

University of Groningen

Clinical relevance of rifampicin-moxifloxacin interaction in isoniazid resistant/intolerant tuberculosis patients

Vogensen, Vanessa B; Bolhuis, Mathieu S; Sturkenboom, Marieke G G; van der Werf, Tjip S; de Lange, Wiel C M; Anthony, Richard M; van Soolingen, Dick; Alffenaar, Jan-Willem; Kerstjens, Huib A M; Akkerman, Onno W

Published in:
Antimicrobial Agents and Chemotherapy

DOI:
[10.1128/AAC.01829-21](https://doi.org/10.1128/AAC.01829-21)

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version
Publisher's PDF, also known as Version of record

Publication date:
2022

[Link to publication in University of Groningen/UMCG research database](#)

Citation for published version (APA):

Vogensen, V. B., Bolhuis, M. S., Sturkenboom, M. G. G., van der Werf, T. S., de Lange, W. C. M., Anthony, R. M., van Soolingen, D., Alffenaar, J-W., Kerstjens, H. A. M., & Akkerman, O. W. (2022). Clinical relevance of rifampicin-moxifloxacin interaction in isoniazid resistant/intolerant tuberculosis patients. *Antimicrobial Agents and Chemotherapy*, 66(2), [e01829-21]. <https://doi.org/10.1128/AAC.01829-21>

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).


The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: <https://www.rug.nl/library/open-access/self-archiving-pure/taverne-amendment>.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.



Clinical Relevance of Rifampicin-Moxifloxacin Interaction in Isoniazid-Resistant/Intolerant Tuberculosis Patients

Vanessa B. Vogensen,^a Mathieu S. Bolhuis,^b Marieke G. G. Sturkenboom,^b Tjip S. van der Werf,^a Wiel C. M. de Lange,^{a,c} Richard M. Anthony,^d Dick van Soolingen,^d  Jan-Willem Alffenaar,^{e,f,g}  Huib A. M. Kerstjens,^a  Onno W. Akkerman^{a,c}

^aUniversity of Groningen, University Medical Center Groningen, Department of Pulmonary Diseases and Tuberculosis, Groningen, the Netherlands

^bUniversity of Groningen, University Medical Center Groningen, Department of Clinical Pharmacy and Pharmacology, Groningen, the Netherlands

^cUniversity of Groningen, University Medical Center Groningen, TB Center Beatrixoord, Groningen, the Netherlands

^dNational Institute for Public Health and the Environment, TB Reference Laboratory (RIVM), Bilthoven, the Netherlands

^eFaculty of Medicine and Health, School of Pharmacy, University of Sydney, Sydney, NSW, Australia

^fWestmead Hospital, Westmead, NSW, Australia

^gMarie Bashir Institute for Infectious Diseases and Biosecurity, University of Sydney, Sydney, NSW, Australia

ABSTRACT Moxifloxacin is an attractive drug for the treatment of isoniazid-resistant rifampicin-susceptible tuberculosis (TB) or drug-susceptible TB complicated by isoniazid intolerance. However, co-administration with rifampicin decreases moxifloxacin exposure. It remains unclear whether this drug–drug interaction has clinical implications. This retrospective study in a Dutch TB center investigated how rifampicin affected moxifloxacin exposure in patients with isoniazid-resistant or -intolerant TB. Moxifloxacin exposures were measured between 2015 and 2020 in 31 patients with isoniazid-resistant or -intolerant TB receiving rifampicin, and 20 TB patients receiving moxifloxacin without rifampicin. Moxifloxacin exposure, i.e., area under the concentration–time curve (AUC_{0-24h}), and attainment of $AUC_{0-24h}/MIC > 100$ were investigated for 400 mg moxifloxacin and 600 mg rifampicin, and increased doses of moxifloxacin (600 mg) or rifampicin (900 mg). Moxifloxacin AUC_{0-24h} and peak concentration with a 400 mg dose were decreased when rifampicin was co-administered compared to moxifloxacin alone (ratio of geometric means 0.61 (90% CI (0.53, 0.70)) and 0.81 (90% CI (0.70, 0.94)), respectively). Among patients receiving rifampicin, 65% attained an $AUC_{0-24h}/MIC > 100$ for moxifloxacin compared to 78% of patients receiving moxifloxacin alone; this difference was not significant. Seven out of eight patients receiving an increased dose of 600 mg moxifloxacin reached the target $AUC_{0-24h}/MIC > 100$. This study showed a clinically significant 39% decrease in moxifloxacin exposure when rifampicin was co-administered. Moxifloxacin dose adjustment may compensate for this drug–drug interaction. Further exploring the impact of higher doses of these drugs in patients with isoniazid resistance or intolerance is paramount.

KEYWORDS *Mycobacterium tuberculosis*, isoniazid, moxifloxacin, pharmacokinetics, rifampicin

Tuberculosis (TB) continues to take a devastating toll on global health with an estimated 10 million new cases and 1.4 million deaths from TB in 2019 (1). The ongoing COVID-19 pandemic is expected to have led to major setbacks in the global elimination of TB (1, 2). Considering these setbacks, mitigating unsuccessful TB treatment remains a priority. While the standard treatment for pan-susceptible disease exhibits favorable outcomes in at least 85% of patients, overall treatment success is complicated by factors such as HIV co-infection, poor adherence, high interindividual variability in drug exposure, and, perhaps most worrisome of all, increasing drug resistance (1).

Copyright © 2022 American Society for Microbiology. All Rights Reserved.

Address correspondence to Onno W. Akkerman, o.w.akkerman@umcg.nl.

The authors declare no conflict of interest.

Received 17 September 2021

Returned for modification 8 October 2021

Accepted 17 November 2021

Accepted manuscript posted online

22 November 2021

Published 15 February 2022

Multi-drug resistant TB (MDR-TB) and extensively drug-resistant TB (XDR-TB) are strongly linked to poorer treatment success rates (1, 3), but isoniazid-resistant rifampicin-susceptible tuberculosis (Hr-TB) should not be overlooked. Hr-TB remains the most common TB resistance pattern worldwide, registered in on average 7.4% of new patients and 11.4% of previously treated patients (4). Hr-TB has also been associated with poorer treatment outcomes than drug-susceptible tuberculosis (DS-TB) and an increased risk of progression to MDR-TB (5–8). A recent study linked treatment of Hr-TB with standard first-line TB drugs to increased rates of treatment relapse for 10%, failure for 11%, and acquired rifampicin resistance for 8% of patients (5). Another study of data from 24 European countries between 2002 and 2014 reported an absolute difference in treatment success (cure or treatment completion) between DS-TB and Hr-TB of 5.3% (6).

Based on available evidence, including an individual-patient meta-analysis, the World Health Organization (WHO) recently issued the recommendation that for Hr-TB, the later-generation fluoroquinolone levofloxacin should be administered with rifampicin, ethambutol, and pyrazinamide (8, 9). However, the WHO graded this as a conditional recommendation, with very low certainty in the estimates of effects. The decision to recommend levofloxacin instead of moxifloxacin, another later-generation fluoroquinolone, in the conditional treatment recommendation was based on its supposed better studied safety profile and fewer known drug interactions (9).

Regimens containing rifampicin have repeatedly been shown to alter moxifloxacin pharmacokinetics (10–12). This can be explained by rifampicin-led induction of UDP glucuronosyltransferase (UGT) and sulfotransferase, both phase II metabolizing enzymes of moxifloxacin (10, 12, 13). Additionally, rifampicin induces P-glycoprotein (P-gp), a membrane protein expressed in intestinal mucosa, among other locations, that is responsible for the efflux of several xenobiotics, possibly including moxifloxacin (10, 14). Rifampicin has reduced the C_{max} of other orally administered P-gp substrates such as talinolol, digoxin, or fexofenadine by 19–70% (15). The key pharmacokinetic parameter area under the concentration-time curve over the span of 24 h (AUC_{0-24h}) of moxifloxacin can decrease by approximately 30% with rifampicin co-administration (10–12). Lower moxifloxacin exposures are likely to result in worse treatment outcomes as its bacteriological activity is known to be concentration-dependent (16).

Moxifloxacin dosing is considered optimal when the pharmacokinetic/pharmacodynamic parameter $fAUC_{0-24h}/MIC$, representing the free (protein unbound) AUC_{0-24h} divided by the MIC, is above 53 (17). Protein binding is often set at approximately 50%, although it can vary considerably per patient (18–20). Since unbound or free concentrations are seldom analyzed in clinical practice, a total (bound and unbound) AUC_{0-24h}/MIC exceeding 100 can also be used instead (10, 11). As earlier studies did not include MIC measurements for moxifloxacin and drug exposure across different rifampicin and moxifloxacin doses, with the exception of Pranger et al., who included data on patients receiving 150 mg, 300 mg, or 600 mg, it is unclear what the net result of drug–drug interaction between rifampicin and moxifloxacin is and whether it is clinically relevant (18). Resolving this uncertainty remains vital considering moxifloxacin may still play a role in the treatment of Hr-TB or of patients with intolerance to isoniazid in settings where levofloxacin is too expensive or inaccessible.

In addition, high-dose rifampicin is steadily gaining a more important role in TB treatment regimens, with some patients receiving dosages up to approximately 40 mg/kg of rifampicin (21–23). There is already some evidence on the effect of high-dose rifampicin on drug–drug interactions involving non-TB drugs (24, 25). Yet it is essential to understand how an increase in rifampicin dose will influence the exposure of other TB drugs such as moxifloxacin.

Therefore, this study aimed to analyze the effect of rifampicin co-administration on moxifloxacin exposure in patients diagnosed with Hr-TB and patients diagnosed with DS-TB who demonstrated intolerance to isoniazid across varying rifampicin and moxifloxacin doses.

TABLE 1 Demographic and clinical characteristics of patients^a

Variable name	Group MFX + RIF (n = 31)	Group MFX (n = 20)	Statistical test results ^b
Male sex, n (%)	24 (77)	14 (70)	<i>P</i> = 0.599
Age, median (IQR)	34 (24–52)	37 (25–43)	<i>P</i> = 0.656
Born in a foreign country, n (%)	28 (90)	16 (80)	<i>P</i> = 0.416
Body weight in kg ^c , median (IQR)	61.6 (52.1–71.2)	63.9 (55.1–76.7)	<i>P</i> = 0.569
Body mass index (BMI) ^c , median (IQR)	22.0 (18.1–24.3)	21.0 (19.0–23.2)	<i>P</i> = 0.883
PG-SGA (malnutrition) score \geq 9 n (%)	12 (75)	4 (44)	<i>P</i> = 0.200
Localization of disease, n (%)			<i>P</i> = 0.407
Pulmonary TB	21 (67)	10 (50)	
Extrapulmonary TB	6 (19)	7 (35)	
Pulmonary + extrapulmonary TB	4 (13)	3 (15)	
Previously treated for TB, n (%)	4 (13)	3 (15)	<i>P</i> = 1.00
Co-infections, n (%)			
HIV	0 (0)	2 (10)	
Hepatitis B	1 (3)	1 (5)	
Hepatitis C	1 (3)	1 (5)	
Comorbidities, n (%)			
Diabetes	8 (26)	3 (15)	<i>P</i> = 0.489
Mental health condition/ symptoms ^d	6 (19)	3 (15)	<i>P</i> = 0.724
Intermittent hemodialysis	1 (3)	0 (0)	NA
Risk factors, n (%)			
Smoking history	14 (45)	10 (50)	<i>P</i> = 0.817
Alcohol abuse	3 (10)	0 (0)	NA
IV drug use	0 (0)	1 (5)	NA

^aPatients who had blood sampled while on 400 mg/day moxifloxacin (Group MFX + RIF and Group MFX) and 600 mg/day rifampicin (Group MFX + RIF). Missing values: PG-SGA (Malnutrition Score), 15 (48.4%) of Group MFX + RIF and 11 (55.0%) of Group MFX. PG-SGA: Patient-Generated Subjective Global Assessment, a score of 9 or above indicates a critical need for a nutritional intervention.

^bResults of Mann-Whitney U test for continuous variables and Chi-Square or Fisher's Exact test for categorical variables.

^cWeight and BMI at time of blood sampling.

^dAlso includes patients without an explicit diagnosis of mental illness but who received medication or psychological support as a result of mental health symptoms.

RESULTS

Thirty-one patients were included in Group MFX + RIF (77% male, median age of 34 and a median BMI 22.0) and 20 patients were included in Group MFX (70% male, median age of 37 and a median BMI of 21.0) (see Table 1). Most patients in both groups were foreign-born, were diagnosed with pulmonary TB, and were treatment naive. The daily dose of moxifloxacin per kg body weight did not differ significantly between the groups (median of 6.47 for Group MFX + RIF versus 6.49 mg/kg for Group MFX, *P* = 0.613).

In Group MFX + RIF, 24/31 patients were intolerant to isoniazid; 15 had previously experienced hepatotoxicity, 7 had peripheral polyneuropathy, 1 had psychiatric complaints, and 1 had a combination of the already mentioned adverse effects. Hr-TB was diagnosed in 6/31 patients, and one patient did not start isoniazid because of failed isoniazid monotherapy for latent tuberculosis infection. In Group MFX, 17 patients had MDR-TB, 1 patient was intolerant to rifampicin, 1 had rifampicin-resistant TB without resistance to isoniazid, and 1 had DS-TB and had temporarily received moxifloxacin due to at first being erroneously diagnosed with rifampicin-resistant TB. One patient in Group MFX + RIF was removed from further analyses because of an extreme moxifloxacin AUC_{0–24h} value of 80 mg*h/liter. Blood sampling was performed on this patient 5 days after beginning co-treatment with rifampicin and moxifloxacin. Therefore, maximal rifampicin-led enzymatic induction would likely not have been achieved yet, although dosing or lab errors cannot be excluded.

The main pharmacokinetic and pharmacodynamic variables for MFX for both groups are shown in Table 2. For a 400 mg moxifloxacin dose, the geometric mean for the moxifloxacin AUC_{0–24h} was 16.8 (geometric standard deviation 1.34) in Group MFX + RIF and 27.6 (geometric standard deviation 1.29) in Group MFX. The geometric

TABLE 2 Main pharmacokinetic and pharmacodynamic variables for MFX^a

Variable name (unit)	Group MFX+RIF (n = 30)		Group MFX (n = 20)		GMR (90% CI)
	Arithmetic Mean (SD)	Geometric Mean (SD ^b)	Arithmetic Mean (SD)	Geometric Mean (SD ^b)	
AUC _{0-24h} (mg*h*L ⁻¹)	17.5 (5.45)	16.8 (1.34)	28.5 (7.30)	27.6 (1.29)	0.61 (0.53–0.70)
Cmax (mg/L)	2.16 (0.67)	2.06 (1.37)	2.62 (0.69)	2.54 (1.30)	0.81 (0.70–0.94)
T _{max} ^c (h)	2.00 (1.03–2.89)		2.02 (1.32–4.06)		
MIC ^d , n%	17 (55)		18 (90)		
MIC ^d (mg/L), n%					
0.125	12 (39)		9 (45)		
0.25	3 (10)		9 (45)		
0.5	2 (6)		0 (0)		
AUC _{0-24h} /MIC ^c (h)	117 (74.1–145)		156 (100–214)		
AUC _{0-24h} /MIC > 100 ^d , n%	11 (65)		14 (78)		

^aBased on a dose of 400 mg moxifloxacin (both groups) and 600 mg rifampicin (Group MFX+RIF). MFX: moxifloxacin; RIF: rifampicin; AUC_{0-24h}: 24-h area under the concentration-time curve; Cmax: peak observed concentration; T_{max}: time when Cmax occurs; GMR: geometric mean ratio (Group MFX+RIF/MFX).

^bGeometric standard deviations.

^cExpressed in median and interquartile range instead of means and standard deviation.

^dExpressed in n (%) instead of means and includes all patients, n = 51.

mean for the moxifloxacin AUC_{0-24h} in Group MFX+RIF was significantly lower than that of Group MFX. Similarly, Cmax was significantly lower in Group MFX+RIF.

In Group MFX+RIF, 11/17 (65%) patients attained the target of AUC_{0-24h}/MIC > 100 for moxifloxacin compared to 14/18 (78%) patients in Group MFX. These proportions were not significantly different (*P* = 0.471). In patients with an unknown MIC in Group MFX+RIF (14/31), 2/14 patients had a moxifloxacin AUC_{0-24h} that would be too low for a MIC of 0.125 mg/liter, 11/14 patients had a moxifloxacin AUC_{0-24h} lower than needed for a MIC of 0.25 mg/liter, and 13/14 had an AUC_{0-24h} lower than needed for a MIC of 0.5 mg/liter.

The stepwise regression analysis with AUC_{0-24h} as the outcome variable resulted in a model with the best overall fit (*R*² = 0.629) composed of group (*B* = 1.55, *P* < 0.001), sex (*B* = 1.31, *P* = 0.001) with female coded as 1, and diabetes mellitus (*B* = 0.79, *P* = 0.006).

The geometric mean of moxifloxacin AUC_{0-24h} in six patients receiving 600 mg moxifloxacin and 600 mg rifampicin did not differ significantly from that of six patients receiving 600 mg moxifloxacin and 900 mg rifampicin (Table 3).

After a dose increase from 400 mg to 600 mg moxifloxacin in 10 patients, the AUC_{0-24h} of moxifloxacin increased significantly resulting in a geometric mean ratio of 1.48 (90% confidence interval (CI) 1.29–1.71), which also indicates a linear dose–exposure relationship (Table 4). When linked to MICs available for 8/10 patients, 7 of these 8 patients (88%) reached an AUC_{0-24h}/MIC > 100 at 600 mg moxifloxacin. For the two patients without MIC data, the AUC_{0-24h} of moxifloxacin was high enough to reach the target AUC_{0-24h}/MIC > 100 if, in the case of the first patient, the MIC was ≤ 0.125 mg/liter and, in the case of the second patient, the MIC was ≤ 0.25 mg/liter.

DISCUSSION

Patients receiving moxifloxacin with rifampicin showed considerably lower moxifloxacin exposure than patients receiving moxifloxacin alone. This effect does not

TABLE 3 Differences in MFX AUC_{0-24h} between patients on 600 mg RIF 600 mg MFX versus patients on 900 mg RIF 600 mg MFX^a

Dose combination	Group size	Arithmetic mean (SD)	Geometric mean (SD ^b)	GMR (90% CI)
900 mg RIF 600 mg MFX	6	24.2 (8.11)	23.1 (1.40)	1.04 (0.72–1.48)
600 mg RIF 600 mg MFX	6	22.7 (4.49)	22.3 (1.23)	

^aAUC_{0-24h}: 24-h area under the concentration-time curve; MFX: moxifloxacin; RIF: rifampicin; GMR: geometric mean ratio.

^bGeometric standard deviation.

TABLE 4 Differences in MFX AUC_{0-24h} between patients that first received 600 mg RIF 400 mg MFX followed by 600 mg RIF 600 mg MFX^a

Dose combination	Arithmetic mean (SD)	Geometric mean (SD) ^b	GMR (90% CI)
600 mg RIF 600 mg MFX	24.1 (4.61)	23.6 (1.22)	1.48 (1.29–1.71)
600 mg RIF 400 mg MFX	16.2 (2.68)	16.0 (1.19)	

^aTen patients in total. AUC_{0-24h} , 24-h area under the concentration-time curve; MFX: moxifloxacin; RIF: rifampicin; GMR: geometric mean ratio.

^bGeometric standard deviation.

seem to be dependent on rifampicin dosing as the six patients receiving a higher dose of rifampicin (900 mg) did not have a significantly lower moxifloxacin exposure when compared to the six patients receiving a standard dose of rifampicin (600 mg). In addition, an increase in moxifloxacin dose from 400 to 600 mg resulted in target attainment of $AUC_{0-24h}/MIC > 100$ in 7/8 patients.

These results are in agreement with previous studies. In crossover or sequential design studies, an absolute decrease of 27–31% has been reported in moxifloxacin AUC_{0-24h} among patients with TB or volunteers receiving rifampicin compared to those receiving moxifloxacin alone (10–12). In 19 TB patients, decreases of 31% and 32% were registered for moxifloxacin AUC_{0-24h} and C_{max} , respectively (11). The difference in moxifloxacin AUC_{0-24h} measured in this study between the two patient groups of 39% was comparable to earlier studies (11, 12). The same can be said about C_{max} with a difference of 19% registered in this study that is slightly lower than previously reported by Nijland et al. (11). However, another study with 101 TB patients reported no difference in C_{max} (26).

Variability in drug exposure across studies can be clarified by different dosing patterns, study designs, and study populations (patients with TB versus healthy volunteers) (10). In other clinical studies among TB patients with smaller sample sizes, no significant differences in AUC_{0-24h} and C_{max} of moxifloxacin between patient groups receiving both moxifloxacin and rifampicin and patients receiving moxifloxacin alone were detected (18, 26, 27). Close to a third of our patients receiving moxifloxacin and rifampicin for whom MICs for moxifloxacin were available did not reach the target $AUC_{0-24h}/MIC > 100$. Our finding that a 400 mg moxifloxacin dose is often inadequate to attain the established PK/PD targets is mirrored in other published studies of drug-resistant TB (10, 17, 28).

Diabetes mellitus significantly contributed to a decrease in moxifloxacin AUC_{0-24h} in the multivariate model, as was also the case in another retrospective study conducted at the same TB center that included most of the patients with diabetes that were included in this study. This effect might be attributed to an increased moxifloxacin clearance in patients with diabetes (29). Interestingly, female sex contributed to an increased moxifloxacin AUC_{0-24h} . However, confounding by other variables cannot be excluded as male patients differed considerably from their female counterparts. Male patients comprised all previously treated cases, most cases with a tobacco smoking history, all cases of alcohol abuse, and received a lower moxifloxacin dose in mg/kg compared to female patients.

Interestingly, patients receiving 600 mg or 900 mg of rifampicin showed no significant difference in moxifloxacin AUC_{0-24h} . Nevertheless, the extent of the effect of a 900 mg dose of rifampicin, usually corresponding to 15 mg/kg, on induction of P-gp or phase II metabolism enzymes and on moxifloxacin exposure could be better ascertained by adopting a 2-period sequential study design. In the majority of studies, only rifampicin doses up to 600 mg, typically equivalent to 10 mg/kg, were assessed (10, 15, 30). Meanwhile, daily doses of rifampicin of up to 35 mg/kg have already been studied or administered in patients with TB (21–23).

An important question deriving from previous studies was whether higher moxifloxacin doses could offset the reduction in drug exposure caused by rifampicin. In our study, a higher dose of 600 mg moxifloxacin per day was paired with an AUC_{0-24h}/MIC that

was satisfactory for seven of eight patients. Although not covered by this study, a dose of 800 mg moxifloxacin, which is already in use in the standardized shorter MDR-TB treatment, has been previously identified as maximizing the chances of resistance suppression, outperforming a 400 mg dose by far as well as performing slightly better than a 600 mg dose (17).

The strength of this study lies in describing the impact of the moxifloxacin–rifampicin interaction on moxifloxacin exposure and AUC_{0-24h}/MIC while using individual MIC values, contrary to other studies that used a preset MIC, usually set at 0.5 mg/liter (11, 31). The addition of levofloxacin in the WHO recommendations for treatment of Hr-TB marked an important step in improving treatment for patients with Hr-TB. Moxifloxacin may be of clinical interest in situations in which levofloxacin is not readily available or when there is a substantial difference in cost. We established that attainment of the AUC_{0-24h}/MIC target did not differ significantly between patients receiving both rifampicin and moxifloxacin and patients receiving moxifloxacin alone and explored the effect of increasing the dose of moxifloxacin or rifampicin on the exposure and AUC_{0-24h}/MIC of moxifloxacin. This could lead to further expanding the exploration of different moxifloxacin and rifampicin dose combinations.

The main limitation of this study is its retrospective design. The risk of selection bias is heightened in the control group because indications for therapeutic drug monitoring (TDM) included malnutrition, comorbidities, or MDR-TB, which may independently contribute to changes in drug exposure. Moreover, individual MIC were unattainable for a sizeable number of patients, especially in Group MFX+RIF, which is important for the interpretation of the resulting $AUC_{0-24h}/MIC > 100$ values. We were not able to evaluate the contribution of genetic variants in the UGT1A and ABCB1 genes coding for one of the main UGT involved in the formation of the moxifloxacin M2 metabolite and for P-gp, respectively, to the variability in moxifloxacin pharmacokinetics (12, 32). Lastly, the patients included were mostly HIV-negative. An interaction between moxifloxacin and efavirenz has been documented resulting in a 30% decrease in moxifloxacin AUC (33). However, potential interactions between moxifloxacin and antiretroviral drugs or supplements commonly used in the treatment of HIV, and how these may potentiate the moxifloxacin–rifampicin interaction, remain to be explored.

In conclusion, this retrospective study showed a 39% absolute decrease in moxifloxacin exposure when rifampicin was co-administered among patients with TB in whom an alternative drug for isoniazid was required. In a small subgroup of patients on moxifloxacin and rifampicin, an increased dose of 600 mg/day moxifloxacin resulted in promising attainment of the $AUC_{0-24h}/MIC > 100$ target. In situations where treatment of Hr-TB with levofloxacin is not possible, (high-dose) moxifloxacin may still be a viable alternative. TDM would then be performed to identify patients in need of a higher moxifloxacin dose. If TDM is not readily available, patients who possess risk factors for low moxifloxacin exposures could receive a higher moxifloxacin dose followed by monitoring for potential signs of toxicity with ECGs, for instance. Further pharmacokinetic/pharmacodynamic research comparing the effectiveness and safety of higher moxifloxacin doses when combined with (high-dose) rifampicin with the currently recommended levofloxacin for patients with TB that require an alternative for isoniazid is warranted.

MATERIALS AND METHODS

Study design. This study was a retrospective analysis of routinely collected data from patients with TB admitted to the Tuberculosis Center in the Beatrixoord location of the University Medical Center of Groningen (UMCG), situated in Haren, the Netherlands.

Ethics. Due to the retrospective nature of the study based on routine care collection of data, the need for the protocol to be reviewed and approved was waived by the Institutional Ethics Review Board of the UMCG (METc 2018/095).

Study subjects. Group MFX+RIF were patients with Hr-TB or intolerance to isoniazid who received moxifloxacin in combination with rifampicin as part of their treatment regimen. Group MFX were patients that received moxifloxacin but no rifampicin. Patients were included regardless of age or disease localization, and if they had one serial blood sampling episode with more than four samples

between 1-1-2015 and 16-9-2020 for the estimation of plasma moxifloxacin exposure while receiving 600 mg/day rifampicin (only Group MFX+RIF) and 400 mg/day moxifloxacin as starting doses.

Resistance to isoniazid was confirmed by means of the BACTEC Mycobacterial Growth Indicator Tube (MGIT) test and molecular detection of mutations in the *inhA* and *katG* genes confidently associated with isoniazid resistance in the *M. tuberculosis* isolates. Low-level resistance to isoniazid, defined as an absence of growth at the breakpoint concentration of 0.4 mg/liter, was also considered. Intolerance was defined as isoniazid being interrupted due to adverse events likely to be caused by this drug, or isoniazid not being started altogether by the treating physician due to an underlying condition that would make the occurrence of adverse events highly probable.

Study procedures. Patients were included if a pharmacokinetic curve consisting of a predose sample and three to seven samples up to 8 h postdose was obtained as part of standard of care. All pharmacokinetic calculations were based on data collected in steady-state conditions. The analysis of moxifloxacin concentrations was performed using validated liquid chromatography-mass spectrometry methods (34). Clinical and demographic data were obtained from patient dossiers. Maximum concentration (C_{max}) was defined as the highest observed concentration, with the time to maximum concentration (T_{max}) as the time to reach C_{max} . The pharmacokinetic variables were calculated by means of one-compartmental population pharmacokinetic models in MWPharm ++ (version 1.87; Medware, Prague, Czech Republic). The model parameters were apparent clearance 19.9 ± 8.8 liters/h and 14.7 ± 5.7 liters/h, apparent volume of distribution 2.82 ± 0.69 L/kg body weight and 2.75 ± 1.01 L/kg body weight, absorption constant 7.4 ± 6.8 h⁻¹ and 6.3 ± 4.8 h⁻¹ and lag time of 0.75 ± 0.11 h and 0.88 ± 0.24 h for the MFX+RIF model and MFX model respectively (26). Individual MICs of moxifloxacin for the included patients were requested from the National TB Reference Laboratory to calculate moxifloxacin AUC_{0-24h}/MIC .

Study endpoints and statistical analysis. The patient characteristics age, sex, country of birth, body weight and body mass index (BMI) at time of blood sampling, localization of disease, history of TB treatment, co-infections, comorbidities, risk factors for poor treatment outcome, and the PG-SGA score were obtained from the electronic health records. PG-SGA score stands for Patient-Generated Subjective Global Assessment score; this is a nutrition assessment tool where nine points or more indicate a critical need for nutritional interventions (35). Patient characteristics were compared between the groups MFX + RIF and MFX. Medians were compared using the Mann-Whitney U test. Proportions were compared using the Chi-Square or Fisher exact test. Characteristics associated ($P < 0.2$) with the main outcome in univariate analyses were included in stepwise regression analyses ($P < 0.05$ to enter, $P > 0.10$ to remove). The main outcome was moxifloxacin AUC_{0-24h} . Secondary outcomes included the pharmacokinetic variables C_{max} and T_{max} .

The variables AUC_{0-24h} and C_{max} were natural log (ln) transformed before analyses and later back-transformed. Arithmetic means with standard deviations, geometric means with geometric standard deviations, and geometric mean ratios with 90% confidence intervals (CI) were calculated for group comparisons. For T_{max} medians and interquartile ranges were presented and the Mann-Whitney U test was applied for comparisons.

The moxifloxacin AUC_{0-24h} of two patient subgroups, both receiving 600 mg moxifloxacin but receiving different rifampicin doses (600 mg or 900 mg), were compared. The moxifloxacin AUC_{0-24h} in patients receiving first 400 mg moxifloxacin and later receiving 600 mg moxifloxacin was also studied.

A significance level of 0.05 was chosen for all analyses. All statistical analyses were performed using IBM SPSS Statistics version 23.

ACKNOWLEDGMENT

No funding was received for the present study.

REFERENCES

- World Health Organization (WHO). 2020. Global tuberculosis report 2020. World Health Organization, Geneva, Switzerland. <https://www.who.int/publications/item/9789240013131>. Accessed 12 March 2021.
- Ong CWM, Migliori GB, Raviglione M, MacGregor-Skinner G, Sotgiu G, Alffenaar J-W, Tiberi S, Adlhoch C, Alonzi T, Archuleta S, Brusin S, Cambau E, Capobianchi MR, Castilletti C, Centis R, Cirillo DM, D'Ambrosio L, Delogu G, Esposito SMR, Figuerola J, Friedland JS, Ho BCH, Ippolito G, Jankovic M, Kim HY, Rosales Klintz S, Ködmön C, Lalle E, Leo YS, Leung C-C, Mårtson A-G, Melazzini MG, Najafi Fard S, Penttinen P, Petrone L, Petruccioli E, Pontali E, Sadari L, Santin M, Spanevello A, van Crevel R, van der Werf MJ, Visca D, Viveiros M, Zellweger J-P, Zumla A, Goletti D. 2020. Epidemic and pandemic viral infections: impact on tuberculosis and the lung. *Eur Respir J* 56:2001727. <https://doi.org/10.1183/13993003.01727-2020>.
- Conradie F, Diacon AH, Ngubane N, Howell P, Everitt D, Crook AM, Mendel CM, Egizi E, Moreira J, Timm J, McHugh TD, Wills GH, Bateson A, Hunt R, Van Niekerk C, Li M, Olugbosi M, Spigelman M, Nix-TB Trial Team. 2020. Treatment of highly drug-resistant pulmonary tuberculosis. *N Engl J Med* 382:893–902. <https://doi.org/10.1056/NEJMoa1901814>.
- Dean AS, Zignol M, Cabibbe AM, Falzon D, Glaziou P, Cirillo DM, Köser CU, Gonzalez-Angulo LY, Tosas-Auget O, Ismail N, Tahseen S, Ama MCG, Skrahina A, Alikhanova N, Kamal SMM, Floyd K. 2020. Prevalence and genetic profiles of isoniazid resistance in tuberculosis patients: a multi-country analysis of cross-sectional data. *PLoS Med* 17:e1003008. <https://doi.org/10.1371/journal.pmed.1003008>.
- Gegia M, Winters N, Benedetti A, van Soolingen D, Menzies D. 2017. Treatment of isoniazid-resistant tuberculosis with first-line drugs: a systematic review and meta-analysis. *Lancet Infect Dis* 17:223–234. [https://doi.org/10.1016/S1473-3099\(16\)30407-8](https://doi.org/10.1016/S1473-3099(16)30407-8).
- Karo B, Kohlenberg A, Hollo V, Duarte R, Fiebig L, Jackson S, Kearns C, Ködmön C, Korzeniewska-Kosela M, Papaventsis D, Solovic I, van Soolingen D, van der Werf MJ. 2019. Isoniazid (INH) mono-resistance and tuberculosis (TB) treatment success: analysis of European surveillance data, 2002 to 2014. *Eurosurveillance* 24:1800392. <https://doi.org/10.2807/1560-7917.ES.2019.24.12.1800392>.
- Stagg HR, Lipman MC, McHugh TD, Jenkins HE. 2017. Isoniazid-resistant tuberculosis: a cause for concern? *Int J Tuberc Lung Dis* 21:129–139. <https://doi.org/10.5588/ijtld.16.0716>.
- Fregonese F, Ahuja SD, Akkerman OW, Arakaki-Sanchez D, Ayakaka I, Baghaei P, Bang D, Bastos M, Benedetti A, Bonnet M, Cattamanchi A, Cegielski P, Chien J-Y, Cox H, Dedicato M, Erkens C, Escalante P, Falzon D, Garcia-Prats AJ, Gegia M, Gillespie SH, Glynn JR, Goldberg S, Griffith D,

- Jacobson KR, Johnston JC, Jones-López EC, Khan A, Koh W-J, Kritski A, Lan ZY, Lee JH, Li PZ, Maciel EL, Galliez RM, Merle CSC, Munang M, Narendran G, Nguyen VN, Nunn A, Ohkado A, Park JS, Phillips PPJ, Ponnuraja C, Reves R, Romanowski K, Seung K, Schaaf HS, Skrahina A, Soilingen D van, Tabarsi P, et al. 2018. Comparison of different treatments for isoniazid-resistant tuberculosis: an individual patient data meta-analysis. *Lancet Respir Med* 6:265–275. [https://doi.org/10.1016/S2213-2600\(18\)30078-X](https://doi.org/10.1016/S2213-2600(18)30078-X).
9. World Health Organization (WHO). 2018. WHO treatment guidelines for isoniazid-resistant tuberculosis: supplement to the WHO treatment guidelines for drug-resistant tuberculosis. World Health Organization, Geneva, Switzerland. https://www.who.int/tb/publications/2018/WHO_guidelines_isoniazid_resistant_TB/en/. Accessed 18 March 2021.
 10. Naidoo A, Naidoo K, McIlleron H, Essack S, Padayatchi N. 2017. A Review of moxifloxacin for the treatment of drug-susceptible tuberculosis. *J Clin Pharmacol* 57:1369–1386. <https://doi.org/10.1002/jcph.968>.
 11. Nijland HMJ, Ruslami R, Suroto AJ, Burger DM, Alisjahbana B, Van Crevel R, Aarnoutse RE. 2007. Rifampicin reduces plasma concentrations of moxifloxacin in patients with tuberculosis. *Clin Infect Dis* 45:1001–1007. <https://doi.org/10.1086/521894>.
 12. Weiner M, Burman W, Luo CC, Peloquin CA, Engle M, Goldberg S, Agarwal V, Vernon A. 2007. Effects of rifampin and multidrug resistance gene polymorphism on concentrations of moxifloxacin. *Antimicrob Agents Chemother* 51:2861–2866. <https://doi.org/10.1128/AAC.01621-06>.
 13. Burman WJ, Galliano K, Peloquin C. 2001. Comparative pharmacokinetics and pharmacodynamics of the rifamycin antibacterials. *Clin Pharmacokinet* 40:327–341. <https://doi.org/10.2165/00003088-200140050-00002>.
 14. Brillault J, De Castro WV, Harnois T, Kitzis A, Olivier J-C, Couet W. 2009. P-Glycoprotein-mediated transport of moxifloxacin in a Calu-3 lung epithelial cell model. *Antimicrob Agents Chemother* 53:1457–1462. <https://doi.org/10.1128/AAC.01253-08>.
 15. Elmeliegy M, Vourvahis M, Guo C, Wang DD. 2020. Effect of P-glycoprotein (P-gp) inducers on exposure of P-gp substrates: review of clinical drug–drug interaction studies. *Clin Pharmacokinet* 59:699–714. <https://doi.org/10.1007/s40262-020-00867-1>.
 16. Hu Y, Coates ARM, Mitchison DA. 2003. Sterilizing activities of fluoroquinolones against rifampin-tolerant populations of *Mycobacterium tuberculosis*. *Antimicrob Agents Chemother* 47:653–657. <https://doi.org/10.1183/09031936.00176610>.
 17. Gumbo T, Louie A, Deziel MR, Parsons LM, Salfinger M, Drusano GL. 2004. Selection of a moxifloxacin dose that suppresses drug resistance in *Mycobacterium tuberculosis*, by use of an in vitro pharmacodynamic infection model and mathematical modeling. *J Infect Dis* 190:1642–1651. <https://doi.org/10.1086/424849>.
 18. Pranger AD, van Alena R, Aarnoutse RE, van Soilingen D, Uges DRA, Kosterink JGW, van der Werf TS, Alffenaar JWC. 2011. Evaluation of moxifloxacin for the treatment of tuberculosis: 3 years of experience. *Eur Respir J* 38:888–894. <https://doi.org/10.1183/09031936.00176610>.
 19. Stass H, Dalhoff A, Kubitzka D, Schühly U. 1998. Pharmacokinetics, safety, and tolerability of ascending single doses of moxifloxacin, a new 8-methoxy quinolone, administered to healthy subjects. *Antimicrob Agents Chemother* 42:2060–2065. <https://doi.org/10.1128/AAC.42.8.2060>.
 20. Wright DH, Brown GH, Peterson ML, Rotschafer JC. 2000. Application of fluoroquinolone pharmacodynamics. *J Antimicrob Chemother* 46:669–683. <https://doi.org/10.1093/jac/46.5.669>.
 21. Seijger C, Hoefsloot W, Guchteneire IB, De Te Brake L, Van Ingen J, Kuipers S, Van Crevel R, Aarnoutse R, Boeree M, Magis-Escorra C. 2019. High-dose rifampicin in tuberculosis: experiences from a Dutch tuberculosis centre. *PLoS One* 14:e0213718. <https://doi.org/10.1371/journal.pone.0213718>.
 22. Peloquin CA, Velásquez GE, Lecca L, Calderón RI, Coit J, Milstein M, Osso E, Jimenez J, Tintaya K, Sanchez Garavito E, Vargas Vasquez D, Mitnick CD, Davies G. 2017. Pharmacokinetic evidence from the HIRIF trial to support increased doses of rifampin for tuberculosis. *Antimicrob Agents Chemother* 61:e00038-17. <https://doi.org/10.1128/AAC.00038-17>.
 23. ClinicalTrials.gov. 2021. Rifampicin at high dose for difficult-to-treat tuberculosis (RIAlta). National Library of Medicine, Bethesda, MD, USA. Identifier: NCT00287391. <https://clinicaltrials.gov/ct2/show/NCT04768231>
 24. Niemi M, Backman JT, Fromm MF, Neuvonen PJ, Kivistö KT. 2003. Pharmacokinetic interactions with rifampicin. *Clin Pharmacokinet* 42:819–850. <https://doi.org/10.2165/00003088-200342090-00003>.
 25. Sekaggya-Wiltshire C, Nabisere R, Musaazi J, Aber F, Otaalo B, Lamorde M, Denti P, Aarnoutse R, Dooley K, Sloan DJ. 2021. PK and safety of high-dose rifampicin in TB/HIV coinfecting patients on efavirenz or dolutegravir, abstr 90. Abstr Virtual Conference on Retroviruses and Opportunistic Infections. <https://www.croiconference.org/abstract/pk-and-safety-of-high-dose-rifampicin-in-tb-hiv-coinfecting-patients-on-efv-or-dtg/>.
 26. van den Elsen SHJ, Sturkenboom MGG, Akkerman OW, Manika K, Kioumis IP, van der Werf TS, Johnson JL, Peloquin C, Touw DJ, Alffenaar JWC. 2019. Limited sampling strategies using linear regression and the Bayesian approach for the therapeutic drug monitoring of moxifloxacin in tuberculosis patients. *Antimicrob Agents Chemother* 63:e00384-19. <https://doi.org/10.1128/AAC.00384-19>.
 27. Manika K, Chatzika K, Papaioannou M, Kontou P, Boutou A, Zarogoulidis K, Kioumis I. 2015. Rifampicin-moxifloxacin interaction in tuberculosis treatment: A real-life study. *Int J Tuberc Lung Dis* 19:1383–1387. <https://doi.org/10.5588/ijtld.14.0935>.
 28. Davies Forsman L, Niward K, Kuhlin J, Zheng X, Zheng R, Ke R, Hong C, Werngren J, Paus J, Simonsson USH, Eliasson E, Hoffner S, Xu B, Alffenaar J-W, Schön T, Hu Y, Bruchfeld J. 2021. Suboptimal moxifloxacin and levofloxacin drug exposure during treatment of patients with multidrug-resistant tuberculosis: results from a prospective study in China. *Eur Respir J* 57:2003463. <https://doi.org/10.1183/13993003.03463-2020>.
 29. Dekkers BG, Bolhuis MS, Ter Beek L, de Lange WC, van der Werf TS, Alffenaar JW, Akkerman OW. 2019. Reduced moxifloxacin exposure in patients with tuberculosis and diabetes. *Eur Respir J* 54:1900373. <https://doi.org/10.1183/13993003.00373-2019>.
 30. Lutz JD, Kirby BJ, Wang L, Song Q, Ling J, Massetto B, Worth A, Kearney BP, Mathias A. 2018. Cytochrome P450 3A induction predicts P-glycoprotein induction; part 1: establishing induction relationships using ascending dose rifampin. *Clin Pharmacol Ther* 104:1182–1190. <https://doi.org/10.1002/cpt.1073>.
 31. Ramachandran G, Kumar AKH, Srinivasan R, Geetharani A, Sugirda P, Nandhakumar B, Nandini R, Tharani CB. 2012. Effect of rifampicin & isoniazid on the steady state pharmacokinetics of moxifloxacin. *Indian J Med Res* 136:979–984. <https://pubmed.ncbi.nlm.nih.gov/23391793/>.
 32. Naidoo A, Ramsuran V, Chirehwa M, Denti P, McIlleron H, Naidoo K, Yende-Zuma N, Singh R, Ngcapu S, Chaudhry M, Pepper MS, Padayatchi N. 2018. Effect of genetic variation in UGT1A and ABCB1 on moxifloxacin pharmacokinetics in South African patients with tuberculosis. *Pharmacogenomics* 19:17–29. <https://doi.org/10.2217/pgs-2017-0144>.
 33. Naidoo A, Chirehwa M, McIlleron H, Naidoo K, Essack S, Yende-Zuma N, Kimba-Phongi E, Adamson J, Govender K, Padayatchi N, Denti P. 2017. Effect of rifampicin and efavirenz on moxifloxacin concentrations when co-administered in patients with drug-susceptible TB. *J Antimicrob Chemother* 72:1441–1449. <https://doi.org/10.1093/jac/dkx004>.
 34. Pranger AD, Alffenaar JWC, Wessels AMA, Greijdanus B, Uges DRA. 2010. Determination of moxifloxacin in human plasma, plasma ultrafiltrate, and cerebrospinal fluid by a rapid and simple liquid chromatography-tandem mass spectrometry method. *J Anal Toxicol* 34:135–141. <https://doi.org/10.1093/jat/34.3.135>.
 35. Jager-Wittenaar H, Ottery FD. 2017. Assessing nutritional status in cancer: role of the patient-generated subjective global assessment. *Curr Opin Clin Nutr Metab Care* 20:322–329. <https://doi.org/10.1097/MCO.0000000000000389>.