Author's Accepted Manuscript

Antihistaminergics and inverse agonism. Potential therapeutic applications

Federico Monczor, Natalia Fernandez, Carlos P. Fitzsimons, Carina Shayo, Carlos Davio



www.elsevier.com/locate/ejphar

PII: S0014-2999(13)00496-2 DOI: 10.1016/j.ejphar.2013.06.027

Reference: EJP68687

To appear in: European Journal of Pharmacology

Received date: 27 February 2013

Revised date: 7 June 2013 Accepted date: 21 June 2013

Cite this article as: Federico Monczor, Natalia Fernandez, Carlos P. Fitzsimons, Carina Shayo, Carlos Davio, Antihistaminergics and inverse agonism. Potential therapeutic applications, European Journal of Pharmacology, 10.1016/j.ejphar.2013.06.027

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting galley proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Antihistaminergics and inverse agonism. Potential therapeutic applications.

Federico Monczor^{1,4}, Natalia Fernandez^{1,4}, Carlos P. Fitzsimons², Carina Shayo^{3,4} and Carlos Davio^{1,4}.

From Laboratorio de Farmacología de Receptores, Cátedra de Química Medicinal, Facultad de Farmacia y Bioquímica, Universidad de Buenos Aires ¹, Center for Neuroscience, Swammerdam Institute for Life Sciences, University of Amsterdam, the Netherlands ², Laboratorio de Farmacología y Patología Molecular, Instituto de Biología y Medicina Experimental ³; and Consejo Nacional de Investigaciones Científicas y Técnicas, Buenos Aires, Argentina ⁴

Running head: G-protein sequestering by GPCR inverse agonists.

Address correspondence to: Federico Monczor, PhD, Junin 956 PP (1113), Buenos Aires, Argentina. Tel: +54-11-4964-8233, E-mail: monczorf@ffyb.uba.ar

Abstract

The accurate characterization of the molecular mechanisms involved in the action of receptor ligands is important for their appropriate therapeutic use and safety.

It is well established that ligands acting at the histamine system currently used in the clinic exert their actions by specifically antagonizing G-protein coupled H₁ and H₂ receptors. However, most of these ligands, assumed to be neutral antagonists, behave as inverse agonists displaying negative efficacy in experimental systems. This suggests that their therapeutic actions may involve not only receptor blockade, but also the decrease of spontaneous receptor activity.

The mechanisms whereby inverse agonists achieve negative efficacy are diverse. Theoretical models predict at least three possible mechanisms, all of which are supported by experimental observations. Depending on the mechanism of action engaged, the inverse agonist could interfere specifically with signaling events triggered by unrelated receptors. This possibility opens up new venues to explain the therapeutic actions of inverse agonists of the histamine receptor and perhaps new therapeutic applications.

Keywords: Histamine receptors, antihistaminergics, inverse agonism, adverse effects.

1. Histamine receptors and its role in human health.

Histamine is a biogenic amine synthesized from L-histidine by histidine decarboxylase. This amine plays an important role in human health and disease. It exerts its biologic effects acting through four receptor subtypes, termed H₁, H₂, H₃, and H₄. These receptors differ in their expression, molecular characteristics, resulting in differences in associated signal transduction pathways and function (Hill et al., 1997; Leurs et al., 2009). Nevertheless, all types of histamine receptors belong to the seven transmembrane spanning family of receptors, also known as GPCRs.

The histamine H_1 receptor is in fact the classical histamine receptor. This is one of many receptors that influence cellular homeostasis by modifying phospholipase C activity via interactions with Gproteins from the $G\alpha q/11$ family. Following the original observations of Hokin and Hokin in 1953 (Hokin and Hokin, 1953), histamine-induced incorporation of ^{32}P in phosphatidylinositol was first reported in 1975 in the rat brain *in vivo* (Friedel and Schanberg, 1975). In a seminal study in the same system, ligand binding with specific H_1 receptor ligands paralleled the development of histamine-induced incorporation of ^{32}P into phosphoinositides, demonstrating for the first time histamine-induced activation of phospholipase C (Subramanian et al., 1981). Currently, H_1 receptor-induced phosphoinositide responses mediated by $G\alpha q/11$ activation have been described in a variety of central and peripheral tissues and in isolated cell systems as well (Smit et al., 1999).

The observation that histamine-evoked gastric acid secretion could not be blocked with classical

antihistamines (H_1 antagonists), led to the conclusion that other histamine receptors were involved. The receptor involved in mediating histamine-dependent gastric acid secretion was subsequently termed histamine H_2 receptor (Black et al., 1972). It is generally accepted that H_2 receptor is coupled to adenylyl cyclase through $G\alpha$ s. However, it has been reported that the H_2 receptor can also activate phospholipase C (Davio et al., 1995; Davio et al., 2002) and perhaps both pathways simultaneously (Delvalle et al., 1992; Wellner-Kienitz et al., 2003). In fact, the activation of signal transduction pathways is highly dependent on the cellular system under study and outcome of H_2 receptor activation is probably mediated by various intracellular mediators.

A third histamine receptor, H_3 , was originally postulated in 1983 as a putative histamine autoreceptor responsible for controlling histamine release in rat cerebral cortical slices (Arrang et al., 1988). Since then, the H_3 receptor has been shown to be coupled to modulation of adenylyl cyclase activity via $G\alpha i/0$ activation, and has been associated to inhibition of neurotransmission in central (Clapham and Kilpatrick, 1992; Schlicker et al., 1993; van der Werf and Timmerman, 1989) and peripheral tissues (Burgaud and Oudart, 1993; Imamura et al., 1995; Ishikawa and Sperelakis, 1987).

More recently, a fourth histamine receptor has been described. The H₄ receptor is also a GPCR and it is coupled to Gαi/0, resulting in modulation of various signal transduction pathways (Oda et al., 2000). The H₄ receptor is prominently expressed in medullary and peripheral hematopoietic cells, namely eosinophils, neutrophils and CD4⁺ T cells (Zhu et al., 2001). The H₄ receptor is involved in histamine-induced increases in intracellular calcium in human eosinophils (Raible et al., 1994), facilitating terminal myeloblast and promonocyte differentiation (Nakaya and Tasaka, 1988).

Until now, the most clinically relevant uses of histamine receptor ligands are achieved through the regulation of H₁ or H₂ receptors, which are widely expressed in most tissues (Bakker et al., 2002). In this regard, H₁ antihistamines are used in the treatment of several allergic conditions, such as rhinoconjunctivitis, urticaria, and atopic dermatitis. Traditionally, the efficacy of H₁ antihistamines in allergic disorders was attributed primarily to inhibition of H₁ receptors in endothelial cells of the postcapillary venules, resulting in decreased vascular permeability, exudation of fluid, protein and cells and increased peripheral resistance (Leurs et al., 2002). Nevertheless, H₁ antagonists also inhibit the effect of histamine at H₁ receptors in the airway smooth muscle and on afferent C fibers, causing bronchodilation and decreased mucosal and cutaneous itch, respectively.

H₂ antagonists have proved to be very active agents for the treatment of duodenal and gastric ulcers, reflux, esophagitis and the Zollinger-Ellison syndrome. Since H₂ receptors are abundant throughout the body, it is rather remarkable that H₂ antagonists have not found other applications. Several studies have claimed beneficial effects of H₂ antagonists in, e.g. viral diseases and alleviation of psoriasis (Nielsen, 1991), with no conclusive results.

The identification of H₃ and H₄ receptors and their pattern of expression has lead to a renewed interest in the potential homeostatic role of histamine in the brain ,, besides their established pleiotropic regulatory functions in the periphery. Although no central nervous system disease has been associated directly to histamine dysfunction until now, the H₃ receptor is recognized as a drug target for neuropathic pain, sleep-wake disorders, including narcolepsy, and cognitive impairment associated with attention deficit hyperactivity disorder, schizophrenia, Alzheimer's, or Parkinson's disease and the first specific H₃ receptor ligands have already entered phase I–III clinical trials (Tiligada et al., 2011). However, before their introduction in the market, H₃R ligands may require further testing to identify the determinants of their clinical efficacy.

Nowadays, more than forty H₁ antihistamines are therapeutically available worldwide. However, some of them, such as astemizole and terfenadine, have been associated with cardiac toxic effects

and are no longer approved for use in most countries (Yap and Camm, 2002). On the other hand, H_2 antagonists, widely used as antiacids, were overcome by proton-pump inhibitors, due to their more favorable efficacy and safety profiles. However, a recent survey revealed that between the twenty most commonly used prescription and over-the-counter drugs are antiallergic formulations acting at H_1 (difenhydramine and loratadine), and antiacids acting at H_2 receptors (ranitidine) (Kaufman et al., 2002).

From the mechanistical point of view, it has been proven *in vitro* that clinically relevant histamine receptor ligands do not act as simple antagonists, but rather as inverse agonists stabilizing inactive forms of the H₁ and H₂ receptors (Leurs et al., 2002; Smit et al., 1998). These observations have lead to the pharmacological reclassification of histamine H₁ and H₂ specific antagonists. The concept of inverse agonism arose from experimental observations showing that certain drugs were able to reduce the activity of receptor systems that were active in the absence of agonists. The inverse agonists possess, in consequence, negative efficacy. According to existing receptor occupancy models, there are several mechanisms whereby inverse agonists exert their negative efficacy, and each one may lead to distinct consequences. However, whether inverse agonism is essential or important for these drugs to exert their medicinal actions has not been clarified yet. This point deserves attention due to the widespread use of histamine receptor ligands reclassified as inverse agonists in clinical treatments. In the following sections we will discuss these aspects of clinically used histamine receptor ligands in the context of their pharmacological safety.

2. Spontaneous receptor activity. The rise of the inverse agonism concept.

Due to the recognition that many receptors are able to spontaneously activate downstream effector protein (reference) pharmacology experienced a rebirth in the understanding of the molecular mechanisms triggered by molecules that modulate receptor function. As a result, there was a significant realization of the importance of explicitly formulating spontaneous receptor activity observed both in natural and artificial systems (Costa and Cotecchia, 2005; Kenakin, 1995). This realization led to the formulation of the two-state model (TSM) (Leff, 1995; Robertson et al., 1994) and related reaction schemes developed later (Bindslev, 2004; Hall, 2000; Weiss et al., 1996a; Weiss et al., 1996b; Weiss et al., 1996c).

Increasing numbers of compounds previously classified as antagonists were shown to inhibit this spontaneous or constitutive receptor activity (Kenakin, 2004). These thought-to-be antagonists with negative efficacy are now known as 'inverse agonists', whereas compounds that antagonize the inhibitory effect of agonists and inverse agonists without having an effect of their own are still termed antagonists, or more precisely neutral antagonists (Chidiac, 2002; Kenakin, 2004; Milligan et al., 1995; Strange, 2002).

Classically, efficacy (whether positive, negative or zero) was thought as a separate property unrelated to affinity. However, in thermodynamic terms, this representation presents a paradox because the molecular forces that control affinity are the same as those controlling efficacy (Kenakin and Onaran, 2002). Taking this into account, it is not surprising that 85% of the ligands formerly known as antagonists have been shown to posses negative efficacy (Kenakin, 2004).

3. Theoretical models of ligand-receptor occupancy and the mechanisms of inverse agonism. Receptor activity measured in functional studies can be either basal (i.e. spontaneous, constitutive and ligand-independent), or agonist-induced. This experimental observation is independent of the model used to interpret the system. However, different spontaneous receptor conformations can be hypothesized to be responsible for the basal activity, comprising a "receptor native ensemble". Hence, as we are going to discuss in the next paragraph, the uniformity of the concept of constitutive receptor activity is apparently challenged when the spontaneous receptor species (and

therefore the source of the receptor basal activity) are explicitly formulated (Kenakin, 2002; Onaran and Costa, 1997) (Fig. 1).

For the TSM, receptors can spontaneously adopt only two forms, the resting or inactive state (R), and the active one (R*), to which the basal activity of the system is formally attributed. However, when accessory proteins (G-proteins) are included in the models, as in the extended ternary complex model (ETCM) (Samama et al., 1993), the native ensemble involves three distinct receptor forms including the inactive (R) and the active species (R*), but also an active G-protein coupled receptor form that is considered responsible for basal activity (R*G). Similarly, the cubic ternary complex model (CTCM) (Weiss et al., 1996a; Weiss et al., 1996b; Weiss et al., 1996c) adds one more receptor form to the native ensemble, allowing receptor to couple to G-

protein in an inactive form (RG).

The CTCM was originally proposed in an attempt to explore the mathematical and pharmacological implications that can be derived from permitting G-proteins to interact with receptors in their inactive and active forms irrespectively. However, it is worth noting that regardless of its theoretical origins, there are empirical observations supporting the CTCM. Some mutants of α_{1B} -adrenergic receptors are able to spontaneously couple to G-protein inactivating it (Chen et al., 2000), and this has also been observed for non-mutant serotonin 5HT₇ and histamine H₂ receptors (Andressen et al., 2006; Tubio et al., 2010).

The three models and their relations are schematically shown in Fig. 2. The fact that these models assume differences in the receptor states that exist spontaneously, implies these models could be used to explain ligand-dependent selection and stabilization of different preexisting receptor conformations. According to the law of mass action, when a factor affects previously established equilibriums, the receptor species are redistributed to re-establish equilibrium. Consequently, for all the models, inverse agonists exert their effect favoring the inactive receptor species at the expense of the active ones. However, the particular receptor species favored changes depending on the model. In this way, according to the TSM, inverse agonists suppress the spontaneous activity of the receptors by stabilizing them in an inactive state. Similarly, in the ETC model, inverse agonists may act by preferentially binding to the R state over the R* state (α <1). In both cases, R*G species will be depleted as more receptor transforms into ligand-bound inactive R (LR), resulting in a decreased constitutive activity. There are several experiments that confirm this model, at least for the α_2 - and β_2 -adrenergic receptors (Samama et al., 1993; Wade et al., 2001). Additionally, for the ETCM, ligands could bind to uncoupled states of the receptor (R and R*) in preference to the coupled state (R*G) (γ<1). This feature was described for 5-HT_{1A} and 5-HT_{2C} receptors, cardiac muscarinic receptors (M₂ receptors), and dopamine D₂ receptors (De Lean et al., 1982; Martin et al., 1984; McLoughlin and Strange, 2000; Westphal and Sanders-Bush, 1994).

Another possibility, only accounted for in the CTCM, is that inverse agonists bias the receptor to a G-protein coupled but inactive conformation of the receptor (RG). It is worth noting that in the ETC model a ligand with high affinity for receptor species coupled to G-proteins would necessarily elicit a response. In contrast, the CTC model allows a ligand with high affinity for a receptor species coupled to G-protein to behave as an inverse agonist.

For the ETC model a ligand that facilitates receptor activation (α >1), promotes that the active receptor has more affinity for G-protein than the inactive form (β >1), and improves G-protein coupling (γ >1), is defined as an agonist. However, a negative cooperativism among those receptor modifications may exist in the sense that ligands might behave as inverse agonists. This point is a distinctive feature of the CTC model and is made possible by proposing the δ parameter, which represents the synergism among the receptor modifications (activation, binding to ligand, or coupling to G-protein). Therefore in the conceptual frame of the CTC it can be theoretically

predicted, and then empirically proved, that an inverse agonist can exert its effect by stabilizing a G-protein coupled but inactive form of the receptor. Consequently, it can be inferred that if the G-protein is in limiting quantities, the ligand will be able to interfere with the signaling of other unrelated GPCRs that share the same signaling cascade. This effect can be interpreted in terms of a G-protein "molecular kidnapping", mediated by the inverse agonist-bound receptor (Fig. 3). This theory explains some experimental observations made not only for ligands acting at H_1 and H_2 histamine receptors (Fitzsimons et al., 2004; Monczor et al., 2003), but also for ligands acting at μ -opioid and CB_1 cannabinoid receptors, that interfere with the signaling of other related receptors (Bouaboula et al., 1997; Brown and Pasternak, 1998). This view can also take account of some striking results obtained for D_2 dopamine and M_3 muscarinic receptor ligands, showing that an inverse agonist can exert its effects without promoting the expected receptor G-protein uncoupling (Dowling et al., 2006; Wilson et al., 2001).

This interference of an inverse agonist of a given GPCR on the signaling of a non related receptor highlights the importance of the study of the mechanistic basis of action of ligands with negative efficacy. Taking into account that this may be a generalized feature of several ligands with clinical uses, a deeper understanding of this phenomenon could help to rationalize otherwise unexpected drug effects. Regarding this, the current as well as potential uses of inverse agonists of the histaminergic receptors in clinical treatment are discussed in the next section.

4. Potential clinical uses of inverse agonists of the histaminergic receptors.

When inverse agonism was first described, there were concerns on whether it was of therapeutic relevance in vivo or just a curious observation induced by laboratory conditions (Milligan et al., 1995). Since then, hundreds of well-known drugs of extensive clinical use have been tested and reclassified for this property. However, questions on the therapeutic relevance of inverse agonism remain.

Prominent examples of extensively used drugs formerly reclassified as antagonists and currently reclassified as inverse agonist due to their negative efficacy in vitro include: propranolol, alprenolol, pindolol, and timolol used for treating hypertension, angina pectoris, and arrhythmia acting on the β_2 -AR; metoprolol and bisoprolol used for treating hypertension, coronary heart disease, and arrhythmias acting on β_1 -AR; cetirizine and loratadine for treating allergies and hay fever acting on histamine H_1 receptor; cimetidine and ranitidine used to treat heartburn and peptic ulcer acting on the histamine H_2 receptor; prazosin and phentolamine for treating hypertension acting on α_1 -adrenergic receptors; atropine used as premedication for anesthesia and pirenzepine used for treatment of peptic ulcer acting on the muscarinic receptors and naloxone used for treating heroin overdose acting on μ -opioid receptor (Kenakin, 2004).

At a first glance, it may seem there is no theoretical or practical reasons to believe that inverse agonists should have an intrinsic benefit over an antagonist in many situations where drugs are clinically used. In general, drugs are simply used to regulate the release of an endogenous agonist or to compete with it and limit its actions. Therefore potency, bioavailability and selectivity seem the key issues to consider. However, in a range of circumstances, specific benefits of using an inverse agonist can be envisaged.

A first requirement to conceive an advantage in using an inverse agonist over an antagonist is that the receptor displays constitutive activity *in vivo*. In this sense, the constitutive activity of GPCRs in native *in vivo* systems has been proved only for few receptors, specifically including H₂ and H₃ histamine receptors, acting at CNS. The H₂ receptor was shown to display spontaneous activity in the *sustancia nigra* region and this activity regulated serotonin independently of the local histamine tone. This observation has implications not only for Parkinson's disease, where histamine levels in the *sustancia nigra* are increased, but also for other neuropsychiatric disorders in which serotonin is pivotal (Threlfell et al., 2008). In turn, constitutive activity of native H₃ receptors was shown in

rodent brain controlling histamine-dependent neuronal activity *in vivo*. The activation of histaminergic neurons, which promotes arousal and attention and improves learning in normal animals, has been proposed as a symptomatic therapeutic approach in human attentional and ageing disorders, and such an effect is more likely to be obtained with H₃ receptor inverse agonists rather than with neutral antagonists (Morisset et al., 2000).

On the other hand, there are clinical instances where the pathological entity is a constitutively active GPCR, which produces physiological response in the absence of endogenous agonists. In these circumstances, the underlying condition may be only effectively treated with inverse agonists. Mutations, which may be preserved in the germ line, have been shown to occur in GPCRs and result in constitutive receptor activity in patients with clinical syndromes, such as Jansen's metaphyseal condrodysplasia or congenital hyperthyroidism (Bastepe et al., 2004; Davies et al., 2005; Seifert and Wenzel-Seifert, 2002). Most of these diseases are associated with the endocrine system, although visual alterations due to rhodopsin mutations (Dryja et al., 1993; Keen et al., 1991) and some viral infections were also related to constitutively active forms of GPCRs (Arvanitakis et al., 1997; Fitzsimons et al., 2006; Maussang et al., 2009).

There are also many examples where inverse agonists may be useful for cancer treatment. For instance, high levels of specific GPCR expression have been described in tumour cells, where it has been shown that their endogenous ligands, present at high levels in the tumour cells (el Battari et al., 1988; Gespach et al., 1988; Korman et al., 1986; Virgolini et al., 1994), enhance cell proliferation (Kroog et al., 1995; Pincus et al., 1990; Zurier et al., 1988). The histamine H₂ receptor represents one interesting example of this property (Davio et al., 1995; Fitzsimons et al., 1999; Molnar et al., 2001). Interestingly, some early studies made in rats bearing experimental mammary adenocarcinomas showed that anti H₂ ligand administration resulted in the remission of a significant number of tumours, while also increased the survival of treated animals (Cricco et al., 1993; Rivera et al., 1993). Moreover, diverse clinical reports suggest that H₂ antagonists/inverse agonists have potential beneficial effects in the treatment of advanced malignant diseases such as colorectal cancer, gastric cancer, liver metastasis, multiple myeloma, chronic lymphocytic leukaemia and melanoma (Burtin et al., 1988; Nielsen, 1996; Nielsen and Kikuchi, 1993). In those cases, inverse agonists would block not only the effects of humoral activation in cancer cells (i.e. secreted histamine) but also increased basal activity of these receptors in the tumour due to receptor overexpression.

The studies summarized above provide supporting evidences for the relevance of pathological constitutive activity, suggesting the potential therapeutic values of its modulation. However, these potential advantages need to be tested for correlation with clinically relevant beneficial or detrimental effects. For this reason, and two decades after the discovery of inverse agonism on GPCRs, we have just begun exploring evidence of the clinical relevance of inverse agonists. However, it is worth noting that none of the above described diseases that involve pathological consequences of a constitutively active GPCR are in fact treated with inverse agonists. As suggested by Kenakin, translational research of inverse agonism will show whether new therapeutic options will emerge (Kenakin, 2004), especially for silencing the constitutively active mutant receptors that cause the diseases described above.

Up to this point, we have described only examples of desirable acute effects of inverse agonists. Alternatively, inverse agonists may require chronic administration and therefore exert some undesirable effects. For instance, receptor up-regulation could occur as it was observed for the H_2 receptor after long-term treatment with cimetidine or ranitidine (Osawa et al., 2005; Smit et al., 1996), providing a plausible explanation for the tolerance to these inverse agonists after prolonged clinical use. In the same way, Milligan and colleagues showed that inverse agonists also up-regulate β_2 - and α_{1B} -ARs (Georgieva et al., 2008; Lee et al., 1997; MacEwan and Milligan, 1996).

Recently, it has been shown that the CB_1 receptor inverse agonist rimonabant suppresses receptor constitutive activity in the ventral tegmental area and basolateral amygdala, reducing weight gain and food intake, but also causing anxiety and reduced motivation for reward, while NESS0327, acting as a neutral antagonist on the same receptor, is equally effective on reducing weight gain and food intake, but lacks the negative effects associated with rimonabant's inverse agonism. Considering that rimonabant has been discontinued for human treatment because its use was occasionally associated with negative effects and suicidality, the findings suggest that neutral CB_1 receptor antagonists can treat obesity more efficiently and safely than inverse agonists (Meye et al., 2012).

In conclusion, in this review we not only emphasize the importance of proper classification of ligands of constitutively active receptors, but also the importance of elucidating the molecular mechanisms by which these ligands exert their actions in terms of their clinical applications and pharmacological effects. An inverse agonist may act by three different ways: biasing the system to an inactive form of the receptor; facilitating the G-protein uncoupled forms; or promoting a G-protein coupled but inactive form of the receptor. If the preferred mechanism for a given ligand would be the third one, it may not only diminish the activity of the specific receptor but it may also interfere with the signalling of other non-related GPCRs. This is the case for some cannabinoid CB_1 (Bouaboula et al., 1997; Georgieva et al., 2008), μ opioid (Brown and Pasternak, 1998) and dopamine D_2 receptor (Wilson et al., 2001) ligands and, of central interest for this review, also for H_1 and H_2 ligands (Fitzsimons et al., 2004; Monczor et al., 2003).

This interference with the signaling of other GPCRs, may bring some unexpected effects that would not be observed with inverse agonists that do not promote G-protein sequestration, and could rationalize the appearance of pharmacological interactions and/or side effects that would be otherwise difficult to explain.

Ligands acting at histaminergic system are among the most widely prescribed and over-the-counter-sold drugs in the world, and there is a trend to use them as long-term therapeutics rather than restricting them to the treatment of short-term manifestations. Within this context, progress in the understanding of their mechanism of action is of crucial importance to improve their safety and specificity. Taking into account the clinically widespread use of histamine antagonists acting at H_1 and H_2 receptors in the treatment of several human diseases, the proper pharmacological classification of these ligands and the accurate characterization of their mechanism(s) of action is of great importance.

Legends for figures

Figure 1.

The figure shows the theoretical receptor species available for ligand binding. Inactive receptor conformations are presented on the left and the active ones on the right of the equilibrium arrows. Depending on the specific ligand-receptor interactions the ligand can redistribute the spontaneous receptor conformations, thereby favoring or disfavoring basal activity. This action of the ligand results from binding preferentially inactive or active species, or both, depending on whether the drug is an agonist, an inverse agonist, or eventually behaves as a neutral antagonist not affecting the spontaneous receptor activity.

Figure 2.

The figure shows the three most common models used to describe GPCR behaviors. The models describe inactive receptor species (R) in spontaneous equilibrium with an active state (R*) responsible for the basal activity of the system. In the extended ternary complex model (center), activation of the receptor could be followed by binding of R* to the G-protein (G). The cubic ternary complex model (right) implies the same asumption, but also allows the ligand-bound or unbound inactive-state receptor species (LR or R) to form a non-signaling complex with the G-protein (LRG or RG, respectively). Dotted arrows point to species that could be favored by inverse agonists, diminishing receptor basal activity. It should be noted that there is a trend in the complexity where following scheme contains the previous, and that the liganded inactive receptor species are different for each model, implying different mechanisms of action accounting for inverse agonism. These different mechanisms are discussed in the main text.

Figure 3.

The figure represents how an inverse agonist, acting on a specific receptor species and promoting the coupling to the G-protein to a receptor inactive state (LRG complex, see figure 2) can interfere with the signaling of a second unrelated receptor that transduces its signal through the same G-protein. The potential implications of such interference are discussed in the main text.

References

- Andressen, K. W., Norum, J. H., Levy, F. O., Krobert, K. A., 2006. Activation of adenylyl cyclase by endogenous G(s)-coupled receptors in Human Embryonic Kidney 293 cells is attenuated by 5-HT(7) receptor expression. Mol Pharmacol 69, 207-215.
- Arrang, J. M., Devaux, B., Chodkiewicz, J. P., Schwartz, J. C., 1988. H3-receptors control Histamine release in human brain. J Neurochem 51, 105-108.
- Arvanitakis, L., Geras-Raaka, E., Varma, A., Gershengorn, M. C., Cesarman, E., 1997. Human herpesvirus kshv encodes a constitutively active G-protein-coupled receptor linked to cell proliferation. Nature 385, 347-350.
- Bakker, R. A., Timmerman, H., Leurs, R. . Histamine receptors: specific ligands, receptor biochemistry, and signal transduction. In *Histamine and H1-antihistamines in allergic disease*. Simons FER (Ed.). 2002. 27-64.
- Bastepe, M., Raas-Rothschild, A., Silver, J., Weissman, I., Wientroub, S., Jüppner, H., Gillis, D., 2004. A form of Jansen's metaphyseal chondrodysplasia with limited metabolic and skeletal abnormalities is caused by a novel activating parathyroid hormone (PTH)/PTH-related peptide receptor mutation. J Clin Endocrinol Metab 89, 3595-3600.
- Bindslev, N., 2004. A homotropic two-state model and auto-antagonism. BMC Pharmacol 4, 11. Black, J. W., Duncan, W. A., Durant, C. J., Ganellin, C. R., Parsons, E. M., 1972. Definition and antagonism of Histamine H2 -receptors. Nature 236, 385-390.
- Bouaboula, M., Perrachon, S., Milligan, L., Canat, X., Rinaldi-Carmona, M., Portier, M., Barth, F., Calandra, B., Pecceu, F., Lupker, J., 1997. A selective inverse agonist for central cannabinoid receptor inhibits mitogen-activated protein kinase activation stimulated by insulin or insulin-like growth factor 1. Evidence for a new model of receptor/ligand interactions. J Biol Chem 272, 22330-22339.
- Brown, G. P., Pasternak, G. W., 1998. 3H-naloxone benzoylhydrazone binding in MOR-1-transfected chinese hamster ovary cells: evidence for G-protein-dependent antagonist binding. J Pharmacol Exp Ther 286, 376-381.
- Burgaud, J. L., Oudart, N., 1993. Effect of an histaminergic H3 agonist on the non-adrenergic non-cholinergic contraction in guinea-pig perfused bronchioles. J Pharm Pharmacol 45, 955-958.
- Burtin, C., Noirot, C., Scheinmann, P., Galoppin, L., Sabolovic, D., Bernard, P., 1988. Clinical improvement in advanced cancer disease after treatment combining Histamine and H2-antihistaminics (ranitidine or cimetidine). Eur J Cancer Clin Oncol 24, 161-167.
- Chen, S., Lin, F., Xu, M., Hwa, J., Graham, R. M., 2000. Dominant-negative activity of an alpha(1b)-adrenergic receptor signal-inactivating point mutation. EMBO J 19, 4265-4271.
- Chidiac, P., 2002. Considerations in the evaluation of inverse agonism and protean agonism at G protein-coupled receptors. Methods Enzymol 343, 3-16.
- Clapham, J., Kilpatrick, G. J., 1992. Histamine H3 receptors modulate the release of [3H]-acetylcholine from slices of rat entorhinal cortex: evidence for the possible existence of H3 receptor subtypes. Br J Pharmacol 107, 919-923.
- Costa, T., Cotecchia, S., 2005. Historical review: negative efficacy and the constitutive activity of G-protein-coupled receptors. Trends Pharmacol Sci 26, 618-624.
- Cricco, G., Davio, C., Bergoc, R., Rivera, E., 1993. Inhibition of tumor growth induced by Histamine: in vivo and in vitro studies. Inflammation Research 38, 75-78.
- Davies, T. F., Ando, T., Lin, R., Tomer, Y., Latif, R., 2005. Thyrotropin receptor-associated diseases: from adenomata to graves disease. J Clin Invest 115, 1972-1983.
- Davio, C., Baldi, A., Mladovan, A., Cricco, G., Fitzsimons, C., Bergoc, R., Rivera, E., 1995. Expression of histamine receptors in different cell lines derived from mammary gland and human breast carcinomas. Inflamm Res 44 Suppl 1, S70-S71.
- Davio, C., Mladovan, A., Lemos, B., Monczor, F., Shayo, C., Rivera, E., Baldi, A., 2002. H1 and

- H2 histamine receptors mediate the production of inositol phosphates but not cAMP in human breast epithelial cells. Inflamm Res 51, 1-7.
- De Lean, A., Kilpatrick, B. F., Caron, M. G., 1982. Dopamine receptor of the porcine anterior pituitary gland. Evidence for two affinity states discriminated by both agonists and antagonists. Mol Pharmacol 22, 290-297.
- Delvalle, J., Wang, L., Gantz, I., Yamada, T., 1992. Characterization of H2 histamine receptor: linkage to both adenylate cyclase and [Ca2+]i signaling systems. Am J Physiol 263, G967-972.
- Dowling, M. R., Willets, J. M., Budd, D. C., Charlton, S. J., Nahorski, S. R., Challiss, R. A. J., 2006. A single point mutation (N514Y) in the human M3 muscarinic acetylcholine receptor reveals differences in the properties of antagonists: evidence for differential inverse agonism. J Pharmacol Exp Ther 317, 1134-1142.
- Dryja, T. P., Berson, E. L., Rao, V. R., Oprian, D. D., 1993. Heterozygous missense mutation in the rhodopsin gene as a cause of congenital stationary night blindness. Nat Genet 4, 280-183.
- el Battari, A., Martin, J. M., Luis, J., Pouzol, O., Secchi, J., Marvaldi, J., Pichon, J., 1988. Solubilization of the active vasoactive intestinal peptide receptor from human colonic adenocarcinoma cells. J Biol Chem 263, 17685-17689.
- Fitzsimons, C. P., Gompels, U. A., Verzijl, D., Vischer, H. F., Mattick, C., Leurs, R., Smit, M. J., 2006. Chemokine-directed trafficking of receptor stimulus to different G proteins: selective inducible and constitutive signaling by human herpesvirus 6-encoded chemokine receptor U51. Mol Pharmacol 69:888-898.
- Fitzsimons, C. P., Monczor, F., Fernández, N., Shayo, C., Davio, C., 2004. Mepyramine, a histamine H1 receptor inverse agonist, binds preferentially to a G protein-coupled form of the receptor and sequesters G protein. J Biol Chem 279, 34431-34439.
- Fitzsimons, C., Durán, H., Engel, N., Molinari, B., Rivera, E., 1999. Changes in H2 receptor expression and coupling during Ca2+-induced differentiation in mouse epidermal keratinocytes. Inflamm Res 48 Suppl 1, S73-S74.
- Friedel, R. O., Schanberg, S. M., 1975. Effects of histamine on phospholipid metabolism of rat brain in vivo. J Neurochem 24, 819-820.
- Georgieva, T., Devanathan, S., Stropova, D., Park, C. K., Salamon, Z., Tollin, G., Hruby, V. J., Roeske, W. R., Yamamura, H. I., Varga, E., 2008. Unique agonist-bound cannabinoid CB1 receptor conformations indicate agonist specificity in signaling. Eur J Pharmacol 581, 19-29.
- Gespach, C., Bawab, W., de Cremoux, P., Calvo, F., 1988. Pharmacology, molecular identification and functional characteristics of vasoactive intestinal peptide receptors in human breast cancer cells. Cancer Res 48, 5079-5083.
- Hall, D. A., 2000. Modeling the functional effects of allosteric modulators at pharmacological receptors: an extension of the two-state model of receptor activation. Mol Pharmacol 58, 1412-1423.
- Hill, S. J., Ganellin, C. R., Timmerman, H., Schwartz, J. C., Shankley, N. P., Young, J. M., Schunack, W., Levi, R., Haas, H. L., 1997. International Union of Pharmacology. XIII. Classification of histamine receptors. Pharmacol Rev 49, 253-278.
- Hokin, M. R., Hokin, L. E., 1953. Enzyme secretion and the incorporation of P32 into phospholipides of pancreas slices. J Biol Chem 203, 967-977.
- Imamura, M., Seyedi, N., Lander, H. M., Levi, R., 1995. Functional identification of histamine H3-receptors in the human heart. Circ Res 77, 206-210.
- Ishikawa, S., Sperelakis, N., 1987. A novel class (H3) of histamine receptors on perivascular nerve terminals. Nature 327, 158-160.
- Kaufman, D. W., Kelly, J. P., Rosenberg, L., Anderson, T. E., Mitchell, A. A., 2002. Recent patterns of medication use in the ambulatory adult population of the united states: the slone survey. JAMA 287, 337-344.
- Keen, T. J., Inglehearn, C. F., Lester, D. H., Bashir, R., Jay, M., Bird, A. C., Jay, B., Bhattacharya, S. S., 1991. Autosomal dominant retinitis pigmentosa: four new mutations in rhodopsin, one of them in the retinal attachment site. Genomics 11, 199-205.

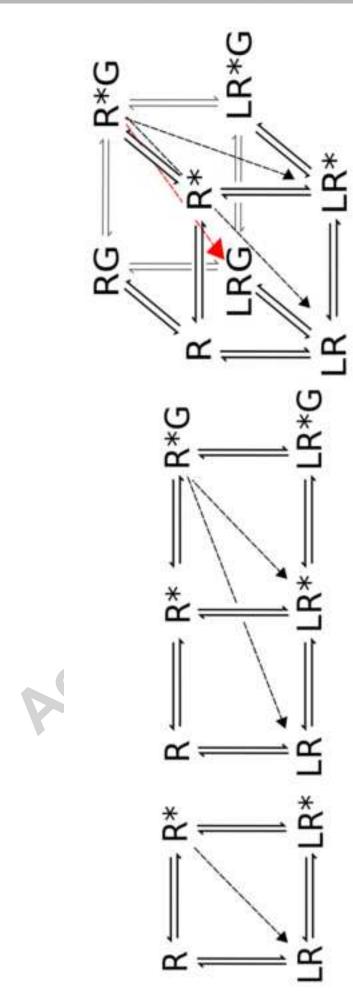
- Kenakin, T., 1995. Agonist-receptor efficacy. I: mechanisms of efficacy and receptor promiscuity. Trends Pharmacol Sci 16, 188-192.
- Kenakin, T., 2002. Efficacy at G-protein-coupled receptors. Nat Rev Drug Discov 1, 103-110.
- Kenakin, T., 2004. Efficacy as a vector: the relative prevalence and paucity of inverse agonism. Mol Pharmacol 65, 2-11.
- Kenakin, T., Onaran, O., 2002. The ligand paradox between affinity and efficacy: can you be there and not make a difference?. Trends Pharmacol Sci 23, 275-280.
- Korman, L. Y., Carney, D. N., Citron, M. L., Moody, T. W., 1986. Secretin/vasoactive intestinal peptide-stimulated secretion of bombesin/gastrin releasing peptide from human small cell carcinoma of the lung. Cancer Res 46, 1214-1218.
- Kroog, G. S., Jensen, R. T., Battey, J. F., 1995. Mammalian bombesin receptors. Med Res Rev 15, 389-417.
- Lee, T. W., Cotecchia, S., Milligan, G., 1997. Up-regulation of the levels of expression and function of a constitutively active mutant of the hamster alpha1b-adrenoceptor by ligands that act as inverse agonists. Biochem J 325 (Pt 3), 733-739.
- Leff, P., 1995. The two-state model of receptor activation. Trends Pharmacol Sci 16, 89-97.
- Leurs, R., Chazot, P. L., Shenton, F. C., Lim, H. D., de Esch, I. J. P., 2009. Molecular and biochemical pharmacology of the histamine H4 receptor. Br J Pharmacol 157, 14-23.
- Leurs, R., Church, M. K., Taglialatela, M., 2002. H1-antihistamines: inverse agonism, anti-inflammatory actions and cardiac effects. Clin Exp Allergy 32, 489-498.
- MacEwan, D. J., Milligan, G., 1996. Inverse agonist-induced up-regulation of the human beta2-adrenoceptor in transfected neuroblastoma X glioma hybrid cells. Mol Pharmacol 50, 1479-1486.
- Martin, M. W., Smith, M. M., Harden, T. K., 1984. Modulation of muscarinic cholinergic receptor affinity for antagonists in rat heart. J Pharmacol Exp Ther 230, 424-430.
- Maussang, D., Langemeijer, E., Fitzsimons, C. P., Stigter-van Walsum, M., Dijkman, R.,Borg, M. K., Slinger, E., Schreiber, A., Michel, D., Tensen, C. P., van Dongen, G. A., Leurs, R., Smit, M. J., 2009. The human cytomegalovirus-encoded chemokine receptor US28 promotes angiogenesis and tumor formation via cyclooxygenase-2. Cancer Res 69, 2861-2869.
- McLoughlin, D. J., Strange, P. G., 2000. Mechanisms of agonism and inverse agonism at serotonin 5-HT1a receptors. J Neurochem 74, 347-357.
- Meye, F. J., Trezza, V., Vanderschuren, L. J. M. J., Ramakers, G. M. J., Adan, R. A. H., 2012. Neutral antagonism at the cannabinoid 1 receptor: a safer treatment for obesity. Mol Psychiatry, . Milligan, G., Bond, R. A., Lee, M., 1995. Inverse agonism: pharmacological curiosity or potential therapeutic strategy?. Trends Pharmacol Sci 16, 10-13.
- Molnar, E. L., Cricco, G., Martin, G., Darvas, Z., Hegyesi, H., Fitzsimons, C., Bergoc, R., Falus, A., Rivera, E., 2001. Histamine as a potential autocrine regulator of melanoma. Inflamm Res 50 Suppl 2, S102-S103.
- Monczor, F., Fernandez, N., Legnazzi, B. L., Riveiro, M. E., Baldi, A., Shayo, C., Davio, C., 2003. Tiotidine, a histamine H2 receptor inverse agonist that binds with high affinity to an inactive G-protein-coupled form of the receptor. Experimental support for the cubic ternary complex model. Mol Pharmacol 64, 512-520.
- Morisset, S., Rouleau, A., Ligneau, X., Gbahou, F., Tardivel-Lacombe, J., Stark, H., Schunack, W., Ganellin, C. R., Schwartz, J. C., Arrang, J. M., 2000. High constitutive activity of native H3 receptors regulates histamine neurons in brain. Nature 408, 860-864.
- Nakaya, N., Tasaka, K., 1988. The influence of histamine on precursors of granulocytic leukocytes in murine bone marrow. Life Sci 42, 999-1010.
- Nielsen, H. J., 1991. Histamine and histamine type-2 receptor antagonists in psoriasis. Mechanisms and speculations. Dan Med Bull 38, 478-480.
- Nielsen, H. J., 1996. Histamine-2 receptor antagonists as immunomodulators: new therapeutic views?. Ann Med 28, 107-113.
- Nielsen, H., Kikuchi, Y. . Histamine H2-antagonists as potential adjuvant treatment of malignant disease. In *Histamine in normal and cancer cell proliferation, advances in the bioscience*. Garcia-

- Caballero M, Brandes L, Hosoda S (Eds.). 1993. 319-334.
- Oda, T., Morikawa, N., Saito, Y., Masuho, Y., Matsumoto, S., 2000. Molecular cloning and characterization of a novel type of histamine receptor preferentially expressed in leukocytes. J Biol Chem 275, 36781-36786.
- Onaran, H. O., Costa, T., 1997. Agonist efficacy and allosteric models of receptor action. Ann N Y Acad Sci 812, 98-115.
- Osawa, S., Kajimura, M., Yamamoto, S., Ikuma, M., Mochizuki, C., Iwasaki, H., Hishida, A., Terakawa, S., 2005. Alteration of intracellular histamine H2 receptor cycling precedes antagonist-induced upregulation. Am J Physiol Gastrointest Liver Physiol 289, G880-G889.
- Pincus, D. W., DiCicco-Bloom, E. M., Black, I. B., 1990. Vasoactive intestinal peptide regulates mitosis, differentiation and survival of cultured sympathetic neuroblasts. Nature 343, 564-567.
- Raible, D. G., Lenahan, T., Fayvilevich, Y., Kosinski, R., Schulman, E. S., 1994. Pharmacologic characterization of a novel histamine receptor on human eosinophils. Am J Respir Crit Care Med 149, 1506-1511.
- Rivera, E., Davio, C., Cricco, G., Bergoc, R. . Histamine regulation of tumour growth. Role of H1 and H2 receptors. In *Histamine in normal and cancer cell proliferation, advances in the bioscience*. Garcia-Caballero M, Brandes L, Hosoda S (Eds.). 1993. 299-317.
- Robertson, M. J., Dougall, I. G., Harper, D., McKechnie, K. C., Leff, P., 1994. Agonist-antagonist interactions at angiotensin receptors: application of a two-state receptor model. Trends Pharmacol Sci 15, 364-369.
- Samama, P., Cotecchia, S., Costa, T., Lefkowitz, R. J., 1993. A mutation-induced activated state of the beta 2-adrenergic receptor. Extending the ternary complex model. J Biol Chem 268, 4625-4636. Schlicker, E., Fink, K., Detzner, M., Göthert, M., 1993. Histamine inhibits dopamine release in the mouse striatum via presynaptic H3 receptors. J Neural Transm Gen Sect 93, 1-10.
- Seifert, R., Wenzel-Seifert, K., 2002. Constitutive activity of G-protein-coupled receptors: cause of disease and common property of wild-type receptors. Naunyn Schmiedebergs Arch Pharmacol 366, 381-416.
- Smit, M. J., Hoffmann, M., Timmerman, H., Leurs, R., 1999. Molecular properties and signalling pathways of the histamine H1 receptor. Clin Exp Allergy 29 Suppl 3, 19-28.
- Smit, M. J., Leurs, R., Alewijnse, A. E., Blauw, J., Van Nieuw Amerongen, G. P., Van De Vrede, Y., Roovers, E., Timmerman, H., 1996. Inverse agonism of histamine H2 antagonist accounts for upregulation of spontaneously active histamine H2 receptors. Proc Natl Acad Sci U S A 93, 6802-6807
- Smit, M. J., Timmerman, H., Alewijnse, A. E., Leurs, R., 1998. From histamine H2 receptor regulation to reclassification of H2 antagonists; inverse agonism as the basis for H2 receptor upregulation. Receptors Channels 5, 99-102.
- Strange, P. G., 2002. Mechanisms of inverse agonism at G-protein-coupled receptors. Trends Pharmacol Sci 23, 89-95.
- Subramanian, N., Whitmore, W. L., Seidler, F. J., Slotkin, T. A., 1981. Ontogeny of histaminergic neurotransmission in the rat brain: concomitant development of neuronal histamine, H-1 receptors, and H-1 receptor-mediated stimulation of phospholipid turnover. J Neurochem 36, 1137-1141.
- Threlfell, S., Exley, R., Cragg, S. J., Greenfield, S. A., 2008. Constitutive histamine H2 receptor activity regulates serotonin release in the substantia nigra. J Neurochem 107, 745-755.
- Tiligada, E., Kyriakidis, K., Chazot, P. L., Passani, M. B., 2011. Histamine pharmacology and new cns drug targets. CNS Neurosci Ther 17, 620-628.
- Tubio, M. R., Fernandez, N., Fitzsimons, C. P., Copsel, S., Santiago, S., Shayo, C., Davio, C., Monczor, F., 2010. Expression of a G protein-coupled receptor (GPCR) leads to attenuation of signaling by other GPCRs: experimental evidence for a spontaneous GPCR constitutive inactive form. J Biol Chem 285, 14990-14998.
- van der Werf, J. F., Timmerman, H., 1989. The histamine H3 receptor: a general presynaptic histaminergic regulatory system?. Trends Pharmacol Sci 10, 159-162.
- Virgolini, I., Raderer, M., Kurtaran, A., Angelberger, P., Banyai, S., Yang, Q., Li, S., Banyai, M.,

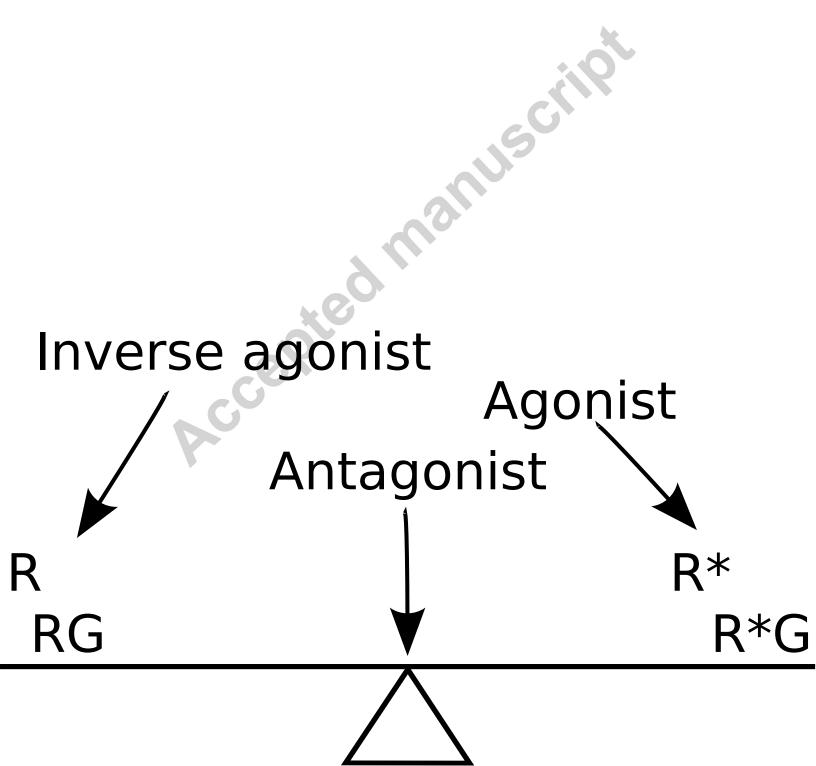
- Pidlich, J., Niederle, B., 1994. Vasoactive intestinal peptide-receptor imaging for the localization of intestinal adenocarcinomas and endocrine tumors. N Engl J Med 331, 1116-1121.
- Wade, S. M., Lan, K., Moore, D. J., Neubig, R. R., 2001. Inverse agonist activity at the alpha(2a)-adrenergic receptor. Mol Pharmacol 59, 532-542.
- Weiss, J. M., Morgan, P. H., Lutz, M. W., Kenakin, T. P., 1996a. The cubic ternary complex receptor-occupancy model. III. resurrecting efficacy. J Theor Biol 181, 381-397.
- Weiss, J. M., Morgan, P. H., Lutz, M. W., Kenakin, T. P., 1996b. The cubic ternary complex receptor-occupancy model. II. understanding apparent affinity. J Theor Biol 178, 169-182.
- Weiss, J. M., Morgan, P. H., Lutz, M. W., Kenakin, T. P., 1996c. The cubic ternary complex receptor-occupancy model. I. model description. J Theor Biol 178, 151-167.
- Wellner-Kienitz, M. C., Bender, K., Meyer, T., Pott, L., 2003. Coupling to Gs and G(q/11) of histamine H2 receptors heterologously expressed in adult rat atrial myocytes. Biochim Biophys Acta 1642, 67-77.
- Westphal, R. S., Sanders-Bush, E., 1994. Reciprocal binding properties of 5-hydroxytryptamine type 2c receptor agonists and inverse agonists. Mol Pharmacol 46, 937-942.
- Wilson, J., Lin, H., Fu, D., Javitch, J. A., Strange, P. G., 2001. Mechanisms of inverse agonism of antipsychotic drugs at the D(2) dopamine receptor: use of a mutant D(2) dopamine receptor that adopts the activated conformation. J Neurochem 77, 493-504.
- Yap, Y., Camm, A. . Potential cardiac toxicity of H1-antihistamines. In *Histamine and h1-antihistamines in allergic disease 2nd ed.* Simos FER (Ed.). 2002. 389-419.

Acceloited.

- Zhu, Y., Michalovich, D., Wu, H., Tan, K. B., Dytko, G. M., Mannan, I. J., Boyce, R., Alston, J., Tierney, L. A., Li, X., 2001. Cloning, expression, and pharmacological characterization of a novel human histamine receptor. Mol Pharmacol 59, 434-441.
- Zurier, R. B., Kozma, M., Sinnett-Smith, J., Rozengurt, E., 1988. Vasoactive intestinal peptide synergistically stimulates dna synthesis in mouse 3t3 cells: role of cAMP, Ca2+, and protein kinase C. Exp Cell Res 176, 155-161.







Accepted manuscript

