# Oxidative stress indicators in populations of the gastropod *Buccinanops globulosus* affected by imposex

M. A. PRIMOST<sup>1</sup>, S. E. SABATINI<sup>2,3</sup>, P. DI SALVATORE<sup>4</sup>, M. C. RÍOS DE MOLINA<sup>2</sup> AND G. BIGATTI<sup>1,5</sup>

¹Instituto de Biología de Organismos Marinos IBIOMAR – CONICET, Bvd. Brown 2915, U9120ACV Puerto Madryn, Chubut, Argentina, ²IQUIBICEN-Dpto Química Biológica, Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires, Pab. II, Intendente Guiraldes 2160, C1428EHA, Ciudad Autónoma de Buenos Aires, Argentina, ³Departamento de Biodiversidad y Biología Experimental, Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires, Pab. II, Intendente Guiraldes 2160, C1428EHA. Ciudad Autónoma de Buenos Aires, Argentina, ⁴Centro Austral de Investigaciones Científicas (CADIC)-CONICET, Houssay 200, V9410CAB. Ushuaia, Argentina, ⁵Facultad de Ciencias Naturales, Universidad Nacional de la Patagonia San Juan Bosco (UNPSJB), Bvd. Brown S/N; U9120ACV Puerto Madryn, Chubut, Argentina

The gastropod Buccinanops globulosus is commonly used as a bioindicator of tributyltin (TBT) contamination due to its high imposex incidence in maritime traffic areas. The aim of this study was to evaluate both oxidative stress in B. globulosus at three sites with different maritime activity, and imposex incidence in Nuevo Gulf, Argentina. Oxidative stress parameters in digestive glands, like superoxide dismutase (SOD) and glutathione-S-transferase (GST) activities, reduced glutathione levels (GSH), and oxidative damage to lipids, estimated as thiobarbituric acid reactive substances (TBARs) as well as imposex parameters (% imposex and female penis length (FPL)) were measured in females. Gastropods from the harbour area showed 100% imposex, the highest FPL and TBARs content, as well as GSH levels and SOD activity.

The different oxidative stress responses and high imposex incidence at the harbour site may indicate a negative effect on the organism's physiological state due to environmental pollution.

Keywords: Oxidative stress, imposex, marine pollution, Buccinanops globulosus, Nassariidae

Submitted 18 June 2015; accepted 23 November 2015; first published online 22 December 2015

## INTRODUCTION

Pollutants such as tributyltin (TBT), polyaromatic hydrocarbons (PAHs), organochlorinated compounds and trace metals are present in areas with intense maritime activity in Patagonian coasts (Gil et al., 1999, 2006; Commendatore et al., 2000; Esteves et al., 2006; Commendatore & Esteves, 2007; Massara Paletto et al., 2008; Bigatti et al., 2009). Aquatic invertebrates, and molluscs in particular, are widely used as bioindicators of polluted environments (Meador et al., 1995; Kim et al., 2002; Antizar-Ladislao, 2008), while biomarkers are powerful tools to detect environmental damage and risk status (Dahlhoff, 2004). Pollutants could affect living organisms by inducing reactive oxygen species (ROS) formation (Winston & Di Giulio, 1991; Cheung et al., 2001; Leonard et al., 2004; Nicholson & Lam, 2005). Oxidative stress is the result of the imbalance between the generation and neutralization of ROS by antioxidant mechanisms (Davies, 1995). Oxidative stress responses (e.g. antioxidant enzyme activities and/or oxidative damage to lipids) have been used as biomarkers in molluscs to test and quantify the toxic effects of pollutants in the aquatic environment (de Almeida et al., 2004; Belcheva et al., 2011; Sabatini et al.,

Corresponding author: S.E. Sabatini Email: sabatini@bg.fcen.uba.ar 2011a). The increased activity or de novo synthesis of antioxidant enzymes to mitigate oxidative damage has been considered as an adaptation of organisms to stress conditions (Young & Woodside, 2001). Among these enzymes are superoxide dismutase (SOD), glutathione peroxidase (GPx) and catalase (CAT) which protect ROS scavenging cells (Karakoc et al., 1997; Borković et al., 2005) and glutathione S-transferase (GST) as well, a phase II detoxifying enzyme, exhibiting a protective mechanism against oxidative stress (Prohaska, 1980; Sheehan & Power, 1999; Doyen et al., Moreover, aquatic organisms also present non-enzymatic antioxidant defences (e.g. vitamin E, reduced glutathione-GSH-, between others) contributing to minimize oxidative damage (Sayeed et al., 2003; Wang et al., 2008).

In molluscs, the digestive gland is the principal site for bio-accumulation and detoxification of pollutants and the main target of oxidative disruption (Malanga *et al.*, 2004). In several bivalve species exposed to pollutants, oxidative damage and an increased/decreased activity of antioxidant enzymes have been registered (Bainy *et al.*, 2000; Sabatini *et al.*, 2009, 2011a, b; Giarratano *et al.*, 2010, 2013; Di Salvatore *et al.*, 2013).

In marine gastropods from the Argentinean coast, the imposex phenomenon (penis or *vas deferens* neoformation) in females exposed to tributyltin (TBT) (Gibbs & Bryan, 1986) has been detected in all the harbour areas (Bigatti *et al.*, 2009). Many gastropod species have been affected by

imposex in Argentina, while the nassarid *Buccinanops globulosus* showed high sensibility to TBT (Bigatti *et al.*, 2009). It has been demonstrated that TBT could induce imposex, shell malformation (Chagot *et al.*, 1990; Alzieu, 2000; Bigatti & Carranza, 2007; Márquez *et al.*, 2011) and also causes oxidative stress (Huang *et al.*, 2005; Wang *et al.*, 2005; Jia *et al.*, 2009; Zhou *et al.*, 2010).

The gastropod Buccinanops globulosus inhabits sandy or muddy bottoms of shallow waters (Pastorino, 1993) in Patagonian coasts, and most of the time lives buried in the sediment (Scarabino, 1977). It is distributed along the South-western Atlantic Ocean (Pastorino, Buccinanops globulosus is dioecious, with internal fertilization. Females attach the egg capsules to their own shells (Penchaszadeh, 1971), and are larger than the males. In general, the populations from Patagonia have shown variability in biological parameters such as growth, shell shape and ageing (Narvarte et al., 2008; Avaca et al., 2013; Bökenhans, 2014; Primost et al., in press). This species is edible and is part of an expanding artisanal fishery (Narvarte et al., 2008; Averbuj et al., 2014). Sublethal effects and bioaccumulation of TBT and other pollutants (trace metals, hydrocarbons) have been detected in harbour areas (Bigatti et al., 2009; Torres et al., 2013; Primost, 2014). While signalling by retinoid X receptors (RXR) (Nishikawa et al., 2004) could be involved on the imposex development in gastropods, although the induction mechanisms are under study, the determination of oxidative stress responses in imposex-affected gastropods still remains inconclusive in Argentina.

The aim of this study was to evaluate oxidative stress responses associated to maritime traffic contamination in imposex-affected *B. globulosus* from Nuevo Gulf, Argentina.

#### MATERIALS AND METHODS

## Study area and imposex incidence

The study was performed in three sites of Nuevo Gulf, with decreasing maritime activity: harbour area at Luis Piedra Buena harbour (LPB) (42°43"57"S 65°1'53.9"W), Punta Cuevas beach (PC) (42°46′45″S 64°59′34″W) and Cerro Avanzado beach (CA) (42°49′37.66″S 64°51′29.19″W) (Figure 1). In the LPB site activity of large vessels is frequently present (~720 vessels per year) (APPM, 2013); in this area 100% imposex was reported in gastropods since 2000 (Bigatti & Penchaszadeh, 2005; Bigatti et al., 2009; del Brío, 2011; Primost, 2014), while moderate pollution by PAHS, trace metals and TBT were previously recorded in sediments and molluscs (Gil et al., 1999; Massara Paletto et al., 2008; Bigatti et al., 2009). The PC site is a recreational public area frequently presenting diving vessels, where low pollution by TBT and trace metals was measured (Primost, 2014) and lesser imposex parameters were reported (Bigatti et al., 2009; Primost, 2014). The CA beach is a recreational area where very low or null imposex incidence was reported as well as no detectable TBT pollution (Bigatti et al., 2009; del Brío, 2011; Primost, 2014); in this area there is low maritime traffic and sport vessels are present only occasionally. Table 1 summarizes the pollution levels previously detected in the sampling sites.

Adult female gastropods *Buccinanops globulosus* (25 approximately at each site) were collected using baited traps.

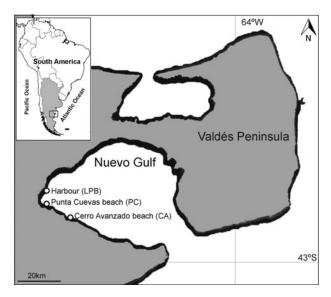


Fig. 1. Location of sampling sites in Nuevo Gulf, Patagonia, Argentina.

The sex was determined *in situ* by presence or absence of the ventral pedal gland (only present in females) used to fix egg capsules on its own shell. Total shell length (TSL) and body weight relative to size (BW) was recorded in the laboratory. Incidence of imposex (% I) was considered as the percentage of females with a penis or *vas deferens* development; correspondingly mean female penis length (FPL) was estimated only in females with penis development and using 0.1 mm precision digital caliper.

## Oxidative stress parameters

In a subsample of nine females per site, oxidative stress parameters were determined. The digestive gland was carefully dissected, weighed (with a digital scale 0.01 g) and frozen at  $-80^{\circ}$ C for later oxidative stress determinations.

Digestive glands were homogenized with 0.154 M KCl (1:5 w v<sup>-1</sup>) containing 0.5 mM phenylmethylsulphonyl fluoride (PMSF) and 0.2 mM benzamidine (protease inhibitors) to study oxidative stress parameters. The homogenates were centrifuged at 12,000  $\times$  g during 30 min (4°C) and the supernatants were stored for later determinations.

Total soluble protein content was measured by the method of Bradford (1976), using bovine serum albumin as standard. The results were expressed as  $\mu g$  of total protein per mL.

Superoxide dismutase (SOD, EC 1.15.1.1) activity was assessed by inhibition of photoreduction of NBT (nitro blue tetrazolium) and monitoring absorbance at 560 nm according to Beauchamp & Fridovich (1971). The standard assay mixture contained 5, 10 and 15  $\mu L$  enzymatic sample, 0.1 mM EDTA, 13 mM DL-methionine, 75  $\mu$ M NBT and 20  $\mu$ M riboflavin, in 50 mM phosphate buffer (pH 7.5), to a final volume of 3 mL. Samples were exposed for 15 min to intense cool-white light, and then kept in the dark until absorbance was measured at 560 nm. Results were expressed as U per mg protein. A SOD unit was defined as the enzyme amount necessary to inhibit the reaction rate by 50%.

Glutathione S-transferase (GST, EC1.11.1.9) activity was measured by monitoring the absorbance at 340 nm using 1-chloro-2 4-dinitrobenzene (CDNB) (100 mM) as substrate according to Habig *et al.* (1974). Briefly, we mixed 10 µL of

Table 1. Maximum values of different pollutants detected in gastropods (whole tissues) and sediments from sampling sites in Nuevo Gulf.

Pollutant	LPB harbour (LPBH)		Punta Cuevas beach (PC)		Cerro Avanzado beach (CA)		Reference	
	Gastropods	Sediments	Gastropods	Sediments	Gastropods	Sediments		
TBTs (ng (Sn)g <sup>-1</sup> dw)								
Tributyltin	171	175		1.9*	Nd	Nd	*Bigatti <i>et al.</i> (2009)	
Dibutyltin	74	19			Nd	Nd		
Monobutyltin	345	72			Nd	Nd	del Brío (2011);	
Booster biocides (ng g <sup>-1</sup> d								
Diuron	Nd	Nd			Nd	Nd		
Irgarol	Nd	Nd			Nd	Nd		
Trace metals (µg g <sup>-1</sup> dw)								
Al	5.5	12,958	16	8664	6	10,541		
Fe	126	13,581	89	12,175	89	10,492	Primost (2014)	
Zn	182	33	119	19	108.5	16.84		
Cu	13	6.1	7.5	3	9	2.99		
Cd	8	Nd	7	Nd	24	Nd		
Pb	1.2	7.5	0.4	Nd	0.4	Nd		
PAHs (ng g <sup>-1</sup> dw)								
Anthracene	174	30			Nd	Nd		
Fluoranthene	141	30			Nd	Nd		
Pyrene	28	20			Nd	Nd		
Benzo(b)fluoranthene	151	30			Nd	Nd	Torres et al. (2013)	
Benzo(k)fluoranthene	44	40			Nd	Nd		
Benzo(a)anthracene	22	50			Nd	Nd		
Chrysene	0	30			Nd	Nd		
Dibenzoanthracene	0	20			Nd	Nd		
Total PAHs		2500		180			Massara Paletto et al. (2008	

Nd: Non detectable.

glutathione (GSH) (100 mM in phosphate buffer) and 20  $\mu L$  of sample in 960  $\mu L$  of 100 mM phosphate buffer (pH 6.5) and 10  $\mu L$  CDNB. One GST Unit was defined as the amount of enzyme needed to catalyse the formation of 1  $\mu$ moL of GS-DNB per minute at 25°C.

Reduced GSH levels were determined monitoring the absorbance at 412 nm after 30 min incubation at room temperature following the Anderson (1985) procedure. Briefly, 100  $\mu L$  supernatant from the 11,000  $\times$  g sample was acidified with 50  $\mu L$  of 10% sulphosalicylic acid. After centrifugation at 8000  $\times$  for 10 min, supernatant (acid-soluble GSH) aliquots were mixed with 6 mM 5,5-dithiobis-(2-nitrobenzoic) acid (DTNB) in 0.143 M buffer sodium sulphate (pH 7.5) (containing 6.3 mM EDTA). Results were expressed as nmol GSH per mg of protein.

Lipid peroxidation was determined measuring thiobarbituric acid reactive substances (TBARs) according to Vavilin *et al.* (1998). Briefly, the 11,000  $\times$  g supernatant (175  $\mu$ L) from total homogenate was mixed with thiobarbituric acid (TBA) (26 mM) solution and incubated at 95–100°C for 45 min. After cooling, the reaction mixture was centrifuged

and the supernatant absorbance was determined at 535 nm. TBARs concentration was estimated using an extinction coefficient of 156 mM $^{-1}$  cm $^{-1}$  and absorbance determination at 535 nm. Results were expressed as  $\mu mol$  TBARs per mg of protein.

## Statistical analysis

Normality and homogeneity of variances were tested by Lilliefors' and Bartlett's tests, respectively (Sokal & Rohlf, 1979). Results from size, weight and oxidative stress parameters were analysed by one way ANOVA followed by a Tukey's post hoc test. Results for imposex analysis were compared between sites by Kruskal–Wallis followed by a Dunn post hoc test. Differences were considered significant with P < 0.05. Statistica7 software was used for statistical analysis. A DistLM multiple correlations was performed using PRIMER software (Clarke & Gorley, 2006) to compare the effect of stress parameters (as co-variable) on penis length (as response variable). Prior to analysis, variables were

**Table 2.** Total shell length, body weight (means + SD) and imposex parameters in *Buccinanops globulosus*.

SITE	Females (n)	Body weight/shell length	Total shell length (mm)	% Imposex	FPL (mm)
LPB harbour (LPBH) Punta Cuevas beach (PC) Cerro Avanzado beach (CA)	25 18 23	$0.34 \pm 0.01^{*}$ $0.20 \pm 0.01$ $0.23 \pm 0.01$	$40.72 \pm 0.69^*$ $32.85 \pm 0.78$ $34.63 \pm 0.95$	100 94.44 4.34	$4.51 \pm 0.23^{*}$ $0.83 \pm 0.11$

FPL: female penis length.

<sup>\*</sup>Significant differences between sites (LPB, PC and CA).

transformed by Z-score using R software (https://www.r-project.org/).

#### RESULTS

# Imposex incidence

A total of 66 females of *Buccinanops globulosus* were analysed for imposex incidence and a subsample of 27 females (nine per site) was used for the determination of oxidative stress parameters. Total shell length (TSL) and body weight (BW) were significantly different between sites (TSL: F = 27.306, P < 0.0001, df = 2, N = 66; BW: F = 46.006, P < 0.0001, df = 2, N = 66). In both cases, the highest values were obtained in the LPB site (Table 2).

The imposex incidence was 100% in LPB (Table 2) and significant differences in female penis length (FPL) between LPB vs. PC sites were observed (U = 450.000, P < 0.0001, N = 43). In CA site, the FPL was not calculated because only one female showed imposex development (with a small incipient penis).

## Antioxidant defences

In order to analyse the antioxidant defences, results firstly showed that total protein content in the digestive gland did not differ between sampling sites (F = 0.220, P = 0.804, df = 2, N = 27) (data not shown). Therefore, all measured variables were standardized as a function of protein content.

Gastropods collected from the harbour area (LPB site) showed higher superoxide dismutase (SOD) activity than those from the other two sites (PC and CA) (F = 13.277, P = 0.0001, df = 2, N = 27) (Figure 2A). Also the reduced glutathione content (GSH) revealed a similar pattern, showing the highest values in the LPB site (F = 8.148, P = 0.002, df = 2, N = 27) (Figure 2C).

On the other hand, glutathione-S-transferase (GST) activity in digestive gland did not show significant differences between sampling sites (F = 2.342, P = 0.118, df = 2, N = 27) (Figure 2B).

# Oxidative damage

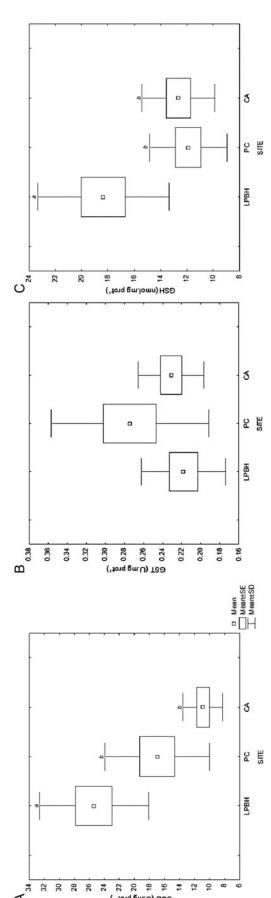
In relation to oxidative damage, significant differences in lipid peroxidation among sites were obtained (TBARs: F = 6.357, P = 0.006, df = 2, N = 27); individuals collected in the LPB site showed the highest values (Figure 3).

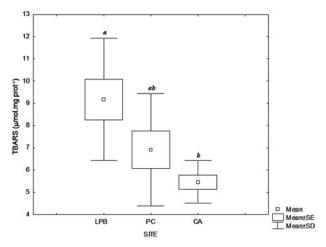
# Stress parameters and imposex response

Significant differences were obtained in DistLM for SOD, GSH and TBAR variables on penis length (as response variable). These results showed that 61.5% of variability in penis length was explained by stress parameters (Table 3).

# DISCUSSION

The imposex incidence and female penis length (FPL) recorded in this work for *Buccinanops globulosus* could be related to maritime traffic and levels of TBT reported previously in Nuevo Gulf (Bigatti *et al.*, 2009; del Brío, 2011). Pollutants could be bioaccumulating in aquatic organisms





**Fig. 3.** Lipid peroxidation, expressed as  $\mu$ mol TBARS mg<sup>-1</sup> prot, in digestive gland of *Buccinanops globulosus*. Results are expressed as mean  $\pm$  SD (N = 9). Letters a and b indicate significant differences between sampling sites (LPB, PC and CA)

and affecting their defence mechanisms (Regoli & Principato, 1995; Chandran *et al.*, 2005; Chen *et al.*, 2011). In the LPB area, del Brío and colleagues detected butyltin levels (TBT + dibutyltin-DBT- + monobutyltin-MBT-) up to 265.8 ng (Sn) g<sup>-1</sup> dry weight (dw) in sediments and up to 567.8 ng (Sn) g<sup>-1</sup> (dw) in the tissues of the marine gastropod *Odontocymbiola magellanica*, the gonads and digestive gland being the organs with the highest TBTs concentration (del Brio *et al.*, in press). Also polyaromatic hydrocarbons such anthracene, benzo(b)fluoranthene (Torres *et al.*, 2013) and trace metals such as copper (Cu), iron (Fe), lead (Pb) and zinc (Zn) were detected in *B. globulosus* (Primost, 2014) in the LPB area confirming the capacity of these gastropod species to bioaccumulate different pollutants and potentially form reactive oxygen species (ROS).

ROS formation and changes in the oxidative balance have been observed as a result of exposure to environmental levels of TBT in bivalves (Huang et al., 2005; An et al., 2009) and gastropods (Jia et al., 2009; Gopalakrishnan et al., 2011). Imposex in B. globulosus was associated with TBT presence (Bigatti et al., 2009) in the LPB area. In this work, the oxidative stress responses registered in imposexed gastropods could be attributed to TBT and other contaminants detected at the LPB harbour area such as trace metals and PAHs (Gil et al., 1988, 1999, 2006; Commendatore et al., 2000; Di Salvatore et al., 2013; Torres et al., 2013; Primost, 2014).

It is well known that a wide range of pollutants enhance enzymatic and non-enzymatic antioxidants in marine

**Table 3.** Results from DistLM multiple correlations between stress parameters (co-variable) and penis length (response variable).

	R <sup>2</sup>	F	P	df
U SOD mg <sup>-1</sup> prot	0.404	16.921	0.0004*	25
Mmol TBARS mg <sup>-1</sup> prot	0.586	10.539	0.0042*	24
U GST mg <sup>-1</sup> prot	0.615	0.176	0.200	23
Nmol GSH mg <sup>-1</sup> prot	0.615	0.015	0.907	22

Best solution:  $R^2 = 0.6152$ ; N° Vars = 4; Selections = All. \* indicates significant differences at P < 0.05 between sites (LPB, PC, CA) for these variables.

invertebrates to protect cells against oxidative damage (Livingstone, 2001; Brown et al., 2004; Valavanidis et al., 2006). Our results show highest SOD activities and highest concentrations of reduced glutathione in the digestive gland of gastropods from the LPB site. However, the glutathione-S-transferase (GST) activity showed a different pattern compared with these former antioxidant responses, where no significant differences in its activity were observed among the three sampling sites. Glutathione-S-transferase is a biotransformation enzyme which catalyses the conjugation of electrophilic pollutants with reduced glutathione (GSH). The resulting conjugates increase their water solubility favouring the excretion processes (Armstrong, 1997; Hayes et al., 2005). In molluscs, the activity of GST usually increases in relation to detoxification processes (Almeida et al., 2005; Huang et al., 2005). However in 2005, Huang and colleagues determined that GST activity in the fish Meretrix meretrix may be increased or inhibited depending on high or low TBT concentrations in water, respectively (Huang et al., 2005). Our results shows that pollution present in the PC site would induce a low increase in GST activity in the digestive gland of B. globulosus, while the pollutant presence in CA environment was not enough to produce changes in GST activity. In the present work, antioxidants (SOD and GSH) increased in the proximity of the harbour area, which was in concordance with higher imposex levels and penis length. Former studies, in the same sampling area, related to oxidative stress responses in the bivalve Aulacomya atra have detected seasonal changes in the antioxidant defences in relation to trace metal exposure and environmental pollution (Di Salvatore et al., 2013; Giarratano et al., 2013). In both studies, animals from the harbour area were the most affected, showing an increase in the antioxidant defences and also suffering higher oxidative damage to lipids. Meanwhile, a study in the fish Sebastiscus marmoratus exposed to TBT also revealed an increase in SOD activity in the liver (Wang et al., 2005). In this sense, SOD increasing in B. globulosus probably could be related to TBT contamination detected recently in the area (Bigatti et al., 2009; del Brío, 2011).

Lipid peroxidation has also been reported as a principal cause of cellular damage induced by oxidative stress conditions (Valavanidis et al., 2006). Membrane alterations in molluscs are the major target of cellular damage in organisms exposed to trace metals and other toxic substances (Viarengo et al., 1990, 1991). In the present work, B. globulosus showed a marked increment in lipid peroxidation in the digestive gland of gastropods collected from the harbour area compared with animals from CA and PC sites. In addition, our results are in accordance with those reported by Zhou et al. (2010), where TBT exposures increase lipid peroxidation (measured as malondialdehyde (MDA) levels) in the abalone Haliotis diversicolor supertexta. Similar results were also observed in laboratory studies where rats exposed to repeated TBT doses showed incremental MDA levels (Liu et al., 2006); Bernat and colleagues also reported the same effect in the filamentous fungus Cunninghamella elegans exposed to TBT (Bernat et al., 2014).

Our results suggest that the differences in terms of oxidative stress responses and high imposex incidence observed in *B. globulosus* at the harbour site indicate a negative effect on its physiological state due to the presence of pollutants in the aquatic environment. The possible relationship between the induction mechanism of imposex and oxidative stress

should be tested in controlled experiments exposing normal and imposexed individuals to TBT, followed by comparative measurement of oxidative stress parameters in experimental groups.

#### CONCLUSIONS

In conclusion, both oxidative stress responses and imposex incidence were increased in gastropods inhabiting the harbour area. While *B. globulosus* suffers an increase of the antioxidant defences (SOD activity and GSH content), an oxidative damage to lipids (TBARs levels) was still observed.

This is the first study on oxidative stress responses associated with marine pollution in an edible gastropod affected by imposex in Argentina. Although TBT is not the only pollutant present in the harbour area, further integrated studies are necessary to evaluate the role of oxidative stress responses in *Buccinanops globulosus* as biomarkers of TBT presence.

#### ACKNOWLEDGEMENTS

Special thanks to Dr E. Marzinelli, Dr L. Arribas, Dr M. Lozada and Dr Pitu Mendez.

#### FINANCIAL SUPPORT

This work was supported by grants: PICT 1232, PICT 2929, PICT 1491, PICT 1476 and PIP 067 (MP and GB), UBACYT20020100300055 and ANPCYTPICT-2010-0260 (SES), UBACYT20020100100985 (MCRM).

#### REFERENCES

- Almeida E.A., Bainy A.C.D., Dafre A.L., Gomes O.F., Medeiros M.H.G. and Di Mascio P. (2005) Oxidative stress in digestive gland and gill of the brown mussel (*Perna perna*) exposed to air and re-submersed. *Journal of Experimental Marine Biology and Ecology* 318, 21–30.
- Alzieu C. (2000) Environmental impact of TBT: the French experience. Science of the Total Environment 258, 99–102.
- An M.I., An K.W. and Choi C.Y. (2009) Changes in antioxidant enzyme activity and physiological responses to cadmium and ributyltin exposure in the ark shell, *Scapharca broughtonii*. *Molecular and Cellular Toxicology* 5, 273–282.
- **Anderson M.E.** (1985) Determination of glutathione and glutathione disulfide in biological samples. *Methods in Enzymology* 113, 548.
- Antizar-Ladislao B. (2008) Environmental levels, toxicity and human exposure to tributyltin (TBT)-contaminated marine environment. A review. Environment International 34, 292–308.
- APPM (2013) Administración Portuaria de Puerto Madryn. http://www.appm.com.ar/. vol. 2013.
- **Armstrong R.N.** (1997) Structure, catalytic mechanism, and evolution of the glutathione transferases. *Chemical Research in Toxicology* 10, 2-18
- **Avaca M.S., Narvarte M. and Martín P.** (2013) Age, growth and mortality in *Buccinanops globulosus* (Gastropoda: Nassariidae) from Golfo Nuevo (Argentina). *Marine Biology Research* 9, 208–219.

- Averbuj A., Rocha M.N. and Zabala S. (2014) Embryonic development and reproductive seasonality of *Buccinanops globulosus* (Nassariidae) (Kiener, 1834) in Patagonia, Argentina. *Invertebrate Reproduction and Development* 58, 138–147.
- Bainy A.C.D., Almeida E.A., Muller I.C., Ventura E.C. and Medeiros I.D. (2000) Biochemical responses in farmed mussel *Perna perna* transplanted to contaminated sites on Santa Catarina Island, SC, Brazil. *Marine Environmental Research* 50, 411–416.
- **Beauchamp C. and Fridovich I.** (1971) Superoxide dismutase: improved assays and an assay applicable to acrylamide gels. *Analytical Biochemistry* 44, 276–287.
- Belcheva N., Zakhartsev M., Dovzhenko N., Zhukovskaya A., Kavun V. and Chelomin V. (2011) Anthropogenic pollution stimulates oxidative stress in soft tissues of mussel *Crenomytilus grayanus* (Dunker 1853). Ocean Science Journal 46, 85-94.
- Bernat P., Gajewska E., Szewczyk R., Słaba M. and Długoński J. (2014)
  Tributyltin (TBT) induces oxidative stress and modifies lipid profile in
  the filamentous fungus *Cunninghamella elegans*. *Environmental*Science and Pollution Research 21, 4228-4235.
- **Bigatti G. and Carranza A.** (2007) Phenotypic variability associated with the occurrence of imposex in *Odontocymbiola magellanica* from Golfo Nuevo, Patagonia. *Journal of the Marine Biological Association of the United Kingdom* 87, 755–759.
- Bigatti G. and Penchaszadeh P.E. (2005) Imposex in Odontocymbiola magellanica (Caenogastropoda: Volutidae) in Patagonia.

  Comunicaciones de la Sociedad Malacológica del Uruguay 9, 371–375.
- Bigatti G., Primost M.A., Cledón M., Averbuj A., Theobald N., Gerwinski W., Arntz W., Morriconi E. and Penchaszadeh P.E. (2009) Biomonitoring of TBT contamination and imposex incidence along 4700 km of Argentinean shoreline (SW Atlantic: From 38S to 54S). Marine Pollution Bulletin 58, 695-701.
- Bökenhans V. (2014) Estimación de la edad y crecimiento de dos poblaciones de Buccinanops globulosus del Golfo Nuevo. Degree thesis. Universidad Nacional de la Patagonia San Juan Bosco, Puerto Madryn.
- Borković S.S., Šaponjić J.S., Pavlović S.Z., Blagojević D.P., Milošević S.M., Kovačević T.B., Radojičić R.M., Spasić M.B., Žikić R.V. and Saičić Z.S. (2005) The activity of antioxidant defence enzymes in the mussel Mytilus galloprovincialis from the Adriatic Sea. Comparative Biochemistry and Physiology Part C: Toxicology and Pharmacology 141, 366–374.
- **Bradford M.M.** (1976) A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry* 72, 248–254.
- Brown R.J., Galloway T.S., Lowe D., Browne M.A., Dissanayake A., Jones M.B. and Depledge M.H. (2004) Differential sensitivity of three marine invertebrates to copper assessed using multiple biomarkers. *Aquatic Toxicology* 66, 267–278.
- Clarke K.R. and Gorley R.N. (2006) Change in marine communities: an approach to statistical analysis and interpretation. PRIMER v6: User Manual/Tutorial. Plymouth: PRIMER-E, 192 pp.
- Chagot D., Alzieu C., Sanjuan J. and Grizel H. (1990) Sublethal and histopathological effects of trace levels of tributyltin fluoride on adult oysters Crassostrea gigas. Aquatic Living Resources 3, 121-130.
- Chandran R., Sivakumar A.A., Mohandass S. and Aruchami M. (2005)

  Effect of cadmium and zinc on antioxidant enzyme activity in the gastropod, Achatina fulica. Comparative Biochemistry and Physiology, Part C: Toxicology and Pharmacology 140, 422-426.
- Chen H.G., Jia X.P., Cai W.G., Lin Q. and Ma S.W. (2011) Antioxidant responses and bioaccumulation in green-lipped mussels (*Perna viridis*) under acute tributyltin chloride exposure. *Bulletin of Environmental Contamination and Toxicology* 87, 506–511.

- Cheung C.C.C., Zheng G.J., Li A.M.Y., Richardson B.J. and Lam P.K.S. (2001) Relationships between tissue concentrations of polycyclic aromatic hydrocarbons and antioxidative responses of marine mussels *Perna viridis. Aquatic Toxicology* 52, 189–203.
- Commendatore M.G. and Esteves J.L. (2007) An assessment of oil pollution in the coastal zone of Patagonia, Argentina. *Environmental Management* 40, 814–821.
- Commendatore M.G., Esteves J.L. and Colombo J.C. (2000) Hydrocarbons in coastal sediments of Patagonia, Argentina: levels and probable sources. *Marine Pollution Bulletin* 40, 989–998.
- Dahlhoff E.P. (2004) Biochemical indicators of stress and metabolism: applications for marine ecological studies. Annual Review of Physiology 66, 183–207.
- Davies K.J.A. (1995) Oxidative stress: the paradox of aerobic life. Biochemical Society Symposia 61, 1-32.
- de Almeida E.A., Miyamoto S., Bainy A.C.D., de Medeiros M.H.G. and Di Mascio P. (2004) Protective effect of phospholipid hydroperoxide glutathione peroxidase (PHGPx) against lipid peroxidation in mussels *Perna perna* exposed to different metals. *Marine Pollution Bulletin* 49, 386–392.
- del Brio F., Castro I.B., Fillmann G., Commendatore M., Gomes Costa P. and Bigatti G. (in press) Distribution and bioaccumulation of butyltins in the edible gastropod *Odontocymbiola magellanica*. *Marine Biology Research*.
- Di Salvatore P., Calcagno J.A., Ortíz N., Ríos de Molina M.C. and Sabatini S.E. (2013) Effect of seasonality on oxidative stress responses and metal accumulation in soft tissues of *Aulacomya atra*, a mussel from the South Atlantic Patagonian coast. *Marine Environmental Research* 92, 244–252.
- Doyen P., Vasseur P. and Rodius F. (2005) cDNA cloning and expression pattern of pi-class glutathione-S-transferase in the fresh water bivalves *Uniotumidus* and *Corbicula fluminea*. *Comparative Biochemistry and Physiology Part C: Toxicology and Pharmacology* 140, 300–308.
- Esteves J.L., Commendatore M., Nievas M.L., Massara Paletto V. and Amín O. (2006) Hydrocarbon pollution in coastal sediments of Tierra del Fuego Islands, Patagonia Argentina. *Marine Pollution Bulletin* 52, 582–590.
- Giarratano E., Duarte C.A. and Amin O.A. (2010) Biomarkers and heavy metal bioaccumulation in mussels transplanted to coastal waters of the Beagle Channel. *Ecotoxicology and Environmental Safety* 73, 270–279.
- Giarratano E., Gil M.N. and Malanga G. (2013) Assessment of antioxidant responses and trace metal accumulation by digestive gland of ribbed mussel *Aulacomya atra atra* from Northern Patagonia. *Ecotoxicology and Environmental Safety* 92, 39-50.
- Gibbs P.E. and Bryan G.W. (1986) Reproductive failure in populations of the dog-whelk, *Nucella lapillus*, caused by imposex induced by tributyltin from antifouling paints. *Journal of the Marine Biological Association of the United Kingdom* 66, 767–777.
- Gil M.N., Harvey M. and Esteves J.L. (1988) Metal content in bivalves molluscs from the San Jose and Nuevo gulfs, Patagonia Argentina. Marine Pollution Bulletin 19, 181–182.
- Gil M.N., Harvey M.A. and Esteves J.L. (1999) Heavy metals in intertidal surface sediments from the Patagonian Coast, Argentina. Bulletin of Environmental Contamination and Toxicology 63, 52-58.
- Gil M.N., Torres A., Harvey M. and Esteves J.L. (2006) Metales pesados en organismos marinos de la zona costera de la Patagonia argentina continental. *Revista de Biologia Marina y Oceanografia* 41, 167–176.
- Gopalakrishnan S., Huang W.B., Wang Q.W., Wu M.L., Liu J. and Wang K.J. (2011) Effects of tributyltin and benzo[a]pyrene on the

- immune-associated activities of hemocytes and recovery responses in the gastropod abalone, *Haliotis diversicolor. Comparative Biochemistry and Physiology, Part C: Toxicology and Pharmacology* 154, 120–128.
- Habig W.H., Pabst M.J. and Jakoby W.B. (1974) Glutathione S-transferases: the first enzymatic step in mercapturic acid formation. *Journal of Biological Chemistry* 249, 7130–7139.
- Hayes J.D., Flanagan J.U. and Jowsey I.R. (2005) Glutathione transferases. Annual Review of Pharmacology and Toxicology 45, 51-88.
- Huang Z., Chen Y., Zhao Y., Zuo Z., Chen M. and Wang C. (2005)

  Antioxidant responses in *Meretrix meretrix* exposed to environmentally relevant doses of tributyltin. *Environmental Toxicology and Pharmacology* 20, 107–111.
- Jia X., Zhang Z., Wang S., Lin P., Zou Z., Huang B. and Wang Y. (2009) Effects of tributyltin (TBT) on enzyme activity and oxidative stress in hepatopancreas and hemolymph of small abalone, *Haliotis diversicolor* supertexta. Chinese Journal of Oceanology and Limnology 27, 816–824.
- Karakoc F.T., Hewer A., Phillips D.H., Gaines A.F. and Yuregir G. (1997) Biomarkers of marine pollution observed in species of mullet living in two eastern Mediterranean harbours. *Biomarkers* 2, 303–309.
- Kim S.K., Oh J.R., Shim W.J., Lee D.H., Yim U.H., Hong S.H., Shin Y.B. and Lee D.S. (2002) Geographical distribution and accumulation features of organochlorine residues in bivalves from coastal areas of South Korea. *Marine Pollution Bulletin* 45, 268–279.
- **Leonard S.S., Harris G.K. and Shi X.** (2004) Metal-induced oxidative stress and signal transduction. *Free Radical Biology and Medicine* 37, 1921–1942.
- Liu H.G., Wang Y., Lian L. and Xu L.H. (2006) Tributyltin induces DNA damage as well as oxidative damage in rats. *Environmental Toxicology* 21, 166-171.
- Livingstone D.R. (2001) Contaminant-stimulated reactive oxygen species production and oxidative damage in aquatic organisms. *Marine Pollution Bulletin* 42, 656–666.
- **Malanga G., Estevez M.S., Calvo J. and Puntarulo S.** (2004) Oxidative stress in limpets exposed to different environmental conditions in the Beagle Channel. *Aquatic Toxicology* 69, 299–309.
- **Márquez F., González R.J. and Bigatti G.** (2011) Combined methods to detect pollution effects on shell shape and structure in Neogastropods. *Ecological Indicators* 11, 248–254.
- Massara Paletto V., Commendatore M.G. and Esteves J.L. (2008) Hydrocarbon levels in sediments and bivalve mollusks from Bahía Nueva (Patagonia, Argentina): an assessment of probable origin and bioaccumulation factors. *Marine Pollution Bulletin* 56, 2082–2105.
- Meador J.P., Stein J.E., Reichert W.L. and Varanasi U. (1995) Bioaccumulation of polycyclic aromatic hydrocarbons by marine organisms. In Ware G.W. (ed.) *Reviews of environmental contamination and toxicology.* New York, NY: Springer, Volume 143, pp. 79– 165.
- Narvarte M.A., Willers V., Avaca M.S. and Echave M.E. (2008)
  Population structure of the snail *Buccinanops globulosum*(Prosobranchia, Nassariidae) in San Matías Gulf, Patagonia
  Argentina: isolated enclaves? *Journal of Sea Research* 60, 144–150.
- **Nicholson S. and Lam P.K.S.** (2005) Pollution monitoring in Southeast Asia using biomarkers in the mytilid mussel *Perna viridis* (Mytilidae: Bivalvia). *Environment International* 31, 121–132.
- Nishikawa J., Mamiya S., Kanayama T., Nishikawa T., Shiraishi F. and Horiguchi T. (2004) Involvement of the retinoid x receptor in the development of imposex caused by organotins in gastropods. *Environmental Science and Technology* 38, 6271–6276.

- Pastorino G. (1993) The taxonomic status of *Buccinanops* d'Orbigny, 1841 (Gastropoda: Nassariidae). *The Veliger* 36, 160–165.
- Penchaszadeh P.E. (1971) Aspectos de la embriogenesis de algunos gasterópodos del genero *Buccinanops* d'Orbigny, 1841 (Gastropoda, Prosobranchiata, Buccinidae). *Physis* 30, 475–482.
- Primost M.A. (2014) Ecotoxicología y alteraciones morfo-funcionales en gasterópodos marinos expuestos a contaminación por organometales y metales. PhD thesis. Universidad Nacional de La Plata, Argentina.
- **Primost M.A., Bigatti G. and Márquez F.** (in press). Shell shape variation as indicator of harbour pollution in marine gastropods affected by imposex. *Marine and Freshwater Research*.
- Prohaska J.R. (1980) The glutathione peroxidase activity of glutathione S-transferases. Biochimica et Biophysica Acta – Enzymology 611, 87-08
- Regoli F. and Principato G. (1995) Glutathione, glutathione-dependent and antioxidant enzymes in mussel, *Mytilus galloprovincialis*, exposed to metals under field and laboratory conditions: implications for the use of biochemical biomarkers. *Aquatic Toxicology* 31, 143–164.
- Sabatini S.E., Rocchetta I., Luquet C.M., Guido M.I. and Ríos de Molina M.C. (2009) Dietary copper effects in the estuarine crab, Neohelice (Chasmagnathus) granulata, maintained at two different salinities. Comparative Biochemistry and Physiology, Part C 150, 521-527.
- Sabatini S.E., Rocchetta I., Luquet C.M., Guido M.I. and Ríos de Molina M.C. (2011a) Effects of sewage pollution and bacterial load on growth and oxidative balance in the freshwater mussel *Diplodon chilensis*. *Limnologica* 41, 356–362.
- Sabatini S.E., Rocchetta I., Nahabedian D.E., Luquet C.M., Eppis M.R., Bianchi L. and Ríos de Molina M.C. (2011b) Oxidative stress and histological alterations produced by dietary copper in the fresh water bivalve Diplodon chilensis. Comparative Biochemistry and Physiology, Part C 154, 391–398.
- Sayeed I., Parvez S., Pandey S., Bin-Hafeez B., Haque R. and Raisuddin S. (2003) Oxidative stress biomarkers of exposure to deltamethrin in freshwater fish, Channa punctatus Bloch. Ecotoxicology and Environmental Safety 56, 295–301.
- Scarabino V. (1977) Moluscos del Golfo San Matías, provincia de Río Negro, República Argentina. Inventario y claves para su identificación. Comisión de la Sociedad Malacológica de Uruguay 4, 177-285.
- Sheehan D. and Power A. (1999) Effects of seasonality on xenobiotic and antioxidant defence mechanisms of bivalve molluscs. *Comparative Biochemistry and Physiology Part C: Pharmacology, Toxicology and Endocrinology* 123, 193–199.
- Sokal R.R. and Rohlf F.J. (1979) Introducción a la bioestadística. Barcelona, España: Reverté.

- Torres P.J., Commendatore M.G., Primost M.A. and Bigatti G. (2013) Hidrocarburos Poliaromáticos (PAHs) en *Buccinanops globulosus* afectados por imposex. *Proceedings of the Abstract book I Congreso Argentino de Malacología (CAM)*, 99 pp.
- Valavanidis A., Vlahogianni T., Dassenakis M. and Scoullos M. (2006) Molecular biomarkers of oxidative stress in aquatic organisms in relation to toxic environmental pollutants. *Ecotoxicology and Environmental Safety* 64, 178–189.
- Vavilin D.V., Ducruet J.M., Matorin D.N., Venediktov P.S. and Rubin A.B. (1998) Membrane lipid peroxidation, cell viability and photosystem II activity in the green alga *Chlorella pyrenoidosa* subjected to various stress conditions. *Journal of Photochemistry and Photobiology*, B: Biology 42, 233-239.
- Viarengo A., Canesi L., Pertica M. and Livingstone D.R. (1991) Seasonal variations in the antioxidant defence systems and lipid peroxidation of the digestive gland of mussels. Comparative Biochemistry and Physiology, Part C: Comparative Pharmacology 100, 187–190.
- Viarengo A., Canesi L., Pertica M., Poli G., Moore M.N. and Orunesu M. (1990) Heavy metal effects on lipid peroxidation in the tissues of Mytilus galloprovincialis Lam. Comparative Biochemistry and Physiology, Part C: Comparative Pharmacology 97, 37–42.
- Wang C.G., Chen Y.X., Li Y., Wei W. and Yu Q. (2005) Effects of low dose tributyltin on activities of hepatic antioxidant and phase II enzymes in Sebastiscus marmoratus. Bulletin of Environmental Contamination and Toxicology 74, 114–119.
- Wang L., Yan B., Liu N., Li Y. and Wang Q. (2008) Effects of cadmium on glutathione synthesis in hepatopancreas of freshwater crab, *Sinopotamon yangtsekiense*. Chemosphere 74, 51-56.
- Winston G.W. and Di Giulio R.T. (1991) Prooxidant and antioxidant mechanisms in aquatic organisms. *Aquatic Toxicology* 19, 137–161.
- Young I. and Woodside J. (2001) Antioxidants in health and disease. Journal of Clinical Pathology 54, 176–186.

and

Zhou J., Zhu X. and Cai Z. (2010) Tributyltin toxicity in abalone (Haliotis diversicolor supertexta) assessed by antioxidant enzyme activity, metabolic response, and histopathology. Journal of Hazardous Materials 183, 428-433.

## Correspondence should be addressed to:

S.E. Sabatini

IQUIBICEN-Dpto Química Biológica, Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires, Pab. II, Intendente Guiraldes 2160, C1428EHA, Ciudad Autónoma de Buenos Aires, Argentina

email: sabatini@bg.fcen.uba.ar