

# Cell Death & Disease



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## [ABOUT THE JOURNAL](#)

### Aims and Scope

*Cell Death & Disease* is a journal devoted to the biology of cell death, survival, stemness and differentiation in the pathogenesis of human diseases or relevant animal models. The journal aims to publish papers that present novel observations in the field of cell death, though with pathophysiological or medical implications.

Particular emphasis will be given to clinical, translational and applied research through its five sections:

- Experimental Medicine
- Cancer
- Immunity
- Internal Medicine
- Neuroscience

To this end, in conjunction with its sister journal *Cell & Differentiation*, *Cell Death & Disease* provides a unified forum for scientists as well as clinicians and members of the pharmaceutical and biotechnology industry. It is committed to the rapid publication of high quality original papers that relate to these subjects, together with topical, usually solicited, reviews, meeting reports, editorial correspondence and occasional commentaries on controversial and scientifically informative issues.

*Cell Death & Disease* is an open access online journal.

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- Abstract
- Introduction
- Results
- Discussion
- Materials (or Subjects) and Methods
- Acknowledgements (including all funding sources)
- Conflict of Interest
- References
- Figure legends
- Tables
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*Journal article, in press:*

Gallardo RL, Juneja HS, Gardner FH. Normal human marrow stromal cells induce clonal growth of human malignant T-lymphoblasts. *Int J Cell Cloning* (in press).

*Complete book:*

Atkinson K, Champlin R, Ritz J, Fibbe W, Ljungman P, Brenner MK (eds). *Clinical Bone Marrow and Blood Stem Cell Transplantation*, 3rd edn. Cambridge University Press: Cambridge, UK, 2004.

*Chapter in book:*

Coccia PF. Hematopoietic cell transplantation for osteopetrosis. In: Blume KG, Forman SJ, Appelbaum FR (eds). *Thomas' Hematopoietic Cell Transplantation*, 3rd edn. Blackwell Publishing Ltd: Malden, MA, USA, 2004, pp 1443–1454.

*Abstract:*

Syrjala KL, Abrams JR, Storer B, Heiman JR. Prospective risk factors for five-year sexuality late effects in men and women after haematopoietic cell transplantation. *Bone Marrow Transplant* 2006; 37(Suppl 1): S4 (abstract 107).

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- Processing (such as changing brightness and contrast) is appropriate only when it is applied equally across the entire image and is applied equally to controls. Contrast should not be adjusted so that data, or the background, disappear. Excessive manipulations, such as processing to emphasize one region in the image at the expense of others (for example, through the use of a biased choice of threshold settings), is unacceptable, as is emphasizing experimental data relative to the control.

For **gels and blots**, positive and negative controls, as well as molecular size markers, should be included on each gel and blot – either in the main figure or an expanded data supplementary figure. The display of cropped gels and blots in the main paper is discouraged. If cropping is necessary to improve the clarity or conciseness of the presentation, it should be done after the manuscript has been seen by reviewers, and in consultation with the editor. In such cases, the cropping must be mentioned in the figure legend.

- Vertically sliced gels that juxtapose lanes that were not contiguous in the experiment must have a clear separation or a black line delineating the boundary between the gels.
- Cropped gels must retain all bands. Irrelevant or cross reactive bands can be indicated as such by using an asterisk and describing them in the legend.
- High-contrast gels and blots are discouraged, as overexposure may mask additional bands. Authors should strive for exposures with grey backgrounds. Artificial (computer generated) backgrounds must not be added. Immunoblots should be surrounded by a black line to indicate the borders of the blot, if the background is faint. Such borders must appear at all places when the images were truncated or cropped
- For quantitative comparisons, appropriate reagents, controls and imaging methods with linear signal ranges should be used.
- The resolution must be a minimum of 300dpi for the original film, blot or capture, and it must be maintained during figure assembly. As a guideline, no squares or blocks indicative of jpeg compression should be visible when the final version is shown at 200% printed size.



**Microscopy** adjustments should be applied to the entire image. Threshold manipulation, expansion or contraction of signal ranges and the altering of high signals should be avoided. If 'pseudo-colouring' and nonlinear adjustment (for example 'gamma changes') are used, this must be disclosed. Adjustments of individual colour channels are sometimes necessary on 'merged' images, but this should be noted in the figure legend. We encourage inclusion of the following with the final revised version of the manuscript for publication:

- In the Methods section, specify the type of equipment (microscopes/objective lenses, cameras, detectors, filter model and batch number) and acquisition software used. Although we appreciate that there is some variation between instruments, equipment settings for critical measurements should also be listed.
- The display lookup table (LUT) and the quantitative map between the LUT and the bitmap should be provided, especially when rainbow pseudo-colour is used. It should be stated if the LUT is linear and covers the full range of the data.
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