



### Cancer immune contexture and immunotherapy

Etienne Becht, Nicolas A Giraldo, Marie-Caroline Dieu-Nosjean, Catherine Sautès-Fridman, Wolf Herman Fridman

#### ▶ To cite this version:

Etienne Becht, Nicolas A Giraldo, Marie-Caroline Dieu-Nosjean, Catherine Sautès-Fridman, Wolf Herman Fridman. Cancer immune contexture and immunotherapy. Current Opinion in Immunology, Elsevier, 2016, 39, pp.7-13. <10.1016/j.coi.2015.11.009>. <hal-01281664>

# HAL Id: hal-01281664 http://hal.upmc.fr/hal-01281664

Submitted on 2 Mar 2016

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

## 1 **Cancer immune contexture and immunotherapy**

- 2
- Etienne Becht<sup>1,2,3,\*</sup>, Nicolas Giraldo<sup>1,2,3,\*</sup>, Marie-Caroline Dieu-Nosjean<sup>1,2,3</sup>, Catherine
   Sautès-Fridman<sup>1,2,3</sup>, Wolf Herman Fridman<sup>1,2,3</sup>
- <sup>5</sup> <sup>1</sup>INSERM UMR\_S 1138, Cancer, Immune Control and Escape, Cordeliers Research Centre, Paris, France
- 6 <sup>2</sup>Université Paris Descartes, Paris, France
- <sup>3</sup>Université Pierre et Marie Curie, Paris, France
- 8 \*These authors contributed equally to this work
- 9

#### 10 Abstract

The immune contexture characterizes the clinical impact of the density, the location, the organization and the functional orientation of tumor-infiltrating immune cells in cancers. It is, in great part, shaped by the malignant cells, as in a given cancer type, tumors presenting different oncogenic processes have different immune contextures. Moreover, the immune contexture in metastatic sites reflects that of the corresponding primary tumors. Finally, the components forming the immune contexture represent

17 targets and markers of efficient anti-cancer immunotherapies.

#### 18

#### 19 Introduction

Cancers grow and spread in tissues where the malignant cells interact with blood and lymphatic vessels, stromal cells and hematopoietic cells involved in immune and inflammatory reactions. This landscape constitutes the immune contexture of a tumor which is of paramount importance for patients' clinical outcome, is predictive of responses to therapies and helps identifying novel targets for immunotherapies [1].

25

#### 26 Clinical impact of the immune contexture in different primary tumors

27

The immune contexture is a concept that emerged from studies mostly performed in 28 human colorectal cancer (CRC). A comprehensive analysis of a large collection of 29 primary CRC tumors revealed that a high density of intratumoral memory T cells 30 correlates with patients' longer disease-free (DFS) and overall (OS) survivals [2], 31 confirming a previous report in ovarian cancer [3]. A closer histopathological analysis 32 of CRC tumors highlighted the fact that the T cells were not stochastically distributed 33 within the tumor microenvironment but were present in the center (CT) and the 34 invasive margin (IM) of the tumor nests [4]. The densities of memory CD8<sup>+</sup> T cells in 35 both the CT and the IM were associated with favorable prognosis, as well as the 36 expression of genes encoding Th1 cytokines (IFN-y, IL2) and cytotoxic mediators 37 (granzymes, granulysin)[4]. A comprehensive approach of all genes having any type of 38 correlation with genes whose expression correlated with both clinical outcome and 39 tumor infiltration by T cells predicted that certain chemokines (CXCL13, CXCL9, 40 CXCL10) were involved in shaping an efficient immune microenvironment [5] and 41 foster T cells activation [6]. 42

In parallel to these studies in CRC, our group has investigated the immune
microenvironment of Non-Small-Cell-Lung Cancer (NSCLC). In addition to confirming
the beneficial effect of high densities of T cells with a Th1 orientation and of cytotoxic
CD8<sup>+</sup> T cells, these studies revealed the essential role of tumor-associated Tertiary
Lymphoid Structures (TLS), lymphoid aggregates that structurally resemble secondary
lymphoid organs [7][8]. These formations are present in the invasive margin and in the

stroma of lung tumors and absent in distant non-tumoral lung tissue. They are 49 organized in a T-cell and a B-cell zone surrounded by High Endothelial Venules (HEV). 50 T cells are in contact with mature DC, and B cells with follicular dendritic cells (FDC) 51 and tingible-body macrophages. B cells express the activation-induced deaminase 52 (AID) enzyme necessary for somatic hypermutation and immunoglobulin isotype 53 switching, a marker of active germinal center (GC) [9]. These data led us to hypothesize 54 that TLS are sites where immune reactions toward tumor-associated antigens are 55 generated [10]. Since they are exclusively present within TLS, naïve T and B cells may 56 have emigrated from peripheral blood via HEV. Thus, TLS may represent an immune-57 naïve lymphocytes may be 58 privileged site where protected from the immunosuppressive and inflammatory milieu of the tumor during their differentiation 59 phase. Indeed, high density of these tumor-associated TLS positively correlates with 60 NSCLC patient's prognosis [8][9][7]. This is in accordance with the fact that HEV are the 61 only type of blood vessels which positively correlates with favorable clinical outcome of 62 cancer [11]. TLS-infiltrating naive B cells differentiate into memory B cells and plasma 63 cells which produce anti-tumor antibodies [9]. T cells in TLS are educated by antigen-64 presenting DC, become memory lymphocytes and migrate into other tumor areas, 65 resulting in local control, and to peripheral lymph nodes where they restrain metastatic 66 spread[10]. TLS likely influence the functional orientation of tumor-infiltrating T cells, 67 as intratumor T cells have a Th1 and cytotoxic CD8 orientation in tumors having high 68 TLS densities [8]. 69

These initial observations in CRC and NSCLC have been extended to most cancer types 70 [1][10]. Indeed the evaluation of the immune contexture involves quantitative 71 immunohistochemistry and gene expression techniques (for evaluation of Th 72 orientation for instance) and both type of assays needs expertise to be performed 73 successfully. Regarding T cell infiltration, an "Immunoscore" has been defined 74 quantifying by immunochemistry two out of the three markers, CD3, CD8 and CD45R0, 75 in the invasive margin and the center of tumors[12]. A general trend emerges that high 76 densities of memory T cells with a Th1 orientation and CD8<sup>+</sup> phenotype correlate with 77 longer DFS and OS also in ovarian, bladder, breast, prostatic, head and neck, and 78 cervical cancers, hepatocellular carcinoma and melanoma (reviewed in [1] and [13]). 79

Regarding TLS or Ectopic Lymphoid Structures, their quantification has been 80 performed using the DC-Lamp marker, lymphoid aggregates in HES sections, CD20 81 positive follicles or PNAd+ marker for HEV. Whatever the methodology used, the data 82 show that high densities of TLS also correlate with favorable clinical outcome in CRC, 83 breast, gastric, pancreatic, colorectal and renal cancers, oral squamous carcinoma, 84 Merkel cell carcinoma, Warthin tumor as well as melanoma (reviewed in [10]). These 85 studies establish the immune contexture as a well-studied, robust and clinically 86 relevant characteristic of human cancers biology. 87

However T and B lymphocytes, and TLS are far from being the only components of the
immune and inflammatory microenvironment of tumors. The densities and
organization of these other cell types also influence patient's clinical outcome although
with less significant power. Indeed the identification of some cell subpopulations such

as Treg[14] or MDSC can be hampered by the lack of consensus markers. Howewer, a 92 trend emerges that high densities of myeloid cells, such as macrophages, MDSCs and 93 particularly M2 macrophages mast cells. and are associated with poor 94 prognosis[13][15][16][17][18]. NK cells infiltrating tumors can be anergic[19][20]. 95 Extensive NK cells infiltration has nonetheless usually been associated with favorable 96 patient outcome[6][21][22][23]. 97

Regarding the other T cell subsets, their impact may depend on the cancer type. It is the case for Th2,  $T_{reg}$  or Th17 (reviewed in [1]). High densities of DC,  $T_{FH}$  and B cells in TLS correlate with favorable prognosis[6]. In presence of low or no TLS, high densities of stromal B cells, CD8<sup>+</sup> T cells and DC are associated with poor prognosis, and may support pro-tumoral inflammation and T cell anergy, underlining again the importance of the location of the immune cells in the tumor landscape [8].

Despite the general agreement on the favorable prognostic value of the density of Th1 104 and CD8<sup>+</sup> T cells, discordant results have been reported in clear cell RCC (ccRCC) [24], 105 Hodgkin lymphoma [25] or uveal melanoma [26]. Intrigued by these results, we 106 recently analyzed a cohort of primary ccRCC and confirmed that high densities of CD8+ 107 T cells in the IM correlated with shorter DFS and OS [27]. Even more intriguingly, high 108 expression of the IFN-γ, TBX21, and granzyme-B genes were a highly significant marker 109 of poor prognosis [27]. These results prompted us to study the immune contexture of 110 ccRCC. We found that there were few TLS in most ccRCC tumors [28][10]. Most CD8<sup>+</sup> T 111 cells presented an exhausted phenotype with concomitant expression of LAG3 and PD-112 1, and tumor cells expressed PD-1 ligands [27]. Patients with tumors containing high 113 densities of PD-1+CD8+ T cells, and PD-L1 or PD-L2 expression by tumor cells had the 114 worst prognosis. Strikingly, in most ccRCC tumors, immature DC-Lamp<sup>+</sup> DC were found 115 outside TLS, close to blood vessels (and not to HEV). It is therefore likely that these DC 116 instructed incoming T cells in an inflammatory and immunosuppressed milieu 117 resulting in an abortive immune response. The IFN- $\gamma$  production by T cells has been 118 reported to induce PD-L1 and/or PD-L2 on tumor cells [29], resulting in an exhaustion 119 of the immune reaction. Indeed, in the few tumors with high densities of mature DC 120 inside TLS, high CD8<sup>+</sup> T cells density correlated with favorable prognosis, supporting 121 the role of TLS in educating HEV-penetrating naive T cells to recognize tumor-122 associated antigens and to control cancer aggressiveness [27]. These data suggest that 123 certain tumor types may be characterized by a disrupted immune contexture, most 124 likely governed by the malignant cells. 125

126

## 127 Clinical impact of the immune contexture in metastatic sites

It is intuitively thought that tumor progression is accompanied by tumor escape from the immune system [30][31]. If this escape linearly followed cancer progression, the immune contexture of metastatic sites should not impact clinical outcome. However, high densities of CD8<sup>+</sup> T cells in hepatic and lung metastases of CRC correlate with longer OS [28], as in primary CRC [4]. Densities of infiltrating immune cells were shown to be correlated between the primary tumors and matched metastases[28]. These

findings both suggest that the malignant cells are prominent in shaping their immune 134 microenvironments. This hypothesis is strengthened by the study of lung metastases of 135 ccRCC. In contrast to lung metastases of CRC in which high TLS and CD8<sup>+</sup> T cell 136 densities correlate with favorable prognosis, lung metastases of ccRCC have few TLS, 137 and high CD8<sup>+</sup> T cell densities correlate with shorter OS, as in primary ccRCC[28]. In 138 addition, transcriptomic analyses revealed a higher expression of genes involved in 139 inflammation, immunosuppression and angiogenesis in lung metastases from ccRCC 140 than in lung metastases from CRC, confirming differences in the functional orientations 141 of these immune contextures [28]. 142

### 143 **Clinical impact of the immune contexture within a given cancer**

Whole genome transcriptomic analyses provide a novel way to classify subgroups in a 144 given cancer type. These unsupervised approaches complement genomic classifications 145 by identifying malignant cell subgroups with distinct functional traits. We undertook an 146 analysis of the expression of immune genes and the concomitant immune cell 147 infiltration in cohorts of various human cancers. To precisely analyze the immune 148 contexture of large collections of human cancers, we established transcriptomic 149 signatures based on the specific expression of genes in a given hematopoietic subset. 150 We identified robust immune metagenes for lymphocytes (T and B cell subsets, NK, Tγδ, 151 cvtotoxic cells), myeloid cells (macrophages, monocytes, granulocytes, DC) as well as 152 endothelial cells and fibroblasts (EB submitted). These signatures were validated in 153 vitro in mixtures of hematopoietic and tumor cells and *ex-vivo* on CRC tumor sections. 154 They were completed by analyses of genes expressed modulating immune functions 155 (cytokines, chemokines, MHC class I) that sign the functional orientation of the immune 156 microenvironments. We review herein our data in two cancers, prototypic for the 157 clinical impacts of their immune contextures, CRC and ccRCC. 158

Different molecular classifications have been proposed for CRC and merged in a four-159 subgroups consensus classification [32]. It identifies a microsatellite-instability (MSI)-160 enriched group with good prognosis, a mesenchymal subgroup with the worst 161 prognosis, and KRAS mutated and canonical subgroups with intermediate prognosis. 162 We applied the immune signatures to these classifications. We analyzed three cohorts 163 of CRC including over 2000 patients and found that molecular subgoups had distinct 164 immune signatures. The hypermutated MSI-enriched subgroup had the highest 165 infiltration of T cells along with cytotoxic lymphocytes, followed by the mesenchymal 166 subgroup which also presented with high lymphocytic infiltration in the context of high 167 myeloid cell infiltration as well as extensive presence of endothelial cells and 168 fibroblasts [33]. The two other groups of tumors had very poor expression of the 169 immune gene signatures. These analyses show that the malignant cells can influence 170 the density of immune and inflammatory cells in tumors. The hypermutated MSI-171 enriched group had the highest expression of genes involved in T cell chemotaxis 172 (CXCL9, CXCL10, CXCL16), T cell activation (IFN-y, IL15), T cell inhibition (PD-L1, PD-173 L2, CTLA-4, LAG3), and TLS formation (CXCL13). The mesenchymal subgroup had the 174 highest expression of genes involved in myeloid cell chemotaxis (CCL2), angiogenesis 175 (VEGF, PDGF), immunosuppression (TGF $\beta$ ), and inflammation (CCL2, complement-176

related genes). The two immune low groups had also the lowest MHC Class I expression 177 [33]. Taken together, these data support the concept that the MSI-tumors, which have 178 defects in the DNA-repair enzymes, produce mutated neo-antigens that activate T cells, 179 concurring to shape a favorable immune contexture. In contrast, the mesenchymal 180 tumors, through a high production of angiogenic and inflammatory molecules shape a 181 disrupted immune contexture where inflammation fuels tumor growth while T cells are 182 attracted, but not properly educated towards tumor antigens, resulting in a poor 183 clinical outcome. Finally, tumors from conventional precursors and with high activation 184 of the Wnt pathway, down-regulate their MHC Class I molecules and escape the T cell 185 attacks, which correlates with intermediate prognosis. 186

We also analyzed the immune signatures in ccRCC primary tumors from metastatic 187 patients treated with Sunitinib, a tyrosine kinase inhibitor. Unsupervised 188 transcriptomic analyses identified four molecular subgroups [34]. Patients of two 189 subgroups responded well to Sunitinib treatment, and consistently had longer 190 Progression-Free Survival (PFS) and OS. The patients of the two other subgroups 191 poorly responded to Sunitinib treatment, and had the worst prognosis. The analyses of 192 the immune signatures revealed that the worst prognostic group had the highest 193 infiltration with lymphoid cells in the context of a high myeloid cell infiltration. Genes 194 involved in T cell chemotaxis (CXCL9, CXCL10, CXCL13), Th1 orientation (IFN-γ, IL12, 195 TBX21). T cell exhaustion (PD1, PD-L1, LAG3) and inflammation (TNF-α, CSF1) were 196 highly expressed. The other bad prognosis group had the highest NK cell infiltration 197 and the lowest expression of genes related to adaptive immunity including MHC Class I 198 [32]. 199

Altogether, this analysis of two prototypic tumors showed that within each cancer type, an immunological classification identifies different subtypes of patients with different clinical outcomes corresponding to different oncogenic processes. Thus, we already identified three cancer types (Fig 1):

- the "immunogenic tumors" with production of immunogenic peptides, and an organized immune contexture that generates anti-tumor T cells and antibodies, resulting in a favorable prognosis,
- the "inflammatory tumors" with production of inflammatory, angiogenic and immunosuppressive molecules that disrupt the immune contexture and attract pro-tumor myeloid and lymphoid cells, resulting in exhausted tumor aggressiveness and poor patient's clinical outcome,
- the "escaping tumors" which down regulate their antigen-presenting machinery
   produce little or no chemokines and cytokines, and are blind to the immune
   attack. Patients with such tumors have an intermediate prognosis.
- These classifications not only reveal pathological features of cancers but also offer targets and predictive markers for immunotherapies (Table 1).

### The immune contexture and response to therapy

The immune microenvironment of tumors not only reflects the oncogenic processes of 217 a cancer in a patient, but is also a constitutive arm of cancer control and thus of 218patient's clinical outcome. It is therefore likely that therapeutic interventions modifying 219 the immune contexture will result in profound changes in cancer evolution. 220 Characterizing the immune contexture or the corresponding molecular profile of a 221 tumor in a patient will allow clinicians to propose the most appropriate therapies. For 222 instance, MSI CRC tumors with high mutational load respond to PD1 axis blockade [35]. 223 Consistent results have been observed in NSCLC [36]. In melanoma patients treated 224 with anti-PD-1 antibodies, responding tumors are characterized by the entry of pre-225 existing CD8<sup>+</sup> T cells from the IM to the CT[37], exemplifying the role of T cell location 226 into tumors[4]. In cervical carcinoma [38] or pancreatic cancer [39], responses to 227 therapeutic vaccination are accompanied by the increase of TLS in the tumor vicinity. 228 In ccRCC patients responding to anti-PD-1 or anti-PD-L1 antibodies, tumors express 229 both molecules on their infiltrating lymphocytes and tumor cells, respectively [40]. In 230 bladder cancer, the expression of PD-L1 on infiltrating immune cells was found 231 essential for response to this immunotherapy [41]. 232

For immune silent tumors, bi-specific antibodies direct to tumor antigens may attract 233 and activate T cells into responding tumors[42]. T cells-based therapies aim also to 234 bring missing memory T cells into tumors. Strikingly, therapies with antibodies 235 recognizing tumor-associated antigens, such as CD20 [43], or HER2-Neu [44], are 236 capable of inducing a memory anti-tumor T cell response responsible for the long-term 237 effect of these therapies. In tumors where class I MHC molecules are downregulated. 238 autologous NK cells transfer, recombinant IL15 or KIR-blocking antibodies could foster 239 NK cells activation and MHC-negative tumor cells elimination [45]. Finally, the immune 240 microenvironment is implicated in the long lasting effects of chemotherapies and 241 radiotherapies, by creating an appropriate microenvironment associated with the 242 release of immunogenic molecules by the tumor cells. 243

- 244
- 245
- 246
- 247
- 248

249

#### 250 **References**

Fridman, W.H., Pagès, F., Sautès-Fridman, C. & Galon, J.: The immune contexture
 in human tumours: impact on clinical outcome. *Nat Rev Cancer* 2012,12:298-306.

Pagès, F., Berger, A., Camus, M., Sanchez-Cabo, F., Costes, A., Molidor, R., Mlecnik,
 B., Kirilovsky, A., Nilsson, M., Damotte, D. et al.: Effector memory T cells, early
 metastasis, and survival in colorectal cancer. *N Engl J Med* 2005,353:2654-2666.

Zhang, L., Conejo-Garcia, J.R., Katsaros, D., Gimotty, P.A., Massobrio, M., Regnani,
G., Makrigiannakis, A., Gray, H., Schlienger, K., Liebman, M.N. et al.: Intratumoral T
cells, recurrence, and survival in epithelial ovarian cancer. N Engl J Med
2003,348:203-213.

Galon, J., Costes, A., Sanchez-Cabo, F., Kirilovsky, A., Mlecnik, B., Lagorce-Pagès, C.,
Tosolini, M., Camus, M., Berger, A., Wind, P. et al.: Type, Density, and Location of
Immune Cells Within Human Colorectal Tumors Predict Clinical Outcome. *Science*2006,313:1960-1964.

5. Mlecnik, B., Tosolini, M., Charoentong, P., Kirilovsky, A., Bindea, G., Berger, A., Camus, M., Gillard, M., Bruneval, P., Fridman, W. et al.: **Biomolecular network reconstruction identifies T-cell homing factors associated with survival in colorectal cancer**. *Gastroenterology* 2010,**138**:1429-1440.

Bindea, G., Mlecnik, B., Tosolini, M., Kirilovsky, A., Waldner, M., Obenauf, A.C.,
Angell, H., Fredriksen, T., Lafontaine, L., Berger, A. et al.: Spatiotemporal dynamics of
intratumoral immune cells reveal the immune landscape in human cancer. *Immunity* 2013,39:782-795.

Dieu-Nosjean, M., Antoine, M., Danel, C., Heudes, D., Wislez, M., Poulot, V., Rabbe,
N., Laurans, L., Tartour, E., de Chaisemartin, L. et al.: Long-term survival for patients
with non-small-cell lung cancer with intratumoral lymphoid structures. *J Clin*Oncol 2008,26:4410-4417.

8. Goc, J., Germain, C., Vo-Bourgais, T.K.D., Lupo, A., Klein, C., Knockaert, S., de
Chaisemartin, L., Ouakrim, H., Becht, E., Alifano, M. et al.: Dendritic cells in tumorassociated tertiary lymphoid structures signal a Th1 cytotoxic immune
contexture and license the positive prognostic value of infiltrating CD8+ T cells. *Cancer Res* 2014,74:705-715.

9. Germain, C., Gnjatic, S., Tamzalit, F., Knockaert, S., Remark, R., Goc, J., Lepelley, A.,
Becht, E., Katsahian, S., Bizouard, G. et al.: Presence of B cells in tertiary lymphoid
structures is associated with a protective immunity in patients with lung cancer.
Am J Respir Crit Care Med 2014,189:832-844.

10. Dieu-Nosjean, M., Goc, J., Giraldo, N.A., Sautès-Fridman, C. & Fridman, W.H.:
Tertiary lymphoid structures in cancer and beyond. *Trends Immunol* 2014,35:571580.

Martinet, L., Garrido, I., Filleron, T., Le Guellec, S., Bellard, E., Fournie, J., Rochaix,
P. & Girard, J.: Human solid tumors contain high endothelial venules: association
with T- and B-lymphocyte infiltration and favorable prognosis in breast cancer. *Cancer Res* 2011,**71**:5678-5687.

12. Becht, E., Giraldo, N.A., Germain, C., de Reyniès, A., Laurent-Puig, P., Zucman-Rossi,
J., Dieu-Nosjean, M., Sautès-Fridman, C. & Fridman, W.H.: Immune contexture,
immunoscore and malignant cell molecular subgroups for prognostic and
theranostic classifications of cancers. Advances in immunology In press,:.

13. Klein, J.L., Nguyen, T.T., Bien-Willner, G.A., Chen, L., Foyil, K.V., Bartlett, N.L.,
Duncavage, E.J., Hassan, A., Frater, J.L. & Kreisel, F.: CD163 immunohistochemistry is
superior to CD68 in predicting outcome in classical Hodgkin lymphoma. *Am J Clin Pathol* 2014,141:381-387.

Jensen, T.O., Schmidt, H., Møller, H.J., Høyer, M., Maniecki, M.B., Sjoegren, P.,
 Christensen, I.J. & Steiniche, T.: Macrophage markers in serum and tumor have
 prognostic impact in American Joint Committee on Cancer stage I/II melanoma. J
 Clin Oncol 2009,27:3330-3337.

Hou, Y., Chao, Y., Tung, H., Wang, H. & Shan, Y.: Coexpression of CD44 positive/CD133-positive cancer stem cells and CD204-positive tumor-associated
 macrophages is a predictor of survival in pancreatic ductal adenocarcinoma.
 *Cancer* 2014,120:2766-2777.

Medrek, C., Pontén, F., Jirström, K. & Leandersson, K.: The presence of tumor
 associated macrophages in tumor stroma as a prognostic marker for breast
 cancer patients. *BMC Cancer* 2012,12:306.

17. Platonova, S., Cherfils-Vicini, J., Damotte, D., Crozet, L., Vieillard, V., Validire, P.,
André, P., Dieu-Nosjean, M., Alifano, M., Régnard, J. et al.: Profound coordinated
alterations of intratumoral NK cell phenotype and function in lung carcinoma. *Cancer Res* 2011,71:5412-5422.

Takeuchi, H., Maehara, Y., Tokunaga, E., Koga, T., Kakeji, Y. & Sugimachi, K.: **Prognostic significance of natural killer cell activity in patients with gastric carcinoma: a multivariate analysis**. *Am J Gastroenterol* 2001,96:574-578.

Rusakiewicz, S., Semeraro, M., Sarabi, M., Desbois, M., Locher, C., Mendez, R.,
Vimond, N., Concha, A., Garrido, F., Isambert, N. et al.: Immune infiltrates are
prognostic factors in localized gastrointestinal stromal tumors. *Cancer Res*2013,73:3499-3510.

Taketomi, A., Shimada, M., Shirabe, K., Kajiyama, K., Gion, T. & Sugimachi, K.:
 Natural killer cell activity in patients with hepatocellular carcinoma: a new
 prognostic indicator after hepatectomy. *Cancer* 1998,83:58-63.

Nakano, O., Sato, M., Naito, Y., Suzuki, K., Orikasa, S., Aizawa, M., Suzuki, Y., 21. 325 Shintaku, I., Nagura, H. & Ohtani, H.: Proliferative activity of intratumoral CD8(+) T-326 a prognostic factor in human renal cell carcinoma: lymphocytes as 327 clinicopathologic demonstration antitumor of immunity. Cancer Res 328 2001,61:5132-5136. 329

Scott, D.W., Chan, F.C., Hong, F., Rogic, S., Tan, K.L., Meissner, B., Ben-Neriah, S.,
 Boyle, M., Kridel, R., Telenius, A. et al.: Gene expression-based model using formalin fixed paraffin-embedded biopsies predicts overall survival in advanced-stage
 classical Hodgkin lymphoma. *J Clin Oncol* 2013,31:692-700.

Whelchel, J.C., Farah, S.E., McLean, I.W. & Burnier, M.N.: Immunohistochemistry
 of infiltrating lymphocytes in uveal malignant melanoma. *Invest Ophthalmol Vis Sci* 1993,34:2603-2606.

Giraldo, N.A., Becht, E., Pagès, F., Skliris, G., Verkarre, V., Vano, Y., Mejean, A., SaintAubert, N., Lacroix, L., Natario, I. et al.: Orchestration and Prognostic Significance of
Immune Checkpoints in the Microenvironment of Primary and Metastatic Renal
Cell Cancer. *Clin Cancer Res* 2015,21:3031-3040.

Remark, R., Alifano, M., Cremer, I., Lupo, A., Dieu-Nosjean, M., Riquet, M., Crozet,
L., Ouakrim, H., Goc, J., Cazes, A. et al.: Characteristics and clinical impacts of the **immune environments in colorectal and renal cell carcinoma lung metastases: influence of tumor origin**. *Clin Cancer Res* 2013,19:4079-4091.

Taube, J.M., Anders, R.A., Young, G.D., Xu, H., Sharma, R., McMiller, T.L., Chen, S.,
Klein, A.P., Pardoll, D.M., Topalian, S.L. et al.: Colocalization of inflammatory **response with B7-h1 expression in human melanocytic lesions supports an**adaptive resistance mechanism of immune escape. *Sci Transl Med* 2012,4:127ra37.

349 27. Hanahan, D. & Weinberg, R.A.: Hallmarks of cancer: the next generation. *Cell*350 2011,144:646-674.

28. Dunn, G.P., Old, L.J. & Schreiber, R.D.: The immunobiology of cancer
immunosurveillance and immunoediting. *Immunity* 2004,21:137-148.

29. Dienstmann, R., Guinney, J., Delorenzi, M., de Reynies, A., Roepman, P.,
Sadanandam, A., Vermeulen, L., Schlicker, A., Missiaglia, E., Soneson, C. et al.: Colorectal
Cancer Subtyping Consortium (CRCSC) identification of a consensus of molecular
subtypes.. J Clin Oncol 2014,32:5s.

30. Becht, E., Giraldo, N.A., Beuselinck, B., Job, S., Marisa, L., Vano, Y., Oudard, S.,
Zucman-Rossi, J., Laurent-Puig, P. & Sautès-Fridman, C.: Prognostic and theranostic
impact of molecular subtypes and immune classifications in Renal Cell Cancer
(RCC) and Colorectal Cancer (CRC). Oncoimmunology 2015,4:12:.

361 31. Beuselinck, B., Job, S., Becht, E., Karadimou, A., Verkarre, V., Couchy, G., Giraldo, N.,
 362 Rioux-Leclercq, N., Molinié, V., Sibony, M. et al.: Molecular subtypes of clear cell renal
 363 cell carcinoma are associated with sunitinib response in the metastatic setting.
 364 *Clin Cancer Res* 2015,21:1329-1339.

365 32. Le, D.T., Uram, J.N., Wang, H., Bartlett, B.R., Kemberling, H., Eyring, A.D., Skora, 366 A.D., Luber, B.S., Azad, N.S., Laheru, D. et al.: **PD-1 Blockade in Tumors with** 367 **Mismatch-Repair Deficiency**. *N Engl J Med* 2015,**372**:2509-2520.

368 33. Rizvi, N.A., Hellmann, M.D., Snyder, A., Kvistborg, P., Makarov, V., Havel, J.J., Lee,
369 W., Yuan, J., Wong, P., Ho, T.S. et al.: Cancer immunology. Mutational landscape
370 determines sensitivity to PD-1 blockade in non-small cell lung cancer. Science
371 2015,348:124-128.

372 34. Tumeh, P.C., Harview, C.L., Yearley, J.H., Shintaku, I.P., Taylor, E.J.M., Robert, L.,
373 Chmielowski, B., Spasic, M., Henry, G., Ciobanu, V. et al.: PD-1 blockade induces
374 responses by inhibiting adaptive immune resistance. *Nature* 2014,515:568-571.

375 35. Maldonado, L., Teague, J.E., Morrow, M.P., Jotova, I., Wu, T.C., Wang, C., Desmarais,
376 C., Boyer, J.D., Tycko, B., Robins, H.S. et al.: Intramuscular therapeutic vaccination
377 targeting HPV16 induces T cell responses that localize in mucosal lesions. *Sci*378 *Transl Med* 2014,6:221ra13.

36. Lutz, E.R., Wu, A.A., Bigelow, E., Sharma, R., Mo, G., Soares, K., Solt, S., Dorman, A.,
Wamwea, A., Yager, A. et al.: Immunotherapy converts nonimmunogenic pancreatic
tumors into immunogenic foci of immune regulation. *Cancer Immunol Res*2014,2:616-631.

37. Taube, J.M., Klein, A., Brahmer, J.R., Xu, H., Pan, X., Kim, J.H., Chen, L., Pardoll, D.M.,
Topalian, S.L. & Anders, R.A.: Association of PD-1, PD-1 Ligands, and Other Features
of the Tumor Immune Microenvironment with Response to Anti-PD-1 Therapy. *Clin Cancer Res* 2014,20:5064-5074.

38. Powles, T., Eder, J.P., Fine, G.D., Braiteh, F.S., Loriot, Y., Cruz, C., Bellmunt, J., Burris,
H.A., Petrylak, D.P., Teng, S. et al.: MPDL3280A (anti-PD-L1) treatment leads to
clinical activity in metastatic bladder cancer. *Nature* 2014,515:558-562.

390 39. Giraldo, N.A., Becht, E., Remark, R., Damotte, D., Sautès-Fridman, C. & Fridman,
391 W.H.: The immune contexture of primary and metastatic human tumours. *Curr*392 *Opin Immunol* 2014,27:8-15.

40. Deligne, C., Metidji, A., Fridman, W. & Teillaud, J.: Anti-CD20 therapy induces a
memory Th1 response through the IFN-γ/IL-12 axis and prevents protumor
regulatory T-cell expansion in mice. *Leukemia* 2015,29:947-957.

41. Mortenson, E.D. & Fu, Y.: Adaptive Immune Responses and HER2/neu Positive
Breast Cancer. *Curr Pathobiol Rep* 2013,1:37-42.

- 398
- 399

### 400 **References of interest and outstanding interest**

401

402 [1] \* A review on the impact of the immune contexture in various cancers

[3] \* A demonstration that CD3<sup>+</sup> T cells densities correlate with favorable outcome in a
 human cancer

[4] \*\* The demonstration of the role of the density, location and functional orientation
of T cells in controlling the outcome of a human cancer

[6] \*\* A comprehensive analysis of the immune contexture in colorectal cancers
 depicting the clinical impact of all immune cell populations, their locations, functional
 orientations and their relationship with cancer stage

- 410 [7] \* The first demonstration of the clinical impact of TLS in cancer patients
- [8] \* The demonstration that TLS shape intratumor CD8<sup>+</sup> T cell responses
- 412 [9] \* The demonstration that intratumor B lymphocytes produce anti-tumor antibodies
- [10] \* A comprehensive review of the role of TLS in cancers
- [27] \* The analysis of the impact of components of the immune contexture in the
- shaping an efficient or deleterious anti-tumor immune reaction
- [28] \* A demonstration of the clinical impact of the immune contexture in metastaticcancer sites
- [35] **\*\*** A clinical trial demonstrating that inhibition of immune checkpoint results in
- longer survival in colorectal-cancer patients with hypermutated MSI tumors



#### 422 **Figure 1**.

421

The immunological wheel. Cartoon depicting the three immune contextures than can 423 be induced in tumors. The 'immunogenic tumors' are characterized by abundant 424 Cytotoxic T-Lymphocyte (CTL) infiltration, the presence of Tertiary Lymphoid 425 Structures (TLS) and low/moderate vascularization while associated with the longest 426 patient's survival. The 'immune neglected' tumors are characterized by lack of 427 infiltration by immune cells, low/moderate vascularization and intermediate 428 prognosis. Finally, the 'inflammatory tumors' are characterized by abundant CTL in the 429 absence of TLS, conspicuous infiltration with M2 macrophages, severe vascularization 430 and poor prognosis. 431

Immune subgroup	Examples of corresponding molecular subgroups	Microenvironment characteristics	Escape mechanisms	Immunotherapeutic goals	Potential immunotherapeutic approach
Immunogenic	CRC Hypermutated	High T cell infiltration with Th1 orientation and cytotoxic lymphocytes	Immune checkpoints : PD1 axis, LAG3, CTLA4	Boost intratumor cytotoxic T cells	Checkpoint-blockade
Inflammatory	CRC Mesenchymal ccrcc4	High lymphocyte and myeloid cells infiltration High angiogenesis Stromal mesenchymal cells	Hypoxia TGFß PD1 axis	Dampen inflammation and associated suppressive mechanisms Establish normoxia Boost intratumor suppressed cytotoxic T cells	Anti-angiogenic Anti TGFβ Checkpoint-blockade
Immune neglected	CRC Canonical and Metabolic ccrcc1 and ccrcc2	Low lymphocytic and myeloid cells infiltration	Low class I MHC expression	Attract cytotoxic T cells in tumors Bypass class I MHC presentation	CAR T cells Bi-specific antibodies

433

434 **Table 1**.

### Immunotherapeutic approaches tailored for tumor immune subgroups.

The three immune subgroups, their hallmarks, immune escape mechanisms are listed,as well as the corresponding immunotherapeutic goals and potential approaches.

- 438 CRC : Colorectal cancer
- 439 ccrcc : clear-cell Renal Cell Carcinoma

Financial support: This work was supported by the 'Institut National de la 441 Santé et de la Recherche Médicale', the University Paris-Descartes, the 442 University Pierre et Marie Curie, the Institut National du Cancer (2009-1-443 PLBIO-07-INSERM 6-1, 2010-1-PLBIO-03-INSERM 6-1, 2011-1-PLBIO-06-444 INSERM 6-1), CARPEM (CAncer Research for PErsonalized Medicine), Labex 445 (LAXE62\_9UMS872 Immuno-Oncology FRIDMAN, 11LAXE62\_9UMS872 446 FRIDMAN), ans the Fondation ARC pour la Rercherche sur le Cancer 447 (SL220110603483), the Universidad de los Andes School of Medicine, 448 449 Colciencias (NAG). EB is supported by CARPEM post-doctorate fellowship and NAG by PPATH doctorate fellowship. 450

Acknowledgements: We thank the members of the teams of I. Cremer/J.L.
Teillaud and J. Galon at the Cordeliers Research Center for their fruitful
discussions and who performed most of the work cited in this review.