

## **RESEARCH ARTICLE**

# How hornbills handle heat: sex-specific thermoregulation in the southern yellow-billed hornbill

Barry van Jaarsveld<sup>1,2,\*</sup>, Nigel C. Bennett<sup>3</sup>, Zenon J. Czenze<sup>1,2,‡</sup>, Ryno Kemp<sup>1,2</sup>, Tanja M. F. N. van de Ven<sup>4,5</sup>, Susan J. Cunningham<sup>4</sup> and Andrew E. McKechnie<sup>1,2</sup>

## **ABSTRACT**

At a global scale, thermal physiology is correlated with climatic variables such as temperature and aridity. There is also evidence that thermoregulatory traits vary with fine-scale microclimate, but this has received less attention in endotherms. Here, we test the hypothesis that avian thermoregulation varies with microclimate and behavioural constraints in a non-passerine bird. Male and female southern yellowbilled hornbills (Tockus leucomelas) experience markedly different microclimates while breeding, with the female sealing herself into a tree cavity and moulting all her flight feathers during the breeding attempt, becoming entirely reliant on the male for provisioning. We examined interactions between resting metabolic rate (RMR), evaporative water loss (EWL) and core body temperature ( $T_b$ ) at air temperatures ( $T_a$ ) between 30°C and 52°C in male and female hornbills, and quantified evaporative cooling efficiencies and heat tolerance limits. At thermoneutral  $T_{\rm a}$ , neither RMR, EWL nor  $T_{\rm b}$  differed between sexes. At  $T_a$  >40°C, however, RMR and EWL of females were significantly lower than those of males, by  ${\sim}13\%$  and  ${\sim}17\%,$  respectively, despite similar relationships between  $T_b$  and  $T_a$ , maximum ratio of evaporative heat loss to metabolic heat production and heat tolerance limits (~50°C). These sex-specific differences in hornbill thermoregulation support the hypothesis that avian thermal physiology can vary within species in response to fine-scale microclimatic factors. In addition,  $Q_{10}$  for RMR varied substantially, with  $Q_{10} \leq 2$  in some individuals, supporting recent arguments that active metabolic suppression may be an underappreciated aspect of endotherm thermoregulation in the heat.

KEY WORDS: Thermoregulation, Microclimate, Sex-specific, Avian, Heat

## INTRODUCTION

An enduring question in evolutionary physiology concerns the extent to which adaptation to local climate contributes to overall

<sup>1</sup>South African Research Chair in Conservation Physiology, South African National Biodiversity Institute, Pretoria 0001, South Africa. <sup>2</sup>DSI-NRF Centre of Excellence at the FitzPatrick Institute, Department of Zoology and Entomology, University of Pretoria, Pretoria, South Africa. <sup>3</sup>Mammal Research Institute, Department of Zoology and Entomology, University of Pretoria, Pretoria 0001, South Africa. <sup>4</sup>FitzPatrick Institute of African Ornithology, DSI-NRF Centre of Excellence, Department of Biological Sciences, University of Cape Town, Private Bag X3, Rondebosch 7701, South Africa. <sup>5</sup>Brain Function Research Group, School of Physiology, Faculty of Health Sciences, University of the Witwatersrand, Parktown 2193, South Africa.

<sup>‡</sup>Present address: Centre for Behavioural and Physiological Ecology, Zoology, University of New England, Armidale, NSW, Australia.

\*Author for correspondence (u04464011@tuks.co.za)

DB.v.J., 0000-0001-5154-5922; N.C.B., 0000-0001-9748-2947; Z.J.C., 0000-0002-1113-7593; R.K., 0000-0002-5339-8783; T.M.F.N.v.d.V., 0000-0003-2486-5378; S.J.C., 0000-0001-9934-2585; A.E.M., 0000-0002-1524-1021

physiological diversity (Gaston et al., 2009; Bozinovic et al., 2011; Muñoz and Bodensteiner, 2019). Traits related to thermal physiology can have important fitness consequences and be under strong selective pressure from the physical environments organisms occupy (Angilletta et al., 2010; Boyles et al., 2012). Thermal adaptation is evident in taxa ranging from plants to endotherms, and understanding how climate drives the evolution of thermal traits is vital for elucidating the processes determining the geographical distribution and range limits of species (Bozinovic et al., 2011; Keller and Seehausen, 2012). How these thermal traits correlate with climatic conditions has been studied at multiple levels of biological organisation and across varying spatiotemporal scales (Addo-Bediako et al., 2000; Sunday et al., 2011; Araújo et al., 2013; Bozinovic et al., 2014; White et al., 2007; Naya et al., 2013).

At the broadest spatial scale, thermoregulatory traits of both endoand ectothermic animals vary with latitudinal gradients in climate. For instance, lower thermal limits in ectotherms decrease with latitude, whereas upper thermal limits do not, resulting in an increased thermal tolerance breadth in species occupying higher latitudes (Addo-Bediako et al., 2000; Sunday et al., 2011; Araújo et al., 2013). Global-scale analyses investigating correlations between climate and endotherm thermoregulation reveal that basal metabolic rate in birds is negatively correlated with air temperature (White et al., 2007). In addition, both maximum and summit metabolism, the maximum rate of resting metabolic thermogenesis during cold exposure, are also negatively correlated with prevailing air temperature (McKechnie and Swanson, 2010).

Thermal physiology can also be correlated with climatic conditions at finer spatial scales. More evidence for physiological adaptation reflecting climate is apparent from studies that have examined physiological variation along aridity gradients. Among insects, reptiles, birds and mammals, taxa and populations inhabiting arid areas display lower rates of evaporative water loss compared with their mesic counterparts (Addo-Bediako et al., 2001; Tieleman et al., 2003; Williams et al., 2004; Van Sant et al., 2012; Guillon et al., 2014; Cox and Cox, 2015; Belasen et al., 2017; Dupoué et al., 2017; Klüg-Baerwald and Brigham, 2017). Similarly, basal metabolic rates are negatively correlated with habitat aridity in endotherms (Lovegrove, 2000; Tieleman et al., 2003; Williams et al., 2004). At the intraspecific level, there is some evidence that avian resting metabolic and evaporative water loss rates and core body temperature  $(T_{\rm b})$  are lower in arid populations than mesic populations (Noakes et al., 2016; Trost, 1972; but see also Smit et al., 2013). The adaptive value of reduced metabolic and evaporative water loss rates in desert animals is thought to be related to increased water and energy savings in response to low and unpredictable availability of resources (Lovegrove, 2000, 2003; Tieleman et al., 2003).

Evidence that thermal physiology evolves in response to climate at multiple spatial scales raises the possibility of similar variation at an intra-population level in response to variation in the fine-scale

microclimatic conditions individuals occupy. Indeed, most of the evidence that individuals within the same populations can occupy different microclimate space stems, indirectly, from literature investigating sex-specific differences in habitat use or behaviour (Cryan and Wolf, 2003; Lailvaux et al., 2003; Beal et al., 2014; Liwanag et al., 2018; Vidal et al., 2008; Cameron et al., 2018). For example, sex-specific habitat use is prevalent in ectothermic western Indian anoles (Butler et al., 2007). Furthermore, sex-specific habitat use amongst endotherms occurs in four species of North American ducks, where females occupy shallower and more vegetated sites compared with males (Bergan and Smith, 1989). Similarly, the great bustard (Otis tarda) exhibits sex-specific behavioural thermoregulation, with males retreating to shaded microsites in response to high temperatures earlier than females (Alonso et al., 2016). Birds exhibiting uniparental care [i.e. one sex entirely responsible for incubation and hatchling care, with the other responsible for food provisioning (Cockburn, 2006)] may experience sex-specific microclimates during breeding seasons. Consequently, males and females may experience substantially different thermal regimes (van de Ven et al., 2020).

One avian taxon that represents a fascinating model for testing hypotheses concerning sex-specific differences in thermal physiology arising from sex-specific microsite selection is the hornbills (Bucerotiformes: Bucerotidae). During the breeding season, female hornbills seal themselves into nest cavities as a protective mechanism against predators (Moreau and Moreau, 1941; Witmer, 1992; Kemp, 1995). Here, females are confined to the nest, undergo a complete wing and tail moult, and males are solely responsible for provisioning females and chicks for much of the breeding attempt (Klaassen et al., 2003; Kemp, 1969). For hornbills occupying hot environments with summer rainfall, females are sealed into cavities during the hottest time of the year and experience a combination of high air temperature, limited convective cooling and humidity above prevailing levels, which together pose substantial thermoregulatory challenges (Fig. 1: Gerson et al., 2019; van Dyk et al., 2019). In addition, the complete, simultaneous wing and tail feather moult undergone by females while confined in the nest cavity means that the heat load experienced by the females includes the metabolic cost of re-growing feathers (Buttemer et al., 2019). In addition, at high temperatures females experience restricted water and food supply because of decreased provisioning rates by male hornbills, reflecting trade-offs between behavioural thermoregulation and foraging (van de Ven et al., 2019, 2020); thus, females experience increased water demands for evaporative cooling combined with reductions in water and food supply.

Here, we investigated thermoregulation in southern yellow-billed hornbills (Tockus leucomelas Lichtenstein 1842) in the Kalahari Desert of southern Africa, where air temperatures can approach 40°C during the mid-summer breeding period (Fig. 1). Using a flowthrough respirometry system, we measured resting metabolic rate (RMR), evaporative water loss (EWL) and core body temperature  $(T_b)$ over air temperatures  $(T_a)$ , ranging from mild to hot. We hypothesised that highly asymmetric parental care in the hornbills, and the related sex-specific differences in microclimates, has led to the evolution of sex differences in thermal physiology under high heat loads. We predicted that compared with males under resting conditions, females: (1) have more efficient evaporative cooling characterised by lower RMR above the upper critical limit of thermoneutrality, ultimately resulting in higher maximum evaporative heat loss (EHL)/metabolic heat production (MHP) ratios [i.e. overall capacity for evaporative cooling and a significant predictor of heat tolerance limits (Czenze et al., 2020)], and (2) have higher heat tolerance limits. In addition, because there is evidence for temperature-independence of avian

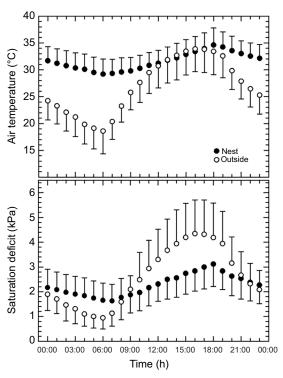


Fig. 1. Mean air temperature and saturation deficit in southern yellow-billed hornbill (*Tockus leucomelas*) tree-cavity nests compared with outside conditions. Means±s.d. for 1 h bins are shown for air temperature and saturation deficit within nests (black circles and error bars) and outside conditions (clear circles and error bars). Error bars for saturation deficit are flipped after 09:00 h. Nest temperatures were recorded using Thermochron iButtons (DS1923, Maxim, Sunnyvale, CA, USA, resolution=0.0625°C) at 5 min intervals. Corresponding ambient temperature and saturation deficit was obtained from a nearby weather station (Vantage Pro2, Davis Instruments, Hayward, USA).

RMR at high  $T_{\rm a}$  in some species (Weathers, 1981) and there have been arguments that metabolic suppression is involved in endotherm thermoregulation in the heat (Song et al., 1997; Lovegrove et al., 2014; Reher et al., 2018), we also compared  $Q_{10}$ , a quantitative descriptor of the Arrhenius effect of temperature on metabolic rate, between male and female hornbills experiencing very high  $T_{\rm a}$  values. An additional aim of the study was to quantify upper limits to heat tolerance and evaporative cooling capacity in a member of the Bucerotiformes, as this order is absent from the data currently available for arid-zone birds (McKechnie et al., 2021a).

## MATERIALS AND METHODS Study site and species

This study was conducted during the hornbills' breeding season, which usually spans from October to November (Kemp, 2005). All sampling took place between 1 October and 2 November 2018 in the southern Kalahari Desert at Radnor farm (26°6′23″S, 22°52′54″E) near Vorstershoop, North West Province, South Africa. The vegetation of this region is characterised as Molopo Bushveld, consisting of a mix of open woodland and closed shrubland dominated by *Vachellia erioloba* and *Boscia albitrunca* trees and *Lycium cinereum*, *Lycium hirsutum* and *Rhigozum trichotomum* shrubs (Rutherford et al., 2006). During the summer months, maximum air temperatures are typically between 33 and 37°C and the site receives ~221 mm precipitation (Fick and Hijmans, 2017).

Ten female and nine male hornbills were captured using spring traps (60×60 cm) baited with superworms (*Zophobas morio*). Prior

to capture, individuals were monitored to ensure that they were not involved in a current breeding attempt; i.e. solitary males, females in a tree cavity, pairs displaying allopreening or allofeeding; and that females were not in moult (Kemp, 2005; Myers, 2000; Mills et al., 2005). Mean±s.d. body mass ( $M_b$ ) for males and females was 242.3±10.7 g (n=9) and 198.9±21.9 g (n=10), respectively. Following capture, the birds were transported to a field laboratory in cotton bags where they were individually housed in cages ( $60\times40\times40$  cm) and provided with an *ad libitum* supply of darkling beetle larvae *Zophobas morio* and water. Birds were sexed based on culmen length, tarsus length and  $M_b$  measurements (Kemp, 2005). Individuals were kept for no longer than 24 h prior to experimentation and were released on the same day.

#### Microclimate data

To test our assumption that females experience a more thermally challenging environment than males do when confined to their tree cavity nests, we analysed data collected from a semi-habituated population of hornbills at Kuruman River Reserve (26°58'S, 21°49'E) in the Northern Cape of South Africa. Occupied tree cavity nest temperatures  $(T_a)$  and humidity were recorded using Hygrochron iButtons (DS1923, Maxim, Sunnyvale, CA, USA, resolution=0.0625°C) at 5 min intervals. To prevent data anomalies arising from females coming into contact with the loggers, iButtons were attached to the upper internal surfaces of cavities. The temperature sensors of the iButtons were calibrated in a circulating water bath against a NiCr-NiAl thermocouple thermometer (Thermocouple HH21A, Omega Engineering, Stanford, USA). The NiCr-NiAl thermocouple thermometer (resolution=0.1°C) was calibrated against a mercury-in-glass thermometer with accuracy traceable to the National Institute of Standard Technology, MD, USA. Recordings were taken from three nests during the summer months of 2015, for a total of 45 nest-days. Corresponding  $T_a$  and humidity were obtained from a nearby weather station (Vantage Pro2, Davis Instruments, Hayward, USA). We acknowledge that  $T_a$ does not equate to operative temperature for males but provides an indication of the minimum operative temperature in the environment available to the birds if they seek deep shade during the hottest part of the day.

## **Gas exchange measurements**

A flow-through respirometry system was used to measure rates of carbon dioxide production (ml min<sup>-1</sup>) and rates of evaporative water loss (EWL; mg min<sup>-1</sup>). A 60 litre airtight container functioned as the metabolic chamber. The chamber was fitted with an air inlet positioned near the top of the chamber and the outlet near the bottom to maximise mixing. A platform constructed from plastic mesh was positioned ~10 cm above a 1 cm layer of mineral oil, which prevented evaporation from excreta. The chamber was placed within a custombuilt temperature-controlled chest (105×81×81 cm), constructed from polystyrene sheets positioned between the interior chipboard and exterior aluminium sheeting. The  $T_a$  within the chest was regulated by a digital controller (TC-720 Temperature Controller, TE Technology, Traverse City, MI, USA) that controlled a thermoelectric air cooler (TC 720 OEM, TE Technology). Atmospheric air, supplied by an oil-free was pushed through a membrane (Champion®CMD3 air dryer and filter, Champion Pneumatic, Quincy, IL, USA) and scrubbed of water vapour. Downstream of the membrane dryer, the airstream was split into a baseline and experimental channel. The flow rate in the baseline channel was regulated at ~1 l min<sup>-1</sup> by a needle valve (Swagelok, Solon, OH, USA). In the experimental channel, the flow rate was regulated by a

mass flow controller (0-60 SLPM, Alicat Scientific Inc., Tucson, AZ, USA), calibrated against a flow rate calibrator (Gilibrator 2, Sensidyne, St Petersburg, FL, USA). Flow rates were selected to maintain absolute humidity levels within the chamber as low as possible (<1 kPa) while still maintaining an accurately measurable difference in CO<sub>2</sub> and H<sub>2</sub>O concentration between the incurrent and excurrent air. Flow rates ranging from 2.3 to  $57.5 \,\mathrm{l\,min^{-1}}$   $(25\pm17 \,\mathrm{l\,min^{-1}})$  were used, depending on  $T_a$ , exposure time and the activity of birds. Excurrent air from the chamber and baseline channel were sequentially subsampled using a respirometry multiplexer (MUX3-1101-18 mol 1<sup>-1</sup>, Sable Systems, Las Vegas, NV, USA) in manual mode. Sub-sampled air was pulled through a CO<sub>2</sub>/H<sub>2</sub>O analyser (LI-840A, LI-COR, Lincoln, NE, USA) regularly zeroed using nitrogen, and spanned for CO<sub>2</sub> using an analytically certified gas with a known CO<sub>2</sub> concentration of 2000 ppm (AFROX, Johannesburg, South Africa) and for H<sub>2</sub>O using a dewpoint generator (DG4, Sable Systems, Las Vegas, NV, USA). All tubing in the system was Bev-A-Line IV tubing (Thermoplastic Processes Inc., Warren, NJ, USA). Voltage outputs from the analysers and thermistor probes were digitised using an analogue-digital converter (UI3, Sable Systems, Las Vegas, NV, USA) and recorded with a sampling interval of 5 s using Expedata software (Sable Systems, Las Vegas, NV, USA).

## Air and body temperature measurements

A 3-mm diameter thermistor probe (Sable Systems, Las Vegas, NV, USA) was inserted through a hole sealed with a rubber grommet in the side of the metabolic chamber to measure  $T_{\rm a}$  during gas exchange measurements. Core body temperature was continuously measured using temperature-sensitive passive integrated transponder (PIT) tags (BioTherm, Biomark, Biose, ID, USA) and a portable transceiver system linked to an antenna (HPR+, Biomark) placed adjacent to the respirometry chamber. The PIT tags were injected intraperitoneally, following Whitfield et al. (2015). Before implantation, PIT tags were calibrated against a digital thermocouple reader (RDXL12SD, Omega, Stamford, CT, USA) in a circulating water bath (F34, Julabo, Seelbach, Baden-Württemberg, Germany), over temperatures of  $30–50^{\circ}$ C. Temperatures measured by PIT tags deviated by  $0.06\pm0.05^{\circ}$ C (n=20) from actual values.

## **Experimental protocol**

Each individual was weighed on an electronic balance (EJ-160, A&D, Tokyo, Japan) and held in the metabolic chamber, without food and water, for at least 1.75 h before commencing measurements. Birds were exposed to a stepped profile of increasing  $T_{\rm a}$  between 30 and 52°C. At  $T_{\rm a}$ =30–40°C,  $T_{\rm a}$  was increased in 5°C increments, whereas at  $T_{\rm a}$ >40°C, it was increased in 2°C increments. All individuals spent >10 min, and the majority 15–20 min, at each  $T_{\rm a}$  value before data were recorded. Runs lasted for approximately 4 h, including the habituation time. Measurements commenced with a baseline air subsample until  $H_{\rm 2}O$  and  $CO_{\rm 2}$  readings were stable (~5 min). Subsequently, the excurrent chamber air was sub-sampled when  $T_{\rm a}$  had stabilised at the target value, followed by another 5 min baseline.

During measurements, behaviour and activity of individuals were continuously monitored using a CCTV video camera and an infrared light source. Trials were terminated, and individuals immediately removed from the chamber if their  $T_b$  exceeded 45°C and/or if they displayed a sudden decrease in EWL or RMR. Trials were also terminated if they displayed behavioural stress such as prolonged escape behaviour (i.e. agitated jumping, pecking and/or wing flapping), or if they exhibited a loss of coordination or balance. Birds that displayed a rapid decrease in EWL, RMR or increase in

 $T_b>45$ °C were considered to have reached their thermal endpoints. Once removed, a cotton pad soaked in 99% ethanol was rubbed on the bird's body to augment rates of heat loss, following Whitfield et al. (2015). Once  $T_b$  stabilised at normothermic levels (40–42°C), the bird was placed back into its cage at room temperature to rest with ad libitum water and food until their release at the site of capture. In almost all cases, the individuals lost less than 5%  $M_{\rm b}$ through faeces and water during measurements and no individuals displayed any adverse effects after experimentation. This experimental protocol has been previously used and, in one instance with opportunistic monitoring for several weeks postrelease, no adverse effects were observed (Kemp and McKechnie, 2019). All experimental protocols were approved by the University of Pretoria Animal Ethics committee (protocol EC023-18) and the South African National Biodiversity Institute's Research Ethics and Scientific Committee (protocol P18/12).

## **Data analysis**

Using equations 9.5 and 9.6 from Lighton (2008), whole animal rates of  $CO_2$  production ( $\dot{V}_{CO_2}$ ) and rates of EWL were calculated from the lowest 5-min steady-state traces, assuming 0.803 mg H<sub>2</sub>O ml<sup>-1</sup> vapour. Evaporative heat loss (W) was calculated assuming a latent heat of vaporisation of water of  $2.406 \,\mathrm{J}\,\mathrm{mg}\,\mathrm{H}_2\mathrm{O}^{-1}$  (Tracy et al., 2010). Given the high flow rates, oxygen consumption was not measured, thus we could not calculate the respiratory exchange ratio (RER). Birds were kept without food or water for a minimum of 1.75 h before measurements, with the allometrically predicted gut passage time for a 240 g bird, the largest individual in our study, being 95 min (Karasov, 1990). We hence assumed RER=0.71, representative of lipid metabolism in post-absorptive birds, and calculated RMR (W) from  $\dot{V}_{\rm CO_2}$  using 27.8 J ml<sup>-1</sup> CO<sub>2</sub> (Walsberg and Wolf, 1995). All statistical analyses were performed in R 3.5.2 (https://www.r-project.org/). Piecewise linear regression models were fitted separately to data from each sex to identify inflection points in EWL, RMR, T<sub>b</sub> and EHL/MHP as a function of  $T_a$ , using the functions described by Muggeo et al. (2014); these functions account for pseudoreplication when identifying inflection points. Furthermore, a piecewise linear regression model was fitted to EHL/MHP as a function of  $T_a-T_b$ . We also calculated  $Q_{10}$  for the RMR of each individual using minimum  $T_b$  and RMR values at  $T_a$ =30–35°C and maximum values at  $T_a$ =48–50°C in order to evaluate the possibility of metabolic suppression at high  $T_a$ .

To test the prediction that male and female hornbills display differing thermoregulatory responses to increases in  $T_a$ , the male and female RMR, EWL, EHL/MHP and T<sub>b</sub> data were pooled, with separate linear mixed-effects models fitted to data above and below the highest of the two sex-specific inflection points for each response variable using *lme4* (Bates et al., 2015). Model selection was conducted using the dredge function in the MuMIn package (https://CRAN.R-project.org/package=MuMIn) The global model contained  $T_a$  (or  $T_a - T_b$ ), sex,  $M_b$  and the  $T_a \times \text{sex}$  interaction. Model goodness of fit was assessed using Akaike information criterion values corrected for small sample size (AICc). Models with  $\Delta$ AICc <2 were considered equally parsimonious, and a final model was constructed by averaging the coefficients of all models below this threshold (Burnham and Anderson, 2002). A single top model explained the variation in EWL, EHL/MHP and  $T_{\rm b}$ , whereas for RMR two competing models provided similar support for data below and above the inflection point (Table S2). Since we were interested in the effect of sex on the response variable, we report conditional averages rather than the full averages, following

Symonds and Moussalli (2011) and Grueber et al. (2011). Multiple measurements from the same individuals required us to account for pseudo-replication by including bird identity as a random term. Body mass did not emerge as a significant predictor for any of the response variables (Table S2). We repeated the above procedure to test whether male and female hornbills displayed differing mass-specific rates of EWL (msEWL) and RMR (msRMR) above and below physiological thresholds (Fig. S1, Table S2). The  $Q_{10}$  of RMR in male and female hornbills was compared using a two-tailed t-test, as were differences between prevailing conditions and those within tree-cavities. Values are presented as means $\pm$ s.d.

## **RESULTS**

#### **Microclimates**

Daily maximum  $T_a$  recorded inside the nests ( $\bar{x}$ =34.36±0.48°C, n=45 days) was similar to the corresponding value outside the nest ( $\bar{x}$ =33.36±0.75°C, n=40 days; t=1.14, d.f.=83, P=0.26; Fig. 1). Minimum saturation deficit was significantly lower inside the nests ( $\bar{x}$ =3.26±0.13 kPa, n=45 days) compared with outside ( $\bar{x}$ =4.21±0.24 kPa, n=40 days; t=3.36, d.f.=83, t<0.001; Fig. 1).

## **Evaporative water loss**

The inflection point in the relationship between EWL and  $T_{\rm a}$  occurred at  $T_{\rm a}$ =40.5°C for males and  $T_{\rm a}$ =39.7°C for females, above which EWL increased with increasing  $T_{\rm a}$ . Below the inflection point, EWL increased significantly with  $T_{\rm a}$  (t=5.56, P<0.001; Fig. 2) but rates of EWL were minimal (0.5±0.2 g h<sup>-1</sup>, t=19) and sex did not emerge as a significant predictor (Fig. 2, Table S1). At t=10 values above the inflection, EWL increased linearly and significantly with t=11 (t=20.22, t=20.001; Fig. 2), and differed between sexes (t=2.36, t=20.3; Fig. 2). Rates of EWL increased to a maximum of 6.2±1.6 g h<sup>-1</sup> (t=9) at t=250°C among females (with one female reaching 7.6 g h<sup>-1</sup> at t=250°C among males. Rates of EWL at t=250°C were 17% lower in females compared with males; equivalent to 3.3±0.5% t=10 for females.

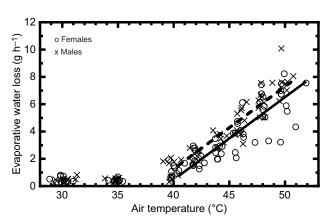


Fig. 2. Relationship between evaporative water loss (EWL) and air temperature ( $T_a$ ) in southern yellow-billed hornbills (*Tockus leucomelas*). Data were obtained from 10 females (circles) and nine males (crosses) using open flow-through respirometry. Lines indicate linear mixed-effects regression models fitted to  $T_a$ >40.5°C, males (dashed): EWL=0.60 $T_a$  –23.72 and females (solid): EWL=0.60 $T_a$ –22.93.

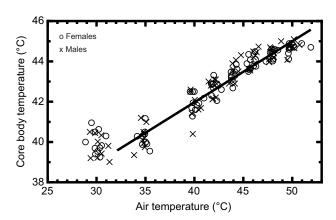


Fig. 3. Relationship between core body temperature ( $T_b$ ) and air temperature ( $T_a$ ) in southern yellow-billed hornbills (Tockus leucomelas). Data were obtained from 10 females (circles) and nine males (crosses) using open flow-through respirometry. Lines indicate linear mixed-effects regression models fitted to  $T_a > 32.1^{\circ}$ C,  $T_b = 0.30 T_a + 29.86$ .

## Core body temperature and thermal endpoints

An inflection in  $T_{\rm b}$  occurred at  $T_{\rm a}$ =32.0°C for females and  $T_{\rm a}$ =32.1°C for males. At  $T_{\rm a}$  above the inflection point,  $T_{\rm b}$  increased linearly and significantly with  $T_{\rm a}$  (t=41.11, P<0.001; Fig. 3), and sex did not emerge as a significant predictor (Fig. 3, Table S1). Below the inflection point,  $T_{\rm a}$  had no effect on  $T_{\rm b}$  (t=1.28, P=0.22; Fig. 3) and sex did not emerge as a significant predictor (Table S1, Fig. 3). At  $T_{\rm a}$ =~30°C, mean normothermic  $T_{\rm b}$  values were 39.9±0.6°C (n=19). Females reached thermal endpoints at  $T_{\rm a}$ =50.6±1.4°C with corresponding  $T_{\rm b}$ =45.0±0.3°C (n=10). Males reached thermal endpoints at  $T_{\rm a}$ =50.4±1.2°C (n=9), at which  $T_{\rm b}$ =44.8±0.3°C (n=9). Nine females reached  $T_{\rm a}$ =50°C and one female reached  $T_{\rm a}$ =51°C; whereas seven males reached 50°C and two males reached 48°C.

# **Resting metabolic rate**

Inflections in RMR occurred at  $T_a$ =39.2°C for males and at  $T_a$ =38.8°C for females, which we interpret as the upper critical limits of thermoneutrality. The onset of panting corresponded closely with these inflection points, occurring at  $T_a$ =37.3±2.5°C (n=9) in males and at  $T_a$ =38.6±2.6°C (n=10) in females. Two

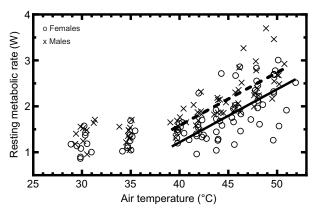


Fig. 4. Relationship between resting metabolic rate (RMR) and air temperature ( $T_a$ ) in southern yellow-billed hornbills (*Tockus leucomelas*). Data were obtained from 10 females (circles) and nine males (crosses) using open flow-through respirometry. Lines indicate linear mixed-effects regression models fitted to  $T_a$ >39.2°C, males (dashed): RMR=0.12 $T_a$ -3.04 and females (solid): RMR=0.12 $T_a$ -3.40.

competing models similarly supported data below and above the inflection point (Table S1). Below the inflection point, a minimum RMR of  $1.3\pm0.2~\rm W~(n=18)$  occurred at  $T_a=\sim30^{\circ}\rm C$  and did not differ between sexes (averaged model: z=1.90, P=0.05; Fig. 4). Above the inflection point, RMR increased linearly and significantly with  $T_a$  (averaged model: z=14.28, P<0.001; Fig. 4) and was lower in females (averaged model: z=2.43, P=0.01; Fig. 4). At  $T_a=\sim50^{\circ}\rm C$ , RMR of males and females was  $2.7\pm0.4~\rm W~(n=7)$  and  $2.3\pm0.6~\rm W~(n=9)$ , respectively, equivalent to a 13% difference. Individuals'  $Q_{10}$  values for RMR varied widely, from 1.68 in a female to 8.05 in a male; however, the mean values for males  $(5.03\pm2.36,~n=9)$  and females  $(4.09\pm1.37,~n=10)$  did not differ  $(t=1.05,~\rm d.f.=12.53,~\rm P=0.31)$ .

## **Evaporative heat loss/metabolic heat production**

An inflection in EHL/MHP occurred at  $T_a$ =37.1°C for males and  $T_a$ =37.3°C for females. When EHL/MHP was plotted against  $T_a$ - $T_b$ , an inflection point was identified at  $T_a$ - $T_b$ =-4.0°C for males and  $T_a$ - $T_b$ =-4.1°C for females. Above the inflection point EHL/MHP increased linearly and significantly with  $T_a$ - $T_b$  (t=23.27, P<0.001; Fig. 5) and sex did not emerge as a significant predictor (Fig. 4, Table S1). Below the inflection point, the null model provided the best fit (Fig. 5, Table S1). The y-intercept of the linear model for  $T_a$  values above the inflection was 0.95, indicating that 95% of MHP is dissipated by evaporative cooling when  $T_b$ = $T_a$ . At  $T_a$  below the inflection point, the minimum ratio of EHL/MHP was 0.2±0.1 (n=18) at  $T_a$ =~30°C and increased to a mean maximum value of 1.8±0.2 (n=18) at  $T_a$ =~50°C (with one female reaching a value of 2.0 at  $T_a$ =~50°C).

## **DISCUSSION**

No sex differences were evident in  $T_b$ , RMR or EWL at thermoneutral  $T_a$ . At higher temperatures, however, female hornbills used significantly less water ( $\sim$ 17%) and energy ( $\sim$ 13%) than males to defend similar  $T_b$  values and to achieve similar maximum EHL/MHP values. These differences broadly support our first prediction, that evaporative cooling is more efficient in female hornbills compared with males; although the maximum EHL/MHP values are similar, absolute water and energy requirements are lower in females compared with males. As rates of EWL and RMR scale allometrically with  $M_b$  (White and Seymour, 2003; McKechnie and Wolf, 2010), a possible

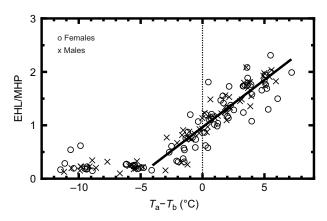


Fig. 5. Relationship between evaporative heat loss/metabolic heat production (EHL/MHP) and the air temperature–body temperature gradient ( $T_a$ – $T_b$ ) in southern yellow-billed hornbills (*Tockus leucomelas*). Data were obtained from 10 females (circles) and nine males (crosses) using open flow-through respirometry. Line indicates linear mixed-effects regression model fitted to  $T_a$ – $T_b$ > –4.04°C, EHL/MHP=0.18( $T_a$ – $T_b$ )+0.96.

explanation is that the sex-specific rates of EWL and RMR in hornbills are driven by sexual dimorphism in  $M_{\rm b}$ . However,  $M_{\rm b}$  did not emerge as a significant predictor for any of the variables we tested. In addition, given a negative relationship of mass-specific EWL and RMR to  $M_{\rm b}$  (McKechnie and Wolf, 2010; White and Seymour, 2003), one would expect female hornbills to display greater mass-specific EWL and RMR compared with males. Yet, there was no significant effect of sex on mass-specific rates of EWL and RMR (Fig. S1, Table S2), further suggesting that sex-specific differences in thermoregulation in the hornbills are not attributable to sexual dimorphism in  $M_{\rm b}$ . Our second prediction was not supported; relationships between  $T_{\rm b}$  and  $T_{\rm a}$  were indistinguishable between sexes, providing no support for the notion that females have greater heat tolerance limits, and maximum EHL/MHP did not differ between sexes.

The heat tolerance limit of ~50°C in male and female hornbills is lower than values of 54-62°C reported for similarly sized nonpasserine species, galah (Eolophus roseicapilla; McWhorter et al., 2018), Burchell's sandgrouse (Pterocles burchelli; McKechnie et al., 2016a) and crested pigeon (Ocyphaps lophotes; McKechnie et al., 2016b). Although male and female hornbills displayed similar heat tolerance limits under laboratory conditions, two aspects of the species' ecology not evaluated in this study may translate into sex-specific differences in heat tolerance limits and maximum evaporative cooling demands under natural conditions. First, although incarcerated female hornbills experience more thermally challenging conditions (i.e. the combined effect of  $T_a$  and humidity) compared with males over time scales of hours to days (Fig. 1), male hornbills potentially experience transient operative temperatures approaching or even exceeding 50°C when foraging on the ground in full sun. Furthermore,  $T_a$  is only an approximation of minimum operative temperature experienced by males. Second, females undergo complete flight feather moult while in the nest, which might enhance non-evaporative heat dissipation when  $T_a < T_b$ , although any such effect would presumably be offset by the increase in MHP associated with moult (e.g. Buttemer et al., 2019). To elucidate whether sex-specific differences in heat tolerance limits are also manifested in this species under natural conditions, future studies should (1) examine free-ranging  $T_{\rm b}$  limits in relation to operative temperatures and (2) evaluate the possibility that the effects of humidity on evaporative cooling at high environmental temperatures differ between males and females, particularly when the latter are sealed in breeding cavities.

Maximum  $T_b$  during acute heat exposure also did not differ between sexes, with maxima of  $\sim 45^{\circ}$ C for both males and females broadly consistent with those reported for most species examined to date (Dawson, 1954; Gerson et al., 2019). Tolerance of unusually high  $T_{\rm b}$  in a central American passerine has been suggested to be functionally linked to impeded EHL in humid environments (Weathers, 1981), the reasoning on which our prediction of more pronounced facultative hyperthermia in female hornbills was based. The observation of Weathers (1981), as well as those of a handful of other authors reporting avian tolerance of T<sub>b</sub>>45°C (Arad and Marder, 1982; Dmi'el and Tel-Tzur, 1985; Freeman et al., 2020), reiterates the need to determine the upper limits to hyperthermia tolerance in many more species, and to evaluate the relative roles of deleterious temperature effects on cell membrane fluidity, transport pathways across membranes and other aspects of macromolecule function (Adolph, 1947; Roti Roti, 2008) and oxygen supply limitation (Teague et al., 2017; Pörtner et al., 2017) in determining

Expressed as percentages of  $M_b$ , maximum rates of EWL were similar in male and female hornbills, suggesting similar risks of

lethal dehydration on extremely hot days. However, the singleparent provisioning employed by this species means that, under natural conditions, breeding females may often be under greater dehydration risk arising from mismatches between supply and demand as a consequence of male foraging decisions. Behavioural observations on breeding hornbills reveal that, during very hot weather, males are unable to meet the energetic requirements of females, resulting from a decreased foraging efficiency under high heat loads, thereby exposing female hornbills to a higher risk of lethal dehydration (van de Ven et al., 2020). Indeed, such mismatches are thought to result in nest abandonment and potentially death if the female leaves her nest in a flightless state (van de Ven, 2017). In addition to direct hyperthermia-related mortality, dehydration is responsible for lethal effects of high  $T_a$ values and is expected to be a critical factor in determining the potential for avian species to persist in arid regions under anticipated global heating (McKechnie and Wolf, 2010; Albright et al., 2017; Conradie et al., 2019).

The sex differences in EWL and RMR at high  $T_a$  we report here for hornbills could conceivably arise in several ways. One is facultative hyperthermia, an often-cited mechanism for water conservation among desert birds (Gerson et al., 2019). However, relationships between  $T_b$  and  $T_a$  were indistinguishable for male and female hornbills, suggesting that facultative hyperthermia is not responsible for sex-specific differences in this species. Similar conclusions were reached concerning intraspecific variation in white-browed sparrow-weavers (*Plocepasser mahali*), even though a desert population had significantly higher heat tolerance limits in summer compared with two populations from cooler habitats (Noakes et al., 2016). Second, evaporative heat dissipation by females could involve lower metabolic heat production, for instance via enhanced cutaneous water loss (Menon et al., 1989; Tieleman and Williams, 2002; Webster and Bernstein, 1987; Williams, 1996), more efficient counter-current heat exchange across respiratory surfaces (Schmidt-Nielson et al., 1970) or more efficient panting via differences in the elastic properties and resonant frequency of the respiratory tract (Crawford and Kampe, 1971; Richards, 1970). A third possibility concerns sex differences in non-evaporative heat dissipation across the surface of the beak, which is smaller in females compared with males (Kemp, 2005). However, van de Ven et al. (2016) found no sex differences in heat dissipation via the beak in this species among individuals exposed to  $T_a$  between 15°C and 45°C, and it is unlikely this avenue of heat dissipation explains the sex differences we observed at  $T_a > 39$ °C.

The notion of active suppression of metabolic rates (i.e. hypometabolism) under conditions of high heat load has received increased attention given recent evidence of 'hyperthermic daily torpor' in some small tropical mammals (Lovegrove et al., 2014; Reher et al., 2018). In Fig. 3, female RMR at  $T_a$  between 45°C and 50°C appears considerably more variable than that of males, with a number of female values remaining similar to the thermoneutral values even at  $T_a$  approaching 50°C. The  $Q_{10}$  values  $\leq 2$  for some individuals in our study support the notion that metabolic suppression may sometimes occur, although we could detect no significant sex difference. Nevertheless, the possibility that females could use metabolic suppression under humid conditions deserves further investigation, as does the apparent lack of temperature effects on metabolism under some conditions in both birds and mammals (Weathers, 1981; Tieleman and Williams, 1999; O'Connor et al., 2017; Lovegrove et al., 2014; Reher et al., 2018).

There is increasing evidence for fine-scale intraspecific variation in the thermal physiology of endotherms (Tieleman

et al., 2009; Glanville et al., 2012; McClennand et al., 2016; Noakes and McKechnie, 2019), which may arise from phenotypic flexibility, developmental plasticity (sensu Piersma and Drent, 2003), epigenetic transmission or genotypic adaptation (Merilä and Hendry, 2014). Far less is known about the contributions of these sources of variation to intraspecific variation in the heat tolerance of endotherms compared with that of ectotherms (Ketola et al., 2012; Xue et al., 2019), although recent work suggests that differences in heat tolerance among white-browed sparrow-weaver populations arise from phenotypic flexibility rather than genotypic adaptation (Noakes and McKechnie, 2019). The present study involved summer-acclimatised hornbills studied at the same time of year as this species breeds, and a future study testing whether these sex differences persist in winter-acclimatised individuals would be informative in terms of whether this variation arises from phenotypic flexibility.

Similar T<sub>b</sub> values yet lower rates of EWL and RMR of female hornbills at high  $T_a$  compared with males likely confers adaptive benefits related to limited energy and water supply for females during hot weather, arising from the strong negative effects of hot weather on male foraging efficiency and hence nest provisioning rates (van de Ven et al., 2019, 2020), as opposed to avoidance of lethal hyperthermia. Resource availability and environmental heat loads are proposed to be important drivers in the evolution of endothermic thermoregulation (Brown et al., 2004; Angilletta et al., 2010; Speakman and Król, 2010; Boyles et al., 2011). Recent work on this species has revealed strong temperature-driven trade-offs between thermoregulation and foraging in male hornbills (van de Ven et al., 2019) and strong negative effects of high  $T_a$  during the breeding season on fitness components, including increased probability of nest abandonment, reduced likelihood of fledging and reduced fledgling mass (van de Ven et al., 2020). These sublethal fitness costs of chronic exposure to hot weather in hornbills and other arid-zone species (du Plessis et al., 2012; Cunningham et al., 2013; Kemp et al., 2020) are anticipated to result in substantial population declines and range contractions during the 21st century in response to global heating in the Kalahari (Conradie et al., 2019; Kemp et al., 2020).

In conclusion, differences in thermoregulation at  $T_{\rm a}$  exceeding normothermic  $T_{\rm b}$ , evident as lower water and energy requirements by female than male hornbills, supports the hypothesis that thermal physiology may evolve from sex-specific microclimates experienced during the breeding season. These findings, taken together with increasing evidence for substantial variation in evaporative cooling capacity and heat tolerance among related species co-occurring at a single site (Czenze et al., 2020) and in conspecific populations occurring along climatic gradients (Trost, 1972; Noakes et al., 2016) emphasise that much remains to be learned about adaptive variation in avian thermoregulation in the heat.

## Acknowledgements

We thank the Mathews family for allowing us to conduct this research at Radnor Farm, and Matthew Symonds, Alex McQueen, Sara Ryding and an anonymous reviewer whose constructive comments improved the quality of the manuscript.

## Competing interests

The authors declare no competing or financial interests.

## Author contributions

Conceptualization: B.v.J., N.C.B., A.E.M.; Methodology: B.v.J., N.C.B., S.J.C., A.E.M.; Formal analysis: B.v.J., R.K., T.M.F.N.V.; Resources: N.C.B., S.J.C., A.E.M.; Data curation: B.v.J., Z.J.C., R.K., T.M.F.N.V.; Writing - original draft: B.v.J., Z.J.C., S.J.C., A.E.M.; Writing - review & editing: B.v.J., N.C.B., Z.J.C., R.K., T.M.F.N.V., S.J.C., A.E.M.; Supervision: N.C.B., S.J.C., A.E.M.; Project administration: N.C.B., S.J.C., A.E.M.; Funding acquisition: N.C.B., S.J.C., A.E.M.

#### **Funding**

This project was funded jointly by the National Research Foundation SARChI chair for Mammal Behavioural Ecology and Physiology (grant number 64756) and the SARChI chair of Conservation Physiology (grant 119754) awarded to N.C.B. and A.E.M., respectively. Any opinions, findings and conclusions or recommendations expressed in this material are those of the authors and do not necessarily reflect the views of the National Research Foundation. T.M.F.N.V. was funded by the Centre of Excellence at the FitzPatrick Institute of African Ornithology and the Claude Leon Foundation.

#### Data availability

Data are available in the Dryad digital repository (McKechnie et al., 2021b): bcc2fqzbq.

#### Supplementary information

Supplementary information available online at https://jeb.biologists.org/lookup/doi/10.1242/jeb.232777.supplemental

#### References

- Addo-Bediako, A., Chown, S. L. and Gaston, K. J. (2000). Thermal tolerance, climatic variability and latitude. *Proc. Roy. Soc. B* 267, 739-745. doi:10.1098/rspb. 2000.1065
- Addo-Bediako, A., Chown, S. L. and Gaston, K. J. (2001). Revisiting water loss in insects: a large scale view. J. Insect. Physiol. 47, 1377-1388. doi:10.1016/S0022-1910(01)00128-7
- Adolph, E. F. (1947). Tolerance to heat and dehydration in several species of mammals. Am. J. Physiol. 151, 564-575. doi:10.1152/ajplegacy.1947.151.2.564
- Albright, T. P., Mutiibwa, D., Gerson, A. R., Smith, E. K., Talbot, W. A., O'Neill, J. J., McKechnie, A. E. and Wolf, B. O. (2017). Mapping evaporative water loss in desert passerines reveals an expanding threat of lethal dehydration. *Proc. Nat. Acad. Sci. USA* 114, 2283-2288. doi:10.1073/pnas.1613625114
- Alonso, J. C., Salgado, I. and Palacín, C. (2016). Thermal tolerance may cause sexual segregation in sexually dimorphic species living in hot environments. Behav. Ecol. 27, 717-724. doi:10.1093/beheco/arv211
- Angilletta, M. J., Cooper, B. S., Schuler, M. S. and Boyles, J. G. (2010). The evolution of thermal physiology in endotherms. *Biosci.* E2, 861-881. doi:10.2741/e148
- Arad, Z. and Marder, J. (1982). Strain differences in heat resistance to acute heat stress, between the bedouin desert fowl, the white leghorn and their crossbreeds. Comp. Biochem. Phys. A 72, 191-193. doi:10.1016/0300-9629(82)90031-7
- Araújo, M. B., Ferri-Yáñez, F., Bozinovic, F., Marquet, P. A., Valladares, F. and Chown, S. L. (2013). Heat freezes niche evolution. *Ecol. Lett.* 16, 1206-1219. doi:10.1111/ele.12155
- Bates, D., Maechler, M., Bolker, B. and Walker, S. (2015). Fitting linear mixedeffects models using Ime4. J. Stat. Softw. 67, 1-48. doi:10.18637/jss.v067.i01
- Beal, M. S., Lattanzio, M. S. and Miles, D. B. (2014). Differences in the thermal physiology of adult yarrow's spiny lizards (*Sceloporus jarrovii*) in relation to sex and body size. *Ecol. Evol.* 4, 4220-4229. doi:10.1002/ece3.1297
- Belasen, A., Brock, K., Li, B., Chremou, D., Valakos, E., Pafilis, P., Sinervo, B. and Foufopoulos, J. (2017). Fine with heat, problems with water: microclimate alters water loss in a thermally adapted insular lizard. *Oikos* 126, 447-457. doi:10. 1111/oik.03712
- Bergan, J. F. and Smith, L. M. (1989). Differential habitat use by dicing ducks wintering in south carolina. J. Wildlife Manag. 53, 117-1126. doi:10.2307/3809618
- Boyles, J. G., Seebacher, F., Smit, B. and McKechnie, A. E. (2011). Adaptive thermoregulation in endotherms may alter responses to climate change. *Integr. Comp. Biol.* 51, 676-690. doi:10.1093/icb/icr053
- Boyles, J. G., Verburgt, L., McKechnie, A. E. and Bennett, N. C. (2012). Heterothermy in two mole-rat species subjected to interacting thermoregulatory challenges. J. Exp. Zool. A 317, 73-82. doi:10.1002/jez.723
- Bozinovic, F., Calosi, P. and Spicer, J. I. (2011). Physiological correlates of geographic range in animals. *Annual Review of Ecol. Evol. Syst.* **42**, 155-179. doi:10.1146/annurev-ecolsys-102710-145055
- Bozinovic, F., Ferri-Yáñez, F., Naya, H., Araújo, M. B. and Naya, D. E. (2014).
  Thermal tolerances in rodents: species that evolved in cold climates exhibit a wider thermoneutral zone. Evol. Ecol. Res. 16, 143-152.
- Brown, J. H., Gillooly, J. F., Allen, A. P., Savage, V. M. and West, G. B. (2004). Toward a metabolic theory of ecology. *Ecology* 85, 1771-1789. doi:10.1890/03-9000
- Burnham, K. P. and Anderson, D. R. (2002). Model Selection and Multimodel Inference: A Practical Information-Theoretic Approach. London: Springer.
- Butler, M. A., Sawyer, S. A. and Losos, J. B. (2007). Sexual dimorphism and adaptive radiation in anolis lizards. *Nature* 447, 202-205. doi:10.1038/pature05774
- Buttemer, W. A., Bauer, S., Emmenegger, T., Dimitrov, D., Peev, S. and Hahn, S. (2019). Moult-related reduction of aerobic scope in passerine birds. *J. Comp. Physiol. B* **189**, 3-4. doi:10.1007/s00360-019-01213-z

- Cameron, S. F., Wheatley, R. and Wilson, R. S. (2018). Sex-specific thermal sensitivities of performance and activity in the asian house gecko, *Hemidactylus* frenatus. J. Comp. Physiol. B 188, 635-647. doi:10.1007/s00360-018-1149-2
- Cockburn, A. (2006). Prevalence of different modes of parental care in birds. *Proc. Roy. Soc. B* **273**, 1375-1383. doi:10.1098/rspb.2005.3458
- Conradie, S. R., Woodborne, S. M., Cunningham, S. J. and McKechnie, A. E. (2019). Chronic, sublethal effects of high temperatures will cause severe declines in southern African arid-zone birds during the 21st century. *Proc. Nat. Acad. Sci. USA* **116**, 14065-14070. doi:10.1073/pnas.1821312116
- Cox, C. L. and Cox, R. M. (2015). Evolutionary shifts in habitat aridity predict evaporative water loss across squamate reptiles. *Evolution* 69, 2507-2516. doi:10. 1111/evo.12742
- Crawford, E. C. and Kampe, G. (1971). Resonant panting in pigeons. *Comp. Biochem. Phys. A* **40**, 549-552, doi:10.1016/0300-9629(71)90048-X
- Cryan, P. M. and Wolf, B. O. (2003). Sex differences in the thermoregulation and evaporative water loss of a heterothermic bat, *Lasiurus cinereus*, during its spring migration. *J. Exp. Biol.* 206, 3381-3390. doi:10.1242/jeb.00574
- Cunningham, S. J., Kruger, A. C., Nxumalo, M. P. and Hockey, P. A. R. (2013). Identifying biologically meaningful hot-weather events using threshold temperatures that affect life-history. *PLoS ONE* 8, 1-11. doi:10.1371/journal.pone.0082492
- Czenze, Z. J., Kemp, R., van Jaarsveld, B., Freeman, M. T., Smit, B., Wolf, B. O. and McKechnie, A. E. (2020). Regularly drinking desert birds have greater evaporative cooling capacity and higher heat tolerance limits than non-drinking species. *Funct. Ecol.* 34, 1589-1600. doi:10.1111/1365-2435.13573
- Dawson, W. R. (1954). Temperature regulation and water requirements of the brown and abert towhees, Pipilo fuscus and Pipilo aberti. In *University of California Publications in Zoology*, Vol. 59 (ed. G. A. Bartholomew, F. Crescitelli, T. H. Bullock, W. H. Furgason and A. M. Schechtman), pp. 343-390. USA: University of California Press.
- Dmi'el, R. and Tel-Tzur, D. (1985) Heat balance of two starling species (Sturnus vulgaris) from temperate and desert habitats. J. Comp. Physiol. B 155, 395-402. doi:10.1007/BF00687484
- Dupoué, A., Rutschmann, A., Le Galliard, J. F., Miles, D. B., Clobert, J., DeNardo, D. F., Brusch, G. A. and Meylan, S. (2017). Water availability and environmental temperature correlate with geographic variation in water balance in common lizards. *Oecologia* 185,561-571. doi:10.1007/s00442-017-3973-6
- Fick, S. E. and Hijmans, R. J. (2017). WorldClim 2: new 1-km spatial resolution climate surfaces for global land areas. *Int. J. Climatol.* 37, 4302-4315. doi:10. 1002/joc.5086
- Freeman, M. T., Czenze, Z. J., Schoeman, K. and McKechnie, A. E. (2020).
  Extreme hyperthermia tolerance in the world's most abundant wild bird. Sci. Rep.
  10, 1-6. doi:10.1038/s41598-020-69997-7
- Gaston, K. J., Chown, S. L., Calosi, P., Bernardo, J., Bilton, D. T., Clarke, A., Clusella-Trullas, S., Ghalambor, C. K., Konarzewski, M., Peck, L. S. et al. (2009). Macrophysiology: a conceptual reunification. *Am. Nat.* 174, 595-612. doi:10.1086/605982
- Gerson, A. R., Smith, E. K., Smit, B., McKechnie, A. E. and Wolf, B. O. (2014). The impact of humidity on evaporative cooling in small desert birds exposed to high air temperatures. *Physiol Biochem. Zool.* 87, 782-795. doi:10.1086/678956
- Gerson, A. R., McKechnie, A. E., Smit, B., Whitfield, M. C., Smith, E. K., Talbot, W. A., McWhorter, T. J., Wolf, B. O. (2019). The functional significance of facultative hyperthermia varies with body size and phylogeny in birds. *Funct. Ecol.* 33, 597-607. doi:10.1111/1365-2435.13274
- **Glanville, E. J., Murray, S. A. and Seebacher, F.** (2012). Thermal adaptation in endotherms: climate and phylogeny interact to determine population-level responses in a wild rat. *Funct. Ecol.* **26**, 390-398. doi:10.1111/j.1365-2435. 2011.01933.x
- Grueber, C. E., Nakagawa, R. J., Lawis, R. J. and Jamieson, I. G. (2011).
  Multimodel inference in ecology and evolution: challenges and solutions. *J. Evol. Biol.* 24, 699-711. doi:10.1111/i.1420-9101.2010.02210.x
- Guillon, M., Guiller, G., DeNardo, D. F. and Lourdais, O. (2014). Microclimate preferences correlate with contrasted evaporative water loss in parapatric vipers at their contact zone. *Can. J. Zool.* **92**, 81-86. doi:10.1139/cjz-2013-0189
- Karasov, W. H. (1990). Digestion in birds: chemical and physiological determinants and implications. In *Studies in Avian Biology* (ed. M. L. Morrison, C. J. Ralph, J. Verner and J. R. Jehl), pp. 391-415. Los Angeles: Cooper Ornithological.
- Keller, I. and Seehausen, O. (2012). Thermal adaptation and ecological speciation.
  Mol. Ecol. 21, 782-799. doi:10.1111/j.1365-294X.2011.05397.x
- Kemp, A. C. (1995). *The Hornbills: Bucerotiformes*. United Kindom: Oxford University Press.
- Kemp, A. C. (1969). Some observations on the sealed-in nesting method of hornbills (family: Bucerotidae). Ostrich 40, 149-152. doi:10.1080/00306525.1969. 9639117
- Kemp, A. C. (2005). Southern Yellow-Billed Hornbill In Roberts Birds of Southern Africa (ed. P. A. R. Hockey, W. R. J. Dean and P. G. Ryan), pp 152-153. Cape Town: The Trustees of the John Voelcker Bird Book Fund.
- Kemp, R. and McKechnie, A. E. (2019). Thermal physiology of a range-restricted desert lark. J. Comp. Physiol. B 189, 131-141. doi:10.1007/s00360-018-1190-1

- Kemp, R., Freeman, M. T., van Jaarsveld, B., Czenze, Z. J., Conradie, S. R. and McKechnie, A. E. (2020). Sublethal fitness costs of chronic exposure to hot weather vary between sexes in a threatened desert lark. *Emu* 120, 216-229. doi:10.1080/01584197.2020.1806082
- Ketola, T., Kristensen, T. N., Kellermann, V. M. and Loeschcke, V. (2012). Can evolution of sexual dimorphism be triggered by developmental temperatures? J. Evol. Biol. 25, 847-855. doi:10.1111/j.1420-9101.2012.02475.x
- Klaassen, M., Brenninkmeijer, A., Boix-hinzen, C. and Mendelsohn, J. (2003). Fathers with highly demanding partners and offspring in a semidesert environment: energetic aspects of the breeding system of monteiro's hornbills (*Tockus montei*) in Namibia. *Auk* 120,866-873. doi:10.1642/0004-8038(2003)120[0866:FWHDPA]2.0.CO;2
- Klüg-Baerwald, B. J. and Brigham, R. M. (2017). Hung out to dry? Intraspecific variation in water loss in a hibernating bat. *Oecologia* 183,977-985. doi:10.1007/ s00442-017-3837-0
- Lailvaux, S. P., Alexander, G. J. and Whiting, M. J. (2003). Sex-based differences and similarities in locomotor performance, thermal preferences, and escape behaviour in the lizard *Platysaurus intermedius wilhelmi*. *Physiol*. *Biochem*. *Zool*. 76, 511-521. doi:10.1086/376423
- **Lighton, J. R. B.** (2008). *Measuring Metabolic Rates: a Manual for Scientists*. United Kingdom: Oxford University Press.
- Liwanag, H. E. M., Haro, D., Callejas, B., Labib, G. and Pauly, G. B. (2018). Thermal tolerance varies with age and sex for the nonnative italian wall lizard (*Podarcis siculus*) in Southern California. *J. Therm. Biol.* **78**, 263-269. doi:10. 1016/j.jtherbio.2018.10.010
- Lovegrove, B. G. (2000). The zoogeography of mammalian basal metabolic rate.
  Am. Nat. 156, 201-219. doi:10.1086/303383
- Lovegrove, B. G. (2003). The influence of climate on the basal metabolic rate of small mammals: a slow-fast continuum. *J. Comp. Physiol. B* **173**, 87-112. doi:10. 1007/s00360-002-0309-5
- Lovegrove, B. G., Canale, C., Levesque, D., Fluch, G. and Rehakova-Petru, M. (2014). Are tropical small mammals physiologically vulnerable to arrhenius effects of climate change? *Physiol. Biochem. Zool.* 87, 000-000. doi:10.1086/673313
- McClennand, G. T. W., McKechnie, A. E. and Chown, S. L. (2016). Basal metabolic rate of the black-faced sheathbill (*Chionis minor*): intraspecific variation in a phylogenetically distinct, island endemic. *Physiol. Biochem. Zool.* 89, 141-150. doi:10.1086/685411
- McKechnie, A. E. and Swanson, D. L. (2010). Sources and significance of variation in basal summit and maximal metabolic rates in birds. *Curr. Biol.* 56, 741-758. doi:10.1093/czoolo/56.6.741
- McKechnie, A. E. and Wolf, B. O. (2010). Climate change increases the likelihood of catastrophic avian mortality events during extreme heat waves. *Biol. Lett.* 6, 253-256. doi:10.1098/rsbl.2009.0702
- McKechnie, A. E., Smit, B., Whitfield, M. C., Noakes, M. J., Talbot, W. A., Garcia, M., Gerson, A. R. and Wolf, B. O. (2016a). Avian thermoregulation in the heat: evaporative cooling capacity in an archetypal desert specialist, burchell's sandgrouse (*Pterocles burchelli*). *J. Exp. Biol.* 219, 2137-2144. doi:10.1242/jeb.
- McKechnie, A. E., Whitfield, M. C., Smit, B., Gerson, A. R., Smith, E. K., Talbot, W. A., McWhorter, T. J. and Wolf, B. O. (2016b). Avian thermoregulation in the heat: efficient evaporative cooling allows for extreme heat tolerance in four southern hemisphere columbids. J. Exp. Biol. 219, 2145-2155. doi:10.1242/jeb. 138776
- McKechnie, A. E., Gerson, A. R. and Wolf, B. O. (2021a). Thermoregulation in desert birds: scaling and phylogenetic variation in heat tolerance and evaporative cooling. J. Exp. Biol. 224, jeb229211. doi:10.1242/jeb.229211
- McKechnie, A. et al. (2021b), How hornbills handle heat: sex-specific thermoregulation in the southern yellow-billed hornbill. *Dryad Dataset*. doi:10. 5061/dryad.bcc2fqzbq
- McWhorter, T. J., Gerson, A. R., Talbot, W. A., Smith, E. K., McKechnie, A. E. and Wolf, B. O. (2018). Avian thermoregulation in the heat: evaporative cooling capacity and thermal tolerance in two australian parrots. *J. Exp. Biol.* 221, jeb168930. doi:10.1242/jeb.168930
- Menon, G. K., Baptista, L. F., Brown, B. E. and Elias, P. M. (1989). Avian epidermal differentiation II adaptive response of permeability barrier to water deprivation\replenishment. *Tissue Cell* **21**, 83-92. doi:10.1016/0040-8166(89)90023-2
- Merilä, J. and Hendry, A. P. (2014). Climate change, adaptation, and phenotypic plasticity: The problem and the evidence. *Evol. Appl.* 7, 1-14. doi:10.1111/eva. 12137
- Mills, M. S. L., Boix-Hinzen, C. and Du Plessis, M. A. (2005). Live or let live: life-history decisions of the breeding female Monteiro's Hornbill *Tockus monteiri*. *Ibis* 147, 48-56. doi:10.1111/i.1474-919X.2004.00340.x
- Moreau, R. E. and Moreau, W. M. (1941). Breeding biology of silvery-cheeked hornbill. Auk 58, 13-27. doi:10.2307/4078893
- Muñoz, M. M. and Bodensteiner, B. L. (2019). Janzen's hypothesis meets the bogert effect: connecting climate variation, thermoregulatory behaviour, and rates of physiological evolution. *Integr. Organ. Biol.* 1, 1-12. doi:10.1093/iob/oby002
- Myers, M. S. (2000). Breeding the Writhe-billed hornbill. *Int. Zoo. Yearb.* 37, 345-354. doi:10.1111/j.1748-1090.2000.tb00740.x

- Muggeo, V., Atkins, D. C., Gallop, R. J. and Dimidjian, S. (2014). Segmented mixed models with random change points: a maximum likelihood approach with application to treatment for depression study. Stat. Model. 14, 293-313. doi:10. 1177/1471082X13504721
- Naya, D. E., Spangenberg, L., Naya, H. and Bozinovic, F. (2013). How does evolutionary variation in basal metabolic rates arise? A statistical assessment and a mechanistic model. *Evolution* 67, 1463-1476. doi:10.1111/evo.12042
- Noakes, M. J. and McKechnie, A. E. (2019). Reaction norms for heat tolerance and evaporative cooling capacity do not vary across a climatic gradient in a passerine bird. *Comp. Biochem. Phys. A* **236**, 110522. doi:10.1016/j.cbpa.2019.06.022
- Noakes, M. J., Wolf, B. O. and McKechnie, A. E. (2016). Seasonal and geographical variation in heat tolerance and evaporative cooling capacity in a passerine bird. J. Exp. Biol. 219, 859-869. doi:10.1242/jeb.132001
- O'Connor, R. S., Wolf, B. O., Brigham, R. M. and McKechnie, A. E. (2017). Avian thermoregulation in the heat: efficient evaporative cooling in two southern African nightjars. J. Comp. Physiol. B 187, 477-491. doi:10.1007/s00360-016-1047-4
- Piersma, T. and Drent, J. (2003). Phenotypic flexibility and the evolution of organismal design. Trends in. Ecol. Evol. 18, 228-233. doi:10.1016/S0169-5347(03)00036-3
- du Plessis, K. L., Martin, R. O., Hockey, P. A. R., Cunningham, S. J. and Ridley, A. R. (2012). The costs of keeping cool in a warming world: implications of high temperatures for foraging, thermoregulation and body condition of an arid-zone bird. Glob. Chang. Biol. 18, 3063-3070. doi:10.1111/j.1365-2486.2012.02778.x
- **Pörtner, H., Bock, C. and Mark, F. C.** (2017). Oxygen- and capacity-limited thermal tolerance: bridging ecology and physiology. *J. Exp. Biol.* **220**, 2685-2696. doi:10. 1242/jeb.134585
- Reher, S., Ehlers, J., Rabarison, H. and Dausmann, K. H. (2018). Short and hyperthermic torpor responses in the malagasy bat *Macronycteris commersoni* reveal a broader hypometabolic scope in heterotherms. *J. Comp. Physiol. B* 188, 1015-1027. doi:10.1007/s00360-018-1171-4
- **Richards, S.-A.** (1970). Physiology of thermal panting in birds. *Ann. Biol. Anim. Biophys.* **10**, 151-168. doi:10.1051/rnd:19700614
- Roti Roti, J. L. (2008). Cellular responses to hyperthermia (40–46°C): cell killing and molecular events. *Int. J. Hyperth.* 24, 3-15. doi:10.1080/02656730701769841
- Rutherford, M. C., Mucina, L. and Powrie, L. W. (2006). Biomes and Bioregions of Southern Africa: the Vegetation of South Africa, Lesotho and Swaziland, South Africa: Strelitzia
- Van Sant, M. J., Oufiero, C. E., Muñoz-Garcia, A., Hammond, K. A. and Williams, J. B. (2012). A phylogenetic approach to total evaporative water loss in mammals. *Physiol. Biochem. Zool.* **85**, 526-532. doi:10.1086/667579
- Schmidt-Nielsen, K., Hainsworth, F. R. and Murrish, D. (1970). Counter-current heat exchange in the respiratory passages: effect on water and heat balance. *Respir. Physiol.* **9**, 263-276. doi:10.1016/0034-5687(70)90075-7
- Smit, B., Harding, C., Hockey, P. and McKechnie, A. E. (2013). Adaptive thermoregulation during summer in two populations of an arid-zone passerine. *Ecology* 94, 1142-1154. doi:10.1890/12-1511.1
- Song, X., Körtner, G. and Geiser, F. (1997). Thermal relations of metabolic rate reduction in a hibernating marsupial. Am. J. Physiol. Regul. Integr. Comp. Physiol. 273, R2097-R2104. doi:10.1152/ajpregu.1997.273.6.R2097
- Speakman, J. R. and Król, E. (2010). Maximal heat dissipation capacity and hyperthermia risk: neglected key factors in the ecology of endotherms. *J. Anim. Ecol.* 79, 726-746. doi:10.1111/j.1365-2656.2010.01689.x
- Sunday, J. M., Bates, A. E. and Dulvy, N. K. (2011). Global analysis of thermal tolerance and latitude in ectotherms. *Proc. Roy. Soc. B* 278,1823-1830. doi:10. 1098/rspb.2010.1295
- Symonds, M. R. E. and Moussalli, A. (2011). A brief guide to model selection, multimodel inference and model averaging in behavioural ecology using Akaike's information criterion. *Behav. Ecol. Sociobiol.* 65, 13-21. doi:10.1007/s00265-010-1037-6
- Teague, C., Youngblood, J. P., Ragan, K., Angilletta, M. J. and VandenBrooks, J. M. (2017). A positive genetic correlation between hypoxia tolerance and heat tolerance supports a controversial theory of heat stress. *Biol. Lett.* 13, 20170309. doi:10.1098/rsbl.2017.0309
- Tieleman, B. and Williams, J. B. (1999). The role of hyperthermia in the water economy of desert birds. *Physiol. Biochem. Zool.* **72**, 87-100. doi:10.1086/316640

- Tieleman, B. I. and Williams, J. B. (2002). Cutaneous and respiratory water loss in larks from arid and mesic environments. *Physiol. Biochem. Zool.* **75**, 590-599. doi:10.1086/344491
- Tieleman, B. I., Versteegh, M. A., Helm, B. and Dingemanse, N. J. (2009). Quantitative genetics parameters show partial independent evolutionary potential for body mass and metabolism in stonechats from different populations. *J. Zool.* 279, 129-136. doi:10.1111/j.1469-7998.2009.00597.x
- Tieleman, B. I., Williams, J. B. and Bloomer, P. (2003). Adaptation of metabolism and evaporative water loss along an aridity gradient. *Proc. Roy. Soc. B* **270**, 207-214. doi:10.1098/rspb.2002.2205
- Tracy, C. R., Welch, W. R., Pinshow, B. and Porter, W. P. (2010). Properties of Air: a Manual for Use in Biophysical Ecology. Madison, USA.
- Trost, C. H. (1972). Adaptations of horned larks (*Eremophila alpestris*) to hot environments. Auk 89,506-527.
- van Dyk, M., Noakes, M. J. and McKechnie, A. E. (2019). Interactions between humidity and evaporative heat dissipation in a passerine bird. *J. Comp. Physiol. B* 2, 299-308. doi:10.1007/s00360-019-01210-2
- van de Ven, T. M. F. N. (2017). Implications of climate change on the reproductive success of the Southern Yellow-billed Hornbill, *Tockus leucomelas*. *PhD thesis*, University of Cape Town, SA.
- van de Ven, T. M. F. N., Martin, R. O., Vink, T. J. F., McKechnie, A. E. and Cunningham, S. J. (2016). Regulation of heat exchange across the hornbill beak: functional similarities with toucans? *PLoS ONE* 11, e0154768. doi:10.1371/journal.pone.0154768
- van de Ven, T. M. F. N., McKechnie, A. E. and Cunningham, S. J. (2019). The costs of keeping cool: behavioural trade-offs between foraging and thermoregulation are associated with significant mass losses in an arid-zone bird. *Oecologia* **191**, 205-215. doi:10.1007/s00442-019-04486-x
- van de Ven, T. M. F. N., McKechnie, A. E. and Cunningham, S. J. (2020). High temperatures are associated with substantial reductions in breeding success and offspring quality in an arid-zone bird. *Oecologia* **193**, 225-235. doi:10.1007/s00442-020-04644-6
- Vidal, M. A., Ortiz, J. C. and Labra, A. (2008). Intraspecific variation in a physiological thermoregulatory mechanism: The case of the lizard *Liolaemus* tenuis (Liolaeminae). Rev. Chil. De. Hist. Nat. 81, 171-178. doi:10.4067/S0716-078X2008000200002
- **Walsberg, G. and Wolf, B. O.** (1995). Variation in the respiratory quotient of birds and implications for indirect calorimetry using measurements of carbon dioxide production. *J. Exp. Biol.* **198**, 213-219.
- Weathers, W. W. (1981). Physiological thermoregulation in heat-stressed birds: consequences of body size. *Physiol. Zool.* **54**, 345-361. doi:10.1086/physzool.54. 3.30159949
- Webster, M. D. and Bernstein, M. H. (1987). Ventilated capsule measurements of cutaneous evaporation in mourning doves. *Condor* 89, 868. doi:10.2307/1368535
   Williams, J. B. (1996). A phylogenetic perspective of evaporative water loss in birds. *Auk* 113, 457-472. doi:10.2307/4088912
- White, C. R., Blackburn, T. M., Martin, G. R. and Butler, P. J. (2007). Basal metabolic rate of birds is associated with habitat temperature and precipitation, not primary productivity. *Proc. Roy. Soc. B* 274, 287-293. doi:10.1098/rspb.2006. 3727
- White, C. R. and Seymour, R. S. (2003). Mammalian basal metabolic rate is proportional to body mass 2/3. Proc. Nat. Acad. Sci. 100, 4046-4049. doi:10.1073/ pnas.0436428100
- Whitfield, M. C., Smit, B., McKechnie, A. E. and Wolf, B. O. (2015). Avian thermoregulation in the heat: scaling of heat tolerance and evaporative cooling capacity in three southern African arid-zone passerines. *J. Exp. Biol.* 218, 1705-1714. doi:10.1242/jeb.121749
- Williams, J. B., Muñoz-Garcia, A., Ostrowski, S. and Tieleman, B. I. (2004). A phylogenetic analysis of basal metabolism, total evaporative water loss, and life-history among foxes from desert and mesic regions. *J. Comp. Physiol. B.* 174, 29-39. doi:10.1007/s00360-003-0386-0
- Witmer, M. C. (1992). Cooperative breeding by Rufous Hornbills on Mindanao Island, Philippines. *Auk* 110, 933-936. doi:10.2307/4088652
- Xue, Q., Majeed, M. Z., Zhang, W. and Sen, M. A. C. (2019). Adaptation of Drosophila species to climate change — A literature review since 2003. J. Integ. Agric. 18, 805-814. doi:10.1016/S2095-3119(18)62042-8