



REVIEW ARTICLE

Current and novel therapeutic opportunities for systemic therapy in biliary cancer

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Biliary tract cancers (BTCs) are a group of rare and aggressive malignancies that arise in the biliary tree within and outside the liver. Beyond surgical resection, which is beneficial for only a small proportion of patients, current strategies for treating patients with BTCs include chemotherapy, as a single agent or combination regimens, in the adjuvant and palliative setting. Increased characterisation of the molecular landscape of these tumours has facilitated the identification of molecular vulnerabilities, such as *IDH* mutations and *FGFR* fusions, that can be exploited for the treatment of BTC patients. Beyond targeted therapies, active research avenues explore the development of novel therapeutics that target the crosstalk between cancer and stroma, the cellular pathways involved in the regulation of cell death, the chemoresistance phenotype and the dysregulation of RNA. In this review, we discuss the therapeutic opportunities currently available in the management of BTC patients, and explore the strategies that can support the implementation of precision oncology in BTCs, including novel molecular targets, liquid biopsies and patient-derived predictive tools.

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BACKGROUND

Biliary tract cancers (BTCs) comprise a group of rare and aggressive malignancies that arise in the biliary tree, a complex system of ducts accounting for the modification and transfer of bile from the canaliculi, where it is initially generated, to the duodenum.

BTCs include cholangiocarcinoma (CCA), gallbladder cancer (GBC) and ampulla of Vater cancer (AVC). The studies mentioned in this paper often include a combination of all biliary cancers. More recently, dedicated trials to CCAs without GBCs and AVC are being conducted. Biliary ampullary cancers are rare tumours, and to date, no dedicated trials have been set up, so their management follows the indication of the rest of BTCs.

According to the updated anatomical classification, CCA can be further subdivided into intrahepatic (iCCA), perihilar (pCCA) and distal (dCCA) cholangiocarcinoma, which also reflects differences in epidemiology, aetiology, embryology, biology, prognosis and strategy for clinical management. Based on previous data, CCA has

also been classified as iCCA, originating from the biliary tree within the liver, and extrahepatic cholangiocarcinoma (eCCA), which occurs outside the liver parenchyma, and includes perihilar and distal ducts.

Comprehensively, BTCs represent 3% of all gastrointestinal cancers, and are the second most common type of primary liver cancer after hepatocellular carcinoma. Worldwide, the incidence and mortality of BTCs are rising.¹ Although the incidence is much higher in Eastern countries (up to 85 per 100,000 in Thailand) compared with the rest of the world due to the liver flukes, studies show that CCA rates are rising in most Western countries. In the United States, a country with one of the lowest incidence rates, BTC incidence increased with an annual percentage change of 4.36% in the last decade, reaching a value of 1.6 per 100,000.² Multiple risk factors are known to be associated with BTC development, including liver fluke, biliary tract disorders, chronic liver diseases and metabolic syndrome.³

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BTCs are characterised by clinical and pathological heterogeneity, showing a poor response to chemotherapy and dismal prognosis. Due to the asymptomatic behaviour of the disease, most of the patients with BTCs are diagnosed at advanced stage. Only patients with localised disease (20%) benefit from surgical resection. However, the recurrence rate is very high, with a median 5-year survival of <50% in resected patients. For patients with advanced unresectable or metastatic BTCs (approximately 60–80%), systemic therapies are the only potential therapeutic options, and the median overall survival (mOS) is poor, ranging from 6 to 18 months.⁴

In an attempt to improve the clinical outcome of patients with BTCs, shared efforts are moving towards two goals: the identification of molecular alterations and prognostic factors that can guide treatment, and the development of novel therapeutics and combination strategies. We begin this review by outlining the currently available therapeutic strategies for BTC patients before discussing personalised oncology as an approach for the management of these patients.

SYSTEMIC THERAPY FOR CHOLANGIOCARCINOMA: WHERE DO WE STAND?

Adjuvant therapy

The incidence of locoregional and distant relapse remains high in patients with resected BTCs. Until 2017, the use of adjuvant treatment was based on meta-analysis data from small and retrospective Phase 2 studies showing an improvement in OS in two high-risk populations: those with node-positive disease and those with R1 resection.⁵ Subsequently, the results of three prospective randomised clinical trials (RCTs) exploring experimental adjuvant chemotherapy arms in resected BTC patients have been published.^{6–8} In the Japanese BCAT trial,⁶ 226 patients with eCCA were randomly assigned to gemcitabine or observation alone following surgery. The study did not meet its primary endpoint, with no significant differences in mOS (62.3 vs. 63.8 months, respectively; hazard ratio [HR] 1.01, 95% confidence interval [CI] 0.70–1.45; $P = 0.964$) or relapse-free survival (RFS; median 36.0 vs. 39.9 months; HR 0.93, 95% CI 0.66–1.32; $P = 0.693$) between the two groups. The French PRODIGE-12/ACCORD-18 study⁷ also failed to show a benefit in response to the adjuvant combination of gemcitabine and oxaliplatin (GEM/OX) compared with observation alone in patients following resection of CCA and GBC; this study did not meet its primary endpoint, with no benefit in terms of RFS in the doublet-chemo arm (30.4 months vs. 18.5 months in observational arm; HR 0.88, 95% CI 0.62–1.25; $P = 0.48$). The BILCAP study,⁸ conducted in the United Kingdom over a period of 9 years, is the largest study so far involving patients with CCA and patients with GBC. Although the study did not meet its primary endpoint in terms of OS in the intention-to-treat population (ITT), the pre-specified ITT sensitivity analysis adjusted for prognostic factors (nodal status, grade of disease and gender) and the per-protocol population analysis did show a longer mOS in the capecitabine arm (53 months vs. 36 months in the observational arm, HR 0.75, 95% CI 0.58–0.97; $P = 0.028$). In the ITT analysis, median RFS was longer with capecitabine (24.4 months, 95% CI 18.6–35.9) compared with observation (17.5 months, 95% CI 12.0–23.8), but no differences in the risk of relapse were demonstrated after 24 months.

As well as differences in BTC subtypes, heterogeneity in the populations enrolled in these three adjuvant trials with regard to node involvement and resection margins should be noted.⁹ The higher proportion of patients with poor prognostic factors could partly explain why the BILCAP trial is the only study that demonstrates a beneficial effect of adjuvant chemotherapy in patients with resected BTCs. Importantly, it should be noted that the three RCTs differ in sample-size calculation, statistical power of study design, maturity of data and follow-up time. Future efforts in

designing multicentre, randomised Phase 3 trials should aim to standardise risk factors and include them in pre-planned analyses to obtain a more optimal patient selection and study design. The largest ongoing study evaluating the efficacy of adjuvant therapy in patients with BTC is the ACTICCA study, which compares gemcitabine and cisplatin chemotherapy (GEM/CIS) with capecitabine alone (ClinicalTrials.gov: NCT02170090).

First-line chemotherapy

GEM/CIS is currently the standard first-line treatment for patients with advanced BTC (aBTC), based on the results of Advanced Biliary Tract Cancer (ABC-02) Phase 3 and the Japanese BT22 Phase 2 trials, which demonstrated the superiority of this combination compared with gemcitabine monotherapy.^{10,11} However, to improve further on the modest survival benefit conferred by GEM/CIS, other first-line chemotherapy options are under investigation. The FUGA-BT trial reported non-inferiority of gemcitabine plus S1 (a fluoropyrimidine derivative) chemotherapy compared with GEM/CIS, suggesting that this treatment could represent another option for aBTC.¹² Furthermore, a Phase 2 study evaluating nanoliposomal irinotecan in combination with fluorouracil (5-FU)/leucovorin versus GEM/CIS is ongoing.¹³ Beyond doublet therapy, a Phase 2 triplet approach with nanoparticle albumin-bound (nab)-paclitaxel plus GEM/CIS attained the highest mOS (19.2 months) reported in this setting;¹⁴ this combination is currently under evaluation in a randomised Phase 3 study versus GEM/CIS (S1815 SWOG clinical trial).

A 2020 post hoc analysis of the results from prospective, randomly assigned ABC-01/02/03 trials of GEM/CIS shows a longer OS (by ~4 months) of patients with iCCA compared with non-iCCA-BTC patients and suggests—albeit with a low level of evidence due to the small size—a more favourable prognosis of iCCA and iCCA with liver-only disease.¹⁵ Such a difference might be of relevance when assessing the suitability of sequential liver-directed therapies on the OS of these patients. Two Phase 2 trials combining gemcitabine and platinum derivatives with concomitant liver-directed therapies (radioembolisation with yttrium-90 [a technique in which microspheres emit β -radiation to block the supply of blood to the tumour] and intra-arterial infusion) yielded interesting median OS figures (22 and 25 months, respectively).^{16,17} Confirmatory Phase 3 studies of radioembolisation are awaited.

When evaluating OS, it is also important to consider the impact of prognostic factors (also relevant for patient stratification). The post hoc analysis of GEM/CIS pivotal trials^{10,11,15} suggests a prognostic role for Eastern Cooperative Oncology Group (ECOG) performance status (PS), white blood cells, haemoglobin, disease status, bilirubin, neutrophil count and gender, but these data have not yet been confirmed.¹⁵ In a real-life setting, a study conducted by the G.I.Co. (Italian Group of Cholangiocarcinoma) involving 940 Italian patients with aBTC captures ECOG, prior resection, tumour grading, baseline carcinoembryonic antigen and carbohydrate antigen 19.9 as factors that are independently associated with OS.¹⁸ Further studies incorporating putative molecular prognostic factors, such as the fibroblast growth factor receptor (*FGFR*)-2 fusions, are needed to identify genomic prognostic variables that might help to identify prognosis and predict treatment outcomes.

Second-line chemotherapy

The benefit of any second-line treatment for patients with BTC has been unclear until the past year. A systematic review published in 2014 showed that studies available in the second-line setting were of limited quality, with 14 out of 25 eligible studies representing Phase 2 clinical trials and no RCTs being identified.¹⁹ Data from a total of 761 individual patients were reported; the pooled mOS, PFS, response rate (RR) and disease-control rate (DCR) were 7.2 months (95% CI 6.2–8.2), 3.2 months (95% CI 2.7–3.7), 7.7% (95% CI 4.6–10.9) and 49.5% (95% CI 41.4–57.7), respectively.

Table 1. Novel opportunities for targeted therapeutics in biliary cancers: ongoing clinical trials.

| Trial number | Phase | Pathways targeted | Drug | Condition | Line of treatment | Primary outcomes | Secondary outcomes |
|--------------|-------|---------------------------------|------------------|--|-------------------|-----------------------------|---|
| NCT03521219 | 2 | VEGFR2 | Apatinib | aCCA | II | DCR | ORR, OS and PFS |
| NCT03427242 | 2 | VEGFR2 | Apatinib | aBTC | II | PFS | OS |
| NCT02520141 | 2 | VEGF | Ramucirumab | aBTC | II | PFS | CR, PR, SD and OS |
| NCT03377179 | 2 | SK2 | Opaganib/ HCQ | aCCA | II | ORR and DDCR | Safety, tolerability, pharmacokinetics, PFS, DCR and OS |
| NCT03185988 | 2 | HER2 | Trastuzumab | HER2-positive aBTC | II | CR, PR and SD | DCR, PFS, OS, TTR and DoR |
| NCT04209465 | 1/2 | EGFR/ErbB | BDTX-189 | EGFR/ErbB mutant aBTC | II | RP2D and ORR | Safety, tolerability and pharmacokinetics |
| NCT03833661 | 2 | PD-L1; TGFβ Trap fusion protein | M7824 | aBTC | II | ORR | AE, DOR, PFS, OS and pharmacokinetics |
| NCT02908451 | 1 | TAA AG7 | AbGn-107 | aBTC | II | AE and safety | Pharmacokinetics and pharmacodynamics |
| NCT02576431 | 2 | NTRK | Larotrectinib | NTRK fusion aBTC | II | ORR | PFS, OS and DoR |
| NCT02568267 | 2 | NTRK | Entrectinib | NTRK1/2/3, ROS1 and ALK rearrangement aCCA | II | ORR | PFS, DoR and TTR |
| NCT03207347 | 2 | PARP | Niraparib | BAP1 and other DDR pathway- deficient CCA | II | ORR | PFS, OS and DoR |
| NCT03422679 | 1/2 | NOTCH | CB-103 | NOTCH alteration CCA | II | DLT and antitumour efficacy | Pharmacokinetics |
| NCT03873532 | 2/3 | MAPK | Surufatinib | aBTC | II | OS | PFS, DCR and DoR |
| NCT03149549 | 1/2 | TAA CD166 | CX-2009 | aBTC | II | DLT | ORR |

aBTC advanced biliary tract cancer, aCCA advanced cholangiocarcinoma, PFS progression-free survival, OS overall survival, DCR disease-control rate, DOR duration of response, ORR overall response rate, PR partial response, CR complete response, SD stable disease, CBR clinical benefit rate, TTR time to response, AE adverse events, QoL quality of life, DDR DNA-damage response, DLT dose-limiting toxicity, TAA tumour-associated antigen, RP2D recommended Phase 2 dose, HCQ hydroxychloroquine sulfate.

Although the available data suggested that a subpopulation of patients, especially young patients and those with a good PS, could benefit from second-line chemotherapy, this benefit seemed limited, and the evidence was considered to be of insufficient quality (level C) to recommend second-line chemotherapy for aCCA as a standard-of-care strategy.²⁰ One of the main challenges for the completion of adequately powered studies was the fact that, due to the aggressive behaviour of CCA, few patients (ranging from 10 to 40% in different series) are considered to be eligible for second-line treatment.^{18,21–23} In 2019, the results from the ABC-06 clinical trial were reported.²⁴ This Phase 3 study recruited 162 patients diagnosed with aBTC (72% of whom had a diagnosis of CCA) following progression on first-line GEM/CIS chemotherapy. Patients were randomly assigned to active symptom control (ASC, 81 patients) or ASC with FOLFOX (5-FU and oxaliplatin, 81 patients). The study met its primary endpoint, showing a benefit from second-line chemotherapy in terms of OS (adjusted HR 0.69, 95% CI 0.50–0.97; $P = 0.031$). Even though absolute differences in mOS were modest (5.3 months [ASC arm] vs. 6.2 months [ASC + FOLFOX arm]), differences in the survival rate at 6 months (35.5% [ASC arm] vs. 50.6% [ASC + FOLFOX arm]) and 12 months (11.4% [ASC arm] vs. 25.9% [ASC + FOLFOX arm]) were clinically meaningful. Therefore, FOLFOX is currently being considered as standard-of-care second-line chemotherapy for patients with aBTC previously treated with GEM/CIS.

Novel chemotherapy strategies, such as FOLFIRINOX (5-FU, irinotecan and oxaliplatin)²⁵ and etoposide toniribate (EDO-S7.1),²⁶ are being tested in the second-line setting, but their efficacy requires confirmation. The Phase 2 studies NALIRICC (ClinicalTrials.gov: NCT03043547) and NAPOLI-2 (ClinicalTrials.gov: NCT04005339) are currently assessing the nanoliposomal

irinotecan/5-FU/leucovorin versus 5-FU/leucovorin in patients previously treated with gemcitabine-based therapies.

Targeted therapies on the horizon

The molecular landscape of BTCs has begun to emerge over the past decade, offering researchers and clinicians the potential to develop novel molecularly targeted therapies (Table 1). Accordingly, molecular profiling of CCA tumours has become increasingly significant over the past few years due to the identification of potentially druggable molecular alterations, such as mutations in *IDH1/2* and *FGFR2* fusions. Mutations in *IDH1/2* disrupt the normal catalytic activity of isocitrate dehydrogenase 1/2, causing the altered protein to produce a new metabolite 2-hydroxyglutarate (2-HG), which induces several oncogenic changes to cellular metabolism. *FGFR2* fusions contain the intact kinase domain fused to a large number of different partners, including *BICC1*, *AHCYL1*, *TACC3*, *MGEA5* and *PPHLN1*,²⁷ leading to the constitutive activation of the *FGFR2* fusion protein (FFP) and its consequent downstream oncogenic pathways.²⁷ The would-be therapeutic effect of acting on these potentially targetable alterations is currently being evaluated.

In the ClarIDHy Phase 3 trial, 185 patients with *IDH1*-mutant CCA following progression on standard-of-care chemotherapy were randomised to receive the *IDH1* inhibitor ivosidenib or placebo. The primary endpoint was met, with a median PFS of 2.7 versus 1.4 months for patients receiving ivosidenib and for placebo group, respectively (HR 0.37, 95% CI, 0.25–0.54; $P < 0.001$). ITT analysis revealed a mOS of 10.8 months in the experimental group versus 9.7 months in the placebo group.²⁸ Ongoing clinical trials are also exploring the efficacy of poly(ADP ribose) polymerase (PARP) inhibitors in *IDH1/2* mutant iCCA (as *IDH1* mutations render tumours sensitive to PARP inhibition) in order to assess their

Table 2. Inhibitors of *IDH1/2* and *FGFR2* fusions: current clinical trials.

| Trial number | Phase | Pathways targeted | Drug | Condition | Line of treatment | Primary outcomes | Secondary outcomes |
|--------------|-------|-------------------|------------------------------------|---|-------------------|------------------|--|
| NCT03656536 | 3 | <i>FGFR</i> | Pemigatinib | <i>FGFR2</i> rearrangement aCCA | I | PFS | ORR, DoR, DCR, AE and QoL |
| NCT03773302 | 3 | <i>FGFR</i> | Infigratinib (BGJ398) | <i>FGFR2</i> gene fusion aCCA | I | PFS | OS, ORR, DOR, DCR and AE |
| NCT04093362 | 3 | <i>FGFR</i> | Futibatinib (TAS-120) | <i>FGFR2</i> gene arrangement aCCA | I | PFS | ORR, DCR, OS, safety and tolerability |
| NCT04256980 | 2 | <i>FGFR</i> | Pemigatinib | <i>FGFR2</i> rearrangement aCCA | II | ORR | PFS, DOR, DCR and OS |
| NCT03230318 | 2 | <i>FGFR</i> | Derazantinib | <i>FGFR</i> fusions, mutations and amplifications advanced iCCA | II | PFS at 3 months | EORTC QLQ-C30, OS and DOR |
| NCT02150967 | 2 | <i>FGFR</i> | Infigratinib | <i>FGFR</i> alteration aCCA | II | ORR | PFS, OS and DCR |
| NCT02052778 | 1/2 | <i>FGFR</i> | Futibatinib (TAS-120) | <i>FGFR</i> aberration CCA | II | ORR | PFS, OS and DCR |
| NCT04238715 | 2 | <i>FGFR2</i> | E7090 | <i>FGFR2</i> gene fusion aCCA | II | ORR | PFS, DOR, TTR, OS, DCR and CBR |
| NCT02699606 | 2 | <i>FGFR</i> | Erdafitinib | <i>FGFR</i> alteration aCCA | II | ORR | PFS, OS and DCR |
| NCT03684811 | 1/2 | <i>IDH1</i> | FT-2102 | <i>IDH1</i> -R132 mutant iCCA | II | DLT | Pharmacokinetics, pharmacodynamics, AE, PFS, TTP, DOR and OS |
| NCT03212274 | 2 | <i>PARP</i> | Olaparib | <i>IDH1</i> or <i>IDH2</i> mutant CCA | II | ORR | PFS, OS and DoR |
| NCT03878095 | 2 | <i>PARP</i> | Olaparib Ceralasertib (AZD6738) | <i>IDH1</i> and <i>IDH2</i> mutant CCA | II | ORR | PFS, OS, DOR and AE |

aCCA advanced cholangiocarcinoma, iCCA intrahepatic cholangiocarcinoma, PFS progression-free survival, OS overall survival, DCR disease-control rate, DOR duration of response, ORR overall response rate, PR partial response, CR complete response, SD stable disease, CBR clinical benefit rate, TTR time to response, AE adverse events, QoL quality of life.

synthetic lethality and to target *IDH1/2*-related dependencies (ClinicalTrials.gov: NCT03212274 and NCT03878095).

Phase 2 clinical trials showed meaningful clinical benefits of *FGFR* inhibitors in the treatment of chemorefractory iCCA patients carrying *FGFR2* fusions, which constitute the most clinically responsive group of patients. In a Phase 2 trial assessing the pan *FGFR* inhibitor BGJ398/infigratinib,²⁹ the objective RR (ORR) and DCR were 18.8% and 83.3%, respectively, while another pan *FGFR* inhibitor, ARQ087/derazantinib, resulted in an ORR and DCR of 20.7% and 82.8%, respectively, in a Phase 2 trial.³⁰ The FIGHT-202 study tested the *FGFR1–3* inhibitor pemigatinib in 107 patients with *FGFR2* fusions, obtaining an impressive 35.5% ORR, with a median duration of response of 7.5 months and PFS of 6.9 months.³¹ Currently there are several *FGFR* inhibitors that differ with respect to their toxicity and specificity through the target range (*FGFR1–4*) under clinical investigation, including Debio 1347, TAS-120/futibatinib and erdafitinib^{29,30,32–35} (Table 2). Infigratinib, pemigatinib and futibatinib have progressed to Phase 3 evaluation as first-line single agents versus the standard-of-care GEM/CIS (ClinicalTrials.gov: NCT03773302, NCT03656536 and NCT04093362), with the trial results eagerly awaited.³⁶

NOVEL OPPORTUNITIES FOR TARGETED THERAPEUTICS IN BILIARY CANCER

Is there more to know about *FGFR2*-aberrant tumours?

FGFR2 fusion transcripts generated by chromosomal rearrangements are found in about 10–15% of patients with iCCA.³⁷ The efficacy of first-generation tyrosine kinase inhibitors (F-TKIs) in iCCA patients is limited by the emergence of secondary resistance, a major genetic determinant of which is represented by on-target mutations that prevent access of F-TKIs to the *FGFR2* ATP-binding pocket.³⁸ Resistance mutations in FFPs are most often polyclonal. In vitro experiments delineated a drug-sensitivity profile of individual FFP mutants congruent with clinical data: thus, while some mutations cause cross-resistance among different F-TKIs

(e.g. N550K, L618V and K660M mutations reduce binding to both BGJ398 and Debio 1347), others appear to be drug-specific (e.g. M538I impairs binding of Debio 1347, but not BGJ398).³⁸ Interestingly, TAS-120 maintains activity against most resistance mutations detected so far in BGJ398-treated patients, but lacks efficacy against the highly prevalent V565F gatekeeper mutation; Debio 1347, on the other hand, loses activity against most resistance mutations, except V565F.³⁸

Rapidly evolving polyclonal FFP mutations represent a clinical challenge. Sequential administration of mutant-specific F-TKIs informed by next-generation sequencing analysis of circulating tumour DNA has been advocated, but its benefit appears to be limited, given the emergence of several clones.³⁸ An alternative strategy could be to prevent the emergence of resistance mutations by upfront combination therapies that incorporate, in addition to the F-TKI of choice, agents that are capable of targeting dependencies shared by wild-type and TKI-resistant FFPs. FFPs, including those with resistance mutations, are heat-shock protein 90 (HSP90) clients and are therefore stabilised by these chaperones; as such, they undergo swift degradation upon HSP90 inhibition.³⁹ Moreover, F-TKIs and HSP90 inhibitors exert synergistic effects against FFP-transformed cells.³⁹ Notably, as latest-generation HSP90 inhibitors lack the liver and ocular toxicities that have limited the clinical development of earlier drugs in this class, they might therefore deserve consideration in the iCCA field.⁴⁰ Along this line, an emerging paradigm postulates that therapeutic targeting of a driver kinase is more efficacious when combined with the blockade of downstream pathway components.⁴¹

Other actionable alterations in CCA

With the advent of improved technologies, it has become apparent that there are multiple potentially actionable alterations in BTCs. In addition to *FGFR2* fusions and *IDH1* mutations, many other alterations, such as amplification of the receptor tyrosine kinase *c-MET*, targetable with savolitinib,⁴² and overexpression of

the epidermal growth factor receptor (*EGFR*),⁴³ require clinical evaluation, although this will always be challenging because of the low number of patients with these changes. Other important events that require further investigation include activation of the Janus kinase/signal transducer and activator of transcription (*JAK/STAT*) signalling pathway through constitutive activation of *STAT3*, which is estimated to occur in 58–77% of patients with iCCA (depending on inflammation or proliferation biological class, respectively),⁴⁴ and gain-of-function mutations in protein tyrosine phosphatase non-receptor type 3 (*PTPN3*), which have been reported in ~41% of patients.⁴⁵ Moreover, it remains to be seen whether therapeutically inhibiting additional promising targets, such as *HER2*,⁴⁶ *BRAF*⁴⁷ and *BRCA*,⁴⁸ confers a similar benefit to that observed in more common cancers, such as breast (*HER2*), melanoma (*BRAF*) and ovarian malignancies (*BRCA*). Preliminary data from patients with *HER2*-positive aBTC have shown that dual *HER2*-targeted treatment with pertuzumab and trastuzumab has activity in this setting.⁴⁹ The combination of *BRAF* and mitogen-activated protein kinase kinase (*MEK*) inhibitors was also tested in a Phase 1 trial, and showed promising results for CCA patients with the activating *BRAF* V600E mutation.⁴⁷

Nevertheless, there remains a large cohort (~50%) of patients with no currently actionable alteration. For instance, some of the most frequent genetic mutations in CCA comprise the proto-oncogene *KRAS* and the tumour suppressor *TP53*, for which the options are limited (Table 3). To date, despite the large number of potential therapeutic targets identified by molecular profiling, more advanced genomic technologies might be required to reveal novel actionable alterations in these difficult-to-treat cancers.

Mutations in DNA-damage repair (*DDR*) genes are present in about 20% of BTCs, especially in extrahepatic BTCs. In these tumours, *PARP* inhibitors may have a therapeutic role as they counteract the activity of the *PARP* enzyme to repair single-strand DNA breaks. However, the benefit of olaparib monotherapy has been limited in other gastrointestinal cancers; thus, it is likely that

combination treatments will be explored in BTC. *PARP* inhibitors may be combined with immunotherapy (see below), with anti-angiogenic therapies (given that hypoxia can reduce *DDR*) or phosphoinositide 3-kinase (*PI3K*)/*MEK* inhibitors (that are over-activated in BTC and have been associated with secondary resistance to *PARP* inhibition).

Epigenetic alterations have also been described in BTCs.⁵⁰ Treatments aimed at reversing these changes have been studied and shown to be promising, such as the histone deacetylase (*HDAC*) inhibitor resminostat in pretreated BTC patients.⁵¹

Immunotherapy: only for the few?

In contrast to the promising data observed with targeted therapies in molecularly defined patients, immunotherapy (given as a monotherapy) has so far been disappointing in patients with anatomically and molecularly uncharacterised aBTC. One of the largest published immunotherapy studies ever is the KEYNOTE-158 Phase 2 clinical trial, which assessed the efficacy of pembrolizumab, an antibody that targets the immune-checkpoint protein programmed death-1 (*PD-1*), in patients with previously treated solid tumours, including those of the biliary tract. The subgroup analysis of 104 patients with aBTC treated with pembrolizumab revealed an RR of 5.8% with a median PFS of 2 months and a mOS of 9.1 months, regardless of *PD-L1* positivity (membranous *PD-L1* expression in ≥1% of tumours and associated inflammatory cells or positive staining in stroma).⁵² Consistent with other studies, pembrolizumab showed durable antitumour activity among the few responsive patients.

So far, a high degree of microsatellite instability (*MSI-High* [*H*]), occurring in 1–3% of CCA patients (with germline mutations in mismatch repair genes), is the only marker that appears to be predictive of clinical response to immunotherapy. The KEYNOTE-158 study evaluating pembrolizumab in previously treated patients with advanced non-colorectal *MSI-H*/deficient mismatch repair (*dMMR*) cancer showed an ORR of 40.9%, median PFS of

Table 3. Targetable mutations in CCA.

| Molecular alteration | Incidence | Primary tumour site | Possible agents | ESCAT | |
|----------------------------------|---|---------------------|-----------------|--|-------|
| 'Established' targets and drugs | <i>FGFR2</i> fusion | 10% | iCCA | Futibatinib (TAS-120), ³⁸ Derazantinib (ARQ087), ³⁰ Infigratinib (BGJ398), ³⁶ Erdafitinib ³³ and Pemigatinib ³¹ | II-B |
| | <i>IDH1</i> mutation | 10% | iCCA | Ivonesidenib ²⁸ FT-1202 | I-B |
| | <i>HER2</i> amplification | 10% | eCCA/GBC | Pertuzumab–Trastuzumab ⁴⁶ | – |
| | <i>HER2</i> mutation | 5% | eCCA/GBC | Neratinib–Trastuzumab ⁴⁶ Pertuzumab–Trastuzumab ⁴⁶ | III-A |
| | <i>MSI-H</i> | 1–3% | eCCA/iCCA/GBC | Durvalumab Pembrolizumab | II-B |
| | <i>BRAF</i> V600E mutation | 3% | eCCA/iCCA/GBC | Dabrafenib–trametinib ⁴⁷ | III-A |
| | <i>BRCA2</i> mutation | 3% | eCCA/iCCA/GBC | Olaparib ⁴⁸ | III-A |
| | <i>EGRF</i> mutation/ amplification | 3% | eCCA/iCCA/GBC | Osimertinib | III-A |
| 'Experimental' targets and drugs | <i>BRAF</i> non-V600E mutation | 1.5% | eCCA/iCCA/GBC | Encorafenib–Binimetinib | IV-A |
| | <i>c-MET</i> amplification | 3% | eCCA/iCCA/GBC | Savolitinib ⁴² | IV-A |
| | <i>BAP1</i> / <i>BRCAness</i> <i>DDR</i> alterations (<i>SMARCA4</i> ; <i>ARID1A</i>) | 10% | eCCA/iCCA/GBC | Olaparib | IV-A |
| | <i>EGFR</i> amplification | <5% | eCCA/iCCA/GBC | Osimertinib ⁴³ | IV-A |
| | <i>NTRK</i> fusions/ <i>ROS1</i> | < 5% | eCCA/iCCA/GBC | Larotrectinib–Entrectinib | IV-A |
| | <i>PIK3CA</i> mutation | < 10% | eCCA/iCCA/GBC | Everolimus–Sirolimus | IV-A |

GBC gallbladder cancer, iCCA intrahepatic cholangiocarcinoma, eCCA extrahepatic cholangiocarcinoma, ESCAT ESMO Scale for Clinical Actionability of molecular Targets, *DDR* DNA-damage repair, *MSI-H* microsatellite instability-high.

4.2 months and mOS of 24.3 months in the BTC cohort of 22 patients,⁵³ demonstrating a clinical benefit of pembrolizumab among these patients, consistent with results from other patients with previously treated MSI-H/dMMR non-colorectal cancer assessed in the study.

In order to increase the efficacy of immunotherapy in BTCs, different therapeutic combinations are currently being tested (Table 4). One approach includes the combination of immunotherapy and chemotherapy. Early clinical data from the combination of nivolumab with GEM/CIS as a first-line treatment showed signs of antitumour activity, with an RR of 37%, a median PFS of 4.2 months and mOS of 15.4 months.⁵⁴ This concept of immunotherapy–chemotherapy combination is currently further evaluated in Phase 3 studies, such as TOPAZ-1 and KEYNOTE-966, in which patients are being treated with GEM/CIS alone or with durvalumab (which targets PD-L1, the PD-1 ligand) or pembrolizumab, respectively.

The use of immunotherapy together with anti-angiogenic agents has shown high efficacy against hepatocellular carcinoma, but has not so far been successful in the treatment of BTC. In one study, pembrolizumab plus ramucirumab, which inhibits vascular endothelial growth factor (VEGF)-induced angiogenesis, showed limited efficacy in patients with previously treated advanced/metastatic BTC (only 4% in 26 patients), with a mOS of 6.4 months and median PFS of 1.6 months.⁵⁵ Similar to VEGF signalling, targeting the transforming growth factor β (TGF- β) pathway has been shown to promote tumour immunosuppression and, based on encouraging efficacy observed in a Phase 1 study, M7824, a first-in-class bifunctional fusion protein comprising two extracellular domains of TGF- β RII (a TGF- β 'trap') fused to a human IgG1 monoclonal antibody against PD-L1, is currently being evaluated in combination with GEM/CIS as a first-line therapy for BTC (clinical trial.gov: NCT04066491). Moreover, the immunogenicity resulted from the increased mutational burden (and thus the neoantigens) caused by the mechanism of action of PARP inhibitors has provided the rationale to assess them with immunotherapy (clinical trial.gov: NCT03639935).

POTENTIAL OPPORTUNITIES TO REVERSE CHEMORESISTANCE IN BILIARY CANCERS

The molecular mechanisms of chemoresistance

The mechanisms of chemoresistance to anticancer drugs,^{56,57} which are classified into seven groups (Fig. 1), can already be present in tumours before the start of treatment (primary resistance), although they usually arise in response to the pharmacological challenge (secondary resistance). Drug resistance occurs due to changes in the expression levels or/and the appearance of genetic variants in genes encoding mechanisms of chemoresistance.

Several transporters play a role in resistance by influencing the bioavailability of drugs, both positively and negatively. For instance, upregulation of the human equilibrative nucleoside transporter 1 (hENT1) in CCA cells is associated with a better response to gemcitabine in patients with resected CCA⁵⁸ and aBTC,⁵⁹ and impaired expression of the organic cation transporter 1 (OCT1) is thought to mediate the poor response to cisplatin and the multikinase inhibitor sorafenib.^{60,61} By contrast, ATP-binding cassette proteins, such as MDR1, MRP1 and MRP3, which are highly expressed in CCA, are able to export a wide variety of antitumour drugs out of cells, thereby limiting their effect. Interestingly, high *MRP1* mRNA levels correlate inversely with OS after the treatment of iCCA.⁶² Chemical modification of some conventional chemotherapy drugs has enabled these compounds to enter the cancer cell independently of the above-mentioned membrane transporters, and may represent a good strategy to overcome chemoresistance (clinicaltrials.gov: NCT04163900).

Regarding detoxifying enzymes, the high expression of aldehyde dehydrogenase 1 family, member A3 (ALDH1A3) correlates with a lower response to gemcitabine-based therapy in patients with advanced iCCA,⁶³ and glutathione S-transferase-pi (GSTP1), also frequently overexpressed in CCA, has similarly been associated with resistance to cisplatin and other alkylating agents.⁶⁴ Downregulation of metallothioneins is accompanied by a better response to cisplatin.⁶⁵ Other components involved in the mechanisms of chemoresistance include orotate phosphoribosyl transferase (OPRT), a key enzyme in the activation pathway of 5-FU;⁶⁶ accordingly, increased expression of OPRT confers increased sensitivity to 5-FU. By contrast, increased expression of thymidylate synthase (TS), which is involved in DNA synthesis and normally inhibited by 5-FU metabolites, results in lower sensitivity to 5-FU.⁶⁷

In terms of apoptosis/survival genes, CCA resistance to the EGFR inhibitor erlotinib has been associated with the upregulation of EGFR in a feedback loop.⁶⁸ Moreover, increased expression of the p53-inducible ribonucleotide reductase (*p53R2*) gene, which is required for normal DNA repair, correlates with, and has been used to predict, gemcitabine resistance.⁶⁹ Downregulation of the pro-apoptotic protein NK4, an antagonist of hepatocyte growth factor (HGF), is responsible for acquired resistance to 5-FU in CCA,⁷⁰ and downregulation of Bax and upregulation of Bcl-2 contribute to evasion of apoptosis in CCA cells resistant to gemcitabine.⁷¹ Furthermore, overexpression of anti-apoptotic proteins, such as extracellular signal-regulated kinase (ERK) and Bcl-2, and overactivation of PI3K/AKT and RAF/MEK/ERK pathways, have been identified to be associated with CCA chemoresistance.

Changes in the tumour microenvironment, such as hypoxia, extracellular fluid acidification and the presence of autocrine and paracrine signals, also affect chemoresistance. Upregulation of the octamer-binding transcription factor 4 (Oct4) in acidic conditions has been shown to be associated with CCA resistance to gemcitabine.⁷² Furthermore, the expression of interleukin (IL)-6 and TGF- β 1 through an autocrine loop involving Smad4 has been involved in the resistance to gemcitabine by inducing epithelial–mesenchymal transition (EMT).⁷³ Moreover, high expression of the mobility group A1 (HMG A1) protein, which promotes EMT, also confers resistance to gemcitabine.⁷⁴ In conclusion, although there continues to be an urgent need to advance our understanding of the mechanisms of chemoresistance, the situation in CCA is starting to be clarified, and novel targets that mediate the contribution of tumour microenvironment in chemoresistance started to be identified for the development of therapeutics that could be clinically investigated.

MicroRNAs as mediators of chemoresistance and potential RNA therapeutics

MicroRNAs (miRNAs or miRs) are single-stranded non-coding RNAs (18–24 nucleotides) that function as post-transcriptional master regulators to modulate the expression of many genes.⁷⁵ Altered miRNA profiles have been described in many tissues and cells under pathological circumstances, including in CCA,^{75,76} and many miRNAs have been implicated in chemoresistance in CCA patients. For instance, miR-21 is highly expressed in CCA cells compared with non-malignant cells, and its experimental inhibition sensitised cells to gemcitabine through the inhibition of phosphatase and tensin homologue (*PTEN*) in vitro and in vivo,⁷⁷ resulting in decreased PI3K signalling.

Downregulation of miR-200b/c has been reported in CCA, and its enforced expression restores 5-FU sensitivity in CCA cells.⁷⁸ Similarly, miR-29b, miR-205 and miR-221 are downregulated in gemcitabine-resistant CCA cells, but their experimental overexpression restores gemcitabine sensitivity.⁷⁹ The levels of miR-320, which targets the anti-apoptotic protein myeloid cell leukaemia 1 (*MCL1*) and contributes to 5-FU resistance, are diminished in iCCA.⁸⁰ Levels of miR-106b are reduced in

Table 4. Immunotherapy combinations: ongoing clinical trials.

| Trial number | Phase | Pathways targeted | Drug | Indication | Line of treatment | Primary outcomes | Secondary outcomes |
|--------------|-------|---|---|--------------------------|-------------------|-------------------------------|-------------------------------------|
| NCT04027764 | 2 | PD-1, chemotherapy | Toripalimab/S1/Albumin Paclitaxel | aBTC | I | ORR | PFS, DCR and OS |
| NCT03796429 | 2 | PD-1, chemotherapy | Toripalimab/Gemcitabine-S1 | aBTC | I | PFS and OS | ORR and safety |
| NCT04191343 | 2 | PD-1, chemotherapy | Toripalimab/GEMOX | aBTC | I | ORR | NA |
| NCT03486678 | 2 | PD-1, chemotherapy | SHR-1210/GEMOX | aBTC | I | PFS and AEs | ORR and OS |
| NCT03111732 | 2 | PD-1, chemotherapy | Pembrolizumab/capecitabine/oxaliplatin | aBTC | II | 5-month survival | ORR, OS, safety and tolerability |
| NCT03785873 | 2 | PD-L1, chemotherapy | Nivolumab/Nal-irinotecan/5-FU, leucovorin | aBTC | II | Phase 1b: DLT; Phase 2: PFS | AE, ORR and OS |
| NCT04004234 | 1/2 | PD-1, chemotherapy | Manganese/anti-PD-1/nab-paclitaxel-gemcitabine | aBTC | I/II | AE and PFS | DCR, ORR and OS |
| NCT03478488 | 3 | PDL-1, chemotherapy | KN035/GEMOX | aBTC | I | OS | PFS and ORR |
| NCT04003636 | 3 | PD-1, chemotherapy | Pembrolizumab/GEM/CIS | aBTC | I | PFS and OS | ORR, DOR and AE |
| NCT03486678 | 2 | PD-1, chemotherapy | SHR-1210/GEMOX | aBTC | I | PFS | ORR |
| NCT03875235 | 3 | PDL-1, chemotherapy | Durvalumab or placebo and GEM/CIS | aBTC | I | OS | PFS, ORR and DoR |
| NCT03046862 | 2 | PDL-1, CTLA-4, chemotherapy | Durvalumab/Tremelimumab/GEM/CIS | aBTC | I | RR | DCR, PFS, DoR and OS |
| NCT02834013 | 2 | PD-1 and CTLA-4 | Nivolumab/ipilimumab | aBTC | II | ORR | Safety, OS, PFS and DCR |
| NCT03849469 | 1 | PD-1, CTLA-4 and LAG-3 | XmAb [®] 22841/Pembrolizumab | iCCA | II | Safety and tolerability | NA |
| NCT03092895 | 2 | PD-1, VEGF, chemotherapy | SHR-1210/Apatinib, GEMOX or FOLFOX | aBTC | I/II | Safety and tolerability | OS, PFS, DCR, and DoR |
| NCT04211168 | 2 | PD-1, VEGF | Toripalimab/Lenvatinib | aBTC | II | ORR and AEs | OS, PFS, OS and CBR |
| NCT03797326 | 2 | PD-1, VEGF | Pembrolizumab/Lenvatinib | Pretreated solid tumours | II | ORR and AEs | DCR, PFS and OS |
| NCT03895970 | 2 | PD-1, VEGF | Pembrolizumab/Lenvatinib | aBTC | II | ORR, DCR and PFS | OS and DoR |
| NCT04066491 | 2/3 | PD-L1; TGFβ trap fusion protein, chemotherapy | GEM/CIS with or without Bintrafusp Alfa (M7824) | aBTC | I | DLT and OS | DoR, AE and pharmacokinetics |
| NCT03937895 | 1/2 | PD-1, SMT-NK | Pembrolizumab/allogeneic NK cell | aBTC | II | DLT and ORR | TTP and toxicity |
| NCT04057365 | 2 | PD-1, DKK1 | Nivolumab/DKN-01 | aBTC | II | ORR (CR and PR) | PFS and OS |
| NCT03250273 | 2 | PD-1, histone deacetylase inhibitor | Nivolumab/Entinostat | aBTC | II | ORR | PFS, OS and DoR |
| NCT03639935 | 2 | PD-1, PARP | Nivolumab/Rucaparib | aBTC | II | 4-month survival and response | CR, PR, SD, PFS and OS |
| NCT03475953 | 1/2 | PD-1, MAPK | Avelumab/Regorafenib | Metastatic solid tumours | I/II | RP2D and antitumour activity | MTD, DLT and toxicity |
| NCT03257761 | 1 | PDL-1, DNMT inhibitor | Durvalumab/Guadecitabine (SGL-110) | aCCA | II | DLT and ORR | Safety and tolerability, PFS and OS |
| NCT03475953 | 1/2 | PD-1, MAPK | Avelumab/Regorafenib | Metastatic solid tumours | I/II | RP2D and antitumour activity | MTD, DLT and toxicity |

Combinations of immunotherapy with chemotherapy and other agents.

PFS progression-free survival, OS overall survival, DCR disease-control rate, ORR overall response rate, PR partial response, CR complete response, SD stable disease, DOR duration of response, CBR clinical benefit rate, DLT dose-limiting toxicities, MTD maximum tolerated dose, AE adverse events, PARP poly-ADP ribose polymerase, GEMOX gemcitabine + oxaliplatin, GEM/CIS gemcitabine + cisplatin, FOLFOX fluorouracil + folic acid + oxaliplatin.

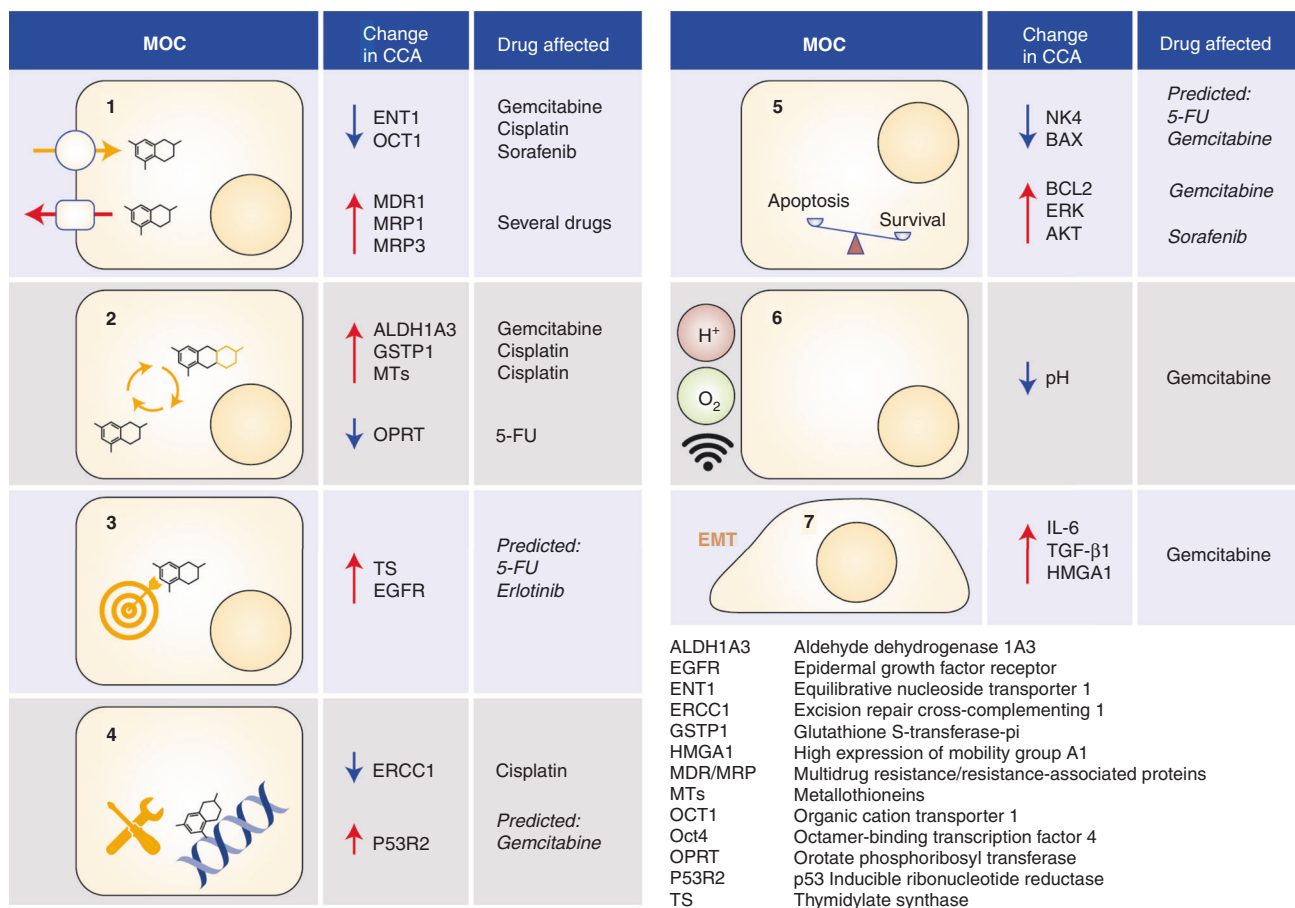


Fig. 1 Schematic representation of the molecular mechanisms of chemoresistance, of which there are seven depicted.^{56,57} (1) Changes in the expression/function of transport proteins involved in drug uptake or efflux. (2) A reduction in the intracellular amount of active drugs due to changes in enzymes involved in metabolism. (3) Changes in the molecular targets of anticancer agents. (4) An increased ability of tumour cells to repair drug-induced DNA damage. (5) Decreased expression/function of pro-apoptotic factors or enhanced expression/function of anti-apoptotic proteins. (6) Changes in tumour-cell microenvironment conditions that affect the effectiveness of drugs. (7) Induction of epithelial-mesenchymal transition (EMT).

5-FU-resistant CCA cells, but the experimental overexpression of this miRNA re-sensitises them to 5-FU, mainly through the modulation of *Zbtb7a*, a proto-oncogenic transcription factor.⁸¹ miR-130a-3p levels mediate resistance to gemcitabine by targeting the expression of another transcription factor, peroxisome proliferator-activated receptor (*PPARG*).⁸² Experimental overexpression of *OCT1* in eCCA and iCCA cells enhanced both the uptake and cytotoxic effects of sorafenib. Notably, miR-141 and miR-330 have been shown to target *OCT1*, but the relevance of the modulation of these miRNAs to sorafenib resistance remains to be unveiled.⁶¹ Functional high-throughput approaches combined with analyses of human tissues have identified miR-1249 as a driver of the expansion of the CD133⁺ subpopulation that is responsible for primary and secondary resistance of CCA cells to cisplatin and gemcitabine.⁷⁶

As the next step for all these findings, it is imperative to evaluate the relevance of these miRNAs in vivo and to correlate their levels with resistance to therapy in patients. Although miRNA-based therapies are already under development, much work needs to be performed in the next few years to improve strategies to synthesise artificial miRNAs and miRNA inhibitors for clinical implementation. It is pivotal to develop and improve new delivery techniques that might help to achieve the best therapeutic efficacy while minimising potential toxic effects.

Targeting death to improve life

Regulated cell death pathways are central in chronic liver disease progression, where the lack of a balance between cell death and regeneration has been shown to lead to carcinogenesis. Failure of regulated cell death in hepatocytes and cholangiocytes is a pivotal step in malignant transformation. This unique relationship between cell death and liver cancer reflects the importance of chronic damage and inflammation, with the release of several mediators that have oncogenic effects. The balance between different types of regulated cell death might influence the type of liver cancer that eventually develops. For instance, a necroptotic microenvironment with high cytokine levels can promote cholangiocarcinogenesis by activating specific oncogenes, while an apoptotic environment appears to increase the risk of hepatocellular carcinogenesis.⁸³ Moreover, a dysregulated equilibrium between anti-apoptotic and pro-apoptotic signals with evasion of both intrinsic and extrinsic apoptosis is a key contributor to the resistance of liver cancer to antitumour drugs, especially in patients with CCA.⁸⁴ The apoptotic mitochondrial pathway is suppressed by overexpression of anti-apoptotic Bcl-2 family proteins, such as Bcl-2⁸⁵ or Mcl-1⁸⁶ in conjunction with downregulation of pro-apoptotic Bcl-2 proteins like Bax.⁸⁷ Similarly, impaired caspase activation caused by overexpression of inhibitors of apoptosis proteins (IAPs) such as XIAP⁸⁸ or survivin,⁶⁰ or abnormal function of death receptors such as Fas

(CD95) and DR4/DR5, contributes to the chemoresistant phenotype in CCA cells.

These mechanisms are also regulated by the surrounding microenvironment.⁸⁴ Indeed, cancer-associated fibroblasts (CAFs) are key cells that support the growth of liver tumours, and are sensitised to apoptotic cell death in a characteristic state termed 'apoptotic priming'.⁸⁹ Pro-apoptotic compounds, such as BH3 mimetics, are being used to exploit this apoptotic priming with encouraging results, reducing tumour growth and metastasis in experimental CCA.⁸⁹ Finally, activation of necroptosis also seems to play a relevant role in CCA by sensitising cells to standard chemotherapy, suggesting novel necroptosis-based therapeutic strategies for CCA patients. Exploring all these different mechanisms of regulated cell death will not only help to understand the powerful mechanisms of chemoresistance, but might also reveal novel opportunities for therapeutic intervention.

Targeting the interaction with the microenvironment

CCA is characterised by marked abundance of tumour stroma, a bioactive connective tissue that not only physically negatively influences drug delivery, but also crosstalks with cancer cells for the activation of a chemoresistant phenotype.⁹⁰ The CCA stroma consists of cancer-associated endothelial cells, CAFs and inflammatory cells—including tumour-associated macrophages (TAMs), neutrophils, natural killer (NK) and T cells—dispersed in a bioactive specialised extracellular matrix (ECM).⁹¹ CAFs are mainly responsible for mediating the composition of the ECM and crosstalk with CCA cells by secreting paracrine factors, such as TGF- β and platelet-derived growth factor (PDGF). Among CCA-infiltrating immune cells, TAMs exert a pivotal role in cancer-related inflammation by promoting tumour-cell proliferation, angiogenesis, matrix turnover and suppression of the adaptive immune response. M2-polarised TAMs communicate in particular with chemoresistant CCA cancer stem cells by releasing numerous soluble mediators, including reactive nitrogen intermediates, cytokines (IL-4, IL-6 and IL-10), chemokines (chemokine ligand [CCL] 17 and CCL18) and metalloproteinases (matrix metalloproteinase [MMP] 9). Together, TAMs and CCA cells create a tumoral niche that constitutes a potential target for therapy. Following the release of CCL2 by tumour cells and TAMs, cytotoxic T lymphocytes acquire CD4/CD25 expression and become immunosuppressive regulators (Treg cells).⁹² By producing TGF- β and IL-10, Treg cells contribute to an immunosuppressive environment through the inhibition of cytotoxic T cells and NK cells. Moreover, by selective binding, Treg cells make IL-2 inaccessible, thus inhibiting the activation of additional immune cells.⁹² Enrichment of Treg cells has also been associated with chemoresistance in BTC.⁹³

As well as cells in the tumour microenvironment, there are other microenvironmental factors linked to the specialised biomatrix components that can significantly impact the behaviour of cancer cells, such as hypoxia, exosomes, proliferative factors and inflammatory cytokines (TGF- β and VEGF).⁹¹ All these factors play different roles in CCA progression, and might be considered as potential targets for therapy. Nevertheless, exploring the dynamics of immunosuppressive cell subpopulations and their interactions with and within the tumour microenvironment will be essential for a better understanding of drug resistance and the subsequent design of novel strategies for innovative anti-CCA therapies.

NOVEL THERAPEUTIC STRATEGIES FOR PERSONALISED MEDICINE

Personalised oncology in BTC

Over the past decade, genomic sequencing technologies have helped to shed light on the molecular landscape of BTCs.^{37,94} However, despite the remarkable steps taken to

unravel the molecular complexity of this heterogeneous disease, the emerging knowledge has only partly been translated into improved clinical management, and hence further studies are needed.

Retracing the path to precision oncology, Verlingue et al.⁹⁴ have demonstrated a tumour-centric approach based on high-throughput genomic analysis of DNA extracted from tumour biopsy samples, selecting potential druggable alterations to match the available target treatments in previously treated BTCs. The prospective MOSCATO-01 trial was successful in determining an outcome improvement (mOS and PFS) in this cohort compared with patients not oriented to molecular targeted agents.⁹⁴ Although preliminary, these results, together with the high frequency of *IDH1/2* and *FGFR2* genetic aberrations confirmed in the trial, have laid the foundation for further investigations. However, as a number of additional targetable molecular alterations have been identified, there is an increasing need to implement our current genetic profiling technologies in clinical practice in order to tailor therapy more appropriately in patients with multiple driver aberrations.

With this information in mind, in the I-PREDICT prospective study, Sicklick et al.⁹⁵ explored the safety and feasibility of a multidrug combination treatment based on a matching score system combining actionable molecular alterations with a corresponding available target therapy or therapies. The most represented population in the study was gastrointestinal refractory tumours (42.2%), including aBTCs. In this study, the 'matching score' rate was higher than in previous studies, with 49% of patients receiving multidrug regimens. The highest matching score rate was associated with significantly improved disease-control rates, as well as longer PFS and OS rates, compared with patients receiving therapy matched to fewer genomic alterations.⁹⁵ Therefore, the current clinical trial paradigm, focused on finding common genomic alterations in patients and targeting them with a single agent, might need to be revised in favour of more tailored combination therapies for specific genetic alterations.

Novel strategies to implement individualisation of treatment: liquid biopsies and patient-derived models

Up to 50% of BTCs are expected to be eligible for targeted therapies, and it has therefore been suggested that genomic profiling is incorporated into routine clinical practice. One of the limiting issues for implementing personalised oncology in BTCs is the lack of tissue for molecular analyses, especially for those BTCs that are diagnosed through cytological sampling. However, this issue might be overcome by the use of liquid biopsies. Mody et al.⁹⁶ presented their experience with a targeted next-generation sequencing panel of 73 genes from the plasma of >120 patients with aBTC. The assessment of molecular alterations was feasible in cell-free DNA (cfDNA), and identification of therapeutically relevant alterations was also successful (*BRAF* and *IDH1/2* mutations, *ERBB2* amplification and *FGFR2* fusions). The limitation of this study was the preponderance of iCCA cases in this cohort, for which lack of tissue is not usually a problem.⁹⁶ Preliminary evidence from only ten patients has demonstrated the possibility of using bile as a source for deep DNA sequencing, showing that cfDNA in bile consists of longer fragments than cfDNA in plasma (with potential higher quality of DNA sequencing), and that there is high correspondence between the mutational profile in bile and BTC tissue.⁹⁷ Further studies are warranted to assess whether bile might be a suitable source of cfDNA for use in the implementation of personalised oncology in patients with advanced pCCA and dCCA. Circulating tumour cells (CTC) are an alternative approach, but to date, low levels of CTC have been detected in BTC limiting their clinical applicability.⁹⁸

DNA sequencing can support precision oncology by identifying targetable molecular alterations. However, it is of no help for

guiding treatment decisions in the case of drugs for which predictive biomarkers have not been identified, such as chemotherapeutic compounds or multityrosine kinases. Patient-derived xenografts (PDXs) have been used for this purpose, but their clinical applicability may be limited by costs and timeframe. Patient-derived organoids (PDOs) are *ex vivo*, organ-like, three-dimensional structures derived from individual patient cells that could be used to predict response to compounds independently on the presence of a molecular biomarker. Notably, cancer PDOs mimic the structure and genomic heterogeneity of their host tumours, and have been demonstrated to mimic in a dish the drug response observed in patients,⁹⁹ generating excitement on the potential use of these PDOs as predictive tools. Growing evidence is supporting the feasibility of establishing biliary cancer PDOs.¹⁰⁰ However, the success rate for generating PDOs from different subtypes of biliary cancer is not yet clear, and so more studies are warranted before this approach can be used to support individualised oncology in patients with BTCs. The next few key steps to validate and promote the use of organoids as clinically relevant tools for the study of biliary cancers include the generation of characterised models representing the different CCA subtypes (intrahepatic, perihilar and distal) and the establishment of a collaborative organoid biobank.

CONCLUSIONS

The current guidelines indicate the use of first-line chemotherapy with cisplatin and gemcitabine in aBTC, followed by FOLFOX chemotherapy. Novel targeted therapies (*IDH* and *FGFR* inhibitors) are being considered for iCCA with selected molecular alterations. An ever-increasing number of molecular alterations is being identified, with different BTC subtypes showing specific molecular profiles. Beyond the role of standard chemotherapy, this approach paves the way to design molecular-orientated clinical trials in which different BTC subtypes can be matched to different targeted inhibitors. One common difficulty encountered when studying rare diseases is the low number of cases that can be investigated in a single institution, and this was indeed the case for BTCs until international CCA-dedicated associations were established, with contributions from both basic and clinical researchers in an attempt to join efforts, skills, information and biological samples to improve research in CCA. Although the situation regarding the available therapeutic options in BTC patients is still limited at present, the increased interest in CCA research and the rapidly growing amount of information in the field support a more optimistic horizon in the near future.

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REFERENCES

1. Khan, S. A., Tavolari, S. & Brandi, G. Cholangiocarcinoma: epidemiology and risk factors. *Liver Int.* **39**, 19–31 (2019).
2. Saha, S. K., Zhu, A. X., Fuchs, C. S. & Brooks, G. A. Forty-year trends in cholangiocarcinoma incidence in the U.S.: intrahepatic disease on the rise. *Oncologist* **21**, 594–599 (2016).
3. Kim, V. N., Han, J. & Siomi, M. C. Biogenesis of small RNAs in animals. *Nat. Rev. Mol. Cell Biol.* **10**, 126–139 (2009).
4. Bridgewater, J., Lopes, A., Palmer, D., Cunningham, D., Anthoney, A., Maraveyas, A. et al. Quality of life, long-term survivors and long-term outcome from the ABC-02 study. *Br. J. Cancer* **114**, 965–971 (2016).
5. Horgan, A. M., Amir, E., Walter, T. & Knox, J. J. Adjuvant therapy in the treatment of biliary tract cancer: a systematic review and meta-analysis. *J. Clin. Oncol.* **30**, 1934–1940 (2012).
6. Ebata, T., Hirano, S., Konishi, M., Uesaka, K., Tsuchiya, Y., Ohtsuka, M. et al. Randomized clinical trial of adjuvant gemcitabine chemotherapy versus observation in resected bile duct cancer. *Br. J. Surg.* **105**, 192–202 (2018).
7. Edeline, J., Benabdelghani, M., Bertaut, A., Watelet, J., Hammel, P., Joly, J.-P. et al. Gemcitabine and oxaliplatin chemotherapy or surveillance in resected biliary tract cancer (PRODIGE 12-ACCORD 18-UNICANCER GI): a randomized phase III study. *J. Clin. Oncol.* **37**, 658–667 (2019).

8. Primrose, J. N., Fox, R. P., Palmer, D. H., Malik, H. Z., Prasad, R., Mirza, D. et al. Capecitabine compared with observation in resected biliary tract cancer (BILCAP): a randomised, controlled, multicentre, phase 3 study. *Lancet Oncol.* **20**, 663–673 (2019).
9. Lamarca, A., Edeline, J., McNamara, M. G., Hubner, R. A., Nagino, M., Bridgewater, J. et al. Current standards and future perspectives in adjuvant treatment for biliary tract cancers. *Cancer Treat. Rev.* **84**, 101936 (2020).
10. Valle, J., Wasan, H., Palmer, D. H., Cunningham, D., Anthoney, A., Maraveyas, A. et al. Cisplatin plus gemcitabine versus gemcitabine for biliary tract cancer. *N. Engl. J. Med.* **362**, 1273–1281 (2010).
11. Okusaka, T., Nakachi, K., Fukutomi, A., Mizuno, N., Ohkawa, S., Funakoshi, A. et al. Gemcitabine alone or in combination with cisplatin in patients with biliary tract cancer: a comparative multicentre study in Japan. *Br. J. Cancer* **103**, 469–474 (2010).
12. Morizane, C., Okusaka, T., Mizusawa, J., Katayama, H., Ueno, M., Ikeda, M. et al. Combination gemcitabine plus S-1 versus gemcitabine plus cisplatin for advanced/recurrent biliary tract cancer: the FUGA-BT (JCOG1113) randomized phase III clinical trial. *Ann. Oncol.* **30**, 1950–1958 (2019).
13. Perkhof, L., Berger, A. W., Beutel, A. K., Gallmeier, E., Angermeier, S., Fischer von Weikersthal, L. et al. Nal-IRI with 5-fluorouracil (5-FU) and leucovorin or gemcitabine plus cisplatin in advanced biliary tract cancer - the NIFE trial (AIO-YMO HEP-0315) an open label, non-comparative, randomized, multicenter phase II study. *BMC Cancer* **19**, 990 (2019).
14. Shroff, R. T., Javle, M. M., Xiao, L., Kaseb, A. O., Varadhachary, G. R., Wolff, R. A. et al. Gemcitabine, cisplatin, and nab-paclitaxel for the treatment of advanced biliary tract cancers. *JAMA Oncol.* **5**, 824–830 (2019).
15. Lamarca, A., Ross, P., Wasan, H. S., Hubner, R. A., McNamara, M. G., Lopes, A. et al. Advanced intrahepatic cholangiocarcinoma: post hoc analysis of the ABC-01, -02, and -03 clinical trials. *JNCI J. Natl. Cancer Inst.* **112**, 200–210 (2019).
16. Edeline, J., Toucheffeu, Y., Guiu, B., Farge, O., Tougeron, D., Baumgaertner, I. et al. Radioembolization plus chemotherapy for first-line treatment of locally advanced intrahepatic cholangiocarcinoma. *JAMA Oncol.* **6**, 51 (2020).
17. Cercek, A., Boerner, T., Tan, B. R., Chou, J. F., Gönen, M., Boucher, T. M. et al. Assessment of hepatic arterial infusion of floxuridine in combination with systemic gemcitabine and oxaliplatin in patients with unresectable intrahepatic cholangiocarcinoma. *JAMA Oncol.* **6**, 60 (2020).
18. Leone, F., Filippi, R., Palloni, A., Fornaro, L., Casadei Gardini, A., Aprile, G. et al. Prognostic factors in unresectable biliary tract cancer: a GICO (Gruppo Italiano COlangiocarcinoma) retrospective analysis. *Ann. Oncol.* **28**, vi48 (2017).
19. Lamarca, A., Hubner, R. A., David Ryder, W. & Valle, J. W. Second-line chemotherapy in advanced biliary cancer: a systematic review. *Ann. Oncol.* **25**, 2328–2338 (2014).
20. Valle, J. W., Borbath, I., Khan, S. A., Huguot, F., Gruenberger, T. & Arnold, D. Biliary cancer: ESMO clinical practice guidelines for diagnosis, treatment and follow-up. *Ann. Oncol.* **27**, v28–v37 (2016).
21. Bridgewater, J., Palmer, D., Cunningham, D., Iveson, T., Gillmore, R., Waters, J. et al. Outcome of second-line chemotherapy for biliary tract cancer. *Eur. J. Cancer* **49**, 1511 (2013).
22. Brieau, B., Dahan, L., De Rycke, Y., Boussaha, T., Vasseur, P., Tougeron, D. et al. Second-line chemotherapy for advanced biliary tract cancer after failure of the gemcitabine-platinum combination: a large multicenter study by the Association des Gastro-Entérologues Oncologues. *Cancer* **121**, 3290–3297 (2015).
23. Schweitzer, N., Kirstein, M. M., Kratzel, A., Mederacke, Y., Fischer, M., Manns, M. P. et al. Second-line chemotherapy in biliary tract cancer: Outcome and prognostic factors. *Liver Int.* **39**, 914–923 (2019).
24. Lamarca, A., Palmer, D. H., Wasan, H. S., Ross, P. J., Ma, Y. T., Arora, A. et al. ABC-06 | A randomised phase III, multi-centre, open-label study of active symptom control (ASC) alone or ASC with oxaliplatin/5-FU chemotherapy (ASC+mFOLFOX) for patients (pts) with locally advanced / metastatic biliary tract cancers (ABC) previously-tr. *J. Clin. Oncol.* **37**, 4003–4003 (2019).
25. Belkouz, A., Vos-Geelen, J., de Eskens, F., Mathot, R. A. A., van Gulik, T., van Oijen, M. G. H. et al. Efficacy and safety of FOLFIRINOX in advanced biliary tract cancer after failure of gemcitabine plus cisplatin: a phase II trial. *J. Clin. Oncol.* **37**, 4086–4086 (2019).
26. Pape, U.-F., Kasper, S., Meiler, J., Sinn, M., Vogel, A., Mueller, L. et al. Post-hoc analyses of a subgroup of patients with advanced biliary tract cancer (BTC) who crossed over to treatment with etoposide tonirbate (EDO-57.1) in a randomized phase II study. *Ann. Oncol.* **30**, v278 (2019).
27. Wu, Y.-M., Su, F., Kalyana-Sundaram, S., Khazanov, N., Ateeq, B., Cao, X. et al. Identification of targetable FGFR gene fusions in diverse cancers. *Cancer Discov.* **3**, 636–647 (2013).
28. Abou-Alfa, G. K., Macarulla Mercade, T., Javle, M., Kelley, R. K., Lubner, S., Adeva, J. et al. ClarIDHy: a global, phase III, randomized, double-blind study of ivosidenib (IVO) vs placebo in patients with advanced cholangiocarcinoma (CC) with an isocitrate dehydrogenase 1 (IDH1) mutation. *Ann. Oncol.* **30**, v872–v873 (2019).
29. Javle, M., Lowery, M., Shroff, R. T., Weiss, K. H., Springfield, C., Borad, M. J. et al. Phase II study of BGJ398 in patients with FGFR-altered advanced cholangiocarcinoma. *J. Clin. Oncol.* **36**, 276–282 (2018).
30. Mazzaferro, V., El-Rayes, B. F., Droz dit Busset, M., Cotsoglou, C., Harris, W. P., Damjanov, N. et al. Derazantinib (ARQ 087) in advanced or inoperable FGFR2 gene fusion-positive intrahepatic cholangiocarcinoma. *Br. J. Cancer* **120**, 165–171 (2019).
31. Vogel, A., Sahai, V., Hollebecque, A., Vaccaro, G., Melisi, D., Al-Rajabi, R. et al. FIGHT-202: a phase II study of pemigatinib in patients (pts) with previously treated locally advanced or metastatic cholangiocarcinoma (CCA). *Ann. Oncol.* **30**, v876 (2019).
32. Goyal, L., Bahleda, R., Furuse, J., Valle, J. W., Moehler, M. H., Oh, D.-Y. et al. FOENIX-101: A phase II trial of TAS-120 in patients with intrahepatic cholangiocarcinoma harboring FGFR2 gene rearrangements. *J. Clin. Oncol.* **37**, TPS468–TPS468 (2019).
33. Chen, Y.-Y., Park, J. O., Su, W.-C., Oh, D.-Y., Kim, K.-P., Feng, Y.-H. et al. Preliminary results of a phase II study to evaluate the clinical efficacy and safety of erdafitinib in Asian patients with biomarker-selected advanced cholangiocarcinoma (CCA). *Ann. Oncol.* **29**, viii209 (2018).
34. Hyman, D. M., Goyal, L., Grivas, P., Meric-Bernstam, F., Taberner, J., Hu, Y. et al. FUZE clinical trial: a phase 2 study of Debio 1347 in FGFR fusion-positive advanced solid tumors irrespectively of tumor histology. *J. Clin. Oncol.* **37**, TPS3157–TPS3157 (2019).
35. Meric-Bernstam, F., Arkenau, H., Tran, B., Bahleda, R., Kelley, R., Hierro, C. et al. Efficacy of TAS-120, an irreversible fibroblast growth factor receptor (FGFR) inhibitor, in cholangiocarcinoma patients with FGFR pathway alterations who were previously treated with chemotherapy and other FGFR inhibitors. *Ann. Oncol.* **29**, v100 (2018).
36. Javle, M. M., Borbath, I., Clarke, S. J., Hitre, E., Louvet, C., Mercade, T. M. et al. Infigratinib versus gemcitabine plus cisplatin multicenter, open-label, randomized, phase 3 study in patients with advanced cholangiocarcinoma with FGFR2 gene fusions/translocations: The PROOF trial. *J. Clin. Oncol.* **37**, TPS4155–TPS4155 (2019).
37. Nakamura, H., Arai, Y., Totoki, Y., Shirota, T., Elzawahry, A., Kato, M. et al. Genomic spectra of biliary tract cancer. *Nat. Genet.* **47**, 1003–1010 (2015).
38. Goyal, L., Shi, L., Liu, L. Y., Fece de la Cruz, F., Lennerz, J. K., Raghavan, S. et al. TAS-120 overcomes resistance to ATP-competitive FGFR inhibitors in patients with FGFR2 fusion-positive intrahepatic cholangiocarcinoma. *Cancer Discov.* **9**, 1064–1079 (2019).
39. Lamberti, D., Cristinziano, G., Porru, M., Leonetti, C., Egan, J. B., Shi, C. et al. HSP90 inhibition drives degradation of FGFR2 fusion proteins: implications for treatment of cholangiocarcinoma. *Hepatology* **69**, hep.30127 (2018).
40. Lampis, A., Carotenuto, P., Vlachogiannis, G., Cascione, L., Hedayat, S., Burke, R. et al. MIR21 drives resistance to heat shock protein 90 inhibition in cholangiocarcinoma. *Gastroenterology* **154**, 1066–1079.e5 (2018).
41. Cocco, E., Schram, A. M., Kulick, A., Misale, S., Won, H. H., Yaeger, R. et al. Resistance to TRK inhibition mediated by convergent MAPK pathway activation. *Nat. Med.* **25**, 1422–1427 (2019).
42. Gu, Y., Sai, Y., Wang, J., Yu, M., Wang, G., Zhang, L. et al. Preclinical pharmacokinetics, disposition, and translational pharmacokinetic/pharmacodynamic modeling of savolitinib, a novel selective cMet inhibitor. *Eur. J. Pharm. Sci.* **136**, 104938 (2019).
43. Zhang, Z., Oyesanya, R. A., Campbell, D. J. W., Almenara, J. A., DeWitt, J. L. & Sirica, A. E. Preclinical assessment of simultaneous targeting of epidermal growth factor receptor (ErbB1) and ErbB2 as a strategy for cholangiocarcinoma therapy. *Hepatology* **52**, 975–986 (2010).
44. Sia, D., Hoshida, Y., Villanueva, A., Roayaie, S., Ferrer, J., Tabak, B. et al. Integrative molecular analysis of intrahepatic cholangiocarcinoma reveals 2 classes that have different outcomes. *Gastroenterology* **144**, 829–840 (2013).
45. Gao, Q., Zhao, Y., Wang, X., Guo, W., Gao, S., Wei, L. et al. Activating mutations in PTPN3 promote cholangiocarcinoma cell proliferation and migration and are associated with tumor recurrence in patients. *Gastroenterology* **146**, 1397–1407 (2014).
46. Yarlaga, B., Kamatham, V., Ritter, A., Shahjehan, F. & Kasi, P. M. Trastuzumab and pertuzumab in circulating tumor DNA ERBB2-amplified HER2-positive refractory cholangiocarcinoma. *npj Precis. Oncol.* **3**, 19 (2019).
47. Wainberg, Z. A., Lassen, U. N., Elez, E., Italiano, A., Curigliano, G., De Braud, F. G. et al. Efficacy and safety of dabrafenib (D) and trametinib (T) in patients (pts) with BRAF V600E-mutated biliary tract cancer (BTC): a cohort of the ROAR basket trial. *J. Clin. Oncol.* **37**, 187–187 (2019).
48. Golan, T., Sella, T., O'Reilly, E. M., Katz, M. H. G., Epelbaum, R., Kelsen, D. P. et al. Overall survival and clinical characteristics of BRCA mutation carriers with stage I/II pancreatic cancer. *Br. J. Cancer* **116**, 697–702 (2017).

49. Javle, M. M., Hainsworth, J. D., Swanton, C., Burris, H. A., Kurzrock, R., Sweeney, C. et al. Pertuzumab + trastuzumab for HER2-positive metastatic biliary cancer: Preliminary data from MyPathway. *J. Clin. Oncol.* **35**, 402–402 (2017).
50. Braconi, C., Roessler, S., Kruk, B., Lammert, F., Krawczyk, M. & Andersen, J. B. Molecular perturbations in cholangiocarcinoma: Is it time for precision medicine? *Liver Int.* **39**, 32–42 (2019).
51. Ikeda, M., Ohno, I., Ueno, H., Mitsunaga, S., Hashimoto, Y., Okusaka, T. et al. Phase I study of resminostat, an HDAC inhibitor, combined with S-1 in patients with pre-treated biliary tract or pancreatic cancer. *Invest. New Drugs* **37**, 109–117 (2019).
52. Bang, Y.-J., Ueno, M., Malka, D., Chung, H. C., Nagrial, A., Kelley, R. K. et al. Pembrolizumab (pembro) for advanced biliary adenocarcinoma: results from the KEYNOTE-028 (KN028) and KEYNOTE-158 (KN158) basket studies. *J. Clin. Oncol.* **37**, 4079–4079 (2019).
53. Marabelle, A., Le, D. T., Ascierto, P. A., Di Giacomo, A. M., De Jesus-Acosta, A., Delord, J.-P. et al. Efficacy of pembrolizumab in patients with noncolorectal high microsatellite instability/mismatch repair-deficient cancer: results from the phase II KEYNOTE-158 study. *J. Clin. Oncol.* **38**, 1–10 (2020).
54. Ueno, M., Ikeda, M., Morizane, C., Kobayashi, S., Ohno, I., Kondo, S. et al. Nivolumab alone or in combination with cisplatin plus gemcitabine in Japanese patients with unresectable or recurrent biliary tract cancer: a non-randomised, multicentre, open-label, phase 1 study. *Lancet. Gastroenterol. Hepatol.* **4**, 611–621 (2019).
55. Arkenau, H., Martin-Liberal, J., Calvo, E., Penel, N., Krebs, M. G., Herbst, R. S. et al. Ramucirumab plus pembrolizumab in patients with previously treated advanced or metastatic biliary tract cancer: nonrandomized, open-label, phase I trial (JVDF). *Oncologist* **23**, 1407 (2018).
56. Marin, J. J. G., Lozano, E., Briz, O., Al-Abdulla, R., Serrano, M. A. & Macias, R. I. R. Molecular bases of chemoresistance in cholangiocarcinoma. *Curr. Drug Targets* **18**, 889–900 (2017).
57. Marin, J. J. G., Lozano, E., Herraiz, E., Asensio, M., Di Giacomo, S., Romero, M. R. et al. Chemoresistance and chemosensitization in cholangiocarcinoma. *Biochim. Biophys. Acta Mol. Basis Dis.* **1864**, 1444–1453 (2018).
58. Brandi, G., Deserti, M., Vasuri, F., Farioli, A., Degiovanni, A., Palloni, A. et al. Membrane localization of human equilibrative nucleoside transporter 1 in tumor cells may predict response to adjuvant gemcitabine in resected cholangiocarcinoma patients. *Oncologist* **21**, 600–607 (2016).
59. Kim, J., Kim, H., Lee, J., Kim, J. W., Paik, W. H., Lee, S. H. et al. Human equilibrative nucleoside transporter 1 (hENT1) expression as a predictive biomarker for gemcitabine chemotherapy in biliary tract cancer. *PLoS ONE* **13**, e0209104 (2018).
60. Martinez-Becerra, P., Vaquero, J., Romero, M. R., Lozano, E., Anadon, C., Macias, R. I. R. et al. No correlation between the expression of FXR and genes involved in multidrug resistance phenotype of primary liver tumors. *Mol. Pharm.* **9**, 1693–1704 (2012).
61. Lozano, E., Macias, R. I. R., Monte, M. J., Asensio, M., Carmen, S., Sanchez-Vicente, L. et al. Causes of hOCT1-dependent cholangiocarcinoma resistance to sorafenib and sensitization by tumor-selective gene therapy. *Hepatology* **70**, 1246–1261 (2019).
62. Srimuntha, U., Sawanyawisuth, K., Kraiklang, R., Pairojkul, C., Puapairoj, A., Titipungul, T. et al. High expression of ABC11 indicates poor prognosis in intrahepatic cholangiocarcinoma. *Asian Pac. J. Cancer Prev.* **13**, 125–130 (2012).
63. Chen, M.-H., Weng, J.-J., Cheng, C.-T., Wu, R.-C., Huang, S.-C., Wu, C.-E. et al. ALDH1A3, the major aldehyde dehydrogenase isoform in human cholangiocarcinoma cells, affects prognosis and gemcitabine resistance in cholangiocarcinoma patients. *Clin. Cancer Res.* **22**, 4225–4235 (2016).
64. Nakajima, T., Takayama, T., Miyaniishi, K., Nobuoka, A., Hayashi, T., Abe, T. et al. Reversal of multiple drug resistance in cholangiocarcinoma by the glutathione S-transferase- π -specific inhibitor O1-hexadecyl- γ -glutamyl- S-benzylcysteinyl-D-phenylglycine ethylester. *J. Pharmacol. Exp. Ther.* **306**, 861–869 (2003).
65. Suksawat, M., Klanrit, P., Phetcharaburanin, J., Namwat, N., Khuntikeo, N., Titapun, A. et al. In vitro and molecular chemosensitivity in human cholangiocarcinoma tissues. *PLoS ONE* **14**, e0222140 (2019).
66. Hahnvajjanawong, C. Orotate phosphoribosyl transferase mRNA expression and the response of cholangiocarcinoma to 5-fluorouracil. *World J. Gastroenterol.* **18**, 3955 (2012).
67. Habara, K., Ajiki, T., Kamigaki, T., Nakamura, T. & Kuroda, Y. High expression of thymidylate synthase leads to resistance to 5-fluorouracil in biliary tract carcinoma in vitro. *Japanese J. Cancer Res.* **92**, 1127–1132 (2001).
68. Jimeno, A., Rubio-Viqueira, B., Amador, M. L., Oppenheimer, D., Bouraoud, N., Kulesza, P. et al. Epidermal growth factor receptor dynamics influences response to epidermal growth factor receptor targeted agents. *Cancer Res.* **65**, 3003–3010 (2005).
69. Sato, J., Kimura, T., Saito, T., Anazawa, T., Kenjo, A., Sato, Y. et al. Gene expression analysis for predicting gemcitabine resistance in human cholangiocarcinoma. *J. Hepatobiliary. Pancreat. Sci.* **18**, 700–711 (2011).
70. Ge, X., Wang, Y., Li, Q., Yu, H., Ji, G. & Miao, L. NK4 regulates 5-fluorouracil sensitivity in cholangiocarcinoma cells by modulating the intrinsic apoptosis pathway. *Oncol. Rep.* **30**, 448–454 (2013).
71. Wattanawongdon, W., Hahnvajjanawong, C., Namwat, N., Kanchanawat, S., Boonmars, T., Jearanaikoon, P. et al. Establishment and characterization of gemcitabine-resistant human cholangiocarcinoma cell lines with multidrug resistance and enhanced invasiveness. *Int. J. Oncol.* **47**, 398–410 (2015).
72. Choodetwattana, P., Prongvitaya, S., Jearanaikoon, P. & Limpiboon, T. The upregulation of OCT4 in acidic extracellular pH is associated with gemcitabine resistance in cholangiocarcinoma cell lines. *Asian Pacific J. Cancer Prev.* **20**, 2745–2748 (2019).
73. Yamada, D., Kobayashi, S., Wada, H., Kawamoto, K., Marubashi, S., Eguchi, H. et al. Role of crosstalk between interleukin-6 and transforming growth factor-beta 1 in epithelial-mesenchymal transition and chemoresistance in biliary tract cancer. *Eur. J. Cancer* **49**, 1725–1740 (2013).
74. Quintavalle, C., Burmeister, K., Piscuoglio, S., Quagliata, L., Karamitopoulou, E., Sepe, R. et al. High mobility group A1 enhances tumorigenicity of human cholangiocarcinoma and confers resistance to therapy. *Mol. Carcinog.* **56**, 2146–2157 (2017).
75. Salati, M. & Braconi, C. Noncoding RNA in cholangiocarcinoma. *Semin. Liver Dis.* **39**, 013–025 (2019).
76. Carotenuto, P., Hedayat, S., Fassan, M., Cardinale, V., Lampis, A., Guzzardo, V., et al. Modulation of biliary cancer chemo-resistance through microRNA-mediated rewiring of the expansion of CD133+ cells. *Hepatology* <https://doi.org/10.1002/hep.31094> (2019).
77. Meng, F., Henson, R., Lang, M., Wehbe, H., Maheshwari, S., Mendell, J. T. et al. Involvement of Human Micro-RNA in Growth and Response to Chemotherapy in Human Cholangiocarcinoma Cell Lines. *Gastroenterology* **130**, 2113–2129 (2006).
78. Peng, F., Jiang, J., Yu, Y., Tian, R., Guo, X., Li, X. et al. Direct targeting of SUZ12/ROCK2 by miR-200b/c inhibits cholangiocarcinoma tumourigenesis and metastasis. *Br. J. Cancer* **109**, 3092–3104 (2013).
79. Okamoto, K., Miyoshi, K. & Murawaki, Y. miR-29b, miR-205 and miR-221 enhance chemosensitivity to gemcitabine in HuH28 human cholangiocarcinoma cells. *PLoS ONE* **8**, e77623 (2013).
80. Chen, L., Yan, H.-X., Yang, W., Hu, L., Yu, L.-X., Liu, Q. et al. The role of microRNA expression pattern in human intrahepatic cholangiocarcinoma. *J. Hepatol.* **50**, 358–369 (2009).
81. Jiao, D., Yan, Y., Shui, S., Wu, G., Ren, J., Wang, Y. et al. miR-106b regulates the 5-fluorouracil resistance by targeting Zbtb7a in cholangiocarcinoma. *Oncotarget* **8**, 52913–52922 (2017).
82. Asukai, K., Kawamoto, K., Eguchi, H., Konno, M., Asai, A., Iwagami, Y. et al. Micro-RNA-130a-3p regulates gemcitabine resistance via PPAR γ in cholangiocarcinoma. *Ann. Surg. Oncol.* **24**, 2344–2352 (2017).
83. Seehawer, M., Heinzmann, F., D'Artista, L., Harbig, J., Roux, P.-F., Hoenicke, L. et al. Necroptosis microenvironment directs lineage commitment in liver cancer. *Nature* **562**, 69–75 (2018).
84. Cadamuro, M., Brivio, S., Spirli, C., Joplin, R., Strazzabosco, M. & Fabris, L. Autocrine and paracrine mechanisms promoting chemoresistance in cholangiocarcinoma. *Int. J. Mol. Sci.* **18**, 149 (2017).
85. Harnois, D. M., Que, F. G., Celli, A., LaRusso, N. F. & Gores, G. J. Bcl-2 is over-expressed and alters the threshold for apoptosis in a cholangiocarcinoma cell line. *Hepatology* **26**, 884–890 (1997).
86. Minagawa, N., Kruglov, E. A., Dranoff, J. A., Robert, M. E., Gores, G. J. & Nathanson, M. H. The anti-apoptotic protein Mcl-1 inhibits mitochondrial Ca²⁺ signals. *J. Biol. Chem.* **280**, 33637–33644 (2005).
87. Yoon, H., Min, J.-K., Lee, J. W., Kim, D.-G. & Hong, H. J. Acquisition of chemoresistance in intrahepatic cholangiocarcinoma cells by activation of AKT and extracellular signal-regulated kinase (ERK)1/2. *Biochem. Biophys. Res. Commun.* **405**, 333–337 (2011).
88. Wehrkamp, C. J., Gutwein, A. R., Natarajan, S. K., Phillippi, M. A. & Mott, J. L. XIAP Antagonist embelin inhibited proliferation of cholangiocarcinoma cells. *PLoS ONE* **9**, e90238 (2014).
89. Mertens, J. C., Fingas, C. D., Christensen, J. D., Smoot, R. L., Bronk, S. F., Werneburg, N. W. et al. Therapeutic effects of deleting cancer-associated fibroblasts in cholangiocarcinoma. *Cancer Res.* **73**, 897–907 (2013).
90. Sirica, A. E. & Gores, G. J. Desmoplastic stroma and cholangiocarcinoma: clinical implications and therapeutic targeting. *Hepatology* **59**, 2397–2402 (2014).
91. Carpino, G., Overi, D., Melandro, F., Grimaldi, A., Cardinale, V., Di Matteo, S. et al. Matrisome analysis of intrahepatic cholangiocarcinoma unveils a peculiar cancer-associated extracellular matrix structure. *Clin. Proteomics* **16**, 37 (2019).

92. Whiteside, T. L. What are regulatory T cells (Treg) regulating in cancer and why? *Semin. Cancer Biol.* **22**, 327–334 (2012).
93. Ghidini, M., Cascione, L., Carotenuto, P., Lampis, A., Trevisani, F., Previdi, M. C. et al. Characterisation of the immune-related transcriptome in resected biliary tract cancers. *Eur. J. Cancer* **86**, 158–165 (2017).
94. Verlingue, L., Malka, D., Allorant, A., Massard, C., Féré, C., Lacroix, L. et al. Precision medicine for patients with advanced biliary tract cancers: an effective strategy within the prospective MOSCATO-01 trial. *Eur. J. Cancer* **87**, 122–130 (2017).
95. Sicklick, J. K., Kato, S., Okamura, R., Schwaederle, M., Hahn, M. E., Williams, C. B. et al. Molecular profiling of cancer patients enables personalized combination therapy: the I-PREDICT study. *Nat. Med.* **25**, 744–750 (2019).
96. Mody, K., Kasi, P. M., Yang, J., Surapaneni, P. K., Bekaii-Saab, T., Ahn, D. H., et al. Circulating tumor DNA profiling of advanced biliary tract cancers. *JCO Precis. Oncol.* <https://doi.org/10.1200/PO.18.00324> (2019).
97. Shen, N., Zhang, D., Yin, L., Qiu, Y., Liu, J., Yu, W. et al. Bile cell-free DNA as a novel and powerful liquid biopsy for detecting somatic variants in biliary tract cancer. *Oncol. Rep.* **42**, 549–560 (2019).
98. Yang, J. D., Campion, M. B., Liu, M. C., Chaiteerakij, R., Giama, N. H., Ahmed Mohammed, H. et al. Circulating tumor cells are associated with poor overall survival in patients with cholangiocarcinoma. *Hepatology* **63**, 148–158 (2016).
99. Vlachogiannis, G., Hedayat, S., Vatsiou, A., Jamin, Y., Fernández-Mateos, J., Khan, K. et al. Patient-derived organoids model treatment response of metastatic gastrointestinal cancers. *Science* **359**, 920–926 (2018).
100. Amato, F., Rae, C., Prete, M. G. & Braconi, C. Cholangiocarcinoma disease modelling through patients derived organoids. *Cells* **9**, 832 (2020).