

IMMUNOLOGICAL RESPONSE TO TOXOPLASMOSIS IN POPULATION GROUPS OF THE STATE OF SÃO PAULO, BRAZIL, AS EVALUATED BY THE DISTRIBUTION OF SERUM TITERS IN THE DYE-TEST

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SUMMARY

The present investigation analyses the pattern of frequencies of serum titers in positive reactants to the SABIN-FELDMAN dye-test from a total of 4,440 individuals resident in three different areas of the State of São Paulo, Brazil, most of which were suspected of having some clinical manifestation of toxoplasmosis. Over three quarters of the observations conformed to a theoretical chance distribution, disclosing one more indication about the specificity of the dye test. Observations with titers over 1:1,024 assembled into a separate distribution, being likely to correspond to recent cases of toxoplasmosis, independently of clinical manifestations.

INTRODUCTION

SABIN & FELDMAN's³⁸ dye-test has been extensively used as a valuable diagnostic tool in clinical medicine, as well as in numerous epidemiological surveys investigating the degree to which population groups harbor infection by *Toxoplasma gondii*.

The intensity of a positive reaction is measured by the highest dilution at which blood serum can induce discoloration of the test organism, under specified conditions. Frequently the intensity of reaction, expressed as serum titer, is used to characterize severity of infection or acuteness of the disease. When the results of a series of tests are considered, the numbers of sera to which ordered titer values are assigned form a frequency distribution. Distributions of such kind have been used in several investigations to correlate results of the dye-test and other immunological means of diagnosis. In other instances, titer distributions have been used to compare groups of individuals of different sex, age, socio-economic condition or envi-

ronment, in what regards the intensity of toxoplasma infection.

Less frequently, other possibilities of the study the distribution of positive titers to the dye-test have been explored. Notwithstanding, the pattern of such distribution of frequencies in fairly extensive surveys could in itself provide clues to the knowledge, on one hand, of the homogeneity of population groups concerning the dissemination of toxoplasma infection and their immune response; on the other hand, of specificity characteristics of the test under the conditions how it was performed in those groups.

With a view to the forementioned possibilities, the present investigation analyses the distribution of serum titers in positive reactants from a total of 4,440 individuals, resident in three different areas of the State of São Paulo, Brazil, most of which were suspected of having some clinical manifestation of toxoplasmosis. Since the indivi-

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duals by no means constitute representative samples of the population groups involved, the observed figures cannot be taken as expressing the incidence of toxoplasma infection in those groups. The figures nevertheless provide subsidiary information concerning the order of magnitude which the prevalence of toxoplasmosis can be expected to assume in this part of the country.

MATERIAL AND METHODS

A) *Origin of the material studied* — All of the 4,440 serum samples were collected in private clinics from subjects suspected of having some form of toxoplasmosis, resident in three different areas of the State of São Paulo, Brazil. This State is situated in the southern area of the country, i.e., within the south-temperate climatic zone (Fig. 1).

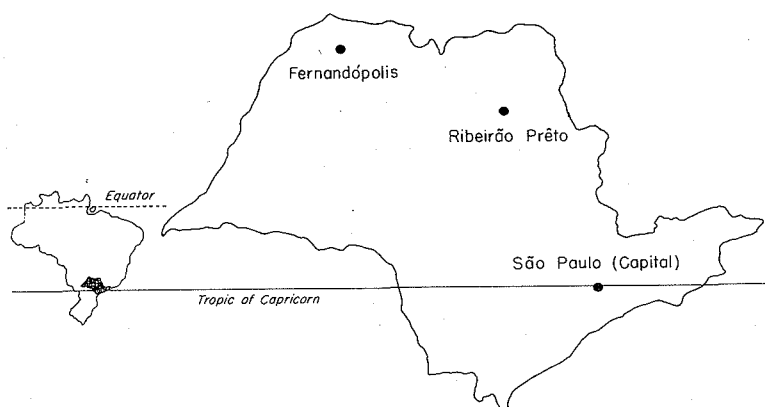


Fig. 1 — Immunological response to toxoplasmosis. Geographical situation of the areas studied. 1 — São Paulo (Capital); 2 — Ribeirão Preto; 3 — Fernandópolis

3,731 Specimens were from São Paulo City, the capital of the State, a 1,509 square kilometer municipality, with a 3,109.9/km² population density (5,849,529 urban and 52,004 rural; 1970). Its altitude varies from 740 to 820 meters above sea level, and its average annual rainfall amounts to 1,640.1 mm.

389 Specimens were from Ribeirão Preto, an agricultural center 289 kilometers westwards from São Paulo, 1,048 square kilometers in area, with a 199.8/km² population density (194,800 urban and 14,627 rural; 1970). The local altitude averages 518 meters, and the annual rainfall 1,109 mm.

The remaining 320 sera were from Fernandópolis, also an agricultural center, 552 kilometers westward from São Paulo, with a 71.5/km² population density (28,480

urban, 10,510 rural; 1970). Its altitude averages 529 meters, and its annual rainfall 1,101 mm.

B) *Laboratory methods*

Test sera — The blood samples were collected aseptically and maintained at room temperature for about two hours, centrifuged at low speed (1,000 r.p.m.) for fifteen minutes, and the sera stored at + 4°C. Sera from residents of São Paulo were tested within two to three days upon collection, but those from Ribeirão Preto and Fernandópolis withstood periods of storage at + 4°C up to fifteen days, plus six to eight hour-periods at room temperature during surface delivery to São Paulo.

All of the sera were non-inactivated.

Toxoplasma suspension — The "N" strain of *Toxoplasma gondii* originally isolated by

TABLE I

Per cent distributions of sera positive to the dye-test (over the total numbers of positive sera)

Area	Year	Number tested	Total positive	Titer (*)										
				2 ¹	2 ²	2 ³	2 ⁴	2 ⁵	2 ⁶	2 ⁷	2 ⁸			
São Paulo	1961	168	76.8	10.9	22.5	34.1	13.9	9.3	3.1	1.5	3.9	0.8	—	—
	1962	461	76.1	4.8	17.9	35.6	21.9	7.4	2.6	2.3	2.9	2.9	1.7	—
	1963	686	70.3	7.5	15.8	29.9	23.8	9.1	0.2	2.1	2.7	2.5	2.1	4.3
	1964	554	75.0	6.7	15.1	31.5	27.7	8.9	1.7	1.9	2.4	1.4	1.7	1.0
	1965	843	68.0	4.5	13.3	34.4	25.7	8.2	2.4	2.6	2.4	1.7	1.6	3.1
	1966	1,019	69.6	3.5	15.4	24.1	24.4	10.6	4.1	3.9	2.7	3.1	3.7	4.5
	1961/66	3,731	71.3	5.5	15.6	30.5	24.2	9.1	2.4	2.7	2.7	2.3	2.2	2.8
Ribeirão Preto ...	1961	60	83.3	14.0	22.0	40.0	12.0	10.0	—	2.0	—	—	—	—
	1962	60	78.3	4.3	17.0	42.6	25.5	8.5	2.1	—	—	—	—	—
	1963	74	79.7	5.1	13.5	37.3	22.0	15.3	—	3.4	1.7	—	1.7	—
	1964	114	84.2	5.2	18.8	28.1	30.2	12.5	—	—	1.0	3.1	—	1.0
	1965	81	86.4	4.3	11.4	50.0	21.4	8.6	4.3	—	—	—	—	—
1961/65	389	82.8	6.2	16.5	38.5	23.3	11.2	1.2	0.9	0.6	0.9	0.3	0.3	
Fernandópolis	1962	62	82.3	11.8	27.4	41.2	11.8	5.9	—	—	—	—	1.9	—
	1963	71	85.9	4.9	19.7	37.7	21.3	8.2	—	—	3.3	3.3	—	1.6
	1964	28	92.9	7.7	23.1	23.1	26.9	11.5	—	7.7	—	—	—	—
	1965	96	84.4	4.9	9.9	32.1	34.6	11.1	3.7	2.5	—	1.2	—	—
	1966	63	65.1	—	22.0	26.8	19.5	17.1	4.9	—	2.4	—	2.4	4.9
1962/66	320	81.3	5.8	18.8	33.5	23.8	10.4	2.0	1.5	1.2	1.2	0.8	1.2	
All areas	1961	228	78.5	11.7	22.4	35.7	13.4	9.5	2.2	1.7	2.8	0.6	—	—
	1962	583	77.0	5.6	18.9	37.0	21.2	7.3	2.2	1.8	2.2	2.2	1.6	—
	1963	831	72.4	7.0	16.0	31.4	23.4	9.6	0.2	2.0	2.7	2.3	1.8	3.6
	1964	696	77.3	6.5	16.2	30.5	28.1	9.7	1.3	1.8	2.0	1.7	1.3	0.9
	1965	1,020	71.0	4.6	12.7	35.6	26.2	8.6	2.8	2.4	1.9	1.5	1.2	2.5
	1966	1,082	69.3	3.3	15.7	24.3	24.1	10.9	4.1	3.7	2.7	2.9	3.6	4.5
1961/66	4,440	73.0	5.6	16.0	31.6	24.1	9.4	2.2	2.4	2.3	2.1	1.9	2.4	

(*) Reciprocal of serum dilution

KIRCHNER, E. & COTRIM, J. X. — Immunological response to toxoplasmosis in population groups of the State of São Paulo, Brazil, as evaluated by the distribution of serum titers in the dye-test. *Rev. Inst. Med. trop. São Paulo* 14:33-50, 1972.

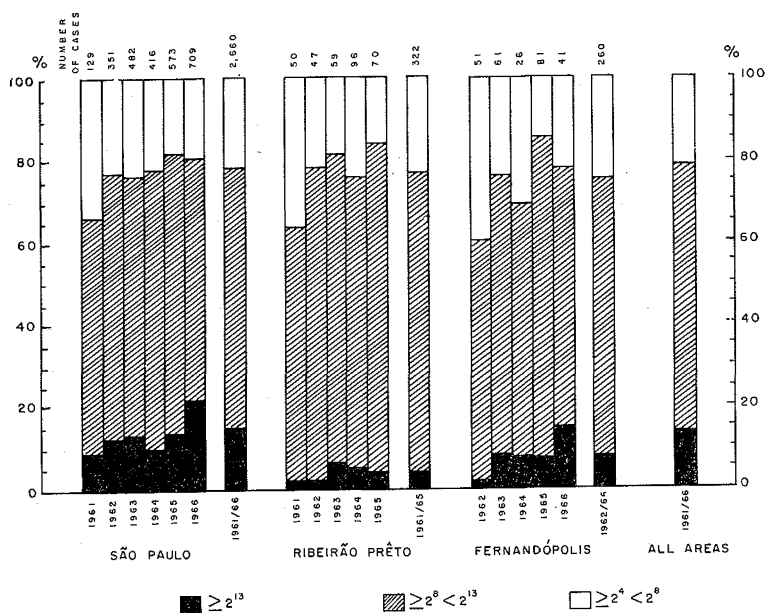


Fig. 2 — Graphical representation of the percent distribution of sera positive to the dye-test

NOBREGA³² during an epizootics of rabbits, and laboratory-maintained by serial mouse-passage, was utilized as antigen. From mice inoculated two days earlier, a peritoneal exudate with an extracellular toxoplasma population ranging 20,000,000/ml was harvested. This exudate was diluted at 1:5 in cooled physiological salt solution added with 3.8 percent sodium citrate as anticoagulant, and kept in a refrigerator. Strict care was taken to have the test set up within one hour after the exudate harvesting.

Accessory factor — Obtained from volunteer donors, and routinely tested previously to storage in 1 ml aliquots, at -20°C .

Positive control serum — The same precautions were observed concerning the positive control serum.

Methylene blue solution — This solution was prepared anew every 2-3 days, from a saturated alcoholic solution of methylene blue and an alkaline soda-borax solution of pH 11.

The test^(*) — The SABIN & FELDMAN³⁸ dye-test technique modified by SABIN et al.³⁷ in 1952, was followed. Every serum was submitted to a screen test covering the range of 2^{-4} to 2^{-12} (in four-fold) dilutions, the sera reacting at the last dilution being subsequently tested in two-fold dilutions from 2^{-12} up to 2^{-18} . Control reactions with known positive sera were carried out simultaneously to each series of tests, the negative control being the accessory factor itself.

C) *Criteria for dye-test positiveness* — The highest serum dilution preventing the stain-intake in 50 per cent of the toxoplasma cells, was considered as the titer of the given serum in the dye-test, these titers being here expressed as the logarithm to base 2 of the corresponding dilution.

RESULTS

Occurrence of positive sera — Sabin & Feldman's dye test was performed in the sera of 4,440 individuals from the three forementioned areas during a period of six

(*) Carried out at the "Laboratório de Análises Brooklin", São Paulo, Brazil

years, from 1961 to 1966. The numbers of positive and negative sera from each area on each year, as well as the distribution of positive reactants according to serum titer, are presented in Table I.

Altogether 73.0 per cent of the sera were found to be positive, with titers varying from 1:16, which was the lowest dilution tested, to 1:262,144 (2^{-18}). As shown by the figures in Table I and the graphical representation in Figure 2, only small

fluctuations were observed from year to year in the percentages that positive tests represented of the total in each area; an exception was the decrease of about 20 per cent observed at Fernandópolis in 1966. The average percentages of positive sera in the 6-year period were 71.3 per cent in São Paulo, 82.8 per cent in Ribeirão Preto and 81.3 per cent in Fernandópolis, the total numbers of tested sera being 3,731, 389 and 320 for each area, respectively.

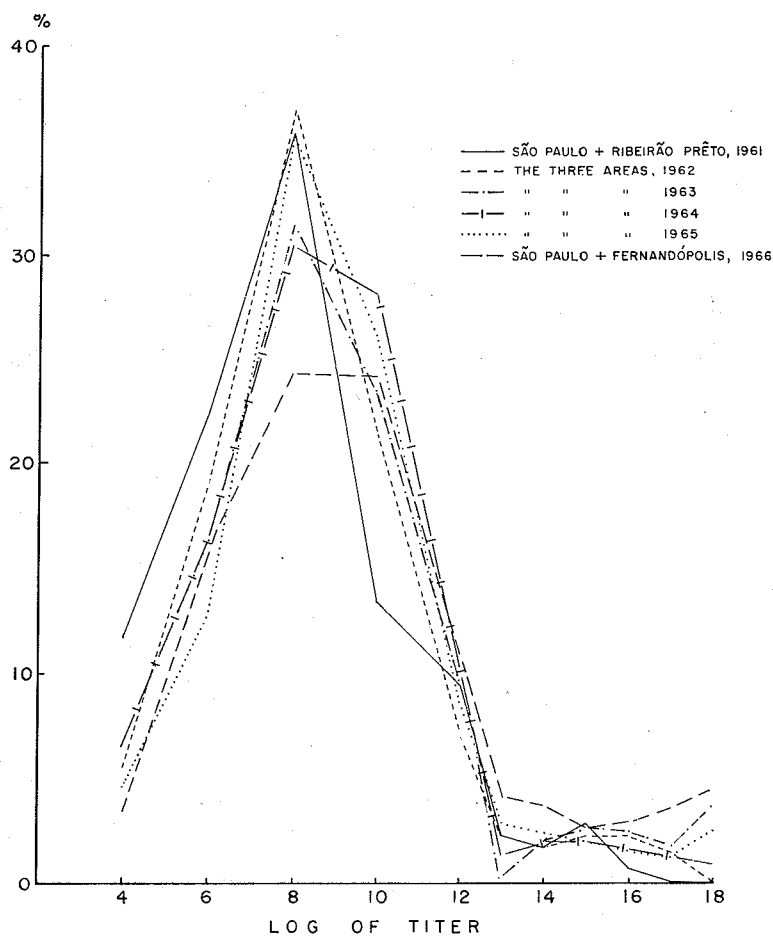


Fig. 3 — Comparison of the frequency distribution of the total sera positive to the dye-test in the three areas studied, corresponding to different years.

In Fig. 2 differently shaded areas represent the numbers of sera with high (higher than 1:4,096), intermediate (from 1:256 to 1:4,096) and low titers (lower than 1:256), expressed as percentages of the total number of sera tested on each year and area.

Considering the whole period of observation, sera with high titers were more frequent in São Paulo (10.7 per cent of the total) than in Ribeirão Preto (3.7 per cent), with intermediary frequency in Fernandópolis (6.3 per cent).

A similar relationship holds true for the separate results corresponding to each year. Furthermore, a slight trend toward an increase of high titers is apparent on successive years in each area of investigation.

Sera with intermediate and low titers were in general less frequent in São Paulo than in Ribeirão Preto for Fernandópolis during the whole period of observation.

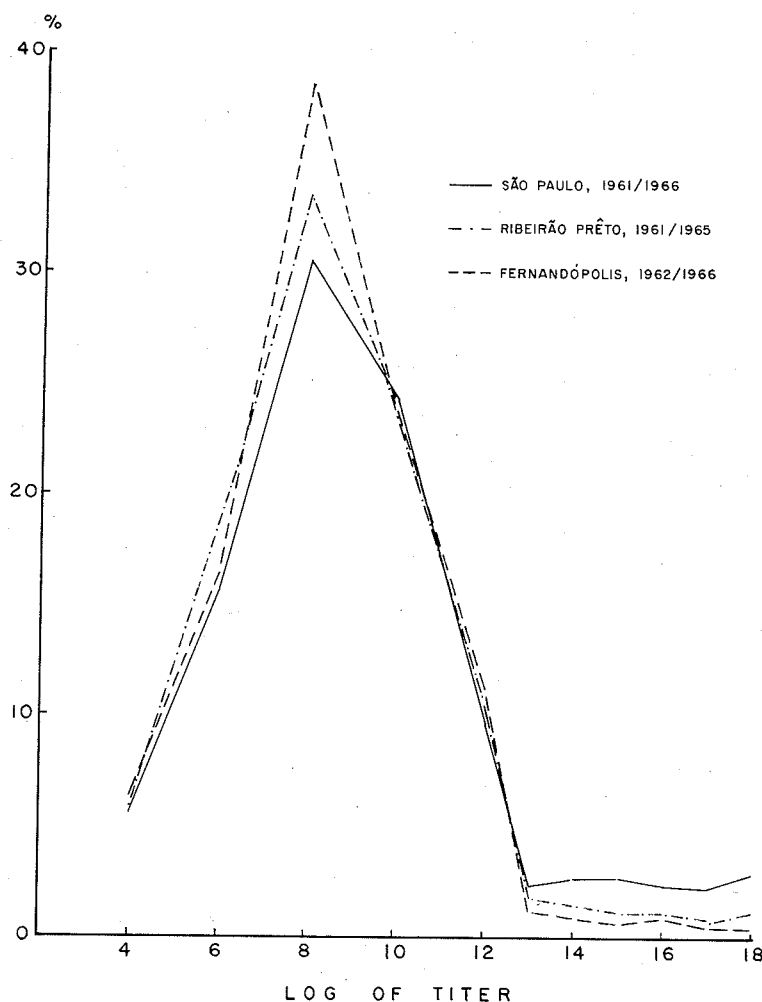


Fig. 4 — Totals of sera positive to the dye-test corresponding to the 6-year period separately for each area, expressed as percent of the total number of positive sera in the respective area

The occurrence of positive sera at different titers can be better analysed when, for each year and area, percentages are taken over the total numbers of positive sera instead of the total numbers of sera tested. Those percentages are also given in Table I, and their distributions are considered in the following section.

Distributions of positive sera according to titer — The frequency distributions of sera positive to the dye-test on each year and area present as a general characteristic an accumulation of cases in correspondence to titers 1:256 and 1:1,024, with gradually decreasing numbers for titers progressively lower or higher than those values.

Since the number of observations is not large for each year and area separately,

especially in Ribeirão Preto and Fernandópolis, the totals of the three areas are first considered to compare the distributions corresponding to different years. The data, expressed as percentages of the total number of positive sera observed on each year, are plotted in Fig. 3 in relation to the logarithm of the serum titer. Apart a few irregularities, the general appearance of the curves is not suggestive of essential differences corresponding to any particular year.

On the other hand, when the totals corresponding to the 6-year period are considered separately for each area and expressed as percentages of the total number of positive sera in the respective area, the curves plotted on Fig. 4 result. Here a striking similarity is apparent, indicating for the different areas a very good agreement in

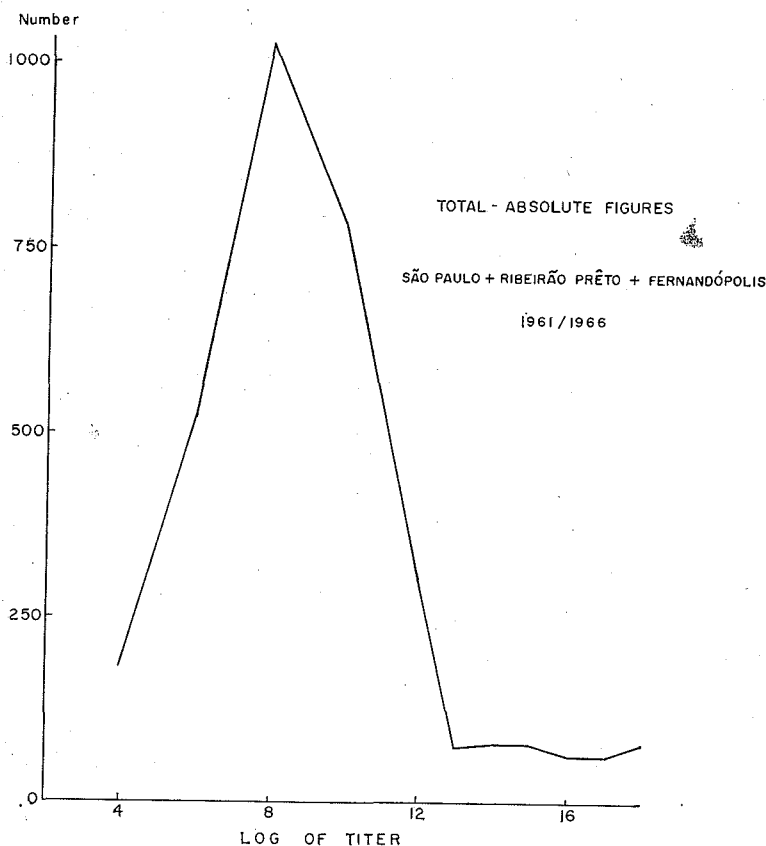


Fig. 5 — Pooled distribution of the 3,242 sera positive to the dye-test

the shape of the distribution of positive sera in relation to the logarithm of the titer.

Inasmuch as the distributions corresponding either to different areas or to different years do not show definite peculiarities characteristic of a certain year or area, but rather seem to exhibit a common pattern, the pooled distribution of 3,242 positive sera according to log titer can be considered as typical of the whole set of data and is plotted in Fig. 5.

The main portion of the resulting curve reminds the bell-shaped configuration of the normal probability curve. However, truncation is apparent in correspondence to the lowest dilution tested, whereas an extended tail corresponds to the highest serum dilutions, rendering the curve definitely asymmetrical. Nevertheless, a major portion of the data could still possibly follow the normal

distribution, partially masked by superposition of the distribution or distributions corresponding to the remaining data. In order to check this possibility, an expedient resource is the graphic test of normality, which in this case consists of plotting as ordinates the probits corresponding to the cumulative percentages of positive sera observed up to each titer, and as abscissae the logarithms of the respective titers: when the graph approximates a straight line, the original data can be considered as normally distributed.

The graphic test of normality was performed with the data of Table II and is shown in Fig. 6. It is seen that the first four points in the graph are disposed on an approximately straight line, whereas the six points corresponding to the highest serum dilutions form a slightly curved, concave up line, at an angle with the straight

TABLE II

Overall distribution of sera positive to the dye-test

Log titer (*)	Observed frequency	Cumulative frequency	Cumulative per cent of total	Probit
4	181	181	5.61	3.41
6	518	699	21.58	4.21
8	1,023	1,722	53.13	5.08
10	782	2,504	77.25	5.75
12	304	2,808	86.62	6.11
13	73	2,881	88.87	6.22
14	78	2,959	91.28	6.36
15	76	3,035	93.62	6.52
16	67	3,102	95.68	6.72
17	61	3,163	97.56	6.98
18	79	3,242	100.00	—

(*) Logarithm to base 2 of the reciprocal of serum dilution

line. As shown by that graph, two different distributions appear to have been superimposed: the observations corresponding to sera which were positive at dilutions varying from 1:16 to 1:1,024 follow an approximately normal distribution; those corresponding

to sera which were positive at higher dilutions diverge from the former.

The bulk of the data, however, corresponds to the normally distributed portion, which represents 77.3 per cent of the total. From the probit graph, estimates of the arithmetic

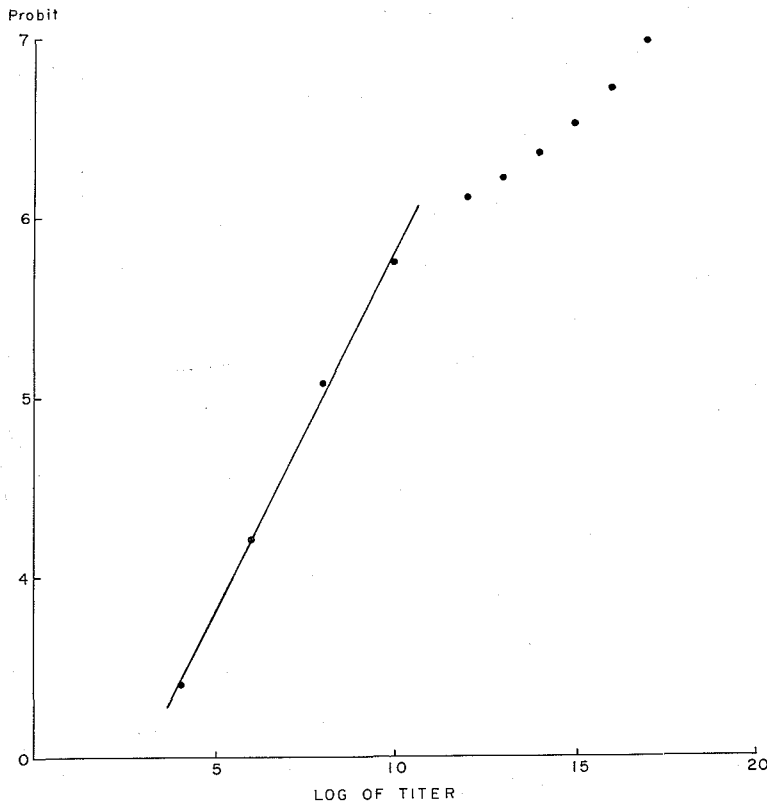


Fig. 6 — Overall distribution of sera positive to the dye-test. Graphic test of normality

mean and standard deviation of the log titers in that portion can be derived. The mean — which is also the logarithm to base 2 of the geometric mean of the titers — is the abscissa value corresponding to probit 5 as the ordinate, and was found to be 7.98. The standard deviation is the reciprocal of the slope of the probit line, and was found to be 2.54.

For the whole set of positive tests, the calculated mean of log titers was 9.31 and the standard deviation was 2.66.

DISCUSSION

The large proportion of positive tests which was found consistently in the three areas investigated, no doubt points out that toxoplasmosis is widely prevalent in those

areas. That finding in itself bears no direct relation to the extent to which cases with clinical symptoms of toxoplasmosis should be expected in the involved communities. Latent infection by *Toxoplasma gondii* is probably responsible for the large majority of positive serological tests which have been found in most surveys of population groups in different countries. In fact, high as well as low rates of positiveness to the dye test appear to have been found independently of the inclusion or not of suspected clinical cases of toxoplasmosis in the investigated group. Considering investigations which included suspected cases, such as the present one, the proportions of positive sera were similarly high in many instances, as shown in the upper section of Table III; however, in other instances, as shown in the lower section of the same table, much lower pro-

TABLE III

Frequency of positive dye test in groups including suspected cases of toxoplasmosis

Authors	Locality	Number tested	Per cent positive	Observations
CAMARGO, M. E. ⁸	São Paulo, Brazil	1,000	76.8	
REUSS, K. ³⁶	Frankfurt/Main, Germany	4,240	62.3	
KUDICKE, H. & POEHLIG, W. ²⁵ (A)	Lower Saxony, Germany	1,234	65.7	
KUDICKE, H. & POEHLIG, W. ²⁵ (B)	Lower Saxony, Germany	2,194	71.4	
HAHN, E. ²¹	North Italy	1,000	77.3	
LEINZIGER, E. et al. ²⁶	Linz, Australia	484	85.5	} Female patients; Landesfrauenklinik } Mothers; Landeskinderkrankenhaus } Children; same hospital
SCHOLZ, R. ³⁹	Linz, Australia	473	97.5	
SCHOLZ, R. ³⁹	Linz, Australia	138	71.0	
GAGLIERIS, A. N. ⁷	Pavia, Italy	93	40.8	
VON ZEIPPEL, G. & LINDER, L. A. ⁴²	Gottenborgh, Sweden	328	38.4	
MERCIER, P. et al. ²⁸	Greece	170	30.6	

portions of positiveness have been found. On the other hand, in surveys of groups consisting of supposedly normal individuals, the proportions of positive sera were often below 30 per cent (Table IV, upper section), but other times well above 60 per cent (lower section, same table). Those discrepancies could be expected if one considers that the chances for the appearance of clinical symptoms of toxoplasmosis do not necessarily run parallel to the positiveness rates indicating spread of the organism in different population groups. But such discrepancies might also raise doubts about the specificity of the dye regarding infection by toxoplasma. Experimental data reported by MUEHLPPFORD³¹, HEIN²³, MICHALZIK²⁹, WILDFUEHR¹⁴, AWAD¹, AWAD & LAINSON² and GROENROOS²⁰, appear to indicate that positive dye tests may result from the presence of other infectious agents or from the action of serum properdin. The evidence in that respect has been denied by FELDMAN

& MILLER¹³ as well as by CATHIE⁹, BORGEN & BJOERNSTADT⁸, and MAS BAKAL²⁷, who on the basis of other experimental findings uphold the reliability of the dye test in the diagnosis of toxoplasmosis.

Significant indications about specificity of the dye test are disclosed by analysis of the titer distribution of positive tests, such as performed in the present paper. As described above, over three quarters of our observations conformed to a theoretical chance distribution, the normal probability curve, when classified according to logarithms of the serum titers.

In what concerns the observations with titers over 1:1,024, which appeared to make up a separate distribution, they are likely to correspond to recent cases of toxoplasmosis, independent of clinical manifestations. The notion that high titers of the dye test are associated with recent infection, rather than with the appearance of symptoms, seems to be well established (SABIN et al.³⁷;

KIRCHNER, E. & COTRIM, J. X. — Immunological response to toxoplasmosis in population groups of the State of São Paulo, Brazil, as evaluated by the distribution of serum titers in the dye-test. *Rev. Inst. Med. trop. São Paulo* 14:33-50, 1972.

TABLE IV

Frequency of positive dye-test in groups of supposedly normal individuals

Authors	Locality	Number tested	Per cent positive	Observations
BERENGO, A. et al. ³	Siena, Italy	1,200	4.1	Blood donors
FELDMAN, H. A. & MILLER, L.T. ¹³	Arizona, USA	236	4.2	Navajo Indians
MIDTWEDT, T. ³⁰	Lapland, Norway	100	5.0	School children
HARBOE, A. ²²	Oslo, Norway	1,599	8.5	
FELDMAN, H. A. & MILLER, L.T. ¹³	Iceland	108	11.1	
MIDTWEDT, T. ³⁰	Norway	200	15.5	School children
FELDMAN, H. A. & MILLER, L.T. ¹³	Portland, Ore., USA	293	17.4	
FELDMAN, H. A. ¹¹	USA	2,294	19.0	Military recruits
BEVERLEY, J. K. A. et al. ⁵	Sheffield, England	581	19.5	
REMINGTON, J. S. et al. ³⁴	USA	359	19.5	Students of Harvard University
GIBSON, C. L. ¹⁷	Tennessee, USA	627	20.4	Urban Negro population
GIBSON, C. L. et al. ¹⁹	Tennessee, USA	987	21.8	Rural Negro population
COOK, I. ¹⁰	Austrália	760	24.0	
GROENROOS, P. ²⁰	Finland	1,176	25.6	
FELDMAN, H. A. & MILLER, L.T. ¹³	St. Louis, Mo., USA	184	25.6	
MAS BAKAL, P. ²⁷	Holland	200	66.0	Blood donors
FELDMAN, H. A. & MILLER, L.T. ¹³	La Lima, Honduras	266	66.2	
JAMRA, L. F. ²⁴	São Paulo, Brazil	300	67.0	
NUSSENZWEIG, R. S. ³³	São Paulo, Brazil	334	71.3	Blood donors
FERRARIS, G. & BERETTA, L. R. ¹⁴	Bari, Italy	676	74.1	Pregnant women
FELDMAN, H. A. & MILLER, L.T. ¹³	Tahiti	121	82.0	
GIBSON, C. L. & COLEMAN, N. ¹⁸	Costa Rica	156	88.5	
GIBSON, C. L. & COLEMAN, N. ¹⁸	Esciuntha, Guatemala	100	94.0	

GROENROOS²⁰; FELDMAN¹²; BEVERLEY⁴; FLECH & LUDLAN¹⁵; SCHOLZ³⁹; TOLENTINO⁴⁰). At any rate, the forementioned observations with high titers, as evidenced by the probit graph, do not fit into the

distribution pattern exhibited by the bulk of our data. The latter form a homogeneous aggregate, suggesting that a stable system of causes was operative in the production of titer variability.

In other words, 77.3 per cent of the observed sera can be considered as samples of the same "universe" of variation, in which the titer values result from the interaction of multiple causes, effective to different degrees in each case, but constantly present. Conformity to one and the same probability distribution, indicating stability of the variation pattern, points out the underlying

operation of a constant-cause system. Such an effect could hardly originate in a system where specificity of the involved test — one of the causes in operation — were not constantly present. Therefore, demonstration that the system of chance causes in operation to produce the observed pattern of titer variability is constant, represents strong evidence in favor of specificity of the dye test to diagnose one and the same infection.

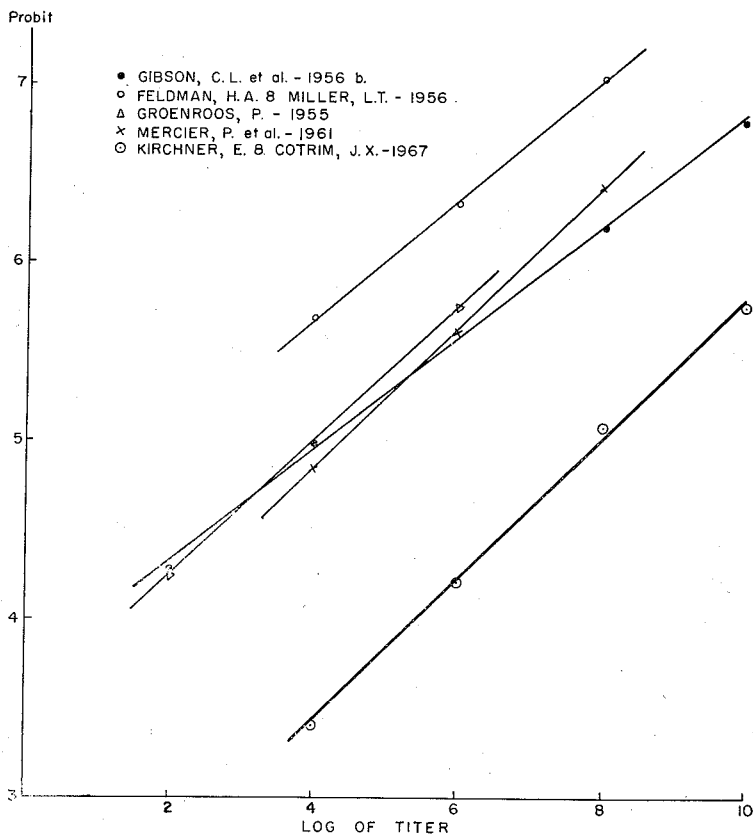


Fig. 7 — Graphic tests of normality. Data reported by:

- ● — GIBSON, C. L. et al.¹⁰ — Rural Negro population, Tennessee, USA
- ○ — FELDMAN, H. A. & MILLER, L. T.⁶³ — Population; Portland, St. Louis and Pittsburgh, USA
- △ — GROENROOS, P.²⁰ — Normal; blood donors and patients, toxoplasmosis excluded; Finland
- × — MERCIER, P. et al.²⁸ — Suspected of toxoplasmosis; Greece
- ○ — KIRCHNER, E. & COTRIM, J. X. — Suspected; São Paulo, Brazil

If such is the case, the titer observed in other dye test surveys should also be expected to conform to the normal probability distribution, at least as far as the considered group is homogeneous in what concerns toxoplasma infection, the corresponding immune response, and the conditions under which the test was performed.

The data of 3 other surveys carried out in São Paulo, Brazil, as well as those corresponding to 14 population groups investigated in other countries were analysed and led to the probit graphs represented in Figures 7-10. As a matter of fact, as shown in Table V in 16 out of the 18 groups considered, over 70 per cent of the

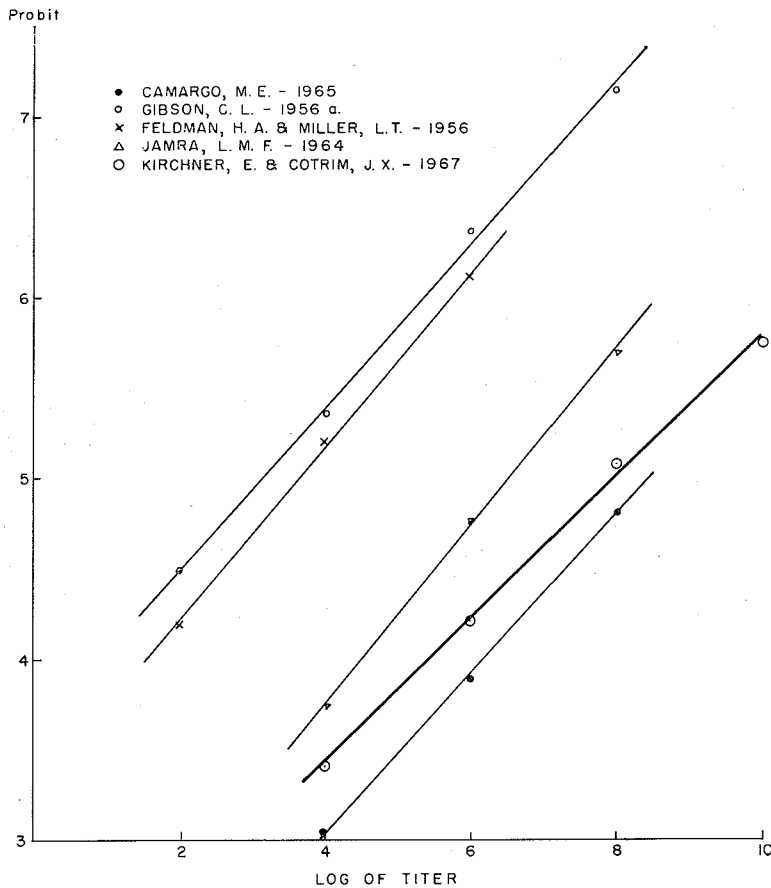


Fig. 8 — Graphic tests of normality. Data reported by:

- ● — CAMARGO, M. E.⁸ — Suspected of toxoplasmosis; São Paulo, Brazil
- ○ — GIBSON, C. L.¹⁷ — Urban Negro population; Tennessee, USA
- × — FELDMAN, H. A. & MILLER, L. T.¹⁸ — Population; New Orleans, La, USA
- △ — JAMRA, L. M. F.²⁴ — Urban population; São Paulo, Brazil
- ○ — KIRCHNER, E. & COTRIM, J. X. — Suspected; São Paulo, Brazil

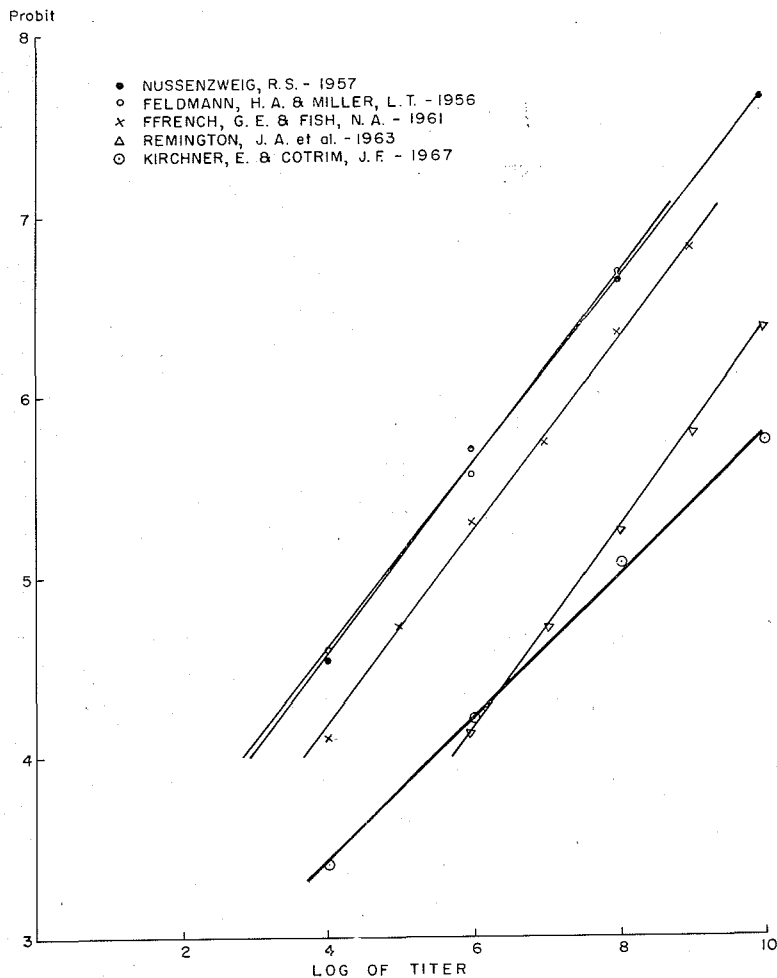


Fig. 9 — Graphic tests of normality. Data reported by:

- ● — NUSSENZWEIG, R. S.³³ — Blood donors; São Paulo, Brazil
- ○ — FELDMAN, H. A. & MILLER, L. T.³² — Population; Honduras
- x — FFRENCH, G. E. & FISH, N. A.¹⁶ — Hospitalized patients; Ontario, Canada
- △ — REMINGTON, J. A. et al.³⁴ — Students at Harvard University; Boston, Mas., USA
- ⊙ — KIRCHNER, E. & COTRIM, J. X. — Suspected of toxoplasmosis; São Paulo, Brazil

sera exhibited titers which could be considered as normally distributed.

As to the possible characterization of these population samples in regard to their including or not actual cases of toxoplas-

mosis, neither the variability range of the titer distributions (b values in Table V or relative inclination of probit lines in Figs. 7-10) nor the remaining values investigated tentatively (s , X_5 , \bar{X} etc.), do allow such delimitation.

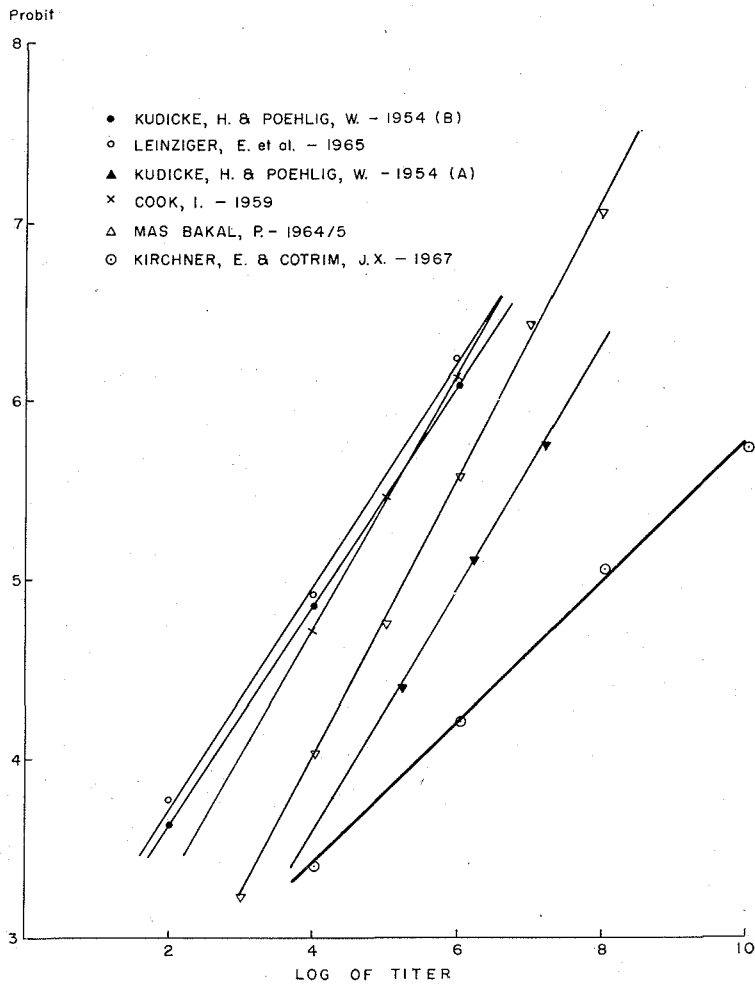


Fig. 10 — Graphic tests of normality. Data reported by:

- ● — KUDICKE, H. & POEHLIG, W.²⁵ (B) — Patients of various diseases, including toxoplasmosis; Saxony, Germany
- ○ — LEINZIGER, E. et al.²⁶ — Obstetric Clinic, Linz, Austria
- ◆ — KUDICKE, H. & POEHLIG, W.²⁵ (A) — Patients of various diseases, including toxoplasmosis; Saxony, Germany
- × — COOK, I.³⁰ — Population; Queensland, Toowoomba and Tableland, Australia
- △ — MAS BAKAL, P.²⁷ — Blood donors; Netherlands
- ○ — KIRCHNER, E. & COTRIM, J. X. — Suspected of toxoplasmosis; São Paulo, Brazil

TABLE V

Immunological response to toxoplasmosis as measured by the dye-test. Analysis of the data corresponding to eighteen population groups

Authors	Sample studied	no. of sera	b	s	X _s	Modal value X	Average X observed	Per cent positive over total	Per cent positive comprised in probit line
GIBSON, C. L. et al. ¹⁹	Rural Negro population; Tennessee, USA	987	.3120	3.205	4.154	4	5.1	21.8	81.4
FELDMAN, H. A. & MILLER, L. T. ¹³	Population; Portland, St. Louis and Pittsburg, USA	621	.3375	2.963	2.010	4	4.4	31.9	73.2
GROENROOS, P. ²⁰	Normal; blood donors and patients, toxoplasmosis excluded; Finland	1,176	.3750	2.667	4.000	6	5.3	25.6	77.3
MERCIER, P. et al. ²⁵	Suspected of toxoplasmosis; Greece	170	.3925	2.548	4.403	4	5.8	30.6	92.2
KIRCHNER, E. & COTRIM, J. X.	Suspected; São Paulo, Brazil	4,440	.3945	2.535	7.982	8	9.3	73.0	77.3
CAMARGO, M. E. ⁸	Suspected; São Paulo, Brazil	1,000	.4400	2.273	8.477	10	9.3	76.8	42.2
GIBSON, C. L. ¹⁷	Urban Negro population; Tennessee, USA	627	.4480	2.232	3.125	4	3.8	20.4	75.0
FELDMAN, H. A. & MILLER, L. T. ¹³	Population; New Orleans, La., USA	270	.4825	2.073	3.648	5	5.0	39.3	86.8
JAMRA, L. M. F. ²⁴	Urban population; São Paulo, Brazil	300	.4875	2.051	6.551	8	7.7	67.0	75.5
NUSSENZWEIG, P. S. ³³	Blood donors; São Paulo, Brazil	334	.5130	1.949	4.788	6	5.9	71.3	99.6
FELDMAN, H. A. & MILLER, L. T. ¹³	Population; Honduras	266	.5225	1.914	4.828	6	6.0	66.2	92.0
FFRENCH, G. E. & FISH, N. A. ⁶⁶	Hospitalized patients; Ontario, Canada	650	.5371	1.862	5.556	6	6.2	40.0	96.5
REMINGTON, J. A. et al. ³⁴	Students at Harvard University; Boston, Mas., USA	359	.5500	1.818	7.535	8	8.1	19.5	75.8
KUDICKE, H. & POEHLIG, W. ^{25(B)}	Patients of various diseases, including toxoplasmosis; Saxony, Germany	2,194	.6150	1.626	4.228	6	5.3	71.4	86.3
LEINZIGER, E. et al. ²⁶	Obstetric Clinic, Linz, Austria	484	.6200	1.613	4.065	6	5.1	85.5	89.4
KUDICKE, H. & POEHLIG, W. ^{25(A)}	Patients of various diseases, including toxoplasmosis; Saxony, Germany	1,234	.6800	1.471	6.068	6.2	6.4	65.7	67.7
COOK, I. ¹⁰	Population; Queensland, Toowoomba and Tableland, Australia	760	.7050	1.418	4.376	4	5.2	24.0	87.1
MAS BAKAL, ²⁷	Blood donors; Netherlands	200	.7760	1.289	5.264	6	5.7	66.0	98.4

KIRCHNER, E. & COTRIM, J. X. — Immunological response to toxoplasmosis in population groups of the State of São Paulo, Brazil, as evaluated by the distribution of serum titers in the dye-test. *Rev. Inst. Med. trop. São Paulo* 14:33-50, 1972.

RESUMO

Resposta imunológica à toxoplasmose em grupos populacionais do Estado de São Paulo, avaliada pela distribuição de títulos da reação de Sabin-Feldman

A presente investigação analisa o padrão de frequências de títulos séricos em reagentes positivos ao teste de Sabin-Feldman, de um total de 4.440 indivíduos residentes em três áreas diversas do Estado de São Paulo, Brasil, a maioria dos quais constituída por casos suspeitos de apresentarem alguma manifestação clínica de toxoplasmose. Mais de três quartas partes das observações enquadram-se como distribuição teórica ao acaso, proporcionando mais uma indicação favorável à especificidade da prova do corante. Verificou-se ainda que os resultados com títulos superiores a 1:1.024 constituem uma distribuição separada, parecendo, assim, representar casos recentes da doença, independentemente de manifestações clínicas.

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REFERENCES

1. AWAD, F. I. — The diagnosis of toxoplasmosis. Lack of specificity of Sabin-Feldman dye test. *Lancet* 2:1054-1056, 1954.
2. AWAD, F. I. & LAINSON, R. — A note on the serology of sarcosporidiosis and toxoplasmosis. *J. Clin. Path.* 7:152-156, 1954.
3. BERENGO, A.; de LALLA, F.; CAVALLINI, F.; CAVALLINI-SAMPIERI, L. & BECHELLI, G. — Ricerche sierologiche sulla diffusione della toxoplasmosi. I. Studio su 1.200 donatori di sangue. *Minerva Med.* 56:4561-4570, 1965.
4. BEVERLEY, J. K. A. — Symposium on toxoplasmosis. *Trans. Roy. Soc. Trop. Med. Hyg.* 51:118-122, 1957.
5. BEVERLEY, J. K. A.; BEATTIE, C. P. & ROSEMAN, C. — Human toxoplasma infection. *J. Hyg. (London)* 52:37-46, 1954.
6. BORGEN, P. H. F. & BJOERNSTADT, R. T. — On the specificity of the toxoplasma dye-test. *Acta Path. Microbiol. Scand.* 41:361-364, 1957.
7. CAGLIERIS, A. N. — Risultati del test tintoriale in individui sani ed ammalati (Ricerca su 466 soggetti). *Boll. Soc. Med. Chir. Pavia* 72:647-661, 1958.
8. CAMARGO, M. E. — *Estudo comparativo das reações de Sabin-Feldman e de imuno-fluorescência indireta, para a toxoplasmose, em 1.000 soros humanos. Comportamento anômalo de alguns soros.* Tese. São Paulo, 1965.
9. CATHIE, J. A. B. — An appraisal of the diagnostic value of the serological tests for toxoplasmosis. *Trans. Roy. Soc. Trop. Med. Hyg.* 51:104-110, 1957.
10. COOK, I. — Toxoplasma in Queensland. IV. Serological surveys of human population. *Austr. J. Exp. Biol. Med. Sci.* 37:581-592, 1959.
11. FELDMAN, H. A. — A nationwide serum survey of United States military recruits, 1962. VI. Toxoplasma antibodies. *Amer. J. Epidem.* 81:385-391, 1965.
12. FELDMAN, H. A. — The clinical manifestations and laboratory diagnosis of toxoplasmosis. *Amer. J. Trop. Med. & Hyg.* 2:420-428, 1952.
13. FELDMAN, H. A. & MILLER, L. T. — Serological study of toxoplasmosis prevalence. *Amer. J. Hyg.* 64:320-335, 1956.
14. FERRARIS, G. & BERETTA, L. R. — La toxoplasmosi in gravidanza. *Minerva Ginec.* 17:745-757, 1965.
15. FLECK, D. G. & LUDLAM, G. B. — Indications for laboratory tests for toxoplasmosis. *Brit. Med. J.* 5472:1239-1240, 1965.
16. FRENCH, G. E. & FISH, N. A. — A survey of toxoplasmosis in an Ontario community. *Canad. M.A.J.* 84:757-767, 1961.
17. GIBSON, C. L. — Distribution of toxoplasma antibodies in comparable urban and rural groups. *Pub. Health Rep.* 71:1119-1123, 1956.

KIRCHNER, E. & COTRIM, J. X. — Immunological response to toxoplasmosis in population groups of the State of São Paulo, Brazil, as evaluated by the distribution of serum titers in the dye-test. *Rev. Inst. Med. trop. São Paulo* 14:33-50, 1972.

18. GIBSON, C. L. & COLEMAN, N. — The prevalence of toxoplasma antibodies in Guatemala and Costa Rica. *Amer. J. Trop. Med. & Hyg.* 7:334-338, 1958.
19. GIBSON, C. L.; EYLES, D. E.; COLEMAN, N. & SMITH, C. S. — Serological response of a rural negro population to the Sabin-Feldman cytoplasm modifying test for toxoplasmosis. *Amer. J. Trop. Med. & Hyg.* 5: 772-783, 1956.
20. GROENROOS, P. — Studies on toxoplasma and the serology of toxoplasmosis. *Ann. Med. Exp. Biol. Fenniae* 33 (Suppl. 11):1-113, 1955.
21. HAHN, E. — Studi serologici sulla toxoplasmosi umana in Italia. *Boll. Inst. Sterot. Milan.* 41:159-162, 1962.
22. HARBOE, A. — Toxoplasma dye test titers of 1,600 blood donors in Oslo. *Acta Path. Microbiol. Scand.* 93 (Suppl.): 325-331, 1952.
23. HEIN, W. — Das Verhalten des Sabin-Feldman Testes und der Westphal'schen Komplementbindungs-Reaktion bei der Lungentuberkulose. *Zchr. Tropenmed.* 3:339-357, 1952.
24. JAMRA, L. M. F. — Contribuição para a epidemiologia da toxoplasmosse. *Inquérito em 100 famílias de uma área da Cidade de São Paulo.* Tese. São Paulo, 1964.
25. KUDICKE, H. & POEHLIG, W. — Erfahrungen und statistische Betrachtungen ueber den Serofarbttest nach Sabin und Feldman. *Zschr. fuer Hygiene und Infektionskrankheiten* 140:350-371, 1954.
26. LEINZIGER, E.; LECHNER, G. & ZWINZ, G. — Fehlbildungen der Frucht bei positiven Toxoplasmosse-testen der Muetter. *Wien. Med. Wschr.* 115:543-546, 1965.
27. MAS BAKAL, P. — The specificity of Sabin and Feldman's dye test in the diagnosis of toxoplasmosis. *Acta Leidensia* 33:148-160, 1964-1965.
28. MERCIER, P.; TZAMOURANIS, N. & CRIMBITHIS, E. — Serological study in Greece. *Arch. Inst. Pasteur Hell.* 7:83-93, 1961.
29. MICHALZIK, K. — *Trichomonas vaginalis* und positive Seroreaktion auf Toxoplasmosse. *Dtsch. Med. Wschr.* 78:307, 1953.
30. MIDTVEDT, T. — The frequency of positive dye test in children from different parts of Norway. *Acta Paediat. Scand.* 54:81-85, 1965.
31. MUEHLPFORD, H. — Das Verhalten Sarcosporidien-infizierter Tiere im Sero-Farbttest auf Toxoplasmosse nach Sabin-Feldman. *Zschr. Tropenmed.* 3:205-215, 1951.
32. NOBREGA, P. — Toxoplasmosse, uma doença de animais transmissível ao homem. *Arq. Inst. Biol.* 13:21-28, 1951.
33. NUSSENZWEIG, R. S. — Toxoplasmosse. Inquérito sorológico feito pela prova do corante em doadores de sangue. *Hospital (Rio)* 51:723-728, 1957.
34. REMINGTON, J. S.; DALRYMPLE, W.; JACOBS, L. & FINLAND, M. — Toxoplasma antibodies among college students. *New England J. Med.* 269:1394-1398, 1963.
35. REMINGTON, J. S. & MERLER, E. — Absence of cross reactivity between heterophile and dye test antibodies. *Proc. Soc. Exp. Biol. Med.* 115:115-118, 1964.
36. REUSS, K. — Die serologische Diagnose der Toxoplasmosse. *Praxis* 53:82-86, 1964.
37. SABIN, A. B.; EICHENWALD, H.; FELDMAN, H. A. & JACOBS, L. — Present status of clinical manifestations of Toxoplasmosis in man. *J.A.M.A.* 150:1063-1069, 1952.
38. SABIN, A. B. & FELDMAN, H. A. — Dyes as microchemical indicators of a new immunity phenomenon affecting a protozoon parasite (Toxoplasma). *Science* 108:660-663, 1948.
39. SCHOLZ, R. — Toxoplasma in Oberoestereich. *Wien. Med. Wschr.* 115:539-543, 1965.
40. TOLENTINO, P. — Epidemiologia e sierologia della toxoplasmosi. *Minerva Ginec.* 17: 758-761, 1965.
41. WILDFUEHR, G. — Ueber den Serofarbttest (Sabin-Feldman) bei Tuberkulose. *Deutsche Med. Wschr.* 5:167-168, 1954.
42. ZEIPEL, G. & LINDER, L. A. — Toxoplasmosis. A serological investigation with the dye test. *Acta Path. Microbiol. Scand.* 29: 229-238, 1951.

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