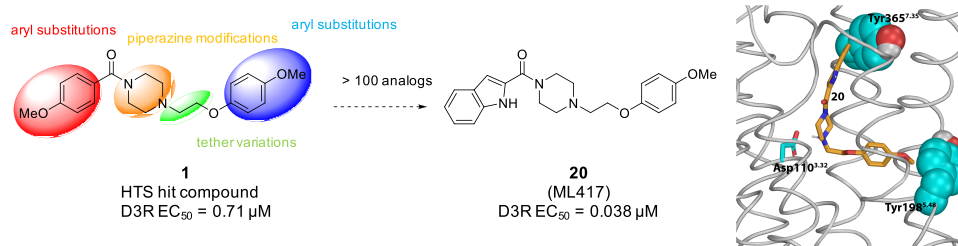


# Discovery, Optimization, and Characterization of ML417: A Novel and Highly Selective D<sub>3</sub> Dopamine Receptor Agonist

Amy E. Moritz, R. Benjamin Free, Warren S. Weiner, Emmanuel O. Akano, Disha Gandhi, Ara Abramyan, Thomas M. Keck, Marc Ferrer, Xin Hu, Noel Southall, Joseph Steiner, Jeffrey Aubé, Lei Shi, Kevin J. Frankowski,\* and David R. Sibley\*



**ABSTRACT:** To identify novel D<sub>3</sub> dopamine receptor (D3R) agonists, we conducted a high-throughput screen using a  $\beta$ -arrestin recruitment assay. Counterscreening of the hit compounds provided an assessment of their selectivity, efficacy, and potency. The most promising scaffold was optimized through medicinal chemistry resulting in enhanced potency and selectivity. The optimized compound, ML417 (**20**), potently promotes D3R-mediated  $\beta$ -arrestin translocation, G protein activation, and ERK1/2 phosphorylation (pERK) while lacking activity at other dopamine receptors. Screening of ML417 against multiple G protein-coupled receptors revealed exceptional global selectivity. Molecular modeling suggests that ML417 interacts with the D3R in a unique manner, possibly explaining its remarkable selectivity. ML417 was also found to protect against neurodegeneration of dopaminergic neurons derived from iPSCs. Together with promising pharmacokinetics and toxicology profiles, these results suggest that ML417 is a novel and uniquely selective D3R agonist that may serve as both a research tool and a therapeutic lead for the treatment of neuropsychiatric disorders.

## INTRODUCTION

Dysregulation of dopamine receptors is linked to the etiology and/or therapy of many neuropsychiatric disorders, including Parkinson's disease (PD), schizophrenia, and substance use disorders.<sup>1–5</sup> These receptors are categorized into two subfamilies: D1-like (D1R and D5R) and D2-like (D2R, D3R, and D4R) based on the protein structure, pharmacology, and physiological signaling.<sup>1,3,5</sup> Dopamine receptor subfamilies exhibit high sequence homology, particularly the D2R and D3R, which share 74% identity between their transmembrane domains (TMs) and 94% identity between their putative orthosteric binding sites, where the endogenous agonist dopamine (DA) binds.<sup>6</sup> Not surprisingly, most currently available drugs that target these receptors, including antipsychotics, anti-PD medications, and research tool compounds, are not highly subtype-selective but instead modulate both D2R and D3R subtypes to varying degrees.<sup>7,8</sup> Further, most known compounds that target the D2R and/or D3R also cross-react with other related G protein-coupled receptors (GPCRs), especially those for biogenic amines, creating the potential for profound off-target side effects. Thus, more selective agents are critically needed not only for the

delineation of dopamine receptor action *in vivo* but also for the development of more selective therapeutics with fewer side effects.

D3R activation is known to have important therapeutic effects. Agonists that are D3R-preferring (e.g., ~10-fold D3R > D2R-selective), including pramipexole and ropinirole, are effective in treating both PD and restless legs syndrome (RLS). These compounds are clinically active in relieving motor deficits and slowing the loss of dopaminergic terminals upon long-term administration to PD patients.<sup>9,10</sup> Further, in animal models, D3R-preferring agonists are the most potent neuroprotective agents identified to date against 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-induced neurodegeneration in mice<sup>11–14</sup> and primates<sup>15</sup> and against 6-hydroxydopamine (6-OHDA)-induced lesions in rats.<sup>16</sup>

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Importantly, studies with D3R-knockout mice support that the neuroprotective effects are mediated directly by the D3R.<sup>14,17,18</sup> Unfortunately, clinical treatment with drugs such as pramipexole and ropinirole is frequently associated with impulse control disorders, including pathological gambling, shopping, eating, and hypersexuality. These side effects typically occur at higher doses that would fully activate the D2R, which may underlie the loss of impulse control.<sup>19,20</sup> Thus, a highly selective D3R agonist that lacks D2R activity may hold significant therapeutic potential, as well as filling an important role as a probe compound for dissecting signaling pathways underlying D3R signaling from those of D2R.

Not surprisingly, extensive efforts have been directed toward the development of selective agonists and antagonists that target the D3R.<sup>21,22</sup> Significant progress has been made in the development of antagonists that are selective for the D3R vs other dopamine receptors, which has been aided by the availability of a crystal structure of the D3R in an inactive state.<sup>6,23</sup> The most selective D3R antagonists are extended-length compounds that are structurally bivalent, in that they possess a primary pharmacophore that binds to the orthosteric site and, connected via a linker, a secondary pharmacophore that binds to a secondary site, which may confer allosteric effects.<sup>23–30</sup> Bivalent compounds that possess allosteric properties are considered to be bitopic in nature.<sup>31–35</sup> No D3R-selective antagonists have yet reached the clinic;<sup>36,37</sup> however, newer antagonists have shown promise, especially for the treatment of substance use disorders.<sup>28,29,38–41</sup>

Less progress has been made in the development of D3R-selective agonists for clinical use. In part, this is due to the unavailability of a crystal structure of the D3R in an active state. As might be expected, virtual screens based on the inactive-state D3R structure have yielded mostly compounds with antagonist-like effects.<sup>42,43</sup> In contrast, synthetic medicinal chemistry approaches to D3R-selective agonists have been more successful with some agents achieving good pharmacological separation between the D3R and D2R.<sup>22,44–50</sup> Various examples of D3R-preferring agonists and their structures have recently been described in Moritz et al.<sup>8</sup> However, nearly all such compounds have been based on a single precursor scaffold, pramipexole, and their global GPCR selectivity and suitability for clinical advancement are unclear. Clearly, novel chemical scaffolds for designing highly selective D3R agonists are greatly needed. Very recently, the approach of designing extended-length bivalent compounds with D3R-selective agonist properties has been reported.<sup>34,51</sup>

Here, we report the identification and development of a novel agonist scaffold that exhibits high selectivity for the D3R. Starting from an unbiased high-throughput screening approach, we identified a hit compound with promising pharmacological and structural characteristics. From this initial hit compound, over 100 structural analogues were synthesized and characterized to develop a comprehensive structure–activity relationship (SAR) investigation and to establish the structural determinants for potency, efficacy, and selectivity at the D3R. An optimized lead compound, **20** (ML417), was identified that promotes potent D3R-mediated  $\beta$ -arrestin translocation, G protein-mediated signaling, and ERK1/2 phosphorylation (pERK) with minimal effects on other GPCR-mediated signaling. In addition, **20** was found to exhibit neuroprotection against toxin-induced neurodegeneration of dopaminergic neurons. In summary, **20** is a novel and uniquely selective D3R agonist that will prove useful as a

research tool and may show utility as a therapeutic lead for the treatment of neuropsychiatric disorders.

## RESULTS AND DISCUSSION

**High-Throughput Screening and Hit Compound Identification.** To identify novel agonist scaffolds for the D3R, we conducted a high-throughput screen (HTS) of the NIH Molecular Libraries Probe Production Centers Network (MLPCN) small molecule library consisting of  $\sim$ 400 000 diverse compounds.<sup>52</sup> The HTS utilized two concentrations (10 and 50  $\mu$ M) of each library compound and measured their ability to stimulate  $\beta$ -arrestin recruitment to the D3R. The primary screen identified 4165 compounds that stimulated  $\beta$ -arrestin recruitment greater than 30% ( $>3$  standard deviations) over basal (vehicle control) at either the lower (10  $\mu$ M) or higher (50  $\mu$ M) concentration of library compound. Primary screening data were deposited in PubChem (AID 652050 and 652048). Hit compounds were first chemoinformatically triaged based on results from a previously executed D2R HTS of the same library<sup>53,54</sup> to eliminate compounds that also possess D2R agonist activity. In addition, compounds were further triaged if they contained known chemically reactive moieties and/or promiscuous scaffolds resulting in  $\sim$ 2500 unique compounds that were then tested in secondary assays. These triaged hit compounds were characterized using 7-point concentration–response assays of  $\beta$ -arrestin recruitment to the D3R and D2R, as well as  $\beta$ -arrestin recruitment to the unrelated prostaglandin E receptor 2 (PTGER2) to assess selectivity and eliminate compounds that modulate  $\beta$ -arrestin recruitment in a nonspecific manner. One hundred and fifty-two unique agonist compounds replicated in a concentration-dependent manner with demonstrated D3R  $>$  D2R selectivity (data not shown) and were selected for further characterization.

All 152 vetted hit compounds were evaluated using D3R radioligand binding competition assays that employed the orthosteric antagonist [<sup>3</sup>H]-methylspiperone. Ninety of these compounds inhibited radioligand binding by greater than 50% at a dose of 40  $\mu$ M and were thus initially classified as orthosteric. In contrast, 62 compounds failed to inhibit binding by greater than 50% (but were functionally active) and thus initially classified as potentially allosteric in nature. As allosteric compounds have the potential for global GPCR selectivity,<sup>55</sup> we focused on those 62 compounds that were relatively ineffective at inhibiting [<sup>3</sup>H]-methylspiperone binding. These compounds were ranked via potency, D3R  $>$  D2R selectivity, and chemical tractability resulting in lead compound **1** (Figure 1).

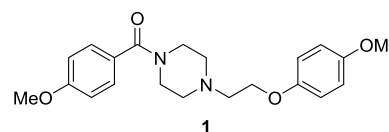
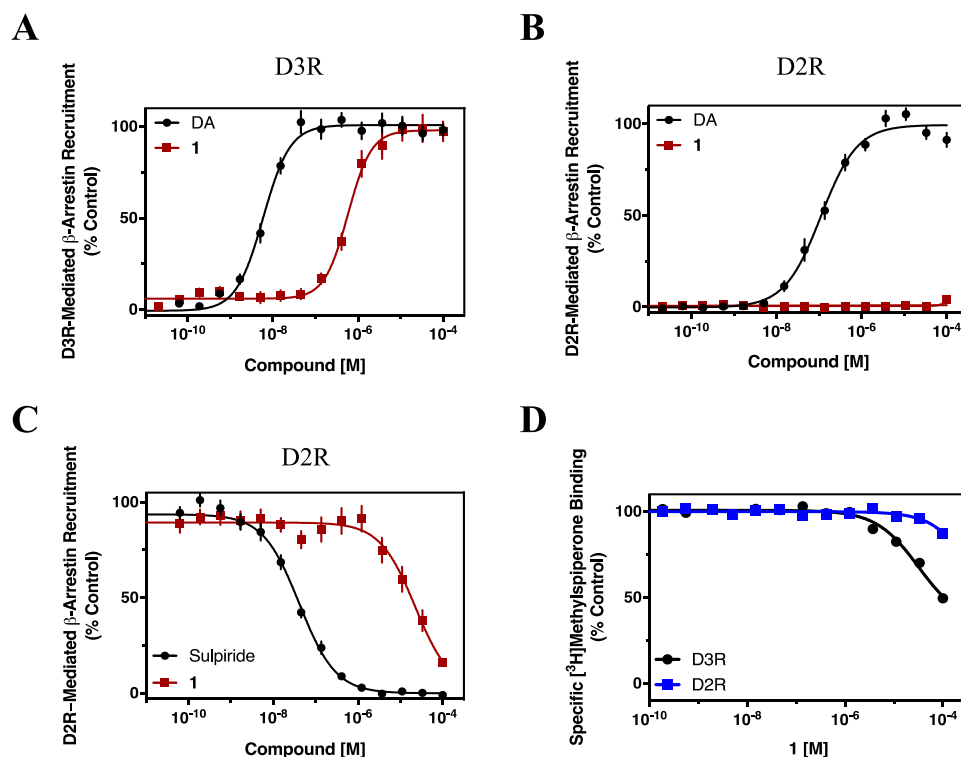


Figure 1. Chemical structure of the HTS lead, compound **1**.

The HTS lead **1** is a full agonist (compared to dopamine) in a D3R-mediated  $\beta$ -arrestin recruitment assay, displaying an EC<sub>50</sub> of 710 nM (Figure 2A). No measurable agonist activity was detected at the D2R using the same  $\beta$ -arrestin recruitment assay at concentrations up to 100  $\mu$ M (Figure 2B). Interestingly, when **1** was tested for its ability to inhibit DA-



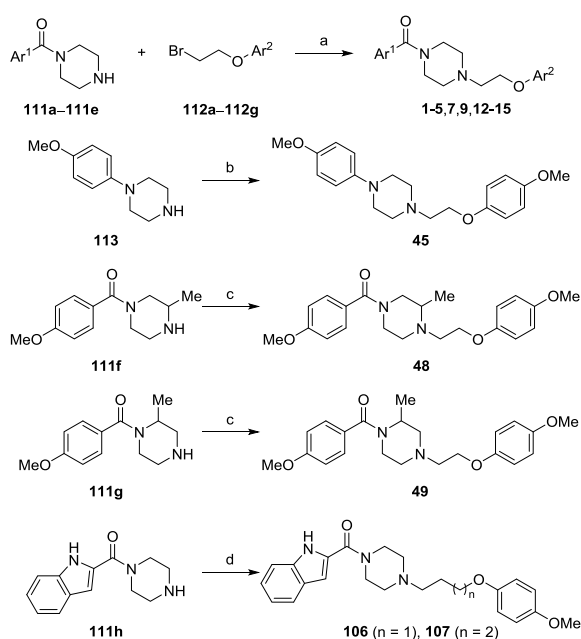
**Figure 2.** Pharmacological activity of lead compound **1** on the D3R and D2R. (A, B) Agonist concentration–response curves for stimulating  $\beta$ -arrestin recruitment in response to either dopamine (DA) or **1** using cells expressing either the D3R (A) or D2R (B). Cells were stimulated as indicated, and  $\beta$ -arrestin recruitment was assessed using the DiscoverX assay as described in the [Experimental Section](#). Data are expressed as a percentage of the maximum dopamine (DA) signal and are shown as means  $\pm$  standard error of the mean (SEM) of at least three experiments performed in triplicate. In (A), the  $EC_{50}$  value for DA is  $6.4 \pm 0.6$  nM (mean  $\pm$  SEM,  $n = 12$ ) and of **1** is  $710 \pm 150$  nM (mean  $\pm$  SEM,  $n = 13$ ). In (B), the  $EC_{50}$  value for DA is  $140 \pm 23$  nM (mean  $\pm$  SEM,  $n = 13$ ). (C) Antagonist concentration–response curves of either sulpiride or **1** for inhibiting  $\beta$ -arrestin recruitment to the D2R. Cells were incubated with the indicated concentrations of the compound in the presence of an  $EC_{80}$  concentration of DA ( $1 \mu\text{M}$ ). Data are expressed as a percentage of signal seen in the absence of test compounds and are shown as means  $\pm$  SEM of at least three experiments performed in triplicate. The  $IC_{50}$  value of sulpiride is  $42 \pm 4.9$  nM (mean  $\pm$  SEM,  $n = 8$ ) and of **1** is  $16 \pm 3.0 \mu\text{M}$  (mean  $\pm$  SEM,  $n = 10$ ). (D) Radioligand binding assays using [ $^3\text{H}$ ]-methylspiperone were performed as described in the [Experimental Section](#). Data are representative of six independent experiments and expressed as a percentage of the specific binding. The  $IC_{50}$  values of **1** were determined to be  $>100 \mu\text{M}$  for the D3R and D2R.

stimulated  $\beta$ -arrestin recruitment to the D2R, we found that it was able to antagonize the receptor with an  $IC_{50}$  of  $16 \mu\text{M}$  (Figure 2C). We next sought to determine the affinity of **1** at the orthosteric binding site by conducting competition binding studies with [ $^3\text{H}$ ]-methylspiperone at both the D2R and D3R. As shown in Figure 2D, the compound weakly inhibited radioligand binding to the D3R with an  $IC_{50} > 10 \mu\text{M}$  and no measurable ability to displace binding to the D2R. This observation suggests an apparent disconnect between the potency of **1** as an agonist in the D3R  $\beta$ -arrestin functional assay and its limited, but demonstrable, ability to inhibit the binding of [ $^3\text{H}$ ]-methylspiperone to the D3R. One possible explanation is that **1** is a weak partial (orthosteric) agonist of the D3R that appears as a potent full agonist in the functional assay due to spare receptors and/or signal amplification. This is not likely, however, as the  $\beta$ -arrestin recruitment assay only produces a signal when the receptor and  $\beta$ -arrestin are complemented 1:1—there are no spare receptors or signal amplification. Regardless of the mechanism, the overall activity profile of **1** appeared to warrant further investigation and its optimization.

**Chemistry.** We employed several complementary synthetic routes to construct the target analogue compounds all based on a central strategy of iterative derivatization of the piperazine core. Our initial strategy to the HTS hit compound resynthesis

and analogue exploration coupled the *N*-acylated piperazines **111** with the alkyl bromides **112** (Scheme 1) utilizing General Procedure A or B (see the [Experimental Section](#) for general procedure details and the synthesis of components **111** and **112**). For specific target analogues, we utilized slightly modified conditions as shown in Scheme 1. Exploration of the *N*-acyl moiety was most efficiently achieved through a reversal in the order of piperazine functionalization. Thus, alkylation of 1-Boc-piperazine and subsequent Boc deprotection afforded the 1-(2-(aryloxy)ethyl)piperazines **114**, which were then acylated utilizing General Procedures D or E (see the [Experimental Section](#)) to provide the final analogues (Scheme 2). Again, specific target analogues required slightly modified conditions, as shown in Scheme 2. We utilized this broadly defined acylation strategy to access a total of 59 analogues, the most common approach that we employed in these SAR studies. The sulfonamide analogue **44** was synthesized using an analogous protocol to acylation by replacing the acid chloride component with 4-methoxybenzene-1-sulfonyl chloride (Scheme 3). The racemic methyl derivative **46** was synthesized via reductive amination of carboxamide fragment **111a** and 1-(4-methoxyphenoxy)propan-2-one (Scheme 4). To efficiently explore the aryl ether composition of (2-indoyl)-substituted piperazine analogues, we employed a nucleophilic displacement approach on

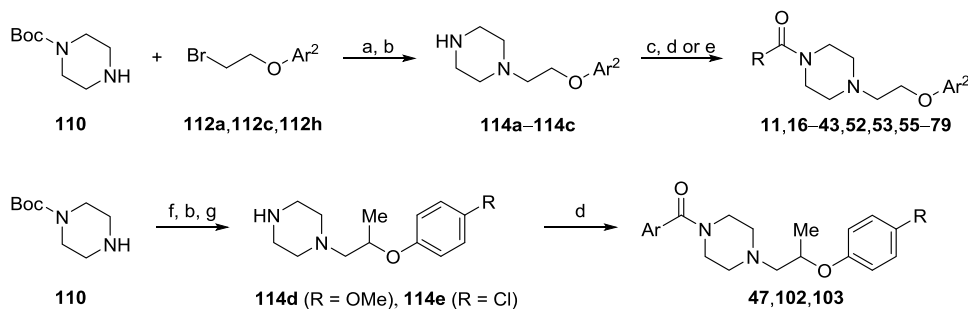
### Scheme 1. Summary of Alkylation Routes to SAR Analogues<sup>a</sup>



<sup>a</sup>Reagents and conditions: (a) potassium iodide (0.1 or 1.0 equiv),  $K_2CO_3$  (3.0 equiv), MeCN or DMF, 60 °C, 14–20 h, 27–81% yield; (b) **112a** (1.1 equiv),  $Et_3N$  (1.4 equiv), MeCN, 60 °C, 18 h, 68% yield; (c) **112a** (1.1 equiv),  $K_2CO_3$  (2.0 equiv), *N,N*-dimethylformamide (DMF), 100 °C, 5 h, 37% yield; and (d) 1-(3-bromopropoxy)-4-methoxybenzene or 1-(4-bromobutoxy)-4-methoxybenzene, potassium iodide (1.0 equiv),  $K_2CO_3$  (3.0 equiv), MeCN, 60 °C, 19 h, 76–83% yield.

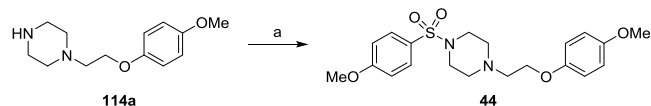
the alkyl chloride **116** (Scheme 5) utilizing General Procedure F or G (see the Experimental Section). This displacement protocol also furnished the piperidine analogue **50** and 1,2-dimethylethylenediamine analogue **51** using the corresponding amine starting materials **117** and **119** (Scheme 5). Select individual analogue syntheses utilized a complementary Mitsunobu reaction protocol on the 1-(2-hydroxyethyl)-piperazine **121** (Scheme 6). Minor modifications to these strategies were utilized to construct other target analogues. Thus, starting with 1-(2-hydroxyethyl)piperidine **117** and

### Scheme 2. Summary of Late-Stage Acylation Routes to SAR Analogues<sup>a</sup>



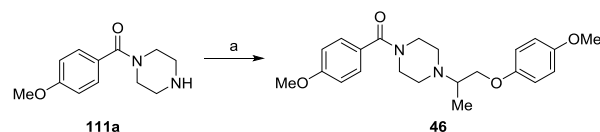
<sup>a</sup>Reagents and conditions: (a)  $K_2CO_3$  (2.0 equiv), potassium iodide (0.1 equiv), MeCN or DMF, 70 or 90 °C, 16–21 h, 70–79% yield; (b)  $Et_3SiH$  (1.5–2.0 equiv), trifluoroacetic acid (15–20 equiv),  $CH_2Cl_2$ , room temperature (rt), 4–26 h, 80–91% yield; (c)  $Ar^1C(O)Cl$  (1.0–1.3 equiv),  $Et_3N$  (1.5 equiv),  $CH_2Cl_2$ , rt, 16–20 h, 61–69% yield; (d)  $Ar^1CO_2H$  (1.0–1.3 equiv), PyBOP (1.2 equiv), *i*- $Pr_2EtN$  (3.0 equiv), DMF, rt, 16–20 h, 15–93% yield; (e) 1*H*-indole-2-carboxylic acid (1.2 equiv), diisopropylcarbodiimide (3.0 equiv), 4-dimethylaminopyridine (DMAP, 0.1 equiv), tetrahydrofuran (THF), rt, 15 h, 73% yield; (f) 2-chloropropanoyl chloride (1.1 equiv),  $K_2CO_3$  (2.5 equiv), 4-methoxyphenol or 4-chlorophenol (1.2 equiv), DMF, 80 °C, 4 h, 24–28% yield; and (g)  $BH_3 \cdot THF$  (3.1–3.2 equiv), THF, 65 °C, 0.5–4 h, 86–88% yield.

### Scheme 3. Synthesis of the Sulfonamide Analogue **44**<sup>a</sup>



<sup>a</sup>Reagents and conditions: (a) 4-methoxybenzene-1-sulfonyl chloride (1.0 equiv),  $Et_3N$  (2.0 equiv), toluene, rt, 19 h, 55% yield.

### Scheme 4. Reductive Amination Protocol for the Synthesis of the Methyl-Substituted Analogue **46**<sup>a</sup>

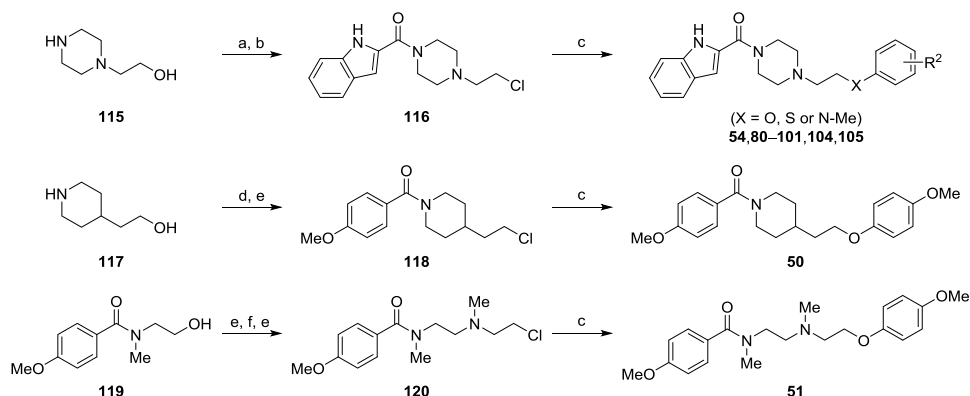


<sup>a</sup>Reagents and conditions: (a) 1-(4-methoxyphenoxy)propan-2-one (1.0 equiv),  $NaBH(OAc)_3$  (1.5 equiv),  $ClCH_2CH_2Cl$ , AcOH (0.3 equiv), 50 °C, 7 d, 57% yield.

using a nucleophilic displacement approach allowed ready access to compound **108** (Scheme 7). In an analogous approach to the late-stage acylation used in Scheme 2, the alkylated piperazine **114a** was subjected to a second alkylation to afford compound **109** (Scheme 8). Overall, the routes summarized here enabled the efficient and modular construction of the numerous analogues synthesized and facilitated the survey of structural modifications on all regions of compound **1**. The activity of these analogues and our optimization strategy will be discussed in detail in the following section.

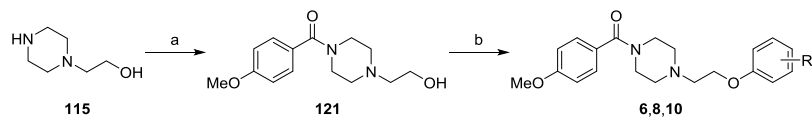
**Structure–Activity Relationships and HTS Hit Compound **1** Optimization.** In an effort to increase the functional potency of **1** at the D3R, while at the same time eliminate its D2R antagonist activity, we explored SAR studies of this scaffold by synthesizing over 100 unique analogues of **1**. These analogues were designed to investigate four regions of the scaffold, including substitutions of the aryl ether and aryl carboxamide groups, modifications of the piperazine core, and changes to the central tether of the molecule, as illustrated in Figure 3. All analogues were analyzed in the D2R and D3R  $\beta$ -arrestin recruitment assays to generate a comprehensive SAR survey around the scaffold.

### Scheme 5. Summary of Alkyl Chloride Displacement Routes to SAR Analogues<sup>a</sup>



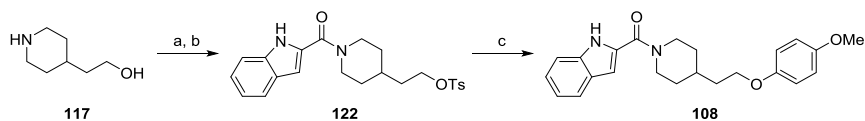
<sup>a</sup>Reagents and conditions: (a) 1*H*-indole-2-carbonyl chloride (1.1 equiv), Et<sub>3</sub>N (1.2 equiv), CH<sub>2</sub>Cl<sub>2</sub>, rt, 20 h, 75% yield; (b) triphosgene (0.5 equiv), CH<sub>2</sub>Cl<sub>2</sub>, 0 °C to rt, 3 h, 53% yield; (c) Ar<sup>2</sup>OH (1.1–1.9 equiv), Ar<sup>2</sup>SH (1.6 equiv) or Ar<sup>2</sup>NHMe (1.6 equiv), K<sub>2</sub>CO<sub>3</sub> (1.8–3.3 equiv), DMF, 50–80 °C, 7–19 h, 5–73% yield; (d) 4-methoxybenzoyl chloride (1.1 equiv), Et<sub>3</sub>N (1.5 equiv), CH<sub>2</sub>Cl<sub>2</sub>, rt, 16 h, 80% yield; (e) thionyl chloride (1.0–1.5 equiv), CHCl<sub>3</sub>, 50–55 °C, 0.5 or 46 h, 54–86% yield; and (f) 2-(methylamino)ethanol (5.1 equiv), MeCN, 80 °C, 25 h, 67% yield.

### Scheme 6. Mitsunobu Reaction Route to SAR Analogues 6, 8, and 10<sup>a</sup>



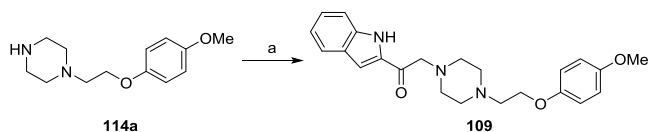
<sup>a</sup>Reagents and conditions: (a) 4-methoxybenzoyl chloride (1.1 equiv), Et<sub>3</sub>N (1.3 equiv), CH<sub>2</sub>Cl<sub>2</sub>, 0 °C to rt, 24 h, 68% yield; and (b) Ar<sup>2</sup>OH (1.0 equiv), Ph<sub>3</sub>P (1.0 equiv), diisopropyl azodicarboxylate (DIAD, 1.3 equiv), CH<sub>2</sub>Cl<sub>2</sub> or THF, 60 °C, 16–40 h, 15–46% yield.

### Scheme 7. Synthesis of the Piperidine Analogue 108<sup>a</sup>

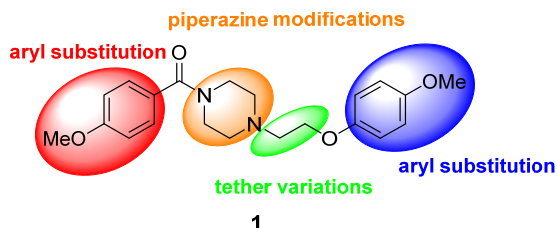


<sup>a</sup>Reagents and conditions: (a) indole-2-carboxylic acid (1.0 equiv), HOBT (1.0 equiv), EDC·HCl (1.0 equiv), CH<sub>2</sub>Cl<sub>2</sub>, rt, 18 h, 49% yield; (b) TsCl (2.0 equiv), Et<sub>3</sub>N (4.0 equiv), CH<sub>2</sub>Cl<sub>2</sub>, rt, 2 h, 46% yield; and (c) 4-methoxyphenol (3.0 equiv), K<sub>2</sub>CO<sub>3</sub> (3.0 equiv), MeCN, 60 °C, 17 h, 70% yield.

### Scheme 8. Synthesis of the Dibasic Piperazine Analogue 109<sup>a</sup>



<sup>a</sup>Reagents and conditions: (a) 2-chloro-1-(1*H*-indol-2-yl)ethan-1-one (1.0 equiv), KI (1.0 equiv), K<sub>2</sub>CO<sub>3</sub> (4.0 equiv), MeCN, 65 °C, 18 h, 51% yield.



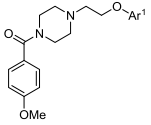
**Figure 3.** Illustration of structural modifications to compound **1** to investigate structure–activity relationships.

We began our SAR studies by examining modifications to the 4-methoxyphenyl groups on the left and right termini of

compound **1** (i.e., aryl carboxamide and aryl ether moieties, respectively). In the latter case, replacing the 4-methoxyphenyl ether with other aryl ethers afforded analogues possessing a wide range of D3R agonist potencies (Table 1). The 2-methoxyphenyl ether **2** was found to be more than twice as potent, while the 3-methoxyphenyl ether **3** was more than 7-fold as potent, compared to compound **1**. Other monosubstituted phenyl ether analogues were less potent (entries 4–6). In fact, the 4-(trifluoromethoxy)phenyl (**6**) and 3,5-dimethoxyphenyl (**7**) ethers were found to be inactive even at the highest tested concentrations. All compounds in this subset possessed no D2R agonism and only negligible, if any, D2R antagonism. The unsubstituted phenyl ether **8** was almost as potent as the 3-methoxyphenyl ether ( $EC_{50} = 115$  nM); however, this analogue was found to also possess D2R agonism ( $EC_{50} = 2300$  nM). Aryl ether **9**, which contained bridged 3,4-dialkoxy substitution, possessed a 2-fold improved D3R agonist potency and excellent selectivity, albeit with reduced agonist efficacy at the D3R. The 3-pyridyl ether analogue **10** was 35-fold more potent than the hit compound **1** in the D3R agonist assay ( $EC_{50} = 17$  nM); however, it also exhibited modestly potent D2R agonist activity ( $EC_{50} = 2900$  nM).

In a more extensive effort than above, we investigated replacements of the 4-methoxybenzamide on the left-hand

**Table 1. Analogues Exploring 2-(4-Methoxyphenylether)ethyl Replacement**

Compound ID		D3R agonist activity <sup>a</sup>		D2R agonist activity <sup>a</sup>		D2R antagonist activity <sup>a</sup>	
		Ar <sup>a</sup> =	EC <sub>50</sub> (nM)	E <sub>max</sub> (% control)	EC <sub>50</sub> (nM)	E <sub>max</sub> (% control)	IC <sub>50</sub> (nM)
1	4-methoxyphen-1-yl	710 ± 150	102 ± 4.2	Inactive	Inactive	15,700 ± 3,000	88 ± 7
2	2-methoxyphen-1-yl	278 ± 62	36 ± 3.1	Inactive	Inactive	9,000 ± 3,700	99 ± 1
3	3-methoxyphen-1-yl	98 ± 21	95 ± 6	>100,000	ND	6,800 ± 1,400	63 ± 6
4	4-ethylphen-1-yl	2,600 ± 550	44 ± 9.3	Inactive	Inactive	>50,000	ND
5	4-chlorophen-1-yl	1,000 ± 275	103 ± 27	Inactive	Inactive	>10,000	101 ± 2
6	4-(trifluoromethoxy)phen-1-yl	Inactive	Inactive	Inactive	Inactive	>50,000	ND
7	3,5-dimethoxyphen-1-yl	Inactive	Inactive	Inactive	Inactive	>50,000	ND
8	phenyl	115 ± 12	64 ± 4	2300 ± 900	38 ± 5	>100,000	ND
9	benzo[d][1,3]dioxol-5-yl	310 ± 150	78 ± 9.4	Inactive	Inactive	>10,000	87 ± 6.8
10	3-pyridyl	17 ± 2.3	110 ± 15	2,900 ± 1,300	96 ± 7.4	>100,000	ND

<sup>a</sup> $\beta$ -arrestin recruitment activity was assessed as described in Figure 2. E<sub>max</sub> values are expressed as a percentage of the maximum dopamine response observed in the same assay. I<sub>max</sub> values are expressed as a percentage of the maximum inhibition of a dopamine (EC<sub>80</sub> concentration) response observed with the antagonist sulpiride in the same assay. <sup>b</sup>ND curve did not plateau.

portion of compound **1** (Table 2). The unsubstituted benzamide analogue **11** was approximately an equipotent D3R agonist compared to the hit compound **1**, along with ablation of the D2R antagonism. The 2-methoxybenzamide (**12**) possessed only modest potency, while the 3-methoxybenzamide (**13**) was marginally more potent. Other monosubstituted benzamides were less promising, with the 4-chloro analogue **14** inactive as a D3R agonist and the 4-ethyl analogue **15** on par in potency compared to the hit. However, both **14** and **15** also possessed limited D2R antagonist activity. We next examined a number of heterocyclic carboxamides at this position, beginning with the two pyridyl analogues **16** and **17**, neither of which possessed significant D3R agonist activity. We next investigated a series of indole carboxamide analogues. While all three of the 5-, 3-, and 2-indole carboxamide analogues (**18**, **19**, and **20**, respectively) tested possessed potent D3R agonist activity, only **20** was completely D3R-selective and also possessed the greatest potency (EC<sub>50</sub> = 38 nM), almost a 20-fold improvement compared to the hit. Thus, we delved more systematically into the substitution of 2-indole carboxamide analogues, examining methoxy, methyl, and chloro substitutions around the benzene of the indole moiety (i.e., 4-, 5-, 6-, and 7-position substitutions). The methoxy-substituted analogues **21–24** were all D3R agonists of reasonably high potency (EC<sub>50</sub>s = 155–980 nM); however, all of these analogues also possessed measurable D2R agonist activity (EC<sub>50</sub>s = 5200–7800 nM). The methyl-substituted analogues **25–28** were also all fairly potent D3R agonists (EC<sub>50</sub>s = 130–611 nM), along with a reduced D2R agonism (EC<sub>50</sub>s = 1100 to >50 000 nM). The 5-methyl analogue **27** was the most selective and possessed D3R agonist potency (EC<sub>50</sub> =

611 nM) on par with the hit compound **1**. Similarly, the chloro-substituted analogues **29–32** were all D3R agonists of modest potency (EC<sub>50</sub>s = 160–2900 nM), with D2R agonist activity similar to the methyl-substituted series (EC<sub>50</sub>s = 3500 to > 50 000 nM). In this series, only the 5-chloro analogue was fully D3R-selective; however, the D3R agonism was not very potent (EC<sub>50</sub> = 2900 nM). None of these substituted analogues approached the potency of the unsubstituted indole **20**, and we did not further investigate substitution of this region of the indole.

Next, we probed the substitution on the nitrogen-containing ring of the indole by testing the *N*- and 3-methyl analogues **33** and **34**, respectively. While the *N*-methyl analogue retained D3R agonist activity on par with compound **1**, it also exhibited D2R antagonism. The 3-methyl analogue was nonselective, showing only weak activity in all three assays (EC<sub>50</sub>s = 1900–4400 nM). We next examined other bicyclic heterocycles attached to the carboxamide, beginning with an additional nitrogen incorporation into the indole ring. Moving the additional nitrogen atom around the ring afforded analogues (**35–39**) of modest D3R agonist potency (EC<sub>50</sub>s = 167–2800 nM), though often with modest (**35**, EC<sub>50</sub> = 413 nM) to weak (**36** and **37**, EC<sub>50</sub>s = 7800 and 5500 nM, respectively) D2R agonism. Only the benzimidazole analogue **39** was both reasonably potent (EC<sub>50</sub> = 192 nM) and D3R-selective. Replacement of the indole with a benzofuran moiety afforded analogue **40**, a D3R agonist of modest potency (EC<sub>50</sub> = 430 nM), though with weak D2R antagonism (IC<sub>50</sub> = 7700 nM). The benzothiophene analogue **41** was both less potent and selective than the benzofuran. The cyclohexyl and methyl carboxamides (**42** and **43**, respectively) did not show any

**Table 2. Analogues Exploring 4-Methoxybenzamide Replacement**

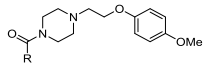
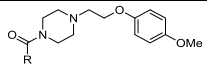
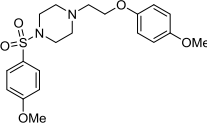
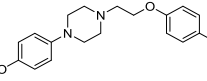
Compound ID	 full structure or R =	D3R agonist activity <sup>a</sup>		D2R agonist activity <sup>a</sup>		D2R antagonist activity <sup>a</sup>	
		EC <sub>50</sub> (nM)	E <sub>max</sub> (% control)	EC <sub>50</sub> (nM)	E <sub>max</sub> (% control)	IC <sub>50</sub> (nM)	I <sub>max</sub> (% control)
11	phenyl	548 ± 165	70 ± 2.6	Inactive	Inactive	>50,000	ND
12	2-methoxyphen-1-yl	2,500 ± 1,000	113 ± 16	Inactive	Inactive	>100,000	ND
13	3-methoxyphen-1-yl	550 ± 57	112 ± 12	24,000 ± 3000	24 ± 7	Inactive	Inactive
14	4-chlorophen-1-yl	Inactive	Inactive	Inactive	Inactive	9,000 ± 3,000	107 ± 8.8
15	4-ethylphen-1-yl	530 ± 130	95 ± 13	Inactive	Inactive	6,000 ± 1,200	106 ± 15
16	4-pyridyl	22,000 ± 5,600	50 ± 7.9	Inactive	Inactive	Inactive	Inactive
17	5-methoxy-2-pyridyl	2,100 ± 375	97 ± 5.7	>50,000	ND	>100,000	ND
18	5-indolyl	208 ± 62	112 ± 11	2,500 ± 383	18.5 ± 3.2	4,600 ± 1,900	64 ± 11.2
19	3-indolyl	210 ± 29	89 ± 9	>100,000	ND	2,800 ± 500	86 ± 2
20	2-indolyl	38 ± 4.0	103 ± 3	>10,000	ND	>10,000	ND
21	7-methoxy-2-indolyl	980 ± 135	115 ± 6.5	6,000 ± 1,300	56 ± 9.7	>10,000	ND
22	6-methoxy-2-indolyl	155 ± 62	91 ± 18	5,200 ± 3,500	62 ± 11.5	Inactive	Inactive
23	5-methoxy-2-indolyl	520 ± 25	109 ± 2.6	7,800 ± 1,900	46 ± 14.2	>100,000	ND
24	4-methoxy-2-indolyl	411 ± 144	116 ± 5.7	5,200 ± 255	33 ± 4.7	>100,000	ND
25	7-methyl-2-indolyl	473 ± 44	103 ± 15.5	10,100 ± 780	49 ± 18.6	>100,000	ND
26	6-methyl-2-indolyl	130 ± 40	117 ± 3	1,100 ± 400	72 ± 7	Inactive	Inactive
27	5-methyl-2-indolyl	611 ± 79	119 ± 7.9	>50,000	ND	>100,000	ND
28	4-methyl-2-indolyl	266 ± 82	106 ± 15	4,300 ± 1,700	37 ± 5.5	744 ± 282	30 ± 8.2
29	7-chloro-2-indolyl	563 ± 138	116 ± 13	13,000 ± 5,200	70 ± 26	>100,000	ND
30	6-chloro-2-indolyl	225 ± 73	110 ± 30	3,500 ± 1,400	58 ± 12.4	>100,000	ND
31	5-chloro-2-indolyl	2,900 ± 1,100	117 ± 14.7	Inactive	Inactive	Inactive	Inactive
32	4-chloro-2-indolyl	160 ± 56	105 ± 4	7,100 ± 3,300	57 ± 17	>100,000	ND
33	1-methyl-2-indolyl	310 ± 52	88 ± 6	Inactive	Inactive	9,600 ± 2,400	105 ± 9
34	3-methyl-2-indolyl	1,900 ± 600	92 ± 4	4,400 ± 1,200	31 ± 6	>100,000	ND
35	pyrrolo[2,3-b]pyridin-2-yl	167 ± 21	109 ± 14	413 ± 105	94 ± 18	Inactive	Inactive
36	pyrrolo[2,3-c]pyridin-2-yl	810 ± 130	105 ± 9.8	7,800 ± 3,300	21 ± 3.1	>100,000	ND
37	pyrrolo[3,2-c]pyridin-2-yl	576 ± 102	107 ± 901	5,500 ± 2,200	28 ± 5.6	>100,000	ND
38	pyrrolo[3,2-b]pyridin-2-yl	2,800 ± 541	103 ± 9.4	>100,000	ND	Inactive	Inactive
39	benzo[d]imidazol-2-yl	192 ± 76	95 ± 12.3	Inactive	Inactive	>50,000	ND

Table 2. continued

Compound ID	 full structure or R =	D3R agonist activity <sup>a</sup>		D2R agonist activity <sup>a</sup>		D2R antagonist activity <sup>a</sup>	
		EC <sub>50</sub> (nM)	E <sub>max</sub> (% control)	EC <sub>50</sub> (nM)	E <sub>max</sub> (% control)	IC <sub>50</sub> (nM)	I <sub>max</sub> (% control)
40	benzofuran-2-yl	430 ± 117	80 ± 14.4	Inactive	Inactive	7,700 ± 2,500	111 ± 7
41	benzothiophen-2-yl	3,300 ± 1,000	51 ± 5.8	Inactive	Inactive	5,100 ± 1,000	96 ± 5.6
42	cyclohexyl	>50,000	ND	Inactive	Inactive	>50,000	ND
43	methyl	9,700 ± 2,500	35 ± 9.2	Inactive	Inactive	Inactive	Inactive
44		Inactive	Inactive	>100,000	ND	Inactive	Inactive
45		Inactive	Inactive	Inactive	Inactive	>50,000	ND

<sup>a</sup> $\beta$ -arrestin recruitment activity was assessed as described in Figure 2.  $E_{\max}$  values are expressed as a percentage of the maximum dopamine response observed in the same assay.  $I_{\max}$  values are expressed as a percentage of the maximum inhibition of a dopamine ( $EC_{50}$  concentration) response observed with the antagonist sulpiride in the same assay. <sup>b</sup>ND curve did not plateau.

appreciable activity, with only **43** possessing any measurable D3R agonist potency ( $EC_{50}$  = 9700 nM). The 4-methoxyphenyl sulfonamide **44** possessed no measurable activity across the three SAR assays, indicating that the sulfonamide isostere is not a suitable replacement for the carboxamide functional group in this instance. The directly attached 4-methoxyphenyl piperazine **45** was similarly inactive, again indicating the importance of the carboxamide for potency.

We explored a number of modifications to the piperazine or alkyl chain of the tethered ether, most often through the addition of a single methyl group to the HTS lead compound **1** structure (Table 3). Thus, analogues **46** and **47** contained an additional methyl group on the two-carbon tether bridging the piperazine and aryl ether groups. While both were D3R-selective agonists, the position of the methyl group was critical to the effect on potency. Analogue **46** (where the methyl group was adjacent to the piperazine) possessed weak D3R agonist potency ( $EC_{50}$  = 9300 nM), while the constitutional isomer **47** (with a methyl group adjacent to the ether moiety) was almost 3-fold more potent than hit compound **1** ( $EC_{50}$  = 160 nM). The effect of methyl substitution on the piperazine ring exhibited an analogous dependence on position. Analogue **48** (with the methyl group adjacent to the basic piperazine nitrogen) displayed greatly diminished potency and efficacy, while analogue **49** (with a methyl group adjacent to the amide nitrogen) was slightly more potent than the hit compound **1** ( $EC_{50}$  = 510 nM), although **49** was not fully selective, possessing weak D2R agonism ( $EC_{50}$  = 4200 nM). The profound detrimental effect observed from the methyl group introduction adjacent to the basic piperazine nitrogen indicates the importance of this moiety for activity, although whether this arises from steric interactions or through the induction of an unfavorable conformation is not immediately clear. Other structural modifications were explored and found to cause an almost complete loss of any activity. The piperidine analogue **50** was inactive in all assays, and the ring-opened analogue **51**

possessed only very weak D2R antagonist activity ( $IC_{50}$  = 18 000 nM).

Having identified potency-enhancing replacements for either the 4-methoxyphenyl ether or 4-methoxybenzamide group of compound **1**, we were interested in exploring simultaneous changes to both ends of the molecule (Table 4). The 3-methoxyphenyl (**3**) and 3-pyridyl ethers (**10**) were two of the most potent aryl ether analogues identified ( $EC_{50}$ s = 98 and 17 nM, respectively) (Table 1), though both also possessed weak D2R activity. We thus explored a range of carboxamide analogues to better identify SAR trends that could lead to potent and selective analogues. The 3-methoxyphenyl ether analogues **52–56** contained some of the most promising carboxamide groups from earlier SAR studies and, gratifyingly, all afforded potent D3R analogues ( $EC_{50}$ s = 34–285 nM). The most potent was the 2-indolyl carboxamide analogue **54** ( $EC_{50}$  = 34 nM), although it also possessed slight D2R agonist activity ( $EC_{50}$  = 2300 nM). The other 3-methoxyphenyl ether analogues were both less potent and even less selective than **54**. The 3-pyridyl ether analogue **10** was the most potent ether replacement identified ( $EC_{50}$  = 17 nM), and we explored a more comprehensive survey of carboxamide replacements paired with this ether, primarily focused on substituted 2-indole carboxamides. The 4-methoxy-2-methylphenyl carboxamide **57** was almost 7-fold less potent than the corresponding 4-methoxyphenyl analogue **10**. In an effort that mirrored the original exploration of carboxamide groups, we tested a series of methoxy-, methyl-, and chloro-substituted 2-indoles. These analogues (**58–70**) possessed good potency ( $EC_{50}$ s = 3–116 nM), with six analogues (**58**, **61**, **63**, **66**, **68**, and **70**) in the single-digit nanomolar range. Unfortunately, none of these analogues were highly selective D3R agonists and the more potent compounds were generally the least selective. Upon comparing the 3-pyridyl ether series to the 4-methoxyphenyl ether series, we also noted poor correlation between indole substitution position and rank order of potency, suggesting that the SAR trends might not be independent of the substitution at other positions of the molecule. Additional



**Table 3. Analogues Exploring the Modification of the Tether or Piperazine Core**

Compound ID	Structure	D3R agonist activity <sup>a</sup>		D2R agonist activity <sup>a</sup>		D2R antagonist activity <sup>a</sup>	
		EC <sub>50</sub> (nM)	E <sub>max</sub> (% control)	EC <sub>50</sub> (nM)	E <sub>max</sub> (% control)	IC <sub>50</sub> (nM)	I <sub>max</sub> (% control)
46		9,300 ± 2,500	106 ± 11	Inactive	Inactive	>100,000	ND
47		160 ± 33	86 ± 7	Inactive	Inactive	18,000 ± 2,900	89 ± 7
48		4,400 ± 2,300	55 ± 7	Inactive	Inactive	>100,000	ND
49		510 ± 110	101 ± 7	4,200 ± 900	29 ± 1	>100,000	ND
50		Inactive	Inactive	Inactive	Inactive	Inactive	Inactive
51		Inactive	Inactive	Inactive	Inactive	18,000 ± 4,800	72 ± 4.6

<sup>a</sup> $\beta$ -arrestin recruitment activity was assessed as described in Figure 2.  $E_{\max}$  values are expressed as a percentage of the maximum dopamine response observed in the same assay.  $I_{\max}$  values are expressed as a percentage of the maximum inhibition of a dopamine (EC<sub>80</sub> concentration) response observed with the antagonist sulpiride in the same assay. <sup>b</sup>ND curve did not plateau.

nitrogen incorporation on the indole moiety afforded a set of 3-pyridyl analogues (71–76) with excellent to good D3R agonist potency (EC<sub>50</sub>s = 13–200 nM) and all but analogue 74 possessing EC<sub>50</sub> values <100 nM. However, as observed with the substituted indole set, none of these analogues were fully D3R > D2R-selective. Three other heterocyclic carboxamide analogues were tested, benzofuran 77, benzothiophene 78, and pyrrole 79, and all three were found to be potent D3R agonists. Contrary to the 4-methoxyphenyl ether series, in this set, 78 was found to be more potent (EC<sub>50</sub> = 13 nM) than 77 (EC<sub>50</sub> = 42 nM), further supporting that SAR trends might be dependent upon all of the groups in the molecule and a cautionary note against the extrapolation of SAR trends across all analogues. Pyrrole 79 was over 15-fold less potent (EC<sub>50</sub> = 118 nM) than the corresponding indole 99 (EC<sub>50</sub> = 7 nM) (Table 5). In line with all other 3-pyridyl ether analogues, these three compounds were not highly D3R > D2R-selective.

We hoped that the potency might be improved through further optimization of the 4-methoxyphenyl ether group. Of

the 35 carboxamide analogues in Table 2, the 2-indolyl analogue 20 was found to be by far the most potent and selective and remained our lead compound. Keeping the 2-indole carboxamide in place, we systematically explored replacements for the 4-methoxyphenyl ether (Table 5). The unsubstituted phenyl ether 80 was indeed more potent (EC<sub>50</sub> = 9 nM); however, it possessed modest D2R agonist and antagonist activities. The 3,4-methylenedioxyphenyl analogue 81 was a competent D3R agonist, though with slight D2R antagonism. The 4-methoxyphenyl thioether 82 was completely inactive across all three SAR assays, while the 4-thiomethoxyphenyl ether 83 was a modestly potent (EC<sub>50</sub> = 630 nM) and a fully selective D3R agonist. The limited data on oxygen-to-sulfur replacement potentially suggest a steric requirement of the ether oxygen for effective interaction with the D3R. This contrasts with the data for the  $\alpha$ -methyl ether analogue 47, which possessed increased potency over the desmethyl hit compound 1. Additional focused analogues and computational experiments would be necessary to further

**Table 4. Analogues Exploring Concurrent 4-Methoxyphenyl Ether and 4-Methoxybenzamide Replacement**

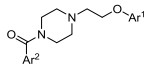
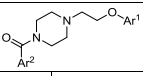
Compound ID			D3R agonist activity <sup>a</sup>		D2R agonist activity <sup>a</sup>		D2R antagonist activity <sup>a</sup>	
	Ar <sup>1</sup> =	Ar <sup>2</sup> =	EC <sub>50</sub> (nM)	E <sub>max</sub> (% control)	EC <sub>50</sub> (nM)	E <sub>max</sub> (% control)	IC <sub>50</sub> (nM)	I <sub>max</sub> (% control)
52	3-methoxyphen-1-yl	pyrrolo[2,3-b]pyridin-2-yl	50.0 ± 12	113 ± 2.8	128 ± 33	76 ± 8.3	Inactive	Inactive
53		pyrrolo[3,2-c]pyridin-2-yl	280 ± 19	111 ± 8	830 ± 170	71 ± 7	7,400 ± 3,700	30 ± 3
54		2-indolyl	33.8 ± 8.7	105 ± 9.3	2,300 nM ± 540	48 ± 5.1	>100,000	ND
55		5-methoxy-2-indolyl	284 ± 100	110 ± 6.5	1,700 ± 324	67 ± 15	Inactive	Inactive
56		benzo[d]imidazol-2-yl	285 ± 58	101 ± 12	660 ± 46	29 ± 7	2,000 ± 1,500	53 ± 10
57		4-methoxy-2-methylphen-1-yl	116 ± 29	102 ± 7.7	4,600 ± 1,100	52 ± 8.3	>100,000	ND
58	3-pyridyl	6-methoxy-2-indolyl	6.4 ± 1.5	107 ± 7	364 ± 58	81 ± 3.0	Inactive	Inactive
59		5-methoxy-2-indolyl	59 ± 23	97 ± 5	2,600 ± 1,200	97 ± 2.8	Inactive	Inactive
60		7-methyl-2-indolyl	25.7 ± 8.1	108 ± 6	786 ± 210	81 ± 8.5	Inactive	Inactive
61		6-methyl-2-indolyl	4.3 ± 2.0	106 ± 13	138 ± 42	74 ± 4.3	Inactive	Inactive
62		5-methyl-2-indolyl	13 ± 3.2	113 ± 8	1,300 ± 270	101 ± 5.1	Inactive	Inactive
63		4-methyl-2-indolyl	6.9 ± 1.7	97 ± 3	180 ± 23	70 ± 7.1	Inactive	Inactive
64		3-methyl-2-indolyl	80 ± 32	92 ± 13	800 ± 80	102 ± 7	Inactive	Inactive
65		1-methyl-2-indolyl	37.6 ± 7.2	112 ± 8	6,200 ± 1,900	53 ± 8.7	Inactive	Inactive
66		1-methyl-3-indolyl	9.5 ± 1.1	94 ± 9	4,400 ± 1,500	91 ± 17.2	Inactive	Inactive
67		7-chloro-2-indolyl	33 ± 4.3	113 ± 3.4	617 ± 144	74 ± 7.8	Inactive	Inactive
68		6-chloro-2-indolyl	3.0 ± 0.3	96 ± 5.5	183 ± 57	89 ± 0.4	Inactive	Inactive
69		5-chloro-2-indolyl	98 ± 45	106 ± 8	1,400 ± 282	90 ± 6.3	Inactive	Inactive
70		4-chloro-2-indolyl	4.4 ± 0.8	107 ± 8.4	217 ± 67	82 ± 6.5	Inactive	Inactive
71		pyrrolo[2,3-b]pyridin-2-yl	13.5 ± 6.5	98.6 ± 6.1	210 ± 83	99 ± 13.7	Inactive	Inactive
72		pyrrolo[2,3-c]pyridin-2-yl	81 ± 26.8	116 ± 9.7	780 ± 240	66 ± 7.0	Inactive	Inactive
73		pyrrolo[3,2-c]pyridin-2-yl	63 ± 20	108 ± 11	1,800 ± 460	61 ± 5.1	Inactive	Inactive
74		pyrrolo[3,2-b]pyridin-2-yl	200 ± 53	105 ± 8	3,000 ± 1,100	64 ± 2	Inactive	Inactive
75		benzo[d]imidazol-2-yl	33 ± 14.6	112 ± 9.8	2,400 ± 1,400	60.6 ± 4.3	Inactive	Inactive
76	1H-indazol-3-yl	45 ± 6.1	97 ± 8.5	4,100 ± 1,500	71 ± 2.8	Inactive	Inactive	

Table 4. continued

Compound ID			D3R agonist activity <sup>a</sup>		D2R agonist activity <sup>a</sup>		D2R antagonist activity <sup>a</sup>	
	Ar <sup>1</sup> =	Ar <sup>2</sup> =	EC <sub>50</sub> (nM)	E <sub>max</sub> (% control)	EC <sub>50</sub> (nM)	E <sub>max</sub> (% control)	IC <sub>50</sub> (nM)	I <sub>max</sub> (% control)
77		benzofuran-2-yl	42 ± 8.5	104 ± 9.5	1,700 ± 63	57 ± 7.4	Inactive	Inactive
78		benzothiophene-2-yl	13.3 ± 2.6	102 ± 7.7	617 ± 40	34 ± 3	2,100 ± 1,200	60 ± 4.9
79		pyrrol-2-yl	118 ± 47	110 ± 17.6	6,100 ± 1,300	34 ± 8.4	>100,000	ND

<sup>a</sup> $\beta$ -arrestin recruitment activity was assessed as described in Figure 2.  $E_{\max}$  values are expressed as a percentage of the maximum dopamine response observed in the same assay.  $I_{\max}$  values are expressed as a percentage of the maximum inhibition of a dopamine ( $EC_{80}$  concentration) response observed with the antagonist sulpiride in the same assay. <sup>b</sup>ND curve did not plateau.

elucidate the contributions of steric bulk and hydrogen-bond acceptor ability to overall analogue potency. We prepared a set of other 4-substituted phenyl ether analogues **84–92**. The 4-(trifluoromethoxy)phenyl ether **85** was a weakly potent ( $EC_{50}$  = 3800 nM) and a fully selective D3R agonist, whereas the corresponding 4-(trifluoromethoxy)phenyl ether on the 4-methoxyphenyl carboxamide scaffold (i.e., **6**) possessed no measurable activity. The 4-ethylphenyl ether analogue **87** possessed comparable D3R potency ( $EC_{50}$  = 1600 nM) to its counterpart on the 4-methoxyphenyl carboxamide scaffold, **4**. However, the 4-chlorophenyl ether analogue **90** did display a significant improvement to potency ( $EC_{50}$  = 114 nM) compared to its 4-methoxyphenyl carboxamide counterpart **5** ( $EC_{50}$  = 1000 nM). Other 4-substituted phenyl ethers also afforded potent and selective D3R agonists, such as the methyl-, fluoro-, and bromo-phenyl ether analogues (**86**, **89**, and **91**, respectively). The 4-ethoxyphenyl ether **84** and the 4-*tert*-butylphenyl ether **88** were completely inactive, possibly revealing a limit to the tolerated substituent size. A limited set of disubstituted phenyl ether analogues were explored (**93–97**), with the 2-chloro-4-methoxyphenyl ether **97** being the most potent ( $EC_{50}$  = 41 nM) and selective analogue among the group. Analogue **97** was only slightly less potent than the original 2-indolyl carboxamide **20**; however, the chlorine atom in **97** did not confer any additional benefit and we continued to focus on **20** for further evaluation. Other examples were notably less potent, including the 3-chloro-4-methylphenyl ether **95**. This is interesting in light of the potent D3R agonism of 4-methylphenyl analogue **86**, further suggesting size limitations of the aryl ether. A set of four pyridyl ether analogues were evaluated, ranging from the very potent 5-chloro-3-pyridyl ether **101** ( $EC_{50}$  = 5 nM) to the weakly potent 2-pyridyl ether **98** ( $EC_{50}$  = 3500 nM). While not an exhaustive set, the potent analogues identified (i.e., **99** and **100**) lacked full D3R > D2R selectivity and only the 4-pyridyl ether **100** was found to be a selective D3R agonist of modest potency ( $EC_{50}$  = 472 nM).

Having earlier noted a slight potency improvement from methyl incorporation adjacent to the aryl ether (i.e., **47**), we tested the effect on two such 2-indole carboxamide exemplars (Table 5). We observed a contradictory effect between the two cases: while the effect was slightly detrimental in the 4-methoxyphenyl ether analogue **102**, it provided a 3-fold potency enhancement for the 4-chlorophenyl ether analogue **103**. In both examples, however, the analogues also exhibited D2R agonist activity. It may be worth noting that methyl incorporation onto the tether afforded analogues that were

racemic mixtures. Separation and testing of the enantiopure analogues could markedly affect the potency and/or selectivity and may be pursued in future SAR investigations. Replacement of the ether with an *N*-methyl tertiary amine afforded the modestly potent ( $EC_{50}$  = 1900 nM) though D3R-selective analogue **104**. The 5-indole ether **105** was found to be a potent D3R agonist ( $EC_{50}$  = 61 nM) as well as a reasonably potent D2R antagonist ( $IC_{50}$  = 440 nM). Although this particular compound does not meet the criteria of a selective D3R agonist sought in the current study, **105** may be of particular interest as a lead in complementary studies examining D3R stimulation and, simultaneously, D2R antagonism.

Finally, within the context of the 2-indole carboxamide analogues, we made additional modifications to the linker between the 2-indole carboxamide and 4-methoxyphenyl moieties. Compounds **106** and **107** have one or two additional methylene groups, respectively, inserted between the piperazine and 4-methoxyphenyl ether (Table 5). These modifications lead to a complete loss of D3R agonist activity, although both compounds retained low potency D2R antagonist activity. In **108**, the piperazine ring was converted to a piperidine ring (Table 5), which, similar to that observed with compound **50** (Table 3), led to a complete loss in D3R and D2R activities, further highlighting the importance of this nitrogen for receptor activity. Finally, we made analogue **109** in which a methylene group was inserted between the 2-indolylcarboxamide and piperazine moieties, which lead to a complete loss of both D3R agonist and D2R antagonist activities (Table 5). Taken together, these results highlight the importance of linker length, especially for D3R agonist potency and efficacy.

In summary, we explored SAR trends for both termini of compound **1** and, to a lesser extent, modifications on the core piperazine and tether. We have identified a number of potent and selective D3R agonists with the greater challenge being to maintain the D3R receptor selectivity without gaining D2R activity. The most potent and selective D3R agonist identified was the 2-indole carboxamide analogue **20**, containing the 4-methoxyphenyl ether found in hit compound **1**. The closely related but slightly less potent 2-chloro-4-methoxyphenyl ether **97** supports the chemotype as a valid D3R agonist scaffold. Therefore, we chose to further investigate compound **20** as a D3R-selective agonist and have designated it as NIH Molecular Libraries Initiative probe molecule ML417 (Figure 4A).

**Pharmacological Characterization of Lead Compound 20.** Our lead compound **20** was subjected to further

Table 5. Analogues Based on the 2-Indolyl Carboxamide Scaffold

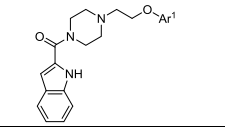
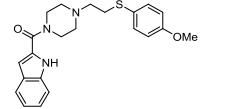
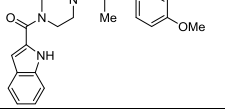
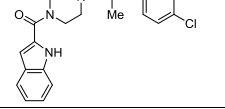
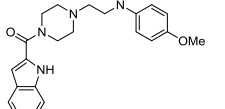
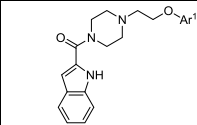
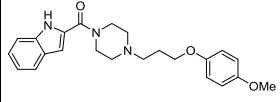
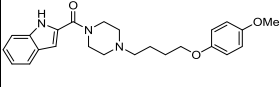
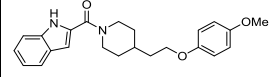
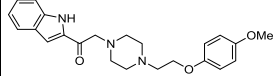
Compound ID		D3R agonist activity <sup>a</sup>		D2R agonist activity <sup>a</sup>		D2R antagonist activity <sup>a</sup>	
		EC <sub>50</sub> (nM)	E <sub>max</sub> (% control)	EC <sub>50</sub> (nM)	E <sub>max</sub> (% control)	IC <sub>50</sub> (nM)	I <sub>max</sub> (% control)
80	phenyl	9.2 ± 0.6	83 ± 3.4	1,800 ± 1,100	49 ± 6.3	535 ± 220	37 ± 8
81	3,4-(methylenedioxy)phenyl	290 ± 130	82 ± 5.2	7,000 ± 1,700	36 ± 8.8	>50,000	ND
82		Inactive	Inactive	Inactive	Inactive	>100,000	ND
83	4-thiomethoxyphen-1-yl	630 ± 149	89 ± 9.3	Inactive	Inactive	>100,000	ND
84	4-ethoxyphen-1-yl	>100,000	ND	Inactive	Inactive	>100,000	ND
85	4-(trifluoromethoxy)phen-1-yl	3,800 ± 770	100 ± 16.7	>100,000	ND	>100,000	ND
86	4-methylphen-1-yl	151 ± 30	91 ± 16.6	>50,000	ND	>100,000	ND
87	4-ethylphen-1-yl	1,600 ± 640	98 ± 16	Inactive	Inactive	>50,000	ND
88	4- <i>tert</i> -butylphen-1-yl	Inactive	Inactive	Inactive	Inactive	>100,000	ND
89	4-fluorophen-1-yl	126 ± 35	90 ± 13.3	>50,000	ND	9,700 ± 5,800	72 ± 4.6
90	4-chlorophen-1-yl	114 ± 27	118 ± 10	Inactive	Inactive	>50,000	ND
91	4-bromophen-1-yl	105 ± 29	106 ± 21	Inactive	Inactive	>100,000	ND
92	4-nitrophen-1-yl	1,300 ± 280	92 ± 9.7	Inactive	Inactive	Inactive	Inactive
93	3,4-dimethoxyphen-1-yl	710 ± 121	111 ± 18	Inactive	Inactive	>50,000	ND
94	3,4-dimethylphenyl	610 ± 37	70 ± 8.1	Inactive	Inactive	13,000 ± 1,600	88 ± 9.4
95	3-chloro-4-methylphen-1-yl	997 ± 300	103 ± 12	Inactive	Inactive	Inactive	Inactive
96	4-chloro-3-methylphen-1-yl	114 ± 20	110 ± 1.3	Inactive	Inactive	3,800 ± 1,100	75 ± 3
97	2-chloro-4-methoxyphen-1-yl	41 ± 12	113 ± 3.5	Inactive	Inactive	>10,000	ND
98	2-pyridyl	3,500 ± 900	94 ± 17	Inactive	Inactive	>100,000	ND
99	3-pyridyl	7.0 ± 1.8	98 ± 7	570 ± 220	70 ± 6	Inactive	Inactive
100	4-pyridyl	472 ± 39	71 ± 6.3	Inactive	Inactive	>100,000	ND
101	5-chloropyridin-3-yl	5.0 ± 1.2	79 ± 9.3	790 ± 220	56 ± 2.5	Inactive	Inactive
102		82 ± 7.8	108 ± 6.4	3,300 ± 1,300	46 ± 7.6	Inactive	Inactive
103		32 ± 8.1	101 ± 4.1	1,900 ± 570	43 ± 4.9	Inactive	Inactive
104		1,900 ± 47	45 ± 3	Inactive	Inactive	>100,000	ND

Table 5. continued

Compound ID		D3R agonist activity <sup>a</sup>		D2R agonist activity <sup>a</sup>		D2R antagonist activity <sup>a</sup>	
		EC <sub>50</sub> (nM)	E <sub>max</sub> (% control)	EC <sub>50</sub> (nM)	E <sub>max</sub> (% control)	IC <sub>50</sub> (nM)	I <sub>max</sub> (% control)
105	5-indolyl	60.9 ± 20.6	84 ± 5.8	Inactive	Inactive	440 ± 138	100 ± 0
106		Inactive	Inactive	Inactive	Inactive	691 ± 75	97 ± 3
107		Inactive	Inactive	Inactive	Inactive	1200 ± 357	97 ± 6
108		Inactive	Inactive	Inactive	Inactive	Inactive	Inactive
109		Inactive	Inactive	Inactive	Inactive	Inactive	Inactive

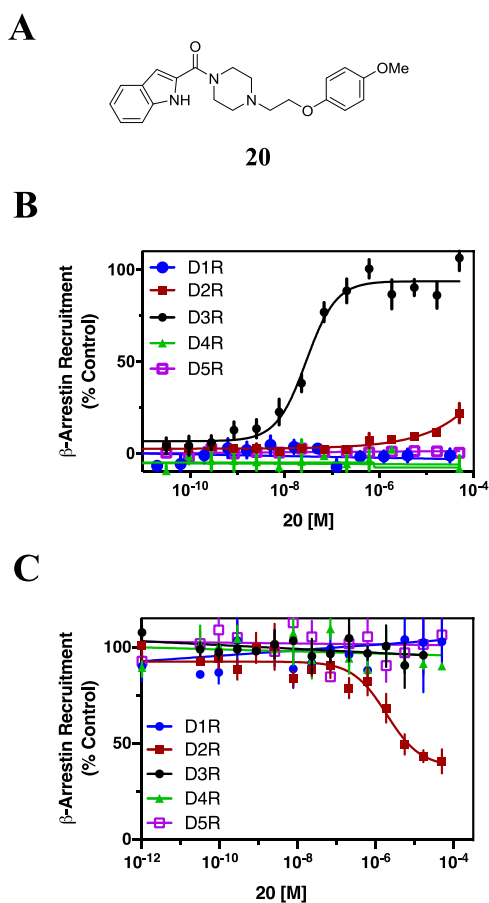
<sup>a</sup> $\beta$ -arrestin recruitment activity was assessed as described in Figure 2.  $E_{\max}$  values are expressed as a percentage of the maximum dopamine response observed in the same assay.  $I_{\max}$  values are expressed as a percentage of the maximum inhibition of a dopamine ( $EC_{80}$  concentration) response observed with the antagonist sulpiride in the same assay. <sup>b</sup>ND curve did not plateau.

characterization using a variety of pharmacological assays. We initially wanted to assess the selectivity of compound **20** among all dopamine receptor subtypes using the  $\beta$ -arrestin recruitment functional assay. As shown in Figure 4B, **20** is a full agonist at the D3R with an  $EC_{50}$  of 38 nM while displaying minimal agonist efficacy at all other receptor subtypes. When tested as an antagonist, **20** displayed very limited activity ( $IC_{50} > 50 \mu M$ ) at all subtypes except for the D2R where it exhibited some weak partial antagonism ( $IC_{50} > 10 \mu M$ ; Figure 4C). This is in contrast to the parent hit compound **1** that showed full antagonism at the D2R, indicating that our medicinal chemistry efforts increased the potency of the compound for the D3R and decreased its activity at the D2R resulting in a potent and highly selective agonist of the D3R.

The activity of **20** on  $\beta$ -arrestin recruitment to the D3R was confirmed using an orthogonal assay that measures bioluminescence resonance energy transfer (BRET) when the D3R and  $\beta$ -arrestin, fused to complimentary biosensors (see the Experimental Section), are in close proximity. As shown in Figure 5A using the BRET-based  $\beta$ -arrestin recruitment assay, **20** displayed full agonist activity at the D3R with an  $EC_{50}$  of 1.2 nM. These results confirm those obtained using the DiscoverX  $\beta$ -arrestin recruitment assay (Figure 4B), although **20** exhibited an  $\sim 30$ -fold greater potency using the BRET-based assay. The reason for this discrepancy is not clear but may be related to differing assay time courses, different cell types, and/or varying levels and types of G protein-coupled receptor kinases (GRKs) that can affect  $\beta$ -arrestin recruitment to GPCRs.<sup>56</sup> Regardless, these findings confirm that **20** is a full and potent agonist for  $\beta$ -arrestin recruitment. We next sought to characterize the effects of **20** on G protein-mediated signaling. Initially, we used another BRET-based assay to examine D3R coupling to Go, an inhibitory G protein that the D3R has previously been shown to activate.<sup>57,58</sup> As shown in

Figure 5B, **20** functioned as a full agonist in the D3R-mediated Go-BRET assay with an  $EC_{50}$  of 0.18 nM. The higher potency of **20** in this assay is likely due to spare receptors resulting in signal amplification in contrast to the  $\beta$ -arrestin recruitment assays, which lack amplification.<sup>59</sup> We next wished to examine G protein-mediated second messenger modulation and turned to a BRET-based cAMP assay using the Epac-based biosensor, CAMYEL.<sup>60–62</sup> Figure 5C shows that **20** potently inhibits cAMP accumulation with an  $EC_{50}$  of 86 nM and an efficacy identical to that of dopamine. Finally, we examined the activity of **20** in an ERK1/2 phosphorylation (pERK) assay (Figure 5D). GPCR-mediated phosphorylation of ERK1/2 has previously been shown to occur through G protein-dependent and -independent signaling pathways,<sup>63,64</sup> which may be related to the GPCR and/or cell type. However, we have found that the pretreatment of our cells with pertussis toxin, which inactivates  $G_{\alpha i/o}$  proteins,<sup>65,66</sup> completely ablates the D3R-mediated pERK response indicating that it is G protein-mediated (data not shown). Similar to the other signaling assays, we found that **20** functioned as a full agonist at the D3R with an  $EC_{50}$  of 21 nM (Figure 5D). Taken together, these data demonstrate that **20** is a full and potent agonist for multiple signaling pathways associated with D3R activation. Notably, in these signaling assays, there are small variances in the potencies of **20** relative to those for dopamine, suggesting that **20** may exhibit biased signaling properties; however, this will need to be evaluated in detail in future experiments.

To further characterize the pharmacological selectivity of **20**, we evaluated its activity in large arrays of GPCRs and several transporters or ion channels. For comparison, we also evaluated the recently described D3R-selective agonist CJ-1639<sup>45</sup> and its parent scaffold, the D3R-preferring agonist pramipexole. Initially, we screened the compounds using the NIMH Psychoactive Drug Screening Program (PDSP),<sup>67</sup>



**Figure 4.** Pharmacological activity of compound **20** on all dopamine receptor subtypes. (A) Chemical structure of **20**. (B) Agonist concentration–response curves for stimulating  $\beta$ -arrestin recruitment in response to **20** for the indicated DA receptor subtypes.  $\beta$ -arrestin recruitment was assessed using the DiscoverX assay as described in the Experimental Section, and the data are expressed as a percentage of the maximum dopamine signal for each receptor (not shown) and represent means  $\pm$  SEM of at least three experiments performed in triplicate. The  $EC_{50}$  for **20** at the D3R is reported in Table 2. (C) Antagonist concentration–response curves of **20** on cells expressing individual dopamine receptor subtypes as indicated.  $\beta$ -arrestin recruitment was stimulated with an  $\sim EC_{80}$  concentration of dopamine for each receptor subtype and incubated with the indicated concentrations of **20**. Data are expressed as a percentage of the signal seen with the  $EC_{80}$  concentration of dopamine and represent means  $\pm$  SEM of at least three experiments performed in triplicate. The  $IC_{50}$  for **20** at the D2R was estimated to be  $>10 \mu M$ .

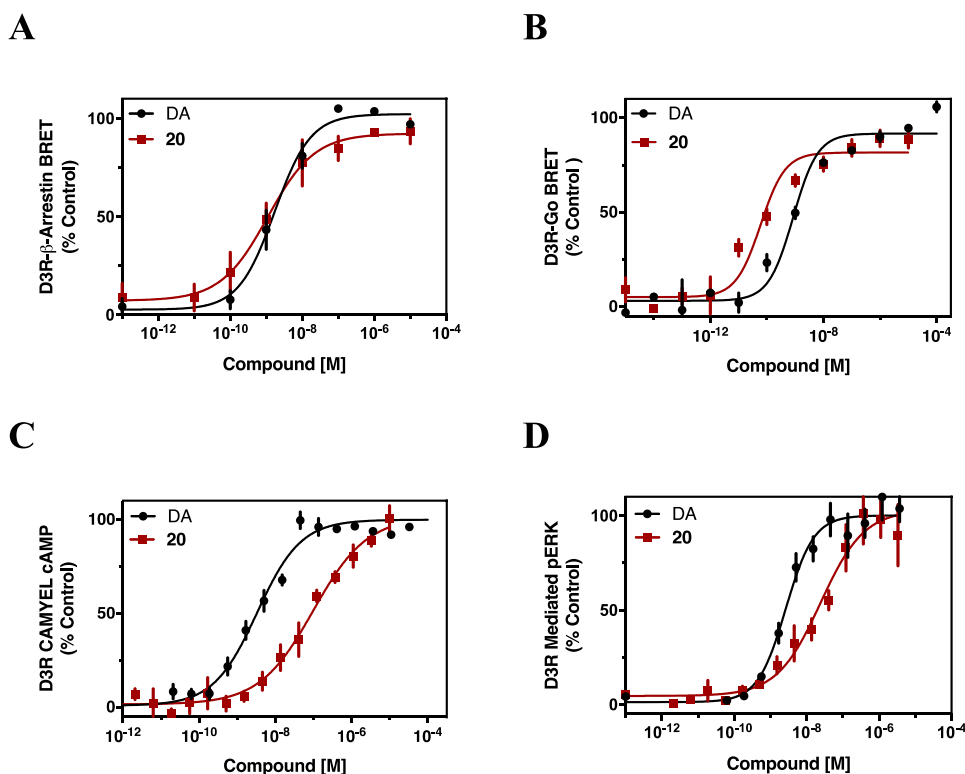
which uses radioligand binding assays to assess affinity values for ligands at 45 unique GPCRs, transporters, and ion channels. The results of this screen are shown in Table 6. For the primary screen, a single high concentration ( $10 \mu M$ ) of test compound was used to inhibit radioligand binding to the targets, and those compounds that exhibited  $>50\%$  inhibition were rescreened in full concentration–response format to estimate their affinity ( $K_i$ ) values. Notably, **20** exhibited submicromolar affinity for only three of the targets, the  $\beta_1$ -adrenergic, 5-HT<sub>2B</sub> serotonergic, and  $\sigma$ -1 receptors. Pramipexole and CJ-1639 exhibited submicromolar affinity for 6 and 13 of the targets, respectively. Among the D2-like receptors, pramipexole was 826-fold D3R  $>$  D2R-selective and 32-fold D3R  $>$  D4R-selective. In parallel studies in our laboratory, we found that pramipexole inhibits [<sup>3</sup>H]-methylspiperone binding

to the D3R with a  $K_i$  of  $8.2 \pm 2.0$  nM (mean  $\pm$  SEM,  $n = 3$ ) and to the D2R with a  $K_i$  of  $4.7 \pm 1.3 \mu M$  (mean  $\pm$  SEM,  $n = 3$ ), exhibiting 573-fold D3R  $>$  D2R selectivity in good agreement with the PDSP data. In the PDSP screen, CJ-1639 exhibited 130-fold D3R  $>$  D2R selectivity and 1.5-fold D3R  $>$  D4R selectivity. In our laboratory, we found that CJ-1639 inhibits [<sup>3</sup>H]-methylspiperone binding to the D3R with a  $K_i$  of  $5.6 \pm 1.0$  nM (mean  $\pm$  SEM,  $n = 3$ ) and to the D2R with a  $K_i$  of  $21 \pm 5.6 \mu M$  (mean  $\pm$  SEM,  $n = 3$ ), exhibiting 3750-fold D3R  $>$  D2R selectivity. Thus, in our hands, CJ-1639 is significantly more D3R  $>$  D2R-selective than was observed in the PDSP screen and closer to the D3R  $>$  D2R selectivity initially described by Chen et al.<sup>45</sup>

Notably, in the PDSP screen, **20** exhibited a  $K_i$  for the D3R of  $1.24 \mu M$ . Thus, as was initially observed with compound **1**, when assessed using [<sup>3</sup>H]-methylspiperone binding in membrane preparations, **20** exhibits an affinity for the D3R that is much weaker than its potency observed using any of the functional assays, even those that lack amplification. In contrast, the agonists pramipexole and CJ-1639 both display similar D3R binding affinities and functional potencies. For instance, when we examined pramipexole and CJ-1639 in the D3R-mediated  $\beta$ -arrestin recruitment assay, we found  $EC_{50}$  values of  $5.4 \pm 1.5$  nM (mean  $\pm$  SEM,  $n = 3$ ) and  $9.3 \pm 1.5$  nM (mean  $\pm$  SEM,  $n = 3$ ), respectively, in good agreement with their affinities observed in the binding assays (see above).

Given the above observations, we decided to investigate the effects of assay buffer conditions on the radioligand binding results (Table 7). Our standard binding buffer is Earle's balanced salt solution (EBSS), which is an isotonic bicarbonate–phosphate buffer containing 117 mM NaCl, 5.37 mM KCl, 1.8 mM CaCl<sub>2</sub>, 0.8 mM MgSO<sub>4</sub>, and 5.55 mM glucose. In addition to EBSS, we utilized Tris buffer with no additional salts, as well as Tris plus 140 mM NaCl, and performed [<sup>3</sup>H]-methylspiperone competition binding with **20** and pramipexole (Table 7). Interestingly, **20** was  $\sim 10$ -fold more potent using the Tris buffer compared to EBSS, whereas **20**'s potency decreased by  $\sim 4$ -fold when 140 mM NaCl was added to the Tris buffer (Tris + Na<sup>+</sup>). Notably, the [<sup>3</sup>H]-methylspiperone binding buffer used by the PDSP is 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid (HEPES) plus 50 mM NaCl and 5 mM MgCl<sub>2</sub>, more similar to that of EBSS with respect to the concentrations of Na<sup>+</sup> and divalent cations. Not surprisingly, the potency of **20** is similar when comparing the EBSS ( $K_i = 3.7 \mu M$ ) and HEPES ( $K_i = 1.24 \mu M$ ) buffer systems. In contrast to **20**, the potency of pramipexole appeared to be relatively unaffected by the assay buffer conditions (Table 7), as is [<sup>3</sup>H]-methylspiperone (data not shown).

Typically, agonist ligands appear more potent when competing for radiolabeled agonist binding as the binding sites represent the high-affinity active G protein-coupled state of the GPCR.<sup>68–70</sup> As such, we repeated our competition binding experiments using [<sup>3</sup>H]-7-OH-DPAT, which is an agonist of the D3R (Table 7). The same potency trends were observed with the various buffer systems, although both **20** and pramipexole appeared more potent using [<sup>3</sup>H]-7-OH-DPAT compared with [<sup>3</sup>H]-methylspiperone, as was expected. In addition, we noted that the addition of 5 mM MgCl<sub>2</sub> to Tris buffer (Tris + Mg<sup>2+</sup>) increased the potency of **20** by  $\sim 2$ -fold. These observations are in line with previous results showing that Na<sup>+</sup> typically decreases agonist binding affinity, likely through stabilizing the inactive state of the GPCR,<sup>71,72</sup> while



**Figure 5.** Pharmacology of compound **20** on D3R-mediated signaling outputs. Agonist-mediated concentration–response curves of dopamine or **20** for stimulating a variety of D3R-mediated signaling pathways. (A) Cells were stimulated as indicated and analyzed using the BRET-based  $\beta$ -arrestin recruitment assay (see the [Experimental Section](#)), resulting in  $EC_{50}$  values of  $2.3 \pm 0.9$  and  $1.2 \pm 0.5$  nM (mean  $\pm$  SEM,  $n = 4$ ) for dopamine and **20**, respectively. (B) Cells were stimulated as indicated and analyzed using the BRET-based Go activation assay as described in the [Experimental Section](#), resulting in  $EC_{50}$  values of  $1.1 \pm 0.04$  and  $0.18 \pm 0.1$  nM (mean  $\pm$  SEM,  $n = 6$ ) for dopamine and **20**, respectively. (C) Cells were incubated with the indicated concentrations of dopamine (DA) or **20**, and inhibition of forskolin-stimulated cAMP accumulation was determined using the CAMYEL biosensor as described in the [Experimental Section](#). Data are presented as the percentage of maximum inhibition by DA. **20** displays a full agonist activity of D3R-mediated inhibition of cAMP production, exhibiting an  $EC_{50}$  of  $86 \pm 26$  nM (mean  $\pm$  SEM,  $n = 4$ ). Dopamine demonstrated an  $EC_{50}$  value of  $3.5 \pm 0.7$  nM (mean  $\pm$  SEM,  $n = 4$ ). (D) Cells were stimulated as indicated, and pERK was assessed using the Alphascreen SureFire assay as described in the [Experimental Section](#).  $EC_{50}$  values for dopamine and **20** are  $2.9 \pm 0.6$  and  $21 \pm 6.6$  nM (mean  $\pm$  SEM,  $n = 4$ ), respectively. All data were analyzed using nonlinear regression curve fits and expressed as a percentage of the maximum dopamine signal.

$Mg^{2+}$  increases agonist binding affinity through promoting GPCR–G protein interactions.<sup>72,73</sup> Taken together, these results suggest that the apparent binding affinity of **20** for the D3R in membrane preparations can vary by as much as 300-fold depending on the buffer constituents and radioligand utilized. Moreover, the potency of **20** using intact cell functional assays ( $EC_{50}$ s = 0.18–86 nM) is more closely aligned with its affinity for the active signaling state of the D3R as detected using [<sup>3</sup>H]-7-OH-DPAT membrane binding, as previously observed for multiple GPCRs.<sup>68–70</sup>

We extended our selectivity screening to the DiscoverX gpcrMAX functional assay, which measures agonist-stimulated  $\beta$ -arrestin recruitment to 168 known GPCRs (<http://www.DiscoverX.com>). We chose to perform this screen using a high concentration (10  $\mu$ M) of **20** to maximize the detection of off-target activities. [Figure 6A](#) shows the results of this screen performed in agonist mode where each GPCR is stimulated with **20**, and  $\beta$ -arrestin recruitment is then measured. Notably, the D3R is the only GPCR in this array that was fully activated, although small, but measurable agonist responses were observed with the short and long isoforms of the D2R. These latter activities were comparable to the low activity of **20** at the D2R observed in [Figure 4B](#) using 10  $\mu$ M and higher concentrations. [Figure 6B](#) shows the results of this screen in

antagonist mode and that **20** exhibits low potency partial antagonist activity at several GPCRs. These include the Epstein–Barr virus-induced GPCR 2 (EBI2), the sphingosine-1-phosphate 5 receptor (EDG8), the cholecystokinin A receptor (CCKAR), the alpha2C-adrenergic receptor (ADRA2C), and the 5HT<sub>2A</sub> serotonergic receptor (HTR2A). Antagonism of these receptors is not associated with any known clinical side effects and, in any case, such interactions would not occur at the nanomolar concentrations employed to selectively activate the D3R. Importantly, **20** was identified as exhibiting submicromolar affinity for the 5-HT<sub>2B</sub> receptor in the PDSP screen ([Table 6](#)), which is a potential liability as drugs that activate the 5-HT<sub>2B</sub> receptor have been associated with cardiac valvulopathy.<sup>74,75</sup> Unfortunately, the DiscoverX gpcrMAX panel does not include the 5-HT<sub>2B</sub> receptor, so this screen did not provide us with a functional profile of **20** at this receptor subtype. Thus, in separate experiments, we assessed the effects of **20** (10  $\mu$ M) on 5-HT<sub>2B</sub> receptor stimulation of inositol 1-phosphate (IP1) accumulation (see the [Experimental Section](#)). Fortunately, no agonist activity was observed, but this concentration of **20** produced an 80% inhibition of the response elicited by 30 nM serotonin (data not shown), suggesting that **20** may be a low potency antagonist of the 5-HT<sub>2B</sub> receptor at high micromolar

**Table 6. Binding Affinities<sup>a</sup> of 20, CJ-1639, and Pramipexole for Inhibiting Radioligand Binding to the Indicated Drug Targets as Determined in the PDSP Screen**

target	compound			target	compound		
	20 ( $K_i$ , nM)	CJ-1639 ( $K_i$ , nM)	Pramipexole ( $K_i$ , nM)		20 ( $K_i$ , nM)	CJ-1639 ( $K_i$ , nM)	Pramipexole ( $K_i$ , nM)
SHT1A	2108	708	6514	D3	1240	30	0.9
SHT1B	NA	NA	3508	D4	NA	45	29
SHT1D	NA	NA	>10 000	D5	NA	NA	>10 000
SHT1E	NA	NA	>10 000	DAT	NA	205	NA
SHT2A	NA	2841	NA	DOR	NA	NA	>10 000
SHT2B	674	1178	NA	GABA <sub>A</sub>	NA	NA	ND
SHT2C	5997	762	NA	H1	NA	110	NA
SHT3	NA	NA	>10 000	H2	NA	224	2683
SHT5A	NA	NA	>10 000	H3	NA	893	NA
SHT6	NA	NA	>10 000	H4	NA	NA	>10 000
SHT7	NA	770	1188	KOR	NA	>10 000	NA
Alpha1A	NA	NA	>10 000	M1	NA	NA	>10 000
Alpha1B	NA	666	NA	M2	NA	NA	>10 000
Alpha1D	NA	1184	NA	M3	NA	NA	>10 000
Alpha2A	>10 000	NA	75.7	M4	NA	2279	NA
Alpha2B	NA	NA	67.7	M5	NA	2297	NA
Alpha2C	2841	NA	52.2	MOR	NA	745	NA
Beta 1	77	NA	NA	NET	NA	436	NA
Beta 2	NA	ND	>10 000	PBR	NA	NA	ND
Beta 3	NA	NA	>10 000	SERT	NA	32	NA
BZP site	NA	NA	ND	Sigma 1	383	531	4446
D1	NA	NA	>10 000	Sigma 2	2750	NA	NA
D2	NA	3902	743.7				

<sup>a</sup> $K_i$  (nM) values for the indicated compounds were determined as described in the [Experimental Section](#). <sup>b</sup>NA indicates that the inhibition of binding was < 50% in the primary assay conducted using a single 10  $\mu$ M concentration. <sup>c</sup>ND indicates that the value was not determined.

**Table 7. Affinities<sup>a</sup> of 20 and Pramipexole for Inhibiting Either [<sup>3</sup>H]-Methylspiperone or [<sup>3</sup>H]-7-OH-DPAT Binding to the D3R in Various Buffer Systems**

	[ <sup>3</sup> H]-methylspiperone $K_i$ (nM)			[ <sup>3</sup> H]-7-OH-DPAT $K_i$ (nM)			
	EBSS	Tris	Tris + Na <sup>+</sup>	EBSS	Tris	Tris + Na <sup>+</sup>	Tris + Mg <sup>2+</sup>
20	3700 $\pm$ 900	350 $\pm$ 130	1500 $\pm$ 370	720 $\pm$ 31	22 $\pm$ 3.7	94 $\pm$ 11	12.5 $\pm$ 3.8
pramipexole	8.2 $\pm$ 2.0	13 $\pm$ 0.8	17 $\pm$ 0.6	1.1 $\pm$ 0.3	0.94 $\pm$ 0.24	5.6 $\pm$ 1.3	ND

<sup>a</sup> $K_i$  (nM) values for the indicated compounds were determined as described in the [Experimental Section](#). <sup>b</sup>Data are expressed as mean  $\pm$  SEM of individual experiments performed 3–16 times. <sup>c</sup>ND indicates that the value was not determined.

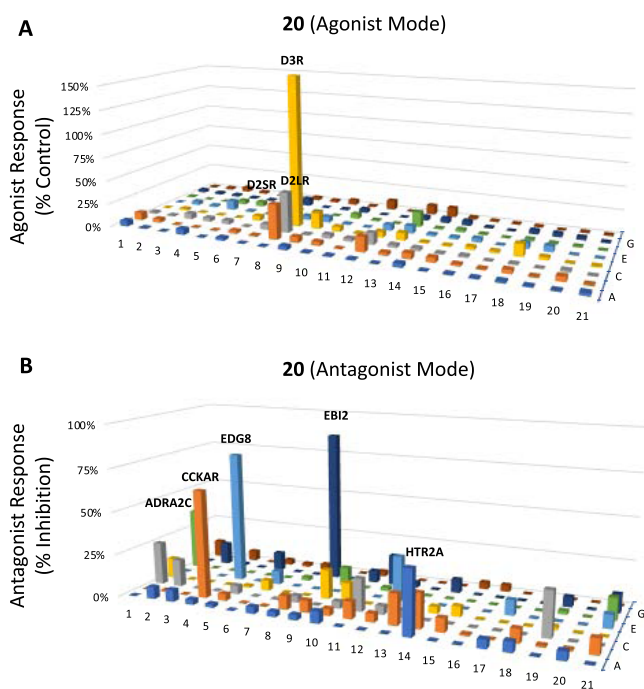
concentrations. Taken together, these findings indicate that **20** is a selective D3R agonist with very limited cross-reactivity at other GPCRs.

For comparison with **20**, the D3R-preferring agonists CJ-1639 and pramipexole were also screened in the DiscoverX gpcrMAX panel. [Figure S1A](#) shows that, at 10  $\mu$ M, CJ-1639 is a full agonist of both the D2R and D3R, with partial agonist activities at the D4R and the chemokine CXCR7 receptor. Conversely, at 10  $\mu$ M, CJ-1639 displays >50% antagonist activity at the  $\alpha_{1B}$ -adrenergic receptor, H<sub>1</sub>, H<sub>2</sub>, and H<sub>3</sub> histamine receptors, and the  $\mu$  opioid receptor ([Figure S1B](#)). Pramipexole also exhibits full agonist activity at the D2R and D3R, as well as partial agonist activity at the  $\alpha_{2A}$ - and  $\alpha_{2B}$ -adrenergic receptors ([Figure S2A](#)). Interestingly, pramipexole did not exhibit antagonist activity at any of the GPCRs tested, even at the 10  $\mu$ M concentration ([Figure S2A](#)). Overall, using these assays, CJ-1639 and pramipexole appear less globally selective than **20**, although they may exhibit greater D3R selectivity if lower screening concentrations are employed.

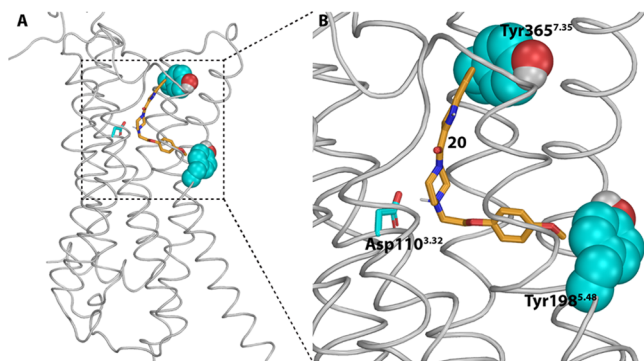
**Molecular Modeling Predicts Unique Interactions of Compound 20 with the D3R.** To characterize the binding pose of **20**, we carried out a computational modeling and

simulation study of a D3R model in complex with **20**. We first docked **20** into a D3R model that we have equilibrated previously<sup>76</sup> and found that the majority of the resulting poses of **20** are with the indole amide moiety pointing away from the orthosteric binding site (OBS, defined in ref 77), similar to other indole amide D3R ligands that we have characterized previously.<sup>6,78</sup> Using three subtly different poses in such an orientation with the lowest docking scores, we collected three molecular dynamics (MD) simulation trajectories. The simulations converged to a **20** pose, in which at one end its anisole moiety binds in the OBS and protrudes into the interface between TM5 and TM6 and interacts with Tyr198<sup>5,48</sup> (superscripts denote Ballesteros and Weinstein numbering),<sup>79</sup> while at the other end its indole amide moiety interacts with Tyr365<sup>7,35</sup> ([Figure 7B](#)). Further, the positively charged piperazine nitrogen of **20** closest to the anisole moiety was found to interact with Asp110<sup>3,32</sup> ([Figure 7B](#)), the highly conserved aspartic acid residue found in TM3 of all biogenic amine GPCRs.<sup>77</sup> Given this observation, it is not surprising that compounds either lacking this nitrogen (**50**) or with immediately adjacent methyl groups (**46** and **48**) that could





**Figure 6.** Functional profiling of compound **20** against an array of 168 known GPCRs. A single high concentration (10  $\mu$ M) of **20** was screened against the DiscoverX gpcrMAX assay panel in both agonist (A) and antagonist (B) modes, as described in the [Experimental Section](#). Data represent the percent maximum stimulation observed by a reference agonist for each GPCR (agonist mode) or the percent inhibition of a response generated by an  $EC_{80}$  concentration of a reference agonist (antagonist mode). A complete key to the GPCR array and numerical results are provided in the [Supplemental Table S1](#). In (B), partial antagonist activity was observed at the ADRA2C (36%), CCKAR (63%), EDG8 (77%), EB12 (88%), and HTR2A (37%) receptors using a 10  $\mu$ M concentration of **20**.



**Figure 7.** Molecular modeling identified residues that uniquely interact with **20**. (A) Side view of a representative D3R model in complex with **20** resulting from our MD simulations. The model was based on the D3R crystal structure (PDB: 3PBL).<sup>6</sup> (B) Zoom-in view of the ligand binding pocket showing that two Tyr residues, Tyr198<sup>5,48</sup> and Tyr365<sup>7,35</sup>, uniquely interact with the two ends of the ligand. The conserved TM3 aspartate residue, Asp110<sup>3,32</sup>, in contact with the basic piperazine nitrogen is also depicted.

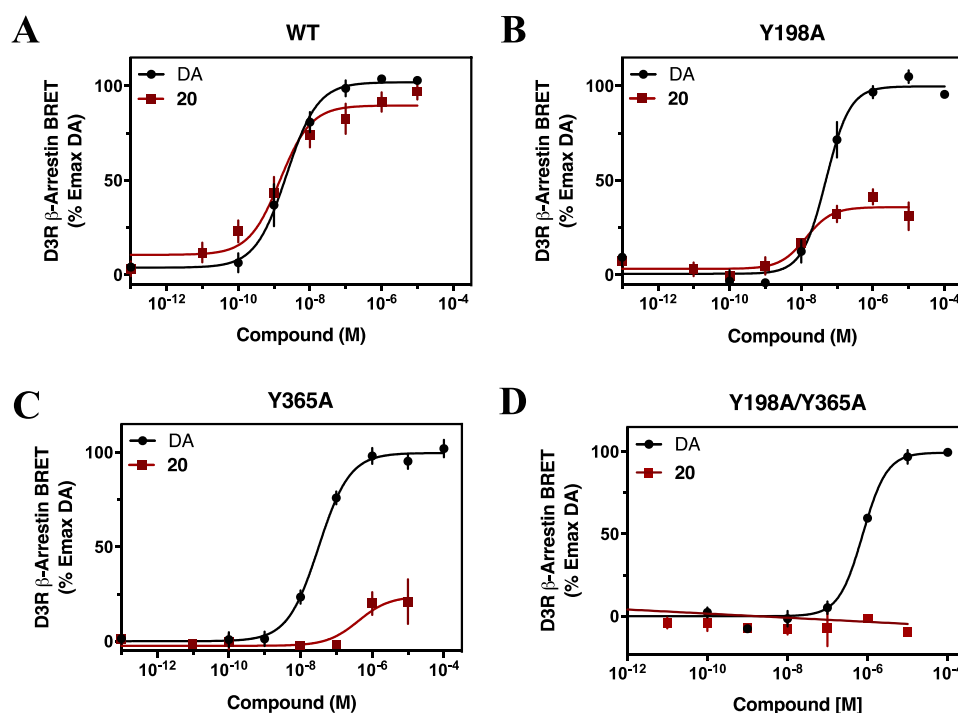
produce steric hindrance exhibited greatly reduced or no activity for interacting with the D3R (Table 3).

We noticed that the two tyrosine residues, Tyr198<sup>5,48</sup> and Tyr365<sup>7,35</sup>, in contact with **20** cannot directly interact with dopamine.<sup>57</sup> Thus, to validate the predicted binding pose of **20** at D3R, we mutated Tyr198<sup>5,48</sup> and Tyr365<sup>7,35</sup> to alanine

residues either individually or in combination and compared the pharmacological profiles of dopamine and **20** at these mutants. The mutant D3R constructs were found to express similarly to D3R-WT as determined using [<sup>3</sup>H]-methylspiperone saturation binding assays ( $B_{max}$  values (fmol/mg): WT =  $3700 \pm 650$ , Y198A =  $4300 \pm 1000$ , Y365A =  $3000 \pm 500$ , Y198A/Y365A =  $2700 \pm 930$ ;  $K_d$  values (nM): WT =  $0.51 \pm 0.08$ , Y198A =  $0.53 \pm 0.06$ , Y365A =  $0.49 \pm 0.11$ , Y198A/Y365A =  $0.35 \pm 0.1$ ; mean  $\pm$  SEM from three independent experiments). We then compared the ability of compound **20** and dopamine to stimulate  $\beta$ -arrestin recruitment to these constructs. Figure 8A shows that they stimulate  $\beta$ -arrestin recruitment to the D3R-WT with similar potencies ( $EC_{50}$  = 1.4 and 3.7 nM for **20** and dopamine, respectively). The Y198A mutation resulted in a reduction in efficacy ( $E_{max}$  = 38%) and a 13-fold reduction in the potency ( $EC_{50}$  = 17 nM) of **20** for stimulating  $\beta$ -arrestin recruitment (Figure 8B). Because the mutation similarly reduced the potency of dopamine by 14-fold ( $EC_{50}$  = 55 nM) but not its efficacy, the drastically reduced efficacy of **20** is a specific effect of the mutation on this ligand. The Y365A mutation resulted in a pronounced 400-fold reduction in the potency ( $EC_{50}$  = 520 nM) and a decrease in efficacy ( $E_{max}$  = 26%) for **20**, whereas dopamine's potency was reduced by only 9-fold ( $EC_{50}$  = 33 nM) without any reduction of its efficacy (Figure 8C). Notably, the double Y198A/Y365A mutation completely abolished the efficacy of **20** for stimulating  $\beta$ -arrestin recruitment to the D3R (Figure 8D), whereas the effects on dopamine's potency ( $EC_{50}$  = 820 nM) appeared to be additive (215-fold) compared to the effects of the single mutants. The unique dependence of **20** on these two residues for full activation of the receptor supports our computational results that show they are in direct contact with **20** but not dopamine. Interestingly, Tyr365<sup>7,35</sup> has previously been suggested as an interaction site for a dopamine receptor agonist with D3R > D2R selectivity.<sup>51</sup>

We thought it would be of interest to also characterize and compare the effects of the Y198A and Y365A mutations on the signaling properties of pramipexole. Figure S3A shows that pramipexole and dopamine have similar potencies in stimulating  $\beta$ -arrestin recruitment to the D3R-WT. Introduction of the Y198A, Y365A, or double Y198A/Y365A mutations into the D3R decreased the potency of pramipexole to a similar extent as that seen with dopamine, while, in contrast to that seen with compound **20**, there was no effect on the efficacy of pramipexole for stimulating  $\beta$ -arrestin recruitment (Figure S3). These results suggest that different efficacy determinants exist for the activation of the D3R by pramipexole and **20**. Taken together, the mutational data lend support to the molecular modeling results that describe unique interactions of **20** with the D3R.

**Neuroprotective Properties of Compound 20.** As D3R-preferring agonists, such as pramipexole and related compounds, have shown neuroprotective properties in several models of neurodegeneration,<sup>11,13–16</sup> we evaluated the effects of compound **20**, using pramipexole as a comparator, in a cellular model of neuroprotection. We genetically engineered a human iPSC cell line so that it stably expresses the human D3R (3–5 pmol/mg) and differentiated these cells into dopaminergic neurons in culture (see the [Experimental Section](#)). The cells were then treated with the dopaminergic neurotoxin 6-OHDA to induce cell death. As shown in Figure 9, 6-OHDA treatment reduced cell viability by  $62 \pm 8\%$  after 24 h. Both compound **20** and pramipexole demonstrated a



**Figure 8.** Mutagenesis studies support the D3R binding site model for compound 20. HEK293 cells were transiently transfected with the following Rluc8-fused receptor constructs: D3R-WT (A), D3R-Y198A (B), D3R-Y365A (C), or D3R-Y198A/Y365A (D), as described in the [Experimental Section](#). Cells were stimulated as indicated and analyzed using the BRET-based  $\beta$ -arrestin recruitment assay (see the [Experimental Section](#)). Data are expressed as a percentage of the maximum dopamine (DA) response. (A) **20** is a full agonist at the D3R-WT;  $EC_{50} = 1.4 \pm 0.7$  nM,  $E_{max} = 92 \pm 10\%$  (mean  $\pm$  SEM,  $n = 8$ ); DA  $EC_{50} = 3.7 \pm 1.2$  nM,  $E_{max} = 100 \pm 0.3\%$  (mean  $\pm$  SEM,  $n = 8$ ). (B) **20** has a reduced potency and efficacy at the D3R-Y198A mutant;  $EC_{50} = 17 \pm 7.6$  nM\*,  $E_{max} = 38 \pm 6.2\%^{**}$  (mean  $\pm$  SEM,  $n = 4$ ); DA  $EC_{50} = 55 \pm 16$  nM\*\*\*,  $E_{max} = 100 \pm 0.2\%$  (mean  $\pm$  SEM,  $n = 4$ ). (C) **20** has a reduced potency and efficacy at the D3R-Y365A;  $EC_{50} = 520 \pm 150$  nM\*\*,  $E_{max} = 26 \pm 11\%^{**}$  (mean  $\pm$  SEM,  $n = 4$ ); DA  $EC_{50} = 33 \pm 3.3$  nM\*\*\*\*,  $E_{max} = 100 \pm 0.1\%$  (mean  $\pm$  SEM,  $n = 4$ ). (D) D3R-Y198A/Y365A mutation abolishes the agonist activity of **20**. DA  $EC_{50} = 820 \pm 56$  nM\*\*\*\*,  $E_{max} = 100 \pm 0.1\%$  (mean  $\pm$  SEM,  $n = 3$ ). Statistical comparisons between WT and mutant parameters were made using a two-tailed t-test: \* $p < 0.05$ , \*\* $p < 0.005$ , \*\*\* $p < 0.0001$ , \*\*\*\* $p < 0.0001$ .

dose-dependent reduction in 6-OHDA-induced cell death. This effect was significant at 50 nM of **20** and maximal at 500 nM. In contrast, pramipexole was somewhat less potent in not achieving a significant level of protection until a concentration of 500 nM was employed ([Figure 9](#)). These data indicate that compound **20** simulates the previously described neuroprotective properties of pramipexole and highlights the utility of pursuing this compound for *in vivo* development, including investigations of additional models of neurodegeneration and as a probe compound to understand the role of the D3R in neuroprotection.

**Pharmacokinetics and Toxicology.** As we are interested in the use of **20** as an *in vivo* probe and as a lead compound for therapeutics development, we performed a number of preliminary toxicology, ADME, and pharmacokinetics experiments. Compound **20** showed no liability ( $AC_{50} > 50 \mu M$ ) using a cytotoxicity screening panel that measures changes in nuclear size, DNA structure, cell membrane permeability, mitochondrial mass, mitochondrial membrane potential, and cytochrome C release (see the [Experimental Section](#)). Further, no toxicity was observed in the AMES reverse mutation assay (data not shown). Compound **20** displays reasonable liver microsomal stability ( $t_{1/2} = 21.2$  min) and excellent permeability using the PAMPA assay ( $541 (10^{-6} \text{ cm}^2/\text{sec})$ ); however, its aqueous solubility is low ( $1.1 \mu\text{g}/\text{mL}$ ). *In vivo* pharmacokinetic experiments in mice (using 20 mg/kg IP) reveal that compound **20** is brain penetrant and exhibits a plasma half-life of 3.44 h and a brain half-life of 4.23 h ([Figure](#)

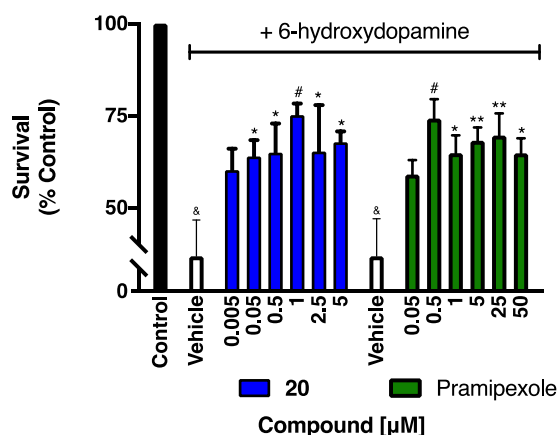
[S4](#), [Tables S2](#) and [S3](#)). Further, compound **20** displays a plasma  $T_{max}$  of 0.5 h and a  $C_{max}$  of 6500 ng/mL, and a brain  $T_{max}$  of 0.25 h and a  $C_{max}$  of 28 000 ng/mL ([Tables S2](#) and [S3](#)). Taken together, these findings are very encouraging for future *in vivo* studies using compound **20** and the potential therapeutic development of this scaffold.

## CONCLUSIONS

In summary, we have identified a novel and highly selective D3R agonist scaffold originating from a high-throughput screening campaign. Focused optimization led to the potent and uniquely selective D3R agonist, **20**. Its exquisite receptor selectivity suggests that **20** will prove to be a valuable pharmacological tool to interrogate the physiological functions of the D3R in normal and pathological conditions. Compound **20** demonstrated neuroprotective effects along with promising toxicology and pharmacokinetic profiles, further suggesting that it may show utility as a therapeutic lead for the treatment of neurodegenerative or other disorders.

## EXPERIMENTAL SECTION

**Materials.** [ $^3\text{H}$ ]-Methylpiperone (80 Ci/mmol) was obtained from PerkinElmer Life Sciences (Waltham, MA). D1R, D2R, D3R, D4R, and D5R expressing CHO-K1 cells and CP2 media were purchased from DiscoverX (Fremont, CA). Other cell culture media and reagents were purchased from MediaTech/Cellgro (Manassas, VA). Cell culture flasks, materials, and all assay plates were purchased from ThermoFisher Scientific (Waltham, MA) and Greiner Bio-One



**Figure 9.** Compound 20 protects D3R-expressing dopaminergic neurons from 6-OHDA-induced cell death. Human iPSCs expressing the D3R were differentiated into dopaminergic neurons, as described in the [Experimental Section](#). Cells were treated with the indicated concentrations of vehicle, 20, or pramipexole for 24 h and then incubated with 30  $\mu$ M of 6-hydroxydopamine (6-OHDA) for 24 h to induce cell death (the control cell group did not receive 6-OHDA). 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) cell viability assays were performed, as described in the [Experimental Section](#). Data represent the means  $\pm$  SEM from six experiments performed in quadruplicate. In the absence of drugs (vehicle), 6-OHDA treatment reduced cell viability by  $62 \pm 8\%$ . Pretreatment with 20 and pramipexole protected against 6-OHDA-induced cell death in a dose-dependent fashion. Statistical significance for differences between the drug- and vehicle-treated groups was assessed using two-way analysis of variance (ANOVA) followed by Dunnett's multiple comparison post hoc test: \* $p < 0.05$ , \*\* $p < 0.01$ , # $p < 0.001$ , & $p < 0.0001$ .

(Monroe, NC). DNA constructs for the BRET assays were kind gifts from Dr. Jonathan A. Javitch. Compound CJ-1639 was a kind gift from Dr. Shaomeng Wang. Receptor mutants were prepared at Bioinnovatise (Rockville, MD), and all mutations were verified by DNA sequencing. Chemicals and buffer components were purchased from Sigma-Aldrich (St. Louis, MO), except where indicated. All tested analogues were synthesized as described below and were characterized as being  $>95\%$  pure.

**Chemistry. General Synthesis and Analysis Experimental Details.** All reagents were used as received from the following suppliers: Alfa Aesar, Sigma-Aldrich, Ark Pharm, Combi-Blocks, Fisher Scientific Oakwood, Matrix, and 1Click Chemistry. Acetonitrile and THF were purified using the Innovative Technology PureSolv solvent purification system using two alumina columns. The  $^1\text{H}$  and  $^{13}\text{C}$  spectra were recorded on a 400 MHz Bruker Avance spectrometer equipped with a broad-band observe probe and a 500 MHz Bruker AVIII spectrometer equipped with a dual cryoprobe, respectively. Chemical shifts are reported in parts per million and were referenced to residual proton solvent signals.  $^{13}\text{C}$  multiplicities were determined with the aid of an APT pulse sequence, differentiating the signals for methyl ( $\text{CH}_3$ ) and methyne ( $\text{CH}$ ) carbons as "d" from methylene ( $\text{CH}_2$ ) and quaternary (C) carbons as "u". The infrared (IR) spectra were acquired as thin films using a universal ATR sampling accessory on a ThermoFisher Nicolet iS5 FTIR spectrometer, and the absorption frequencies are reported in  $\text{cm}^{-1}$ . Melting points were determined on a Stanford Research Systems Optimelt automated melting point system interfaced through a PC and are uncorrected. Microwave syntheses were conducted in a Biotage Initiator constant temperature microwave synthesizer. Flash column chromatography separations were performed using the Teledyne Isco CombiFlash Rf using RediSep Rf silica gel columns. Thin-layer chromatography (TLC) was performed on Analtech UNIPLATE silica gel GHLF plates (gypsum inorganic hard layer with fluorescence). TLC plates were developed using iodine vapor or ceric ammonium molybdate

stain, as required. Automated preparative RP HPLC purification was performed using an Agilent 1200 Mass-Directed Fractionation system (Prep Pump G1361 with gradient extension, make-up pump G1311A, pH modification pump G1311A, HTS PAL autosampler, UV-DAD detection G1315D, fraction collector G1364B, and Agilent 6120 quadrupole spectrometer G6120A). HRMS determinations for compounds 106–109 were analyzed with a ThermoFisher Q Exactive HF-X (ThermoFisher, Bremen, Germany) mass spectrometer coupled with a Waters Acquity H-class liquid chromatograph system. Samples were introduced via a heated electrospray ionization (HESI) source at a flow rate of 0.6 mL/min. Electrospray source conditions were set as follows: spray voltage 3.0 kV, sheath gas (nitrogen) 60 arb, auxiliary gas (nitrogen) 20 arb, sweep gas (nitrogen) 0 arb, nebulizer temperature 375  $^\circ\text{C}$ , capillary temperature 380  $^\circ\text{C}$ , and RF funnel 45 V. The mass range was set to 150–2000  $m/z$ . All measurements were recorded at a resolution setting of 120 000. The preparative chromatography conditions included a Waters X-Bridge  $\text{C}_{18}$  column ( $19 \times 150 \text{ mm}^2$ , 5  $\mu\text{m}$ , with  $19 \times 10 \text{ mm}^2$  guard column), elution with a water and acetonitrile gradient, which increases 20% in acetonitrile content over 4 min at a flow rate of 20 mL/min (modified to pH 9.8 through the addition of  $\text{NH}_4\text{OH}$  by auxiliary pump), and sample dilution in dimethyl sulfoxide (DMSO). The preparative gradient, triggering thresholds, and UV wavelength were selected according to the analytical reverse-phase high-performance liquid chromatography (RP HPLC) analysis of each crude sample. The analytical method used an Agilent 1200 RRCL system with UV detection (Agilent 1200 DAD SL) and mass detection (Agilent 6224 TOF). The analytical method conditions included a Waters Acquity BEH  $\text{C}_{18}$  column ( $2.1 \times 50 \text{ mm}^2$ , 1.7  $\mu\text{m}$ ) and elution with a linear gradient of 5% acetonitrile in pH 9.8 buffered aqueous ammonium formate to 100% acetonitrile at a 0.4 mL/min flow rate. Compound purity was measured on the basis of peak integration (area under the curve) from UV/vis absorbance (at 214 nm), and compound identity was determined on the basis of mass analysis. All compounds used for assays or biological studies possessed HPLC purity  $>95\%$ . The analytical HPLC system used is a dedicated instrument for assessing compound purity and routinely detects impurities as low as 0.1%. Any compounds with a measured HPLC purity of 100% were thus conservatively assigned a purity of " $>99.5\%$ ". Any compounds purified by automated preparative RP HPLC purification utilized the same solvent gradient and column material in the analytical conditions to minimize the possibility of undetected impurities carrying over from the purification run.

All final compounds were inspected for functional groups known to contribute PAINS liabilities, and none were found.

**General Procedure A: Piperazine Coupling with Alkyl Bromides.** The monocarboxamide piperazine substrate, alkyl bromide (1.1–1.2 equiv), potassium carbonate (3.0 equiv), and potassium iodide (1.0 equiv) were charged in a reaction vial and slurred with MeCN (50 mL/mmol substrate) and stirred at 60  $^\circ\text{C}$  for 14–20 h. The reaction was filtered, and the solids were washed with  $\text{CH}_2\text{Cl}_2$  ( $2 \times 5 \text{ mL}$ ). The combined filtrates were evaporated, and the residue was purified by silica gel chromatography to afford the alkylated piperazine product.

**General Procedure B: Piperazine Coupling with Alkyl Bromides: High-Throughput Synthesis Protocol.** The monocarboxamide piperazine substrate as a solution in DMF (0.2 M) and alkyl bromide (1.0 equiv) as a solution in DMF (0.2 M) were added to a reaction tube (Bohdan MiniBlock) containing potassium carbonate (3.0 equiv) and potassium iodide (0.1 equiv). The vial was sealed, and the reaction was stirred at 60  $^\circ\text{C}$  for 14–20 h. The reaction was filtered, and the solids were washed with  $\text{CH}_2\text{Cl}_2$  ( $2 \times 5 \text{ mL}$ ). The combined filtrates were evaporated, and the residue was purified by mass-directed, reverse-phase HPLC to afford the alkylated piperazine product.

**General Procedure C: Mitsunobu Reaction Route to Aryl Ether Analogues.** A solution of (2-hydroxyethyl)piperazine substrate, phenol (1.0 equiv), and triphenyl phosphine (1.0 equiv) in THF or  $\text{CH}_2\text{Cl}_2$  (10 mL/mmol substrate) was cooled in an ice/water bath, and DIAD (1.25 equiv) was added in a single portion. After 1 min, the

ice/water bath was removed, and the reaction was stirred at room temperature (rt) for an additional 5 min and then stirred at 60 °C for 16–40 h. The reaction was monitored for conversion by liquid chromatography/mass spectrometer (LC–MS) and declared complete when no further change was observed in the chromatogram. For reactions in THF, the solvent was removed in vacuo, and the residue was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (5 mL). For reactions in CH<sub>2</sub>Cl<sub>2</sub>, additional CH<sub>2</sub>Cl<sub>2</sub> was added to adjust the volume to 5 mL. The CH<sub>2</sub>Cl<sub>2</sub> solution was washed with aqueous 1 N NaOH (2 × 1 mL) and then water (1 mL), and the organic layer was purified by silica gel chromatography to afford the aryl ether product.

**General Procedure D: Late-Stage Acylation Route to Disubstituted Piperazine Analogues.** A solution of piperazine **114** and triethylamine (1.5 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (20 mL/mmol substrate) was cooled in an ice/water bath, and the appropriate acid chloride (1.0–1.3 equiv) was added in a single portion. The reaction was capped and stirred for 16–20 h, slowly warming to rt. The reaction was washed with water (2 × 5 mL), then dried over Na<sub>2</sub>SO<sub>4</sub>, and the organic layer was purified by silica gel chromatography to afford the aryl ether product.

**General Procedure E: PyBOP-Mediated Coupling Route to Disubstituted Piperazine Analogues.** To a solution of arylcarboxylic acid (1.0–1.3 equiv) in DMF (10 mL/mmol substrate) was added PyBOP (1.2 equiv), and the reaction was stirred at rt for 10 min. A solution of 1-(2-(aryloxy)ethyl)piperazine **114** (1.0 equiv) and diisopropylethyl amine (3.0 equiv) in DMF (10 mL/mmol substrate) was added, and the reaction was stirred at rt for 16–20 h. The reaction solvent was removed in vacuo, and the residue was partitioned between saturated aqueous NaHCO<sub>3</sub> and CH<sub>2</sub>Cl<sub>2</sub> (2 × 5 mL). The combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, concentrated in vacuo, and purified by silica gel chromatography or reverse-phase, mass-directed preparative HPLC to afford the disubstituted piperazine product.

**General Procedure F: Piperazine Alkyl Chloride Displacement Route to Aryl Ether Piperazine Analogues.** A solution of alkyl chloride substrate, phenol nucleophile (1.6–1.9 equiv), and potassium carbonate (2.5 equiv) in DMF (10 mL/mmol substrate) was heated at 60 °C for the stated reaction time and cooled to rt. The reaction was diluted with water (10 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 × 2 mL). The combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, concentrated in vacuo, and purified by silica gel chromatography or reverse-phase, mass-directed preparative HPLC to afford the disubstituted piperazine product.

**General Procedure G: Piperazine Alkyl Chloride Displacement Route to Aryl Ether Piperazine Analogues, Parallel Synthesis Protocol.** A solution of piperazine alkyl chloride substrate in DMF (0.528 M, 1.0 equiv) was added to a 24-position Bohdan MiniBlock reaction tube containing a mixture of phenol, aniline, or thiol nucleophile (1.6–1.9 equiv) and potassium carbonate (3.9 equiv) in DMF (800 μL). The reaction was heated at 60 °C for 16 h and concentrated on a Genevac centrifugal evaporator for 3 h at 35 °C. The residue was partitioned between water (2 mL) and CH<sub>2</sub>Cl<sub>2</sub> (3 mL) and filtered through a Biotage phase separator tube into a 16 × 100 mm tube. The organic layer was concentrated on a Genevac centrifugal evaporator and purified by preparative, mass-directed, reverse-phase HPLC to afford the aryl ether product.

**General Procedure H: Alkylation of Phenols with 1,2-Dibromoethane.** To a solution of phenol **112** and potassium hydroxide (2.0 equiv) in water (3 mL/mmol of phenol) were added sequentially tetrabutylammonium bromide (0.2 equiv) and then 1,2-dibromoethane (5 equiv). The reaction was heated at reflux for 19 h, cooled to rt, and extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 × 50 mL). The combined organic layers were dried with Na<sub>2</sub>SO<sub>4</sub> and concentrated in vacuo. The residue was purified by silica chromatography to afford the bromide product **112**.

**(4-(2-(4-Methoxyphenoxy)ethyl)piperazin-1-yl)(4-methoxyphenyl)methanone (1).** (4-Methoxyphenyl)(piperazin-1-yl)methanone (49 mg, 0.22 mmol) and 1-(2-bromoethoxy)-4-methoxybenzene (61 mg, 0.27 mmol, 1.2 equiv) were reacted according to General Procedure A to afford the alkylated piperazine as

a white solid (66 mg, 0.18 mmol, 81% yield). *R*<sub>f</sub> = 0.29 (EtOAc); mp = 87–89 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 2.51–2.68 (complex, 4H), 2.81 (t, *J* = 5.6 Hz, 2H), 3.51–3.74 (complex, 4H), 3.76 (s, 3H), 3.82 (s, 3H), 4.06 (t, *J* = 5.6 Hz, 2H), 6.80–6.86 (complex, 4H), 6.89–6.92 (m, 2H), 7.36–7.40 (m, 2H); <sup>13</sup>C NMR (101 MHz, APT pulse sequence, CDCl<sub>3</sub>) δ d: 55.3, 55.7, 113.7, 114.6, 115.6, 129.1; u: 53.5, 53.7, 57.3, 66.5, 127.8, 152.8, 154.0, 160.7, 170.2; FTIR (neat): 1623, 1607, 1506 cm<sup>-1</sup>; HRMS (*m/z*): calcd for C<sub>21</sub>H<sub>27</sub>N<sub>2</sub>O<sub>4</sub> [*M* + *H*]<sup>+</sup> 371.1965; found 371.1973; HPLC purity = 99.2%.

**(4-(2-(2-Methoxyphenoxy)ethyl)piperazin-1-yl)(4-methoxyphenyl)methanone (2).** (4-Methoxyphenyl)(piperazin-1-yl)methanone (44 mg, 0.20 mmol) and 1-(2-bromoethoxy)-2-methoxybenzene (46 mg, 0.20 mmol, 1.0 equiv) were reacted according to General Procedure B to afford the alkylated piperazine as a light yellow solid (28 mg, 0.076 mmol, 38% yield). *R*<sub>f</sub> = 0.14 (EtOAc); mp = 119–121 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 2.52–2.68 (complex, 4H), 2.86 (t, *J* = 5.9 Hz, 2H), 3.45–3.88 (complex, 4H), 3.81 (s, 3H), 3.83 (s, 3H), 4.14 (t, *J* = 5.9 Hz, 2H), 6.85–6.94 (complex, 6H), 7.35–7.39 (m, 2H); <sup>13</sup>C NMR (101 MHz, APT pulse sequence, CDCl<sub>3</sub>) δ d: 55.3, 55.9, 111.9, 113.7, 113.9, 120.8, 121.6, 129.1; u: 53.5, 53.7, 57.0, 67.0, 127.8, 148.1, 149.6, 160.7, 170.2; FTIR (neat): 1608, 1504 cm<sup>-1</sup>; HRMS (*m/z*): calcd for C<sub>21</sub>H<sub>27</sub>N<sub>2</sub>O<sub>4</sub> [*M* + *H*]<sup>+</sup> 371.1965; found 371.1985; HPLC purity = 96%.

**(4-(2-(3-Methoxyphenoxy)ethyl)piperazin-1-yl)(4-methoxyphenyl)methanone (3).** (4-Methoxyphenyl)(piperazin-1-yl)methanone (44 mg, 0.20 mmol) and 1-(2-bromoethoxy)-3-methoxybenzene (46 mg, 0.20 mmol, 1.0 equiv) were reacted according to General Procedure B to afford the alkylated piperazine as a pale yellow oil (47 mg, 0.13 mmol, 63% yield). *R*<sub>f</sub> = 0.24 (EtOAc). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 2.51–2.66 (complex, 4H), 2.81 (t, *J* = 5.6 Hz, 2H), 3.47–3.82 (complex, 4H), 3.76 (s, 3H), 3.80 (s, 3H), 4.08 (t, *J* = 5.6 Hz, 2H), 6.44–6.51 (complex, 3H), 6.87–6.91 (m, 2H), 7.17 (t, *J* = 8.2 Hz, 1H), 7.36–7.39 (m, 2H); <sup>13</sup>C NMR (101 MHz, APT pulse sequence, CDCl<sub>3</sub>) δ d: 55.25, 55.32, 101.1, 106.5, 106.6, 113.7, 129.1, 129.9; u: 53.57, 53.63, 57.1, 65.8, 127.8, 159.8, 160.7, 160.8, 170.2; FTIR (neat): 1603, 1492 cm<sup>-1</sup>; HRMS (*m/z*): calcd for C<sub>21</sub>H<sub>27</sub>N<sub>2</sub>O<sub>4</sub> [*M* + *H*]<sup>+</sup> 371.1965; found 371.1988; HPLC purity = 99%.

**(4-(2-(4-Ethylphenoxy)ethyl)piperazin-1-yl)(4-methoxyphenyl)methanone (4).** (4-Methoxyphenyl)(piperazin-1-yl)methanone (44 mg, 0.20 mmol) and 1-(2-bromoethoxy)-4-ethylbenzene (46 mg, 0.20 mmol, 1.0 equiv) were reacted according to General Procedure B to afford the alkylated piperazine as a waxy white solid (44 mg, 0.12 mmol, 59% yield). *R*<sub>f</sub> = 0.19 (EtOAc); mp = 57–64 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.20 (t, *J* = 7.6 Hz, 3H), 2.52–2.65 (complex, 4H), 2.58 (q, *J* = 5.6 Hz, 2H), 2.82 (t, *J* = 5.6 Hz, 2H), 3.43–3.87 (complex, 4H), 3.81 (s, 3H), 4.08 (t, *J* = 5.6 Hz, 2H), 6.79–6.84 (m, 2H), 6.88–6.92 (m, 2H), 7.07–7.12 (m, 2H), 7.36–7.39 (m, 2H); <sup>13</sup>C NMR (101 MHz, APT pulse sequence, CDCl<sub>3</sub>) δ d: 15.9, 55.3, 113.7, 114.4, 128.7, 129.1; u: 28.0, 53.5, 53.6, 57.2, 65.9, 127.8, 136.7, 156.6, 160.7, 170.2; FTIR (neat): 1625, 1607, 1510 cm<sup>-1</sup>; HRMS (*m/z*): calcd for C<sub>22</sub>H<sub>29</sub>N<sub>2</sub>O<sub>3</sub> [*M* + *H*]<sup>+</sup> 369.2173; found 369.2195; HPLC purity = 98%.

**(4-(2-(4-Chlorophenoxy)ethyl)piperazin-1-yl)(4-methoxyphenyl)methanone (5).** (4-Methoxyphenyl)(piperazin-1-yl)methanone (44 mg, 0.20 mmol) and 1-(2-bromoethoxy)-4-chlorobenzene (46 mg, 0.20 mmol, 1.0 equiv) were reacted according to General Procedure B to afford the alkylated piperazine as a white solid (28 mg, 0.08 mmol, 38% yield). *R*<sub>f</sub> = 0.18 (EtOAc); mp = 69–72 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 2.47–2.64 (complex, 4H), 2.81 (t, *J* = 5.6 Hz, 2H), 3.48–3.79 (complex, 4H), 3.81 (s, 3H), 4.06 (t, *J* = 5.6 Hz, 2H), 6.79–6.82 (m, 2H), 6.87–6.91 (m, 2H), 7.18–7.23 (m, 2H), 7.35–7.38 (m, 2H); <sup>13</sup>C NMR (101 MHz, APT pulse sequence, CDCl<sub>3</sub>) δ d: 55.3, 113.7, 115.8, 129.1, 129.3; u: 53.6, 53.7, 57.0, 66.2, 125.8, 127.7, 157.2, 160.8, 170.2; FTIR (neat): 1623, 1607 cm<sup>-1</sup>; HRMS (*m/z*): calcd for C<sub>20</sub>H<sub>24</sub>ClN<sub>2</sub>O<sub>3</sub> [*M* + *H*]<sup>+</sup> 375.1470; found 375.1501; HPLC purity = 99%.

**(4-(2-(4-(Trifluoromethoxy)phenoxy)ethyl)piperazin-1-yl)(4-methoxyphenyl)methanone (6).** (4-(2-Hydroxyethyl)piperazin-1-yl)(4-methoxyphenyl)methanone (96 mg, 0.36 mmol) and 4-

(trifluoromethoxy)phenol (65 mg, 0.36 mmol, 1.0 equiv) in  $\text{CH}_2\text{Cl}_2$  were reacted according to General Procedure C for 46 h to afford the aryl ether as a light orange oil (56 mg, 0.13 mmol, 36% yield).  $R_f = 0.27$  (EtOAc).  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.50–2.68 (complex, 4H), 2.84 (t,  $J = 5.6$  Hz, 2H), 3.47–3.82 (complex, 4H), 3.83 (s, 3H), 4.10 (t,  $J = 5.6$  Hz, 2H), 6.86–6.92 (complex, 4H), 7.11–7.16 (m, 2H), 7.37–7.41 (m, 2H);  $^{13}\text{C NMR}$  (101 MHz, APT pulse sequence,  $\text{CDCl}_3$ )  $\delta$  d: 55.4, 114.0, 115.5, 122.7, 129.5; u: 52.7, 56.4, 63.26, 63.30, 120.4 (q,  $J = 257.9$  Hz), 125.6, 143.6, 155.5, 161.6, 170.4;  $^{19}\text{F NMR}$  (376 MHz,  $\text{CDCl}_3$ )  $\delta$  –58.4; FTIR (neat): 1634, 1607, 1507  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{24}\text{F}_3\text{N}_2\text{O}_4$  [ $\text{M} + \text{H}$ ] $^+$  425.1683; found 425.1696; HPLC purity >99.5%.

(4-(2-(3,5-Dimethoxyphenoxy)ethyl)piperazin-1-yl)(4-methoxyphenyl)methanone (7). (4-Methoxyphenyl)(piperazin-1-yl)methanone (44 mg, 0.20 mmol) and 1-(2-bromoethoxy)-3,5-dimethoxybenzene (52 mg, 0.20 mmol, 1.0 equiv) were reacted according to General Procedure B to afford the alkylated piperazine as a yellow oil (57 mg, 0.14 mmol, 71% yield).  $R_f = 0.17$  (EtOAc).  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.48–2.67 (complex, 4H), 2.81 (t,  $J = 5.5$  Hz, 2H), 3.74 (s, 6H), 3.81 (s, 3H), 3.45–3.79 (complex, 4H), 4.05 (t,  $J = 5.5$  Hz, 2H), 6.04–6.08 (complex, 3H), 6.89 (d,  $J = 8.7$  Hz, 2H), 7.37 (d,  $J = 8.7$  Hz, 2H);  $^{13}\text{C NMR}$  (101 MHz, APT pulse sequence,  $\text{CDCl}_3$ )  $\delta$  d: 55.3, 93.1, 93.5, 113.7, 129.1; u: 53.5, 53.6, 57.0, 65.8, 127.8, 160.5, 160.7, 161.5, 170.2; FTIR (neat): 1592, 1456, 1425  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{29}\text{N}_2\text{O}_5$  [ $\text{M} + \text{H}$ ] $^+$  401.2071; found 401.2102; HPLC purity = 98%.

(4-Methoxyphenyl)(4-(2-phenoxyethyl)piperazin-1-yl)methanone (8). (4-(2-Hydroxyethyl)piperazin-1-yl)(4-methoxyphenyl)methanone (76 mg, 0.29 mmol) and phenol (27 mg, 0.29 mmol, 1.0 equiv) in THF were reacted according to General Procedure C for 22 h to afford the aryl ether as an orange oil (31 mg, 0.091 mmol, 32% yield).  $R_f = 0.17$  (EtOAc).  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.53–2.72 (complex, 4H), 2.85 (t,  $J = 5.6$  Hz, 2H), 3.43–3.82 (complex, 4H), 3.83 (s, 3H), 4.12 (t,  $J = 5.6$  Hz, 2H), 6.86–6.93 (complex, 4H), 6.95 (tt,  $J = 1.0, 7.4$  Hz, 1H), 7.25–7.31 (m, 2H), 7.37–7.40 (m, 2H);  $^{13}\text{C NMR}$  (101 MHz, APT pulse sequence,  $\text{CDCl}_3$ )  $\delta$  d: 55.3, 113.7, 114.5, 120.9, 129.1, 129.4; u: 53.57, 53.60, 57.1, 65.7, 127.8, 158.5, 160.7, 170.2; FTIR (neat): 1622, 1599, 1456, 1427  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{20}\text{H}_{25}\text{N}_2\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  341.1860; found 341.1872; HPLC purity = 99%.

(4-(2-(Benzo[d][1,3]dioxol-5-yloxy)ethyl)piperazin-1-yl)(4-methoxyphenyl)methanone (9). (4-Methoxyphenyl)(piperazin-1-yl)methanone (44 mg, 0.20 mmol) and 5-(2-bromoethoxy)benzo[d][1,3]dioxole (49 mg, 0.20 mmol, 1.0 equiv) were reacted according to General Procedure B to afford the alkylated piperazine as a tan solid (50 mg, 0.13 mmol, 65% yield).  $R_f = 0.16$  (EtOAc); mp = 97–101 °C.  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.48–2.64 (complex, 4H), 2.77 (t,  $J = 5.6$  Hz, 2H), 3.44–3.86 (complex, 4H), 3.80 (s, 3H), 4.01 (t,  $J = 5.6$  Hz, 2H), 5.87 (s, 2H), 6.29 (dd,  $J = 2.5, 8.5$  Hz, 1H), 6.47 (d,  $J = 2.4$  Hz, 1H), 6.67 (d,  $J = 8.4$  Hz, 1H), 6.88 (d,  $J = 8.3$  Hz, 1H), 7.36 (d,  $J = 8.8$  Hz, 1H);  $^{13}\text{C NMR}$  (101 MHz, APT pulse sequence,  $\text{CDCl}_3$ )  $\delta$  d: 55.3, 98.2, 105.7, 107.9, 113.7, 129.1; u: 53.5, 53.6, 57.1, 66.8, 127.8, 141.8, 148.2, 154.1, 160.7, 170.2; FTIR (neat): 1622, 1607, 1486  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{25}\text{N}_2\text{O}_5$  [ $\text{M} + \text{H}$ ] $^+$  385.1758; found 385.1789; HPLC purity = 99%.

(4-Methoxyphenyl)(4-(2-(pyridin-3-yloxy)ethyl)piperazin-1-yl)methanone (10). (4-(2-Hydroxyethyl)piperazin-1-yl)(4-methoxyphenyl)methanone (81 mg, 0.31 mmol) and 3-hydroxypyridine (29 mg, 0.31 mmol, 1.0 equiv) in  $\text{CH}_2\text{Cl}_2$  were reacted according to General Procedure C for 22 h to afford the aryl ether as a colorless oil (16 mg, 0.047 mmol, 15% yield).  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.53–2.65 (complex, 4H), 2.86 (t,  $J = 5.6$  Hz, 2H), 3.45–3.80 (complex, 4H), 3.83 (s, 3H), 4.16 (t,  $J = 5.6$  Hz, 2H), 6.89–6.93 (m, 2H), 7.19–7.23 (m, 2H), 7.37–7.41 (m, 2H), 8.23 (dd,  $J = 2.0, 4.0$  Hz, 1H), 8.32 (dd,  $J = 1.0, 2.4$  Hz, 1H);  $^{13}\text{C NMR}$  (126 MHz,  $\text{CDCl}_3$ )  $\delta$  53.6, 53.7, 55.4, 57.0, 66.2, 113.7, 121.2, 123.8, 127.7, 129.2, 138.0, 142.4, 154.8, 160.8, 170.3; HRMS ( $m/z$ ): calcd for  $\text{C}_{19}\text{H}_{24}\text{N}_3\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  342.1812; found 342.1822; HPLC purity = 99%.

(4-(2-(4-Methoxyphenoxy)ethyl)piperazin-1-yl)(phenyl)methanone (11). 1-(2-(4-Methoxyphenoxy)ethyl)piperazine (107 mg, 0.45 mmol) and benzoyl chloride (83 mg, 0.59 mmol, 1.3 equiv) were reacted according to General Procedure D for 16 h to afford the acylated product as a viscous, pale yellow oil (106 mg, 0.31 mmol, 69% yield).  $R_f = 0.28$  (EtOAc).  $^1\text{H NMR}$  (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.49–2.70 (complex, 4H), 2.82 (t,  $J = 5.5$  Hz, 2H), 3.39–3.50 (m, 2H), 3.75–3.86 (m, 2H), 3.77 (s, 3H), 4.07 (t,  $J = 5.6$  Hz, 2H), 6.79–6.86 (complex, 4H), 7.37–7.44 (complex, 5H);  $^{13}\text{C NMR}$  (126 MHz,  $\text{CDCl}_3$ )  $\delta$  53.3, 53.9, 55.7, 57.2, 66.5, 114.6, 115.6, 127.0, 128.5, 129.7, 135.8, 152.7, 154.0, 170.3; FTIR (neat): 1626, 1506, 1431  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{20}\text{H}_{25}\text{N}_2\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  341.1860; found 341.1874; HPLC purity = 98%.

(4-(2-(4-Methoxyphenoxy)ethyl)piperazin-1-yl)(2-methoxyphenyl)methanone (12). (2-Methoxyphenyl)(piperazin-1-yl)methanone (44 mg, 0.20 mmol) and 1-(2-bromoethoxy)-4-methoxybenzene (46 mg, 0.20 mmol, 1.0 equiv) were reacted according to General Procedure B to afford the alkylated piperazine as an orange oil (22 mg, 0.059 mmol, 30% yield).  $R_f = 0.21$  (EtOAc).  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.37–2.71 (complex, 4H), 2.79 (t,  $J = 5.6$  Hz, 2H), 3.19–3.33 (m, 2H), 3.74 (s, 3H), 3.77–3.92 (m, 2H), 3.80 (s, 3H), 4.04 (t,  $J = 5.6$  Hz, 2H), 6.78–6.84 (complex, 4H), 6.88 (d,  $J = 6.4$  Hz, 1H), 6.96 (t,  $J = 7.5$  Hz, 1H), 7.20–7.25 (m, 1H), 7.32 (tt,  $J = 1.1, 7.9$  Hz, 1H);  $^{13}\text{C NMR}$  (101 MHz, APT pulse sequence,  $\text{CDCl}_3$ )  $\delta$  d: 55.5, 55.7, 110.8, 114.6, 115.5, 120.9, 128.0, 130.3; u: 41.5, 46.7, 53.3, 53.8, 57.3, 66.5, 125.7, 152.8, 153.9, 155.3, 167.7; FTIR (neat): 1628, 1600, 1506  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{27}\text{N}_2\text{O}_4$  [ $\text{M} + \text{H}$ ] $^+$  371.1965; found 371.1969; HPLC purity = 97%.

(4-(2-(4-Methoxyphenoxy)ethyl)piperazin-1-yl)(3-methoxyphenyl)methanone (13). (3-Methoxyphenyl)(piperazin-1-yl)methanone (44 mg, 0.20 mmol) and 1-(2-bromoethoxy)-4-methoxybenzene (46 mg, 0.20 mmol, 1.0 equiv) were reacted according to General Procedure B to afford the alkylated piperazine as an orange oil (44 mg, 0.12 mmol, 59% yield).  $R_f = 0.23$  (EtOAc).  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.45–2.66 (complex, 4H), 2.78 (t,  $J = 5.6$  Hz, 2H), 3.36–3.49 (m, 2H), 3.71–3.84 (m, 2H), 3.73 (s, 3H), 3.79 (s, 3H), 4.03 (t,  $J = 5.6$  Hz, 2H), 6.76–6.83 (complex, 4H), 6.89–6.95 (complex, 3H), 7.23–7.30 (m, 1H);  $^{13}\text{C NMR}$  (101 MHz, APT pulse sequence,  $\text{CDCl}_3$ )  $\delta$  d: 55.3, 55.7, 112.4, 114.6, 115.5, 115.5, 119.0, 129.5; u: 53.2, 53.8, 57.2, 66.5, 137.0, 152.7, 153.9, 159.6, 170.0; FTIR (neat): 1628, 1578, 1506  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{27}\text{N}_2\text{O}_4$  [ $\text{M} + \text{H}$ ] $^+$  371.1965; found 371.1987; HPLC purity = 99%.

(4-Chlorophenyl)(4-(2-(4-methoxyphenoxy)ethyl)piperazin-1-yl)methanone (14). (4-Chlorophenyl)(piperazin-1-yl)methanone (45 mg, 0.20 mmol) and 1-(2-bromoethoxy)-4-methoxybenzene (46 mg, 0.20 mmol, 1.0 equiv) were reacted according to General Procedure B to afford the alkylated piperazine as a light yellow solid (27 mg, 0.073 mmol, 37% yield).  $R_f = 0.29$  (EtOAc); mp = 78–80 °C.  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.47–2.67 (complex, 4H), 2.80 (t,  $J = 5.6$  Hz, 2H), 3.36–3.49 (m, 2H), 3.72–3.84 (m, 2H), 3.74 (s, 3H), 4.04 (t,  $J = 5.6$  Hz, 2H), 6.78–6.85 (complex, 4H), 7.32–7.38 (complex, 4H);  $^{13}\text{C NMR}$  (101 MHz, APT pulse sequence,  $\text{CDCl}_3$ )  $\delta$  d: 55.7, 114.6, 115.5, 128.6, 128.7; u: 53.2, 53.8, 57.2, 66.5, 134.1, 135.7, 152.7, 154.0, 169.1; FTIR (neat): 1628, 1594, 1505  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{20}\text{H}_{24}\text{ClN}_2\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  375.1470; found 375.1478; HPLC purity >99.5%.

(4-Ethylphenyl)(4-(2-(4-methoxyphenoxy)ethyl)piperazin-1-yl)methanone (15). (4-Ethylphenyl)(piperazin-1-yl)methanone (44 mg, 0.20 mmol) and 1-(2-bromoethoxy)-4-methoxybenzene (46 mg, 0.20 mmol, 1.0 equiv) were reacted according to General Procedure B to afford the alkylated piperazine as a light yellow oil (40 mg, 0.11 mmol, 54% yield).  $R_f = 0.29$  (EtOAc).  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  1.21 (t,  $J = 7.6$  Hz, 3H), 2.46–2.68 (complex, 4H), 2.64 (q,  $J = 7.6$  Hz, 2H), 2.79 (t,  $J = 5.6$  Hz, 2H), 3.39–3.54 (m, 2H), 3.67–3.84 (m, 2H), 3.73 (s, 3H), 4.04 (t,  $J = 5.6$  Hz, 2H), 6.77–6.84 (complex, 4H), 7.18–7.22 (m, 2H), 7.28–7.32 (m, 2H);  $^{13}\text{C NMR}$  (101 MHz, APT pulse sequence,  $\text{CDCl}_3$ )  $\delta$  d: 15.4, 55.7, 114.6, 115.5, 127.2, 127.9; u: 28.7, 53.7, 53.9, 57.2, 66.5, 133.0, 146.1, 152.7, 153.9, 170.5; FTIR

(neat): 1627, 1506, 1426  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{29}\text{N}_2\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  369.2173; found 369.2196; HPLC purity = 99%.

**(4-(2-(4-Methoxyphenoxy)ethyl)piperazin-1-yl)pyridine-4-yl-methanone (16).** 1-(2-(4-Methoxyphenoxy)ethyl)piperazine (108 mg, 0.46 mmol) and isonicotinoyl chloride hydrochloride (99 mg, 0.56 mmol, 1.2 equiv) were reacted according to General Procedure D for 16 h to afford the acylated product as an orange-brown oil (95 mg, 0.28 mmol, 61% yield).  $R_f = 0.11$  (EtOAc).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.51–2.56 (m, 2H), 2.65–2.71 (m, 2H), 2.83 (t,  $J = 5.5$  Hz, 2H), 3.36–3.41 (m, 4H), 3.77 (s, 3H), 3.79–3.85 (m, 2H), 4.06 (t,  $J = 5.5$  Hz, 2H), 6.81–6.87 (complex, 4H), 7.28 (d,  $J = 6.0$  Hz, 2H), 8.70 (d,  $J = 6.0$  Hz, 2H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  42.0, 47.4, 53.1, 53.7, 55.7, 57.2, 66.5, 114.6, 115.5, 121.2, 143.4, 150.3, 152.7, 154.0, 167.6; FTIR (neat): 1632, 1506, 1436  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{19}\text{H}_{24}\text{N}_3\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  342.1812; found 342.1827; HPLC purity >99.5%.

**(4-(2-(4-Methoxyphenoxy)ethyl)piperazin-1-yl)(5-methoxy-pyridin-2-yl)methanone (17).** 5-Methoxypicolinic acid (19 mg, 0.12 mmol, 1.2 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as a light yellow oil (38 mg, 0.10 mmol, 93% yield).  $R_f = 0.13$  (EtOAc).  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ )  $\delta$  2.42–2.51 (m, 2H), 2.52–2.58 (m, 2H), 2.70 (t,  $J = 5.8$  Hz, 2H), 3.48–3.53 (m, 2H), 3.60–3.66 (m, 2H), 3.69 (s, 3H), 3.87 (s, 3H), 4.02 (t,  $J = 5.7$  Hz, 2H), 6.82–6.89 (complex, 4H), 7.49 (dd,  $J = 2.9, 8.7$  Hz, 1H), 7.59 (d,  $J = 8.6$  Hz, 1H), 8.27 (dd,  $J = 0.6, 2.9$  Hz, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{DMSO}-d_6$ )  $\delta$  41.8, 46.7, 52.9, 53.4, 55.4, 55.8, 56.6, 65.8, 114.6, 115.4, 121.2, 124.8, 135.7, 146.0, 152.4, 153.4, 155.9, 166.3; FTIR (neat): 1628, 1507, 1458  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{20}\text{H}_{26}\text{N}_3\text{O}_4$  [ $\text{M} + \text{H}$ ] $^+$  372.1918; found 372.1926; HPLC purity >99.5%.

**(1H-Indol-5-yl)(4-(2-(4-methoxyphenoxy)ethyl)piperazin-1-yl)-methanone (18).** 1H-Indole-5-carboxylic acid (21 mg, 0.13 mmol, 1.2 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as a white solid (29 mg, 0.08 mmol, 69% yield).  $R_f = 0.17$  (EtOAc); mp = 123–127  $^{\circ}\text{C}$ ;  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.50–2.69 (complex, 4H), 2.83 (t,  $J = 5.6$  Hz, 2H), 3.50–3.85 (complex, 4H), 3.77 (s, 3H), 4.07 (t,  $J = 5.7$  Hz, 2H), 6.57–6.60 (m, 1H), 6.81–6.85 (complex, 4H), 7.24–7.28 (m, 2H), 7.38 (d,  $J = 8.4$  Hz, 1H), 7.73 (d,  $J = 1.6$  Hz, 1H), 8.52 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  46.5, 53.8, 55.9, 57.4, 66.6, 103.3, 111.1, 114.8, 115.7, 120.4, 121.6, 125.5, 127.3, 127.5, 136.5, 152.9, 154.1, 171.9; FTIR (neat): 1599, 1506, 1431  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{26}\text{N}_3\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  380.1969; found 380.1971; HPLC purity = 96%.

**(1H-Indol-3-yl)(4-(2-(4-methoxyphenoxy)ethyl)piperazin-1-yl)-methanone (19).** 1H-Indole-3-carboxylic acid (21 mg, 0.13 mmol, 1.2 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as an off-white solid (6.4 mg, 0.017 mmol, 15% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.59–2.66 (complex, 4H), 2.84 (t,  $J = 5.6$  Hz, 2H), 3.72–3.81 (complex, 4H), 3.76 (s, 3H), 4.08 (t,  $J = 5.6$  Hz, 2H), 6.79–6.86 (complex, 4H), 7.19–7.25 (m, 2H), 7.40–7.43 (m, 1H), 7.55 (d,  $J = 2.7$  Hz, 1H), 7.71 (d,  $J = 7.7$  Hz, 1H), 8.47 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  46.4, 54.0, 55.9, 57.5, 66.7, 111.7, 112.4, 114.8, 115.7, 120.6, 121.3, 123.1, 125.5, 127.0, 135.6, 152.9, 154.1, 166.6; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{26}\text{N}_3\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  380.1969; found 380.1973; HPLC purity = 99%.

**(1H-Indol-2-yl)(4-(2-(4-methoxyphenoxy)ethyl)piperazin-1-yl)-methanone (20).** To a mixture of 1-(2-(4-methoxyphenoxy)ethyl)piperazine (146 mg, 0.618 mmol), indole-2-carboxylic acid (119 mg, 0.741 mmol, 1.2 equiv), and DMAP (8 mg, 0.062 mmol, 0.1 equiv) in THF (10 mL) was added diisopropylcarbodiimide (0.29 mL, 234 mg, 1.85 mmol, 3.0 equiv). The reaction was stirred at rt for 15 h, and the solvents were removed under vacuum. The residue was purified via silica gel chromatography to afford the acylated product as an off-white solid (172 mg, 0.453 mmol, 73% yield).  $R_f = 0.54$  (MeOH (10%) and  $\text{NH}_4\text{OH}$  (2%) in  $\text{CH}_2\text{Cl}_2$ ); mp = 163–165  $^{\circ}\text{C}$ .  $^1\text{H}$  NMR

(400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.68 (t,  $J = 5.1$  Hz, 4H), 2.85 (t,  $J = 5.6$  Hz, 2H), 3.77 (s, 3H), 3.90–4.05 (m, 4H), 4.10 (t,  $J = 5.5$  Hz, 2H), 6.78 (dd,  $J = 1.0, 2.1$  Hz, 1H), 6.82–6.89 (complex, 4H), 7.13 (ddd,  $J = 1.0, 7.0, 8.0$  Hz, 1H), 7.24–7.32 (m, 1H), 7.43 (dd,  $J = 1.1, 8.3$  Hz, 1H), 7.65 (dd,  $J = 1.2, 8.0$  Hz, 1H), 9.65 (br s, 1H);  $^{13}\text{C}$  NMR (101 MHz, APT pulse sequence,  $\text{CDCl}_3$ )  $\delta$  d: 55.7, 105.2, 111.8, 114.7, 115.6, 120.5, 121.8, 124.4; u: 53.6, 57.3, 66.6, 127.4, 129.2, 135.7, 152.8, 154.0, 162.3; FTIR (neat): 3258, 1597, 1506, 1437  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{26}\text{N}_3\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  380.1969; found 380.1995; HPLC purity = 96% (for LC/MS chromatogram and spectrum, see the Supporting Information, Figures S5 and S6).

**(7-Methoxy-1H-indol-2-yl)(4-(2-(4-methoxyphenoxy)ethyl)piperazin-1-yl)methanone (21).** 7-Methoxy-1H-indole-2-carboxylic acid (22 mg, 0.11 mmol, 1.1 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as a tan solid (31.6 mg, 0.077 mmol, 71% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.67 (t,  $J = 5.1$  Hz, 4H), 2.84 (t,  $J = 5.6$  Hz, 2H), 3.77 (s, 3H), 3.89–4.01 (complex, 4H), 3.96 (s, 3H), 4.09 (t,  $J = 5.6$  Hz, 2H), 6.69 (dd,  $J = 0.8, 7.7$  Hz, 1H), 6.74 (d,  $J = 2.3$  Hz, 1H), 6.82–6.87 (complex, 4H), 7.05 (t,  $J = 7.9$  Hz, 1H), 7.21–7.25 (m, 1H), 9.22 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  41.1, 53.7, 55.6, 55.9, 57.4, 66.6, 103.6, 105.5, 114.2, 114.8, 115.7, 121.2, 126.8, 128.7, 129.1, 146.6, 152.9, 154.1, 162.3; HRMS ( $m/z$ ): calcd for  $\text{C}_{23}\text{H}_{28}\text{N}_3\text{O}_4$  [ $\text{M} + \text{H}$ ] $^+$  410.2074; found 410.2073; HPLC purity = 99%.

**(6-Methoxy-1H-indol-2-yl)(4-(2-(4-methoxyphenoxy)ethyl)piperazin-1-yl)methanone (22).** 6-Methoxy-1H-indole-2-carboxylic acid (22 mg, 0.11 mmol, 1.1 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as an off-white solid (30 mg, 0.073 mmol, 67% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.67 (t,  $J = 5.1$  Hz, 4H), 2.84 (t,  $J = 5.6$  Hz, 2H), 3.77 (s, 3H), 3.86 (s, 3H), 3.90–4.01 (complex, 4H), 4.09 (t,  $J = 5.6$  Hz, 2H), 6.72 (dd,  $J = 0.9, 2.2$  Hz, 1H), 6.81 (dd,  $J = 2.3, 8.7$  Hz, 1H), 6.82–6.88 (complex, 5H), 7.44–7.62 (m, 1H), 9.10 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  53.7, 55.7, 55.9, 57.4, 66.7, 93.8, 105.8, 112.1, 114.8, 115.7, 122.0, 122.8, 128.3, 136.7, 152.9, 154.1, 158.4, 162.3; FTIR (neat): 1596, 1505  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{23}\text{H}_{28}\text{N}_3\text{O}_4$  [ $\text{M} + \text{H}$ ] $^+$  410.2074; found 410.2074; HPLC purity = 98%.

**(5-Methoxy-1H-indol-2-yl)(4-(2-(4-methoxyphenoxy)ethyl)piperazin-1-yl)methanone (23).** 5-Methoxy-1H-indole-2-carboxylic acid (22 mg, 0.11 mmol, 1.1 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as a white solid (32 mg, 0.078 mmol, 72% yield).  $R_f = 0.20$  (EtOAc); mp = 154–155  $^{\circ}\text{C}$ .  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.67 (t,  $J = 5.1$  Hz, 4H), 2.85 (t,  $J = 5.6$  Hz, 2H), 3.77 (s, 3H), 3.84 (s, 3H), 3.89–4.04 (complex, 4H), 4.09 (t,  $J = 5.6$  Hz, 2H), 6.70 (dd,  $J = 0.9, 2.2$  Hz, 1H), 6.82–6.89 (complex, 4H), 6.95 (dd,  $J = 2.4, 8.9$  Hz, 1H), 7.05 (d,  $J = 2.4$  Hz, 1H), 7.32 (td,  $J = 0.8, 9.1$  Hz, 1H), 9.22 (s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  41.1, 53.7, 55.9, 57.4, 66.7, 102.4, 105.1, 112.8, 114.8, 115.7, 115.8, 127.9, 129.8, 131.1, 152.9, 154.1, 154.7, 162.4; FTIR (neat): 1595, 1525, 1506, 1437  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{23}\text{H}_{28}\text{N}_3\text{O}_4$  [ $\text{M} + \text{H}$ ] $^+$  410.2074; found 410.2076; HPLC purity >99.5%.

**(4-Methoxy-1H-indol-2-yl)(4-(2-(4-methoxyphenoxy)ethyl)piperazin-1-yl)methanone (24).** 4-Methoxy-1H-indole-2-carboxylic acid (22 mg, 0.11 mmol, 1.1 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as a white solid (30 mg, 0.072 mmol, 67% yield).  $R_f = 0.64$  (10% MeOH in  $\text{CH}_2\text{Cl}_2$ ); mp = 127–129  $^{\circ}\text{C}$ .  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.67 (t,  $J = 5.1$  Hz, 4H), 2.84 (t,  $J = 5.6$  Hz, 2H), 3.77 (s, 3H), 3.90–4.05 (complex, 4H), 3.96 (s, 3H), 4.09 (t,  $J = 5.6$  Hz, 2H), 6.51 (dd,  $J = 0.6, 7.8$  Hz, 1H), 6.90 (dd,  $J = 0.9, 2.3$  Hz, 1H), 7.04 (ddd,  $J = 1.8, 8.0, 79.5$  Hz, 1H), 7.20 (t,  $J = 7.9$  Hz, 1H), 9.29 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  41.1, 53.7, 55.4, 55.9, 57.4, 66.7, 99.7, 102.8, 105.0, 114.8, 115.7, 118.8,

125.5, 128.1, 137.1, 152.9, 154.1, 154.2, 162.2; FTIR (neat): 1595, 1580, 1505, 1433  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{23}\text{H}_{28}\text{N}_3\text{O}_4$  [ $\text{M} + \text{H}$ ] $^+$  410.2074; found 410.2076; HPLC purity >99.5%.

**(4-(2-(4-Methoxyphenoxy)ethyl)piperazin-1-yl)(7-methyl-1H-indol-2-yl)methanone (25).** 7-Methyl-1H-indole-2-carboxylic acid (20 mg, 0.11 mmol, 1.1 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as an off-white solid (28.0 mg, 0.071 mmol, 66% yield).  $R_f$  = 0.64 (10% MeOH in  $\text{CH}_2\text{Cl}_2$ ); mp = 134–138 °C.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.50 (s, 3H), 2.68 (t,  $J$  = 5.1 Hz, 4H), 2.85 (t,  $J$  = 5.6 Hz, 2H), 3.77 (s, 3H), 3.87–4.05 (complex, 4H), 4.10 (t,  $J$  = 5.5 Hz, 2H), 6.78 (d,  $J$  = 2.1 Hz, 1H), 6.82–6.91 (complex, 4H), 7.01–7.11 (m, 2H), 7.49 (dd,  $J$  = 2.2, 6.7 Hz, 1H), 9.12 (s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  16.9, 41.1, 53.7, 55.4, 57.2, 66.0, 101.2, 105.9, 106.7, 106.8, 119.6, 121.0, 121.2, 124.8, 127.1, 129.0, 130.1, 135.5, 160.0, 161.0, 162.5; FTIR (neat): 1598, 1534, 1505, 1436  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{23}\text{H}_{28}\text{N}_3\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  394.2125; found 394.2124; HPLC purity >99.5%.

**(4-(2-(4-Methoxyphenoxy)ethyl)piperazin-1-yl)(6-methyl-1H-indol-2-yl)methanone (26).** 6-Methyl-1H-indole-2-carboxylic acid (19 mg, 0.11 mmol, 1.0 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as a white solid (26.7 mg, 0.068 mmol, 63% yield).  $R_f$  = 0.60 (10% MeOH in  $\text{CH}_2\text{Cl}_2$ ); mp = 130–131 °C.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.46 (s, 3H), 2.67 (t,  $J$  = 5.1 Hz, 4H), 2.85 (t,  $J$  = 5.6 Hz, 2H), 3.77 (s, 3H), 3.90–4.03 (complex, 4H), 4.09 (t,  $J$  = 5.6 Hz, 2H), 6.73 (dd,  $J$  = 0.9, 2.1 Hz, 1H), 6.81–6.89 (complex, 4H), 6.97 (ddd,  $J$  = 0.6, 1.4, 8.2 Hz, 1H), 7.21 (m, 1H), 7.52 (dd,  $J$  = 0.9, 8.2 Hz, 1H), 9.08 (br s, 1H);  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  22.1, 53.7, 55.9, 57.4, 66.7, 105.4, 111.6, 114.8, 115.7, 121.6, 122.8, 125.5, 128.8, 134.6, 136.2, 152.9, 154.1, 162.5; FTIR (neat): 1596, 1523, 1507, 1438  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{23}\text{H}_{28}\text{N}_3\text{O}_4$  [ $\text{M} + \text{H}$ ] $^+$  394.2125; found 394.2128; HPLC purity = 99%.

**(4-(2-(4-Methoxyphenoxy)ethyl)piperazin-1-yl)(5-methyl-1H-indol-2-yl)methanone (27).** 5-Methyl-1H-indole-2-carboxylic acid (20 mg, 0.11 mmol, 1.1 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as a white solid (29 mg, 0.074 mmol, 68% yield).  $R_f$  = 0.60 (10% MeOH in  $\text{CH}_2\text{Cl}_2$ ); mp = 175–179 °C.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.44 (s, 3H), 2.67 (t,  $J$  = 5.1 Hz, 4H), 2.85 (t,  $J$  = 5.6 Hz, 2H), 3.77 (s, 3H), 3.91–4.02 (complex, 4H), 4.09 (t,  $J$  = 5.5 Hz, 2H), 6.69 (dd,  $J$  = 0.9, 2.2 Hz, 1H), 6.81–6.89 (complex, 4H), 7.11 (dd,  $J$  = 1.3, 8.4 Hz, 1H), 7.31 (dt,  $J$  = 0.8, 8.4 Hz, 1H), 7.41 (dd,  $J$  = 0.8, 1.7 Hz, 1H), 9.10 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  53.7, 55.7, 55.9, 57.4, 66.7, 93.8, 105.8, 112.1, 114.8, 115.7, 122.0, 122.8, 128.3, 136.7, 152.9, 154.1, 158.4, 162.3; FTIR (neat): 1592, 1536, 1506, 1434  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{23}\text{H}_{28}\text{N}_3\text{O}_4$  [ $\text{M} + \text{H}$ ] $^+$  394.2125; found 394.2127; HPLC purity = 99%.

**(4-(2-(4-Methoxyphenoxy)ethyl)piperazin-1-yl)(4-methyl-1H-indol-2-yl)methanone (28).** 4-Methyl-1H-indole-2-carboxylic acid (21 mg, 0.12 mmol, 1.1 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as an off-white solid (31.3 mg, 0.080 mmol, 74% yield).  $R_f$  = 0.68 (10% MeOH in  $\text{CH}_2\text{Cl}_2$ ); mp = 134–136 °C.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.56 (s, 2H), 2.69 (t,  $J$  = 5.1 Hz, 4H), 2.85 (t,  $J$  = 5.5 Hz, 2H), 3.77 (s, 3H), 3.91–4.06 (complex, 4H), 4.10 (t,  $J$  = 5.5 Hz, 2H), 6.78 (dd,  $J$  = 1.0, 2.2 Hz, 1H), 6.81–6.89 (complex, 4H), 6.93 (td,  $J$  = 0.9, 7.1 Hz, 1H), 7.19 (dd,  $J$  = 7.0, 8.3 Hz, 1H), 7.24–7.27 (m, 1H), 9.20 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  18.9, 41.1, 53.7, 55.9, 57.4, 66.7, 103.9, 109.4, 114.8, 115.7, 120.7, 124.7, 127.7, 128.8, 131.6, 135.5, 152.9, 154.1, 162.4; FTIR (neat): 1597, 1506, 1456, 1437  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{23}\text{H}_{28}\text{N}_3\text{O}_4$  [ $\text{M} + \text{H}$ ] $^+$  394.2125; found 394.2125; HPLC purity = 99%.

**(7-Chloro-1H-indol-2-yl)(4-(2-(4-methoxyphenoxy)ethyl)piperazin-1-yl)methanone (29).** 7-Chloro-1H-indole-2-carboxylic acid (23 mg, 0.12 mmol, 1.1 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as a yellow solid (31.8 mg, 0.077 mmol, 71% yield).  $R_f$  = 0.68 (10% MeOH in  $\text{CH}_2\text{Cl}_2$ ); mp = 125–130 °C.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.68 (t,  $J$  = 5.1 Hz, 4H), 2.85 (t,  $J$  = 5.5 Hz, 2H), 3.77 (s, 3H), 3.87–4.04 (complex, 4H), 4.09 (t,  $J$  = 5.5 Hz, 2H), 6.79 (d,  $J$  = 2.2 Hz, 1H), 6.83–6.90 (m, 4H), 7.08 (t,  $J$  = 7.8 Hz, 1H), 7.28 (dd,  $J$  = 0.9, 7.6 Hz, 1H), 7.54 (td,  $J$  = 0.9, 8.0 Hz, 1H), 9.23 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  41.1, 53.7, 55.9, 57.4, 66.7, 105.9, 114.8, 115.7, 117.3, 120.6, 121.5, 123.7, 128.8, 130.1, 133.2, 152.9, 154.1, 161.8; FTIR (neat): 1604, 1505, 1435  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{25}\text{ClN}_3\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  414.1579; found 414.1578; HPLC purity >99.5%.

**(6-Chloro-1H-indol-2-yl)(4-(2-(4-methoxyphenoxy)ethyl)piperazin-1-yl)methanone (30).** 6-Chloro-1H-indole-2-carboxylic acid (24 mg, 0.12 mmol, 1.1 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as a white solid (31.2 mg, 0.075 mmol, 70% yield).  $R_f$  = 0.61 (10% MeOH in  $\text{CH}_2\text{Cl}_2$ ); mp = 165–168 °C.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.69 (t,  $J$  = 5.1 Hz, 4H), 2.85 (t,  $J$  = 5.5 Hz, 2H), 3.77 (s, 3H), 3.89–4.04 (m, 4H), 4.10 (t,  $J$  = 5.5 Hz, 2H), 6.75 (dd,  $J$  = 1.0, 2.1 Hz, 1H), 6.82–6.88 (complex, 4H), 7.10 (dd,  $J$  = 1.8, 8.5 Hz, 1H), 7.40–7.44 (m, 1H), 7.55 (d,  $J$  = 8.7 Hz, 1H), 9.46 (s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  53.7 ( $\times 2$  C), 55.9, 57.4, 66.7, 105.4, 111.7, 114.8, 115.7, 121.7, 122.9, 126.1, 130.1, 130.4, 136.0, 152.9, 154.1, 162.0; FTIR (neat): 1591, 1506, 1456, 1437  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{25}\text{ClN}_3\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  414.1579; found 414.1575; HPLC purity = 95%.

**(5-Chloro-1H-indol-2-yl)(4-(2-(4-methoxyphenoxy)ethyl)piperazin-1-yl)methanone (31).** 5-Chloro-1H-indole-2-carboxylic acid (22 mg, 0.11 mmol, 1.1 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as an off-white solid (30.6 mg, 0.074 mmol, 68% yield).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.68 (t,  $J$  = 5.1 Hz, 4H), 2.85 (t,  $J$  = 5.5 Hz, 2H), 3.77 (s, 3H), 3.88–4.02 (m, 4H), 4.09 (t,  $J$  = 5.5 Hz, 2H), 6.70 (dd,  $J$  = 0.9, 2.2 Hz, 1H), 6.81–6.88 (complex, 4H), 7.23 (dd,  $J$  = 2.0, 8.8 Hz, 1H), 7.35 (d,  $J$  = 8.7 Hz, 1H), 7.60 (d,  $J$  = 2.0 Hz, 1H), 9.41 (s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  53.7 ( $\times 2$  C), 55.9, 57.4, 66.7, 104.7, 113.0, 114.8, 115.7, 121.2, 125.0, 126.3, 128.5, 130.6, 134.1, 152.9, 154.1, 162.0; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{25}\text{ClN}_3\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  414.1579; found 414.1578; HPLC purity = 99.5%.

**(4-Chloro-1H-indol-2-yl)(4-(2-(4-methoxyphenoxy)ethyl)piperazin-1-yl)methanone (32).** 4-Chloro-1H-indole-2-carboxylic acid (21 mg, 0.11 mmol, 1.1 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as a white solid (29.9 mg, 0.072 mmol, 67% yield).  $R_f$  = 0.64 (10% MeOH in  $\text{CH}_2\text{Cl}_2$ ); mp = 139–141 °C.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ )  $\delta$  2.57 (t,  $J$  = 5.1 Hz, 4H), 2.73 (t,  $J$  = 5.7 Hz, 2H), 3.69 (s, 3H), 3.71–3.82 (m, 4H), 4.04 (t,  $J$  = 5.7 Hz, 2H), 6.76 (s, 1H), 6.82–6.91 (complex, 4H), 7.13 (dd,  $J$  = 1.0, 7.6 Hz, 1H), 7.16–7.23 (m, 1H), 7.40 (d,  $J$  = 8.0 Hz, 1H), 11.97 (s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{DMSO}-d_6$ )  $\delta$  53.1 ( $\times 2$  C), 55.4, 56.6, 65.9, 101.6, 111.3, 114.6, 115.4, 119.4, 124.0, 125.2, 125.4, 131.0, 136.6, 152.5, 153.4, 161.3; FTIR (neat): 1599, 1505, 1458, 1437  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{25}\text{ClN}_3\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  414.1579; found 414.1581; HPLC purity >99.5%.

**(4-(2-(4-Methoxyphenoxy)ethyl)piperazin-1-yl)(1-methyl-1H-indol-2-yl)methanone (33).** 1-(2-(4-Methoxyphenoxy)ethyl)piperazine (109 mg, 0.461 mmol) and 1-methyl-1H-indole-2-carbonyl chloride (107 mg, 0.554 mmol, 1.2 equiv) were reacted according to General Procedure D to afford the acylated piperazine product as a tan oil (117 mg, 0.297 mmol, 65% yield).  $R_f$  = 0.81 (10% MeOH in  $\text{CH}_2\text{Cl}_2$ ).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.53–2.72 (m, 4H), 2.84

(t, *J* = 5.5 Hz, 2H), 3.77 (s, 3H), 3.84 (s, 3H), 3.74–3.87 (m, 4H), 4.08 (t, *J* = 5.5 Hz, 2H), 6.60 (d, *J* = 0.9 Hz, 1H), 6.80–6.88 (complex, 4H), 7.14 (ddd, *J* = 1.0, 6.9, 8.0 Hz, 1H), 7.30 (ddd, *J* = 1.2, 6.9, 8.3 Hz, 1H), 7.36 (td, *J* = 0.9, 8.4 Hz, 1H), 7.62 (d, *J* = 8.0 Hz, 1H); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 31.1, 53.7 (×2 C), 55.7, 57.2, 66.5, 103.6, 109.8, 114.7, 115.6, 120.3, 121.5, 123.3, 126.4, 131.6, 137.9, 152.7, 154.0, 163.0; FTIR (neat): 1624, 1506, 1462, 1438 cm<sup>-1</sup>; HRMS (*m/z*): calcd for C<sub>23</sub>H<sub>28</sub>N<sub>3</sub>O<sub>3</sub> [M + H]<sup>+</sup> 394.2125; found 394.2146; HPLC purity = 98.2%.

(4-(2-(4-Methoxyphenoxy)ethyl)piperazin-1-yl)(3-methyl-1H-indol-2-yl)methanone (34). 3-Methyl-1H-indole-2-carboxylic acid (19 mg, 0.11 mmol, 1.0 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as a light yellow solid (20.4 mg, 0.052 mmol, 48% yield). *R*<sub>f</sub> = 0.73 (10% MeOH in CH<sub>2</sub>Cl<sub>2</sub>); mp = 105–123 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 2.26 (s, 3H), 2.48–2.55 (m, 4H), 2.72 (t, *J* = 5.7 Hz, 2H), 3.52–3.61 (m, 4H), 3.69 (s, 3H), 4.02 (t, *J* = 5.7 Hz, 2H), 6.84–6.89 (complex, 4H), 7.03 (ddd, *J* = 1.0, 6.9, 7.9 Hz, 1H), 7.16 (ddd, *J* = 1.2, 7.0, 8.2 Hz, 1H), 7.33 (d, *J* = 8.2 Hz, 1H), 7.51–7.58 (m, 1H), 11.19 (s, 1H); <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>) δ 9.3, 53.2 (×2 C), 55.3, 56.5, 65.8, 109.9, 111.6, 114.5, 115.3, 118.9, 119.2, 122.8, 127.4, 127.7, 135.6, 152.4, 153.3, 163.3; FTIR (neat): 1603, 1506, 1451, 1440 cm<sup>-1</sup>; HRMS (*m/z*): calcd for C<sub>23</sub>H<sub>28</sub>N<sub>3</sub>O<sub>3</sub> [M + H]<sup>+</sup> 394.2125; found 394.2126; HPLC purity = 99.2%.

(4-(2-(4-Methoxyphenoxy)ethyl)piperazin-1-yl)(1H-pyrrolo[2,3-*b*]pyridin-2-yl)methanone (35). 1H-Pyrrolo[2,3-*b*]pyridine-2-carboxylic acid (20 mg, 0.12 mmol, 1.1 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as a white solid (27.3 mg, 0.072 mmol, 66% yield). *R*<sub>f</sub> = 0.71 (10% MeOH in CH<sub>2</sub>Cl<sub>2</sub>); mp = 167–176 °C; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 2.69 (t, *J* = 5.1 Hz, 4H), 2.85 (t, *J* = 5.5 Hz, 2H), 3.77 (s, 3H), 3.90–4.03 (m, 4H), 4.09 (t, *J* = 5.5 Hz, 2H), 6.72 (d, *J* = 1.4 Hz, 1H), 6.81–6.92 (complex, 4H), 7.13 (dd, *J* = 4.7, 7.9 Hz, 1H), 7.98 (dd, *J* = 1.6, 7.9 Hz, 1H), 8.50 (dd, *J* = 1.6, 4.7 Hz, 1H), 10.73 (s, 1H); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 53.6, 55.7, 57.2, 66.5, 103.4, 114.7, 115.6, 116.9, 119.8, 129.9, 130.3, 146.0, 147.7, 152.8, 154.0, 162.0; FTIR (neat): 1618, 1579, 1506, 1436 cm<sup>-1</sup>; HRMS (*m/z*): calcd for C<sub>21</sub>H<sub>25</sub>N<sub>4</sub>O<sub>3</sub> [M + H]<sup>+</sup> 381.1921; found 381.1921; HPLC purity >99.5%.

(4-(2-(4-Methoxyphenoxy)ethyl)piperazin-1-yl)(1H-pyrrolo[2,3-*c*]pyridin-2-yl)methanone (36). 1H-Pyrrolo[2,3-*c*]pyridine-2-carboxylic acid (20 mg, 0.12 mmol, 1.1 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as a white solid (30.5 mg, 0.080 mmol, 74% yield). *R*<sub>f</sub> = 0.28 (10% MeOH in CH<sub>2</sub>Cl<sub>2</sub>); mp = 163–165 °C. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 2.69–2.74 (m, 4H), 2.86 (t, *J* = 5.4 Hz, 2H), 3.77 (s, 3H), 3.90–4.03 (m, 4H), 4.10 (t, *J* = 5.5 Hz, 2H), 6.70–6.97 (complex, 5H), 7.34 (d, *J* = 5.8 Hz, 1H), 8.37 (d, *J* = 5.8 Hz, 1H), 8.99 (d, *J* = 1.1 Hz, 1H), 10.02 (s, 1H); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 53.6, 55.7, 57.2, 66.5, 104.3, 106.8, 114.7, 115.6, 124.6, 130.3, 139.1, 142.7, 145.5, 152.7, 154.0, 161.7; FTIR (neat): 1609, 1578, 1506 cm<sup>-1</sup>; HRMS (*m/z*): calcd for C<sub>21</sub>H<sub>25</sub>N<sub>4</sub>O<sub>3</sub> [M + H]<sup>+</sup> 381.1921; found 381.1926; HPLC purity = 99.5%.

(4-(2-(4-Methoxyphenoxy)ethyl)piperazin-1-yl)(1H-pyrrolo[3,2-*c*]pyridin-2-yl)methanone (37). 1H-Pyrrolo[3,2-*c*]pyridine-2-carboxylic acid (23 mg, 0.14 mmol, 1.3 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as a light yellow solid (30.3 mg, 0.080 mmol, 74% yield). *R*<sub>f</sub> = 0.28 (10% MeOH in CH<sub>2</sub>Cl<sub>2</sub>); mp = 160–164 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 2.57 (t, *J* = 5.1 Hz, 4H), 2.74 (t, *J* = 5.7 Hz, 2H), 3.69 (s, 3H), 3.70–3.83 (m, 4H), 4.04 (t, *J* = 5.7 Hz, 2H), 6.82–6.91 (complex, 4H), 6.92–7.01 (m, 1H), 7.36 (d, *J* = 5.8 Hz, 1H), 8.22 (d, *J* = 5.8 Hz, 1H), 8.90 (d, *J* = 1.2 Hz, 1H), 11.99 (s, 1H); <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>)

δ 53.1, 55.3, 56.5, 65.8, 103.1, 107.0, 114.5, 115.3, 124.0, 131.0, 138.9, 141.5, 144.7, 152.4, 153.3, 161.3; FTIR (neat): 1608, 1575, 1505 cm<sup>-1</sup>; HRMS (*m/z*): calcd for C<sub>21</sub>H<sub>25</sub>N<sub>4</sub>O<sub>3</sub> [M + H]<sup>+</sup> 381.1921; found 381.1926; HPLC purity = 99.7%.

(4-(2-(4-Methoxyphenoxy)ethyl)piperazin-1-yl)(1H-pyrrolo[3,2-*b*]pyridin-2-yl)methanone (38). 1H-Pyrrolo[3,2-*b*]pyridine-2-carboxylic acid (19 mg, 0.12 mmol, 1.1 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as a white solid (26.9 mg, 0.071 mmol, 65% yield). *R*<sub>f</sub> = 0.34 (10% MeOH in CH<sub>2</sub>Cl<sub>2</sub>); mp = 165–166 °C. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 2.70 (t, *J* = 5.1 Hz, 4H), 2.86 (t, *J* = 5.5 Hz, 2H), 3.77 (s, 3H), 3.85–4.14 (m, 4H), 4.10 (t, *J* = 5.6 Hz, 2H), 6.75–6.92 (complex, 4H), 6.98 (d, *J* = 1.1 Hz, 1H), 7.20 (dd, *J* = 4.6, 8.3 Hz, 1H), 7.74 (d, *J* = 8.2 Hz, 1H), 8.53 (dd, *J* = 1.4, 4.6 Hz, 1H), 9.69 (s, 1H); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 53.6, 55.7, 57.2, 66.5, 105.7, 114.7, 115.6, 119.1, 119.2, 128.8, 131.7, 144.9, 145.3, 152.7, 154.0, 161.9; FTIR (neat): 1601, 1505, 1437, 1407 cm<sup>-1</sup>; HRMS (*m/z*): calcd for C<sub>21</sub>H<sub>25</sub>N<sub>4</sub>O<sub>3</sub> [M + H]<sup>+</sup> 381.1921; found 381.1929; HPLC purity = 99.0%.

(1H-Benzo[*d*]imidazol-2-yl)(4-(2-(4-methoxyphenoxy)ethyl)piperazin-1-yl)methanone (39). 1H-Benzo[*d*]imidazole-2-carboxylic acid (20 mg, 0.12 mmol, 1.1 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (26 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as a white solid (29.3 mg, 0.077 mmol, 71% yield). *R*<sub>f</sub> = 0.78 (10% MeOH in CH<sub>2</sub>Cl<sub>2</sub>); mp = 139–141 °C. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 2.70–2.81 (m, 4H), 2.86 (t, *J* = 5.6 Hz, 2H), 3.77 (s, 3H), 3.94 (t, *J* = 5.0 Hz, 2H), 4.11 (t, *J* = 5.6 Hz, 2H), 4.82 (t, *J* = 5.2 Hz, 2H), 6.75–6.95 (complex, 4H), 7.30–7.43 (m, 2H), 7.48–7.58 (m, 1H), 7.82 (d, *J* = 8.0 Hz, 1H), 11.00 (s, 1H); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 43.2, 46.7, 53.4, 54.2, 55.7, 57.2, 66.5, 111.7, 114.6, 115.6, 121.0, 123.1, 125.1, 132.5, 143.2, 145.3, 152.8, 154.0, 158.2; FTIR (neat): 1608, 1506, 1440, 1406 cm<sup>-1</sup>; HRMS (*m/z*): calcd for C<sub>21</sub>H<sub>25</sub>N<sub>4</sub>O<sub>3</sub> [M + H]<sup>+</sup> 381.1921; found 381.1919; HPLC purity = 99.8%.

Benzofuran-2-yl(4-(2-(4-methoxyphenoxy)ethyl)piperazin-1-yl)methanone (40). Benzofuran-2-carboxylic acid (21.0 mg, 0.13 mmol, 1.2 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (25.7 mg, 0.11 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as a white solid (25.3 mg, 0.067 mmol, 61% yield). *R*<sub>f</sub> = 0.68 (10% MeOH in CH<sub>2</sub>Cl<sub>2</sub>); mp = 113–116 °C. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 2.70–2.81 (m, 4H), 2.86 (t, *J* = 5.6 Hz, 2H), 3.77 (s, 3H), 3.94 (t, *J* = 5.0 Hz, 2H), 4.11 (t, *J* = 5.6 Hz, 2H), 4.82 (t, *J* = 5.2 Hz, 2H), 6.75–6.95 (complex, 4H), 7.30–7.43 (m, 2H), 7.48–7.58 (m, 1H), 7.82 (d, *J* = 8.0 Hz, 1H), 11.00 (s, 1H); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 53.4, 54.0, 55.7, 57.2, 66.5, 111.9, 112.0, 114.6, 115.6, 122.2, 123.6, 126.4, 127.0, 149.0, 152.8, 154.0, 154.6, 159.7; FTIR (neat): 1630, 1561, 1506, 1433 cm<sup>-1</sup>; HRMS (*m/z*): calcd for C<sub>22</sub>H<sub>25</sub>N<sub>2</sub>O<sub>4</sub> [M + H]<sup>+</sup> 381.1809; found 381.1814; HPLC purity = 98.6%.

Benzo[*b*]thiophen-2-yl(4-(2-(4-methoxyphenoxy)ethyl)piperazin-1-yl)methanone (41). Benzo[*b*]thiophen-2-carboxylic acid (25 mg, 0.15 mmol, 1.2 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (30 mg, 0.13 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as a white solid (27.0 mg, 0.077 mmol, 60% yield). *R*<sub>f</sub> = 0.68 (10% MeOH in CH<sub>2</sub>Cl<sub>2</sub>); mp = 57–59 °C. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 2.62–2.69 (m, 4H), 2.84 (t, *J* = 5.5 Hz, 2H), 3.77 (s, 3H), 3.78–3.88 (m, 4H), 4.08 (t, *J* = 5.6 Hz, 2H), 6.80–6.89 (m, 4H), 7.37–7.42 (m, 2H), 7.48 (d, *J* = 0.8 Hz, 1H), 7.79–7.83 (m, 1H), 7.84–7.87 (m, 1H); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 55.7, 57.2, 66.5, 114.6, 115.6, 122.4, 124.6, 124.8, 125.2, 125.8, 136.6, 138.6, 140.2, 152.7, 154.0, 163.8; FTIR (neat): 1619, 1505, 1458, 1437 cm<sup>-1</sup>; HRMS (*m/z*): calcd for C<sub>22</sub>H<sub>25</sub>N<sub>2</sub>O<sub>3</sub>S [M + H]<sup>+</sup> 397.1580; found 397.1587; HPLC purity = 99.5%.

Cyclohexyl(4-(2-(4-methoxyphenoxy)ethyl)piperazin-1-yl)methanone (42). Cyclohexanecarbonyl chloride (75 μL, 0.56 mmol, 1.2 equiv) and 1-(2-(4-methoxyphenoxy)ethyl)piperazine (110 mg,



0.47 mmol) were reacted according to General Procedure D to afford the acylated piperazine product as a yellow solid (101.3 mg, 0.293 mmol, 63% yield);  $R_f = 0.58$  (10% MeOH in  $\text{CH}_2\text{Cl}_2$ ); mp = 54–58 °C.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  1.17–1.32 (m, 4H), 1.44–1.59 (m, 2H), 1.69–1.87 (m, 4H), 2.39–2.61 (m, 5H), 2.80 (t,  $J = 5.6$  Hz, 2H), 3.58 (dt,  $J = 5.2, 60.8$  Hz, 4H), 3.77 (s, 3H), 4.07 (t,  $J = 5.6$  Hz, 2H), 6.78–6.88 (m, 4H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  25.8, 25.9, 29.4, 40.4, 41.5, 45.3, 53.4, 54.1, 55.7, 57.3, 66.5, 114.6, 115.6, 152.8, 154.0, 174.5; FTIR (neat): 1634, 1506, 1433  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{20}\text{H}_{31}\text{N}_2\text{O}_3$  [ $M + H$ ] $^+$  347.2329; found 347.2349; HPLC purity = 99.2%.

**1-(4-(2-(4-Methoxyphenoxy)ethyl)piperazin-1-yl)ethan-1-one (43).** Acetyl chloride (32.8  $\mu\text{L}$ , 0.461 mmol, 1.0 equiv), 1-(2-(4-methoxyphenoxy)ethyl)piperazine (109 mg, 0.461 mmol), and triethylamine (100  $\mu\text{L}$ , 0.717 mmol, 1.5 equiv) were reacted according to General Procedure D to afford the acylated piperazine product as a brown oil (79.6 mg, 0.287 mmol, 62% yield).  $R_f = 0.57$  (10% MeOH in  $\text{CH}_2\text{Cl}_2$ ).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.09 (s, 3H), 2.54 (t,  $J = 5.2$  Hz, 2H), 2.58 (t,  $J = 5.0$  Hz, 2H), 2.81 (t,  $J = 5.6$  Hz, 2H), 3.49 (t,  $J = 5.3$  Hz, 2H), 3.64 (t,  $J = 4.6$  Hz, 2H), 3.77 (s, 3H), 4.06 (t,  $J = 5.6$  Hz, 2H), 6.77–6.87 (m, 4H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  21.3, 41.4, 46.3, 53.2, 53.7, 55.7, 57.2, 66.5, 114.6, 115.6, 152.8, 154.0, 168.9; FTIR (neat): 1637, 1505, 1461, 1426  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{15}\text{H}_{23}\text{N}_2\text{O}_3$  [ $M + H$ ] $^+$  279.1703; found 279.1716; HPLC purity >99.5%.

**1-(2-(4-Methoxyphenoxy)ethyl)-4-((4-methoxyphenyl)sulfonyl)piperazine (44).** To a solution of 1-(2-(4-methoxyphenoxy)ethyl)piperazine (76 mg, 0.32 mmol) and triethylamine (90  $\mu\text{L}$ , 0.64 mmol, 2.0 equiv) in toluene (25 mL) at rt was added 4-methoxybenzene-1-sulfonyl chloride (67 mg, 0.32 mmol, 1.0 equiv). The reaction was capped and stirred for 19 h at rt. The reaction solvents were removed in vacuo, and the residue was partitioned between saturated aqueous  $\text{NaHCO}_3$  (40 mL) and  $\text{CH}_2\text{Cl}_2$  (2  $\times$  20 mL). The combined organic phases were dried over  $\text{Na}_2\text{SO}_4$  and purified by silica gel chromatography to afford the sulfonamide product as an off-white solid (71.4 mg, 0.176 mmol, 55% yield).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.65 (t,  $J = 5.0$  Hz, 4H), 2.76 (t,  $J = 5.5$  Hz, 2H), 3.03 (t,  $J = 4.9$  Hz, 2H), 3.75 (s, 3H), 3.86 (s, 3H), 3.98 (t,  $J = 5.5$  Hz, 2H), 6.77–6.81 (m, 4H), 6.98 (d,  $J = 8.9$  Hz, 2H), 7.68 (d,  $J = 9.0$  Hz, 2H);  $^{13}\text{C}$  NMR (101 MHz, APT pulse sequence,  $\text{CDCl}_3$ )  $\delta$  d: 55.7, 55.8, 114.3, 114.7, 115.7, 130.1; u: 46.1, 52.7, 57.0, 66.6, 127.0, 152.8, 154.1; HRMS ( $m/z$ ): calcd for  $\text{C}_{20}\text{H}_{27}\text{N}_2\text{O}_5\text{S}$  [ $M + H$ ] $^+$  407.1635; found 407.1635; HPLC purity >99.5%.

**1-(2-(4-Methoxyphenoxy)ethyl)-4-(4-methoxyphenyl)piperazine (45).** To a solution of 1-(4-methoxyphenyl)piperazine (39 mg, 0.20 mmol) and 1-(2-bromoethoxy)-4-methoxybenzene (52 mg, 0.22 mmol, 1.1 equiv) in MeCN (10 mL) was added triethylamine (40  $\mu\text{L}$ , 0.29 mmol, 1.4 equiv), and the reaction was stirred at 60 °C for 18 h. The reaction was filtered, and the solids were washed with  $\text{CH}_2\text{Cl}_2$  (2  $\times$  5 mL). The combined filtrates were evaporated, and the residue was purified by silica gel chromatography to afford the alkylated piperazine product as a tan solid (47.5 mg, 0.139 mmol, 68% yield).  $R_f = 0.45$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 156–159 °C.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.72–2.78 (m, 4H), 2.85 (t,  $J = 5.8$  Hz, 2H), 3.08–3.16 (m, 4H), 3.76 (s, 3H), 3.78 (s, 3H), 4.10 (t,  $J = 5.8$  Hz, 2H), 6.81–6.88 (m, 6H), 6.89–6.92 (m, 2H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  50.6, 53.8, 55.6, 55.7, 57.4, 66.6, 114.4, 114.6, 115.6, 118.2, 145.7, 152.9, 153.8, 153.9; FTIR (neat): 1508, 1457, 1441  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{20}\text{H}_{27}\text{N}_2\text{O}_3$  [ $M + H$ ] $^+$  343.2016; found 343.2032; HPLC purity = 97.1%.

**(4-(1-(4-Methoxyphenoxy)propan-2-yl)piperazin-1-yl)(4-methoxyphenyl)methanone (46).** A solution of (4-methoxyphenyl)-(piperazin-1-yl)methanone-2,2,2-trifluoroacetate (78 mg, 0.23 mmol, 1.0 equiv), 1-(4-methoxyphenoxy)propan-2-one (44 mg, 0.24 mmol, 1.0 equiv), acetic acid (4  $\mu\text{L}$ , 0.07 mmol, 0.3 equiv), and sodium triacetoxyborohydride (76 mg, 0.36 mmol, 1.5 equiv) in dichloroethane (2 mL) was heated at 50 °C for 7 days and cooled to rt. The reaction was diluted with  $\text{CH}_2\text{Cl}_2$  (4 mL) and washed with saturated aqueous  $\text{NaHCO}_3$  (2  $\times$  2 mL) and water (1  $\times$  2 mL). The combined organic layers were dried over  $\text{MgSO}_4$ , concentrated in vacuo, and

purified by silica gel chromatography to afford the reductive amination product as a tan solid (51.1 mg, 0.132 mmol, 57% yield).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  1.17 (d,  $J = 6.8$  Hz, 3H), 2.54–2.75 (m, 4H), 3.03 (dtd,  $J = 5.6, 6.8, 12.4$  Hz, 1H), 3.41–3.75 (br m, 4H), 3.77 (s, 3H), 3.83 (s, 3H), 3.83–3.86 (m, 1H), 4.00 (dd,  $J = 5.6, 9.6$  Hz, 1H), 6.83 (m, 4H), 6.87–6.95 (m, 2H), 7.35–7.42 (m, 2H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  13.3, 55.3, 55.7, 58.4, 70.6, 113.7, 114.6, 115.5, 127.9, 129.2, 152.9, 153.9, 160.7, 170.2; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{29}\text{N}_2\text{O}_4$  [ $M + H$ ] $^+$  385.2122; found 385.2135; HPLC purity = 99.6%.

**(4-(2-(4-Methoxyphenoxy)propyl)piperazin-1-yl)(4-methoxyphenyl)methanone (47).** 4-Methoxybenzoic acid (22 mg, 0.14 mmol, 1.2 equiv) and 1-(2-(4-methoxyphenoxy)propyl)piperazine 114d (29 mg, 0.12 mmol, 1.0 equiv) were reacted according to General Procedure E to afford the acylated piperazine product 47 as a yellow oil (30.3 mg, 0.0789 mmol, 68% yield).  $R_f = 0.41$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  1.28 (d,  $J = 6.1$  Hz, 3H), 2.50 (dd,  $J = 4.5, 13.3$  Hz, 1H), 2.53–2.63 (m, 4H), 2.71 (dd,  $J = 6.8, 13.3$  Hz, 1H), 3.28–3.74 (m, 4H), 3.77 (s, 3H), 3.83 (s, 3H), 4.42 (td,  $J = 4.6, 6.5$  Hz, 1H), 6.83 (qt,  $J = 2.8, 9.4$  Hz, 4H), 6.88–6.92 (m, 2H), 7.34–7.39 (m, 2H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  18.6, 55.3, 55.7, 63.5, 73.3, 113.7, 114.6, 117.6, 127.9, 129.1, 151.7, 154.1, 160.7, 170.2; FTIR (neat): 1625, 1606, 1503, 1427  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{29}\text{N}_2\text{O}_4$  [ $M + H$ ] $^+$  385.2122; found 385.2121; HPLC purity = 98.4%.

**(4-(2-(4-Methoxyphenoxy)ethyl)-3-methylpiperazin-1-yl)(4-methoxyphenyl)methanone (48).** (4-Methoxyphenyl)(3-methylpiperazin-1-yl)methanone (45.0 mg, 0.19 mmol) and 1-(2-bromoethoxy)-4-methoxybenzene (48.8 mg, 0.21 mmol, 1.1 equiv) were reacted according to General Procedure A and then purified by preparative, reverse-phase HPLC to afford the alkylated piperazine as a sticky, golden solid (20.1 mg, 0.052 mmol, 27% yield).  $R_f = 0.38$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  1.01–1.19 (m, 3H), 2.44–2.64 (m, 4H), 2.78 (dt,  $J = 5.8, 13.9$  Hz, 1H), 2.90–3.02 (m, 2H), 3.10 (dt,  $J = 5.9, 12.7$  Hz, 1H), 3.32 (ddd,  $J = 3.1, 9.9, 13.0$  Hz, 1H), 3.76 (s, 3H), 3.83 (s, 3H), 4.03 (t,  $J = 5.9$  Hz, 2H), 6.80–6.83 (m, 4H), 6.90 (d,  $J = 8.7$  Hz, 2H), 7.37 (d,  $J = 8.7$  Hz, 2H);  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  52.4, 55.1, 55.3, 55.7, 66.4, 113.7, 114.6, 115.5, 127.8, 129.1, 152.8, 153.9, 160.7, 170.2; FTIR (neat): 1627, 1608, 1507, 1429  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{29}\text{N}_2\text{O}_4$  [ $M + H$ ] $^+$  385.2122; found 385.2140; HPLC purity = 95.1%.

**(4-(2-(4-Methoxyphenoxy)ethyl)-2-methylpiperazin-1-yl)(4-methoxyphenyl)methanone (49).** A mixture of 1-(2-bromoethoxy)-4-methoxybenzene (51.0 mg, 0.221 mmol, 1.1 equiv), (4-methoxyphenyl)(2-methylpiperazin-1-yl)methanone 111f (47.0 mg, 0.201 mmol, 1.0 equiv), and potassium carbonate (56.0 mg, 0.405 mmol, 2.0 equiv) in DMF (1 mL) was heated at 100 °C for 5 h and cooled to rt. The reaction was diluted with water (10 mL) and extracted with  $\text{CH}_2\text{Cl}_2$  (4  $\times$  2 mL). The combined organic layers were dried over  $\text{Na}_2\text{SO}_4$ , concentrated in vacuo, and purified by silica gel chromatography to afford the aryl ether product as a light yellow oil (28.4 mg, 0.0741 mmol, 37% yield).  $R_f = 0.48$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  1.35 (d,  $J = 6.8$  Hz, 3H), 2.14–2.24 (m, 1H), 2.34 (dd,  $J = 3.8, 11.3$  Hz, 1H), 2.67–2.84 (m, 3H), 2.90 (d,  $J = 11.3$  Hz, 1H), 3.29 (t,  $J = 12.5$  Hz, 1H), 3.77 (s, 3H), 3.83 (s, 3H), 3.93–4.15 (m, 3H), 4.24–4.60 (m, 1H), 6.80–6.87 (m, 4H), 6.88–6.94 (m, 2H), 7.31–7.38 (m, 2H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  16.6, 54.1, 55.3, 55.7, 57.2, 58.1, 66.8, 113.7, 114.6, 115.6, 128.5, 128.7, 152.8, 153.9, 160.5, 170.3; FTIR (neat): 1624, 1607, 1506, 1421  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{29}\text{N}_2\text{O}_4$  [ $M + H$ ] $^+$  385.2122; found 385.2121; HPLC purity = 99.7%.

**(4-(2-(4-Methoxyphenoxy)ethyl)piperidin-1-yl)(4-methoxyphenyl)methanone (50).** A mixture of (4-(2-chloroethyl)piperidin-1-yl)(4-methoxyphenyl)methanone (64.9 mg, 0.230 mmol, 1.0 equiv), 4-methoxyphenol (32.0 mg, 0.258 mmol, 1.1 equiv), and potassium carbonate (57.0 mg, 0.412 mmol, 1.8 equiv) in DMF (1 mL) was heated at 55 °C for 19 h, followed by 80 °C for 45 h and then cooled to rt. The reaction was diluted with  $\text{CH}_2\text{Cl}_2$  (10 mL) and washed with 1 M NaOH (3  $\times$  2 mL) and water (4  $\times$  2 mL). The organic layer was dried over  $\text{Na}_2\text{SO}_4$ , concentrated in vacuo, and

purified by silica gel chromatography to afford the aryl ether product as an off-white solid (12.2 mg, 0.033 mmol, 14% yield).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  1.12–1.35 (br m, 3H), 1.74 (q,  $J = 6.3$  Hz, 3H), 1.78–1.90 (m, 2H), 2.69–3.07 (br m, 2H), 3.77 (s, 3H), 3.83 (s, 3H), 3.97 (t,  $J = 6.2$  Hz, 2H), 4.47–4.79 (m, 1H), 6.79–6.85 (m, 4H), 6.88–6.93 (m, 2H), 7.35–7.40 (m, 2H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  33.2, 35.8, 55.3, 55.7, 65.8, 113.6, 114.7, 115.3, 128.4, 128.9, 153.0, 153.8, 160.6, 170.3; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{28}\text{NO}_4$  [ $\text{M} + \text{H}$ ] $^+$  370.2013; found 370.2004; HPLC purity = 98.1%.

**4-Methoxy-*N*-(2-((4-methoxyphenoxy)ethyl)(methylamino)ethyl)-*N*-methylbenzamide (51).** *N*-(2-((2-Chloroethyl)(methylamino)ethyl)-4-methoxy-*N*-methylbenzamide hydrochloride (70.7 mg, 0.220 mmol, 1.0 equiv), 4-methoxyphenol (38.0 mg, 0.306 mmol, 1.4 equiv), and potassium carbonate (101 mg, 0.731 mmol, 3.3 equiv) in DMF (1 mL) was heated at 50 °C for 7 h and cooled to rt. The reaction was diluted with  $\text{CH}_2\text{Cl}_2$  (10 mL) and washed with water (4  $\times$  2 mL). The organic layer was dried over  $\text{Na}_2\text{SO}_4$ , concentrated in vacuo, and purified by silica gel chromatography to afford the aryl ether product as a colorless oil (56.5 mg, 0.161 mmol, 73% yield).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.11–2.52 (br m, 3H), 2.58–2.92 (br m, 4H), 3.05 (s, 3H), 3.36–3.69 (br m, 2H), 3.77 (s, 3H), 3.81 (s, 3H), 3.97 (d,  $J = 17.9$  Hz, 2H), 6.78–6.84 (m, 4H), 6.84–6.90 (m, 2H), 7.32–7.41 (m, 2H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  43.0, 55.3, 55.7, 56.7, 66.8, 113.6, 114.6, 115.4, 128.8, 128.9, 152.9, 153.8, 160.4; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{29}\text{N}_2\text{O}_4$  [ $\text{M} + \text{H}$ ] $^+$  373.2122; found 373.2132; HPLC purity = 97.9%.

**(4-(2-(3-Methoxyphenoxy)ethyl)piperazin-1-yl)(1*H*-pyrrolo[2,3-*b*]pyridin-2-yl)methanone (52).** 1*H*-Pyrrolo[2,3-*b*]pyridine-2-carboxylic acid (19 mg, 0.12 mmol, 1.2 equiv) and 1-(2-(3-methoxyphenoxy)ethyl)piperazine (200  $\mu\text{L}$ , 0.488 M, 0.0980 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as a white solid (24.8 mg, 0.0653 mmol, 67% yield).  $R_f = 0.54$  (10% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 136–139 °C.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.68 (t,  $J = 5.1$  Hz, 4H), 2.86 (t,  $J = 5.5$  Hz, 2H), 3.78 (s, 3H), 3.85–4.03 (m, 4H), 4.12 (t,  $J = 5.5$  Hz, 2H), 6.43–6.55 (m, 3H), 6.68–6.74 (m, 1H), 7.09–7.22 (m, 2H), 7.98 (dd,  $J = 1.5, 7.9$  Hz, 1H), 8.54 (dd,  $J = 1.5, 4.7$  Hz, 1H), 11.57 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  26.4, 26.5, 46.3, 46.3, 55.3, 57.1, 65.8, 101.1, 105.7, 106.58, 106.63, 119.1, 119.2, 128.8, 129.9, 131.7, 144.9, 145.3, 159.8, 160.9, 161.9; FTIR (neat): 1602, 1530, 1492  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{25}\text{N}_4\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  381.1921; found 381.1928; HPLC purity = 99.4%.

**(4-(2-(3-Methoxyphenoxy)ethyl)piperazin-1-yl)(1*H*-pyrrolo[3,2-*c*]pyridin-2-yl)methanone (53).** 1*H*-Pyrrolo[3,2-*c*]pyridine-2-carboxylic acid (18 mg, 0.11 mmol, 1.1 equiv) and 1-(2-(3-methoxyphenoxy)ethyl)piperazine (200  $\mu\text{L}$ , 0.488 M, 0.0980 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as a sticky, off-white solid (22.9 mg, 0.0598 mmol, 61% yield);  $R_f = 0.11$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.70 (t,  $J = 5.1$  Hz, 4H), 2.87 (t,  $J = 5.4$  Hz, 2H), 3.78 (s, 3H), 3.85–4.06 (m, 4H), 4.12 (t,  $J = 5.4$  Hz, 2H), 6.45–6.56 (m, 3H), 6.82–6.89 (m, 1H), 7.18 (t,  $J = 8.2$  Hz, 1H), 7.31–7.37 (m, 1H), 8.36 (d,  $J = 5.9$  Hz, 1H), 8.96–9.02 (m, 1H), 10.32 (br s, 1H);  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  55.3, 57.0, 65.9, 101.2, 104.3, 106.5, 106.7, 106.9, 124.5, 129.9, 130.4, 139.2, 142.5, 145.4, 159.8, 160.9, 161.8; FTIR (neat): 1603, 1534, 1492, 1451, 1435  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{25}\text{N}_4\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  381.1921; found 381.1928; HPLC purity = 96.4%.

**(1*H*-Indol-2-yl)(4-(2-(3-methoxyphenoxy)ethyl)piperazin-1-yl)methanone (54).** (4-(2-Chloroethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and 4-methoxyphenol (19  $\mu\text{L}$ , 0.17 mmol, 1.6 equiv) were reacted according to General Procedure G to afford the aryl ether product as a light yellow solid (24.1 mg, 0.0638 mmol, 60% yield).  $R_f = 0.59$  (10% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 99–102 °C.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.64–2.71 (m, 4H), 2.87 (t,  $J = 5.5$  Hz, 2H), 3.79 (s, 3H), 3.88–4.04 (m, 4H), 4.13 (t,  $J = 5.5$  Hz, 2H), 6.49 (t,  $J = 2.4$  Hz, 1H), 6.50–6.54 (m, 2H), 6.78 (dd,  $J = 0.9, 2.1$  Hz, 1H), 7.14 (ddd,  $J = 1.0, 7.0, 8.0$  Hz, 1H), 7.19 (t,  $J = 8.2$  Hz, 1H), 7.28 (ddd,  $J = 1.2, 7.0, 8.2$  Hz, 1H), 7.43 (dq,  $J = 0.9, 8.2$  Hz, 1H), 7.65 (dq,  $J = 0.9, 8.1$  Hz, 1H), 9.25 (br s, 1H);  $^{13}\text{C}$  NMR

(126 MHz,  $\text{CDCl}_3$ )  $\delta$  41.0, 53.6, 55.3, 57.1, 65.8, 101.1, 105.3, 106.5, 106.7, 111.7, 120.6, 121.9, 124.4, 127.5, 129.2, 129.9, 135.6, 159.9, 160.8, 162.2; FTIR (neat): 1592, 1526, 1491  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{26}\text{N}_3\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  380.1969; found 380.1980; HPLC purity = 99.2%.

**(5-Methoxy-1*H*-indol-2-yl)(4-(2-(3-methoxyphenoxy)ethyl)piperazin-1-yl)methanone (55).** 5-Methoxy-1*H*-indole-2-carboxylic acid (20 mg, 0.10 mmol, 1.0 equiv) and 1-(2-(3-methoxyphenoxy)ethyl)piperazine (200  $\mu\text{L}$ , 0.488 M, 0.0980 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as an off-white solid (24.0 mg, 0.0588 mmol, 60% yield).  $R_f = 0.64$  (10% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 116–118 °C.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.63–2.73 (m, 4H), 2.86 (t,  $J = 5.6$  Hz, 2H), 3.79 (s, 3H), 3.85 (s, 3H), 3.90–4.02 (m, 4H), 4.13 (t,  $J = 5.5$  Hz, 2H), 6.48 (t,  $J = 2.4$  Hz, 1H), 6.50–6.55 (m, 2H), 6.70 (dd,  $J = 0.9, 2.2$  Hz, 1H), 6.96 (dd,  $J = 2.5, 8.9$  Hz, 1H), 7.05 (d,  $J = 2.5$  Hz, 1H), 7.19 (t,  $J = 8.2$  Hz, 1H), 7.32 (dt,  $J = 0.8, 8.9$  Hz, 1H), 9.21 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  53.6, 55.3, 55.7, 57.1, 65.8, 101.1, 102.2, 104.9, 106.5, 106.7, 112.6, 115.7, 127.8, 129.7, 129.9, 130.9, 154.6, 159.9, 160.8, 162.2; FTIR (neat): 1591, 1525, 1492  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{23}\text{H}_{28}\text{N}_3\text{O}_4$  [ $\text{M} + \text{H}$ ] $^+$  410.2074; found 410.2071; HPLC purity >99.5%.

**(1*H*-Benzo[*d*]imidazol-2-yl)(4-(2-(3-methoxyphenoxy)ethyl)piperazin-1-yl)methanone (56).** 1*H*-Benzo[*d*]imidazole-2-carboxylic acid (17 mg, 0.11 mmol, 1.1 equiv) and 1-(2-(3-methoxyphenoxy)ethyl)piperazine (200  $\mu\text{L}$ , 0.488 M, 0.0980 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as a white solid (22.7 mg, 0.0598 mmol, 61% yield).  $R_f = 0.38$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 50–65 °C.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.75 (q,  $J = 4.6$  Hz, 4H), 2.88 (t,  $J = 5.6$  Hz, 2H), 3.79 (s, 3H), 3.96 (t,  $J = 5.1$  Hz, 2H), 4.14 (t,  $J = 5.6$  Hz, 2H), 4.82 (t,  $J = 5.1$  Hz, 2H), 6.46–6.55 (m, 3H), 7.18 (t,  $J = 8.2$  Hz, 1H), 7.27–7.39 (m, 2H), 7.52 (d,  $J = 7.6$  Hz, 1H), 7.82 (d,  $J = 8.0$  Hz, 1H), 11.53 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  43.1, 46.6, 53.3, 54.1, 55.2, 57.0, 65.7, 101.0, 106.4, 106.5, 111.6, 121.0, 123.0, 125.0, 129.8, 132.5, 143.1, 145.2, 158.2, 159.8, 160.7; FTIR (neat): 1604, 1588, 1491, 1447  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{25}\text{N}_4\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  381.1921; found 381.1924; HPLC purity = 99.4%.

**(4-Methoxy-2-methylphenyl)(4-(2-(pyridin-3-yloxy)ethyl)piperazin-1-yl)methanone (57).** 4-Methoxy-2-methylbenzoic acid (26 mg, 0.15 mmol, 1.2 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (545  $\mu\text{L}$ , 0.235 M, 0.128 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as a light yellow solid (29.5 mg, 0.083 mmol, 65% yield).  $R_f = 0.29$  (10% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 91–93 °C.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.30 (s, 3H), 2.48 (t,  $J = 5.1$  Hz, 2H), 2.65 (t,  $J = 5.2$  Hz, 2H), 2.85 (t,  $J = 5.5$  Hz, 2H), 3.26–3.34 (m, 2H), 3.78–3.89 (m, 2H), 3.80 (s, 3H), 4.15 (t,  $J = 5.5$  Hz, 2H), 6.71–6.75 (m, 2H), 7.08–7.13 (m, 1H), 7.17–7.24 (m, 2H), 8.23 (dd,  $J = 1.7, 4.3$  Hz, 1H), 8.31–8.34 (m, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  19.4, 41.5, 46.9, 53.4, 54.1, 55.3, 57.0, 66.3, 111.2, 115.8, 121.2, 123.8, 127.4, 128.5, 136.3, 138.0, 142.4, 154.8, 159.8, 170.1; FTIR (neat): 1625, 1606, 1574, 1459  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{20}\text{H}_{26}\text{N}_3\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  356.1969; found 356.1970; HPLC purity = 98.6%.

**(6-Methoxy-1*H*-indol-2-yl)(4-(2-(pyridin-3-yloxy)ethyl)piperazin-1-yl)methanone (58).** 6-Methoxy-1*H*-indole-2-carboxylic acid (26 mg, 0.14 mmol, 1.1 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (200  $\mu\text{L}$ , 0.645 M, 0.129 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as a white solid (42.2 mg, 0.111 mmol, 86% yield).  $R_f = 0.40$  (10% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 142–144 °C.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.67 (t,  $J = 5.1$  Hz, 4H), 2.87 (t,  $J = 5.5$  Hz, 2H), 3.83 (s, 3H), 3.88–4.04 (m, 4H), 4.17 (t,  $J = 5.5$  Hz, 2H), 6.68–6.75 (m, 1H), 6.79 (dd,  $J = 1.4, 8.8$  Hz, 1H), 6.83–6.88 (m, 1H), 7.17–7.24 (m, 2H), 7.49 (d,  $J = 8.7$  Hz, 1H), 8.19–8.28 (m, 1H), 8.30–8.37 (m, 1H), 9.44 (s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  26.4, 26.5, 46.2, 46.3, 55.7, 57.1, 66.3, 93.7, 106.1, 111.9, 121.3, 121.8, 122.6, 123.9, 128.1, 136.6, 138.0, 142.4, 154.8, 158.3, 162.2; FTIR (neat): 1623, 1607, 1506  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{23}\text{H}_{28}\text{N}_3\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  381.1921; found 381.1905; HPLC purity = 95%.

(5-Methoxy-1H-indol-2-yl)(4-(2-(pyridin-3-yloxy)ethyl)piperazin-1-yl)methanone (59). 5-Methoxy-1H-indole-2-carboxylic acid (26 mg, 0.14 mmol, 1.1 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (200  $\mu$ L, 0.645 M, 0.129 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as a pale yellow solid (36.6 mg, 0.0963 mmol, 75% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.67 (t,  $J = 5.1$  Hz, 4H), 2.87 (t,  $J = 5.5$  Hz, 2H), 3.83 (s, 3H), 3.88–4.03 (m, 4H), 4.17 (t,  $J = 5.5$  Hz, 2H), 6.67–6.72 (m, 1H), 6.94 (dd,  $J = 2.4, 8.9$  Hz, 1H), 7.04 (d,  $J = 2.4$  Hz, 1H), 7.19–7.23 (m, 2H), 7.31 (d,  $J = 8.9$  Hz, 1H), 8.23 (dd,  $J = 2.2, 3.7$  Hz, 1H), 8.30–8.37 (m, 1H), 9.57 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  53.7, 55.7, 57.1, 66.3, 102.3, 104.9, 112.7, 115.7, 121.3, 123.9, 127.8, 129.7, 131.1, 138.0, 142.4, 154.6, 154.9, 162.4; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{25}\text{N}_4\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  381.1921; found 381.1922; HPLC purity = 98.7%.

(7-Methyl-1H-indol-2-yl)(4-(2-(pyridin-3-yloxy)ethyl)piperazin-1-yl)methanone (60). 7-Methyl-1H-indole-2-carboxylic acid (24 mg, 0.14 mmol, 1.1 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (200  $\mu$ L, 0.645 M, 0.129 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as an orange solid (33.5 mg, 0.0919 mmol, 71% yield).  $R_f = 0.26$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 49–60  $^\circ\text{C}$ .  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.49 (s, 3H), 2.67 (t,  $J = 4.8$  Hz, 4H), 2.88 (t,  $J = 5.5$  Hz, 2H), 3.83–4.06 (m, 4H), 4.18 (t,  $J = 5.5$  Hz, 2H), 6.78 (d,  $J = 2.0$  Hz, 1H), 6.99–7.10 (m, 2H), 7.18–7.24 (m, 2H), 7.44–7.51 (m, 1H), 8.24 (q,  $J = 2.5, 3.7$  Hz, 1H), 8.29–8.36 (m, 1H), 9.24 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  16.7, 53.7, 57.1, 66.3, 105.8, 119.5, 120.9, 121.2, 121.4, 123.9, 124.7, 127.0, 128.9, 135.4, 138.0, 142.4, 154.9, 162.5; FTIR (neat): 1601, 1574, 1536, 1429  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{25}\text{N}_4\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  365.1972; found 365.1974; HPLC purity = 98.2%.

(6-Methyl-1H-indol-2-yl)(4-(2-(pyridin-3-yloxy)ethyl)piperazin-1-yl)methanone (61). 6-Methyl-1H-indole-2-carboxylic acid (24 mg, 0.14 mmol, 1.1 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (200  $\mu$ L, 0.645 M, 0.129 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as a white solid (31.8 mg, 0.0872 mmol, 68% yield).  $R_f = 0.29$  (10% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 163–166  $^\circ\text{C}$ .  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.47 (s, 3H), 2.62–2.72 (m, 4H), 2.89 (t,  $J = 5.5$  Hz, 2H), 3.85–4.02 (m, 4H), 4.19 (t,  $J = 5.5$  Hz, 2H), 6.74 (dd,  $J = 0.9, 2.2$  Hz, 1H), 6.86–7.01 (m, 1H), 7.19–7.24 (m, 3H), 7.46–7.64 (m, 1H), 8.23–8.25 (m, 1H), 8.33–8.35 (m, 1H), 9.07 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  21.9, 53.6, 57.1, 66.3, 105.3, 111.4, 121.3, 121.5, 122.7, 123.9, 125.3, 128.6, 134.6, 136.0, 138.0, 142.4, 154.8, 162.3; FTIR (neat): 1596, 1573, 1522, 1427  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{25}\text{N}_4\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  365.1972; found 365.1974; HPLC purity >99.5%.

(5-Methyl-1H-indol-2-yl)(4-(2-(pyridin-3-yloxy)ethyl)piperazin-1-yl)methanone (62). 5-Methyl-1H-indole-2-carboxylic acid (24 mg, 0.14 mmol, 1.1 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (200  $\mu$ L, 0.645 M, 0.129 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as an off-white solid (38.2 mg, 0.105 mmol, 81% yield).  $R_f = 0.29$  (10% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 145–147  $^\circ\text{C}$ .  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.44 (s, 3H), 2.64–2.73 (m, 4H), 2.89 (t,  $J = 5.5$  Hz, 2H), 3.82–4.02 (m, 4H), 4.19 (t,  $J = 5.5$  Hz, 2H), 6.69 (dd,  $J = 0.9, 2.2$  Hz, 1H), 7.11 (dd,  $J = 1.6, 7.1$  Hz, 1H), 7.21–7.24 (m, 2H), 7.29–7.33 (m, 1H), 7.40–7.42 (m, 1H), 8.22–8.26 (m, 1H), 8.33–8.35 (m, 1H), 9.14 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  21.5, 53.6, 57.0, 66.2, 104.8, 111.4, 121.2, 121.3, 123.9, 126.4, 127.7, 129.2, 129.9, 134.0, 138.0, 142.4, 154.8, 162.4; FTIR (neat): 1597, 1574, 1529, 1427  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{25}\text{N}_4\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  365.1972; found 365.1977; HPLC purity = 99.5%.

(4-Methyl-1H-indol-2-yl)(4-(2-(pyridin-3-yloxy)ethyl)piperazin-1-yl)methanone (63). 4-Methyl-1H-indole-2-carboxylic acid (24 mg, 0.14 mmol, 1.1 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (200  $\mu$ L, 0.645 M, 0.129 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as a tan solid (38.4 mg, 0.105 mmol, 82% yield).  $R_f = 0.28$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 52–58  $^\circ\text{C}$ .  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.55 (s, 3H), 2.68

(t,  $J = 5.3$  Hz, 4H), 2.87 (t,  $J = 5.5$  Hz, 2H), 3.91–4.09 (m, 4H), 4.17 (t,  $J = 5.5$  Hz, 2H), 6.77 (d,  $J = 1.4$  Hz, 1H), 6.91 (d,  $J = 7.0$  Hz, 1H), 7.13–7.22 (m, 3H), 7.24–7.29 (m, 1H), 8.21–8.26 (m, 1H), 8.32–8.37 (m, 1H), 9.99 (s, 1H);  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  18.7, 53.6, 57.0, 66.3, 103.7, 109.4, 120.5, 121.3, 123.9, 124.4, 127.5, 128.6, 131.3, 135.7, 138.0, 142.4, 154.8, 162.6; FTIR (neat): 1600, 1585, 1519, 1427  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{25}\text{N}_4\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  365.1972; found 365.1976; HPLC purity = 98.1%.

(3-Methyl-1H-indol-2-yl)(4-(2-(pyridin-3-yloxy)ethyl)piperazin-1-yl)methanone (64). 3-Methyl-1H-indole-2-carboxylic acid (24 mg, 0.14 mmol, 1.1 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (200  $\mu$ L, 0.645 M, 0.129 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as a golden oil (40.9 mg, 0.112 mmol, 87% yield).  $R_f = 0.17$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.37 (s, 3H), 2.57–2.69 (m, 4H), 2.85 (t,  $J = 5.5$  Hz, 2H), 3.69–3.81 (m, 4H), 4.15 (t,  $J = 5.5$  Hz, 2H), 7.10–7.16 (m, 1H), 7.17–7.28 (m, 3H), 7.36 (d,  $J = 8.2$  Hz, 1H), 7.59 (d,  $J = 7.9$  Hz, 1H), 8.23 (dd,  $J = 1.8, 4.1$  Hz, 1H), 8.32 (d,  $J = 2.5$  Hz, 1H), 8.86 (br s, 1H);  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  10.2, 53.8, 57.0, 66.2, 111.5, 112.1, 119.8, 119.8, 121.3, 123.9, 124.0, 127.0, 128.2, 135.9, 138.0, 142.4, 154.8, 164.7; FTIR (neat): 1607, 1574, 1472, 1450, 1424  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{25}\text{N}_4\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  365.1972; found 365.1973; HPLC purity = 98.1%.

(1-Methyl-1H-indol-2-yl)(4-(2-(pyridin-3-yloxy)ethyl)piperazin-1-yl)methanone (65). 1-Methyl-1H-indole-2-carboxylic acid (24 mg, 0.14 mmol, 1.1 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (200  $\mu$ L, 0.645 M, 0.129 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as a yellow oil (41.6 mg, 0.114 mmol, 88% yield).  $R_f = 0.34$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.54–2.72 (m, 4H), 2.87 (t,  $J = 5.5$  Hz, 2H), 3.77–3.82 (m, 4H), 3.83 (s, 3H), 4.16 (t,  $J = 5.5$  Hz, 2H), 6.60 (d,  $J = 0.7$  Hz, 1H), 7.10–7.17 (m, 1H), 7.20–7.22 (m, 1H), 7.24–7.39 (m, 3H), 7.62 (d,  $J = 7.9$  Hz, 1H), 8.23 (dd,  $J = 2.1, 3.8$  Hz, 1H), 8.28–8.35 (m, 1H);  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  26.4, 31.1, 46.3, 57.0, 66.3, 103.7, 109.8, 120.3, 121.4, 121.5, 123.4, 123.9, 126.4, 131.5, 137.9, 142.3, 154.8, 163.1; FTIR (neat): 1625, 1574, 1522, 1463, 1422  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{25}\text{N}_4\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  365.1972; found 365.1972; HPLC purity = 98.7%.

(1-Methyl-1H-indol-3-yl)(4-(2-(pyridin-3-yloxy)ethyl)piperazin-1-yl)methanone (66). 1-Methyl-1H-indole-3-carboxylic acid (27 mg, 0.15 mmol, 1.2 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (545  $\mu$ L, 0.235 M, 0.128 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as a golden oil (29.5 mg, 0.0803 mmol, 63% yield).  $R_f = 0.23$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.63 (t,  $J = 5.1$  Hz, 4H), 2.87 (t,  $J = 5.6$  Hz, 2H), 3.77 (t,  $J = 4.9$  Hz, 4H), 3.82 (s, 3H), 4.17 (t,  $J = 5.6$  Hz, 2H), 7.18–7.24 (m, 3H), 7.27–7.31 (m, 1H), 7.35 (dt,  $J = 1.0, 8.2$  Hz, 1H), 7.43 (s, 1H), 7.69 (dt,  $J = 1.0, 7.9$  Hz, 1H), 8.23 (dd,  $J = 2.0, 4.0$  Hz, 1H), 8.32 (dd,  $J = 1.1, 2.5$  Hz, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  33.2, 41.0, 53.9, 57.1, 66.2, 109.8, 110.4, 120.5, 120.9, 121.2, 122.4, 123.8, 125.9, 131.7, 136.4, 138.0, 142.4, 154.8, 166.6; FTIR (neat): 1604, 1532, 1472, 1424  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{25}\text{N}_4\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  365.1972; found 365.1972; HPLC purity = 99.0%.

(7-Chloro-1H-indol-2-yl)(4-(2-(pyridin-3-yloxy)ethyl)piperazin-1-yl)methanone (67). 7-Chloro-1H-indole-2-carboxylic acid (27 mg, 0.14 mmol, 1.1 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (200  $\mu$ L, 0.645 M, 0.129 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as a golden oil (37.9 mg, 0.0978 mmol, 76% yield).  $R_f = 0.29$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.69 (t,  $J = 5.1$  Hz, 4H), 2.89 (t,  $J = 5.5$  Hz, 2H), 3.87–4.03 (m, 4H), 4.19 (t,  $J = 5.5$  Hz, 2H), 6.80 (d,  $J = 2.2$  Hz, 1H), 7.08 (t,  $J = 7.8$  Hz, 1H), 7.20–7.24 (m, 2H), 7.28 (dd,  $J = 0.9, 7.6$  Hz, 1H), 7.55 (dt,  $J = 0.9, 8.0$  Hz, 1H), 8.24 (dd,  $J = 2.3, 3.7$  Hz, 1H), 8.34 (dd,  $J = 1.3, 2.4$  Hz, 1H), 9.24 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  41.0, 57.0, 66.3, 105.8, 117.2, 120.4, 121.3, 121.4, 123.6, 123.9, 128.7, 129.9, 133.0, 138.0, 142.4, 154.8, 161.7; FTIR (neat): 1614, 1574, 1532, 1430  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{20}\text{H}_{22}\text{ClN}_4\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  386.1426; found 386.1431; HPLC purity = 98.7%.

(6-Chloro-1H-indol-2-yl)(4-(2-(pyridin-3-yloxy)ethyl)piperazin-1-yl)methanone (68). 6-Chloro-1H-indole-2-carboxylic acid (28 mg, 0.14 mmol, 1.1 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (200  $\mu$ L, 0.645 M, 0.129 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as an off-white solid (39.1 mg, 0.101 mmol, 78% yield).  $R_f$  = 0.25 (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 161–163 °C.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.70 (t,  $J$  = 5.2 Hz, 4H), 2.89 (t,  $J$  = 5.5 Hz, 2H), 3.83–4.09 (m, 4H), 4.19 (t,  $J$  = 5.5 Hz, 2H), 6.75 (dd,  $J$  = 0.9, 2.2 Hz, 1H), 7.10 (dd,  $J$  = 1.8, 8.5 Hz, 1H), 7.19–7.24 (m, 2H), 7.42 (dt,  $J$  = 0.8, 1.7 Hz, 1H), 7.55 (dt,  $J$  = 0.7, 8.5 Hz, 1H), 8.25 (dd,  $J$  = 2.3, 3.7 Hz, 1H), 8.34 (dd,  $J$  = 1.2, 2.5 Hz, 1H), 9.52 (br s, 1H).;  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  41.0, 53.6, 57.0, 66.3, 105.3, 111.6, 121.3, 121.6, 122.8, 123.9, 126.0, 129.9, 130.3, 136.0, 138.0, 142.4, 154.8, 162.0; FTIR (neat): 1606, 1574, 1522, 1430  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{20}\text{H}_{22}\text{ClN}_4\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  386.1426; found 386.1423; HPLC purity = 99.7%.

(5-Chloro-1H-indol-2-yl)(4-(2-(pyridin-3-yloxy)ethyl)piperazin-1-yl)methanone (69). 5-Chloro-1H-indole-2-carboxylic acid (25.2 mg, 0.129 mmol, 1.0 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (200  $\mu$ L, 0.645 M, 0.129 mmol) were reacted according to General Procedure E and purified by mass-directed, preparative HPLC to afford the acylated product as a white solid (33.6 mg, 0.087 mmol, 67% yield).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.69 (t,  $J$  = 5.1 Hz, 4H), 2.89 (t,  $J$  = 5.5 Hz, 2H), 3.89–4.04 (m, 4H), 4.19 (t,  $J$  = 5.5 Hz, 2H), 6.71 (dd,  $J$  = 0.8, 2.1 Hz, 1H), 7.21–7.24 (m, 3H), 7.35 (td,  $J$  = 0.6, 8.8 Hz, 1H), 7.60–7.62 (m, 1H), 8.24–8.25 (m, 1H), 8.32–8.35 (m, 1H), 9.39 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  53.6 (br, 2C), 57.0, 66.2, 104.6, 112.8, 121.0, 121.3, 123.9, 124.9, 126.1, 128.4, 130.4, 133.9, 138.0, 142.4, 154.8, 161.9; HRMS ( $m/z$ ): calcd for  $\text{C}_{20}\text{H}_{22}\text{ClN}_4\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  385.1426; found 385.1418; HPLC purity = 98.8%.

(4-Chloro-1H-indol-2-yl)(4-(2-(pyridin-3-yloxy)ethyl)piperazin-1-yl)methanone (70). 4-Chloro-1H-indole-2-carboxylic acid (27 mg, 0.14 mmol, 1.1 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (200  $\mu$ L, 0.645 M, 0.129 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as an off-white solid (36.4 mg, 0.0939 mmol, 73% yield).  $R_f$  = 0.22 (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 155–158 °C.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.65–2.76 (m, 4H), 2.90 (t,  $J$  = 5.5 Hz, 2H), 3.81–4.10 (m, 4H), 4.19 (t,  $J$  = 5.5 Hz, 2H), 6.86 (dd,  $J$  = 0.9, 2.4 Hz, 1H), 7.11–7.25 (m, 4H), 7.33 (dt,  $J$  = 1.0, 8.0 Hz, 1H), 8.20–8.28 (m, 1H), 8.35 (dd,  $J$  = 1.3, 2.4 Hz, 1H), 9.61 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  41.0, 53.6, 57.0, 66.3, 103.6, 110.4, 120.2, 121.3, 123.9, 124.9, 126.4, 127.0, 129.7, 136.2, 138.0, 142.4, 154.8, 161.8; FTIR (neat): 1604, 1572, 1527, 1427  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{20}\text{H}_{22}\text{ClN}_4\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  386.1426; found 386.1424; HPLC purity = 99.4%.

(4-(2-(Pyridin-3-yloxy)ethyl)piperazin-1-yl)(1H-pyrrolo[2,3-*b*]pyridin-2-yl)methanone (71). 1H-Pyrrolo[2,3-*b*]pyridine-2-carboxylic acid (25 mg, 0.15 mmol, 1.2 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (200  $\mu$ L, 0.645 M, 0.129 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as a white solid (39.9 mg, 0.113 mmol, 88% yield).  $R_f$  = 0.13 (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 133–135 °C.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.70 (t,  $J$  = 5.1 Hz, 4H), 2.90 (t,  $J$  = 5.4 Hz, 2H), 3.85–4.08 (m, 4H), 4.19 (t,  $J$  = 5.5 Hz, 2H), 6.73 (d,  $J$  = 1.7 Hz, 1H), 7.14 (dd,  $J$  = 4.7, 7.9 Hz, 1H), 7.22 (dt,  $J$  = 1.5, 4.5 Hz, 2H), 7.99 (dd,  $J$  = 1.6, 7.9 Hz, 1H), 8.24 (dd,  $J$  = 2.0, 3.9 Hz, 1H), 8.34 (dd,  $J$  = 1.2, 2.5 Hz, 1H), 8.51 (dd,  $J$  = 1.6, 4.7 Hz, 1H), 10.81 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  41.0, 53.6, 57.0, 66.3, 103.4, 116.9, 119.8, 121.3, 123.9, 129.9, 130.4, 138.0, 142.4, 146.0, 147.7, 154.8, 162.0; FTIR (neat): 1618, 1575, 1521, 1430  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{19}\text{H}_{22}\text{N}_5\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  352.1768; found 352.1769; HPLC purity >99.5%.

(4-(2-(Pyridin-3-yloxy)ethyl)piperazin-1-yl)(1H-pyrrolo[2,3-*c*]pyridin-2-yl)methanone (72). 1H-Pyrrolo[2,3-*c*]pyridine-2-carboxylic acid (24 mg, 0.15 mmol, 1.1 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (200  $\mu$ L, 0.645 M, 0.129 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as a light yellow solid (35.7 mg, 0.101 mmol, 79% yield).  $R_f$  = 0.07 (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 166–172 °C.  $^1\text{H}$  NMR (400 MHz,

$\text{CDCl}_3$ )  $\delta$  2.71 (t,  $J$  = 5.2 Hz, 4H), 2.90 (t,  $J$  = 5.4 Hz, 2H), 3.86–4.07 (m, 4H), 4.20 (t,  $J$  = 5.4 Hz, 2H), 6.87 (dd,  $J$  = 1.0, 1.9 Hz, 1H), 7.20–7.25 (m, 2H), 7.34 (dt,  $J$  = 1.1, 5.8 Hz, 1H), 8.24–8.26 (m, 1H), 8.35 (dd,  $J$  = 1.4, 2.3 Hz, 1H), 8.38 (d,  $J$  = 5.8 Hz, 1H), 8.96–9.01 (m, 1H), 9.75 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  41.0, 53.6, 57.0, 66.3, 104.3, 106.8, 121.3, 123.9, 124.6, 130.3, 138.0, 139.0, 142.5, 142.8, 145.5, 154.8, 161.6; FTIR (neat): 1611, 1573, 1534, 1428  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{19}\text{H}_{22}\text{N}_5\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  352.1768; found 352.1766; HPLC purity >99.5%.

(4-(2-(Pyridin-3-yloxy)ethyl)piperazin-1-yl)(1H-pyrrolo[3,2-*c*]pyridin-2-yl)methanone (73). 1H-Pyrrolo[3,2-*c*]pyridine-2-carboxylic acid (22 mg, 0.14 mmol, 1.0 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (200  $\mu$ L, 0.645 M, 0.129 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as an off-white solid (35.9 mg, 0.103 mmol, 80% yield).  $R_f$  = 0.07 (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 163–170 °C.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.71 (t,  $J$  = 5.1 Hz, 4H), 2.91 (t,  $J$  = 5.5 Hz, 2H), 3.80–4.13 (m, 4H), 4.20 (t,  $J$  = 5.5 Hz, 2H), 6.87 (d,  $J$  = 1.4 Hz, 1H), 7.20–7.26 (m, 2H), 7.34 (dt,  $J$  = 1.1, 5.9 Hz, 1H), 8.24 (dd,  $J$  = 1.2, 1.7 Hz, 1H), 8.35 (dd,  $J$  = 1.2, 2.5 Hz, 1H), 8.38 (d,  $J$  = 5.8 Hz, 1H), 8.99 (d,  $J$  = 1.1 Hz, 1H), 9.90 (s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  41.0, 53.6, 57.0, 66.3, 104.3, 106.8, 121.3, 123.9, 124.6, 130.3, 138.0, 139.0, 142.5, 142.8, 145.5, 154.8, 161.7; FTIR (neat): 1610, 1573, 1534, 1427  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{19}\text{H}_{22}\text{N}_5\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  352.1768; found 352.1768; HPLC purity >99.5%.

(4-(2-(Pyridin-3-yloxy)ethyl)piperazin-1-yl)(1H-pyrrolo[3,2-*b*]pyridin-2-yl)methanone (74). 1H-Pyrrolo[3,2-*b*]pyridine-2-carboxylic acid (25 mg, 0.15 mmol, 1.2 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (200  $\mu$ L, 0.645 M, 0.129 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as a white solid (35.8 mg, 0.102 mmol, 79% yield).  $R_f$  = 0.11 (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 173–177 °C.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.71 (t,  $J$  = 5.1 Hz, 4H), 2.90 (t,  $J$  = 5.5 Hz, 2H), 3.76–4.13 (m, 4H), 4.19 (t,  $J$  = 5.5 Hz, 2H), 6.98 (dd,  $J$  = 0.9, 2.1 Hz, 1H), 7.18–7.25 (m, 3H), 7.74 (dt,  $J$  = 1.2, 8.2 Hz, 1H), 8.24 (dd,  $J$  = 1.9, 4.2 Hz, 1H), 8.35 (dd,  $J$  = 1.1, 2.5 Hz, 1H), 8.53 (dd,  $J$  = 1.4, 4.6 Hz, 1H), 9.72–9.82 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  53.5, 57.0, 66.2, 105.7, 119.1, 119.2, 121.3, 123.9, 128.8, 131.7, 137.9, 142.4, 145.0, 145.2, 154.8, 161.9; FTIR (neat): 1617, 1574, 1526, 1428  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{19}\text{H}_{22}\text{N}_5\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  352.1768; found 352.1766; HPLC purity = 99.5%.

(1H-Benzo[d]imidazol-2-yl)(4-(2-(pyridin-3-yloxy)ethyl)piperazin-1-yl)methanone (75). 1H-Benzo[d]imidazole-2-carboxylic acid (23 mg, 0.14 mmol, 1.1 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (200  $\mu$ L, 0.645 M, 0.129 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as an off-white solid (34.2 mg, 0.0972 mmol, 75% yield).  $R_f$  = 0.14 (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 163–164 °C.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.75 (dt,  $J$  = 5.1, 10.1 Hz, 4H), 2.90 (t,  $J$  = 5.5 Hz, 2H), 3.93 (t,  $J$  = 5.1 Hz, 2H), 4.20 (t,  $J$  = 5.5 Hz, 2H), 4.82 (t,  $J$  = 5.1 Hz, 2H), 7.23 (dd,  $J$  = 1.5, 3.5 Hz, 2H), 7.29–7.40 (m, 2H), 7.53 (d,  $J$  = 8.0 Hz, 1H), 7.82 (d,  $J$  = 8.0 Hz, 1H), 8.25 (dd,  $J$  = 2.5, 3.6 Hz, 1H), 8.35 (dd,  $J$  = 1.4, 2.3 Hz, 1H), 10.84 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  41.0, 43.2, 46.6, 53.4, 54.2, 57.0, 66.2, 111.7, 121.1, 121.3, 123.2, 123.9, 125.1, 132.5, 138.0, 142.4, 143.2, 145.3, 154.8, 158.2; FTIR (neat): 1618, 1574, 1489, 1428  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{19}\text{H}_{22}\text{N}_5\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  352.1768; found 352.1764; HPLC purity = 99.6%.

(1H-Indazol-3-yl)(4-(2-(pyridin-3-yloxy)ethyl)piperazin-1-yl)methanone (76). 1H-Indazole-3-carboxylic acid (25 mg, 0.15 mmol, 1.2 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (545  $\mu$ L, 0.235 M, 0.128 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as a tan solid (25.8 mg, 0.0739 mmol, 57% yield).  $R_f$  = 0.11 (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ). mp = 121–123 °C.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.64–2.76 (m, 4H), 2.89 (t,  $J$  = 5.6 Hz, 2H), 3.86–4.15 (m, 4H), 4.19 (t,  $J$  = 5.6 Hz, 2H), 7.20–7.23 (m, 2H), 7.25 (dd,  $J$  = 1.1, 6.9 Hz, 1H), 7.39–7.44 (m, 1H), 7.49 (dt,  $J$  = 0.9, 8.5 Hz, 1H), 8.13 (dt,  $J$  = 1.0, 8.2 Hz, 1H), 8.24 (dd,  $J$  = 1.9, 4.1 Hz, 1H), 8.34 (dd,  $J$  = 1.2, 2.5 Hz, 1H), 10.86 (br s, 1H).;  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  42.4, 46.9, 53.5, 54.2, 57.0, 66.2, 109.7,

121.3, 122.2, 122.4, 123.4, 123.9, 127.3, 138.0, 140.0, 140.5, 142.3, 154.9, 162.7; FTIR (neat): 1613, 1574, 1486, 1429  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{19}\text{H}_{22}\text{N}_3\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  352.1768; found 352.1768; HPLC purity = 97.5%.

**Benzofuran-2-yl(4-(2-(pyridin-3-yloxy)ethyl)piperazin-1-yl)methanone (77).** Benzofuran-2-carboxylic acid (25 mg, 0.15 mmol, 1.2 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (545  $\mu\text{L}$ , 0.235 M, 0.128 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as a sticky yellow solid (40.1 mg, 0.114 mmol, 89% yield).  $R_f$  = 0.27 (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.63–2.70 (m, 4H), 2.86 (t,  $J$  = 5.5 Hz, 2H), 3.76–3.96 (m, 4H), 4.16 (t,  $J$  = 5.5 Hz, 2H), 7.18–7.21 (m, 2H), 7.24–7.28 (m, 1H), 7.34–7.41 (m, 1H), 7.50 (d,  $J$  = 8.3 Hz, 1H), 7.63 (d,  $J$  = 7.8 Hz, 1H), 8.17–8.24 (m, 1H), 8.31 (br s, 1H);  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  26.3, 26.4, 46.2, 46.3, 57.0, 66.2, 111.8, 112.1, 121.3, 122.2, 123.6, 123.8, 126.5, 126.9, 138.0, 142.4, 148.9, 154.6, 154.8, 159.7; FTIR (neat): 1630, 1573, 1475, 1428  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{20}\text{H}_{22}\text{N}_3\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  352.1656; found 352.1663; HPLC purity = 99.2%.

**Benzo[*b*]thiophen-2-yl(4-(2-(pyridin-3-yloxy)ethyl)piperazin-1-yl)methanone (78).** Benzo[*b*]thiophene-2-carboxylic acid (27 mg, 0.15 mmol, 1.2 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (545  $\mu\text{L}$ , 0.235 M, 0.128 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as a light yellow oil (31.4 mg, 0.0855 mmol, 67% yield).  $R_f$  = 0.38 (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.59–2.73 (m, 4H), 2.86 (t,  $J$  = 5.5 Hz, 2H), 3.75–3.86 (m, 4H), 4.15 (t,  $J$  = 5.5 Hz, 2H), 7.20 (d,  $J$  = 2.7 Hz, 2H), 7.34–7.41 (m, 2H), 7.45–7.49 (m, 1H), 7.76–7.87 (m, 2H), 8.18–8.26 (m, 1H), 8.27–8.36 (m, 1H);  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  46.26, 46.30, 53.6, 57.0, 66.3, 121.3, 122.4, 123.8, 124.6, 124.8, 125.2, 125.8, 136.5, 138.0, 138.6, 140.1, 142.4, 154.8, 163.8; FTIR (neat): 1617, 1573, 1521, 1421  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{20}\text{H}_{22}\text{N}_3\text{O}_2\text{S}$  [ $\text{M} + \text{H}$ ] $^+$  368.1427; found 368.1427; HPLC purity = 99.0%.

**4-(2-(Pyridin-3-yloxy)ethyl)piperazin-1-yl(1*H*-pyrrol-2-yl)methanone (79).** 1*H*-Pyrrole-2-carboxylic acid (17 mg, 0.15 mmol, 1.2 equiv) and 1-(2-(pyridin-3-yloxy)ethyl)piperazine (545  $\mu\text{L}$ , 0.235 M, 0.128 mmol) were reacted according to General Procedure E to afford the acylated piperazine product as an off-white solid (19.7 mg, 0.0646 mmol, 50% yield).  $R_f$  = 0.17 (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 131–133  $^{\circ}\text{C}$ .  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.64 (t,  $J$  = 5.2 Hz, 4H), 2.87 (t,  $J$  = 5.5 Hz, 2H), 3.80–3.97 (m, 4H), 4.18 (t,  $J$  = 5.6 Hz, 2H), 6.25 (dt,  $J$  = 2.7, 3.8 Hz, 1H), 6.52 (ddd,  $J$  = 1.3, 2.5, 3.8 Hz, 1H), 6.92 (td,  $J$  = 1.2, 2.7 Hz, 1H), 7.16–7.25 (m, 2H), 8.24 (dd,  $J$  = 2.1, 3.7 Hz, 1H), 8.34 (dd,  $J$  = 1.3, 2.3 Hz, 1H), 9.53 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  53.6, 57.1, 66.2, 109.6, 112.1, 120.9, 121.3, 123.8, 124.5, 138.0, 142.4, 154.8, 161.6; FTIR (neat): 1584, 1573, 1465, 1426  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{16}\text{H}_{21}\text{N}_4\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  301.1659; found 301.1667; HPLC purity = 98.3%.

**1*H*-Indol-2-yl(4-(2-phenoxyethyl)piperazin-1-yl)methanone (80).** (4-(2-Chloroethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (37.1 mg, 0.128 mmol) and phenol (19.3 mg, 0.205 mmol, 1.6 equiv) were reacted according to General Procedure F and purified by silica gel chromatography to afford the aryl ether product as a light yellow solid (26.3 mg, 0.075 mmol, 59% yield).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.69 (t,  $J$  = 5.1 Hz, 4H), 2.88 (t,  $J$  = 5.6 Hz, 2H), 3.89–4.04 (m, 4H), 4.15 (t,  $J$  = 5.6 Hz, 2H), 6.78 (dd,  $J$  = 0.9, 2.1 Hz, 1H), 6.90–6.98 (m, 3H), 7.12–7.15 (m, 1H), 7.25–7.32 (m, 3H), 7.43 (dd,  $J$  = 0.8, 8.3 Hz, 1H), 7.65 (dd,  $J$  = 0.9, 8.0 Hz, 1H), 9.22 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  53.6 (br, 2C), 57.2, 65.8, 105.3, 111.7, 114.6, 120.6, 121.0, 121.8, 124.4, 127.4, 129.2, 129.5, 135.5, 158.6, 162.2; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{24}\text{N}_3\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  350.1863; found 350.1860; HPLC purity = 99.0%.

**4-(2-(Benzo[*d*][1,3]dioxol-5-yloxy)ethyl)piperazin-1-yl(1*H*-indol-2-yl)methanone (81).** (4-(2-Chloroethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and benzo[*d*][1,3]-dioxol-5-ol (25 mg, 0.18 mmol, 1.7 equiv) were reacted according to General Procedure G to afford the aryl ether product as a white solid (20.5 mg, 0.052 mmol, 49% yield).  $R_f$  = 0.34 (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 142–146  $^{\circ}\text{C}$ .  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ )  $\delta$  2.56 (t,  $J$  = 5.0

Hz, 4H), 2.72 (t,  $J$  = 5.7 Hz, 2H), 3.69–3.83 (m, 4H), 4.03 (t,  $J$  = 5.7 Hz, 2H), 5.95 (s, 2H), 6.38 (dd,  $J$  = 2.5, 8.5 Hz, 1H), 6.65 (d,  $J$  = 2.5 Hz, 1H), 6.76–6.85 (m, 2H), 7.04 (ddd,  $J$  = 1.0, 6.9, 8.0 Hz, 1H), 7.18 (ddd,  $J$  = 1.2, 7.0, 8.2 Hz, 1H), 7.41 (dd,  $J$  = 0.9, 8.2 Hz, 1H), 7.60 (dd,  $J$  = 1.0, 8.0 Hz, 1H), 11.56 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{DMSO}-d_6$ )  $\delta$  53.11, 53.13, 56.4, 66.1, 97.8, 100.9, 103.9, 105.7, 108.0, 112.0, 119.7, 121.3, 123.1, 126.7, 129.8, 135.8, 141.1, 147.9, 153.8, 161.9; FTIR 1600, 1527, 1487, 1437  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{24}\text{N}_3\text{O}_4$  [ $\text{M} + \text{H}$ ] $^+$  394.1761; found 394.1782; HPLC purity = 99.4%.

**1*H*-Indol-2-yl(4-(2-((4-methoxyphenyl)thio)ethyl)piperazin-1-yl)methanone (82).** (4-(2-Chloroethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and 4-methoxybenzenethiol (21  $\mu\text{L}$ , 0.17 mmol, 1.6 equiv) were reacted according to General Procedure G to afford the aryl thioether product as a tan solid (19.3 mg, 0.0487 mmol, 46% yield).  $R_f$  = 0.39 (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 114–122  $^{\circ}\text{C}$ .  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ )  $\delta$  2.46 (t,  $J$  = 5.1 Hz, 4H), 2.51–2.56 (m, 2H), 2.96–3.03 (m, 2H), 3.62–3.81 (m, 7H), 6.76 (dd,  $J$  = 0.9, 2.2 Hz, 1H), 6.88–6.94 (m, 2H), 7.04 (ddd,  $J$  = 1.0, 7.0, 8.0 Hz, 1H), 7.18 (ddd,  $J$  = 1.2, 7.0, 8.2 Hz, 1H), 7.30–7.37 (m, 2H), 7.41 (dq,  $J$  = 1.0, 8.3 Hz, 1H), 7.59 (dq,  $J$  = 0.9, 8.0 Hz, 1H), 11.55 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{DMSO}-d_6$ )  $\delta$  30.3, 51.4, 54.0, 55.9, 102.8, 110.9, 113.6, 118.6, 120.1, 122.0, 124.8, 125.6, 128.7, 130.8, 134.7, 157.1, 160.7; FTIR (neat) 1595, 1526, 1492, 1437  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{26}\text{N}_3\text{O}_2\text{S}$  [ $\text{M} + \text{H}$ ] $^+$  396.1740; found 396.1739; HPLC purity >99.5%.

**1*H*-Indol-2-yl(4-(2-(4-(methylthio)phenoxy)ethyl)piperazin-1-yl)methanone (83).** (4-(2-Chloroethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and 4-(methylthio)phenol (24 mg, 0.17 mmol, 1.6 equiv) were reacted according to General Procedure G to afford the aryl ether product as a tan residue (10.3 mg, 0.0265 mmol, 25% yield).  $R_f$  = 0.38 (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ).  $^1\text{H}$  NMR (500 MHz,  $\text{DMSO}-d_6$ )  $\delta$  2.40–2.42 (s, 3H), 2.57 (t,  $J$  = 5.1 Hz, 4H), 2.75 (t,  $J$  = 5.7 Hz, 2H), 3.72–3.82 (m, 4H), 4.09 (t,  $J$  = 5.7 Hz, 2H), 6.78 (dd,  $J$  = 0.9, 2.2 Hz, 1H), 6.92–6.96 (m, 2H), 7.04 (ddd,  $J$  = 1.0, 6.9, 7.9 Hz, 1H), 7.18 (ddd,  $J$  = 1.2, 6.9, 8.2 Hz, 1H), 7.22–7.25 (m, 2H), 7.41 (dq,  $J$  = 1.0, 8.2 Hz, 1H), 7.60 (dq,  $J$  = 0.9, 8.0 Hz, 1H), 11.57 (br s, 1H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{DMSO}-d_6$ )  $\delta$  16.5, 53.1, 56.4, 65.5, 103.9, 112.0, 115.3, 119.7, 121.3, 123.1, 126.7, 128.3, 129.0, 129.8, 135.8, 156.7, 161.9; FTIR (neat) 3247, 1598, 1574, 1525, 1427  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{26}\text{N}_3\text{O}_2\text{S}$  [ $\text{M} + \text{H}$ ] $^+$  396.1740; found 396.1754; HPLC purity = 98%.

**4-(2-(4-Ethoxyphenoxy)ethyl)piperazin-1-yl(1*H*-indol-2-yl)methanone (84).** (4-(2-Chloroethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and 4-ethoxyphenol (25 mg, 0.18 mmol, 1.7 equiv) were reacted according to General Procedure G to afford the aryl ether product as a white residue (18.6 mg, 0.0477 mmol, 45% yield).  $R_f$  = 0.35 (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ).  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ )  $\delta$  1.29 (t,  $J$  = 7.0 Hz, 3H), 2.56 (t,  $J$  = 5.1 Hz, 4H), 2.73 (t,  $J$  = 5.7 Hz, 2H), 3.67–3.82 (m, 4H), 3.94 (q,  $J$  = 7.0 Hz, 2H), 4.04 (t,  $J$  = 5.7 Hz, 2H), 6.78 (dd,  $J$  = 0.9, 2.2 Hz, 1H), 6.80–6.90 (m, 4H), 7.04 (ddd,  $J$  = 1.0, 6.9, 8.0 Hz, 1H), 7.18 (ddd,  $J$  = 1.2, 7.0, 8.2 Hz, 1H), 7.41 (dq,  $J$  = 1.0, 8.3 Hz, 1H), 7.60 (dq,  $J$  = 0.9, 8.0 Hz, 1H), 11.56 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{DMSO}-d_6$ )  $\delta$  14.6, 53.1, 56.4, 63.2, 65.7, 103.8, 111.9, 115.1, 115.2, 119.6, 121.2, 123.1, 126.7, 129.8, 135.8, 152.2, 152.5, 161.8; FTIR (neat) 1598, 1526, 1506, 1437  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{23}\text{H}_{28}\text{N}_3\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  394.2125; found 394.2139; HPLC purity >99.5%.

**1*H*-Indol-2-yl(4-(2-(4-(trifluoromethoxy)phenoxy)ethyl)piperazin-1-yl)methanone (85).** (4-(2-Chloroethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and 4-(trifluoromethoxy)phenol (30.3 mg, 0.170 mmol, 1.6 equiv) were reacted according to General Procedure G to afford the aryl ether product as an off-white solid (27.7 mg, 0.0647 mmol, 61% yield).  $R_f$  = 0.35 (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 147–150  $^{\circ}\text{C}$ .  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.65–2.70 (m, 4H), 2.87 (t,  $J$  = 5.5 Hz, 2H), 3.85–4.02 (m, 4H), 4.13 (t,  $J$  = 5.5 Hz, 2H), 6.78 (dd,  $J$  = 0.9, 2.2 Hz, 1H), 6.88–6.93 (m, 2H), 7.12–7.17 (m, 3H), 7.29 (ddd,  $J$  = 1.2, 7.0, 8.2 Hz, 1H), 7.43 (dq,  $J$  = 0.9, 8.2 Hz, 1H), 7.65 (dq,  $J$  = 0.9, 8.0 Hz, 1H), 9.23 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  53.6, 57.0,

66.4, 105.3, 111.7, 115.3, 117.5, 119.5, 120.6, 121.6, 121.9, 122.5, 123.6, 124.5, 127.5, 129.2, 135.6, 142.9 (q,  $J = 1.6, 2.5$  Hz), 157.1, 162.2; FTIR (neat) 1598, 1526, 1506, 1437  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{23}\text{F}_3\text{N}_3\text{O}_3$  [ $M + H$ ] $^+$  434.1686; found 434.1685; HPLC purity >99.5%.

**(1*H*-Indol-2-yl)(4-(2-(*p*-tolylloxy)ethyl)piperazin-1-yl)methanone (86).** (4-(2-Chloroethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and 4-methylphenol (18.6 mg, 0.172 mmol, 1.6 equiv) were reacted according to General Procedure G to afford the aryl ether product as a white solid (19.3 mg, 0.0535 mmol, 50% yield).  $R_f = 0.36$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 151–153 °C.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.29 (s, 3H), 2.65–2.71 (m, 4H), 2.86 (t,  $J = 5.6$  Hz, 2H), 3.85–4.04 (m, 4H), 4.12 (t,  $J = 5.6$  Hz, 2H), 6.78 (dd,  $J = 1.0, 2.2$  Hz, 1H), 6.79–6.84 (m, 2H), 7.06–7.11 (m, 2H), 7.14 (ddd,  $J = 1.0, 7.0, 8.0$  Hz, 1H), 7.26–7.31 (m, 1H), 7.42 (dq,  $J = 0.9, 8.2$  Hz, 1H), 7.65 (dq,  $J = 1.0, 8.0$  Hz, 1H), 9.20 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  20.5, 41.0, 53.6, 57.2, 65.9, 105.3, 111.7, 114.4, 120.6, 121.9, 124.4, 127.5, 129.2, 129.9, 130.2, 135.5, 156.5, 162.2; FTIR (neat) 1599, 1527, 1510, 1437  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{26}\text{N}_3\text{O}_2$  [ $M + H$ ] $^+$  364.2020; found 364.2038; HPLC purity = 99.5%.

**(4-(2-(4-Ethylphenoxy)ethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (87).** (4-(2-Chloroethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and 4-ethylphenol (22 mg, 0.18 mmol, 1.7 equiv) were reacted according to General Procedure G to afford the aryl ether product as a white solid (20.1 mg, 0.0535 mmol, 50% yield).  $R_f = 0.32$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 135–138 °C.  $^1\text{H}$  NMR (500 MHz,  $\text{DMSO}-d_6$ )  $\delta$  1.14 (t,  $J = 7.6$  Hz, 3H), 2.52–2.59 (m, 6H), 2.75 (t,  $J = 5.7$  Hz, 2H), 3.70–3.82 (m, 4H), 4.07 (t,  $J = 5.7$  Hz, 2H), 6.78 (dd,  $J = 0.9, 2.2$  Hz, 1H), 6.84–6.88 (m, 2H), 7.04 (ddd,  $J = 1.0, 6.9, 7.9$  Hz, 1H), 7.07–7.13 (m, 2H), 7.18 (ddd,  $J = 1.2, 6.9, 8.2$  Hz, 1H), 7.42 (dq,  $J = 0.9, 8.3$  Hz, 1H), 7.60 (dq,  $J = 0.9, 7.8$  Hz, 1H), 11.57 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{DMSO}-d_6$ )  $\delta$  15.9, 27.2, 53.1, 56.5, 65.3, 103.9, 112.0, 114.3, 119.7, 121.3, 123.1, 126.7, 128.6, 129.8, 135.7, 135.8, 156.4, 161.9; FTIR (neat) 1598, 1526, 1510, 1436  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{23}\text{H}_{28}\text{N}_3\text{O}_2$  [ $M + H$ ] $^+$  378.2176; found 378.2183; HPLC purity = 99.6%.

**(4-(2-(4-*tert*-Butylphenoxy)ethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (88).** (4-(2-Chloroethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and 4-(*tert*-butyl)phenol (27 mg, 0.18 mmol, 1.7 equiv) were reacted according to General Procedure G to afford the aryl ether product as a white solid (18.2 mg, 0.0477 mmol, 45% yield).  $R_f = 0.34$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 171–175 °C.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ )  $\delta$  1.25 (s, 9H), 2.57 (t,  $J = 5.1$  Hz, 4H), 2.75 (t,  $J = 5.7$  Hz, 2H), 3.70–3.83 (m, 4H), 4.08 (t,  $J = 5.7$  Hz, 2H), 6.78 (dd,  $J = 0.9, 2.2$  Hz, 1H), 6.84–6.90 (m, 2H), 7.04 (ddd,  $J = 1.0, 7.0, 8.0$  Hz, 1H), 7.18 (ddd,  $J = 1.2, 7.0, 8.3$  Hz, 1H), 7.25–7.30 (m, 2H), 7.41 (dq,  $J = 0.9, 8.3$  Hz, 1H), 7.60 (dq,  $J = 0.9, 8.1$  Hz, 1H), 11.57 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{DMSO}-d_6$ )  $\delta$  31.2, 33.6, 53.0, 56.4, 65.2, 103.8, 111.9, 113.8, 119.6, 121.2, 123.1, 125.9, 126.7, 129.8, 135.8, 142.6, 156.0, 161.8; FTIR (neat) 1599, 1512, 1459, 1436  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{25}\text{H}_{32}\text{N}_3\text{O}_2$  [ $M + H$ ] $^+$  406.2489; found 406.2488; HPLC purity = 99.6%.

**(4-(2-(4-Fluorophenoxy)ethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (89).** (4-(2-Chloroethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and 4-fluorophenol (22 mg, 0.19 mmol, 1.8 equiv) were reacted according to General Procedure G to afford the aryl ether product as an off-white solid (24.3 mg, 0.0668 mmol, 63% yield).  $R_f = 0.44$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 162–164 °C.  $^1\text{H}$  NMR (500 MHz,  $\text{DMSO}-d_6$ )  $\delta$  2.57 (t,  $J = 5.1$  Hz, 4H), 2.75 (t,  $J = 5.7$  Hz, 2H), 3.72–3.80 (m, 4H), 4.09 (t,  $J = 5.7$  Hz, 2H), 6.78 (dd,  $J = 0.9, 2.2$  Hz, 1H), 6.95–6.99 (m, 2H), 7.04 (ddd,  $J = 1.0, 7.0, 8.0$  Hz, 1H), 7.09–7.13 (m, 2H), 7.18 (ddd,  $J = 1.2, 6.9, 8.2$  Hz, 1H), 7.42 (dq,  $J = 0.9, 8.3$  Hz, 1H), 7.60 (dq,  $J = 0.9, 8.0$  Hz, 1H), 11.58 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{DMSO}-d_6$ )  $\delta$  52.8, 56.4, 65.9, 103.9, 112.0, 115.7 (d,  $J = 7.3$  Hz), 115.9, 119.7, 121.3, 123.1, 126.7, 129.8, 135.8, 154.7 (d,  $J = 1.6$  Hz), 161.9; FTIR (neat): 1596, 1527, 1504, 1436  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{23}\text{FN}_3\text{O}_2$  [ $M + H$ ] $^+$  368.1769; found 368.1780; HPLC purity = 99.7%.

**(4-(2-(4-Chlorophenoxy)ethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (90).** (4-(2-Chloroethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and 4-chlorophenol (25 mg, 0.19 mmol, 1.8 equiv) were reacted according to General Procedure G to afford the aryl ether product as a light yellow solid (24.1 mg, 0.0625 mmol, 59% yield).  $R_f = 0.43$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 170–173 °C.  $^1\text{H}$  NMR (500 MHz,  $\text{DMSO}-d_6$ )  $\delta$  2.56 (t,  $J = 5.1$  Hz, 4H), 2.75 (t,  $J = 5.6$  Hz, 2H), 3.64–3.86 (m, 4H), 4.11 (t,  $J = 5.7$  Hz, 2H), 6.78 (dd,  $J = 0.9, 2.2$  Hz, 1H), 6.96–7.01 (m, 2H), 7.04 (ddd,  $J = 1.0, 7.0, 8.0$  Hz, 1H), 7.18 (ddd,  $J = 1.2, 7.0, 8.3$  Hz, 1H), 7.30–7.34 (m, 2H), 7.41 (dq,  $J = 0.9, 8.2$  Hz, 1H), 7.60 (dq,  $J = 1.0, 8.0$  Hz, 1H), 11.57 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{DMSO}-d_6$ )  $\delta$  53.0, 56.2, 65.6, 103.8, 111.9, 116.2, 119.6, 121.2, 123.1, 124.1, 126.6, 129.1, 129.7, 135.8, 157.2, 161.8; FTIR (neat) 1596, 1526, 1490, 1436  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{23}\text{ClN}_3\text{O}_2$  [ $M + H$ ] $^+$  384.1473; found 384.1474; HPLC purity = 99.7%.

**(4-(2-(4-Bromophenoxy)ethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (91).** (4-(2-Chloroethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and 4-bromophenol (31 mg, 0.18 mmol, 1.7 equiv) were reacted according to General Procedure G to afford the aryl ether product as a light yellow solid (27.4 mg, 0.0646 mmol, 61% yield).  $R_f = 0.41$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 165–168 °C.  $^1\text{H}$  NMR (500 MHz,  $\text{DMSO}-d_6$ )  $\delta$  2.56 (q,  $J = 5.2$  Hz, 4H), 2.75 (t,  $J = 5.7$  Hz, 2H), 3.68–3.82 (m, 4H), 4.10 (t,  $J = 5.7$  Hz, 2H), 6.78 (dd,  $J = 0.9, 2.2$  Hz, 1H), 6.92–6.96 (m, 2H), 7.04 (ddd,  $J = 1.0, 6.9, 7.9$  Hz, 1H), 7.18 (ddd,  $J = 1.2, 6.9, 8.2$  Hz, 1H), 7.40–7.46 (m, 3H), 7.60 (dq,  $J = 0.9, 8.1$  Hz, 1H), 11.57 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{DMSO}-d_6$ )  $\delta$  53.1, 56.3, 65.7, 103.9, 111.9, 112.0, 116.8, 119.7, 121.3, 123.1, 126.7, 129.8, 132.1, 135.8; FTIR (neat) 1597, 1526, 1487, 1436  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{23}\text{BrN}_3\text{O}_2$  [ $M + H$ ] $^+$  429.0968; found 430.0954; HPLC purity = 99.6%.

**(1*H*-Indol-2-yl)(4-(2-(4-nitrophenoxy)ethyl)piperazin-1-yl)methanone (92).** (4-(2-Chloroethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and 4-nitrophenol (25 mg, 0.18 mmol, 1.7 equiv) were reacted according to General Procedure G to afford the aryl ether product as an off-white solid (5.3 mg, 0.0138 mmol, 13% yield).  $R_f = 0.40$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 188–204 °C.  $^1\text{H}$  NMR (500 MHz,  $\text{DMSO}-d_6$ )  $\delta$  2.55–2.60 (m, 4H), 2.81 (t,  $J = 5.6$  Hz, 2H), 3.68–3.82 (m, 4H), 4.28 (t,  $J = 5.6$  Hz, 2H), 6.78 (dd,  $J = 0.9, 2.2$  Hz, 1H), 7.04 (ddd,  $J = 1.0, 6.9, 7.9$  Hz, 1H), 7.16–7.21 (m, 3H), 7.42 (dq,  $J = 1.0, 8.2$  Hz, 1H), 7.60 (dq,  $J = 0.9, 8.0$  Hz, 1H), 8.19–8.24 (m, 2H), 11.57 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{DMSO}-d_6$ )  $\delta$  53.0, 56.1, 66.4, 103.9, 112.0, 115.1, 119.7, 121.3, 123.2, 125.9, 126.7, 129.8, 135.8, 140.8, 161.9, 163.8; FTIR (neat) 1592, 1509, 1437  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{21}\text{H}_{23}\text{N}_4\text{O}_4$  [ $M + H$ ] $^+$  395.1714; found 395.1712; HPLC purity >99.5%.

**(4-(2-(3,4-Dimethoxyphenoxy)ethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (93).** (4-(2-Chloroethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and 3,4-dimethoxyphenol (28 mg, 0.18 mmol, 1.7 equiv) were reacted according to General Procedure G to afford the aryl ether product as a colorless, sticky solid (25.5 mg, 0.0625 mmol, 59% yield).  $R_f = 0.36$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.63–2.73 (m, 4H), 2.85 (t,  $J = 5.5$  Hz, 2H), 3.84 (s, 3H), 3.85 (s, 3H), 3.92–4.05 (m, 4H), 4.09 (t,  $J = 5.5$  Hz, 2H), 6.40 (td,  $J = 2.8, 7.2, 8.0$  Hz, 1H), 6.54 (d,  $J = 2.8$  Hz, 1H), 6.75–6.81 (m, 2H), 7.13 (t,  $J = 7.5$  Hz, 1H), 7.27 (t,  $J = 7.6$  Hz, 1H), 7.43 (d,  $J = 8.3$  Hz, 1H), 7.64 (d,  $J = 8.0$  Hz, 1H), 9.59 (br s, 1H);  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  53.6, 55.9, 56.40, 56.44, 57.2, 66.3, 69.4, 101.0, 101.4, 103.7, 104.4, 105.2, 111.7, 111.8, 120.5, 121.8, 124.3, 127.4, 129.2, 143.7, 149.9, 153.2; FTIR (neat) 1596, 1510, 1437  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{23}\text{H}_{28}\text{N}_3\text{O}_4$  [ $M + H$ ] $^+$  410.2074; found 410.2088; HPLC purity >99.5%.

**(4-(2-(3,4-Dimethylphenoxy)ethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (94).** (4-(2-Chloroethyl)piperazin-1-yl)(1*H*-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and 3,4-dimethylphenol (24 mg, 0.20 mmol, 1.9 equiv) were reacted according to General Procedure G to afford the aryl ether product as a white solid (20.7 mg, 0.0548 mmol, 52% yield).  $R_f = 0.42$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 129–137 °C.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ )  $\delta$  2.13 (s, 3H), 2.18 (s, 3H), 2.56 (t,  $J = 5.0$  Hz, 4H), 2.73 (t,  $J = 5.7$  Hz, 2H), 3.65–3.85 (m, 4H),

4.05 (t,  $J = 5.7$  Hz, 2H), 6.66 (dd,  $J = 2.7, 8.3$  Hz, 1H), 6.74–6.81 (m, 2H), 6.98–7.09 (m, 2H), 7.18 (ddd,  $J = 1.2, 7.0, 8.2$  Hz, 1H), 7.41 (dd,  $J = 1.0, 8.2$  Hz, 1H), 7.56–7.70 (m, 1H), 11.56 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz, DMSO- $d_6$ )  $\delta$  18.4, 19.6, 53.1, 56.5, 65.2, 103.9, 111.4, 112.0, 115.8, 119.7, 121.3, 123.1, 126.7, 127.9, 129.8, 130.1, 135.8, 137.2, 156.5, 161.9; FTIR (neat) 1600, 1526, 1502, 1437  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{23}\text{H}_{28}\text{N}_3\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  378.2176; found 378.2181; HPLC purity = 99.2%.

(4-(2-(3-Chloro-4-methylphenoxy)ethyl)piperazin-1-yl)(1H-indol-2-yl)methanone (**95**). (4-(2-Chloroethyl)piperazin-1-yl)(1H-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and 3-chloro-4-methylphenol (25 mg, 0.17 mmol, 1.7 equiv) were reacted according to General Procedure G to afford the aryl ether product as a white solid (25.0 mg, 0.0636 mmol, 60% yield).  $R_f = 0.53$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 156–158 °C.  $^1\text{H}$  NMR (500 MHz, DMSO- $d_6$ )  $\delta$  2.24 (s, 3H), 2.26 (t,  $J = 5.1$  Hz, 4H), 2.74 (t,  $J = 5.6$  Hz, 2H), 3.68–3.83 (m, 4H), 4.10 (t,  $J = 5.7$  Hz, 2H), 6.78 (dd,  $J = 0.9, 2.2$  Hz, 1H), 6.86 (dd,  $J = 2.6, 8.4$  Hz, 1H), 7.02–7.06 (m, 2H), 7.18 (ddd,  $J = 1.2, 6.9, 8.2$  Hz, 1H), 7.24 (dd,  $J = 0.8, 8.4$  Hz, 1H), 7.41 (dq,  $J = 0.9, 8.2$  Hz, 1H), 7.60 (dq,  $J = 0.9, 8.0$  Hz, 1H), 11.57 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz, DMSO- $d_6$ )  $\delta$  18.6, 53.1, 56.3, 65.8, 103.9, 112.0, 113.8, 114.7, 119.7, 121.3, 123.1, 126.7, 127.0, 129.8, 131.6, 133.5, 135.8, 157.3, 161.9; FTIR (neat) 1599, 1526, 1495, 1437  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{25}\text{ClN}_3\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  398.1630; found 398.1640; HPLC purity = 99.3%.

(4-(2-(4-Chloro-3-methylphenoxy)ethyl)piperazin-1-yl)(1H-indol-2-yl)methanone (**96**). (4-(2-Chloroethyl)piperazin-1-yl)(1H-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and 4-chloro-3-methylphenol (26 mg, 0.18 mmol, 1.7 equiv) were reacted according to General Procedure G to afford the aryl ether product as a white solid (25.1 mg, 0.0636 mmol, 60% yield).  $R_f = 0.49$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 146–148 °C.  $^1\text{H}$  NMR (500 MHz, DMSO- $d_6$ )  $\delta$  2.29 (s, 3H), 2.56 (t,  $J = 5.1$  Hz, 4H), 2.75 (t,  $J = 5.7$  Hz, 2H), 3.68–3.83 (m, 4H), 4.09 (t,  $J = 5.7$  Hz, 2H), 6.78 (dd,  $J = 0.9, 2.2$  Hz, 1H), 6.79–6.83 (m, 1H), 6.98 (dd,  $J = 0.8, 3.1$  Hz, 1H), 7.04 (ddd,  $J = 1.0, 6.9, 8.0$  Hz, 1H), 7.18 (ddd,  $J = 1.1, 6.9, 8.2$  Hz, 1H), 7.29 (d,  $J = 8.8$  Hz, 1H), 7.41 (dq,  $J = 1.0, 8.4$  Hz, 1H), 7.60 (dq,  $J = 0.9, 8.0$  Hz, 1H), 11.57 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz, DMSO- $d_6$ )  $\delta$  19.7, 53.1, 56.3, 65.6, 103.9, 112.0, 113.6, 117.2, 119.7, 121.3, 123.1, 124.5, 126.7, 129.4, 129.8, 135.8, 136.4, 157.2, 161.9; FTIR (neat) 1597, 1526, 1480, 1436  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{25}\text{ClN}_3\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  398.1630; found 398.1630; HPLC purity >99.5%.

(4-(2-(2-Chloro-4-methoxyphenoxy)ethyl)piperazin-1-yl)(1H-indol-2-yl)methanone (**97**). (4-(2-Chloroethyl)piperazin-1-yl)(1H-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and 2-chloro-4-methoxyphenol (30 mg, 0.19 mmol, 1.8 equiv) were reacted according to General Procedure G to afford the aryl ether product as a white solid (26.6 mg, 0.0647 mmol, 61% yield).  $R_f = 0.46$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 157–159 °C.  $^1\text{H}$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  2.60 (t,  $J = 5.1$  Hz, 4H), 2.78 (t,  $J = 5.7$  Hz, 2H), 3.72 (s, 3H), 3.73–3.82 (m, 4H), 4.13 (t,  $J = 5.7$  Hz, 2H), 6.78 (dd,  $J = 0.9, 2.2$  Hz, 1H), 6.87 (dd,  $J = 3.0, 9.0$  Hz, 1H), 7.01–7.07 (m, 2H), 7.12 (d,  $J = 9.1$  Hz, 1H), 7.18 (ddd,  $J = 1.2, 7.0, 8.2$  Hz, 1H), 7.41 (dq,  $J = 0.9, 8.3$  Hz, 1H), 7.60 (dq,  $J = 0.9, 8.0$  Hz, 1H), 11.56 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz, DMSO- $d_6$ )  $\delta$  52.8, 55.7, 56.3, 67.5, 103.9, 112.0, 113.4, 115.3, 115.5, 119.7, 121.3, 122.0, 123.1, 126.7, 129.8, 135.8, 148.0, 153.5, 161.9; FTIR (neat) 1596, 1526, 1496, 1437  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{25}\text{ClN}_3\text{O}_3$  [ $\text{M} + \text{H}$ ] $^+$  414.1579; found 414.1594; HPLC purity = 99.5%.

(1H-Indol-2-yl)(4-(2-(pyridin-2-yloxy)ethyl)piperazin-1-yl)methanone (**98**). (4-(2-Chloroethyl)piperazin-1-yl)(1H-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and pyridine-2-ol (16 mg, 0.17 mmol, 1.6 equiv) were reacted according to General Procedure G to afford the pyridyl ether product as a light yellow, sticky solid (21.4 mg, 0.0615 mmol, 58% yield).  $R_f = 0.27$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ).  $^1\text{H}$  NMR (500 MHz, DMSO- $d_6$ )  $\delta$  2.49–2.54 (m, 4H), 2.61 (t,  $J = 6.4$  Hz, 2H), 3.68–3.78 (m, 4H), 4.01 (t,  $J = 6.4$  Hz, 2H), 6.21 (td,  $J = 1.4, 6.6$  Hz, 1H), 6.37 (ddd,  $J = 0.6, 1.4, 9.1$  Hz, 1H), 6.77 (dd,  $J = 0.9, 2.2$  Hz, 1H), 7.04 (ddd,  $J = 1.1, 6.9, 8.0$  Hz, 1H), 7.18 (ddd,  $J = 1.2, 6.9, 8.2$  Hz, 1H), 7.37–7.43 (m, 2H), 7.59 (dq,  $J = 0.9, 8.0$  Hz,

1H), 7.67 (ddd,  $J = 0.7, 2.1, 6.8$  Hz, 1H), 11.57 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz, DMSO- $d_6$ )  $\delta$  45.5, 52.8, 55.9, 103.9, 104.8, 112.0, 119.3, 119.7, 121.3, 123.2, 126.7, 129.8, 135.8, 139.7, 139.9, 161.4, 161.9; FTIR (neat) 1652, 1572, 1538, 1435  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{20}\text{H}_{23}\text{N}_4\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  351.1816; found 351.1844; HPLC purity = 99.4%.

(1H-Indol-2-yl)(4-(2-(pyridin-3-yloxy)ethyl)piperazin-1-yl)methanone (**99**). (4-(2-Chloroethyl)piperazin-1-yl)(1H-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and pyridine-2-ol (19 mg, 0.20 mmol, 1.9 equiv) were reacted according to General Procedure G to afford the pyridyl ether product as a tan solid (18.1 mg, 0.0517 mmol, 49% yield).  $R_f = 0.50$  (10% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 129–132 °C.  $^1\text{H}$  NMR (500 MHz, DMSO- $d_6$ )  $\delta$  2.58 (t,  $J = 5.1$  Hz, 5H), 2.78 (t,  $J = 5.6$  Hz, 2H), 3.69–3.85 (m, 4H), 4.19 (t,  $J = 5.6$  Hz, 2H), 6.78 (dd,  $J = 0.9, 2.2$  Hz, 1H), 7.04 (ddd,  $J = 1.0, 6.9, 7.9$  Hz, 1H), 7.18 (ddd,  $J = 1.2, 6.9, 8.2$  Hz, 1H), 7.33 (ddd,  $J = 0.7, 4.6, 8.4$  Hz, 1H), 7.41 (dd,  $J = 1.2, 2.8$  Hz, 1H), 7.43 (dd,  $J = 1.2, 2.9$  Hz, 1H), 7.60 (dd,  $J = 1.0, 8.0$  Hz, 1H), 8.17 (dd,  $J = 1.3, 4.6$  Hz, 1H), 8.31 (dd,  $J = 0.7, 3.0$  Hz, 1H), 11.58 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz, DMSO- $d_6$ )  $\delta$  53.1, 56.3, 65.6, 103.9, 112.0, 119.7, 121.0, 121.3, 123.1, 124.1, 126.7, 129.8, 135.8, 137.8, 141.7, 154.7, 161.9; FTIR (neat) 1598, 1574, 1525, 1427  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{20}\text{H}_{23}\text{N}_4\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  351.1816; found 351.1815; HPLC purity >99.5%.

(1H-Indol-2-yl)(4-(2-(pyridin-4-yloxy)ethyl)piperazin-1-yl)methanone (**100**). (4-(2-Chloroethyl)piperazin-1-yl)(1H-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and pyridin-4-ol (19 mg, 0.20 mmol, 1.9 equiv) were reacted according to General Procedure G to afford the pyridyl ether product as an off-white residue (9.0 mg, 0.025 mmol, 24% yield).  $R_f = 0.29$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ).  $^1\text{H}$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  2.57 (t,  $J = 5.1$  Hz, 4H), 2.78 (t,  $J = 5.6$  Hz, 2H), 3.67–3.85 (m, 4H), 4.20 (t,  $J = 5.6$  Hz, 2H), 6.78 (dd,  $J = 0.9, 2.2$  Hz, 1H), 6.96–7.01 (m, 2H), 7.04 (ddd,  $J = 1.0, 6.9, 8.0$  Hz, 1H), 7.18 (ddd,  $J = 1.2, 6.9, 8.2$  Hz, 1H), 7.41 (dq,  $J = 1.0, 8.2$  Hz, 1H), 7.58–7.62 (m, 1H), 8.34–8.41 (m, 2H), 11.56 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz, DMSO- $d_6$ )  $\delta$  53.0, 56.1, 65.3, 103.9, 110.4, 112.0, 119.7, 121.3, 123.1, 126.7, 129.8, 135.8, 150.9, 161.9, 164.2; FTIR (neat) 1638, 1608, 1549, 1436  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{20}\text{H}_{23}\text{N}_4\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  351.1816; found 351.1808; HPLC purity >99.5%.

(4-(2-(5-Chloropyridin-3-yloxy)ethyl)piperazin-1-yl)(1H-indol-2-yl)methanone (**101**). (4-(2-Chloroethyl)piperazin-1-yl)(1H-indol-2-yl)methanone (30.9 mg, 0.106 mmol) and 5-chloropyridin-3-ol (26 mg, 0.20 mmol, 1.9 equiv) were reacted according to General Procedure G to afford the pyridyl ether product as a white solid (23.1 mg, 0.0604 mmol, 57% yield).  $R_f = 0.38$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 184–186 °C.  $^1\text{H}$  NMR (500 MHz, DMSO- $d_6$ )  $\delta$  2.57 (t,  $J = 5.0$  Hz, 4H), 2.78 (t,  $J = 5.6$  Hz, 2H), 3.68–3.83 (m, 4H), 4.24 (t,  $J = 5.6$  Hz, 2H), 6.78 (dd,  $J = 0.9, 2.3$  Hz, 1H), 7.04 (ddd,  $J = 1.0, 6.9, 8.0$  Hz, 1H), 7.18 (ddd,  $J = 1.1, 6.9, 8.2$  Hz, 1H), 7.42 (dq,  $J = 1.0, 8.2$  Hz, 1H), 7.59–7.62 (m, 1H), 7.64–7.67 (m, 1H), 8.22 (d,  $J = 1.9$  Hz, 1H), 8.29 (d,  $J = 2.5$  Hz, 1H), 11.57 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz, DMSO- $d_6$ )  $\delta$  51.9, 55.0, 65.1, 102.8, 110.9, 118.6, 119.9, 120.2, 122.0, 125.6, 128.7, 130.0, 134.7, 135.7, 138.8, 154.1, 160.7; FTIR (neat) 1611, 1576, 1535, 1429  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{20}\text{H}_{22}\text{ClN}_4\text{O}_2$  [ $\text{M} + \text{H}$ ] $^+$  385.1426; found 385.1422; HPLC purity >99.5%.

(1H-Indol-2-yl)(4-(2-(4-methoxyphenoxy)propyl)piperazin-1-yl)methanone (**102**). 1H-Indole-2-carboxylic acid (24 mg, 0.15 mmol, 1.3 equiv) and 1-(2-(4-methoxyphenoxy)propyl)piperazine **114d** (29 mg, 0.11 mmol, 1.0 equiv) were reacted according to General Procedure E to afford the aryl ether product as an off-white solid (27.1 mg, 0.0684 mmol, 59% yield).  $R_f = 0.54$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 136–138 °C.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  1.30 (d,  $J = 6.1$  Hz, 3H), 2.53 (dd,  $J = 4.4, 13.3$  Hz, 1H), 2.65 (tq,  $J = 5.6, 6.3, 11.5$  Hz, 4H), 2.74 (dd,  $J = 6.9, 13.3$  Hz, 1H), 3.77 (s, 3H), 3.86–3.98 (m, 4H), 4.45 (pd,  $J = 4.3, 6.2$  Hz, 1H), 6.76 (dd,  $J = 0.9, 2.2$  Hz, 1H), 6.81–6.90 (m, 4H), 7.13 (ddd,  $J = 0.9, 6.9, 7.9$  Hz, 1H), 7.26–7.30 (m, 1H), 7.42 (dq,  $J = 0.9, 8.4$  Hz, 1H), 7.64 (dq,  $J = 0.9, 8.1$  Hz, 1H), 9.23 (br s, 1H);  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  18.6, 29.7, 53.8, 55.7, 63.5, 73.4, 105.2, 111.7, 114.7, 117.6, 120.6, 121.8, 124.4, 127.5, 129.3, 135.5, 151.8, 154.1, 162.2; FTIR (neat) 1597, 1526,

1504, 1438  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{23}\text{H}_{28}\text{N}_3\text{O}_3$  [ $\text{M} + \text{H}$ ]<sup>+</sup> 394.2125; found 394.2124; HPLC purity >99.5%.

(4-(2-(4-Chlorophenoxy)propyl)piperazin-1-yl)(1H-indol-2-yl)methanone (**103**). 1H-Indole-2-carboxylic acid (22.0 mg, 0.137 mmol, 1.2 equiv) and 1-(2-(4-chlorophenoxy)propyl)piperazine **114e** (29.0 mg, 0.116 mmol, 1.0 equiv) were reacted according to General Procedure E to afford the acylated product **103** as a white solid (22.3 mg, 0.0558 mmol, 49% yield).  $R_f = 0.46$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 149–151 °C. <sup>1</sup>H NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  1.32 (d,  $J = 6.2$  Hz, 3H), 2.55 (dd,  $J = 4.4, 13.4$  Hz, 1H), 2.64 (tq,  $J = 5.6, 6.4, 11.5$  Hz, 4H), 2.74 (dd,  $J = 6.8, 13.4$  Hz, 1H), 3.82–4.01 (m, 4H), 4.53 (pd,  $J = 4.3, 6.2$  Hz, 1H), 6.76 (dd,  $J = 0.9, 2.2$  Hz, 1H), 6.83–6.87 (m, 2H), 7.13 (ddd,  $J = 1.0, 7.0, 8.0$  Hz, 1H), 7.21–7.25 (m, 2H), 7.26–7.30 (m, 1H), 7.42 (dq,  $J = 0.9, 8.3$  Hz, 1H), 7.64 (dq,  $J = 0.9, 8.0$  Hz, 1H), 9.26 (br s, 1H); <sup>13</sup>C NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  18.4, 29.7, 53.8, 63.3, 72.8, 105.2, 111.7, 117.3, 120.6, 121.8, 124.4, 125.7, 127.4, 129.2, 129.4, 135.5, 156.4, 162.2; FTIR (neat) 1595, 1526, 1487, 1436  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{22}\text{H}_{25}\text{ClN}_3\text{O}_2$  [ $\text{M} + \text{H}$ ]<sup>+</sup> 398.1630; found 398.1633; HPLC purity = 99.7%.

(1H-Indol-2-yl)(4-(2-((4-methoxyphenyl)(methyl)amino)ethyl)piperazin-1-yl)methanone (**104**). (4-(2-Chloroethyl)piperazin-1-yl)-(1H-indol-2-yl)methanone in DMF (200  $\mu\text{L}$ , 0.528 M, 0.106 mmol) and 4-methoxy-*N*-methylaniline (24 mg, 0.18 mmol, 1.6 equiv) were reacted according to General Procedure G to afford the tertiary aniline product as a white solid (1.9 mg, 0.0048 mmol, 5% yield). HRMS ( $m/z$ ): calcd for  $\text{C}_{23}\text{H}_{29}\text{N}_4\text{O}_2$  [ $\text{M} + \text{H}$ ]<sup>+</sup> 393.2285; found 393.2283; HPLC purity >99.5%.

(4-(2-((1H-Indol-5-yl)oxy)ethyl)piperazin-1-yl)(1H-indol-2-yl)methanone (**105**). (4-(2-Chloroethyl)piperazin-1-yl)(1H-indol-2-yl)methanone (30.8 mg, 0.106 mmol) and 5-hydroxyindole (22.6 mg, 0.170 mmol, 1.6 equiv) were reacted according to General Procedure F and purified by mass-directed, preparative HPLC to afford the aryl ether product as a colorless oil (9.8 mg, 0.025 mmol, 24% yield). <sup>1</sup>H NMR (500 MHz,  $\text{DMSO}-d_6$ )  $\delta$  2.59 (t,  $J = 5.1$  Hz, 4H), 2.77 (t,  $J = 5.8$  Hz, 2H), 3.70–3.85 (m, 4H), 4.10 (t,  $J = 5.7$  Hz, 2H), 6.31–6.33 (m, 1H), 6.74 (dd,  $J = 2.4, 8.7$  Hz, 1H), 6.79 (dd,  $J = 0.8, 2.2$  Hz, 1H), 7.03–7.07 (m, 2H), 7.16–7.20 (m, 1H), 7.26–7.28 (m, 2H), 7.41 (dd,  $J = 0.9, 8.3$  Hz, 1H), 7.61 (dd,  $J = 0.6, 8.0$  Hz, 1H), 10.90 (br s, 1H), 11.57 (br s, 1H); <sup>13</sup>C NMR (126 MHz,  $\text{DMSO}-d_6$ )  $\delta$  53.17, 53.19, 56.7, 65.9, 100.8, 102.8, 103.9, 111.6, 111.9, 112.0, 119.7, 121.3, 123.1, 125.7, 126.7, 127.9, 129.8, 131.0, 135.8, 152.2, 161.9; HRMS ( $m/z$ ): calcd for  $\text{C}_{23}\text{H}_{25}\text{N}_4\text{O}_2$  [ $\text{M} + \text{H}$ ]<sup>+</sup> 389.1972; found 389.1971; HPLC purity = 96.8%.

(1H-Indol-2-yl)(4-(3-(4-methoxyphenoxy)propyl)piperazin-1-yl)methanone (**106**). (1H-Indol-2-yl)(piperazin-1-yl)methanone (43 mg, 0.19 mmol) and 1-(3-bromopropoxy)-4-methoxybenzene (55 mg, 0.23 mmol, 1.2 equiv) were reacted according to General Procedure A to afford the alkylated piperazine as a tan solid (61 mg, 0.16 mmol, 83% yield).  $R_f = 0.36$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 122–127 °C. <sup>1</sup>H NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  1.95–2.02 (m, 2H), 2.53–2.61 (complex, 6H), 3.77 (s, 3H), 3.94–4.09 (complex, 6H), 6.78 (d,  $J = 1.9$  Hz, 1H), 6.83–6.88 (complex, 4H), 7.13 (t,  $J = 7.5$  Hz, 1H), 7.25–7.29 (m, 1H), 7.45 (d,  $J = 8.3$  Hz, 1H), 7.65 (d,  $J = 8.0$  Hz, 1H), 10.02 (br s, 1H); <sup>13</sup>C NMR (101 MHz, APT pulse sequence,  $\text{CDCl}_3$ )  $\delta$  d 55.7, 105.2, 111.9, 114.7, 115.5, 120.4, 121.7, 124.2; u 26.8, 53.19, 53.23, 55.0, 66.6, 127.4, 129.3, 135.9, 153.1, 153.8, 162.5; FTIR (neat) 1595, 1526, 1505, 1436  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{23}\text{H}_{28}\text{N}_3\text{O}_3$  [ $\text{M} + \text{H}$ ]<sup>+</sup> 394.2125; found 394.2117; HPLC purity = 97.2%.

(1H-Indol-2-yl)(4-(4-(4-methoxyphenoxy)butyl)piperazin-1-yl)methanone (**107**). (1H-Indol-2-yl)(piperazin-1-yl)methanone (38 mg, 0.17 mmol) and 1-(4-bromobutoxy)-4-methoxybenzene (52 mg, 0.20 mmol, 1.2 equiv) were reacted according to General Procedure A to afford the alkylated piperazine as a tan solid (51 mg, 0.13 mmol, 76% yield).  $R_f = 0.36$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 111–113 °C. <sup>1</sup>H NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  1.64–1.72 (m, 2H), 1.77–1.84 (m, 2H), 2.44 (t,  $J = 7.4$  Hz, 2H), 2.53 (t,  $J = 5.0$  Hz, 4H), 3.75 (s, 3H), 3.91–3.99 (complex, 6H), 6.75–6.77 (m, 1H), 6.80–6.85 (m, 1H), 7.09–7.13 (m, 1H), 7.23–7.27 (m, 1H), 7.42 (dd,  $J = 0.7, 8.3$  Hz, 1H), 7.63 (dd,  $J = 0.6, 7.4$  Hz, 1H), 9.92 (br s, 1H); <sup>13</sup>C NMR (101 MHz,

$\text{CDCl}_3$ )  $\delta$  23.3, 27.3, 53.2 (br, 2 C), 55.7, 58.1, 68.2, 105.2, 111.9, 114.6, 115.4, 120.4, 121.7, 124.2, 127.4, 129.3, 135.8, 153.1, 153.7, 162.5; FTIR (neat) 1593, 1506, 1463, 1437  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{24}\text{H}_{30}\text{N}_3\text{O}_3$  [ $\text{M} + \text{H}$ ]<sup>+</sup> 408.2282; found 408.2265; HPLC purity = 98.1%.

(1H-Indol-2-yl)(4-(2-(4-methoxyphenoxy)ethyl)piperidin-1-yl)methanone (**108**). To a mixture of 2-(1-(1H-indole-2-carbonyl)-piperidin-4-yl)ethyl 4-methylbenzenesulfonate **122** (82 mg, 0.19 mmol) and potassium carbonate (80 mg, 0.58 mmol, 3.0 equiv) in MeCN (3 mL) was added 4-methoxyphenol (72 mg, 0.58 mmol, 3 equiv) in THF (5 mL), and the reaction was stirred at 60 °C for 17 h. After cooling to rt, the reaction was filtered and the solids washed with  $\text{CH}_2\text{Cl}_2$  ( $2 \times 5$  mL). The combined filtrates were evaporated, and the residue was purified by silica gel chromatography to afford the aryl ether product **108** as an off-white solid (51 mg, 0.14 mmol, 70% yield).  $R_f = 0.58$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ); mp = 176–179 °C. <sup>1</sup>H NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  1.29–1.41 (m, 2H), 1.74–1.80 (m, 2H), 1.87–1.95 (complex, 3H), 2.97–3.17 (br m, 2H), 3.78 (s, 3H), 4.00 (t,  $J = 6.2$  Hz, 2H), 4.70–4.4.76 (m, 2H), 6.77–6.79 (m, 1H), 6.84–6.85 (m, 4H), 7.11–7.15 (m, 1H), 7.25–7.30 (m, 1H), 7.43 (dd,  $J = 0.7, 8.3$  Hz, 1H), 7.65 (dd,  $J = 0.6, 8.0$  Hz, 1H), 9.35 (br s, 1H); <sup>13</sup>C NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  32.4 (br, 2 C), 33.2, 35.8, 55.7, 65.8, 104.9, 111.7, 114.7, 115.4, 120.5, 121.8, 124.2, 127.5, 129.6, 135.6, 153.0, 153.8, 162.2; FTIR (neat) 1596, 1534, 1505, 1440  $\text{cm}^{-1}$ ; HRMS ( $m/z$ ): calcd for  $\text{C}_{23}\text{H}_{27}\text{N}_2\text{O}_3$  [ $\text{M} + \text{H}$ ]<sup>+</sup> 379.2016; found 379.2020; HPLC purity = 97.8%.

1-(1H-Indol-2-yl)-2-(4-(2-(4-methoxyphenoxy)ethyl)piperazin-1-yl)ethan-1-one (**109**). 1-(2-(4-Methoxyphenoxy)ethyl)piperazine (60 mg, 0.25 mmol), 2-chloro-1-(1H-indol-2-yl)ethan-1-one (49 mg, 0.25 mmol, 1.0 equiv), potassium carbonate (140 mg, 1.02 mmol, 4.0 equiv), and potassium iodide (42 mg, 0.25 mmol, 1.0 equiv) were charged in a reaction vial, slurried with MeCN (4 mL), and stirred at 65 °C for 18 h. The reaction was filtered and the solids washed with  $\text{CH}_2\text{Cl}_2$  ( $2 \times 5$  mL). The combined filtrates were evaporated, and the residue was purified by silica gel chromatography to afford the aryl ether product as a tan residue (51 mg, 0.13 mmol, 51% yield).  $R_f = 0.30$  (5% MeOH/ $\text{CH}_2\text{Cl}_2$ ). <sup>1</sup>H NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  2.64–2.75 (complex, 8H), 2.82 (t,  $J = 5.8$  Hz, 2H), 3.75–3.77 (complex, 5H), 4.06 (t,  $J = 5.8$  Hz, 2H), 6.79–6.85 (complex, 4H), 7.12–7.17 (m, 1H), 7.32–7.38 (m, 2H), 7.43 (d,  $J = 8.3$  Hz, 1H), 7.71 (d,  $J = 8.1$  Hz, 1H), 9.78 (br s, 1H); <sup>13</sup>C NMR (101 MHz, APT pulse sequence,  $\text{CDCl}_3$ )  $\delta$  d 55.7, 109.4, 112.3, 114.6, 115.6, 121.0, 123.1, 126.4; u 53.4, 53.6, 57.3, 64.8, 66.5, 127.4, 134.4, 137.1, 152.9, 153.9, 189.6; HRMS ( $m/z$ ): calcd for  $\text{C}_{23}\text{H}_{28}\text{N}_3\text{O}_3$  [ $\text{M} + \text{H}$ ]<sup>+</sup> 394.2125; found 394.2132; HPLC purity = 96.9%.

(4-Methoxyphenyl)(piperazin-1-yl)methanone (**111a**). This material was purchased from Oakwood Chemical.

(2-Methoxyphenyl)(piperazin-1-yl)methanone (**111b**). This material was purchased from 1Click Chemistry Inc.

(3-Methoxyphenyl)(piperazin-1-yl)methanone (**111c**). This material was purchased from Combi-Blocks Inc.

(4-Chlorophenyl)(piperazin-1-yl)methanone (**111d**). This material was synthesized according to the protocol of Wang and co-workers.<sup>80</sup> Thus, a solution of piperazine (1.26 g, 14.6 mmol) in THF (100 mL) was cooled in an rt water bath, and butyllithium (13.8 mL, 30.7 mmol) was added. The reaction was stirred at rt for 30 min, and 4-chlorobenzoyl chloride (1.9 mL, 14.6 mmol) was added as a solution in THF (10 mL). The reaction was stirred for 10 additional min and quenched with MeOH (10 mL). All solvents were removed in vacuo, and the residue was partitioned between saturated aqueous  $\text{NaHCO}_3$  (50 mL) and EtOAc (25 mL). The aqueous layer was extracted with EtOAc ( $2 \times 25$  mL), and the combined organic layers were dried ( $\text{Na}_2\text{SO}_4$ ), concentrated in vacuo, and purified by silica chromatography to afford the previously reported acylated product.<sup>81</sup> (818 mg, 2.72 mmol, 68% yield). <sup>1</sup>H NMR (400 MHz, MeOD- $d_4$ )  $\delta$  2.68–2.96 (m, 4H), 3.34–3.78 (m, 4H), 7.36–7.44 (m, 2H), 7.44–7.51 (m, 2H); <sup>13</sup>C NMR (101 MHz, MeOD- $d_4$ )  $\delta$  42.1, 45.6, 128.4, 128.5, 134.0, 135.6, 169.9.

(4-Ethylphenyl)(piperazin-1-yl)methanone (**111e**). To a solution of *tert*-butyl piperazine-1-carboxylate (1.44 g, 7.73 mmol) and



triethylamine (2.69 mL, 19.33 mmol, 2.5 equiv) in THF (120 mL) at 0 °C was added 4-ethylbenzoyl chloride (1.25 mL, 8.50 mmol, 1.1 equiv). The reaction was stirred overnight (19 h), slowly warming to rt. The reaction was quenched with saturated aqueous NaHCO<sub>3</sub>, and the aqueous layer was extracted with EtOAc (2 × 30 mL). The combined organic layers were dried (Na<sub>2</sub>SO<sub>4</sub>), evaporated, and purified by silica chromatography to afford the acylated product, *tert*-butyl 4-(4-ethylbenzoyl)piperazine-1-carboxylate, as a white solid (2.14 g, 6.72 mmol, 87% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.23 (t, *J* = 7.6 Hz, 3H), 1.45 (s, 9H), 2.66 (q, *J* = 7.6 Hz, 2H), 3.31–3.74 (m, 8H), 7.19–7.27 (m, 2H), 7.31 (d, *J* = 8.1 Hz, 2H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 15.2, 15.4, 28.4, 28.7, 29.0, 80.3, 127.2, 127.8, 128.0, 130.2, 132.7, 146.4, 154.6, 170.9.

To a solution of *tert*-butyl 4-(4-ethylbenzoyl)piperazine-1-carboxylate (1.90 g, 5.97 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (60 mL) was added trifluoroacetic acid (9.19 mL, 119 mmol, 20 equiv). The reaction was stirred at rt for 4 h and concentrated under vacuum. The residue was partitioned between aqueous saturated sodium bicarbonate (100 mL) and CH<sub>2</sub>Cl<sub>2</sub> (3 × 30 mL). The combined organic layers were concentrated under vacuum to afford the piperazine product **111e** as a white solid (1.87 g, 5.63 mmol, 94% yield). <sup>1</sup>H NMR (400 MHz, MeOD-*d*<sub>4</sub>) δ 1.24 (t, *J* = 7.6 Hz, 3H), 2.68 (q, *J* = 7.6 Hz, 2H), 2.72–2.94 (m, 4H), 3.35–3.78 (m, 4H), 7.25–7.35 (m, 4H); <sup>13</sup>C NMR (101 MHz, MeOD-*d*<sub>4</sub>) δ 14.5, 28.3, 42.3, 45.4, 126.8, 127.7, 132.6, 146.5, 171.3.

**(4-Methoxyphenyl)(2-methylpiperazin-1-yl)methanone (111f).** To a solution of 2-methylpiperazine (0.99 g, 9.88 mmol) in THF (100 mL) was added butyllithium (2.22 M in hexanes, 9.35 mL, 20.76 mmol, 2.1 equiv), and the reaction was stirred at rt for 30 min. This preformed anion solution was cooled to 0 °C, and a solution of 4-methoxybenzoyl chloride (1.68 g, 9.88 mmol, 1.0 equiv) in THF (10 mL) was added. After stirring at 0 °C for 10 min, the reaction was quenched by the addition of MeOH (10 mL), and the solvents were removed in vacuo. The residue was partitioned between saturated aqueous NaHCO<sub>3</sub> (50 mL) and EtOAc (3 × 25 mL). The combined organic layers were dried (Na<sub>2</sub>SO<sub>4</sub>), concentrated in vacuo, and purified on C18-functionalized silica gel chromatography to afford the acylated product **111f** as a tan solid (427.9 mg, 1.83 mmol, 18% yield).

**(4-Methoxyphenyl)(2-methylpiperazin-1-yl)methanone (111g).** To a solution of *tert*-butyl 3-methylpiperazine-1-carboxylate (1.20 g, 6.01 mmol) and triethylamine (1.0 mL, 7.17 mmol, 1.2 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (25 mL) was added 4-methoxybenzoyl chloride (1.12 g, 6.54 mmol, 1.1 equiv). The reaction was stirred at rt for 4 h and quenched with aqueous citric acid (1 M, 15 mL). The organic layer was washed with water (5 mL) and then aqueous NaOH (1 M, 5 mL) and dried with MgSO<sub>4</sub>. The solvent was removed in vacuo, and the residue was purified by silica chromatography to afford the acylated product, *tert*-butyl 4-(4-methoxybenzoyl)-3-methylpiperazine-1-carboxylate, as a colorless oil (1.98 g, 5.92 mmol, 98% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.25 (d, *J* = 6.7 Hz, 3H), 1.47 (s, 9H), 2.69–3.27 (complex, 5H), 3.84 (s, 3H), 3.96–4.15 (m, 2H), 6.92 (d, *J* = 8.8 Hz, 2H), 7.35 (d, *J* = 8.8 Hz, 2H).

To a solution of *tert*-butyl 4-(4-methoxybenzoyl)-3-methylpiperazine-1-carboxylate (1.98 g, 5.91 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (10 mL) was added trifluoroacetic acid (10 mL, 129 mmol, 22 equiv). The reaction was stirred at rt for 5 h and concentrated under vacuum. The residue was partitioned between aqueous sodium hydroxide (1 M, 50 mL) and CH<sub>2</sub>Cl<sub>2</sub> (4 × 10 mL). The combined organic layers were dried with Na<sub>2</sub>SO<sub>4</sub>, concentrated under vacuum, and purified by silica chromatography to afford the piperazine product **111g** as a viscous, tan oil (1.30 g, 5.56 mmol, 94% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.36 (d, *J* = 6.9 Hz, 3H), 2.76 (dt, *J* = 3.5, 12.3 Hz, 1H), 2.82–2.87 (m, 1H), 2.97 (dd, *J* = 4.0, 12.4 Hz, 1H), 3.00–3.05 (m, 1H), 3.16–3.27 (m, 1H), 3.83 (s, 3H), 3.91–4.05 (m, 1H), 4.34–4.48 (m, 1H), 6.88–6.96 (m, 2H), 7.32–7.38 (m, 2H); <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>) δ 15.5, 40.7, 46.1, 47.6, 50.2, 55.4, 113.8, 128.3, 128.6, 160.6, 170.6.

**(1H-Indol-2-yl)(piperazin-1-yl)methanone (111h).** 1H-Indole-2-carboxylic acid (1.55 g, 9.62 mmol) and HATU (4.39 g, 11.55 mmol,

1.2 equiv) were dissolved in EtOAc (80 mL) and stirred at rt for 10 min followed by the addition of a *tert*-butyl piperazine-1-carboxylate (1.79 g, 9.62 mmol, 1.0 equiv) solution in THF (80 mL). After stirring at rt for an additional 10 min, Et<sub>3</sub>N (4.0 mL, 28.87 mmol, 3.0 equiv) was added, and the reaction was stirred at rt for 43 h. The solvents were removed in vacuo, and the residue was washed with water (4 × 50 mL). The crude solid product was further purified by silica chromatography to afford the amide piperazine product, *tert*-butyl 4-(1H-indole-2-carbonyl)piperazine-1-carboxylate, as a white solid (2.65 g, 9.62 mmol, 84% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.49 (s, 9H), 3.56 (dd, *J* = 4.2, 6.4 Hz, 4H), 3.91 (s, 4H), 6.76–6.81 (m, 1H), 7.14 (ddd, *J* = 1.0, 7.0, 8.0 Hz, 1H), 7.25–7.34 (m, 1H), 7.43 (dd, *J* = 1.0, 8.4 Hz, 1H), 7.62–7.69 (m, 1H), 9.29 (br s, 1H); <sup>13</sup>C NMR (101 MHz, APT pulse sequence, CDCl<sub>3</sub>) δ d 28.4, 105.4, 111.7, 120.7, 121.9, 124.6; u 37.2, 41.9, 80.4, 127.4, 128.9, 135.6, 154.6, 162.6.

A solution of *tert*-butyl 4-(1H-indole-2-carbonyl)piperazine-1-carboxylate (1.66 g, 5.03 mmol) in hexafluoroisopropanol (15 mL) was heated under microwave irradiation at 145 °C for 2.5 h and cooled to rt. The solvent was removed in vacuo, and the residue was recrystallized from methyl *tert*-butyl ether to afford the free base piperazine product **111h** as a white solid (998 mg, 4.35 mmol, 87% yield). Mp 185–187 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.49 (s, 9H), 3.56 (dd, *J* = 4.2, 6.4 Hz, 4H), 3.91 (s, 4H), 6.76–6.81 (m, 1H), 7.14 (ddd, *J* = 1.0, 7.0, 8.0 Hz, 1H), 7.25–7.34 (m, 1H), 7.43 (dd, *J* = 1.0, 8.4 Hz, 1H), 7.62–7.69 (m, 1H), 9.29 (br s, 1H); <sup>13</sup>C NMR (101 MHz, APT pulse sequence, CDCl<sub>3</sub>) δ d 105.1, 111.7, 120.2, 121.5, 124.0; u 45.5 (2C), 127.0, 128.7, 136.0, 163.1.

**1-(2-Bromoethoxy)-4-methoxybenzene (112a).** This material was synthesized as previously described.<sup>82</sup>

**1-(2-Bromoethoxy)-2-methoxybenzene (112b).** 2-Methoxyphenol (2.84 g, 22.88 mmol) was reacted according to General Procedure H to afford bromide as a white solid (2.0 g, 8.65 mmol, 38% yield). *R*<sub>f</sub> = 0.70 (25% EtOAc in hexanes); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 3.62 (t, *J* = 6.6 Hz, 2H), 3.87 (s, 3H), 4.33 (t, *J* = 6.9 Hz, 2H), 6.88–7.03 (complex, 4H); <sup>13</sup>C NMR (101 MHz, APT pulse sequence, CDCl<sub>3</sub>) δ d 56.0, 112.3, 115.0, 120.9, 122.4; u 28.9, 69.3, 147.5, 149.9. These data are in agreement with those previously reported.<sup>83</sup>

**1-(2-Bromoethoxy)-3-methoxybenzene (112c).** 3-Methoxyphenol (2.05 g, 16.51 mmol) was reacted according to General Procedure H to afford bromide as a colorless oil (1.90 g, 8.22 mmol, 50% yield). *R*<sub>f</sub> = 0.71 (25% EtOAc in hexanes); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 3.62 (t, *J* = 6.3 Hz, 2H), 3.78 (s, 3H), 4.26 (t, *J* = 6.3 Hz, 2H), 6.47–6.55 (complex, 3H), 7.16–7.21 (m, 1H); <sup>13</sup>C NMR (101 MHz, APT pulse sequence, CDCl<sub>3</sub>) δ d 55.3, 101.3, 106.7, 107.1, 130.0; u 29.2, 67.9, 159.3, 160.9; IR (thin film): 1589, 1491, 1450 cm<sup>-1</sup>.

**1-(2-Bromoethoxy)-4-ethylbenzene (112d).** 4-Ethylphenol (3.55 g, 29.10 mmol) was reacted according to General Procedure H to afford bromide as a dark golden oil (2.10 g, 9.17 mmol, 31% yield). *R*<sub>f</sub> = 0.79 (25% EtOAc in hexanes); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.19 (t, *J* = 7.6 Hz, 3H), 2.57 (q, *J* = 7.6 Hz, 2H), 3.56 (t, *J* = 6.3 Hz, 2H), 4.20 (t, *J* = 6.3 Hz, 2H), 6.78–6.86 (m, 2H), 7.07–7.10 (m, 2H); <sup>13</sup>C NMR (101 MHz, APT pulse sequence, CDCl<sub>3</sub>) δ d 16.0, 114.8, 128.9; u 28.1, 29.4, 68.1, 137.3, 156.2. These data are in agreement with those previously reported.<sup>84</sup>

**1-(2-Bromoethoxy)-4-chlorobenzene (112e).** 4-Chlorophenol (3.18 g, 24.74 mmol) was reacted according to General Procedure H to afford bromide as a white solid (2.80 g, 11.89 mmol, 48% yield). *R*<sub>f</sub> = 0.73 (25% EtOAc in hexanes); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 3.61 (t, *J* = 6.2 Hz, 2H), 4.23 (t, *J* = 6.2 Hz, 2H), 6.80–6.87 (m, 2H), 7.21–7.25 (m, 2H); <sup>13</sup>C NMR (101 MHz, APT pulse sequence, CDCl<sub>3</sub>) δ d 116.1, 129.5; u 29.0, 68.2, 126.4, 156.7; IR (neat): 1594, 1581, 1488, 1457 cm<sup>-1</sup>. These data are in agreement with those previously reported.<sup>85</sup>

**1-(2-Bromoethoxy)-3,5-dimethoxybenzene (112f).** 3,5-Dimethoxyphenol (3.02 g, 19.6 mmol) was reacted according to General Procedure H to afford bromide as a colorless oil (2.70 g, 10.3 mmol, 53% yield). *R*<sub>f</sub> = 0.43 (25% EtOAc in hexanes); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) 3.61 (t, *J* = 6.2 Hz, 2H), 3.76 (s, 6H), 4.24 (t, *J* = 6.2 Hz, 2H), 6.08 (d, *J* = 2.1 Hz, 2H), 6.10–6.12 (m, 1H); <sup>13</sup>C NMR (101

MHz, APT pulse sequence, CDCl<sub>3</sub>)  $\delta$  d 55.4, 93.61, 93.62; u 29.1, 67.9, 160.0, 161.6; IR (thin film): 1590, 1474, 1456 cm<sup>-1</sup>. These data are in agreement with those previously reported.<sup>86</sup>

**5-(2-Bromoethoxy)benzo[d][1,3]dioxole (112g).** 3,4-(Methylenedioxy)phenol (2.31 g, 16.72 mmol) was reacted according to General Procedure H to afford bromide as a faintly purple solid (2.70 g, 11.0 mmol, 66% yield).  $R_f = 0.55$  (25% EtOAc in hexanes); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  3.50 (t,  $J = 6.3$  Hz, 2H), 4.11 (t,  $J = 6.2$  Hz, 2H), 5.82 (s, 2H), 6.24 (dd,  $J = 2.5, 8.4$  Hz, 1H), 6.42 (d,  $J = 2.5$  Hz, 1H), 6.61 (d,  $J = 8.4$  Hz, 1H); <sup>13</sup>C NMR (101 MHz, APT pulse sequence, CDCl<sub>3</sub>)  $\delta$  d 98.6, 106.3, 108.0; u 29.3, 69.0, 101.3, 142.3, 148.4, 153.5; IR (thin film): 1632, 1494, 1481 cm<sup>-1</sup>.

**3-(2-Chloroethoxy)pyridine (112h).** To a mixture of 3-hydroxypyridine (872 mg, 9.17 mmol) and potassium carbonate (2.54 mg, 18.35 mmol, 2.0 equiv) in DMF (15 mL) was added 1-dibromo-2-chloroethane (3.0 mL, 5.19 g, 36.2 mmol, 4.0 equiv). The reaction was heated at 60 °C for 22 h, cooled to rt, and partitioned between water (150 mL) and CH<sub>2</sub>Cl<sub>2</sub> (5 × 10 mL). The combined organic layers were washed with aqueous sodium hydroxide (1 M, 2 × 10 mL) and then water (2 × 10 mL) and dried with Na<sub>2</sub>SO<sub>4</sub>. The organic layer was adsorbed onto silica and purified by silica chromatography to afford the ether product as a dark orange oil (284 mg, 1.80 mmol, 20% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  3.84 (t,  $J = 5.8$  Hz, 2H), 4.29 (t,  $J = 5.8$  Hz, 2H), 7.22–7.25 (m, 2H), 8.27 (dd,  $J = 2.2, 3.8$  Hz, 1H), 8.34 (dd,  $J = 1.3, 2.4$  Hz, 1H).

**1-(3-Bromopropoxy)-4-methoxybenzene (112i).** This material was purchased from 1Click Chemistry Inc.

**1-(4-Bromobutoxy)-4-methoxybenzene (112j).** This material was purchased from 1Click Chemistry Inc.

**1-(4-Methoxyphenyl)piperazine (113).** This material was purchased from Combi-Blocks Inc.

**1-(2-(4-Methoxyphenoxy)ethyl)piperazine (114a).** A mixture of *tert*-butyl piperazine-1-carboxylate (490 mg, 2.63 mmol), 1-(2-bromoethoxy)-4-methoxybenzene **112a** (608 mg, 2.63 mmol), potassium carbonate (727 mg, 5.26 mmol, 2.0 equiv), and potassium iodide (44 mg, 0.263 mmol) in acetonitrile (35 mL) was heated at 70 °C for 16 h. The reaction was cooled to rt, filtered, and the solids were washed with acetonitrile (2 × 15 mL). The combined organics were adsorbed onto celite and purified by silica chromatography to afford the ether product, *tert*-butyl 4-(2-(4-methoxyphenoxy)ethyl)piperazine-1-carboxylate, as a colorless oil (622 mg, 1.85 mmol, 70% yield).  $R_f = 0.57$  (EtOAc); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.46 (s, 9 H), 2.51 (t,  $J = 4.8$  Hz, 4 H), 2.79 (t,  $J = 6.0$  Hz, 2 H), 3.45 (t,  $J = 4.8$  Hz, 4 H), 3.76 (s, 3 H), 4.05 (t,  $J = 6.0$  Hz, 2 H), 6.83 (d,  $J = 2.4$  Hz, 4 H); <sup>13</sup>C NMR (101 MHz, APT pulse sequence, CDCl<sub>3</sub>)  $\delta$  d 28.5, 55.8, 114.7, 115.7; u 53.5, 57.5, 66.7, 79.6, 153.0, 154.1, 154.8; HRMS ( $m/z$ ): calcd for C<sub>18</sub>H<sub>29</sub>N<sub>2</sub>O<sub>4</sub> ([M]<sup>+</sup> + H) 337.2122; found 337.2122; HPLC purity = 99.0%.

To a solution of *tert*-butyl 4-(2-(4-methoxyphenoxy)ethyl)piperazine-1-carboxylate (185 mg, 0.55 mmol) and triethylsilane (96 mg, 0.825 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) was added trifluoroacetic acid (0.85 mL, 1.254 mg, 11.00 mmol). The reaction was stirred at rt for 4 h and concentrated under vacuum. The residue was partitioned between aqueous sodium bicarbonate (10 mL) and CH<sub>2</sub>Cl<sub>2</sub> (3 × 10 mL). The combined organic layers were concentrated under vacuum to afford the piperazine product **114a** as a colorless oil (118 mg, 0.499 mmol, 91% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  2.54–2.60 (m, 4 H), 2.78 (t,  $J = 6.0$  Hz, 2 H), 2.94 (t,  $J = 4.8$  Hz, 4 H), 3.76 (s, 3 H), 4.05 (t,  $J = 6.0$  Hz, 2 H), 6.83 (d,  $J = 2.8$  Hz, 4 H); <sup>13</sup>C NMR (101 MHz, APT pulse sequence, CDCl<sub>3</sub>)  $\delta$  d 55.8, 114.8, 115.7; u 45.9, 54.6, 58.0, 66.6, 153.0, 154.0; HRMS ( $m/z$ ): calcd for C<sub>13</sub>H<sub>21</sub>N<sub>2</sub>O<sub>2</sub> ([M]<sup>+</sup> + H) 237.1598; found 237.1573; HPLC purity = 94.1%.

**1-(2-(3-Methoxyphenoxy)ethyl)piperazine (114b).** This material was purchased from 1Click Chemistry Inc.

**1-(2-(Pyridin-3-yloxy)ethyl)piperazine (114c).** A slurry of *tert*-butyl piperazine-1-carboxylate (699 mg, 3.59 mmol), 3-(2-chloroethoxy)pyridine **112h** (283 mg, 1.80 mmol), and potassium carbonate (499 mg, 3.61 mmol, 2.0 equiv) in DMF (2 mL) was heated at 90 °C for 21 h. The reaction was cooled to rt, diluted with water (20 mL), and extracted with CH<sub>2</sub>Cl<sub>2</sub> (4 × 5 mL). The

combined organics were dried with Na<sub>2</sub>SO<sub>4</sub>, concentrated under vacuum, and purified by silica chromatography to afford the alkylated product, *tert*-butyl 4-(2-(4 pyridin-3-yloxy)ethyl)piperazine-1-carboxylate, as a golden oil (434 mg, 1.41 mmol, 79% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  2.53 (t,  $J = 5.0$  Hz, 4H), 2.83 (t,  $J = 5.6$  Hz, 2H), 3.46 (t,  $J = 5.1$  Hz, 4H), 4.15 (t,  $J = 5.7$  Hz, 2H), 7.19–7.23 (m, 2H), 8.20–8.26 (m, 1H), 8.32–8.34 (m, 1H).

To a solution of *tert*-butyl 4-(2-(4 pyridin-3-yloxy)ethyl)piperazine-1-carboxylate (1.54 g, 5.02 mmol) and triethylsilane (0.874 g, 7.51 mmol, 1.5 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (6 mL) was added trifluoroacetic acid (6.0 mL, 8.94 g, 78.4 mmol). The reaction was stirred at rt for 19 h and concentrated under vacuum. The residue was partitioned between aqueous sodium hydroxide (5 M, 11 mL) and CH<sub>2</sub>Cl<sub>2</sub> (5 × 10 mL). The combined organic layers were dried with Na<sub>2</sub>SO<sub>4</sub>, concentrated under vacuum, and purified by chromatography on basic alumina (50 g) to afford the piperazine product **114c** as a light orange oil (915 mg, 4.41 mmol, 88% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  2.49–2.61 (m, 4H), 2.81 (t,  $J = 5.8$  Hz, 2H), 2.92 (t,  $J = 4.9$  Hz, 4H), 4.15 (t,  $J = 5.8$  Hz, 2H), 7.18–7.24 (m, 2H), 8.22 (dd,  $J = 2.4, 3.6$  Hz, 1H), 8.32 (dd,  $J = 1.4, 2.3$  Hz, 1H).

**1-(2-(4-Methoxyphenoxy)propyl)piperazine (114d).** To a mixture of *tert*-butyl piperazine-1-carboxylate (931 mg, 5.00 mmol) and potassium carbonate (1.73 g, 12.5 mmol, 2.5 equiv) in DMF (5 mL) was added 2-chloropropanoyl chloride (698 mg, 5.50 mmol, 1.1 equiv), and the reaction was stirred at rt for 20 min. 4-Methoxyphenol (746 mg, 6.01 mmol, 1.2 equiv) was added, and the reaction was stirred at rt for 5 min and then at 80 °C for 4 h. The reaction was partitioned between water (50 mL) and CH<sub>2</sub>Cl<sub>2</sub> (3 × 10 mL). The combined organic layers were sequentially washed with saturated aqueous NaHCO<sub>3</sub> (3 mL), water (3 mL), aqueous citric acid (1 M, 3 mL), and water (2 × 3 mL). The organic layer was dried over Na<sub>2</sub>SO<sub>4</sub>, adsorbed onto silica gel (4 g), and purified by silica gel chromatography to afford the acylated ether product, *tert*-butyl 4-(2-(4-methoxyphenoxy)propyl)piperazine-1-carboxylate, as a colorless oil (430 mg, 1.18 mmol, 24% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.44 (s, 9H), 1.54–1.64 (complex, 5H), 3.06–3.27 (m, 2H), 3.39–3.69 (m, 4H), 3.76 (s, 3H), 4.88 (q,  $J = 6.8$  Hz, 1H), 6.72–6.93 (m, 4H); HRMS ( $m/z$ ): calcd for C<sub>19</sub>H<sub>29</sub>N<sub>2</sub>O<sub>5</sub> [M + H]<sup>+</sup> 365.2071; found 365.2082.

To a solution of *tert*-butyl 4-(2-(4-methoxyphenoxy)propyl)piperazine-1-carboxylate (376 mg, 1.03 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (4 mL) was added sequentially triethylsilane (240 mg, 2.07 mmol, 2.0 equiv) and then trifluoroacetic acid (4 mL), and the reaction was stirred at rt for 26 h. The reaction was concentrated in vacuo and partitioned between aqueous NaOH (2 M, 6 mL) and CH<sub>2</sub>Cl<sub>2</sub> (3 × 5 mL). The combined organic layers were washed with water (2 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated, and purified by silica gel chromatography to afford the deprotected product, 2-(4-methoxyphenoxy)-1-(piperazin-1-yl)propan-1-one, as a colorless oil (219 mg, 0.829 mmol, 80% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.59 (d,  $J = 6.9$  Hz, 3H), 2.58–2.91 (m, 4H), 3.47–3.73 (m, 4H), 3.76 (s, 3H), 4.88 (q,  $J = 6.8$  Hz, 1H), 6.77–6.88 (m, 4H).

To a solution of 2-(4-methoxyphenoxy)-1-(piperazin-1-yl)propan-1-one (219 mg, 0.829 mmol) in THF (2 mL) at 0 °C was added a THF solution of borane (2.8 mL, 2.8 mmol, 3.2 equiv). After bubbling had ceased (6 min), the reaction was stirred at 65 °C for 4 h. MeOH (2 mL) was added, and the reaction was concentrated in vacuo. The residue was dissolved in MeOH (3 mL), and concentrated HCl (0.25 mL) was added, and the solution was stirred at 65 °C for 30 min. The solution was concentrated in vacuo, and the residue was partitioned between aqueous HCl (1 M, 4 mL) and CH<sub>2</sub>Cl<sub>2</sub> (4 mL). Aqueous sodium hydroxide (10 M, 3 mL) was added to increase the pH to >9, and the aqueous layer was extracted with CH<sub>2</sub>Cl<sub>2</sub> (4 × 2 mL). The combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated, and purified by silica gel chromatography to afford the reduced product **114d** as a colorless oil (178 mg, 0.711 mmol, 86% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.27 (d,  $J = 6.2$  Hz, 3H), 2.38–2.57 (complex, 5H), 2.68 (dd,  $J = 6.7, 13.1$  Hz, 1H), 2.88 (t,  $J = 4.9$  Hz, 4H), 3.77 (s, 3H), 4.35–4.48 (m, 1H), 6.75–6.92 (m, 4H).

**1-(2-(4-Chlorophenoxy)propyl)piperazine (114e).** To a mixture of *tert*-butyl piperazine-1-carboxylate (931 mg, 5.00 mmol) and potassium carbonate (1.73 g, 12.5 mmol, 2.5 equiv) in DMF (5 mL) was added 2-chloropropanoyl chloride (698 mg, 5.50 mmol, 1.1 equiv), and the reaction was stirred at rt for 20 min. 4-Chlorophenol (771 mg, 6.00 mmol, 1.2 equiv) was added, and the reaction was stirred at rt for 5 min and then at 80 °C for 4 h. The reaction was partitioned between water (50 mL) and CH<sub>2</sub>Cl<sub>2</sub> (3 × 10 mL). The combined organic layers were sequentially washed with saturated aqueous NaHCO<sub>3</sub> (3 mL), water (3 mL), aqueous citric acid (1 M, 3 mL) and water (2 × 3 mL). The organic layer was dried over MgSO<sub>4</sub>, adsorbed onto silica gel (4 g), and purified by silica gel chromatography to afford the acylated ether product, *tert*-butyl 4-(2-(4-chlorophenoxy)propyl)piperazine-1-carboxylate, as a white solid (515 mg, 1.40 mmol, 28% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.45 (s, 9H), 1.62 (d, *J* = 6.9 Hz, 3H), 3.09–3.26 (m, 3H), 3.41–3.79 (m, 5H), 4.91 (q, *J* = 6.8 Hz, 1H), 6.83 (d, *J* = 9.1 Hz, 2H), 7.23 (d, *J* = 9.0 Hz, 2H); HRMS (*m/z*): calcd for C<sub>14</sub>H<sub>18</sub>ClN<sub>2</sub>O<sub>4</sub> [M + *tert*-butyl + 2H]<sup>+</sup> 313.0950; found 313.0944.

To a solution of *tert*-butyl 4-(2-(4-chlorophenoxy)propyl)piperazine-1-carboxylate (458 mg, 1.24 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (4 mL) was added sequentially triethylsilane (291 mg, 2.50 mmol, 2.0 equiv) and then trifluoroacetic acid (4 mL), and the reaction was stirred at rt for 19 h. The reaction was concentrated in vacuo and partitioned between aqueous NaOH (2 M, 6 mL) and CH<sub>2</sub>Cl<sub>2</sub> (3 × 5 mL). The combined organic layers were washed with water (2 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated, and purified by silica gel chromatography to afford the deprotected product, 2-(4-chlorophenoxy)-1-(piperazin-1-yl)propan-1-one, as a colorless oil (299 mg, 1.11 mmol, 90% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.61 (d, *J* = 6.8 Hz, 3H), 2.56–2.91 (m, 5H), 3.45–3.76 (m, 3H), 4.91 (q, *J* = 6.8 Hz, 1H), 6.83 (d, *J* = 9.0 Hz, 2H), 7.23 (d, *J* = 9.1 Hz, 2H).

To a solution of 2-(4-chlorophenoxy)-1-(piperazin-1-yl)propan-1-one (308 mg, 1.15 mmol) in THF (2 mL) at 0 °C was added a THF solution of borane (1 M, 3.5 mL, 3.5 mmol, 3.1 equiv). After bubbling had ceased (6 min), the reaction was stirred at 65 °C for 4 h. MeOH (2 mL) was added, and the reaction was concentrated in vacuo. The residue was dissolved in MeOH (3 mL), and concentrated HCl (0.25 mL) was added, and the solution was stirred at 65 °C for 30 min. The solution was concentrated in vacuo, and the residue was partitioned between aqueous HCl (1 M, 4 mL) and CH<sub>2</sub>Cl<sub>2</sub> (4 mL). Aqueous sodium hydroxide (10 M, 3 mL) was added to increase the pH to >9, and the aqueous was extracted with CH<sub>2</sub>Cl<sub>2</sub> (4 × 2 mL). The combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated, and purified by silica gel chromatography to afford the amide reduction product **114e** as a colorless oil (257 mg, 1.10 mmol, 88% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.29 (d, *J* = 6.2 Hz, 3H), 2.39–2.55 (complex, 5H), 2.68 (dd, *J* = 6.6, 13.2 Hz, 1H), 2.86 (t, *J* = 4.9 Hz, 4H), 4.49 (td, *J* = 4.8, 6.3 Hz, 1H), 6.84 (d, *J* = 9.0 Hz, 2H), 7.21 (d, *J* = 9.0 Hz, 2H).

**4-(2-(2-Chloroethyl)piperazin-1-yl)(1H-indol-2-yl)methanone (116).** To a solution of 2-(piperazin-1-yl)ethanol (904 mg, 6.94 mmol) and triethylamine (843 mg, 8.33 mmol, 1.2 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (18 mL) was added 1H-indole-2-carbonyl chloride (1,367 mg, 7.61 mmol, 1.1 equiv) while cooling the reaction solution in an ice/water bath. The reaction was stirred for 20 h, slowly warming to rt. The reaction was concentrated in vacuo, and the residue was suspended in 50% saturated aqueous NaHCO<sub>3</sub> (40 mL). The solids were collected by filtration, washed with water (2 × 20 mL), and dried under vacuum to afford the acylated product, 4-(2-(hydroxyethyl)piperazin-1-yl)(1H-indol-2-yl)methanone, as a tan solid (1.41 g, 5.17 mmol, 75% yield), which was used without further purification. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 2.59 (t, *J* = 5.3 Hz, 2H), 3.65 (t, *J* = 5.4 Hz, 2H), 3.84 (s, 3H), 6.91 (d, *J* = 8.7 Hz, 2H), 7.39 (d, *J* = 8.8 Hz, 2H); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 53.0 (br), 55.4, 57.8, 59.3, 113.7, 127.7, 129.2, 160.8, 170.3; HRMS (*m/z*): calcd for C<sub>14</sub>H<sub>21</sub>N<sub>2</sub>O<sub>3</sub> [M + H]<sup>+</sup> 265.1547; found 265.1575.

Chlorination of the primary alcohol was accomplished utilizing the protocol of Kartika and co-workers.<sup>87</sup> Thus, a solution of 4-(2-(hydroxyethyl)piperazin-1-yl)(1H-indol-2-yl)methanone (1.96 g, 1.18

mmol) and triethylamine (2.50 mL, 1.82 g, 17.94 mmol, 2.5 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (65 mL) was cooled to 0 °C. Triphosgene (1.07 g, 3.60 mmol, 0.5 equiv) was added as a solid, and the reaction was stirred for 5 min at 0 °C and then at rt for 3 h. The reaction was quenched with saturated aqueous NaHCO<sub>3</sub> (50 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 × 10 mL). The combined organic layers were washed sequentially with saturated aqueous NaHCO<sub>3</sub> (25 mL) and then water (25 mL), dried with Na<sub>2</sub>SO<sub>4</sub>, and concentrated in vacuo. The residue was purified by silica chromatography to afford chloride **116** as a tan solid (1.10 g, 3.77 mmol, 53% yield). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 2.44 (t, *J* = 6.2 Hz, 2H), 2.46–2.53 (m, 4H), 3.53 (q, *J* = 6.1 Hz, 2H), 3.64–3.83 (m, 4H), 4.47 (t, *J* = 5.3 Hz, 1H), 6.77 (dd, *J* = 0.9, 2.2 Hz, 1H), 7.04 (ddd, *J* = 1.0, 6.9, 7.9 Hz, 1H), 7.18 (ddd, *J* = 1.2, 7.0, 8.2 Hz, 1H), 7.41 (dd, *J* = 1.0, 8.2 Hz, 1H), 7.60 (dd, *J* = 1.0, 8.0 Hz, 1H), 11.56 (br s, 1H).

**4-(2-(2-Chloroxyethyl)piperidin-1-yl)(4-methoxyphenyl)methanone (118).** To a solution of 2-(piperidin-4-yl)ethanol (518 mg, 4.01 mmol) and triethylamine (508 mg, 5.02 mmol, 1.25 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (20 mL) was added 4-methoxybenzoyl chloride (0.60 mL, 4.36 mmol, 1.1 equiv) while cooling the solution in an ice/water bath. The reaction was stirred for 16 h, slowly warming to rt, and then washed sequentially with aqueous HCl (1 M, 2 × 5 mL), water (1 × 5 mL), and aqueous NaHCO<sub>3</sub> (1 × 4 mL). The organic layer was adsorbed onto silica gel (1.5 g) and purified by silica chromatography to afford the acylated product, 4-(2-(hydroxyethyl)piperidin-1-yl)(4-methoxyphenyl)methanone, as a colorless oil (845 mg, 3.21 mmol, 80% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.12–1.30 (m, 1H), 1.50–1.59 (m, 2H), 1.64–1.86 (m, 4H), 2.69–3.06 (m, 4H), 3.71 (t, *J* = 6.5 Hz, 3H), 3.83 (s, 5H), 6.90 (d, *J* = 8.7 Hz, 2H), 7.37 (d, *J* = 8.8 Hz, 2H); HRMS (*m/z*): calcd for C<sub>15</sub>H<sub>22</sub>N<sub>2</sub>O<sub>3</sub> [M + H]<sup>+</sup> 264.1594; found 264.1605.

To a solution of 4-(2-(hydroxyethyl)piperidin-1-yl)(4-methoxyphenyl)methanone (144 mg, 0.547 mmol) in CHCl<sub>3</sub> (3 mL) at 0 °C was added thionyl chloride (65.1 mg, 0.547 mmol, 1.0 equiv). The reaction was stirred at 0 °C for 18 min, rt for 2 h, heated at 50 °C for 46 h, and then cooled to rt. The reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub> (4 mL) and washed sequentially with aqueous NaHCO<sub>3</sub> (2 × 2 mL) and then water (1 × 2 mL). The organic layer was dried with MgSO<sub>4</sub> and purified by silica gel chromatography to afford the chlorinated product **118** as a beige oil (141 mg, 0.470 mmol, 86% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.10–1.29 (m, 1H), 1.68–1.88 (complex, 6H), 2.72–3.09 (m, 4H), 3.59 (t, *J* = 6.6 Hz, 2H), 3.83 (s, 3H), 6.90 (d, *J* = 8.7 Hz, 2H), 7.37 (d, *J* = 8.7 Hz, 2H).

**N-(2-((2-Chloroethyl)(methylamino)ethyl)-4-methoxy-N-methylbenzamide Hydrochloride (120).** To a solution of *N*-(2-(hydroxyethyl)-4-methoxy-*N*-methylbenzamide<sup>88</sup> **119** (572 mg, 2.73 mmol) in CHCl<sub>3</sub> (6 mL) at 0 °C was added thionyl chloride (489 mg, 4.11 mmol, 1.5 equiv). The reaction was stirred at 0 °C for 3 min, heated at 55 °C for 35 min, and then cooled to rt. The reaction mixture was adsorbed onto silica gel (1.0 g) and purified by silica gel chromatography to afford the chlorinated product, *N*-(2-(chloroethyl)-4-methoxy-*N*-methylbenzamide, as a colorless oil (336 mg, 1.48 mmol, 54% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.53–1.69 (m, 2H), 3.12 (s, 3H), 3.68–3.88 (m, 2H), 3.84 (s, 3H), 6.92 (d, *J* = 8.7 Hz, 2H), 7.41 (d, *J* = 8.2 Hz, 2H).

To a solution of *N*-(2-(chloroethyl)-4-methoxy-*N*-methylbenzamide (332 mg, 1.45 mmol) in MeCN (6 mL) was added 2-(methylamino)ethanol (563 mg, 7.50 mmol, 5.1 equiv). The reaction was stirred at 80 °C for 25 h and then cooled to rt. The reaction mixture was adsorbed onto silica gel (1.4 g) and purified by silica gel chromatography to afford the alkylated product, *N*-(2-((2-(hydroxyethyl)(methylamino)ethyl)-4-methoxy-*N*-methylbenzamide, as a colorless oil (259 mg, 0.972 mmol, 67% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 2.29–2.74 (complex, 7H), 3.04 (s, 3H), 3.39–3.68 (m, 4H), 3.83 (s, 3H), 6.90 (d, *J* = 8.7 Hz, 2H), 7.39 (d, *J* = 8.7 Hz, 2H); HRMS (*m/z*): calcd for C<sub>14</sub>H<sub>23</sub>N<sub>2</sub>O<sub>3</sub> [M + H]<sup>+</sup> 267.1703; found 267.1726.

To a solution of *N*-(2-((2-(hydroxyethyl)(methylamino)ethyl)-4-methoxy-*N*-methylbenzamide (121 mg, 0.454 mmol) in CHCl<sub>3</sub> (3

mL) at 0 °C was added thionyl chloride (82.0 mg, 0.685 mmol, 1.5 equiv). The reaction was stirred at 0 °C for 3 min, rt for 30 min, heated at 50 °C for 25 min, then cooled to rt, and concentrated in vacuo. The residue was partitioned between aqueous sodium hydroxide (1 M, 3 mL) and CH<sub>2</sub>Cl<sub>2</sub> (2 × 10 mL). The combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated, and purified by silica gel chromatography to afford the hydroxy product **120** as a beige oil (109 mg, 0.38 mmol, 84% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 3.00 (s, 3H), 3.17 (s, 3H), 3.38–3.54 (m, 2H), 3.60–3.71 (m, 2H), 3.84 (s, 3H), 3.92–4.23 (m, 4H), 6.93 (d, *J* = 8.8 Hz, 2H), 7.47 (d, *J* = 8.7 Hz, 2H).

**(4-(2-Hydroxyethyl)piperazin-1-yl)(4-methoxyphenyl)methanone (121).** To a solution of 2-(piperazin-1-yl)ethanol (521 mg, 4.00 mmol) and triethylamine (508 mg, 5.02 mmol, 1.3 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (20 mL) was added 4-methoxybenzoyl chloride (0.60 mL, 4.36 mmol, 1.1 equiv) while cooling the solution in an ice/water bath. The reaction was stirred for 24 h, slowly warming to rt. The reaction was adsorbed onto silica gel (2 g) and purified by silica chromatography to afford the previously reported acylated product<sup>89</sup> as a white solid (818 mg, 2.72 mmol, 68% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 2.42–2.66 (m, 4H), 2.59 (t, *J* = 5.3 Hz, 2H), 3.46–3.79 (m, 4H), 3.65 (t, *J* = 5.4 Hz, 2H), 3.84 (s, 3H), 6.91 (d, *J* = 8.7 Hz, 2H), 7.39 (d, *J* = 8.8 Hz, 2H); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 53.0 (br), 55.4, 57.8, 59.3, 113.7, 127.7, 129.2, 160.8, 170.3; HRMS (*m/z*): calcd for C<sub>14</sub>H<sub>21</sub>N<sub>2</sub>O<sub>3</sub> [M + H]<sup>+</sup> 265.1547; found 265.1575.

**2-(1-(1H-Indole-2-carbonyl)piperidin-4-yl)ethyl 4-methylbenzenesulfonate (122).** To a solution of 1H-indole-2-carboxylic acid (374 mg, 2.32 mmol, 1.0 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (40 mL) were added HOBt (314 mg, 2.32 mmol, 1.0 equiv) and EDC·HCl (445 mg, 2.32 mmol, 1.0 equiv), and the reaction was stirred at rt for 10 min. 2-(Piperidin-4-yl)ethan-1-ol (300 mg, 2.32 mmol) was added, and the reaction was stirred at rt for 18 h. The reaction was partitioned between water (40 mL) and CH<sub>2</sub>Cl<sub>2</sub> (3 × 10 mL). The combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub> and evaporated, and the residue was purified by silica gel chromatography to afford the previously reported amide product, (4-(2-hydroxyethyl)piperidin-1-yl)(1H-indol-2-yl)methanone, as a white solid (310 mg, 1.14 mmol, 49% yield).<sup>90</sup> <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.13–1.32 (m, 2H), 1.45–1.52 (m, 2H), 1.67–1.85 (m, 3H), 2.68–3.21 (complex, 4H), 3.60 (t, *J* = 6.6 Hz, 2H), 4.56 (d, *J* = 13.4 Hz, 1H), 6.68 (s, 1H), 7.05 (t, *J* = 7.5 Hz, 1H), 7.19 (t, *J* = 7.7 Hz, 1H), 7.37 (d, *J* = 8.3 Hz, 1H), 7.56 (d, *J* = 8.0 Hz, 1H), 10.00 (br s, 1H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 32.2, 32.7, 38.7, 43.5, 59.5, 104.9, 111.8, 120.3, 121.5, 124.0, 127.1, 129.2, 135.8, 162.9.

To (4-(2-hydroxyethyl)piperidin-1-yl)(1H-indol-2-yl)methanone (166 mg, 0.61 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (13 mL) were added Et<sub>3</sub>N (0.34 mL, 2.44 mmol, 4.0 equiv) and then *p*-toluenesulfonyl chloride (232 mg, 1.22 mmol, 2.0 equiv). The reaction was stirred at rt until TLC indicated the conversion of alcohol (2 h). The reaction was quenched with saturated aqueous ammonium chloride (10 mL), and the layers were separated. The aqueous layer was extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 × 10 mL), and the combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub> and evaporated, and the residue was purified by silica gel chromatography to afford tosylate **122** as a white solid (120 mg, 0.28 mmol, 46% yield). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.16–1.28 (m, 2H), 1.61–1.66 (m, 2H), 1.70–1.83 (complex, 3H), 2.45 (s, 3H), 2.86–3.12 (m, 2H), 4.10 (t, *J* = 6.2 Hz, 2H), 4.66–4.73 (m, 2H), 6.73–6.75 (m, 1H), 7.10–7.14 (m, 1H), 7.24–7.28 (m, 1H), 7.36 (d, *J* = 8.0 Hz, 2H), 7.42 (dd, *J* = 0.8, 8.3 Hz, 1H), 7.64 (dd, *J* = 0.7, 8.0 Hz, 1H), 7.81 (d, *J* = 8.3 Hz, 1H), 9.66 (s, 1H); <sup>13</sup>C NMR (101 MHz, APT pulse sequence, CDCl<sub>3</sub>) δ d 21.7, 32.4, 104.9, 111.8, 120.4, 121.7, 124.2, 127.9, 129.9; u 32.1, 35.1, 67.8, 127.4, 129.5, 132.9, 135.7, 144.9, 162.3.

**Transient Transfections.** HEK293 cells were grown in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% fetal bovine serum and maintained at 37 °C in a humidified incubator containing 5% CO<sub>2</sub>. The day before transfection, cells were plated in 100 mm tissue culture dishes at 4 × 10<sup>6</sup> cells/dish in serum-free DMEM. The cells were transfected with the indicated DNA constructs using polyethylenimine (PEI) as the transfection reagent at

a ratio of 3:1 (μL PEI: μg DNA). Twenty-four hours after transfection, the media was changed to DMEM supplemented with 10% fetal bovine serum, and cells were used for experiments the next day.

**Radioligand Binding Assays.** HEK293 cells that were stably transfected with the human D2RL or D3R (Codex Biosolutions, Inc., Gaithersburg, MD) or parental HEK293 cells that were transiently transfected with either the D2RL, D3R, or indicated point mutants, as described above, were removed mechanically using calcium-free Earle's balanced salt solution (EBSS-). Intact cells were collected by centrifugation and then lysed with 5 mM Tris-HCl and 5 mM MgCl<sub>2</sub> at pH 7.4. Homogenates were centrifuged at 30 000g for 30 min. The membranes were resuspended in EBSS pH 7.4. For competition binding studies, membrane preparations were incubated for 90 min at room temperature with various concentrations of compound and a single or various concentration(s) of [<sup>3</sup>H]-methylspiperone in a reaction volume of 250 μL. Nonspecific binding was determined in the presence of 4 μM (+)-butaclamol. Bound ligand was separated from unbound by filtration through GF/C filters using a PerkinElmer cell harvester and quantified using a TopCount instrument (PerkinElmer). *K<sub>i</sub>* values were calculated using the Cheng-Prusoff equation<sup>91</sup> from the observed IC<sub>50</sub> values and the radioligand *K<sub>d</sub>* value determined from separate saturation binding experiments.

**β-Arrestin Recruitment DiscoverX PathHunter Assay.** In this assay, a CHO-K1 cell line expressing the human D3R fused with a small 42 amino acid fragment of β-galactosidase called ProLink and also expressing a fusion protein consisting of β-arrestin and a larger N-terminal deletion mutant of β-galactosidase was employed. Upon receptor activation, β-arrestin is recruited to the ProLink-tagged D3R, resulting in complementation of β-galactosidase to form a functional enzyme; addition of the substrate (PathHunter Detection Reagent) generates a chemiluminescent signal. Assays were conducted as previously described by our laboratory.<sup>53,92</sup> Briefly, CHO-K1 DiscoverX cells stably expressing various dopamine receptors were plated in CP2 media (DiscoverX, Fremont, CA) in 384-well black, clear-bottom plates at a density of 2625 cells/well and incubated overnight at 37 °C. Cells were then incubated with indicated compounds for 90 min at 37 °C for agonist mode or for 120 min in the presence of an EC<sub>80</sub> concentration of dopamine for antagonist mode. Cells were then incubated with a DiscoverX detection reagent for 30 min at room temperature, and luminescence was read on an FDSS μCell (Hamamatsu, Bridgewater, NJ). Data were collected as relative luminescence units (RLUs) and normalized as a percentage of the luminescence produced by a maximum concentration of dopamine.

**β-Arrestin Recruitment BRET Assay.** HEK293 cells were transiently transfected with D3R-Rluc8, β-arrestin2-mVenus, and G protein-coupled receptor kinase 3. GRK3 was used because it provides the greatest enhancement of β-arrestin recruitment for the D3R compared to other GRKs (unpublished observations). This GRK preference has also been noted by other labs.<sup>56</sup> Cells were harvested with EBSS-, plated in 96-well white plates at 20 000 cells/well in Dulbecco's phosphate-buffered saline (DPBS) and incubated at room temperature for 45 min. Cells were incubated with 5 μM coelenterazine h (Nanolight Technology, Pinetop, AZ) for 5 min and then stimulated with the indicated concentrations of the test compound for 5 min. The BRET signal was determined by quantifying and calculating the ratio of the light emitted by mVenus (525 nm) over that emitted by Rluc8 (485 nm) using a PHERAstar FSX Microplate Reader (BMG Labtech, Cary, NC).

**Go-BRET Activation Assay.** HEK293 cells transiently expressing the D3R and Gα<sub>oA</sub>-Rluc8, untagged-β<sub>1</sub>, and mVenus-γ<sub>2</sub> were harvested with EBSS-, plated in 96-well white plates at 20,000 cells/well in DPBS and incubated at room temperature for 45 min. Cells were incubated with 5 μM coelenterazine h (Nanolight Technology, Pinetop, AZ) for 5 min and then stimulated with the indicated concentrations of the test compound for 5 min. The BRET signal was determined by quantifying and calculating the ratio of the light emitted by mVenus (525 nm) over that emitted by RLuc8 (485

nm) using a PHERAstar FSX Microplate Reader (BMG Labtech, Cary, NC).

**ERK1/2 Phosphorylation Assay.** ERK1/2 phosphorylation was measured using the AlphaScreen SureFire Ultra ERK kit (PerkinElmer, Waltham, USA). CHO-K1 DiscoverX cells stably expressing the D3R were seeded into 384-well small volume white plates at a density of 40,000 cells/well in serum-free Ham's F12 media overnight. Cells were stimulated with the indicated concentration of the test compound for 15 min, followed by cell lysis as specified by the manufacturer's protocol. The plate was shaken for 10 min at room temperature, followed by the addition of Surefire activation buffer, Surefire reaction buffer, AlphaScreen acceptor beads, and AlphaScreen donor beads in ratios specified by the manufacturer. The plate was incubated in the dark for 2 h and then read using a PHERAstar FSX Microplate Reader (BMG Labtech, Cary, NC).

**CAMP CAMYEL Biosensor Assay.** HEK293 cells transiently expressing the D3R and the CAMYEL cAMP biosensor (yellow fluorescence protein-Epac-Rluc)<sup>93</sup> were harvested with EBSS-, plated in 96-well white plates at 20 000 cells/well in DPBS and incubated at room temperature for 45 min. Cells were pretreated for 5 min with 10  $\mu$ M forskolin and 10  $\mu$ M propranolol (to block endogenous  $\beta$ -adrenergic receptors) and then incubated with 5  $\mu$ M coelenterazine h (Nanolight Technology, Pinetop, AZ) for 5 min, followed by stimulation with the indicated concentrations of the test compound for 5 min. The BRET signal was determined by quantifying and calculating the ratio of the light emitted by mVenus (525 nm) over that emitted by Rluc8 (485 nm) using a PHERAstar FSX Microplate Reader (BMG Labtech, Cary, NC).

**DiscoverX gpcrMAX GPCR Panel.** To determine the selectivity profiles of ML417, CJ-1639, and pramipexole, these compounds were screened using the DiscoverX gpcrMAX GPCR panel, which measures the GPCR activation of  $\beta$ -arrestin recruitment to different GPCRs in either agonist or antagonist modes using a single high concentration (10  $\mu$ M) of the test compound. This study was conducted by DiscoverX, Inc. (Fremont, CA). Reference standards were run for each GPCR in the panel as an integral part of each assay to ensure the validity of the results. Assay results are presented as the mean percent activation or inhibition of the indicated GPCRs (for  $n = 2$  replicates) for each compound tested. For a full description of the DiscoverX gpcrMAX GPCR panel and the [Experimental Section](#), see <http://www.DiscoverX.com>.

**Psychoactive Drug Screening Program (PDSP) Radioligand Binding Panel.** 20, CJ-1639, and pramipexole were screened using the National Institute of Mental Health (NIMH) Psychoactive Drug Screening Program (PDSP) directed by Dr. Bryan L. Roth (University of North Carolina, Chapel Hill, NC). For experimental details, including radioligands used and associated  $K_d$  values for each individual target, please refer to the PDSP website <http://pdsp.med.unc.edu/>. Primary screening is performed using 10  $\mu$ M of the test compound, and, if >50% inhibition of radioligand binding is observed, then secondary screening is performed in full concentration–response format to derive affinity values.

**Inositol 1-Phosphate (IP1) Accumulation Assay.** Compound 20 was tested as both an agonist and antagonist of the 5-HT<sub>2B</sub> receptor using the Eurofins Cerep service (Celle l'Evescault, France). Briefly, recombinant CHO cells expressing the human 5-HT<sub>2B</sub> were used to measure IP1 accumulation in response to an agonist. Serotonin was used as the control agonist, while SB206553<sup>94</sup> was used as the control antagonist. In these assays, serotonin had an EC<sub>50</sub> of 4.5 nM, while SB206553 had an IC<sub>50</sub> of 13 nM. Antagonist assays were conducted in the presence of an EC<sub>80</sub> concentration (30 nM) of serotonin. The results obtained were the means of two separate assays.

**Molecular Dynamics Simulations.** The binding mode of 20 at the D3R was predicted by computational docking and molecular dynamics (MD) simulations. The ligand was docked to an equilibrated model of the D3R, which was built<sup>76</sup> based on the D3R crystal structure (PDB: 3PBL).<sup>6</sup> Briefly, the missing N terminus was predicted *de novo*, and a truncated poly-Gly segment was used in place of the ICL3.<sup>76</sup> The initial poses of the ligand were acquired by

using an induced-fit docking protocol<sup>95</sup> in the Schrodinger software (release 2015-3; Schrodinger Inc., New York, NY). The MD simulations were performed in the explicit water–POPC lipid bilayer environment using Desmond MD System (version 3.8; D. E. Shaw Research, New York, NY) with the CHARMM36 protein and lipid force field<sup>96,97</sup> and the TIP3P water model. The ligand parameters were acquired from the GAAMP server.<sup>98</sup>

As the predicted pKa of 20 is ambiguous using both empirical and quantum mechanics approaches, we experimentally titrated the molecule using a Sirius T3 instrument with a pH electrode and onboard titration tubes (Sirius Analytical Inc., Beverly, MA). Reference spectra without a compound and with controls containing either piroxicam or sulfathiazole were collected prior to the assays (data not shown). Samples were titrated using onboard robotics via the addition of HCl or KOH to generate the spectra, and pKa values were calculated using the onboard Sirius T3 software. The pKa of 20 was determined to be 8.62, and thus the basic piperazine nitrogen is predicted to be largely protonated at physiological pH. Consequently, we used the protonated, i.e., the charged form of the ligand in our simulations. The system charges were neutralized, and a solvent concentration of 0.15 M NaCl was added. The system was initially minimized and equilibrated with restraints on the ligand heavy atoms and protein backbone atoms, followed by a production stage in an isothermal–isobaric (NPT) ensemble at 310 K and 1 atm with all atoms unrestrained, as described previously.<sup>71,76</sup> We used the Langevin constant pressure and temperature dynamical system<sup>99</sup> to maintain the pressure and the temperature, on an anisotropic flexible periodic cell with a constant-ratio constraint applied on the lipid bilayer in the X–Y plane.

**Insertion of Human D3R into NSCs.** The human D3R open reading frame (ORF) was cloned into a donor vector containing homologous arms for the CLYBL safe harbor site and a neomycin resistance selection cassette. The ORF was synthesized as a codon-optimized gBlock (Integrated DNA Technologies) and inserted into the BsrGI site of the donor vector via Gibson assembly. The donor vector with the DRD3 knock-in and two TALEN pairs targeting the CLYBL locus were delivered by nucleofection (2  $\mu$ g of each vector) into the human neural stem cells (NSCs) derived from the NCRM-1 iPSC line. Forty-eight hours after transfection, the drug selection was initiated in media containing 600  $\mu$ g/mL of G418. Selection was continued for 7 days at which point the surviving NSCs were expanded in media without G418. Details about the TALEN-based safe harbor targeting approach are described by Cerbini et al. (2015).<sup>100</sup> Targeted cell lines were characterized for D3R expression by [<sup>3</sup>H]-methylspiperone binding assays.

**iPSC Cell Culture and Differentiation into Dopaminergic Neurons.** Neural stem cells (NSCs) were maintained in a growth medium composed of neurobasal medium (Gibco), B-27 (Gibco), nonessential amino acid (Sigma-Aldrich), GlutaMAX (Gibco), and 10 ng/mL of fibroblast growth factor (FGF). Differentiation into a dopaminergic phenotype was carried out over 10 days in a medium composed of neurobasal medium, 1X B-27 supplements, 1X nonessential amino acids, 20 ng/mL brain-derived neurotrophic factor (BDNF), 20 ng/mL glial-derived neurotrophic factor (GDNF), and 20 ng/mL TGF $\beta$ 3.<sup>101</sup> All growth factors were purchased from Peprotech (Rocky Hill, NJ). NSCs were grown in plates/flasks coated with Geltrex (ThermoFisher Scientific, Carlsbad, CA). Fifty percent of the differentiation media was replaced with fresh media every 72 h.

**MTT Assay.** 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) was purchased from ThermoFisher Scientific (Carlsbad, CA) (catalog number: M6494). MTT was dissolved in PBS at a concentration of 5 mg/mL (12 mM) and filtered. NSCs were plated at 10,000 cells per well in a 96-well plate and differentiation as described above for 10 days. On day 10 of the differentiation protocol, the cells were treated with the indicated concentrations of DMSO vehicle, 20, or pramipexole. Twenty-four hours later, the cells were treated with 30  $\mu$ M of 6-hydroxydopamine (6-OHDA). MTT assays were performed 24 h after 6-OHDA treatment. Each dilution was done in quadruplicates, and the experiments were performed at least 6 times. MTT assays were performed per the manufacturer's protocol

(ThermoFisher Scientific, Waltham, MA). Briefly, all media were pipetted out of the wells and fresh 100  $\mu$ L of media was added. Ten percent (10  $\mu$ L) of 5 mg/mL MTT solution was added to each well and incubated at 37 °C for 4 h. All but 25  $\mu$ L of the MTT/media solution was pipetted out of each well. Fifty microliters of DMSO was added to dissolve the resultant formazan crystals. The plate was agitated on a shaker for 10 min. Optical density (OD) was read at a 540 nm wavelength on PHERAstar FX (BMG Labtech, Cary, NC) with a reference wavelength at 650 nm.

**Ames Assay.** The mutagenicity potential of **20** was tested using the Ames reverse mutation assay by Cyprotex Inc. (Watertown, MA). Briefly, 10 million bacteria were exposed in triplicate to **20** for 90 min in a medium containing a low concentration of histidine. The cultures were then diluted into an indicator medium lacking histidine, dispensed into a 384-well plate, and incubated for 48 h at 37 °C. Cells that have undergone a reversion will grow, resulting in a color change. The number of wells showing growth were counted and compared to the vehicle control. An increase in the number of colonies of at least 2-fold over baseline and a dose–response indicated a positive response. Data were analyzed using an unpaired, one-sided Student's *t* test.

**Cytotoxicity Screening Panel.** HepG2 cells were plated on 384-well tissue culture-treated black-walled clear-bottomed polystyrene plates. The cells were treated with the test compound using a range of concentrations. At the end of the incubation period, the cells were loaded with the relevant dye/antibody for each cell health marker. The plates were then scanned using an automated fluorescent cellular imager, ArrayScan (Thermo Scientific Cellomics, Waltham, MA). Cytotoxicity was assessed using a multiparametric approach using high content screening (HCS).

**Mouse Plasma and Brain Tissue Sampling.** All animal studies were conducted by Cyprotex Inc. (Watertown, MA). The levels of **20** in mouse plasma and brain tissue samples were assessed as follows. A single IP dose (20 mg/kg) of **20** was administered to 6–8 week old male C57BL/6 mice. The formulation consisted of 10% dimethylacetamide (DMA) and 60% PEG400, balanced with 30% saline. Plasma and brain samples were collected across eight time points (5, 15, 30, 60, 120, 240, 480, and 1440 min). Brain samples were homogenized in 2 volumes (1:2 w/v dilution) of phosphate buffer solution (PBS). Once homogenized, the samples were precipitated with 3 volumes of acetonitrile containing an analytical internal standard (bucetin). Samples were then centrifuged to remove the precipitated protein, and the supernatant was analyzed by LC–MS/MS. All brain samples were compared to a calibration curve prepared from a control mouse brain. Plasma samples (5, 15, 30, 60, and 120 min time points) were diluted 10-fold with the control plasma. No dilutions were made for the 240, 480, and 1440 min plasma samples. All plasma samples were precipitated with 3 volumes of acetonitrile containing an analytical internal standard (bucetin). Samples were then centrifuged to remove the precipitated protein, and the supernatant was analyzed by LC–MS/MS. All plasma samples were compared to a calibration curve prepared from control mouse plasma. Samples were analyzed by LC–MS/MS using a Waters Xevo TQ mass spectrometer coupled with an Acquity UPLC and a CTC PAL chilled autosampler, all controlled by MassLynx software (Waters). After separation on a C18 reverse-phase HPLC column (Waters Acquity HSS T3 2.1  $\times$  50 mm<sup>2</sup> 1.8  $\mu$ M) using an acetonitrile–water gradient system, peaks were analyzed by mass spectrometry (MS) using ESI ionization in MRM mode. All work was performed with appropriate local health regulations and ethical approval. Three mice were used for each time point collected, and the data represent means  $\pm$  SEM.

**Statistical Analyses.** Nonlinear regression analyses were conducted using GraphPad Prism version 8.0.1 (GraphPad Software, Inc., La Jolla, CA). Results are expressed as means  $\pm$  SEM. EC<sub>50</sub>, IC<sub>50</sub>, E<sub>max</sub>, and I<sub>max</sub> values were calculated from individual concentration–response curves and then averaged to generate means and SEM values. Other statistical tests were performed as described in the legends.

## ■ ASSOCIATED CONTENT

### SI Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.jmedchem.0c00424>.

Bioanalysis of compound **20** in samples from mouse plasma and brain; functional profiling of CJ-1639 and pramipexole against an array of 168 known GPCRs; MS analysis for key analog **20**; HPLC and NMR spectra; and images of <sup>1</sup>H and <sup>13</sup>C NMR spectra for final analog compounds **1–109** (PDF)

Molecular formula strings and SAR data (CSV)

Protein data bank for D3R homology model (PDB)

## ■ AUTHOR INFORMATION

### Corresponding Authors

**Kevin J. Frankowski** – University of Kansas Specialized Chemistry Center, University of Kansas, Lawrence, Kansas 66047, United States; Center for Integrative Chemical Biology and Drug Discovery, UNC Eshelman School of Pharmacy, Chapel Hill, North Carolina 27599, United States; Phone: (919)-966-1659; Email: [kevinf@unc.edu](mailto:kevinf@unc.edu); Fax: (919)-843-8465

**David R. Sibley** – Molecular Neuropharmacology Section, National Institute of Neurological Disorders and Stroke, Intramural Research Program, National Institutes of Health, Bethesda, Maryland 20892-3723, United States; [orcid.org/0000-0002-0624-962X](https://orcid.org/0000-0002-0624-962X); Phone: (301)-496-9316; Email: [sibleyd@ninds.nih.gov](mailto:sibleyd@ninds.nih.gov); Fax: (301)-480-3726

### Authors

**Amy E. Moritz** – Molecular Neuropharmacology Section, National Institute of Neurological Disorders and Stroke, Intramural Research Program, National Institutes of Health, Bethesda, Maryland 20892-3723, United States

**R. Benjamin Free** – Molecular Neuropharmacology Section, National Institute of Neurological Disorders and Stroke, Intramural Research Program, National Institutes of Health, Bethesda, Maryland 20892-3723, United States; [orcid.org/0000-0002-2428-0487](https://orcid.org/0000-0002-2428-0487)

**Warren S. Weiner** – University of Kansas Specialized Chemistry Center, University of Kansas, Lawrence, Kansas 66047, United States

**Emmanuel O. Akano** – Molecular Neuropharmacology Section, National Institute of Neurological Disorders and Stroke, Intramural Research Program, National Institutes of Health, Bethesda, Maryland 20892-3723, United States

**Disha Gandhi** – Center for Integrative Chemical Biology and Drug Discovery, UNC Eshelman School of Pharmacy, Chapel Hill, North Carolina 27599, United States

**Ara Abramyan** – Computational Chemistry and Molecular Biophysics Unit, Molecular Targets and Medications Discovery Branch, National Institute on Drug Abuse, Intramural Research Program, National Institutes of Health, Baltimore, Maryland 21224, United States

**Thomas M. Keck** – Department of Chemistry & Biochemistry, Department of Molecular & Cellular Biosciences, College of Science and Mathematics, Rowan University, Glassboro, New Jersey 08028, United States; [orcid.org/0000-0003-1845-9373](https://orcid.org/0000-0003-1845-9373)

**Marc Ferrer** – NIH Chemical Genomics Center, Division of Preclinical Innovation, National Center for Advancing

Translational Sciences, National Institutes of Health, Rockville, Maryland 20850, United States

**Xin Hu** – NIH Chemical Genomics Center, Division of Preclinical Innovation, National Center for Advancing Translational Sciences, National Institutes of Health, Rockville, Maryland 20850, United States

**Noel Southall** – NIH Chemical Genomics Center, Division of Preclinical Innovation, National Center for Advancing Translational Sciences, National Institutes of Health, Rockville, Maryland 20850, United States

**Joseph Steiner** – NeuroTherapeutics Development Unit, National Institute for Neurological Disorders and Stroke, Intramural Research Program, National Institutes of Health, Bethesda, Maryland 20892, United States

**Jeffrey Aubé** – University of Kansas Specialized Chemistry Center, University of Kansas, Lawrence, Kansas 66047, United States; Center for Integrative Chemical Biology and Drug Discovery, UNC Eshelman School of Pharmacy, Chapel Hill, North Carolina 27599, United States; [orcid.org/0000-0003-1049-5767](https://orcid.org/0000-0003-1049-5767)

**Lei Shi** – Computational Chemistry and Molecular Biophysics Unit, Molecular Targets and Medications Discovery Branch, National Institute on Drug Abuse, Intramural Research Program, National Institutes of Health, Baltimore, Maryland 21224, United States

Complete contact information is available at:  
<https://pubs.acs.org/10.1021/acs.jmedchem.0c00424>

#### Author Contributions

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#### Notes

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#### ABBREVIATIONS USED

(MPTP), 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine; (HTR2A), 5HT<sub>2A</sub> serotonergic receptor; (6-OHDA), 6-hydroxydopamine; (ADRA2C), alpha<sub>2C</sub>-adrenergic receptor; (BRET), bioluminescence resonance energy transfer; (BDNF), brain-derived neurotrophic factor; (CCKAR), cholecystokinin A receptor; (DA), dopamine; (DAR), dopamine receptor; (DPBS), Dulbecco's phosphate-buffered saline; (EBSS), Earle's balanced salt solution; (EBI2), Epstein-Barr virus-induced GPCR 2; (pERK), ERK1/2 phosphorylation; (FGF), fibroblast growth factor; (GRKs), G protein-coupled receptor kinases; (GDNF), glial-derived neurotrophic factor; (GPCR), G protein-coupled receptor; (HTS), high-throughput screen; (IP1), inositol 1-phosphate; (MD), molecular dynamics; (MLPCN), Molecular Libraries Probe Production Centers Network; (OBS), orthosteric binding site; (PD), Parkinson's disease; (PEI), polyethylenimine; (PTGER2), prostaglandin E receptor 2; (PDSP), Psychoactive Drug Screening Program; (RLU), relative luminescence unit; (RLS), restless legs syndrome; (EDG8), sphingosine-1-phosphate 5 receptor; (SAR), structure-activity relationship; (TM), transmembrane domain

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