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Three isomeric 4-[(n-bromophenyl)carbamoyl]butanoic acids (n = 2, 3, and 4) as DNA intercalator: Synthesis, physiochemical characterization, antimicrobial activity, antioxidant potential and in silico study --Manuscript Draft--

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Dear Editor,

I am submitting a manuscript entitled "Three isomeric 4-[(nbromophenyl)carbamoyl]butanoic acids (n = 2, 3, and 4) as DNA intercalator: Synthesis, physiochemical characterization, antimicrobial activity, antioxidant potential, and *in silico* study" by Bibi Hanifa, Muhammad Sirajuddin, Maciej Kubicki, Edward R. T. Tiekink for possible publication in Journal of Molecular Structure. The art of this paper is in accordance to the standard format of this journal and contains a complete set of informations for the readers working in the fields of chemistry.

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The manuscript has neither been previously published, nor submitted to any other journal. I trust that this manuscript will prove acceptable for publication. Yours sincerely,

M. Sirajuddin

Highlights

- Synthesis of three isomeric 4-[(n-bromophenyl)carbamoyl]butanoic acids
- Structural and spectroscopic characterization
- Interaction with SSDNA via intercalative mode of interaction
- Antioxidant and antimicrobial activities
- In silico study

Three isomeric 4-[(n-bromophenyl)carbamoyl]butanoic acids (n = 2, 3, and 4) as DNA intercalator: Synthesis, physiochemical characterization, antimicrobial activity, antioxidant potential and *in silico* study

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Abstract

A series of three isomeric 4-[(n-bromophenyl)carbamoyl]butanoic acids (n = 2, 3, and 4) were synthesized and their structures confirmed by FTIR, NMR, MS, micro elemental analysis (CHN) and single crystal X-ray crystallography. Kinks are noted in the molecular structures of the n = 2 and n = 3 compounds, namely about the methylene-C–C(carbonyl) and N–C(phenyl) bonds; no such twist about the former bond is seen in the n = 4 molecule. In the molecular packing, supramolecular tapes are evident, being constructed by orthogonally orientated carboxylic acid-O-H…O(carbonyl) and amide-N-H…O(amide) hydrogen bonds. The compounds were evaluated for interaction with salmon sperm DNA and found that they bind *via* an intercalative mode resulting in hypochromism and bathochromic shift as confirmed by the UV-visible spectroscopic and viscometric techniques. The results of antimicrobial activity performed against five bacterial and two fungal strains show that these compounds have < 50% inhibition. The DPPH antioxidant activity results revealed 78% maximum scavenging activity. An *in silico* study performed using the SwissADME webserver revealed the compounds obey the rules of drug-likeness and exhibit good potential for bioavailability.

Keywords: 4-[(n-bromophenyl) carbamoyl]butanoic acids; X-ray crystallography; DNA intercalator; antimicrobial activity; DPPH scavenging; *in silico* study

1. Introduction

Carboxylic acid derivatives of amides play an important role in prodrug design due to their easy enzymatic hydrolysis [1] as well as feature as a valuable part of pharmacophores of diverse classes of therapeutic agents [2] despite the carboxylic functional group having several potential shortcomings such as metabolic instability, toxicity, as well as limited passive diffusion across biological membranes [3]. It is estimated about > 450 drugs contain carboxylic acid functional groups which aremarketed Worldwide, e.g., nonsteroidal anti-inflammatory drugs (NSAIDs) such as naproxen, ibuprofen, indomethacin, cetylsalicylic acid (aspirin), salicylic acid, diclofenac, mefenamic acid, flufenamic acid, etc., antibiotics such as cephalosporin antibiotics (e.g., cephalothin, cephacetrile, cephapirin, cephaloridine, etc.) and penicillin antibiotics (e.g., benzylpenicillin, phenethicillin, methicillin, nafcillin, etc.), quinolone antibiotics (e.g., ciprofloxacin, norfloxacin, acrosoxacin, pipemidic acid, nalidixic acid, etc.), anticoagulants (e.g., heparin), cholesterol-lowering statins (e.g., omega-3 carboxylic acids), among others [3]. The importance of these compounds relates to favorable drug-target interaction owing to their acidity and ability to form a strong electrostatic as well as hydrogen bonding interactions [4, 5].

Due to the flexible binding nature of carboxylic acids, they form various supramolecular structures [6]. As a ligand, the carboxylate moiety plays an important role in the formation of coordination and organometallic compounds, often with desirable properties [7]. The carboxylate moiety can bind to metal in complexation either in ionic or covalent modes. In the later circumstances, coordination may be in monodentate, bridging or bidentate (which may be either *syn-syn* or syn-anti) modes [8].

In the present study, glutaric acid-amide based carboxylic derivatives have been synthesized in order to generate compounds with enhanced solubility and bioavailability. Relatively limited literature is available for this class of compound. Thus, in 1973, Skinner and Sargent reported the syntheses of the *N*-(substituted)glutaramic acids by the condensation of equimolar amounts of the substituted amine and glutaric anhydride at 100°C for 3 h under anhydrous conditions; supporting spectroscopic and crystallographic data of these synthesized compounds were not provided. The original application for these compounds was as plant growth regulators [9]. Herein, an easy and economical method for the synthesis of glutaric anhydride based carboxylic derivatives is described requiring only room temperature conditions and with a maximum of 3-5 min reaction time.

Small organic molecules can bind to DNA either in covalent or non-covalent mode resulting the alteration or inhibition of DNA function. Those molecules which bind to nitrogenous bases of DNA lead to the inhibition of cell division through interference with DNA replication and transcription processes. Another possible mechanism for their anticancer activity is that

proliferation of the cells is halted by affecting the multienzyme complexes with are responsible for the DNA replication and transcription [1, 10-11].

Keeping in view the importance of these compounds as well as in continuation of previous research in this area, in this article are reported the synthesis, characterization and biological evaluation, namely DNA binding, antimicrobial and antioxidant potentials, of three isomeric 4-[(n-bromophenyl)carbamoyl]butanoic acids (n = 2, 3, and 4) compounds. The compositions of the synthesized compounds are confirmed by FTIR, NMR, MS, elemental analysis and X-ray crystallography. An *in silico* study of these compounds relating to their biological potential was also performed.

2. Materials and Methods

The substituted anilines (2-, 3-, and 4-bromoaniline) and glutaric anhydride were purchased from Macklin (Shanghai, China) and solvents were obtained Sigma (Saint Louis, MO, USA) and used without further purification. The melting points were determined on a BioCote melting point apparatus (Staffordshire, UK). Elemental analyses were performed on a PerkinElmer CHNS 2400 instrument (Waltham, MA, USA). The FTIR spectra were measured on a Thermo Nicolet-6700 spectrophotometer (Vienna, Austria) from 4000 to 450 cm⁻¹. The ¹H and ¹³C NMR spectra were recorded in DMSO-d₆ solution on a Bruker Avance 500-MHz NMR (Billerica, MA, USA) spectrometer. The DNA binding experiments were performed using a UH-5300 UV/Vis. spectrophotometer (Hitachi High Tech Science Corporation, Kyoto, Japan) and PSL ASTM Ubbelohde viscometer (Model: 65922, Essex, UK). The GC-MS spectrum was obtained using a Thermo Scientific TRACETM 1310 Gas Chromatograph and Thermo Scientific ISQTM Series Quadrupole GC-MS (Waltham, MA, USA) with conditions: carrier gas: helium; column gas flow: 1.2 mL/s; constant flow; injection mode: splitless injection; column: Agilent HP 5MS, (30 m x 0.25 mm x 25 µm); inlet temp.: 270 °C; oven temperature program: 40 °C (1 min) at 10 °C/min to 300 °C (7 min); transfer line temp.: 300 °C; solvent delay: 2.5 min; ionization energy: 70 eV; ion source temp.: 230 °C; mass range: 35–550 amu, scan rate: 3 scan/s; software: Xcalibur.

2.1 X-ray crystallography

Diffraction data were collected by the ω -scan technique, at room temperature for on a Rigaku SuperNova four-circle diffractometer with Atlas CCD detector, equipped with Nova microfocus

CuK_a radiation source ($\lambda = 1.54184$ Å) for **1** and **3**; for **2**, data were collected on a Rigaku/Oxford Diffraction XtaLAB Synergy diffractometer (Dualflex, AtlasS2) also fitted with CuK_a radiation. The data were processed, including correcting for absorption effects with CrysAlis PRO [12]. The structures were solved with SHELXT (**1** and **3**)/SHELXS (**2**) [13] and refined by the full-matrix least-squares procedure on F² by SHELXL-2018/3 [14]. For **1** and **3**, all non-hydrogen atoms were refined anisotropically, hydrogen atoms were placed in idealized positions and refined as 'riding model' with isotropic displacement parameters set at 1.2 (1.5 for hydroxyl groups) times U_{eq} of appropriate carrier atoms. For **2**, the O- and N-bound atoms were located from a difference map and refined with but, were refined with O–H = 0.82±0.01 and N–H = 0.86±0.01 Å distance restraints, and with U_{iso}(H) set to 1.5U_{equiv}(O) and 1.2U_{equiv}(N), respectively. A weighting scheme of the form $w = 1/[\sigma^2(F_o^2) + (aP)^2 + bP]$ where $P = (F_o^2 + 2F_c^2)/3$ was introduced in each refinement. The programs ORTEP-3 for Windows [15], PLATON [16], and DIAMOND [17] were also used in the study. Crystal data and refinement details are given in Table 1.

Table 1

Crystal data, data collection, and details of structure refinement

Compound	1	2	3
Formula	$C_{11}H_{12}BrNO_3$	$C_{11}H_{12}BrNO_3$	$C_{11}H_{12}BrNO_3$
Formula weight	286.13	286.13	286.13
Crystal system	triclinic	triclinic	monoclinic
Space group	PI	PI	$P2_{1}/c$
<i>a</i> (Å)	4.8317(2)	4.9260(2)	24.6256(2)
<i>b</i> (Å)	10.2446(6)	8.3664(5)	4.80980(5)
<i>c</i> (Å)	12.0871(7)	14.6791(6)	9.79495(13)
α (°)	93.050(5)	98.247(4)	90
β (°)	98.725(4)	96.176(3)	97.4563(10)
γ (°)	99.408(4)	99.697(4)	90
$V(Å^3)$	581.55(5)	584.75(5)	1150.34(2)
Ζ	2	2	4
$D_{\rm x}$ (g cm ⁻³)	1.634	1.625	1.652
<i>F</i> (000)	288	288	576
μ (mm ⁻¹)	4.774	4.748	4.827
Reflections:			
collected	4094	12804	17562
unique (R _{int})	2256 (0.032)	2078 (0.040)	2383 (0.033)
with $I > 2\sigma(I)$	2101	1918	2289
$R(F) [I > 2\sigma(I)]$	0.042	0.041	0.033
$wR(F^2) [I > 2\sigma(I)]$	0.116	0.107	0.094
a, b in weighting scheme	0.078, 0.162	0.047, 0.659	0.052, 0.577
R(F) [all data]	0.045	0.044	0.034
$wR(F^2)$ [all data]	0.118	0.110	0.095
Goodness of fit	1.07	1.06	1.06
max/min $\Delta \rho$ (e·Å ⁻³)	0.37/-0.79	0.92/-0.93	0.49/-0.52
CCDC number	2111806	2115862	2111807

2.2 Synthesis

Glutaric anhydride (1.141 g, 10 mmol) dissolved in toluene (25 mL) was reacted with the respective n-bromoaniline (1.720 g, 10 mmol) also taken in toluene (25 mL) at room temperature. After the mixing two reactants, each precipitate appeared after a few minutes mixing, as depicted in Scheme 1. The precipitate was washed with toluene to remove any unreacted reactants and then

with distilled H_2O to remove any glutaric acid. The product was then air dried to get the desired compound, **1-3** [18-23]. Recrystallization of each compound was from its acetone/ethanol (1:1 v/v) solution.



Scheme 1: Schematic representation of the reactions to give 1-3 along with atom numbering for NMR data interpretation

2.2.1 4-[(2-bromophenyl) carbamoyl]butanoic acid (1)

Melting point: 138-140 °C. Color: Red. Molecular formula: $C_{11}H_{12}BrNO_3$. Molecular Weight: 286.13. **CHN** data in % (calculated/found): 46.18/46.36 (C); 4.23/4.16 (H); 4.90/4.87 (N). **FTIR** data (v cm⁻¹): 3275 (NH stretching); 2888 (H-bonded OH of the carboxylic moiety); 1689 (COO stretching); 1649 (amide carbonyl); 1578 (C=C); 1461 (OH bending); 1437 (COO bending) 1261 (NH bending); 1028 (C-N stretching). ¹**HNMR** (DMSO-d₆, 500 MHz) with chemical shift in ppm: 11.00, s (OH of carboxylic moiety); 9.45, s (NH of amide moiety); H-2, t (2.39, ³*J*_{H-H} = 7.0 Hz); H-3, quint. (1.83, ³*J*_{H-H} = 7.3 Hz); H-4, t (2.30, ³*J*_{H-H} = 7.3 Hz); H-8, d (7.56, ³*J*_{H-H} = 8.0 Hz); H-9, t (7.15, ³*J*_{H-H} = 7.3 Hz); H-10, t (7.36, ³*J*_{H-H} = 7.5 Hz); H-11, d (7.64, ³*J*_{H-H} = 8.0 Hz). ¹³CNMR (DMSO-d₆, 125 MHz) with chemical shift in ppm: 174.6 (C-1); 35.3 (C-2); 21.0 (C-3); 33.5 (C-2); 2

4); 171.4 (C-5); 136.8 (C-6); 118.8 (C-7); 133.1 (C-8); 128.0 (C-9); 128.4 (C-10); 127.5 (C-11). MS: [M]⁺: 285 m/e (⁷⁹Br); base peak ([C₆H₆NBr]⁺): 171 m/e (⁷⁹Br).

2.2.2 4-[(3-bromophenyl) carbamoyl]butanoic acid (2)

Melting point: 130-132 °C. Color: Red-brown. Molecular formula: $C_{11}H_{12}BrNO_3$. Molecular Weight: 286.13. **CHN** data in % (calculated/found): 46.18/46.20 (C); 4.23/4.20 (H); 4.90/4.92 (N). **FTIR** data (v cm⁻¹): 3295 (NH stretching); 2882 (H-bonded OH of the carboxylic moiety); 1688 (COO stretching); 1656 (amide carbonyl); 1583 (C=C); 1476 (OH bending); 1438 (COO bending) 1260 (NH bending); 1066 (C-N stretching). ¹**HNMR** (DMSO-d₆, 500 MHz) with chemical shift in ppm: 12.07, s (OH of carboxylic moiety); 10.06, s (NH of amide moiety); H-2, t (2.36, ³*J*_{H-H} = 7.0 Hz); H-3, quint. (1.81, ³*J*_{H-H} = 7.0 Hz); H-4, t (2.26, ³*J*_{H-H} = 7.5 Hz); H-7, s (7.96); H-9, d (7.47, ³*J*_{H-H} = 7.0 Hz); H-10, t (7.25, ³*J*_{H-H} = 7.5 Hz); H-11, d (7.21, ³*J*_{H-H} = 7.0 Hz). ¹³**CNMR** (DMSO-d₆, 125 MHz) with chemical shift in ppm: 174.6 (C-1); 35.7 (C-2); 20.8 (C-3); 33.4 (C-4); 171.6 (C-5); 141.3 (C-6); 121.8 (C-7); 122.0 (C-8); 126.0 (C-9); 131.1 (C-10); 118.2 (C-11). MS: [M]⁺: 285 m/e (⁷⁹Br); base peak ([C₆H₆NBr]⁺): 171 m/e (⁷⁹Br).

2.2.3 4-[(3-bromophenyl) carbamoyl]butanoic acid (3)

Melting point: 160-162 °C. Color: Red-brown. Molecular formula: $C_{11}H_{12}BrNO_3$. Molecular Weight: 286.13. **CHN** data in % (calculated/found): 46.18/46.95 (C); 4.23/4.10 (H); 4.90/5.01 (N). **FTIR** data (v cm⁻¹): 3307 (NH stretching); 2894 (H-bonded OH of the carboxylic moiety); 1689 (COO stretching); 1662 (amide carbonyl); 1586 (C=C); 1485 (OH bending); 1436 (COO bending); 1271 (NH bending); 1010 (C-N stretching). ¹**HNMR** (DMSO-d₆, 500 MHz) with chemical shift in ppm: 12.08, s (OH of carboxylic moiety); 10.03, s (NH of amide moiety); H-2, t (2.35, ³*J*_{H-H} = 7.5 Hz); H-3, quint. (1.80, ³*J*_{H-H} = 7.5 Hz); H-4, t (2.27, ³*J*_{H-H} = 7.3 Hz); H-7,7', d (7.56, ³*J*_{H-H} = 9.0 Hz); H-8,8', d (7.46, ³*J*_{H-H} = 8.0 Hz). ¹³**CNMR** (DMSO-d₆, 125 MHz) with chemical shift in ppm: 174.6 (C-1); 35.8 (C-2); 20.8 (C-3); 33.4 (C-4); 171.4 (C-5); 139.0 (C-6); 121.4 (C-7,7'); 131.9 (C-8,8'); 114.9 (C-9). MS: [M]⁺: 285 m/e (⁷⁹Br); base peak ([C₆H₆NBr]⁺): 171 m/e (⁷⁹Br).

2.3 Antimicrobial screening

The antibacterial data were determined against five bacterial strains, as specified in Table 2. All bacteria were cultured in Cation-adjusted Mueller Hinton broth (CAMHB) at 37 °C overnight. A

sample of each culture was then diluted 40-fold in fresh broth and incubated at 37 °C for 1.5-3 h. The resultant mid-log phase cultures were diluted then added to each well of the compound containing plates, giving a cell density of 5×10^5 CFU/mL. Plates were covered and incubated at 37 °C for 18 h without shaking. Inhibition of bacterial growth was determined measuring absorbance at 600 nm (OD600), using a Tecan M1000 Pro monochromator plate reader (Equipped with premium quad4 monochromators). The percentage of growth inhibition was calculated for each well, using the negative control (media only) and positive control (bacteria without inhibitors) on the same plate as references. For the screening, the significance of the inhibition values was determined by modified Z-scores, calculated using the median and mean absolute deviation (MAD) of the samples (no controls) on the same plate. Samples with inhibition value above 80% and Z-Score > 2.5 for either replicate (n=2 on different plates) were classed as actives while sample with inhibition below 50.9% - 79.9% and Z-Score < 2.5 were classed as inactive [24].

The antifungal data was collected against two fungal strains as detailed in Table 2. Fungi strains were cultured for 3 days on Yeast Extract-Peptone Dextrose (YPD) agar at 30 °C. A yeast suspension of 1 x 10^6 to 5 x 10^6 CFU/mL (as determined by OD530) was prepared from five colonies. The suspension was subsequently diluted and added to each well of the compoundcontaining plates giving a final cell density of fungi suspension of 2.5 $\times 10^3$ CFU/mL and a total volume of 50 µL. All plates were covered and incubated at 35 °C for 24 h without shaking. Growth inhibition of C. albicans was determined measuring absorbance at 530 nm (OD530), while the growth inhibition of C. neoformans was determined measuring the difference in absorbance between 600 and 570 nm (OD600-570), after the addition of resazurin (0.001% final concentration) and incubation at 35 °C for additional 2 h. The absorbance was measured using a Biotek Synergy HTX plate reader (Light source: Xenon flash lamp Detector: Photodiode Wavelength selection: Monochromator Wavelength range: 200 - 999 nm, 1 nm increments Monochromator bandwidth: 2.4 nm Dynamic range: 0 - 4.0 OD Resolution: 0.0001 OD Pathlength correction: Yes Monochromator wavelength accuracy: ±2 nm Monochromator wavelength repeatability: ±0.2 nm OD linearity). The percentage of growth inhibition was calculated for each well, using the negative control (media only) and positive control (bacteria without inhibitors) on the same plate as references [24].

Colistin and Vancomycin were used as positive bacterial inhibitor standards for Gram-negative and Gram-positive bacteria, respectively. Fluconazole was used as a positive fungal inhibitor standard

for *C. albicans and C. neoformans*. The antibiotics were provided in 4 concentrations, with 2 above and 2 below its MIC value, and plated into the first 8 wells of column 23 of the 384-well NBS plates.

Table 2

Information of the bacterial and fungal strain and standards investigated in the present study

Abbr.	Code	Name	Descript	tion	Stra	in	Organism	Туре
Sa	GP_020	Staphylococcus	MRSA		ATCC -	43300	Bacteria	G+ve
		aureus						
Ec	GN_001	Escherichia coli	FDA cor	ntrol	ATCC	25922	Bacteria	G-ve
Кр	GN_003	Klebsiella	MDR	2	ATC	CC	Bacteria	G-ve
		pneumoniae			7006	503		
Ab	GN_034	Acinetobacter	Type str	ain	ATCC	19606	Bacteria	G-ve
		baumannii						
Pa	GN_042	Pseudomonas	Type str	ain	ATCC 2	27853	Bacteria	G-ve
		aeruginosa						
Ca	FG_001	Candida albicans	CLSI refe	rence	ATCC	90028	Fungi	Yeast
Cn	FG_002	Cryptococcus	Type str	ain	H99; A	TCC	Fungi	Yeast
		neoformans var.			2088	821		
		grubii						
Standards								
Samp	ole name	Sample ID	Full MW	Stocl	c Conc.	Solve	ent Sou	rce
				(mg	g/mL)			
Colisti	n-Sulfate	MCC_000094:02	1400.63	1	0.0	DMS	O Sigma;	C4461
Vancon	nycin-HCL	MCC_000095:02	485.71	1	0.0	DMS	O Sigma;	861987
Fluc	onazole	MCC_008383:01	306.27	2	2.56	DMS	O Sigma;	F8929

2.4 DNA binding assay by UV-visible spectroscopy and viscometry

A solution of Salmon sperm DNA was prepared in distilled water by dissolving 2 mg of the sodium salts of SS-DNA and stirred at room temperature overnight. The concentration of the solution was determined on a UH-5300 UV/Vis. spectrophotometer using based on $\varepsilon = 6600 \text{ M}^{-1} \text{ cm}^{-1}$ and found

to be 1.4 x 10^{-4} M. The nature of DNA free from protein was checked from its absorbance ratio $A_{260}/A_{280} = 1.8$. A solution of each of **1-3** (1 mM) was prepared in 70% absolute EtOH. During the DNA binding study, the concentration of **1-3** was kept constant while that of the DNA was changed [25-27].

The viscosity was measured at a temperature of 29 ± 1 °C using an Ubbelohde viscometer. A digital timer was used to measure the flow time. The average flow time was determined after measuring each sample three times. The results were shown as a plot of relative viscosity $[(\eta/\eta_o)^{1/3}]$, vs. binding ratio [(compound)/(DNA)], where η indicates the viscosity of DNA in the presence of **1-3** and η_o the viscosity of DNA only. Viscosity results were determined from the experimental flow rate of DNA containing solution, $\eta = t - t_o$ [28-30].

2.5 DPPH scavenging activity

DPPH (3.94 mg) was dissolved in MeOH (100 mL). The DPPH and solutions of **1-3** were prepared as follows: to methanolic solutions of DPPH (2800 μ L) was added 0.2 μ L of **1-3** (also prepared in methanol) with concentrations of **1-3** ranging from 10 to 150 mg/mL. The decrease in DPPH absorbance was noted at 517 nm after 10 min *via* UV–visible spectrophotometer (provide details). The same protocol was followed for well-known antioxidant, ascorbic acid, as a control. All the measurements were performed in triplicate and average results reported. The percent scavenging activity of screened compounds was measured as per [31-33]:

% Scavenging activity =
$$\frac{A_o - A_s}{A_o}$$

 A_o and A_s represent the DPPH absorbance in the absence (control) and presence of sample (1-3), respectively.

2.6 Drug-likeness and ADME studies

ADME properties such as physicochemical and pharmacokinetics of the synthesized compounds and drug similarity properties were performed on the SwissADME webserver [34, 35]. These studies provide properties such as TPSA, molecular weight, hydrogen bond acceptor/donor atoms and lipophilicity (logP). A BOILED-Egg model of the compounds, the absorption in the gastrointestinal system and the crossing of the blood-brain barrier were also evaluated [36]. The oral bioavailability of the compounds was also examined on six parameters employing a radar image. Drug-likeness analysis of the compounds, "Lipinski's Rule of Five" compatibility was examined using the same SwissADME webserver [34, 35].

Results and Discussion

Three new isomeric 4-[(n-bromophenyl)carbamoyl]butanoic acids (n = 2, 3, and 4) compounds, 1-3, have been synthesized from the 1:1 (nucleophilic addition) reaction of glutaric anhydride with respective n-bromoaniline. The lone pair of electrons on the N atom of the n-bromoaniline attack on the electrophilic carbonyl carbon of the glutaric anhydride, followed by the abstraction of H⁺ from the electropositive N atom by the negatively charged oxygen atom resulting in the formation of the desired 4-[(n-bromophenyl)carbamoyl]butanoic acids [37, 38]. The compounds are air stable, soluble in common organic solvents including MeOH, EtOH, DMSO, acetone and chloroform. Details of the spectroscopic and crystallographic characterization are given below. In addition, certain medicinal potential, namely DNA binding, antimicrobial, antioxidant potentials, and the theoretical determination of medicinally relevant attributes are reported.

3.1 FTIR

The FTIR spectra of **1-3** were recorded in the range 4000–450 cm⁻¹. A broad peak of weak intensity centered at 2888, 2882, 2894 cm⁻¹ for **1-3**, respectively, is attributed to the hydrogen bonded O–H stretching vibrations of the carboxylic acid. The strong N–H stretching appeared at 3275, 3295, and 3307 cm⁻¹ for **1-3**, respectively, while the N–H bending vibration were observed at 1261, 1260, and 1271 cm⁻¹, respectively. A peak at 1649, 1656, and 1662 cm⁻¹ was assigned to the amide carbonyl stretching in **1-3**. A medium intensity peak at 1028, 1066, and 1010 cm⁻¹ is attributed to the C–N stretching vibration, while the asymmetric COO stretching band appeared as a strong peak at 1689, 1688, and 1689 cm⁻¹ in **1-3** [19, 20].

3.2 NMR

The ¹H NMR spectra of **1-3** displayed a singlet resonance at 12.07, 11.00, and 12.08 ppm, respectively, which is assigned to the carboxylic acid OH confirming the presence of the carboxylic acid residue in each case. Similarly, another characteristic resonance is ascribed to the NH proton i.e., a singlet in the range 9.45-10.03 ppm. Additional resonances, multiplicity and integration, are as expected, see **2.2.1** – **2.2.3**.

The most notable features in the ¹³C NMR spectra of the compounds **1-3** were downfield resonances at δ 174.6 ppm (**1-3**) and at δ 171.4, 171.6, and 171.4 ppm, which are assigned to the carboxylic acid (C-1) and amide (C-4) nuclei. The resonances of the remaining nuclei fall in their anticipated regions [19, 20], as detailed in **2.2.1** – **2.2.3**.

3.3 X-ray crystallography

The crystal and molecular structures of isomeric **1-3** have been established by X-ray crystallography. The molecular structures are illustrated in Figure 1 and selected geometric parameters are collated in Table 3. The three molecules differ in the position of the bromide substituent in the terminal phenyl ring, and bear a close resemblance to each other so the discussion will focus on the 2-bromo derivative, **1**. Evidence for the carboxylic acid assignment is readily gained through the large difference between the C–O1, O2 bond lengths, Table 3, and in the pattern of supramolecular association between molecules (see below); the angles subtended at the C1 atom involving the O2 atom are wider, again consistent with the presence of a C1=O2 bond. The bond lengths in the amide functionality, which adopts an anti-conformation, are as expected and the angles at C5 involving the carbonyl-O5 atom are systematically wider. Based on the torsion angle data included in Table 3, there is a kink in the chain linking the carboxylic acid and amide residues. The twist occurs about the C4–C5 bond with the torsion angle (-145.3(3)°) indicative of a -anticlinal (-ac) conformation. A second twist in the molecule occurs about the C6–N1 bond, consistent with a +anti-clinal (-ac) conformation.



Fig. 1. The molecular structures of (a) **1**, (b) **2**, and (c) **3**, showing the atom-labelling scheme and displacement ellipsoids at the 35% probability level.

Table 3

Selected geometric parameters (Å, °) for 1-3

Parameter	1	2	3
C101	1.311(4)	1.307(4)	1.302(2)
C1–O2	1.213(4)	1.211(4)	1.232(2)
C5–O3	1.218(4)	1.221(3)	1.218(2)
C5–N1	1.340(4)	1.343(4)	1.350(2)
C6N1	1.417(4)	1.416(4)	1.422(2)
C5-N1-C6	122.8(2)	125.3(2)	124.68(16)
O1–C1–O2	123.5(3)	123.0(3)	123.49(17)
O1C1C2	113.0(3)	113.2(3)	113.61(17)
O2C1C2	123.5(3)	123.8(3)	122.87(18)
O3-C5-N1	122.9(3)	122.8(3)	122.68(18)
O3–C5–C4	120.7(3)	121.5(3)	123.25(18)
N1-C5-C4	116.3(2)	115.8(2)	114.06(16)
C1C2C3C4	-177.7(3)	-175.6(3)	-171.5(2)
C2-C3-C4-C5	-176.6(3)	176.2(3)	-173.3(2)
C3-C4-C5-N1	-145.3(3)	-141.1(3)	-171.4(2)
C4-C5-N1-C6	177.1(3)	-177.9(3)	178.7(2)
C5-C6-N1-C7	127.5(3)	135.5(3)	-135.0(2)
CO_2/C_6	14.3(3)	8.0(3)	75.83(11)

The molecular structure of **2** mimics that just described for **1**, an observation entirely consistent with the isostructural relationship between the crystals, Table 1. While the bond lengths and angles in **3** match those of the previous isomers, a difference in the chain is noted with the C3–C4–C5–N1 torsion angle $(-171.4(2)^{\circ})$ indicative of a -anti-periplanar (-ap) conformation. In addition to the difference in the conformation of the side-chains, the dihedral angle between the terminal carboxylic acid and phenyl residues is close to perpendicular in **3** in contrast the those in **1** and **2**. The conformational differences between the three molecules are highlighted in Figure 2. The conformational flexibility of related molecules has already been noted in the literature [22, 23].



Fig. 2. An overlay diagram of **1** (red image), **2** (green image), and **3** (blue image), highlighting the similarity in the conformations of **1** and **2**, and the difference between these and that of **3**. The molecules have been overlapped so the amide residues are coincident.

As anticipated from the chemical composition of 1-3, conventional hydrogen bonding interactions are prominent in the supramolecular association in their crystals. Indeed, all three crystals feature comparable hydrogen bonding patterns, namely the self-association of the carboxylic acid resides about a center of inversion to generate eight-membered $\{...OCOH\}_2$ synthons, and the subsequent linking of the dimeric aggregates into tapes via amide-N-H...O(amide) hydrogen bonds; geometric data characterizing the specified contacts are included in Table 3. A representative tape is shown in Figure 3a for 1. Also shown is a side-on view which indicates a significant deviation from planarity in the tape. A similar situation pertains for isostructural 2, Figure 3b, in contrast to the near to planar tape noted in the crystal of 3, Figure 3c.

Table 4

Hydrogen bonding (A-H...B) interactions in the crystals of 1-3

А	Н	В	HB	AB	A–H…B	Symmetry
			(Å)	(Å)	(°)	operation
1						
01	H1o	O2	1.86	2.679(3)	174	1-x, 2-y, 2-z
N1	H1n	03	2.15	2.983(4)	164	1+x, y, z
2						
01	H1o	O2	1.86(5)	2.680(4)	178(7)	2-x, 1-y, 2-z
N1	H1n	03	2.119(18)	2.963(3)	169(3)	-1+x, y, z
3						
01	H1o	O2	1.84	2.653(2)	172	1-x, 1-y, 2-z
N1	H1n	O3	2.20	3.028(2)	160	x, 1+y, z

Based on an assessment of the molecular packing employing PLATON [17], the tapes in the crystal of **1** assemble without directional interactions between them. There is evidence of a close Br1...Br1ⁱ contact in the crystal of **2** with the separation being 3.6097(7) Å, close to the sum of the van der Waals radii of 3.70 Å [17]; symmetry operation (i): -x, -y, -z. This interaction has the character of a type-I halogen bonding contact which often equates to a van der Waals contact arising a result of global molecular packing considerations rather than being a directional interaction. In continuation of the theme of a lack of directional interactions between the supramolecular tapes in **1-3**, Br...Br contacts are evident in the crystal of **3** but, with the Br1...Br1ⁱⁱ separations are longer than the sum of the van der Waals radii, with the separation being 3.7552(6) Å; symmetry operation (ii): 2-x, - $\frac{1}{2}$ +y, $\frac{1}{2}$ -z.



3.4 Mass spectrometry

Mass spectral data for isomeric **1-3** were obtained by electron impact (EI) at 70 eV. As anticipated from the isomeric composition, the general mass fragmentation patterns are similar and are summarized in Scheme 2, with a representative mass spectrum for **3** shown in Figure 4. The molecular ion peak $[M]^+$ with isotopic ⁷⁹Br was noted at m/z = 285, and the molecular ion peak loses the COOH and CO(CH₂)₃ units to give the baseline peak at m/z =171.



Scheme 2. General mass fragmentation pattern for compounds 1-3



Fig. 4. Mass spectrum for 3.

3.5 Antimicrobial activity

Compounds 1-3, being soluble in DMSO, were submitted to CO-ADD (the Community for Open Antimicrobial Drug Discovery). The compounds undergo a primary screen in duplicate at a single concentration ($32 \mu g/mL$) in 384-well format to test their killing ability against broth solutions of key ESKAPE bacterial pathogens: *S. aureus* (MRSA), *E. coli, K. pneumoniae, A. baumannii, P. aeruginosa*, and as well as fungal pathogens *C. neoformans* and *C. albicans*; see Table 2 for details. It can be seen from Table 5 that the maximum antibacterial activity exhibited by 1 and 2 is against *P. aeruginosa* while that of 3 shows is against *K. pneumoniae*. In terms of antifungal activity, all compounds were most active against the *C. neoformans* compared to *C. albicans*. The Z-score of the screened compounds against both bacterial and fungal strains was below [2.5]. Since the antimicrobial inhibition exhibited by 1-3 is lower than 50%, they are considered inactive. Accordingly, none of the compounds was selected for further dose response studies or Hitconfirmation.

Table 5

Compd.				% Inhibitio)n		
	Antibacterial						ifungal
	S.	Е.	К.	Р.	А.	С.	С.
	aureus	coli	pneumoniae	aeruginosa	baumannii	albicans	neoformans
	(MRSA)						
1	-1.63	5.02	12.01	16.77	5.61	7.45	-20.86
2	0.16	2.41	10.89	13.32	11.44	5.25	-18.95
3	2.04	7.36	14.89	9.67	12.85	4.14	-13.51

Percentage inhibition exhibited by 1-3 against selected bacteria and fungi^a

a Active compound: Inhibition \geq 80% and abs(Z-Score) > |2.5|. Partial Active: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inactive Compound: Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inhibition = 50.9 - 79.9% and abs(Z-Score) < |2.5|. Inhibition = 50.9\% abs(Z-Score) < |2.5|. Inhibition = 50.9\% abs(Z-Score) < |2.5|. Inhibition = 50.9\% abs(Z-Score) < |2.5|. I

compounds: Inhibition < 50% and abs(Z-Score) < |2.5|

3.6 DNA interaction

The interactions between 1-3 and DNA were studies via UV-visible spectroscopic with the results summarized in Figures 5-7, respectively. Each compound exhibited two strong bands at approximately 200-230 and 235-300 nm ascribed to π - π * and n- π * transitions, respectively. Upon

interaction with DNA, two phenomena were observed: hypochromism along with a bathochromic effect of 2-5 nm. After intercalating the base pairs of DNA, the π^* orbital of the intercalated molecule couples with the π orbital of the base pairs, thereby decreasing the π - π^* transition energy, resulting in the bathochromic shift. Further, the coupling of the respective, partially filled π orbital decreases the transition probabilities resulting in the observed hypochromic shift. When these two phenomena occur then the dominant mode of interaction is the intercalation, which involves strong stacking between the chromophore and the base pairs of DNA [31-33]. The binding constant (K_b) was determined form the intercept to slope ratio of the plot of A_o/(A-A_o) vs. 1/[DNA] as shown in the inset of Figures 5-7. The K_b value was then used to determine the Gibb's free energy value (which relates to the spontaneity of DNA-compound adduct formation) from the equation $\Delta G = -RTlnK_b$, where the negative sign of ΔG indicate the spontaneity of the DNA-compound adduct formation [31-33].

The intercalative binding mode of between **1-3** and DNA was further confirmed by the viscometric method. During the viscosity measurements, the viscosity of the DNA in the presence of various concentration of **1-3** was increased due to the entrance of the respective compound between the DNA bases resulting in the lengthening of DNA. The increase in the viscosity of DNA upon the addition of various concentration is the sign of the intercalative binding mode [34]. The plot of relative viscosity *vs*. ratio of the concentration of **1-3**/DNA is shown in Figure 8.



Fig. 5. DNA binding spectra of **1** (1 mM), in the absence and presence of various DNA concentrations. The arrowhead represents the increasing concentration of DNA.



б

Fig. 6. DNA binding spectra of **2** (1 mM), in the absence and presence of various DNA concentrations. The arrowhead represents the increasing concentration of DNA.



Fig. 7. DNA binding spectra of **3** (1 mM), in the absence and presence of various DNA concentrations. The arrowhead represents the increasing concentration of DNA.



Fig. 8. Viscosity plot for the interaction of 1-3 with DNA.

3.7 Antioxidant activity

DPPH is the most widely used radical to determine the reducing or antioxidant potential of a compound. The antioxidant potential of the compounds 1-3 was evaluated form their interaction with DPPH. Compounds having antioxidant potential must reduce the free 1,1-diphenyl-2- picryl-hydrazyl (DPPH) radical into 1,1-diphenyl-2-picryl-hydrazine by giving an electron to DPPH radial. The DPPH radical has a strong absorption peak at 517 nm with a deep violet color. Upon the addition of various concentration of the compound, the radical nature of the DPPH changes as its unpaired electron gets paired and as a result the decrease in absorbance as well as decolorization occurs [28, 29]. The effect of the various concentration of compounds 1-3 as well as that of ascorbic acid (AA) used as a standard is shown in Figure 9. The activity of all the three isomeric compounds is almost similar and the maximum activity of compounds 1-3 at 1800 μ L/mL concentration are 72, 75, 78%, respectively. The activity of the ascorbic acid at 1800 μ L/mL concentration is 99%.

However, the activity of the compounds 1-3 is lower as compared to ascorbic acid but they possess the antioxidant potential.



Fig. 9. DPPH scavenging activity of 1-3 using AA (ascorbic acid) as standard.

3.8 Physicochemical properties and ADME parameters

The physicochemical properties, ADME parameters and the violations of drug-likeness rules of the synthesized compounds 1-3 were listed in Tables 7 and 8. The evaluated physicochemical properties are: the molecular weight, topological polar surface area (tPSA), Molar Refractivity, fraction of sp³ carbon atoms (Fsp³) and some Hydrogen Bond properties. tPSA is the sum of surface areas of polar atoms in a molecule and is used to estimate drug transport properties. Low tPSA values in molecules correspond to a higher propensity for transport and tPSA values obtained for the three isomeric compounds 1-3 is 66.40 Å² which is within the range of values recommended by various drug-likeness filters. Fsp³ is a newer parameter [40] used to evaluate drug-likeness properties of molecules and its values for the three isomeric compounds 1-3 is 0.27. Molar Refractivity is the overall polarity of a molecule and is expected to be in the range from 40 to 130. Molar Refractivity value for the three isomeric compounds 1-3 is 64.65.

Lipophilicity is a valuable parameter that affects drug activity in the human body. LogP values are the most widely used measure of lipophilicity and represents an indicator of drugs permeability to reach the target tissue in the body. The LogP values used by the different drug-likeness filters (MLogP for Lipinski filter [41], WLogP for Ghose [42] and Egan filters [43], XLogP for Muegge filter [44]) and their mean values (consensus LogP) were shown in the Table 8. All other LogP values for the compounds 1-3 obeys general standards (< 5).

ESOL is aqueous solubility parameter of molecules proposed by Delenay [45] and is considered one of the key physical properties in drug discovery. ESOL values for the compounds 1-3 belongs to soluble class as shown in Table 7.

There are a lot of filter approach in the literature that suggest a set of rules to evaluate druglikeness profiles of molecules. The filters discussed in this paper and their rules are as follows.

• Lipinski (Pfizer) filter [41]: $MW \le 500$; $MLogP \le 4.15$; $HBA \le 10$; $HBD \le 5$

- Ghose filter [42]: $160 \le MW \le 480$; $-0.4 \le WLogP \le 5.6$; $40 \le MR \le 130$; $20 \le atoms \le 70$
- Veber (GSK) filter [46]: $RB \le 10$; $tPSA \le 140$
- Egan (Pharmacia) filter [43]: $WLogP \le 5.88$; $tPSA \le 131.6$
- Muegge (Bayer) filter [44]: $200 \le MW \le 600$, $-2 \le XLogP \le 5$; tPSA ≤ 157 ; HBA ≤ 10 ; HBD \le

5; $RB \le 15$; No. of rings ≤ 7 ; No. of carbons > 4; No. of heteroatoms > 1

The filters generally state that an orally active drug should not violate the above criteria more than once. According to Table 8, it is observed that the compounds **1-3** obey all the filters and has zero violation.

Bioavailability score estimate the probability of a compound to have oral bioavailability in rat or measurable Caco-2 permeability and the bioavailability score value of a compound in the rat is expected to be > 0.1068. A poor bioavailability results in lower activity of the molecule and higher inter-individual variability, and thus causes an unexpected response of a drug [47]. The bioavailability score value for the three isomeric compounds is 0.85.

Log K_p is skin permeation parameter suggested by Potts *et al.* [48]; high negative Log K_p value of the molecule indicates that the molecule has less penetration into the skin. The value of Log K_p value for compounds 1-3 is -7.24 cm/s, -6.55cm/s and -6.92 cm/s, respectively.

In summary, Tables 7 and 8 show the physicochemical properties, lipophilicity and water solubility values of the compounds 1-3 used by various drug filters and have zero violation. Moreover, the favorable bioavailability scores and the higher skin absorption indicate that these

compounds can be potential drug candidates. The three isomeric compounds **1-3** are Nonmutagenic, non-tumorigenic, non-irritant and have no reproductive effect.

The radar image and the BOILED-Egg model of the synthesized compounds are shown in Figure 10. The obtained radar image identifies substances that can be considered drug-like in the pink area through 6 different physicochemical parameters. These parameters are defined as lipophilicity (LIPO), molecular size (SIZE), polarity (POLAR), solubility (INSOLU), flexibility (FLEX) and saturation (INSATU). The radar image of the three isomeric compounds completely falls in the pink area in accordance with 5 different parameters. In the BOILED-Egg model, the yellow area represents the crossing of the blood-brain barrier, and the white area represents absorption in the gastrointestinal system. This model is a distribution graph of commercial drugs by defining the X-axis as the TPSA value of the molecule and the WLOGP value of the Y-axis. The red dot represents the molecule selected, and inferences about pharmacokinetic properties are made according to the status of this point in the yellow, white and grey area. As seen in Figure 5, it was predicted that the compounds 1-3 can be a candidate for such drug molecules with its absorption status in the blood-brain barrier. On the other hand, there are many factors affecting the passage of a drug candidate molecule into the lymph and blood circulation. Some of these factors are the size of the molecule, molecular weight, hydrophilic and lipophilic structure.

Table 7

Property	1	2	3					
Physicochemical Properties								
M. Weight (g/mol)	286.12	286.12	286.12					
Fraction Csp ³	0.27	0.27	0.27					
No. rotatable bonds	6	6	6					
No. H-bond acceptors/donor	3/2	3/2	3/2					
Molar Refractivity	64.65	64.65	64.65					
Total polar surface area (Å ²)	66.40	66.40	66.40					
Lipophilicity								
Log <i>P</i> _{o/w} (iLOGP)	1.83	1.84	1.73					
$\text{Log } P_{\text{o/w}} (\text{XLOGP})$	1.13	2.10	1.58					
Log P _{o/w} (WLOGP)	2.45	2.45	2.45					
Log P _{o/w} (MLOGP)	2.12	2.12 2.12						
$\log P_{o/w}$ (SILICOS-IT)	2.09	2.09	2.09					
Consensus Log P _{o/w}	1.92	2.12	2.00					
	Water Solubilit	y						
Log S (ESOL)	-2.21	-2.82	-2.49					
Solubility (mg/mL), Class	1.77, Soluble	4.35e-01, Soluble	9.24e-01, Soluble					
Log S (Ali)	-2.12	-3.12	-2.59					
Solubility (mg/mL), Class	2.18, Soluble	2.15e-01, Soluble	7.44e-01, Soluble					
Log S (SILICOS-IT)	-3.83	-3.83	-3.83					
Solubility (mg/mL), Class	4.20e-02, Soluble	4.20e-02, Soluble	4.20e-02, Soluble					

Physicochemical properties, lipophilicity, and solubility for 1-3

Table 8

D			-1	- · · · · · · · ·		1 f 1 7
Imuglikeness	nnarmacokinetics	medicinal	cnemistry	and to	v101tv r1s	KC OT I-1
Diaginconcos,	pharmaconneuros,	moulomai	chemistry	and to	AICILY IIG	
0 /	1				2	

Property	1	2	3
	Druglikeness		
Lipinski	Yes; 0 violation	Yes; 0 violation	Yes; 0 violation
Ghose	Yes	Yes	Yes
Veber	Yes	Yes	Yes
Egan	Yes	Yes	Yes
Muegge	Yes	Yes	Yes
Bioavailability Score	0.85	0.85	0.85
Drug Score	0.44	0.44	0.26
	Pharmacokinetic	S	
Gatrointestinal absorption	High	High	High
BBB permeant	Yes	Yes	Yes
P-glycoprotein substrate	No	No	No
Cytochrome P450 1A2 inhibitor	No	Yes	No
Cytochrome P450 2C19 inhibitor	No	No	No
Cytochrome P450 2C9 inhibitor	No	No	No
Cytochrome P450 2D6 inhibitor	No	No	No
Cytochrome P450 3A4 inhibitor	No	No	No
Log K _p (skin permeation)	-7.24 cm/s	-6.55 cm	/s -6.92 cm/s
	Medicinal Chemist	try	
PAINS	0 alert	0 alert	0 alert
Brenk	0 alert	0 alert	0 alert
Leadlikeness	Yes	Yes	Yes
Synthetic accessibility	1.55	1.78	1.49
	Toxicity Risks		
Mutagenic	Not	Not	Not
Tumorigenic	Not	Not	Not
Irritant	Not	Not	Not



Not



Fig. 10. Radar image and the BOILED-Egg model of 1-3.

Conclusions

Three isomeric carboxylic acid derivatives obtained from the reaction of glutaric anhydride with *ortho, meta* and *para* bromoaniline at room temperature just in 2-3 min reaction time. The formation of supramolecular packing structures as a result of intermolecular H-bonding of carboxylic O–H…O and amide–N–H …O(amide) is the most important characteristics of these compounds. An intercalative binding mode was assigned for the interaction of these compounds with DNA as confirmed by UV-Visible spectroscopic and viscometric methods. The antimicrobial activity of the synthesized compounds performed against 5 bacterial and 2 fungal strains fall in the category of inactive compounds as their inhibition is less than 50%. A maximum of 78% DPPH

scavenging at 1800 μ L/mL concentration was shown by the compound-**3**. The compounds possess high gatrointestinal absorption and blood brain brayer (BBB) permeant properties. Similarly good bioavailability scores (0.85), drug score (0.44) and the higher skin absorption indicate that these compounds can be potential drug candidates.

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Graphical Abstract: Pictograph



Synopsis Graphical Abstract:

The three isomeric carboxylic acid derivatives interact with SS-DNA via intercalative mode which was confirmed by UV/Vis. Spectroscopy and viscometery. They have shown moderate antimicrobial and good antioxidant potenmitals.

checkCIF/PLATON report

Structure factors have been supplied for datablock(s) 1

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No syntax errors found. CIF dictionary Interpreting this report

Datablock: 1

Bond precision:	C-C = 0.0049 A	Wavelength	Wavelength=1.54184			
Cell:	a=4.8317(2)	b=10.2446(6)	c=12.0871(7)			
	alpha=93.050(5)	beta=98.725(4)	gamma=99.408(4)			
Temperature:	295 K					
	Calculated	Reported				
Volume	581.55(5)	581.55(5))			
Space group	P -1	P -1				
Hall group	-P 1	-P 1				
Moiety formula	C11 H12 Br N O3	C11 H12 I	Br N O3			
Sum formula	C11 H12 Br N O3	C11 H12 I	Br N O3			
Mr	286.12	286.13				
Dx,g cm-3	1.634	1.634				
Z	2	2				
Mu (mm-1)	4.774	4.774				
F000	288.0	288.0				
F000′	287.37					
h,k,lmax	6,12,15	6,12,15				
Nref	2429	2256				
Tmin,Tmax	0.349,0.620	0.395,1.0	000			
Tmin'	0.264					
Correction meth	od= # Reported T L	imits: Tmin=0.395 Tr	max=1.000			
ADSCOLL - MOLLI	JCAN					
Data completene	ss= 0.929	Theta(max) = 76.21	12			
R(reflections)=	0.0420(2101)		wR2(reflections)=			
			0.1181(2256)			
S = 1.073	Npar= 1	46				

The following ALERTS were generated. Each ALERT has the format test-name_ALERT_alert-type_alert-level.

Click on the hyperlinks for more details of the test.

🍛 Alert level C

PLAT029_AI	LERT_3_	C _diffrn_	_measured	_fraction	n_theta_	full value	Low .	0.963 Why?
PLAT911_AI	LERT_3_	C Missing	FCF Refl	Between	Thmin &	STh/L=	0.600	79 Report

Alert level G

PLAT007_ALERT_5_G Number of Unrefined Donor-H Atoms	2 Report
PLAT912_ALERT_4_G Missing # of FCF Reflections Above STh/L= 0.600	94 Note
PLAT913_ALERT_3_G Missing # of Very Strong Reflections in FCF	1 Note
PLAT933_ALERT_2_G Number of OMIT Records in Embedded .res File	8 Note
PLAT941_ALERT_3_G Average HKL Measurement Multiplicity	1.8 Low
PLAT978_ALERT_2_G Number C-C Bonds with Positive Residual Density.	1 Info

0 ALERT level A = Most likely a serious problem - resolve or explain 0 ALERT level B = A potentially serious problem, consider carefully 2 ALERT level C = Check. Ensure it is not caused by an omission or oversight 6 ALERT level G = General information/check it is not something unexpected 0 ALERT type 1 CIF construction/syntax error, inconsistent or missing data 2 ALERT type 2 Indicator that the structure model may be wrong or deficient 4 ALERT type 3 Indicator that the structure quality may be low 1 ALERT type 4 Improvement, methodology, query or suggestion 1 ALERT type 5 Informative message, check It is advisable to attempt to resolve as many as possible of the alerts in all categories. Often the minor alerts point to easily fixed oversights, errors and omissions in your CIF or refinement strategy, so attention to these fine details can be worthwhile. In order to resolve some of the more serious problems it may be necessary to carry out additional measurements or structure refinements. However, the purpose of your study may justify the reported deviations and the more serious of these should normally be commented upon in the discussion or experimental section of a paper or in the "special_details" fields of the CIF. checkCIF was carefully designed to identify outliers and unusual parameters, but every test has its limitations and alerts that are not important in a particular case may appear. Conversely, the absence of alerts does not guarantee there are no aspects of the results needing attention. It is up to the individual to critically assess their own results and, if necessary, seek expert advice.

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PLATON version of 13/07/2021; check.def file version of 13/07/2021

Datablock 1 - ellipsoid plot



checkCIF/PLATON report

Structure factors have been supplied for datablock(s) shelx

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No syntax errors found. CIF dictionary Interpreting this report

Datablock: shelx

Bond precision:	C-C = 0.0046 A	Wavelengt	Wavelength=1.54184	
Cell:	a=4.9260(2)	b=8.3664(5)	c=14.6791(6)	
	alpha=98.247(4)	beta=96.176(3)	gamma=99.697(4)	
Temperature:	294 K			
	Calculated	Reported		
Volume	584.75(5)	584.75(5)	
Space group	P -1	P -1		
Hall group	-P 1	-P 1		
Moiety formula	C11 H12 Br N O3	C11 H12	Br N O3	
Sum formula	C11 H12 Br N O3	C11 H12	Br N O3	
Mr	286.12	286.13		
Dx,g cm-3	1.625	1.625		
Z	2	2		
Mu (mm-1)	4.748	4.748		
F000	288.0	288.0		
F000′	287.37			
h,k,lmax	5,9,17	5,9,17		
Nref	2079	2078		
Tmin,Tmax	0.597,0.717	0.790,1.	000	
Tmin'	0.541			
Correction meth AbsCorr = GAUSS	od= # Reported T L IAN	imits: Tmin=0.790 T	max=1.000	
Data completene	ss= 1.000	Theta(max) = 67.0°	78	
R(reflections)=	0.0413(1918)		wR2(reflections) = 0.1100(2078)	
S = 1.062	Npar= 1	.51		

The following ALERTS were generated. Each ALERT has the format test-name_ALERT_alert-type_alert-level.

Click on the hyperlinks for more details of the test.

Alert level C

PLAT242_ALERT_2_C Low 'MainMol' Ueq as Compared to Neighbors of C1 Check

Alert level G

PLAT002_ALERT_2_G Number of Distance or Angle Restraints on AtSite4 NotePLAT172_ALERT_4_G The CIF-Embedded .res File Contains DFIX Records2 ReportPLAT860_ALERT_3_G Number of Least-Squares Restraints2 NotePLAT909_ALERT_3_G Percentage of I>2sig(I) Data at Theta(Max) Still76% NotePLAT910_ALERT_3_G Missing # of FCF Reflection(s) Below Theta(Min).1 NotePLAT933_ALERT_2_G Number of OMIT Records in Embedded .res File ...1 NotePLAT978_ALERT_2_G Number C-C Bonds with Positive Residual Density.6 Info

0 ALERT level A = Most likely a serious problem - resolve or explain
0 ALERT level B = A potentially serious problem, consider carefully
1 ALERT level C = Check. Ensure it is not caused by an omission or oversight
7 ALERT level G = General information/check it is not something unexpected
0 ALERT type 1 CIF construction/syntax error, inconsistent or missing data
4 ALERT type 2 Indicator that the structure model may be wrong or deficient
3 ALERT type 3 Indicator that the structure quality may be low
1 ALERT type 4 Improvement, methodology, query or suggestion
0 ALERT type 5 Informative message, check

It is advisable to attempt to resolve as many as possible of the alerts in all categories. Often the minor alerts point to easily fixed oversights, errors and omissions in your CIF or refinement strategy, so attention to these fine details can be worthwhile. In order to resolve some of the more serious problems it may be necessary to carry out additional measurements or structure refinements. However, the purpose of your study may justify the reported deviations and the more serious of these should normally be commented upon in the discussion or experimental section of a paper or in the "special_details" fields of the CIF. checkCIF was carefully designed to identify outliers and unusual parameters, but every test has its limitations and alerts that are not important in a particular case may appear. Conversely, the absence of alerts does not guarantee there are no aspects of the results needing attention. It is up to the individual to critically assess their own results and, if necessary, seek expert advice.

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Datablock shelx - ellipsoid plot



checkCIF/PLATON report

Structure factors have been supplied for datablock(s) 2

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No syntax errors found. CIF dictionary Interpreting this report

Datablock: 2

Bond precision:	C-C = 0.0030 A	Wavelength	=1.54184
Cell:	a=24.6256(2)	b=4.80980(5)	c=9.79495(13)
	alpha=90	beta=97.4563(10)	gamma=90
Temperature:	295 К		
	Calculated	Reported	
Volume	1150.35(2)	1150.34(2	2)
Space group	P 21/c	P 1 21/c	1
Hall group	-P 2ybc	-P 2ybc	
Moiety formula	C11 H12 Br N O3	C11 H12 E	Br N 03
Sum formula	C11 H12 Br N O3	C11 H12 E	8r N 03
Mr	286.12	286.13	
Dx,g cm-3	1.652	1.652	
Z	4	4	
Mu (mm-1)	4.827	4.827	
F000	576.0	576.0	
F000′	574.74		
h,k,lmax	31,6,12	31,6,12	
Nref	2408	2383	
Tmin,Tmax	0.588,0.713	0.387,1.0	000
Tmin'	0.285		
Correction metho AbsCorr = MULTI-	od= # Reported T -SCAN	Limits: Tmin=0.387 Tn	max=1.000
Data completene:	ss= 0.990	Theta(max) = 76.51	2
R(reflections)=	0.0329(2289)		wR2(reflections)=
S = 1.056	Npar=	147	0.0000 (2000)

The following ALERTS were generated. Each ALERT has the format test-name_ALERT_alert-type_alert-level.

Click on the hyperlinks for more details of the test.

🤪 Alert level C

PLAT250_ALERT_2_C Large U3/U1 Ratio for Average U(i,j) Tensor 2.3 Note

Alert level G

PLAT007_ALERT_5_G Number of Unrefined Donor-H Atoms	2	Report
PLAT910_ALERT_3_G Missing # of FCF Reflection(s) Below Theta(Min).	1	Note
PLAT912_ALERT_4_G Missing # of FCF Reflections Above STh/L= 0.600	23	Note
PLAT913_ALERT_3_G Missing # of Very Strong Reflections in FCF	1	Note
PLAT978_ALERT_2_G Number C-C Bonds with Positive Residual Density.	4	Info

0 ALERT level A = Most likely a serious problem - resolve or explain 0 ALERT level B = A potentially serious problem, consider carefully 1 ALERT level C = Check. Ensure it is not caused by an omission or oversight 5 ALERT level G = General information/check it is not something unexpected 0 ALERT type 1 CIF construction/syntax error, inconsistent or missing data 2 ALERT type 2 Indicator that the structure model may be wrong or deficient 2 ALERT type 3 Indicator that the structure quality may be low 1 ALERT type 4 Improvement, methodology, query or suggestion 1 ALERT type 5 Informative message, check It is advisable to attempt to resolve as many as possible of the alerts in all categories. Often the minor alerts point to easily fixed oversights, errors and omissions in your CIF or refinement strategy, so attention to these fine details can be worthwhile. In order to resolve some of the more serious problems it may be necessary to carry out additional measurements or structure refinements. However, the purpose of your study may justify the reported deviations and the more serious of these should normally be commented upon in the discussion or experimental section of a paper or in the "special_details" fields of the CIF. checkCIF was carefully designed to identify outliers and unusual parameters, but every test has its limitations and alerts that are not important in a particular case may appear. Conversely, the absence of alerts does not guarantee there are no aspects of the results needing attention. It is up to the individual to critically assess their own results and, if necessary, seek expert advice.

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Datablock 2 - ellipsoid plot



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Declaration of Interest Statement

The authors declare no conflict of interest.