### **Edith Cowan University**

## **Research Online**

Research outputs 2022 to 2026

3-1-2022

# Development of 1,2,4-oxadiazole antimicrobial agents to treat enteric pathogens within the gastrointestinal tract

Noel P. Pitcher

Jitendra R. Harjani

Yichao Zhao

Jianwen Jin

Daniel Knight Edith Cowan University

See next page for additional authors

Follow this and additional works at: https://ro.ecu.edu.au/ecuworks2022-2026



Part of the Medicine and Health Sciences Commons

#### 10.1021/acsomega.1c06294

Pitcher, N. P., Harjani, J. R., Zhao, Y., Jin, J., Knight, D. R., Li, L., ... & Baell, J. B. (2022). Development of 1, 2, 4-Oxadiazole Antimicrobial Agents to Treat Enteric Pathogens within the Gastrointestinal Tract. ACS omega, 7, 8, 6737-6759. https://doi.org/10.1021/acsomega.1c06294

This Journal Article is posted at Research Online.

https://ro.ecu.edu.au/ecuworks2022-2026/408

<b>Authors</b> Noel P. Pitcher, Jitendra R. Harjani, Yichao Zhao, Jianwen Jin, Daniel Knight, Lucy Li, Papanin Putsathit, Thomas V. Riley, Glen P. Carter, and Jonathan B. Baell	_





http://pubs.acs.org/journal/acsodf Article

## Development of 1,2,4-Oxadiazole Antimicrobial Agents to Treat Enteric Pathogens within the Gastrointestinal Tract

Noel P. Pitcher, Jitendra R. Harjani, Yichao Zhao, Jianwen Jin, Daniel R. Knight, Lucy Li, Papanin Putsathit, Thomas V. Riley, Glen P. Carter,\*, and Jonathan B. Baell\*, Daniel R. Knight, Lucy Li, Papanin Putsathit, Thomas V. Riley, Glen P. Carter,\*



Cite This: ACS Omega 2022, 7, 6737-6759



**ACCESS** I

III Metrics & More

Article Recommendations

s Supporting Information

ABSTRACT: Colonization of the gastrointestinal (GI) tract with pathogenic bacteria is an important risk factor for the development of certain potentially severe and life-threatening healthcare-associated infections, yet efforts to develop effective decolonization agents have been largely unsuccessful thus far. Herein, we report modification of the 1,2,4-oxadiazole class of antimicrobial compounds with poorly permeable functional groups in order to target bacterial pathogens within the GI tract. We have identified that the quaternary ammonium functionality of analogue 26a results in complete impermeability in Caco-2 cell monolayers while retaining activity against GI pathogens *Clostridioides difficile* and

SAR exploration with poorly permeable groups
28 Compounds

26a

C. difficile MIC - 8 µg/mL

Efm MIC - 8 µg/mL

Permeability across

Caco-2 cell

monolayers abolished

multidrug-resistant (MDR) Enterococcus faecium. Low compound recovery levels after oral administration in rats were observed, which suggests that the analogues may be susceptible to degradation or metabolism within the gut, highlighting a key area for optimization in future efforts. This study demonstrates that modified analogues of the 1,2,4-oxadiazole class may be potential leads for further development of colon-targeted antimicrobial agents.

#### INTRODUCTION

The gastrointestinal (GI) tract serves as an important site of colonization for many pathogenic bacteria prior to the initiation of infection. The "healthy" microbiota that resides in the GI tract is normally considered to offer "colonization resistance" against invading pathogens but this can be compromised by exposure to antimicrobials or in certain illnesses.<sup>1-3</sup> Such disruption can lead to the overgrowth of GI pathogens, which is a major risk factor in the development of several different healthcare-associated infections. 4-8 Among the most significant nosocomial pathogens that colonize the GI tract is Clostridioides difficile, which causes a range of colonic infections collectively referred to as C. difficile infections (CDI). CDI may lead to mild to severe diarrhoea and can lead to potentially fatal complications such as pseudomembranous colitis and toxic mega-colon. 9,10 Indeed, *C. difficile* has been classified as an urgent threat to public health by the United States Centers for Disease Control and Prevention (CDC).<sup>11</sup> Traditional first-line treatments including metronidazole and vancomycin are effective for most CDI cases but approximately 15-30% of patients will suffer a recurrence of infection after cessation of treatment. 12,13 More recently, fidaxomicin, a narrow spectrum macrocyclic antimicrobial from the tiacumicin family, has become a recommended treatment for CDI. This antimicrobial is associated with lower rates of recurrence than vancomycin or metronidazole. 14,15 Also, metronidazole is

no longer recommended as a first line therapy of CDI due to treatment failures in the  ${\rm US.}^{14}$ 

Another important nosocomial pathogen is Enterococcus faecium. 16 Colonization of the GI tract by E. faecium is recognized as a major risk factor for the onset of potentially life-threatening diseases such as bacteraemia and infective <sup>7</sup> E. faecium has a high propensity for the endocarditis.<sup>8,7</sup> development of antimicrobial resistance due to its malleable genome which has resulted in the emergence of multidrugresistant (MDR) strains, 18 most notably vancomycin-resistant E. faecium (VREfm). The CDC has declared VREfm a serious threat to human health<sup>11</sup> and it has been included in the 2017 World Health Organization priority pathogen list for the development of new antimicrobials, 19 which highlights the limited therapeutic options available for treating this pathogen. High rates of VREfm infection have rendered the traditional first-line treatment of vancomycin increasingly ineffective against E. faecium infections, with clinicians now increasingly using linezolid and daptomycin. 17 Reports documenting the emergence of linezolid and daptomycin resistance in VREfm

Received: November 8, 2021 Accepted: February 3, 2022 Published: February 18, 2022





Figure 1. (a) Reported antimicrobials of a new oxadiazole class, with leading compounds 1 and 75b; (b) modifications reported here, leading to 26a that exhibits potent anti-C. difficile and anti-VREfm activity. (a) Previous work by Chang and Mobashery et al. 26,28-30 (b) This work.

are therefore of great clinical concern. Furthermore, the development of decolonization agents that can be used to eradicate *E. faecium* from the gut has been an aim for several years without success.  $^{22-25}$ 

Despite exhibiting different clinical manifestations, the disease states caused by both of these pathogens arise following initial colonization of the GI tract. There is therefore an urgent need for novel antimicrobials that selectively target pathogens within the GI tract while maintaining the integrity of gut microbiota. Such agents would undoubtedly improve treatment outcomes for difficult cases of *C. difficile* and *E. faecium* infections.

The 1,2,4-oxadiazole antimicrobial compound class was reported in 2014 by Chang and Mobashery et al. and displayed potent activity against important Gram-positive pathogens in the ESKAPE panel. Analogue 1 emerged as a lead compound in the study with activity against Staphylococcus aureus, Enterococcus faecalis, and E. faecium, and this activity was maintained across multiple strains, including those exhibiting vancomycin resistance. Analogue 75b was another leading compound. Significantly, there was no activity observed against the Gram-negative bacteria in the ESKAPE group, potentially demonstrating a selectivity profile that may be sparing of important members of the GI tract microbiota. Chang and Mobashery et al. have advanced several analogues in the class with a focus on developing effective treatments for methicillin-resistant S. aureus (MRSA) (Figure 1a). 28–30

Here, we explore the utility of the 1,2,4-oxadiazole compound class for the treatment of GI pathogens by attaching substituents that are predicted to reduce permeability in the GI tract. The idea is that such compounds may therefore reach the colon in relatively higher concentrations (Figure 1b). In an important proof-of-concept, several analogues modified to be less Caco-2 permeable maintained antimicrobial activity. Quaternary ammonium derivative **26a** is of particular note with complete impermeability to Caco-2 cells and notable anti-C. difficile and anti-VREfm activity.

#### RESULTS AND DISCUSSION

Minimum Inhibitory Concentration and Time-Kill Assays of Lead Compound 1. The lead 1,2,4-oxadiazole

compound 1 was assessed by minimum inhibitory concentration (MIC) assay and exhibited a modest potency of 6  $\mu$ g/mL against *C. difficile*, comparable to that of vancomycin (2  $\mu$ g/mL). Compound 1 also showed concentration-dependent killing kinetics in time-kill assays and rapid bactericidal activity when using  $\geq$ 2 × MIC (12–48  $\mu$ g/mL), which was similar to the activity observed for the positive control compound benzalkonium chloride (a quaternary ammonium compound with known membrane lysing activity) and superior to the bacteriostatic activity of vancomycin (Figure 2).

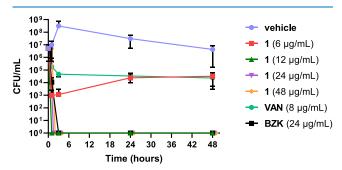


Figure 2. Time-kill analysis of 1 against *C. difficile* strain EDN0008. Blue circles are DMSO-containing untreated cultures (vehicle), red squares are cultures treated with 6  $\mu$ g/mL 1, green triangles are cultures treated with 12  $\mu$ g/mL 1, purple triangles are cultures treated with 24  $\mu$ g/mL 1, orange diamonds are cultures treated with 48  $\mu$ g/mL 1, turquoise circles are cultures treated with 8  $\mu$ g/mL vancomycin, and black squares are cultures treated with 24  $\mu$ g/mL benzalkonium chloride. Compounds were tested using three independent biological replicates with the data shown representing the mean  $\pm$  SEM. VAN—vancomycin, BZK—benzalkonium chloride, and CFU—colony-forming units.

Our previous findings demonstrated that compound 1 also displayed potent activity against a range of clinically relevant *E. faecium* strains, including both VREfm and daptomycinresistant strains. Compound 1 was not toxic against the human cell line HepG2 at the *E. faecium* MIC<sub>90</sub> (2  $\mu$ g/mL); however, reduced viability was observed at higher concentrations. This was deemed to represent a potentially narrow therapeutic window, but analogues of compound 1 have been

reported by Spink et al. in 2015 with activity against *S. aureus* and reduced toxicity against HepG2 cells. Thus, we hypothesized that optimization of the therapeutic index for this compound class would be highly feasible.

**General Strategy.** To address the high bioavailability reported for the 1,2,4-oxadiazole compound class, <sup>26</sup> we proposed that their physicochemical properties could be modified to reduce "drug-likeness", thereby decreasing systemic exposure and leading to accumulation in the colon. Amine and carboxylic acid functional groups that are charged at physiological pH were linked to the oxadiazole structure to induce ionization after oral administration and reduce compound permeability across the small intestine. <sup>32,33</sup> Indeed, similar approaches have shown success previously by utilizing the poorly permeable tetramic acid moiety to localize metronidazole in the GI tract. <sup>34</sup>

Suitable positions for linking to the oxadiazole structure needed to be carefully selected as to not decrease antimicrobial activity. It was noted in similar oxadiazole analogues reported for the treatment of MRSA<sup>28</sup> that trifluoromethyl groups at the 4 position of ring A had minimal effect on *in vitro* activity but improved oral bioavailability (Figure 3). Therefore, the

**Figure 3.** General structure of the oxadiazole class, consisting of four main aromatic rings (A-D). R' and R'' represent the positions explored to link poorly permeable groups.

trifluoromethyl group was excluded from our oxadiazole analogues and ring A was instead used to introduce the poorly permeable groups. Our structure—activity relationship (SAR) investigation probed all aromatic positions of ring A while exploring different linkers to attach the ionizable functional groups. The hydroxyl in compound 1 was maintained consistent for the analogues in the series as it was previously recognized that a hydrogen bond donor at the position was essential for activity against MRSA.<sup>28</sup>

Additionally, we explored replacement of the phenol with an aniline to probe modification of ring D while maintaining the critical hydrogen bond donor. Primary and secondary aniline groups were synthesized to investigate if substitution from ring D was tolerated and if it could offer an alternative site to link the poorly permeable groups.

**Synthetic Chemistry.** Modified oxadiazole analogues were synthesized as outlined in Scheme 1. Reagents 2a—h were used to prepare *t*-Bu protected carboxylic acid substrates 3a—h, followed by installation of a benzonitrile moiety using a known Ullmann-type reaction procedure. Hydroxylamine addition to the nitrile yielded amidoximes 6a—h which then underwent coupling, cyclization, and dehydration with benzoic acid 7 in a "one-pot" process to form the 1,2,4-oxadiazole ring C. An acetyl group was used as a transient protecting group to inhibit the ring D phenol from reacting during coupling and was

subsequently cleaved in situ at high temperature to reveal the free phenol in compounds 8a-h. Protecting group removal was achieved under acidic conditions to afford final analogues 9a-h.

A more concise scheme was also devised to introduce the derivatized phenols at the penultimate step, as outlined in Scheme 2. Hydroxylamine addition to 4-iodobenzonitrile 4 and then reaction with acyl chloride 11 yielded the key intermediate 12.<sup>26</sup> Optimized Ullmann-type reaction conditions were used to couple the derivatized phenols, during which the *tert*-butyldimethylsilyl (TBS) provided a transient protecting group that was cleaved in situ. The derivatized phenol reagents 13a–i were individually prepared from commercially available material as described in the Supporting Information. Deprotection of the boc groups under acidic conditions yielded final analogues 15a–i.

Aniline series analogues were synthesized as outlined in Scheme 3. Amine- and nitro-substituted aryl carboxylate substrates 17a—d were subjected to coupling, cyclization, and dehydration with amidoxime intermediate 18 to form the 1,2,4-oxadiazole ring C and afford analogues 19c and 19d.<sup>35</sup> Treatment with 4 M HCl in 1,4-dioxane then afforded compounds 20a and 20b. Compound 22 was synthesized *via* sodium hydride-mediated alkylation of intermediate 19a using 2-bromoethyl methyl ether, followed by protecting group removal. Alkylation of 20a using *tert*-butyl bromoacetate and subsequent acidic removal of the protecting group was undertaken to synthesize 24.

Dimethylamine and quaternary ammonium substituted analogues 25a, 25b, 26a, and 26b were synthesized from their corresponding primary amine analogues, as outlined in Scheme 4. Boc-deprotection reactions of 14e and 14h were undertaken using standard conditions followed by careful neutralization to remove the trifluoroacetic acid (TFA) salts of the amines while also avoiding removal of the acidic phenolic protons. Treatment with formaldehyde and NaBH(OAc)<sub>3</sub> was undertaken to convert the primary amine intermediates to the dimethylamine groups,<sup>36</sup> and subsequent reaction with iodomethane selectively yielded the quaternary ammonium analogues without phenol methylation.<sup>37</sup>

MIC and Time-Kill Assays of Oxadiazole Analogues. The antimicrobial activity of the modified oxadiazole analogues against C. difficile and E. faecium was evaluated using standardized broth microdilution MIC assays. The initial SAR exploration sought to identify the best position for substitution from ring A and which poorly permeable groups were suitable for maintaining potency. The attachment of certain amine and carboxylic acid groups was tolerated in terms of C. difficile activity but was dependent on linker length and aromatic position (Table 1). Carboxylic acid substituents with ethylene linkers 9b (4  $\mu$ g/mL) and 9d (8  $\mu$ g/mL) were favored from the 2 and 3 positions, displaying similar activities to analogue 1 (6  $\mu$ g/mL). However, a 4-fold loss in potency was observed for the same substituent in the 4 position (9g, 25  $\mu$ g/mL). Decreasing the linker length from ethyl (9b, 9d, and 9g) to methyl (9a, 9c, and 9f) was not favored with at least a 2-fold loss in potency against *C. difficile* for the carboxylic acid groups at all aromatic positions on ring A. The methyl ether linkers (9e and 9h) also caused a loss of activity compared with their corresponding ethyl linkers (9d and 9g), thereby demonstrating that additional hydrogen bond accepting or hydrophilic interactions were not favored from the benzylic site. The trend of a 2-fold increase in potency for the ethyl

Scheme 1. Synthesis for Modified Oxadiazole Analogues 9a-ha

"Reagents and conditions: (a) (i) 2-tert-butyl-1,3-diisopropylisourea, DMAP, DCM, rt, 96 h; (ii) 1,1'-carbonyldiimidazole, 1,8-diazabicyclo[5.4.0]-undec-7-ene, tert-butanol, DMF, 65 °C, 72 h; (iii) tert-butyl bromoacetate, K<sub>2</sub>CO<sub>3</sub>, DMF, rt, 16 h; (b) 4, CuI, Cs<sub>2</sub>CO<sub>3</sub>, N,N-dimethylglycine, 1,4-dioxane, 100 °C, 48 h; (c) NH<sub>2</sub>OH·HCl, Et<sub>3</sub>N, MeOH, 60 °C, 6 h; (d) 7, HOBt, EDC·HCl, DMF, rt for 16 h to 100 °C for 16 h; (e) TFA, DCM, rt, 8 h.

linkers (15b and 15e) compared with methyl (15a and 15d) was also observed for the amine substituents at both the 2 position and 4 position. Ethylamine substitution was favored at the 3 and 4 positions with analogues 15c (4  $\mu$ g/mL) and 15e (3  $\mu$ g/mL) giving slight increases in potency against *C. difficile* as compared to analogue 1 (6  $\mu$ g/mL).

The replacement of the phenol ring D with an unsubstituted aniline (20a, >50  $\mu g/mL$ ) was detrimental to *C. difficile* activity. Interestingly, the activity was able to be somewhat restored upon functionalization of the aniline with methyl (20b,  $8 \mu g/mL$ ), methoxyethyl (22,  $8 \mu g/mL$ ), acetic acid (24,  $13 \mu g/mL$ ), or conversion to nitro groups (19d,  $8 \mu g/mL$ ). The analogue bearing a nitro substituent (19d) had similar potency to the unsubstituted phenol analogue (1), and it was apparent that a hydrogen bond donor on ring D was not essential for activity against *C. difficile*.

Modification of the oxadiazole structure was less tolerated in terms of *E. faecium* activity, with analogue 1 remaining the most potent at 2  $\mu$ g/mL. Carboxylic acid substitution led to a complete loss of potency, regardless of the aromatic position or linker variant. The decrease in activity was less pronounced when attaching the amine substituents, with only a 2-fold loss in activity at the 4 position (15d, 15e, 4  $\mu$ g/mL) and a 4-fold loss at the 2 position (15a, 15b, 8  $\mu$ g/mL). Linking with methyl or ethyl groups had no effect on the activity for these analogues. There was a complete loss of potency for all analogues from the aniline series which suggested that the hydroxyl on ring D was important for activity against *E. faecium*.

The initial SAR exploration suggested that modification with amino groups could be made while maintaining antimicrobial activity. Specifically, analogue 15e with ethylamine substitution from the 4-position of ring A appeared to be suitable, exhibiting modest activity against both C. difficile (3  $\mu$ g/mL) and E. faecium (4  $\mu$ g/mL). Further, analogue 15e displayed rapid killing when assessed in time-kill assays, with a 100% kill of C. difficile at  $4 \times MIC$  (12  $\mu g/mL$ ) within 3 h and VREfm at  $2 \times MIC (8 \mu g/mL)$  within 1 h (Figure 4). This rapid rate of killing was comparable to the positive control benzalkonium chloride. When benchmarked against 1, we observed superior killing activity of 15e against C. difficile at 6  $\mu$ g/mL and indistinguishable kill kinetics at 12 µg/mL. Against VREfm, 15e displayed comparable killing kinetics to 1 at 4  $\mu$ g/mL despite the 2-fold loss of activity identified above, with both compounds killing 100% of VREfm within 24 h. Of note, while growth of VREfm remained undetectable after exposure to 4  $\mu g/mL$  of 15e for the duration of the assays, rebounding growth of VREfm was observed after 48 h in cultures exposed to 1. At a concentration of 8  $\mu$ g/mL, however, the killing kinetics of 1 and 15e were indistinguishable, with 100% of VREfm being killed within 1 h of exposure and no rebounding growth observed for either compound.

1 has previously been shown to possess membrane-lytic activity, with concentrations in excess of  $64 \,\mu\text{g/mL}$  leading to haemolysis in human erythrocytes.<sup>31</sup> To determine whether 15e displayed a similar membrane-lytic activity as 1, the compound was tested against human erythrocytes as before (Figure 4c). Limited haemolysis was observed following incubation of human erythrocytes with  $4 \,\mu\text{g/mL}$  15e

Scheme 2. Synthesis for Modified Oxadiazole Analogues 15a-i

"Reagents and conditions: (a) NH<sub>2</sub>OH·HCl, Et<sub>3</sub>N, MeOH, 60 °C, 6 h; (b) 11 (synthesis previously described<sup>26</sup>), toluene, 120 °C, 27 h; (c) 13a—i, 1,3-diphenyl-1,3-propanedione, Cs<sub>2</sub>CO<sub>3</sub>, copper(II) acetylacetonate, DMSO, 90 °C, 24 h; (d) 4 M HCl/1,4-dioxane, rt, 7 h

HCL HN

(approximate MIC of *C. difficile* and *E. faecium*), with only 2.5% (range: 0–7%) of erythrocytes undergoing lysis. As previously observed for 1, increasing concentrations of 15e resulted in increased levels of hemolysis in a dose-dependent manner, with a concentration of  $\geq$ 64  $\mu$ g/mL leading to substantial levels of haemolysis. These data therefore provide evidence to show that 15e most likely operates through a membrane-lysing mechanism.

22%

Further SAR studies of analogue 15e were undertaken to identify the optimal linker for use and explore functionalization of the amine (Table 2). Against C. difficile, the introduction of a branched methyl group at the benzylic position of the linker (15f, 16  $\mu$ g/mL) saw a 5-fold loss in potency compared with the ethyl linker (15e, 3  $\mu$ g/mL), which suggested that flexibility of the linkers may be important for activity. Conversely, activity was retained when further restricting the linker flexibility by adding an N-methylene bridge to the 3 position of ring A to form a tetrahydroisoquinoline moiety (15i, 4  $\mu$ g/mL). Methylation of the ethylamine group to form both secondary (15g) and tertiary amines (25a) caused slight decreases in activity to 4  $\mu$ g/mL. The observation that the dimethylamine group retained activity suggested that favorable interactions were made via the amine basicity or a hydrogen bond acceptor effect, rather than through hydrogen bond donating. The conversion of the amine group into a quaternary ammonium (26a, 8  $\mu$ g/mL) was also tolerated but with another slight drop in C. difficile activity observed. Nonetheless,

the analogue was of particular interest because it represented an active compound that also possessed a permanently cationic group which would greatly reduce permeability across the small intestine.<sup>32,33</sup> A further look at modifying the linker identified that a slight increase in activity against *C. difficile* was obtained when the length was increased to a propyl (15h, 2  $\mu$ g/mL). Unfortunately, this increase in potency was not maintained when forming the tertiary amine 25b (16  $\mu$ g/mL) or quaternary ammonium 26b (8  $\mu$ g/mL).

The SAR for *E. faecium* was generally similar to the trends observed for changes in activity against *C. difficile*. Restriction of the linkers in analogues **15f** and **15i** had no effect on *E. faecium* activity. Methylation of the ethylamine to form secondary and tertiary amines **15g** and **25a** also saw no change in potency, and quaternization (**26a**, 8  $\mu$ g/mL) caused only a 2-fold decrease in activity compared with the primary amine **15e** (4  $\mu$ g/mL). Extension of the linker to a propylamine (**15h**, 2  $\mu$ g/mL) gave a 2-fold increase in activity; however, again this increased activity was not translated when forming the quaternary ammonium (**26b**, 16  $\mu$ g/mL).

**SAR Summary.** Modification of the oxadiazole analogues with amine substituents was generally more favored than the carboxylic acid groups. Exploring changes to the linkers indicated that while different variations were tolerated, they did not lead to significant increases in activity. With the discovery of analogue **26a**, we identified that a poorly permeable quaternary ammonium group could be linked to

#### Scheme 3. Synthesis of Aniline Series Analogues<sup>a</sup>

<sup>a</sup>Reagents and conditions: (a) Boc<sub>2</sub>O, NaHCO<sub>3</sub>, MeOH/H<sub>2</sub>O, rt, 24 h; (b) **18** (synthesis previously described<sup>26</sup>), HOBt, EDC·HCl, DMF, rt for 16 h to 100 °C for 16 h; (c) 4 M HCl/1,4-dioxane, rt, 6 h; (d) NaH (60% in mineral oil), DMF, 0 °C to rt, 1 h, 1-bromo-2-methoxyethane, 0 °C to rt, 2 h; (e) *tert*-butyl bromoacetate, DIPEA, acetonitrile, 80 °C, 48 h; (f) TFA, DCM, rt, 8 h

#### Scheme 4. Synthesis of Methylated-Amine Substituted Analogues<sup>a</sup>

"Reagents and conditions: (a) TFA, DCM, rt, 7 h; (b) 37% aq formaldehyde, NaBH(OAc)<sub>3</sub>, acetonitrile/H<sub>2</sub>O, rt, 50 min; (c) MeI, DCM/MeOH, rt, 24 h.

Table 1. Structure and Activity of Oxadiazole Analogues

Entry	R	C. difficile MIC (μg/mL) <sup>a</sup>	VREfm MIC (μg/mL) <sup>a</sup>	
	$\begin{array}{c} R \\ \\ \end{array} \begin{array}{c} \\ \\ N \\ \end{array} \begin{array}{c} \\ \\ \end{array} \begin{array}{c} \\ \\ \end{array} \begin{array}{c} \\ \\ \\ \\ \\ \\ \\ \end{array} \begin{array}{c} \\ \\ \\ \\ \\ \\ \\ \end{array} \begin{array}{c} \\ \\ \\ \\ \\ \\ \\ \end{array} \begin{array}{c} \\ \\ \\ \\ \\ \\ \\ \end{array} \begin{array}{c} \\ \\ \\ \\ \\ \\ \\ \end{array} \begin{array}{c} \\ \\ \\ \\ \\ \\ \\ \\ \end{array} \begin{array}{c} \\ \\ \\ \\ \\ \\ \\ \\ \end{array} \begin{array}{c} \\ \\ \\ \\ \\ \\ \\ \\ \\ \end{array} \begin{array}{c} \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \end{array} \begin{array}{c} \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \end{array} \begin{array}{c} \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \end{array} \begin{array}{c} \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\$			
1	Н	6	2	
9a	2-CH₂COOH	>50	>32	
9b	2-CH <sub>2</sub> CH <sub>2</sub> COOH	4	>32	
9c	3-CH <sub>2</sub> COOH	50	>32	
9d	3-CH <sub>2</sub> CH <sub>2</sub> COOH	8	>32	
9e	3-OCH <sub>2</sub> COOH	50	>32	
9f	4-CH <sub>2</sub> COOH	50	>32	
9g	4-CH <sub>2</sub> CH <sub>2</sub> COOH	25	>32	
9h	4-OCH <sub>2</sub> COOH	50	>32	
	HCI.R—OH	16		
	15a 2-CH <sub>2</sub> NH <sub>2</sub>		8	
15b	$2-CH_2CH_2NH_2$	8	8	
15c	$3-CH_2CH_2NH_2$	4	n.d.	
	4-CH2NH2		4	
15e	4-CH <sub>2</sub> CH <sub>2</sub> NH <sub>2</sub>	3	4	
	O O PR			
19c	-NHAc	>50	>32	
19d	$-NO_2$	8	>32	
20a	-NH <sub>2</sub>	>50	>32	
20b	-NHCH <sub>3</sub>	8	n.d.	
22	-NHCH <sub>2</sub> CH <sub>2</sub> OCH <sub>3</sub>	8	>32	
24	-NHCH₂COOH	13	n.d.	
Vancomycin		0.5 - 2	n.d.	
Daptomycin		n.d.	2	

an.d.—not determined.

the oxadiazole structure while retaining moderate MICs of 8  $\mu$ g/mL for both *C. difficile* and *E. faecium*.

In Vitro Permeability Studies. We sought to measure the effect that each modification had on compound permeability by using differentiated Caco-2 cell monolayers. Four analogues were assessed to represent the different types of ionizable groups and linkers (Table 3). Analogues 9e and 9g bearing carboxylic acid groups displayed moderate permeability with  $P_{\text{app}}$  (10<sup>-6</sup> cm/s) values of 5.3 ± 0.20 and 18 ± 1.0. Measurable concentrations of the primary amine-substituted analogue 15e could not be detected in the acceptor chamber, prohibiting the determination of a  $P_{\rm app}$  value. However, the low mass balance indicated that the analogue was likely being retained in the Caco-2 cell monolayer rather than having intrinsically low permeability.<sup>38</sup> Quaternary ammoniumsubstituted analogue 26a could also not be detected in the acceptor chamber but was fully recovered in the donor chamber and therefore displayed no detectable permeability across the Caco-2 cell monolayer. This finding strongly suggested that the quaternary ammonium-linked analogue would be ideally suited to exhibit minimal absorption across the small intestine.

In Vivo Pharmacokinetic Studies. To investigate compound exposure to the colon, the *in vivo* pharmacokinetic properties of analogues 15e and 26a were assessed after oral administration in healthy male Sprague Dawley rats (Table 4). Importantly, there were no adverse effects observed in the rats as a result of oxadiazole compound administration. Plasma concentrations of the oxadiazole analogues were measurable

but low after oral administration, with  $AUC_{0-inf}$  values of only 0.90 and 0.28 h\* $\mu$ M reached by analogues 15e and 26a, respectively. However, the cumulative recoveries of unchanged compounds in the faeces were also very low for both compounds. Of the oral doses, only 0.42% of 15e and 1.5% of 26a were recovered in the faeces over a 48 h period and even less detected in the urine. Based on the poor recovery levels, it was thought that the compounds may have been degraded or metabolized in the GI lumen by chemical or enzymatic processes.

#### CONCLUSIONS

The synthesis and evaluation of colon-targeted antimicrobial agents were accomplished via the attachment of poorly permeable functional groups to the 1,2,4-oxadiazole class of compounds. Modified oxadiazole analogues were produced which displayed antimicrobial activity against two important Gram-positive pathogens. Rapid bactericidal activity was a hallmark feature of these analogues. Our study identified that modification with the quaternary ammonium moiety at ring A of the oxadiazole scaffold led to a suitable physicochemical profile for colon-targeted delivery, as these compounds were completely impermeant toward Caco-2 cell monolayers yet maintained good antimicrobial potency. The recovery of analogues 15e and 26a after oral administration in rats was low and this was identified as a key target for future optimization in order to progress the analogues toward exhibiting in vivo efficacy. Analysis of the resulting metabolites may be a reasonable approach toward identifying the structural

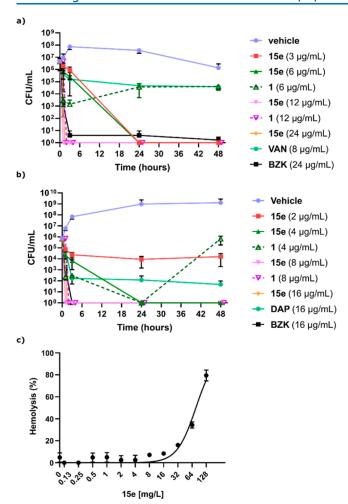


Figure 4. Time-kill analyses of 15e against GI pathogens: (a) C. difficile strain EDN0008, blue circles are DMSO-containing untreated cultures (vehicle), red squares are cultures treated with 3  $\mu$ g/mL 15e, closed green triangles are cultures treated with 6  $\mu$ g/mL 15e, open green triangles are cultures treated with 6  $\mu$ g/mL 1, closed purple triangles are cultures treated with 12  $\mu$ g/mL 15e, open purple triangles are culture treated with 12  $\mu$ g/mL 1, orange diamonds are cultures treated with 24  $\mu$ g/mL 15e, turquoise circles are cultures treated with 8 µg/mL vancomycin, and black squares are cultures treated with 24 µg/mL benzalkonium chloride; (b) E. faecium strain Aus0085, blue circles are DMSO-containing untreated cultures (vehicle), red squares are cultures treated with 2  $\mu$ g/mL 15e, closed green triangles are cultures treated with 4  $\mu$ g/mL 15e, open green triangles are cultures treated with 4  $\mu$ g/mL 1, closed purple triangles are cultures treated with 8  $\mu$ g/mL 15e, open purple triangles are cultures treated with 8  $\mu$ g/mL 1, orange diamonds are cultures treated with 16  $\mu$ g/mL 15e, turquoise circles are cultures treated with 16  $\mu$ g/ mL daptomycin, and black squares are cultures treated with 16  $\mu$ g/ mL benzalkonium chloride; (c) haemolysis assays of 15e against human erythrocytes. All assays were performed using three independent biological replicates with the data shown representing the mean ± SEM. VAN—vancomycin, DAP—daptomycin, BZK benzalkonium chloride, and CFU-colony-forming units.

changes needed to avoid such metabolism or degradation. In summary, we believe our results contribute important data toward the future development of decolonization agents with utility against difficult-to-treat infections caused by *C. difficile* and *E. faecium*.

#### EXPERIMENTAL SECTION

General Chemistry. Reactions were monitored by analytical thin-layer chromatography (TLC) using silica gel 60/F254 pre-coated aluminium sheets (0.25 mm, Merck). Flash column chromatography was performed with silica gel 60, 0.63–0.20 mm (70–230 mesh, Merck). The solvents; ethyl acetate (EtOAc); dichloromethane (DCM); dimethyl formamide (DMF); 1,4-dioxane; dimethyl sulfoxide (DMSO); methanol (MeOH); tetrahydrofuran; and diethyl ether (Et<sub>2</sub>O) were of analytical grade or distilled laboratory grade. <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded at 400 and 100 Hz, respectively, using an Avance III Nanobay 400 MHz Bruker spectrometer coupled to the BACS 60 automatic sample changer. Chemical shifts  $(\delta, ppm)$  are reported relative to the solvent peak [CDCl<sub>3</sub>: 7.26 (<sup>1</sup>H NMR), 77.16 (<sup>13</sup>C NMR), DMSO-d<sub>6</sub>: 2.50 (<sup>1</sup>H NMR), 39.52 (<sup>13</sup>C NMR), methanol-d<sub>4</sub>: 3.31 (<sup>1</sup>H NMR), 49.00 (<sup>13</sup>C NMR)]. Each proton resonance was assigned with the following annotations; chemical shift ( $\delta$ , ppm), singlet (s), doublet (d), triplet (t), quartet (q), multiplet (m), broad signal (br), coupling constant (J, Hz), and number of protons. Analytical HPLC was performed on an Agilent 1260 Infinity analytical HPLC coupled with a G1322A degasser, G1312B binary pump, G1367E high-performance autosampler, and G4212B diode array detector. The conditions were Zorbax Eclipse Plus C18 Rapid Resolution column (4.6 × 100 mm) with UV detection at 254 and 214 nm, 30 °C; the sample was eluted using a gradient system. Solvent A: water 0.1% TFA. Solvent B: acetonitrile 0.1% TFA. PP-gradient method—gradient: 5-95% B over 9 min and 100% B over 1 min. Detection: 254 or 214 nm. Hydrophobic PP method—gradient: 5-80% B over 0.1 min, 80-100% B from 0.6 to 9 min, and 100% B for 1 min. The PP-gradient method was used for all characterization unless otherwise specified. LCMS was obtained on an Agilent 6100 Series Single Quad LC/MS coupled with an Agilent 1200 Series HPLC, 1200 Series G1311A quaternary pump, 1200 series G1329A thermostatted autosampler and 1200 series G1314B variable wavelength detector. The liquid chromatography conditions were reverse-phase HPLC analysis fitted with a Phenomenex Luna C8(2) 5  $\mu$ m (50 × 4.6 mm) 100 Å column; column temperature: 30 °C; injection volume: 5  $\mu$ L; solvent: 99.9% acetonitrile, 0.1% formic acid; gradient: 5-100% of solvent over 10 min; detection: 254 nm. The mass spectrometry conditions were quadrupole ion source; ion mode: multimode-ES; drying gas temp: 300 °C; vaporizer temperature: 200 °C; capillary voltage: 2000 V (positive), 4000 (negative); scan range:  $100-1000 \ m/z$ ; step size: 0.1 s Acquisition time: 10 min. HRMS were obtained on an Agilent 6224 TOF LC/MS mass spectrometer coupled to an Agilent 1290 Infinity (Agilent, Palo Alto, CA). All data were acquired, and reference mass was corrected via a dual-spray electrospray ionization (ESI) source. Each scan or data point on the total ion chromatogram (TIC) is an average of 13,700 transients, producing a spectrum every second. Mass spectra were created by averaging the scans across each peak and background subtracted against the first 10 s of the TIC. Acquisition was performed using the Agilent Mass Hunter Data Acquisition software version B.05.00 Build 5.0.5042.2 and analysis was performed using MassHunter Qualitative Analysis version B.05.00 Build 5.0.519.13. Mass spectrometer conditions: ionization mode: ESI. Drying gas flow: 11 L/min; nebulizer: 45 psi; drying gas temperature: 325 °C; capillary voltage

Table 2. Structure and Activity of Oxadiazole Analogues

Entry	Structure or R  C. dij		VREfm MIC (μg/mL) <sup>a</sup>
	HCI.R N-O		
15f	4-CH(CH <sub>3</sub> )CH <sub>2</sub> NH <sub>2</sub>	16	4
15g	4-CH <sub>2</sub> CH <sub>2</sub> NHCH <sub>3</sub>	4	4
15h	4-CH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> NH <sub>2</sub>	2	2
15i	15i HN O NO OH		4
	R—N—OH		
25a	4-CH <sub>2</sub> CH <sub>2</sub> N(CH <sub>3</sub> ) <sub>2</sub>	4	4
25b	4-CH2CH2CH2N(CH3)2	16	n.d.
26a	$4-CH_2CH_2N^+(CH_3)_3$	8	8
26b	4-CH2CH2CH2N+(CH3)3	8	16
Vancomycin		0.5 - 2	n.d.
Daptomycin		n.d.	2

an.d.—not determined.

Table 3. Caco-2 Cell Permeability Assay of Key Oxadiazole Analogues

Entry	R	A-B P <sub>app</sub> (10 <sup>-6</sup> cm/s) <sup>a</sup>	Mass balance (%) <sup>a</sup>
	$R \longrightarrow 0$ $N \longrightarrow 0$ $N \longrightarrow 0$ $N \longrightarrow 0$		
9e	3-OCH <sub>2</sub> COOH	$5.3 \pm 0.20$	$96 \pm 5.3$
9g	4-CH <sub>2</sub> CH <sub>2</sub> COOH	$18 \pm 1.0$	$55 \pm 1.7$
15e	4-CH <sub>2</sub> CH <sub>2</sub> NH <sub>2</sub>	c.n.d.	$13 \pm 2.1$
26a	$4-CH_2CH_2N^+(CH_3)_3$	c.n.d.	$105 \pm 7.6$

<sup>&</sup>lt;sup>a</sup>Values are presented as the mean  $\pm$  SD of n=3 transwells. C.n.d.—could not be determined.

Table 4. Plasma Exposure Parameters and Urinary and Faecal Recovery from Rats of Unchanged Analogues after Oral Administration

entry	$t_{1/2} (h)^a$	$C_{\text{max}} (\mu M)^a$	$T_{\text{max}} (h)^a$	$AUC_{0-inf} (h*\mu M)^a$	% dose in urine $(0-48 \text{ h})^a$	% dose in faeces $(0-48 \text{ h})^a$
15e	$2.0 \pm 0.30$	$0.16 \pm 0.038$	$2.5 \pm 0.0$	$0.90 \pm 0.22$	$0.08 \pm 0.03$	$0.42 \pm 0.47$
26a	$5.3 \pm 2.1$	$0.039 \pm 0.046$	$2.8 \pm 2.0$	$0.28 \pm 0.27$	$0.06 \pm 0.03$	$1.5 \pm 0.98$
<sup>a</sup> Values are presented as the mean $\pm$ SD of $n = 3$ . Measured dose—30 mg/kg.						

 $(V_{\rm cap})$ : 4000 V; fragmentor: 160 V; skimmer: 65 V; OCT RFV: 750 V; and scan range acquired: 100–1500 m/z. Internal reference ions: positive ion mode = m/z = 121.050873 & 922.009798. Chromatographic separation was performed using an Agilent Zorbax SB-C18 Rapid Resolution HT 2.1 × 50 mm, 1.8  $\mu$ m column (Agilent Technologies, Palo Alto, CA) using an acetonitrile gradient (5–100%) over 3.5 min at 0.5 mL/min. Solvent A = aqueous 0.1% formic acid. Solvent B = acetonitrile/0.1% formic acid. Compounds **6a**–**h** were obtained as unpurified intermediates and were used directly in subsequent reactions. Therefore, these intermediates were not fully characterized. Characterization and syntheses of the

derivatized phenol reagents 13a–i in Scheme 2 are described in the Supporting Information. The purity of all compounds submitted for biological testing was  $\geq$ 95% as determined using HPLC analysis.

General Procedure A. A solution of phenol (1.2 equiv) in DCM (0.27 M) was treated with 2-tert-butyl-1,3-diisopropylisourea (1 equiv) and a catalytic amount of DMAP and then stirred at rt for 96 h. The reaction mixture was concentrated in vacuo, diluted with a 10% solution of aq Na<sub>2</sub>CO<sub>3</sub>, and then, Et<sub>2</sub>O was added to extract the product. The separated organic layer was dried over anhydrous MgSO<sub>4</sub> and concentrated in vacuo. The residue was purified using flash chromatography

eluting with 15% EtOAc in petroleum ether to yield the product.

**General Procedure B.** Phenol (1 equiv), 4-iodobenzonitrile (1 equiv),  $Cs_2CO_3$  (2 equiv), and N,N-dimethylglycine (0.3 equiv) were dissolved in 1,4-dioxane (0.59 M) in a sealed tube and purged with  $N_2$  for 15 min. CuI (0.2 equiv) was then added, the reaction vessel was sealed, and the solution was stirred at 100 °C for 48 h. The reaction mixture was filtered, washed with  $Et_2O$ , and the collected filtrate was evaporated in vacuo. The residue was purified using flash chromatography eluting with 2–15% EtOAc in petroleum ether to yield the product.

**General Procedure C.**  $NH_2OH \cdot HCl$  (2 equiv) and  $Et_3N$  (2 equiv) were added to a solution of benzonitrile (1 equiv) in MeOH (0.2 M). The solution was heated to 60 °C and allowed to stir for 6 h. The solvent was removed in vacuo, and the residue was diluted with water and then extracted with DCM. The organic layers were combined, dried over  $MgSO_4$ , and the solvent removed in vacuo to yield the product that was used in the subsequent step without further purification.

General Procedure D. A solution of amidoxime (1 equiv) in DMF (0.22 M) was treated with 4-acetoxybenzoic acid 7 (1.5 equiv), HOBt (2 equiv), and EDC·HCl (2 equiv) then the solution was stirred at rt for 16 h. The temperature was increased to 100 °C, and the stirring was continued for 16 h. The reaction mixture was concentrated in vacuo, diluted with a 10% solution of aq Na<sub>2</sub>CO<sub>3</sub>, and then, DCM was added to extract the product. The separated organic layer was dried over anhydrous MgSO<sub>4</sub> and concentrated in vacuo. The residue was purified using flash chromatography eluting with 0–15% EtOAc and 1% acetic acid in petroleum ether to yield the product.

**General Procedure E.** A solution of the *tert*-butyl ester substrate in DCM (1 mL per 100 mg substrate) was treated with TFA (0.1 mL per 100 mg substrate) and stirred at rt for 8 h. The reaction mixture was concentrated in vacuo, and the remaining residue suspended in water and sonicated. The solid was filtered, washed with water, and air-dried to yield the pure product.

4-(3-(4-Phenoxyphenyl)-1,2,4-oxadiazol-5-yl)phenol (1). The title compound was synthesized as previously reported to yield the product as a yellow solid (287 mg, 84%). <sup>26</sup>  $^{1}$ H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.13–8.07 (m, 4H), 7.41–7.35 (m, 2H), 7.17 (t, J = 7.4 Hz, 1H), 7.12–7.06 (m, 4H), 7.01–6.96 (m, 2H); spectroscopic data consistent with the literature. <sup>26</sup>

tert-Butyl 2-(2-Hydroxyphenyl)acetate (3a). The title compound was synthesized from 2-(2-hydroxyphenyl)acetic acid 2a (9.12 mmol, 1.39 g) according to general procedure A and obtained as a yellow solid (800 mg, 42%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.00 (br, 1H), 7.19 (td, J = 7.9, 1.7 Hz, 1H), 7.07 (dd, J = 7.5, 1.4 Hz, 1H), 6.96 (dd, J = 8.1, 1.0 Hz, 1H), 6.87 (td, J = 7.4, 1.2 Hz, 1H), 3.60 (s, 2H), 1.47 (s, 9H); spectroscopic data consistent with the literature.<sup>39</sup>

tert-Butyl 3-(2-Hydroxyphenyl)propanoate (3b). 3-(2-Hydroxyphenyl) propionic acid 2b (1 equiv, 18.1 mmol, 3.00 g) was dissolved in DMF (16 mL), and 1,1'-carbonyldiimidazole (1 equiv, 18.1 mmol, 2.93 g) was added carefully. The solution was stirred for 2 h at 40 °C. 1,8-Diazabicyclo[5.4.0]undec-7-ene (2 equiv, 36.1 mmol, 5.40 mL) and anhydrous tert-butanol (2.5 equiv, 45.2 mmol, 4.30 mL) were added, and the solution was stirred at 65 °C for 3 days. Solvents were removed in vacuo, and the residue was

diluted with water and aq 2 M HCl solution. The aqueous layer was washed with EtOAc, then the organic layers were combined, dried over MgSO<sub>4</sub>, and the solvent was removed in vacuo. The residue was purified using flash chromatography (10% EtOAc in petroleum ether) to yield the product as light yellow oil (804 mg, 20%).  $^1\mathrm{H}$  NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.54 (br, 1H), 7.14–7.04 (m, 2H), 6.93–6.83 (m, 2H), 2.90–2.81 (m, 2H), 2.66–2.61 (m, 2H), 1.42 (s, 9H); spectroscopic data consistent with the literature.

tert-Butyl 2-(3-Hydroxyphenyl)acetate (3c). The title compound was synthesized from 2-(3-hydroxyphenyl)acetic acid 2c (15.9 mmol, 2.42 g) according to general procedure A and obtained as clear oil (0.796 g, 29%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.16 (t, J = 7.8 Hz, 1H), 6.81 (d, J = 7.4 Hz, 1H), 6.74 (s, 1H), 6.72 (d, J = 8.1 Hz, 1H), 5.47 (br, 1H), 3.47 (s, 2H), 1.44 (s, 9H); spectroscopic data consistent with the literature. <sup>41</sup>

tert-Butyl 3-(3-Hydroxyphenyl)propanoate (**3d**). 3-(3-Hydroxyphenyl) propionic acid 2d (1 equiv, 18.1 mmol, 3.00 g) was dissolved in DMF (16 mL) and 1,1'-carbonyldiimidazole (1 equiv, 18.1 mmol, 2.93 g) was added carefully. The solution was stirred for 2 h at 40 °C. 1,8-Diazabicyclo [5.4.0] undec-7-ene (2 equiv, 36.1 mmol, 5.40 mL) and anhydrous tert-butanol (2.5 equiv, 45.2 mmol, 4.30 mL) were added, and the solution was stirred at 65 °C for 3 days. Solvents were removed in vacuo, and the residue was diluted with water and aq 2 M HCl solution. The aqueous layer was washed with EtOAc, then the organic layers were combined, dried over MgSO<sub>4</sub>, and the solvent was removed in vacuo. The residue was purified using flash chromatography (10% EtOAc in petroleum ether) to yield the product as clear oil (1.95 g, 49%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  9.26 (br, 1H), 7.04 (t, J = 7.7 Hz, 1H), 6.64–6.53 (m, 3H), 2.70 (t, J =7.5 Hz, 2H), 2.45 (t, J = 7.5 Hz, 2H), 1.36 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 172.5, 155.8, 142.8, 129.7, 120.9, 115.4, 113.2, 80.6, 37.0, 31.1, 28.2. HPLC:  $t_R = 6.6 \text{ min}$ , 99% at 254 nm. HRMS m/z: calcd for  $2 \times (C_{13}H_{18}O_3) - H^+$  [2M - H<sup>+</sup>], 443.2439; found, 443.2440.

tert-Butyl 2-(3-Hydroxyphenoxy)acetate (3e). A solution of resorcinol 2e (50 mmol, 5.50 g, 5 equiv) in DMF (50 mL) was treated with anhydrous K<sub>2</sub>CO<sub>3</sub> (50 mmol, 6.91 g, 5 equiv) and tert-butyl bromoacetate (10 mmol, 1.48 mL, 1 equiv); then, the solution was stirred at rt for 16 h. The reaction mixture was concentrated in vacuo, and a 6 M solution of aq HCl was added while stirring until the solution became until acidic. DCM was added to extract the product, and the separated organic layer was dried over anhydrous MgSO<sub>4</sub> and then concentrated in vacuo. The residue was purified using flash chromatography (25% EtOAc in petroleum ether) to yield the product as orange oil (1.18 g, 53%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.10 (t, J = 8.1 Hz, 1H), 6.50–6.35 (m, 3H), 5.76 (br, 1H), 4.48 (s, 2H), 1.49 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>):  $\delta$  168.6, 159.2, 157.2, 130.2, 109.1, 106.4, 102.8, 82.9, 65.8, 28.2. HPLC:  $t_{\rm R}$  = 4.8 min, 99% at 254 nm. HRMS m/z: calcd for  $C_{12}H_{16}O_4 + H^+ [M + H^+]$ , 225.1121; found, 225.1119.

tert-Butyl 2-(4-Hydroxyphenyl)acetate (3f). The title compound was synthesized from 2-(4-hydroxyphenyl)acetic acid 2f (24 mmol, 3.65 g) according to general procedure A and obtained as a white solid (1.15 g, 28%).  $^1$ H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.12–7.08 (m, 2H), 6.75–6.70 (m, 2H), 5.33 (br, 1H), 3.45 (s, 2H), 1.44 (s, 9H); spectroscopic data consistent with the literature.  $^{39}$ 

tert-Butyl 3-(4-Hydroxyphenyl)propanoate (3*g*). The title compound was synthesized from 3-(4-hydroxyphenyl)-propanoic acid 2*g* (24.0 mmol, 3.98 g) according to general procedure A and obtained as yellow oil (1.17 g, 26%).  $^{1}$ H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.05 (d, J = 8.4 Hz, 2H), 6.76–6.70 (m, 2H), 5.31 (br, 1H), 2.83 (t, J = 7.7 Hz, 2H), 2.50 (t, J = 7.7 Hz, 2H), 1.42 (s, 9H); spectroscopic data consistent with the literature.  $^{40}$ 

tert-Butyl 2-(4-Hydroxyphenoxy)acetate (**3h**). A solution of hydroquinone 2h (50 mmol, 5.50 g, 5 equiv) in DMF (50 mL) was treated with anhydrous K<sub>2</sub>CO<sub>3</sub> (50 mmol, 6.91 g, 5 equiv) and tert-butyl bromoacetate (10 mmol, 1.48 mL, 1 equiv); then, the solution was stirred at rt for 16 h. The reaction mixture was concentrated in vacuo and a 6 M solution of aq HCl was added while stirring until the solution became until acidic. DCM was added to extract the product, and the separated organic layer was dried over anhydrous MgSO<sub>4</sub> and then concentrated in vacuo. The residue was purified using flash chromatography eluting with 25% EtOAc in petroleum ether to yield the product as an off-white amorphous solid (1.75 g, 78%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  8.96 (br, 1H), 6.75–6.69 (m, 2H), 6.68–6.63 (m, 2H), 4.49 (s, 2H), 1.41 (s, 9H); spectroscopic data consistent with the literature.<sup>42</sup>

tert-Butyl 2-(2-(4-Cyanophenoxy)phenyl)acetate (5a). The title compound was synthesized from tert-butyl 2-(2-hydroxyphenyl)acetate 3a (2.89 mmol, 600 mg) according to general procedure B and purified using flash chromatography (15% EtOAc in petroleum ether) to yield the product as clear oil (454 mg, 51%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.60–7.56 (m, 2H), 7.36 (dd, J = 7.5, 1.5 Hz, 1H), 7.31 (td, J = 7.8, 1.8 Hz, 1H), 7.21 (td, J = 7.5, 1.3 Hz, 1H), 7.00–6.95 (m, 3H), 3.52 (s, 2H), 1.33 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 170.3, 161.6, 153.0, 134.2, 132.2, 129.1, 127.7, 125.6, 120.7, 119.0, 117.8, 105.9, 81.2, 37.1, 28.1. HPLC: t<sub>R</sub> = 6.3 min, 99% at 254 nm. HRMS m/z: calcd for C<sub>19</sub>H<sub>19</sub>NO<sub>3</sub> + H<sup>+</sup> [M + H<sup>+</sup>], 310.1438; found, 310.1433.

tert-Butyl 3-(2-(4-Cyanophenoxy)phenyl)propanoate (5b). The title compound was synthesized from tert-butyl 3-(2-hydroxyphenyl)propanoate 3b (3.54 mmol, 785 mg) according to general procedure B and purified using flash chromatography (2% EtOAc in petroleum ether) to yield the product as light yellow oil (208 mg, 18%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.62–7.56 (m, 2H), 7.32 (dd, J = 7.5, 1.7 Hz, 1H), 7.25 (td, J = 7.7, 1.8 Hz, 1H), 7.17 (td, J = 7.5, 1.3 Hz, 1H), 6.98–6.92 (m, 3H), 2.83 (t, J = 7.7 Hz, 2H), 2.49 (dd, J = 9.0, 6.5 Hz, 2H), 1.39 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 172.1, 161.8, 152.7, 134.3, 133.1, 131.2, 128.3, 125.7, 121.0, 119.0, 117.4, 105.8, 80.6, 35.6, 28.2, 25.8. HPLC:  $t_R$  = 7.6 min, 99% at 254 nm. HRMS m/z: calcd for  $C_{20}H_{21}NO_3$  + H<sup>+</sup> [M + H<sup>+</sup>], 324.1594; found, 324.1597.

tert-Butyl 2-(3-(4-Cyanophenoxy)phenyl)acetate (**5c**). The title compound was synthesized from tert-butyl 2-(3-hydroxyphenyl)acetate **3c** (3.10 mmol, 645 mg) according to general procedure B and purified using flash chromatography (15% EtOAc in petroleum ether) to yield the product as clear oil (786 mg, 82%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.62–7.57 (m, 2H), 7.35 (t, J = 7.9 Hz, 1H), 7.13 (d, J = 8.1 Hz, 1H), 7.02–6.99 (m, 3H), 6.96 (dd, J = 8.1, 1.7 Hz, 1H), 3.53 (s, 2H), 1.43 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 170.4, 161.7, 155.0, 137.3, 134.3, 130.3, 126.2, 121.4, 119.0, 118.9, 118.1, 106.0, 81.3, 42.5, 28.2. HPLC:  $t_R = 6.4$  min, 96% at 254

nm. HRMS m/z: calcd for  $C_{19}H_{19}NO_3 + H^+$  [M + H<sup>+</sup>], 310.1438; found, 310.1428.

*tert-Butyl* 3-(3-(4-Cyanophenoxy)phenyl)propanoate (*5d*). The title compound was synthesized from *tert*-butyl 3-(3-hydroxyphenyl)propanoate 3d (3.94 mmol, 875 mg) according to general procedure B and purified using flash chromatography (2% EtOAc in petroleum ether) to yield the product as light yellow oil (600 mg, 47%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.61–7.57 (m, 2H), 7.31 (dd, J = 9.1, 6.5 Hz, 1H), 7.07 (dd, J = 7.6, 0.5 Hz, 1H), 7.01–6.96 (m, 2H), 6.90 (ddd, J = 8.1, 2.9, 1.7 Hz, 2H), 2.91 (t, J = 7.7 Hz, 2H), 2.54 (t, J = 7.7 Hz, 2H), 1.41 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 172.0, 161.8, 155.0, 143.7, 134.7, 130.3, 125.4, 120.5, 119.0, 118.3, 118.0, 105.9, 80.7, 36.9, 31.0, 28.2. HPLC: t<sub>R</sub> = 8.4 min, 99% at 254 nm. HRMS m/z: calcd for C<sub>20</sub>H<sub>21</sub>NO<sub>3</sub> + H<sup>+</sup> [M + H<sup>+</sup>], 324.1594; found, 324.1594.

tert-Butyl 2-(3-(4-Cyanophenoxy)phenoxy)acetate (**5e**). The title compound was synthesized from tert-butyl 2-(3-hydroxyphenoxy)acetate **3e** (4.69 mmol, 1.05 g) according to general procedure B and purified using flash chromatography (15% EtOAc in petroleum ether) to yield the product as clear oil (1.08 g, 71%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.62–7.57 (m, 2H), 7.30 (t, J = 8.2 Hz, 1H), 7.04–6.99 (m, 2H), 6.75 (ddd, J = 8.4, 2.5, 0.8 Hz, 1H), 6.68 (ddd, J = 8.1, 2.2, 0.8 Hz, 1H), 6.60 (t, J = 2.3 Hz, 1H), 4.50 (s, 2H), 1.47 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 167.7, 161.4, 159.6, 156.1, 134.3, 130.9, 118.9, 118.3, 113.3, 111.3, 107.3, 106.2, 82.8, 65.8, 28.2. HPLC:  $t_R = 6.3$  min, 99% at 254 nm. HRMS m/z: calcd for  $C_{19}H_{19}NO_4 + H^+ [M + H^+]$ , 326.1387; found, 326.1375.

tert-Butyl 2-(4-(4-Cyanophenoxy)phenyl)acetate (*5f*). The title compound was synthesized from *tert*-butyl 2-(4-hydroxyphenyl)acetate 3f (4.66 mmol, 970 mg) according to general procedure B and purified using flash chromatography (5% EtOAc in petroleum ether) to yield the product as white crystals (1.20 g, 84%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.62–7.56 (m, 2H), 7.31 (d, J = 8.5 Hz, 2H), 7.05–6.96 (m, 4H), 3.54 (s, 2H), 1.46 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>):  $\delta$  170.9, 161.8, 153.8, 134.3, 131.8, 131.2, 120.5, 119.0, 118.0, 105.9, 81.3, 41.9, 28.2. HPLC:  $t_R$  = 7.2 min, 99% at 254 nm. HRMS m/z: calcd for C<sub>19</sub>H<sub>19</sub>NO<sub>3</sub> + H<sup>+</sup> [M + H<sup>+</sup>], 310.1438; found, 310.1435.

tert-Butyl 3-(4-(4-Cyanophenoxy)phenyl)propanoate (**5g**). The title compound was synthesized from *tert*-butyl 3-(4-hydroxyphenyl)propanoate **3g** (4.90 mmol, 1.09 g) according to general procedure B and purified using flash chromatography (15% EtOAc in petroleum ether) to yield the product as a white solid (1.39 g, 88%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.60–7.55 (m, 2H), 7.26–7.21 (m, 2H), 7.00–6.94 (m, 4H), 2.92 (t, J = 7.7 Hz, 2H), 2.55 (t, J = 7.7 Hz, 2H), 1.42 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>):  $\delta$  172.2, 162.0, 153.1, 138.0, 134.2, 130.2, 120.6, 119.0, 117.8, 105.7, 80.7, 37.1, 30.6, 28.2. HPLC:  $t_R$  = 6.7 min, 99% at 254 nm. HRMS m/z: calcd for C<sub>20</sub>H<sub>21</sub>NO<sub>3</sub> + H<sup>+</sup> [M + H<sup>+</sup>], 324.1594; found, 324.1585.

tert-Butyl 2-(4-(4-Cyanophenoxy)phenoxy)acetate (5h). The title compound was synthesized from tert-butyl 2-(4-hydroxyphenoxy)acetate 3h (4.69 mmol, 1.05 g) according to general procedure B and purified using flash chromatography (15% EtOAc in petroleum ether) to yield the product as white crystals (1.40 g, 92%).  $^1$ H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.60–7.54 (m, 2H), 7.02–6.89 (m, 6H), 4.52 (s, 2H), 1.50 (s, 9H).  $^1$ C NMR (101 MHz, CDCl<sub>3</sub>): δ 168.0, 162.4, 155.4, 148.7, 134.2, 121.9, 119.1, 117.4, 116.2, 105.6, 82.7, 66.2, 28.2.

HPLC:  $t_R = 6.3$  min, 99% at 254 nm. HRMS m/z: calcd for  $C_{19}H_{19}NO_4 + H^+$  [M + H<sup>+</sup>], 326.1387; found, 326.1373.

tert-Butyl (Z)-2-(2-(4-(N'-Hydroxycarbamimidoyl)-phenoxy)phenyl)acetate (6a). The title compound was synthesized from tert-butyl 2-(2-(4-cyanophenoxy)phenyl)-acetate 5a (1.25 mmol, 385 mg) according to general procedure C and obtained as yellow oil (426 mg, 100%). The product was used in the next step without further purification. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.59–7.54 (m, 2H), 7.32 (dd, J = 7.5, 1.6 Hz, 1H), 7.24 (dt, J = 8.0, 2.2 Hz, 1H), 7.13 (td, J = 7.5, 1.2 Hz, 1H), 6.98–6.93 (m, 2H), 6.91 (dd, J = 8.1, 1.1 Hz, 1H), 4.86 (br, 2H), 3.56 (s, 2H), 1.36 (s, 9H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 170.5, 159.1, 154.2, 131.6, 129.3, 128.6, 127.3, 127.0, 126.9, 124.3, 119.6, 117.8, 117.2, 80.8, 67.1, 37.0, 27.9, 27.9. LCMS (ESI<sup>+</sup>) m/z: 343.8 [M + H]<sup>+</sup>,  $t_R = 4.9$  min.

tert-Butyl (*Z*)-3-(2-(4-(N'-Hydroxycarbamimidoyl)-phenoxy)phenyl)propanoate (**6b**). The title compound was synthesized from tert-butyl 3-(2-(4-cyanophenoxy)phenyl)-propanoate **5b** (0.529 mmol, 171 mg) according to general procedure C and obtained as yellow oil (155 mg, 85%). The product was used in the next step without further purification. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ): δ 9.55 (br, 1H), 7.69–7.61 (m, 2H), 7.34 (dd, J = 7.6, 1.6 Hz, 1H), 7.25 (td, J = 7.8, 1.7 Hz, 1H), 7.13 (td, J = 7.5, 1.2 Hz, 1H), 6.92–6.87 (m, 3H), 5.76 (br, 2H), 2.78 (t, J = 7.6 Hz, 2H), 2.50–2.45 (m, 2H), 1.34 (s, 9H). <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ): δ 171.9, 157.9, 156.6, 150.8, 143.5, 130.3, 128.8, 127.6, 124.3, 119.4, 118.2, 117.1, 55.4, 36.5, 30.7, 28.2. LCMS (ESI<sup>+</sup>) m/z: 301.2 [M –  $(C_4H_{10}) + H]^+$ ,  $t_R = 0.5$  min.

tert-Butyl (Z)-2-(3-(4-(N'-Hydroxycarbamimidoyl)-phenoxy)phenyl)acetate (**6c**). The title compound was synthesized from tert-butyl 2-(3-(4-cyanophenoxy)phenyl)-acetate **5c** (2.30 mmol, 712 mg) according to general procedure C and obtained as clear oil (787 mg, 100%). The product was used in the next step without further purification. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.62–7.56 (m, 2H), 7.30 (t, J = 7.8 Hz, 1H), 7.05 (d, J = 7.6 Hz, 1H), 7.00 (dd, J = 8.6, 1.1 Hz, 2H), 6.95 (s, 1H), 6.93 (d, J = 8.1 Hz, 1H), 4.87 (br, 2H), 3.51 (s, 2H), 1.43 (s, 9H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 170.5, 158.9, 156.5, 152.3, 136.8, 129.8, 127.4, 127.2, 124.8, 120.2, 118.5, 117.8, 42.5, 28.0. LCMS (ESI<sup>+</sup>) m/z: 343.2 [M + H]<sup>+</sup>,  $t_R$  = 5.0 min.

tert-Butyl (*Z*)-3-(3-(4-(*N*′-Hydroxycarbamimidoyl)-phenoxy)phenyl)propanoate (**6d**). The title compound was synthesized from tert-butyl 3-(3-(4-cyanophenoxy)phenyl)propanoate **5d** (1.77 mmol, 570 mg) according to general procedure C and obtained as brown oil (628 mg, 100%). The product was used in the next step without further purification. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ): δ 9.57 (br, 1H), 7.69–7.64 (m, 2H), 7.30 (t, J = 7.9 Hz, 1H), 7.02 (d, J = 7.9 Hz, 1H), 6.97–6.93 (m, 2H), 6.92–6.90 (m, 1H), 6.85 (dd, J = 7.8, 2.1 Hz, 1H), 5.78 (br, 2H), 2.80 (t, J = 7.4 Hz, 2H), 2.51 (t, J = 6.9 Hz, 2H), 1.33 (s, 9H). <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ): δ 172.0, 158.2, 154.8, 150.8, 136.6, 130.5, 130.4, 128.7, 127.6, 119.5, 119.5, 117.9, 80.1, 55.4, 36.8, 30.3, 28.2. LCMS (ESI<sup>+</sup>) m/z: 357.2 [M + H]<sup>+</sup>,  $t_R = 5.1$  min.

tert-Butyl (Z)-2-(3-(4-(N'-Hydroxycarbamimidoyl)-phenoxy)phenoxy)acetate (6e). The title compound was synthesized from tert-butyl 2-(3-(4-cyanophenoxy)phenoxy)-acetate 5e (2.53 mmol, 823 mg) according to general procedure C and obtained as viscous yellow oil (906 mg,

100%). The product was used in the next step without further purification.  $^1$ H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.61–7.56 (m, 2H), 7.24 (t, J = 8.3 Hz, 1H), 7.03–6.97 (m, 2H), 6.68–6.62 (m, 2H), 6.56 (t, J = 2.3 Hz, 1H), 4.93 (br, 2H), 4.47 (s, 2H), 1.46 (s, 9H).  $^{13}$ C NMR (101 MHz, CDCl<sub>3</sub>):  $\delta$  167.8, 159.2, 158.6, 157.7, 130.4, 127.5, 127.4, 118.7, 112.3, 109.9, 106.1, 82.5, 65.7, 45.8, 28.0, 8.6. LCMS (ESI<sup>+</sup>) m/z: 359.2 [M + H]<sup>+</sup>,  $t_R$  = 5.0 min.

tert-Butyl (Z)-2-(4-(4-(N'-Hydroxycarbamimidoyl)-phenoxy)phenyl)acetate (6f). The title compound was synthesized from tert-butyl 2-(4-(4-cyanophenoxy)phenyl)acetate 5f (3.05 mmol, 943 mg) according to general procedure C and obtained as light yellow oil (1.04 g, 100%). The product was used in the next step without further purification. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ): δ 9.57 (br, 1H), 7.67 (d, J = 8.8 Hz, 2H), 7.28 (t, J = 9.2 Hz, 2H), 6.98 (t, J = 8.6 Hz, 4H), 5.78 (br, 2H), 3.55 (s, 2H), 1.40 (s, 9H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 171.0, 159.0, 155.3, 130.7, 127.4, 119.4, 118.4, 41.8, 28.1. LCMS (ESI<sup>+</sup>) m/z: 343.9 [M + H]<sup>+</sup>,  $t_R = 5.1$  min.

tert-Butyl (Z)-3-(4-(4-(N'-Hydroxycarbamimidoyl)-phenoxy)phenyl)propanoate (6g). The title compound was synthesized from tert-butyl 3-(4-(4-cyanophenoxy)phenyl)propanoate 5g (3.81 mmol, 1.23 g) according to general procedure C and obtained as yellow oil (1.35 g, 99%). The product was used in the next step without further purification. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.61–7.54 (m, 2H), 7.21–7.16 (m, 2H), 6.99–6.93 (m, 4H), 4.86 (br, 2H), 2.90 (t, J = 7.7 Hz, 2H), 2.54 (t, J = 7.7 Hz, 2H), 1.42 (s, 9H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 172.2, 159.3, 154.6, 152.3, 136.5, 129.8, 127.4, 127.0, 119.6, 118.1, 80.5, 37.1, 30.4, 28.1. LCMS (ESI<sup>+</sup>) m/z: 357.2 [M + H]<sup>+</sup>,  $t_R$  = 5.1 min.

tert-Butyl (Z)-2-(4-(4-(N'-Hydroxycarbamimidoyl)-phenoxy)phenoxy)acetate (6h). The title compound was synthesized from tert-butyl 2-(4-(4-cyanophenoxy)phenoxy)-acetate 5h (2.53 mmol, 823 mg) according to general procedure C and obtained as a yellow amorphous solid (860 mg, 95%). The product was used in the next step without further purification.  $^1$ H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.57–7.53 (m, 2H), 6.99–6.86 (m, 6H), 4.87 (br, 2H), 4.50 (s, 2H), 1.49 (s, 9H).  $^{13}$ C NMR (100 MHz, CDCl<sub>3</sub>): δ 168.0, 159.9, 154.6, 150.1, 127.4, 126.6, 121.1, 117.5, 115.9, 82.5, 66.2, 28.1. LCMS (ESI<sup>+</sup>) m/z: 359.2 [M + H]<sup>+</sup>,  $t_R$  = 4.9 min.

tert-Butyl 2-(2-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3yl)phenoxy)phenyl)acetate (8a). The title compound was synthesized from tert-butyl (Z)-2-(2-(4-(N'hydroxycarbamimidoyl)phenoxy)phenyl)acetate 6a (1.17 mmol, 400 mg) according to general procedure D and purified using flash chromatography (15% EtOAc and 1% acetic acid in petroleum ether) to yield the product as a tan solid (169 mg, 33%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.11–8.05 (m, 4H), 7.35 (dd, I = 7.5, 1.4 Hz, 1H), 7.31–7.25 (m, 1H), 7.16 (t, I =7.5 Hz, 1H), 7.06-7.02 (m, 2H), 7.00-6.93 (m, 3H), 5.96 (br, 1H), 3.59 (s, 2H), 1.37 (s, 9H). 13C NMR (101 MHz, CDCl<sub>3</sub>):  $\delta$  175.6, 170.9, 168.5, 160.2, 159.8, 154.2, 131.9, 130.5, 129.4, 128.9, 127.3, 124.7, 121.7, 120.0, 118.0, 117.1, 116.2, 81.2, 37.3, 28.1. HPLC:  $t_R = 6.9 \text{ min}$ , 97% at 254 nm. HRMS m/z: calcd for  $C_{26}H_{24}N_2O_5 + H^+[M + H^+]$ , 445.1758; found, 445.1751.

tert-Butyl 3-(2-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenyl)propanoate (8b). The title compound was synthesized from tert-butyl (Z)-3-(2-(4-(N'-hydroxycarbamimidoyl)phenoxy)phenyl)propanoate 6b

(0.374 mmol, 133 mg) according to general procedure D and purified using flash chromatography (0–5% EtOAc and 1% acetic acid in petroleum ether) to yield the product as an amorphous white solid (49.0 mg, 29%).  $^1\mathrm{H}$  NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.57 (br, 1H), 8.08–8.03 (m, 2H), 8.03–7.99 (m, 2H), 7.38 (dd, J=7.6, 1.6 Hz, 1H), 7.30 (td, J=7.8, 1.7 Hz, 1H), 7.19 (td, J=7.5, 1.2 Hz, 1H), 7.10–7.05 (m, 2H), 7.02 (dd, J=8.1, 1.1 Hz, 1H), 7.01–6.97 (m, 2H), 2.78 (t, J=7.6 Hz, 2H), 2.52–2.45 (m, 2H), 1.33 (s, 9H).  $^{13}\mathrm{C}$  NMR (101 MHz, CDCl<sub>3</sub>):  $\delta$  175.7, 173.2, 168.3, 160.4, 160.4, 154.0, 132.5, 130.9, 130.4, 129.4, 128.1, 124.8, 121.4, 120.3, 117.7, 116.6, 116.2, 81.1, 35.8, 28.2, 26.1. HPLC:  $t_\mathrm{R}=9.3$  min, 100% at 254 nm. HRMS m/z: calcd for  $\mathrm{C_{27}H_{26}N_2O_5} + \mathrm{H^+}$  [M + H^+], 459.1914; found, 459.1927.

tert-Butyl 2-(3-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3yl)phenoxy)phenyl)acetate (8c). The title compound was synthesized from tert-butyl (Z)-2-(3-(4-(N'hydroxycarbamimidoyl)phenoxy)phenyl)acetate 6c (2.19 mmol, 750 mg) according to general procedure D and purified using flash chromatography (15% EtOAc and 1% acetic acid in petroleum ether) to yield the product as an off-white solid (153 mg, 16%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  8.07 (d, J = 8.6 Hz, 2H), 8.02 (d, J = 8.6 Hz, 2H), 7.40 (t, J = 7.8 Hz, 2H)1H), 7.16 (d, J = 8.6 Hz, 2H), 7.10 (d, J = 7.7 Hz, 1H), 7.06– 6.95 (m, 4H), 3.58 (s, 2H), 1.37 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>):  $\delta$  175.8, 171.2, 168.4, 160.4, 160.1, 156.4, 136.8, 130.5, 130.1, 129.4, 125.1, 121.8, 120.7, 118.6, 118.4, 116.7, 116.3, 81.7, 42.7, 28.2. HPLC:  $t_R = 7.0 \text{ min}$ , 96% at 254 nm. HRMS m/z: calcd for  $C_{26}H_{24}N_2O_5 + H^+$  [M + H<sup>+</sup>], 445.1758; found, 445.1770.

tert-Butyl 3-(3-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3yl)phenoxy)phenyl) Propanoate (8d). The title compound was synthesized from tert-butyl (Z)-3-(3-(4-(N'hydroxycarbamimidoyl)phenoxy)phenyl)propanoate 6d (1.53 mmol, 543 mg) according to general procedure D and purified using flash chromatography (5–10% EtOAc and 1% acetic acid in petroleum ether) to yield the product as an off-white solid (330 mg, 47%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  11.59 (br, 1H), 8.08-8.04 (m, 2H), 8.04-8.00 (m, 2H), 7.36 (t, J = 7.9Hz, 1H), 7.16-7.11 (m, 2H), 7.09 (d, J = 7.9 Hz, 1H), 7.02-6.98 (m, 3H), 6.96 (dd, J = 7.7, 2.1 Hz, 1H), 2.83 (t, J = 7.4Hz, 2H), 2.55-2.51 (m, 2H), 1.34 (s, 9H). <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ ):  $\delta$  175.4, 171.4, 167.5, 162.1, 159.8, 155.3, 143.3, 130.1, 130.1, 129.1, 124.6, 121.0, 119.6, 118.2, 117.4, 116.3, 114.1, 79.7, 36.0, 30.2, 27.7. HPLC:  $t_R = 8.2 \text{ min}$ , 99% at 254 nm. HRMS m/z: calcd for  $C_{27}H_{26}N_2O_5 + H^+$  [M + H<sup>+</sup>], 459.1914; found, 459.1927.

tert-Butyl 2-(3-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenoxy)acetate (**8e**). The title compound was synthesized from tert-butyl (Z)-2-(3-(4-(N'-hydroxycarbamimidoyl)phenoxy)phenoxy)acetate **6e** (2.42 mmol, 867 mg) according to general procedure D and purified using flash chromatography (15% EtOAc and 1% acetic acid in petroleum ether) to yield the product as a yellow solid (312 mg, 28%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ): δ 10.56 (br, 1H), 8.09–8.04 (m, 2H), 8.04–7.99 (m, 2H), 7.35 (t, J = 8.2 Hz, 1H), 7.19–7.13 (m, 2H), 7.02–6.97 (m, 2H), 6.77 (dd, J = 8.2, 2.2 Hz, 1H), 6.72 (dd, J = 8.0, 2.0 Hz, 1H), 6.65 (t, J = 2.3 Hz, 1H), 4.66 (s, 2H), 1.39 (s, 9H). <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ ): δ 175.5, 167.6, 167.5, 162.1, 159.4, 159.2, 156.5, 130.8, 130.1, 129.1, 121.3, 118.5, 116.3, 114.1, 112.1, 110.8, 106.0, 81.5, 65.1, 27.6. HPLC:  $t_R$  = 6.8 min, 98% at 254 nm.

HRMS m/z: calcd for  $C_{26}H_{24}N_2O_6 + H^+$  [M + H<sup>+</sup>], 461.1707; found, 461.1699.

tert-Butyl 2-(4-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3yl)phenoxy)phenyl)acetate (8f). The title compound was synthesized from tert-butyl (Z)-2-(4-(4-(N'hydroxycarbamimidoyl)phenoxy)phenyl)acetate 6f (1.53 mmol, 522 mg) according to general procedure D and purified using flash chromatography (15% EtOAc and 1% acetic acid in petroleum ether) to yield the product as off-white crystals (316 mg, 40%).  ${}^{1}$ H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.58 (br, 1H), 8.06 (d, J = 8.8 Hz, 2H), 8.02 (d, J = 8.7 Hz, 2H), 7.32 (d, J =8.5 Hz, 2H), 7.14 (d, J = 8.8 Hz, 2H), 7.08 (d, J = 8.5 Hz, 2H), 6.99 (d, J = 8.7 Hz, 2H), 3.58 (s, 2H), 1.41 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 175.7, 171.8, 168.4, 160.3, 160.2, 155.2, 131.0, 130.5, 130.4, 129.4, 121.7, 119.9, 118.5, 116.6, 116.3, 81.7, 42.0, 28.2. HPLC:  $t_R = 7.9$  min, 99% at 254 nm. HRMS m/z: calcd for  $C_{26}H_{24}N_2O_5 + H^+[M + H^+]$ , 445.1758; found, 445.1748.

tert-Butyl 3-(4-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenyl)propanoate (8g). The title compound was synthesized from tert-butyl (Z)-3-(4-(4-(N'-hydroxycarbamimidoyl)phenoxy)phenyl)propanoate 6g (2.39 mmol, 851 mg) according to general procedure D and purified using flash chromatography (15% EtOAc and 1% acetic acid in petroleum ether) to yield the product as a tan solid (303 mg, 28%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.12–8.08 (m, 4H), 7.21 (d, J = 8.4 Hz, 2H), 7.06 (d, J = 8.6 Hz, 2H), 6.99 (t, J = 8.4 Hz, 4H), 2.92 (t, J = 7.7 Hz, 2H), 2.57 (t, J = 7.7 Hz, 2H), 1.43 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 175.6, 172.6, 168.5, 160.5, 160.0, 154.5, 136.8, 130.5, 130.0, 129.4, 121.6, 120.0, 118.2, 117.0, 116.2, 80.8, 37.3, 30.6, 28.2. LCMS:  $t_R$  = 7.2 min, 84% at 254 nm. HRMS m/z: calcd for  $C_{27}H_{26}N_2O_5$  +  $H^+$  [M +  $H^+$ ], 459.1914; found, 459.1909.

tert-Butyl 2-(4-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenoxy)acetate (8h). The title compound was synthesized from tert-butyl (Z)-2-(4-(4-(N'-hydroxycarbamimidoyl)phenoxy)phenoxy)acetate 6h (2.42 mmol, 867 mg) according to general procedure D and purified using flash chromatography (15% EtOAc and 1% acetic acid in petroleum ether) to yield the product as white powder (323 mg, 29%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ): δ 10.61 (br, 1H), 8.09–7.96 (m, 4H), 7.13–7.05 (m, 4H), 7.02–6.95 (m, 4H), 4.67 (s, 2H), 1.44 (s, 9H). <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ ): δ 175.4, 167.9, 167.5, 162.1, 160.7, 154.6, 148.7, 130.1, 129.1, 121.4, 120.5, 117.3, 116.3, 116.0, 114.1, 81.4, 65.4, 27.7. HPLC:  $t_R$  = 6.8 min, 99% at 254 nm. HRMS m/z: calcd for  $C_{26}H_{24}N_2O_6$  + H<sup>+</sup> [M + H<sup>+</sup>], 461.1707; found, 461.1720.

2-(2-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3-yl)-phenoxy)phenyl)acetic Acid (9a). The title compound was synthesized from tert-butyl 2-(2-(4-(5-(4-hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenyl)acetate 8a (0.225 mmol, 100 mg) according to general procedure E and obtained as white powder (78 mg, 89%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ): δ 10.56 (br, 1H), 8.03 (t, J = 8.8 Hz, 4H), 7.42 (d, J = 7.4 Hz, 1H), 7.34 (t, J = 7.4 Hz, 1H), 7.21 (t, J = 7.3 Hz, 1H), 7.09 (d, J = 8.6 Hz, 2H), 7.00 (t, J = 8.7 Hz, 3H), 3.59 (s, 2H). <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ ): δ 175.4, 172.0, 167.6, 162.0, 159.7, 153.6, 132.2, 130.1, 129.0, 128.8, 127.3, 124.6, 120.9, 119.8, 117.8, 116.3, 114.2, 35.4. HPLC:  $t_R = 5.7$  min, 98% at 254 nm. HRMS m/z: calcd for  $C_{22}H_{16}N_2O_5 + H^+$  [M + H<sup>+</sup>], 389.1132; found, 389.1125.

3-(2-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3-yl)-phenoxy)phenyl)propanoic Acid (9b). The title compound

was synthesized *tert*-butyl 3-(2-(4-(5-(4-hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenyl)propanoate **8b** (0.058 mmol, 27 mg) according to general procedure E and obtained as an off-white solid (23 mg, 98%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ): δ 12.14 (br, 1H), 10.55 (br, 1H), 8.05 (d, J = 8.8 Hz, 2H), 8.02 (d, J = 8.7 Hz, 2H), 7.40 (d, J = 7.6 Hz, 1H), 7.30 (td, J = 7.8, 1.5 Hz, 1H), 7.20 (t, J = 7.1 Hz, 1H), 7.08 (d, J = 8.8 Hz, 2H), 7.03 (d, J = 8.0 Hz, 1H), 6.99 (d, J = 8.7 Hz, 2H), 2.79 (t, J = 7.7 Hz, 2H), 2.54–2.47 (m, 2H). <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ ): δ 175.4, 173.6, 167.6, 162.1, 160.1, 152.9, 132.6, 130.7, 130.1, 129.2, 128.2, 125.1, 120.7, 120.5, 117.3, 116.3, 114.2, 33.7, 24.9. HPLC:  $t_R$  = 6.6 min, 100% at 254 nm. HRMS m/z: calcd for  $C_{23}H_{18}N_2O_5$  + H<sup>+</sup> [M + H<sup>+</sup>], 403.1288; found, 403.1300.

2-(3-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3-yl)-phenoxy)phenyl)acetic Acid (9c). The title compound was synthesized from tert-butyl 2-(3-(4-(5-(4-hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenyl)acetate 8c (0.225 mmol, 100 mg) according to general procedure E and obtained as white powder (85 mg, 97%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ): δ 10.56 (br, 1H), 8.10–8.05 (m, 2H), 8.05–7.99 (m, 2H), 7.39 (t, J = 7.8 Hz, 1H), 7.18–7.14 (m, 2H), 7.12 (d, J = 7.7 Hz, 1H), 7.06–7.02 (m, 1H), 7.02–6.97 (m, 3H), 3.61 (s, 2H). <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ ): δ 175.5, 172.4, 167.5, 162.1, 159.6, 155.3, 137.5, 130.1, 130.0, 129.2, 125.6, 121.1, 120.7, 118.4, 117.8, 116.3, 114.2, (1 peak missing). HPLC:  $t_R$  = 5.8 min, 99% at 254 nm. HRMS m/z: calcd for  $C_{22}H_{16}N_2O_5$  + H<sup>+</sup> [M + H<sup>+</sup>], 389.1132; found, 389.1123.

3-(3-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3-yl)-phenoxy)phenyl)propanoic Acid (9d). The title compound was synthesized from tert-butyl 3-(3-(4-(5-(4-hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenyl) propanoate 8d (0.22 mmol, 100 mg) according to general procedure E and obtained as an off-white solid (84 mg, 95%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ): δ 12.12 (br, 1H), 10.55 (br, 1H), 8.09–8.04 (m, 2H), 8.05–8.00 (m, 2H), 7.36 (t, J = 7.9 Hz, 1H), 7.17–7.12 (m, 2H), 7.10 (d, J = 8.0 Hz, 1H), 7.04–6.97 (m, 3H), 6.97–6.92 (m, 1H), 2.84 (t, J = 7.5 Hz, 2H), 2.55 (t, J = 7.6 Hz, 2H). <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ ): δ 175.4, 173.7, 167.6, 162.1, 159.7, 155.3, 143.6, 130.1, 130.1, 129.1, 124.5, 120.9, 119.6, 118.2, 117.3, 116.3, 114.1, 35.0, 30.1. HPLC:  $t_R$  = 7.2 min, 100% at 254 nm. HRMS m/z: calcd for  $C_{23}H_{18}N_2O_5+Na^+$  [M + Na $^+$ ], 425.1108; found, 425.1122.

2-(3-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3-yl)-phenoxy)phenoxy)acetic Acid (*9e*). The title compound was synthesized from *tert*-butyl 2-(3-(4-(5-(4-hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenoxy)acetate 8e (0.217 mmol, 100 mg) according to general procedure E and obtained as off-white powder (84 mg, 96%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ): δ 10.57 (br, 1H), 8.07 (d, J = 8.7 Hz, 2H), 8.02 (d, J = 8.7 Hz, 2H), 7.35 (t, J = 8.5 Hz, 1H), 7.17 (d, J = 8.7 Hz, 2H), 7.00 (d, J = 8.7 Hz, 2H), 6.81–6.75 (m, 1H), 6.73–6.67 (m, 2H), 4.70 (s, 2H). <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ ): δ 175.5, 170.0, 167.5, 162.1, 159.4, 159.3, 156.5, 130.8, 130.1, 129.2, 121.2, 118.5, 116.4, 114.1, 112.0, 110.7, 106.2, 64.7. HPLC:  $t_R$  = 5.8 min, 99% at 254 nm. HRMS m/z: calcd for  $C_{22}H_{16}N_2O_6$  +  $H^+$  [M +  $H^+$ ], 405.1081; found, 405.1067.

2-(4-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3-yl)-phenoxy)phenyl)acetic Acid (9f). The title compound was synthesized from *tert*-butyl 2-(4-(4-(5-(4-hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenyl)acetate 8f (0.225 mmol, 100 mg) according to general procedure E and obtained as a

white solid (78 mg, 89%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  12.38 (br, 1H), 10.56 (br, 1H), 8.09–8.05 (m, 2H), 8.04–8.00 (m, 2H), 7.34 (d, J = 8.6 Hz, 2H), 7.18–7.12 (m, 2H), 7.12–7.06 (m, 2H), 7.03–6.96 (m, 2H), 3.60 (s, 2H). <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ ):  $\delta$  175.5, 172.7, 167.6, 162.1, 159.9, 154.0, 131.3, 130.2, 129.2, 121.0, 119.6, 118.2, 116.3, 114.2, 39.9, (1 peak missing). HPLC:  $t_{\rm R}$  = 6.4 min, 99% at 254 nm. HRMS m/z: calcd for  ${\rm C}_{22}{\rm H}_{16}{\rm N}_2{\rm O}_4$  + H<sup>+</sup> [M + H<sup>+</sup>], 389.1132; found, 389.1146.

3-(4-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3-yl)-phenoxy)phenyl)propanoic Acid (9g). The title compound was synthesized from tert-butyl 3-(4-(4-(5-(4-hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenyl)propanoate 8g (0.437 mmol, 200 mg) according to general procedure E and obtained as yellow powder (160 mg, 91%). <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>): δ 10.56 (br, 1H), 8.03 (dd, J = 13.9, 8.7 Hz, 4H), 7.30 (d, J = 8.4 Hz, 2H), 7.11 (d, J = 8.7 Hz, 2H), 7.04 (d, J = 8.4 Hz, 2H), 6.99 (d, J = 8.7 Hz, 2H), 2.84 (t, J = 7.5 Hz, 2H), 2.56 (t, J = 7.6 Hz, 2H). <sup>13</sup>C NMR (101 MHz, DMSO): δ 175.4, 173.8, 167.6, 162.1, 160.0, 153.5, 137.2, 130.1, 130.1, 129.1, 120.9, 119.8, 118.0, 116.3, 114.2, 35.2, 29.7. HPLC:  $t_R$  = 5.9 min, 99% at 254 nm. HRMS m/z: calcd for  $C_{23}H_{18}N_2O_5$  + H<sup>+</sup> [M + H<sup>+</sup>], 403.1288; found, 403.1280.

2-(4-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3-yl)-phenoxy)phenoxy)acetic Acid (9h). The title compound was synthesized from tert-butyl 2-(4-(4-(5-(4-hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenoxy)acetate 8h (0.217 mmol, 100 mg) according to general procedure E and obtained as off-white powder (83 mg, 95%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ): δ 10.56 (br, 1H), 8.03 (t, J = 9.1 Hz, 4H), 7.14–7.05 (m, 4H), 7.02–6.95 (m, 4H), 4.69 (s, 2H). <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ ): δ 175.4, 170.2, 167.6, 162.1, 160.7, 154.7, 148.6, 130.1, 129.1, 121.4, 120.5, 117.3, 116.3, 116.0, 114.2, 64.9. HPLC:  $t_R$  = 5.8 min, 99% at 254 nm. HRMS m/z: calcd for  $C_{22}H_{16}N_2O_6 + H^+$  [M + H<sup>+</sup>], 405.1081; found, 405.1073.

(*Z*)-*N'-Hydroxy-4-iodobenzimidamide* (*10*). The title compound was synthesized from 4-iodobenzonitrile 4 (5.24 mmol, 1.20 g) according to general procedure C and obtained as off-white crystals (1.16 g, 84%).  $^{1}$ H NMR (400 MHz, DMSO- $d_{6}$ ):  $\delta$  9.72 (br, 1H), 7.77–7.68 (m, 2H), 7.49–7.42 (m, 2H), 5.84 (br, 2H); spectroscopic data consistent with the literature.  $^{43}$ 

4-((tert-Butyldimethylsilyl)oxy)benzoyl Chloride (11). The title compound was synthesized as previously reported and used directly in the next step without further purification.<sup>26</sup>

5-(4-((tert-Butyldimethylsilyl)oxy)phenyl)-3-(4-iodophenyl)-1,2,4-oxadiazole (12). A suspension of 4-((tertbutyldimethylsilyl)oxy)benzoyl chloride 11 (1 equiv, 9.11 mmol) in toluene (30 mL) was added to a suspension of (Z)-N'-hydroxy-4-iodobenzimidamide 10 (1 equiv, 9.11 mmol, 2.39 g) in toluene (20 mL) and allowed to stir at reflux (130 °C) for 48 h. The solution was filtered while still hot, and the filtrate was concentrated in vacuo. The residue was recrystallized from EtOH to yield the product as off-white crystals (2.62 g, 60%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.12–8.07 (m, 2H), 7.92-7.83 (m, 4H), 7.00-6.95 (m, 2H), 1.02-0.99 (m, 9H), 0.28-0.24 (m, 6H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>):  $\delta$ 176.0, 168.4, 160.2, 138.2, 130.2, 129.2, 126.9, 120.9, 117.4, 97.9, 25.7, 18.4, -4.2. HPLC: (hydrophobic PP method)  $t_R =$ 5.0 min, 92% at 254 nm. HRMS m/z: calcd for  $C_{20}H_{23}IN_2O_2Si$ + H<sup>+</sup> [M + H<sup>+</sup>], 479.0646; found, 479.0661.

**General Procedure F.** 5-(4-((*tert*-Butyldimethylsilyl)oxy)-phenyl)-3-(4-iodophenyl)-1,2,4-oxadiazole **12** (1 equiv), phe-

nol (4–5 equiv),  $K_2CO_3$  (2 equiv) and 1,3-diphenyl-1,3-propanedione (1 equiv) were dissolved in DMSO (0.93 M) in a sealed tube and purged with  $N_2$  several times. Copper(II) acetylacetonate (0.5 equiv) was then added, the reaction vessel was sealed, and the solution was stirred at 90 °C for 24 h. The reaction mixture was diluted with EtOAc and filtered through Celite. The filtrate was washed with aq  $NH_4Cl$  solution and brine; then, the organic layers were dried over  $MgSO_4$  and solvent was removed in vacuo. The desired product was isolated using flash chromatography eluting with 10-35% EtOAc in petroleum ether. 1,3-Diphenyl-1,3-propanedione impurities that were present with the obtained product were removed after the subsequent reaction.

**General Procedure G.** The boc-protected substrate was dissolved in 4 M HCl/1,4-dioxane ( $\sim$ 1 mL/30 mg substrate) and stirred at rt for 6 h. After reaction completion as determined by TLC, the solution was diluted with Et<sub>2</sub>O, filtered, and further washed with Et<sub>2</sub>O. The filtered solid was air-dried and collected to yield the HCl salt of the product.

tert-Butyl (2-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)benzyl)carbamate (14a). The title compound was synthesized from tert-butyl (2-hydroxybenzyl)carbamate 13a (4 equiv, 1.6 mmol, 350 mg) according to general procedure F and purified using flash chromatography (10–25% EtOAc in petroleum ether) to yield the product as a yellow solid (48 mg, 27%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.05–8.00 (m, 2H), 8.00–7.93 (m, 2H), 7.41 (d, J = 7.3 Hz, 1H), 7.31–7.26 (m, 1H), 7.17 (td, J = 7.5, 0.8 Hz, 1H), 7.00–6.88 (m, 5H), 5.06 (br, 1H), 4.37 (d, J = 5.8 Hz, 2H), 1.45 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 175.8, 168.2, 160.9, 160.0, 156.5, 153.9, 130.4, 130.3, 129.9, 129.5, 129.3, 124.9, 121.9, 120.1, 117.8, 116.2, 116.2, 80.3, 40.2, 28.5. HPLC:  $t_R$  = 8.1 min, 95% at 254 nm. HRMS m/z: calcd for  $C_{26}H_{25}N_3O_5$  +  $H^+$  [M +  $H^+$ ], 460.1867; found, 460.1883.

tert-Butyl (2-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3yl)phenoxy)phenethyl)carbamate (14b). The title compound was synthesized from tert-butyl (2-hydroxyphenethyl)carbamate 13b (5 equiv, 3.0 mmol, 700 mg) according to general procedure F and purified using flash chromatography (10-25% EtOAc in petroleum ether) to yield the product as a yellow solid (78 mg, 28%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$ 8.05 (t, J = 8.7 Hz, 4H), 7.31-7.25 (m, 1H), 7.23 (dd, J = 7.9)1.8 Hz, 1H), 7.14 (td, J = 7.4, 1.2 Hz, 1H), 7.00 (d, J = 8.8 Hz, 2H), 6.98–6.92 (m, 3H), 4.75 (br, 1H), 3.41 (dd, *J* = 12.4, 6.1 Hz, 2H), 2.84 (t, J = 6.9 Hz, 2H), 1.43 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>):  $\delta$  175.8, 168.4, 160.6, 160.2, 156.4, 154.3, 131.5, 130.8, 130.4, 129.5, 128.4, 124.8, 121.8, 120.1, 117.9, 116.5, 116.3, 79.8, 41.0, 30.9, 28.6. HPLC:  $t_R = 7.5 \text{ min}$ , 83% at 254 nm. HRMS m/z: calcd for  $C_{27}H_{27}N_3O_5 + H^+$  [M + H<sup>+</sup>], 474.2023; found, 474.2035.

tert-Butyl (4-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)benzyl)carbamate (14d). The title compound was synthesized from tert-butyl (4-hydroxybenzyl)carbamate 13d (5 equiv, 2.09 mmol, 467 mg) according to general procedure F and purified using flash chromatography (10–25% EtOAc in petroleum ether) to yield the product as a flaky yellow solid (69 mg, 36%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.12–8.04 (m, 4H), 7.28 (d, J = 8.5 Hz, 2H), 7.08–7.00 (m, 4H), 6.97 (d, J = 8.8 Hz, 2H), 4.98 (br, 1H), 4.33 (d, J = 5.3 Hz, 2H), 1.49 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 175.8, 168.4, 160.5, 160.1, 156.4, 155.6, 134.6, 130.5, 129.4, 129.2, 121.9, 120.0, 118.5, 116.7, 116.3, 80.2, 44.3, 28.6. HPLC:  $t_R$  =

6.6 min, 99% at 254 nm. HRMS m/z: calcd for  $C_{26}H_{25}N_3O_5 + H^+$  [M + H<sup>+</sup>], 460.1867; found, 460.1870.

tert-Butyl (4-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenethyl)carbamate (14e). The title compound was synthesized from *N*-boc-tyramine (5 equiv, 4.71 mmol, 1.12 g) according to general procedure F (using Cs<sub>2</sub>CO<sub>3</sub> instead of K<sub>2</sub>CO<sub>3</sub>) and purified using flash chromatography (10–35% EtOAc in petroleum ether) to yield the product as a yellow solid (319 mg, 72%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.15–8.07 (m, 4H), 7.20 (d, J = 8.4 Hz, 2H), 7.09–7.05 (m, 2H), 7.04–6.96 (m, 4H), 6.06 (br, 1H), 4.61 (br, 1H), 3.40 (d, J = 6.1 Hz, 2H), 2.80 (t, J = 7.0 Hz, 2H), 1.46 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 175.7, 168.5, 160.3, 160.3, 156.3, 154.9, 134.8, 130.5, 130.4, 129.4, 121.8, 120.0, 118.4, 116.9, 116.3, 79.8, 42.1, 35.8, 28.6. HPLC: t<sub>R</sub> = 7.6 min, 99% at 254 nm. HRMS m/z: calcd for  $2\times(C_{27}H_{27}N_3O_5) + H^+$  [2M + H<sup>+</sup>], 947.3974; found, 947.3977.

(RS)-tert-Butyl (2-(4-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenyl)propyl)carbamate (14f). The title compound was synthesized from (RS)-tert-butyl (2-(4hydroxyphenyl)propyl)carbamate 13f (4.5 equiv, 1.9 mmol, 470 mg) according to general procedure F and purified using flash chromatography (10-25% EtOAc in petroleum ether) to yield the product as a yellow solid (71 mg, 35%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.12–8.05 (m, 4H), 7.20 (d, J = 8.3 Hz, 2H), 7.07 (d, J = 8.7 Hz, 2H), 7.03-6.97 (m, 4H), 4.59 (br, 1H), 3.39 (dt, J = 13.0, 6.5 Hz, 1H), 3.21 (ddd, J = 13.5, 8.1, 5.5 Hz, 1H), 2.93 (dd, *J* = 13.6, 6.7 Hz, 1H), 1.44 (s, 9H), 1.27 (d, J = 7.0 Hz, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>):  $\delta$  175.8, 168.4, 160.6, 160.2, 156.4, 154.9, 140.0, 130.5, 129.4, 128.8, 121.8, 119.9, 118.5, 116.6, 116.3, 79.9, 47.6, 39.7, 28.6, 19.3. HPLC:  $t_R$  = 7.8 min, 91% at 254 nm. HRMS m/z: calcd for  $2\times(C_{28}H_{29}N_3O_5) + H^+ [2M + H^+], 975.4287;$  found, 975.4271.

tert-Butyl (4-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenethyl)(methyl)carbamate (14g). The title compound was synthesized from tert-butyl (4-hydroxyphenethyl) (methyl) carbamate 13g (4 equiv, 1.8 mmol, 450 mg) according to general procedure F and purified using flash chromatography (10–25% EtOAc in petroleum ether) to yield the product as a yellow solid (44 mg, 21%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.12–8.04 (m, 4H), 7.18 (d, J = 7.8 Hz, 2H), 7.08–6.94 (m, 6H), 3.51–3.41 (m, 2H), 2.89–2.77 (m, 5H), 1.44 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 175.8, 168.4, 160.8, 160.4, 156.1, 154.8, 135.1, 130.5, 130.4, 129.4, 121.8, 120.0, 118.3, 116.5, 116.3, 80.1, 51.1, 34.8, 33.9, 28.6. HPLC:  $t_R$  = 8.8 min, 97% at 254 nm. HRMS m/z: calcd for  $[M + H]^+$   $C_{28}H_{29}N_3O_5$ , 488.2180; found, 488.2190.

tert-Butyl (3-(4-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenyl)propyl)carbamate (14h). The title compound was synthesized from tert-butyl (3-(4-hydroxyphenyl)propyl)carbamate 13h (4.8 equiv, 3.52 mmol, 885 mg) according to general procedure F (using Cs<sub>2</sub>CO<sub>3</sub> instead of K<sub>2</sub>CO<sub>3</sub>) and purified using flash chromatography (10–25% EtOAc in petroleum ether) to yield the product as an amorphous yellow solid (257 mg, 75%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.14–8.07 (m, 4H), 7.18 (d, J = 8.6 Hz, 2H), 7.09–7.04 (m, 2H), 7.02–6.95 (m, 4H), 5.69 (br, 1H), 4.56 (br, 1H), 3.18 (d, J = 6.6 Hz, 2H), 2.70–2.60 (m, 2H), 1.84 (dq, J = 14.6, 7.1 Hz, 2H), 1.46 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 175.8, 168.4, 160.7, 160.5, 156.6, 154.4, 137.4, 130.5, 129.9, 129.4, 121.6, 120.0, 118.2, 116.6, 116.3, 79.9, 40.4, 32.5, 31.8, 28.6. HPLC: t<sub>R</sub> = 8.5 min, 92% at 254 nm.

HRMS m/z: calcd for  $C_{28}H_{29}N_3O_5 + H^+$  [M + H<sup>+</sup>], 488.2180; found, 488.2185.

tert-Butyl 7-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3yl)phenoxy)-3,4-dihydroisoquinoline-2(1H)-carboxylate (14i). The title compound was synthesized from tert-butyl 7hydroxy-3,4-dihydroisoquinoline-2(1H)-carboxylate 13i (4 equiv, 1.4 mmol, 350 mg) according to general procedure F and purified using flash chromatography (10-20% EtOAc in petroleum ether) to yield the product as a yellow solid (37 mg, 22%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.09 (dd, J = 8.3, 4.8 Hz, 4H), 7.14 (d, J = 8.3 Hz, 1H), 7.06 (d, J = 8.0 Hz, 2H), 7.00 (d, J = 8.7 Hz, 2H), 6.90 (dd, J = 8.2, 2.0 Hz, 1H), 6.82 (d, J = 2.3 Hz, 1H), 4.57 (d, J = 11.3 Hz, 2H), 3.68 (t, J = 5.8)Hz, 2H), 2.84 (t, J = 5.8 Hz, 2H), 1.51 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>):  $\delta$  175.8, 168.4, 160.6, 160.3, 155.3, 154.7, 135.4, 130.6, 130.5, 130.3, 129.4, 121.9, 118.4, 118.3, 117.5, 116.6, 116.3, 80.5, 46.1, 42.0, 28.7, 28.5. HPLC:  $t_R = 8.1 \text{ min}$ , 75% at 254 nm. HRMS m/z: calcd for  $C_{28}H_{27}N_3O_5 + Na^+$  [M + Na<sup>+</sup>], 508.1843; found, 508.1853.

4-(3-(4-(2-(Aminomethyl)phenoxy)phenyl)-1,2,4-oxadiazol-5-yl)phenol Hydrochloride (15a). The title compound was synthesized from tert-butyl (2-(4-(5-(4-hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)benzyl)carbamate 14a (0.102 mmol, 47 mg) according to general procedure G and obtained as off-white powder (30 mg, 74%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ): δ 10.67 (br, 1H), 8.47 (br, 3H), 8.14–8.09 (m, 2H), 8.05–7.99 (m, 2H), 7.66 (dd, J = 7.7, 1.5 Hz, 1H), 7.44 (td, J = 8.1, 1.7 Hz, 1H), 7.28 (td, J = 7.6, 1.0 Hz, 1H), 7.26–7.21 (m, 2H), 7.05–6.99 (m, 3H), 4.06 (q, J = 5.7 Hz, 2H). <sup>13</sup>C NMR (101 MHz, methanol- $d_4$ ): δ 177.5, 169.3, 163.6, 160.2, 156.4, 132.4, 132.2, 131.2, 130.5, 125.7, 125.4, 124.3, 120.3, 119.8, 117.2, 116.4, 39.6. HPLC:  $t_R$  = 5.4 min, 98% at 254 nm. HRMS m/z: calcd for  $C_{21}H_{17}N_3O_3 + H^+$  [M + H<sup>+</sup>], 360.1343; found, 360.1347.

4-(3-(4-(2-(2-Aminoethyl)phenoxy)phenyl)-1,2,4-oxadiazol-5-yl)phenol Hydrochloride (15b). The title compound was synthesized from tert-butyl (2-(4-(5-(4-hydroxyphenyl)-1,2,4oxadiazol-3-yl)phenoxy)phenethyl)carbamate 14b (0.13 mmol, 63 mg) according to general procedure G and obtained as tan powder (9.1 mg, 17%). <sup>1</sup>H NMR (400 MHz, DMSO $d_6$ ):  $\delta$  10.61 (br, 1H), 8.10–8.06 (m, 2H), 8.04–7.99 (m, 2H), 7.88 (br, 3H), 7.43 (dd, J = 7.6, 1.6 Hz, 1H), 7.37 (td, J = 7.8, 1.8 Hz, 1H), 7.25 (td, J = 7.5, 1.2 Hz, 1H), 7.15–7.11 (m, 2H), 7.05 (dd, J = 8.1, 1.0 Hz, 1H), 7.03-6.98 (m, 2H), 3.04-3.00 (m, 2H), 2.89 (dd, I = 9.6, 6.3 Hz, 2H). <sup>13</sup>C NMR (101 MHz, methanol- $d_4$ ):  $\delta$  177.4, 169.3, 163.5, 161.4, 155.5, 132.4, 131.2, 130.4, 130.4, 129.6, 126.3, 123.3, 121.4, 118.9, 117.2, 116.4, 40.8, 29.6. HPLC:  $t_R$  = 6.0 min, 95% at 254 nm. HRMS m/z: calcd for  $C_{22}H_{19}N_3O_3 + H^+[M + H^+]$ , 374.1499; found, 374.1503.

4-(3-(4-(3-(2-Aminoethyl)phenoxy)phenyl)-1,2,4-oxadiazol-5-yl)phenol Hydrochloride (15c). The title compound was synthesized from tert-butyl (3-hydroxyphenethyl)carbamate 13c (5 equiv, 1.6 mmol, 370 mg) according to general procedure F, and flash chromatography (5–20% EtOAc in petroleum ether) was carried out to yield the impure product. The crude material was dissolved in 4 M HCl/1,4-dioxane (0.5 mL) and stirred at rt for 6 h. The solution was diluted with Et<sub>2</sub>O, filtered, and further washed with Et<sub>2</sub>O. The collected solid was air-dried, and the products were separated using preparative HPLC. The recovered material was treated with 1.25 M HCl/1,4-dioxane (0.50 mL) to convert the product to the HCl salt. The reaction mixture was concentrated in vacuo,

diluted with Et<sub>2</sub>O, and the resulting solid was collected and dried to yield the product as a white solid (7.2 mg, 6% over two steps).  $^1\mathrm{H}$  NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  8.11–8.05 (m, 2H), 8.02 (d, J = 8.8 Hz, 2H), 7.75 (br, 3H), 7.42 (t, J = 7.9 Hz, 1H), 7.21–7.16 (m, 2H), 7.14 (d, J = 7.7 Hz, 1H), 7.07 (s, 1H), 7.05–6.99 (m, 3H), 3.10–3.03 (m, 2H), 2.93–2.86 (m, 2H).  $^{13}\mathrm{C}$  NMR (101 MHz, methanol- $d_4$ ):  $\delta$  177.4, 169.4, 163.5, 161.5, 158.1, 140.4, 131.7, 131.2, 130.3, 125.7, 123.2, 121.1, 119.6, 117.2, 116.4, 41.8, 34.4 (1 peak missing). HPLC:  $t_\mathrm{R}$  = 6.1 min, 99% at 254 nm. HRMS m/z: calcd for  $\mathrm{C}_{22}\mathrm{H}_{19}\mathrm{N}_3\mathrm{O}_3$  + H\* [M + H\*], 374.1499; found, 374.1503.

4-(3-(4-(4-(Aminomethyl)phenoxy)phenyl)-1,2,4-oxadiazol-5-yl)phenol Hydrochloride (15d). The title compound was synthesized from tert-butyl (4-(4-(5-(4-hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)benzyl)carbamate 14d (0.124 mmol, 57.0 mg) according to general procedure G and obtained as off-white powder (46 mg, 94%). <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>): δ 10.59 (br, 1H), 8.23 (br, 3H), 8.12–8.07 (m, 2H), 8.04–8.00 (m, 2H), 7.55 (d, J = 8.7 Hz, 2H), 7.23–7.18 (m, 2H), 7.18–7.13 (m, 2H), 7.03–6.97 (m, 2H), 4.05 (q, J = 5.8 Hz, 2H). <sup>13</sup>C NMR (101 MHz, methanol-d<sub>4</sub>): δ 177.5, 169.4, 163.6, 161.0, 158.6, 132.1, 131.2, 130.4, 130.1, 123.7, 121.0, 119.9, 117.2, 116.4, 43.8. HPLC:  $t_R$  = 5.0 min, 97% at 254 nm. HRMS m/z: calcd for  $C_{21}H_{17}N_3O_3 + H^+$  [M + H<sup>+</sup>], 360.1343; found, 360.1342.

4-(3-(4-(4-(2-Aminoethyl)phenoxy)phenyl)-1,2,4-oxadiazol-5-yl)phenol Hydrochloride (15e). The title compound was synthesized from tert-butyl (4-(4-(5-(4-hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenethyl)carbamate 14e (0.32 mmol, 149 mg) according to general procedure G and obtained as a tan solid (128 mg, 100%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ): δ 10.63 (br, 1H), 8.10–7.94 (m, 7H), 7.35 (d, J = 8.4 Hz, 2H), 7.13 (dd, J = 12.6, 8.7 Hz, 4H), 7.01 (d, J = 8.7 Hz, 2H), 3.11–3.00 (m, 2H), 2.95–2.86 (m, 2H). <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ ): δ 175.5, 167.5, 162.1, 159.8, 154.1, 133.5, 130.5, 130.1, 129.1, 121.0, 119.9, 118.1, 116.3, 114.1, 32.3. HPLC:  $t_R$  = 5.2 min, 100% at 254 nm. HRMS m/z: calcd for  $C_{22}H_{19}N_3O_3 + H^+$  [M + H $^+$ ], 374.1499; found, 374.1507.

(RS)-4-(3-(4-(4-(1-Aminopropan-2-yl)phenoxy)phenyl)-1,2,4-oxadiazol-5-yl)phenol Hydrochloride (15f). The title compound was synthesized from (RS)-tert-butyl (2-(4-(4-(5-(4-hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenyl)propyl)carbamate 14f (0.094 mmol, 46 mg) according to general procedure G and obtained as tan powder (30 mg, 75%).  $^{1}$ H NMR (400 MHz, DMSO- $d_{6}$ ):  $\delta$  10.65 (br, 1H), 8.07 (d, J = 8.1 Hz, 5H), 8.01 (d, J = 8.3 Hz, 2H), 7.38 (d, J = 8.5)Hz, 2H), 7.16 (d, J = 8.8 Hz, 2H), 7.12 (d, J = 8.5 Hz, 2H), 7.01 (d, J = 8.7 Hz, 2H), 3.09 (dt, J = 13.4, 6.8 Hz, 1H), 3.01 (d, J = 5.5 Hz, 2H), 1.28 (d, J = 6.8 Hz, 3H). <sup>13</sup>C NMR (101 MHz, methanol- $d_4$ ):  $\delta$  177.4, 169.4, 163.5, 161.6, 156.9, 139.2, 131.2, 130.3, 130.0, 123.1, 121.5, 119.4, 117.2, 116.4, 46.9, 39.1, 20.0. HPLC:  $t_R = 5.8 \text{ min}$ , 99% at 254 nm. HRMS m/z: calcd for  $C_{23}H_{21}N_3O_3 + H^+ [M + H^+]$ , 388.1656; found, 388.1661.

*4-*(*3-*(*4-*(*4-*(*2-*(*Methylamino*)*ethyl*)*phenoxy*)*phenyl*)*-1,2,4-oxadiazol-5-yl*)*phenol Hydrochloride* (*15g*). The title compound was synthesized from *tert*-butyl (4-(4-(5-(4-hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenethyl)(methyl)-carbamate **14g** (0.067 mmol, 34 mg) according to general procedure G and obtained as tan powder (23 mg, 77%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ): *δ* 10.62 (br, 1H), 8.83 (br, 2H), 8.07 (d, J = 8.8 Hz, 2H), 8.02 (d, J = 8.7 Hz, 2H), 7.35 (d, J = 8.5 Hz, 2H), 7.13 (t, J = 9.0 Hz, 4H), 7.01 (d, J = 8.7 Hz, 2H),

3.14 (s, 2H), 2.99–2.91 (m, 2H), 2.58 (s, 3H).  $^{13}$ C NMR (101 MHz, methanol- $d_4$ ):  $\delta$  177.4, 169.4, 163.5, 161.6, 156.9, 133.5, 131.6, 131.2, 130.3, 123.1, 121.3, 119.4, 117.2, 116.4, 51.4, 33.8, 32.6. HPLC:  $t_R$  = 5.8 min, 100% at 254 nm. HRMS m/z: calcd for  $C_{23}H_{21}N_3O_3$  +  $H^+$  [M +  $H^+$ ], 388.1656; found, 388.1658.

4-(3-(4-(4-(3-Aminopropyl)phenoxy)phenyl)-1,2,4-oxadiazol-5-yl)phenol Hydrochloride (15h). The title compound was synthesized from tert-butyl (3-(4-(4-(5-(4-hydroxyphenyl)1,2,4-oxadiazol-3-yl)phenoxy)phenyl)propyl)carbamate 14h (0.032 mmol, 16 mg) according to general procedure G and obtained as tan powder (12 mg, 89%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ): δ 8.07 (d, J = 8.8 Hz, 2H), 8.02 (d, J = 8.7 Hz, 2H), 7.96–7.53 (br, 3H), 7.30 (d, J = 8.5 Hz, 2H), 7.11 (dd, J = 15.3, 8.7 Hz, 4H), 7.00 (d, J = 8.8 Hz, 2H), 2.83–2.78 (m, 2H), 2.67 (t, J = 7.6 Hz, 2H), 1.90–1.82 (m, 2H). <sup>13</sup>C NMR (101 MHz, methanol- $d_4$ ): δ 177.4, 169.4, 163.5, 161.9, 156.0, 137.9, 131.2, 131.0, 130.2, 122.9, 121.1, 119.1, 117.2, 116.4, 40.3, 32.8, 30.4. HPLC:  $t_R$  = 5.9 min, 100% at 254 nm. HRMS m/z: calcd for  $C_{23}H_{21}N_3O_3 + H^+$  [M + H $^+$ ], 388.1656; found, 388.1666.

4-(3-(4-((1,2,3,4-Tetrahydroisoquinolin-7-yl)oxy)phenyl)-1,2,4-oxadiazol-5-yl)phenol Hydrochloride (15i). The title compound was synthesized from tert-butyl 7-(4-(5-(4-hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)-3,4-dihydroisoquinoline-2(1H)-carboxylate 14i (0.068 mmol, 33 mg) according to general procedure G and obtained as a tan solid (21 mg, 75%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ): δ 10.61 (br, 1H), 9.28 (br, 2H), 8.11–8.06 (m, 2H), 8.04–8.00 (m, 2H), 7.32 (d, J = 8.2 Hz, 1H), 7.17–7.12 (m, 2H), 7.10–7.04 (m, 2H), 7.03–6.98 (m, 2H), 4.26 (s, 2H), 3.42–3.34 (m, 2H), 3.01 (t, J = 6.1 Hz, 2H). <sup>13</sup>C NMR (101 MHz, methanol- $d_4$ ): δ 177.4, 169.4, 163.5, 161.3, 156.6, 132.0, 131.2, 131.1, 130.3, 128.5, 123.4, 120.8, 119.5, 118.7, 117.2, 116.4, 45.7, 43.0, 25.6. HPLC:  $t_R$  = 5.7 min, 100% at 254 nm. HRMS m/z: calcd for C<sub>23</sub>H<sub>19</sub>N<sub>3</sub>O<sub>3</sub> + H<sup>+</sup> [M + H<sup>+</sup>], 386.1499; found, 386.1508.

4-((tert-Butoxycarbonyl)amino)benzoic Acid (17a). 4-Aminobenzoic acid 16a (1 equiv, 21.9 mmol, 3.00 g) was solubilized in MeOH (24 mL) and water (24 mL) and NaHCO<sub>3</sub> (2 equiv, 43.8 mmol, 3.72 g) was added. Boc<sub>2</sub>O (1.5 equiv, 32.8 mmol, 7.15 g) dissolved in MeOH (24 mL) was added dropwise, and the solution was stirred at rt overnight. The solvent was removed in vacuo, and the residue was extracted with EtOAc and then washed with aq 1 M HCl solution and brine. The organic layers were combined, dried over MgSO<sub>4</sub>, and the solvent was removed in vacuo to yield the product as a white solid (4.47 g, 86%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.06–8.01 (m, 2H), 7.46 (d, J = 8.8 Hz, 2H), 6.73 (br, 1H), 1.54 (s, 9H); spectroscopic data consistent with the literature. <sup>44</sup>

4-((tert-Butoxycarbonyl)(methyl)amino)benzoic Acid (17b). 4-(Methylamino)benzoic acid 16b (1 equiv, 13.2 mmol, 2.0 g) was solubilized in MeOH (16 mL) and water (16 mL) and NaHCO<sub>3</sub> (2 equiv, 26.5 mmol, 2.25 g) was added. Boc<sub>2</sub>O (1.5 equiv, 19.9 mmol, 4.32 g) dissolved in MeOH (16 mL) was added dropwise, and the solution was stirred at rt overnight. The solvent was removed in vacuo, and the residue was extracted with EtOAc and then washed with aq 1 M HCl solution and brine. The organic layers were combined, dried over MgSO<sub>4</sub>, and the solvent was removed in vacuo to yield the product as a white solid (2.61 g, 79%).  $^1$ H NMR (400 MHz, DMSO- $d_6$ ): δ 7.95–7.82 (m, 2H), 7.47–

7.33 (m, 2H), 3.22 (s, 3H), 1.41 (s, 9H); spectroscopic data consistent with the literature.<sup>45</sup>

(*Z*)-*N'-Hydroxy-4-phenoxybenzimidamide* (*18*). The title compound was synthesized as previously reported and obtained as a white solid (1.06 g, 80%). <sup>26</sup> <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.63–7.56 (m, 2H), 7.40–7.33 (m, 2H), 7.18–7.12 (m, 1H), 7.06–6.98 (m, 4H), 4.84 (br, 2H); spectroscopic data consistent with the literature. <sup>26</sup>

General Procedure H. A solution of (Z)-N'-hydroxy-4-phenoxybenzimidamide 18 (1 equiv) in DMF (0.11 M) was treated with benzoic acid (1.5 equiv), HOBt (2 equiv), and EDC·HCl (2 equiv); then, the solution was stirred at rt for 16 h. The temperature was increased to 100 °C, and the stirring was continued for 16 h. The reaction mixture was concentrated in vacuo, diluted with a 10% solution of aq Na<sub>2</sub>CO<sub>3</sub>, and then, EtOAc was added to extract the product. The separated organic layer was dried over anhydrous MgSO<sub>4</sub> and concentrated in vacuo; then, the residue was purified using flash chromatography eluting with 5–20% EtOAc in petroleum ether to yield the product.

*tert*-Butyl (4-(3-(4-Phenoxyphenyl)-1,2,4-oxadiazol-5-yl)phenyl)carbamate (19a). The title compound was synthesized from 4-((*tert*-butoxycarbonyl)amino)benzoic acid 17a (4.22 mmol, 1.00 g) according to general procedure H and purified using flash chromatography (5–10% EtOAc in petroleum ether) to yield the product as a white solid (252 mg, 21%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.17–8.09 (m, 4H), 7.55 (d, J = 8.8 Hz, 2H), 7.43–7.35 (m, 2H), 7.21–7.15 (m, 1H), 7.12–7.06 (m, 4H), 6.69 (br, 1H), 1.55 (s, 9H). <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ ): δ 212.8, 205.2, 197.4, 193.0, 190.1, 181.8, 167.9, 166.8, 166.6, 162.1, 158.6, 157.3, 155.9, 155.7, 154.2, 117.5, 65.6. HPLC:  $t_R$  = 8.6 min, 98% at 254 nm. HRMS m/z: calcd for  $C_{25}H_{23}N_3O_4 + H^+$  [M + H<sup>+</sup>], 430.1761; found, 430.1769.

tert-Butyl Methyl(4-(3-(4-phenoxyphenyl)-1,2,4-oxadiazol-5-yl)phenyl)carbamate (19b). The title compound was synthesized from 4-((tert-butoxycarbonyl)(methyl)amino)-benzoic acid (3.58 mmol, 900 mg) 17b (4.22 mmol, 1.00 g) according to general procedure H and purified using flash chromatography (10–20% EtOAc and 1% Et<sub>3</sub>N in petroleum ether) to yield the product as yellow crystals (216 mg, 14%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.19–8.10 (m, 4H), 7.49–7.43 (m, 2H), 7.42–7.35 (m, 2H), 7.21–7.15 (m, 1H), 7.13–7.06 (m, 4H), 3.34 (s, 3H), 1.50 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 175.4, 168.6, 160.3, 156.4, 154.2, 148.0, 130.1, 129.5, 128.7, 125.2, 124.3, 121.8, 120.7, 119.9, 118.6, 81.4, 37.0, 28.5. HPLC:  $t_R$  = 8.9 min, 90% at 254 nm. HRMS m/z: calcd for C<sub>26</sub>H<sub>25</sub>N<sub>3</sub>O<sub>4</sub> + H<sup>+</sup> [M + H<sup>+</sup>], 444.1918; found, 444.1927.

*N-*(*4-*(*3-*(*4-Phenoxyphenyl*)-*1,2,4-oxadiazol-5-yl*)*phenyl*)-*acetamide* (*19c*). The title compound was synthesized from 4-acetamidobenzoic acid 17c (1.04 mmol, 186 mg) according to general procedure H and purified using flash chromatography (20% EtOAc in petroleum ether) to yield the product as an off-white solid (54 mg, 21%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): *δ* 8.19–8.09 (m, 4H), 7.72 (d, *J* = 8.5 Hz, 2H), 7.54 (br, 1H), 7.42–7.35 (m, 2H), 7.21–7.14 (m, 1H), 7.11–7.04 (m, 4H), 2.23 (s, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): *δ* 175.3, 168.7, 168.6, 160.3, 156.3, 142.1, 130.1, 129.5, 129.4, 124.3, 121.7, 120.0, 119.9, 119.6, 118.5, 24.9. HPLC:  $t_R$  = 6.6 min, 99% at 254 nm. HRMS m/z: calcd for  $C_{22}H_{17}N_3O_3 + H^+$  [M + H<sup>+</sup>], 372.1343; found, 372.1339.

5-(4-Nitrophenyl)-3-(4-phenoxyphenyl)-1,2,4-oxadiazole (19d). A solution of (Z)-N'-hydroxy-4-phenoxybenzimidamide 18 (1.5 equiv, 2.07 mmol, 473 mg) in DMF (5 mL) was treated with 4-nitrobenzoic acid 17d (1 equiv, 1.38 mmol, 231 mg), HOBt (2 equiv, 2.76 mmol, 373 mg), and EDC·HCl (2 equiv, 2.76 mmol, 531 mg); then, the solution was stirred at rt for 16 h. The temperature was increased to 100 °C, and the stirring was continued for 16 h. The reaction mixture was concentrated in vacuo, diluted with a 10% solution of aq Na<sub>2</sub>CO<sub>3</sub>, and then, EtOAc was added to extract the product. The separated organic layer was dried over anhydrous MgSO<sub>4</sub> and concentrated in vacuo; then, the residue was purified using flash chromatography (20% EtOAc in petroleum ether) to yield the product as a yellow solid (290 mg, 58%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  8.28–8.23 (m, 1H), 8.20–8.15 (m, 1H), 8.06 (d, J = 8.7 Hz, 2H), 8.02-7.96 (m, 2H), 7.47 (t, J =7.7 Hz, 2H), 7.27–7.21 (m, 1H), 7.19–7.12 (m, 4H). <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ ):  $\delta$  172.4, 167.8, 160.0, 155.3, 148.2, 134.2, 133.9, 131.6, 130.3, 129.3, 124.9, 124.5, 120.1, 119.7, 118.4, 117.5. HPLC:  $t_R = 6.9$  min, 99% at 254 nm. HRMS m/z: calcd for  $C_{20}H_{13}N_3O_4 + H^+[M + H^+]$ , 360.0979; found, 360.0980.

4-(3-(4-Phenoxyphenyl)-1,2,4-oxadiazol-5-yl)aniline (**20a**). A solution of *tert*-butyl (4-(3-(4-phenoxyphenyl)-1,2,4oxadiazol-5-yl)phenyl)carbamate 19a (0.50 mmol, 214 mg) in 4 M HCl/1,4-dioxane (4.0 mL) was stirred at rt for 6 h. After reaction completion as determined by TLC, the solution was diluted with Et<sub>2</sub>O, filtered, and further washed with Et<sub>2</sub>O. The recovered precipitate was suspended in saturated ag sodium bicarbonate solution and washed with EtOAc. The organic layers were combined and dried over MgSO4 and then concentrated in vacuo to yield the product as yellow powder (128 mg, 78%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.15–8.08 (m, 2H), 8.04–7.97 (m, 2H), 7.42–7.35 (m, 2H), 7.20–7.14 (m, 1H), 7.12–7.06 (m, 4H), 6.78–6.73 (m, 2H), 4.13 (br, 2H).  $^{13}$ C NMR (101 MHz, CDCl<sub>3</sub>):  $\delta$  176.1, 168.3, 160.1, 156.4, 150.8, 130.2, 130.1, 129.4, 124.2, 122.2, 119.8, 118.5, 114.7, 114.2. HPLC:  $t_R = 7.3$  min, 99% at 254 nm. HRMS m/z: calcd for  $C_{20}H_{15}N_3O_2 + H^+ [M + H^+]$ , 330.1237; found, 330.1245.

N-Methyl-4-(3-(4-phenoxyphenyl)-1,2,4-oxadiazol-5yl)aniline (20b). A solution of tert-butyl methyl(4-(3-(4phenoxyphenyl)-1,2,4-oxadiazol-5-yl)phenyl)carbamate 19b (0.23 mmol, 100 mg) in 4 M HCl/1,4-dioxane (2.0 mL) was stirred at rt for 6 h. After reaction completion as determined by TLC, the solution was diluted with Et<sub>2</sub>O, filtered, and further washed with Et2O. The collected precipitate was air-dried to yield the product as a white solid (66 mg, 86%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  8.10–8.01 (m, 2H), 7.93–7.85 (m, 2H), 7.49–7.42 (m, 2H), 7.26–7.20 (m, 1H), 7.17–7.10 (m, 4H), 6.73–6.66 (m, 2H), 2.77 (s, 3H). <sup>13</sup>C NMR (101 MHz, methanol- $d_4$ ):  $\delta$  176.5, 169.7, 162.0, 157.5, 147.1, 131.2, 131.1, 130.3, 125.5, 122.6, 121.1, 120.9, 120.0, 119.3, 34.6. HPLC:  $t_R = 7.9 \text{ min}$ , 99% at 254 nm. HRMS m/z: calcd for  $C_{21}H_{17}N_3O_2 + H^+[M + H^+]$ , 344.1394; found, 344.1399.

tert-Butyl (2-Methoxyethyl)(4-(3-(4-phenoxyphenyl)-1,2,4-oxadiazol-5-yl)phenyl)carbamate (21). tert-Butyl (4-(3-(4-phenoxyphenyl)-1,2,4-oxadiazol-5-yl)phenyl)carbamate 19a (0.12 mmol, 50 mg) was dissolved in anhydrous DMF (3 mL) under  $N_2$  atmosphere and NaH (60% in mineral oil, 1.2 equiv, 0.14 mmol, 5.8 mg) was added in small portions at 0 °C. The reaction mixture was stirred for

1 h at rt. 1-Bromo-2-methoxyethane (1.5 equiv, 0.18 mmol, 16  $\mu$ L) was added slowly at 0 °C; then, the mixture was stirred at rt for 2 h. The reaction mixture was diluted with aq NH<sub>4</sub>Cl solution and extracted with EtOAc. The organic layers were combined, washed with aq NH<sub>4</sub>Cl solution and brine, then dried over MgSO<sub>4</sub>, and concentrated in vacuo. The residue was purified using flash chromatography (5-10% EtOAc in petroleum ether) to yield the product as opaque oil (16 mg, 28%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.19–8.11 (m, 4H), 7.52-7.46 (m, 2H), 7.42-7.36 (m, 2H), 7.18 (ddd, J = 8.5, 2.2, 1.1 Hz, 1H), 7.12-7.07 (m, 4H), 3.86 (t, J = 5.7 Hz, 2H), 3.59 (t, J = 5.7 Hz, 2H), 3.34 (s, 3H), 1.47 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 175.4, 168.7, 160.3, 156.3, 154.2, 147.3, 130.1, 129.5, 128.8, 127.2, 124.3, 121.8, 121.4, 119.9, 118.6, 81.3, 70.56, 58.9, 49.8, 28.5. HPLC:  $t_R = 9.3 \text{ min}$ , 94% at 254 nm. HRMS m/z: calcd for  $C_{28}H_{29}N_3O_5 + H^+ [M + H^+]$ , 488.2180; found, 488.2186.

N-(2-Methoxyethyl)-4-(3-(4-phenoxyphenyl)-1,2,4-oxadiazol-5-yl)aniline (22). A solution of tert-butyl (2methoxyethyl)(4-(3-(4-phenoxyphenyl)-1,2,4-oxadiazol-5-yl)phenyl)carbamate 21 (0.031 mmol, 15 mg) in 4 M HCl/1,4dioxane (1.5 mL) was stirred at rt for 6 h. After reaction completion as determined by TLC, the solution was diluted with Et<sub>2</sub>O, filtered, and further washed with Et<sub>2</sub>O. The obtained precipitate was dissolved in acetonitrile and washed with petroleum benzines; then, the solvent was removed in vacuo. The residue was dissolved in EtOAc and washed with water; then, the organic layer dried over MgSO<sub>4</sub> and concentrated in vacuo to yield the product as a yellow solid (7.4 mg, 87%). <sup>1</sup>H NMR (400 MHz, methanol- $d_4$ ):  $\delta$  8.10– 8.05 (m, 2H), 7.95-7.90 (m, 2H), 7.41 (tt, J = 7.6, 2.2 Hz, 2H), 7.22-7.17 (m, 1H), 7.11-7.05 (m, 4H), 6.78-6.72 (m, 2H), 3.61 (t, J = 5.5 Hz, 2H), 3.39 (s, 3H), 3.37 (t, J = 5.6 Hz, 2H). <sup>13</sup>C NMR (101 MHz, methanol- $d_4$ ):  $\delta$  177.9, 169.2, 161.7, 157.6, 154.4, 131.1, 130.8, 130.2, 125.4, 123.1, 120.8, 119.3, 113.1, 112.3, 72.0, 59.0, 43.7. HPLC:  $t_R = 8.4 \text{ min}$ , 95% at 254 nm. HRMS m/z: calcd for  $C_{23}H_{21}N_3O_3 + H^+[M + H^+]$ , 388.1656; found, 388.1656.

tert-Butyl (4-(3-(4-Phenoxyphenyl)-1,2,4-oxadiazol-5-yl)-phenyl)glycinate (23). The title compound was synthesized from 4-(3-(4-phenoxyphenyl)-1,2,4-oxadiazol-5-yl)aniline 20a (0.40 mmol, 130 mg) according to a previously reported procedure and obtained as a yellow solid (56 mg, 32%). <sup>46</sup> <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.11 (d, J = 8.7 Hz, 2H), 8.03 (d, J = 8.7 Hz, 2H), 7.38 (t, J = 7.9 Hz, 2H), 7.17 (t, J = 7.4 Hz, 1H), 7.08 (dd, J = 8.6, 1.8 Hz, 4H), 6.66 (d, J = 8.7 Hz, 2H), 4.82 (bt, J = 4.6 Hz, 1H), 3.88 (d, J = 4.9 Hz, 2H), 1.51 (s, 9H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 176.1, 169.5, 168.3, 160.0, 156.5, 150.7, 130.2, 130.1, 129.4, 124.2, 122.2, 119.8, 118.5, 113.6, 112.6, 82.8, 45.9, 28.2. HPLC:  $t_R$  = 7.4 min, 98% at 254 nm. HRMS m/z: calcd for  $C_{26}H_{25}N_3O_4 + H^+$  [M + H<sup>+</sup>], 444.1918; found, 444.1915.

(4-(3-(4-Phenoxyphenyl)-1,2,4-oxadiazol-5-yl)phenyl)-glycine (24). The title compound was synthesized from *tert*-butyl (4-(3-(4-phenoxyphenyl)-1,2,4-oxadiazol-5-yl)phenyl)-glycinate 23 (0.047 mmol, 21 mg) according to general procedure E and obtained as a yellow solid (17 mg, 92%). <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ): δ 8.06 (d, J = 8.6 Hz, 2H), 7.89 (d, J = 8.7 Hz, 2H), 7.46 (t, J = 7.8 Hz, 2H), 7.23 (t, J = 7.4 Hz, 1H), 7.14 (dd, J = 8.2, 2.8 Hz, 4H), 6.99 (br, 1H), 6.74 (d, J = 8.7 Hz, 2H), 3.94 (d, J = 3.9 Hz, 2H). <sup>13</sup>C NMR (101 MHz, DMSO- $d_6$ ): δ 175.8, 171.9, 167.4, 159.6, 155.5, 152.5, 130.3, 129.5, 129.1, 124.5, 121.3, 119.7, 118.3, 112.2, 110.3,

44.0. HPLC:  $t_R = 6.3 \text{ min}$ , 95% at 254 nm. HRMS m/z: calcd for  $C_{22}H_{17}N_3O_4 + H^+[M + H^+]$ , 388.1292; found, 388.1279. 4-(3-(4-(4-(2-(Dimethylamino)ethyl)phenoxy)phenyl)-1,2,4-oxadiazol-5-yl)phenol (**25a**). tert-Butyl (4-(4-(5-(4hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenethyl)carbamate 14e (0.42 mmol, 200 mg) was dissolved in DCM (2.5 mL), and TFA (0.2 mL) was added to stir at rt for 6 h. The solvent was removed in vacuo, and the residue was treated with saturated aq sodium bicarbonate solution and then sonicated until a precipitate formed. The suspension was treated with 10% aq citric acid solution until pH 6 and then extracted with EtOAc three times. The organic layers were combined, dried over MgSO<sub>4</sub>, and the solvent was removed in vacuo to yield 4-(3-(4-(4-(2-aminoethyl)phenoxy)phenyl)-1,2,4-oxadiazol-5-yl)phenol 15e as the neutral species (158 mg, 100%). <sup>1</sup>H NMR (400 MHz, methanol- $d_4$ ):  $\delta$  8.11–8.06 (m, 2H), 8.06-8.01 (m, 2H), 7.38-7.27 (m, 2H), 7.14-7.03 (m, 4H), 7.00-6.92 (m, 2H), 3.25-3.15 (m, 2H), 3.03-2.94 (m, 2H).

To a suspension of 15e (1 equiv, 0.13 mmol, 50 mg) in acetonitrile (2 mL) and water (0.5 mL) was added 37% aq formaldehyde solution (12 equiv, 1.6 mmol, 130  $\mu$ L). NaBH(OAc)<sub>3</sub> (4.8 equiv, 0.64 mmol, 140 mg) was added slowly, and the reaction was stirred at rt for 50 min. The reaction mixture was quenched with saturated aq sodium bicarbonate solution and then extracted several times with a 9:1 solution of EtOAc/isopropanol. The organic layers were combined and concentrated in vacuo. The residue was treated with small amounts of EtOAc and saturated ag sodium bicarbonate solution until a precipitate formed. A sodium citrate buffer of pH  $\sim$  6.5 was added slowly until the solution became neutral and the precipitate dissolved. The aqueous layer was washed with EtOAc; then, the organic layers were combined and set aside. The aqueous layer was concentrated in vacuo to obtain cruder residue. The sequential process of saturated ag sodium bicarbonate solution and sodium citrate buffer additions followed by EtOAc extraction was repeated until no further precipitation was observed. The combined organic layers were dried over MgSO<sub>4</sub>, and the solvent was removed in vacuo to yield the product as an off-white solid (39 mg, 74%). <sup>1</sup>H NMR (400 MHz, methanol- $d_4$ ):  $\delta$  8.11–8.01 (m, 4H), 7.32–7.27 (m, 2H), 7.10–7.05 (m, 2H), 7.05–7.01 (m, 2H), 6.99-6.94 (m, 2H), 2.90-2.82 (m, 2H), 2.77-2.71 (m, 2H), 2.44 (s, 6H). <sup>13</sup>C NMR (101 MHz, methanol- $d_4$ ):  $\delta$ 177.4, 169.5, 163.7, 162.0, 155.9, 136.8, 131.4, 131.2, 130.2, 122.8, 121.1, 119.1, 117.2, 116.4, 62.0, 45.1, 33.6. HPLC:  $t_{\rm R}$  = 6.0 min, 99% at 254 nm. HRMS m/z: calcd for  $C_{24}H_{23}N_3O_3 +$  $H^{+}$  [M +  $H^{+}$ ], 402.1812; found, 402.1812.

2-(4-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3-yl)-phenoxy)phenyl)-N,N,N-trimethylethan-1-aminium lodide (26a). 4-(3-(4-(4-(2-(Dimethylamino)ethyl)phenoxy)-phenyl)-1,2,4-oxadiazol-5-yl)phenol 25a (1 equiv, 0.092 mmol, 37 mg) was dissolved in a 2:1 solution of DCM/MeOH (3 mL) under N<sub>2</sub> atmosphere, and iodomethane (5 equiv, 0.46 mmol, 29 μL) was added. The reaction mixture was stirred for 24 h at rt. The solvent was removed in vacuo, and then, the residue was suspended in Et<sub>2</sub>O and filtered. The precipitate was collected to yield the product as pink crystals (50 mg, 100%). <sup>1</sup>H NMR (400 MHz, methanol- $d_4$ ): δ 8.15 – 8.10 (m, 2H), 8.10 – 8.04 (m, 2H), 7.42 (d, J = 8.6 Hz, 2H), 7.11 (dd, J = 9.8, 1.7 Hz, 4H), 7.01 – 6.96 (m, 2H), 3.65 – 3.59 (m, 2H), 3.26 (s, 9H), 3.22 – 3.15 (m, 2H). <sup>13</sup>C NMR (101 MHz, methanol- $d_4$ ): δ 177.4, 169.4, 163.6, 161.5, 156.9, 132.8,

131.8, 131.2, 130.3, 123.2, 121.3, 119.4, 117.2, 116.4, 68.4, 53.8, 53.8, 53.7, 29.6. HPLC:  $t_R = 6.1$  min, 100% at 254 nm. HRMS m/z: calcd for  $C_{25}H_{26}N_3O_3$  [M<sup>+</sup>], 416.1969; found, 416.1981.

4-(3-(4-(4-(3-(Dimethylamino)propyl)phenoxy)phenyl)-1,2,4-oxadiazol-5-yl)phenol Hydrochloride (25b). tert-Butyl (3-(4-(4-(5-(4-hydroxyphenyl)-1,2,4-oxadiazol-3-yl)phenoxy)phenyl)propyl)carbamate 14h (0.20 mmol, 100 mg) was dissolved in DCM (1 mL) and TFA (0.1 mL) was added to stir at rt for 7 h. The solvent was removed in vacuo, and the residue was treated with saturated aq sodium bicarbonate solution and then sonicated until a precipitate was formed. The suspension was treated with 10% aq citric acid solution until pH 6 and then was extracted with EtOAc three times. The organic layers were combined, dried over MgSO<sub>4</sub>, and the solvent was removed in vacuo to yield 4-(3-(4-(4-(3aminopropyl)phenoxy)phenyl)-1,2,4-oxadiazol-5-yl)phenol 15h as the neutral species (79 mg, 100%). <sup>1</sup>H NMR (400 MHz, methanol- $d_4$ ):  $\delta$  8.11–8.03 (m, 4H), 7.30 (d, J = 8.6 Hz, 2H), 7.10-7.02 (m, 4H), 7.00-6.94 (m, 2H), 2.99-2.92 (m, 2H), 2.79-2.71 (m, 2H), 1.99 (dt, J = 15.5, 7.7 Hz, 2H). To a suspension of 15h (1 equiv, 0.052 mmol, 20 mg) in acetonitrile (0.9 mL) and water (0.1 mL) was added 37% aq formaldehyde solution (12 equiv, 0.62 mmol, 50  $\mu$ L). NaBH(OAc)<sub>3</sub> (4.8 equiv, 0.25 mmol, 53 mg) was added slowly, and the reaction was stirred at rt for 50 min. The reaction mixture was quenched with saturated aq sodium bicarbonate solution and then extracted several times with a 9:1 solution of EtOAc/ isopropanol. The organic layers were combined and concentrated in vacuo. The residue was treated with small amounts of EtOAc and saturated aq sodium bicarbonate solution until a precipitate formed. A sodium citrate buffer of pH  $\sim$  6.5 was added slowly until the solution became neutral and the precipitate dissolved. The aqueous layer was washed with EtOAc; then the organic layers were combined and set aside. The aqueous layer was concentrated in vacuo to obtain cruder residue. The sequential process of saturated aq sodium bicarbonate solution and sodium citrate buffer additions followed by EtOAc extraction was repeated until no further precipitation was observed. The combined organic layers were dried over MgSO<sub>4</sub>, and the solvent was removed in vacuo to obtain the impure product as the neutral species. The residue was dissolved in 4 M HCl/1,4-dioxane (1.5 mL) and stirred at rt for 6 h. The solution was diluted with Et<sub>2</sub>O, filtered, and the solid was further washed with Et<sub>2</sub>O. The filtered solid was collected to yield the product as an off-white solid (2.4 mg, 10%). <sup>1</sup>H NMR (400 MHz, methanol- $d_4$ ):  $\delta$  8.14–8.04 (m, 4H), 7.33 (d, J = 8.5 Hz, 2H), 7.13-7.04 (m, 4H), 7.02-6.96(m, 2H), 3.21-3.14 (m, 2H), 2.92 (s, 6H), 2.76 (t, J = 7.6 Hz,2H), 2.13–2.03 (m, 2H).  $^{13}$ C NMR (101 MHz, methanol- $d_4$ ):  $\delta$  177.4, 169.4, 163.6, 161.9, 156.1, 137.6, 131.2, 131.1, 130.2, 122.9, 121.2, 119.1, 117.2, 116.4, 58.6, 43.5, 32.7, 27.5. HPLC:  $t_{\rm R} = 6.3$  min, 96% at 254 nm. HRMS m/z: calcd for  $C_{25}H_{25}N_3O_3 + H^+ [M + H^+]$ , 416.1969; found, 416.1979.

3-(4-(4-(5-(4-Hydroxyphenyl)-1,2,4-oxadiazol-3-yl)-phenoxy)phenyl-N,N,N-trimethylpropan-1-aminium lodide (26b). The neutral species of 4-(3-(4-(4-(3-(dimethylamino)-propyl)phenoxy)phenyl)-<math>1,2,4-oxadiazol-5-yl)phenol 25b (1 equiv, 0.072 mmol, 30 mg) was dissolved in a 2:1 solution of DCM/MeOH (2 mL) under  $N_2$  atmosphere and iodomethane (5 equiv, 0.36 mmol, 23  $\mu$ L) was added. The reaction mixture was stirred for 24 h at rt. The solvent was removed in vacuo; then, the residue was suspended in Et<sub>2</sub>O and filtered.

The precipitate was washed with EtOAc, air-dried, and then collected to yield the product as yellow powder (23 mg, 58%).  $^{1}$ H NMR (400 MHz, methanol- $d_4$ ):  $\delta$  8.14–8.03 (m, 4H), 7.38–7.33 (m, 2H), 7.13–7.05 (m, 4H), 7.02–6.96 (m, 2H), 3.44–3.39 (m, 2H), 3.17 (s, 9H), 2.76 (t, J=7.6 Hz, 2H), 2.22–2.12 (m, 2H).  $^{13}$ C NMR (101 MHz, methanol- $d_4$ ):  $\delta$  177.4, 169.4, 163.6, 161.9, 156.1, 137.4, 131.2, 131.2, 130.2, 122.9, 121.2, 119.1, 117.2, 116.4, 67.4, 67.4, 67.4, 53.7, 53.7, 53.6, 32.5, 25.8. HPLC:  $t_{\rm R}=6.4$  min, 95% at 254 nm. HRMS m/z: calcd for  ${\rm C_{26}H_{28}N_3O_3}$  [M $^+$ ], 430.2125; found, 430.2111.

Antimicrobial Susceptibility Testing. Antimicrobial susceptibility testing was performed using CLSI methods.<sup>47</sup> Specifically, MICs were determined by broth microdilution assays as described previously for C. difficile<sup>48</sup> and E. faecium.<sup>31</sup> In brief, a 2-fold dilution series (from 128 to 1  $\mu$ g/mL) of 1,2,4 oxadiazole analogues, vancomycin (C. difficile comparator control), or daptomycin (E. faecium comparator control) was made in 100  $\mu$ L volumes of either SDW (C. difficile) or cationadjusted Mueller-Hinton broth (CAMHB) (E. faecium) purchased from Thermo Fisher (additionally supplemented with 50  $\mu$ g/mL Ca<sup>2+</sup> for daptomycin assays) in a 96-well plate (Corning) and an inoculum of 100  $\mu$ L from an overnight C. difficile or E. faecium broth culture adjusted to  $5 \times 10^5$  CFU/ mL in supplemented, pre-reduced Brucella broth (C. difficile) or CAMHB (E. faecium) added. After 48 h incubation under anaerobic conditions (C. difficile) or 24 h incubation under aerobic conditions (E. faecium), the MIC was defined as the lowest antimicrobial concentration that inhibited visible growth. All MIC testing was performed in biological triplicate.

**Time-Kill Assays.** Time-kill assays were performed with *C*. difficile strain EDN0008 (NAP1/027) or E. faecium strain Aus0085 (ST203), using supplemented brain heart infusion broth (BHIBS) for C. difficile or CAMHB for E. faecium. BHIBS—BHIB supplemented with 5 g/L yeast extract, 0.1% (w/v) L-cysteine, and 0.3% (w/v) glucose. Broths were supplemented with either DMSO (vehicle) or with 1× MIC, 2× MIC, 4× MIC, and 8× MIC of 1,2,4-oxadiazole or vancomycin (8 µg/mL), daptomycin (16 µg/mL) (additionally supplemented with 50  $\mu$ g/mL Ca<sup>2+</sup> for daptomycin assays) or benzalkonium chloride (24 or 16  $\mu$ g/mL) as controls. Broths were inoculated with overnight bacterial broth cultures and adjusted to  $1 \times 10^6$  CFU/mL. Samples withdrawn at 0, 1, 3, 24, and 48 h post-inoculation were serially diluted in phosphate-buffered saline and plated onto either supplemented brain heart infusion agar and incubated overnight at 37 °C in a DG250 anaerobic chamber (Don Whitley Scientific) for C. difficile or on BHIA (Oxoid) and grown overnight at 37 °C aerobically for E. faecium before colony enumeration was performed. All assays were performed using three independent biological replicates, with each test strain being purity plated from frozen glycerol stocks three independent times and each bacterial culture used in the time-kill assay then being derived from a single colony picked from one of the three separate purity plates.

**Haemolysis Assay.** Fresh whole human blood was collected into K2-EDTA-coated Vacutainer tubes and pelleted at 500 g for 5 min. The red blood cell pellet was washed twice in saline solution followed by a third wash in phosphate-buffered saline (PBS) before being diluted 1:50 in PBS. Compound **15e** was prepared to  $20\times$  concentration in DMSO, and a  $10~\mu$ L aliquot of each concentration was added to  $190~\mu$ L of diluted blood in a 96-well U-bottom plate (Corning) giving a final concentration range of 0.125-128~mg/L. The plate was

then incubated at 37 °C for 1 h. Negative (vehicle) controls were incubated with DMSO, and positive controls were incubated with 1% (v/v) Triton X-100. The plate was centrifuged at 500g after the incubation to pellet intact erythrocytes, and 100  $\mu$ L of the supernatant from each well was then collected and transferred to a 96-well flat bottom plate (Corning). The absorbance of each supernatant sample was then measured at 450 nm using CLARIOstar plus (BMG LABTECH). All assays were performed using three independent biological replicates.

Caco-2 Cell Permeability. The apical to basolateral apparent permeability (A–B  $P_{\rm app}$ ) was assessed in Caco-2 cells (passage 39) seeded onto 0.3 cm<sup>2</sup> polycarbonate filter transwells at a density of  $6 \times 10^4$  cells/well. Experiments were conducted 23 days post-seeding. Experiments were performed using pH 7.4 Hanks balanced salt solution (containing 20 mM N-(2-hydroxyethyl)piperazine-N'-ethanesulfonic acid) with the donor solution spiked with the test compound. Sample aliquots were taken 5-6 times over 120 min, and the compound flux were determined by assessing compound concentration using LCMS. The apical to basolateral apparent permeability coefficient (A-B P<sub>app</sub>) for each compound was determined using the equation,  $P_{\text{app}}(\text{cm/s}) = dQ/dt \times 1/(A \times 1)$  $C_{\text{initial donor}}$ ), where dQ/dt = apparent steady-state transport rate ( $\mu$ mol/s), A = surface area of Caco-2 monolayer (0.3) cm<sup>2</sup>), and  $C_{\text{initial donor}}$  = donor chamber concentration at the start of the experiment ( $\mu$ mol/cm<sup>3</sup>). Transepithelial electrical resistance (TEER) readings, high and low permeability markers, and a multidrug resistance protein-mediated efflux marker were included in the study to assess the integrity of the cell monolayers. The TEER values for transwells used in the Caco-2 permeability study ranged from 457 to 551  $\Omega$ cm<sup>2</sup>, indicating the presence of confluent monolayers. For lucifer yellow,  $P_{\rm app}$  values of <1  $\times$  10<sup>-6</sup> cm/s were yielded in the acceptor solution. For propranolol, an A–B  $P_{\rm app}$  value of 42  $\pm$  $2.5 \times 10^{-6}$  cm/s was obtained and for rhodamine 123, an A–B  $P_{\rm app}$  value of 1.6  $\pm$  0.13  $\times$  10<sup>-6</sup> cm/s, B-A  $P_{\rm app}$  value of 12  $\pm$  $2.7 \times 10^{-6}$  cm/s, and an efflux ratio of 7.3 were obtained, all of which compared well with in-house validation data. Values are presented as the mean  $\pm$  SD of n = 3 transwells.

**Pharmacokinetic Studies.** Experimental procedures are summarized in the Supporting Information.

#### ASSOCIATED CONTENT

#### Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acsomega.1c06294.

Syntheses and characterization of intermediates 13a-i, example NMR spectra, HPLC profiles, and study design and formulations for the pharmacokinetic studies (PDF) Molecular formula strings for final compounds (XLSX)

#### **AUTHOR INFORMATION**

#### **Corresponding Authors**

Glen P. Carter — Department of Microbiology & Immunology, Peter Doherty Institute for Infection and Immunity, The University of Melbourne, Melbourne, Victoria 3000, Australia; Email: glen.carter@unimelb.edu.au

Jonathan B. Baell — School of Pharmaceutical Sciences, Nanjing Tech University, Nanjing 211816, People's Republic of China; Medicinal Chemistry, Monash Institute of Pharmaceutical Sciences and Australian Translational Medicinal Chemistry Facility (ATMCF), Monash University, Parkville, Victoria 3052, Australia; orcid.org/0000-0003-2114-8242; Phone: +61 3 9903 9044; Email: jonathan.baell@monash.edu

#### **Authors**

Noel P. Pitcher – Medicinal Chemistry, Monash Institute of Pharmaceutical Sciences, Monash University, Parkville, Victoria 3052, Australia

Jitendra R. Harjani – Medicinal Chemistry, Monash Institute of Pharmaceutical Sciences, Monash University, Parkville, Victoria 3052, Australia

Yichao Zhao – Medicinal Chemistry, Monash Institute of Pharmaceutical Sciences, Monash University, Parkville, Victoria 3052, Australia

Jianwen Jin – Medicinal Chemistry, Monash Institute of Pharmaceutical Sciences, Monash University, Parkville, Victoria 3052, Australia

Daniel R. Knight — School of Medical and Health Sciences, Edith Cowan University, Joondalup, Western Australia 6027, Australia; School of Biomedical Sciences, Faculty of Health and Medical Sciences, The University of Western Australia, Queen Elizabeth II Medical Centre, Nedlands, Western Australia 6009, Australia; Medical, Molecular and Forensic Sciences, Murdoch University, Murdoch, Western Australia 6150, Australia; Department of Microbiology, PathWest Laboratory Medicine, Queen Elizabeth II Medical Centre, Nedlands, Western Australia 6009, Australia

Lucy Li — Department of Microbiology & Immunology, Peter Doherty Institute for Infection and Immunity, The University of Melbourne, Melbourne, Victoria 3000, Australia

Papanin Putsathit — School of Medical and Health Sciences, Edith Cowan University, Joondalup, Western Australia 6027, Australia; School of Biomedical Sciences, Faculty of Health and Medical Sciences, The University of Western Australia, Queen Elizabeth II Medical Centre, Nedlands, Western Australia 6009, Australia; Medical, Molecular and Forensic Sciences, Murdoch University, Murdoch, Western Australia 6150, Australia; Department of Microbiology, PathWest Laboratory Medicine, Queen Elizabeth II Medical Centre, Nedlands, Western Australia 6009, Australia

Thomas V. Riley — School of Medical and Health Sciences, Edith Cowan University, Joondalup, Western Australia 6027, Australia; School of Biomedical Sciences, Faculty of Health and Medical Sciences, The University of Western Australia, Queen Elizabeth II Medical Centre, Nedlands, Western Australia 6009, Australia; Medical, Molecular and Forensic Sciences, Murdoch University, Murdoch, Western Australia 6150, Australia; Department of Microbiology, PathWest Laboratory Medicine, Queen Elizabeth II Medical Centre, Nedlands, Western Australia 6009, Australia

Complete contact information is available at: https://pubs.acs.org/10.1021/acsomega.1c06294

#### **Author Contributions**

 $^{\circ}$ G.P.C and J.B.B. contributed equally to this paper.

#### Notes

The authors declare no competing financial interest.

#### ACKNOWLEDGMENTS

The National Health and Medical Research Council of Australia (NHMRC) is thanked for Fellowship support for J.B.B. (2012–2016 Senior Research Fellowship #1020411, 2017-Principal Research Fellowship #1117602) and funding for G.P.C. (Ideas grant GNT1185213). The Australian Translational Medicinal Chemistry Facility (ATMCF) within Monash Institute of Pharmaceutical Sciences (MIPS), acknowledges the support of the Australian Government's National Collaborative Research Infrastructure Strategy (NCRIS) program via Therapeutic Innovation Australia (TIA).

#### ABBREVIATIONS

Aq, aqueous; Boc, tert-butyloxycarbonyl; CDC, Centers for Disease Control and Prevention; CDI, C. difficile infections; DCM, dichloromethane; DIPEA, N,N-diisopropylethylamine; DMAP, 4-(N,N-dimethylamino)pyridine; DMF, dimethylformamide; DMSO, dimethyl sulfoxide; EDC·HCl, N-(3dimethylaminopropyl)-N'-ethylcarbodiimide hydrochloride; ESKAPE, Enterococcus faecium, Staphylococcus aureus, Klebsiella pneumoniae, Acinetobacter baumanni, Pseudomonas aeruginosa, and Enterobacter species; Et<sub>3</sub>N, triethylamine; Et<sub>2</sub>O, diethyl ether; EtOAc, ethyl acetate; equiv, equivalents; GI, gastrointestinal; HOBt, 1-hydroxybenzotriazole hydrate; MDR, multidrug-resistant; MeOH, methanol; MIC, minimum inhibitory concentration; MRSA, methicillin-resistant S. aureus; rt, room temperature; SAR, structure-activity relationship; TBS, tert-butyldimethylsilyl; t-Bu, tert-butyl; TFA, trifluoroacetic acid; TLC, thin-layer chromatography; VREfm, vancomycin-resistant E. faecium

#### REFERENCES

- (1) Van der Waaij, D.; Berghuis-de Vries, J. M.; Lekkerkerk-van der Wees, J. E. C. Colonization Resistance of the Digestive Tract in Conventional and Antibiotic-Treated Mice. J. Hyg. 1971, 69, 405–411.
- (2) Vollaard, E. J.; Clasener, H. A. Colonization Resistance. *Antimicrob. Agents Chemother.* **1994**, 38, 409–414.
- (3) Sekirov, I.; Russell, S. L.; Antunes, L. C. M.; Finlay, B. B. Gut Microbiota in Health and Disease. *Physiol. Rev.* **2010**, *90*, 859–904.
- (4) Lange, K.; Buerger, M.; Stallmach, A.; Bruns, T. Effects of Antibiotics on Gut Microbiota. *Dig. Dis.* **2016**, *34*, 260–268.
- (5) Owens, R. C.; Donskey, C. J.; Gaynes, R. P.; Loo, V. G.; Muto, C. A. Antimicrobial-Associated Risk Factors for Clostridium Difficile Infection. *Clin. Infect. Dis.* **2008**, *46*, S19—S31.
- (6) Theriot, C. M.; Young, V. B. Interactions Between the Gastrointestinal Microbiome and Clostridium Difficile. *Annu. Rev. Microbiol.* **2015**, *69*, 445–461.
- (7) Ubeda, C.; Taur, Y.; Jenq, R. R.; Equinda, M. J.; Son, T.; Samstein, M.; Viale, A.; Socci, N. D.; van den Brink, M. R. M.; Kamboj, M.; Pamer, E. G. Vancomycin-Resistant *Enterococcus* Domination of Intestinal Microbiota Is Enabled by Antibiotic Treatment in Mice and Precedes Bloodstream Invasion in Humans. *J. Clin. Invest.* 2010, 120, 4332–4341.
- (8) Arias, C. A.; Murray, B. E. The Rise of the *Enterococcus*: Beyond Vancomycin Resistance. *Nat. Rev. Microbiol.* **2012**, *10*, 266–278.
- (9) Smits, W. K.; Lyras, D.; Lacy, D. B.; Wilcox, M. H.; Kuijper, E. J. Clostridium Difficile Infection. *Nat. Rev. Dis. Primers* **2016**, *2*, 16020. (10) Czepiel, J.; Dróżdż, M.; Pituch, H.; Kuijper, E. J.; Perucki, W.; Mielimonka, A.; Goldman, S.; Wultańska, D.; Garlicki, A.; Biesiada, G. *Clostridium Difficile* Infection: Review. *Eur. J. Clin. Microbiol. Infect. Dis.* **2019**, *38*, 1211–1221.
- (11) CDC. Antibiotic Resistance Threats in the United States, 2019. https://www.cdc.gov/drugresistance/pdf/threats-report/2019-arthreats-report-508.pdf (accessed Feb 18, 2020).
- (12) Zar, F. A.; Bakkanagari, S. R.; Moorthi, K. M. L. S. T.; Davis, M. B. A Comparison of Vancomycin and Metronidazole for the

- Treatment of Clostridium Difficile-Associated Diarrhea, Stratified by-Disease Severity. Clin. Infect. Dis. 2007, 45, 302–307.
- (13) Vardakas, K. Z.; Polyzos, K. A.; Patouni, K.; Rafailidis, P. I.; Samonis, G.; Falagas, M. E. Treatment Failure and Recurrence of *Clostridium Difficile* Infection Following Treatment with Vancomycin or Metronidazole: A Systematic Review of the Evidence. *Int. J. Antimicrob. Agents* **2012**, *40*, 1–8.
- (14) McDonald, L. C.; Gerding, D. N.; Johnson, S.; Bakken, J. S.; Carroll, K. C.; Coffin, S. E.; Dubberke, E. R.; Garey, K. W.; Gould, C. V.; Kelly, C.; Loo, V.; Shaklee Sammons, J.; Sandora, T. J.; Wilcox, M. H. Clinical Practice Guidelines for *Clostridium Difficile* Infection in Adults and Children: 2017 Update by the Infectious Diseases Society of America (IDSA) and Society for Healthcare Epidemiology of America (SHEA). *Clin. Infect. Dis.* 2018, 66, 987–994.
- (15) Louie, T. J.; Miller, M. A.; Mullane, K. M.; Weiss, K.; Lentnek, A.; Golan, Y.; Gorbach, S.; Sears, P.; Shue, Y.-K. Fidaxomicin versus Vancomycin for *Clostridium Difficile* Infection. N. Engl. J. Med. 2011, 364, 422–431.
- (16) Weiner, L. M.; Webb, A. K.; Limbago, B.; Dudeck, M. A.; Patel, J.; Kallen, A. J.; Edwards, J. R.; Sievert, D. M. Antimicrobial-Resistant Pathogens Associated With Healthcare-Associated Infections: Summary of Data Reported to the National Healthcare Safety Network at the Centers for Disease Control and Prevention, 2011-2014. *Infect. Control Hosp. Epidemiol.* 2016, 37, 1288–1301.
- (17) O'Driscoll, T.; Crank, C. W. Vancomycin-Resistant Enterococcal Infections: Epidemiology, Clinical Manifestations, and Optimal Management. *Infect. Drug Resist.* **2015**, *8*, 217–230.
- (18) Hegstad, K.; Mikalsen, T.; Coque, T. M.; Werner, G.; Sundsfjord, A. Mobile Genetic Elements and Their Contribution to the Emergence of Antimicrobial Resistant *Enterococcus Faecalis* and *Enterococcus Faecium*. Clin. Microbiol. Infect. 2010, 16, 541–554.
- (19) WHO. Global Priority List of Antibiotic-Resistant Bacteria to Guide Research, Discovery, and Development of New Antibiotics. https://www.who.int/medicines/publications/global-priority-list-antibiotic-resistant-bacteria/en/#.X0ScqvDj7S0.mendeley (accessed Feb 18, 2020).
- (20) Deshpande, L. M.; Ashcraft, D. S.; Kahn, H. P.; Pankey, G.; Jones, R. N.; Farrell, D. J.; Mendes, R. E. Detection of a New Cfr-like Gene, Cfr(B), in *Enterococcus Faecium* Isolates Recovered from Human Specimens in the United States as Part of the SENTRY Antimicrobial Surveillance Program. *Antimicrob. Agents Chemother.* 2015, 59, 6256–6261.
- (21) Lellek, H.; Franke, G. C.; Ruckert, C.; Wolters, M.; Wolschke, C.; Christner, M.; Büttner, H.; Alawi, M.; Kröger, N.; Rohde, H. Emergence of Daptomycin Non-Susceptibility in Colonizing Vancomycin-Resistant *Enterococcus Faecium* Isolates during Daptomycin Therapy. *Int. J. Med. Microbiol.* **2015**, 305, 902–909.
- (22) Hachem, R.; Raad, I. Failure of Oral Antimicrobial Agents in Eradicating Gastrointestinal Colonization with Vancomycin-Resistant Enterococci. *Infect. Control Hosp. Epidemiol.* **2002**, 23, 43–44.
- (23) Stiefel, U.; Pultz, N. J.; Helfand, M. S.; Donskey, C. J. Efficacy of Oral Ramoplanin for Inhibition of Intestinal Colonization by Vancomycin-Resistant Enterococci in Mice. *Antimicrob. Agents Chemother.* **2004**, *48*, 2144–2148.
- (24) AbdelKhalek, A.; Abutaleb, N. S.; Elmagarmid, K. A.; Seleem, M. N. Repurposing Auranofin as an Intestinal Decolonizing Agent for Vancomycin-Resistant Enterococci. *Sci. Rep.* **2018**, *8*, 8353.
- (25) AbdelKhalek, A.; Abutaleb, N. S.; Mohammad, H.; Seleem, M. N. Repurposing Ebselen for Decolonization of Vancomycin-Resistant Enterococci (VRE). *PLoS One* **2018**, *13*, No. e0199710.
- (26) O'Daniel, P. I.; Peng, Z.; Pi, H.; Testero, S. A.; Ding, D.; Spink, E.; Leemans, E.; Boudreau, M. A.; Yamaguchi, T.; Schroeder, V. A.; Wolter, W. R.; Llarrull, L. I.; Song, W.; Lastochkin, E.; Kumarasiri, M.; Antunes, N. T.; Espahbodi, M.; Lichtenwalter, K.; Suckow, M. A.; Vakulenko, S.; Mobashery, S.; Chang, M. Discovery of a New Class of Non- $\beta$ -Lactam Inhibitors of Penicillin-Binding Proteins with Gram-Positive Antibacterial Activity. *J. Am. Chem. Soc.* **2014**, *136*, 3664–3672.

- (27) Rice, L. B. Federal Funding for the Study of Antimicrobial Resistance in Nosocomial Pathogens: No ESKAPE. *J. Infect. Dis.* **2008**, *197*, 1079–1081.
- (28) Spink, E.; Ding, D.; Peng, Z.; Boudreau, M. A.; Leemans, E.; Lastochkin, E.; Song, W.; Lichtenwalter, K.; O'Daniel, P. I.; Testero, S. A.; Pi, H.; Schroeder, V. A.; Wolter, W. R.; Antunes, N. T.; Suckow, M. A.; Vakulenko, S.; Chang, M.; Mobashery, S. Structure-Activity Relationship for the Oxadiazole Class of Antibiotics. *J. Med. Chem.* 2015, 58, 1380–1389.
- (29) Ding, D.; Boudreau, M. A.; Leemans, E.; Spink, E.; Yamaguchi, T.; Testero, S. A.; O'Daniel, P. I.; Lastochkin, E.; Chang, M.; Mobashery, S. Exploration of the Structure-Activity Relationship of 1,2,4-Oxadiazole Antibiotics. *Bioorg. Med. Chem. Lett.* **2015**, 25, 4854–4857.
- (30) Janardhanan, J.; Chang, M.; Mobashery, S. The Oxadiazole Antibacterials. *Curr. Opin. Microbiol.* **2016**, 33, 13–17.
- (31) Carter, G. P.; Harjani, J. R.; Li, L.; Pitcher, N. P.; Nong, Y.; Riley, T. V.; Williamson, D. A.; Stinear, T. P.; Baell, J. B.; Howden, B. P. 1,2,4-Oxadiazole Antimicrobials Act Synergistically with Daptomycin and Display Rapid Kill Kinetics against MDR *Enterococcus Faecium*. J. Antimicrob. Chemother. 2018, 73, 1562–1569.
- (32) Schanker, L. S.; Tocco, D. J.; Brodie, B. B.; Hogben, C. A. Absorption of Drugs from the Rat Small Intestine. *J. Pharmacol. Exp. Ther.* **1958**, 123, 81–88.
- (33) Manallack, D. T.; Prankerd, R. J.; Yuriev, E.; Oprea, T. I.; Chalmers, D. K. The Significance of Acid/Base Properties in Drug Discovery. *Chem. Soc. Rev.* **2013**, 42, 485–496.
- (34) Cherian, P. T.; Wu, X.; Yang, L.; Scarborough, J. S.; Singh, A. P.; Alam, Z. A.; Lee, R. E.; Hurdle, J. G. Gastrointestinal Localization of Metronidazole by a Lactobacilli-Inspired Tetramic Acid Motif Improves Treatment Outcomes in the Hamster Model of *Clostridium Difficile* Infection. *J. Antimicrob. Chemother.* **2015**, *70*, 3061–3069.
- (35) Poulain, R. F.; Tartar, A. L.; Déprez, B. P. Parallel Synthesis of 1,2,4-Oxadiazoles from Carboxylic Acids Using an Improved, Uronium-Based, Activation. *Tetrahedron Lett.* **2001**, 42, 1495–1498.
- (36) Manglik, A.; Lin, H.; Aryal, D. K.; McCorvy, J. D.; Dengler, D.; Corder, G.; Levit, A.; Kling, R. C.; Bernat, V.; Hübner, H.; Huang, X.-P.; Sassano, M. F.; Giguère, P. M.; Löber, S.; Da Duan, R. C.; Scherrer, G.; Kobilka, B. K.; Gmeiner, P.; Roth, B. L.; Shoichet, B. K.; Scherrer, G.; Kling, R. C. Structure-Based Discovery of Opioid Analgesics with Reduced Side Effects. *Nature* **2016**, *537*, 185–190.
- (37) Wach, J.-Y.; Malisova, B.; Bonazzi, S.; Tosatti, S.; Textor, M.; Zürcher, S.; Gademann, K. Protein-Resistant Surfaces through Mild Dopamine Surface Functionalization. *Chem. Eur. J.* **2008**, *14*, 10579–10584.
- (38) Heikkinen, A. T.; Mönkkönen, J.; Korjamo, T. Kinetics of Cellular Retention during Caco-2 Permeation Experiments: Role of Lysosomal Sequestration and Impact on Permeability Estimates. *J. Pharmacol. Exp. Ther.* **2009**, 328, 882–892.
- (39) Hama, T.; Liu, X.; Culkin, D. A.; Hartwig, J. F. Palladium-Catalyzed α-Arylation of Esters and Amides under More Neutral Conditions. *J. Am. Chem. Soc.* **2003**, *125*, 11176–11177.
- (40) Meier, C.; Ruppel, M. F. H.; Vukadinović, D.; Balzarini, J. Second Generation of CycloSal-Pro-Nucleotides with Esterase-Cleavable Sites: The "Lock-In"-Concept. *Nucleos Nucleot. Nucleic Acids* **2004**, *23*, 89–115.
- (41) Hama, T.; Ge, S.; Hartwig, J. F. Palladium-Catalyzed  $\alpha$ -Arylation of Zinc Enolates of Esters: Reaction Conditions and Substrate Scope. *J. Org. Chem.* **2013**, *78*, 8250–8266.
- (42) Winiarski, Ł.; Oleksyszyn, J.; Sieńczyk, M. Human Neutrophil Elastase Phosphonic Inhibitors with Improved Potency of Action. *J. Med. Chem.* **2012**, *55*, 6541–6553.
- (43) Charton, J.; Deprez-Poulain, R.; Hennuyer, N.; Tailleux, A.; Staels, B.; Deprez, B. Novel Non-Carboxylic Acid Retinoids: 1,2,4-Oxadiazol-5-One Derivatives. *Bioorg. Med. Chem. Lett.* **2009**, 19, 489–492.
- (44) Zhang, X.; Zhang, J.; Su, M.; Zhou, Y.; Chen, Y.; Li, J.; Lu, W. Design, Synthesis and Biological Evaluation of 4'-Demethyl-4-

- Deoxypodophyllotoxin Derivatives as Novel Tubulin and Histone Deacetylase Dual Inhibitors. RSC Adv. 2014, 4, 40444–40448.
- (45) Gillespie, R. J.; Bamford, S. J.; Botting, R.; Comer, M.; Denny, S.; Gaur, S.; Griffin, M.; Jordan, A. M.; Knight, A. R.; Lerpiniere, J.; Leonardi, S.; Lightowler, S.; McAteer, S.; Merrett, A.; Misra, A.; Padfield, A.; Reece, M.; Saadi, M.; Selwood, D. L.; Stratton, G. C.; Surry, D.; Todd, R.; Tong, X.; Ruston, V.; Upton, R.; Weiss, S. M. Antagonists of the Human A2A Adenosine Receptor. 4. Design, Synthesis, and Preclinical Evaluation of 7-Aryltriazolo[4,5-d]-Pyrimidines. J. Med. Chem. 2009, 52, 33–47.
- (46) Roy, O.; Caumes, C.; Esvan, Y.; Didierjean, C.; Faure, S.; Taillefumier, C. The Tert-Butyl Side Chain: A Powerful Means to Lock Peptoid Amide Bonds in the Cis Conformation. *Org. Lett.* **2013**, 15, 2246–2249.
- (47) Clinical Laboratory Standards Institute (CLSI). *Performance Standards for Antimicrobial Susceptibility Testing*, 26th ed.; Clinical and Laboratory Standards 416 Institute: Wayne, PA, 2015, CLSI Document M100S.
- (48) Roshan, N.; Riley, T. V.; Hammer, K. A. Antimicrobial Activity of Natural Products against Clostridium Difficile in Vitro. J. Appl. Microbiol. 2017, 123, 92–103.