

**CHARACTERIZATIONS OF SAPONIN AND
PHENOLIC BIOACTIVE COMPOUNDS
EXTRACTED FROM FENUGREEK SEED AND
ALOE VERA LEAVES VIA MICROWAVE-
ASSISTED EXTRACTION METHOD**

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Thesis submitted in fulfillment of the requirements
for the award of the degree of
Doctor of Philosophy

College of Engineering
UNIVERSITI MALAYSIA PAHANG

FEBRUARY 2021

ACKNOWLEDGEMENTS

First and foremost, I would like to thank almighty Allah for giving me health, knowledge, strength, opportunity and ability to undertake my studies and complete this work.

During this journey, the Almighty Allah also blessed me to carry out this work under the supervision of Professor Dr. Abdurahman Hamid Nour, a role model, a friend, an inspiration and a hand of support. He was always eager to support me and provide me invaluable guidance, supports, great ideas, and suggestions that have helped me to complete this thesis. His knowledge and advice supported me to never give-up when faced life's problems during my studies. This thesis would not have been possible without his guidance and supports. I would really like to thank him for his assistance during my PhD journey. My special thanks also go to my co-supervisor Professor Dato Dr. Rosli Bin Mohd Yunus for his supports and guidance during my PhD studies.

I would also like to thank my husband Dr. Fahim Fayaz for his supports and helps in entire of my PhD journey. My thanks also go to my parents for their prayers and giving me the strength while being far from them. Thanks to my brothers and my only sister for encouraging and supporting me.

My special thanks go to University Malaysia Pahang for rewarding me Doctoral Research Scheme (DRS) for financial assistance. I would like to thank the honorable Dean, Deputy Dean, faculty staff and technical staff of Faculty of Chemical and Process Engineering Technology, University Malaysia Pahang.

ABSTRAK

Dalam kajian ini, potensi benih fenugreek dan Aloe vera sebagai sumber saponin, fenolik dan antioksidan telah dikaji. Pengekstrakan dibantu oleh ketuhar gelombang mikro (MAE) dan pengekstrakan Soxhlet digunakan untuk mendapatkan ekstrak. Kesan faktor eksperimen MAE seperti masa pengekstrakan (2-12 min), kuasa gelombang mikro (300-700 W), kepekatan etanol (20-100%), nisbah pelarut (F:S) (1:8-1:16 dan 1:18-1:22 g/mL) dan suhu pengekstrakan (40-80°C) dinilai menggunakan (OFAT). Faktor pengekstrakan Soxhlet termasuk masa pengekstrakan (1-5 h), kepekatan etanol (20-100%) dan F: S (1:14-1:24 g/mL) juga dikaji. Keputusan menunjukkan hasil pengekstrakan yang lebih tinggi, jumlah kandungan saponin (TSC) dan jumlah kandungan fenolik (TPC) diperoleh melalui MAE berbanding pengekstrakan Soxhlet dalam penjimatan masa. Hasil maksimum dalam pengekstrakan Soxhlet diperoleh pada 3 jam masa pengekstrakan, kepekatan etanol 60% dan nisbah 1:20 g/mL F:S yang $19.35 \pm 0.75\%$, 125.04 ± 1.55 mg DE/g d.w. dan 60.13 ± 2.04 mg GAE/g d.w. untuk benih fenugreek dan $22.45 \pm 0.76\%$, 44.78 ± 1.01 mg OAE/g d.w. dan 49.99 ± 0.56 mg GAE/g d.w. untuk Aloe vera. Penyaringan terhadap “factors via two-level factorial design” dijalankan untuk parameter MAE. Keputusan menunjukkan bahawa kepekatan etanol dan suhu pengekstrakan adalah faktor yang paling penting dan paling tidak penting dalam mencapai hasil maksimum. Pemilihan tahap faktor untuk proses pengoptimuman diperoleh berdasarkan faktor prapenilaian (OFAT). Faktor optimum yang terbaik adalah masa pengekstrakan (2-4 min), kuasa gelombang mikro (500-700 W dan 400-600 W), kepekatan etanol (40-80% dan 20-60%), F:S (1:8-1:12 dan 1:18-1:22 g/mL) dan suhu ialah 70 °C. Kondisi optimum untuk benih fenugreek dan Aloe vera menghasilkan hasil pengekstrakan, TSC dan TPC berada pada 2.84 min, 572.50 W, 63.68%, dan 1:9 g/mL. Berdasarkan keadaan optimum, hasil pengekstrakan, TSC dan TPC benih fenugreek adalah $26.04 \pm 0.88\%$, 195.89 ± 1.07 mg DE/g dw, 81.85 ± 0.61 mg GAE/g dw, dan ekstrak daun Aloe vera di MAE adalah 2.79 min, 478.95 W, 43.38% etanol, dan 1:19 g/mL. Berdasarkan kepada kondisi tersebut, hasil ekstrak daun TSC dan TPC dari Aloe vera adalah $36.17 \pm 1.13\%$, 65.89 ± 0.77 mg OAE/g d.w dan 73.05 ± 1.05 mg GAE/g d.w. Ekstrak yang diperoleh melalui MAE dan Soxhlet juga diuji untuk kapasiti antioxidant melalui DPPH dan ABTS, struktur dan ikatan menggunakan FTIR dan kajian morfologi menggunakan SEM. Keputusan menunjukkan bahawa ekstrak yang diperoleh melalui MAE menunjukkan kapasiti antioksidan yang lebih tinggi dengan nilai IC₅₀ yang rendah (195.27 ± 0.56 µg/mL; DPPH), (157.92 ± 1.11 µg/mL; ABTS), 12 puncak yang dikenal pasti dalam FTIR untuk benih fenugreek dan (275 ± 1.45 µg/mL; DPPH) (215.58 ± 0.57 µg/mL; ABTS), 11 puncak di FTIR dan masing-masing tekstur kemas dan tekstur terbuka melalui SEM. Walau bagaimanapun, ekstrak Soxhlet (224.47 ± 0.77 µg/mL; DPPH), (199.67 ± 0.96 µg/mL; ABTS) untuk benih fenugreek dan (305.79 ± 0.66 µg/mL; DPPH), (263.29 ± 1.21 µg/mL, ABTS), dengan 6 puncak yang dikenal pasti melalui FTIR dan liang yang tidak tertutup dan liang tertutup ditunjukkan melalui SEM pada kedua-dua tanaman. kajian kinetik dan sifat dielektrik untuk MAE juga dilakukan. Hasil LC-QTOF-MS dari optimum ekstrak mengesahkan kehadiran 58 saponin dan 27 fenolik dalam benih fenugreek dan 29 saponin dengan 32 fenolik dalam ekstrak Aloe. Optimum ekstrak juga menunjukkan sifat surfaktan seperti pembasahan, pengurangan ketegangan permukaan air, sifat berbuih dan emulsifikasi. Oleh itu, ekstrak ini boleh menjadi sumber saponin, fenolik, antioksidan dan pengemulsi semula jadi untuk makanan, kosmetik dan produk farmaseutikal.

ABSTRACT

In this study, the potential of fenugreek seed and Aloe vera leaves as a source of saponins, phenolics and antioxidants were investigated. Microwave-assisted extraction (MAE) and Soxhlet extraction (SE) were used to obtain the extracts. The effects of experimental factors in MAE such as extraction time (2-12 min), microwave power (300-700 W), ethanol concentration (20-100%), feed-to-solvent ratio (1:8-1:16 and 1:18-1:22 g/mL) and extraction temperature (40-80 °C) were evaluated using one-factor-at-a-time (OFAT), respectively. The SE factors including extraction time (1-5 h), ethanol concentration (20-100%) and feed-to-solvent ratio (1:14-1:24 g/mL) were also investigated. Results indicated the higher extraction yield, total saponin content (TSC) and Total phenolic content (TPC) were obtained via MAE compared to SE in a time saving process. The maximum yields in SE were obtained at 3 h of extraction time, 60 % ethanol concentration and 1:20 g/mL F:S ratio which were $19.35 \pm 0.75\%$, 125.04 ± 1.55 mg DE/g d.w. and 60.13 ± 2.04 mg GAE/g d.w. for fenugreek seed and $22.45 \pm 0.76\%$, 44.78 ± 1.01 mg OAE/g d.w. and 49.99 ± 0.56 mg GAE/g d.w. for Aloe vera leaves, respectively. Further screening of the factors via two-level factorial design was carried out for MAE parameters. Results indicated that ethanol concentration and extraction temperature were the most and least significant factors in achieving maximum recoveries of the yields, respectively. The selection of factor levels for optimization process was obtained based on the pre-evaluation of factors (OFAT). The best points for optimizing the factors were extraction time (2-4 min), microwave power (500-700 W and 400-600 W), ethanol concentration (40-80% and 20-60%), feed-to-solvent ratio (1:8-1:12 and 1:18-1:22 g/mL) and constant temperature of 70 °C, respectively. The optimal MAE conditions for fenugreek seed and Aloe vera leaves extraction yield, TSC and TPC were at 2.84 min, 572.50 W, 63.68%, and 1:9 g/mL. Based on the optimum condition, the responses of extraction yield, TSC and TPC of fenugreek seed were $26.04 \pm 0.88\%$, 195.89 ± 1.07 mg DE/g d.w., 81.85 ± 0.61 mg GAE/g d.w. and for Aloe vera leaves extracts in MAE were 2.79 min, 478.95 W, 43.38% ethanol, and 1:19 g/mL. Where, based on these conditions, the extraction yield, TSC and TPC of Aloe vera leaves extract were $36.17 \pm 1.13\%$, 65.89 ± 0.77 mg OAE/g d.w and 73.05 ± 1.05 mg GAE/g d.w, respectively. The extracts obtained via MAE and SE were also tested for its antioxidant capacity via 2,2-diphenyl-1-picrylhydrazyl (DPPH) and 2,20-azino- bis (3-ethylbenzothiazoline-6-sulfonic acid) (ABTS), structure and bonding using Fourier transform infrared (FTIR) and morphological studies using SEM. Results indicated that extracts obtained via MAE showed higher antioxidant capacity with low IC₅₀ values of (195.27 ± 0.56 µg/mL; DPPH), (157.92 ± 1.11 µg/mL; ABTS), 12 identified peaks in FTIR for fenugreek seed and (275 ± 1.45 µg/mL; DPPH) (215.58 ± 0.57 µg/mL; ABTS), 11 peaks in FTIR and more wrapped and opened texture via scanning electron microscope (SEM), respectively. However, in SE it was (224.47 ± 0.77 µg/mL; DPPH), (199.67 ± 0.96 µg/mL; ABTS) for fenugreek seed and (305.79 ± 0.66 µg/mL; DPPH), (263.29 ± 1.21 µg/mL; ABTS), with 6 identified peaks via FTIR and closed pores showed via SEM in both plants, respectively. kinetic studies and dielectric properties for MAE were also carried out. The LC-QTOF-MS result of optimized extracts also confirmed the presence of 58 saponins and 27 phenolic compounds in fenugreek seed and 29 saponin with 32 phenolic compounds in Aloe extract. The optimized extracts also indicated surfactant properties such as wetting, reduction of water surface tension, foaming and emulsification properties. Thus, these extracts can be a promising source of saponins, phenolics, antioxidants and natural co-emulsifier for food, cosmetics and pharmaceutical products.

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LIST OF SYMBOLS

R2	R- square
Adj. R2	Adjacent coefficient of determination
C	Carbon
A	Alpha
Θ	Theta
Y	Response
β_0	Constant term
β_i	Coefficient of linear factor
β_{ii}	Coefficient of quadratic parameter
β_{ij}	Coefficient of interaction parameters
ϵ'	Dielectric constant
ϵ''	Dielectric loss
τ	Relaxation time
ω	Angular frequency in radians per second
ϵ_∞	Complex permittivity
ϵ_s	Static permittivity
λ	Wavelength
λ_0	Wavelength in free space

LIST OF ABBREVIATIONS

MAE	Microwave-assisted extraction
UAE	Ultrasonic-assisted extraction
DCM	dichloromethane
TSC	Total saponin content
TPC	Total phenolic content
DPPH	2,2-diphenyl-1-picrylhydrazyl (DPPH),
ABTS	2,20-azino- bis (3-ethylbenzothiazoline-6-sulfonic acid
RSM	Response surface methodology
ANOVA	Analysis of variance
CV	Coefficient of variation
FCCCD	Face-centred central composite design
FTIR	Fourier transform infrared (FTIR) spectroscopy
SEM	Scanning electron microscopy
LC-QTOF-MS	Liquid chromatography-mass spectrometry quadrupole time-of-flight
OFAT	One-factor-at-a-time
DNA	Deoxyribonucleic acid
WHO	World health organization
DOE	Design expert
<i>D_P</i>	Penetration depth

REFERENCES

- Adão, C. R., Da Silva, B. P., & Parente, J. P. (2011). A new steroidal saponin with antiinflammatory and antiulcerogenic properties from the bulbs of *Allium ampeloprasum* var. *porrum*. *Fitoterapia*, 82(8), 1175–1180. <https://doi.org/10.1016/j.fitote.2011.08.003>
- Aguilar-Reynosa, A., Romani, A., Rodriguez-Jasso, R. M., Aguilar, C. N., Garrote, G., & Ruiz, H. A. (2017). Microwave heating processing as alternative of pretreatment in second-generation biorefinery: An overview. *Energy Conversion and Management*, 136, 50-65. doi: 10.1016/j.enconman.2017.01.004
- Akbari, S., Abdurahman, N. H., Yunus, R. M., Alara, O. R., & Abayomi, O. O. (2019). Extraction, characterization and antioxidant activity of fenugreek (*Trigonella foenum graecum*) seed oil. *Materials Science for Energy Technologies*, 2(2), 349-355. doi:10.1016/j.mset.2018.12.001
- Akbari, S., Nour, A. H., Jamari, S. S., & Rajabi, A Q., (2016). Demulsification of water-in-crude oil emulsion via conventional heating and microwave heating technology in their optimum conditions. *Australian Journal of Basic and Applied Sciences*, 10(4), 66-74. https://papers.ssrn.com/sol3/papers.cfm?abstract_id=2792246
- Al-Asadi, J. N. (2014). Therapeutic uses of fenugreek (*Trigonella foenum-graecum L.*). *American Journal Society Issuesnand Humanities*, 21-36.
- Alara, O. R., Abdurahman, N. H., Mudalip, S. K. A., & Olalere, O. A. (2018). Microwave-assisted extraction of *Vernonia amygdalina* leaf for optimal recovery of total phenolic content. *Journal of Applied Research on Medicinal and Aromatic Plants*, 10, 16-24. doi:10.1016/j.jarmap.2018.04.004
- Alara, O. R., Abdurahman, N. H., & Olalere, O. A. (2018a). Ethanolic extraction of bioactive compounds from *Vernonia amygdalina* leaf using response surface methodology as an optimization tool. *Journal of Food Measurement and Characterization*, 12(2), 1107-1122. doi:10.1007/s11694-018-9726-3
- Alara, O. R., Abdurahman, N. H., & Olalere, O. A. (2017). Ethanolic extraction of flavonoids, phenolics and antioxidants from *Vernonia amygdalina* leaf using two-level factorial design. *Journal of King Saud University-Science*, 32(1), 7-16. doi: 10.1016/j.jksus.2017.08.001

- Alara, O. R., & Abdurahman, N. H. (2019). Kinetics studies on effects of extraction techniques on bioactive compounds from Vernonia cinerea leaf. *Journal of Food Science and Technology*, 56(2), 580–588. <https://doi.org/10.1007/s13197-018-3512-4>
- Alara, O. R., Abdurahman, N. H., Ukaegbu, C. I., & Azhari, N. H. (2018b). Vernonia cinerea leaves as the source of phenolic compounds, antioxidants, and anti-diabetic activity using microwave-assisted extraction technique. *Industrial Crops and Products*, 122, 533-544. doi: 10.1016/j.indcrop.2018.06.034
- Almutairi M. S., & Ali M. (2014). Direct detection of saponins in crude extracts of soapnuts by FTIR, Natural Product Research, 29(13), 1271-1275. doi: 10.1080/14786419.2014.992345
- Altemimi, A., Lakhssassi, N., Baharlouei, A., Watson, D. G., & Lightfoot, D. A. (2017). Phytochemicals: Extraction, isolation, and identification of bioactive compounds from plant extracts. *Plants*, 6(4), 1-23. doi:10.3390/plants6040042
- Amid, B.T., Mirhosseini, H. (2012). Effect of different purification techniques on the characteristics of heteropolysaccharide-protein biopolymer from durian (*Durio zibethinus*) seed. *Molecules* 2012, 17(9), 10875-10892; doi:10.3390/molecules170910875
- Amin, A., Alkaabi, A., Al-Falasi, S., Daoud, S.A. (2005). Chemopreventive activities of *Trigonella foenum graecum* (Fenugreek) against breast cancer. *Cell Biology International* 29 (2005) 687-694. doi:10.1016/j.cellbi.2005.04.004
- Anderson, M. (1997). Design of experiments. Ind. Phys. 3, 24. doi/10.1201/9781420055085-c15
- Arivalagan, M., Gangopadhyay, K. K., & Kumar, G. (2013). Determination of steroidal saponins and fixed oil content in fenugreek (*Trigonella foenum-graecum*) genotypes. *Indian journal of pharmaceutical sciences*, 75(1), 110. doi: 10.4103/0250-474X.113542
- Arunkumar, S., & Muthuselvam, M. (2009). Analysis of phytochemical constituents and antimicrobial activities of *Aloe vera* L. against clinical pathogens. *World Journal of Agricultural Sciences*, 5(5), 572-576.
- Ashraf, M. F., Abd Aziz, M., Stanslas, J., Ismail, I., & Abdul Kadir, M. (2013). Assessment of antioxidant and cytotoxicity activities of saponin and crude extracts of *Chlorophytum borivilianum*. *The Scientific World Journal*, 1-7. doi:10.1155/2013/216894

- Atirah, N., Abbas, M., & Zaini, A. (2017). Solvent selection in microwave assisted extraction of castor oil. *Chemical Engineering Transactions*, 56, 865–870. <https://doi.org/10.3303/CET1756145>
- Augustin, J. M., Kuzina, V., Andersen, S. B., & Bak, S. (2011). Molecular activities, biosynthesis and evolution of triterpenoid saponins. *Phytochemistry*, 72(6), 435–457. <https://doi.org/10.1016/j.phytochem.2011.01.015>
- Azwanida, N. N. (2015). A review on the extraction methods use in medicinal plants, principle, strength and limitation. *Med Aromat Plants*, 4(196), 2167-0412. doi:10.4172/2167-0412.1000196
- Bahrami, Y., Zhang, W., Chataway, T., & Franco, C. (2014). Structural elucidation of novel saponins in the sea cucumber Holothuria lessoni. *Marine drugs*, 12(8), 4439-4473. doi:10.3390/md12084439
- Bale, A. S., & Shinde, N. H. (2013). Microwave-assisted extraction of essential oil from lemon leaves. *Int J Recent Sci Res*, 4(9), 1414-1417.
- Barba, F. J., Zhu, Z., Koubaa, M., Sant'Ana, A. S., & Orlien, V. (2016). Green alternative methods for the extraction of antioxidant bioactive compounds from winery wastes and by-products: A review. *Trends in Food Science & Technology*, 49, 96-109. doi:10.1016/j.tifs.2016.01.006
- Barbosa-Cánovas, G. V. (2013). Microwave-assisted extraction for bioactive compounds. Washington State University, USA. Springer. 238.
- Barthel, J., & Buchner, R. (1991). High frequency permittivity and its use in the investigation of solution properties. *Pure and Applied Chemistry*, 63(10), 1473–1482. <https://doi.org/10.1351/pac199163101473>
- Baruah, A., Bordoloi, M., & Baruah, H. P. D. (2016). Aloe vera: A multipurpose industrial crop. *Industrial Crops and Products*, 94, 951-963. doi:10.1016/j.indcrop.2016.08.034
- Belguith-Hadriche, O., Bouaziz, M., Jamoussi, K., El Feki, A., Sayadi, S., & Makni-Ayedi, F. (2010). Lipid-lowering and antioxidant effects of an ethyl acetate extract of fenugreek seeds in high-cholesterol-fed rats. *Journal of agricultural and food chemistry*, 58(4), 2116-2122. doi:10.1021/jf903186w
- Bezerra, M. A., Santelli, R. E., Oliveira, E. P., Villar, L. S., & Escaleira, L. A. (2008). Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta*, 76(5), 965-977. doi:10.1016/j.talanta.2008.05.019

- Bhadoriya, S. S., Madoriya, N., Shukla, K., & Parihar, M. S. (2013). Biosurfactants: A new pharmaceutical additive for solubility enhancement and pharmaceutical development. *Biochem Pharmacol*, 2(2), 113. doi:10.4172/2167-0501.1000113
- Boruah, B., & Gogoi, M. (2013). Plant based natural surfactants. *Asian Journal of Home Science*, 8(2), 759–762.
- Bowman, J. M., Braxton, M. S., Churchill, M. A., Hellie, J. D., Starrett, S. J., Causby, G. Y., Ellis, D.J., Ensley, S.D., Maness, S.J., Meyer, C.D., Sellers, J.R., Hua, Y., Woosley, R.S., Butcher, D.J., (1997). Extraction method for the isolation of terpenes from plant tissue and subsequent determination by gas chromatography. *Microchemical journal*, 56(1), 10-18. doi:10.1006/mchj.1996.1422
- Bracco, G., Holst, B., (2013). Surface science techniques, Springer Series in Surface Sciences. doi:10.1007/978-3-642-34243-1
- Chakraborty, S., Ghosh, M., Chakraborti, S., Jana, S., Sen, K. K., Kokare, C., & Zhang, L. (2015). Biosurfactant produced from Actinomycetes nocardiosis A17: Characterization and its biological evaluation. *International journal of biological macromolecules*, 79, 405-412. doi:10.1016/j.ijbiomac.2015.04.068
- Chan, C.-H. (2013). Optimization and modelling of microwave-assisted extraction of active compounds from Cocoa leaves. University of Malaya.
- Chan, K. W., Iqbal, S., Khong, N. M., Ooi, D. J., & Ismail, M. (2014). Antioxidant activity of phenolics-saponins rich fraction prepared from defatted kenaf seed meal. *LWT-Food Science and Technology*, 56(1), 181-186. doi:10.1016/j.lwt.2013.10.028
- Chan, K. W., Khong, N. M., Iqbal, S., & Ismail, M. (2013). Isolation and antioxidative properties of phenolics-saponins rich fraction from defatted rice bran. *Journal of Cereal Science*, 57(3), 480-485. doi:10.1016/j.jcs.2013.02.002
- Chang, H. J., Lu, X., Bonnett, J. F., Canfield, N. L., Han, K., Engelhard, M. H., Jung, K., Sprenkle, V. L., Li, G. (2018). Decorating β'' -alumina solid-state electrolytes with micron Pb spherical particles for improving Na wettability at lower temperatures. *Journal of Materials Chemistry A*, 6(40), 19703-19711. doi:10.1039/x0xx00000x
- Chassaing, B., Koren, O., Goodrich, J. K., Poole, A. C., Srinivasan, S., Ley, R. E., & Gewirtz, A. T. (2015). Dietary emulsifiers impact the mouse gut microbiota promoting colitis and metabolic syndrome. *Nature*, 519(7541), 92-96. doi:10.1038/nature14232.

Cheeke, P. R. (2000). Actual and potential applications of *Yucca schidigera* and *Quillaja saponaria* saponins in human and animal nutrition. In *Saponins in food, feedstuffs and medicinal plants* (pp. 241-254). Springer, Dordrecht. doi:10.1007/978-94-015-9339-7_25

Chen, D. Z., Chen, L. Q., Lin, M. X., Gong, Y. Q., Ying, B. Y., & Wei, D. Z. (2017). Esculetoside A inhibits LPS-induced acute kidney injury by activating PPAR- γ . *Microbial pathogenesis*, 110, 208-213. doi:10.1016/j.micpath.2017.06.037

Chen, H. F., Wang, G. H., Luo, Q., Wang, N. L., & Yao, X. S. (2009). Two new steroidal saponins from *Allium macrostemon* Bunge and their cytotoxicity on different cancer cell lines. *Molecules*, 14(6), 2246-2253. doi:10.3390/molecules14062246

Chen, X. F., Wu, H. T., Tan, G. G., Zhu, Z. Y., & Chai, Y. F. (2011). Liquid chromatography coupled with time-of-flight and ion trap mass spectrometry for qualitative analysis of herbal medicines. *Journal of pharmaceutical analysis*, 1(4), 235-245. doi: 10.1016/j.jpha.2011.09.008

Chen, Y., Xie, M. Y., & Gong, X. F. (2007). Microwave-assisted extraction used for the isolation of total triterpenoid saponins from *Ganoderma atrum*. *Journal of Food Engineering*, 81(1), 162-170. doi: /10.1016/j.jfoodeng.2006.10.018

Chen, Y. F., Yang, C. H., Chang, M. S., Ciou, Y. P., & Huang, Y. C. (2010). Foam properties and detergent abilities of the saponins from *Camellia oleifera*. *International Journal of Molecular Sciences*, 11(11), 4417-4425. Doi: 10.3390/ijms11114417

Cheng, H., Feng, S., Jia, X., Li, Q., Zhou, Y., & Ding, C. (2013). Structural characterization and antioxidant activities of polysaccharides extracted from *Epimedium acuminatum*. *Carbohydrate polymers*, 92(1), 63-68. doi: 10.1016/j.carbpol.2012.09.051

Cheok, C. Y., Salman, H. A. K., & Sulaiman, R. (2014). Extraction and quantification of saponins: A review. *Food Research International*, 59, 16-40. doi: 10.1016/j.foodres.2014.01.057

Dahmoune, F., Nayak, B., Moussi, K., Remini, H., & Madani, K. (2015). Optimization of microwave-assisted extraction of polyphenols from *Myrtus communis* L . leaves. *Food Chemistry*, 166, 585–595. <https://doi.org/10.1016/j.foodchem.2014.06.066>

De Almeida, D. G., Soares Da Silva, R. D. C. F., Luna, J. M., Rufino, R. D., Santos, V. A., Banat, I. M., & Sarubbo, L. A. (2016). Biosurfactants: promising molecules for petroleum biotechnology advances. *Frontiers in microbiology*, 7, 1718. Doi: 10.3389/fmicb.2016.01718

Demirel, M., & Kayan, B. (2012). Application of response surface methodology and central composite design for the optimization of textile dye degradation by wet air oxidation. *International Journal of Industrial Chemistry*, 3(1), 24. doi: 10.1186/2228-5547-3-24

Desai, M., Parikh, J., & Parikh, P. A. (2010). Extraction of natural products using microwaves as a heat source. *Separation & Purification Reviews*, 39(1-2), 1-32. doi: 10.1080/15422111003662320

Devaraj, A., & Karpagam, T. (2011). Evaluation of anti-inflammatory activity and analgesic effect of Aloe vera leaf extract in rats. *International Research Journal of Pharmacy*, 2(3), 103-110.

Dhobi, M., Mandal, V., & Hemalatha, S. (2009). Optimization of microwave assisted extraction of bioactive flavonolignan-silybinin. *Journal of chemical metrology*, 3(1), 13-23.

Doyle, J.J., (2001) Leguminosae. Encycl. Genet. 1081–1085.
doi:10.1006/rwgn.2001.1642

Duan, S., Jiang, Y., Geng, T., Ju, H., & Wang, Y. (2020). Wetting, foaming, and emulsification properties of novel methyltriphenylphosphonium carboxylate ionic liquid surfactants. *Journal of Dispersion Science and Technology*, 41(1), 47-53. doi:10.1080/01932691.2018.1541416

Dudonne, S., Vitrac, X., Coutiere, P., Woillez, M., & Mérillon, J. M. (2009). Comparative study of antioxidant properties and total phenolic content of 30 plant extracts of industrial interest using DPPH, ABTS, FRAP, SOD, and ORAC assays. *Journal of agricultural and food chemistry*, 57(5), 1768-1774. Doi: 10.1021/jf803011r

Elazzazy, A. M., Abdelmoneim, T. S., & Almaghrabi, O. A. (2015). Isolation and characterization of biosurfactant production under extreme environmental conditions by alkali-halo-thermophilic bacteria from Saudi Arabia. *Saudi Journal of Biological Sciences*, 22(4), 466-475. doi: 10.1016/j.sjbs.2014.11.018

Fang, X., Wang, J., Hao, J., Li, X., & Guo, N. (2015). Simultaneous extraction, identification and quantification of phenolic compounds in Eclipta prostrata using microwave-assisted extraction combined with HPLC–DAD–ESI–MS/MS. *Food chemistry*, 188, 527-536. doi: 10.1016/j.foodchem.2015.05.037

Farhan Aslam, M., Majeed, S., Aslam, S., & Irfan, J. A. (2017). Vitamins: key role players in boosting up immune response-a mini review. *Vitamins & Minerals*, 6(1), 1–8. <https://doi.org/10.4172/2376-1318.1000153>

- Feng, J., Zeng, Y., Ma, C., Cai, X., Zhang, Q., Tong, M., Tong, M., Yu, B., Xu, P. (2006). The surfactant tween 80 enhances biodesulfurization. *Appl. Environ. Microbiol.*, 72(11), 7390-7393. doi: 10.1128/AEM.01474-06
- Floegel, A., Kim, D., Chung, S., Koo, S.I., Chun, O.K. (2011). Comparison of ABTS / DPPH assays to measure antioxidant capacity in popular antioxidant-rich US foods. *Journal of Food Composition and Analysis*, 24, 1043–1048. doi:10.1016/j.jfca.2011.01.008
- Fracchia, L., Cavallo, M., Martinotti, M. G., & Banat, I. M. (2012). Biosurfactants and bioemulsifiers biomedical and related applications—present status and future potentials. *Biomedical Science, Engineering and Technology*, 14, 326-335. doi:10.5772/23821
- Fracchia, L., Banat, J. J., Cavallo, M., & Banat, I. M. (2015). Potential therapeutic applications of microbial surface-activecompounds. *AIMS Bioengineering*, 2(3), 144-162. doi: 10.3934/bioeng.2015.3.144
- Frey, D. D., & Jugulum, R. (2006). The mechanisms by which adaptive one-factor-at-a-time experimentation leads to improvement. *Journal of Mechanical Design*, 128(5), 1050. doi:10.1115/1.2216733
- Garg, C., Verma, S., Satija, S., Mehta, M., Dureja, H., & Garg, M. (2016). Microwave assisted extraction of bioactive compound phyllanthin from Phyllanthus amarus and optimization using central composite design. *International Journal of Pharmaceutical Science and Research*, 1(7), 30-35.
- Gholamreza, D.N., Fariba, S., Payam, K., Mohajeri, E., & Jahanbakhsh, J. (2011). Formulation of herbal conditioner shampoo by using extract of fenugreek seeds and evaluation of its physicochemical parameters. *African Journal of Pharmacy and Pharmacology*, 5(22), 2420–2427. <https://doi.org/10.5897/AJPP11.121>
- Ghosh, D., Pradhan, A. K., Mondal, S., Begum, N. A., & Mandal, D. (2014). Proton transfer reactions of 4'-chloro substituted 3-hydroxyflavone in solvents and aqueous micelle solutions. *Physical Chemistry Chemical Physics*, 16(18), 8594-8607. doi: 10.1039/c3cp52209a
- Giada, M. de L. R. (2013). Food phenolic compounds: main classes, sources and their antioxidant power. *Oxidative stress and chronic degenerative diseases—A role for antioxidants* (pp. 87–112).
- Gil-Ramirez, A., Salas-Veizaga, D. M., Grey, C., Karlsson, E. N., Rodriguez-Meizoso, I., & Linares-Pastén, J. A. (2018). Integrated process for sequential extraction of saponins, xylan and cellulose from quinoa stalks (*Chenopodium quinoa* Willd.). *Industrial Crops and Products*, 121, 54-65.

Goel, P.K., Panchkula. (2010). European Patent Office No. EP2285821A1, Novel process for the extraction of furostanolic saponins from fenugreek seeds.

Güçlü-Üstündağ, Ö., & Mazza, G. (2007). Saponins: properties, applications and processing. *Critical Reviews in Food Science and Nutrition*, 47(3), 231-258. doi:10.1080/10408390600698197

Gupta, P., Khanday, W. A., Majid, S. A., Kushwa, V., Tomar, S. S., & Tomar, R. (2013). Study of sorption of metal oxoanions from waste water on surfactant modified analog of laumontite. *Journal of Environmental Chemical Engineering*, 1(3), 510-515. doi:10.1016/j.jece.2013.06.016

Hameed, B. S., Bhatt, C. S., Nagaraj, B., & Suresh, A. K. (2018). Chromatography as an efficient technique for the separation of diversified nanoparticles. *Nanomaterials in Chromatography*, 503-518. doi: 10.1016/b978-0-12-812792-6.00019-4.

Haritha, K., Ramesh, B., & Saralakumari, D. (2014). Effect of Aloe vera gel on antioxidant enzymes in streptozotocin-induced cataractogenesis in male and female Wistar rats. *Journal of Acute Medicine*, 4(1), 38-44. doi: 0.1016/j.jacme.2014.01.005

Hierroa, J. N. del, Herrera, T., García-Risco, M. R., Fornari, T., Reglero, G., & Martin, D. (2018). Ultrasound-assisted extraction and bioaccessibility of saponins from edible seeds: quinoa, lentil, fenugreek, soybean and lupin. *Food Research International*, 109, 440–447. <https://doi.org/10.1016/j.foodres.2018.04.058>

Hu, T., Guo, Y. Y., Zhou, Q. F., Zhong, X. K., Zhu, L., Piao, Chen, J. H., Jiang, J. G. (2012). Optimization of ultrasonic-assisted extraction of total saponins from Eclipta prostrata L. using response surface methodology. *Journal of Food Science*, 77(9), C975-C982. Doi: 10.1111/j.1750-3841.2012.02869.x

Ibe, C., Jacobs, C. C., Imo, C., Osuocha, K. U., & Okoronkwo, M. U. (2014). Evaluation of the antioxidant activities of Psidium guajava and Aloe vera. *Journal of Pharmaceutical Research International*, 397-406. doi: 10.9734/bjpr/2014/6989

Ince, A. E., Sahin, S., & Sumnu, G. (2014). Comparison of microwave and ultrasound-assisted extraction techniques for leaching of phenolic compounds from nettle. *Journal of food science and technology*, 51(10), 2776-2782. doi: 10.1007/s13197-012-0828-3

Iness Bettaieb, Soumaya Bourgou, Wissem Aidiwannes, Ibtissem Hamrouni, Ferid Limam, B. (2010). Essential oils , phenolics , and antioxidant activities of different parts of cumin (Cuminum cyminum L .). *Journal of Agricultural and Food Chemistry*, 58, 10410–10418. <https://doi.org/10.1021/jf102248j>

Iqbal, J., Abbasi, B. A., Mahmood, T., Kanwal, S., Ali, B., Shah, S. A., & Khalil, A. T. (2017). Plant-derived anticancer agents: A green anticancer approach. *Asian Pacific Journal of Tropical Biomedicine*, 7(12), 1129–1150. <https://doi.org/10.1016/j.apjtb.2017.10.016>

Jang, G. H., Kim, H. W., Lee, M. K., Jeong, S. Y., Bak, A. R., Lee, D. J., & Kim, J. B. (2018). Characterization and quantification of flavonoid glycosides in the *Prunus* genus by UPLC-DAD-QTOF/MS. *Saudi Journal of Biological Sciences*, 25(8), 1622-1631. doi: 10.1016/j.sjbs.2016.08.001

Jha, S. S., Joshi, S. J., & SJ, G. (2016). Lipopeptide production by *Bacillus subtilis* R1 and its possible applications. *Brazilian Journal of Microbiology*, 47(4), 955-964. doi: 10.1016/j.bjm.2016.07.006

Jose S. Dambolena, Maria P. Zunino, Enrique I. Lucini, Ruben Olmedo, Erika Banchio, Paula J. Bima, J. A. Z. (2010). Total phenolic content, radical scavenging properties , and essential oil composition of *origanum* species from different populations. *Journal of Agricultural and Food Chemistry*, 58, 1115–1120. <https://doi.org/10.1021/jf903203n>

Karabegović, I. T., Stojičević, S. S., Veličković, D. T., Nikolić, N. Č., & Lazić, M. L. (2013). Optimization of microwave-assisted extraction and characterization of phenolic compounds in cherry laurel (*Prunus laurocerasus*) leaves. *Separation and Purification Technology*, 120, 429-436. doi: 10.1016/j.seppur.2013.10.021

Karami, Z., Emam-Djomeh, Z., Mirzaee, H. A., Khomeiri, M., Mahoonak, A. S., & Aydani, E. (2015). Optimization of microwave assisted extraction (MAE) and soxhlet extraction of phenolic compound from licorice root. *Journal of food science and technology*, 52(6), 3242-3253. doi: 10.1007/s13197-014-1384-9

Karimi, A., Majlesi, M., & Rafieian-Kopaei, M. (2015). Herbal versus synthetic drugs; beliefs and facts. *Journal of Nephropharmacology*, 4(1), 27-30

Kaur, R., Arora, S., & Thukral, A. (2015). Quantitative and qualitative analysis of saponins in different plant parts of *Chlorophytum borivilianum*. *International Journal of Pharma and Bio Sciences* 6(1), 826-35.

Kaviarasan, S., Ramamurty, N., Gunasekaran, P., Varalakshmi, E., & Anuradha, C. V. (2006). Fenugreek (*Trigonella foenum graecum*) seed extract prevents ethanol-induced toxicity and apoptosis in Chang liver cells. *Alcohol and Alcoholism*, 41(3), 267-273. doi: 10.1093/alcalc/agl020

- Keshari, A. K., Srivastava, R., Singh, P., Yadav, V. B., & Nath, G. (2018). Antioxidant and antibacterial activity of silver nanoparticles synthesized by Cestrum nocturnum. *Journal of Ayurveda and Integrative Medicine*. 1–8. doi:10.1016/j.jaim.2017.11.003
- Khan, M. I., Ahhmed, A., Shin, J. H., Baek, J. S., Kim, M. Y., & Kim, J. D. (2018). Green tea seed isolated saponins exerts antibacterial effects against various strains of gram positive and gram negative bacteria, a comprehensive study in vitro and in vivo. *Evidence-Based Complementary and Alternative Medicine*, doi: 10.1155/2018/3486106
- Khaniabadi, Y. O., Heydari, R., Nourmoradi, H., Basiri, H., & Basiri, H. (2016). Low-cost sorbent for the removal of aniline and methyl orange from liquid-phase: Aloe Vera leaves wastes. *Journal of the Taiwan institute of chemical engineers*, 68, 90-98. doi: 10.1016/j.jtice.2016.09.025
- Khuri, A. I., & Mukhopadhyay, S. (2010). Response surface methodology. *Wiley Interdisciplinary Reviews: Computational Statistics*, 2(2), 128-149.
- Kothekar, S. C., Ware, A. M., Waghmare, J. T., & Momin, S. A. (2007). Comparative analysis of the properties of Tween-20, Tween-60, Tween-80, Arlacel-60, and Arlacel-80. *Journal of dispersion science and technology*, 28(3), 477-484. doi: 10.1080/01932690601108045
- Kregiel, D., Berlowska, J., Witonska, I., Antolak, H., Proestos, C., Babic, M., & Zhang, B. (2017). Saponin-based, biological-active surfactants from plants. *Application and characterization of surfactants*, 183-205.
- Kumar, S., Yadav, A., Yadav, M., & Yadav, J. P. (2017). Effect of climate change on phytochemical diversity, total phenolic content and in vitro antioxidant activity of Aloe vera (L.) Burm.f. *BMC Research Notes*, 10(1), 1–12. <https://doi.org/10.1186/s13104-017-2385-3>
- Kumarasamy, Y., Nahar, L., & Sarker, S. D. (2003). Bioactivity of gentiopicroside from the aerial parts of Centaurium erythraea. *Fitoterapia*, 74(1-2), 151-154. doi: 10.1016/S0367-326X(02)00319-2
- Kusuma, H. S., & Mahfud, M. (2017). Comparison of kinetic models of oil extraction from sandalwood by microwave-assisted hydrodistillation. *International Food Research Journal*, 24(4), 1697–1702.
- Lee, B., Jung, K., & Kim, D. H. (2009). Timosaponin AIII, a saponin isolated from Anemarrhena asphodeloides, ameliorates learning and memory deficits in mice. *Pharmacology Biochemistry and Behavior*, 93(2), 121-127. doi: 10.1016/j.pbb.2009.04.021

- Lee, K. J., Oh, Y. C., Cho, W. K., & Ma, J. Y. (2015). Antioxidant and anti-inflammatory activity determination of one hundred kinds of pure chemical compounds using offline and online screening HPLC assay. *Evidence-Based Complementary and Alternative Medicine*, 1-13. doi:10.1155/2015/165457
- Li, D. C., & Jiang, J. G. (2010). Optimization of the microwave-assisted extraction conditions of tea polyphenols from green tea. *International Journal of Food Sciences and Nutrition*, 61(8), 837-845. doi:10.3109/09637486.2010.489508
- Li, H. B., Jiang, Y., Wong, C. C., Cheng, K. W., & Chen, F. (2007). Evaluation of two methods for the extraction of antioxidants from medicinal plants. *Analytical and Bioanalytical Chemistry*, 388(2), 483-488. doi: 10.1007/s00216-007-1235-x
- Li, J., Zu, Y. G., Fu, Y. J., Yang, Y. C., Li, S. M., Li, Z. N., & Wink, M. (2010). Optimization of microwave-assisted extraction of triterpene saponins from defatted residue of yellow horn (*Xanthoceras sorbifolia* Bunge.) kernel and evaluation of its antioxidant activity. *Innovative Food Science & Emerging Technologies*, 11(4), 637-643. doi: 10.1016/j.ifset.2010.06.004
- Liao, Z. G., Wang, G. F., Liang, X. L., Zhao, G. W., & Jiang, Q. Y. (2008). Optimization of microwave-assisted extraction of active components from Yuanhu Zhitong prescription. *Separation and Purification Technology*, 63(2), 424-433. doi: 10.1016/j.seppur.2008.06.004
- Liazid, A., Guerrero, R. F., Cantos, E., Palma, M., & Barroso, C. G. (2011). Microwave assisted extraction of anthocyanins from grape skins. *Food Chemistry*, 124(3), 1238-1243. doi: 10.1016/j.foodchem.2010.07.053
- Lidia, D.-A., Delia, M.-H., Nancy, O.-T., & Rosa Isela, G.-G. (2017). Microwave-assisted extraction of phytochemicals and other bioactive compounds. *Reference Module in Food Science*. doi:10.1016/b978-0-08-100596-5.21437-6
- Liu, J. L., Li, L. Y., & He, G. H. (2016). Optimization of microwave-assisted extraction conditions for five major bioactive compounds from *Flos Sophorae immaturus* (cultivars of *Sophora japonica* L.) using response surface methodology. *Molecules*, 21(3), 296. doi: 10.3390/molecules21030296
- Luque de Castro, M. D., & Priego-Capote, F. (2010). Soxhlet extraction: Past and present panacea. *Journal of Chromatography A*, 1217(16), 2383–2389. <https://doi.org/10.1016/j.chroma.2009.11.027>
- Maan, A. A., Nazir, A., Khan, M. K. I., Ahmad, T., Zia, R., Murid, M., & Abrar, M. (2018). The therapeutic properties and applications of aloe vera: a review. *Journal of Herbal Medicine*, 12, 1-10. doi: 10.1016/j.hermed.2018.01.002

- Maeng, J. H., Muhammad Shahbaz, H., Ameer, K., Jo, Y., & Kwon, J. H. (2017). Optimization of microwave-assisted extraction of bioactive compounds from coriolus versicolor mushroom using response surface methodology. *Journal of Food Process Engineering*, 40(2), e12421. Doi: 10.1111/jfpe.12421
- Maenthaisong, R., Chaiyakunapruk, N., Niruntraporn, S., & Kongkaew, C. (2007). The efficacy of aloe vera used for burn wound healing: a systematic review. *Burns*, 33(6), 713-718. Doi: 10.1016/j.burns.2006.10.384
- Marchant, R., & Banat, I. M. (2012). Biosurfactants: a sustainable replacement for chemical surfactants?. *Biotechnology letters*, 34(9), 1597-1605. Doi: 10.1007/s10529-012-0956-x
- Matthews, A., Haas, D. M., O'Mathúna, D. P., & Dowswell, T. (2015). Interventions for nausea and vomiting in early pregnancy. *Cochrane Database of Systematic Reviews*, 1-118. doi: 10.1002/14651858.CD007575.pub4
- Mnif, I., & Ghribi, D. (2016). Glycolipid biosurfactants: main properties and potential applications in agriculture and food industry. *Journal of the Science of Food and Agriculture*, 96(13), 4310-4320. doi: 10.1002/jsfa.7759
- Mondal, M. H., Malik, S., Roy, A., Saha, R., & Saha, B. (2015). Modernization of surfactant chemistry in the age of gemini and bio-surfactants: a review. *RSC Advances*, 5(112), 92707-92718. doi: 10.1039/C5RA18462B
- Monrroy, M., García, E., Ríos, K., & García, J. R. (2017). Extraction and physicochemical characterization of mucilage from Opuntia cochenillifera (L.) Miller. *Journal of Chemistry*, 2017. doi: 10.1155/2017/4301901
- Morton, J. F. (1990). Mucilaginous plants and their uses in medicine. *Journal of Ethnopharmacology*, 29(3), 245-266. doi: 10.1016/0378-8741(90)90036-S
- Moyo, M., Amoo, S. O., Ncube, B., Ndhlala, A. R., Finnie, J. F., & Van Staden, J. (2013). Phytochemical and antioxidant properties of unconventional leafy vegetables consumed in southern Africa. *South African Journal of Botany*, 84, 65-71. doi: 10.1016/j.sajb.2012.09.010
- Mukherjee, P. K., Nema, N. K., Maity, N., Mukherjee, K., & Harwansh, R. K. (2013). Phytochemical and therapeutic profile of Aloe vera. *Journal of Natural Remedies*, 14(1), 1-26.
- Mulligan, C.N. (2009). Recent advances in the environmental applications of biosurfactants. *Current Opinion in Colloid & Interface Science*, 14, 372–378. Doi:10.1016/j.cocis.2009.06.005

- Mulligan, C. N., Cooper, D. G., & NEUFELD, R. J. (1984). Selection of microbes producing biosurfactants in media without hydrocarbons. *Journal of Fermentation Technology*, 62(4), 311-314.
- Mulligan, C.N., 2005. Environmental applications for biosurfactants. *Environmental Pollution*, 133, 183–198. doi:10.1016/j.envpol.2004.06.009
- Nakilcioglu-Taş, E., & Otles, S. (2018). Degradation kinetics of bioactive compounds and antioxidant capacity of Brussels sprouts during microwave processing. *International Journal of Food Properties*, 20(3), S2798–S2809. <https://doi.org/10.1080/10942912.2017.1375944>
- Nandal, U., & Bhardwaj, R. L. (2012). Aloe vera: a valuable wonder plant for food, medicine and cosmetic use-a review. *International Journal of Pharmaceutical Sciences Review and Research*, 13(1), 59-67.
- Neelesh K. Nema, 2012. Hyaluronidase, elastase and MMP-1 inhibition activity of standardized juice of Aloe vera leaves. Tractates Sabbat 'Eruvin. *Jadavpur University*. doi:10.1515/9783110289039.734
- Nickel, J., Spanier, L. P., Botelho, F. T., Gularate, M. A., & Helbig, E. (2016). Effect of different types of processing on the total phenolic compound content, antioxidant capacity, and saponin content of Chenopodium quinoa Willd grains. *Food Chemistry*, 209, 139-143. doi: 10.1016/j.foodchem.2016.04.031
- Nielsen, C. K., Kjems, J., Mygind, T., Snabe, T., & Meyer, R. L. (2016). Effects of Tween 80 on growth and biofilm formation in laboratory media. *Frontiers in Microbiology*, 7, 1878. doi: 10.3389/fmicb.2016.01878
- Nitschke, M., & Costa, S. G. V. A. O. (2007). Biosurfactants in food industry. *Trends in Food Science & Technology*, 18(5), 252-259. doi: 10.1016/j.tifs.2007.01.002
- Oleszek, W., & Hamed, A. (2010). Saponin-based surfactants. *Surfactants from Renewable Resources*, 239-249. doi: 10.1002/9780470686607.ch12
- OM, M., X, L., Quiral, & Cardemil. (2015). Extraction, characterization and properties of the gel of Aloe vera (Aloe barbadensis Miller) cultivated in Chile. *Medicinal & Aromatic Plants*, 4(3), 1–7. <https://doi.org/10.4172/2167-0412.1000199>
- Pacheco, G. J., Ciapina, E. M. P., Gomes, E. D. B., & Pereira Junior, N. (2010). Biosurfactant production by Rhodococcus erythropolis and its application to oil removal. *Brazilian Journal of Microbiology*, 41(3), 685-693. doi: 10.1590/S1517-83822010000300019

Pacwa-Płociniczak, M., Plaza, G. A., Piotrowska-Seget, Z., & Cameotra, S. S. (2011). Environmental applications of biosurfactants: recent advances. *International Journal of Molecular Sciences*, 12(1), 633-654. doi: 10.3390/ijms12010633

Pagureva, N., Tcholakova, S., Golemanov, K., Denkov, N., Pelan, E., & Stoyanov, S. D. (2016). Surface properties of adsorption layers formed from triterpenoid and steroid saponins. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 491, 18–28. <https://doi.org/10.1016/j.colsurfa.2015.12.001>

Pająk, P., Socha, R., Broniek, J., Królikowska, K., & Fortuna, T. (2019). Antioxidant properties, phenolic and mineral composition of germinated chia, golden flax, evening primrose, phacelia and fenugreek. *Food chemistry*, 275, 69-76. doi:10.1016/j.foodchem.2018.09.081

Pan, X., Liu, H., Jia, G., & Shu, Y. Y. (2000). Microwave-assisted extraction of glycyrrhetic acid from licorice root. *Biochemical Engineering Journal*, 5(3), 173-177. doi:10.1016/S1369-703X(00)00057-7

Peak, D. (2005). Fourier transform infrared spectroscopy. *Encyclopedia of Soils in the Environment*, 4, 80-85. doi:10.1016/B0-12-348530-4/00174-0

Pem, D., & Jeewon, R. (2015). Fruit and vegetable intake: Benefits and progress of nutrition education interventions-narrative review article. *Iranian Journal of Public Health*, 44(10), 1309–1321.

Peter S. Piispanen, Marcus Persson, Per Claesson, and T. N. (2004). Surface properties of surfactants derived from natural products. Part 1: Syntheses and structure/property relationships—Solubility and emulsification. *Journal of Surfactants and Detergents*, 7(2), 147–159. <https://doi.org/10.1007/s11743-004-0298-6>

Poojary, M. M., Vishnumurthy, K. A., & Vasudeva Adhikari, A. (2015). Extraction, characterization and biological studies of phytochemicals from Mammea suriga. *Journal of Pharmaceutical Analysis*, 5(3), 182–189. <https://doi.org/10.1016/j.jpha.2015.01.002>

Prabhu, A., & Krishnamoorthy, M. (2010). Anticancer activity of Trigonella foenum-graecum on Ehrlich Ascites carcinoma in Mus musculus system. *Journal of Pharmacy Research*, 3(6), 1181-3.

Pradhan, A., & Bhattacharyya, A. (2017). Quest for an eco-friendly alternative surfactant: Surface and foam characteristics of natural surfactants. *Journal of Cleaner Production*, 150, 127-134. doi: 10.1016/j.jclepro.2017.03.013

Proestos, C., & Komaitis, M. (2008). Application of microwave-assisted extraction to the fast extraction of plant phenolic compounds. *LWT-Food Science and Technology*, 41(4), 652-659. doi: 10.1016/j.lwt.2007.04.013

Rababah, T. M., Hettiarachchy, N. S., & Horax, R. (2004). Total phenolics and antioxidant activities of fenugreek, green tea, black tea, grape seed, ginger, rosemary, gotu kola, and ginkgo extracts, vitamin E, and tert-butylhydroquinone. *Journal of Agricultural and Food Chemistry*, 52(16), 5183-5186. doi:10.1021/jf049645z

Rakholiya, K. D., Kaneria, M. J., & Chanda, S. V. (2013). Medicinal plants as alternative sources of therapeutics against multidrug-resistant pathogenic microorganisms based on their antimicrobial potential and synergistic properties. In *Fighting Multidrug resistance with Herbal Extracts, Essential Oils and their Components* (pp. 165-179). doi:10.1016/B978-0-12-398539-2.00011-2

Raphael, E. (2012). Phytochemical constituents of some leaves extract of Aloe vera and Azadirachta indica plant species. *Global Advanced Research Journal of Environmental Science and Toxicology*, 1(2), 014-017.

Ravishankar Rai, V., Bai, J.A. (2014). Microbial food safety and preservation techniques, CRC Press. doi:10.1201/b17465

Ray, A., Ghosh, S., Ray, A., & Aswatha, S. M. (2015). An analysis of the influence of growth periods on potential functional and biochemical properties and thermal analysis of freeze-dried Aloe vera L. gel. *Industrial Crops and Products*, 76, 298-305. doi: 10.1016/j.indcrop.2015.05.015

Rekiel, E., Smułek, W., Zdziennicka, A., Kaczorek, E., & Jańczuk, B. (2020). Wetting properties of Saponaria officinalis saponins. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 584, 123980. doi: 10.1016/j.colsurfa.2019.123980

Riasat, M., Heidari, B., Pakniyat, H., & Jafari, A. A. (2018). Assessment of variability in secondary metabolites and expected response to genotype selection in fenugreek (*Trigonella* spp.). *Industrial Crops and Products*, 123, 221-231. doi: 10.1016/j.indcrop.2018.06.068

Ross, J., & Miles, G. (1941). An apparatus for comparison of foaming properties of soaps and detergents. *Oil Soap*, 18, 99-102. doi:10.1007/BF02545418

Rufino, R. D., de Luna, J. M., de Campos Takaki, G. M., & Sarubbo, L. A. (2014). Characterization and properties of the biosurfactant produced by *Candida lipolytica* UCP 0988. *Electronic Journal of Biotechnology*, 17(1), 34-38. <https://doi.org/10.1016/j.ejbt.2013.12.006>

Sagratini, G., Zuo, Y., Caprioli, G., Cristalli, G., Giardina, D., Maggi, F., Molin, L., Ricciutelli, M., Traldi, P., Vittori, S. (2009). Quantification of soyasaponins I and β g in Italian lentil seeds by solid-phase extraction (SPE) and high-performance liquid chromatography– mass spectrometry (HPLC-MS). *Journal of Agricultural and Food Chemistry*, 57(23), 11226-11233. doi: 10.1021/jf901707z

Saikat K. Basu, P. Z. and W. C.-I. (2019). Fenugreek (*Trigonella foenum-graecum* L.): distribution, genetic diversity, and potential to serve as an industrial crop for the global pharmaceutical, nutraceutical, and functional food industries. In *The Role of Functional Food Security in Global Health*. Elsevier Inc. <https://doi.org/10.1016/B978-0-12-813148-0.00028-1>

Samal, K., Das, C., & Mohanty, K. (2017). Eco-friendly biosurfactant saponin for the solubilization of cationic and anionic dyes in aqueous system. *Dyes and Pigments*, 140, 100-108. doi:10.1016/j.dyepig.2017.01.031

Sasidharan, S., Chen, Y., Saravanan, D., Sundram, K. M., & Latha, L. Y. (2011). Extraction, isolation and characterization of bioactive compounds from plants' extracts. *African Journal of Traditional, Complementary and Alternative Medicines*, 8(1)1-10. doi: 10.4314/ajtcam.v8i1.60483

Semmar, N., Farman, M., & McCullagh, J. S. O. (2017). Measurement of total phenolic content and antioxidant activity of aerial parts of medicinal plant Coronopus didymus. *Asian Pacific Journal of Tropical Medicine*, 10(8), 792–801. <https://doi.org/10.1016/j.apjtm.2017.07.024>

Shah, A., Shahzad, S., Munir, A., Nadagouda, M. N., Khan, G. S., Shams, D. F., Dionysiou, D.D., Rana, U. A. (2016). Micelles as soil and water decontamination agents. *Chemical Reviews*, 116(10), 6042-6074. doi: 10.1021/acs.chemrev.6b00132

Shao, P., He, J., Sun, P., & Zhao, P. (2012). Analysis of conditions for microwave-assisted extraction of total water-soluble flavonoids from *Perilla Frutescens* leaves. *Journal of Food Science and Technology*, 49(1), 66-73. doi: 10.1007/s13197-011-0265-8

Sharp, O., Wong, K. Y., & Johnston, P. (2018). Segmental fracture of the scaphoid. *BMJ Case Reports*, Elsevier Inc. doi:10.1136/bcr-2017-223556

Singh, A.K. (2016). Experimental methodologies for the characterization of nanoparticles, Engineered Nanoparticles. doi:10.1016/b978-0-12-801406-6.00004-2

Sooi, L. K., & Keng, S. L. (2013). Herbal Medicines : Malaysian Women ' s Knowledge and Practice. *Evidence-based Complementary and Alternative Medicine*.

Sookjitsumran, W., Devahastin, S., Mujumdar, A. S., & Chiewchan, N. (2016). Comparative evaluation of microwave-assisted extraction and preheated solvent extraction of bioactive compounds from a plant material: a case study with cabbages. *International Journal of Food Science & Technology*, 51(11), 2440-2449. doi:10.1111/ijfs.13225

Sperandio, O., Fan, B. T., Zakrzewska, K., Jia, Z. J., Zheng, R. L., Panaye, A., El Fassi, N. (2002). Theoretical study of fast repair of DNA damage by cistanoside C and analogs: mechanism and docking. *SAR and QSAR in Environmental Research*, 13(2), 243-260. doi: 10.1080/10629360290002749

Sporring, S., Bøwadt, S., Svensmark, B., & Björklund, E. (2005). Comprehensive comparison of classic Soxhlet extraction with Soxtec extraction, ultrasonication extraction, supercritical fluid extraction, microwave assisted extraction and accelerated solvent extraction for the determination of polychlorinated biphenyls in soil. *Journal of Chromatography A*, 1090(1-2), 1-9. doi: 10.1016/j.arabjc.2011.06.022

Subramanian, R., Subbramaniyan, P., Noorul, J., & Raj, V. Double bypasses soxhlet apparatus for extraction of piperine from *Piper nigrum*, Arab J Chem. 2016; 9: S537-S540. doi: 10.1016/j.arabjc.2011.06.022

Snese, M., Bot, F., Panozzo, A., Mirolo, G., & Lippe, G. (2015). Effect of ultrasound treatment, oil addition and storage time on lycopene stability and in vitro bioaccessibility of tomato pulp. *Food chemistry*, 172, 685-691

Sultana, B., Anwar, F., Przybylski, R. (2007). Antioxidant activity of phenolic components present in barks of *Azadirachta indica*, *Terminalia arjuna*, *Acacia nilotica*, and *Eugenia jambolana* Lam. trees. *Food Chemistry*. 104, 1106–1114. doi:10.1016/j.foodchem.2007.01.019

Sultana N., Alsarhan, A., Al-Khatib, A., & Kadir, M. R. A. (2014). Review on some Malaysian traditional medicinal plants with therapeutic properties. *Journal of Basic and Applied Sciences*, 10, 149-159. doi: 10.6000/1927-5129.2014.10.20

Tan, S.N., Yong, J.W.H., Teo, C.C., Ge, L., Chan, Y.W., Hew, C.S. (2011). Determination of metabolites in *Uncaria sinensis* by HPLC and GC-MS after green solvent microwave-assisted extraction. *Talanta*, 83, 891-898. doi:10.1016/j.talanta.2010.10.048

Tavakoly, R., Maracy, M. R., Karimifar, M., & Entezari, M. H. (2018). Does fenugreek (*Trigonella foenum-graecum*) seed improve inflammation, and oxidative stress in patients with type 2 diabetes mellitus? A parallel group randomized clinical trial. *European Journal of Integrative Medicine*, 18, 13-17. doi: 10.1016/j.eujim.2018.01.005

Tmáková, L., Sekretár, S., & Schmidt, Š. (2016). Plant-derived surfactants as an alternative to synthetic surfactants: surface and antioxidant activities. *Chemical Papers*, 70(2), 188-196. doi: 10.1515/chempap-2015-0200

Van Middlesworth, F., & Cannell, R. J. (1998). Dereplication and partial identification of natural products. In *Natural products isolation* (pp. 279-327). Humana Press. doi: 10.1007/978-1-59259-256-2_10

Varjani, S. J., & Upasani, V. N. (2017). Critical review on biosurfactant analysis, purification and characterization using rhamnolipid as a model biosurfactant. *Bioresource Technology*, 232, 389-397. doi: 10.1016/j.biortech.2017.02.047

Veggi, P. C., Martinez, J., & Meireles, M. A. A. (2013). Microwave-assisted extraction for bioactive compounds: theory and practice. *Food Engineering, Series*, 15-52. doi:10.1007/978-1-4614-4830-3

Venegas-Calerón, M., Ruíz-Méndez, M. V., Martínez-Force, E., Garcés, R., & Salas, J. J. (2017). Characterization of Xanthoceras sorbifolium Bunge seeds: lipids, proteins and saponins content. *Industrial Crops and Products*, 109, 192-198. doi: 10.1016/j.indcrop.2017.08.022

Wang, G., Su, P., Zhang, F., Hou, X., Yang, Y., Guo, Z. (2011). Comparison of microwave-assisted extraction of aloe-emodin in aloe with Soxhlet extraction and ultrasound-assisted extraction. *Science China Chemistry*, 54, 231–236. doi:10.1007/s11426-010-4017-9

Wang, L., Qin, P., & Hu, Y. (2010). Study on the microwave-assisted extraction of polyphenols from tea. *Frontiers of Chemical Engineering in China*, 4(3), 307-313. doi: /10.1007/s11705-009-0282-6

Wang, P., Zhang, Q., Wang, Y., Wang, T., Li, X., Ding, L., & Jiang, G. (2010). Evaluation of Soxhlet extraction, accelerated solvent extraction and microwave-assisted extraction for the determination of polychlorinated biphenyls and polybrominated diphenyl ethers in soil and fish samples. *Analytica Chimica Acta*, 663(1), 43-48. doi: 10.1016/j.aca.2010.01.035

Wang, T.X., Zhang, Z.Q., Cong, Y., Shi, X.Y., Liu, Y.H., Zhao, F.L. (2013). Prosapogenin A induces apoptosis in human cancer cells in vitro via inhibition of the STAT3 signaling pathway and glycolysis. *Oncology Letters*, 6, 1323–1328. doi:10.3892/ol.2013.1561

Wani, S. A., Bishnoi, S., & Kumar, P. (2016). Ultrasound and microwave assisted extraction of diosgenin from fenugreek seed and fenugreek-supplemented cookies. *Journal of Food Measurement and Characterization*, 10(3), 527-532. doi: 10.1007/s11694-016-9331-2

Wei, S., Fukuhara, H., Chen, G., Kawada, C., Kurabayashi, A., Furihata, M., & Shuin, T. (2014). Terrestrosin D, a steroidal saponin from *Tribulus terrestris* L., inhibits growth and angiogenesis of human prostate cancer in vitro and in vivo. *Pathobiology*, 81(3), 123-132. doi: 10.1159/000357622

Worsfold, P.J., Zagatto, E.A.G. (2017). Spectrophotometry: Overview, 3rd ed, Reference Module in Chemistry, *Molecular Sciences and Chemical Engineering*. Elsevier Inc. doi:10.1016/b978-0-12-409547-2.14265-9

Xiao, X., Song, W., Wang, J., & Li, G. (2012). Microwave-assisted extraction performed in low temperature and in vacuo for the extraction of labile compounds in food samples. *Analytica chimica acta*, 712, 85-93. doi: 10.1016/j.aca.2011.11.034

Xiong, W., Chen, X., Lv, G., Hu, D., Zhao, J., & Li, S. (2016). Optimization of microwave-assisted extraction of bioactive alkaloids from lotus plumule using response surface methodology. *Journal of Pharmaceutical Analysis*, 6(6), 382-388. doi: 10.1016/j.jpha.2016.05.007

Xu, H. J., Shi, X. W., Ji, X., Du, Y. F., Zhu, H., & Zhang, L. T. (2012). A rapid method for simultaneous determination of triterpenoid saponins in *Pulsatilla turczaninovii* using microwave-assisted extraction and high performance liquid chromatography–tandem mass spectrometry. *Food chemistry*, 135(1), 251-258. doi: 10.1016/j.foodchem.2012.04.081

Yadav, U. C., & Baquer, N. Z. (2014). Pharmacological effects of *Trigonella foenum-graecum* L. in health and disease. *Pharmaceutical Biology*, 52(2), 243-254. doi:10.3109/13880209.2013.826247

Yang, Y. C., Li, J., Zu, Y. G., Fu, Y. J., Luo, M., Wu, N., & Liu, X. L. (2010). Optimisation of microwave-assisted enzymatic extraction of corilagin and geraniin from *Geranium sibiricum* Linne and evaluation of antioxidant activity. *Food Chemistry*, 122(1), 373-380. doi: /10.1016/j.foodchem.2010.02.061

Yang-Hua, Y., & Fu-Bao, D. (1991). A new triterpenoid and its glycoside from *Phytolacca esculenta*. *Planta Medica*, 57(02), 162-164.

Yu, H. B., Ding, L. F., Wang, Z., & Shi, L. X. (2014). Study on extraction of polyphenol from grape peel microwave-assisted activity. In *Advanced Materials Research*, 864, 520-525. Trans Tech Publications Ltd. doi: 10.4028/www.scientific.net/AMR.864-867.520

Zhang, F., Wang, X., Qiu, X., Wang, J., Fang, H., Wang, Z., & Xia, Z. (2014). The protective effect of Esculentoside A on experimental acute liver injury in mice. *PloS one*, 9(11). doi: 10.1371/journal.pone.0113107

Zhang, H. F., Yang, X. H., & Wang, Y. (2011). Microwave assisted extraction of secondary metabolites from plants: Current status and future directions. *Trends in Food Science & Technology*, 22(12), 672-688. doi: 10.1016/j.tifs.2011.07.003

Zhong, L., Zhang, Y., Chi, R., & Yu, J. (2016). Optimization of microwave-assisted ethanol reflux extraction process of flavonoids and saponins simultaneously from radix astragali using response surface methodology. *Food Science and Technology Research*, 22(6), 759-770. doi: 10.3136/fstr.22.759

Zhong, Z., Han, J., Zhang, J., Xiao, Q., Hu, J., & Chen, L. (2018). Pharmacological activities, mechanisms of action, and safety of salidroside in the central nervous system. *Drug Design, Development and Therapy*, 12, 1479. doi: 10.2147/DDDT.S160776

Zielinski, A.F., Silva, M. V, Pontes, P.V.D.A., Iora, S.R.F., Maciel, G.M., Haminiuk, C.W.I., Granato, D. (2014). Original article ewvaluation of the bioactive compounds and the antioxidant capacity of grape pomace. *Int. J. Food Sci. Technol.* 50, 62–69. doi:10.1111/ijfs.12583