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Two dephosphorylation pathways of inositol 1,4,5-trisphosphate in homogenates of the cellular slime mould *Dictyostelium discoideum*

Michiel M. VAN LOOKEREN CAMPAGNE,*† Cristophe ERNEUX,† Ronald VAN EIJK* and Peter J. M. VAN HAASTERT*

*Cell Biology and Genetics Unit, Zoological Laboratory, University of Leiden, Kaiserstraat 63, NL-2311 GP Leiden, The Netherlands, and †Institute of Interdisciplinary Research (IRIBHN), School of Medicine, Free University of Brussels (U.L.B.), Campus Erasme (Bât. C), Route de Lennik 808, B-1070 Brussels, Belgium

Dictyostelium discoideum homogenates contain phosphatase activity which rapidly dephosphorylates Ins(1,4,5) P_3 (D-*myo*-inositol 1,4,5-trisphosphate) to Ins (*myo*-inositol). When assayed in Mg^{2+} , Ins(1,4,5) P_3 is dephosphorylated by the soluble *Dictyostelium* cell fraction to 20% Ins(1,4) P_2 (D-*myo*-inositol 1,4-bisphosphate) and 80% Ins(4,5) P_2 (D-*myo*-inositol 4,5-bisphosphate). In the particulate fraction Ins(1,4,5) P_3 5-phosphatase is relatively more active than the Ins(1,4,5) P_3 1-phosphatase. $CaCl_2$ can replace $MgCl_2$ only for the Ins(1,4,5) P_3 5-phosphatase activity. Ins(1,4) P_2 and Ins(4,5) P_2 are both further dephosphorylated to Ins4P (D-*myo*-inositol 4-monophosphate), and ultimately to Ins. Li^+ ions inhibit Ins(1,4,5) P_3 1-phosphatase, Ins(1,4) P_2 1-phosphatase, Ins4P phosphatase and L-Ins1P (L-*myo*-inositol 1-monophosphate) phosphatase activities; Ins(1,4,5) P_3 1-phosphatase is 10-fold more sensitive to Li^+ (half-maximal inhibition at about 0.25 mM) than are the other phosphatases (half-maximal inhibition at about 2.5 mM). Ins(1,4,5) P_3 5-phosphatase activity is potently inhibited by 2,3-bisphosphoglycerate (half-maximal inhibition at 3 μM). Furthermore, 2,3-bisphosphoglycerate also inhibits dephosphorylation of Ins(4,5) P_2 . These characteristics point to a number of similarities between *Dictyostelium* phospho-inositol phosphatases and those from higher organisms. The presence of an hitherto undescribed Ins(1,4,5) P_3 1-phosphatase, however, causes the formation of a different inositol bisphosphatase isomer [Ins(4,5) P_2] from that found in higher organisms [Ins(1,4) P_2]. The high sensitivity of some of these phosphatases for Li^+ suggests that they may be the targets for Li^+ during the alteration of cell pattern by Li^+ in *Dictyostelium*.

INTRODUCTION

The pivotal role of Ins(1,4,5) P_3 as the second messenger for receptor-mediated Ca^{2+} mobilization has been firmly established in a wide variety of systems (reviews: Downes & Michell, 1985; Berridge, 1987). In the best studied mammalian systems, such as human erythrocytes, platelets, rat brain, liver, pancreas and parotid gland, the Ins(1,4,5) P_3 response is attenuated by a specific phosphatase which removes the phosphate from the 5-position to yield Ins(1,4) P_2 (Downes *et al.*, 1982; Storey *et al.*, 1984; Connolly *et al.*, 1985; Erneux *et al.*, 1986; Shears *et al.*, 1987). Ins(1,4) P_2 is then further dephosphorylated to Ins4P in rat liver and brain and calf brain (Delvaux *et al.*, 1987a; Ackermann *et al.*, 1987; Inhorn *et al.*, 1987; Ragan *et al.*, 1988), and finally to Ins. The Ins formed in this way can then be re-used for the synthesis of inositol phospholipids, thus closing the cyclic metabolic pathway characteristic for this signalling system.

Dephosphorylation of Ins(1,4) P_2 and InsP has been shown to be sensitive to Li^+ ions (Hallcher & Sherman, 1980; Storey *et al.*, 1984; Takimoto *et al.*, 1985; Delvaux *et al.*, 1987a,b; Gee *et al.*, 1988), and it has been suggested that the pharmacological effect of Li^+ as a drug

against manic-depressive illness might be due to inhibition of these enzymes by Li^+ (Drummond, 1987).

In the cellular slime mould *Dictyostelium discoideum*, which is frequently used as a model for studying signal transduction and differentiation, a similar second-messenger function has been proposed for Ins(1,4,5) P_3 ; Ins(1,4,5) P_3 can elicit Ca^{2+} release from non-mitochondrial Ca^{2+} stores in saponin-permeabilized *Dictyostelium* cells (Europe-Finner & Newell, 1986), and more recently it was shown that the chemoattractant cyclic AMP can trigger the accumulation of intracellular Ins P_3 *in vivo* (Europe-Finner & Newell, 1987). Furthermore, Li^+ ions can alter the pattern in *Dictyostelium* slugs and direct differentiation to the stalk pathway (Maeda, 1970; Sakai, 1973; Van Lookeren Campagne *et al.*, 1988).

Very little is known about the enzymes involved in the turnover of inositol phospholipids and inositol phosphates in *D. discoideum*. The only enzymes that have been described are the CDP-diacylglycerol: inositol phosphatidyltransferase and the Mn^{2+} -catalysed phosphatidylinositol:*myo*-inositol exchange activity (Machon *et al.*, 1980), a kinase which phosphorylates phosphatidylinositol to phosphatidylinositol 4-phosphate (Varela *et al.*, 1987) and a kinase which phos-

Abbreviations used: Ins, *myo*-inositol; Ins1P, D-*myo*-inositol 1-phosphate; L-Ins1P, L-*myo*-inositol 1-phosphate; Ins4P, D-*myo*-inositol 4-phosphate; Ins(1,4) P_2 , D-*myo*-inositol 1,4-bisphosphate; Ins(4,5) P_2 , D-*myo*-inositol 4,5-bisphosphate; Ins(1,5) P_2 , D-*myo*-inositol 1,5-bisphosphate; Ins(1,4,5) P_3 , D-*myo*-inositol 1,4,5-trisphosphate; InsP, Ins P_2 and Ins P_3 , D-*myo*-inositol phosphates without specification of the phosphate position(s).

† Present address: Department of Anatomy and Cell Biology, College of Physicians and Surgeons, Columbia University, 630 West 168th Street, New York, NY 10032, U.S.A.

phorylates diacylglycerol to phosphatidic acid (Jimenez *et al.*, 1988). Phosphatidylinositol 4-phosphate kinase is also present in *Dictyostelium* (M. M. Van Lookeren Campagne, unpublished work), but no phospholipase C activity has been demonstrated up to now (Irvine *et al.*, 1980). It was not known whether *Dictyostelium* cells have enzymes which dephosphorylate $\text{Ins}(1,4,5)\text{P}_3$. Here we report that $\text{Ins}(1,4,5)\text{P}_3$ can be dephosphorylated to Ins in *D. discoideum* homogenates. Furthermore, we show that this dephosphorylation can occur by two different routes, with as intermediates either $\text{Ins}(1,4)\text{P}_2$ or $\text{Ins}(4,5)\text{P}_2$, which are both dephosphorylated, through $\text{Ins}4\text{P}$, to Ins .

MATERIALS AND METHODS

Materials

$[2\text{-}^3\text{H}]\text{Ins}(1,4,5)\text{P}_3$ (1.0 Ci/mmol) and $\text{L-}[U\text{-}^{14}\text{C}]\text{Ins}1\text{P}$ (55 mCi/mmol) were from Amersham International. $[2\text{-}^3\text{H}]\text{Ins}(1,4)\text{P}_2$ (2.0 Ci/mmol), $[4,5\text{-}^{32}\text{P}]\text{Ins}(1,4,5)\text{P}_3$ (130 Ci/mmol), $[2\text{-}^3\text{H}]\text{Ins}1\text{P}$ (5.4 Ci/mmol) and $[2\text{-}^3\text{H}]\text{Ins}4\text{P}$ (1.5 Ci/mmol) were from New England Nuclear. Dowex 1 (200–400 mesh) and 2,3-bisphosphoglycerate were from Sigma. The h.p.l.c. columns were from Waters ($\mu\text{Bondapak NH}_2$; 30 cm \times 0.39 cm), Whatman (Partisil SAX; 25 cm \times 0.49 cm) and Chrompack (LiChrosorb 10RP18; 25 cm \times 0.49 cm).

Organism and culture conditions

Dictyostelium discoideum strain NC-4(H) was grown in association with *Escherichia coli* 281 on glucose/peptone agar as described previously (Van Lookeren Campagne *et al.*, 1986). Amoebae were harvested in 10 mM-phosphate buffer, pH 6.5, and freed from bacteria by repeated centrifugation. Cells were then plated on non-nutrient agar plates at a density of 10^7 cells/cm² and incubated overnight at 6 °C to induce full aggregation-competence (Konijn, 1970).

Homogenate

Aggregation-competent cells were harvested in 10 mM-phosphate buffer, pH 6.5, washed once in ice-cold buffer A (20 mM-Hepes/NaOH, 0.5 mM-EDTA, 200 mM-sucrose, pH 7.0), and resuspended to 2×10^9 cells/ml in the same buffer. Homogenates were made by passing the cells through a Nucleopore filter (3 μm pore size) (Das & Henderson, 1983). Lysates were then centrifuged for 3 min at 10000 g. The particulate cell fraction was prepared by washing the pellet once in buffer A and resuspending it in the same buffer to the original volume of the homogenate. The soluble fraction was prepared by re-centrifuging the 10000 g supernatant for 5 min in a Beckman Airfuge at 150000 g. Soluble and particulate fractions thus obtained were immediately used for the phosphatase assay.

Phosphatase assay

Dephosphorylation of $[2\text{-}^3\text{H}]\text{Ins}(1,4,5)\text{P}_3$, $[4,5\text{-}^{32}\text{P}]\text{Ins}(1,4,5)\text{P}_3$, $[2\text{-}^3\text{H}]\text{Ins}(1,4)\text{P}_2$, $[2\text{-}^3\text{H}]\text{Ins}(4,5)\text{P}_2$, $[4,5\text{-}^{32}\text{P}]\text{Ins}(4,5)\text{P}_2$ and/or $\text{L-}[U\text{-}^{14}\text{C}]\text{Ins}1\text{P}$ was assayed in buffer A, in the presence of either 5 mM-MgCl₂ or 2.5 mM-CaCl₂,

at 22 °C. Incubations were started by adding 5 μl of either the soluble or particulate fraction of the *Dictyostelium* homogenate to 15 μl assay mixture, containing 1000–3000 c.p.m. of radiolabelled substrate. Reactions were stopped after 5–30 min by adding 0.5 ml of chloroform/methanol/conc. HCl (20:40:1, by vol.). Phases were separated by adding 200 μl of water. After vigorous shaking and centrifugation (1 min, 10000 g), the aqueous phase was applied to 0.5 ml Dowex-1 anion-exchange columns (formate form). The different reaction products were separated by stepwise elution with: (1) 10 ml of water (Ins); (2) 10 ml of 150 mM-ammonium formate/5 mM-Na₂B₄O₇ (InsP and P_i); (3) 10 ml of 300 mM-ammonium formate/100 mM-formic acid (InsP_2); and (4) 10 ml of 750 mM-ammonium formate/100 mM-formic acid (InsP_3). Radioactivity of the fractions was measured by liquid-scintillation counting after adding 13 ml of Instagel (Packard).

When it was necessary to separate $\text{Ins}4\text{P}$ from $\text{Ins}1\text{P}$ and P_i reaction products (usually with the internal standard of $\text{L-}[U\text{-}^{14}\text{C}]\text{Ins}1\text{P}$) were separated by anion-exchange h.p.l.c. as described in the Figure legends. $\text{Ins}(1,4,5)\text{P}_3$ 5-phosphatase activity from human erythrocyte membranes was assayed as described previously (Erneux *et al.*, 1986). Enzyme activities were approximately linear with time and enzyme concentrations, provided that not more than about 25 % of the substrate was utilized. The s.d. of the phosphatase assay was about 10 %. Experiments were performed at least three times with similar results; the analysis of the InsP isomers by h.p.l.c. was performed twice with identical results.

Preparation and purification of $\text{Ins}(4,5)\text{P}_2$

Aggregation-competent *Dictyostelium* cells were lysed in buffer B [50 mM-Tris/HCl, pH 7.2, 10 % (v/v) glycerol, 10 mM-dithiothreitol, leupeptin (6.5 $\mu\text{g}/\text{ml}$), 100 μM -phenylmethanesulphonyl fluoride, soya-bean trypsin inhibitor (50 $\mu\text{g}/\text{ml}$) and 5 mM-benzamidine]. The high-speed supernatant from 3×10^9 cells was chromatographed on a DEAE-cellulose column (10 ml; 8 cm \times 1.3 cm), which was equilibrated and eluted in buffer B. The $\text{Ins}(1,4,5)\text{P}_3$ 5-phosphatase activity binds to the column, whereas the $\text{Ins}(1,4,5)\text{P}_3$ 1-phosphatase is eluted from the column between 1.5 and 2 column vol. (P. J. M. Van Haastert & E. Rovers, unpublished work).

A mixture of ³²P- and ³H-labelled $\text{Ins}(4,5)\text{P}_2$ was prepared in an incubation (100 μl) containing 50 nCi of $[4,5\text{-}^{32}\text{P}]\text{Ins}(1,4,5)\text{P}_3$, 100 nCi of $[2\text{-}^3\text{H}]\text{Ins}(1,4,5)\text{P}_3$, 5 mM-MgCl₂, buffer A and 40 μl of enzyme from the DEAE-cellulose column. After 60 min the incubation was terminated by the addition of 100 μl of 0.1 M-tributylammonium phosphate, pH 6.5. The sample was centrifuged immediately for 5 min at 10000 g, and the supernatant was chromatographed by h.p.l.c. on a reversed-phase LiChrosorb 10RP18 column, which was eluted isocratically with 1.5 mM-tributylammonium phosphate / 4.5 mM-triethylammonium formate / 16 % (v/v) methanol, pH 6.5, at a flow rate of 1.2 ml/min. Fractions of volume 0.6 ml were collected; the radioactivity of 6 μl samples was determined by using a dual-label program. Peak fractions were combined and concentrated to dryness under reduced pressure at 10 °C. $[^3\text{H}]\text{Ins}(4,5)\text{P}_2$ was prepared in parallel from 100 nCi of $[^3\text{H}]\text{Ins}(1,4,5)\text{P}_3$.

Table 1. Relative Ins(1,4,5) P_3 phosphatase activity in soluble and particulate cell fractions measured under different conditions

Soluble or particulate cell fractions, derived from 5×10^7 cells/ml in the assay, were incubated with 5 nCi of [2- ^3H]Ins(1,4,5) P_3 (0.25 μM) in the presence of either MgCl_2 or CaCl_2 and the phosphatase inhibitors LiCl and 2,3-bisphosphoglycerate as indicated. The phosphatase activity data are expressed as percentages (not standardized for protein) relative to the value measured in the soluble fraction with MgCl_2 . When standardized for protein content, phosphatase activity assayed with 5 mM- MgCl_2 in soluble and particulate fractions was 9.8 and 9.2 pmol/min per mg of protein respectively.

Assay with:	Activity (%)		
	No addition	25 mM-LiCl	0.25 mM-2,3-Bisphosphoglycerate
Soluble			
5 mM- MgCl_2	100	21	82
2.5 mM- CaCl_2	19	17	1
Particulate			
5 mM- MgCl_2	21	11	13
2.5 mM- CaCl_2	11	10	1

RESULTS

General properties of Ins(1,4,5) P_3 dephosphorylation in *Dictyostelium*

Ins(1,4,5) P_3 can be rapidly dephosphorylated by a *D. discoideum* homogenate to Ins P_2 . Activity is optimal at pH 7.0 and 5 mM- MgCl_2 (results not shown) and is located predominantly in the soluble cell fraction (Table 1). LiCl and 2,3-bisphosphoglycerate, which are known inhibitors of phospho-inositol phosphatases (Downes *et al.*, 1982; Storey *et al.*, 1984; Delvaux *et al.*, 1987a) have differential potencies in the soluble and particulate fractions, Li $^+$ being a more potent inhibitor of the soluble Ins(1,4,5) P_3 phosphatase activity and 2,3-bisphosphoglycerate being more potent on the particulate activity (Table 1). The particulate enzyme is probably not located on the cell surface, because intact cells express little enzyme activity (results not shown).

CaCl_2 (2.5 mM) can replace MgCl_2 to a certain extent, more so in the particulate fraction (to about 50%) than in the soluble fraction (to about 20%). Furthermore, replacement of Ca^{2+} for Mg^{2+} changes the sensitivity towards Li $^+$ and 2,3-bisphosphoglycerate, Li $^+$ becoming ineffective and 2,3-bisphosphoglycerate almost completely inhibiting all phosphatase activity (Table 1).

Identification of the first phosphate group removed by Ins(1,4,5) P_3 phosphatase

The studies with the Ins(1,4,5) P_3 phosphatase inhibitors Li $^+$ and 2,3-bisphosphoglycerate suggest that in 5 mM- Mg^{2+} Ins(1,4,5) P_3 is dephosphorylated by a route different from that in 2.5 mM- Ca^{2+} . To see which phosphate group is removed first under the different conditions, we measured the hydrolysis of a mixture of [4,5- ^{32}P]Ins(1,4,5) P_3 and [2- ^3H]Ins(1,4,5) P_3 . The distribution of ^{32}P label between the 4- and 5-phosphates of

Table 2. Ratio between ^{32}P and ^3H radioactivity in Ins P_2 produced by hydrolysis of a mixture of [4,5- ^{32}P]Ins(1,4,5) P_3 and [2- ^3H]Ins(1,4,5) P_3

Ins P_3 phosphatase activity was assayed in the soluble fraction from 5×10^7 cells/ml with 0.5 nCi of [4,5- ^{32}P]Ins(1,4,5) P_3 and 2.5 nCi of [2- ^3H]Ins(1,4,5) P_3 under the conditions indicated and described in the Materials and methods section. Incubations were terminated after 10 min when assayed in MgCl_2 or after 20 min when assayed in CaCl_2 and chromatographed on Dowex columns. The ratio between the ^{32}P and ^3H radioactivity in the Ins P_2 fractions was determined by liquid-scintillation counting, with a dual-label program. The $^{32}\text{P}/^3\text{H}$ ratios in Ins P_2 relative to that in Ins(1,4,5) P_3 are given in parentheses. The incubation with human erythrocyte membranes, which contains only 5-phosphatase, revealed that the ^{32}P label was distributed over the 4- and 5-positions in the ratio 12:88. A low $^{32}\text{P}:^3\text{H}$ ratio in Ins P_2 therefore indicates 5-phosphatase activity, whereas a high ratio indicates 1- and/or 4-phosphatase activity.

	$^{32}\text{P}/^3\text{H}$ ratio
Ins(1,4,5) P_3 substrate	0.74 (1.00)
Ins P_2 formed in 5 mM- MgCl_2	
No additions	0.61 (0.82)
25 mM-LiCl	0.12 (0.16)
0.25 mM-2,3-bisphosphoglycerate	0.68 (0.92)
Ins P_2 formed in 2.5 mM- CaCl_2	
No additions	0.11 (0.15)
Ins P_2 formed by 5-phosphatase from human erythrocyte membranes	0.09 (0.12)

the commercial [4,5- ^{32}P]Ins(1,4,5) P_3 was 12% at the 4- and 88% at the 5-position (as determined with specific 5-phosphatase in human erythrocyte membranes; Downes *et al.*, 1982). After incubation of this [4,5- ^{32}P]Ins(1,4,5) P_3 and [2- ^3H]Ins(1,4,5) P_3 mixture with the *D. discoideum* soluble cell fraction, and subsequent fractionation of the inositol phosphates formed on Dowex columns, the ratio of ^{32}P label recovered with respect to the ^3H label in the Ins P_2 column fraction was determined. As the [4,5- ^{32}P]Ins(1,4,5) P_3 is labelled predominantly in the 5-position, a low $^{32}\text{P}:^3\text{H}$ ratio in Ins P_2 indicates 5-phosphatase activity, whereas a high $^{32}\text{P}:^3\text{H}$ ratio in Ins P_2 indicates 1- and/or 4-phosphatase activity.

Table 2 shows that the $^{32}\text{P}:^3\text{H}$ ratio in Ins P_2 is 82% of the original value in the Ins(1,4,5) P_3 substrate, when dephosphorylation is assayed in 5 mM- Mg^{2+} . In the presence of 2,3-bisphosphoglycerate the ratio even approaches the original ratio in Ins(1,4,5) P_3 (92%). This suggests that, in 5 mM- Mg^{2+} , Ins(1,4,5) P_3 is dephosphorylated predominantly at the 1- and/or the 4-position, thus yielding Ins(1,5) P_2 and/or Ins(4,5) P_2 . In contrast, the Li $^+$ -insensitive part of the activity in Mg^{2+} dephosphorylates Ins(1,4,5) P_3 at the 5-position, as the $^{32}\text{P}:^3\text{H}$ ratio is similar to that found for the 5-phosphatase from erythrocyte membrane (Table 2). The same is the case for the activity measured in Ca^{2+} .

From this we conclude that in the soluble fraction of *Dictyostelium* homogenates, assayed for Ins(1,4,5) P_3 phosphatase in Mg^{2+} , about 80% of the activity is accounted for by either a 1- and/or a 4-phosphatase, which is sensitive to Li $^+$, and about 20% of the activity

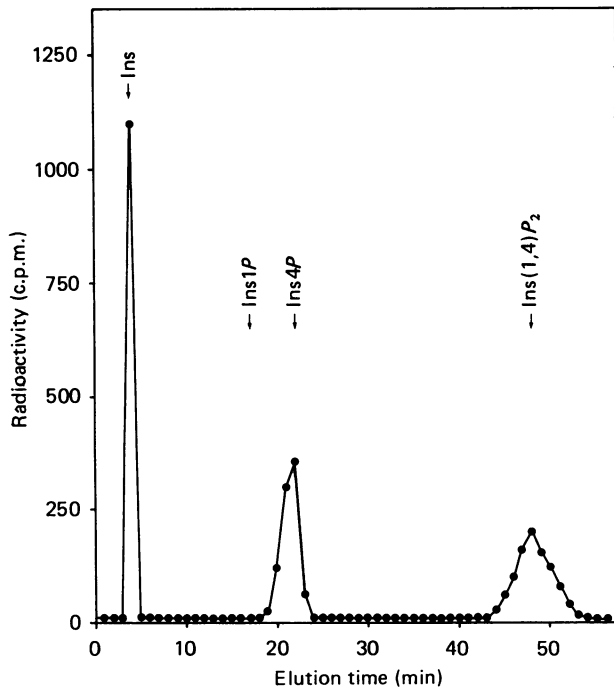


Fig. 1. H.p.l.c. analysis of the products of $\text{Ins}(1,4)\text{P}_2$ dephosphorylation

$[2\text{-}^3\text{H}]\text{Ins}(1,4)\text{P}_2$ (10 nCi; $0.25\ \mu\text{M}$) was dephosphorylated by incubation with *Dictyostelium* soluble cell fraction for 10 min, as described in the Materials and methods section in the presence of $5\ \text{mM-MgCl}_2$. After filtration on a Centricon TM microconcentrator, a sample was loaded on a $\mu\text{Bondapak NH}_2$ column. Separation was carried out by isocratic elution for 10 min with $20\ \text{mM-ammonium acetate/acetic acid}$, pH 4.0, followed by a 60 min linear gradient to $1\ \text{M-ammonium acetate/acetic acid}$, pH 4.0. Fractions were collected every 1 min (flow rate $1\ \text{ml/min}$), and radioactivity was determined by liquid-scintillation spectroscopy. The arrows indicate the elution times of commercial $[2\text{-}^3\text{H}]\text{Ins}$, commercial $[2\text{-}^3\text{H}]\text{Ins1P}$ and $[2\text{-}^3\text{H}]\text{Ins4P}$ (prepared by dephosphorylation of $[2\text{-}^3\text{H}]\text{Ins}(1,4)\text{P}_2$ with rat brain soluble fraction; Delvaux *et al.*, 1987b).

by a 5-phosphatase, which is sensitive to 2,3-bis-phosphoglycerate. Ca^{2+} can only replace Mg^{2+} for the 5-phosphatase. In the particulate fraction the 5-phosphatase and 1- and/or 4-phosphatase are about equally active.

Analysis of $^{32}\text{P}:^3\text{H}$ ratios does not give a clear-cut answer about whether the Li^+ -sensitive $\text{Ins}(1,4,5)\text{P}_3$ phosphatase activity dephosphorylates $\text{Ins}(1,4,5)\text{P}_3$ at the 1- or at the 4-position, as the fraction of ^{32}P label in the 4-position is too small for accurate analysis. As we do know that the InsP_2 thus formed still contains the 5-phosphate, the two possible isomers of this InsP_2 are $\text{Ins}(4,5)\text{P}_2$ or $\text{Ins}(1,5)\text{P}_2$, and it is therefore termed $\text{Ins}(x,5)\text{P}_2$.

Dephosphorylation of InsP_2

The two distinct pathways for the first step in $\text{Ins}(1,4,5)\text{P}_3$ dephosphorylation, measured in $5\ \text{mM-MgCl}_2$, have been shown above to yield $\text{Ins}(x,5)\text{P}_2$ (80%) and $\text{Ins}(1,4)\text{P}_2$ (20%). We used $[2\text{-}^3\text{H}]\text{Ins}(1,4)\text{P}_2$ as a

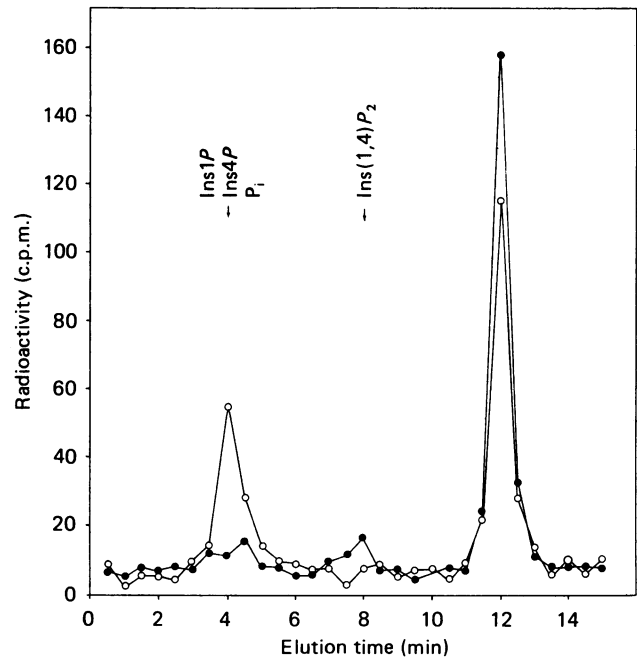


Fig. 2. Purification by h.p.l.c. of a $^{32}\text{P}/^3\text{H}$ -labelled mixture of $\text{Ins}(x,5)\text{P}_2$ synthesized from $[4,5\text{-}^{32}\text{P}]\text{Ins}(1,4,5)\text{P}_3$ and $[2\text{-}^3\text{H}]\text{Ins}(1,4,5)\text{P}_3$ with a partially purified phosphatase

A mixture of $[4,5\text{-}^{32}\text{P}]\text{Ins}(1,4,5)\text{P}_3$ and $[^3\text{H}]\text{Ins}(1,4,5)\text{P}_3$ was incubated with a partially purified phosphatase from *Dictyostelium* as described in the Materials and methods section. The reaction mixture was chromatographed on a LiChrosorb reversed-phase column, and 0.6 ml fractions were collected; the radioactivity in $6\ \mu\text{l}$ was determined with a dual-label program. The three fractions eluted around 12 min were combined. The $^{32}\text{P}/^3\text{H}$ ratio in the substrate $\text{Ins}(1,4,5)\text{P}_3$ was 0.60, and that in the product $\text{Ins}(x,5)\text{P}_2$ was 0.61. The elution of commercial standards is indicated by the arrows; $\text{Ins}(1,4,5)\text{P}_3$ is eluted later than 15 min. \circ , ^{32}P radioactivity; \bullet , ^3H radioactivity.

substrate to characterize the second dephosphorylation step of the latter pathway. H.p.l.c. analysis of the products formed after incubating $\text{Ins}(1,4)\text{P}_2$ with the *Dictyostelium* soluble cell fraction shows that the InsP formed from $\text{Ins}(1,4)\text{P}_2$ is exclusively Ins4P (Fig. 1).

To study the dephosphorylation of $\text{Ins}(x,5)\text{P}_2$ to InsP , we synthesized and purified a $^{32}\text{P}/^3\text{H}$ -labelled mixture of $\text{Ins}(x,5)\text{P}_2$ from $[4,5\text{-}^{32}\text{P}]\text{Ins}(1,4,5)\text{P}_3$ and $[2\text{-}^3\text{H}]\text{Ins}(1,4,5)\text{P}_3$ (see the Materials and methods section and Fig. 2). The $^{32}\text{P}/^3\text{H}$ ratio of the $\text{Ins}(1,4,5)\text{P}_3$ substrate was 0.60; the ratio in the $\text{Ins}(x,5)\text{P}_2$ product was 0.61. This purified $^{32}\text{P}/^3\text{H}$ mixture of $\text{Ins}(x,5)\text{P}_2$ was incubated with *Dictyostelium* soluble cell fraction, and the dephosphorylation products were subsequently analysed by h.p.l.c. (Fig. 3). The InsP formed is co-eluted with Ins4P , is not co-eluted with Ins1P , and has a low $^{32}\text{P}/^3\text{H}$ ratio (0.05) compared with the synthesized $\text{Ins}(x,5)\text{P}_2$ substrate (0.61), indicating that the 5-phosphate has been removed. From this we infer that the InsP formed is Ins4P , and thus the synthesized $\text{Ins}(x,5)\text{P}_2$ must have been $\text{Ins}(4,5)\text{P}_2$ [and not $\text{Ins}(1,5)\text{P}_2$]. In summary, we can conclude that $\text{Ins}(1,4,5)\text{P}_3$ is dephosphorylated by *Dictyostelium* soluble cell fraction, through $\text{Ins}(1,4)\text{P}_2$ and predominantly $\text{Ins}(4,5)\text{P}_2$, to Ins4P .

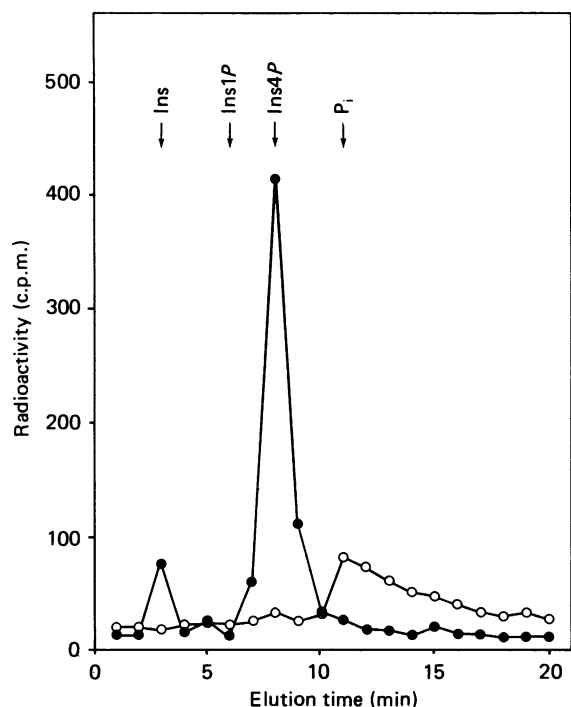


Fig. 3. H.p.l.c. analysis of the InsP isomer(s) formed after dephosphorylation of a $^{32}\text{P}/^3\text{H}$ -labelled mixture of $\text{Ins}(x,5)\text{P}_2$ synthesized from $[4,5\text{-}^{32}\text{P}]\text{Ins}(1,4,5)\text{P}_3$ and $[2\text{-}^3\text{H}]\text{Ins}(1,4,5)\text{P}_3$

H.p.l.c.-purified $[^{32}\text{P}, ^3\text{H}]\text{Ins}(x,5)\text{P}_2$ (see Fig. 2) was incubated with *Dictyostelium* soluble cell fraction for 60 min in the presence of 5 mM- MgCl_2 . The incubation was terminated by adding 0.5 ml of ice-cold 150 mM-ammonium acetate/acetic acid (pH 4.0) and immediate filtration on a Centricon TM microconcentrator. The reaction products were separated on a Partisil SAX column by isocratic elution with 150 mM-ammonium acetate/acetic acid, pH 4.0. Fractions were collected every 1 min (flow rate 1.5 ml/min), and radioactivity was determined by liquid-scintillation spectrometry. The $^{32}\text{P}/^3\text{H}$ ratio in the InsP peak is low (0.05, relative to 0.61 in substrate), indicating that it does not contain a 5-phosphate group. Co-elution with authentic Ins4P and absence of a 5-phosphate in the InsP identifies the product as Ins4P , and the $\text{Ins}(x,5)\text{P}_2$ substrate as $\text{Ins}(4,5)\text{P}_2$. \circ , ^{32}P radioactivity; \bullet , ^3H radioactivity.

Inhibition of $\text{Ins}(1,4,5)\text{P}_3$ phosphatase by Li^+

As shown in Table 1, the inhibitor-sensitivity of $\text{Ins}(1,4,5)\text{P}_3$ dephosphorylation is rather complex. We have therefore studied the sensitivity of the different dephosphorylation reactions in more detail. In mammalian tissues, Li^+ does not affect the 5-phosphatase activity acting on $\text{Ins}(1,4,5)\text{P}_3$ (Downes *et al.*, 1982; Storey *et al.*, 1984; Connolly *et al.*, 1985; Erneux *et al.*, 1986), but is a potent uncompetitive inhibitor of 1- and 4-phosphatase activities acting on $\text{Ins}(1,4)\text{P}_2$, Ins4P and Ins1P (Hallcher & Sherman, 1980; Inhorn & Majerus, 1987; Gee *et al.*, 1988). As shown in Fig. 4, Li^+ effectively inhibits $\text{Ins}(1,4,5)\text{P}_3$ phosphatase activity to a maximum of about 80%, if measured in 5 mM- Mg^{2+} with 0.25 μM - $\text{Ins}(1,4,5)\text{P}_3$. Half-maximal inhibition of this activity occurs at about 0.25 mM- LiCl . When phosphatase activity was measured in Ca^{2+} , Li^+ had no effect (Fig. 4).

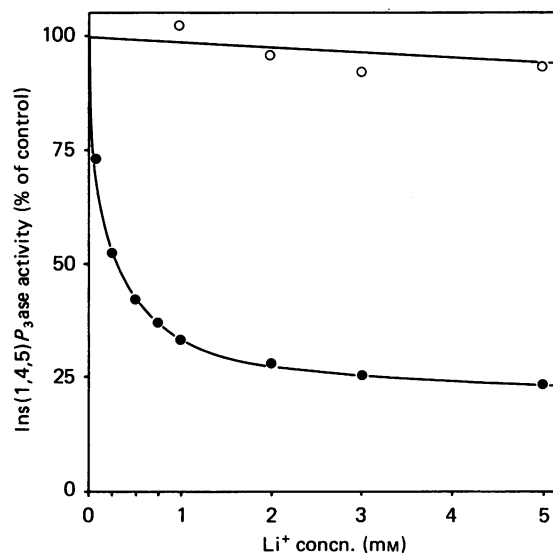


Fig. 4. Effect of Li^+ on $\text{Ins}(1,4,5)\text{P}_3$ phosphatase activity in the soluble cell fraction of *Dictyostelium* homogenates

Dephosphorylation of $[2\text{-}^3\text{H}]\text{Ins}(1,4,5)\text{P}_3$ (5 nCi; 0.25 μM) was assayed in the presence of different LiCl concentrations as described in the Materials and methods section, with either 5 mM- MgCl_2 (\bullet) or 2.5 mM- CaCl_2 (\circ). The phosphatase activity data are expressed as percentages of the control without LiCl .

Inhibition of $\text{Ins}(1,4,5)\text{P}_3$ phosphatase by 2,3-bisphosphoglycerate

2,3-Bisphosphoglycerate is a potent competitive inhibitor of 5-phosphatase activities from erythrocyte membranes (Downes *et al.*, 1982) and soluble and particulate rat brain fraction (Delvaux *et al.*, 1987a). Furthermore, concentrations of up to 1 mM have no effect on $\text{Ins}(1,4)\text{P}_2$ and Ins1P phosphatases (Delvaux *et al.*, 1987a).

In the *Dictyostelium* soluble cell fraction 2,3-bisphosphoglycerate inhibits the $\text{Ins}(1,4,5)\text{P}_3$ phosphatase activity biphasically, when measured with 0.25 mM- $\text{Ins}(1,4,5)\text{P}_3$ in 5 mM- Mg^{2+} (Fig. 5). Under these conditions, about 20% of the activity is inhibited with high sensitivity, whereas the remaining 80% is inhibited only at high concentrations (above 0.5 mM) of 2,3-bisphosphoglycerate. When measured in Ca^{2+} , however, all the activity can be inhibited with high sensitivity, half-maximal inhibition occurring at about 3 μM (Fig. 5).

The combination of the effects of Li^+ and 2,3-bisphosphoglycerate on the *Dictyostelium* $\text{Ins}(1,4,5)\text{P}_3$ phosphatase activity (assuming that the inhibitor-sensitivities of the *Dictyostelium* and rat brain enzymes are similar) supplements the evidence presented above that in Mg^{2+} $\text{Ins}(1,4,5)\text{P}_3$ is dephosphorylated by two enzymes: 20% of the activity is due to a 5-phosphatase, as this 20% of the activity is insensitive to Li^+ and highly sensitive to 2,3-bisphosphoglycerate, and 80% of the activity is due to a 1-phosphatase, as this activity is less sensitive to 2,3-bisphosphoglycerate and highly sensitive to Li^+ . Ca^{2+} apparently can only replace Mg^{2+} in that part of the activity which is sensitive to 2,3-bisphosphoglycerate, so in Ca^{2+} all activity is apparently due to a 5-phosphatase which has some similarities to the 5-phosphatase from higher organisms.

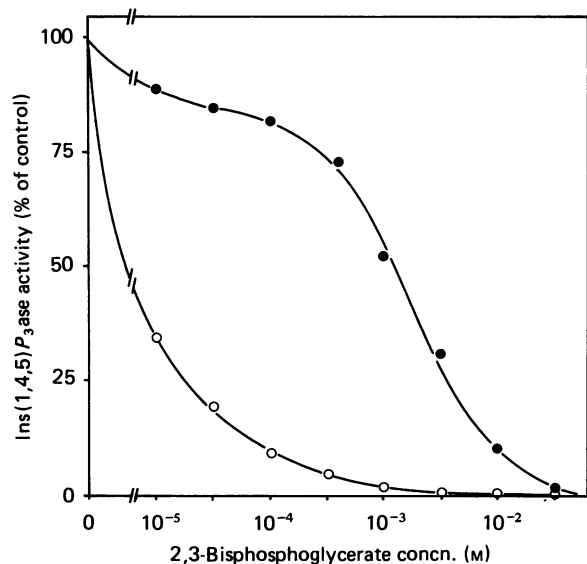


Fig. 5. Effect of 2,3-bisphosphoglycerate on Ins(1,4,5) P_3 phosphatase activity in the soluble cell fraction of *Dictyostelium homogenates*

Dephosphorylation of [$2\text{-}^3\text{H}$]Ins(1,4,5) P_3 (5 nCi; 0.25 μM) was assayed in the presence of different 2,3-bisphosphoglycerate concentrations as described in the Materials and methods section, with either 5 mM-MgCl $_2$ (●) or 2.5 mM-CaCl $_2$ (○). The phosphatase activity data are expressed as percentages of the control without 2,3-bisphosphoglycerate.

Sensitivity of Ins P_2 dephosphorylation to Li $^+$ and 2,3-bisphosphoglycerate

Dephosphorylation of Ins(1,4) P_2 to Ins4P has been shown to be Li $^+$ -sensitive and 2,3-bisphosphoglycerate-insensitive in rat brain (Inhorn *et al.*, 1987; Delvaux *et al.*, 1987a). The same is the case for [$2\text{-}^3\text{H}$]Ins(1,4) P_2 dephosphorylation in *Dictyostelium*; Li $^+$ inhibits the 1-phosphatase activity with half-maximal inhibition at about 2.5 mM (Fig. 6), and 0.25 mM-2,3-bisphosphoglycerate has no effect on this activity (results not shown).

To measure the sensitivities of Ins(4,5) P_2 dephosphorylation to Li $^+$ and 2,3-bisphosphoglycerate, we synthesized and purified [$2\text{-}^3\text{H}$]Ins(4,5) P_2 from [$2\text{-}^3\text{H}$]Ins(1,4,5) P_3 (see the Materials and methods section) and incubated the compound with the high-speed supernatant from *Dictyostelium*: 25 mM-LiCl inhibits the dephosphorylation of [$2\text{-}^3\text{H}$]Ins(4,5) P_2 by only 7%, whereas 0.25 mM-2,3-bisphosphoglycerate inhibits the dephosphorylation by 76% (results not shown).

Li $^+$ -sensitivity of Ins4P and L-Ins1P dephosphorylation

myo-Inositol monophosphates can be derived from two different sources: (1) L-Ins1P formed from isomerization of D-glucose 6-phosphate catalysed by L-*myo*-inositol 1-phosphate synthase, which is required for Ins synthesis *de novo*, and (2) InsP in the D-conformation, formed through the action of phospholipase C on phosphatidylinositol, or through dephosphorylation of the D-*myo*-inositol polyphosphates. In high organisms, InsP is dephosphorylated by a phosphatase which is not very specific; the enzyme can hydrolyse all InsP isomers with an equatorial phosphate group, as well as 2'-AMP

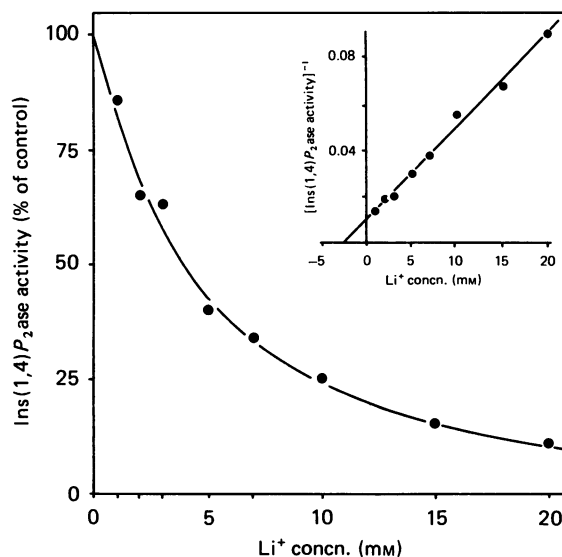


Fig. 6. Effect of Li $^+$ on Ins(1,4) P_2 1-phosphatase activity in the soluble cell fraction of *Dictyostelium homogenates*

Dephosphorylation of [$2\text{-}^3\text{H}$]Ins(1,4) P_2 (5 nCi; 0.125 μM) was assayed in the presence of different LiCl concentrations as described in the Materials and methods section. The phosphatase activity data are expressed as percentages of the control without LiCl. The inset shows a Dixon plot of the same data.

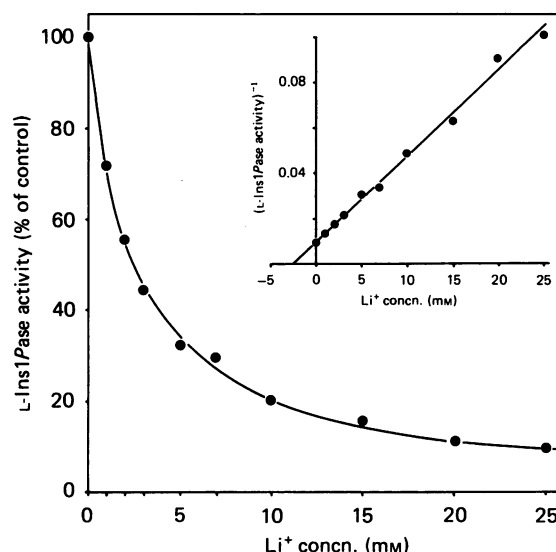


Fig. 7. Effect of Li $^+$ on L-Ins1P phosphatase activity in the soluble cell fraction of *Dictyostelium homogenates*

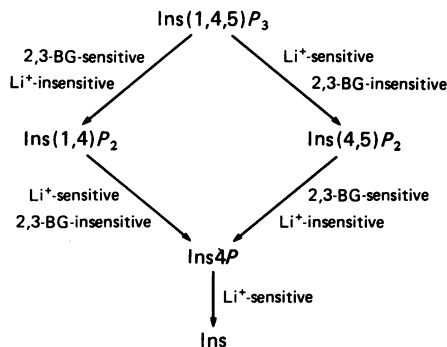
Dephosphorylation of L-[U- ^{14}C]Ins1P (6.3 nCi; 5.7 μM) was assayed as described for Ins P_2 phosphatase in Fig. 6.

and (–)-*chiro*-Ins(3)P [but not Ins(1,4,5) P_3] and does not discriminate between the two enantiomeric conformations of Ins1P (Eisenberg, 1967; Hallcher & Sherman, 1980; Ackermann *et al.*, 1987). Furthermore, Li $^+$ is a potent inhibitor of InsP phosphatase activities (Naccarato *et al.*, 1974; Hallcher & Sherman, 1980; Sherman *et al.*, 1984; Ackermann *et al.*, 1987; Delvaux *et al.*, 1987b; Gee *et al.*, 1988).

In *Dictyostelium* homogenates, L-Ins1P and Ins4P are dephosphorylated under conditions similar to those described for the above systems. Li⁺ inhibits the dephosphorylation of L-[U-¹⁴C]Ins1P in the soluble fraction of these homogenates, with half-maximal inhibition at about 2.5 mM (Fig 7). Furthermore, when [2-³H]Ins(1,4)P₂ is incubated with this cell fraction until almost no substrate is left, subsequent addition of Li⁺ inhibits the further dephosphorylation of the Ins4P thus formed: by 51% with 4 mM-LiCl and by 74% with 20 mM-LiCl.

DISCUSSION

Our results show that *Dictyostelium discoideum* homogenates possess phosphatases which can rapidly dephosphorylate Ins(1,4,5)P₃. The two presumptive dephosphorylation pathways can be summarized by Scheme 1.



Scheme 1.

Abbreviation: 2,3-BG, 2,3-bisphosphoglycerate.

Although at first sight the phosphatase activities appear to be very different from those in higher organisms, on closer inspection many similarities can be found. *Dictyostelium* contains an Ins(1,4,5)P₃ 5-phosphatase activity which is Mg²⁺-dependent, sensitive to 2,3-bisphosphoglycerate, insensitive to Li⁺ and present in both particulate and soluble cell fractions. This is similar to the 5-phosphatase from rat liver and brain (Erneux *et al.*, 1986; Shears *et al.*, 1987), which also contains both soluble and particulate 5-phosphatase activities. Further dephosphorylation of Ins(1,4)P₂ by the *Dictyostelium* soluble cell fraction is also similar to the rat or bovine brain systems; Ins(1,4)P₂ is dephosphorylated in both systems by a 1-phosphatase to form Ins4P, which is then dephosphorylated to Ins (Inhorn *et al.*, 1987; Delvaux *et al.*, 1987b; Ragan *et al.*, 1988). Furthermore, the Ins(1,4)P₂ 1-phosphatase and the Ins4P phosphatases of both systems are sensitive to Li⁺, with half-maximal inhibition at about 2.5 mM-LiCl, and are 10–100 times less sensitive to 2,3-bisphosphoglycerate than are their Ins(1,4,5)P₃ 5-phosphatase activities (Delvaux *et al.*, 1987a,b). The *Dictyostelium* 5-phosphatase enzyme differs from these systems in that Ca²⁺ can replace Mg²⁺. In platelets and erythrocytes Ca²⁺ cannot replace Mg²⁺, and Ca²⁺ inhibits the Mg²⁺-activated activity with high affinity (K_i = 70 μM) (Downes *et al.*, 1982; Connolly *et al.*, 1985). We have used Ca²⁺ as a tool to elucidate the dephosphorylation pathway of Ins(1,4,5)P₃ in *Dictyos-*

stelium, but have no indication for its physiological importance.

The major difference between Ins(1,4,5)P₃ dephosphorylation in *Dictyostelium* and that of higher organisms is, however, that in *Dictyostelium* most of the Ins(1,4,5)P₃ phosphatase activity is due to a predominantly soluble 1-phosphatase, which is highly Li⁺-sensitive [10-fold more sensitive than Ins(1,4)P₂ and L-Ins1P 1-phosphatase activities]. This shows that, although many of the *Dictyostelium* Ins(1,4,5)P₃-dephosphorylating enzymes could be very similar to those of higher organisms, the metabolic pathway of Ins(1,4,5)P₃ dephosphorylation is very different, and a new InsP₂ isomer, Ins(4,5)P₂, is formed.

It is now important to investigate which InsP₂ is present *in vivo*. Nothing is known about the identity of any of the isomers of the inositol phosphates that are present *in vivo* in *Dictyostelium*, but this can be investigated by h.p.l.c. The high sensitivity of Ins(1,4,5)P₃ dephosphorylation for Li⁺ *in vitro* makes it interesting to see whether the dramatic effects that Li⁺ has on the determination of cell differentiation and pattern formation *in vivo* (Maeda, 1970; Sakai, 1973; Van Lookeren Campagne *et al.*, 1988) are mediated through interference with the metabolism of the inositol (poly)phosphates.

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