Developing a Method for Rapid and Accurate Identification of *Bacillus* Species in Clinical Isolates Using Polymerase Chain Reaction

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Abstract

Bacillus species have caused a variety of diseases in humans throughout history. These include foodborne illness, wound infections, and anthrax poisoning. With over 266 identified species within the Bacillus genus, isolates previously described as independent species (e.g. B. anthracis, B. cereus, B. thuringiensis) have been discovered to be so genetically and phenotypically similar that they are often very difficult to discriminate between. Therefore, there have been instances of misidentification reported in scientific literature. Therefore, the majority of clinical tests done may not be precise enough to distinguish between some species of Bacillus, as they are so genetically similar. A new software, called MALDI-TOF, has been very successful in species discrimination. However, this technology is relatively new and not widely available for all clinical settings. The misidentification of clinical Bacillus isolates from infected patients could have medical significance and negatively affect treatment. By finding improved approaches for correctly identifying and distinguishing between Bacillus species, the scientific community can increase its understanding of the pathogenic potential of certain Bacillus species and positively impact patient diagnosis and treatment



of Bacillus thuringlensis HER1410 reveals a cry-containing dromosome, two megaplasmids & amp; a integrative plasmidial prophage. Published online May 8, 2020-2020.05.05.080028. doi:10.1101/2020.05.05.080028

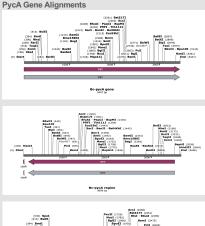
This study was intended to develop a technique for better distinguishing between *Bacillus* species. Recent research has suggested that the pycA gene may be the key to separating Bacillus species. A series of specially designed primers will be used to target the pycA gene in *Bacillus thuringiensis* and *Bacillus cereus* strains grown from isolates obtained from the NRRL repository. With the use of these primers, PCR testing will be performed on the various Bacillus strains. This will allow for identification of the isolate strains based on the PCR products and comparison to the initial repository identification. If successful, this approach could prove to be a more rapid and precise way to distinguish between *Bacillus* species in clinical settings, possibly improving the speed of diagnosis and treatment. Data collection and analysis are currently being executed.

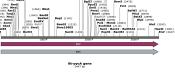
Introduction

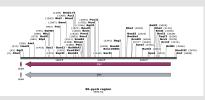
Designing the primers was a crucial part of the experimental design process. The gene of focus was the pycA gene. After reviewing the literature, there seemed to be the most variability between *Bacillus* species in the pycA gene region of the genomes. The pycA gene encodes for pyruvate carboxylase protein.

10 primers were carefully chosen using multiple sequence alignment software. The pycA gene sequences of *Bacillus cereus* and *Bacillus thuringiensis* were aligned, and primers were chosen to cover the most variable areas.

Each primer will be used to preform a PCR analysis followed by gel electrophoresis in order to compare the results from the *Bacillus cereus* and *Bacillus thuringiensis* strains from the NRRL repository.







Materials & Methods

Designed Primer Alignment

	T sA _A .I _ I
Consensus	NNNNANATCNANNGNNNNN
Primer_Bc-pycalignment_(45	CTAAAATCCCTTGCCAATTC- 20
Primer_Bc-pyclignment_(702	TCGAAATCTACTGGCTCT 18
Primer_Bc-pyclignment_(726	-AACAGTAAGACCAGGTGA 18
Primer_Bc-pycignment_(1011	TCTCCTAAACCAACCGCC 18
Primer_Bc-pycignment_(1250	GTGAGTTCTATGGCTGGT 18
Primer_Bc-pycignment_(2042	TTGGATTGGCTCTGAATGT 19
Primer_Bc-pycignment_(2043)	AAACATTCAGAGCCAATCCA 20
Primer_Bc-pycignment_(2773	TCACAGCAGCATCACAAA 18
Primer_Bc-pycignment_(2774	ATTTGTGATGCTGCTGTG 18
Primer_Bc-pycignment_(3575	AGGA <mark>A</mark> AG <mark>T</mark> AG <mark>A</mark> GG <mark>G</mark> GGAAA 19

Procedure

- I. Grow each of the 20 strains on nutrient agar plates for 12h
- . Transfer loopful of 1 cell colony to 0.1mL H2O in conical tube Boil each tube for 10min
- . Centrifuge (10sec @ 10,000rpm)
- Combine PCR mixture in tube
- 15 µL supernatant
- 0.5-2.5 U tag polymerase
- 0.1-0.5 µM primer
- 2.5 mM dNTPS
- Amplification - Single denaturation step (2min @ 95°C)
- 30-cvcle program
- Denaturation (1min @ 95°C)
- Annealing (1min @ 48°C)
- Extension (1min @ 72°C)
- Extension (5min @ 72°C)
- Electrophorese 15µL of each PCR mixture on 3% agarose in 0.5x trisborate buffer @250V for 30-35min
- Stain with ethidium bromide

NRRL Isolates

	NRRL Bacillus Isolate Strains			
	NRRL ID	Species	Source	
1	B-3711	Bacillus cereus	Type Strain	
2	B-14724	Bacillus cereus	Food Poisoning Incident	
3	B-1425	Bacillus cereus	Food Poisoning Incident	
4	B-1426	Bacillus cereus	Food Poisoning Incident	
5	B-1427	Bacillus cereus	Food Poisoning Outbreak	
6	NRS-1232	Bacillus cereus	Lesion on NRS-243 Injected Mouse	
7	NRS-1261	Bacillus cereus	Infection Post-Mastectomy	
8	NRS-1595	Bacillus cereus	Bovine Mastitis	
9	NRS-202	Bacillus cereus	Blood Culture	
10	NRS-599	Bacillus cereus	Spinal Fluid	
11	NRS-720	Bacillus cereus	Blood Culture	
12	NRS-785	Bacillus cereus	Puncture Wound	
13	HD-735	Bacillus thuringiensis	Type Strain	
14	HD-1	Bacillus thuringiensis	Diseased Pink Bollworm	
15	HD-119	Bacillus thuringiensis	Insect	
16	HD-121	Bacillus thuringiensis	Insect	
17	HD-15	Bacillus thuringiensis	Silkworm	
18	HD-17	Bacillus thuringiensis	Moth	
19	HD-203	Bacillus thuringiensis	Diseased Moth	
20	HD-27	Bacillus thuringiensis	Insect	

References

1. Baek I, Lee K, Goodfellow M, Chun J. Comparative Genomic and Phylogenomic Analyses Clarify Relationships Within and Between Bacillus cereus and Bacillus thuringiensis: Proposal for the Recognition of Two Bacillus thuringiensis Genomovars. Frontiers in Microbiology. 2019:10:1978. doi:10.3389/micb.2019.01978

 Cerón J, Ortiz A, Quintero R, Güereca L, Bravo A. Specific PCR primers directed to identify cryl and crylli genes within a Bacillus thuringiensis strain collection. Appl Environ Microbiol. 1995;61(11):3826-3831. doi:10.1128/aem.61.11.3826-3831.1995

3. Chen M I., Tsen H y. Discrimination of Bacillus cereus and Bacillus thuringiensis with 16S rRNA and gyrB gene based PCR primers and sequencing of their annealing sites. Journal of Applied Microbiology. 2002;92(5):912-919. doi:10.1046/j.1365-2672.2002.01606.x 4. Han CS, Xie G, Challacombe JF, et al. Pathogenomic Sequence Analysis of Bacillus cereus and Bacillus thuringiensis Isolates Closely

Related to Bacillus anthracis. Journal of Bacteriology. 2006;188(9):3382-3390. doi:10.1128/JB.188.9.3382-3390.2006

5. Ivanova N, Sorokin A, Anderson I, et al. Genome sequence of Bacillus cereus and comparative analysis with Bacillus anthracis. Nature. 2003;423(6935):87-91. doi:10.1038/nature01582

 Jackson S g., Goodbrand R b., Ahmed R, Kasatiya S. Bacillus cereus and Bacillus thuringiensis isolated in a gastroenteritis outbreak investigation. Letters in Applied Microbiology. 1995;21(2):103-105. doi:10.1111/j.1472-765X.1995.tb/1017.x

 Kolstø AB, Tourasse NJ, Økstad OA. What Sets Bacillus anthracis Apart from Other Bacillus Species? Annual Review of Microbiology. 2009;63(1):41-476. doi:10.1146/annurew.nicro.091208.073255
Lechuga A, Lood C, Salas M, van Noort V, Lavigne R, Redrejo-Rodriguez M. Compileted Genomic Sequence of Bacillus thuringiensis HER1410 Reveals a Cry-Containing Chromosome, Two Megaplasmids, and an Integrative Plasmidial Prophage. G3 Genes/Genomes/Genetics. 2020;10(9):2927-2939. doi:10.1534/g3.120.401361
Lechuga A, Lood C, Salas M, Noort VV van, Lavigne R, Redrejo-Rodriguez M. The fully resolved genome of Bacillus thuringiensis HER1410 reveals a cry-containing chromosome, two megaplasmids &: an integrative plasmidial prophage. Published online May 8, 2020:2020.05.05.080028. doi:10.1101/2020.05.05.080028
Liu Y, Lai Q, Göker M, et al. Genomic insights into the taxonomic status of the Bacillus cereau group. Sci Rev. 2015;5(1):14082.

doi:10.1038/srep14082 11. McIntyre L, Bernard K, Beniac D, Isaac-Renton JL, Naseby DC. Identification of Bacillus cereus Group Species Associated with Food Poisoning Outbreaks in British Columbia, Canada. Applied and Environmental Microbiology. 2008;74(23):7451-7453. doi:10.1128/AEM.01284-08

12. Patel, R. (2021, November 8). Re: Winona State Bacillus Research Project

 Raymond B, Federici BA. In defence of Bacillus thuringiensis, the safest and most successful microbial insecticide available to humanity—a response to EFSA. FEMS Microbiology Ecology. 2017;93(7). doi:10.1093/femsecfix084

14. Read, T. (2021, November 10). Re: Undergraduate Research Inquiry. 15. Read TD, Peterson SN, Tourasse N, et al. The genome sequence of Bacillus anthracis Ames and comparison to closely related bacteria. Nature. 2003;423(6935);81-86. doi:10.1038/nature01586 16. Shu LJ, Yang YL. Bacillus Classification Based on Matrix-Assisted Laser Desorption Ionization Time-of-Flight Mass Spectrometry—Effects of Culture Conditions. Sci Rep. 2017;7(1):15546. doi:10.1038/s41598-017-15808-5