PATTERN OF CLASS-SPECIFIC FLUORESCENT ANTIBODIES ACCORDING TO THE STAGE OF THE INFECTION IN HUMAN SCHISTOSOMIASIS MANSONI

Hermínia Y. KANAMURA, Sumie HOSHINO-SHIMIZU and L. C. da SILVA

SUMMARY

The possible association of class specific antibodies to S. mansoni and stage of infection was studied by indirect fluorescent antibody test (IFT) in twenty-five selected sera from patients with acute and chronic schistosomiasis mansoni. IgA antibodies were detected only in sera from acute cases, but IgG, IgM and IgE antibodies were found in acute and chronic cases. Rheumatoid factor eventually present in the sera was removed by adsorbing the sera with immunoadsorbent of IgG, before the IFT was performed. In the acute stage the hemagglutination test (HAT) showed to be less sensitive than IFT.

INTRODUCTION

Immunofluorescence, hemagglutination, complement fixation and immunoprecipitation tests can detect anti-S. mansoni antibodies and are a very useful tool for the diagnosis of the disease. However, the correlation between serological findings and the stage of the infection has not definitely been stablished.

The levels of different immunoglobulin classes in the sera of patients with schistosomiasis mansoni were studied by single radial immunodiffusion test 1,3,5,7,10,18. However, as far as the clinical forms of the disease are concerned the results obtained by those Authors do not agree. Therefore, it might be more useful to determine the class-specific antibodies to S. mansoni instead of studying the total amount of immunoglobulin classes by radial immunodiffusion.

Recently, the immunofluorescence test carried out with class-specific conjugates to immunoglobulins detected IgG, IgM, IgA and IgE antibodies to S. mansoni in sera of infected patients 9,15,16, but these findings were not related to the different stages of the disease.

This report shows the results of a study based on class-specific fluorescent anti-S. mansoni antibodies in patients with different

stages of the infection, clinically characterized as acute and chronic (intestinal, hepatointestinal and hepatosplenic) forms. Indirect hemagglutination test was also performed.

MATERIAL AND METHODS

Sera — They were collected from 25 schistosomotic patients, with the following clinical diagnosis ¹⁹:

Acute form: 5 patients

Chronic form: 20 patients (intestinal: 6, hepatointestinal: 5, hepatosplenic: 9 patients).

Conjugates — Commercially available conjugates to human IgG, IgM, IgA and IgE (Hyland Div. Travenol Lab. U.S.A.) were used after testing their specificity by immuno-electrophoresis against selected sera with high level of immunoglobulins as determined by radial immunodiffusion test. In order to test the specificity of anti-IgE conjugate, one serum was lyophilized and concentrated 3 times.

Indirect fluorescent antibody test (IFT)

Two S. mansoni antigens were prepared for cryostat sectioning as follows:

1) adult worms were embedded in Tissue-Tek O.C.T. medium (*) and frozen in liquid nitrogen according to WILSON et al. ²¹.

^(*) Ames Co., Miles Lab., U.S.A.

Instituto de Medicina Tropical de São Paulo, São Paulo, Brasil. Supported by a Grant from «Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq)» (SIP/08-110)

2) liver specimen with eggs and granulomata obtained from infected hamster was also frozen as described by COUDERT et al. 6.

Sections were cut at 4 \mu thickness, fixed on microscope slides and stored at -20°C until used. The slides were then fixed in cold acetone for 5 minutes before the immunofluorescent technique. Serum dilutions from 1/10 to 1/5,120 were added on tissue sections and. after incubating for 30 minutes at 37°C, these were washed for 20 minutes with two changes of PBS (0.15M NaCl; 0.01M phosphates; pH An appropriate dilution of conjugate was placed on the specimens, and the slides were again incubated for 30 minutes at 37°C. twice washed in PBS and mounted with glicerol (pH 8.5) and a coverslip. All immunofluorescence tests included both positive and negative controls.

The slides were examined under a Zeiss fluorescence-microscope provided with a HBO-200 bulb, KP-500 exciter filter, Zeiss 50 barrier filter and dark field condenser.

Passive hemagglutination test — For the

hemagglutination test a preserved reagent was used according to HOSHINO et al. ¹³. This was prepared by lyophilizing formalin treated cells which had been preserved by aldehyde fixation after tannic acid treatment and sensitization with S. mansoni extracts. Sera were serially diluted from 1/20 to 1/5,120 in 0.85% NaCl and plastic microtitration plates used for this test.

Rheumatoid factor — A search for rheumatoid factor was performed in all sera by latex test (Behringwerke AG, Germany).

Ten selected sera, in which IgM was detected, were also adsorbed with immunoadsorbent of human IgG² and retested by immunofluorescence technique, in order to identify the immunological specificity of IgM.

RESULTS

Monospecific anti-human globulins immunofluorescence tests and hemagglutination test titers obtained in 25 sera from patients with acute and chronic schistosomiasis are presented in Table I. Figure 1 shows the fin-

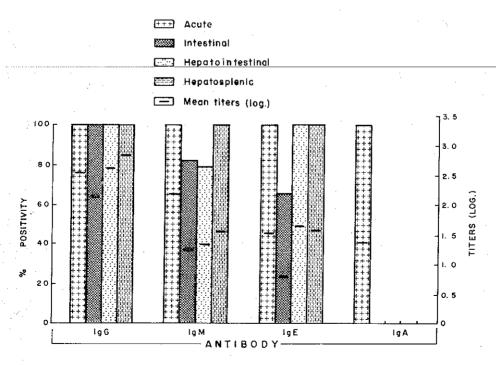


Fig. 1 — Distribution and mean titers of class specific antibodies found in sera from schistosomotic patients according to the clinical form of the disease.

TABLEI

Titers of 25 sera from patients with acute and chronic schistosomiasis mansoni obtained by immunofluorescence test (IFT), using monospecific conjugates, and by passive hemagglutination test (HAT)

			IFT								
Patient		Clinical forms	IgG Ab Liver Worm (**)		IgM Ab		IgE Ab Liver Worm		IgA Ab		HAT
1. 2. 3. 4. 5.	CSS IGO JAL JRA WSS	ACUTE	320 320 640 640 640	320 80 320 640 320	80 40 320 320 160	160 80 320 640 160	20 40 80 80 80	20 (***) 80 80 40	40 20 40 40 40 80	20 20 80 40	40 80 80 160 80
6. 7. 8. 9. 10. 11. 12. 13. 14. 15.	AFN ARL ATS ESN FAM VAS CRX DDS DMJ MCN ZLS	CHRONIC (*)	320 40 20 640 640 80 640 640 320 2560 640	320 80 40 640 320 80 320 320 320 320 80	160 20 10 40 40 40 	80 20 10 40 80 	40 	10 10 40 40 40 40 20 80 40			320 320 40 320 40 320 320 320 320 640 320
17. 18. 19. 20. 21. 22. 23. 24.	AMD ASS EAV HVS JBA JJS LVS MAS MDX		640 320 1280 2560 2560 640 1280 640	640 160 320 1280 640 640 820 320 640	20 40 20 80 20 80 80 80 20	40 20 80 160 80 160 40 —	80 20 80 320 160 80 40 40	40 40 80 160 80 40 20 10			160 320 320 2560 640 320 640 1280 160

(*) Chronic forms: intestinal (No. 6 to 11)
hepatointestinal (No. 12 to 16)
hepatosplenic (No. 17 to 25)

(**) Liver = liver sections with egg and granulomata worm = adult worm sections

(***) = negative result

dings of IgG, IgM, IgA and IgE antibodies to S. mansoni, according to clinical form of disease and estimated in percentages and mean titers.

Fluorescent antibody to S. mansoni

IgG — All sera gave a positive reaction to adult worm and to egg (liver section) antigens, respectively with mean titers (*) 280 and 480 in acute cases, and 290 and 500 in chronic cases.

IgM — 17 Out of 20 sera from chronic cases gave a positive reaction to adult worm and egg antigens, and both mean titers were 30. Two sera gave negative results to worm and egg antigens and one, showed positive result only to egg antigen.

All 5 sera from acute cases were positive to IgM antibodies, and the titers tended to be higher than in chronic cases. The mean titers 210 and 140 were respectively obtained with worm and egg antigens.

IgE — Most sera (23) gave a positive anti-IgE fluorescent reaction. However, in 3 sera,

^(*) mean titers = geometric mean titers

2 from chronic and one from acute schistosomiasis, positive reactions were observed with one antigen, either worm or egg. Moreover, there were 2 chronic cases in which IgE antibodies could not be detected. In acute cases, mean titers were 20 for worm and 50 for egg antigen. In chronic cases, mean titers were respectively 20 and 30. Negative and lower titers were mostly found in intestinal forms.

IgA — IgA antibodies were detected only in the acute form of the disease, with worm and egg antigens (mean titers 20 and 40, respectively).

Hemagglutination test (HAT) — Mean titers were lower in acute (80) than in chronic schistosomiasis (320).

Rheumatoid factor — The latex test revealed antibodies to IgG in two sera (AFN e LVS).

The adsorption of 10 selected sera (IGO, JAL, AFN, FAM, MCN, ZLS, HVS, JJS, LVS and MAS) with immunoadsorbent of IgG indicated that IgM antibodies first detected by immunofluorescent test were specific to S. mansoni. The rheumatoid factor detected in those two sera did not interfere with the titration of specific antibody.

DISCUSSION

Although some Authors 8,9,15,16 have already studied immunoglobulin classes of anti-S. mansoni antibodies in infected patients and animals, to our knowledge there is no investigation about the significance of these antibodies in different stages of the disease in humans.

The fluorescent antibody test showed IgG, IgM and IgE anti-S. mansoni antibodies in sera from acute and chronic cases. The finding of IgA antibodies only in acute cases differed strikingly from the chronic cases in which these antibodies were absent.

The presence of IgA antibodies in schistosomiasis seems to be significant since in other parasitic infections such as toxoplasmosis, trichinosis and hydatidosis IgA antibodies were found to be an indication of recent antigenic stimuli ^{15,17}.

In S. mansoni infection, HULDT et al. ¹⁵ have found IgA antibodies in 6 out of 103 patients and DEELDER et al.⁹ in every case from 19 selected patients. Also in S. haematobium infection IgA antibodies were detected in all 24 patients studied by KANE et al. ¹⁶. However, the stage of the infection was not mentioned by those Authors.

Unlike virus or protozoan infection, IgM response in schistosomiasis does not necessarily reflect a recent infection. However, it was observed that in acute cases IgM levels were slightly higher than in chronic cases.

A continuous stimulation with different antigenic determinants might elicit IgM antibody response in the host even at a late stage of the disease. IgM antibodies to S. mansoni in chronic cases of human schistosomiasis were also described by SILVA & FERRI ²⁰ and HILLYER ¹².

Since rheumatoid factors are referred as usually occurring in many parasitic diseases 4.11.14, we considered the possibility of IgM reactions seen in the IFT being due to this factor. However, the tests showed that IgM detected in the sera of acute and chronic cases were specific to S. mansoni. For the latex test, only two revealed the presence of rheumatoid factor. Besides, after complete removal of rheumatoid factor from these two sera, both from patients with chronic infection, IgM antibodies to S. mansoni could still be detected. The other sera also showed no change in IgM antibody titers after treatment with insoluble IgG.

IgE antibodies to **S. mansoni** were demonstrated in 23 out of 25 sera. Some Authors ^{9,10} have found IgE in about 50% to 70% of infected patients. In our hands IgE as well as IgM and IgG antibodies did not differentiate acute from chronic infections.

Our results suggest distribution of class specific antibodies to vary in the course of the infection. The observed modifications permit, thus, to select more appropriate tests to be applied either for diagnostic purposes or to follow the course of the infection. More detailed studies are under way in a larger group of patients from an endemic area.

As far as HAT is concerned, it showed to be a less sensitive test than IFT in the acute stage of the disease.

RESUMO

Comportamento das imuneglobulinas específicas nas diferentes formas clínicas da esquistossomose mansônica

A possível associação de anticorpos anti-S. mansoni de diferentes classes de imuneglobulinas com o estágio de infecção foi estudada por técnica de imunofluorescência (IF), em 25 soros selecionados de pacientes com esquistossomose aguda e crônica. Anticorpos IgA foram detectados somente em soro de casos agudos, tendo-se encontrado anticorpos IgG, IgM e IgE, tanto em casos agudos como crônicos. O fator reumatóide, eventualmente presente nos soros, foi removido por adsorção com imunoadsorvente de IgG, antes de se efetuar o teste de IF. A reação de hemaglutinação passiva mostrou-se menos sensível do que a IF para detecção de anticorpos em casos agudos da doença.

ACKNOWLEDGEMENT

We wish to express our gratitude to Maria do Carmo Berthe Rosa for the elaboration of the bibliographical references.

REFERENCES

- ANTUNES, L. J.; REIS, A. P.; PELLEGRINO, J.: TAVARES, C. A. & KATZ, N. — Immunoglobulins in human schistosomiasis mansoni. J. Parasit. 57:539-542, 1971.
- AVRAMEAS, A. Coupling of enzymes of proteins with glutaraldehyde. Use of conjugate for the detection of antigens and antibodies. Immunochemistry 6:43-55, 1969.
- BASSILY, S.; HIGASHI, G. I.; FARID, Z. & WILLIAMS, R. E. — Serum immunoglobulins in schistosomiasis mansoni. J. Trop. Med. Hyg. 75: 73-75, 1972.
- CAMARGO, M. E.; LESER, P. G. & ROCCA, A.
 — Rheumatoid factors as a cause for false positive IgM anti-toxoplasma fluorescent tests. A technique for specific results. Rev. Inst. Med. trop. São Paulo 14:310-313, 1972.
- COOK, J. A.; WOODSTOCK, L. & JORDAN, P.
 Immunological studies in Schistosoma mansoni

- infection in St. Lucia. Ann. Trop. Med. Parasit. 66:869-373, 1972.
- 6. COUDERT, J.; GARIN, J. P.; AMBRIOSE-THOMAS, P. & POTHIER, M. A. Premiers résultats à propos du diagnostic sérologique de la bilharziose par immunofluorescence sur coupes à la congelation de Schistosoma mansoni. Ann. Parasit. Hum. Comp. 42:483-492, 1967.
- CURIEL, M.; CHAVES, J. & TORREALBA, J.
 W. Immunoglobulin levels in schistosomiasis.
 Rev. Inst. Med. trop. São Paulo 14:384-387, 1972.
- DEELDER, A. M. Immunology of experimental infections with Schistosoma mansoni in the swiss mouse and Fasciola hepatica in the rabbit. Acta Leidensia 39:1-107, 1973.
- DEELDER, A. M.; SNOIJINK, J. J. & PLOEM, J. S. — Immunoprecipitation and class-specific immunofluorescence titration of human serum antibodies to Schistosoma mansoni antigens. Z. Parasitenk. 46:195-201, 1975.
- DESSAINT, J. P.; CAPRON, M.; BOUT, D. & CAPRON, A. Quantitative determination of specific IgE antibodies to schistosome antigens and serum IgE levels in patients with schistosomiasis (S. mansoni or S. haematobium). Clin. Exp. Immunol. 20:427-436, 1975.
- GREENWOOD, B. M.; MULLER, A. S. & VAL-KENBURG, H. A. — Rheumatoid factor in Nigerian sera. Clin. Exp. Immunol. 8:161-173, 1971.
- HILLYER, G. V. Immunoprecipitins in Schistosoma mansoni infections. IV Human infections. Exp. Parasit. 25:376-381, 1969.
- HOSHINO, S., CAMARGO, M. E. & SILVA, L. C. da Standardization of a hemagglutination test for schistosomiasis with formalin-treated human erythrocytes. Amer. J. Trop. Med. Hyg. 19:463-470, 1970.
- HOUBA, V. & ALLISON, A. C. M-antiglobulins (rheumatoid-factor-like globulins) and other gamma-globulins in relation to tropical parasitic infections. Lancet 1:843-852, 1966.
- HULDT, G.; LJUNGSTROM, I. & AUST-KETTIS,
 A. Detection by immunofluorescence of antibodies to parasitic agents. Use of class-specific conjugates. Ann. N.Y. Acad. Sci. 254:304-313, 1975.
- 16. KANE, G. J.; MATOSSIAN, R. & BATTY, I. Fluorochrome-labelled anti-immunoglobulin fractions used with stabilized antigen preparations for the assessment of parasitic diseases. Ann. N.Y Acad. Sci. 177:134-145, 1971.
- MATOSSIAN, R. M.; KANE, G. J.; CHANTLER, S. M.; BATTY, I. & SARHADIAN, H. — The specific immunoglobulin in hydatic disease. Immunology 22:423-430, 1972.

- KANAMURA, H. Y.; HOSHINO-SHIMIZU, S. & SILVA, L. C. da Pattern of class-specific fluorescent antibodies according to the stage of the infection in human schistosomiasis mansoni. Rev. Inst. Med. trop. São Paulo 20:76-81, 1978.
- MORIEARTY, P. L. & LEWERT R. M. Delayed hypersentivity in Ugandan schistosomiasis mansoni. III — Examination of serological responses and clinical states. Amer. J. Trop. Med. Hyg. 23:190-196, 1974.
- 19. SILVA, L. C. da Anticorpos e eosinófilos circulantes na esquistossomose mansônica. Contribuição ao estudo de efeitos da quimioterapia [Tese de livre-docência]. São Paulo, Faculdade de Medicina da Universidade de São Paulo, 1974.
- 20. SILVA, L. C. da & FERRI, R. G. Immuno-

- diffusion studies in human schistosomiasis mansoni. II — Localization of antibodies by immunoelectrophoresis. Rev. Inst. Med. trop. São Paulo 7:7-10, 1965.
- WILSON, M.; SULZER A. J. & WALLS, K. W.
 — Modified antigens in the indirect immunofluorescence test for schistosomiasis. Amer. J. Trop.
 Med. Hyg. 23:1072-1076, 1974.

Recebido para publicação em 30/5/1977.