 4).
- $\binom{x}{y} = \frac{x!}{y!(x-y)!}$  binomial coefficient.

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PROGRAM FUNFIT(INPUT,OUTPUT,TAPE5=INPUT,TAPE6=OUTPUT)          10
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FUNFIT IS AN INTERACTIVE TIME-SHARING PROGRAM FOR GENERAL      20
NONLINEAR REGRESSION AND CURVE FITTING                          30
THE PROGRAM CONSISTS OF-                                       40
MAIN PROGRAM / FUNFIT                                          50
SUBROUTINES / NELDOR,CHOL,SYMINV,LSQ,XYPLOT,PLOT,              60
/ SEARCH,READ,PLACE,PROB1,KOLMIR                               70
FUNCTIONS / SIGNOF,WEIGHT,SEPRO,NBC,SDF                         80
A SUBROUTINE MODEL(Y,X,P,IPRINT) MUST BE SUPPLIED BY THE USER 100
TO DEFINE THE FUNCTION(S) TO BE FITTED. FOR DETAILS ABOUT THIS 110
SUBROUTINE CONTACT P.VENG PEDERSEN,THE PHARMACY DEPARTMENT    120
THE UNIVERSITY OF SYDNEY,N.S.W. 2006,AUSTRALIA.                130
-----
DIMENSION F(20),F1(20),F2(20),FSAVE(20),NF(20),FS(20),NT(20), 140
*STPS(20),STEP(20),STEPS(20),VAR(20),TCRIT(30),X(100),Z(100), 150
*Y(100),WY(100),WYN(100),RES(100),DIFFCT(100),YEST(100),IOUTL(10), 160
*VARCOV(55),INDEX(100),WRRES(100),FNEW(7),STPNEW(7),NRECOM(7), 170
*NCRIT(7),VC(55),A(650),FMIN(20),AR(12),XX(9,100),G(9),LABEL(100), 180
*ITHSET(10),LIM1(10),LIM2(10),RSS(10),WRSS(10)                190
DOUBLE PRECISION VC                                           200
COMMON XMIN,XMAX                                              210
COMMON /FUNCOM/ITHFUN                                         220
COMMON /DATA/ XX,Y,WYN,NOBS                                   230
COMMON /PARLM/ F1,F2                                          240
COMMON /CONSTR/LIMITS                                         250
COMMON /B1/ X                                                 260
COMMON /B2/ Z                                                 270
COMMON /B3/VARCOV,VC,NEVAL1,NEVAL2,AMINO                     280
COMMON /B4/ NIND                                              290
COMMON /B5/NG,NS,IGO                                          300
COMMON /B6/ NOP                                               310
COMMON /B7/ ITABLE                                            320
COMMON /B8/NFINC                                              330
COMMON /B9/LIM1,LIM2                                          340
COMMON /B10/LABEL                                             350
COMMON /B11/AR                                                360
COMMON /B12/NPR                                               370
COMMON /B13/IGNORE                                            380
COMMON /B14/NVIOL                                             390
COMMON /B15/SKIP                                              400
LOGICAL SKIP,SKIP                                            410
EQUIVALENCE (AR(1),ANS)                                       420
EXTERNAL LSQ                                                  430
DATA TCRIT/12.7,4.30,3.18,2.78,2.57,2.45,2.36,2.31,2.26,2.23, 440
*2.20,2.18,2.16,2.14,2.13,2.12,2.11,2.10,2.09,2.09,2.08,    450
*2.07,2.07,2.06,2.06,2.06,2.05,2.05,2.04,2.04/              460
DATA NRECOM /0,0,343,625,1024,729,2187 /                      470
DATA NCRIT /0,0,8,16,32,64,128 /                              480
WRITE(6,500)                                                  490
LINES=30                                                       500
NRUN=0                                                         510
ITABLE=0                                                       520
LIMITS=0                                                       530
NCOVR=0                                                        540
NDUMMY=0                                                       550
NS=1                                                           560
NFUNC=-1                                                       570
NOP=-1                                                         580
NOP2=-1                                                        590
NOBS2=-1                                                       600

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IQUAD=1                                                       620
SIMP=1E-6                                                      630
ANS4=2HNO                                                      640
PLOTRES=2HNO                                                  650
MAX=800                                                        660
NLOOP=20                                                       670
IPRINT=-1                                                      680
ANS11=2HNO                                                     690
ANS12=2HNO                                                     700
STOPCR=0.001                                                  710
NIND=1                                                         720
PLT=2HNO                                                       730
ANS16=2HNO                                                     740
5 NO=1                                                         750
WRITE(6,505)                                                  760
IGNORE=0                                                       770
CALL READ(1,AR)                                               780

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	J=IOUTL(I)	9300
370	WRITE(6,822)J	9310
	IF(NOUTL.EQ.1) WRITE(6,824)	9320
	IF(NOUTL.GT.1) WRITE(6,826)	9330
3701	CONTINUE	9340
	IF(ANS16.EQ.2HNO.OR.NOHWGT.EQ.1) GO TO 3704	9350
	WRITE(6,8241)T	9360
	NOUTL=0	9370
	STAR=2H *	9380
	NSAVE=1	9390
	DO 3703 I=1,NOBS	9400
	N=LABEL(I)	9410
	IF(N.EQ.NSAVE+1) WRITE(6,8240) N	9420
	NSAVE=N	9430
	WNRDV=WRES(I)/WSDVRS	9440
	IF(ABS(WNRDV).LT.T) GO TO 3702	9450
	NOUTL=NOUTL+1	9460
	WRITE(6,8242)I,STAR,WY(I),WYN(I),WRES(I),WNRDV	9470
	GO TO 3703	9480
3702	WRITE(6,8243)I,WY(I),WYN(I),WRES(I),WNRDV	9490
3703	CONTINUE	9500
	WRITE(6,8244)	9510
	IF(NOUTL.EQ.1) WRITE(6,8245)	9520
	IF(NOUTL.GT.1) WRITE(6,8246)	9530
3704	CONTINUE	9540
	IF(NIND.GT.1.OR.PLT.NE.3HYES) GO TO 371	9550
	CALL XYPL0T(F)	9560
	DO 3709 II=1,NFUNC	9570
	N1=LIM1(II)	9580
	N2=N1+1	9590
	N3=LIM2(II)	9600
	NN=N3-1	9610
	XMAX=X(N1)	9620
	XMIN=X(N1)	9630
	DO 3705 I=N2,N3	9640
	IF(X(I).GT.XMAX) XMAX=X(I)	9650
	IF(X(I).LT.XMIN) XMIN=X(I)	9660
3705	DELTA=(XMAX-XMIN)/50.	9670
	IF(II.EQ.1) WRITE(6,8261)	9680
	IF(II.GT.1) WRITE(6,8264)II	9690
	DO 3706 I=1,26	9700
	J=26+I	9710
	XP1=XMIN+FLOAT(I-1)*DELTA	9720
	XP2=XMIN+FLOAT(J-1)*DELTA	9730
	DO 3708 L=N1,NN	9740
	IF(XP1.GT.X(L).AND.XP1.LE.X(L+1))	9750
	YP1=Y(L)+(Y(L+1)-Y(L))*(XP1-X(L))/(X(L+1)-X(L))	9760
3708	IF(XP2.GT.X(L).AND.XP2.LE.X(L+1))	9770
	YP2=Y(L)+(Y(L+1)-Y(L))*(XP2-X(L))/(X(L+1)-X(L))	9780
	ITHFUN=II	9790
	CALL MODEL(YP1,XP1,F,0)	9800
	CALL MODEL(YP2,XP2,F,0)	9810
	IF(I.EQ.26) GO TO 3707	9820
3706	WRITE(6,8262) I,XP1,YP1,J,XP2,YP2	9830
3707	WRITE(6,8262) I,XP1,YP1	9840
3709	CONTINUE	9850
	WRITE(6,8263)	9860
371	CONTINUE	9870
	IF(IQUAD.NE.1) GO TO 376	9880
	WRITE(6,827)	9890
	DO 375 K=1,2	9900
	IF(K.EQ.2) WRITE(6,8270)	9910

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	L=1	9920
372	IF(L.GT.NPR) GO TO 375	9930
	II=L+(L-1)/2	9940
	DO 373 I=L,NPR	9950
	IA=II+L	9960
	II=II+I	9970
	IB=MIND(II,IA*5)	9980
	IF(K.EQ.2) GO TO 3721	9990
	DO 3720 J=IA,IB	10000
3720	VARCOV(J)=2.*FUNC*VARCOV(J)/FLOAT(NRSDF)	10010
	WRITE(6,829)(VARCOV(J),J=IA,IB)	10020
	GO TO 373	10030
3721	DO 3722 KK=IA,IB	10040
3722	VARCOV(KK)=VC(KK)	10050
	WRITE(6,829)(VARCOV(J),J=IA,IB)	10060
373	CONTINUE	10070
	L=L+6	10080

L=I*(I-1)/2+J	18600
BMAT(L)=2D0*((HSTST+A0)-(AVAL(I)+AVAL(J)))	18610
208 CONTINUE	18620
207 CONTINUE	18630
L=0	18640
DO 210 I=1,NAP	18650
I1=I+1	18660
L=L+1	18670
BMAT(L)=2D0*((H(I1)+A0)-2D0*AVAL(I))	18680
210 CONTINUE	18690
DO 237 I=1,NAP	18700
I1=I+1	18710
237 AVAL(I)=2D0*AVAL(I)-(H(I1)+3D0*A0)/2D0	18720
DO 219 I=1,NOP	18730
219 PMIN(I)=G(I,I)	18740
DO 211 I=1,NAP	18750
I1=I+1	18760
DO 211 J=1,NOP	18770
G(I1,J)=G(I1,J)-G(I,J)	18780
211 CONTINUE	18790
DO 212 I=1,NAP	18800
I1=I+1	18810
DO 212 J=1,NOP	18820
G(I,J)=G(I1,J)	18830
212 CONTINUE	18840
CALL SYMINV(BMAT,NAP,BMAT,TEMP,NULLTY,IFAU)LT)	18850
IF(IFAU)LT.NE.0) GO TO 450	18860
IRANK = NAP-NULLTY	18870
GO TO 441	18880
450 WRITE(IW,302)	18890
302 FORMAT(49H MATRIX TO BE INVERTED NOT POSITIVE SEMI-DEFINITE)	18900
IF(NVIOL.GT.0.AND.IPRINT.EQ.-1) WRITE(IW,8031)NVIOL	18910
IF(NFIX.GT.0) WRITE(6,3020) NFIX	18920
3020 FORMAT(' NO. OF VERTICES OF SIMPLEX CONSTRAINED BY THE '	18930
' PARAMETER LIMITS =',I2/ )	18940
IFAU)LT = 2	18950
RETURN	18960
441 DO 213 I=1,NAP	18970
H(I)=0D0	18980
DO 214 J=1,NAP	18990
IF(J-I)216,216,215	19000
216 IJ=I*(I-1)/2+J	19010
GO TO 217	19020
215 IJ=J*(J-1)/2+I	19030
217 H(I)=H(I)+BMAT(IJ)*AVAL(J)	19040
214 CONTINUE	19050
213 CONTINUE	19060
YMIN=0D0	19070
DO 218 I=1,NAP	19080
218 YMIN=YMIN+H(I)+AVAL(I)	19090
YMIN=A0-YMIN	19100
DO 220 I=1,NOP	19110
PSTST(I)=0.0	19120
DO 220 J=1,NAP	19130
220 PSTST(I)=PSTST(I)+H(J)*G(J,I)	19140
DO 221 I=1,NOP	19150
221 PMIN(I)=PMIN(I)-PSTST(I)	19160
AMIN=YMIN	19170
IF(IPRINT)223,222,222	19180
222 WRITE(IW,303) YMIN,(PMIN(I),I=1,NPR)	19190
303 FORMAT(42H MINIMUM OF FITTED QUADRATIC SURFACE IS ,D15.8,3H AT//	19200
'(5E14.6))	19210

WRITE(IW,304)FUNC,(F(I),I=1,NPR)	19220
304 FORMAT(42H COMPARE WITH MINIMUM FOUND BY ITERATION ,D15.8,3H AT//	19230
'(5E14.6))	19240
D=SNGL(100-YMIN/FUNC)	19250
IF(ABS(D).GT..1) WRITE(IW,305)	19260
305 FORMAT(/	19270
' IF DIFFERENCE IS LARGE, INFORMATION MATRIX IS INACCURATE AND A //	19280
' NEW RUN WITH A DIFFERENT STOPPING CRITERION, EXPANSION ,	19290
' CRITERION // OR PARAMETER RANGE IS RECOMMENDED. / )	19300
223 CONTINUE	19310
DO 2230 I=1,NOP	19320
IF(PMIN(I).LT.F1(I))PMIN(I)=F1(I)	19330
2230 IF(PMIN(I).GT.F2(I))PMIN(I)=F2(I)	19340
CALL FUNCTN(PMIN,QMIN)	19350
IF(QMIN.GE.FUNC) GO TO 2234	19360
FUNC=QMIN	19370
DO 2231 I=1,NOP	19380
2231 F(I)=PMIN(I)	19390

DO 260 I7=1,NP	27900
PP(7,I7)=F1(7)+FLOAT(I7-1)*DE'(7)	27910
FF(7)=PP(7,I7)	27920
DO 250 I6=1,NP	27930
PP(6,I6)=F1(6)+FLOAT(I6-1)*DEL(6)	27940
FF(6)=PP(6,I6)	27950
DO 240 I5=1,NP	27960
PP(5,I5)=F1(5)+FLOAT(I5-1)*DEL(5)	27970
FF(5)=PP(5,I5)	27980
DO 230 I4=1,NP	27990
PP(4,I4)=F1(4)+FLOAT(I4-1)*DEL(4)	28000
FF(4)=PP(4,I4)	28010
DO 220 I3=1,NP	28020
PP(3,I3)=F1(3)+FLOAT(I3-1)*DEL(3)	28030
FF(3)=PP(3,I3)	28040
DO 210 I2=1,NP	28050
PP(2,I2)=F1(2)+FLOAT(I2-1)*DEL(2)	28060
FF(2)=PP(2,I2)	28070
DO 200 I1=1,NP	28080
PP(1,I1)=F1(1)+FLOAT(I1-1)*DEL(1)	28090
FF(1)=PP(1,I1)	28100
CALL LSQ(FF,SUM)	28110
NEVAL=NEVAL+1	28120
IF(SUM.GE.SSMIN) GO TO 195	28130
SSMIN=SUM	28140
IS(1)=I1	28150
IS(2)=I2	28160
IS(3)=I3	28170
IS(4)=I4	28180
IS(5)=I5	28190
IS(6)=I6	28200
IS(7)=I7	28210
195 IF(F1(1).EQ.F2(1)) GO TO 205	28220
200 CONTINUE	28230
205 IF(F1(2).EQ.F2(2)) GO TO 215	28240
210 CONTINUE	28250
215 IF(F1(3).EQ.F2(3)) GO TO 225	28260
220 CONTINUE	28270
225 IF(NOP.EQ.3) GO TO 270	28280
IF(F1(4).EQ.F2(4)) GO TO 235	28290
230 CONTINUE	28300
235 IF(NOP.EQ.4) GO TO 270	28310
IF(F1(5).EQ.F2(5)) GO TO 245	28320
240 CONTINUE	28330
245 IF(NOP.EQ.5) GO TO 270	28340
IF(F1(6).EQ.F2(6)) GO TO 255	28350
250 CONTINUE	28360
255 IF(NOP.EQ.6) GO TO 270	28370
IF(F1(7).EQ.F2(7)) GO TO 270	28380
260 CONTINUE	28390
270 CONTINUE	28400
DO 280 I=1,7	28410
FNEW(I)=PP(I,IS(I))	28420
280 STPNEW(I)=DEL(I)/2.	28430
RETURN	28440
1000 FORMAT(// 11X,	28450
*" PLOT OF RESIDUAL SUM OF SQUARES VERSUS PARAMETER NO.",I2/	28460
* 11X," RANGING FROM ",E11.4," TO ",E11.4 )	28470
1010 FORMAT(// 6X,	28480
*" PLOT OF WEIGHTED RESIDUAL SUM OF SQUARES VERSUS PARAMETER",	28490
*" NO.",I2/6X," RANGING FRM ",E11.4," TO ",E11.4 )	28500
1020 FORMAT(// " TABLE OF PLOT ABOVE //	28510

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*2C" NO. SUM OF SQUARES PARAMETER " //1X,68(1H-))	28520
1025 FORMAT(// " TABLE OF PLOT ABOVE //	28530
*2C" NO. WGH.T.SUM OF SQUARES PARAMETER " //1X,68(1H-))	28540
1026 FORMAT(I4,2E14.4,3X,I4,2E14.4)	28550
1027 FORMAT(1X,68(1H-))	28560
1030 FORMAT(// " THE SEARCH GRID CONSISTS OF 100 POINTS GIVEN BY",	28570
*" PAIRED " COMBINATION OF FOLLOWING PARAMETER VALUES //	28580
*" PAR.NO.",I1," (J=1-10) //(5( " ",I2," ),E10.4) )	28590
1040 FORMAT(// " PAR.NO.",I1," (I=1-10) //(5( " ",I2," ),E10.4) )	28600
1050 FORMAT(//	28610
*" MATRIX OF RESIDUAL SUM OF SQUARES AT POINTS IN SEARCH GRID." )	28620
1060 FORMAT(// " MATRIX OF WEIGHTED RESIDUAL SUM OF SQUARES AT POINTS",	28630
*" IN SEARCH GRID." )	28640
1070 FORMAT(//	28650
*" THE I,J-TH ELEMENT IN THE MATRIX CORRESPONDS TO THE J-TH /	28660
*" VALUE OF PAR.NO.",I1," AND THE I-TH VALUE OF PAR.NO.",I1// )	28670
1075 FORMAT(5(2X," J=",I2),3X,5(2X," J=",I2) / )	28680
1080 FORMAT(5(E10.4,5X))	28690

COMMON /B11/AR	370
COMMON /B12/NPR	380
COMMON /B13/IGNORE	390
COMMON /B14/NVIOL	400
COMMON /B15/SKIP	410
LOGICAL SKIP,SKIPS	420
EQUIVALENCE (AR(1),ANS)	430
EXTERNAL LSO	440
DATA TCRIT/12.7,4.30,3.18,2.78,2.57,2.45,2.36,2.31,2.26,2.23,	450
+2.20,2.18,2.16,2.14,2.13,2.12,2.11,2.10,2.09,2.09,2.08,	460
+2.07,2.07,2.06,2.06,2.06,2.05,2.05,2.04,2.04/	470
DATA NRECOH /0,0,343,623,1024,729,2187 /	480
DATA NCRIT /0,0,8,16,32,64,128 /	490
WRITE(6,500)	500
LINES=30	510
NRUN=0	520
ITABLE=0	530
LIMITS=0	540
NCONR=0	550
NDUMMY=0	560
NS=1	570
NFUNC=-1	580
NOP=-1	590
NOP2=-1	600
NOBS2=-1	610

B01

IQUAD=1	620
SIMP=1E-6	630
ANS4=2HNO	640
PLOTSR=2HNO	650
MAX=800	660
NLOOP=20	670
IPRINT=-1	680
ANS11=2HNO	690
ANS12=2HNO	700
STOPCR=0.001	710
NIND=1	720
PLT=2HNO	730
ANS16=2HNO	740
5 NO=1	750
WRITE(6,505)	760
IGNORE=0	770
CALL READ(1,AR)	780
GO TO (1000,1001) IGO	790
6 IF(ANS.NE.3HYES) GO TO 8	800
WRITE(6,511)	810
WRITE(6,512)	820
8 IF(ANS.GT.NQ) GO TO 1006	830
9 CONTINUE	840
10 NO=2	850
WRITE(6,520)	860
READ(5,525)	870
IF(EOF(5))10,11,10	880
11 IF(ANS.GT.NQ) GO TO 1006	890
15 NO=3	900
WRITE(6,530)	910
CALL READ(1,AR)	920
GO TO (1000,1001) IGO	930
16 IF(ANS.EQ.3HYES) IGO=1	940
IF(ANS.EQ.2HNO) IQUAD=0	950
IF(ANS.EQ.2HNO) GO TO 18	960
WRITE(6,535)	970
CALL READ(1,AR)	980
IF(IGO.EQ.2) GO TO 1001	990
IF(ANS.EQ.4HBACK) GO TO 1004	1000
SIMP=ANS	1010
IF(SIMP.LT.0..OR.SIMP.GT.10.)GO TO 1007	1020
18 IF(ANS.GT.NQ) GO TO 1006	1030
20 NO=4	1040
WRITE(6,540)	1050
CALL READ(1,AR)	1060
GO TO (1000,1001) IGO	1070
21 ANS=-ANS	1080
IF(ANS4.EQ.2HNO) GO TO 24	1090
WRITE(6,541)	1100
22 CALL READ(1,AR)	1110
IF(IGO.EQ.2) GO TO 1001	1120
IF(ANS.EQ.4HBACK) GO TO 1004	1130
PLOTSR=ANS	1140
IF(PLOTSR.NE.2HNO.AND.PLOTSR.NE.3HYES) GO TO 23	1150
GO TO 24	1160
23 WRITE(6,1020)	1170
GO TO 22	1180
24 IF(ANS.GT.NQ) GO TO 1006	1190
25 NO=5	1200
WRITE(6,545)	1210
CALL READ(1,AR)	1220
GO TO (1003,1001)IGO	1230

C01

26 MAX=IFIX(ANS+.1)	1240
IF(MAX.LT.1.OR.MAX.GT.2000) GO TO 1007	1250
IF(ANS.GT.NQ) GO TO 1006	1260
30 NO=6	1270
WRITE(6,550)	1280
CALL READ(1,AR)	1290
GO TO (1003,1001)IGO	1300
31 NLOOP=IFIX(ANS+.1)	1310
IF(NLOOP.LT.1) GO TO 1007	1320
IF(NLOOP.GT.MAX.AND.MAX.NE.1) GO TO 32	1330
GO TO 33	1340
32 NAGR=5	1350
GO TO 1008	1360
33 IF(ANS.GT.NQ) GO TO 1006	1370
35 NO=7	1380

DELTA=(XMAX-XMIN)/30	9670
IF(II.EQ.1) WRITE(6,8261)	9680
IF(II.GT.1) WRITE(6,8264)II	9690
DO 3706 I=1,26	9700
J=26+I	9710
XP1=XMIN+FLOAT(I-1)*DELTA	9720
XP2=XMIN+FLOAT(J-1)*DELTA	9730
DO 3708 L=M1,NN	9740
IF(XP1.GT.X(L).AND.XP1.LE.X(L+1))	9750
*YP1=Y(L)+(Y(L+1)-Y(L))*(XP1-X(L))/(X(L+1)-X(L))	9760
3708 IF(XP2.GT.X(L).AND.XP2.LE.X(L+1))	9770
*YP2=Y(L)+(Y(L+1)-Y(L))*(XP2-X(L))/(X(L+1)-X(L))	9780
ITHFUN=11	9790
CALL MODEL(YP1,XP1,F,0)	9800
CALL MODEL(YP2,XP2,F,0)	9810
IF(I.EQ.26) GO TO 3707	9820
3706 WRITE(6,8262) I,XP1,YP1,J,XP2,YP2	9830
3707 WRITE(6,8262) I,XP1,YP1	9840
3709 CONTINUE	9850
WRITE(6,8263)	9860
371 CONTINUE	9870
IF(IQUAD.NE.1) GO TO 376	9880
WRITE(6,827)	9890
DO 375 K=1,2	9900
IF(K.EQ.2) WRITE(6,8270)	9910

802

L=1	9920
372 IF(L.GT.NPR) GO TO 375	9930
II=L+(L-1)/2	9940
DO 373 I=L,NPR	9950
IA=II+L	9960
II=II+1	9970
IB=MINO(II,IA+5)	9980
IF(K.EQ.2) GO TO 3721	9990
DO 3720 J=IA,IB	10000
3720 VARCOV(J)=2.*FUNC+VARCOV(J)/FLOAT(NRSDF)	10010
WRITE(6,829)(VARCOV(J),J=IA,IB)	10020
GO TO 373	10030
3721 DO 3722 KK=IA,IB	10040
3722 VARCOV(KK)=VC(KK)	10050
WRITE(6,829)(VARCOV(J),J=IA,IB)	10060
373 CONTINUE	10070
L=L+6	10080
GO TO 372	10090
375 CONTINUE	10100
376 IF(ANS4.EQ.2HNO) GO TO 395	10110
DWSTAT=0.	10120
DO 380 I=2,NOBS	10130
380 DWSTAT=DWSTAT+(RES(I-1)-RES(I))*2	10140
DWSTAT=DWSTAT/SSRES	10150
WRITE(6,830)DWSTAT	10160
N=0	10170
DO 381 I=1,NOBS	10180
IF(NYN(I).EQ.0.) GO TO 381	10190
N=N+1	10200
VARCOV(N)=RES(I)	10210
381 CONTINUE	10220
PR=SEQPRO(VARCOV,N)	10230
WRITE(6,832)PR	10240
IF(PR.LT.0.05) WRITE(6,833)	10250
IF(NOBS.GT.3) CALL KOLMIR(VARCOV,N,DN,CRIT,1)	10260
CALL PROB1(VARCOV,N,MM,PR)	10270
WRITE(6,8320)MM,N,PR	10280
IF(PR.LT.0.05) WRITE(6,833)	10290
RATIO=FLOAT(NRSDF)/FLOAT(NOBS-NDUMMY)	10300
IF(RATIO.LE..5) WRITE(6,834)RATIO	10310
IGNORE=1	10320
IF(PLOTRES.NE.3HYES) GO TO 395	10330
IF(NIND.GT.1) GO TO 390	10340
WRITE(6,836)	10350
CALL PLOT(X,PES,NOBS,LINES)	10360
WRITE(6,838)	10370
CALL PLOT(YEST,RES,NOBS,LINES)	10380
GO TO 391	10390
390 WRITE(6,840)	10400
CALL PLOT(X,RES,NOBS,LINES)	10410
WRITE(6,842)	10420
CALL PLOT(Z,RES,NOBS,LINES)	10430
WRITE(6,838)	10440
CALL PLOT(YEST,RES,NOBS,LINES)	10450
391 IF(ANS16.EQ.2HNO.OR.NOWGHT.EQ.1) GO TO 395	10460
IF(NIND.GT.1) GO TO 392	10470
WRITE(6,8360)	10480
CALL PLOT(X,WRES,NOBS,LINES)	10490
WRITE(6,8380)	10500
CALL PLOT(YEST,WRES,NOBS,LINES)	10510
GO TO 395	10520
392 WRITE(6,8400)	10530

C02

CALL PLOT(X,WRES,NOBS,LINES)	10540
WRITE(6,8420)	10550
CALL PLOT(Z,WRES,NOBS,LINES)	10560
WRITE(6,8380)	10570
CALL PLOT(YEST,WRES,NOBS,LINES)	10580
395 CONTINUE	10590
WRITE(6,8401)STOPCR	10600
IF(IQUAD.NE.1) GO TO 396	10610
WRITE(6,8402)SIMP	10620
D=ABS(1.-AMINO/FUNC)	10630
IF(IPRINT.EQ.-1.AND.D.GT.0.1) WRITE(6,8403)AMINO,FUNC	10640
396 NEVAL1=NEVAL1-NVIOL	10650
WRITE(6,842?)NEVAL1,NVIOL	10660
IF(IQUAD.EQ.1.AND.IPRINT.EQ.-1)WRITE(6,8424)NEVAL2	10670
ITABLE=1	10680
ITHFUN=NFUNC	10690



H(I)=000	18980
DO 214 J=1,NAP	18990
IF(J-1)216,216,215	19000
216 IJ=I*(I-1)/2+J	19010
GO TO 217	19020
215 IJ=J*(J-1)/2+I	19030
217 H(I)=H(I)+BMAT(IJ)*AVAL(J)	19040
214 CONTINUE	19050
213 CONTINUE	19060
YMIN=000	19070
DO 218 I=1,NAP	19080
218 YMIN=YMIN+H(I)*AVAL(I)	19090
YMIN=A0-YMIN	19100
DO 220 I=1,NOP	19110
PSTST(I)=0.0	19120
DO 220 J=1,NAP	19130
220 PSTST(I)=PSTST(I)+H(J)*G(J,I)	19140
DO 221 I=1,NOP	19150
221 PMIN(I)=PMIN(I)-PSTST(I)	19160
AMINQ=YMIN	19170
IF(IPRINT)223,222,222	19180
222 WRITE(IW,303) YMIN,(PMIN(I),I=1,NPR)	19190
303 FORMAT(42H MINIMUM OF FITTED QUADRATIC SURFACE IS ,D15.8,3H AT//	19200
*(5E14.6))	19210

803

WRITE(IW,304)FUNC,(F(I),I=1,NPR)	19220
304 FORMAT(/42H COMPARE WITH MINIMUM FOUND BY ITERATION ,D15.8,3H AT//	19230
*(5E14.6))	19240
D=SNGL(100-YMIN/FUNC)	19250
IF(ABS(D).GT..1) WRITE(IW,305)	19260
305 FORMAT(/	19270
*" IF DIFFERENCE IS LARGE, INFORMATION MATRIX IS INACCURATE AND A"	19280
*" NEW RUN WITH A DIFFERENT STOPPING CRITERION, EXPANSION,	19290
*" CRITERION"/" OR PARAMETER RANGE IS RECOMMENDED."/ )	19300
223 CONTINUE	19310
DO 2230 I=1,NOP	19320
IF(PMIN(I).LT.F1(I))PMIN(I)=F1(I)	19330
2230 IF(PMIN(I).GT.F2(I))PMIN(I)=F2(I)	19340
CALL FUNCTN(PMIN,QMIN)	19350
IF(QMIN.GE.FUNC) GO TO 2234	19360
FUNC=QMIN	19370
DO 2231 I=1,NOP	19380
2231 F(I)=PMIN(I)	19390
IF(IPRINT.LT.0) GO TO 2234	19400
WRITE(IW,2232)FUNC	19410
2232 FORMAT(/" IMPROVED FUNCTION VALUE =" ,D15.8//	19420
IF(NPR.LE.5) WRITE(IW,1080)(F(I),I=1,NPR)	19430
IF(NPR.GT.5) WRITE(IW,108)(F(I),I=1,NPR)	19440
2234 CONTINUE	19450
DO 224 I=1,NOP	19460
DO 225 J=1,NAP	19470
H(J)=000	19480
DO 226 K=1,NAP	19490
IF(K-J)227,227,228	19500
227 JK=J*(J-1)/2+K	19510
GO TO 229	19520
228 JK=K*(K-1)/2+J	19530
229 H(J)=H(J)+BMAT(JK)*DBLE(G(K,I))/2D0	19540
226 CONTINUE	19550
225 CONTINUE	19560
DO 230 J=1,NOP	19570
IJ = J*(J-1)/2+I	19580
VC(IJ)=000	19590
DO 231 K=1,NAP	19600
231 VC(IJ)=VC(IJ)+H(K)*DBLE(C(K,J))	19610
230 CONTINUE	19620
224 CONTINUE	19630
J = 0	19640
DO 234 I=1,NOP	19650
J = J+1	19660
234 VAR(I)=VC(J)	19670
236 IF(IPRINT.GE.0) WRITE(IW,306) IRANK	19680
306 FORMAT(/" INFORMATION MATRIX HAS RANK",I3//	19690
*" GENERALIZED INVERSE OF INFORMATION MATRIX"/ )	19700
IJK=1	19710
GO TO 710	19720
717 CONTINUE	19730
IF(IPRINT.GE.0) WRITE(IW,308)	19740
308 FORMAT(	19750
*" THE MATRIX ABOVE MUST BE MULTIPLIED BY TWICE THE ESTIMATE OF"	19760
*" THE RESIDUAL"/" VARIANCE TO GIVE THE VARANCE-COVARIANCE MATRIX"	19770
*" OF THE PARAMETERS."/ )	19780
CALL SYMINV(VC,NAP,BMAT,TEMP,NULLTY,IFALT)	19790
IF(IPRINT.GE.0) WRITE(IW,320)	19800
320 FORMAT(/" INFORMATION MATRIX (HESSIAN MATRIX)" / )	19810
IJK=3	19820
GO TO 710	19830

C03

711 IJK=2	19840
II = 0	19850
IJ = 0	19860
DO 701 I=1,NPR	19870
II = II+1	19880
IF(VC(II).LE.000) GO TO 702	19890
VC(II)=100/DSQRT(VC(II))	19900
GO TO 703	19910
702 VC(II)=000	19920
703 IF(I.EQ.1) GO TO 701	19930
IM1 = I-1	19940
JJ = 0	19950
DO 705 J=1,IM1	19960
JJ = JJ+J	19970
IJ = IJ+1	19980
705 VC(IJ) = VC(IJ)+VC(II)*VC(JJ)	19990
704 II = II+1	20000

223	IF(NOP.EQ.5) GO TO 270	28280
	IF(F1(4).EQ.F2(4)) GO TO 235	28290
230	CONTINUE	28300
235	IF(NOP.EQ.4) GO TO 270	28310
	IF(F1(5).EQ.F2(5)) GO TO 245	28320
240	CONTINUE	28330
245	IF(NOP.EQ.5) GO TO 270	28340
	IF(F1(6).EQ.F2(6)) GO TO 255	28350
250	CONTINUE	28360
255	IF(NOP.EQ.6) GO TO 270	28370
	IF(F1(7).EQ.F2(7)) GO TO 270	28380
260	CONTINUE	28390
270	CONTINUE	28400
	DO 280 I=1,7	28410
	FNEW(I)=PP(I,IS(I))	28420
280	STPNEW(I)=DEL(I)/2.	28430
	RETURN	28440
1000	FORMAT(/ / 11X,	28450
	* PLOT OF RESIDUAL SUM OF SQUARES VERSUS PARAMETER NO., I2 /	28460
	* 11X, RANGING FROM ,E11.4, TO ,E11.4 )	28470
1010	FORMAT(/ / 6X,	28480
	* PLOT OF WEIGHTED RESIDUAL SUM OF SQUARES VERSUS PARAMETER,	28490
	* NO., I2/6X, RANGING FROM ,E11.4, TO ,E11.4 )	28500
1020	FORMAT(/ / TABLE OF PLOT ABOVE//	28510

804

	*2( NO. SUM OF SQUARES PARAMETER )/1X,68(1H-))	28520
1025	FORMAT(/ / TABLE OF PLOT ABOVE//	28530
	*2( NO. WGT.SUM OF SQUARES PARAMETER )/1X,68(1H-))	28540
1026	FORMAT(14,2E14.4,3X,14,2E14.4)	28550
1027	FORMAT(1X,68(1H-))	28560
1030	FORMAT(/ / THE SEARCH GRID CONSISTS OF 100 POINTS GIVEN BY,	28570
	* PAIRED / COMBINATION OF FOLLOWING PARAMETER VALUES //	28580
	* PAR.NO., I1, (J=1-10) // (5( " (, I2, " E10.4) ) )	28590
1040	FORMAT(/ / PAR.NO., I1, (I=1-10) // (5( " (, I2, " E10.4) ) )	28600
1050	FORMAT(/	28610
	* MATRIX OF RESIDUAL SUM OF SQUARES AT POINTS IN SEARCH GRID. )	28620
1060	FORMAT( / / MATRIX OF WEIGHTED RESIDUAL SUM OF SQUARES AT POINTS,	28630
	* IN SEARCH GRID. )	28640
1070	FORMAT(	28650
	* THE I, J-TH ELEMENT IN THE MATRIX CORRESPONDS TO THE J-TH /	28660
	* VALUE OF PAR.NO., I1, AND THE I-TH VALUE OF PAR.NO., I1 // )	28670
1075	FORMAT(5(2X, " J=", I2), 3X, 5(2X, " J=", I2) / )	28680
1080	FORMAT(5(E10.4, 5X))	28690
1085	FORMAT(2X, 5(5X, E10.4))	28700
1090	FORMAT(/ / 6X, SIGN OF THE PARTIAL DERIVATIVES AT THE POINTS IN,	28710
	* THE SEARCH GRID. // 6X,	28720
	* WITH RESPECT TO PAR.NO., I1, 15X, WITH RESPECT TO PAR.NO., I1 /	28730
1100	FORMAT(10X, 10A2, 20X, 10A2)	28740
1110	FORMAT(/ / 12X, CONTOUR CHART / 45X, 12(1H-), KEY ", 13(1H-))	28750
1120	FORMAT(5X, 10A3 )	28760
1130	FORMAT(45X, I1, " = ,E10.4, TO ,E10.4 )	28770
	END	28780
	*****	28790
C	FUNCTION SIGNOF(A)	28800
	IF(A.LT.0.)SIGNOF=2H-	28810
	IF(A.GT.0.)SIGNOF=2H+	28820
	IF(ABS(A).LT.1.E-15) SIGNOF=2H0	28830
	RETURN	28840
	END	28850

C04

WRITE(6,553)	970
CALL READ(1,AR)	980
IF(IGO.EQ.2) GO TO 1001	990
IF(ANS.EQ.4HBACK) GO TO 1004	1000
SIMP=ANS	1010
IF(SIMP.LT.0..OR.SIMP.GT.10.)GO TO 1007	1020
18 IF(NS.GT.NQ) GO TO 1006	1030
20 NO=4	1040
WRITE(6,540)	1050
CALL READ(1,AR)	1060
GO TO (1000,1001) IGO	1070
21 ANS=-ANS	1080
IF(ANS4.EQ.2HNO) GO TO 24	1090
WRITE(6,541)	1100
22 CALL READ(1,AR)	1110
IF(IGO.EQ.2) GO TO 1001	1120
IF(ANS.EQ.4HBACK) GO TO 1004	1130
PLOTS=ANS	1140
IF(PLOTS.NE.2HNO.AND.PLOTS.NE.3HYES) GO TO 23	1150
GO TO 24	1160
23 WRITE(6,1020)	1170
GO TO 22	1180
24 IF(NS.GT.NQ) GO TO 1006	1190
25 NO=5	1200
WRITE(6,545)	1210
CALL READ(1,AR)	1220
GO TO (1003,1001)IGO	1230

C01

26 MAX=IFIX(ANS+.1)	1240
IF(MAX.LT.1.OR.MAX.GT.2000) GO TO 1007	1250
IF(NS.GT.NQ) GO TO 1006	1260
30 NO=6	1270
WRITE(6,550)	1280
CALL READ(1,AR)	1290
GO TO (1003,1001)IGO	1300
31 NLOOP=IFIX(ANS+.1)	1310
IF(NLOOP.LT.1) GO TO 1007	1320
IF(NLOOP.GT.MAX.AND.MAX.NE.1) GO TO 32	1330
GO TO 33	1340
32 NAGR=5	1350
GO TO 1008	1360
33 IF(NS.GT.NQ) GO TO 1006	1370
35 NO=7	1380
WRITE(6,555)	1390
CALL READ(1,AR)	1400
GO TO (1003,1001)IGO	1410
36 IPRINT=IFIX(ANS+.1)	1420
IF(ANS.LT.0..AND.ANS.GE.-1.) IPRINT=-1	1430
IF(IPRINT.LT.-1.OR.IPRINT.GE.MAX.AND.MAX.NE.1) GO TO 1007	1440
IF(NS.GT.NQ) GO TO 1006	1450
40 NO=8	1460
WRITE(6,560)	1470
CALL READ(1,AR)	1480
GO TO (1003,1001)IGO	1490
41 NFUNC=IFIX(ANS+.1)	1500
IF(NFUNC.LT.1.OR.NFUNC.GT.10) GO TO 1007	1510
IF(NFUNC.EQ.1) GO TO 4405	1520
WRITE(6,561)NFUNC	1530
NOBS=0	1540
DO 4400 I=1,NFUNC	1550
42 CALL READ(2,AR)	1560
IF(IGO.EQ.2) GO TO 1001	1570
IF(ANS.EQ.4HBACK) GO TO 1004	1580
J=IFIX(ANS+.1)	1590
IF(J.LT.1.OR.J.GT.10) GO TO 43	1600
GO TO 44	1610
43 WRITE(6,1024)	1620
GO TO 42	1630
44 ITHSET(J)=IFIX(AR(2)+.1)	1640
IF(ITHSET(J).LT.1.OR.ITHSET(J).GT.100) GO TO 43	1650
4400 NOBS=NOBS+ITHSET(J)	1660
GO TO 4410	1670
4405 WRITE(6,562)	1680
CALL READ(1,AR)	1690
IF(IGO.EQ.2) GO TO 1001	1700
IF(ANS.EQ.4HBACK) GO TO 1004	1710
NOBS=IFIX(ANS+.1)	1720
ITHSET(1)=NOBS	1730
4410 IF(NOBS.LT.2.OR.NOBS.GT.200) GO TO 1007	1740
N=1	1750
DO 4415 I=1,NFUNC	1760
LIM1(I)=N	1770
N1=N	1780
LIM2(I)=LIM1(I)+ITHSET(I)-1	1790
N2=LIM2(I)	1800
N=N2+1	1810
DO 4415 J=N1,N2	1820
4415 LABEL(J)=I	1830
IF(NS.GT.NQ) GO TO 1006	1840
45 NO=9	1850

D01

WRITE(6,565)	1860
CALL READ(1,AR)	1870
GO TO (1003,1001)IGO	1880
46 NOP=IFIX(ANS+.1)	1890
IF(NOP.LT.1) GO TO 1007	1900
IF(NS.GT.NQ) GO TO 1006	1910
50 NO=10	1920
WRITE(6,570)NOP	1930
NOP2=NOP+NOP/2	1940
DO 54 I=1,NOP2	1950
CALL READ(3,AR)	1960
GO TO (1003,1001)IGO	1970
51 J=IFIX(ANS+.1)	1980
IF(AR(2).GT.AR(3)) GO TO 1009	1990

CALL PLOT(X,VARLOV,N,PR)	10270
WRITE(6,8320)MM,N,PR	10280
IF(PR.LT.0.05) WRITE(6,833)	10290
RATIO=FLOAT(NRSDF)/FLOAT(NOBS-NDUMMY)	10300
IF(RATIO.LE..5) WRITE(6,834)RATIO	10310
IGNORE=1	10320
IF(PLOTRES.NE.3) GO TO 395	10330
IF(NIND.GT.1) GO TO 390	10340
WRITE(6,836)	10350
CALL PLOT(X,RES,NOBS,LINES)	10360
WRITE(6,838)	10370
CALL PLOT(YEST,RES,NOBS,LINES)	10380
GO TO 391	10390
390 WRITE(6,840)	10400
CALL PLOT(X,RES,NOBS,LINES)	10410
WRITE(6,842)	10420
CALL PLOT(Z,RES,NOBS,LINES)	10430
WRITE(6,838)	10440
CALL PLOT(YEST,RES,NOBS,LINES)	10450
391 IF(ANS16.EQ.2HNO.OR.NOWHGT.EQ.1) GO TO 395	10460
IF(NIND.GT.1) GO TO 392	10470
WRITE(6,8360)	10480
CALL PLOT(X,WRES,NOBS,LINES)	10490
WRITE(6,8380)	10500
CALL PLOT(YEST,WRES,NOBS,LINES)	10510
GO TO 395	10520
392 WRITE(6,8400)	10530

C02

CALL PLOT(X,WRES,NOBS,LINES)	10540
WRITE(6,8420)	10550
CALL PLOT(Z,WRES,NOBS,LINES)	10560
WRITE(6,8380)	10570
CALL PLOT(YEST,WRES,NOBS,LINES)	10580
395 CONTINUE	10590
WRITE(6,8401)STOPCR	10600
IF(IQUAD.NE.1) GO TO 396	10610
WRITE(6,8402)SIMP	10620
D=ABS(1.-AMINO/FUNC)	10630
IF(IPRINT.EQ.-1.AND.D.GT.0.1) WRITE(6,8403)AMINO,FUNC	10640
396 NEVAL1=NEVAL1-NVIOL	10650
WRITE(6,8422)NEVAL1,NVIOL	10660
IF(IQUAD.EQ.1.AND.IPRINT.EQ.-1)WRITE(6,8424)NEVAL2	10670
ITABLE=1	10680
ITHFUN=NFUNC	10690
N=LIM1(NFUNC)	10700
AA=YEST(N)	10710
XMIN=X(1)	10720
XMAX=X(1)	10730
DO 3960 I=2,NOBS	10740
IF(X(I).GT.XMAX) XMAX=X(I)	10750
3960 IF(X(I).LT.XMIN) XMIN=X(I)	10760
DO 397 J=1,NIND	10770
397 G(J)=X(J,N)	10780
WRITE(6,8425)	10790
IGNORE=0	10800
IF(NIND.EQ.1) CALL MODEL(AA,G(1),F,1)	10810
IF(NIND.GT.1) CALL MODEL(AA,G,F,1)	10820
ITABLE=0	10830
WRITE(6,844)NRUN	10840
ANS11=2HNO	10850
ANS12=2HNO	10860
NS=19	10870
GO TO 160	10880
500 FORMAT(/13(1H+),	10890
*" YOU ARE NOW LINKED TO FUNFIT",14(1H+)	10900
*66X," VERSION 7-76"/ )	10910
505 FORMAT(" (1) DO YOU WANT SOME INFORMATION ABOUT THIS PROGRAM",	10920
*" (TYPE YES OR NO)" // )	10930
511 FORMAT(/	10940
*" IN THE YES-OR-NO TYPE QUESTIONS THAT FOLLOW,ONLY FOUR ANSWERS"/	10950
*" ARE ALLOWED,NAMELY YES NO BACK REQUEST"/	10960
*" OR IN SHORT FORM Y N B R"/	10970
*" BACK (B) CAN ALSO BE ENTERED AT ANY TIME ON REQUESTS THAT ASK"/	10980
*" FOR NUMERICAL DATA. IT ALLOWS YOU TO GO BACK AND CORRECT ANY"/	10990
*" OF YOUR PREVIOUS INPUTS AND THEN CONTINUE FROM CURRENT INPUT"/	11000
*" REQUEST. SIMPLY TYPE BACK** (OR B**) ANYWHERE ON A LINE IF YOU"/	11010
*" WANT TO GO BACK AND CORRECT THE INPUT UNDER REQUEST NUMBER **"/	11020
*" THE B** FEATURE ALSO MAKES IT EASY TO REPEAT A RUN WITH ONLY"/	11030
*" A FEW CHANGES IN THE INPUTS. BY TYPING R** YOU WILL BE"/	11040
*" TRANSFERRED"/ BACKWARD OR FORWARD TO REQUEST NUMBER **"/	11050
*" THE R-COMMAND AND THE DEFAULT VALUES ENABLE A FAST INPUT."/	11060
*" INPUT ERRORS IN REQUEST NO.10 OR 17 CAN BE CORRECTED"/	11070
*" (BY RETYPING"/ THE WRONG LINE(S) IN CORRECT FORM. THE LAST LINE"/	11080
*" STARTS THE NEXT REQUEST."/)	11090
512 FORMAT(/	11100
*" A PARAMETER CAN BE CHANGED TO A CONSTANT IN REQUEST NO.10 BY"/	11110
*" ASSIGNING IT THE SAME LOWER AND UPPER LIMITS. THIS ENABLES ANY"/	11120
*" SUM-OF-SQUARES CONTOUR TO BE EVALUATED (REQUEST NO.19) BY"/	11130
*" FIXING ALL EXCEPT 2 PARAMETERS."/	11140
*" A LATTICE SEARCH FOR OPTIMAL INITIAL PARAMETER ESTIMATES CAN"/	11150

D02

*" BE MADE BEFORE THE START OF THE FITTING PROCEDURE BY ENTERING"/	11160
*" 1 AS THE MAX. NO. OF SUM-OF-SQUARES FUNCTION EVALUATIONS"/	11170
*" ALLOWED IN REQUEST NO.5."/	11180
*" THE DEFAULT VALUE SPECIFIED IN SOME REQUESTS CAN BE ENTERED IF"/	11190
*" THERE IS DOUBT ABOUT WHICH VALUE TO ENTER."/	11200
*" FOR MORE DETAILED INFORMATION ABOUT THE PROGRAM CONTACT"/	11210
*" P.VENG PEDERSEN."/)	11220
520 FORMAT(" (2) ENTER A HEADING BUT NOT MORE THAN ONE LINE"/)	11230
525 FORMAT(80H	11240
*)	11250
530 FORMAT(" (3) DO YOU WANT STATISTICAL EVALUATIONS OF THE",	11260
*" PARAMETERS (DEFAULT=YES)"/)	11270
535 FORMAT(" ENTER THE EXPANSION CRITERION TO AVOID ROUNDING-OFF",	11280
*" ERRORS"/ IN THE STATISTICAL EVALUATION OF THE PARAMETERS",	11290

IJ = J*(J-1)/2+1	19580
VC(IJ)=0D0	19590
DO 231 K=1,NAP	19600
231 VC(IJ)=VC(IJ)+H(K)*DBLE(C(K,J))	19610
230 CONTINUE	19620
224 CONTINUE	19630
J = 0	19640
DO 234 I=1,NOP	19650
J = J+1	19660
234 VAR(I)=VC(J)	19670
236 IF(IPRINT.GE.0) WRITE(IW,306) IRANK	19680
306 FORMAT(/ INFORMATION MATRIX HAS RANK ,I3//	19690
* " GENERALIZED INVERSE OF INFORMATION MATRIX" / )	19700
IJK=1	19710
GO TO 710	19720
717 CONTINUE	19730
IF(IPRINT.GE.0) WRITE(IW,308)	19740
308 FORMAT(	19750
* " THE MATRIX ABOVE MUST BE MULTIPLIED BY TWICE THE ESTIMATE OF "	19760
* " THE RESIDUAL / " VARIANCE TO GIVE THE VARANCE-COVARIANCE MATRIX "	19770
* " OF THE PARAMETERS " / )	19780
CALL SYMINV(VC,NAP,BMAT,TEMP,NULLTY,IFAU)LT)	19790
IF(IPRINT.GE.0) WRITE(IW,320)	19800
320 FORMAT(/ INFORMATION MATRIX (HESSIAN MATRIX)" / )	19810
IJK=3	19820
GO TO 710	19830

C03

711 IJK=2	19840
II = 0	19850
IJ = 0	19860
DO 701 I=1,NPR	19870
II = II+1	19880
IF(VC(II).LE.0D0) GO TO 702	19890
VC(II)=1D0/DSORT(VC(II))	19900
GO TO 703	19910
702 VC(II)=0D0	19920
703 IF(I.EQ.1) GO TO 701	19930
IM1 = I-1	19940
JJ = 0	19950
DO 705 J=1,IM1	19960
JJ = JJ+J	19970
IJ = IJ+1	19980
705 VC(IJ) = VC(IJ)+VC(II)+VC(JJ)	19990
701 IJ = IJ+1	20000
IF(IPRINT.GE.0) WRITE(IW,312)	20010
312 FORMAT(" CORRELATION MATRIX" / )	20020
II = 0	20030
DO 706 I=1,NPR	20040
II = II+1	20050
IF(VC(II).NE.0D0) VC(II)=1D0	20060
706 CONTINUE	20070
GO TO 710	20080
712 IF(IPRINT.GE.0) WRITE(IW,310)NEVAL	20090
310 FORMAT( " A FURTHER",I4," FUNCTION EVALUATIONS HAVE BEEN USED"/)	20100
NEVAL2=NEVAL	20110
IF(NFIX.GT.0.AND.IPRINT.GE.0) WRITE(6,3020)NFIX	20120
235 RETURN	20130
710 L=1	20140
716 IF(L.GT.NPR) GO TO (717,712,711),IJK	20150
II=L*(L-1)/2	20160
DO 713 I=L,NPR	20170
I1 = II+L	20180
II=II+I	20190
I2=MIN0(II,I1+5)	20200
IF(IJK.EQ.3) GO TO 718	20210
IF(IPRINT.GE.0) WRITE(IW,714)(VC(J),J=I1,I2)	20220
IF(IJK.NE.1) GO TO 713	20230
DO 500 J=I1,I2	20240
500 VARCOV(J)=VC(J)	20250
GO TO 713	20260
718 IF(IPRINT.GE.0) WRITE(IW,714)(BMAT(J),J=I1,I2)	20270
713 CONTINUE	20280
714 FORMAT(1X,6D13.6)	20290
WRITE(IW,715)	20300
715 FORMAT(1H, /)	20310
L=L+6	20320
GO TO 716	20330
END	20340
C .....	20350
SUBROUTINE CHOL (A,N,U,NU)LLTY,IFAU)LT)	20360
DIMENSION A(55),U(55)	20370
DOUBLE PRECISION A,U,W,ETA	20380
ETA=1D-16	20390
NN=0	20400
IFAU)LT)=1	20410
IF (N.LE.0) GO TO 100	20420
IFAU)LT)=2	20430
NU)LLTY=0	20440
J=1	20450

D03

K=0	20460
DO 10 ICOL = 1,N	20470
L=0	20480
DO 11 IROW = 1,ICOL	20490
K=K+1	20500
W=A(K)	20510
M=J	20520
DO 12 I = 1,IROW	20530
L=L+1	20540
IF (I.EQ.IROW) GO TO 13	20550
W=W-U(L)*U(M)	20560
M=M+1	20570
12 CONTINUE	20580
13 IF (IROW.EQ.ICOL) GO TO 14	20590

GO TO 44	1610
43 WRITE(6,1024)	1620
GO TO 42	1630
44 ITHSET(J)=IFIX(AR(2)+.1)	1640
IF(ITHSET(J).LT.1.OR.ITHSET(J).GT.100) GO TO 43	1650
4400 NOBS=NOBS+ITHSET(J)	1660
GO TO 4410	1670
4405 WRITE(6,562)	1680
CALL READ(1,AR)	1690
IF(IGO.EQ.2) GO TO 1001	1700
IF(ANS.EQ.4HBACK) GO TO 1004	1710
NOBS=IFIX(ANS+.1)	1720
ITHSET(1)=NOBS	1730
4410 IF(NOBS.LT.2.OR.NOBS.GT.200) GO TO 1007	1740
N=1	1750
DO 4415 I=1,NFUNC	1760
LIM1(I)=N	1770
N1=N	1780
LIM2(I)=LIM1(I)+ITHSET(I)-1	1790
NZ=LIM2(I)	1800
N=N2+1	1810
DO 4415 J=N1,NZ	1820
4415 LABEL(J)=I	1830
IF(NS.GT.NQ) GO TO 1006	1840
45 NQ=9	1850

001

WRITE(6,565)	1860
CALL READ(1,AR)	1870
GO TO (1003,1001)IG0	1880
46 NOP=IFIX(ANS+.1)	1890
IF(NOP.LT.1) GO TO 1007	1900
IF(NS.GT.NQ) GO TO 1006	1910
50 NQ=10	1920
WRITE(6,570)NOP	1930
NOP2=NOP+NOP/2	1940
DO 54 I=1,NOP2	1950
CALL READ(3,AR)	1960
GO TO (1003,1001)IG0	1970
51 J=IFIX(ANS+.1)	1980
IF(AR(2).GT.AR(3)) GO TO 1009	1990
IF(J.LT.1.OR.J.GT.NOP) GO TO 52	2000
GO TO 53	2010
52 NAGR=9	2020
GO TO 1008	2030
53 F1(J)=AR(2)	2040
F2(J)=AR(3)	2050
IF(J.EQ.NOP) GO TO 5400	2060
54 CONTINUE	2070
WRITE(6,571)NOP	2080
GO TO 50	2090
5400 IF(NS.GT.NQ) GO TO 1006	2100
55 NQ=11	2110
WRITE(6,575)	2120
CALL READ(1,AR)	2130
GO TO (1000,1001) IGO	2140
56 ANS11=ANS	2150
IF(ANS.EQ.2HNO) GO TO 66	2160
WRITE(6,580)	2170
CALL READ(1,AR)	2180
IF(IGO.EQ.2) GO TO 1001	2190
IF(ANS.EQ.4HBACK) GO TO 1004	2200
N11=IFIX(ANS+.1)	2210
IF(N11.LT.1) GO TO 1007	2220
IF(N11.GT.NOP) GO TO 57	2230
GO TO 58	2240
57 NAGR=9	2250
GO TO 1008	2260
58 IF(N11.GT.1) GO TO 59	2270
WRITE(6,585)	2280
GO TO 60	2290
59 WRITE(6,590)N11	2300
60 CONTINUE	2310
DO 65 I=1,N11	2320
CALL READ(2,AR)	2330
IF(IGO.EQ.2) GO TO 1001	2340
IF(ANS.EQ.4HBACK) GO TO 1004	2350
J=IFIX(ANS+.1)	2360
IF(J.LT.1) GO TO 1007	2370
IF(J.GT.NOP) GO TO 61	2380
GO TO 62	2390
61 NAGR=9	2400
GO TO 1008	2410
62 IF(AR(2).LT.F1(J).OR.AR(2).GT.F2(J)) GO TO 63	2420
GO TO 64	2430
63 NAGR=10	2440
GO TO 1008	2450
64 NF(I)=J	2460
65 FS(J)=AR(2)	2470

E01

66 IF(NS.GT.NQ) GO TO 1006	2480
70 NQ=12	2490
WRITE(6,595)	2500
CALL READ(1,AR)	2510
GO TO (1000,1001) IGO	2520
71 ANS12=ANS	2530
IF(ANS.EQ.2HNO) GO TO 81	2540
WRITE(6,600)	2550
CALL READ(1,AR)	2560
IF(IGO.EQ.2) GO TO 1001	2570
IF(ANS.EQ.4HBACK) GO TO 1004	2580
N12=IFIX(ANS+.1)	2590
IF(N12.LT.1) GO TO 1007	2600
IF(N12.GT.NOP) GO TO 72	2610
GO TO 73	2620

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*66X, " VERSION 7-76" / ) 10910
505 FORMAT(" (1) DO YOU WANT SOME INFORMATION ABOUT THIS PROGRAM", 10920
      * (TYPE YES OR NO) // ) 10930
511 FORMAT(/ 10940
      * " IN THE YES-OR-NO TYPE QUESTIONS THAT FOLLOW, ONLY FOUR ANSWERS" / 10950
      * " ARE ALLOWED, NAMELY YES NO BACK REQUEST" / 10960
      * " OR IN SHORT FORM Y N B R" / 10970
      * " BACK (B) CAN ALSO BE ENTERED AT ANY TIME ON REQUESTS THAT ASK" / 10980
      * " FOR NUMERICAL DATA. IT ALLOWS YOU TO GO BACK AND CORRECT ANY" / 10990
      * " OF YOUR PREVIOUS INPUTS AND THEN CONTINUE FROM CURRENT INPUT" / 11000
      * " REQUEST. SIMPLY TYPE BACK** (OR B**) ANYWHERE ON A LINE IF YOU" / 11010
      * " WANT TO GO BACK AND CORRECT THE INPUT UNDER REQUEST NUMBER **" / 11020
      * " THE B** FEATURE ALSO MAKES IT EASY TO REPEAT A RUN WITH ONLY" / 11030
      * " A FEW CHANGES IN THE INPUTS. BY TYPING R** YOU WILL BE " / 11040
      * " TRANSFERRED" /" BACKWARD OR FORWARD TO REQUEST NUMBER **" / 11050
      * " THE R-COMMAND AND THE DEFAULT VALUES ENABLE A FAST INPUT." / 11060
      * " INPUT ERRORS IN REQUEST NO.10 OR 17 CAN BE CORRECTED" / 11070
      * " BY RETYPING /" THE WRONG LINE(S) IN CORRECT FORM. THE LAST LINE" / 11080
      * " STARTS THE NEXT REQUEST." /) 11090
512 FORMAT( 11100
      * " A PARAMETER CAN BE CHANGED TO A CONSTANT IN REQUEST NO.10 BY" / 11110
      * " ASSIGNING IT THE SAME LOWER AND UPPER LIMITS. THIS ENABLES ANY" / 11120
      * " SUM-OF-SQUARES CONTOUR TO BE EVALUATED (REQUEST NO.19) BY" / 11130
      * " FIXING ALL EXCEPT 2 PARAMETERS." / 11140
      * " A LATTICE SEARCH FOR OPTIMAL INITIAL PARAMETER ESTIMATES CAN" / 11150

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D02

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      * " BE MADE BEFORE THE START OF THE FITTING PROCEDURE BY ENTERING" / 11160
      * " 1 AS THE MAX. NO. OF SUM-OF-SQUARES FUNCTION EVALUATIONS" / 11170
      * " ALLOWED IN REQUEST NO.5." / 11180
      * " THE DEFAULT VALUE SPECIFIED IN SOME REQUESTS CAN BE ENTERED IF" / 11190
      * " THERE IS DOUBT ABOUT WHICH VALUE TO ENTER." / 11200
      * " FOR MORE DETAILED INFORMATION ABOUT THE PROGRAM CONTACT" / 11210
      * " P.VENG PEDERSEN. /) 11220
520 FORMAT(" (2) ENTER A HEADING BUT NOT MORE THAN ONE LINE" // ) 11230
525 FORMAT(80H 11240
      * 11250
530 FORMAT(" (3) DO YOU WANT STATISTICAL EVALUATIONS OF THE", 11260
      * " PARAMETERS (DEFAULT=YES) // ) 11270
535 FORMAT(" ENTER THE EXPANSION CRITERION TO AVOID ROUNDING-OFF", 11280
      * " ERRORS" /" IN THE STATISTICAL EVALUATION OF THE PARAMETERS", 11290
      * " (DEFAULT=1E-6) // ) 11300
540 FORMAT(" (4) DO YOU WANT AN ANALYSIS OF THE RESIDUALS", 11310
      * " (DEFAULT=NO) // ) 11320
541 FORMAT(" DO YOU WANT PLOTS OF THE RESIDUALS" // ) 11330
545 FORMAT(" (5) HOW MANY SUM-OF-SQUARES FUNCTION EVALUATIONS WILL", 11340
      * " YOU ALLOW" /" TO REACH CONVERGENCE ON MINIMUM (DEFAULT=800)" // ) 11350
550 FORMAT(" (6) THE CHECK FOR FINAL CONVERGENCE IS DONE EVERY NLOOP" / 11360
      * " TIMES THAT THE PARAMETERS ARE CHANGED BY THE ITERATIVE PROCESS" / 11370
      * " ENTER THE VALUE YOU WANT FOR NLOOP (DEFAULT=20)" // ) 11380
555 FORMAT(" (7) ENTER THE DEGREE OF OUTPUT YOU WANT FROM / 11390
      * " THE FUNCTION MINIMIZATION PROCESS (DEFAULT=-1)" // 11400
      * " 1X, NUMBER= FULL PROGRESS REPORT EVERY *NUMBER* FUNCTION", 11410
      * " EVALUATIONS" / 11420
      * " 5X, " 0= PARTIAL REPORT" / 11430
      * " 5X, "-1= NO OUTPUT" // ) 11440
560 FORMAT(" (8) HOW MANY FUNCTIONS OR RESPONSE SYSTEMS ARE TO BE", 11450
      * " FITTED" /" SIMULTANEOUSLY BY LEAST SQUARES (DEFAULT=1)" // ) 11460
561 FORMAT(" ENTER ,I2, LINES CONTAINING THESE TWO NUMBERS IN THE", 11470
      * " FOLLOWING ORDER" //5X, " (I) THE NUMBER OF THE FUNCTION TO BE", 11480
      * " FITTED (I.E. 1,2,3,...)" /5X, " (II) THE NUMBER OF OBSERVATIONS", 11490
      * " FOR THE FUNCTION" // ) 11500
562 FORMAT(" HOW MANY OBSERVATIONS DO YOU HAVE" // ) 11510
565 FORMAT(" (9) HOW MANY PARAMETERS (I.E. PARAMETERS+CONSTANTS)", 11520
      * " DOES THE FUNCTION(S)" /" HAVE. (A CONSTANT IS A PARAMETER", 11530
      * " WITH LOWER LIMIT = UPPER LIMIT)" // ) 11540
570 FORMAT(" (10) ENTER ,I2, LINES EACH CONTAINING THESE 3 NUMBERS", 11550
      * " IN THE FOLLOWING ORDER" // 11560
      * " 5X, " (I) THE INDEX OF THE PARAMETER (I.E. 1,2,3,...ETC)" / 11570
      * " 5X, " (II) ITS LOWER LIMIT /5X, " (III) ITS UPPER LIMIT" // ) 11580
571 FORMAT(" YOU FORGOT TO ENTER THE LAST PARAMETER (NO., I2, 11590
      * " ) -TRY AGAIN-" /) 11600
575 FORMAT(" (11) DO YOU WANT TO GIVE AN INITIAL ESTIMATE OF ONE OR" / 11610
      * " MORE OF YOUR PARAMETERS (DEFAULT=NO)" // ) 11620
580 FORMAT(" FOR HOW MANY PARAMETERS" // ) 11630
585 FORMAT(" ENTER ONE LINE CONTAINING THE INDEX OF THE PARAMETER" / 11640
      * " FOLLOWED BY ITS VALUE." // ) 11650
590 FORMAT(" ENTER FOR EACH OF THESE ,I2, " PARAMETERS ONE" / 11660
      * " LINE CONTAINING ITS INDEX FOLLOWED BY ITS VALUE." // ) 11670
595 FORMAT(" (12) DO YOU WANT TO GIVE A STARTING STEP SIZE FOR ONE" / 11680
      * " OR MORE OF YOUR PARAMETERS (DEFAULT=NO)" // ) 11690
600 FORMAT(" FOR HOW MANY PARAMETERS" // ) 11700
605 FORMAT(" ENTER ONE LINE CONTAINING THE INDEX OF THE PARAMETER" / 11710
      * " FOLLOWED BY ITS STEP SIZE." // ) 11720
610 FORMAT(" ENTER FOR EACH OF THESE ,I2, " DIFFERENT PARAMETERS ONE" / 11730
      * " LINE CONTAINING ITS INDEX FOLLOWED BY ITS STEP SIZE." // ) 11740
615 FORMAT(" (13) ENTER THE CONVERGENCE CRITERION FOR THE SUM OF", 11750
      * " SQUARES" /" MINIMIZATION PROCESS (DEFAULT=0.001)" // ) 11760
620 FORMAT(" (14) HOW MANY INDEPENDENT VARIABLES DOES YOUR FUNCTION", 11770

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E02

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      * " HAVE (DEFAULT=1)" // ) 11780
625 FORMAT(" (15) DO YOU WANT A PLOT OF THE FITTED CURVE (DEFAULT=NO)" 11790
      * // ) 11800
635 FORMAT(" (16) DO YOU WANT TO WEIGHT YOUR DATA (DEFAULT=NO)" // ) 11810
636 FORMAT(" ENTER ONE OF FOLLOWING 4 DIGITS ACCORDING TO THE", 11820
      * " WEIGHTING YOU PREFER" /" (Y= DEPENDENT VARIABLE. YOU CAN", 11830
      * " SPECIFY R IN NEXT INPUT REQUEST)" // 11840
      * " 1 THE WEIGHT TO BE SPECIFIED FOR EACH OBSERVATION" 11850
      * " /" 2 WEIGHT = Y**R ( R FOR EXAMPLE = -2,1,1/2 ETC. )" / 11860
      * " 3 WEIGHT = LOG( R*Y ) 11870
      * " 4 WEIGHT = EXP( R*Y ) 11880
637 FORMAT(" ENTER THE VALUE FOR R IN THE PREFERRED WEIGHTING SCHEME", 11890
      * " NO., I2, " ABOVE" // ) 11900
640 FORMAT(" (17) ENTER A LINE FOR EACH OF YOUR ,I3, " OBSERVATIONS." / 11910
      * " EACH LINE MUST CONTAIN ,I2, " NUMBERS IN THE FOLLOWING ORDER" 11920
      * " //10X, " (I) THE OBSERVATION NUMBER" / 11930

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IF(IJK.EQ.3) GO TO 710	20210
IF(IPRINT.GE.0) WRITE(IW,714)(VC(J),J=11,12)	20220
DO 500 J=11,12	20230
500 VARCV(J)=VC(J)	20240
GO TO 713	20250
718 IF(IPRINT.GE.0) WRITE(IW,714)(BMAT(J),J=11,12)	20260
713 CONTINUE	20270
714 FORMAT(1X,6D13.6)	20280
WRITE(IW,715)	20290
715 FORMAT(1H, /)	20300
L=L+6	20310
GO TO 716	20320
END	20330
C *****	20340
SUBROUTINE CHOL (A,N,U, NULLTY, IFAULT)	20350
DIMENSION A(55),U(55)	20360
DOUBLE PRECISION A,U,M,ETA	20370
ETA=1D-16	20380
NN=0	20390
IFAU=1	20400
IF (N.LE.0) GO TO 100	20410
IFAU=2	20420
NULLTY=0	20430
J=1	20440
	20450

D03

K=0	20460
DO 10 ICOL = 1,N	20470
L=0	20480
DO 11 IROW = 1,ICOL	20490
K=K+1	20500
M=A(K)	20510
M=J	20520
DO 12 I = 1,IROW	20530
L=L+1	20540
IF (I.EQ.IROW) GO TO 13	20550
M=W-U(L)*U(M)	20560
M=M+1	20570
12 CONTINUE	20580
13 IF (IROW.EQ.ICOL) GO TO 14	20590
IF(U(L).EQ.0D0) GO TO 21	20600
U(K)=M/U(L)	20610
GO TO 11	20620
21 U(K)=0D0	20630
11 CONTINUE	20640
14 IF(M.LE.DABS(ETA*A(K)))GO TO 20	20650
U(K)=DSQRT(M)	20660
GO TO 15	20670
20 U(K)=0D0	20680
IF(M.LT.0D0)NN=NN+1	20690
NULLTY=NULLTY+1	20700
15 J=ICOL	20710
10 CONTINUE	20720
IFAU=0	20730
IF(N.LE.4.AND.NN.GT.1) IFAU=2	20740
IF(N.LE.6.AND.NN.GT.2) IFAU=2	20750
IF(NN.GT.3.OR.N.EQ.NN) IFAU=2	20760
100 RETURN	20770
END	20780
C *****	20790
SUBROUTINE SYMINV (A,N,C,W, NULLTY, IFAULT)	20800
DIMENSION A(55),C(55),W(20)	20810
DOUBLE PRECISION A,C,W,X	20820
NROW=N	20830
IFAU=1	20840
IF (NROW.LE.0) GO TO 100	20850
IFAU=0	20860
CALL CHOL(A,NROW,C, NULLTY, IFAULT)	20870
IF (IFAU.NE.0) GO TO 100	20880
NN=(NROW*(NROW+1))/2	20890
IROW=NROW	20900
NDIAG=NN	20910
16 IF(C(NDIAG).EQ.0D0) GO TO 11	20920
L=NDIAG	20930
DO 10 I=IROW,NROW	20940
W(I)=C(L)	20950
L=L+1	20960
10 CONTINUE	20970
ICOL=NROW	20980
JCOL=NN	20990
MDIAG=NN	21000
15 L=JCOL	21010
X=0D0	21020
IF(ICOL.EQ.IROW)X=1D0/W(ICOL)	21030
K=NROW	21040
13 IF (K.EQ.IROW) GO TO 12	21050
X=X-W(K)*C(L)	21060
K=K-1	21070

E03

L=L-1	21080
IF (L.GT.MDIAG) L=L-K+1	21090
GO TO 13	21100
12 C(L)=X/W(ICOL)	21110
IF (ICOL.EQ.IROW) GO TO 14	21120
MDIAG=MDIAG-ICOL	21130
ICOL=ICOL-1	21140
JCOL=JCOL-1	21150
GO TO 15	21160
11 L=NDIAG	21170
DO 17 J=IROW,NROW	21180
C(L)=0D0	21190
L=L+J	21200
17 CONTINUE	21210
14 NDIAG=NDIAG-IROW	21220
IROW=IROW-1	21230



GO TO 58	2240
57 NAGR=9	2250
GO TO 1008	2260
58 IF(N11.GT.1) GO TO 59	2270
WRITE(6,585)	2280
GO TO 60	2290
59 WRITE(6,590)N11	2300
60 CONTINUE	2310
DO 65 I=1,N11	2320
CALL READ(2,AR)	2330
IF(IGO.EQ.2) GO TO 1001	2340
IF(ANS.EQ.4HBACK) GO TO 1004	2350
J=IFIX(ANS+.1)	2360
IF(J.LT.1) GO TO 1007	2370
IF(J.GT.NOP) GO TO 61	2380
GO TO 62	2390
61 NAGR=9	2400
GO TO 1008	2410
62 IF(AR(2).LT.F1(J).OR.AR(2).GT.F2(J)) GO TO 63	2420
GO TO 64	2430
63 NAGR=10	2440
GO TO 1008	2450
64 NF(1)=J	2460
65 FS(J)=AR(2)	2470

E01

66 IF(NS.GT.NQ) GO TO 1006	2480
70 NQ=12	2490
WRITE(6,595)	2500
CALL READ(1,AR)	2510
GO TO (1000,1001) IGO	2520
71 ANS12=ANS	2530
IF(ANS.EQ.2HNO) GO TO 81	2540
WRITE(6,600)	2550
CALL READ(1,AR)	2560
IF(IGO.EQ.2) GO TO 1001	2570
IF(ANS.EQ.4HBACK) GO TO 1004	2580
N12=IFIX(ANS+.1)	2590
IF(N12.LT.1) GO TO 1007	2600
IF(N12.GT.NOP) GO TO 72	2610
GO TO 73	2620
72 NAGR=9	2630
GO TO 1008	2640
73 IF(N12.GT.1) GO TO 74	2650
WRITE(6,605)	2660
GO TO 75	2670
74 WRITE(6,610)N12	2680
75 CONTINUE	2690
DO 80 I=1,N12	2700
CALL READ(2,AR)	2710
IF(IGO.EQ.2) GO TO 1001	2720
IF(ANS.EQ.4HBACK) GO TO 1004	2730
J=IFIX(ANS+.1)	2740
IF(J.LT.1) GO TO 1007	2750
IF(J.GT.NOP) GO TO 76	2760
GO TO 77	2770
76 NAGR=9	2780
GO TO 1008	2790
77 IF(ABS(AR(2)).GE.(F2(J)-F1(J))) GO TO 78	2800
GO TO 79	2810
78 NAGR=10	2820
GO TO 1008	2830
79 NT(1)=J	2840
80 STPS(J)=AR(2)	2850
81 IF(NS.GT.NQ) GO TO 1006	2860
85 NQ=13	2870
WRITE(6,615)	2880
CALL READ(1,AR)	2890
GO TO (1003,1001)IGO	2900
86 IF(ANS.LT.0.OR.ANS.GT.1.) GO TO 1007	2910
STOPCR=ANS	2920
IF(NS.GT.NQ) GO TO 1006	2930
90 NQ=14	2940
WRITE(6,620)	2950
CALL READ(1,AR)	2960
GO TO (1003,1001)IGO	2970
91 NIND=IFIX(ANS+.1)	2980
IF(NIND.LT.1.OR.NIND.GT.9) GO TO 93	2990
GO TO 94	3000
93 WRITE(6,1024)	3010
GO TO 92	3020
94 IF(NS.GT.NQ) GO TO 1006	3030
95 NQ=15	3040
IF(NIND.GT.1) GO TO 97	3050
WRITE(6,625)	3060
CALL READ(1,AR)	3070
GO TO (1000,1001) IGO	3080
96 PLT=ANS	3090

F01

97 IF(NS.GT.NQ) GO TO 1006	3100
100 NQ=15	3110
WRITE(6,635)	3120
CALL READ(1,AR)	3130
GO TO (1000,1001) IGO	3140
101 ANS16=ANS	3150
IMGHT=0	3160
IF(ANS16.EQ.2HNO) GO TO 102	3170
WRITE(6,636)	3180
CALL READ(1,AR)	3190
IF(IGO.EQ.2) GO TO 1001	3200
IF(ANS.EQ.4HBACK) GO TO 1004	3210
IMGHT=IFIX(ANS+.1)	3220
IF(IMGHT.LT.1.OR.IMGHT.GT.4) GO TO 1007	3230
IF(IMGHT.EQ.1) GO TO 102	3240
WRITE(6,637)IMGHT	3250
CALL READ(1,AR)	3260

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* WITH LOWER LIMIT = UPPER LIMIT) // ) 11540
570 FORMAT( (10) ENTER ,12, LINES EACH CONTAINING THESE 3 NUMBERS", 11550
* IN THE FOLLOWING ORDER // 11560
* ,5X, (I) THE INDEX OF THE PARAMETER (I.E. 1,2,3...ETC) / 11570
* ,5X, (II) ITS LOWER LIMIT /5X, (III) ITS UPPER LIMIT // 11580
571 FORMAT( YOU FORGOT TO ENTER THE LAST PARAMETER (NO. ,12, 11590
* ) -TRY AGAIN- // ) 11600
575 FORMAT( (11) DO YOU WANT TO GIVE AN INITIAL ESTIMATE OF ONE OR / 11610
* MORE OF YOUR PARAMETERS (DEFAULT=NO) // ) 11620
580 FORMAT( FOR HOW MANY PARAMETERS // ) 11630
585 FORMAT( ENTER ONE LINE CONTAINING THE INDEX OF THE PARAMETER / 11640
* FOLLOWED BY ITS VALUE. // ) 11650
590 FORMAT( ENTER FOR EACH OF THESE ,12, PARAMETERS ONE / 11660
* LINE CONTAINING ITS INDEX FOLLOWED BY ITS VALUE. // ) 11670
595 FORMAT( (12) DO YOU WANT TO GIVE A STARTING STEP SIZE FOR ONE / 11680
* OR MORE OF YOUR PARAMETERS (DEFAULT=NO) // ) 11690
600 FORMAT( FOR HOW MANY PARAMETERS // ) 11700
605 FORMAT( ENTER ONE LINE CONTAINING THE INDEX OF THE PARAMETER / 11710
* FOLLOWED BY ITS STEP SIZE. // ) 11720
610 FORMAT( ENTER FOR EACH OF THESE ,12, DIFFERENT PARAMETERS ONE / 11730
* LINE CONTAINING ITS INDEX FOLLOWED BY ITS STEP SIZE. // ) 11740
615 FORMAT( (13) ENTER THE CONVERGENCE CRITERION FOR THE SUM OF , 11750
* SQUARES / MINIMIZATION PROCESS (DEFAULT=0.001) // ) 11760
620 FORMAT( (14) HOW MANY INDEPENDENT VARIABLES DOES YOUR FUNCTION", 11770

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E02

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* HAVE (DEFAULT=1) // ) 11780
625 FORMAT( (15) DO YOU WANT A PLOT OF THE FITTED CURVE (DEFAULT=NO) 11790
* // ) 11800
635 FORMAT( (16) DO YOU WANT TO WEIGHT YOUR DATA (DEFAULT=NO) // ) 11810
636 FORMAT( ENTER ONE OF FOLLOWING 4 DIGITS ACCORDING TO THE", 11820
* WEIGHTING YOU PREFER / (Y= DEPENDENT VARIABLE. YOU CAN", 11830
* SPECIFY R IN NEXT INPUT REQUEST) // 11840
* 1 THE WEIGHT TO BE SPECIFIED FOR EACH OBSERVATION" 11850
* / 2 WEIGHT = Y+R ( R FOR EXAMPLE = -2,1,1/2 ETC. ) / 11860
* 3 WEIGHT = LOG( R+Y ) / 11870
* 4 WEIGHT = EXP( R+Y ) // ) 11880
637 FORMAT( ENTER THE VALUE FOR R IN THE PREFERRED WEIGHTING SCHEME", 11890
* NO. ,12, ABOVE // ) 11900
640 FORMAT( (17) ENTER A LINE FOR EACH OF YOUR ,13, OBSERVATIONS. / 11910
* EACH LINE MUST CONTAIN ,12, NUMBERS IN THE FOLLOWING ORDER" 11920
* //10X, (I) THE OBSERVATION NUMBER / 11930
* 10X, (II) THE VALUE OF THE DEPENDENT VARIABLE (RESPONSE) / 11940
* 10X, (III) THE WEIGHT OF THE OBSERVATION / 11950
* 10X, (IV) THE VALUE(S) OF THE ,12, INDEPENDENT VARIABLE(S) // ) 11960
641 FORMAT( ENTER NOW THE ,13, OBSERVATIONS FOR RESPONSE SYSTEM", 11970
* NO. ,12, REMEMBER THE FIRST OBSERVATION FOR THIS RESPONSE", 11980
* SYSTEM MUST BE NUMBERED ,13, IN CONTINUATION OF THE PREVIOUS", 11990
* OBSERVATION NUMBER. // ) 12000
642 FORMAT( YOU FORGOT TO ENTER THE LAST OBSERVATION (NO. ,13, ) / 12010
* REENTER YOUR DATA STRICTLY ACCORDING TO THE FOLLOWING REQUEST // ) 12020
645 FORMAT( (18) ENTER A LINE FOR EACH OF YOUR ,13, OBSERVATIONS. / 12030
* EACH LINE MUST CONTAIN ,12, NUMBERS IN THE FOLLOWING ORDER" 12040
* //10X, (I) THE OBSERVATION NUMBER / 12050
* 10X, (II) THE VALUE OF THE DEPENDENT VARIABLE (RESPONSE) / 12060
* 10X, (III) THE VALUE(S) OF THE ,12, INDEPENDENT VARIABLE(S) // ) 12070
660 FORMAT( (18) ENTER-BACK-FOR CORRECTION OR-OK-FOR A FINAL", 12080
* CHECK ON ALL INPUTS // ) 12090
662 FORMAT( YOUR INPUTS HAVE BEEN CHECKED. DO YOU WANT A SUMMARY", 12100
* OF THEM // ) 12110
663 FORMAT( DO YOU WANT A PLOT OF YOUR DATA POINTS INCLUDED IN", 12120
* THE SUMMARY // ) 12130
664 FORMAT( ENTER -BACK- FOR CORRECTION OR -OK- FOR ACTIVATION OF", 12140
* THE COMPUTATIONS // ) 12150
670 FORMAT( WAIT NOW FOR THE RESULTS // ) 12160
6700 FORMAT( (19) DO YOU WANT A ,11, -DIMENSIONAL SEARCH // ) 12170
6705 FORMAT( HOW MANY FUNCTION EVALUATIONS DO YOU ALLOW IN THE", 12180
* SEARCH (DEFAULT= ,14, ) // ) 12190
6710 FORMAT( //20(1H),12, -D I M E N S I O N A L S E A R C H", 12200
* 20(1H) ) 12210
6715 FORMAT( // 15, LATTICE POINTS HAVE BEEN EVALUATED ( , 12220
* 12, POINTS PER PARAMETER ) ) 12230
6720 FORMAT( // THE SMALLEST WEIGHTED RESIDUAL SUM OF SQUARES VALUE / 12240
* FOUND IN SEARCH = ,E10.4, AT / ) 12250
6725 FORMAT( // THE SMALLEST RESIDUAL SUM OF SQUARES VALUE", 12260
* FOUND IN THE SEARCH = ,E10.4, AT / ) 12270
6730 FORMAT( 5X, PAR.NO. ,11, = ,E11.4,5X,40A1 ) 12280
6731 FORMAT( WHICH IS SMALLER THAN THE VALUE ( ,E10.4, ) OBTAINED", 12290
* USING THE FOLLOWING // INITIAL PARAMETER ESTIMATES // ) 12300
6735 FORMAT( WHICH IS SMALLER THAN THE VALUE ( ,E10.4, ) FOUND", 12310
* BY THE MINIMIZATION // PROCESS. THE MINIMIZATION IS THEREFORE", 12320
* RESTARTED IN NEW MINIMUM REGION // USING THE PARAMETERS ABOVE // ) 12330
6740 FORMAT( WHICH IS LARGER THAN THE VALUE ( ,E10.4, ) FOUND BY", 12340
* THE MINIMIZATION PROCESS // AT // ) 12350
6741 FORMAT( WHICH IS LARGER THAN THE VALUE ( ,E10.4, ) OBTAINED", 12360
* USING THE FOLLOWING // INITIAL PARAMETER ESTIMATES // ) 12370
6745 FORMAT( // 77(1H) ) 12380
6747 FORMAT( DO YOU WANT TO MAKE ANY ADDITIONAL CORRECTIONS BEFORE", 12390

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* THE RESTART // ) 12400
6748 FORMAT( DO YOU WANT TO START THE MINIMIZATION PROCESS / 12410
* USING THE PARAMETER VALUES FOUND IN THE SEARCH ABOVE // ) 12420
675 FORMAT( (20) DO YOU WANT TO REPEAT THE COMPUTATIONS ABOVE BUT / 12430
* WITH SOME CHANGE IN THE INPUTS // ) 12440
680 FORMAT( DO YOU WANT ANOTHER RUN WITH A NEW SET OF OBSERVATIONS // 12450
* ) 12460
685 FORMAT( I HOPE YOU HAVE BEEN PLEASED WITH THE PROGRAM. / 12470
* IF YOU HAVE ANY SUGGESTIONS FOR ALTERATIONS PLEASE CONTACT ME // 12480
* REGARDS // 12490
* P.VENG PEDERSEN // ) 12500
690 FORMAT( (21) DO YOU WANT TO USE THE FINAL COMPUTED PARAMETERS / 12510
* FROM THE RUN ABOVE AS STARTING PARAMETERS FOR THE NEXT RUN. // ) 12520
692 FORMAT( YOU CAN NOW MAKE ALTERATIONS IN PREVIOUS INPUTS", 12530
* FOR THE NEXT RUN. ) 12540
694 FORMAT( (22) DO YOU WANT TO MAKE MORE ALTERATIONS // ) 12550

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IF (NROW.LE.0) GO TO 100	20840
IF (NROW.LE.0) GO TO 100	20850
IF (NROW.LE.0) GO TO 100	20860
CALL CHOL(A,NROW,C, NULLTY,IFAU)LT)	20870
IF (IFAU)LT.NE.0) GO TO 100	20880
NN=(NROW*(NROW+1))/2	20890
IROW=NROW	20900
NDIAG=NN	20910
16 IF (C(NDIAG).EQ.000) GO TO 11	20920
L=NDIAG	20930
DO 10 I=IROW,NROW	20940
W(I)=C(L)	20950
L=L-I	20960
10 CONTINUE	20970
ICOL=NROW	20980
JCOL=NN	20990
MDIAG=NN	21000
15 L=JCOL	21010
X=000	21020
IF (ICOL.EQ.IROW)X=100/W(IROW)	21030
K=NROW	21040
13 IF (K.EQ.IROW) GO TO 12	21050
X=X-W(K)*C(L)	21060
K=K-1	21070

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L=L-1	21080
IF (L.GT.MDIAG) L=L-K+1	21090
GO TO 13	21100
12 C(L)=X/W(IROW)	21110
IF (ICOL.EQ.IROW) GO TO 14	21120
MDIAG=MDIAG-ICOL	21130
ICOL=ICOL-1	21140
JCOL=JCOL-1	21150
GO TO 15	21160
11 L=NDIAG	21170
DO 17 J=IROW,NROW	21180
C(L)=000	21190
L=L+J	21200
17 CONTINUE	21210
14 NDIAG=MDIAG-IROW	21220
IROW=IROW-1	21230
IF (IROW.NE.0) GO TO 16	21240
100 RETURN	21250
END	21260
C *****	21270
SUBROUTINE LSQ(F, FUNC)	21280
DIMENSION F(20), F1(20), F2(20), Y(100), WYN(100), XX(9, 100), A(9),	21290
* LABEL(100)	21300
DOUBLE PRECISION FUNC, YOBS, YCAL, W	21310
COMMON /DATA/ XX, Y, WYN, NOBS	21320
COMMON /PARIM/ F1, F2	21330
COMMON /CONSTR/LIMITS	21340
COMMON /FUNNUM/ITHFUN	21350
COMMON /B4/ NIND	21360
COMMON /B6/ NOP	21370
COMMON /B10/LABEL	21380
COMMON /B14/NVIOL	21390
S=0.	21400
FUNC=000	21410
IF (LIMITS.EQ.0) GO TO 6	21420
DO 5 I=1, NOP	21430
IF (F(I).GE.F1(I).AND.F(I).LE.F2(I).OR.F1(I).EQ.F2(I)) GO TO 5	21440
B=ABS(F1(I)-F(I))	21450
B2=ABS(F2(I)-F(I))	21460
B=1.E50*(1.+AMIN1(B, B2))/AMAX1(B, B2)	21470
IF (B.GT.S)S=B	21480
5 CONTINUE	21490
IF (S.GE.1E50) GO TO 20	21500
6 CONTINUE	21510
DO 15 I=1, NOBS	21520
DO 10 J=1, NIND	21530
10 A(J)=XX(J, I)	21540
ITHFUN=LABEL(I)	21550
S=Y(I)	21560
IF (NIND.EQ.1) CALL MODEL(S, A(1), F, 0)	21570
IF (NIND.GT.1) CALL MODEL(S, A, F, 0)	21580
IF (S.GE.1E50) GO TO 20	21590
YCAL=S	21600
YOBS=Y(I)	21610
W=WYN(I)	21620
15 FUNC=FUNC+W*(YOBS-YCAL)*(YOBS-YCAL)	21630
RETURN	21640
20 FUNC=S+1.000000001	21650
NVIOL=NVIOL+1	21660
RETURN	21670
END	21680
C *****	21690

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SUBROUTINE READ(N, AR)	21700
DIMENSION A(78), AR(12), AL(10)	21710
COMMON/BS/NO, NS, IGO	21720
COMMON /B15/SKIP	21730
LOGICAL SKIP	21740
DATA YES, AND, BACK, AD, AK, BLANK, PLUS, AMINUS, COMMA, DOT, E	21750
* /1HY, 1HN, 1HB, 1HO, 1HK, 1H, 1H+, 1H-, 1H, 1H, 1HE/	21760
DATA AL/1HO, 1H1, 1H2, 1H3, 1H4, 1H5, 1H6, 1H7, 1H8, 1H9 /	21770
5 IGO=1	21780
SKIP=.FALSE.	21790
DO 6 L=1, 78	21800
6 A(L)=BLANK	21810
DO 7 L=1, 12	21820
7 AR(L)=0.0	21830
READ(5, 200)A	21840
IF (EOF(5))8, 9, 8	21850
8 WRITE(6, 225)	21860

90	NO=14	2940
	WRITE(6,620)	2950
92	CALL READ(1,AR)	2960
	GO TO (1003,1001)IGO	2970
91	NIND=FIX(ANS+.1)	2980
	IF(NIND.LT.1.OR.NIND.GT.9) GO TO 93	2990
	GO TO 94	3000
93	WRITE(6,1024)	3010
	GO TO 92	3020
94	IF(NS.GT.NQ) GO TO 1006	3030
95	NQ=15	3040
	IF(NIND.GT.1) GO TO 97	3050
	WRITE(6,625)	3060
	CALL READ(1,AR)	3070
	GO TO (1000,1001) IGO	3080
96	PLT=ANS	3090

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97	IF(NS.GT.NQ) GO TO 1006	3100
100	NQ=15	3110
	WRITE(6,635)	3120
	CALL READ(1,AR)	3130
	GO TO (1000,1001) IGO	3140
101	ANS16=ANS	3150
	IWGHT=0	3160
	IF(ANS16.EQ.2HNO) GO TO 102	3170
	WRITE(6,636)	3180
	CALL READ(1,AR)	3190
	IF(IGO.EQ.2) GO TO 1001	3200
	IF(ANS.EQ.4HBACK) GO TO 1004	3210
	IWGHT=FIX(ANS+.1)	3220
	IF(IWGHT.LT.1.OR.IWGHT.GT.4) GO TO 1007	3230
	IF(IWGHT.EQ.1) GO TO 102	3240
	WRITE(6,637)IWGHT	3250
	CALL READ(1,AR)	3260
	IF(IGO.EQ.2) GO TO 1001	3270
	IF(ANS.EQ.4HBACK) GO TO 1004	3280
102	ANS16B=ANS	3290
	IF(NS.GT.NQ) GO TO 1006	3300
105	NQ=17	3310
	NSAVE=1	3320
	J=0	3330
	NOBS2=NOBS+5	3340
	IF(ANS16.EQ.2HNO.OR.IWGHT.GT.1) GO TO 120	3350
106	NN=NIND+3	3360
	WRITE(6,640)NOBS,NN,NIND	3370
	IW=1	3380
	DO 109 I=1,NOBS2	3390
	J=J+1	3400
	N=LABEL(J)	3410
	IF(N.EQ.NSAVE+1) WRITE(6,641)ITHSET(N),N,J	3420
	NSAVE=N	3430
1060	CALL READ(NN,AR)	3440
	IF(IGO.EQ.2) GO TO 1001	3450
	IF(ANS.EQ.4HBACK) GO TO 1004	3460
	IF(AR(3).LT.0.) WRITE(6,1033)	3470
	IF(AR(3).LT.0.) GO TO 1060	3480
	J=FIX(ANS+.1)	3490
	IF(J.LT.1) GO TO 1007	3500
	IF(J.GT.NOBS) GO TO 107	3510
	GO TO 108	3520
107	NAGR=8	3530
	GO TO 1008	3540
108	CONTINUE	3550
	DO 1080 L=1,NIND	3560
1080	XX(L,J)=AR(L+3)	3570
	INDEX(J)=J	3580
	Y(J)=AR(2)	3590
	WY(J)=AR(3)	3600
	IF(J.EQ.NOBS)GO TO 125	3610
109	CONTINUE	3620
	WRITE(6,642)NOBS	3630
	GO TO 105	3640
120	NN=NIND+2	3650
	WRITE(6,655)NOBS,NN,NIND	3660
	IW=2	3670
	DO 123 I=1,NOBS2	3680
	J=J+1	3690
	N=LABEL(J)	3700
	IF(N.EQ.NSAVE+1) WRITE(6,641)ITHSET(N),N,J	3710

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	NSAVE=N	3720
	CALL READ(NN,AR)	3730
	IF(IGO.EQ.2) GO TO 1001	3740
	IF(ANS.EQ.4HBACK) GO TO 1004	3750
	J=FIX(ANS+.1)	3760
	IF(J.LT.1) GO TO 1007	3770
	IF(J.GT.NOBS) GO TO 121	3780
	GO TO 122	3790
121	NAGR=8	3800
	GO TO 1008	3810
122	CONTINUE	3820
	DO 1220 L=1,NIND	3830
1220	XX(L,J)=AR(L+2)	3840
	INDEX(J)=J	3850
	Y(J)=AR(2)	3860
	IF(J.EQ.NOBS) GO TO 125	3870
123	CONTINUE	3880
	WRITE(6,642)NOBS	3890
	GO TO 120	3900
125	CONTINUE	3910
	DO 126 I=1,NOBS	3920
	X(I)=XX(1,I)	3930
126	IF(NIND.GT.1) Z(I)=XX(2,I)	3940
	IF(NS.GT.NQ) GO TO 1006	3950
130	NQ=18	3960

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6720 FORMAT(// " THE SMALLEST WEIGHTED RESIDUAL SUM OF SQUARES VALUE" / 12240
* " FOUND IN SEARCH =" ,E10.4, " AT" / ) 12250
6725 FORMAT(// " THE SMALLEST RESIDUAL SUM OF SQUARES VALUE" , 12260
* " FOUND IN THE SEARCH =" ,E10.4, " AT" / ) 12270
6730 FORMAT( 5X, " PAR.NO. ",I1, " =" ,E11.4,5X,40A1 ) 12280
6731 FORMAT(// WHICH IS SMALLER THAN THE VALUE (" ,E10.4,") OBTAINED" , 12290
* " USING THE FOLLOWING" / " INITIAL PARAMETER ESTIMATES" / ) 12300
6735 FORMAT(// WHICH IS SMALLER THAN THE VALUE (" ,E10.4,") FOUND" , 12310
* " BY THE MINIMIZATION" / " PROCESS.THE MINIMIZATION IS THEREFORE" , 12320
* " RESTARTED IN NEW MINIMUM REGION" / " USING THE PARAMETERS ABOVE" / ) 12330
6740 FORMAT(// WHICH IS LARGER THAN THE VALUE (" ,E10.4,") FOUND BY" , 12340
* " THE MINIMIZATION PROCESS" / " AT" ) 12350
6741 FORMAT(// WHICH IS LARGER THAN THE VALUE (" ,E10.4,") OBTAINED" , 12360
* " USING THE FOLLOWING" / " INITIAL PARAMETER ESTIMATES" / ) 12370
6745 FORMAT(// 77(1H*)) 12380
6747 FORMAT(// " DO YOU WANT TO MAKE ANY ADDITIONAL CORRECTIONS BEFORE" , 12390

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F02

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* " THE RESTART" / / ) 12400
6748 FORMAT(// " DO YOU WANT TO START THE MINIMIZATION PROCESS" / 12410
* " USING THE PARAMETER VALUES FOUND IN THE SEARCH ABOVE" / / ) 12420
675 FORMAT(// (20) DO YOU WANT TO REPEAT THE COMPUTATIONS ABOVE BUT" / 12430
* " WITH SOME CHANGE IN THE INPUTS" / / ) 12440
680 FORMAT( " DO YOU WANT ANOTHER RUN WITH A NEW SET OF OBSERVATIONS" / / 12450
* ) 12460
685 FORMAT( " I HOPE YOU HAVE BEEN PLEASED WITH THE PROGRAM." / 12470
* " IF YOU HAVE ANY SUGGESTIONS FOR ALTERATIONS PLEASE CONTACT ME" / / 12480
* " REGARDS" / / 12490
* " P.VENG PEDERSEN" / / ) 12500
690 FORMAT( " (21) DO YOU WANT TO USE THE FINAL COMPUTED PARAMETERS" / 12510
* " FROM THE RUN ABOVE AS STARTING PARAMETERS FOR THE NEXT RUN." / / ) 12520
692 FORMAT( " YOU CAN NOW MAKE ALTERATIONS IN PREVIOUS INPUTS " , 12530
* " FOR THE NEXT RUN. " ) 12540
694 FORMAT( " ( 22) DO YOU WANT TO MAKE MORE ALTERATIONS" / / ) 12550
750 FORMAT(5(// ,1X,77(1H*)) / ) 12560
752 FORMAT( /1X,29(1H*) " I N P U T D A T A " ,28(1H*) / / ) 12570
* " NUMBER OF VARIABLE PARAMETERS =" ,I4 / 12580
* " NUMBER OF PARAMETERS HELD CONSTANT =" ,I4 / 12590
* " NUMBER OF OBSERVATIONS =" ,I4 / 12600
* " NUMBER OF DUMMY OBSERVATIONS =" ,I4 / 12610
* " NUMBER OF RESPONSE SYSTEMS =" ,I4 / 12620
* " NUMBER OF INDEPENDENT VARIABLES =" ,I4 / / 12630
* " PAR.NO. " ,7X, " ALLOWED RANGE" ,9X, " INITIAL ESTIMATE " , 12640
* " INITIAL STEP SIZE" /1X,77(1H-) ) 12650
754 FORMAT(14,4(6X,E10.4) ) 12660
756 FORMAT(1X,77(1H-) / / " OBS.NO. " X1",12X," X2",10X," RESPONSE",7X, 12670
* " WEIGHT",4X," NORM.WEIGHT" /1X,77(1H-) ) 12680
758 FORMAT(14,4X,E13.4,15X,E13.4 ) 12690
760 FORMAT(14,4X,E13.4,15X,3(E13.4,1X) ) 12700
762 FORMAT(14,4X,3(E13.4,1X) ) 12710
764 FORMAT(14,4X,5(E13.4,1X) ) 12720
765 FORMAT(/9X, 12730
* " PLOT OF RESPONSE VERSUS INDEPENDENT VARIABLE X1 (ARB.UNITS)" ) 12740
766 FORMAT(// 12750
* " MAXIMUM NUMBER OF FUNCTION EVALUATIONS ALLOWED =" ,I4 / 12760
* " STOPPING CRITERION FOR CONVERGENCE =" ,E10.4 ) 12770
767 FORMAT(//14X, 12780
* " PLOT OF RESPONSE VERSUS SECOND INDEPENDENT VARIABLE X2" , 12790
* " (ARB.UNITS)" ) 12800
768 FORMAT( " STATISTICS ON THE PARAMETERS HAVE BEEN REQUESTED" / 12810
* " CRITERION FOR EXPANSION OF SIMPLEX BEFORE QUADRATIC FITTING =" , 12820
* " E10.4 ) 12830
770 FORMAT( " ANALYSIS OF RESIDUALS HAS BEEN REQUESTED" ) 12840
772 FORMAT( " RESIDUAL PLOTS HAVE BEEN REQUESTED" ) 12850
774 FORMAT( 12860
* " NO OUTPUT FROM THE FUNCTION MINIMIZATION HAS BEEN REQUESTED" ) 12870
776 FORMAT( " PARTIAL PROGRESS REPORT OF THE FUNCTION MINIMIZATION" , 12880
* " HAS BEEN REQUESTED" ) 12890
778 FORMAT( " FULL PROGRESS REPORT OF THE FUNCTION MINIMIZATION" , 12900
* " HAS BEEN REQUESTED" ) 12910
780 FORMAT( " PLOT OF THE FITTED CURVE AND DATA HAS BEEN REQUESTED" ) 12920
781 FORMAT( " WEIGHTING HAS BEEN DONE ACCORDING TO SCHEME NO." ,I2, 12930
* " (WITH R=" ,E10.4,") ) 12940
789 FORMAT(/1X,16(1H*) " SUM-OF-SQUARES FUNCTION MINIMIZATION REPORT " 12950
* ,16(1H*) / / ) 12960
790 FORMAT(/1X,77(1H*) / / ) 12970
791 FORMAT(// " IT IS NECESSARY TO MAKE CORRECTION IN YOUR INPUT" , 12980
* " TO REACH CONVERGENCE " ) 12990
792 FORMAT( " PROBABLY BECAUSE OF POSSIBLE CONSTRAIN(S) AS SEEN" , 13000
* " FROM " ) 13010

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G02

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793 FORMAT(// " THE REASON CAN ALSO BE UNSUITABLE VALUE(S) OF" ) 13020
794 FORMAT(// " TRY NEW INITIAL ESTIMATES OR RESCALE THE PARAMETERS" , 13030
* " OR CHANGE THE VALUE OF" ) 13040
795 FORMAT(// " OR CHANGE THE LIMITS OF ONE OR MORE OF THE PARAMETERS" ) 13050
800 FORMAT(/1X,31(1H*) " R E S U L T S " ,31(1H*) / / 13060
* 1X,77(1H*) / / / / 13070
* " PAR.NO. ALLOWED RANGE" ,5X, " INITIAL VALUE FINAL VALUE STD." 13080
* " DEV. " ,4X, " C.V.(PCT)" /1X,77(1H-) ) 13090
8010 FORMAT(/1X,31(1H*) " R E S U L T S " ,31(1H*) / / 13100
* 1X,77(1H*) / / / / 13110
* " PAR.NO. ALLOWED RANGE" ,5X, " INITIAL VALUE FINAL VALUE" / 13120
* 1X,57(1H-) ) 13130
8011 FORMAT(14,3X,4(E11.5,1X)) 13140
8012 FORMAT(1X,57(1H-)) 13150
802 FORMAT(14,3X,6(E11.5,1X) ) 13160
803 FORMAT(1X,77(1H-) / / / / 13170
* " PAR.NO. APPROX. 95 PCT. CONF.LIMITS" ,/1X,38(1H-) ) 13180
8030 FORMAT(// " THE MINIMUM FOUND BY THE MINIMIZATION" , 13190
* " PROCESS APPEARS" / " TO BE CONSTRAINED BY THE " ,A5, 13200
* " LIMIT OF PARAMETER NUMBER " ,I2 ) 13210
8031 FORMAT(// 13220
* " *** ENTER B10 OR B11 IF YOU WANT A NEW RUN WITH DIFFERENT" / 13230
* " *** PARAMETER LIMITS OR DIFFERENT PARAMETER ESTIMATES" / 13240
* " *** OTHERWISE TYPE -OK- TO CONTINUE" / / ) 13250
804 FORMAT(// 13260

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IFHFN=LABEL(1)	21550
S=Y(I)	21560
IF(NIND.EQ.1) CALL MODEL(S,A(1),F,0)	21570
IF(NIND.GT.1) CALL MODEL(S,A,F,0)	21580
IF(S.GE.1E50) GO TO 20	21590
YCAL=S	21600
YOBS=Y(I)	21610
W=MYN(I)	21620
15 FUNC=FUNC+W*(YOBS-YCAL)*(YOBS-YCAL)	21630
RETURN	21640
20 FUNC=S+1.000000001	21650
NVIOL=NVIOL+1	21660
RETURN	21670
END	21680
*****	21690

F03

SUBROUTINE READ(N,AR)	21700
DIMENSION A(78),AR(12),AL(10)	21710
COMMON/B5/NO,NS,IGO	21720
COMMON /B15/SKIP	21730
LOGICAL SKIP	21740
DATA YES,ANO,BACK,AD,AK,BLANK,PLUS,AMINUS,COMMA,DOT,E	21750
*/1HY,1HN,1HB,1HD,1HK,1H,1H+,1H-,1H.,1H.,1HE/	21760
DATA AL/1HO,1H1,1H2,1H3,1H4,1H5,1H6,1H7,1H8,1H9 /	21770
5 IGO=1	21780
SKIP=.FALSE.	21790
DO 6 L=1,78	21800
6 A(L)=BLANK	21810
DO 7 L=1,12	21820
7 AR(L)=0.0	21830
READ(5,200)A	21840
IF(EOF(5))8,9,8	21850
8 WRITE(6,225)	21860
GO TO 5	21870
9 CONTINUE	21880
DO 11 I=1,78	21890
IF(A(I).EQ.1HR) GO TO 139	21900
IF(A(I).EQ.BACK) GO TO 140	21910
11 CONTINUE	21920
IF(A(I).EQ.YES) GO TO 130	21930
IF(A(I).EQ.ANO) GO TO 135	21940
IF(A(I).EQ.AD.AND.A(2).EQ.AK) GO TO 145	21950
I=0	21960
J=1	21970
K=0	21980
FAC=1.	21990
GO TO 15	22000
10 FAC=-1.	22010
15 I=I+1	22020
IF(I.EQ.79) GO TO 125	22030
DO 20 L=1,10	22040
IF(A(I).EQ.AL(L)) GO TO 50	22050
20 CONTINUE	22060
IF(K.EQ.0) GO TO 30	22070
IF(A(I).EQ.BLANK.OR.A(I).EQ.COMMA.OR.A(I).EQ.DOT.OR.A(I).EQ.E)	22080
*GO TO 55	22090
25 WRITE(6,205)A(I)	22100
GO TO 5	22110
30 IF(A(I).EQ.BLANK) GO TO 15	22120
IF(J.GT.1.AND.A(I).EQ.COMMA) GO TO 15	22130
IF(A(I).EQ.AMINUS) GO TO 10	22140
IF(A(I).EQ.DOT) GO TO 71	22150
GO TO 25	22160
50 IF(K.EQ.0) IFLAG=I	22170
AR(J)=AR(J)+FLOAT(L-1)*(10.**(-K))	22180
K=K+1	22190
GO TO 15	22200
55 AR(J)=FAC*AR(J)*(10.**(-IFLAG-1))	22210
IF(A(I).EQ.DOT.AND.A(I+1).EQ.COMMA.OR.A(I+1).EQ.BLANK) GO TO 60	22220
IF(A(I).EQ.DOT) GO TO 70	22230
IF(A(I).EQ.E) GO TO 90	22240
60 J=J+1	22250
IF(J.GT.N) GO TO 65	22260
FAC=1.	22270
K=0	22280
GO TO 15	22290
65 I=I+1	22300
IF(I.EQ.79) RETURN	22310

G03

IF(A(I).EQ.BLANK) GO TO 65	22320
WRITE(6,215)N	22330
GO TO 5	22340
70 K=0	22350
71 I=I+1	22360
K=K+1	22370
DO 75 L=1,10	22380
IF(A(I).EQ.AL(L)) GO TO 85	22390
75 CONTINUE	22400
IF(A(I).EQ.BLANK.OR.A(I).EQ.COMMA) GO TO 60	22410
IF(A(I).EQ.E) GO TO 90	22420
GO TO 25	22430
85 AR(J)=AR(J)+FAC*FLOAT(L-1)*(10.**(-K))	22440
GO TO 71	22450
90 KK=0	22460
NEXP=0	22470
NFAC=1	22480
NCOUNT=0	22490
GO TO 100	22500
95 NFAC=-1	22510
100 I=I+1	22520
IF(I.EQ.79) GO TO 126	22530
NCOUNT=NCOUNT+1	22540
IF(NCOUNT.EQ.5) GO TO 126	22550
IF(NCOUNT.GT.1) GO TO 105	22560
IF(A(I).EQ.BLANK.OR.A(I).EQ.COMMA) GO TO 100	22570

109	CONTINUE	3620
	WRITE(6,642)NOBS	3630
	GO TO 105	3640
120	NN=NIND+2	3650
	WRITE(6,655)NOBS,NN,NIND	3660
	IW=2	3670
	DO 123 I=1,NOBS2	3680
	J=J+1	3690
	N=LABEL(J)	3700
	IF(N.EQ.NSAVE+1) WRITE(6,641)ITHSET(N),N,J	3710

G01

	NSAVE=N	3720
	CALL READ(NN,AR)	3730
	IF(IG0.EQ.2) GO TO 1001	3740
	IF(ANS.EQ.4HBACK) GO TO 1004	3750
	J=IFIX(ANS+.1)	3760
	IF(J.LT.1) GO TO 1007	3770
	IF(J.GT.NOBS) GO TO 121	3780
	GO TO 122	3790
121	NAGR=8	3800
	GO TO 1008	3810
122	CONTINUE	3820
	DO 1220 L=1,NIND	3830
1220	XX(L,J)=AR(L+2)	3840
	INDEX(J)=J	3850
	Y(J)=AR(2)	3860
	IF(J.EQ.NOBS) GO TO 125	3870
123	CONTINUE	3880
	WRITE(6,642)NOBS	3890
	GO TO 120	3900
125	CONTINUE	3910
	DO 126 I=1,NOBS	3920
	X(I)=XX(1,I)	3930
126	IF(NIND.GT.1) Z(I)=XX(2,I)	3940
	IF(NS.GT.ND) GO TO 1006	3950
130	NQ=18	3960
	WRITE(6,660)	3970
	CALL READ(1,AR)	3980
	IF(IG0.EQ.2) GO TO 1001	3990
	IF(ANS.EQ.4HBACK) GO TO 1004	4000
	IF(ANS.EQ.2HOK) GO TO 131	4010
	WRITE(6,1027)	4020
	GO TO 130	4030
131	IF(NFUNC.EQ.-1.OR.NOP.EQ.-1.OR.NOP2.EQ.-1.OR.NOBS2.EQ.-1)GOTO 1013	4040
	DO 133 I=1,NOBS	4050
	IF(ANS16.NE.3HYES) GO TO 1310	4060
	IF(IWGHT.GT.1) WY(I)=WEIGHT(Y(I),IWGHT,ANS16B)	4070
	IF(WY(I).LT.0.) GO TO 1012	4080
1310	CONTINUE	4090
	DO 132 J=1,NOBS2	4100
	IF(INDEX(J).EQ.I) GO TO 133	4110
132	CONTINUE	4120
	J=0	4130
	NSAVE=1	4140
	WRITE(6,1032)I	4150
	GO TO (106,120) IW	4160
133	CONTINUE	4170
	IF(NLOOP.GT.MAX.AND.MAX.NE.1) GO TO 1010	4180
	DO 144 I=1,NOP	4190
	IF(ANS11.EQ.2HND) GO TO 143	4200
	DO 141 J=1,N11	4210
	K=NF(J)	4220
	IF(K.EQ.I) GO TO 142	4230
141	CONTINUE	4240
	GO TO 143	4250
142	IF(F1(K).EQ.F2(K)) GO TO 143	4260
	IF(FS(K).LT.F1(K).OR.FS(K).GT.F2(K)) GO TO 1420	4270
	F(K)=FS(K)	4280
	FSAVE(K)=F(K)	4290
	GO TO 144	4300
1420	WRITE(6,1030)K	4310
	CALL READ(1,AR)	4320
	IF(IG0.EQ.2) GO TO 1001	4330

H01

	IF(ANS.EQ.4HBACK) GO TO 1004	4340
	IF(ANS.EQ.2HOK) GO TO 143	4350
	WRITE(6,1027)	4360
	GO TO 1420	4370
143	F(I)=F1(I)+0.5*(F2(I)-F1(I))	4380
	FSAVE(I)=F(I)	4390
144	CONTINUE	4400
	NEWNOP=NOP	4410
	DO 150 I=1,NOP	4420
	IF(ANS12.EQ.2HND) GO TO 148	4430
	DO 145 J=1,N12	4440
	K=NT(J)	4450
	IF(K.EQ.I) GO TO 146	4460
145	CONTINUE	4470
	GO TO 148	4480
146	IF((STPS(I)+F(I)).LT.F1(I).OR.(STPS(I)+F(I)).GT.F2(I)) GO TO 147	4490
	STEP(I)=STPS(I)	4500
	GO TO 149	4510
147	WRITE(6,1031)I	4520
	CALL READ(1,AR)	4530
	IF(IG0.EQ.2) GO TO 1001	4540
	IF(ANS.EQ.4HBACK) GO TO 1004	4550
	IF(ANS.EQ.2HOK) GO TO 148	4560
	WRITE(6,1027)	4570
	GO TO 147	4580
148	DEL1=F(I)-F1(I)	4590
	DEL2=F2(I)-F(I)	4600
	IF(DEL1.GT.DEL2) STEP(I)=-(.3+.1/FLOAT(I))*DEL1	4610
	IF(DEL1.LE.DEL2) STEP(I)=(.3+.1/FLOAT(I))*DEL2	4620
	STPS(I)=STEP(I)	4630
149	STEPS(I)=STEP(I)	4640
	IF(DEL1.EQ.F2(I)) STEP(I)=0	4650

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781 FORMAT(" WEIGHTING HAS BEEN DONE ACCORDING TO SCHEME NO.",I2, 12930
* (WITH R=,E10.4,") ) 12940
789 FORMAT(/1X,16(1H+), " SUM-OF-SQUARES FUNCTION MINIMIZATION REPORT " 12950
* ,16(1H+)// ) 12960
790 FORMAT(/1X,77(1H+)// ) 12970
791 FORMAT(" IT IS NECESSARY TO MAKE CORRECTION IN YOUR INPUT", 12980
* " TO REACH CONVERGENCE. " ) 12990
792 FORMAT(" PROBABLY BECAUSE OF POSSIBLE CONSTRAINT(S) AS SEEN", 13000
* " FROM " ) 13010

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793 FORMAT(" THE REASON CAN ALSO BE UNSUITABLE VALUE(S) OF " ) 13020
794 FORMAT(" TRY NEW INITIAL ESTIMATES OR RESCALE THE PARAMETERS", 13030
* " OR CHANGE THE VALUE OF " ) 13040
795 FORMAT(" OR CHANGE THE LIMITS OF ONE OR MORE OF THE PARAMETERS" ) 13050
800 FORMAT(/1X,31(1H+), " R E S U L T S ",31(1H+)// 13060
* 1X,77(1H+)// ) 13070
* " PAR.NO. ALLOWED RANGE",5X," INITIAL VALUE FINAL VALUE STD." 13080
* " DEV.",4X," C.V.(PCT)"/1X,77(1H-)) 13090
8010 FORMAT(/1X,31(1H+), " R E S U L T S ",31(1H+)// 13100
* 1X,77(1H+)// ) 13110
* " PAR.NO. ALLOWED RANGE",5X," INITIAL VALUE FINAL VALUE"/ 13120
* 1X,57(1H-)) 13130
8011 FORMAT(14,3X,4(E11.5,1X)) 13140
8012 FORMAT(1X,57(1H-)) 13150
802 FORMAT(14,3X,6(E11.5,1X) ) 13160
803 FORMAT(1X,77(1H-)// ) 13170
* " PAR.NO. APPROX. 95 PCT. CONF.LIMITS",/1X,38(1H-)) 13180
8030 FORMAT(" THE MINIMUM FOUND BY THE MINIMIZATION", 13190
* " PROCESS APPEARS TO BE CONSTRAINED BY THE ",A5, 13200
* " LIMIT OF PARAMETER NUMBER ",I2 ) 13210
8031 FORMAT( 13220
* " *** ENTER B10 OR B11 IF YOU WANT A NEW RUN WITH DIFFERENT"/ 13230
* " *** PARAMETER LIMITS OR DIFFERENT PARAMETER ESTIMATES"/ 13240
* " *** OTHERWISE TYPE -OK- TO CONTINUE"// ) 13250
804 FORMAT( 13260
* " RES. SUM OF SQUARES",4X,E12.6,5X," REG. SUM OF SQUARES",5X, 13270
* E12.6/ 13280
* " TOT. SUM OF SQUARES",4X,E12.6,5X," SUM OF SQUARED RESPONSE ", 13290
* E12.6/ 13300
* " MEAN OF RESPONSE",7X,E12.6,5X," RES. MEAN SQUARE",7X,E12.6/ 13310
* " REG. MEAN SQUARE",7X,E12.6,5X," MEAN OF RESIDUALS",7X,E12.6/ 13320
* " STD. DEV. OF RESIDUALS",E12.6,5X," PCT. RESPONSE EXPLAINED", 13330
* E12.6/ " CORR. COEFFICIENT R",2X,E12.6/ ) 13340
805 FORMAT(14,6X,2E13.4) 13350
8050 FORMAT(1X,38(1H-)) 13360
8051 FORMAT(" RELATIVE POSITION OF THE CALCULATED PARAMETER", 13370
* " IN ITS ALLOWED RANGE. / ) 13380
8052 FORMAT(" RELATIVE POSITIONS OF THE CALCULATED PARAMETERS", 13390
* " IN THEIR ALLOWED RANGE. / ) 13400
8053 FORMAT(" PAR.NO.",I2,3X,65A1) 13410
8054 FORMAT(" RESIDUAL SUM OF SQUARES =",E12.6/ ) 13420
8055 FORMAT(" WEIGHTED RESIDUAL SUM OF SQUARES =",E12.6/ ) 13430
806 FORMAT( 13440
* " WGT. RES. SUM OF SQUARES",E12.6,5X," WGT. RES. MEAN SQUARE",3X, 13450
* E12.6/ 13460
* " MEAN OF WGT. RESIDUALS",E12.6,5X," STD. DEV. OF WGT. RES.",2X, 13470
* E12.6/ ) 13480
807 FORMAT(" THE RESIDUAL SUM OF SQUARES FOR RESPONSE SYSTEM NO." 13490
* ,I2," IS",E12.6 ) 13500
808 FORMAT(" THE WEIGHTED RESIDUAL SUM OF SQUARES FOR RESPONSE", 13510
* " SYSTEM NO.",I2," IS",E12.6) 13520
810 FORMAT(" ---THE REGRESSION IS NOT SIGNIFICANT---") 13530
812 FORMAT(" OBS.NO. X1",9X,"X2 RESPONSE CALC.RESPONSE DIFF.", 13540
* " DIFF(PCT) NORM.DEVIATE"/1X,77(1H-)) 13550
813 FORMAT(/28X," RESPONSE SYSTEM NO.",I2/ ) 13560
814 FORMAT(14,3X,E10.4,10X,5E10.4) 13570
816 FORMAT(14,3X,7E10.4) 13580
817 FORMAT(" OBS.NO.",7(" X",11,4X)/1X,77(1H-)) 13590
818 FORMAT(1X,77(1H-)) 13600
819 FORMAT(15,2X,7E10.4) 13610
820 FORMAT(" ASSUMING A NORMAL DISTRIBUTION OF ERRORS,THE UNIT NORMAL" 13620
* " DEVIATE FORM"/ OF THE RESIDUALS TOGETHER WITH THE T-DIS", 13630

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H02

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* "TRIBUTION WITH",I3," DEGREES OF FREEDOM"/ INDICATES THAT") 13640
822 FORMAT(20X," OBSERVATION NUMBER",I3) 13650
824 FORMAT(" IS AN OUTLIER (P.LT.0.05)"/ ) 13660
8240 FORMAT(/21X," RESPONSE SYSTEM NO.",I2/ ) 13670
8241 FORMAT(" OBS.NO. WEIGHT NORM.WEIGHT WEIGHTED RESIDUAL", 13680
* " UNIT NORMAL DEVIATE"/47X," (CRIT.VALUES="+",F4.2,")"/ 13690
* 1X,67(1H-)) 13700
8242 FORMAT(14,A2,2E12.4,2E15.4) 13710
8243 FORMAT(14,2X,2E12.4,2E15.4) 13720
8244 FORMAT(1X,67(1H-)) 13730
8245 FORMAT(" THE OBSERVATION MARKED * IN THE TABLE ABOVE"/ 13740
* " APPEARS TO BE AN OUTLIER. / ) 13750
8246 FORMAT(" THE OBSERVATIONS MARKED * IN THE TABLE ABOVE"/ 13760
* " APPEAR TO BE OUTLIERS. / ) 13770
826 FORMAT(" ARE OUTLIERS (P.LT.0.05)"/ ) 13780
8261 FORMAT(" THE FITTED CURVE CAN BE PLOTTED FROM FOLLOWING 51 POINTS" 13790
* "/2(" PT.NO. X",8X," RESPONSE")/1X,58(1H-)) 13800
8262 FORMAT(2(14,3X,2E11.4)) 13810
8263 FORMAT(1X,58(1H-)) 13820
8264 FORMAT(/18X," RESPONSE SYSTEM NO.",I2/ ) 13830
827 FORMAT(" VARIANCE-COVARIANCE MATRIX OF THE PARAMETERS. / ) 13840
8270 FORMAT(" CORRELATION MATRIX / ) 13850
829 FORMAT(1X,6E13.6) 13860
830 FORMAT(" DURBIN WATSON STATISTIC FOR SERIAL CORRELATION OF", 13870
* " RESIDUALS =",E10.4/ (REF. J.DURBIN AND G.S.WATSON, 13880
* " BIOMETRICA,PP 157-178,1951) / ) 13890
832 FORMAT(" THE PROBABILITY OF RANDOMNESS OF RUNS =",E12.4/ 13900
* " (REF. F.S.SWED AND C.EISENHART, ANN. OF MATH. STATS.,14," 13910
* " PP66-87,43) / ) 13920
8320 FORMAT(" ASSUMING EQUAL PROBABILITY (0.5) OF GETTING NEGATIVE AND" 13930
* " POSITIVE"/ RESIDUALS, THE PROBABILITY OF GETTING",I2, 13940
* " OR LESS RESIDUALS OF SAME SIGN"/ OF A TOTAL OF",I3, 13950

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IF(A(I).EQ.001) GO TO 70	22230
IF(A(I).EQ.E) GO TO 90	22240
60 J=J+1	22250
IF(J.GT.N) GO TO 65	22260
FAC=1.	22270
K=0	22280
GO TO 15	22290
65 I=I+1	22300
IF(I.EQ.79) RETURN	22310

G03

IF(A(I).EQ.BLANK) GO TO 65	22320
WRITE(6,215)N	22330
GO TO 5	22340
70 K=0	22350
71 I=I+1	22360
K=K+1	22370
DO 75 L=1,10	22380
IF(A(I).EQ.AL(L)) GO TO 85	22390
75 CONTINUE	22400
IF(A(I).EQ.BLANK.OR.A(I).EQ.COMMA) GO TO 60	22410
IF(A(I).EQ.E) GO TO 90	22420
GO TO 25	22430
85 AR(J)=AR(J)+FAC*FLOAT(L-1)*(10.**(-K))	22440
GO TO 71	22450
90 KK=0	22460
NEXP=0	22470
NFAC=1	22480
NCOUNT=0	22490
GO TO 100	22500
95 NFAC=-1	22510
100 I=I+1	22520
IF(I.EQ.79) GO TO 126	22530
NCOUNT=NCOUNT+1	22540
IF(NCOUNT.EQ.5) GO TO 126	22550
IF(NCOUNT.GT.1) GO TO 105	22560
IF(A(I).EQ.BLANK.OR.A(I).EQ.PLUS) GO TO 100	22570
IF(A(I).EQ.AMINUS) GO TO 95	22580
105 CONTINUE	22590
DO 110 L=1,10	22600
IF(A(I).EQ.AL(L)) GO TO 115	22610
110 CONTINUE	22620
IF(KK.EQ.0) GO TO 25	22630
IF(A(I).EQ.BLANK.OR.A(I).EQ.COMMA) GO TO 120	22640
GO TO 25	22650
115 IF(KK.EQ.1) NEXP=10*NEXP	22660
NEXP=NEXP+NFAC*(L-1)	22670
KK=KK+1	22680
GO TO 100	22690
120 AR(J)=AR(J)*(10.**NEXP)	22700
GO TO 60	22710
125 WRITE(6,210)N	22720
GO TO 5	22730
126 WRITE(6,220)	22740
GO TO 5	22750
130 AR(1)=3HYES	22760
RETURN	22770
135 AR(1)=2HNO	22780
RETURN	22790
139 SKIP=.TRUE.	22800
140 AR(1)=4HBACK	22810
J1=I+1	22820
J2=I+2	22830
IF(A(I+3).NE.1HK) GO TO 141	22840
J1=I+4	22850
J2=I+5	22860
141 N1=11	22870
N2=11	22880
DO 142 I=1,10	22890
IF(A(J1).EQ.AL(I)) N1=I-1	22900
142 IF(A(J2).EQ.AL(I)) N2=I-1	22910
IF(N1.EQ.11.OR.N2.EQ.11.AND.A(J2).NE.BLANK.OR.	22920
*N1.EQ.0.AND.N2.EQ.0) RETURN	22930

H03

NS=NO	22940
NO=10*N1+N2	22950
IF(A(J2).EQ.BLANK) NO=N1	22960
IGD=2	22970
IF(SKIP.AND.NO.GE.1.AND.NO.LE.22) RETURN	22980
IF(NO.GE.1.AND.NO.LE.NS) RETURN	22990
NO=NS	23000
WRITE(6,230)	23010
GO TO 5	23020
145 AR(1)=ZHOK	23030
RETURN	23040
200 FORMAT(78A1)	23050
205 FORMAT(/" INPUT ERROR. UNRECOGNIZED CHARACTER ( ",A1," )"/	23060
* REENTER LAST LINE IN CORRECT FORM. "/)	23070
210 FORMAT(/" INPUT ERROR. TOO FEW NUMBERS ON LINE ABOVE.",I2,	23080
* ARE EXPECTED. "/ REENTER LAST LINE IN CORRECT FORM ACCORDING",	23090
* TO REQUEST. "/)	23100
215 FORMAT(/" INPUT ERROR. TOO MANY NUMBERS ON LINE ABOVE.",I2,	23110
* EXPECTED. "/ REENTER LAST LINE IN CORRECT FORM ACCORDING",	23120
* TO REQUEST. "/)	23130
220 FORMAT(/" INPUT ERROR. -WRONG EXPONENT-"/	23140
* REENTER LAST LINE IN CORRECT FORM ACCORDING TO REQUEST. "/)	23150
225 FORMAT(/" INPUT ERROR, YOU FORGOT TO ENTER DATA. -TRY AGAIN- / )	23160
230 FORMAT(/" INPUT ERROR, ILLIGAL TRANSFER COMMAND - TRY AGAIN -"/)	23170
END	23180
*****	23190
C SUBROUTINE PLACE(F1,F2,F,NP,NOP,A)	23200
DIMENSION A(650),F1(20),F2(20),F(20)	23210
DATA DOT,X,BLANK /1H-,1HX,1H /	23220
DO 20 J=1,NOP	23230
N1=NP*(J-1)+1	23240
N2=N1+NP-1	23250

IF(FS(K).LT.F1(K)).OR.FS(K).GT.F2(K)) GO TO 1420	4270
F(K)=FS(K)	4280
FSAVE(K)=F(K)	4290
GO TO 144	4300
1420 WRITE(6,1030)K	4310
CALL READ(1,AR)	4320
IF(IGO.EQ.2) GO TO 1001	4330

H01

IF(ANS.EQ.4HBACK) GO TO 1004	4340
IF(ANS.EQ.2HOK) GO TO 143	4350
WRITE(6,1027)	4360
GO TO 1420	4370
143 F(I)=F1(I)+0.5*(F2(I)-F1(I))	4380
FSAVE(I)=F(I)	4390
144 CONTINUE	4400
NEWNOP=NOP	4410
DO 150 I=1,NOP	4420
IF(ANS12.EQ.2HNO) GO TO 148	4430
DO 145 J=1,N12	4440
K=NT(J)	4450
IF(K.EQ.I) GO TO 146	4460
145 CONTINUE	4470
GO TO 148	4480
146 IF((STPS(I)+F(I)).LT.F1(I).OR.(STPS(I)+F(I)).GT.F2(I)) GO TO 147	4490
STEP(I)=STPS(I)	4500
GO TO 149	4510
147 WRITE(6,1031)I	4520
CALL READ(1,AR)	4530
IF(IGO.EQ.2) GO TO 1001	4540
IF(ANS.EQ.4HBACK) GO TO 1004	4550
IF(ANS.EQ.2HOK) GO TO 148	4560
WRITE(6,1027)	4570
GO TO 147	4580
148 DEL1=F(I)-F1(I)	4590
DEL2=F2(I)-F(I)	4600
IF(DEL1.GT.DEL2) STEP(I)=-(.3+0.1/FLOAT(I))*DEL1	4610
IF(DEL1.LE.DEL2) STEP(I)=(.3+0.1/FLOAT(I))*DEL2	4620
STPS(I)=STEP(I)	4630
149 STEPS(I)=STEP(I)	4640
IF(F1(I).EQ.F2(I)) STEP(I)=0.	4650
IF(F1(I).EQ.F2(I)) NEWNOP=NEWNOP-1	4660
150 CONTINUE	4670
IF(NEWNOP.GT.NOBS) GO TO 1011	4680
NOWGHT=0	4690
IF(ANS16.NE.3HYES) GO TO 1525	4700
IF(IWGHT.NE.1) GO TO 1510	4710
N=0	4720
NDUMMY=0	4730
DO 1500 I=1,NOBS	4740
IF(WY(I).EQ.0.) NDUMMY=NDUMMY+1	4750
1500 IF(WY(I).EQ.0..OR.WY(I).EQ.1.) N=N+1	4760
IF(NDUMMY.EQ.0..OR.N.NE.NOBS) GO TO 1510	4770
NOWGHT=1	4780
DO 1505 I=1,NOBS	4790
1505 WYN(I)=WY(I)	4800
GO TO 155	4810
1510 SUMM=0.	4820
DO 1515 I=1,NOBS	4830
1515 SUMM=SUMM+WY(I)	4840
FACTOR=FLOAT(NOBS-NDUMMY)/SUMM	4850
DO 1520 I=1,NOBS	4860
1520 WYN(I)=FACTOR*WY(I)	4870
GO TO 155	4880
1525 DO 1530 I=1,NOBS	4890
WY(I)=1.	4900
1530 WYN(I)=1.	4910
155 CONTINUE	4920
157 WRITE(6,662)	4930
CALL READ(1,AR)	4940
IF(IGO.EQ.2) GO TO 1001	4950

I01

IF(ANS.EQ.4HBACK) GO TO 1004	4960
IF(ANS.EQ.3HYES) GO TO 300	4970
IF(ANS.EQ.2HNO) GO TO 159	4980
WRITE(6,1020)	4990
GO TO 157	5000
158 WRITE(6,664)	5010
CALL READ(1,AR)	5020
IF(IGO.EQ.2) GO TO 1001	5030
IF(ANS.EQ.4HBACK) GO TO 1004	5040
IF(ANS.EQ.2HOK) GO TO 159	5050
WRITE(6,1027)	5060
GO TO 158	5070
159 IF(NRUN.EQ.0.AND.MAX.NE.1) WRITE(6,670)	5080
GO TO 350	5090
160 NO=19	5100
NEWNOP=NOP	5110
DO 1600 I=1,NOP	5120
1600 IF(F1(I).EQ.F2(I)) NEWNOP=NEWNOP-1	5130
IF(NEWNOP.EQ.0..OR.NOP.GT.7) GO TO 1666	5140
IF(MAX.EQ.1) GO TO 162	5150
WRITE(6,6700)NEWNOP	5160
CALL READ(1,AR)	5170
GO TO (1000,1001) IGO	5180
161 IF(ANS.EQ.2HNO) GO TO 1666	5190
162 IF(NEWNOP.LT.3) GO TO 165	5200
WRITE(6,6705) NRECOM(NEWNOP)	5210
163 CALL READ(1,AR)	5220
IF(IGO.EQ.2) GO TO 1001	5230
IF(ANS.EQ.4HBACK) GO TO 1004	5240
NPOINT=1+IX(ANS+.1)	5250
IF(NPOINT.NE.1.NCRIT(NEWNOP).OR.NPOINT.GT.2187) GO TO 164	5260
GO TO 165	5270
164 WRITE(6,162)	5280
GO TO 163	5290

814	FORMAT( I4,3X,E10.4,10X,5E10.4 )	13570
816	FORMAT( I4, 3X,7E10.4 )	13580
817	FORMAT( / OBS.NO.,7( " X",11,4X)/1X,77(1H-) )	13590
818	FORMAT( 1X,77(1H-) / )	13600
819	FORMAT(15,2X,7E10.4 )	13610
820	FORMAT( " ASSUMING A NORMAL DISTRIBUTION OF ERRORS,THE UNIT NORMAL"	13620
	*, " DEVIATE FORM" / OF THE RESIDUALS TOGETHER WITH THE T-DIS",	13630

H02

	*"TRIBUTION WITH",13," DEGREES OF FREEDOM" /" INDICATES THAT")	13640
822	FORMAT(20X," OBSERVATION NUMBER ",I3)	13650
824	FORMAT( / " IS AN OUTLIER (P.LT.0.05)" / )	13660
8240	FORMAT(/21X," RESPONSE SYSTEM NO.",I2/ )	13670
8241	FORMAT( / " OBS.NO. WEIGHT NORM.WEIGHT WEIGHTED RESIDUAL",	13680
	*" UNIT NORMAL DEVIATE"/47X," (CRIT.VALUES=+-",F4.2,") /	13690
	*1X,67(1H-) )	13700
8242	FORMAT(I4,A2,2E12.4,2E15.4 )	13710
8243	FORMAT(I4,2X,2E12.4,2E15.4 )	13720
8244	FORMAT(1X,67(1H-) / )	13730
8245	FORMAT( " THE OBSERVATION MARKED * IN THE TABLE ABOVE"/	13740
	*" APPEARS TO BE AN OUTLIER." / )	13750
8246	FORMAT( " THE OBSERVATIONS MARKED * IN THE TABLE ABOVE"/	13760
	*" APPEAR TO BE OUTLIERS." / )	13770
826	FORMAT( / " ARE OUTLIERS (P.LT.0.05)" / )	13780
8261	FORMAT( " THE FITTED CURVE CAN BE PLOTTED FROM FOLLOWING 51 POINTS"	13790
	*//2( " PT.NO. X",8X," RESPONSE")/1X,58(1H-) )	13800
8262	FORMAT(2(I4,3X,2E11.4))	13810
8263	FORMAT(1X,58(1H-) )	13820
8264	FORMAT( /18X," RESPONSE SYSTEM NO.",I2/ )	13830
827	FORMAT( / " VARIANCE-COVARIANCE MATRIX OF THE PARAMETERS." / )	13840
8270	FORMAT( / " CORRELATION MATRIX" / )	13850
829	FORMAT(1X,6E13.6)	13860
830	FORMAT( / " DURBIN WATSON STATISTIC FOR SERIAL CORRELATION OF",	13870
	*" RESIDUALS =" ,E10.4/" (REF. J.DURBIN AND G.S.WATSON,"	13880
	*" BIOMETRICA,PP 157-178,1951)" / )	13890
832	FORMAT( " THE PROBABILITY OF RANDOMNESS OF RUNS =" ,E12.4/	13900
	*" (REF. S.SWED AND C.EISENHART, ANN. OF MATH. STATS.,14,"	13910
	*" PP66-87,1943)" / )	13920
8320	FORMAT( " ASSUMING EQUAL PROBABILITY (0.5) OF GETTING NEGATIVE AND"	13930
	*" POSITIVE" /" RESIDUALS, THE PROBABILITY OF GETTING ",I2,	13940
	*" OR LESS RESIDUALS OF SAME SIGN" /" OF A TOTAL OF",I3,	13950
	*" RESIDUALS IS =" ,E11.4 / )	13960
833	FORMAT( " THIS INDICATES THAT THE ERRORS ARE NOT RANDOM AT THE",	13970
	*" MINIMUM FOUND" / )	13980
834	FORMAT( " THE RATIO OF THE DEGREES OF FREEDOM OF RESIDUALS TO THE",	13990
	*" NUMBER" /" OF RESIDUALS =" ,E10.4," A SIGNIFICANT CORRELATION"	14000
	*" BETWEEN THE RESIDUALS" /" IS THEREFORE EXPECTED,VIOLATING THE",	14010
	*" ASSUMPTION BEHIND THE STATISTICAL" /" EVALUATION OF THE",	14020
	*" PARAMETERS." )	14030
836	FORMAT(/8X,	14040
	*" PLOT OF RESIDUALS VERSUS INDEPENDENT VARIABLE X1 (SCL.UNITS)" )	14050
8360	FORMAT(/2X," PLOT OF WEIGHTED RESIDUALS VERSUS",	14060
	*" INDEPENDENT VARIABLE X1 (SCL.UNITS)" )	14070
838	FORMAT(/11X,	14080
	*" PLOT OF RESIDUALS VERSUS RESPONSE ESTIMATE (SCL.UNITS)" )	14090
8380	FORMAT(/5X," PLOT OF WEIGHTED RESIDUALS",	14100
	*" VERSUS RESPONSE ESTIMATE (SCL.UNITS)" )	14110
840	FORMAT(/5X," PLOT OF RESIDUALS VERSUS 1ST. INDEPENDENT",	14120
	*" VARIABLE X1 (SCL.UNITS)" )	14130
8400	FORMAT(/2X," PLOT OF WEIGHTED RESIDUALS VERSUS",	14140
	*" 1ST. INDEPENDENT VARIABLE X1 (SCL.UNITS)" )	14150
842	FORMAT(/5X," PLOT OF RESIDUALS VERSUS 2ND. INDEPENDENT VARIABLE",	14160
	*" X2 (SCL.UNITS)" )	14170
8420	FORMAT(/2X," PLOT OF WEIGHTED RESIDUALS VERSUS",	14180
	*" 2ND. INDEPENDENT VARIABLE X2 (SCL.UNITS)" )	14190
8401	FORMAT( " CONVERGENCE CRITERION ",E10.4 )	14200
8402	FORMAT( " EXPANSION CRITERION ",E12.4 )	14210
8403	FORMAT( " MINIMUM OF FITTED QUADRATIC SURFACE IS ",E15.8/	14220
	*" COMPARE WITH MINIMUM FOUND BY ITERATION ",E15.8/	14230
	*" IF DIFFERENCE IS LARGE THE STATISTICAL EVALUATION OF THE",	14240
	*" PARAMETERS IS" /" INACCURATE AND A NEW RUN WITH A DIFFERENT",	14250

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	*" STOPPING CRITERION,EXPANSION" /" CRITERION OR DIFFERENT RANGE",	14260
	*" OF THE PARAMETERS IS RECOMMENDED." )	14270
8404	FORMAT( /" TYPE BACK (8*) IF YOU WANT TO MAKE CORRECTIONS OR",	14280
	*" OK IF YOU WANT" /" TO CONTINUE BUT WITHOUT STATISTICAL",	14290
	*" EVALUATION OF THE PARAMETERS" / )	14300
8422	FORMAT( " NO. OF SS EVALUATIONS ",I4,17X,	14310
	*" NO. OF CONSTR. VIOLATIONS ",I4,	14320
8424	FORMAT( " THE QUADRATIC SURFACE FITTING REQUIRED A FURTHER",I4,	14330
	*" SS EVALUATIONS" )	14340
8425	FORMAT(/1H )	14350
844	FORMAT(	14360
	*//1X,27(1H+)," END OF FITTING NO.",I2,2X,26(1H+) // )	14370
1000	IF(ANS.EQ.3HYES.OR.ANS.EQ.2HNO) GO TO 1002	14380
	IF(ANS.EQ.4HBACK) GO TO 1004	14390
	WRITE(6,1020)	14400
1001	IF(SKIP) NS=NO	14410
	GO TO (5,10,15,20,25,30,35,40,45,50,55,70,85,90,95,100,105,	14420
	*130,160,170,175,185 )NO	14430
1002	GO TO (6,11,16,21,26,31,36,41,46,51,56,71,86,91,96,101,105,	14440
	*130,161,171,175,185 )NO	14450
1003	IF(ANS.EQ.4HBACK) GO TO 1004	14460
	GO TO 1002	14470
1004	NS=NO	14480
	WRITE(6,1021)	14490
	SKIPS=SKIP	14500
	CALL READ(1,AR)	14510
	IF(ANS.EQ.4HBACK) GO TO 1004	14520
	SKIP=SKIPS	14530
	NO=IFIX(ANS+.1)	14540
	IF(SKIP.AND.NO.GE.1.AND.NO.LE.22) GO TO 1001	14550
	DO 1005 I=1,NS	14560
	IF(I.EQ.NO) GO TO 1001	14570
1005	CONTINUE	14580
	WRITE(6,1022)	14590

814	FORMAT( I4,3X,E10.4,10X,5E10.4 )	13570
816	FORMAT( I4, 3X,7E10.4 )	13580
817	FORMAT( / OBS.NO.,7( " X",11,4X)/1X,77(1H-) )	13590
818	FORMAT( 1X,77(1H-) / )	13600
819	FORMAT(15,2X,7E10.4 )	13610
820	FORMAT( " ASSUMING A NORMAL DISTRIBUTION OF ERRORS,THE UNIT NORMAL "	13620
	*, " DEVIATE FORM " / OF THE RESIDUALS TOGETHER WITH THE T-DIS "	13630

H02

	*"TRIBUTION WITH",13," DEGREES OF FREEDOM" /" INDICATES THAT")	13640
822	FORMAT(20X," OBSERVATION NUMBER " ,I3 )	13650
824	FORMAT( / " IS AN OUTLIER (P.LT.0.05) " / )	13660
8240	FORMAT(/21X," RESPONSE SYSTEM NO.",I2/ )	13670
8241	FORMAT( / " OBS.NO. WEIGHT NORM.WEIGHT WEIGHTED RESIDUAL",	13680
	*" UNIT NORMAL DEVIATE"/47X," (CRIT.VALUES=+-",F4.2," ) /	13690
	*1X,67(1H-) )	13700
8242	FORMAT(I4,A2,2E12.4,2E15.4 )	13710
8243	FORMAT(I4,2X,2E12.4,2E15.4 )	13720
8244	FORMAT(1X,67(1H-) / )	13730
8245	FORMAT( " THE OBSERVATION MARKED * IN THE TABLE ABOVE"/	13740
	*" APPEARS TO BE AN OUTLIER. " / )	13750
8246	FORMAT( " THE OBSERVATIONS MARKED * IN THE TABLE ABOVE"/	13760
	*" APPEAR TO BE OUTLIERS. " / )	13770
826	FORMAT( / " ARE OUTLIERS (P.LT.0.05) " / )	13780
8261	FORMAT( " THE FITTED CURVE CAN BE PLOTTED FROM FOLLOWING 51 POINTS"	13790
	*//2( " PT.NO. " X",8X," RESPONSE" )/1X,58(1H-) )	13800
8262	FORMAT(2(I4,3X,2E11.4))	13810
8263	FORMAT(1X,58(1H-) )	13820
8264	FORMAT( /18X," RESPONSE SYSTEM NO.",I2/ )	13830
827	FORMAT( / " VARIANCE-COVARIANCE MATRIX OF THE PARAMETERS. " / )	13840
8270	FORMAT( / " CORRELATION MATRIX " / )	13850
829	FORMAT(1X,6E13.6 )	13860
830	FORMAT( / " DURBIN WATSON STATISTIC FOR SERIAL CORRELATION OF",	13870
	*" RESIDUALS =" ,E10.4/" (REF. J.DURBIN AND G.S.WATSON,"	13880
	*" BIOMETRICA,PP 157-178,1951) / )	13890
832	FORMAT( " THE PROBABILITY OF RANDOMNESS OF RUNS =" ,E12.4/	13900
	*" (REF. S.SWED AND C.EISENHART, ANN. OF MATH. STATS.,14,"	13910
	*" PP66-87,1943) " / )	13920
8320	FORMAT( " ASSUMING EQUAL PROBABILITY (0.5) OF GETTING NEGATIVE AND"	13930
	*" POSITIVE " /" RESIDUALS, THE PROBABILITY OF GETTING " ,I2,	13940
	*" OR LESS RESIDUALS OF SAME SIGN " /" OF A TOTAL OF",I3,	13950
	*" RESIDUALS IS =" ,E11.4 / )	13960
833	FORMAT( " THIS INDICA "S THAT THE ERRORS ARE NOT RANDOM AT THE",	13970
	*" MINIMUM FOUND " / )	13980
834	FORMAT( " THE RATIO OF ) E DEGREES OF FREEDOM OF RESIDUALS TO THE",	13990
	*" NUMBER " /" OF RESIDUALS =" ,E10.4," A SIGNIFICANT CORRELATION"	14000
	*" BETWEEN THE RESIDUALS " /" IS THEREFORE EXPECTED,VIOLATING THE",	14010
	*" ASSUMPTION BEHIND THE STATISTICAL " /" EVALUATION OF THE",	14020
	*" PARAMETERS. " )	14030
836	FORMAT(/8X,	14040
	*" PLOT OF RESIDUALS VERSUS INDEPENDENT VARIABLE X1 (SCL.UNITS)" )	14050
8360	FORMAT(/2X," PLOT OF WEIGHTED RESIDUALS VERSUS",	14060
	*" INDEPENDENT VARIABLE X1 (SCL.UNITS)" )	14070
838	FORMAT(/11X,	14080
	*" PLOT OF RESIDUALS VERSUS RESPONSE ESTIMATE (SCL.UNITS)" )	14090
8380	FORMAT(/5X," PLOT OF WEIGHTED RESIDUALS",	14100
	*" VERSUS RESPONSE ESTIMATE (SCL.UNITS)" )	14110
840	FORMAT(/5X," PLOT OF RESIDUALS VERSUS 1ST. INDEPENDENT",	14120
	*" VARIABLE " X1 (SCL.UNITS)" )	14130
8400	FORMAT(/2X," PLOT OF WEIGHTED RESIDUALS VERSUS",	14140
	*" 1ST. INDEPENDENT VARIABLE X1 (SCL.UNITS)" )	14150
842	FORMAT(/5X," PLOT OF RESIDUALS VERSUS 2ND. INDEPENDENT VARIABLE",	14160
	*" X2 (SCL.UNITS)" )	14170
8420	FORMAT(/2X," PLOT OF WEIGHTED RESIDUALS VERSUS",	14180
	*" 2ND. INDEPENDENT VARIABLE X2 (SCL.UNITS)" )	14190
8401	FORMAT( " CONVERGENCE CRITERION " ,E10.4 )	14200
8402	FORMAT( " EXPANSION CRITERION " ,E12.4 )	14210
8403	FORMAT( " MINIMUM OF FITTED QUADRATIC SURFACE IS " ,E15.8/	14220
	*" COMPARE WITH MINIMUM FOUND BY ITERATION " ,E15.8/	14230
	*" IF DIFFERENCE IS LARGE THE STATISTICAL EVALUATION OF THE",	14240
	*" PARAMETERS IS " /" INACCURATE AND A NEW RUN WITH A DIFFERENT",	14250

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	*" STOPPING CRITERION,EXPANSION" /" CRITERION OR DIFFERENT RANGE",	14260
	*" OF THE PARAMETERS IS RECOMMENDED. " )	14270
8404	FORMAT( /" TYPE BACK (8*) IF YOU WANT TO MAKE CORRECTIONS OR",	14280
	*" OK IF YOU WANT " /" TO CONTINUE BUT WITHOUT STATISTICAL",	14290
	*" EVALUATION OF THE PARAMETERS " / )	14300
8422	FORMAT( " NO. OF SS EVALUATIONS " ,I4,17X,	14310
	*" NO. OF CONSTR. VIOLATIONS " ,I4 )	14320
8424	FORMAT( " THE QUADRATIC SURFACE FITTING REQUIRED A FURTHER",I4,	14330
	*" SS EVALUATIONS " )	14340
8425	FORMAT(/1H )	14350
844	FORMAT(	14360
	*//1X,27(1H+)," END OF FITTING NO.",I2,2X,26(1H+) // )	14370
1000	IF(ANS.EQ.3HYES.OR.ANS.EQ.2HNO) GO TO 1002	14380
	IF(ANS.EQ.4HBACK) GO TO 1004	14390
	WRITE(6,1020)	14400
1001	IF(SKIP) NS=NO	14410
	GO TO (5,10,15,20,25,30,35,40,45,50,55,70,85,90,95,100,105,	14420
	*130,160,170,175,185 )NO	14430
1002	GO TO (6,11,16,21,26,31,36,41,46,51,56,71,86,91,96,101,105,	14440
	*130,161,171,175,185 )NO	14450
1003	IF(ANS.EQ.4HBACK) GO TO 1004	14460
	GO TO 1002	14470
1004	NS=NO	14480
	WRITE(6,1021)	14490
	SKIPS=SKIP	14500
	CALL READ(1,AR)	14510
	IF(ANS.EQ.4HBACK) GO TO 1004	14520
	SKIP=SKIPS	14530
	NO=IFIX(ANS+.1)	14540
	IF(SKIP.AND.NO.GE.1.AND.NO.LE.22) GO TO 1001	14550
	DO 1005 I=1,NS	14560
	IF(I.EQ.NO) GO TO 1001	14570
1005	CONTINUE	14580
	WRITE(6,1022)	14590

N2=11	22870
DO 142 I=1,10	22880
IF(A(J1).EQ.AL(I)) N1=I-1	22900
142 IF(A(J2).EQ.AL(I)) N2=I-1	22910
IF(N1.EQ.11.OR.N2.EQ.11.AND.A(J2).NE.BLANK.OR.	22920
*N1.EQ.0.AND.N2.EQ.0) RETURN	22930

H03

NS=NO	22940
NO=10*N1+N2	22950
IF(A(J2).EQ.BLANK) NO=N1	22960
IGO=2	22970
IF(SKIP.AND.NO.GE.1.AND.NO.LE.22) RETURN	22980
IF(NO.GE.1.AND.NO.LE.NS) RETURN	22990
NO=NS	23000
WRITE(6,230)	23010
GO TO 5	23020
145 AR(1)=ZHOK	23030
RETURN	23040
200 FORMAT(78A1)	23050
205 FORMAT(/" INPUT ERROR. UNRECOGNIZED CHARACTER ( ",A1," )"/	23060
*" REENTER LAST LINE IN CORRECT FORM."/)	23070
210 FORMAT(/" INPUT ERROR. TOO FEW NUMBERS ON LINE ABOVE.",I2,	23080
*" ARE EXPECTED."/) REENTER LAST LINE IN CORRECT FORM ACCORDING",	23090
*" TO REQUEST."/)	23100
215 FORMAT(/" INPUT ERROR. TOO MANY NUMBERS ON LINE ABOVE.",I2,	23110
*" EXPECTED."/) REENTER LAST LINE IN CORRECT FORM ACCORDING",	23120
*" TO REQUEST."/)	23130
220 FORMAT(/" INPUT ERROR. -WRONG EXPONENT-"/	23140
*" REENTER LAST LINE IN CORRECT FORM ACCORDING TO REQUEST."/)	23150
225 FORMAT(/" INPUT ERROR. YOU FORGOT TO ENTER DATA. -TRY AGAIN- / )	23160
230 FORMAT(/" INPUT ERROR, ILLIGAL TRANSFER COMMAND - TRY AGAIN -"/)	23170
END	23180
C *****	23190
SUBROUTINE PLACC(F1,F2,F,NP,NOP,A)	23200
DIMENSION A(650),F1(20),F2(20),F(20)	23210
DATA DOT,X,BLANK /1H-,1HX,1H /	23220
DO 20 J=1,NOP	23230
N1=NP*(J-1)+1	23240
N2=N1+NP-1	23250
IF(F1(J).EQ.F2(J)) GO TO 10	23260
DO 5 I=N1,N2	23270
5 A(I)=DOT	23280
NX=N1+IFIX((NP-1)*(F(J)-F1(J))/(F2(J)-F1(J))+.5)	23290
IF(NX.GT.N2.OR.NX.LT.N1) GO TO 20	23300
A(NX)=X	23310
GO TO 20	23320
10 CONTINUE	23330
DO 15 I=N1,N2	23340
15 A(I)=BLANK	23350
20 CONTINUE	23360
RETURN	23370
END	23380
C *****	23390
FUNCTION WEIGHT(Y,INHT,R)	23400
I=INHT-1	23410
GO TO (5,10,15)I	23420
5 IF(Y.LT.0..OR.Y.EQ.0..AND.R.LT.0.) GO TO 20	23430
WEIGHT=Y**R	23440
RETURN	23450
10 IF(R*Y.LT.1) GO TO 20	23460
WEIGHT=ALOG(R*Y)	23470
RETURN	23480
15 WEIGHT=EXP(R*Y)	23490
RETURN	23500
20 WEIGHT=-1.	23510
RETURN	23520
END	23530
C *****	23540
SUBROUTINE XYPLOT(F)	23550

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DIMENSION X(100),Y(100),WYN(100),LX(100),LY(100),LIM1(10),LIM2(10)	23560
*LXF(75),LYF(75),XF(75),YF(75),F(20),A(75),XC(9,100)	23570
COMMON /DATA/ XX,Y,WYN,NOBS	23580
COMMON /FUNNUM/ITHFUN	23590
COMMON /LXLY/LX,LY	23600
COMMON /B1/ X	23610
COMMON /B4/ NIND	23620
COMMON /B6/ NOP	23630
COMMON /B8/NFUNC	23640
COMMON /B9/LIM1,LIM2	23650
DATA BLANK,POINT1,POINT2,DOT,STAR /1H ,1H*,1H0,1H.,1HX/	23660
DO 45 I=1,NFUNC	23670
N1=LIM1(I)	23680
N2=N1+1	23690
N3=LIM2(I)	23700
NN=N3-1	23710
XMIN=X(N1)	23720
XMAX=X(N1)	23730
YMIN=Y(N1)	23740
YMAX=Y(N1)	23750
IF(I.EQ.1) WRITE(6,100)	23760
IF(I.GT.1) WRITE(6,101)II	23770
DO 5 I=N2,N3	23780
IF(X(I).GT.XMAX) XMAX=X(I)	23790
IF(X(I).LT.XMIN) XMIN=X(I)	23800
IF(Y(I).GT.YMAX) YMAX=Y(I)	23810
5 IF(Y(I).LT.YMIN) YMIN=Y(I)	23820
SPANX=XMAX-XMIN	23830
IF(SPANX.EQ.0.)RETURN	23840
DELX=SPANX/74.	23850
DO 10 I=1,75	23860
XF(I)=XMIN+FLOAT(I-1)*DELX	23870
DO 7 L=N1,NN	23880
7 IF(XF(I).GT.X(L).AND.XF(I).LE.X(L+1))	23890
*YF(I)=Y(L)+(Y(L+1)-Y(L))*(XF(I)-X(L))/(X(L+1)-X(L))	23900

FACTOR=FLOAT(NOBS-NDUMMY)/SUMM	4850
DO 1520 I=1,NOBS	4860
1520 WYN(I)=FACTOR*WY(I)	4870
GO TO 155	4880
1525 DO 1530 I=1,NOBS	4890
WY(I)=1.	4900
1530 WYN(I)=1.	4910
155 CONTINUE	4920
157 WRITE(6,662)	4930
CALL READ(1,AR)	4940
IF(IGO.EQ.2) GO TO 1001	4950

101

IF(ANS.EQ.4HBACK) GO TO 1004	4960
IF(ANS.EQ.3HYES) GO TO 300	4970
IF(ANS.EQ.2HNO) GO TO 159	4980
WRITE(6,1020)	4990
GO TO 157	5000
158 WRITE(6,664)	5010
CALL READ(1,AR)	5020
IF(IGO.EQ.2) GO TO 1001	5030
IF(ANS.EQ.4HBACK) GO TO 1004	5040
IF(ANS.EQ.2HOK) GO TO 159	5050
WRITE(6,1027)	5060
GO TO 158	5070
159 IF(NRUN.EQ.0.AND.MAX.NE.1) WRITE(6,670)	5080
GO TO 350	5090
160 NQ=19	5100
NEWNOP=NOP	5110
DO 1600 I=1,NOP	5120
1600 IF(F1(I).EQ.F2(I)) NEWNOP=NEWNOP-1	5130
IF(NEWNOP.EQ.0.OR.NOP.GT.7) GO TO 1666	5140
IF(MAX.EQ.1) GO TO 162	5150
WRITE(6,6700)NEWNOP	5160
CALL READ(1,AR)	5170
GO TO (1000,1001) IGO	5180
161 IF(ANS.EQ.2HNO) GO TO 1666	5190
162 IF(NEWNOP.LT.3) GO TO 165	5200
WRITE(6,6705) NRECOM(NEWNOP)	5210
163 CALL READ(1,AR)	5220
IF(IGO.EQ.2) GO TO 1001	5230
IF(ANS.EQ.4HBACK) GO TO 1004	5240
NPOINT=FIX(ANS+.1)	5250
IF(NPOINT*.1.NCRIT(NEWNOP).OR.NPOINT.GT.2187) GO TO 164	5260
GO TO 165	5270
164 WRITE(6,1024)	5280
GO TO 163	5290
165 WRITE(6,6710)NEWNOP	5300
IF(MAX.EQ.1) CALL LSQ(F, FUNC1)	5310
IF(MAX.EQ.1) FUNC=FUNC1	5320
CALL SEARCH(FNEW,STPNEW,SSMIN,ANS16,NPOINT,NEVAL,NEWNOP,STEP,NP)	5330
IF(NEWNOP.GT.2) WRITE(6,6715)NEVAL,NP	5340
IF(ANS16.EQ.3HYES) WRITE(6,6720)SSMIN	5350
IF(ANS16.EQ.2HNO) WRITE(6,6725)SSMIN	5360
CALL PLACE(F1,F2,FNEW,40,NPR,A)	5370
DO 1650 J=1,NPR	5380
IF(F1(J).EQ.F2(J)) GO TO 1650	5390
N1=40*(J-1)+1	5400
N2=N1+39	5410
WRITE(6,6730)J,FNEW(J),(A(I),I=N1,N2)	5420
1650 CONTINUE	5430
IF(SSMIN.GE.FUNC) GO TO 1653	5440
IF(MAX.EQ.1) GO TO 1652	5450
WRITE(6,6735) FUNC	5460
DO 1651 I=1,NOP	5470
F(I)=FNEW(I)	5480
FSAVE(I)=F(I)	5490
IF(F(I)+STPNEW(I).GT.F2(I)) STPNEW(I)=-STPNEW(I)	5500
1651 STEP(I)=STPNEW(I)	5510
GO TO 1664	5520
1652 WRITE(6,6731) FUNC	5530
GO TO 1655	5540
1653 IF(MAX.EQ.1) GO TO 1654	5550
WRITE(6,6740) FUNC	5560
CALL PLACE(F1,F2,FMIN,40,NPR,A)	5570

J01

GO TO 1656	5580
1654 WRITE(6,6741) FJNC	5590
1655 CALL PLACE(F1,F2,F,40,NPR,A)	5600
1656 CONTINUE	5610
DO 1657 J=1,NPR	5620
IF(F1(J).EQ.F2(J)) GO TO 1657	5630
N1=40*(J-1)+1	5640
N2=N1+39	5650
IF(MAX.EQ.1) WRITE(6,6730)J,F(J),(A(I),I=N1,N2)	5660
IF(MAX.NE.1) WRITE(6,6730)J,FMIN(J),(A(I),I=N1,N2)	5670
1657 CONTINUE	5680
WRITE(6,6745)	5690
1660 WRITE(6,6748)	5700
CALL READ(1,AR)	5710
IF(IGO.EQ.2) GO TO 1001	5720
IF(ANS.EQ.4HBACK) GO TO 1004	5730
IF(ANS.EQ.3HYES) GO TO 1661	5740
IF(ANS.EQ.2HNO.AND.MAX.EQ.1) GO TO 1663	5750
IF(ANS.EQ.2HNO) GO TO 1666	5760
WRITE(6,1020)	5770
GO TO 1660	5780
1661 ANS11=3HYES	5790
ANS12=3HYES	5800
N11=NOP	5810
N12=NOP	5820
DO 1662 I=1,NOP	5830
NF(I)=1	5840
NT(I)=1	5850
F(I)=FNEW(I)	5860
FS(I)=F(I)	5870

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* 1ST. INDEPENDENT VARIABLE X1 (SCL.UNITS) ) 14150
842 FORMAT(/5X, " PLOT OF RESIDUALS VERSUS 2ND. INDEPENDENT VARIABLE", 14160
* X2 (SCL.UNITS) " ) 14170
8420 FORMAT(/2X, " PLOT OF WEIGHTED RESIDUALS VERSUS", 14180
* 2ND. INDEPENDENT VARIABLE X2 (SCL.UNITS) " ) 14190
8401 FORMAT(" CONVERGENCE CRITERION ",E10.4 ) 14200
8402 FORMAT(" EXPANSION CRITERION ",E12.4 ) 14210
8403 FORMAT(" MINIMUM OF FITTED QUADRATIC SURFACE IS ",E15.8/ 14220
* COMPARE WITH MINIMUM FOUND BY ITERATION ",E15.8/ 14230
* IF DIFFERENCE IS LARGE THE STATISTICAL EVALUATION OF THE", 14240
* PARAMETERS IS /" INACCURATE AND A NEW RUN WITH A DIFFERENT", 14250

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* " STOPPING CRITERION,EXPANSION"/" CRITERION OR DIFFERENT RANGE", 14260
* " OF THE PARAMETERS IS RECOMMENDED." ) 14270
8404 FORMAT(/" TYPE BACK (B**) IF YOU WANT TO MAKE CORRECTIONS OR", 14280
* " OK IF YOU WANT"/" TO CONTINUE BUT WITHOUT STATISTICAL", 14290
* " EVALUATION OF THE PARAMETERS" // ) 14300
8422 FORMAT(" NO. OF SS EVALUATIONS ",I4,17X, 14310
* " NO. OF CONSTR. VIOLATIONS ",I4 ) 14320
8424 FORMAT(" THE QUADRATIC SURFACE FITTING REQUIRED A FURTHER",I4, 14330
* " SS EVALUATIONS" ) 14340
8425 FORMAT(/1H ) 14350
844 FORMAT( 14360
* /1X,27(1H*), " END OF FITTING NO.",I2,2X,26(1H*) // ) 14370
1000 IF(ANS.EQ.3HYES.OR.ANS.EQ.2HNO) GO TO 1002 14380
IF(ANS.EQ.4HBACK) GO TO 1004 14390
WRITE(6,1020) 14400
1001 IF(SKIP) NS=NO 14410
GO TO (5,10,15,20,25,30,35,40,45,50,55,70,85,90,95,100,105, 14420
*130,160,170,175,185 )NO 14430
1002 GO TO (6,11,16,21,26,31,36,41,46,51,56,71,86,91,96,101,105, 14440
*130,161,171,175,185 )NO 14450
1003 IF(ANS.EQ.4HBACK) GO TO 1004 14460
GO TO 1002 14470
1004 NS=NO 14480
WRITE(6,1021) 14490
SKIPS=SKIP 14500
CALL READ(1,AR) 14510
IF(ANS.EQ.4HBACK) GO TO 1004 14520
SKIP=SKIPS 14530
NQ=IFIX(ANS+.1) 14540
IF(SKIP.AND.NQ.GE.1.AND.NQ.LE.22) GO TO 1001 14550
DO 1005 I=1,NS 14560
IF(I.EQ.NO) GO TO 1001 14570
1005 CONTINUE 14580
WRITE(6,1022) 14590
NQ=NS 14600
GO TO 1004 14610
1006 NCCORR=NCCORR+1 14620
IF(NCCORR.EQ.1.AND.NS.NE.NQ.AND.NS.NE.22) WRITE(6,1023)NS 14630
NQ=NS 14640
GO TO 1001 14650
1007 WRITE(6,1024) 14660
GO TO 1001 14670
1008 WRITE(6,1025) NAGR 14680
GO TO 1001 14690
1009 WRITE(6,1026) 14700
GO TO 1001 14710
1010 WRITE(6,1028) 14720
GO TO 1004 14730
1011 WRITE(6,1029) 14740
GO TO 1004 14750
1012 WRITE(6,1034)I,Y(I),IWGT 14760
GO TO 1004 14770
1013 WRITE(6,1035) 14780
GO TO 1004 14790
1020 FORMAT(/," INPUT ERROR, YOUR ANSWER MUST BE ONE OF THE FOLLOWING"/ 14800
* " FOUR, Y N B** R** - TRY AGAIN -"/ ) 14810
1021 FORMAT(/" WHAT REQUEST NUMBER DO YOU WANT TO GO TO"// ) 14820
1035 FORMAT(" YOU FORGOT AN ESSENTIAL REQUEST (I.E. 8,9,10 OR 17)" ) 14830
1022 FORMAT(/," INPUT ERROR, THE ENTERED NUMBER IS NOT AMONG"/ 14840
* " THE REQUEST NUMBERS ABOVE. - TRY AGAIN -"/) 14850
1023 FORMAT(" CONTINUE NOW AFTER CORRECTION FROM REQUEST NO."I3//) 14860
1024 FORMAT(/," INPUT ERROR, THE ENTERED NUMBER IS NOT", 14870

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* " IN THE ALLOWED RANGE. - TRY AGAIN -"/ ) 14880
1025 FORMAT(/," INPUT ERROR, YOUR INPUT DOES NOT AGREE WITH YOUR"/ 14890
* " INPUT ABOVE UNDER REQUEST NUMBER",I3, " - TRY AGAIN -"/) 14900
1026 FORMAT(/," INPUT ERROR, THE INPUT IS NOT IN THE RIGHT ORDER. ", 14910
* " - TRY AGAIN -"/ ) 14920
1027 FORMAT(/," INPUT ERROR, YOUR ANSWER MUST BE OK B OR R", 14930
* " - TRY AGAIN -"/ ) 14940
1028 FORMAT(" INPUT ERROR, YOUR INPUT UNDER REQUEST NUMBER 6 DOES"/ 14950
* " NOT AGREE WITH THE INPUT UNDER REQUEST NUMBER 5") 14960
1029 FORMAT(" INPUT ERROR, YOUR INPUT UNDER REQUEST NUMBER 9 DOES"/ 14970
* " NOT AGREE WITH THE INPUT UNDER REQUEST NUMBER 8") 14980
1030 FORMAT(/" INPUT ERROR, THE INITIAL ESTIMATE OF PARAMETER NO.", 14990
* I2, " IS NOT WITHIN"/" ITS LIMITS. ENTER -BACK- IF YOU WANT TO", 15000
* " MAKE CORRECTION OR -OK- IF YOU"/" WANT THE PROGRAM TO CHOOSE", 15010
* " AN ACCEPTABLE INITIAL PARAMETER ESTIMATE" // ) 15020
1031 FORMAT(/" INPUT ERROR, THE STEP SIZE FOR PARAMETER NO.",I2, 15030
* " DOES NOT AGREE WITH"/" THE LIMITS FOR THIS PARAMETER. ENTER", 15040
* " -BACK- IF YOU WANT TO MAKE CORRECTION"/" OR -OK- IF YOU WANT", 15050
* " THE PROGRAM TO CHOOSE AN ACCEPTABLE STEP SIZE" // ) 15060
1032 FORMAT(" INPUT ERROR, YOU FORGOT TO ENTER OBSERVATION NUMBER",I3/ 15070
* " REENTER YOUR DATA STRICTLY ACCORDING TO FOLLOWING REQUEST"/) 15080
1033 FORMAT(" INPUT ERROR, THE WEIGHT FOR THE OBSERVATION ", 15090
* " IS NEGATIVE."/ " REENTER THE LAST LINE IN CORRECT FORM."//) 15100
1034 FORMAT(/" INPUT ERROR, THE WEIGHT FOR OBSERVATION NO.",I3, 15110
* " (Y=",E10.4,") "/" IS NOT DEFINED BY THE WEIGHTING SCHEME NO.", 15120
* I2 ) 15130
END 15140
***** 15150
C SUBROUTINE NELDRC(F,STEP,NOP,FUNC,MAX,IPRINT,STOPCR,NLOOP,IQUAD, 15160
* SIMP,VAR,FUNCTN,IFALT) 15170

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10	IF(R*Y.LT.1) GO TO 20	23460
	WEIGHT=ALOG(R*Y)	23470
	RETURN	23480
15	WEIGHT=EXP(R*Y)	23490
	RETURN	23500
20	WEIGHT=-1.	23510
	RETURN	23520
	END	23530
C	*****	23540
	SUBROUTINE XYPLOT(F)	23550

103

	DIMENSION X(100),Y(100),WYN(100),LX(100),LY(100),LIM1(10),LIM2(10)	23560
	*LXF(75),LYF(75),XF(75),YF(75),F(20),A(75),X(9,100)	23570
	COMMON /DATA/ XX,Y,WYN,NOBS	23580
	COMMON /FUNNUM/ ITHFUN	23590
	COMMON /LXLY/ LX,LY	23600
	COMMON /B1/ X	23610
	COMMON /B4/ NIND	23620
	COMMON /B6/ NOP	23630
	COMMON /B8/ NFUNC	23640
	COMMON /B9/ LIM1,LIM2	23650
	DATA BLANK,POINT1,POINT2,DOT,STAR /1H ,1H*,1H0,1H.,1HX/	23660
	DO 45 I=1,NFUNC	23670
	N1=LIM1(I)	23680
	N2=N1+1	23690
	N3=LIM2(I)	23700
	NN=N3-1	23710
	XMIN=X(N1)	23720
	XMAX=X(N1)	23730
	YMIN=Y(N1)	23740
	YMAX=Y(N1)	23750
	IF(I.EQ.1) WRITE(6,100)	23760
	IF(I.GT.1) WRITE(6,101)I	23770
	DO 5 I=N2,N3	23780
	IF(X(I).GT.XMAX) XMAX=X(I)	23790
	IF(X(I).LT.XMIN) XMIN=X(I)	23800
	IF(Y(I).GT.YMAX) YMAX=Y(I)	23810
	IF(Y(I).LT.YMIN) YMIN=Y(I)	23820
5	SPANX=XMAX-XMIN	23830
	IF(SPANX.EQ.0.)RETURN	23840
	DELX=SPANX/74.	23850
	DO 10 I=1,75	23860
	XF(I)=XMIN+FLOAT(I-1)*DELX	23870
	DO 7 L=N1,NN	23880
7	IF(XF(I).GT.X(L).AND.XF(I).LE.X(L+1))	23890
	*YF(I)=Y(L)*(XF(I)-X(L))/(X(L+1)-X(L))	23900
	ITHFUN=I	23910
	CALL MODEL(YF(I),XF(I),F,0)	23920
	IF(YF(I).GT.YMAX) YMAX=YF(I)	23930
10	IF(YF(I).LT.YMIN) YMIN=YF(I)	23940
	SPANY=YMAX-YMIN	23950
	IF(SPANY.EQ.0.)RETURN	23960
	DO 15 I=N1,N3	23970
	IF(WYN(I).EQ.0.) GO TO 12	23980
	LY(I)=FIX(49.*(YMAX-Y(I))/SPANY+.5) +1	23990
	LX(I)=FIX(74.*(X(I)-XMIN)/SPANX+.5) +1	24000
	GO TO 15	24010
12	LY(I)=0	24020
	LX(I)=0	24030
15	CONTINUE	24040
	DO 20 I=1,75	24050
	LYF(I)=FIX(49.*(YMAX-YF(I))/SPANY+.5) +1	24060
20	LXF(I)=FIX(74.*(XF(I)-XMIN)/SPANX+.5) +1	24070
	NZERO=FIX(49.*(YMAX/SPANY+.5) +1	24080
	IF(NZERO.EQ.50.AND.YMIN.GT.0.)NZERO=51	24090
	IF(NZERO.EQ.1.AND.YMAX.LT.0.) NZERO=0	24100
	DO 40 I=1,50	24110
	DO 35 J=1,75	24120
	A(J)=BLANK	24130
	IF(I.EQ.NZERO)A(J)=DOT	24140
	N=0	24150
	DO 25 K=1,75	24160
25	IF(LXF(K).EQ.J.AND.LYF(K).EQ.I) A(J)=POINT1	24170

J03

	IF(A(J).EQ.POINT1)N=1	24180
	DO 30 L=N1,N3	24190
30	IF(LX(L).EQ.J.AND.LY(L).EQ.I) A(J)=POINT2	24200
35	IF(N.EQ.1.AND.A(J).EQ.POINT2)A(J)=STAR	24210
40	WRITE(6,105)A	24220
	WRITE(6,110)	24230
45	CONTINUE	24240
	WRITE(6,115)	24250
100	FORMAT(/15X," PLOT OF FITTED CURVE AND DATA POINTS (SCL.UNITS)"/	24260
	*1X,77(1H-))	24270
101	FORMAT(/26X," RESPONSE SYSTEM NO.",I2//1X,77(1H-))	24280
105	FORMAT(2H I,75A1,1H I)	24290
110	FORMAT(1X,77(1H-)/" 0 =OBSERVED POINTS * =CALCULATED POINTS",	24300
	*" X =CALCULATED AND OBSERVED POINTS"/)	24310
115	FORMAT(" THE ACCURACY OF THE ABOVE PLOT(S) IS +-1/2 CHARACTER",	24320
	*" AND LINE DISTANCE."/)	24330
	RETURN	24340
	END	24350
C	*****	24360
	SUBROUTINE PLOT(X,Y,NPOINT,LINES)	24370
	DIMENSION X(100),Y(0),A(75),LX(100),LY(100),LIM1(10),LIM2(10)	24380
	COMMON /LXLY/ LX,LY	24390
	COMMON /B7/ ITABLE	24400
	COMMON /B8/ NFUNC	24410
	COMMON /B9/ LIM1,LIM2	24420
	COMMON /B13/ IGNORE	24430
	DATA BLANK,POINT,DOT/1H ,1H0,1H./	24440
	N1=1	24450
	N2=2	24460
	N3=NPOINT	24470
	NZ=1	24480



F(I)=FNEW(I)	5480
FSAVE(I)=F(I)	5490
IF(F(I)+STPNEW(I).GT.F2(I)) STPNEW(I)=-STPNEW(I)	5500
1651 STEP(I)=STPNEW(I)	5510
GO TO 1664	5520
1652 WRITE(6,6731) FUNC	5530
GO TO 1655	5540
1653 IF(MAX.EQ.1) GO TO 1654	5550
WRITE(6,6740) FUNC	5560
CALL PLACE(F1,F2,FMIN,40,NPR,A)	5570

J01

GO TO 1656	5580
1654 WRITE(6,6741) FJNC	5590
1655 CALL PLACE(F1,F2,F,40,NPR,A)	5600
1656 CONTINUE	5610
DO 1657 J=1,NPR	5620
IF(F1(J).EQ.F2(J)) GO TO 1657	5630
N1=40+(J-1)+1	5640
N2=N1+39	5650
IF(MAX.EQ.1) WRITE(6,6730) J,F(J),(A(I),I=N1,N2)	5660
IF(MAX.NE.1) WRITE(6,6730) J,FMIN(J),(A(I),I=N1,N2)	5670
1657 CONTINUE	5680
WRITE(6,6745)	5690
1660 WRITE(6,6748)	5700
CALL READ(1,AR)	5710
IF(IGO.EQ.2) GO TO 1001	5720
IF(ANS.EQ.4HBACK) GO TO 1004	5730
IF(ANS.EQ.3HYES) GO TO 1661	5740
IF(ANS.EQ.2HNO.AND.MAX.EQ.1) GO TO 1663	5750
.F(ANS.EQ.2HNO) GO TO 1666	5760
WRITE(6,1020)	5770
GO TO 1660	5780
1661 ANS11=3HYES	5790
ANS12=3HYES	5800
N11=NOP	5810
N12=NOP	5820
DO 1662 I=1,NOP	5830
NF(I)=I	5840
NT(I)=I	5850
F(I)=FNEW(I)	5860
FS(I)=F(I)	5870
FSAVE(I)=F(I)	5880
IF(F(I)+STPNEW(I).GT.F2(I)) STPNEW(I)=-STPNEW(I)	5890
STEP(I)=STPNEW(I)	5900
IF(F1(I).EQ.F2(I)) STEP(I)=0.	5910
1662 STPS(I)=STEP(I)	5920
IF(MAX.NE.1) GO TO 1664	5930
1663 NS=18	5940
GO TO 25	5950
1664 WRITE(6,6747)	5960
CALL READ(1,AR)	5970
IF(IGO.EQ.2) GO TO 1001	5980
IF(ANS.EQ.4HBACK) GO TO 1004	5990
IF(ANS.EQ.2HNO) GO TO 350	6000
IF(ANS.NE.3HYES) GO TO 1665	6010
NQ=22	6020
GO TO 1004	6030
1665 WRITE(6,1020)	6040
GO TO 1664	6050
1666 IF(NS.GT.NQ) GO TO 1006	6060
170 NQ=20	6070
WRITE(6,675)	6080
CALL READ(1,AR)	6090
IF(IGO.EQ.2) GO TO 1001	6100
IF(ANS.EQ.4HBACK) GO TO 1004	6110
IF(ANS.EQ.2HNO) GO TO 171	6120
IF(ANS.EQ.3HYES) GO TO 173	6130
WRITE(6,1020)	6140
GO TO 170	6150
171 WRITE(6,680)	6160
CALL READ(1,AR)	6170
IF(IGO.EQ.2) GO TO 1001	6180
IF(ANS.EQ.4HBACK) GO TO 1004	6190

K01

IF(ANS.EQ.3HYES) NS=8	6200
IF(ANS.EQ.3HYES) GO TO 9	6210
IF(ANS.EQ.2HNO) GO TO 172	6220
WRITE(6,1020)	6230
GO TO 171	6240
172 WRITE(6,685)	6250
STOP	6260
173 IF(NS.GT.NQ) GO TO 1006	6270
175 NQ=21	6280
WRITE(6,690)	6290
CALL READ(1,AR)	6300
IF(IGO.EQ.2) GO TO 1001	6310
IF(ANS.EQ.4HBACK) GO TO 1004	6320
IF(ANS.EQ.3HYES) GO TO 178	6330
IF(ANS.EQ.2HNO) GO TO 176	6340
WRITE(6,1020)	6350
GO TO 175	6360
176 CONTINUE	6370
ANS11=3HYES	6380
ANS12=2HNO	6390
N11=NOP	6400
DO 177 I=1,NOP	6410
NF(I)=I	6420
177 FS(I)=FSAVE(I)	6430
GO TO 179	6440
178 ANS12=2HNO	6450
ANS11=3HYES	6460
N11=NOP	6470
DO 1780 I=1,NOP	6480
NF(I)=I	6490
1780 FS(I)=F(I)	6500

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1015 WRITE(6,1033) 14780
GO TO 1004 14790
1020 FORMAT(/," INPUT ERROR, YOUR ANSWER MUST BE ONE OF THE FOLLOWING"/ 14800
* " FOUR, Y N B** R** - TRY AGAIN -"/ ) 14810
1021 FORMAT(/," WHAT REQUEST NUMBER DO YOU WANT TO GO TO"/ ) 14820
1035 FORMAT(/," YOU FORGOT AN ESSENTIAL REQUEST (I.E. 8,9,10 OR 17) " ) 14830
1022 FORMAT(/," INPUT ERROR, THE ENTERED NUMBER IS NOT AMONG"/ 14840
* " THE REQUEST NUMBERS ABOVE. - TRY AGAIN -"/ ) 14850
1023 FORMAT(/," CONTINUE NOW AFTER CORRECTION FROM REQUEST NO."13/) 14860
1024 FORMAT(/," INPUT ERROR, THE ENTERED NUMBER IS NOT", 14870

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J02

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* " IN THE ALLOWED RANGE. - TRY AGAIN -"/ ) 14880
1025 FORMAT(/," INPUT ERROR, YOUR INPUT DOES NOT AGREE WITH YOUR"/ 14890
* " INPUT ABOVE UNDER REQUEST NUMBER 13," - TRY AGAIN -"/ ) 14900
1026 FORMAT(/," INPUT ERROR, THE INPUT IS NOT IN THE RIGHT ORDER. " , 14910
* " - TRY AGAIN -"/ ) 14920
1027 FORMAT(/," INPUT ERROR, YOUR ANSWER MUST BE OK B OR R" , 14930
* " - TRY AGAIN -"/ ) 14940
1028 FORMAT(/," INPUT ERROR, YOUR INPUT UNDER REQUEST NUMBER 6 DOES"/ 14950
* " NOT AGREE WITH THE INPUT UNDER REQUEST NUMBER 5") 14960
1029 FORMAT(/," INPUT ERROR, YOUR INPUT UNDER REQUEST NUMBER 9 DOES"/ 14970
* " NOT AGREE WITH THE INPUT UNDER REQUEST NUMBER 8") 14980
1030 FORMAT(/," INPUT ERROR, THE INITIAL ESTIMATE OF PARAMETER NO." , 14990
* " 12 " IS NOT WITHIN"/ " ITS LIMITS. ENTER -BACK- IF YOU WANT TO" , 15000
* " MAKE CORRECTION OR -OK- IF YOU"/ " WANT THE PROGRAM TO CHOOSE" , 15010
* " AN ACCEPTABLE INITIAL PARAMETER ESTIMATE"/ " ) 15020
1031 FORMAT(/," INPUT ERROR, THE STEP SIZE FOR PARAMETER NO." ,12, 15030
* " DOES NOT AGREE WITH"/ " THE LIMITS FOR THIS PARAMETER. ENTER" , 15040
* " -BACK- IF YOU WANT TO MAKE CORRECTION"/ " OR -OK- IF YOU WANT" , 15050
* " THE PROGRAM TO CHOOSE AN ACCEPTABLE STEP SIZE"/ " ) 15060
1032 FORMAT(/," INPUT ERROR, YOU FORGOT TO ENTER OBSERVATION NUMBER" ,13/ 15070
* " REENTER YOUR DATA STRICTLY ACCORDING TO FOLLOWING REQUEST"/ ) 15080
1033 FORMAT(/," INPUT ERROR, THE WEIGHT FOR THE OBSERVATION " ,13/ ) 15090
* " IS NEGATIVE."/ " REENTER THE LAST LINE IN CORRECT FORM."/ ) 15100
1034 FORMAT(/," INPUT ERROR, THE WEIGHT FOR OBSERVATION NO." ,13, 15110
* " (Y=" ,E10.4,") "/ " IS NOT DEFINED BY THE WEIGHTING SCHEME NO." , 15120
* " 12 ) 15130
END 15140
C 15150
***** 15150
SUBROUTINE WELDR(F,STEP,NOP,FUNCTN,MAX,IPRINT,STOPCR,NLOOP,IQUAD, 15160
* SIMP,VAR,FUNCTN,IFAUULT) 15170
DOUBLE PRECISION DMEAN,DMEANS,DHCV,FUNCT,FUNCS,H,HMIN,HMAX, 15180
* HSTAR,HSTST,YMIN,A0,AVAL,BMAT,VC,TEMP,OMIN,PBAR 15190
DIMENSION F(20),STEP(20),AR(12),F1(20),F2(20) 15200
DIMENSION G(11,20),H(11),PBAR(20),PSTAR(20),PSTST(20),VARCOV(55) 15210
DIMENSION AVAL(20),BMAT(55),PMIN(20),VC(55),VAR(10),TEMP(20) 15220
EQUIVALENCE(PMIN,PSTAR),(AVAL,PBAR) 15230
COMMON /PARLM/F1,F2 15240
COMMON /CONSTR/LIMITS 15250
COMMON/B3/VARCOV,VC,NEVAL1,NEVAL2,AMINO 15260
COMMON /B4/ NIND 15270
COMMON /B5/NO,NS,IGO 15280
COMMON /B11/AR 15290
COMMON /B12/NPR 15300
COMMON /B14/NVIOI 15310
COMMON /B15/ SKIP 15320
LOGICAL SKIP 15330
A=1 15340
B=0.5 15350
C=2.5 15360
LIMITS=1 15370
CALL FUNCTN(F,FUNCT) 15380
NEVAL=1 15390
IF(FUNCT.GT.1050) WRITE(6,1000) 15400
1000 FORMAT(/," ONE OR MORE OF THE INITIAL PARAMETERS DOES NOT LIE"/ 15410
* " WITHIN THE SPECIFIED CONSTRAINED PARAMETER SPACE."/ ) 15420
IW=6 15430
IF(IPRINT)5,5,55 15440
55 WRITE(IW,100)IPRINT 15450
100 FORMAT(/," PROGRESS REPORT EVERY" ,14, " FUNCTION EVALUATIONS."/ 15460
* " EVAL.NO. FUNC.VALUE PARAMETERS"/ ) 15470
5 IFAULT = 0 15480
IF(NOP.LE.0) IFAULT=3 15490

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K02

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IF(NLOOP.LE.0) IFAULT=4 15500
IF (IFAUULT.NE.0) RETURN 15510
NAP=0 15520
LOOP=0 15530
IFLAG=0 15540
DO 1 I=1,NOP 15550
IF(STEP(I).NE.0.0) NAP=NAP+1 15560
1 CONTINUE 15570
IF(NAP.EQ.0)RETURN 15580
MORE=0.20*((FLOAT(NAP+1))*2.5) 15590
MORE=MIN0(50,MORE) 15600
MORE=MAX0(20,MORE) 15610
DO 6 I=1,NOP 15620
6 G(1,I)=F(I) 15630
IROW=2 15640
DO 7 I=1,NOP 15650
IF(STEP(I).EQ.0.0) GO TO 7 15660
DO 9 J = 1,NOP 15670
9 G(IROW,J)=F(J) 15680
G(IROW,I)=G(IROW,I)+STEP(I) 15690
IROW=IROW+1 15700
7 CONTINUE 15710
NP1=NAP+1 15720
NEVAL=0 15730
DO 10 I=1,NP1 15740
DO 11 J=1,NOP 15750
11 F(J)=G(I,J) 15760
CALL FUNCTN(F,H(I)) 15770
NEVAL=NEVAL+1 15780
IF(IPRINT)10,10,12 15790
12 IF(NPR.LE.5)WRITE(IW,1010)NEVAL,H(I),(F(J),J=1,NPR) 15800

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IF(NZERO.EQ.50.AND.YMIN.GT.0.)NZERO=51      24090
IF(NZERO.EQ.1.AND.YMAX.LT.0.)NZERO=0        24100
DO 40 I=1,50                                24110
DO 35 J=1,75                                24120
A(J)=BLANK                                  24130
IF(I.EQ.NZERO)A(J)=DOT                      24140
N=0                                          24150
DO 25 K=1,75                                24160
25 IF(LXF(K).EQ.J.AND.LYF(K).EQ.I) A(J)=POINT1 24170

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J03

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IF(A(J).EQ.POINT1)N=1                       24180
DO 30 L=N1,N3                               24190
30 IF(LX(L).EQ.J.AND.LY(L).EQ.I) A(J)=POINT2 24200
35 IF(N.EQ.1.AND.A(J).EQ.POINT2)A(J)=STAR   24210
40 WRITE(6,105)A                             24220
WRITE(6,110)                                 24230
45 CONTINUE                                  24240
WRITE(6,115)                                 24250
100 FORMAT(/15X," PLOT OF FITTED CURVE AND DATA POINTS (SCL.UNITS)"/ 24260
*1X,77(1H-))                                24270
101 FORMAT(/26X," RESPONSE SYSTEM NO.",I2//1X,77(1H-)) 24280
105 FORMAT(2H I,75A1,1H1)                   24290
110 FORMAT(1X,77(1H-)/" 0 =OBSERVED POINTS * =CALCULATED POINTS", 24300
*" X =CALCULATED AND OBSERVED POINTS"/)     24310
115 FORMAT(" THE ACCURACY OF THE ABOVE PLOT(S) IS +/-1/2 CHARACTER", 24320
*" AND LINE DISTANCE."/)                   24330
RETURN                                       24340
END                                          24350
C *****                                     24360
SUBROUTINE PLOT(X,Y,NPOINT,LINES)           24370
DIMENSION X(100),Y(100),A(75),LX(100),LY(100),LIM1(10),LIM2(10) 24380
COMMON /LXLY/LX,LY                          24390
COMMON /B7/ ITABLE                          24400
COMMON /B8/NFUNC                             24410
COMMON /B9/LIM1,LIM2                        24420
COMMON /B13/IGNORE                           24430
DATA 'BLANK,POINT,DOT/1H ,1H0,1H./        24440
N1=1                                         24450
N2=2                                         24460
N3=NPOINT                                    24470
N4=1                                         24480
IF(ITABLE.EQ.0) N4=NFUNC                    24490
DO 25 I=1,N4                                24500
IF(ITABLE.EQ.1) GO TO 1                     24510
IF(I.GT.1) WRITE(6,125)I                   24520
N1=LIM1(I)                                  24530
N2=N1+1                                     24540
N3=LIM2(I)                                  24550
1 XMIN=X(N1)                                24560
XMAX=X(N1)                                  24570
YMIN=Y(N1)                                  24580
YMAX=Y(N1)                                  24590
IF(IGNORE.EQ.0) GO TO 3                    24600
DO 2 I=N2,N3                                24610
IF(Y(I).EQ.0.) GO TO 2                    24620
XMIN=X(I)                                  24630
XMAX=X(I)                                  24640
YMIN=Y(I)                                  24650
YMAX=Y(I)                                  24660
2 CONTINUE                                  24670
3 CONTINUE                                  24680
DO 5 I=N2,N3                                24690
IF(Y(I).EQ.0..AND.IGNORE.NE.0) GO TO 5    24700
IF(X(I).GT.XMAX) XMAX=X(I)                 24710
IF(X(I).LT.XMIN) XMIN=X(I)                 24720
IF(Y(I).GT.YMAX) YMAX=Y(I)                 24730
IF(Y(I).LT.YMIN) YMIN=Y(I)                 24740
5 CONTINUE                                  24750
SPANX=XMAX-XMIN                            24760
SPANY=YMAX-YMIN                            24770
IF(SPANX.EQ.0..OR.SPANY.EQ.0.)RETURN       24780
B=FLOAT(LINES-1)                           24790

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K03

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DO 10 I=N1,N3                               24800
IF(Y(I).EQ.0..AND.IGNORE.NE.0) GO TO 9    24810
LY(I)=FIX(B*(YMAX-Y(I))/SPANY+.5)+1       24820
LX(I)=FIX(74.*(X(I)-XMIN)/SPANX+.5)+1     24830
GO TO 10                                    24840
9 LY(I)=0                                   24850
LX(I)=0                                     24860
10 CONTINUE                                  24870
NZERO=FIX(B*YMAX/SPANY+.5)+1              24880
IF(NZERO.EQ.LINES.AND.YMIN.GT.0.)NZERO=LINES+1 24890
IF(NZERO.EQ.1.AND.YMAX.LT.0.)NZERO=0      24900
WRITE(6,110)                                24910
DO 20 I=1,LINES                             24920
DO 15 J=1,75                                24930
A(J)=BLANK                                  24940
IF(I.EQ.NZERO)A(J)=DOT                      24950
DO 15 K=N1,N3                               24960
15 IF(LX(K).EQ.J.AND.LY(K).EQ.I) A(J)=POINT 24970
20 WRITE(6,105)A                             24980
WRITE(6,100)                                 24990
WRITE(6,115)YMAX,YMIN,XMAX,XMIN            25000
100 FORMAT(1H ,77(1H-))                     25010
105 FORMAT(2H I,75A1,1H1)                   25020
110 FORMAT(/1X,77(1H-))                     25030
115 FORMAT(" YMAX=",E11.4," YMIN=",E11.4," XMAX=",E11.4, 25040
*" XMIN=",E11.4)                             25050
IF(ITABLE.EQ.0) GO TO 25                    25060
WRITE(6,120) NPOINT,(X(I),Y(I),I=N1,N3)    25070
120 FORMAT(" THE COORDINATES OF THE ",I3," POINTS ARE"/ 25080
*(3(" ",E9.3," ",E9.3," ),2X))            25090
125 FORMAT(/ 28X," RESPONSE SYSTEM NO.",I2) 25100
CONTINUE                                    25110

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1665	WRITE(6,1023)	6020
	GO TO 1664	6030
1666	IF(NS.GT.NQ) GO TO 1006	6040
170	NO=20	6050
	WRITE(6,675)	6060
	CALL READ(1,AR)	6070
	IF(IGO.EQ.2) GO TO 1001	6080
	IF(ANS.EQ.4HBACK) GO TO 1004	6090
	IF(ANS.EQ.2HNO) GO TO 171	6100
	IF(ANS.EQ.3HYES) GO TO 173	6110
	WRITE(6,1020)	6120
	GO TO 170	6130
171	WRITE(6,680)	6140
	CALL READ(1,AR)	6150
	IF(IGO.EQ.2) GO TO 1001	6160
	IF(ANS.EQ.4HBACK) GO TO 1004	6170
		6180
		6190

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	IF(ANS.EQ.3HYES) NS=8	6200
	IF(ANS.EQ.3HYES) GO TO 9	6210
	IF(ANS.EQ.2HNO) GO TO 172	6220
	WRITE(6,1020)	6230
	GO TO 171	6240
172	WRITE(6,685)	6250
	STOP	6260
173	IF(NS.GT.NQ) GO TO 1006	6270
175	NO=21	6280
	WRITE(6,690)	6290
	CALL READ(1,AR)	6300
	IF(IGO.EQ.2) GO TO 1001	6310
	IF(ANS.EQ.4HBACK) GO TO 1004	6320
	IF(ANS.EQ.3HYES) GO TO 178	6330
	IF(ANS.EQ.2HNO) GO TO 176	6340
	WRITE(6,1020)	6350
	GO TO 175	6360
176	CONTINUE	6370
	ANS11=3HYES	6380
	ANS12=2HNO	6390
	N11=NOP	6400
	DO 177 I=1,NOP	6410
	NF(I)=I	6420
177	FS(I)=FSAVE(I)	6430
	GO TO 179	6440
178	ANS12=2HNO	6450
	ANS11=3HYES	6460
	N11=NOP	6470
	DO 1780 I=1,NOP	6480
	NF(I)=I	6490
1780	FS(I)=F(I)	6500
179	IF(NS.GT.NQ) GO TO 1006	6510
	WRITE(6,692)	6520
	NO=22	6530
	GO TO 1004	6540
185	NO=22	6550
	WRITE(6,694)	6560
	CALL READ(1,AR)	6570
	IF(IGO.EQ.2) GO TO 1001	6580
	IF(ANS.EQ.3HYES.OR.ANS.EQ.4HBACK) GO TO 1004	6590
	IF(ANS.EQ.2HNO) GO TO 186	6600
	WRITE(6,1020)	6610
	GO TO 185	6620
186	NO=18	6630
	GO TO 131	6640
300	WRITE(6,663)	6650
	CALL READ(1,AR)	6660
	IF(IGO.EQ.2) GO TO 1001	6670
	IF(ANS.EQ.4HBACK) GO TO 1004	6680
	IF(ANS.NE.3HYES.AND.ANS.NE.2HNO) GO TO 301	6690
	PLOTDP=ANS	6700
	GO TO 302	6710
301	WRITE(6,1020)	6720
	GO TO 300	6730
302	NCON=NOP-NEWNOP	6740
	WRITE(6,750)	6750
	WRITE(6,525)	6760
	WRITE(6,752) NEWNOP,NCON,NOBS,NDUMMY,NFUNC,NIND	6770
	DO 305 I=1,NOP	6780
305	WRITE(6,754)I,F1(I),F2(I),F(I),STEP(I)	6790
	WRITE(6,756)	6800
	NSAVE=1	6810

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	DO 310 I=1,NOBS	6820
	N=LABEL(I)	6830
	IF(N.EQ.NSAVE+1) WRITE(6,813)N	6840
	NSAVE=N	6850
	IF(NIND.GT.1) GO TO 308	6860
	IF(ANS16.EQ.3HYES) GO TO 307	6870
	WRITE(6,758)I,X(I),Y(I)	6880
	GO TO 310	6890
307	WRITE(6,760)I,X(I),Y(I),WY(I),WYN(I)	6900
	GO TO 310	6910
308	IF(ANS16.EQ.3HYES) GO TO 309	6920
	WRITE(6,762)I,X(I),Z(I),Y(I)	6930
	GO TO 310	6940
309	WRITE(6,764)I,X(I),Z(I),Y(I),WY(I),WYN(I)	6950
310	CONTINUE	6960
	WRITE(6,818)	6970
	IF(NIND.LT.3) GO TO 312	6980
	WRITE(6,817)(I,I=3,9)	6990
	DO 311 I=1,NOBS	7000
311	WRITE(6,819)I,(XX(J,I),J=3,NIND)	7010
	WRITE(6,818)	7020
312	IF(PLOTDP.EQ.2HNO) GO TO 313	7030
	WRITE(6,765)	7040
	CALL PRINT(V,WRITE(6,818))	7050

COMMON /B12/NPR	15300
COMMON /B14/NVIOL	15310
COMMON /B15/ SKIP	15320
LOGICAL SKIP	15330
A=1.	15340
B=0.5	15350
C=2.5	15360
LIMITS=1	15370
CALL FUNCTN(F, FUNC)	15380
NEVAL=1	15390
IF(FUNC.GT.1D50) WRITE(6,1000)	15400
1000 FORMAT(/ " ONE OR MORE OF THE INITIAL PARAMETERS DOES NOT LIE"/	15410
* " WITHIN THE SPECIFIED CONSTRAINED PARAMETER SPACE." / )	15420
IW=6	15430
IF(IPRINT)5,5,55	15440
55 WRITE(IW,100)IPRINT	15450
100 FORMAT(" PROGRESS REPORT EVERY",I4," FUNCTION EVALUATIONS."//	15460
* " EVAL_NO. FUNC.VALUE PARAMETERS" /)	15470
5 IFAULT = 0	15480
IF(NOP.LE.0) IFAULT=3	15490

K02

IF(NLOOP.LE.0) IFAULT=4	15500
IF (IFAILT.NE.0) RETURN	15510
NAP=0	15520
LOOP=0	15530
IFLAG=0	15540
DO 1 I=1,NOP	15550
IF(STEP(I).NE.0.0) NAP=NAP+1	15560
1 CONTINUE	15570
IF(NAP.EQ.0)RETURN	15580
MORE=0.20*((FLOAT(NAP+1))*2.5)	15590
MORE=MINO( 50,MORE)	15600
MORE=MAXO(20,MORE)	15610
DO 6 I=1,NOP	15620
6 G(1,I)=F(I)	15630
IROW=2	15640
DO 7 I=1,NOP	15650
IF(STEP(I).EQ.0.) GO TO 7	15660
DO 9 J = 1,NOP	15670
9 G(IROW,J)=F(J)	15680
G(IROW,I)=G(IROW,I)+STEP(I)	15690
IROW=IROW+1	15700
7 CONTINUE	15710
NP1=NAP+1	15720
NEVAL=0	15730
DO 10 I=1,NP1	15740
DO 11 J=1,NOP	15750
11 F(J)=G(I,J)	15760
CALL FUNCTN(F,H(I))	15770
NEVAL=NEVAL+1	15780
IF(IPRINT)10,10,12	15790
12 IF(NPR.LE.5)WRITE(IW,1010)NEVAL,H(I),(F(J),J=1,NPR)	15800
1010 FORMAT(1X,I4,D16.8,1X,5E11.5)	15810
IF(NPR.GT.5) WRITE(IW,101)NEVAL,H(I),(F(J),J=1,NPR)	15820
101 FORMAT(1X,I4,D16.8,1X,5E11.5/(22X,5E11.5))	15830
10 CONTINUE	15840
45 LOOP=LOOP+1	15850
IMAX=1	15860
IMIN=1	15870
DO 13 I=2,NP1	15880
IF(H(I)-H(IMAX))15,15,14	15890
14 IMAX=I	15900
15 IF(H(I)-H(IMIN))16,13,13	15910
16 IMIN = I	15920
13 CONTINUE	15930
HMAX = H(IMAX)	15940
HMIN = H(IMIN)	15950
DO 17 I=1,NOP	15960
17 PBAR(I)=0.0	15970
DO 18 I=1,NP1	15980
IF(I-IMAX)19,18,19	15990
19 DO 20 J=1,NOP	16000
20 PBAR(J) = PBAR(J)+G(I,J)	16010
18 CONTINUE	16020
DO 602 J=1,NOP	16030
602 PBAR(J) = PBAR(J)/NAP	16040
DO 21 I=1,NOP	16050
21 PSTAR(I)=A*(PBAR(I)-G(IMAX,I))+PBAR(I)	16060
CALL FUNCTN (PSTAR,HSTAR)	16070
NEVAL=NEVAL+1	16080
IF(IPRINT)57,57,56	16090
56 IF(NPR.LE.5.AND.MOD(NEVAL,IPRINT).EQ.0) WRITE(IW,1010)	16100
*NEVAL,HSTAR,(PSTAR(J),J=1,NPR)	16110

L02

IF(NPR.GT.5.AND.MOD(NEVAL,IPRINT).EQ.0) WRITE(IW,101)	16120
*NEVAL,HSTAR,(PSTAR(J),J=1,NPR)	16130
57 IF(HSTAR-HMIN)22,23,23	16140
22 DO 24 I=1,NOP	16150
24 PSTST(I)=C*(PSTAR(I)-PBAR(I))+PBAR(I)	16160
CALL FUNCTN (PSTST,HSTST)	16170
NEVAL=NEVAL+1	16180
IF(IPRINT)60,60,59	16190
59 IF(NPR.LE.5.AND.MOD(NEVAL,IPRINT).EQ.0) WRITE(IW,1010)	16200
*NEVAL,HSTST,(PSTST(J),J=1,NPR)	16210
IF(NPR.GT.5.AND.MOD(NEVAL,IPRINT).EQ.0) WRITE(IW,101)	16220
*NEVAL,HSTST,(PSTST(J),J=1,NPR)	16230
60 IF(HSTST-HMIN)25,26,26	16240
25 DO 27 I=1,NOP	16250
IF(STEP(I).NE.0.0) G(IMAX,I)=PSTST(I)	16260
27 CONTINUE	16270
H(IMAX)=HSTST	16280
GO TO 41	16290
23 DO 28 I=1,NP1	16300
IF(I-IMAX)29,28,29	16310
29 IF(HSTAR-H(I))26,28,28	16320

```

IF(LINE.EQ.0) GO TO 5
DO 2 I=N2,N3
IF(Y(I).EQ.0.) GO TO 2
XMIN=X(I)
XMAX=X(I)
YMIN=Y(I)
YMAX=Y(I)
2 CONTINUE
3 CONTINUE
DO 5 I=N2,N3
IF(Y(I).EQ.0. .AND. IGNORE.NE.0) GO TO 5
IF(X(I).GT.XMAX) XMAX=X(I)
IF(X(I).LT.XMIN) XMIN=X(I)
IF(Y(I).GT.YMAX) YMAX=Y(I)
IF(Y(I).LT.YMIN) YMIN=Y(I)
5 CONTINUE
SPANX=XMAX-XMIN
SPANY=YMAX-YMIN
IF(SPANX.EQ.0. .OR. SPANY.EQ.0.)RETURN
B=FLOAT(LINES-1)

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K03

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DO 10 I=N1,N3
IF(Y(I).EQ.0. .AND. IGNORE.NE.0) GO TO 9
LY(I)=FIX(B*(YMAX-Y(I))/SPANY+.5)+1
LX(I)=FIX(74.*(X(I)-XMIN)/SPANX+.5)+1
GO TO 10
9 LY(I)=0
LX(I)=0
10 CONTINUE
NZERO=FIX(B*YMAX/SPANY+.5)+1
IF(NZERO.EQ.LINES .AND. YMIN.GT.0.)NZERO=LINES+1
IF(NZERO.EQ.1 .AND. YMAX.LT.0.) NZERO=0
WRITE(6,110)
DO 20 I=1,LINES
DO 15 J=1,75
A(J)=BLANK
IF(I.EQ.NZERO)A(J)=DOT
DO 15 K=N1,N3
15 IF(LX(K).EQ.J .AND. LY(K).EQ.I) A(J)=POINT
20 WRITE(6,105)A
WRITE(6,100)
WRITE(6,115)YMAX,YMIN,XMAX,XMIN
100 FORMAT(1H ,77(1H-))
105 FORMAT(2H I,75A1,1H1)
110 FORMAT(/1X,77(1H-))
115 FORMAT(" YMAX=",E11.4," YMIN=",E11.4," XMAX=",E11.4,
* " XMIN=",E11.4 )
IF(ITABLE.EQ.0) GO TO 25
WRITE(6,120) NPOINT,(X(I),Y(I),I=N1,N3)
120 FORMAT(/ " THE COORDINATES OF THE ",I3," POINTS ARE"//
*(3(" (",E9.3," ",E9.3," )",2X)))
125 FORMAT(/ 28X," RESPONSE SYSTEM NO.",I2 )
25 CONTINUE
RETURN
END
C
*****
SUBROUTINE KOLMIR(Z,N,DN,CRIT,IPRINT)
DIMENSION X(100),Z(100),DNCRIT(30)
DATA DNCRIT /0.,0.,0.,.381,.337,.319,.300,.285,.271,.258,.249,
*.242,.234,.227,.220,.213,.206,.200,.195,.190,.188,186,.184,.182,
*.180,.176,.172,.168,.164,.161 /
IF(N.LT.31) CRIT=DNCRIT(N)
IF(N.GT.30) CRIT=.886/SQRT(FLOAT(N))
SX=0.
SXX=0.
DO 1 I=1,N
X(I)=Z(I)
SX=SX+X(I)
1 SXX=SXX + X(I)*X(I)
AN=N
BAR=SX/AN
SIGMA=SQRT((SXX-AN*BAR*BAR)/FLOAT(N-1))
IF(SIGMA.EQ.0.)RETURN
DO 6 I=2,N
IF(X(I)-X(I-1))2,6,6
2 TEMP=X(I)
IM=I-1
DO 4 J=1,IM
L=I-J
IF(TEMP-X(L))3,5,5
3 X(L+1)=X(L)
4 CONTINUE
X(1)=TEMP

```

L03

```

GO TO 6
5 X(L+1)=TEMP
6 CONTINUE
AN=N
DN=0.
DO 7 I=1,N
XT=(X(I)-BAR)/SIGMA
FSTAR=SDF(XT)
D1=ABS(FSTAR-FLOAT(I-1)/AN)
D2=ABS(FSTAR-FLOAT(I)/AN)
7 DN=AMAX1(DN,D1,D2)
IF(IPRINT.EQ.0) RETURN
WRITE(6,100) DN,CRIT
IF(DN.GT.CRIT) WRITE(6,101)
100 FORMAT(" KOLMOGOROV-SMIRNOV STATISTIC FOR TEST OF NORMALITY",
* " OF THE RESIDUALS =",F7.4/" (CRIT.VALUE =",F6.3,
* ",P.LT.0.05--REF. H.W.LILLIEFORS,J.A.S.A.,62,P399,1967)"/ )
101 FORMAT(" THE STATISTIC INDICATES THAT THE RESIDUALS ARE NOT",
* " NORMALLY DISTRIBUTED."/ )
RETURN
END

```

	IF(IGO.EQ.2) GO TO 1001	6670
	IF(ANS.EQ.4HBACK) GO TO 1004	6680
	IF(ANS.NE.3HYES.AND.ANS.NE.2HNO) GO TO 301	6690
	PLOTDP=ANS	6700
	GO TO 302	6710
301	WRITE(6,1020)	6720
	GO TO 300	6730
302	NCON=NOP-NEWNOP	6740
	WRITE(6,750)	6750
	WRITE(6,525)	6760
	WRITE(6,752) NEWNOP,NCON,NOBS,NDUMMY,NFUNC,NIND	6770
	DO 305 I=1,NOP	6780
305	WRITE(6,754) I,F1(I),F2(I),F(I),STEP(I)	6790
	WRITE(6,756)	6800
	NSAVE=1	6810

L01

	DO 310 I=1,NOBS	6820
	N=LABEL(I)	6830
	IF(N.EQ.NSAVE+1) WRITE(6,813)N	6840
	NSAVE=N	6850
	IF(NIND.GT.1) GO TO 308	6860
	IF(ANS16.EQ.3HYES) GO TO 307	6870
	WRITE(6,758) I,X(I),Y(I)	6880
	GO TO 310	6890
307	WRITE(6,760) I,X(I),Y(I),WY(I),WYN(I)	6900
	GO TO 310	6910
308	IF(ANS16.EQ.3HYES) GO TO 309	6920
	WRITE(6,762) I,X(I),Z(I),Y(I)	6930
	GO TO 310	6940
309	WRITE(6,764) I,X(I),Z(I),Y(I),WY(I),WYN(I)	6950
310	CONTINUE	6960
	WRITE(6,818)	6970
	IF(NIND.LT.3) GO TO 312	6980
	WRITE(6,817) (I,I=3,9)	6990
	DO 311 I=1,NOBS	7000
311	WRITE(6,819) I,(XX(J,I),J=3,NIND)	7010
	WRITE(6,818)	7020
312	IF(PLOTDP.EQ.2HNO) GO TO 313	7030
	WRITE(6,765)	7040
	CALL PLOT(X,Y,NOBS,50)	7050
	IF(NIND.GT.1) WRITE(6,767)	7060
	IF(NIND.GT.1) CALL PLOT(Z,Y,NOBS,50)	7070
313	WRITE(6,766) MAX,STOPCR	7080
	IF(IQUAD.EQ.1) WRITE(6,768) SIMP	7090
	IF(ANS4.EQ.3HYES) WRITE(6,770)	7100
	IF(PLOTS.EQ.3HYES) WRITE(6,772)	7110
	IF(IPRINT.EQ.-1) WRITE(6,774)	7120
	IF(IPRINT.EQ.0) WRITE(6,776)	7130
	IF(IPRINT.GT.0) WRITE(6,778)	7140
	IF(PLT.EQ.3HYES) WRITE(6,780)	7150
	IF(ANS16.EQ.3HYES.AND.IWGHT.GT.1) WRITE(6,781) IWGHT,ANS16B	7160
	WRITE(6,790)	7170
	GO TO 158	7180
350	NPR=NOP	7190
	DO 3500 I=1,NOP	7200
	N=NOP+1-I	7210
	IF(STEP(N).NE.0.) GO TO 3501	7220
3500	NPR=NPR-1	7230
3501	IF(MAX.EQ.1) GO TO 160	7240
	NRUN=NRUN+1	7250
	NVIOL=0	7260
	IF(IPRINT.EQ.-1) GO TO 351	7270
	WRITE(6,790)	7280
	WRITE(6,525)	7290
	WRITE(6,789)	7300
351	CALL WELDR(F,STEP,NOP,FUNC,MAX,IPRINT,STOPCR,NLOOP,IQUAD,SIMP,	7310
	*VAR,LSQ,IFAUULT)	7320
	SKIP=.FALSE.	7330
	IF(IFAUULT.EQ.5.AND.IGO.EQ.2) GO TO 1001	7340
	IF(IFAUULT.EQ.5.AND.AR(1).EQ.4HBACK) GO TO 1004	7350
	LIMITS=0	7360
	DO 3510 I=1,NOP	7370
3510	FMIN(I)=F(I)	7380
	IF(IFAUULT.EQ.3) GO TO 3514	7390
	IF(IFAUULT.NE.1.AND.IFAUULT.NE.2) GO TO 352	7400
	ANS12=2HNO	7410
	IF(IFAUULT.EQ.2) GO TO 3511	7420
	WRITE(6,791)	7430

M01

	NQ=18	7440
	GO TO 1004	7450
3511	IFLAG=1	7460
	DO 3512 I=1,NOP	7470
	IF(F2(I).EQ.F1(I)) GO TO 3512	7480
	DEL=(F2(I)-F1(I))*0.001	7490
	IF(F(I)-F1(I).GT.DEL.AND.F2(I)-F(I).GT.DEL) GO TO 3512	7500
	IFLAG=2	7510
	GO TO 3601	7520
3512	CONTINUE	7530
	GO TO 3602	7540
3513	IF(IFLAG.EQ.2) WRITE(6,793)	7550
	IF(IFLAG.EQ.1) WRITE(6,794)	7560
	WRITE(6,8401) STOPCR	7570
	WRITE(6,8402) SIMP	7580
	IF(IFLAG.EQ.1) WRITE(6,795)	7590
	IF(IFAUULT.NE.2) GO TO 3514	7600
3520	WRITE(6,8404)	7610
	CALL READ(1,AR)	7620
	IF(IGO.EQ.2) GO TO 1001	7630
	IF(ANS.EQ.4HBACK.OR.ANS.EQ.2HNO) GO TO 3514	7640
	IF(ANS.EQ.2HOK.OR.ANS.EQ.3HYES) GO TO 3521	7650
	WRITE(6,1020)	7660
	GO TO 3520	7670
3521	IQUAD=0	7680
	GO TO 352	7690

DO 17 I=1,NOP	15960
17 PBAR(I)=0.0	15970
DO 18 I=1,NP1	15980
IF(I-IMAX)19,18,19	15990
19 DO 20 J=1,NOP	16000
20 PBAR(J) = PBAR(J)+G(I,J)	16010
18 CONTINUE	16020
DO 602 J=1,NOP	16030
602 PBAR(J) = PBAR(J)/NAP	16040
DO 21 I=1,NOP	16050
21 PSTAR(I)=A*(PBAR(I)-G(IMAX,I))+PBAR(I)	16060
CALL FUNCTN (PSTAR,HSTAR)	16070
NEVAL=NEVAL+1	16080
IF(IPRINT)57,57,56	16090
56 IF(NPR.LE.5.AND.MOD(NEVAL,IPRINT).EQ.0) WRITE(IW,1010)	16100
*NEVAL,HSTAR,(PSTAR(J),J=1,NPR)	16110

L02

IF(NPR.GT.5.AND.MOD(NEVAL,IPRINT).EQ.0) WRITE(IW,101)	16120
*NEVAL,HSTAR,(PSTAR(J),J=1,NPR)	16130
57 IF(HSTAR-HMIN)22,23,23	16140
22 DO 24 I=1,NOP	16150
24 PSTST(I)=C*(PSTAR(I)-PBAR(I))+PBAR(I)	16160
CALL FUNCTN (PSTST,HSTST)	16170
NEVAL=NEVAL+1	16180
IF(IPRINT)60,60,59	16190
59 IF(NPR.LE.5.AND.MOD(NEVAL,IPRINT).EQ.0) WRITE(IW,1010)	16200
*NEVAL,HSTST,(PSTST(J),J=1,NPR)	16210
IF(NPR.GT.5.AND.MOD(NEVAL,IPRINT).EQ.0) WRITE(IW,101)	16220
*NEVAL,HSTST,(PSTST(J),J=1,NPR)	16230
60 IF(HSTST-HMIN)25,26,26	16240
25 DO 27 I=1,NOP	16250
IF(STEP(I).NE.0.0) G(IMAX,I)=PSTST(I)	16260
27 CONTINUE	16270
H(IMAX)=HSTST	16280
GO TO 41	16290
23 DO 28 I=1,NP1	16300
IF(I-IMAX)29,28,29	16310
29 IF(HSTAR-H(I))26,28,28	16320
28 CONTINUE	16330
IF(HSTAR-HMAX)30,30,31	16340
30 DO 32 I=1,NOP	16350
IF(STEP(I).NE.0.0) G(IMAX,I)=PSTAR(I)	16360
32 CONTINUE	16370
HMAX=HSTAR	16380
H(IMAX)=HSTAR	16390
31 DO 33 I=1,NOP	16400
33 PSTST(I)=B+G(IMAX,I)*(1.0-B)+PBAR(I)	16410
CALL FUNCTN(PSTST,HSTST)	16420
NEVAL=NEVAL+1	16430
IF(IPRINT)63,63,62	16440
62 IF(NPR.LE.5.AND.MOD(NEVAL,IPRINT).EQ.0)WRITE(IW,1010)	16450
*NEVAL,HSTST,(PSTST(J),J=1,NPR)	16460
IF(NPR.GT.5.AND.MOD(NEVAL,IPRINT).EQ.0)WRITE(IW,101)	16470
*NEVAL,HSTST,(PSTST(J),J=1,NPR)	16480
63 IF(HSTST-HMAX)35,35,34	16490
35 DO 36 I=1,NOP	16500
IF(STEP(I).NE.0.0) G(IMAX,I)=PSTST(I)	16510
36 CONTINUE	16520
H(IMAX)=HSTST	16530
GO TO 41	16540
34 DO 38 I=1,NP1	16550
IF(I.EQ.IMIN) GO TO 38	16560
DO 39 J=1,NOP	16570
IF(STEP(J).NE.0.0) G(I,J)=(G(I,J)+G(IMIN,J))/2.0	16580
39 F(J) = G(I,J)	16590
CALL FUNCTN (F,H(I))	16600
NEVAL=NEVAL+1	16610
IF(IPRINT)38,38,65	16620
65 IF(NPR.LE.5.AND.MOD(NEVAL,IPRINT).EQ.0)WRITE(IW,1010)	16630
*NEVAL,H(I),(F(J),J=1,NPR)	16640
IF(NPR.GT.5.AND.MOD(NEVAL,IPRINT).EQ.0)WRITE(IW,101)	16650
*NEVAL,H(I),(F(J),J=1,NPR)	16660
38 CONTINUE	16670
GO TO 41	16680
26 DO 40 I=1,NOP	16690
IF(STEP(I).NE.0.0) G(IMAX,I)=PSTAR(I)	16700
40 CONTINUE	16710
H(IMAX)=HSTAR	16720
41 IF(NEVAL-MAX.GT.0) GO TO 46	16730

M02

IF(IFLAG.NE.1) GO TO 4100	16740
IF(LOOP-MAX0(NLOOP,MORE))45,46,45	16750
4100 IF(LOOP-NLOOP)45,46,45	16760
46 DHCV=000	16770
DMEAN=000	16780
DO 42 I=1,NP1	16790
IF(H(I).GE.1050) GO TO 45	16800
42 DMEAN=DMEAN+H(I)	16810
DMEAN=DMEAN/FLOAT(NP1)	16820
DO 601 I=1,NP1	16830
601 DHCV=DHCV+(H(I)-DMEAN)*(H(I)-DMEAN)	16840
DHCV=102*DSQRT(DHCV/FLOAT(NP1))/DMEAN	16850
DO 53 I=1,NOP	16860
IF(STEP(I).EQ.0.0) GO TO 53	16870
F(I)=0.0	16880
DO 54 J=1,NP1	16890
54 F(I)=F(I)+G(J,I)	16900
F(I)=F(I)/NP1	16910
53 CONTINUE	16920
CALL FUNCTN (F,FUNC)	16930
NEVAL=NEVAL+1	16940
IF(IPRINT)LE.0) GO TO 700	16950
IF(NPR.LE.5.AND.MOD(NEVAL,IPRINT).EQ.0)WRITE(IW,1010)	16960
*NEVAL,FUNC,(F(J),J=1,NPR)	16970
IF(NPR.GT.5.AND.MOD(NEVAL,IPRINT).EQ.0)WRITE(IW,101)	16980



1	SX=SX+X(I)	25260
	AN=N	25270
	BAR=SX/AN	25280
	SIGMA=SQRT((SX-AN*BAR*BAR)/FLOAT(N-1))	25290
	IF(SIGMA.EQ.0.)RETURN	25300
	DO 6 I=2,N	25310
	IF(X(I)-X(I-1))2,6,6	25320
2	TEMP=X(I)	25330
	IM=I-1	25340
	DO 4 J=1,IM	25350
	L=I-J	25360
	IF(TEMP-X(L))3,5,5	25370
3	X(L+1)=X(L)	25380
4	CONTINUE	25390
	X(1)=TEMP	25400
		25410

L03

	GO TO 6	25420
5	X(L+1)=TEMP	25430
6	CONTINUE	25440
	AN=N	25450
	DN=0.	25460
	DO 7 I=1,N	25470
	XT=(X(I)-BAR)/SIGMA	25480
	FSTAR=SDF(XT)	25490
	D1=ABS(FSTAR-FLOAT(I-1)/AN)	25500
	D2=ABS(FSTAR-FLOAT(I)/AN)	25510
7	DN=AMAX1(DN,D1,D2)	25520
	IF(IPRINT.EQ.0) RETURN	25530
	WRITE(6,100) DN,CRIT	25540
	IF(DN.GT.CRIT) WRITE(6,101)	25550
100	FORMAT(' KOLMOGOROV-SHIRNOV STATISTIC FOR TEST OF NORMALITY',	25560
	' OF THE RESIDUALS =',F7.4/ '(CRIT.VALUE =',F6.3,	25570
	' P.LT.0.05--REF. H.W.LILLIEFORS,J.A.S.A.,62,P399,1967)'/ )	25580
101	FORMAT(' THE STATISTIC INDICATES THAT THE RESIDUALS ARE NOT',	25590
	' NORMALLY DISTRIBUTED.'/ )	25600
	RETURN	25610
	END	25620
C	*****	25630
	FUNCTION SEQPRO(E,NE)	25640
	DIMENSION E(100)	25650
	N=0	25660
	NU=1	25670
	NFU=0	25680
	IF(E(1).GT.0.)N=1	25690
	DO 5 J=2,NE	25700
	IF(E(J).GT.0.)N=N+1	25710
5	IF(E(J).LT.0. .AND. E(J-1).GT.0.	25720
	' .OR. E(J).GT.0. .AND. E(J-1).LT.0.)NU=NU+1	25730
	IF(NU.EQ.1) GO TO 20	25740
	IF(N.LT.NE-N)N=NE-N	25750
	M=NE-N	25760
	MM=M-1	25770
	NN=N-1	25780
	IF(NE.GT.25) GO TO 25	25790
	DO 15 I=2,NU	25800
	IF(MOD(I,2).EQ.0) GO TO 10	25810
	KK=(I+1)/2-1	25820
	NFU=NFU+NBC(MM,KK)*NBC(NN,KK-1)+NBC(MM,KK-1)*NBC(NN,KK)	25830
	GO TO 15	25840
10	KK=I/2-1	25850
	NFU=NFU+2*NBC(MM,KK)*NBC(NN,KK)	25860
15	CONTINUE	25870
	SEQPRO=FLOAT(NFU)/FLOAT(NBC(NE,M))	25880
	RETURN	25890
20	SEQPRO=1.	25900
	RETURN	25910
25	AM=M	25920
	AN=N	25930
	BAR=2.*AM/AN/(AM+AN)+1.	25940
	SIGMA=(BAR-1.)*(BAR-2.)/(AM+AN-1.)	25950
	Z=(FLOAT(NU)+.5-BAR)/SIGMA	25960
	SEQPRO=SDF(Z)	25970
	RETURN	25980
	END	25990
C	*****	26000
	FUNCTION NBC(N,M)	26010
	DOUBLE PRECISION NC,ND,NE,NF	26020
	IF(M.GT.N) GO TO 6	26030

M03

	IF(N.EQ.M.OR.M.EQ.0) GO TO 5	26040
	IF(M.EQ.1.OR.N-M.EQ.1) GO TO 4	26050
	IF(2*M-N.GT.0) GO TO 1	26060
	NC=N-M	26070
	ND=NC+1	26080
	K=M-1	26090
	GO TO 2	26100
1	NC=M	26110
	ND=M+1	26120
	K=N-M-1	26130
2	NE=1	26140
	DO 3 J=1,K	26150
	NF=J+1	26160
	ND=ND+(NC+NF)	26170
3	NE=NE+NF	26180
	NBC=ND/NE	26190
	RETURN	26200
4	NBC=N	26210
	RETURN	26220
5	NBC=1	26230
	RETURN	26240
6	NBC=0	26250
	RETURN	26260
	END	26270
C	*****	26280
	FUNCTION NBC(N)	26290

IF(FAULT.EQ.5.AND.IGD.EQ.2) GO TO 1004	7340
IF(FAULT.EQ.5.AND.AR(1).EQ.4HBACK) GO TO 1004	7350
LIMITS=0	7360
DO 3510 I=1,NOP	7370
3510 FMIN(I)=F(I)	7380
IF(FAULT.EQ.3) GO TO 3514	7390
IF(FAULT.NE.1.AND.FAULT.NE.2) GO TO 352	7400
ANS12=2HNO	7410
IF(FAULT.EQ.2) GO TO 3511	7420
WRITE(6,791)	7430

M01

NO=18	7440
GO TO 1004	7450
3511 IFLAG=1	7460
DO 3512 I=1,NOP	7470
IF(F2(I).EQ.F1(I)) GO TO 3512	7480
DEL=(F2(I)-F1(I))*0.001	7490
IF(F(I)-F1(I).GT.DEL.AND.F2(I)-F(I).GT.DEL) GO TO 3512	7500
IFLAG=2	7510
GO TO 3601	7520
3512 CONTINUE	7530
GO TO 3602	7540
3513 IF(IFLAG.EQ.2) WRITE(6,793)	7550
IF(IFLAG.EQ.1) WRITE(6,794)	7560
WRITE(6,8401)STOPCR	7570
WRITE(6,8402)SIMP	7580
IF(IFLAG.EQ.1) WRITE(6,795)	7590
IF(FAULT.NE.2) GO TO 3514	7600
3520 WRITE(6,8404)	7610
CALL READ(1,AR)	7620
IF(IGD.EQ.2) GO TO 1001	7630
IF(ANS.EQ.4HBACK.OR.ANS.EQ.2HNO) GO TO 3514	7640
IF(ANS.EQ.2HOK.OR.ANS.EQ.3HYES) GO TO 3521	7650
WRITE(6,1020)	7660
GO TO 3520	7670
3521 IQUAD=0	7680
GO TO 352	7690
3514 NO=18	7700
GO TO 1004	7710
352 SSRES=0.	7720
SUMRES=0.	7730
WSUMRS=0.	7740
SSY=0.	7750
SY=0.	7760
SSYC=0.	7770
SYC=0.	7780
SYCY=0.	7790
IFLAG=0	7800
DO 355 II=1,NFUNC	7810
RSS(II)=0.	7820
WRSS(II)=0.	7830
N1=LIM1(II)	7840
N2=LIM2(II)	7850
DO 355 I=N1,N2	7860
DO 353 J=1,NIND	7870
353 G(J)=YX(J,I)	7880
B=Y(I)	7890
ITHFUN=II	7900
IF(NIND.EQ.1) CALL MODEL(B,G(1),F,0)	7910
IF(NIND.GT.1) CALL MODEL(B,G,F,0)	7920
YEST(I)=B	7930
RES(I)=Y(I)-YEST(I)	7940
IF(WYN(I).EQ.0.)RES(I)=0.	7950
WRRES(I)=RES(I)*SQRT(WYN(I))	7960
SUMRES=SUMRES+RES(I)	7970
WSUMRS=WSUMRS+WRRES(I)	7980
RESQ=RES(I)*RES(I)	7990
SSRES=SSRES+RESQ	8000
RSS(II)=RSS(II)+RESQ	8010
WRSS(II)=WRSS(II)+WYN(I)*RESQ	8020
IF(ABS(YEST(I)).LT.1E-50) DIFPCT(I)=9.999E+98	8030
IF(ABS(YEST(I)).GE.1E-50) DIFPCT(I)=100.*RES(I)/YEST(I)	8040
IF(WYN(I).EQ.0.) GO TO 355	8050

M01

SSY=SSY+Y(I)*Y(I)	8060
SY=SY+Y(I)	8070
SSYC=SSYC+B*B	8080
SYC=SYC+B	8090
SYCY=SYCY+B*Y(I)	8100
355 CONTINUE	8110
NOBS=NOBS	8120
NOBS=NOBS-NDUMMY	8130
YBAR=SY/FLOAT(NOBS)	8140
SSTOT=SSY-SY*SY/FLOAT(NOBS)	8150
B=NOBS	8160
R=ABS(SYCY-SY*SYC/B)/SQRT(ABS((SSY-SY*SY/B)*(SSYC-SYC*SYC/B)))	8170
RSQ=100*R*R	8180
SSREG=SSTOT-SSRES	8190
RESBAR=SUMRES/FLOAT(NOBS)	8200
WRBAR=WSUMRS/FLOAT(NOBS)	8210
NRGDF=NEWNOP-1	8220
IF(NRGDF.LE.0) NRGDF=1	8230
NRSDF=NOBS-NEWNOP	8240
NTODF=NOBS-1	8250
REGMSQ=SSREG/FLOAT(NRGDF)	8260
RESMSQ=SSRES/FLOAT(NRSDF)	8270
WRMSQ=FUNC/FLOAT(NRSDF)	8280
SDVRES=SQRT(RESMSQ)	8290
SDVRS=SQRT(WRMSQ)	8300
OVERF=REGMSQ/RESMSQ	8310
T=1.95	8320
IF(NRSDF.LE.30) OVERF=OVERF/T	8330
WRITE(6,790)	8340
WRITE(6,525)	8350
NOBS=NOBS	8360

```

IF(NPR.GT.5.AND.MOD(NEVAL,IPRINT).EQ.0)WRITE(IW,101) 16650
*NEVAL,H(I),(F(J),J=1,NPR) 16660
38 CONTINUE 16670
GO TO 41 16680
26 DO 40 I=1,NOP 16690
IF(STEP(I).NE.0.0) G(IMAX,I)=PSTAR(I) 16700
40 CONTINUE 16710
H(IMAX)=HSTAR 16720
41 IF(NEVAL-MAX.GT.0) GO TO 46 16730

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M02

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IF(IFLAG.NE.1) GO TO 4100 16740
IF(LOOP-MAX0(NLOOP,MORE))45,46,45 16750
4100 IF(LOOP-NLOOP)45,46,45 16760
46 DHCV=0D0 16770
DMEAN=0D0 16780
DO 42 I=1,NP1 16790
IF(H(I).GE.1D50) GO TO 45 16800
42 DMEAN=DMEAN+H(I) 16810
DMEAN=DMEAN/FLOAT(NP1) 16820
DO 601 I=1,NP1 16830
601 DHCV=DHCV+(H(I)-DMEAN)*(H(I)-DMEAN) 16840
DHCV=1D2*DSQRT(DHCV/FLOAT(NP1))/DMEAN 16850
DO 53 I=1,NOP 16860
IF(STEP(I).EQ.0.0) GO TO 53 16870
F(I)=0.0 16880
DO 54 J=1,NP1 16890
54 F(I)=F(I)+G(J,I) 16900
F(I)=F(I)/NP1 16910
53 CONTINUE 16920
CALL FUNCTN (F,FUNC) 16930
NEVAL=NEVAL+1 16940
IF(IPRINT.LE.0) GO TO 700 16950
IF(NPR.LE.5.AND.MOD(NEVAL,IPRINT).EQ.0)WRITE(IW,1010) 16960
*NEVAL,FUNC,(F(J),J=1,NPR) 16970
IF(NPR.GT.5.AND.MOD(NEVAL,IPRINT).EQ.0)WRITE(IW,101) 16980
*NEVAL,FUNC,(F(J),J=1,NPR) 16990
700 IF(NEVAL-MAX) 44,44,43 17000
43 GO TO 67 17010
67 WRITE(IW,102)MAX 17020
102 FORMAT(40H NUMBER OF FUNCTION EVALUATIONS EXCEEDS ,I4) 17030
IF(NVIOL.GT.10) WRITE(IW,8031)NVIOL 17040
WRITE(IW,103)DHCV 17050
103 FORMAT(" COEFFICIENT OF VARIATION (PCT) OF FUNCTION VALUES", 17060
* " OF LAST SIMPLEX",D10.4 ) 17070
IF(NPR.LE.5)WRITE(IW,1040)(F(I),I=1,NPR) 17080
1040 FORMAT(" CENTROID ",5E12.6) 17090
IF(NPR.GT.5)WRITE(IW,104)(F(I),I=1,NPR) 17100
104 FORMAT(" CENTROID ",5E12.6/(12X,5E12.6)) 17110
WRITE(IW,105)FUNC 17120
105 FORMAT(" FUNCTION VALUE AT CENTROID ",D15.8 / ) 17130
68 IFAULT = 1 17140
RETURN 17150
44 IF(DHCV-DBLE(STOPCR))72,48,48 17160
48 IFLAG=0 17170
LOOP=0 17180
GO TO 45 17190
72 IF(IPRINT)47,70,70 17200
70 WRITE(IW,106)DHCV 17210
106 FORMAT(" * INITIAL EVIDENCE OF CONVERGENCE COEFF. OF VARIATION", 17220
* "(PCT)",D10.4 ) 17230
IF(NPR.LE.5) WRITE(IW,1010)NEVAL,FUNC,(F(I),I=1,NPR) 17240
IF(NPR.GT.5) WRITE(IW,101)NEVAL,FUNC,(F(I),I=1,NPR) 17250
47 IF(IFLAG)49,50,49 17260
50 IFLAG=1 17270
51 DMEANS=DMEAN 17280
FUNCS=FUNC 17290
LOOP=0 17300
GO TO 45 17310
49 IF(DABS(1D1*(DMEANS-DMEAN)/DMEANS).GT.DBLE(STOPCR).OR. 17320
*DABS(1D2*(FUNC-FUNCS)/FUNCS).GT.DBLE(STOPCR)) GO TO 51 17330
NEVAL1=NEVAL 17340
IF(IQUAD.NE.0.OR.H(IMIN).GT.FUNC) GO TO 4905 17350

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M02

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FUNC=H(IMIN) 17360
DO 4900 J=1,NOP 17370
4900 F(I)=G(IMIN,J) 17380
4905 CONTINUE 17390
IF(IPRINT)74,73,73 17400
73 IF(NVIOL.GT.0) WRITE(IW,107)NEVAL,NVIOL 17410
107 FORMAT(" PROCESS CONVERGES AFTER ,I4,"-",",I3," FUNCTION", 17420
* " EVALUATIONS / ) 17430
IF(NPR.LE.5)WRITE(IW,1080)(F(I),I=1,NPR) 17440
1080 FORMAT(" MINIMUM AT ",5E12.6) 17450
IF(NPR.GT.5) WRITE(IW,108)(F(I),I=1,NPR) 17460
108 FORMAT(" MINIMUM AT ",5E12.6/(12X,5E12.6)) 17470
WRITE(IW,109)FUNC 17480
WRITE(IW,103)DHCV 17490
109 FORMAT(" 26H MINIMUM FUNCTION VALUE ,D15.8 / ) 17500
74 CONTINUE 17510
IF(IPRINT.GT.0.AND.NVIOL.GT.NPR) WRITE(IW,8035) 17520
8035 FORMAT(" (FUNC.VALUES GREATER THAN 1D+50 INDICATE VIOLATION", 17530
* " OF PARAMETER CONSTRAINS" ) 17540
IF(IPRINT.GE.0.AND.NVIOL.GT.1) WRITE(IW,8031)NVIOL 17550
8031 FORMAT(" * NO. OF VIOLATIONS OF PARAMETER CONSTRAINS", 17560
* " DURING MINIMIZATION ",I4 ) 17570
IF(IQUAD)200,75,200 17580
75 RETURN 17590
200 LIMITS=0 17600
ALOW=5HLOWER 17610
AUPP=5HUPPER 17620
ICNSTR=0 17630
DO 806 I=1,NOP 17640
IF(F1(I).EQ.F2(I)) GO TO 806 17650
DEL=(F2(I)-F1(I))*0.001 17660
IF(F1(I)-DEL.GT.F2(I)) GO TO 17670

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```

SIGMA=(BAR-1.)*(BAR-2.)/(AM+AN-1.)
Z=(FLOAT(NU)+.5-BAR)/SIGMA
SEOPRO=SDF(Z)
RETURN
END
C
*****
FUNCTION NBC(N,M)
DOUBLE PRECISION NC,ND,NE,NF
IF(M.GT.N) GO TO 6

```

```

25950
25960
25970
25980
25990
26000
26010
26020
26030

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N03

```

IF(N.EQ.M.OR.M.EQ.0) GO TO 5
IF(M.EQ.1.OR.N-M.EQ.1) GO TO 4
IF(2+M-N.GT.0) GO TO 1
NC=N-M
ND=NC+1
K=M-1
GO TO 2
1 NC=M
ND=M+1
K=N-M-1
2 NE=1
DO 3 J=1,K
NF=J+1
ND=ND+(NC+NF)
3 NE=NE+NF
NBC=ND/NE
RETURN
4 NBC=N
RETURN
5 NBC=1
RETURN
6 NBC=0
RETURN
END

```

```

26040
26050
26060
26070
26080
26090
26100
26110
26120
26130
26140
26150
26160
26170
26180
26190
26200
26210
26220
26230
26240
26250
26260
26270

```

```

C
*****
FUNCTION SDF(X)
AX=ABS(X)
T=1./(1+.2316419*AX)
D=.3989423*EXP(-X*X/2.)
SDF=1.-D*T*(((1.330274*T-1.821256)*T+1.781478)*T-
+.3565638)*T+.3193815)
IF(X)1,2,2
1 SDF=1.-SDF
2 RETURN
END

```

```

26280
26290
26300
26310
26320
26330
26340
26350
26360
26370
26380

```

```

C
*****
SUBROUTINE PROB1(E,NE,M,P)
DIMENSION E(100)
NS=0
N=0
DO 5 J=1,NE
5 IF(E(J).GT.0.) N=N+1
IF(N.LT.NE-N) N=NE-N
M=NE-N
MM=M+1
IF(NE.GT.25) GO TO 15
DO 10 J=1,MM
10 NS=NS+NBC(ME,J-1)
P=FLOAT(NS)*(2.**(-NE))
RETURN
15 Z=FLOAT(2*M+1-NE)/SQRT(FLOAT(NE))
P=SDF(Z)
RETURN
END

```

```

26390
26400
26410
26420
26430
26440
26450
26460
26470
26480
26490
26500
26510
26520
26530
26540
26550
26560
26570

```

```

C
*****
SUBROUTINE SEARCH(FNEW,STPNEW,SSMIN,ANS16,NPOINT,NEVAL,
*NEWNOP,STEP,NP)
DIMENSION FNEW(7),F1(20),F2(20),STPNEW(7),P1(75),P2(75),XX(9,100),
*SS(75),SSS(10,10),PDEV(10,20),FF(7),Y(100),LABEL(100),
*WYN(100),DEL(7),PP(7,12),IS(7),STEP(20),A(10),AL(9),CON(10,10)
COMMON /DATA/ XX,Y,WYN,NOBS
COMMON /CONSTR/LIMITS

```

```

26580
26590
26600
26610
26620
26630
26640
26650

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N03

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COMMON /PARLIN/ F1,F2
COMMON /FUNNUM/ITHFUN
COMMON /B4/ NIND
COMMON /B6/ NOP
COMMON /B10/LABEL
DATA AL/3H1 ,3H2 ,3H3 ,3H4 ,3H5 ,3H6 ,3H7 ,3H8 ,3H9 /
LIMITS=0
IF(NEWNOP-2) 5,50,135
5 CONTINUE
DO 10 I=1 NOP
FNEW(I)=F1(I)
STPNEW(I)=STEP(I)
10 IF(F1(I).NE.F2(I)) IP=I
DEL(IP)=(F2(IP)-F1(IP))/74.
DO 20 I=1,75
FNEW(IP)=F1(IP)+F2(IP)-DEL(IP)
CALL LSQ(FNEW,SS(I))
20 P1(I)=FNEW(IP)
IF(ANS16.EQ.2HNO) WRITE(6,1000)IP,F1(IP),F2(IP)
IF(ANS16.EQ.3HYES) WRITE(6,1010)IP,F1(IP),F2(IP)
CALL PLOT(P1,SS,75,50)
IF(ANS16.EQ.2HNO) WRITE(6,1020)
IF(ANS16.EQ.3HYES) WRITE(6,1025)
DO 30 I=1,37
J=I+37
30 WRITE(6,1026)I,SS(I),P1(I),J,SS(J),P1(J)
WRITE(6,1027)
ISAVE=1
SSMIN=SS(1)
DO 40 I=2,75
IF(SS(I).GE.SSMIN) GO TO 40
SSMIN=SS(I)

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26660
26670
26680
26690
26700
26710
26720
26730
26740
26750
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26800
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26870
26880
26890
26900
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26960
26970

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YEST(1)=B	7930
RES(1)=Y(1)-YEST(1)	7940
IF(WYN(1).EQ.0)RES(1)=0.	7950
WRRES(1)=RES(1)*SQRT(WYN(1))	7960
SUMRES=SUMRES+RES(1)	7970
WSUMRS=WSUMRS+WRRES(1)	7980
RESQ=RES(1)*RES(1)	7990
SSRES=SSRES+RESQ	8000
RSS(1)=RSS(1)+RESQ	8010
WRSS(1)=WRSS(1)+WYN(1)*RESQ	8020
IF(ABS(YEST(1)).LT.1E-50) DIFPCT(1)=9.999E+98	8030
IF(ABS(YEST(1)).GE.1E-50) DIFPCT(1)=100.*RES(1)/YEST(1)	8040
IF(WYN(1).EQ.0.) GO TO 355	8050

NO1

SSY=SSY+Y(1)*Y(1)	8060
SY=SY+Y(1)	8070
SSYC=SSYC+B*B	8080
SYC=SYC+B	8090
SYCY=SYCY+B*Y(1)	8100
355 CONTINUE	8110
NOBSS=NOBSS	8120
NOBS=NOBS-NDUMMY	8130
YBAR=SY/FLOAT(NOBS)	8140
SSTOT=SSY-SY*SY/FLOAT(NOBS)	8150
B=NOBS	8160
R=ABS(SYCY-SY*SYC/B)/SQRT(ABS((SSY-SY*SY/B)*(SSYC-SYC*SYC/B)))	8170
RSQ=100*R*R	8180
SSREG=SSTOT-SSRES	8190
RESBAR=SUMRES/FLOAT(NOBS)	8200
WRSBAR=WSUMRS/FLOAT(NOBS)	8210
NRGDF=NEWHOP-1	8220
IF(NRGDF.LE.0) NRGDF=1	8230
NRSDF=NOBS-NEWHOP	8240
NTODF=NOBS-1	8250
REGMSQ=SSREG/FLOAT(NRGDF)	8260
RESMSQ=SSRES/FLOAT(NRSDF)	8270
WRSMSQ=FUNC/FLOAT(NRSDF)	8280
SDVRES=SQRT(RESMSQ)	8290
SDVRS=SQRT(WRSMSQ)	8300
OVERF=REGMSQ/RESMSQ	8310
T=1.95	8320
IF(NRSDF.LE.30) T=EXPIT(1/NRSDF)	8330
WRITE(6,790)	8340
WRITE(6,525)	8350
NOBS=NOBSS	8360
IF(IQUAD.EQ.0) GO TO 3610	8370
WRITE(6,800)	8380
FAC=SQRT(2.*FUNC/FLOAT(NRSDF))	8390
DO 360 I=1,NOP	8400
SDV=FAC*SQRT(VAR(I))	8410
IF(ABS(F(I)).LT.1E-50) CV=9.999E+98	8420
IF(ABS(F(I)).GE.1E-50) CV=100.*SDV/ABS(F(I))	8430
IF(SDV.EQ.0.) CV=0.	8440
360 WRITE(6,802)I,F1(I),F2(I),FSAVE(I),F(I),SDV,CV	8450
WRITE(6,803)	8460
DO 361 I=1,NOP	8470
E=T*FAC*SQRT(VAR(I))	8480
E1=F(I)-E	8490
E2=F(I)+E	8500
361 IF(F(I).NE.F2(I)) WRITE(6,805)I,E1,E2	8510
WRITE(6,8050)	8520
3600 CONTINUE	8530
IFLAG=0	8540
GO TO 3602	8550
3601 WRITE(6,792)	8560
3602 CALL PLACE(F1,F2,F,65,NPR,A)	8570
IF(NPR.EQ.1) WRITE(6,8051)	8580
IF(NPR.GT.1) WRITE(6,8052)	8590
DO 3605 J=1,NPR	8600
IF(F1(J).EQ.F2(J)) GO TO 3605	8610
N1=65*(J-1)+1	8620
N2=N1+64	8630
WRITE(6,8053) J,(A(1),I=N1,N2)	8640
3605 CONTINUE	8650
IF(IFLAG.NE.0.AND.ANS16.EQ.2HNO) WRITE(6,8054)FUNC	8660
IF(IFLAG.NE.0.AND.ANS16.NE.2HNO) WRITE(6,8055)FUNC	8670

001

GO TO 3612	8680
3610 WRITE(6,8010)	8690
DO 3611 I=1,NOP	8700
3611 WRITE(6,8011)I,F1(I),F2(I),FSAVE(I),F(I)	8710
WRITE(6,8012)	8720
GO TO 3600	8730
3612 CONTINUE	8740
ALOW=5HLOWER	8750
AUPP=5HUPPER	8760
ICNSTR=0	8770
DO 3621 I=1,NOP	8780
IF(F1(I).EQ.F2(I)) GO TO 3621	8790
DEL=(F2(I)-F1(I))*0.001	8800
IF(F1(I)-F1(I).GT.DEL) GO TO 3620	8810
ICNSTR=ICNSTR+1	8820
WRITE(6,8030) ALOW,I	8830
3620 IF(F2(I)-F1(I).GT.DEL) GO TO 3621	8840
ICNSTR=ICNSTR+1	8850
WRITE(6,8030) AUPP,I	8860
3621 CONTINUE	8870
IF(ICNSTR.EQ.0) GO TO 3622	8880
WRITE(6,8031)	8890
CALL READ(1,AR)	8900
IF(IGD.EQ.2.OR.ANS.EQ.4HBACK) WRITE(6,844) NRUN	8910
IF(IGD.EQ.2) GO TO 1001	8920
IF(ANS.EQ.4HBACK) GO TO 1004	8930
3622 CONTINUE	8940
IF(IFLAG.NE.0) GO TO 3513	8950

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IF(NPR.LE.5) WRITE(IW,1010)NEVAL,FUNC,(F(I),I=1,NPR) 17240
IF(NPR.GT.5) WRITE(IW,1011)NEVAL,FUNC,(F(I),I=1,NPR) 17250
47 IF(IFLAG)49,50,49 17260
50 IFLAG=1 17270
51 DMEANS=DMEAN 17280
FUNC=FUNC 17290
LOOP=0 17300
GO TO 45 17310
49 IF(DABS(1D1+(DMEANS-DMEAN)/DMEANS).GT.DBLE(STOPCR).OR. 17320
*DABS(1D2+(FUNC-FUNCS)/FUNCS).GT.DBLE(STOPCR)) GO TO 51 17330
NEVAL1=NEVAL 17340
IF(IQUAD.NE.0.OR.H(IMIN).GT.FUNC) GO TO 4905 17350

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402

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FUNC=H(IMIN) 17360
DO 4900 J=1,NOP 17370
4900 F(I)=G(IMIN,J) 17380
4905 CONTINUE 17390
IF(IPRINT)74,73,73 17400
73 IF(NVIOL.GT.0) WRITE(IW,107)NEVAL,NVIOL 17410
107 FORMAT(' PROCESS CONVERGES AFTER ',I4,'-',I3,' FUNCTION', 17420
* ' EVALUATIONS / ') 17430
IF(NPR.LE.5)WRITE(IW,1080)(F(I),I=1,NPR) 17440
1080 FORMAT(' MINIMUM AT ',5E12.6) 17450
IF(NPR.GT.5) WRITE(IW,108)(F(I),I=1,NPR) 17460
108 FORMAT(' MINIMUM AT ',5E12.6/(12X,5E12.6)) 17470
WRITE(IW,109)FUNC 17480
WRITE(IW,103)DMCV 17490
109 FORMAT(/ 26H MINIMUM FUNCTION VALUE ,D15.8/ ) 17500
74 CONTINUE 17510
IF(IPRINT.GT.0.AND.NVIOL.GT.NPR) WRITE(IW,8035) 17520
8035 FORMAT(' (FUNC.VALUES GREATER THAN 1D+50 INDICATE VIOLATION', 17530
* ' OF PARAMETER CONSTRAINS' ) 17540
IF(IPRINT.GE.0.AND.NVIOL.GT.1) WRITE(IW,8031)NVIOL 17550
8031 FORMAT(' * NO. OF VIOLATIONS OF PARAMETER CONSTRAINS', 17560
* ' DURING MINIMIZATION ',I4 ) 17570
IF(IQUAD)200,75,200 17580
75 RETURN 17590
200 LIMITS=0 17600
ALOW=3HLOWER 17610
AUPP=3HUPPER 17620
ICNSTR=0 17630
DO 806 I=1,NOP 17640
IF(F1(I).EQ.F2(I)) GO TO 806 17650
DEL=(F2(I)-F1(I))*0.001 17660
IF(F(I)-F1(I).GT.DEL) GO TO 806 17670
ICNSTR=ICNSTR+1 17680
IF(ICNSTR.NE.1) GO TO 800 17690
WRITE(6,8000) FUNC,(F(L),L=1,NPR) 17700
WRITE(6,8005) 17710
800 WRITE(6,8010) ALOW,F1(I),I,F(I) 17720
802 IF(F2(I)-F(I).GT.DEL) GO TO 306 17730
ICNSTR=ICNSTR+1 17740
IF(ICNSTR.NE.1) GO TO 804 17750
WRITE(6,8000) FUNC,(F(L),L=1,NPR) 17760
WRITE(6,8005) 17770
804 WRITE(6,8010) AUPP,F2(I),I,F(I) 17780
806 CONTINUE 17790
IF(ICNSTR.EQ.0) GO TO 814 17800
IF(IPRINT.EQ.-1.AND.NVIOL.GT.1) WRITE(IW,8031)NVIOL 17810
WRITE(6,8015) 17820
808 CALL READ(1,AR) 17830
IF(AR(1).EQ.4HBACK.OR.1GO.EQ.2) GO TO 810 17840
IF(AR(1).EQ.2HNO) GO TO 812 17850
IF(AR(1).EQ.3HYES) GO TO 814 17860
WRITE(6,8030) 17870
GO TO 808 17880
810 IFAULT=5 17890
WRITE(6,8020) 17900
RETURN 17910
812 IQUAD=0 17920
WRITE(6,8025) 17930
RETURN 17940
814 CONTINUE 17950
8000 FORMAT(' * THE RESIDUAL SUM OF SQUARES MINIMUM (' ,D12.6, 17960
* ') AT THE PARAMETER VALUES / * / (' ,7E10.4) ) 17970

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002

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8005 FORMAT(' * / * SEEM TO BE CONSTRAINED BY THE "/ * " ) 17980
8010 FORMAT(' * ,5X,A5, LIMIT (' ,E11.5, ') OF PAR.NO.,I2, 17990
* (' ,E11.5, ') ) 18000
8015 FORMAT(' * / * A STATISTICAL EVALUATION OF THE PARAMETERS UNDER', 18010
* ' THESE CONDITIONS MAY FAIL.' / 18020
* ' * ENTER B10 OR B11 IF YOU WANT A NEW RUN WITH DIFFERENT' / 18030
* ' * PARAMETER LIMITS OR DIFFERENT INITIAL PARAMETER ESTIMATES' / 18040
* ' * ENTER -YES- IF YOU WANT TO CONTINUE OR' / 18050
* ' * ENTER -NO- IF YOU WANT TO CONTINUE BUT WITHOUT A STATISTICAL', 18060
* ' EVALUATION' // ) 18070
8020 FORMAT(/1X,77(1H-)/) 18080
8025 FORMAT(/' * REMEMBER YOUR INPUT UNDER REQUEST NO.3 HAS NOW', 18090
* ' BEEN CHANGED TO -NO-' / ) 18100
8030 FORMAT(/' INPUT ERROR, YOUR ANSWER MUST BE ONE OF THE FOLLOWING' / 18110
* ' FOUR, Y N B R - TRY AGAIN -' / ) 18120
IF(IPRINT)233,232,232 18130
232 WRITE(IW,301) 18140
301 FORMAT(/1X,13(1H*), ' FITTING OF QUADRATIC SURFACE IN REGION OF', 18150
* ' MINIMUM ',13(1H*) / ) 18160
233 NEVAL=0 18170
NFIX=0 18180
B=SIMP/100. 18190
SIMP2=FUNC*B 18200
SIMP2=AMAX1(B,SIMP2) 18210
DO 201 I=1,NP1 18220
204 IF(H(I).GE.1D50) GO TO 2030 18230
TEST=DABS(H(I)-FUNC) 18240
IF(TEST-SIMP2)202,201,201 18250
202 DO 203 J=1,NOP 18260

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Z=FLOAT(Z*P+1-NE)/SQRT(FLOAT(NE))	26540
P=SDF(Z)	26550
RETURN	26560
END	26570
*****	26580
C SUBROUTINE SEARCH(FNEW,STPNEW,SSMIN,ANS16,NPOINT,NEVAL,	26590
*NEMNOP,STEP,NP)	26600
DIMENSION FNEW(7),F1(20),F2(20),STPNEW(7),P1(75),P2(75),XX(9,100),	26610
*SS(75),SSS(10,10),PDEV(10,20),FF(7),Y(100),LABEL(100),	26620
*MYN(100),DEL(7),PP(7,12),IS(7),STEP(20),A(10),AL(9),CON(10,10)	26630
COMMON /DATA/ XX,Y,MYN,NOBS	26640
COMMON /CONSTR/LIMITS	26650

003

COMMON /PARLIM/ F1,F2	26660
COMMON /FUNNUM/ITHFUN	26670
COMMON /B4/ NIND	26680
COMMON /B6/ NOP	26690
COMMON /B10/LABEL	26700
DATA AL/3H1 ,3H2 ,3H3 ,3H4 ,3H5 ,3H6 ,3H7 ,3H8 ,3H9 /	26710
LIMITS=0	26720
IF(NEMNOP-2) 5,50,135	26730
5 CONTINUE	26740
DO 10 I=1,NOP	26750
FNEW(I)=F1(I)	26760
STPNEW(I)=STEP(I)	26770
10 IF(F1(I).NE.F2(I)) IP=1	26780
DEL(IP)=(F2(IP)-F1(IP))/74.	26790
DO 20 I=1,75	26800
FNEW(IP)=F1(IP)+FLOAT(I-1)*DEL(IP)	26810
CALL LSO(FNEW,SS(I))	26820
20 P1(I)=FNEW(IP)	26830
IF(ANS16.EQ.2HND) WRITE(6,1000)IP,F1(IP),F2(IP)	26840
IF(ANS16.EQ.3HYES) WRITE(6,1010)IP,F1(IP),F2(IP)	26850
CALL PLOT(P1,SS,75,50)	26860
IF(ANS16.EQ.2HND) WRITE(6,1020)	26870
IF(ANS16.EQ.3HYES) WRITE(6,1025)	26880
DO 30 I=1,37	26890
J=I+37	26900
30 WRITE(6,1026)I,SS(I),P1(I),J,SS(J),P1(J)	26910
WRITE(6,1027)	26920
ISAVE=1	26930
SSMIN=SS(I)	26940
DO 40 I=2,75	26950
IF(SS(I).GE.SSMIN) GO TO 40	26960
SSMIN=SS(I)	26970
ISAVE=I	26980
40 CONTINUE	26990
FNEW(IP)=P1(ISAVE)	27000
STPNEW(IP)=DEL(IP)/2.	27010
RETURN	27020
50 IP1=7	27030
DO 55 I=1,NOP	27040
FNEW(I)=F1(I)	27050
STPNEW(I)=STEP(I)	27060
IF(I.GT.IP1) GO TO 55	27070
IF(F1(I).NE.F2(I)) IP1=I	27080
55 IF(F1(I).NE.F2(I)) IP2=I	27090
DEL(IP1)=(F2(IP1)-F1(IP1))/9.	27100
DEL(IP2)=(F2(IP2)-F1(IP2))/9.	27110
ISAVE=1	27120
JSAVE=1	27130
SSMIN=1.E99	27140
SSMAX=0.	27150
DO 60 J=1,10	27160
FNEW(IP1)=F1(IP1)+FLOAT(J-1)*DEL(IP1)	27170
P1(J)=FNEW(IP1)	27180
DO 60 I=1,10	27190
FNEW(IP2)=F1(IP2)+FLOAT(I-1)*DEL(IP2)	27200
P2(I)=FNEW(IP2)	27210
CALL LSO(FNEW,SSS(I,J))	27220
IF(SSS(I,J).GE.SSMIN) GO TO 60	27230
SSMIN=SSS(I,J)	27240
ISAVE=I	27250
JSAVE=J	27260
60 IF(SSS(I,J).GT.SSMAX) SSMAX=SSS(I,J)	27270

003

FNEW(IP1)=P1(JSAVE)	27280
FNEW(IP2)=P2(ISAVE)	27290
STPNEW(IP1)=DEL(IP1)/2.	27300
STPNEW(IP2)=DEL(IP2)/2.	27310
D=(SSMAX-SSMIN)/9.	27320
DO 65 I=1,10	27330
65 A(I)=SSMIN+(I-1)*D	27340
A(10)=A(10)+0.01*D	27350
WRITE(6,1030)IP1,(I,P1(I),I=1,10)	27360
WRITE(6,1040)IP2,(I,P2(I),I=1,10)	27370
IF(ANS16.EQ.2HND) WRITE(6,1050)	27380
IF(ANS16.EQ.3HYES) WRITE(6,1060)	27390
WRITE(6,1070)IP1,IP2	27400
WRITE(6,1075)(I,I=1,10)	27410
DO 67 I=1,10	27420
WRITE(6,1080)(SSS(I,J),J=1,9,2)	27430
67 WRITE(6,1085)(SSS(I,J),J=2,10,2)	27440
DO 120 J=1,10	27450
DO 120 I=1,10	27460
IF(J.EQ.1) GO TO 70	27470
IF(J.EQ.10) GO TO 80	27480
PDEV(I,J)=SIGNOF(SSS(I,J+1)-SSS(I,J-1))	27490
GO TO 90	27500
70 PDEV(I,J)=SIGNOF(4.*SSS(I,J+1)-3.*SSS(I,J)-SSS(I,J+2))	27510
GO TO 90	27520
80 PDEV(I,J)=SIGNOF(-4.*SSS(I,J-1)+3.*SSS(I,J)+SSS(I,J-2))	27530
90 K=J+10	27540
IF(I.EQ.1) GO TO 100	27550
IF(I.EQ.10) GO TO 110	27560

IF(SDV.EQ.0.) CV=0.	8440
360 WRITE(6,802)I,F1(I),F2(I),FSAVE(I),F(I),SDV,CV	8450
WRITE(6,803)	8460
DO 361 I=1,NOP	8470
E=T*FAC*SQRT(VAR(I))	8480
E1=F(I)-E	8490
E2=F(I)+E	8500
361 IF(F1(I).NE.F2(I)) WRITE(6,805)I,E1,E2	8510
WRITE(6,8050)	8520
3600 CONTINUE	8530
IFLAG=0	8540
GO TO 3602	8550
3601 WRITE(6,792)	8560
3602 CALL PLACE(F1,F2,F.65,NPR,A)	8570
IF(NPR.EQ.1) WRITE(6,8051)	8580
IF(NPR.GT.1) WRITE(6,8052)	8590
DO 3605 J=1,NPR	8600
IF(F1(J).EQ.F2(J)) GO TO 3605	8610
N1=65*(J-1)+1	8620
N2=N1+64	8630
WRITE(6,8053) J,(A(I),I=N1,N2)	8640
3605 CONTINUE	8650
IF(IFLAG.NE.0.AND.ANS16.EQ.2HND) WRITE(6,8054)FUNC	8660
IF(IFLAG.NE.0.AND.ANS16.NE.2HND) WRITE(6,8055)FUNC	8670

001

GO TO 3612	8680
3610 WRITE(6,8010)	8690
DO 3611 I=1,NOP	8700
3611 WRITE(6,8011)I,F1(I),F2(I),FSAVE(I),F(I)	8710
WRITE(6,8012)	8720
GO TO 3600	8730
3612 CONTINUE	8740
ALOW=5MLOWER	8750
AUPP=5MUPPER	8760
ICNSTR=0	8770
DO 3621 I=1,NOP	8780
IF(F1(I).EQ.F2(I)) GO TO 3621	8790
DEL=(F2(I)-F1(I))*0.001	8800
IF(F(I)-F1(I).GT.DEL) GO TO 3620	8810
ICNSTR=ICNSTR+1	8820
WRITE(6,8030) ALOW,I	8830
3620 IF(F2(I)-F1(I).GT.DEL) GO TO 3621	8840
ICNSTR=ICNSTR+1	8850
WRITE(6,8030) AUPP,I	8860
3621 CONTINUE	8870
IF(ICNSTR.EQ.0) GO TO 3622	8880
WRITE(6,8031)	8890
CALL READ(1,AR)	8900
IF(IGD.EQ.2.OR.ANS.EQ.4HBACK) WRITE(6,844) NRUN	8910
IF(IGD.EQ.2) GO TO 1001	8920
IF(ANS.EQ.4HBACK) GO TO 1004	8930
3622 CONTINUE	8940
IF(IFLAG.NE.0) GO TO 3513	8950
WRITE(6,804)SSRES,SSREG,SSTOT,SSY,YBAR,RESMSQ,REGMSQ,RESBAR,	8960
*SDVRES,RSQ,R	8970
IF(ANS16.EQ.3HYES.AND.NOWGHT.NE.1) WRITE(6,806)FUNC,WRSMSQ,	8980
*WRSBAR,MSDVRS	8990
IF(NFUNC.EQ.1) GO TO 364	9000
DO 363 I=1,NFUNC	9010
WRITE(6,807)II,RSS(II)	9020
363 IF(ANS16.EQ.3HYES.AND.NOWGHT.NE.1) WRITE(6,808)II,WRSS(II)	9030
364 IF(RSQ.LT.50.) WRITE(6,810)	9040
WRITE(6,812)	9050
NOUTL=0	9060
NSAVE=1	9070
DO 365 I=1,NOBS	9080
N=LABEL(I)	9090
IF(N.EQ.NSAVE+1) WRITE(6,813)N	9100
NSAVE=N	9110
ANRDV=RES(I)/SDVRES	9120
IF(NIND.EQ.1)WRITE(6,814)I,X(I),Y(I),YEST(I),RES(I),DIFPCT(I)	9130
*ANRDV	9140
IF(NIND.GT.1)WRITE(6,816)I,X(I),Z(I),Y(I),YEST(I),RES(I),	9150
*DIFPCT(I),ANRDV	9160
IF(ABS(ANRDV).LT.T) GO TO 365	9170
NOUTL=NOUTL+1	9180
IOUTL(IOUTL)=I	9190
365 CONTINUE	9200
WRITE(6,818)	9210
IF(NIND.LT.3) GO TO 367	9220
WRITE(6,817)(I,I=3,9)	9230
DO 366 I=1,NOBS	9240
366 WRITE(6,819)I,(XX(J,I),J=3,NIND)	9250
WRITE(6,818)	9260
367 IF(NOUTL.EQ.0) GO TO 3701	9270
WRITE(6,820) NRSDF	9280
DO 370 I=1,NOUTL	9290

P01



IF(ICNSTR.NE.1) GO TO 804	17750
WRITE(6,8000) FUNC,(F(L),L=1,NPR)	17760
WRITE(6,8005)	17770
804 WRITE(6,8010) AUPP,F2(I),I,F(I)	17780
806 CONTINUE	17790
IF(ICNSTR.EQ.0) GO TO 814	17800
IF(IPRINT.EQ.-1.AND.NVIOL.GT.1) WRITE(IW,8031)NVIOL	17810
WRITE(6,8015)	17820
808 CALL READ(1,AR)	17830
IF(AR(1).EQ.4HBACK.OR.IGQ.EQ.2) GO TO 810	17840
IF(AR(1).EQ.2HND) GO TO 812	17850
IF(AR(1).EQ.3HYES) GO TO 814	17860
WRITE(6,8030)	17870
GO TO 808	17880
810 IFAULT=5	17890
WRITE(6,8020)	17900
RETURN	17910
812 IQUAD=0	17920
WRITE(6,8025)	17930
RETURN	17940
814 CONTINUE	17950
8000 FORMAT( ' * THE RESIDUAL SUM OF SQUARES MINIMUM (',D12.6,	17960
' * ) AT THE PARAMETER VALUES /' * /(' * ',7E10.4))	17970

002

8005 FORMAT( ' * /' * SEEM TO BE CONSTRAINED BY THE' /' * ' )	17980
8010 FORMAT( ' * ,5X,A5, " LIMIT (',E11.5," ) OF PAR.NO.',I2,	17990
' * (',E11.5," ) )	18000
8015 FORMAT( ' * /' * A STATISTICAL EVALUATION OF THE PARAMETERS UNDER' ,	18010
' * THESE CONDITIONS MAY FAIL.' /	18020
' * * ENTER B10 OR B11 IF YOU WANT A NEW RUN WITH DIFFERENT' /	18030
' * * PARAMETER LIMITS OR DIFFERENT INITIAL PARAMETER ESTIMATES' /	18040
' * * ENTER -YES- IF YOU WANT TO CONTINUE OR' /	18050
' * * ENTER -NO- IF YOU WANT TO CONTINUE BUT WITHOUT A STATISTICAL' ,	18060
' * EVALUATION' / / )	18070
8020 FORMAT(/1X,77(1H-)/ / )	18080
8025 FORMAT(/' * REMEMBER YOUR INPUT UNDER REQUEST NO.3 HAS NOW' ,	18090
' * BEEN CHANGED TO -NO-' / )	18100
8030 FORMAT(/' INPUT ERROR, YOUR ANSWER MUST BE ONE OF THE FOLLOWING' /	18110
' * FOUR, Y N B R - TRY AGAIN -' / )	18120
IF(IPRINT)233,232,232	18130
232 WRITE(IW,301)	18140
301 FORMAT(/1X,13(1H+), " FITTING OF QUADRATIC SURFACE IN REGION OF",	18150
' * MINIMUM ',13(1H+ ) / )	18160
233 NEVAL=0	18170
NFIX=0	18180
B=SIMP/100.	18190
SIMP2=FUNC*B	18200
SIMP2=AMAX1(B,SIMP2)	18210
DO 201 I=1,NP1	18220
204 IF(H(I).GE.1D50) GO TO 2030	18230
TEST=DABS(H(I)-FUNC)	18240
IF(TEST-SIMP2)202,201,201	18250
202 DO 203 J=1,NOP	18260
IF(STEP(J).NE.0.0) G(I,J)=(G(I,J)-F(J))+G(I,J)	18270
203 PSTST(J)=G(I,J)	18280
CALL FUNCTN(PSTST,H(I))	18290
NEVAL=NEVAL+1	18300
GO TO 204	18310
2030 DO 2031 J=1,NOP	18320
IF(STEP(J).EQ.0.) GO TO 2031	18330
IF(G(I,J).GT.F2(J)) G(I,J)=F2(J)	18340
IF(G(I,J).LT.F1(J)) G(I,J)=F1(J)	18350
2031 PSTST(J)=G(I,J)	18360
CALL FUNCTN(PSTST,H(I))	18370
NEVAL=NEVAL+1	18380
NFIX=NFIX+1	18390
201 CONTINUE	18400
A0 = H(1)	18410
DO 205 I=1,NAP	18420
I1=I+1	18430
DO 206 J=1,NOP	18440
206 PSTAR(J)=(G(I,J)+G(I1,J))/2.0	18450
CALL FUNCTN(PSTAR,AVAI(I))	18460
IF(AVAI(I).GT.(H(I)+H(I1))/2D0)AVAI(I)=H(I)+H(I1)-AVAI(I)	18470
205 NEVAL=NEVAL+1	18480
DO 207 I=1,NAP	18490
I1=I-1	18500
I2=I+1	18510
IF(I1.LT.1)GOTO207	18520
DO 208 J=1,I1	18530
J1=J+1	18540
DO 209 K=1,NOP	18550
209 PSTST(K)=(G(I2,K)+G(J1,K))/2.0	18560
CALL FUNCTN(PSTST,HSTST)	18570
IF(HSTST.GT.(H(I2)+H(J1))/2D0)HSTST=H(I2)+H(J1)-HSTST	18580
NEVAL=NEVAL+1	18590

P02

FNEW(I)=F1(I)	27050
STPNEM(I)=STEP(I)	27060
IF(I.GT.IP1) GO TO 55	27070
IF(F1(I).NE.F2(I)) IP1=I	27080
55 IF(F1(I).NE.F2(I)) IP2=I	27090
DEL(IP1)=(F2(IP1)-F1(IP1))/9.	27100
DEL(IP2)=(F2(IP2)-F1(IP2))/9.	27110
ISAVE=1	27120
JSAVE=1	27130
SSMIN=1.E99	27140
SSMAX=0.	27150
DO 60 J=1,10	27160
FNEW(IP1)=F1(IP1)+FLOAT(J-1)*DEL(IP1)	27170
P1(J)=FNEW(IP1)	27180
DO 60 I=1,10	27190
FNEW(IP2)=F1(IP2)+FLOAT(I-1)*DEL(IP2)	27200
P2(I)=FNEW(IP2)	27210
CALL LSO(FNEW,SSS(I,J))	27220
IF(SSS(I,J).GE.SSMIN) GO TO 60	27230
SSMIN=SSS(I,J)	27240
ISAVE=I	27250
JSAVE=J	27260
60 IF(SSS(I,J).GT.SSMAX) SSMAX=SSS(I,J)	27270

003

FNEW(IP1)=P1(JSAVE)	27280
FNEW(IP2)=P2(JSAVE)	27290
STPNEM(IP1)=DEL(IP1)/2.	27300
STPNEM(IP2)=DEL(IP2)/2.	27310
D=(SSMAX-SSMIN)/9.	27320
DO 65 I=1,10	27330
65 A(I)=SSMIN+(I-1)*D	27340
A(10)=A(10)+0.01*D	27350
WRITE(6,1030)IP1,(I,P1(I),I=1,10)	27360
WRITE(6,1040)IP2,(I,P2(I),I=1,10)	27370
IF(ANS16.EQ.2HND) WRITE(6,1050)	27380
IF(ANS16.EQ.3HYES) WRITE(6,1060)	27390
WRITE(6,1070)IP1,IP2	27400
WRITE(6,1075)(I,I=1,10)	27410
DO 67 I=1,10	27420
WRITE(6,1080)(SSS(I,J),J=1,9,2)	27430
67 WRITE(6,1085)(SSS(I,J),J=2,10,2)	27440
DO 120 J=1,10	27450
DO 120 I=1,10	27460
IF(J.EQ.1) GO TO 70	27470
IF(J.EQ.10) GO TO 80	27480
PDEV(I,J)=SIGNOF(SSS(I,J+1)-SSS(I,J-1))	27490
GO TO 90	27500
70 PDEV(I,J)=SIGNOF(4.*SSS(I,J+1)-3.*SSS(I,J)-SSS(I,J+2))	27510
GO TO 90	27520
80 PDEV(I,J)=SIGNOF(-4.*SSS(I,J-1)+3.*SSS(I,J)+SSS(I,J-2))	27530
90 K=J+10	27540
IF(I.EQ.1) GO TO 100	27550
IF(I.EQ.10) GO TO 110	27560
PDEV(I,K)=SIGNOF(SSS(I+1,J)-SSS(I-1,J))	27570
GO TO 115	27580
100 PDEV(I,K)=SIGNOF(4.*SSS(I+1,J)-3.*SSS(I,J)-SSS(I+2,J))	27590
GO TO 115	27600
110 PDEV(I,K)=SIGNOF(-4.*SSS(I-1,J)+3.*SSS(I,J)+SSS(I-2,J))	27610
115 CONTINUE	27620
DO 120 L=1,9	27630
120 IF(SSS(I,J).GE.A(L).AND.SSS(I,J).LE.A(L+1)) CON(I,J)=AL(L)	27640
WRITE(6,1090)IP1,IP2	27650
WRITE(6,1100)(PDEV(I,J),J=1,20),I=1,10	27660
WRITE(6,1110)	27670
DO 130 I=1,10	27680
WRITE(6,1120)(CON(I,J),J=1,10)	27690
IF(I.EQ.10) GO TO 130	27700
WRITE(6,1130)I,A(I),A(I+1)	27710
130 CONTINUE	27720
RETURN	27730
135 NP=FIX(NPOINT*(1./FLOAT(NEWNOP))*.001)	27740
140 NEVAL=NP*NEWNOP	27750
IF(NEVAL.LE.2187) GO TO 150	27760
NP=NP-1	27770
GO TO 140	27780
150 CONTINUE	27790
IF(NOP.EQ.7) GO TO 170	27800
K=NOP+1	27810
DO 160 I=K,7	27820
F1(I)=0.	27830
160 F2(I)=0.	27840
170 CONTINUE	27850
DO 180 I=1,7	27860
180 DEL(I)=(F2(I)-F1(I))/FLOAT(NP-1)	27870
SSMIN=1.E98	27880
NEVAL=0	27890

P03

FNEW(I)=F1(I)	27050
STPNEM(I)=STEP(I)	27060
IF(I.GT.IP1) GO TO 55	27070
IF(F1(I).NE.F2(I)) IP1=I	27080
55 IF(F1(I).NE.F2(I)) IP2=I	27090
DEL(IP1)=(F2(IP1)-F1(IP1))/9.	27100
DEL(IP2)=(F2(IP2)-F1(IP2))/9.	27110
ISAVE=1	27120
JSAVE=1	27130
SSMIN=1.E99	27140
SSMAX=0.	27150
DO 60 J=1,10	27160
FNEW(IP1)=F1(IP1)+FLOAT(J-1)*DEL(IP1)	27170
P1(J)=FNEW(IP1)	27180
DO 60 I=1,10	27190
FNEW(IP2)=F1(IP2)+FLOAT(I-1)*DEL(IP2)	27200
P2(I)=FNEW(IP2)	27210
CALL LSO(FNEW,SSS(I,J))	27220
IF(SSS(I,J).GE.SSMIN) GO TO 60	27230
SSMIN=SSS(I,J)	27240
ISAVE=I	27250
JSAVE=J	27260
60 IF(SSS(I,J).GT.SSMAX) SSMAX=SSS(I,J)	27270

003

FNEW(IP1)=P1(JSAVE)	27280
FNEW(IP2)=P2(ISAVE)	27290
STPNEM(IP1)=DEL(IP1)/2.	27300
STPNEM(IP2)=DEL(IP2)/2.	27310
D=(SSMAX-SSMIN)/9.	27320
DO 65 I=1,10	27330
65 A(I)=SSMIN+(I-1)*D	27340
A(10)=A(10)+0.01*D	27350
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GO TO 90	27520
80 PDEV(I,J)=SIGNOF(-4.*SSS(I,J-1)+3.*SSS(I,J)+SSS(I,J-2))	27530
90 K=J+10	27540
IF(I.EQ.1) GO TO 100	27550
IF(I.EQ.10) GO TO 110	27560
PDEV(I,K)=SIGNOF(SSS(I+1,J)-SSS(I-1,J))	27570
GO TO 115	27580
100 PDEV(I,K)=SIGNOF(4.*SSS(I+1,J)-3.*SSS(I,J)-SSS(I+2,J))	27590
GO TO 115	27600
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115 CONTINUE	27620
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120 IF(SSS(I,J).GE.A(L).AND.SSS(I,J).LE.A(L+1)) CON(I,J)=AL(L)	27640
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IF(I.EQ.10) GO TO 130	27700
WRITE(6,1130)I,A(I),A(I+1)	27710
130 CONTINUE	27720
RETURN	27730
135 NP=IFIX(NPOINT*(1./FLOAT(NEWNOP))*.001)	27740
140 NEVAL=NP*NEWNOP	27750
IF(NEVAL.LE.2187) GO TO 150	27760
NP=NP-1	27770
GO TO 140	27780
150 CONTINUE	27790
IF(NOP.EQ.7) GO TO 170	27800
K=NOP+1	27810
DO 160 I=K,7	27820
F1(I)=0.	27830
160 F2(I)=0.	27840
170 CONTINUE	27850
DO 180 I=1,7	27860
180 DEL(I)=(F2(I)-F1(I))/FLOAT(NP-1)	27870
SSMIN=1.E98	27880
NEVAL=0	27890

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