

CASE REPORT

Open Access



# Genetic prescreening of a candidate for laser refractive surgery identifies risk for inadequate tissue response: a case report

Andrea Cusumano<sup>1</sup> , Hila Roshanravan<sup>2</sup>, Connie Chao-Shern<sup>2</sup>, Jacopo Sebastiani<sup>3</sup>, Jung Hee Levaldi Ghiron<sup>3</sup>, Larry DeDionisio<sup>2\*†</sup> and Tara Moore<sup>2,4†</sup>

## Abstract

**Background:** Inadequate response to corneal laser refractive surgery, e.g., ectatic corneal diseases, may not be identified by conventional examinations, hence creating therapeutic uncertainty. Herein we demonstrate the application of genetic prescreening to augment preassessment for corneal laser refractive surgery and highlight the ability to prevent the possibility of enrolling a subject at risk for developing ectatic corneal diseases.

**Case presentation:** Preoperative tests were performed alongside deoxyribonucleic acid (DNA) sequencing of 75 genes specific to the structure and health of the eye of a 44-year-old Caucasian male candidate for corneal laser refractive surgery. The patient had no medical, family, or psychosocial history, nor symptoms that could lead to suspect any corneal abnormalities, and conventional preoperative tests confirmed that no corneal abnormalities were present. The sequencing results uncovered rare DNA variants within the *ADGRV1*, *PTK2*, *ZNF469*, and *KRT15* genes. These variants were considered potential risk factors for inadequate response in the patient post corneal laser refractive surgery. Subsequent reevaluation with three different last-generation corneal tomographers identified in the left eye a “warning” for a deformity of the posterior profile of the cornea.

**Conclusions:** Genetic prescreening identifies potential risk of inadequate response to corneal laser refractive surgery where current technologies in use may lead to a hazardous predictive diagnostic uncertainty.

**Keywords:** Refractive surgery, Corneal ectatic disease, Genetic testing

## Background

The possibility of postoperative ectasia following corneal laser refractive surgery (CLRS) is an ongoing concern within the ophthalmic community [1, 2]. Although uncommon, occurrences of ectasia have been observed after laser-assisted *in situ* keratomileusis (LASIK) and have also been reported after less disruptive procedures such as small incision lenticule extraction (SMILE) [3,

4]. A predisposition to develop an ectatic corneal disease (ECD) such as keratoconus (KC) can lead to inadequate responses to CLRS. During the preoperative phase for patients considering CLRS, conventional examinations, including corneal topography, may not easily identify the propensity to develop an ECD.

Important information can be obtained regarding a patient’s risk for developing an ECD through genetic testing. Genetic factors are known to play a role in the etiology of ECDs such as KC [5, 6], and a wide range of evidence exists indicating a link between central corneal thickness (CCT), a highly heritable trait associated with ECD, and multiple loci within the human genome [7–11]. Thus, augmenting conventional examinations

<sup>†</sup>Larry DeDionisio and Tara Moore joint senior authorship.

\*Correspondence: larry@avellino.com

<sup>2</sup> Avellino Labs, Menlo Park, CA, USA

Full list of author information is available at the end of the article



with a genetic prescreening (GP) test provides a powerful tool in determining risk for postoperative ectasia. Herein, we present a representative case illustrating the utility of GP in uncovering genetic variants within the deoxyribonucleic acid (DNA) of the patient and his children. We advocate GP to assess a patient’s candidacy for CLRS, thus preventing the possibility of enrolling subjects at risk of developing ECDs or inadequate responses to CLRS with poor postoperative visual outcomes and complex corneal healing.

**Case presentation**

**Initial clinical examinations and genetic testing**

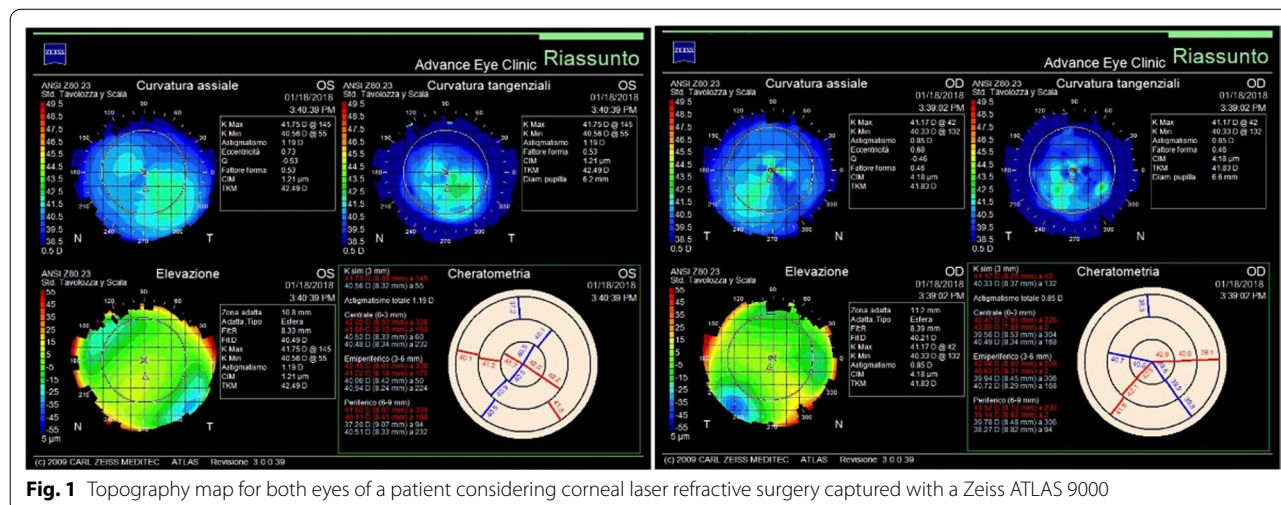
Preoperative tests, including corneal topography utilizing a Zeiss ATLAS 9000 (Carl Zeiss AG, Oberkochen, Germany), were performed on a 44-year-old Caucasian male patient considering CLRS. The patient did not have medical, family, or psychosocial history that could indicate predisposition or existence of corneal abnormalities, nor did he show any symptoms that could suggest corneal aberrations. Moreover, the patient had never undergone previous eye interventions. No financial, language, or cultural challenges occurred regarding the management of this case. The results from the initial presentation (T = 0) (that is, topographic examination) did not indicate corneal abnormalities in either eye of the patient (Fig 1), nor any other ocular pathologies. For GP, a buccal mucosa sample was collected by using an iSWAB collection kit (Mawi DNA Technologies, Hayward, CA, USA). After DNA was extracted (QIAamp DNA mini kit, QIAGEN Inc. Hilden, Germany) from the sample, a next-generation sequencing (NGS) test (AvaGen, Avellino Lab, Menlo Park, CA, USA) targeting 75 genes specific to the structure and health of the eye was used to analyze the patient’s

DNA. The sequencing results identified four potentially pathogenic variants located on chr 5 (*ADGRV1*), chr 8 (*PTK2*), chr 16 (*ZNF469*), and chr 17 (*KRT15*) within the patient’s genome (Table 1). These DNA mutations were found to carry a risk factor for ECD based on the following analysis.

**Sequence variant analysis**

Adhering to the guidelines put forward by the American College of Medical Genetics and Genomics (ACMG) for the interpretation of sequence variants [12], the following criteria were applied to determine which variants within the patient’s test results were most likely damaging and thus carry a risk for ECD. Sequence variants were filtered based on their minor allele frequency (MAF) in the general population and protein coding changes, categorized as missense, STOP gain/loss, nonsense, or frameshift/non-frameshift indels. Variants with an MAF < 0.01 based on The Genome Aggregation Database (<https://gnomad.broadinstitute.org/>) were considered to be possibly pathogenic. The pathology for each variant was also gauged by using three in silico prediction tools: PolyPhen2-HDIV, LRT, and PROVEAN. Each tool aims to determine the likely impact on the transcribed amino acid sequence and translated protein due to a change in the DNA sequence. All genome positions were based on the Genome Reference Consortium Human genome build 37 (GRCh37.p13).

The filtering criteria resulted in the four heterozygous, missense mutations detailed in Table 1. The *ADGRV1* and *KRT15* variants were classified as possibly damaging, deleterious, and damaging by PolyPhen2-HDIV, LRT, and PROVEAN respectively, while the variants in the *PTK2*



**Fig. 1** Topography map for both eyes of a patient considering corneal laser refractive surgery captured with a Zeiss ATLAS 9000

**Table 1** Variants found in the patient's and his children's deoxyribonucleic acid

	Chr	Position	Ref. allele	Allele 1	Allele 2	Gene	dbSNP	Protein/amino acid	PolyPhen2 prediction	HumDiv	LRT prediction	PROVEAN prediction
Father	5	90079091	A	A	G	ADGRV1	rs182698253	p.H4461R	Probably damaging		Deleterious	Damaging
	8	141771274	T	T	C	PTK2	N/A	p.Y526C	N/A		N/A	N/A
Daughter	16	88504716	G	G	T	ZNF469	rs568704426	p.G3585V	Benign		N/A	Neutral
	17	39674737	G	G	A	KRT15	rs201164162	p.R115C	Probably damaging		Deleterious	Damaging
Son	17	39674737	G	G	A	KRT15	rs201164162	p.R115C	Probably damaging		Deleterious	Damaging
	5	90079091	A	A	G	ADGRV1	rs182698253	p.H4461R	Probably damaging		Deleterious	Damaging
	16	88504716	G	G	T	ZNF469	rs568704426	p.G3585V	Benign		N/A	Neutral
	17	39674737	G	G	A	KRT15	rs201164162	p.R115C	Probably damaging		Deleterious	Damaging

and *ZNF469* genes were classified as benign, neutral, or no data available by the in silico prediction tools we used.

**Gene network analysis**

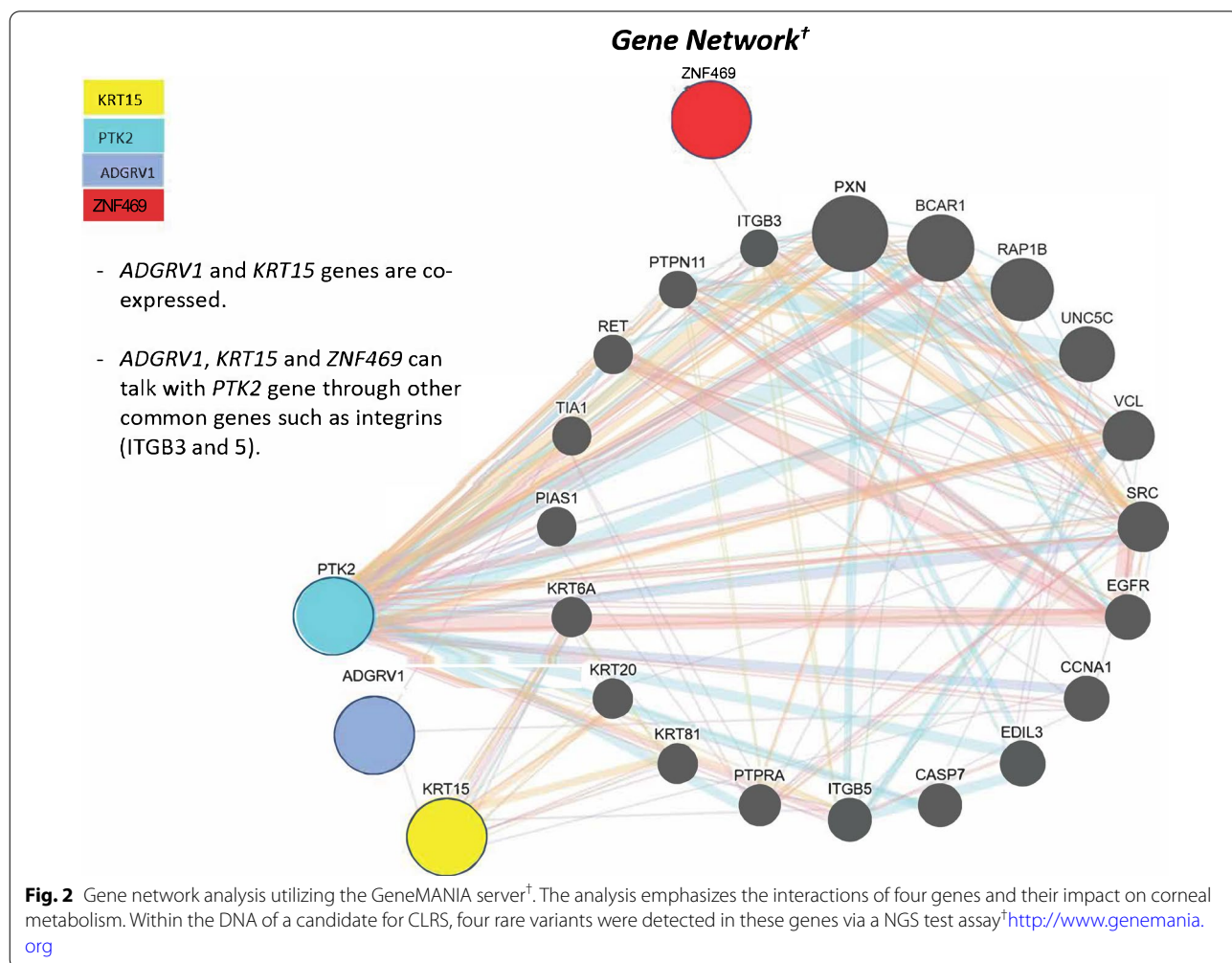
A further analysis was conducted by using the GeneMANIA server (<http://www.genemania.org>) [13] to provide insight of gene function for the four genes and how these variants within each gene could affect the health of the cornea via coordinated molecular pathways (Fig 2). This analysis found that *ADGRV1* and *KRT15* are co-expressed and that *ADGRV1*, *KRT15*, and *ZNF469* interact with *PTK2* via integrins, transmembrane cell adhesion receptors that play important roles in the regulation of cell migration during development, wound healing, and inflammatory response [14]. All four genes are integral to eye development and various molecular pathways. A few have been indicated in eye diseases of various types.

**Adhesion G protein-coupled receptor V1 (*ADGRV1*) variant ID: rs182698253**

The missense variant located in *ADGRV1* results in a change in the amino acid sequence of histidine to arginine at position 4661 (H4661R) within a region of the *ADGRV1* protein that overlaps with a Calx-beta domain. The Calx-beta motif is a protein motif that is used for calcium binding and regulation, and this variant may lead to conformational changes in the protein that affects the binding of the *ADGRV1* protein to its interacting partners by disrupting calcium binding sites. *ADGRV1* is part of the *USH2* complex, which plays an essential role in the development of hearing and vision via cell surface receptor signaling pathways [15]. A variant within this gene has previously been reported in two brothers suffering from KC [16].

**Protein tyrosine kinase 2 (*PTK2*) variant: no rsID**

Currently, there is no evidence that links *PTK2* to ECD. However, the *PTK2* variant found in the proband induces





an amino acid change of tyrosine to cystine (Y726C) in the functional domain of the PTK2 protein. *PTK2* plays an essential role in regulating cell migration, adhesion, spreading, and reorganization of the actin cytoskeleton [17].

#### **Zinc finger 469 (ZNF469) variant ID: rs568704426**

This missense variant in the *ZNF469* gene results in the conversion of amino acid 3585 from glycine to valine (G3585V) within the ZNF469 protein. It is considered benign and neutral by PolyPhen2-HDIV and PROVEAN, respectively; however, the region where it is located overlaps with two regulatory regions that may affect binding of certain transcription factors (TFs) to their sites. There are numerous studies that have linked mutations in *ZNF469* to KC and to brittle cornea syndrome (BCS), an extreme form of ECD [18–22].

#### **Keratin 15 (KRT15) variant ID: rs201164162**

*KRT15* is expressed in corneal limbal epithelial cells and is involved in wound healing [23, 24]. The missense variant found in this gene, which results in an amino acid change from arginine to cysteine (R115C), overlaps with an intermediate filament rod domain in the protein structure. This plays a significant role during the protein assembly process, and mutations in this functionally important area disrupt end-to-end keratin interactions that could subsequently affect the corneal epithelia [25]. This variant also falls within a regulatory region that serves as a TF binding site [26]. Based on tissue and upstream signaling pathways, the related TF either enhances or suppresses the transcription of *KRT15*. Any dysregulation of *KRT15* at this TF binding site caused by this variant could affect changes in the cytoskeleton and could possibly affect the development of the lacrimal gland [26].

#### **Subsequent reevaluation with three different corneal tomographers**

After the results of GP determined the presence of potentially pathogenic variants, the patient was recalled and reevaluated with three different advanced tomographers (T = 52 days after initial presentation): a Pentacam (OCULUS, Wetzlar, Germany), an Orbscan (Bausch & Lomb, Rochester, NY, USA), and a SIRIUS (CSO, Florence, Italy). A deformity of the posterior profile of the cornea in the left eye (LE) was identified with all three instruments. The tomography measurements are shown in Fig. 3. The Pentacam produced a Belin/Ambrósio Enhanced Ectasia Display that highlighted a “warning” signal (yellow) that showed an elevation of

12 microns in the differential map of the back surface of the cornea (Fig 3A). The Orbscan showed the presence of an alteration of the corneal posterior elevation of 0.058 mm (Fig 3B). The results of the Sirius further confirmed the presence of an asymmetry of the curvature expressed in D (diopters) between the front and back face of the cornea in the LE, Slb = 0.24 D (Fig 3C).

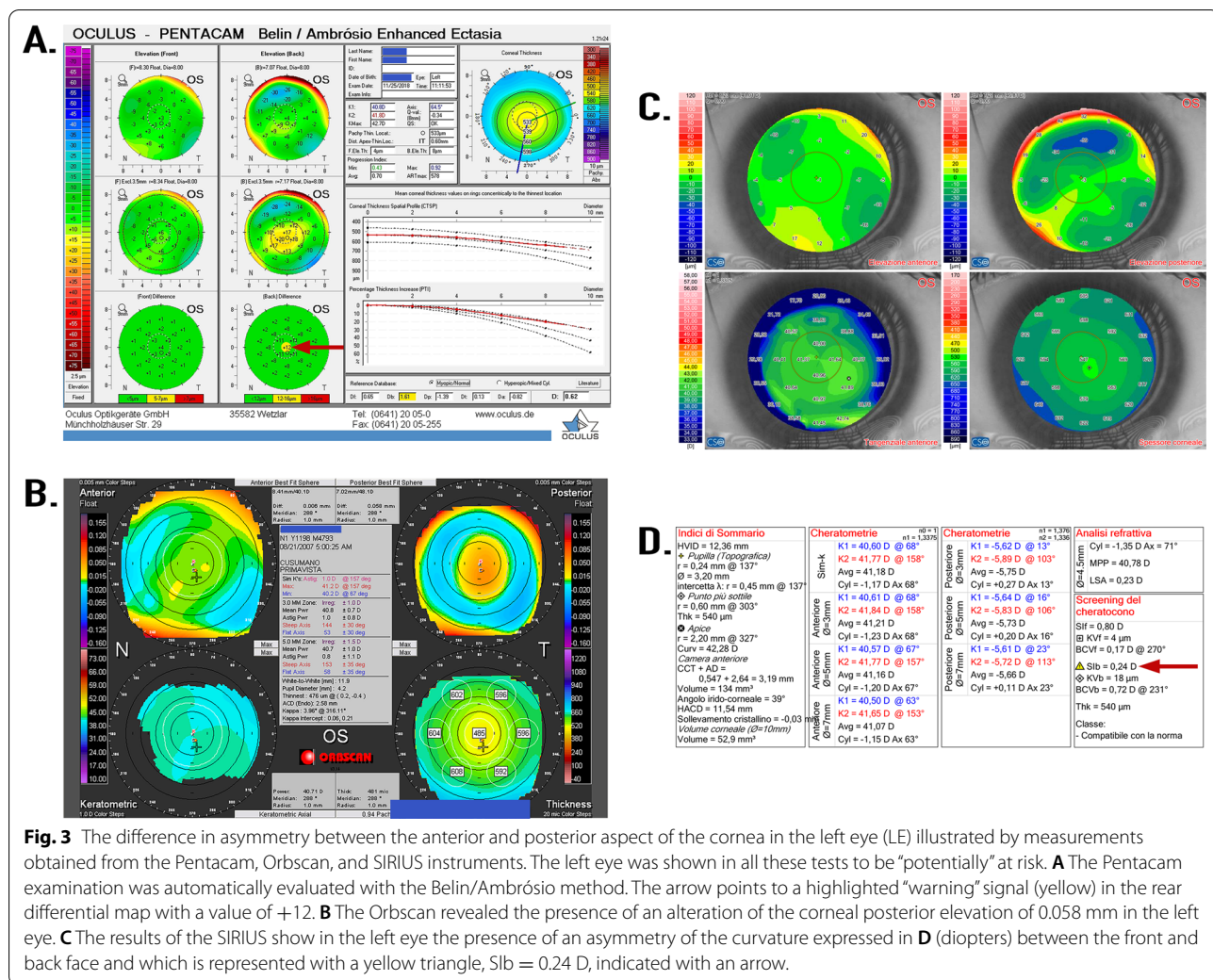
#### **Discussion and conclusions**

Genetic screening identified four rare variants in four genes within the DNA of a candidate for CLRS. Further clinical tests found an abnormality in the cornea of the patient’s LE. These results led to the decision to not perform any type of corneal laser refractive surgery due to the existing risk of inadequate responses in the patient’s cornea. Not having received any treatment, the patient was suggested to undergo follow-up visits every 12 months. Prior to GP, an initial corneal topography examination did not find any aberrations that would have warned the physician about possible poor postoperative results. DNA sequencing of the patient’s children uncovered, within the son, the variants located in the *ADGRV1*, *ZNF469*, and *KRT15* genes, while the daughter inherited the *KRT15* gene variant (Table 1). Based solely on the genetic inheritance pattern, the son is therefore at higher risk for developing an ECD than the daughter, since his genotype showed that he shared three of the four variants found in the father’s DNA. We must emphasize that the most common form of ECD, KC, is known to be a multifactorial disease [5, 6]. Environmental, behavioral, and genetic factors contribute to the onset of KC, and each of these factors should be considered in evaluating a risk for the disease.

#### **Evidence of a genetic risk for ECD**

It is worth noting that the *KRT15* variant uncovered here could deleteriously affect other molecular pathways beyond the structure of the keratin 15 protein for which it encodes. Apart from its expression pattern in corneal limbal epithelial cells [23], the gene network analysis revealed that the region where this variant lies serves as a regulatory region crucial for lacrimal gland development [26]. We also note that *KRT15* is the host gene for *MIR6510*, a noncoding microRNA that which is involved in posttranscriptional regulation of gene expression and is part of the keratinization pathway. Further research is needed to fully understand how the R115C variant in *KRT15* might impact the functionality of *MIR6510*.

We determined that the *KRT15* gene is directly linked to *ADGRV1* through coexpression of their proteins. Given that the H4661R variant uncovered in *ADGRV1* could disrupt its functionality, we postulate that this variant, paired with the potential deleterious effects of



**Fig. 3** The difference in asymmetry between the anterior and posterior aspect of the cornea in the left eye (LE) illustrated by measurements obtained from the Pentacam, Orbscan, and SIRIUS instruments. The left eye was shown in all these tests to be “potentially” at risk. **A** The Pentacam examination was automatically evaluated with the Belin/Ambrósio method. The arrow points to a highlighted “warning” signal (yellow) in the rear differential map with a value of +12. **B** The Orbscan revealed the presence of an alteration of the corneal posterior elevation of 0.058 mm in the left eye. **C** The results of the SIRIUS show in the left eye the presence of an asymmetry of the curvature expressed in **D** (diopeters) between the front and back face and which is represented with a yellow triangle, Slb = 0.24 D, indicated with an arrow.

*KRT15* variant, could ultimately alter the integrity of the cornea and play a role in the development of ECD. Furthermore, *KRT15* and *ADGRV1* indirectly interact with *ZNF469* and *PTK2* through cell surface transmembrane receptors known as integrins. Integrins recognize and bind to extracellular matrix (ECM) proteins. They comprise various  $\alpha$ - and  $\beta$ -subunits, resulting in 24 different protein combinations that have overlapping substrate specificity and cell-type-specific expression patterns [27]. The gene network analysis (Fig. 2) indicated that *PTK2* activates the integrins *ITGB3* and *ITGB5*, and that *ZNF469* is part of an integrin complex with *ITGB3*. Since the Y726C variant discovered in *PTK2* could alter the structure of the encoded protein, it could affect its ability to activate its integrin substrates and subsequently alter interactions between the *ADGRV1* and keratin proteins with the ECM.

In addition, *ZNF469* plays a role in collagen homeostasis and corneal structure. The *ZNF469* protein

expressed in the human cornea shares 30% homology with the helical regions of collagen I and collagen IV, suggesting that it may function as a transcription factor or extranuclear regulator factor for the synthesis or organization of collagen fibers [21, 28]. *ZNF469* may also be involved in the transforming growth factor (TGF)- $\beta$  pathway [29], whose disturbance would lead to disarray of collagen in human cornea. During wound healing, TGF- $\beta$ 1 induces the expression of major ECM proteins such as fibronectin and collagen [30]. *ZNF469* has been linked to CCT and its association to KC [7, 9, 22] and is implicated in the etiology for BCS [20, 21]. Nonetheless, there is evidence suggesting that *ZNF469* is not causative for KC in some population groups of European descent [31, 32]. The evidence indicates that, when viewed in isolation of the other variants, the G3585V variant in *ZNF469* is most likely benign (Table 1); however, a combination of this variant in conjunction with the other damaging and deleterious

variants in *KRT15* and *ADGRV1* most likely contributed to the phenotype illustrated in the patient's corneal tomography (Fig. 3).

The gene network analysis (Fig. 2) also indicates that the *PTK2* variant may also participate in the morphologic phenotype. Overall, the evidence presented here points to the likelihood that corneal aberrations that could lead to ECD can be attributed to a combination of different variants within a patient's genome, a composite of both benign and damaging, rare and common variants, a polygenic etiology. Given the various gene interactions among the four genes under scrutiny, we must not conclude that the patient's son is at lower risk for ECD because the *PTK2* variant was not present in his DNA, nor can we state that the daughter is free of risk for ECD simply because she inherited only one potentially pathogenic variant from her father (R115C in *KRT15*).

The four variants identified in this report are in genes that are part of a molecular network crucial to a healthy cornea. Their predicted interdependency, and in the case of the *KRT15* and *ADGRV1* variants, potential pathogenicity, can affect the expected refractive correction and corneal healing after CLRS is performed. We demonstrate here how GP has the potential to identify risk of inadequate response to CLRS where even the most advanced technologies in use today may leave uncertainty for "borderline cases," without being able to give a true indication of the potential risk for corneal ectasia.

#### Abbreviations

CLRS: Corneal laser refractive surgery; ECD: Ectatic corneal disease; GP: Genetic prescreening; KC: Keratoconus.

#### Acknowledgements

None.

#### Author contributions

AC conducted the clinical examinations and wrote the clinical interpretation. HR provided the genetic interpretation and gene network analysis. CCS conducted the genetic testing in the laboratory. JS and JHLG assisted in the clinical examination of the patient and contributed to the interpretation of the clinical findings. LD drafted the manuscript and contributed to the genetic interpretation. TM provided critical revision of the manuscript. TM and LD jointly supervised the study. All authors read and approved the final manuscript.

#### Funding

No specific funding was received for this study.

#### Availability of data and materials

The datasets generated for this study are not publicly available; however, data are available from the corresponding author on reasonable request.

#### Declarations

##### Ethics approval and consent to participate

Ethics approval is not necessary for case presentations at our institution. In adherence to the tenets of the Declaration of Helsinki, the protocol of our study was explained to the subjects and consent was obtained.

#### Consent for publication

Written informed consent was obtained from the patient for publication of this case report and any accompanying images. A copy of the written consent is available for review by the Editor-in-Chief of this journal.

#### Competing interests

AC, Avellino Labs, non-remunerative; HR and CCS, former employees of Avellino Labs; JS and JHLG, none; LD, employee of Avellino Labs; TM, consultant to Avellino Labs

#### Author details

<sup>1</sup>University of Rome Tor Vergata, Rome, Italy. <sup>2</sup>Avellino Labs, Menlo Park, CA, USA. <sup>3</sup>Macula & Genoma Foundation Onlus, Rome, Italy. <sup>4</sup>Ulster University, Coleraine, UK.

Received: 26 February 2021 Accepted: 31 March 2022

Published online: 16 May 2022

#### References

- Rabinowitz YS. Ectasia after laser *in situ* keratomileusis. *Curr Opin Ophthalmol.* 2006;17(5):421–6.
- Jin SX, Dackowski E, Chuck RS. Risk factors for postlaser refractive surgery corneal ectasia. *Curr Opin Ophthalmol.* 2020;31(4):288–92.
- Randleman JB. Ectasia after corneal refractive surgery: nothing to SMILE about. *J Refract Surg.* 2016;32(7):434–5.
- Spadea L, Cantera E, Cortes M, Conocchia NE, Stewart CW. Corneal ectasia after myopic laser *in situ* keratomileusis: a long-term study. *Clin Ophthalmol (Auckland, NZ).* 2012;6:1801.
- Rong SS, Ma STU, Yu XT, Ma L, Chu WK, Chan TCY, et al. Genetic associations for keratoconus: a systematic review and meta-analysis. *Sci Rep.* 2017;7(1):1–11.
- Gordon-Shaag A, Millodot M, Shneur E, Liu Y. The genetic and environmental factors for keratoconus. *Biomed Res Int.* 2015;2015:795738.
- Lu Y, Vitart V, Burdon KP, Khor CC, Bykhovskaya Y, Mirshahi A, et al. Genome-wide association analyses identify multiple loci associated with central corneal thickness and keratoconus. *Nat Genet.* 2013;45(2):155–63.
- Cornes BK, Khor CC, Nongpiur ME, Xu L, Tay WT, Zheng Y, et al. Identification of four novel variants that influence central corneal thickness in multi-ethnic Asian populations. *Hum Mol Genet.* 2012;21(2):437–45.
- Hoehn R, Zeller T, Verhoeven VJ, Grus F, Adler M, Wolfs RC, et al. Population-based meta-analysis in Caucasians confirms association with COL5A1 and ZNF469 but not COL8A2 with central corneal thickness. *Hum Genet.* 2012;131(11):1783–93.
- Gao X, Gauderman WJ, Liu Y, Marjoram P, Torres M, Haritunians T, et al. A genome-wide association study of central corneal thickness in Latinos. *Invest Ophthalmol Vis Sci.* 2013;54(4):2435–43.
- Vitart V, Bencic G, Hayward C, Skunca Herman J, Huffman J, Campbell S, et al. New loci associated with central cornea thickness include COL5A1, AKAP13 and AVGR8. *Hum Mol Genet.* 2010;19(21):4304–11.
- Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet Med.* 2015;17(5):405–23.
- Warde-Farley D, Donaldson SL, Comes O, Zuberi K, Badrawi R, Chao P, et al. The GeneMANIA prediction server: biological network integration for gene prioritization and predicting gene function. *Nucleic Acids Res.* 2010. <https://doi.org/10.1093/nar/gkq537>.
- Hsia DA, Lim S-T, Bernard-Trifilo JA, Mitra SK, Tanaka S, Den Hertog J, et al. Integrin  $\alpha 4 \beta 1$  promotes focal adhesion kinase-independent cell motility via  $\alpha 4$  cytoplasmic domain-specific activation of c-Src. *Mol Cell Biol.* 2005;25(21):9700–12.
- Dona M, Slijkerman R, Lerner K, Broekman S, Wegner J, Howat T, et al. Usherin defects lead to early-onset retinal dysfunction in zebrafish. *Exp Eye Res.* 2018;173:148–59.
- Dimacali V, Koronis S, Koukoulas S, Rasoglou A, Adamopoulou A, Siskou E, et al. Keratoconus and cone-rod dystrophy among brothers: clinical case study and genetic analysis. *TOOAJ.* 2019. <https://doi.org/10.32474/TOOAJ.2019.02.000133>.

17. Mitra SK, Hanson DA, Schlaepfer DD. Focal adhesion kinase: in command and control of cell motility. *Nat Rev Mol Cell Biol.* 2005;6(1):56–68.
18. Davidson AE, Borasio E, Liskova P, Khan AO, Hassan H, Cheetham ME, et al. Brittle cornea syndrome ZNF469 mutation carrier phenotype and segregation analysis of rare ZNF469 variants in familial keratoconus. *Invest Ophthalmol Vis Sci.* 2015;56(1):578–86.
19. Vincent AL, Jordan CA, Cadzow MJ, Merriman TR, McGhee CN. Mutations in the zinc finger protein gene, ZNF469, contribute to the pathogenesis of keratoconus. *Invest Ophthalmol Vis Sci.* 2014;55(9):5629–35.
20. Lu Y, Dimasi DP, Hysi PG, Hewitt AW, Burdon KP, Toh T, et al. Common genetic variants near the Brittle Cornea Syndrome locus ZNF469 influence the blinding disease risk factor central corneal thickness. *PLoS Genet.* 2010;6(5):e1000947.
21. Abu A, Frydman M, Marek D, Pras E, Nir U, Reznik-Wolf H, et al. Deleterious mutations in the Zinc-Finger 469 gene cause brittle cornea syndrome. *Am J Hum Genet.* 2008;82(5):1217–22.
22. Yu X, Chen B, Zhang X, Shentu X. Identification of seven novel ZNF469 mutations in keratoconus patients in a Han Chinese population. *Mol Vis.* 2017;23:296.
23. Sartaj R, Zhang C, Wan P, Pasha Z, Guaiquil V, Liu A, et al. Characterization of slow cycling corneal limbal epithelial cells identifies putative stem cell markers. *Sci Rep.* 2017;7(1):1–14.
24. Miyashita H, Niwano H, Yoshida S, Hatou S, Inagaki E, Tsubota K, et al. Long-term homeostasis and wound healing in an *in vitro* epithelial stem cell niche model. *Sci Rep.* 2017;7:43557.
25. McLean WI, Irvine AD. Disorders of keratinisation: from rare to common genetic diseases of skin and other epithelial tissues. *Ulster Med J.* 2007;76(2):72.
26. Hirayama M, Ko SB, Kawakita T, Akiyama T, Goparaju SK, Soma A, et al. Identification of transcription factors that promote the differentiation of human pluripotent stem cells into lacrimal gland epithelium-like cells. *NPJ Aging Mech Dis.* 2017;3(1):1–9.
27. Legate KR, Wickström SA, Fässler R. Genetic and cell biological analysis of integrin outside-in signaling. *Genes Dev.* 2009;23(4):397–418.
28. Burkitt Wright EMM, Spencer HL, Daly SB, Manson FDC, Zeef LAH, Urquhart J, et al. Mutations in PRDM5 in brittle cornea syndrome identify a pathway regulating extracellular matrix development and maintenance. *Am J Hum Genet.* 2011;88(6):767–77.
29. Rohrbach M, Spencer HL, Porter LF, Burkitt-Wright EM, Bürer C, Janecke A, et al. ZNF469 frequently mutated in the brittle cornea syndrome (BCS) is a single exon gene possibly regulating the expression of several extracellular matrix components. *Mol Genet Metab.* 2013;109(3):289–95.
30. Morikawa M, Derynck R, Miyazono K. TGF- $\beta$  and the TGF- $\beta$  family: context-dependent roles in cell and tissue physiology. *Cold Spring Harbor Perspect Biol.* 2016;8(5):a021873.
31. Karolak JA, Gambin T, Rydzanicz M, Szaflik JP, Polakowski P, Frajdemberg A, et al. Evidence against ZNF 469 being causative for keratoconus in Polish patients. *Acta Ophthalmol.* 2016;94(3):289–94.
32. Lucas SE, Zhou T, Blackburn NB, Mills RA, Ellis J, Leo P, et al. Rare, potentially pathogenic variants in ZNF469 are not enriched in keratoconus in a large Australian cohort of European descent. *Invest Ophthalmol Vis Sci.* 2017;58(14):6248–56.

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more [biomedcentral.com/submissions](https://biomedcentral.com/submissions)

