



Resolution Enhancement in Multidimensional Solid-State NMR Spectroscopy of Proteins using Spin-State Selection

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the spectral resolution in the CO dimension to the maximal CT of 9.2 ms. It can be further increased by lengthening t_1 in an additional non-constant acquisition time or by processing using stronger apodization functions and linear prediction (LP). Despite the CT frequency encoding, advanced prediction algorithms like mirror-image LP can not be used here. Indeed, in solid-state NMR, the ^{13}C linewidth is dominated by refocusing interactions¹⁴ and the signal therefore still decays during the CT evolution period.

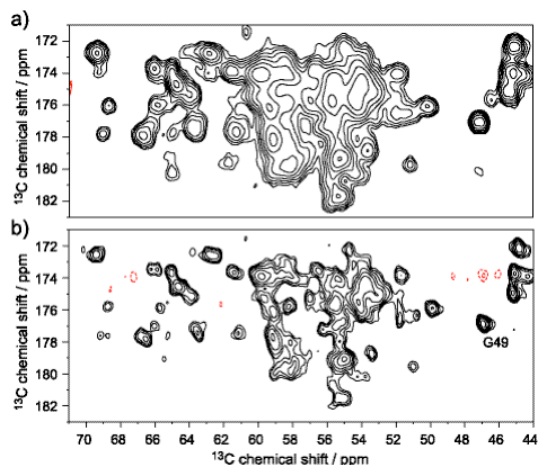


Figure 2. CO- C^α region of the standard (a) and the ($\alpha\alpha$)-spin-state selective (b) PDS spectrum of microcrystalline Crh. The second spectrum was obtained from the linear combination $A1 + B1 + k(A2 + B2)$, with $k = 0.7$. Both experiments were performed on a Bruker Avance spectrometer operating at a ^1H frequency of 500.13 MHz with a 4 mm double-resonance CP/MAS probe. The temperature was set to 269 K, and the MAS frequency was 11 kHz. SPINAL¹⁵ ^1H decoupling was used with $\omega_1 = 78$ kHz, $\tau_{\text{mix}} = 30$ ms, and t_2^{max} was 25 ms. For the standard and the spin-state-selective spectra, a total of 64 and 4×24 transients respectively were accumulated for each of 240 complex points in t_1 . Cosine apodization was applied in both dimensions prior to Fourier transformation. For both spectra, the first contour level was set to 15% of the intensity of the G49 resonance, with a factor of 1.4 between levels.

The efficiency of the spin-state-selective experiment compared to the standard PDS experiment was estimated from measured cross peak intensities to be about 35%. This loss is mainly due to the $2T \approx 9\text{ms}$ IPAP filter. As we have shown recently,⁶ this signal loss is greatly reduced by using high-power ^1H decoupling during the filter delay and faster spinning of the sample. This is possible for protein samples if a better cooling system than we have is available. In addition, the four sub-spectra can be added after appropriate shifting of the spectrum by $\pm J_{\text{CO}^\alpha}/2 \approx 27$ Hz along the CO and/or C^α dimensions, resulting in an additional gain of a factor of two in signal to noise. Therefore we expect that for an optimized experimental setup the spin-state-selective experiment not only increases spectral resolution but also sensitivity.

Figure 3 illustrates how the recorded 2D data sets are combined. The linear combinations $(A1 + \delta \times B1) \pm k(A2 + \delta \times B2)$ yield all four single-transition-to-single-transition correlation spectra ($\alpha\alpha$), ($\alpha\beta$), ($\beta\alpha$), and ($\beta\beta$). For $\tau_{\text{mix}} = 30$ ms, the scaling factor was found to be $k = 0.7$. For spectra with $\tau_{\text{mix}} = 15$ ms, a scaling factor $k = 0.5$ was found.

At longer mixing times some polarization is transferred by PDS from the CO to other side chain carbons (C^β , C^γ , C^δ). Since there is no direct scalar coupling between the CO and the side chain carbons, there is no frequency shift along the detection dimension (ω_2) between the ($\alpha\alpha$) and ($\alpha\beta$), or ($\beta\alpha$) and ($\beta\beta$) sub-spectra. Thus the new experiment provides a simple way of

distinguishing C^α from other side chain carbons (e.g. C^β of Thr and Ser residues) by comparison of sub-spectra.

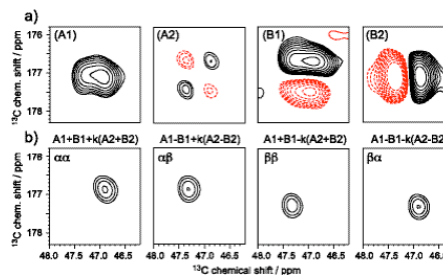


Figure 3. a) Sub-spectra (A1), (A2), (B1), and (B2) recorded using the spin-state-selective CO- C^α -PDS experiment of Figure 1. Experimental details are given in the caption of Figure 2. b) Separation of the four cross-peak transitions obtained by linear combination of the spectra in (a). For clarity, only the G49 cross peak region is shown. Contours are drawn at the same levels for all spectra.

In conclusion, we have introduced a new experiment providing significant resolution enhancement in multi-dimensional solid-state NMR correlation experiments. Resolution enhancement is achieved by using transition-selective excitation and transfer techniques. Spin-state-selective polarization transfer is obtained using standard ZQ solid-state NMR mixing sequences. Similar results are expected for transfer sequences based on DQ rotations. The new experiment can be easily extended to higher-dimensional experiments. In addition, spin-state-selective correlation experiments allow the distinction of ‘direct’ transfer peaks, involving covalently-bound nuclei, and ‘relayed’ transfer peaks. The new experiment is expected to be very useful for the assignment of solid-state NMR spectra of proteins.

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