Biochimica et Biophysica Acta xxx (2013) xxx-xxx

Contents lists available at ScienceDirect

Biochimica et Biophysica Acta

journal homepage: www.elsevier.com/locate/bbamem



Atypical surface behavior of ceramides with nonhydroxy and 2-hydroxy very long-chain (C28–C32) PUFAs

Daniel A. Peñalva ^a, Gerardo M. Oresti ^a, Fernando Dupuy ^b, Silvia S. Antollini ^a, Bruno Maggio ^b, Marta I. Aveldaño ^a, María L. Fanani ^{b,*}

- ^a Instituto de Investigaciones Bioquímicas de Bahía Blanca (INIBIBB), Universidad Nacional del Sur and Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), 8000 Bahía Blanca, Argentina
- b Departamento de Química Biológica, Centro de Investigaciones en Química Biológica de Córdoba (CIQUIBIC), Facultad de Ciencias Químicas,
- Universidad Nacional de Córdoba and CONICET, 5000 Córdoba, Argentina

ARTICLE INFO

Article history:

10

11

16

19

41

42

43

44

45 46

47

48

49 50

51

52

- 12 Received 15 August 2013
- 13 Received in revised form 7 November 2013
 - Accepted 26 November 2013
- 15 Available online xxxx

Keywords:

- 20 2-Hydroxy fatty acids
- 21 Brewster angle microscopy
- 22 Langmuir monolayers
- 23 Two dimensional phase transitions
- 24 Sphingolipids

ABSTRACT

Unique species of ceramide (Cer) with very-long-chain polyunsaturated fatty acid (VLCPUFA), mainly 28–32 carbon atoms, 4–5 double bonds, in nonhydroxy and 2-hydroxy forms (n-V Cer and h-V Cer, respectively), are generated in rat spermatozoa from the corresponding sphingomyelins during the acrosomal reaction. The aim of this 27 study was to determine the properties of these sperm-distinctive ceramides in Langmuir monolayers. Individual 28 Cer species were isolated by HPLC and subjected to analysis of surface pressure, surface potential, and Brewster 29 angle microscopy (BAM) as a function of molecular packing. In comparison with known species of Cer, n-V Cer 30 and h-V Cer species showed much larger mean molecular areas and increased molecular dipole moments in liquid expanded phases, which suggest bending and partial hydration of the double bonded portion of the VLCPUFA. 32 The presence of the 2-hydoxyl group induced a closer molecular packing in h-V Cer than in their chain-matched 33 n-V Cer. In addition, all these Cer species showed liquid-expanded to liquid-condensed transitions at room temperature. Existence of domain segregation was confirmed by BAM. Additionally, thermodynamic analysis suggests a phase transition close to the physiological temperature for VLCPUFA-Cers if organized as bulk dispersions. 36 © 2013 Published by Elsevier B.V. 37

1. Introduction

Intensive research over the past decades has implicated ceramide (Cer) in the regulation of a great variety of key cellular functions. However, the paradigm that a single Cer species is responsible for many different cell functions has been challenged by the recognition that "ceramide" constitutes a family of related molecules, subject to metabolism by nearly 30 enzymes and with dozens of structurally distinct molecular species [15]. Saturated and monounsaturated fatty acids from C_{14} to C_{24} are typical acyl chains of sphingolipids in most mammalian tissues, although shorter chain ceramides originated from various transacylation reactions also occur naturally [6]. Notable exceptions are skin and testis sphingolipids, which contains glucosyl-Cer species

with fatty acids up to C_{34} long [39] and mammalian testis and spermatozoa, which have sphingomyelin (SM) and Cer with high proportions of very long chain (C_{24} to C_{36}) polyunsaturated fatty acids (VLCPUFA) of the n-6 or the n-3 series [12,35], i.e., elongated versions of well-known C20 or C22 PUFA of the n-6 or the n-3 series, such as arachidonic, docosapentaenoic or docosahexaenoic acids.

In many tissues, of which myelin and skin sphingolipids are known 60 examples, an important part of the sphingolipid species contains a 2- 61 hydroxyl group at the second carbon atom of their fatty acyl chain 62 [14]. This biochemical modification also occurs in the VLCPUFA of ro- 63 dent testis sphingolipids including SM [37], gangliosides [36,40], and 64 Cer [43]. In rat testis, the ratio between 2-hydroxy and nonhydroxy 65 VLCPUFA (h-V and n-V, respectively) in SM and Cer increases from 66 the onset of spermatogenesis to adulthood [43]. This is due to the fact 67 that n-V Cer and SM species are exclusive components of precursor 68 spermatocytes, whereas h-V Cer and SM species appear as they differ- 69 entiate to spermatids, as well as in spermatozoa [43]. In mature gam- 70 etes, n-V SM and h-V SM, but no Cer, occur endogenously on the 71 sperm head and, intriguingly, considerable amounts of solely h-V Cer 72 species are located on the tail [30]. Almost complete hydrolysis of the 73 head-located SM occurs after inducing the acrosomal reaction [44], 74 this leading to gametes that are considerably enriched in n-V, and 75 especially in h-V, ceramides. Thus, whereas in testicular germ cells 76 VLCPUFA-containing ceramides play a role as biosynthetic precursors 77

E-mail address: lfanani@fcq.unc.edu.ar (M.L. Fanani).

0005-2736/\$ – see front matter © 2013 Published by Elsevier B.V. http://dx.doi.org/10.1016/j.bbamem.2013.11.018

Abbreviations: Cer, ceramide; VLCPUFA, very long chain polyunsaturated fatty acid; h-V, 2-hydroxy VLCPUFA; n-V, nonhydroxy VLCPUFA; SM, sphingomyelin; μ_{\perp} , dipole moment perpendicular to the interface; π, surface pressure; π_b transition pressure; ΔV , surface potential; BAM, Brewster angle microscopy; Cs $^{-1}$, compressibility modulus; LC, liquid-condensed; LE, liquid-expanded; MMA, mean molecular area; S, solid phase; L, semi-empirical average molecular length; V, semi-empirical molecular volume; V, semi-empirical film thickness

^{*} Corresponding author at: CIQUIBIC, UNC-CONICET, Haya de la Torre y Medina Allende, Ciudad Universitaria, X5000HUA Córdoba, Argentina. Tel.: +54 351 4334168; fax: +54 351 4334074.

2

78 79

80 81

82

83

84

85 86

87

88

89

90

91

92

93

94 95

96

97

98

99

100

102

103

104

 $105 \\ 106$

107

108

109

110

111

112

113

114

115

116

117

118 119

120

121

122

123

124

125

126

of complex sphingolipids including SM, in acrosome-reacted spermato-zoa they are final products. Based on previous studies on different species of SM being converted to Cer by the action of sphingomyelinase [8,19,38], such a massive increase in the Cer/SM mole ratio may be envisaged to have an important impact on the sperm surface properties.

Although there are studies extensively describing Cer properties in Langmuir monolayer [4,6,7,9,18,23], the effects on lipid bilayers of diverse Cer species in pure and mixed form [34], and the impact of actively generating Cer from SM by sphingomyelinase on different features of the membrane physical state [8,19,38], the properties and behavior of the relatively novel n-V and h-V Cer species remain to be established. The aim of this study was to define specific molecular parameters of individual molecular species of these sperm-associated Cers, as well as their thermodynamic properties. In Langmuir monolayers, surface pressure, surface potential, and imaging by Brewster angle microscopy (BAM) were measured to determine their average molecular organization at the membrane interface. Our results showed that n-V Cer species differ significantly from h-V Cer species in most of their surface properties and that both behave atypically if compared with the more ubiquitous molecular species of Cer previously described in the literature [6].

2. Materials and methods

2.1. Chemicals

16:0 Cer, 18:1 Cer, 24:1 Cer and 2-hydroxy 24:0 Cer were from Avanti Polar Lipids Co (Alabaster, AL). The n-V Cer and h-V Cer were obtained from adult Wistar rat testes using a combination of TLC and HPLC procedures. The silica gel was from Merck, the HPLC column was from Rainin LC. The gas chromatography and HPLC equipments were both from Varian Inc., USA. All solvents were HPLC grade (JT Baker; UVE, Dorwill, Argentina).

2.2. Separation of ceramides

Lipid extracts were prepared from rat testes [2] and spotted on 500 μm-silica gel TLC plates under N₂ along with commercial standards. Chloroform:methanol:ammonia (90:10:2, by vol.) resolved rat testis Cer into two bands. The lower one was almost exclusively made up by h-V Cer species, whereas the upper one contained all Cer species with nonhydroxy fatty acids, this including common Cer species with C₁₆ to C₂₆ fatty acids and those with nonhydroxy VLCPUFA [43]. The separated h-V and n-V fractions were recovered and treated with N2-saturated 0.5 N NaOH in anhydrous methanol at 50 °C for 10 min in order to remove any potential lipid contaminant with ester-bounded fatty acids from the Cer samples, neutralized, partitioned into chloroform, and subjected again to TLC. This time chloroform:methanol:ammonia:water (90:10:05:0.5, by vol.) was used, as it allows the fraction containing Cer species with C₁₆ to C₂₆ fatty acids to lag behind the Cer species containing nonhydroxy VLCPUFA [11]; this propensity facilitates partial purification of the latter.

Silica gel plates impregnated with 10% AgNO₃ and chloroform:methanol (80:20, by vol.) were then used to separate the Cer species with C_{16} to C_{26} fatty acids from those containing VLCPUFA. This resulted in partial

resolution of the latter ceramides into two bands: the upper one 127 contained the main very long chain tetraenoic fatty acid (n-28:4) 128 and the lower one mainly the pentaenoic fatty acids (n-30:5) and 129 (n-32:5). Although with the expectedly lower Rf values due to the hy- 130 droxyl group, an essentially similar separation resulted when h-V Cers 131 were subjected to this procedure. After this pre-separation, each of the 132 Cer subfractions was subjected to HPLC to obtain the six major 133 VLCPUFA-containing Cer species from rat testis (Fig. 1).

Lipids were located on TLC plates under UV light after spraying with 135 2,7-dichlorofluorescein, and rapidly transferred to glass tubes for elu-136 tion. This was done by successively extracting the silica with nitrogen-137 saturated chloroform:methanol:water (5:5:1), centrifuging, collecting 138 the supernatants, and partitioning them with 4.5 volumes of water [2]. 139 The eluates were finally washed with methanol:1 M NaCl (1:1 by vol.) 140 to remove Ag⁺ ions. The organic phases were reduced in volume 141 under N₂, filtered to remove traces of particulate matter, dried, and dissolved in methanol for HPLC injection.

HPLC was performed at 35 °C using a stainless steel column 144 (250 mm \times 4.6 mm ID) packed with 5 μ m particles covered with 145 octylsilane (C8). The solvent, at a flow rate of 1 mL/min was 146 methanol:1 mM potassium phosphate buffer, pH 7.4, in a 96:4 (v/v) 147 ratio. Cer peaks were detected at 205 nm using a Prostar 335 photodi- 148 ode array detector and collected as they emerged from the column. 149 Chloroform and water were then added to the eluates and Cer species 150 were recovered in the chloroform phase resulting after phase partition. 151

Individual species were identified and quantified by means of gas 152 chromatography after adding appropriate amounts of n-16:0 Cer 153 and h-24:0 (2S–OH) as internal standards to the samples, subjecting 154 them to methanolysis, and recovering by TLC the nonhydroxy and 2- 155 hydroxy fatty acid methyl esters to be subjected to GC. The former were analyzed directly and the latter after conversion into trimethyl- 158 silyl derivatives [29,43].

2.3. Monolayer compression isotherms

Langmuir isotherms were obtained using a 90 cm 2 compartment of 160 a specially designed circular Monofilmmeter Teflon trough (Mayer 161 Feintechnik, Germany) filled with 80 mL of 145 mM NaCl, pH \sim 5.6, 162 equipped with a platinized Pt plate for measuring the surface pressure. 163 The surface potential of the film was simultaneously measured by a surface ionizing electrode formed by a 241 Am plate positioned 5 mm above the monolayer surface, and a reference calomel electrode connected to 166 the aqueous subphase.

159

The whole system was enclosed in an acrylic box, saturated with N $_2$ 168 gas to prevent lipid peroxidation, and surrounded by a metallic grid 169 connected to ground to reduce external interference in surface potential 170 measurements. Experiments were performed using a subphase of 171 145 mM NaCl kept at 21 °C and 8 °C (\pm 0.5 °C) by means of an external 172 circulating water bath (Haake F3C). Absence of surface active impurities 173 in the subphase solution or the spreading solvent was ascertained as described elsewhere [32].

Lipid monolayers were obtained by spreading adequate aliquots 176 of Cer solutions onto the aqueous surface. After allowing solvent 177 evaporation for 5 min, the surface pressure—area isotherms were 178

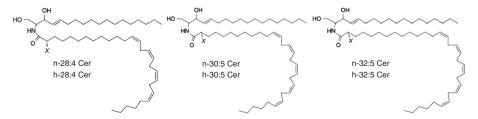


Fig. 1. Schematic structures of the ceramides (Cer) containing nonhydroxy (n) and 2-hydroxy (h) very-long-chain polyunsaturated fatty acids (VLCPUFA) studied in this work. X represents the presence of – H or – OH, respectively, in the second carbon atom of the amide-bound fatty acid. The depictions are shown for illustrative purposes and are not meant to represent the molecular organization adopted by the chains at the interface.

Please cite this article as: D.A. Peñalva, et al., Atypical surface behavior of ceramides with nonhydroxy and 2-hydroxy very long-chain (C28–C32) PUFAs, Biochim. Biophys. Acta (2013), http://dx.doi.org/10.1016/j.bbamem.2013.11.018

recorded while compressing the monolayers at a constant speed of $2 \text{ Å}^2 \text{ molecule}^{-1} \text{ min}^{-1}$. The collapse pressures and limiting mean molecular areas of the Cer films were estimated by the third derivative methods [3]. Compressibility modulus (Cs⁻¹) [13] was as:

$$Cs^{-1} = -MMA(\partial \pi/\partial MMA)_{T}, \tag{1}$$

where MMA is the mean molecular area at a given surface pressure π . The resultant dipole moment perpendicular to the interface (μ_{\perp}) [13] of Cer at each molecular area was calculated from the isotherm data as:

$$\mu_{\perp} = \frac{1}{12\pi} MMA\Delta V, \tag{2}$$

189 where ΔV is the surface potential of the monolayer at the corresponding MMA, and π in this case is the mathematical Euclidean constant taken as 3.14159.

2.4. Brewster angle microscopy

179 180

181 182

182

185

186

187

192

193

194 195

196

197

198

199 200

201

202

203

204

205

206

BAM visualization was performed with an auto-nulling imaging ellipsometer (Nanofilm EP3sw imaging ellipsometer, Accurion GmbH, Germany) working in the BAM mode. Zero reflection was set with a polarized 532 nm laser incident on the bare aqueous surface at the experimentally calibrated Brewster angle (53.1°). After monolayer formation and compression, the reflected light was collected with a $20\times$ objective and a CCD camera, which operates at a resolution of 1 μ m. The gray level of each section of the micrograph is proportional to the reflected light intensity, which is a function of both the film thickness and its refraction index [21].

3. Results and discussion

3.1. Isolation of VLCPUFA-containing ceramide species

The procedure we used to isolate the six major VLCPUFA-containing Cer species (Fig. 1) whose properties are explored in this study was

based on a recently described method applied to isolate nonhydroxy 207 and 2-hydroxy VLCPUFA-containing species of SM from rat testis [31]. 208 Whereas the high polarity of the phosphoryl-choline headgroup in 209 SM imposed a difficulty for the separation of the species containing a 210 hydroxyl group from their non-hydroxylated analogs by silica-based 211 TLC, the h-V Cer and h-V Cer species were easily separated using silica 212 gel as support. As previously shown [43], the h-V Cer species lag well be- 213 hind the subfraction containing nonhydroxy fatty acids, and are detect- 214 ed as a single sharp band because it is composed almost exclusively by 215 Cer species containing 2-hydroxy VLCPUFA. In contrast, the Cer species 216 with nonhydroxy fatty acids tend to separate into bands according to 217 the length of their fatty chains. Of these bands, the one grouping the 218 Cer species with nonhydroxy VLCPUFA tends to migrate ahead from 219 species containing shorter, mostly saturated and some monoenoic 220 $(C_{16}-C_{26})$ fatty acids [12]. In preliminary HPLC separations, we found 221 that Cer species with 24:0 or 24:1 tended to co-elute with some of the 222 species containing C₂₈–C₃₀ VLCPUFA of our interest; for this reason we 223 decided to directly eliminate them from our samples using argentation 224 TLC (Ag-TLC).

The high degree of unsaturation of the VLCPUFA-rich Cer species 226 with respect to the common (C_{16} – C_{26}) ceramides was advantageous, 227 not only to separate the former from the latter using Ag–TLC, but to detect the former with a good sensitivity when subjected to HPLC (Fig. 2). 229 In contrast to the case of SM, for which species with very-long-chain 230 tetraenoic and pentaenoic fatty acids are hard to separate by Ag–TLC, 231 Cer species containing these fatty acids tended to separate according 232 to the degree of unsaturation of the fatty acid when subjected to this 233 procedure. Although this pre-separation implied a more laborious procedure, since more samples had to be eluted and injected, it facilitated 235 the isolation of Cer species according to their fatty acid chain length 236 and number of double bonds, as shown in Fig. 2.

The reverse-phase HPLC procedure previously devised to separate 238 SMs also separated Cer species in the same order, mostly determined 239 by their fatty acids. In both cases, elution of each of the h-V species pre- 240 ceded that of its corresponding chain-matched n-V counterpart (e. g., 241 h - 30:5 Cer eluted before n-30:5 Cer), as expected because of the 242 higher polarity conferred to the former by the hydroxyl group. Also, 243

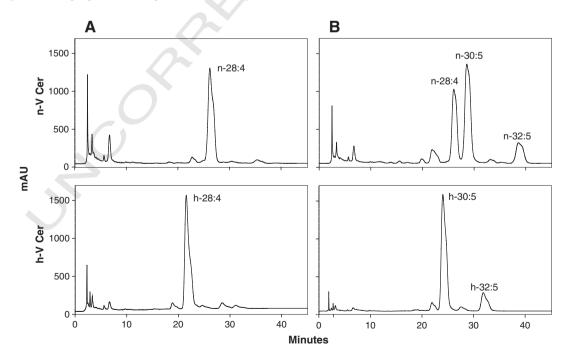


Fig. 2. Preparative isolation of VLCPUFA-containing ceramides. The nonhydroxy and 2-hydroxy VLCPUFA-containing Cer (n-V Cer and h-V Cer, respectively) were separated from rat testis by TLC and subjected to argentation TLC to isolate them from Cer species with saturated and monoenoic fatty acids. This partially resolved each the VLCPUFA-containing Cer into tetraenoic (A) and pentaenoic (B) fatty acid-containing fractions. Each of these was then separately subjected to HPLC to collect the six major (C28–C32) molecular species shown in Fig. 1. Chromatography was performed on an octylsilane column using a mobile phase of methanol–phosphate buffer flowing at 1 mL/min and a detector set at $\lambda = 203$ nm.

244 245

 $\frac{246}{247}$

248

249

250

251

252

253

254

255

256

257

258

259

260

261

262

263

264

265

266

within h-V or n-V subfractions, species with 28:4n – 6 preceded those with 30.5n - 6 in elution order, just as free 20.4n - 6 precedes free 22.5n-6 when they are subjected together to HPLC on reverse phase columns [1]. Thus, an additional segment of two carbon atoms in the fatty acid is more powerful in increasing the reverse phase retention of Cer than the presence of an additional double bond is in decreasing it. Most interestingly, retention times of each of the Cer species studied here (Fig. 2) were somewhat shorter than those of the corresponding SM species having chain-matched fatty acids [31], despite the high polarity the phosphorylcholine headgroup confers to the latter. Because the hydrophobicity associated to the fatty acids amide-bound to sphingosine is the same in these SM and Cer species, this relatively stronger retention may be ascribed to additional forces between the polar head group of SM and the material of the column. These could involve polar interaction of the SM charged phosphate group with free silanols or hydrophobic interactions between SM methyl groups with the covalently bound hydrophobic material. This could explain the higher symmetry of the peaks of the present Cer (Fig. 2) than of the previous SM [31] species having the same fatty acids.

3.2. Langmuir monolayers of VLCPUFA-containing Cer species

In this section, Langmuir compression isotherms of the six Cer species of this study were performed at room temperature. As shown in Fig. 3, all of them underwent a phase transition in the 5–20 mN/m

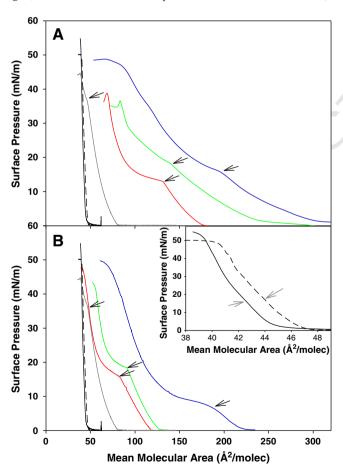


Fig. 3. Compression isotherms of n-V Cer (A) and h-V Cer (B) species. In each of the panels, the mean molecular area vs. surface pressure curves were plotted using the same symbol for each nV-Cer and the corresponding h-V Cer, as follows: n-28:4 Cer and h-28:4 Cer (red lines), n-30:5 Cer and h-30:5 Cer (green lines) and n-32:5 Cer and h-32:5 Cer (blue lines). For comparison, the isotherms of 24:1 Cer (black dashed line), 16:0 Cer (full black line) and 18:1 Cer (gray line) are also shown. The inset shows 16:0 Cer and 18:1 Cer in an expanded x-axis scale. The black arrows indicate the beginning of a LE \rightarrow LC phase transition and the gray arrows a LC \rightarrow S transition.

pressure range, as evidenced by a kink in the plots relating mean molecular area (MMA) and surface pressure (π). At π values below 10 mN/m 268 (or even lower in the case of h - 32:5Cer), and relatively large values of 269 MMA, the films showed a liquid-expanded (LE) character, as emphasized by the relatively low value of the corresponding compressibility 271 modulus (Cs $^{-1}$) (Table 1).

Cs⁻¹ partially reflects the surface elasticity of the films, giving values 273 lower than 130 mN/m for typical LE phases [22,41,42]. When the transi- 274 tion pressure (π_t) is reached and a LE \rightarrow LC coexistence region is 275 attained, Cs⁻¹ typically drops to lower values (20–30 mN/m) and rises 276 again when a homogeneous liquid-condensed (LC) phase is established 277 (Table 1, see 30 mN/m). For the present VLCPUFA-containing ceramides, 278 the π_t followed the trend: 30:5Cer > 28:4Cer > 32:5Cerin both n-V and 279 h-V cases (Fig. 3). It is worth noting that the Cs⁻¹ values for the LC phases 280 formed by all Cer species containing VLCPUFA were lower than those ob- 281 tained for other Cers, as shown for 16:0Cer and 24:1Cer in Table 1 and in 282 [6,9,18], and also lower than that for phospholipid-LC phases, for which 283 $Cs^{-1} > 200 \text{ mN/m}$ [22,41,42]. In comparison, at 21 °C saturated 284 16:0Cer organizes in a LC phase [18], which undergoes a LC \rightarrow solid 285 (S) phase transition at ~16–18 mN/m and a MMA of ~43 Å^2 /molecule, 286 as shown as an inset in Fig. 3, being able to form LE phases only at higher 287 temperatures (~35to ~48 °C) [9].

Shortening of the acyl chain, as in 12:0Cer or 10:0Cer [6], or introduc- 289 tion of a double bond at C9 in the amide-bound fatty acid of ceramide, as 290 in 9Δ18:1Cer,leads to expanded monolayers which transform to con- 291 densed phases at higher π_t values than those of any of the VLCPUFA- 292 containing ceramides examined here (see Fig. 3 and [23]). However, 293 when the double bond occurs at the C15 of the N-linked acyl hydrocar- 294 bon chain, as in $15\Delta 24$:1Cer, the molecules can still pack into an LC 295 phase at room temperature [18,23] and exhibit a LC \rightarrow S transition 296 quite similar to that shown by the saturated 16:0Cer (Fig. 3 and 297 Table 1). Counting from the amide-bound carbon, the first of the series 298 of 4 or 5 methylene-interrupted cis double bonds of the Cer species of 299 this study are located at C12, C13 and C14 in the 30:5, 28:4 and 32:5 300 acyl chains, respectively (Fig. 1), i.e., at an intermediate location between 301 the two above-mentioned monounsaturated Cers. Our results indicate 302 that VLCPUFA-containing Cer species also behave in an intermediate 303 manner between these two species in monolayer, showing both an LE 304 and an LC phase at room temperature (Fig. 3). The LC phase may be ascribed to their saturated portion facilitating Van der Waals interactions 306 among acyl chains, and the LE phase to their several double bonds, pro- 307 moting a more expanded behavior of these species by inducing hydro- 308 carbon chain disordering and reducing the tightness of their packing.

A remarkable characteristic of VLCPUFA-containing Cer species was 310 their large MMA (Fig. 3 and Table 1). Ordinary ceramides in their LC 311 or S state typically occupy an area of $\sim 40-42 \text{ Å}^2/\text{molecule}$ at high π 312 (Fig. 3, inset and [4]) which is roughly the lower limiting cross- 313 sectional area for lipids with two-tailed fully extended saturated acyl 314 chains. However, only h – 28:4Cer and h – 30:5Cer showed MMA that 315 were close to such value at 30 mN/m. The other Cer species with 316 VLCPUFAshowed larger MMA values that increased with the chain 317 length. At 10 mN/m, VLCPUFA-containing Cer followed the same ten- 318 dency, occupying an area 1.5- to 3.4-fold larger than that occupied by 319 18:1Cer in its LE state. A comparison between each of the n-V Cer and 320 the h-V Cer species showed that the latter packed into smaller areas 321 (Table 1) and had more defined LE \rightarrow LC transitions (Fig. 3) than the 322 former. This reflects a better capacity for close molecular packing in h- 323 V than in n-V Cer species, which may be ascribed to the presence of 324 the 2-hydroxyl group near the air-water interphase.

The surface (dipole) potential (ΔV) in lipid monolayers is the resultant of several components. It includes contributions of the chemical 327 groups of the lipid molecule to the overall resultant dipole moment perpendicular to the interface, as well as the contribution from the hydration shield associated to the polar headgroups [4,13]. As expected, ΔV 330 measurements of the VLCPUFA-containing Cer films increased with 331 the acyl chain length and with π . This may be ascribed to the attainment 332

Table 1Characteristic parameters of VLCPUFA-containing ceramide monolayers.

t3.3	Molecular species	10 mN/m			30 mN/m				
t3.4		MMA	Cs ^{-1a}	Phase state	ΔV	MMA	Cs ^{-1 a}	Phase state	ΔV
t3.5	n-28:4Cer	141 ± 4	62 ± 4	LE	248 ± 5	78 ± 4	80 ± 4	LC	379 ± 9
t3.6	n-30:5Cer	182 ± 3	37 ± 2	LE	258 ± 7	102 ± 6	46 ± 14	LE-LC	301 ± 10
t3.7	n-32:5Cer	222 ± 3	37 ± 2	LE	306 ± 7	125 ± 2	50 ± 2	LC	380 ± 7
t3.8	h-28:4Cer	101 ± 3	56 ± 6	LE	172 ± 1	58 ± 4	74 ± 9	LC	248 ± 1
t3.9	h-30:5Cer	106 ± 2	69 ± 9	LE	168 ± 4	60 ± 1	93 ± 7	LC	257 ± 4
t3.10	h-32:5Cer	140 ± 5	20 ± 6	LE-LC	215 ± 5	97 ± 3	71 ± 8	LC	266 ± 4
t3.11	n – 16:0Cer	43 ± 2	307 ± 10	LC	475 ± 6	41 ± 1	433 ± 10	S	513 ± 20
t3.12	n-18:1Cer	65 ± 3	59 ± 7	LE	237 ± 5	50 ± 2	91 ± 6	LE	274 ± 6
t3.13	n-24:1Cer	45 ± 2	644 ± 12	LC	428 ± 10	42 ± 1	904 ± 15	S	431 ± 10

^a Calculated after Eq. (1).

t3 14

333

334 335

336

337

338

339

340

342

343

344

345

346

348

349

350

351

352

353

354

355

356

357

358

359

360

361

362

363

364

365

366

367

368

369

370

371

372

373

374

375

376 377 of an orientation more perpendicular to the interface of the Cer molecules as the molecular density increases. However, hydration—dehydration processes under increased packing cannot be excluded, since these are also contained in this parameter. The ΔV values for the n-V Cer species in the LE state were situated in the same range as the ΔV of 18:1Cer (which is also able to form an LE phase at room temperature), whereas in the LC state they were lower than those of the condensed 16:0 Cer and 24:1 Cer (Table 1 and [18]). It is remarkable that the n-V Cer species showed higher ΔV values than the h-V Cer species even when such values corresponded to lower molecular densities, considering that the MMA of the n-V Cer species were in all cases larger than those of the corresponding h-V Cer species. This phenomenon may reflect chain conformational differences between those two groups and is discussed further below.

3.3. BAM visualization of monolayers of VLCPUFA-containing ceramides

The surface topography of monolayers of VLCPUFA Cer at the airwater interface was inspected by Brewster angle microscopy (BAM). This technique allows visualization and partial identification of the different lipid phases because of contrasts derived from intrinsic physical properties such as refraction index and interfacial thickness. In general, LC phases appear brighter than LE phases (increased surface reflectivity) as they show a higher refraction index and/or film thickness [6]. Fig. 4 shows BAM images of the VLCPUFA-containing Cer films at the π region where there is LE–LC coexistence. The appearance of LC domains was observed in all but in the n-30:5 Cer film, which showed a very diffuse phase transition at 23 \pm 2 mN/m. This may correspond either to two optically similar phases, not different enough in reflectivity to be visualized, or to the formation of LC domains that were too small to be resolved by the microscope.

The rest of the VLCPUFA-containingCer monolayers revealed the occurrence of small LC domains a few mN/m above their π_t which grew upon film compression forming flower-like bright domains that remained separated until most of the film area became LC (Fig. 4). This behavior was observed previously for other Cer species [6] and is characteristic of the formation of an LC phase with a higher surface potential than that of the LE phase (see Table 1). This induces an intradomain molecular dipole repulsion that leads to adoption of noncircular domain shapes [28], with higher order dipolar repulsion being sensed at long ranges as inducing an inter-domain repulsion that prevents Cer domains to coalesce and leads them to dispose in a rather organized lattice [16].

3.4. Estimation of ceramide organization in monolayers

At this point the question arose whether these lipids, having such large acyl chains, are organized as rather thick films. BAM reflectivity is proportional to the film thickness, but since it also depends on the

film refraction index and their separate contributions cannot be 378 assessed with certainty, this question was approached from an indirect 379 point of view. The average volume and length of an acyl chain has been 380 empirically evaluated in early studies for both glycophospholipids and 381 sphingolipids [20,24] and found to be expressed by the following 382 semi-empirical laws:

$$v = 27.4 + 26.9n; \tag{3}$$

$$l = 0.8(1.5 + 1.265n) - 0.9n_{db}, (4)$$

where v is the average volume of a single hydrocarbon chain, n is the 386 number of carbon atoms in such chain, l is the length of a single chain 388 in its maximal extension, and n_{db} is the number of double bonds in 389 the chain.

For the present Cer species with VLCPUFA, we calculated their total 391 average volume (V) as the sum of the volumes of their two asymmetrical chains (ν) after Eq. (3), and their average chain lengths (L) as the average between the lengths of their two hydrocarbon chains (*l*) after 394 Eq. (4) (Table 2). Also, taking into account the MMA measured experimentally, a semi-empirical thickness (Th) for each Cer species was calculated, as V/MMA. The Th values were smaller for all VLCPUFA- 397 containing Cer than for 16:0 or 24:1 Cer, and smaller for n-V Cer than 398 for their corresponding chain-matched h-V Cer (Table 2). A comparison 399 of *Th* with *L* values should highlight some structural properties of Cer 400 molecular organization in films. Cer Th values should closely match 401 Cer L if molecules self-organized at their LC state with a fully extended 402 conformation, as is the case of common saturated Cer species, whereas 403 a low Th value with respect to L would indicate curved dispositions of 404 their acyl chains. The lowest values of L/Th ratio were displayed by 405 16:0 and 24:1 Cer, followed by h – 28:4 Cer and h – 30:5 Cer in their 406 LC state (Table 2). For all the other VLCPUFA Cer species in the LC 407 phase Th values were considerably smaller than L, even by a factor of 408 two (Table 2). This indicates that these species self-organize with a 409 bent structure of their acyl chains in monolayer films.

Whereas the polar headgroup of Cer is small enough to fit under a 411 transverse area section of scarcely 40 Ų/molecule [6,9], the MMA 412 values for these Cers ranged between 58 and 120 Ų/molecule in the 413 LC phase and 100–222 Ų/molecule in the LE state (Table 1). Thus, an 414 important portion of the hydrocarbon chains may be exposed to the 415 water interface for some Cer species and phase conditions. Due to the 416 conjugated π -orbitals, the unsaturated second portion of the VLCPUFA 417 acyl chains bears a more polarizable nature and higher conformational 418 degrees of freedom at the studied temperatures than the first saturated 419 portion, and it could become partially in contact with the aqueous inter-420 face. Table 2 shows the resultant dipole moment perpendicular to the 421 interface (μ_{\perp}) calculated from ΔV and MMA data according to Eq. (2), 422 which reflects the overall molecular dipole contribution to the total 423 ΔV measurement. Two of the Cer that appeared to be in a straight con-424 formation of their hydrocarbon chains in the LC phase as suggested by 425

426

427

428

429

430

431

432

433

D.A. Peñalva et al. / Biochimica et Biophysica Acta xxx (2013) xxx-xxx

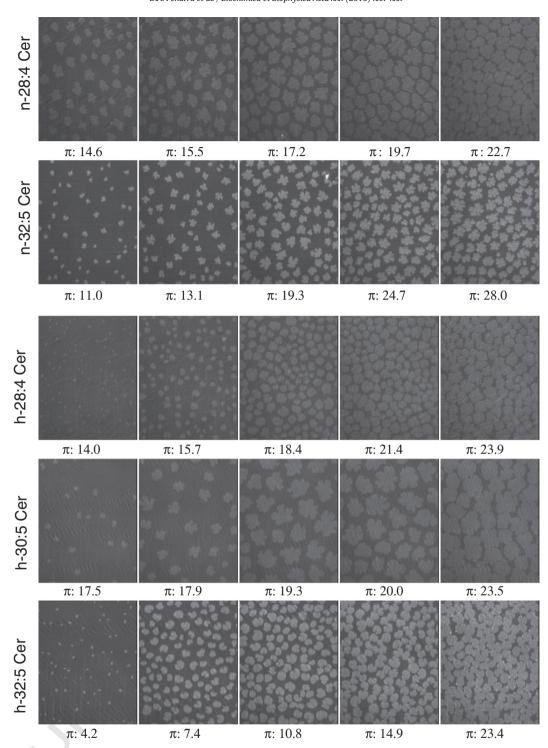


Fig. 4. Brewster angle microscopy visualization of monolayers of VLCPUFA-containing ceramides through the LE-LC transition region. The images correspond to each of the Cer species of this study at the indicated surface pressure π values. For a better visualization, the lower 0–100 gray level range (from the 0–255 original scale) was selected. The pictures are representative of two independent experiments. Image size 200 \times 250 μm.

the L/Th analysis, h - 28:4 and h - 30:5 Cer, showed the lowest μ_{\perp} values of the table (~360–400 mD). These values were even lower than those exhibited for 16:0Cer and 24:1Cer in the LC state ([6,18] and Table 2) indicating a strong contribution of the chain-OH group in opposition to that of the hydrocarbon moiety. Remarkably, the n-V Cer species showed very high μ_{\perp} values, almost duplicating those of the h-V Cer species and correlating with $L/Th \gg 1$ (Table 2). It has been previously demonstrated that the introduction of a double bound in the middle portion of the acyl chain of Cer, where an isotropic

electronic environment is sensed, do not affect substantially the dipole 435 properties. However, when the double bond is sited in a region that 436 present an asymmetric electronic environment, such as in the vicinity 437 of the interface, the double bound contributes to enhance the dipole 438 potential of the molecule, due both by polarization of the conjugated 439 π -orbitals of the double bond and, likely, by interaction with water molecules [4]. Thus, the high resultant dipole moment measured for the 441 longest VLCPUFA-Cers supports that these Cer molecules adopt a bent 442 conformation when organized at the interface.

 Table 2

 Estimation of the organization of VLCPUFA-containing ceramides in monolayers.

t1.3	Molecular species	Molecular dimension		LC phase (30 mN/m)			
t1.4		Acyl chains volume $(V)^a$ (\mathring{A}^2)	Acyl chain length $(L)^{b}$ (Å)	Semi-empirical thickness ^c (<i>Th</i>) (Å)	L/Th	$\mu_{\perp}^{\mathbf{d}}$ (mD)	
t1.5	n-28:4Cer	1292	22.7	16.6 ± 0.8	1.4 ± 0.1	832 ± 40	
t1.6	n-30:5Cer	1346	23.2	13.2 ± 0.8	1.8 ± 0.1	777 ± 25	
t1.7	n-32:5Cer	1400	24.3	11.2 ± 0.2	2.2 ± 0.1	1255 ± 10	
t1.8	h-28:4Cer	1292	22.7	22.0 ± 2.0	1.0 ± 0.1	360 ± 20	
t1.9	h-30:5Cer	1346	23.2	22.4 ± 0.4	1.0 ± 0.1	400 ± 18	
t1.10	h-32:5Cer	1400	24.3	14.4 ± 0.4	1.7 ± 0.1	668 ± 22	
t1.11	n – 16:0Cer	969	18.4	23.6 ± 0.8	0.8 ± 0.1	551 ± 33	
t1.12	n-24:1Cer	1185	22.0	28.2 ± 0.2	0.8 ± 0.1	463 ± 25	

- ^a Calculated as the sum of the average volumes of the two asymmetrical hydrophobic Cer chains, obtained after Eq. (3).
- ^b Calculated as the average of the two asymmetrical hydrophobic chain lengths, obtained after Eq. (4).
- 1.15 c Calculated as V/MMA.

t1.13 t1.14

444

445

446

447 448

449

450

451

452 453

454

455

456 457

458

459

460

461

462

463

464

465

 $466 \\ 467$

468

469 470

471

472

473

474

475

476 477

t2.1

t2.2

t2.11

t1.16 d Calculated after Eq. (2).

3.5. Thermodynamic analysis of VLCPUFA-containing ceramides in monolayers

Lowering the temperature from 21 °C to 8 °C resulted, as expected, in a decrease of the π_t of VLCPUFA-containing Cer films since the latter condition favors the LC regime; this was complete in the case of the species with the longest fatty acids, n-32:5 Cer and h-32:5Cer (Table 3). The collapse pressure was less affected by this temperature difference. The integrated area under the π versus MMA curve gives information of the free energy of the compression process [10,13]. Since this parameter involves the entropy lost upon the increase of molecular packing order in the monolayer under compression, it is expected to be smaller at lower temperatures, as a more condensed state is favored than at higher temperatures. These effects, as well as an increase of the free energy of compression with the acyl chain length, are observed in Table 3. This supports the logical implication that the longer and more unsaturated the Cer acyl chains are, the larger the entropy lost upon compression is

As a comparison, in early work [10] a free energy of compression of $\sim 0.15-0.25$ kcal mol⁻¹ was reported for bovine brain Cer in the temperature range studied here. Our own calculations give values of 0.3 ± 0.1 kcal mol⁻¹ for the more expanded 18:1 Cer and a very low value for the condensed 16:0 and 24:1 Cers (0.07 \pm 0.01 kcal mol⁻¹). These values are far lower than those found here for the VLCPUFA Cer species, even for monolayers that are completely in the LC state $(h-32:5 \text{ Cer and } n-32:5 \text{ Cer at } 8 ^{\circ}\text{C})$. Most interesting was the comparison between the n-V and h-V Cer species, as the n-V Cer species showed larger free energy values than their h-V counterparts at the two temperatures studied. The larger free energies of compression observed for the n-V Cer species were consistent not only with their larger MMA, but also with their more diffuse phase transitions (see Fig. 2). This may reflect a lower capacity for close packing of the n-V Cer than of the h-V Cer species (Fig. 3 and Table 1). It is important to recall that the free energy of compression, besides entropic contributions, also contains a component related to the enthalpy change associated to the phase transformation. Thus, the difference 478 may be a consequence of stronger interactions between the polar 479 headgroups, in addition to a tighter acyl chain packing, of the h-V 480 Cer than of the n-V Cer species.

One of the advantages of using the monolayer technique to study thermodynamic properties of lipids is that it requires small amounts of sample. The isolation in pure form of the Cer species containing nand h-VLCPUFAfrom a natural source by the method described above is laborious and may result in very scarce amounts of the rarer species. This largely limits the possibility of performing thermodynamic analysis of these molecules organized as bilayer structures. However, their behavior in Langmuir monolayers may shed some light into their possible behavior if organized as bulk dispersions.

As previously proposed [10] the lower temperature at which a com- 491 plete LE behavior is observed in monolayers (as obtained from the inter-492 cepts between the collapse pressure- and the π_t -temperature linear 493 regressions) should be roughly in keeping with the melting temperature 494 observed in bilayers (T_m). Thus, for a series of glycosphingolipids the 495 agreement between the $T_{\rm m}$ experimentally determined by differential $\,$ 496 scanning calorimetry (DSC) and that estimated from monolayer mea- 497 surements is remarkable [25]. On these bases, for 16:0 Cer suspended 498 in bulk aqueous solution, a T_m of 93.2 $\,^\circ\text{C}$ was previously reported using $\,^{499}$ DSC [5] and a T_m of 91.4 \pm 1.4 °C can be estimated from extrapolation 500 of monolayer transitions by the above-mentioned interception method 501 [9]. This quite close agreement (within about 2 °C) led us to estimate 502 an approximate T_m for the present VLCPUFA-containing Cer (Table 3). 503 This method suggested T_m values of 45–46 °C for n- and h – 28:4Cer, a 504 higher value for the h-30:5Cer (53 °C) and a lower value for its n-V 505 counterpart (36 °C).

As the n-V Cer is expected to undergo its phase transition at ~36 °C, 507 it should presumably be organized as a low curvature interface, this es- 508 timation positions the T_m of the n - 30:5Cer within the physiological 509 temperature range and the rest of the VLCPUFA-containing Cer analyzed 510 not far from such range. This suggests that in the cell membrane envi- 511 ronment subtle changes of temperature, and/or a drop in π due to 512

Table 3Thermodynamic parameters of VLCPUFA-containing ceramides in monolayer.

t2.3	Molecular species	21 °C			8 °C	_		
t2.4		$LE \rightarrow LC$ transition pressure (π_t) (mN/m)	Collapse pressure (mN/m)	Compression free energy ^a (kcal/mol)	$LE \rightarrow LC$ transition pressure (π_t) (mN/m)	Collapse pressure (mN/m)	Compression free energy ^a (kcal/mol)	π_{t} -T vs. collapse π -T intersection (°C)
t2.5	n – 28:4 Cer	15 ± 2	39 ± 1	0.99 ± 0.05	6 ± 1	44 ± 1	0.72 ± 0.01	45 ± 4
t2.6	n-30:5 Cer	23 ± 2	39 ± 2	1.64 ± 0.04	14 ± 1	45 ± 2	1.58 ± 0.05	36 ± 6
t2.7	n-32:5 Cer	16 ± 1	47 ± 1	1.90 ± 0.03	_	45 ± 1	1.53 ± 0.01	_
t2.8	h-28:4 Cer	15 ± 1	45 ± 2	0.72 ± 0.02	5 ± 1	48 ± 1	0.50 ± 0.01	46 ± 5
t2.9	h-30:5 Cer	19 ± 1	42 ± 2	0.78 ± 0.04	11 ± 1	45 ± 1	0.53 ± 0.01	53 ± 3
t2.10	h – 32:5 Cer	5 ± 2	44 ± 1	1.00 ± 0.07	-	41 ± 1	0.66 ± 0.05	-

^a Obtained by integrating the compression isotherms from 1 to 35 mN/m.

Please cite this article as: D.A. Peñalva, et al., Atypical surface behavior of ceramides with nonhydroxy and 2-hydroxy very long-chain (C28–C32) PUFAs, Biochim. Biophys. Acta (2013), http://dx.doi.org/10.1016/j.bbamem.2013.11.018

thermal fluctuation [33], could induce an LC \rightarrow LE phase transition of 513 these molecules, which, according to our results (Table 1), involves a 514 large molecular lateral expansion. Since the Cer polar headgroup is 515 516 very small (Fig. 1), such lateral expansion would concomitantly translate into changes of the overall geometry of the Cer molecules. As it 517 was previously proposed [20,24], the geometrical properties of lipids 518 are of paramount importance for the adoption of defined supramolecular structure. All Cers fall into the group of "inverted cone" shaped mol-520 521 ecules; as a general rule, this characteristic should favor negatively curved structures, some of which promote membrane fusion events 522 523 [17,25–27]. It remains to be evaluated if the long and bulk acyl chains of VLCPUFA-Cer may further enhance this effect compared to the 524 more common species. Studies in this direction will provide evidences 525 526 for a deeper understanding of the physiological importance of VLCPUFA-containing Cers during the acrosomal reaction, a process 527 that requires the involvement of structural lipid intermediates with 528 529 negative curvature.

Acknowledgments

530

531

532

533

534

535

536

537

538

539

540

541

542

543

544 545

546

547

549

550

551

553

554

555

556

557

558

559

560

561

562

563

564

565

566

567

568

569

570

571

572

573

574 575

576

577

578 579

580

663

This work was supported by FONCyT, CONICET, SECyT-Universidad Nacional de Córdoba, and SGCyT-Universidad Nacional del Sur (Argentina). FD, SSA, BM, MIA and MLF are career investigators and DAP is fellow of CONICET.

References

- [1] M.I. Aveldano, M. VanRollins, L.A. Horrocks, Separation and quantitation of free fatty acids and fatty acid methyl esters by reverse phase high pressure liquid chromatography, J. Lipid Res. 24 (1983) 83–93.
- E.G. Bligh, W.J. Dyer, A rapid method of total lipid extraction and purification, Can. J. Biochem. Physiol. 37 (1959) 911-917.
- H.L. Brockman, C.M. Jones, C.J. Schwebke, J.M. Smaby, D.E. Jarvis, Application of microcomputer-controlled film balance system to collection and analysis of data from mixed monolayers, J. Colloid Interface Sci. 78 (1980) 502-512.
- H.L. Brockman, M.M. Momsen, R.E. Brown, L. He, J. Chun, H.S. Byun, R. Bittman, The 4,5-double bond of ceramide regulates its dipole potential, elastic properties, and packing behavior, Biophys. J. 87 (2004) 1722-1731.
- J.V. Busto, M.L. Fanani, T.L. De, J. Sot, B. Maggio, F.M. Goni, A. Alonso, Coexistence of immiscible mixtures of palmitoylsphingomyelin and palmitoylceramide in monolayers and bilayers, Biophys. J. 97 (2009) 2717-2726.
- F. Dupuy, M.L. Fanani, B. Maggio, Ceramide N-acyl chain length: a determinant of bidimensional transitions, condensed domain morphology, and interfacial thickness, Langmuir 27 (2011) 3783-3791.
- [7] F. Dupuy, B. Maggio, The hydrophobic mismatch determines the miscibility of ceramides in lipid monolayers, Chem. Phys. Lipids 165 (2012) 615-629.
- M.L. Fanani, S. Hartel, B. Maggio, T.L. De, J. Jara, F. Olmos, R.G. Oliveira, The action of sphingomyelinase in lipid monolayers as revealed by microscopic image analysis, Biochim. Biophys. Acta 1798 (2010) 1309-1323.
- M.L. Fanani, B. Maggio, Phase state and surface topography of palmitoyl-ceramide monolayers, Chem. Phys. Lipids 163 (2010) 594-600.
- [10] G.D. Fidelio, B. Maggio, F.A. Cumar, Molecular parameters and physical state of neutral glycosphingolipids and gangliosides in monolayers at different temperatures, Biochim. Biophys. Acta 854 (1986) 231-239.
- N.E. Furland, G.M. Oresti, S.S. Antollini, A. Venturino, E.N. Maldonado, M.I. Aveldano, Very long-chain polyunsaturated fatty acids are the major acyl groups of sphingomyelins and ceramides in the head of mammalian spermatozoa, J. Biol. Chem. 282 (2007) 18151-18161.
- N.E. Furland, S.R. Zanetti, G.M. Oresti, E.N. Maldonado, M.I. Aveldano, Ceramides and sphingomyelins with high proportions of very long-chain polyunsaturated fatty acids in mammalian germ cells, J. Biol. Chem. 282 (2007) 18141-18150.
- G.L. Gaines, Insoluble Monolayers at Liquid-Gas Interfaces, Interscience Publishers, New York, 1966.
- H. Hama, Fatty acid 2-hydroxylation in mammalian sphingolipid biology, Biochim. Biophys. Acta 1801 (2010) 405-414.
- Y.A. Hannun, L.M. Obeid, Many ceramides, J. Biol. Chem. 286 (2011) 27855-27862. S. Hartel, M.L. Fanani, B. Maggio, Shape transitions and lattice structuring of ceramide-enriched domains generated by sphingomyelinase in lipid monolayers,
- I.M. Holopainen, M.I. Angelova, P.K. Kinnunen, Vectorial budding of vesicles by asymmetrical enzymatic formation of ceramide in giant liposomes, Biophys. J. 78 (2000) 830-838.

Biophys. J. 88 (2005) 287-304.

- [18] J.M. Holopainen, H.L. Brockman, R.E. Brown, P.K. Kinnunen, Interfacial interactions of 581 ceramide with dimyristoylphosphatidylcholine: impact of the N-acyl chain, Biophys. 582 I. 80 (2001) 765-775.
- [19] I.M. Holopainen, M. Subramanian, P.K. Kinnunen, Sphingomyelinase induces lipid 584 microdomain formation in a fluid phosphatidylcholine/sphingomyelin membrane, 585 Biochemistry 37 (1998) 17562-17570.

586

588

592

593

594

608

- I.N. Israelachvili, S. Marcelja, R.G. Horn, Physical principles of membrane organiza-587 tion, O. Rev. Biophys. 13 (1980) 121-200.
- [21] C. Lheveder, I. Meunier, S. Henon, Brewster angle microscopy, in: A. Baszkin, W. 589 Norde (Eds.), Physical Chemistry of Biological Interfaces, Marcel Dekker, Inc., NY, 590 2000
- X.M. Li, I.M. Smaby, M.M. Momsen, H.L. Brockman, R.E. Brown, Sphingomyelin interfacial behavior: the impact of changing acyl chain composition, Biophys. J. 78 (2000) 1921-1931
- H. Lofgren, I. Pascher, Molecular arrangements of sphingolipids. The monolayer be-595 haviour of ceramides, Chem. Phys. Lipids 20 (1977) 273-284. 596
- [24] B. Maggio, Geometric and thermodynamic restrictions for the self-assembly of 597 glycosphingolipid-phospholipid systems, Biochim. Biophys. Acta 815 (1985) 598 245-258
- [25] B. Maggio, The surface behavior of glycosphingolipids in biomembranes: a new 600 frontier of molecular ecology, Prog. Biophys. Mol. Biol. 62 (1994) 55-117. 601
- [26] B. Maggio, J. Albert, R.K. Yu, Thermodynamic-geometric correlations for the mor- 602 phology of self-assembled structures of glycosphingolipids and their mixtures 603 with dipalmitoylphosphatidylcholine, Biochim. Biophys. Acta 945 (1988) 145-160. 604
- [27] B. Maggio, F.A. Cumar, R. Caputto, Induction of membrane fusion by 605 polysialogangliosides, FEBS Lett. 90 (1978) 149-152. 606 607
- H.M. McConnell, Structures and transitions in lipid monolayers at the air-water interface, Annu. Rev. Phys. Chem. 42 (1991) 171-195.
- G.M. Oresti, P.L. Ayuza Aresti, G. Gigola, L.E. Reyes, M.I. Aveldano, Sequential depletion of rat testicular lipids with long-chain and very long-chain polyenoic fatty acids 610 after X-ray-induced interruption of spermatogenesis, J. Lipid Res. 51 (2010) 611 2600-2610.
- [30] G.M. Oresti, J.M. Luquez, N.E. Furland, M.I. Aveldano, Uneven distribution of 613 ceramides, sphingomyelins and glycerophospholipids between heads and tails of 614 rat spermatozoa, Lipids 46 (2011) 1081-1090.
- D.A. Peñalva, N.E. Furland, G.H. Lopez, M.I. Aveldaño, S.S. Antollini, Unique thermal behavior of sphingomyelin species with nonhydroxy and 2-hydroxy 617 very-long-chain (C28-C32) polyunsaturated fatty acids, J. Lipid Res. (2013)
- [32] M.A. Perillo, A. Guidotti, E. Costa, R.K. Yu, B. Maggio, Modulation of phospholipases 619 A2 and C activities against dilauroylphosphorylcholine in mixed monolayers with 620 semisynthetic derivatives of ganglioside and sphingosine, Mol. Membr. Biol. 11 621 (1994) 119-126.
- M.C. Phillips, The physical state of phospholipid and cholesterol in monolayers, bilayers, and membranes, in: J.F. Danielli, M.D. Rosenberg, D.A. Cadenhead (Eds.), Progress in Surface and Membrane science, Academic Press, New York, 1972, 625
- [34] S.N. Pinto, L.C. Silva, A.H. Futerman, M. Prieto, Effect of ceramide structure on membrane biophysical properties: the role of acyl chain length and unsaturation, Biochim, Biophys. Acta 1808 (2011) 2753-2760.
- [35] A. Poulos, D.W. Johnson, K. Beckman, I.G. White, C. Easton, Occurrence of unusual 630 molecular species of sphingomyelin containing 28-34-carbon polyenoic fatty acids 631 in ram spermatozoa, Biochem. J. 248 (1987) 961–964.
- M. Rabionet, A.C. van der Spoel, C.C. Chuang, B. von Tumpling-Radosta, M. Litjens, D. 633 Bouwmeester, C.C. Hellbusch, C. Korner, H. WIEGANDT, K. Gorgas, F.M. Platt, H.J. 634 Grone, R. Sandhoff, Male germ cells require polyenoic sphingolipids with complex glycosylation for completion of meiosis: a link to ceramide synthase-3, J. Biol. Chem. 283 (2008) 13357-13369.
- [37] B.S. Robinson, D.W. Johnson, A. Poulos, Novel molecular species of sphingomyelin 638 containing 2-hydroxylated polyenoic very-long-chain fatty acids in mammalian testes and spermatozoa, J. Biol. Chem. 267 (1992) 1746-1751.
- [38] M.B. Ruiz-Arguello, M.P. Veiga, J.L. Arrondo, F.M. Goni, A. Alonso, Sphingomyelinase 641 cleavage of sphingomyelin in pure and mixed lipid membranes. Influence of the physical state of the sphingolipid, Chem. Phys. Lipids 114 (2002) 11-20. 643 644
- R. Sandhoff, Very long chain sphingolipids: tissue expression, function and synthesis, FEBS Lett. 584 (2010) 1907-1913. 645
- R. Sandhoff, R. Geyer, R. Jennemann, C. Paret, E. Kiss, T. Yamashita, K. Gorgas, T.P. 646 Sijmonsma, M. Iwamori, C. Finaz, R.L. Proia, H. Wiegandt, H.J. Grone, Novel class 647 of glycosphingolipids involved in male fertility, J. Biol. Chem. 280 (2005) 27310-27318. 649
- [41] J.M. Smaby, V.S. Kulkarni, M. Momsen, R.E. Brown, The interfacial elastic packing interactions of galactosylceramides, sphingomyelins, and phosphatidylcholines, 651 Biophys. I. 70 (1996) 868-877. 652
- [42] J.M. Smaby, M.M. Momsen, H.L. Brockman, R.E. Brown, Phosphatidylcholine acyl 653 unsaturation modulates the decrease in interfacial elasticity induced by cholesterol, 654 Biophys. J. 73 (1997) 1492-1505. 655
- S.R. Zanetti, M.M. de Los Angeles, D.E. Rensetti, M.W. Fornes, M.I. Aveldano, 656 Ceramides with 2-hydroxylated, very long-chain polyenoic fatty acids in rodents: 657 From testis to fertilization-competent spermatozoa, Biochimie 92 (2010) 658 1778-1786. 659
- [44] S.R. Zanetti, M.L. Monclus, D.E. Rensetti, M.W. Fornes, M.I. Aveldano, Differential in-660 volvement of rat sperm choline glycerophospholipids and sphingomyelin in capac-661 itation and the acrosomal reaction, Biochimie 92 (2010) 1886-1894. 662