





Draft Genome Sequence of the Carboxydotrophic Alphaproteobacterium *Aminobacter carboxidus* Type Strain DSM 1086

Paolo Turrini, a Irene Artuso, a Marco Tescari, a Gabriele Andrea Lugli, b Emanuela Frangipani, c 📵 Marco Ventura, b 📵 Paolo Visca

Paolo Turrini and Irene Artuso contributed equally to this work, and the order of these authors was determined on the basis of seniority.

ABSTRACT Aminobacter carboxidus is a soil Gram-negative alphaproteobacterium belonging to the physiological group of carboxydobacteria which aerobically oxidize CO to CO_2 . Here, we report the draft genome sequence of the A. carboxidus DSM 1086 type strain and the identification of both form I and form II CO dehydrogenase systems in this strain.

minobacter carboxidus DSM 1086^T (basonym Carbophilus carboxidus), formerly known as Achromobacter carboxydus (1) or Alcaligenes carboxydus (2), is the type strain and the unique known member of the species (3, 4). It was isolated from soil near a stream in Moscow, Russia (1), and was assigned to the physiological group of carboxydobacteria due to its ability to grow aerobically on carbon monoxide (CO) as the sole carbon and energy source (1, 5). Carbon monoxide dehydrogenase (CODH) activity, which is responsible for the oxidation of CO to carbon dioxide (CO₂), was formerly detected in A. carboxidus DSM 1086^T (6). Two CODH forms are known; form I specifically oxidizes CO, whereas form II is a putative CODH with a lower affinity for CO and still uncertain function (7, 8). Carbon dioxide produced by CO oxidation can be assimilated through the Calvin-Benson-Bassham cycle, although A. carboxidus is a facultative chemolithotroph able to utilize a wide variety of organic substrates for heterotrophic growth (5). Here, the genome sequence of A. carboxidus DSM 1086^T is reported with the aim of providing helpful insights into the genetic basis of CO oxidation in this monotypic strain.

A. carboxidus DSM 1086^T was obtained from DSMZ and aerobically grown at 30°C in Trypticase soy broth. DNA extraction was performed using a QIAamp DNA minikit (Qiagen). A genomic library of A. carboxidus was obtained with the TruSeq DNA PCR-free sample preparation kit (Illumina, Inc., San Diego, CA, USA). Genome sequencing was performed with a NextSeq 500 sequencing system (Illumina, UK) according to the manufacturer's protocol, and library samples were loaded into a midoutput kit v2.5 (300 cycles) (Illumina, UK), producing 1,416,277 pairs of reads. Raw sequence reads were filtered and trimmed using the command-line fastq-mcf software (https://expressionanalysis.github.io/ea-utils/). Fastq files of Illumina paired-end reads (150 bp) were used as input in the MEGAnnotator pipeline for microbial genome assembly and annotation (9). This pipeline employed the SPAdes program v3.14.0 for de novo assembly of the genome sequence with the option "--careful" and a list of k-mer sizes of 21, 33, 55, 77, 99, and 127 (10). The genome quality was evaluated with the program CheckM (11), estimating a genome completeness of 99.3%. The contigs were then submitted to the National Center for Biotechnology

Citation Turrini P, Artuso I, Tescari M, Lugli GA, Frangipani E, Ventura M, Visca P. 2020. Draft genome sequence of the carboxydotrophic alphaproteobacterium *Aminobacter carboxidus* type strain DSM 1086. Microbiol Resour Announc 9:e01170-20. https://doi.org/10.1128/MRA.01170-20.

Editor Kenneth M. Stedman, Portland State University

Copyright © 2020 Turrini et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Paolo Visca, paolo.visca@uniroma3.it.

Received 8 October 2020 Accepted 22 October 2020 Published 5 November 2020

^aDepartment of Science, Roma Tre University, Rome, Italy

Laboratory of Probiogenomics, Department of Chemistry, Life Sciences, and Environmental Sustainability, University of Parma, Parma, Italy

^cDepartment of Biomolecular Sciences, University of Urbino Carlo Bo, Urbino, Italy

	Locus tag	Product annotation	KEGG	Predicted	Closest ortholog (organism)
			orthology	gene name	
Form I	IHE39_24790	XdhC family protein	K07402	coxI	WP_095779620.1 (Mesorhizobium sp. WSM3868)
	IHE39_24795	XdhC family protein	K07402	coxH	WP_027028142.1 (Mesorhizobium sp. URHA0056)
	IHE39_24800	Carbon monoxide dehydrogenase subunit G	K09386	coxG	WP_148914117.1 (Mesorhizobium sp. MaA-C15)
	IHE39_24805	XdhC family protein	K07402	coxF	WP_148914116.1 (Mesorhizobium sp. MaA-C15)
	IHE39_24810	VWA domain-containing protein	K07161	coxE	WP_148914115.1 (Mesorhizobium sp. MaA-C15)
	IHE39_24815	MoxR family AAA+ ATPase	-	coxD	WP_148914114.1 (Mesorhizobium sp. MaA-C15)
	IHE39_24820	Carbon-monoxide dehydrogenase large subunit	K03520	coxL	WP_148914113.1 (Mesorhizobium sp. MaA-C15)
	IHE39_24825	(2Fe-2S)-binding protein	K03518	coxS	WP_148914112.1 (Mesorhizobium sp. MaA-C15)
	IHE39_24830	Xanthine dehydrogenase family protein subunit M	K03519	coxM	WP_148914111.1 (Mesorhizobium sp. MaA-C15)
Form II	IHE39_20405	Xanthine dehydrogenase family protein subunit M	K03519	coxM	WP_184767396.1 (Aminobacter lissarensis)
	IHE39_20410	Xanthine dehydrogenase family protein molybdopterin-binding subunit	K03520	coxL	WP_184767395.1 (Aminobacter lissarensis)
	IHE39_20415	(2Fe-2S)-binding protein	K03518	coxS	WP_184767394.1 (Aminobacter lissarensis)

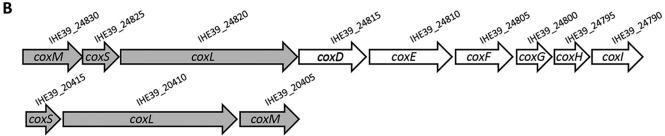


FIG 1 Forms I and II of the cox gene clusters in A. carboxidus DSM 1086^{T} . (A) The putative cox genes encoding form I and form II of the heterotrimeric $(\alpha\beta\chi)_2$ CODH enzyme complex (CoxL, CoxM, and CoxS subunits) of A. carboxidus DSM 1086^{T} with their GenBank annotation, gene name, and closest ortholog. KEGG Orthology numbers were assigned with the KEGG Automatic Annotation Server (KAAS) (13). (B) Physical map of the A. carboxidus DSM 1086^{T} genomic regions encompassing the form I (contig 11) and form II (contig 7) cox gene clusters. Form I is characterized by three structural genes, in the order coxMSL (gray), followed by six accessory genes (white), and by the presence of the AYXCSFR motif in the predicted CoxL active site. A different order of the three structural genes, coxSLM (gray), is characteristic of form II, and the CoxL active site contains the typical AYRGAGR motif.

Information (NCBI) for the prediction of protein-encoding open reading frames (ORFs) and tRNA and rRNA genes using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) (12). All tools were run with default parameters unless otherwise specified.

The draft genome of *A. carboxidus* is 6,291,275 bp long. It was assembled into 31 contigs with an N_{50} value of 458,931 bp, an average coverage of 65×, and a mean GC content of 62.96%. Genome annotation identified 6,023 ORFs, 49 tRNA genes, and 3 rRNA genes. Two gene clusters predicted to encode both forms of heterotrimeric CODH were identified (Fig. 1). Form I showed the specific AYXCSFR signature in CoxL and six accessory genes (coxDEFGHI) flanking the coxMSL structural genes, whereas form II showed the typical coxSLM structural gene arrangement and the specific AYRGAGR signature in CoxL.

Data availability. This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under accession number JACZEP000000000. The version described in this paper is JACZEP000000000.1. The raw sequencing reads are available at the Sequence Read Archive under accession number SRR12759717 and are associated with BioProject number PRJNA666410.

ACKNOWLEDGMENTS

This work was supported by the Excellence Departments grant (art. 1, commi 314-337 Legge 232/2016) to the Department of Science, Roma Tre University, and grant PRIN 2017 (Prot. 20177J5Y3P) to P.V., both from the Italian Ministry of Education, University and Research (MIUR).

Downloaded from https://journals.asm.org/journal/mra on 02 May 2022 by 90.147.2.114.

REFERENCES

- 1. Zavarzin GA, Nozhevnikova AN. 1977. Aerobic carboxydobacteria. Microb Ecol 3:305-326. https://doi.org/10.1007/BF02010738.
- 2. Cypionka H, Meyer O. 1983. The cytochrome composition of carboxydotrophic bacteria. Arch Microbiol 135:293-298. https://doi.org/10.1007/ BF00413484.
- 3. Meyer O, Stackebrandt E, Auling G. 1993. Reclassification of ubiquinone Q-10 containing carboxidotrophic bacteria: transfer of "[Pseudomonas] carboxydovorans" OM5T to Oligotropha, gen. nov., as Oligotropha carboxidovorans, comb. nov., transfer of "[Alcaligenes] carboxydus" DSM 1086T to Carbophilus, gen. nov., as Carbophilus carboxidus, comb. nov., transfer of "[Pseudomonas] compransoris" DSM 1231T to Zavarzinia, gen. nov., as Zavarzinia compransoris, comb. nov., and amended descriptions of the new genera. Syst Appl Microbiol 16:390-395. https://doi.org/10.1016/ 50723-2020(11)80271-7.
- 4. Oren A, Garrity GM. 2020. List of new names and new combinations previously effectively, but not validly, published. Int J Syst Evol Microbiol 70:4043-4049. https://doi.org/10.1099/ijsem.0.004244.
- 5. Meyer O, Schlegel HG. 1983. Biology of aerobic carbon monoxide oxidizing bacteria. Annu Rev Microbiol 37:277-310. https://doi.org/10.1146/annurev .mi.37.100183.001425.
- 6. Kraut M, Hugendieck I, Herwig S, Meyer O. 1989. Homology and distribution of CO dehydrogenase structural genes in carboxydotrophic bacteria. Arch Microbiol 152:335-341. https://doi.org/10.1007/
- 7. King GM. 2003. Molecular and culture-based analyses of aerobic carbon

- monoxide oxidizer diversity. Appl Environ Microbiol 69:7257-7265. https:// doi.org/10.1128/aem.69.12.7257-7265.2003.
- 8. King GM, Weber CF. 2007. Distribution, diversity and ecology of aerobic CO-oxidizing bacteria. Nat Rev Microbiol 5:107–118. https://doi.org/10 .1038/nrmicro1595.
- 9. Lugli GA, Milani C, Mancabelli L, van Sinderen D, Ventura M. 2016. MEGAnnotator: a user-friendly pipeline for microbial genomes assembly and annotation. FEMS Microbiol Lett 363:fnw049. https://doi.org/10.1093/femsle/
- 10. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455-477. https://doi.org/10.1089/cmb .2012.0021.
- 11. Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. Genome Res 25:1043-1055. https://doi.org/10.1101/gr.186072.114.
- 12. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614-6624. https://doi .org/10.1093/nar/gkw569.
- 13. Moriya Y, Itoh M, Okuda S, Yoshizawa A, Kanehisa M. 2007. KAAS: an automatic genome annotation and pathway reconstruction server. Nucleic Acids Res 35:W182-W185. https://doi.org/10.1093/nar/gkm321.