



University of Groningen

Clinical monitoring of 'autoimmune' chronic active hepatitis

Hoek, Bart van

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version Publisher's PDF, also known as Version of record

Publication date: 1989

Link to publication in University of Groningen/UMCG research database

Citation for published version (APA): Hoek, B. V. (1989). Clinical monitoring of 'autoimmune' chronic active hepatitis. s.n.

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: https://www.rug.nl/library/open-access/self-archiving-pure/taverneamendment.

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): http://www.rug.nl/research/portal. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

CLINICAL MONITORING OF 'AUTOIMMUNE' CHRONIC ACTIVE HEPATITIS

A HISTOLOGICAL, BIOCHEMICAL AND IMMUNOLOGICAL STUDY



BART VAN HOEK

CLINICAL MONITORING OF 'AUTOIMMUNE' CHRONIC ACTIVE HEPATITIS

A histological, biochemical and immunological study

STELLINGEN

 Although physicians are intuitively aware of the impact of different chronic conditions on functioning and well-being, they need precise measures of these outcomes that are also practical for use in the office setting... We still know very little about the relative impact of different chronic conditions on patients' functioning and well-being.

(Stewart AL, Greenfield S, Hays R, et al. JAMA 1989; 262: 907-913).

- 2. Het is zeer wel mogelijk dat idiopathische 'autoimmune' CAH een 'final common pathway' is met een uiteenlopende etiologie, zoals wellicht: hepatitis C, Epstein Barr virus, mazelenvirus en 'inborn errors of metabolism'.
- 3. Een klassiek geval van 'piecemeal necrose' en regeneratie van de lever vinden we in de Griekse mythologie bij Prometheus.
- De bepaling van de meeste niet orgaan-specifieke antistoffen heeft bij CAH slechts een beperkte waarde. (dit proefschrift)
- Bepaling van N-terminale procollageen-III-peptide in serum maakt het, in tegenstelling tot vele gangbare methoden, vaak mogelijk de diagnose remissie of actieve CAH te stellen zonder leverbiopsie. (dit proefschrift)
- 6. Het is gewenst om bij het verstrekken van immunosuppressieve therapie aan een patiënt met 'autoimmune' chronisch actieve hepatitis te streven naar: een histologische activiteitsscore (0-18) gelijk aan of lager dan 1, normalisering van Nterminale procollageen-III-peptide in serum, afwezigheid van detecteerbare anti-LSP antistoffen, en normale eiwitsynthese door de lever. (dit proefschrift)
- 7. Indien gepoogd wordt immunosuppressie geleidelijk te stoppen of op een lager niveau te brengen bij een patiënt met 'autoimmune' chronisch actieve hepatitis is het de beste methode dit in korte tijd te doen (bijv. 6-12 weken) opdat frequente (bijv. 2-wekelijkse) controle met meting van anti-LSP, PIIINP en ALT (met een individuele referentie limiet) mogelijk is. (dit proefschrift)

- Na het bereiken van histologische remissie van idiopathische 'autoimmune' CAH dient deze remissie veelal twee jaar gehandhaafd te blijven met behulp van immunosuppressieve medicatie, teneinde volledig herstel van de synthesefunctie van de lever te bewerkstelligen. (dit proefschrift)
- 9. Bij 'autoimmuun' chronisch actieve hepatitis lijkt een vroeg stadium van hepatorenaal falen, ook wel functionele nierinsufficientie genoemd, reversibel te ziin, niet alleen door middel van levertransplantatie, maar ook met behulp van immunosuppressieve therapie indien de hepatitis daarop in remissie komt. (dit proefschrift)
- Antistoffen tegen cardiolipine komen vaak voor bij chronisch actieve hepatitis, de kruisreactie met anti-DNA antistoffen is beperkt, en er is bij deze ziekte geen correlatie met thrombose of recidiverende spontane abortus aangetoond. (dit proefschrift)
- Er zijn geen aanwijzingen dat het CMV een etiologische rol speelt via 'molecular mimicry' tussen door CMV op de celmembraan geïnduceerde antigenen en antigenen in LSP. (dit proefschrift)
- De bepaling van anti-DNA antistoffen met Crithidia luciliae als substraat kan behulpzaam zijn in de differentiaaldiagnose van SLE met leverproefstoornissen en CAH met extrahepatische manifestaties. (dit proefschrift)
- 13. Bij controle van patiënten met leverziekten wordt vaak te weinig aandacht geschonken aan de gestoorde functies van dit orgaan.
- 14. Het vragen naar nachtblindheid hoort bij de anamnese van een patiënt met leverziekte.
- 15. Het is nog zeer de vraag of de 74 kD antimitochondriale antistoffen van het type M2, gericht tegen pyruvaat-dehydrogenase E2, een (additionele) rol spelen in de pathogenese van primaire biliaire cirrose.
- 16. Daar waar de verstrekking van delen van de gezondheidszorg het monopolie wordt van enkelen, dreigt het belang voor de gemeenschap verloren te gaan.
- 17. Een 'muzikaal gehoor' is de stemvork voor slechts één persoon.
- 18. Het kind van 'tweeverdieners' krijgt vaak niet wat het verdient.

- 19. Aanwezigheid van het 'human immune deficiency virus' (HIV) bij de recipient is geen absolute contra-indicatie tegen orgaantransplantatie. Het kan wel een relatieve contra-indicatie hiertegen zijn in een situatie van beperkte beschikbaarheid van donororganen.
- In de jaren 1965 tot 1980 was er in de Duitse Bondsrepubliek een evidente correlatie tussen de daling van het aantal menselijke geboorten per jaar en van het aantal broedende paren ooievaars. (Nature, 7 april 1988)
- 21. Het toenemend aantal giframpen werkt 'riooljournalistiek' in de hand.
- 22. Bij het voornemen tot opheffing van de grenzen tussen de EEG-lidstaten per 1992 lijkt het gewenst om te anticiperen op mogelijke protectionistische neigingen bij EEG-handelspartners.
- 23. Vele clinici zijn slechts in letterlijke zin 'voordeurdelers'.
- 24. Kunst wordt gemaakt door 'kenners'.
- 25. Nederlanders die in het buitenland verblijven tellen voor verkiezingen niet mee.
- 26. De functie van paranimfen bij een promotie komt pas ten volle tot zijn recht als de promovendus zich in het buitenland bevindt.

Stellingen behorende bij proefschrift

CLINICAL MONITORING OF 'AUTOIMMUNE' CHRONIC ACTIVE HEPATITIS

A histological, biochemical and immunological study

BART VAN HOEK 22 november 1989

CLINICAL MONITORING OF 'AUTOIMMUNE' CHRONIC ACTIVE HEPATITIS

A histological, biochemical and immunological study

PROEFSCHRIFT

ter verkrijging van het doctoraat in de Geneeskunde aan de Rijksuniversiteit Groningen op gezag van Rector Magnificus Dr. L.J. Engels, in het openbaar te verdedigen op woensdag 22 november 1989 des namiddags te 14.45 uur precies.

door

BART VAN HOEK geboren te Voorburg

PROMOTORES:	prof. dr. C.H. Gips
	prof. dr. T.H. The

REFERENTEN:	dr. A.J.K. Grond
	dr. C.G.M. Kallenberg

PROMOTIECOMMISSIE: prof. dr. J.D. Elema prof. dr. H.S.A. Heymans prof. J.H.P. Wilson

aan mijn ouders

© COPYRIGHT 1989 by BART VAN HOEK all rights reserved.

No part of this book may be reproduced, stored in a retrieval system, or transmitted in any form or by any means, electronic, mechanical, photocopying, recording or otherwise, without the prior permission in writing by the author.

Cataloging-in-Publication DATA ROYAL LIBRARY, THE HAGUE, THE NETHERLANDS.

Hoek, Bart van

Clinical monitoring of 'autoimmune' chronic active hepatitis : a histological, biochemical and immunological study / Bart van Hoek. - [S.l. : s.n.] (Groningen : Computekst). -Ill.

Thesis Groningen. - With ref. ISBN 90-9003047-6 SISO 605.13 UDC 616.36-002(043.3) 'Subject heading: hepatitis.

Cover drawing: Huib Waterbolk. (after Arkesilas-painter, Prometheus and Atlas, interior of black-figure Laconian kylix from Cerveteri, 565-555 B.C., Vatican Museums, Rome)

Computer Typeset by COMPUTEKST tekstverwerking, Groningen, The Netherlands. Printed and bound in The Netherlands by Dijkhuizen van Zanten, Groningen.

The printing costs were supported by kind contributions of Merck, Sharp & Dohme and Glaxo.

CONTENTS

Voorwoord /	Ackn	owledgements	XI
Collaboratio	n		XVII
Abbreviatio	ns		XVIII
Scope of this	s study		XXI
Introduction	I		1
	§1	Chronic active hepatitis in a historical perspective	3
	§2	Terminology	3
	§3	Histology	4
	§4	Definition	5
	§5	Clinical symptoms and physical findings	6
	§6	Prognosis of untreated severe CAH	6
	§7	Treatment regimens in CAH	8
	§8	Prognosis of treated CAH	12
	§9	Immunology of IAI-CAH	13
	§10	Problems in the diagnosis and management of chronic active hepatitis	17
	§11	Monitoring activity of disease and liver function in CAH	23

Section 1

Patients and Methods

Chapter 1	Patients	31
Chapter 2	Methods	36
Chapter 3	Numerical versus conventional histological scoring of activity in 'autoimmune' chronic active hepatitis	37
Chapter 4	Serum N-terminal propeptide of collagen type III in 'autoim- mune' chronic active hepatitis. Comparison of three radio- immunoassays and relation with histology	45
Chapter 5	Establishment of a modified quantitive radioimmunoassay for the determination of antibodies against 'liver-specific protein' (LSP)	51

Section 2

Analysis of survival in chronic active hepatitis of various aetiologies

Chapter 6	Long-term prognosis in chronic active hepatitis	59

Section 3

'Autoimmune' chronic active hepatitis: Activity of disease and repair during standardized treatment

Chapter 7	Routine blood tests in standardized immunosuppressive therapy of 'autoimmune' chronic active hepatitis		
	§7.1 Routine 'liver biochemistry' in standardized immuno- suppressive therapy of 'autoimmune' chronic active hepa- titis	75	
	§7.2 Serum sodium, depressed in 'autoimmune' chronic active hepatitis, normalizes during immunosuppressive therapy. Relation to histology, synthesizing capacity of the liver and routine biochemistry	30	
	 Routine haematological parameters in 'autoimmune' chronic active hepatitis, and changes during immunosuppression 	35	
Chapter 8	Serial histology and routine liver biochemistry in standardized immunosuppressive therapy of idiopathic 'autoimmune' chronic active hepatitis		
Chapter 9	Repair and normalization of hepatic functional capacity of protein synthesis despite presence of cirrhosis in 'autoimmune' chronic active hepatitis		
Chapter 10	Acute-phase reactants, C-reactive protein and serum amyloid A, in chronic active hepatitis. Relation to histology and synthesizing capacity of the liver and influence of immunosuppressive therapy		
Chapter 11	Serum N-terminal propeptide of collagen type III in 'auto- immune' chronic active hepatitis. Relation to histology and func- tional capacity of the liver and effects of standardized immuno- suppression		
Chapter 12	The serum N-terminal propeptide of collagen type III as a pre- dictor of relapse in 'autoimmune' chronic active hepatitis		

Section 4

Humoral immunology in chronic active hepatitis

'Autoimmune' chronic active hepatitis

- Chapter 13 Antibodies against 'liver-specific membrane lipoprotein' (LSP) in 147 relation to histology in 'autoimmune' chronic active hepatitis before and during standardized immunosuppression
- Chapter 14 Antibodies against 'liver-specific membrane lipoprotein' (LSP) as 155 an early predictor of relapse of 'autoimmune' chronic active hepatitis during therapy-withdrawal
- Chapter 15 Clinical significance of serologically defining subgroups of idiopathic 'autoimmune' chronic active hepatitis, by presence/ absence of antibodies to soluble liver antigen (SLA) and other autoantibodies

Chronic active hepatitis of various aetiologies:

- Chapter 16 Anti-cardiolipin antibodies in chronic active hepatitis. Prevalence in subgroups and lack of correlation with anti-DNA antibodies or extrahepatic disease
- Chapter 17 Humoral responses against defined cytomegalovirus antigens and liver-specific membrane lipoprotein' in chronic active hepatitis. Lack of evidence for aetiologic involvement of cytomegalovirus by molecular mimicry

General Discussion	201
Summary	205
Samenvatting	211
References	217
Curriculum vitae	265

VOORWOORD/ACKNOWLEDGEMENTS

Het onderzoek, beschreven in dit proefschrift, werd verricht binnen de afdeling Hepatologie (hoofd: Prof. Dr. C.H. Gips) en de afdeling Klinische Immunologie (hoofd: Prof. Dr. T.H. The) van de Kliniek voor Inwendige Geneeskunde (hoofd: Prof. Dr. G.K. van der Hem) van het Academisch Ziekenhuis te Groningen.

Bij het tot stand komen van dit proefschrift waren de hulp en de bijdragen van velen onontbeerlijk. Een aantal van hen ben ik bijzondere dank verschuldigd.

Mijn ouders dank ik voor hun warme belangstelling, steun en aanmoediging die zij altijd gegeven hebben, waardoor ik het gestelde doel kon bereiken.

De fundamenten voor dit proefschrift zijn reeds vele jaren geleden, ruim voor de periode waarin dit onderzoek werd verricht, gelegd door Prof. Dr. C.H. Gips. Beste Chris, je aanstekelijk enthousiasme in combinatie met je diepgaande kennis van de pathofysiologie van de lever hebben mij getroffen en steeds aangespoord mij verder te verdiepen in dit interessante gebied van de interne geneeskunde. Je vriendschap waardeer ik zeer.

Prof. Dr. T.H. The ben ik zeer erkentelijk voor de vele stimulerende gesprekken en het scheppen van ruime mogelijkheden voor het doen van onderzoek. Hij introduceerde mij in het fascinerende gebied van de Klinische Immunologie. De plezierige en persoonlijke wijze waarop hij mij steeds begeleidde was een belangrijke stimulans op vele momenten.

Dr. A.J.K. Grond (afdeling Pathologische Anatomie) was een voortdurende bron van tomeloze energie. Beste Joris, voor de wijze waarop jij mede de aanzet en later de uitwerking van dit proefschrift mogelijk maakte wil ik je hartelijk danken. Je kennis van de leverpathologie bewonder ik zeer. Ik ben je erkentelijk voor de wijze waarop je mijn huidige verblijf in de Mayo Clinic gestimuleerd hebt.

Dr. C.G.M. Kallenberg (afdeling Klinische Immunologie) opende wegen voor nieuw onderzoek, en was verantwoordelijk voor vele suggesties en verbeteringen. Beste Cees, van je kennis van autoimmuunziekten heb ik veel geleerd.

Ik ben Prof. Dr. E. Mandema, Prof. J.H. Scholten en Prof. Dr. W.D. Reitsma bijzonder erkentelijk voor mijn opleiding tot internist, en ik dank Prof. Dr. G.K. van der Hem voor de genoten gastvrijheid binnen de Kliniek voor Inwendige Geneeskunde van het Academisch Ziekenhuis te Groningen.

De leden van de promotie-commissie, Prof. Dr. J.H.P. Wilson (Rotterdam), Prof. Dr. J.D. Elema (Groningen) en Prof. Dr. H.S.A. Heymans (Groningen) dank ik voor het kritisch doornemen van het manuscript.

En speciaal woord van dank betreft de heer J.R. (Reint) Huizenga. Reint, de plezierige sfeer die jij wist te creëren in het door jou geleide laboratorium hepatologie was voor mij en anderen steeds een stimulans om door te gaan. De nauwgezette wijze waarop je sinds 1969 een uitgebreid serumarchief hebt opgezet is uniek en was een belangrijke basis voor dit onderzoek. Jij zorgde ervoor dat vanuit de U.S.A. toch steeds alles geregeld kon worden. In de loop der jaren heb je een uitgebreide kennis van de hepatologie verworven. Aan de vele discussies die wij voerden zowel in werksfeer als daarbuiten denk ik met plezier terug. Dit alles heeft geresulteerd in een mij dierbare vriendschap.

Aan mevrouw T.H. (Tineke) Lijnema wil ik mijn bewondering uitspreken voor de snelle en intelligente wijze waarop zij zich nieuwe onderwerpen en technieken eigen wist te maken. Tineke, je was een onmisbare steun, en ik hoop dat je huidige aanstelling in het kader van het project "chronisch actieve hepatitis" een vastere basis krijgt.

Dat het Academisch Ziekenhuis de heer drs. A.J.(Arja) Mackor ook in de gelegenheid stelt om binnen de afdeling hepatologie onderzoek naar dit ziektebeeld te verrichten verheugt mij zeer.

Mevr. Mr. A.K. (Annie) van Zanten wil ik bijzonder hartelijk danken voor de gastvrijheid en plezierige samenwerking bij het opzetten van de anti-LSP bepaling en het verrichten van vele PIIINP bepalingen.

De heer Dr. V. Fidler verzorgde op deskundige wijze de statische analyse van gegevens betreffende overleving en prognostische parameters. Zeer hartelijk dank.

Mevrouw E. (Els) Hummel-Tappel, mevrouw C. (Carla) Terpstra en de heer C. (Chairul) Effendi wil ik hartelijk danken voor het verrichten van een groot deel van de anticardiolipine, ANA, a-ENA en anti-DNA-bepalingen, alsmede de plezierige sfeer waarin dit verliep.

Van de heer Dr. P.C. (Piet) Limburg en collega Drs. B.P.C. (Bauke) Hazenberg kreeg ik steeds op plezierige wijze een acute fase respons op nieuwe versies van het artikel betreffende CRP en SAA. Prof. Dr. M.H. (Martin) van Rijswijk dank ik voor verhelderende discussies over dit onderwerp, alsmede voor het scheppen van de mogelijkheden voor dit gezamenlijk onderzoek. De heer J. (Johan) Bijzet dank ik voor de vele bepalingen die hij hiervoor verrichtte. De heer S. (Siep) Postma en mevrouw J. (Coba) van Zanten dank ik voor het binnen zeer korte tijd verrichten van vele ELISA's voor het onderzoek betreffende CMV. Mevrouw Dr. M. (Marijke) van der Giessen speelde een belangrijke rol in het interpreteren van deze uitslagen. Dr. L.F.M.H. (Lou) de Leij, mevrouw J.G. (Anita) ter Haar, en mevrouw H.A. (Harmke) de Vries-Huiges dank ik voor mijn introductie in het laboratorium van de afdeling Klinische Immunologie.

I would like to thank Dr. R. (Roger) Williams, FRCP, for the opportunity he gave me to work in his excellent laboratory in King's College Hospital, London, Great-Brittain. Dr. I.G. (Ian) McFarlane and Dr. B.M. (Barbara) McFarlane-Wojcicka showed me how to perform the radioimmunoassay for anti-LSP and anti-ASGR. I very much appreciate their help and friendliness during and after my stay in London.

De Nederlandse Lever Darm Stichting ondersteunde dit verblijf in London financieel.

I wish to thank Prof. Dr. Dr. K.-H. Meyer zum Büschenfelde and Prof. Dr. M. (Michael) Manns for their hospitality and help during the weeks we spent in their well-known laboratory in the Clinic of the Johannes Gutenberg University in Mainz, West-Germany. I also thank mrs. S. Bratfish and Mr. Hodawand for technical assistance.

I thank Dr. J. (Juha) and Dr. L. (Leila) Risteli, Collagen Research Unit, Department of Medical Biochemistry, University of Oulu, Oulu, Finland, who did part of the determinations with the new human PIIINP assay they developed.

De dames K. (Karin) Cazemier en A.M. (Anne Marie) Zoelman en de heren H. (Henk) Oost en K.H.P. (Peter) Geres verrichten als stagiaires vele cholinesterase bepalingen en hielpen met het serum-archief. Dank!

Mevrouw H.J. (Rina) de Jong (laboratorium immunochemie, hoofd Dr. J. Marrink) assisteerde op deskundige wijze bij het isoleren van LSP.

De heer J.G.J. (Joan) Vos dank ik voor het vliegensvlug verrichten van, onder andere, hepatitis B serologie.

De heer F. (Frans) Broekhuizen (polikliniek hepatologie) en de registratie-assitentes van afdeling A2 waren essentieel voor het continueren van dit archief.

Dr. J. Trommel, voormalig geneesheer-direkteur van het Talmahuis te Veenwouden, verstrekte gegevens betreffende een relatief groot aantal met oxyphenisatine behandelde patiënten, waarvan degenen bij wie een in het AZG verrichte leverbiopsie CAH liet zien in dit onderzoek zijn opgenomen. De heer Dr. J.A. Kramps, Academisch Ziekenhuis te Leiden, (lab. longziekten) dank ik voor het verrichten van α_1 -antitrypsine fenotyperingen bij alle patiënten.

Op mevrouw M.H. (Kiki) Bugter, mevrouw J.A. (Jetty) Renkema, mevrouw M.G.H. (Riet) van den Broek en mevrouw M. (Martha) Messchendorp deed ik nooit een vergeefs beroep. Mevrouw R. (Renske) Wekema was een belangrijke schakel in de overzeese communicatie.

De dames en heren van het statusarchief zorgden ervoor dat gegevens snel beschikbaar waren.

De heren K. (Klaas) van der Linde en G. (Gerard) Glade verrichtten, naast hun studie in de geneeskunde, belangrijk werk bij het verzamelen van gegevens. Dr. H. (Herman) Kreeftenberg en Drs. J. (Jan) Jager dank ik voor de opbouwende discussies tijdens diverse besprekingen. Dit gold ook voor mevrouw P. (Pauline) Schuur en de heren N. (Niele) Bhairo, G.M. (Go) van Dam, R. (Rik) Reekers, en drs G.J. (Gerrit Jan) Westerveld, die hun studie goed wisten te combineren met het verrichten van onderzoek in de afdeling hepatologie.

De patiënten waarop dit onderzoek gebaseerd is wil ik hartelijk danken voor hun medewerking en de toestemming een serumarchief aan te leggen voor later onderzoek. Ik hoop van ganser harte dat de resultaten van dit onderzoek hun behandeling ten goede zal komen.

Vele internisten, huisartsen en patholoog-anatomen uit Nederland en daarbuiten stelden gegevens en leverbiopsieën te beschikking van patiënten die zowel afwisselend elders als in het Academisch Ziekenhuis te Groningen behandeld werden. Ik dank hen zeer hartelijk voor deze medewerking. Ook de medici die in het Academisch Ziekenhuis Groningen in de loop der jaren de zorg voor de beschreven patiënten hadden dank ik voor de verstrekte gegevens. Diverse patiënten verstrekten gegevens. Gemeentehuizen gaven waardevolle informatie. Dank daarvoor.

Alle medewerkers van het CKCL (in het bijzonder G. (Greetje) Sienot en de heer E.E. (Eddie) Ligon), het haematologie laboratorium, en het stollings-laboratorium dank ik voor het verrichten van de routine bepalingen die opgenomen zijn in dit proefschrift.

De heer A. Huizer van de afdeling Medische Fotodienst van de RUG dank ik voor de snelle en correcte wijze waarop hij grafieken "drukklaar" maakte.

De heer A.J. (Bert) Schaalma assisteerde waar mogelijk bij het verzamelen van literatuur. Dr. K. H. Brandt, internist, wijlen Dr. F. Bronkhorst, patholoog-anatoom (Ziekenhuis Rijnstate, lokatie Gemeenteziekenhuis te Arnhem), Prof. Dr. S.W. Schalm (Academisch Ziekenhuis Dijkzigt, Rotterdam), Prof. Dr. G.P. van Berge Henegouwen (Academisch Ziekenhuis Utrecht), Dr. R.A. Janssens (voorheen Academisch Ziekenhuis Leiden) en Dr. P. Niermeyer (Majella Ziekenhuis, Bussum) dank ik voor de vele stimulerende discussies omtrent chronisch actieve hepatitis, onder andere tijdens de voorbereiding van een cursus en een symposium.

De firma's Behring en Imphos dank ik voor het gratis ter beschikking stellen voor radioimmunoassay-kits voor de bepaling van PIIINP. De uitkomst dat resultaten van beider assays goed correleren lijkt mij voor beide firma's een verheugend gegeven.

De Nederlandse Organisatie voor Wetenschappelijk Onderzoek (NWO) dank ik voor het ter beschikking stellen van een beurs, die mij momenteel in de gelegenheid stelt onderzoek naar, onder andere, chronisch leverziekten en rejectie na levertransplantatie te verrichten in de Mayo Clinic te Rochester, Minnesota, U.S.A.

I wish to thank Dr. A.J. Czaja, M.D. and Dr. R.H. Wiesner, M.D., Dept. of Gastroenterology and Internal Medicine, Mayo Clinic, Rochester MN, U.S.A., for valuable comments after critically reviewing the manuscript of this book.

I thank Suzan and Anthony Hayes for editing the English language in this thesis. You did a marvelous job, and I hope our friendship will last forever.

De vriendschap van mevrouw Mr. J. (Jeanne) Wesseling-Lubberink en van de heer J.R. (Reint) Huizenga is mij zeer waardevol, en hun vele inspanningen als paranimfen waardeer ik zeer.

"Last but not least" wil ik de gecompliceerde interactie noemen die Beryl, dit proefschrift en ik met elkaar hadden tijdens de wording van dit boek. Steeds wist je mij op originele wijze ervan te overtuigen dat er nog vele andere belangrijke zaken in het leven bestaan. Dit heeft op paradoxale wijze bijgedragen aan de voltooiïng van dit werk.

De vriendschap en hulp van velen worden in dankbaarheid gememoreerd.

COLLABORATION

Chapter 4	J. Risteli, L. Risteli, J.R. Huizenga
Chapter 5	T.H. Lijnema, A.K. van Zanten, J.R. Huizenga, I.G. McFarlane, B.M.
	McFarlane
Chapter 6	G. Glade, V. Fidler
Chapter 9	J.R. Huizenga, T.H. Lijnema
Chapter 10	B.P.C. Hazenberg, P.C. Limburg, J. Bijzet, J.R. Huizenga, M.H. van
	Rijswijk
Chapter 11	J. Risteli, J.R. Huizenga, L. Risteli
Chapter 12	J.R. Huizenga, T.H. Lijnema
Chapter 13	T.H. Lijnema, A.K. van Zanten, J.R. Huizenga
Chapter 14	T.H. Lijnema, A.K. van Zanten, J.R. Huizenga
Chapter 15	M. Manns, J.R. Huizenga, KH. Meyer zum Büschenfelde
Chapter 16	C.G.M. Kallenberg, C. Effendi, P.C. Limburg, E. Hummel-Tappel
Chapter 17	T.H. Lijnema, C.G.M. Kallenberg, M. van der Giessen, A.K. van
	Zanten, J.R. Huizenga

LIST OF ABBREVIATIONS

AAA	=	aromatic amino acids
ABT	=	aminopyrine breath test
AI-CAH	=	'autoimmune' CAH
ALB	=	albumin
A1AT	=	alpha ₁ antitrypsin
ALT	=	alanine aminotransferase
AMA	=	antimitochondrial antibodies
ANA	=	antinuclear antibodies
anti-DNA	=	antibodies against dsDNA
anti-HBs	=	antibodies against HBsAg
anti-HBs	=	antibodies against HBcAg
anti-M2	=	AMA against M2 antigen
anti-Mx	=	AMA against Mx antigen
APTT	=	activated partial prothrombin time
ASGR	=	asialoglycoprotein receptor
AST	=	aspartate aminotransferase
ATIII	=	antithrombin III
AZA	=	azathioprine
BCAA	=	branched chain amino acids
BUN	=	blood urea nitrogen
CAH	=	chronic active hepatitis
CB	_	conjugated bilirubin
CHE	_	(nseudo-)cholinesterase
СТН	_	chronic lobular henatitis
CMV	_	cytomegalovirus
CPH	_	chronic persistent henatitis
CREAT	_	creatinin
CRP	_	C-reactive protein
CTI	_	cytotoxic T-lymphocyte
CvA	_	cyclosporin A
	_	drug-induced
	_	double stronded desoruribonucleic acid
USDINA E A	=	double-suanded desoxymboliticiele acid
	=	cally allight
ELISA	=	enzyme-mikeu minunosorbem assay
ENA	=	
EO	=	eosinophii granulocytes
GG	=	
GEC	=	galaktose elimination clearance
HAS	=	Histological Activity Score (= P+PP+L)
Hb	=	nemoglobin
HBcAg	=	hepatitis B virus core antigen
HBD	=	healthy blood donors
HBeAg	=	hepatitis B virus e-antigen
HBsAg	=	hepatitis B virus surface antigen
HDL ₃	=	high density lipoprotein type 3
HFS	=	Histological Fibrosis Score
Ht	=	hematocrit

IAI-CAH	=	idiopathic 'autoimmune' chronic active hepatitis	
ICH	=	immunecompromised host	
ΙĒΑ	=	immediate early antigen	
IFN	=	interferon	
IL-1B	=	interleukin 1ß	
IL-2	=	interleukin 2	
IL-2R	=	interleukin 2 receptors	
IL-6	=	interleukin 6	
K cells	=	killer cells	
kD	=	kiloDalton	
K-M	=	Kaplan-Meier	
L	=	lobular inflammation score	
LA	=	late antigen	
LAK cells	=	lymphokine-activated killer cells	
LEU	=	leukocytes	
LC-1	=	liver cytosol antigen(s) type 1	
LKM-1	=	liver kidney microsomal antigen type 1	
LMA	=	liver membrane antigen	
LSP	=	'liver-specific membrane lipoprotein'	
LT	=	liver transplantation	
LYMPHO	=	lymphocytes	
MA	=	membrane antigen	
Na	=	sodium	
NK cells	=	natural killer cells	
nRNP	=	an extractable nuclear antigen	
OLT	=	orthotopic liver transplantation	
Ρ	=	portal inflammation score	
PP	=	periportal inflammation score	
PLASMA	=	plasma cells	
PMN	=	polymorphonuclear leucocytes	
PRED	=	prednisolone	
PT	=	prothrombin time	
PIIINP	=	N-terminal propeptide of collagen type III	
RIA	=	radioimmunoassay	
SAA	=	serum amyloid precursor A	
SAP	=	serum amyloid P-component	
SBA	=	serum bile acids	
SLA	=	soluble liver antigen	
Sm	=	an extractable nuclear antigen	
SS-A	=	an extractable nuclear antigen	
SS-B	=	an extractable nuclear antigen	
UB	=	unconjugated bilirubin	
ТВ	=	total bilirubin (=UB+CB)	
THR	=	thrombocytes	
TNF-α	=	tumor necrosis factor α	
	_		

SCOPE OF THIS STUDY

The scope of this work was to gain insight into the effect of an accepted standardized therapeutic regimen for 'autoimmune' chronic active hepatitis, consisting of prednisolone and azathioprine in a small group of patients treated according to the same standardized protocol in a specialized hepatological unit of a university hospital. In some parts of the study these patients were analyzed against the background of a larger group of patients with CAH referred within the same period of time. This represented a limited number of measurements as survival, antibodies to cytomegalovirus, anticardiolipin, anti-DNA and antinuclear antibodies. The items we studied can be divided into three groups:

- 1) parameters of inflammation and derived parameters such as fibrogenesis,
- 2) parameters of repair and restoration of liver function, and
- 3) immunological parameters.

With regard to immunological parameters we studied pathogenetic factors like anti-LSP autoantibodies, as well as other (epiphenomenal?) autoantibodies. Antiviral antibodies were determined in a search into etiology of 'autoimmune' chronic active hepatitis. The value of these parameters for diagnosis, therapy and follow-up of 'autoimmune' chronic active hepatitis is discussed. The work is introduced by a review of the literature and is concluded with an analysis of our observations and suggestions for future lines of research.

INTRODUCTION

§1 CHRONIC ACTIVE HEPATITIS IN A HISTORICAL PERSPECTIVE

In 1907 Noël Fissinger described a disease in which the immune system does no longer defend but attacks the liver ("le sujet ne défend plus son foie, il se défend contre son foie"), raising the concept of autoimmune liver disease.¹ The concept of idiopathic, often recurrent, jaundice in association with severe chronic necrosis and inflammation of the liver emerged in the late 1930's and early 1940's.^{2,3} The components of a syndrome, however, were not identified until the 1950's, when cirrhosis with plasma cell infiltration of the liver was recognised in young women who had hypergammaglobulinaemia, fever, arthralgias, acne, and delayed menstruation and amenorrhoea, splenomegaly, obscure febrile reactions, and Cushingoid facies.⁴⁻⁹ The term active chronic hepatitis was first applied in 1953¹⁰ and was later modified to lupoid hepatitis in 1956^{11} when the frequency of association with the lupus erythematosis cell phenomenon was appreciated.¹² Subsequently, immunoserologic findings were regarded as having etiologic importance and an 'autoimmune' concept of the disease developed.13-¹⁹ The emergence of additional immunoserologic tests, including assays for smooth muscle antibodies (SMA), antimitochondrial antibodies (AMA), and hepatitis B surface antigen (HBsAg), allowed subclassification of patients with the same generic disease but different clinical and laboratory manifestations. Subgroups were distinguished by, for instance, specific immunoserologic findings.¹⁸⁻²¹ presence of HBsAg,²²⁻²⁴ associated autoimmune diseases²⁵⁻²⁶ and sex.²⁷ Prognosis and multiple morphologic patterns were also described.²⁸ Eventually a single classification was proposed which comprised all disorders of similar clinical, biochemical, and histological features of periportal hepatitis,^{29,30,31} and the term *chronic active hepatitis* (CAH) became current, Although incorrectly regarded as specific for CAH, the histologic pattern of *piecemeal necrosis* is regarded as the sine qua non of all forms of CAH.

§2 TERMINOLOGY

Although chronic viral hepatitis was described in 1947,³² the first histopathological description of CAH dates from 1966.³³ De Groote *et al.* defined and classified chronic hepatitis based on histopathological findings into chronic aggressive hepatitis (CAH), chronic persistent hepatitis (CPH), and chronic lobular hepatitis (CLH).^{29,31} With the gaining of knowledge the classification of chronic liver diseases underwent some modifications, the most important being the recognition of primary biliary cirrhosis (PBC) and primary sclerosing cholangitis (PSC) as distinct entities.³⁴ This implies that in the earlier descriptions of chronic hepatitis incorporates CAH, CPH and CLH but not PBC or PSC. We will use this terminology, although CAH, CPH and CLH probably belong to the spectrum of one disease. Moreover, the distinction between CAH

and PBC or PSC can be difficult or even impossible, suggesting some overlap as well. Another problem in interpreting the early studies on CAH is that in these trials patients with chronic hepatitis B and other causes of CAH or 'CAH-related liver disease' (table 1) were included. Terminology like "chronic active liver disease", "chronic active cirrhosis", "chronic aggressive hepatitis", and "subacute hepatitis" has been abandoned now, because it does not serve a special purpose and produces confusion.^{35,36}

disorders'.	
	Abbreviation used:
САН	
'Autoimmune', aetiology unknown	Idiopathic 'autoimmune' CAH
'Autoimmune', after hepatitis B (anti-HBs positive)	Anti-HBs positive 'autoimmune' CAH
Hepatitis B virus (HBsAg positive)	HBsAg-positive CAH
Hepatitis C virus (and other parenterally transmitted, but unknown non-A,non-B viruses)	HCV-CAH or NANB-CAH
Hepatitis delta	HDV-CAH
CAH-related disorders	
drug-induced	DI-CAH
Wilson's disease	Wilson-CAH
α_1 -antitrypsin deficiency	A1AT-CAH
'small-duct primary sclerosing cholangitis'/	
CAH/PSC 'overlap'	CAH/PSC
cholestatic CAH ('overlap' of CAH with	
primary biliary cirrhosis)	CAH/PBC
alcohol related	
haemochromatosis	
anti-chymotrypsin deficiency	

 Table 1
 Common causes of chronic active hepatitis (CAH) and 'CAH-related disorders'.

§3 HISTOLOGY

The traditional description of the histology of CAH has its roots in the autoimmune form.³⁷ The hallmark of CAH is portal and periportal inflammation with erosion of the limiting plate, piecemeal necrosis, and fibrosis. The classic or lymphocytic form of piecemeal necrosis was originally described by Popper *et al.*³⁸ It may be defined as inflammatory destruction of single or small groups of liver cells at a mesenchymal/

parenchymal interface (periportal, periseptal or at the margin of confluent necrosis) associated with lymphohistiocytic inflammatory cells.³¹ Bianchi states that a close contact between the membranes of clustered lymphocytes/macrophages and hepatocytes (peripolesis³⁹) is a prerequisite for the diagnosis of piecemeal necrosis and hence of chronic active hepatitis, while periportal hepatitis without peripolesis is referred to as 'CAH-related liver disease'.⁴⁰ Others do not include peripolesis in the definition of CAH. One of the reasons is that recognizing the presence or absence of peripolesis can be difficult, even for experienced pathologists.^{35,36,41} In chronic persistent hepatitis (CPH), the predominantly lymphocytic infiltrate is restricted to widened portal tracts, leaving the parenchymal limiting plate preserved.⁴² A special form of CPH, chronic septal hepatitis⁴³ is characterized by the formation of septa with conspicuous inflammatory activity but without piecemeal necrosis. This may represent a regressive stage of CAH, as seen spontaneously or after immunosuppressive therapy⁴⁴ and will be referred to as CPH or CAH in remission. When acute lobular features of the degree seen in acute hepatitis are present over a period of more than 6 months, this is termed chronic lobular hepatitis (CLH).45,46 For more extensive descriptions of histology the reader is referred to Ludwig, Bianchi and Scheuer.35,40,41

§4 DEFINITION

In general, the definition of CAH includes only CAH with parenteral hepatitis viruses as cause, or with unknown cause, with or without 'autoimmune' phenomena.

Conditions such as α_1 -antitrypsin deficiency or Wilson's disease are excluded, and are labelled *CAH-related disorders*.^{35,36,40}. Other authors however, state that these diseases are other causes of the same pattern of periportal hepatitis.⁴⁷

We will adhere to the first definition of CAH, being:

- a primarily hepatocytic liver disease
- characterized by a common pattern of progression, that is: (episodes of) periportal and portal inflammation and piecemeal necrosis, with the potential to progress to cirrhosis or being associated with cirrhosis
- a condition which will last 6 months or longer,
- caused by a presumed autoimmune disorder, idiopathic or cryptogenic CAH caused by unknown aetiology, or by hepatitis virus type B (HBV) (with or without type D), or a parenteral hepatitis virus type non-A,non-B (e.g. hepatitis C).

This definition of CAH includes 1) morphology, 2) various aetiologies as mentioned above and 3) a clinical observation time.

Certain liver diseases, listed in table 1, can have histopathological features resembling CAH with periportal hepatitis and piecemeal necrosis. Such cases are termed *chronic* hepatopathy with features of CAH, CAH-related disorder, or CAH-related liver disease.^{35,36,40}

We will define 'autoimmune' CAH as

- idiopathic CAH, in the absence of known causes of CAH
- in the presence of at least one immunologic disorder.

This excludes patients with e.g. HB_sA_g , but includes patients with only anti-HBs that fulfill these criteria. Patients can have AMA, but not of subtype M₂. The immunologic disorder can include presence of other autoantibodies, like ANA or SMA or elevated gamma globulin, and can also be associated extrahepatic autoimmune disease.

§5 CLINICAL SYMPTOMS AND PHYSICAL FINDINGS

Initial reports by Waldenström and Kunkel emphasized the occurrence of CAH in young women with amenorrhoea, hepatosplenomegaly, hyperglobulinaemia, cushingoid facies, pigmented abdominal striae and arthritis.^{4,5} 70-80% of the patients with idiopathic 'autoimmune' CAH (IAI-CAH) are women.⁴⁸⁻⁵⁰ The disease is usually detected in the 3th to 5th decade of life, but young children and adults in their 60s and 70s may also be affected.⁴⁸⁻⁵⁰ In approximately one third of the patients the disease begins abruptly and mimics acute viral hepatitis. The majority of the patients described in the literature had a more insidious onset of disease , however, with progressive jaundice (61%), abdominal pain (35%), epistaxis (21%), acne (19%), persistent fever (11%) and arthralgia (11%).⁴⁸ Fatigue, jaundice, arthralgias, and abdominal pain are the most frequently occurs.⁴⁸⁻⁵⁰ Sometimes patients present with extrahepatic manifestations of the disease, or develop such features later in the disease.^{25,48-51} Up to 20% of the CAH-patients fulfill the revised ARA-criteria for SLE.⁵²

§6 PROGNOSIS OF UNTREATED SEVERE CAH

Early assessments of natural history and prognosis of severe CAH were recently summarized.⁵³ Mortality ranged from 20% in 3 years,⁵⁴ 40% in 4 years,⁵⁵ 80% within 5 years,⁵⁶ or 70% in 6 years,⁵⁷ to 50% within 10 years.⁸ Prospective studies have indicated that mortality approximates 40 percent within 6 months of diagnosis if corticosteroids are not administered in severe CAH.⁵⁰

Prognostic factors

- Histology and presence/absence of cirrhosis:

It has been demonstrated that cirrhosis occurred in 90% of untreated patients within 4 years.^{48,58,59} Cirrhosis at presentation was associated with the highest mortality.⁶⁰ Untreated patients with morphologic evidence of bridging or multilobular necrosis had an 82% incidence of cirrhosis within 5 years and a mortality of 45%. In contrast, patients with only periportal necrosis had less than a 17% incidence of cirrhosis and a normal life expectancy.⁶¹ Cirrhosis frequently develops in early survivors,⁵⁷ and late mortality usually is related to the consequences of portal hypertension rather than liver failure, the main cause of early death in CAH.⁴⁸ Survival beyond 3 years in untreated CAH was frequently associated with a spontaneous reduction in inflammatory activity, and with transition to an inactive macronodular cirrhosis in most survivors.⁴⁸

The pattern of fluctuating inflammatory activity was later recognized as a behavioral characteristic of CAH. Spontaneous clinical improvement occurs in 40% of untreated patients within 6 months of diagnosis, and histologic resolution has been described in 20% of patients surviving from 1 to 3 years after diagnosis.⁵⁰ Of the patients entering remission, 67% sustained the improvement. Death from liver failure eventuated in 41% and cirrhosis developed in 59%.⁶¹ Patients who improved without corticosteroids could not be distinguished from others on accession by disease duration, age, sex, initial histologic findings, associated autoimmune disease (with exception of colitis ulcerosa, which probably indicates that PSC and not CAH is present), immunoserologic studies (presence of LE-cell phenomenon, antinuclear antibody, smooth muscle antibody), or biochemical abnormalities (AST, bilirubin, gamma globulin, albumin).53.62 Patients with mild disease improve without therapy as frequently as those with severe disease (13 percent versus 20 percent).⁶³ Although less than severe CAH has a better prognosis than the more severe forms of CAH, its course is not always a benign one. In a study of 80 asymptomatic patients, including 34 patients with HBsAg, 20% developed cirrhosis and 9% died of liver failure, while the 5- and 10-year survival probabilities were 91 and 81 percent respectively.⁶⁴ The presence of nonsuppurative cholangitis seems to have prognostic significance by increasing the likelihood of cirrhotic transformation.65

- Aetiology:

Aetiology is an important prognostic factor in CAH: The prognosis of untreated IAI-CAH has been mentioned above. Although it has been described,^{66,67} HCC in patients with IAI-CAH is rare. 70% of HBsAg-positive patients developed cirrhosis and 30 percent died within 10 years.^{68,69} These HBsAg-positive patients have an increased risk for the development of hepatocellular carcinoma (HCC),⁷⁰ even if a patient has become an asymptomatic carrier.⁷¹ In HCV-hepatitis most patients are anicteric and

35% asymptomatic during the initial episode. Nevertheless, the majority (55-65%) develop progressive liver disease with CAH, with or without cirrhosis^{72,73} In transfusion-related HCV-CAH however, spontaneous improvement is usual, a complete resolution having been described in half of the patients within 3 years after diagnosis.^{74,75} Cirrhosis developed in 10-60 percent, immediate survival was excellent and HCC was absent.^{73,75,76} HCC after HCV-hepatitis has been reported, however.^{77,78}

- Other determinants of prognosis

The abrupt onset of symptoms, persistent jaundice, hepatic encephalopathy, ascites, and ulcerative colitis were associated with poor prognosis.⁵⁸ Serum aminotransferase elevation of greater than tenfold normal, or fivefold normal in conjunction with twofold elevation of gamma globulin level, was associated with mortality of greater than 50% within 3 years of diagnosis.³⁰ The presence of extensive hepatocellular necrosis with or without cirrhosis presaged a fatal outcome within 5 years in untreated patients.^{58,59} Patients presenting on first admission with bleeding from varices, hepatic encephalopathy, ascites, greater prolongation of prothrombin time, cirrhosis or confluent necrosis in the liver biopsy, or HBsAg-positivity, have a worse prognosis than patients without these features.⁷⁹

§7 TREATMENT REGIMENS IN CAH

In 1964, Page et al demonstrated that therapy with 6-mercaptopurine (6-MP) could improve the clinical, biochemical and histologic manifestations of CAH, without reducing the level of hypergammaglobulinaemia.⁶⁰ Cook et al.,⁸⁰ Kirk et al.,⁸¹ and Vogten⁸² showed that variable doses of prednis(ol)one improved certain liver tests and survival,^{80,81} Appropriate downward *titration* of corticosteroids was used to reduce drug-toxicity. The Copenhagen Study Group for Liver Diseases comparing 10 mg prednisone daily and placebo in histologically confirmed cirrhosis showed that survival was not enhanced by prednisone (45% after 4 years and 20% after 8 years), although extrapolation from these results suggested that a subgroup of patients with clinical characteristics commonly associated with IAI-CAH could be improved by corticosteroid therapy.^{83,84} Therapy with a *fixed dose of prednisone* (15 mg daily) appeared to be more effective than azathioprine (75 mg daily) for increasing the life expectancy of patients with CAH with or without cirrhosis.⁸⁵ Soloway et al. proved that therapy with a fixed dose of prednisone 20 mg, or a combination of prednisone 10 mg with azathioprine 50 mg improved survival and secured clinical, biochemical and histological remission of CAH significantly better than azathioprine (100 mg) or placebo.⁵⁰ Subsequently Summerskill *et al.* showed that this combination therapy with prednisone 10 mg and azathioprine 50 mg is the initial treatment of choice for CAH (fulfilling criteria for treatment. The resolution of disease activity occurred as often as
with prednisone 20 mg, whereas side-effects were significantly less frequent, and because with both treatment regimens histological remission occurred more often than in the group receiving 'dose-titration' of prednisone.⁸⁶ Re-analysis of this study, however, showed no difference between the remission rates in fixed dose prednisone and the titration regimen. Remission was more frequent in patients receiving fixed dose prednisone and azathioprine, however.⁸⁷ Although this retrospective analysis also showed that in the untreated control group cirrhosis was significantly more common in patients with treatment failure (65%) than in those who achieved remission (37.5%). This might have made therapy appear more effective than it would have been with an equal distribution of cirrhosis between the control and treatment groups. The main conclusions of the Mayo study remain unaffected,⁸⁷ and this study is the basis of most current immunosuppressive treatment regimens in CAH.

Current absolute criteria for treatment of CAH (in the absence of contra indications) are AST>10N or AST>5N with GG>2N (N = upper reference limit). Arthritis, fatigue, itching or other severe subjective complaints are all relative indication for treatment. At present it is uncertain whether less severe CAH should be treated or not. It is also unknown whether compromised liver synthetic function or the presence of functional heparorenal failure should be included in the criteria for treatment. Further problems will be mentioned in 10 of this introduction.

Two treatment options have been recommended in the management of idiopathic 'autoimmune' CAH: Prednisone in a high initial dose (60 mg daily) tapered over a 4-week period to a fixed maintenance dose (20 mg daily) is as effective as prednisone (30 mg daily) tapered to a smaller dose (10 mg daily) in conjuction with a fixed maintenance dose of azathioprine (50 mg daily).^{88,89} Hepatic conversion of prednisone to prednisolone, the metabolically active compound, is not greatly impaired in patients with and without cirrhosis and there is no reason to prefer either one.^{90,91} Withdrawal of azathioprine in patients maintained on a combination of azathioprine and prednisolone, leaving the dose of prednisolone unchanged, increases the probability of relapse.^{92,93} After induction of remission some patients can be maintained on azathioprine alone to prevent relapse.⁹⁴ Relapse occurs in up to 80% of patients when an attempt to withdraw corticosteroid therapy is made, while a minority enters sustained remission.⁹⁵⁻⁹⁷

Immunosuppression for viral CAH ?

The clinical data regarding steroid treatment of HBsAg-positive CAH are contradictory. Controlled studies have suggested that steroids are ineffective and even deleterious in HBsAg-positive hepatitis.^{98,99} Prednisone delays loss of HBeAg and seroconversion to anti-HBe⁹⁸ and does not affect⁹⁸ (and possibly increases^{99,100}) mortality in HBsAgpositive patients, especially in those with cirrhosis. Corticosteroid treatment in HBsAgpositive patients may even delay biochemical remission and hastened biochemical relapse.¹⁰⁰ Some reports indicate that short courses of corticosteroid treatment may be beneficial in some patients with acute clinical and biochemical exacerbations of chronic HBsAg-positive hepatitis.¹⁰¹⁻¹⁰⁴ These reports should be interpreted with caution, however: Immunosuppression may lead to an increase in amount of hepatitis B virus in the hepatocyte and/or expression of virally coded antigens on the hepatocyte, 103, 105 frequently resulting in a hyperimmune rebound state after withdrawal of immunosuppression.^{103,106} This presumed effect is used in current evaluation of treatment of hepatitis B, in combination with subsequent immunostimulatory therapy. A recent controlled trial of a 4-week course of prednisolone in chronic hepatitis-B demonstrated that even short courses of corticosteroid treatment can be potentially detrimental in hepatitis B.¹⁰⁷ While it is uncertain whether prednisone increases the level of viral replication, it has been demonstrated that corticosteroid therapy increases the average duration of active HBV replication,^{108,109} which might result in progression of the liver disease and might increase the likelihood of integration of viral DNA sequences into the host genome.¹¹⁰ These data combined with the fact that steroid therapy can give rise to serious side effects, lead to the conclusion that steroids should be only considered in patients with severe and potentially life-threatening hepatitis B, and even then may be contra-indicated.

In anti-HBe-positive patients with liver disease, however, steroids plus azathioprine may be beneficial. This may also apply in less active disease,⁹⁸ especially if associated immunoserologic markers are present.^{110,111}

An uncontrolled trial of prednisolone and azathioprine in patients with HCV-hepatitis, and of cyclophosphamide in chimpanzees suggested some beneficial effect in HCV-CAH.¹¹²⁻¹¹⁴

Preliminary data indicate that treatment of HCV-hepatitis by immunostimulation with alpha interferon may be beneficial.^{115,116} It is possible that a small subgroup of patients with severe HCV-hepatitis, but without active viral replication, benefit from corticosteroids, as do some anti-HBe positive patients in HBV-hepatitis. Recently the genome of a specific HCV-antigen has been cloned¹¹⁷ and an assay for circulating antibodies to the antigen of this toga- or flavivirus will become available.¹¹⁸ Thus future trials will probably indicate the best strategy in HCV-CAH.

Treatment options for viral CAH have been summarized recently.^{119,120}

Other treatment options for non-viral CAH

Alternate-day steroids were originally tested by suddenly lowering the dose of steroids to 5 mg every other day after 3 months of treatment, and treating biochemical relapses by increasing therapy only on alternate days.⁸⁶ The present method for treatment with alternate-day steroids in other diseases is to double the dose on one day and then gradually reduce the other day's dose to zero. This approach has not been fully tested in CAH and may need examination, for example in patients with osteoporosis.¹²¹

Pulse prednisone therapy with high-dose prednisone (90 mg daily) given in 3- to 5-day courses repeated at 4-week intervals can inhibit B-cell synthesis of immunoglobulin G

and reduce inflammatory activity. Experience with this regimen is limited, however,^{122,123} and it was recently reported that such pulse therapy failed to control the symptoms and biochemical findings of relapse in severe CAH, and is less effective than combination therapy with 10 mg prednisone and 50 mg azathioprine daily.¹²³

Cyclos porine (CsA) was effective in some young patients with 'autoimmune' CAH,^{124,125} and may be tried in a clinical setting if high-dose combination-therapy with prednisolone and azathioprine fails, or if contraindications to the use of corticosteroids exist (e.g. severe growth failure in children). Close monitoring, especially of renal function and CsA blood levels will be mandatory, and will probably preclude its routine use in the long term, although further evaluation of CsA therapy in corticosteroid-unresponsive CAH may be justified. A trial with cyclophosphamide yielded disappoint-ing results.¹²⁶

Cytoprotective therapies like polyunsaturated phosphatidyl choline or arginine thiazolidinecarboxylate suggested some effectiveness in limited trials, but the risks and advantages of such treatment are uncertain.^{127,128}

Experience with *antifibrotic therapy* (other than corticosteroids and penicillamine) in liver disease is limited. Almost every report on D-penicillamine in CAH has demonstrated some efficacy, and it was used for several years. Its high toxicity and lack of superiority over prednisolone, however, did not allow this treatment to be an option.¹²⁹ Colchicine appeared effective in a heterogeneous group of patients with liver disease,¹³⁰ also in primary biliary cirrhosis,¹³¹ although it probably does not have a direct inhibitory effect on collagen synthesis. Other promising drugs for the treatment of hepatic fibrosis, like 2-oxoglutarate analogs and prostaglandins are currently tested *i n vitro* and in animal models.¹³² The value of antifibrotic drugs in CAH remains to be established. Since ursodeoxycholic acid appears an effective therapy for PBC,¹³³ the rare patients with cholestatic CAH ('CAH/PBC overlap') might benefit from such therapy if a therapy trial with corticosteroids¹³⁴ is not, or only partially, effective. However, these treatment options require further investigation.

The value of bed rest in CAH

This item remains controversial. Some studies report that exercise does not have any harmful effect on the course of hepatitis, even in the acute phase,¹³⁵ or in patients with CAH in remission while receiving immunosuppressive therapy.¹³⁶ This confirms earlier observations.^{137,138} Others, however, report deterioration of patients with CAH after exercise.¹³⁹⁻¹⁴¹ Currently bedrest is advised only during active disease.

Liver transplantation in CAH

This item will be discussed in the section on therapy failure.

§8 PROGNOSIS OF TREATED CAH

Corticosteroid therapy induces remission in 60 percent of patients with severe autoimmune CAH within 3 years and 5-year survival is approximately 90 percent.⁵¹ The cumulative remission rate for therapy of IAI-CAH with a combination of fixed dose prednisolone and azathioprine is 90%.⁸⁷ From 151 treated HBsAg-negative patients, 92% survived 5 years and 86% survived 10 years, while in 22 HBsAg-positive (65% also HBeAg-positive) patients, survival was lower. 73% and 65% at the same intervals (p<0.01). While the survival curve in treated HBsAg-negative CAH declines gradually, highest mortality in HBsAg-positive CAH occurred within the first 2 to 3 years and was quite low thereafter.¹¹⁰ In retrospect, several patients with severe HBsAg-positive CAH who entered the Mayo study had in all likelihood an exacerbation of CAH in relation to 'immune clearance' of virally infected hepatocytes.¹¹⁰ These results underline what has already been mentioned, namely that the indication for immunosuppressive therapy in viral CAH is very limited, while this therapy leads to a marked enhancement of survival in non-viral (idiopathic 'autoimmune') CAH.

Determinants of prognosis in treated CAH

- Prognostic value of histology:

In 1964 Page *et al.* demonstrated that patients without cirrhosis had the greatest chance of improvement during immunosuppressive therapy.⁶⁰ The presence of bridging necrosis (moderate CAH) or multilobular necrosis (severe CAH) in the initial biopsy was associated with a worse prognosis than periportal hepatitis in the absence of these features (minimal CAH).⁶¹ In fact, some authors even state that the sole presence of periportal hepatitis is a benign condition. In the Mayo experience, the 10-year probability of survival without hepatic failure is 98 percent for patients without cirrhosis at presentation. For patients with cirrhosis prior to treatment, the likelihood of survival without hepatic failure is 80 percent after 5 years and 65 percent after 10 years.^{142,143} Interestingly, patients who develop cirrhosis during therapy have a 5-year survival after the development of cirrhosis that is similar to that of patients who do not develop cirrhosis,¹⁴⁴ indicating that the propensity to develop cirrhosis in IAI-CAH depends more on the rapidity and completeness of the response to treatment than on any histologic feature at presentation.¹⁴⁴ This observation suggests that the ability to suppress inflammatory activity may be the most important prognostic factor.

- Prognostic value of markers for 'autoimmunity':

Page *et al.* also observed that patients with an acute onset of disease, evidence of multisystemic involvement, the LE cell phenomenon, and extreme hypergammaglobulinaemia were more likely to respond to corticosteroids than others.⁶⁰ Later on it was demonstrated, however, that the presence or absence of autoimmune markers does not influence the response to corticosteroids in patients with severe CAH, screened for drug-induced or virus-related disease. The frequency of remission, treatment failure, hepatic death, and progression to cirrhosis were similar in patients with and without the LE cell phenomenon or antinuclear antibody (ANA). Therefore the same program of management for all patients with idiopathic, presumably autoimmune disease is recommended, regardless of immunologic findings, emphasizing that treatment decisions be based on clinical, biochemical and histologic assessments of disease severity.¹⁴⁵ Patients with LE cell or ANA who entered remission, however, were less likely to relapse after medication was discontinued, suggesting that corticosteroids had a greater ability to disrupt the underlying pathogenetic mechanisms in these patients. However, death from hepatic failure (4 versus 12 percent) and 5-year survival (92 versus 87 percent) were not statistically different between the two groups with and without LE cell or ANA.¹⁴⁵

- To conclude, the degree of inflammation will determine the extent of liver failure and is connected with the induction of cirrhosis and its complications. The inflammation influences hepatocyte-function via cytokines and causes cell-necrosis and hence reduction of the number of functional hepatocytes. The duration of the hepatitis will especially influence prognosis via these mechanisms. This means that in idiopathic 'autoimmune' CAH, the initial reaction to immunosuppressive therapy must be the most important prognostic factor. Indeed, a recent study confirmed that patients who resolved at least one pretreatment laboratory abnormality, improved a pretreatment hyperbilirubinaemia, or did not experience biochemical deterioration after two weeks of corticosteroid therapy (with or without azathioprine) survived for at least 6 months in 98 percent of instances.¹⁴⁶

§9 IMMUNOLOGY OF IAI-CAH

Pathogenetic mechanisms leading to CAH and 'CAH-related liver disease' can be diverse,¹⁴⁷ as has been pointed out earlier. In IAI-CAH, there is considerable evidence that cytotoxic reactions of the antibody dependant (ADCC-)type are directed at hepatocyte plasma membrane-derived epitopes in the macromolecular, lipid-associated complex called 'liver specific membrane lipoprotein' (LSP).^{148,149} In 1972, Meyer zum Büschenfelde *et al.* reported the finding that rabbits repeatedly immunized with a high molecular weight fraction (LP1) of normal human liver developed liver lesions remarkably similar to those seen in CAH in man.¹⁵⁰ LP1 appeared to be a lipoprotein containing a liver-specific antigen derived from the hepatocellular plasma membrane, and consequently this fraction became known as liver-specific membrane lipoprotein (LSP).¹⁵¹ Lymphocytes from patients with a variety of acute and chronic liver diseases are able to 'kill' heterologous¹⁵²⁻¹⁵⁶ or autologous¹⁵⁷⁻¹⁶⁰ hepatocytes, or LSP-coated

avian erythrocytes¹⁶¹ *in vitro*. These cytotoxic reactions are apparently directed at antigens in LSP, while cytotoxicity is blocked by addition of LSP.¹⁶⁰ The hepatic asialoglycoprotein receptor (ASGR) is an important constituent of LSP.¹⁶² This is a liver-specific, species cross-reactive hepatic receptor, also known as hepatic lectin. Anti-LSP antibodies appear to be predominantly directed at the ASGR, although a role for anti-LSP directed at other antigenic specificities has not yet been excluded. Anti-LSP is usually present in IAI-CAH patients requiring immunosuppressive therapy according to accepted criteria, and anti-ASGR is found in many of these anti-LSP positive CAH-patients.¹⁶³ It has also been shown that anti-ASGR antibodies preferentially coat periportal hepatocytes in the ante- and retrogradely perfused rat liver,¹⁶⁴ which is consistent with the concept of ADCC.

Composition of LSP

At present LSP is perhaps most accurately described as a macromolecular, lipidassociated complex containing a number of hepatocyte plasma membrane-derived antigens. Some of these antigens are non-organ-specific and others are liver-specific (including at least one that is species-specific and at least one other that is species crossreactive).¹⁶⁵ There is now convincing evidence^{166,167} indicating that LSP is composed to a large extent of vesicles (40-1600 nm diameter) and other fragments of the hepatocellular plasma membrane. It also contains some cytoplasmic fragments,168 Furthermore, it has been demonstrated that LSP contains liver-specific antigens and non-specific determinants. Both of these are localized in the low-density subfraction after CsCl density gradient centrifugation, using both heterologous and autologous sera.¹⁶⁹ Attempts have been made to employ hybridoma techniques to identify antigens in LSP170-174 and one monoclonal antibody has been produced¹⁷¹ that reacts with a liverspecific, species-specific, plasma membrane antigen in rabbit LSP. However, it might eventually be proved that liver-specific but species cross-reactive determinants are the most important target antigens in CAH. The molecular weights of four LSPdeterminants, recognized by monoclonal antibodies, in the study of Wiedmann et al.¹⁷² ranged from 22 kiloDalton (kD) to 164 kD. LSP contains lecithin, lysolecithin, sphingomyelin, phosphatidyl ethanolamine, fatty acids, and cholesterol. SDS-PAGE of LSP from man, cattle, and rabbits shows essentially identical patterns in each, with four main bands corresponding to molecular weights of 50, 55, 60 and 66 kD, and a number of minor bands of 29 to 350 kD.¹⁷⁵ In addition it is now known that LSP contains the ASGR, as has been mentioned above.¹⁶². This lectin was found to comprise only 0.25%of the total protein in LSP, which may explain why it is difficult to demonstrate liverspecific LSP components by such techniques as sodium dodecyl sulphate/ polyacrylamide (SDS-PAGE)-gel electrophoresis.¹⁷⁶

Regulation of the immune response in IAI-CAH

Con A induced non-specific T-suppressor cell activity in monocytes from blood of patients with IAI-CAH or HBV-CAH is impaired.177-180 If IAI-CAH is treated with corticosteroids, this T-suppressor activity normalizes¹⁸¹. Some authors doubt whether this T-suppressor defect is antigen-specific or not. If the monocytes isolated from IAI-CAH patients are preincubated with prednisolone, T-suppressor function improves, while the function of T-suppressor cells from HBV-CAH patients does not improve on addition of corticosteroids.182 This may indicate that the T-suppressor defect in IAI-CAH and HBV-CAH may be of a different nature, and it reflects the difference in response to corticosteroids between these diseases. In the indirect migration inhibition test, monocytes of an IAI-CAH patient produce T-lymphocyte migration inhibition factors in the presence of LSP. These factors were absent if these cells were cultured in the presence of T-cells of healthy donors or HBV-CAH patients. These inhibitory factors were also absent in the same assay with monocytes derived from HBV-CAH patients. This implies a defect in the LSP-specific suppressor T-cell function in IAI-CAH,¹⁸³ and recent studies indicate that the basal defect probably lies in the inducers of these suppressor cells.¹⁸⁴ These observations were supported by findings in thymectomized mice with experimental IAI-CAH.185

In addition to triggering factors, a genetic predisposition appears a prerequisite for the development of IAI-CAH. The histocompatibility antigens A1, B8 and D3 and the GM-ax allotype marker are all more frequent in IAI-CAH.¹⁸⁶⁻¹⁹⁰ Relatives of patients with IAI-CAH show an increased incidence of serological abnormalities, various autoimmune diseases, and suppressor T-cell defects.^{184,191} Whether these suppressor T-cell defects in relatives, linked with HLA-type, are only non-antigen specific or also antigen specific is still uncertain.

To summarize^{192,193} in IAI-CAH a genetically determined defect exists in T-cell inducers of antigen-specific T-suppressor lymphocytes. It is possible that these patients are provoked into ADCC by LSP, or more specifically, the ASGR. Corticosteroids can improve the non-antigen specific T-suppressor cell functions in IAI-CAH, resulting in reversal of histological and biochemical manifestations of disease. This is sufficient to induce 'remission' according to currently accepted criteria, but from the King's College's experience it appears that corticosteroids have no effect on the genetically determined LSP-specific T-suppressor cell defect. This latter defect could allow for continued stimulation of anti-LSP producing B lymphocytes.

No defects in the secretion of interleukins and interpherons have been reported to play a role in IAI-CAH. This is unlike chronic HBV-infection.¹⁸⁴⁻¹⁸⁶ A number of immuno-regulatory factors have been identified in human liver and sera, including liver-derived inhibitory protein (LIP),¹⁹⁷⁻¹⁹⁹ rosette inhibitory factor (RIF),²⁰⁰⁻²⁰³ serum inhibition factors (SIF),²⁰⁴⁻²⁰⁶ and an immunoregulatory subset of low-density lipoprotein (LDL)

which has been designated as LDL-In.²⁰⁷⁻²⁰⁹ The functions of these factors have been reviewed recently.²¹⁰ It is not currently known whether altered secretion of these factors might be pathogenetically involved in CAH.

Effector mechanisms of the immune response in CAH

Evidence for cell-mediated immunity to LSP has been found in 26 out of 29 patients with IAI-CAH, but in only one out of 21 patients with HBV-CAH.¹⁸³ The presence of cytotoxic T-cells and natural killer cells (Pit cells) in contact with hepatocytes in the periportal inflammatory infiltrate of CAH has been demonstrated,²¹¹⁻²¹⁴ suggesting a pathogenetic role for these cells in both IAI-CAH, HBV-CAH and HCV-CAH. Cytotoxicity in CAH shows antigen-¹⁵² and target cell²¹⁵ specificity. Although direct evidence for the role of these effector cells *in vivo* is lacking at present, indirect evidence for their role is abundant in *in vitro* experiments.^{153,216-225} Unfortunately many of the *in vitro* studies have been plagued by methodological difficulties, such as the small numbers of autologous cells available for testing, the problems related to different methods for evaluating target cell death, and the failure in some studies to allow for the constraints of HLA restriction when testing for T-cell cytotoxicity.²²⁶ The properties of these mononuclear cells *in vitro* do not necessarily reflect those of the cells in the inflammatory infiltrate of the liver, of course.

To summarize, CAH can be the final common pathway of several aetiologies, and partly shared pathogenetic mechanisms. Cytotoxicity for autologous hepatocytes by mononuclear cells from the peripheral blood has been found in both viral-associated and idiopathic autoimmune CAH. Cytotoxicity mediated by lymphokine-activated killer (LAK)-cells directed to LSP (probably of the ADCC type) has been shown in most patients with IAI-CAH and HBV-CAH. The main type of cytotoxicity in HBV-CAH is an attack by cytotoxic T-cells (CTL), possibly directed to HBcAg and HBeAg on the membrane of hepatocytes.

Putative role for exogenic factors in triggering IAI-CAH

Besides a genetically determined immunological abnormality, an exogenous factor may be necessary to induce 'autoimmune' CAH. Increased antibody titres to exogenous antigens, notably bacterial and viral, were reported.^{227,228} High antibody titres against gut-derived bacteria such as E. Coli, Bacterioides, and Salmonella can occur.^{227,229} Elevated antibody titres were described for measles, rubella²³⁰⁻²³³ and cytomegalovirus (CMV).²³³ Elevated antibody titres to CMV early antigen²³⁴ were also described in oxyphenisatin-induced CAH.²³⁵ Anti-measles antibodies in CAH can belong to the IgM-class of antibodies,²³⁶ suggesting persistent production of measles virus antigens. Recently, persistent measles virus genome was identified in the lymphocytes from 12/18 patients with IAI-CAH, in contrast to 1/45 controls.²³⁷ This finding correlated strongly with the presence of high antibody titres to measles.

These findings suggest the possibility that viruses and bacteria may be involved in the aetiology and pathogenesis of IAI-CAH.

Other authors postulated as intrinsic defect caused by spontaneous mutation of immunocytes. A third theory involves as environmental toxin. This theory was - like the infections theory - supported by the presence of autoantibodies in the sera of spouses of CAH-patients.

§10 PROBLEMS IN THE DIAGNOSIS AND MANAGEMENT OF CHRONIC ACTIVE HEPATITIS

These have been extensively dealt with by Czaja.²³⁸ The most important problems he mentions are:

1. Determining chronicity

No reliable methods exist to determine whether hepatitis will follow a chronic course or not, even when a histology of CAH is present.²³⁸ Spontaneous improvement of CAH can occur, as already mentioned. Determination of chronicity is necessary to prevent administration of potentially toxic or unnecessary medication. Only a subgroup of CAH-patients requires immunosuppressive therapy, while institution of such therapy in other CAH-subgroups can be contra-indicated, as discussed. Therefore, duration of illness was relied on as a criterion of chronicity.

Continuation of hepatitis for at least 10 weeks without clinical or biochemical improvement excludes self-limited acute hepatitis in most instances and suggests chronicity.²³⁹ Demonstration of ongoing activity for at least 6 months stresses further the unresolving nature of the disease and satisfies international criteria for chronic disease.^{34,36,240} However, morphologic features of acute viral infection in a patient with disease of more than 6 months duration can challenge classification. The presence of hypoalbuminaemia, hypergammaglobulinaemia, and ascites in a patient with disease of acute onset and of less than 6 months' duration may justify corticosteroid treatment, even though the disease is not fully chronic by international criteria. Initially, exclusion of a viral aetiology of hepatitis was difficult, because sensitive and specific markers were lacking. Now that sensitive and specific methods for detection of hepatitis B and recently- hepatitis C have been developed, known causes of CAH (including viral) can be excluded earlier in the disease, and with greater reliability. Immunosuppressive therapy might therefore, when indicated, be instituted earlier in a patient fulfilling criteria for the diagnosis of idiopathic 'autoimmune' CAH (duration excluded).

2. Establishing the diagnosis

Known causes of CAH and CAH-related liver disorders, as mentioned in table 1, must be excluded to allow diagnosis of idiopathic 'autoimmune' CAH.

3. Deciding to treat

Patients who most clearly deserve immunosuppressive therapy are those with IAI-CAH and sustained elevation of AST at least 10-fold normal or 5-fold increases in conjunction with at least twice-normal serum gamma-globulin concentrations.⁵⁰ There is a high mortality among these patients if untreated.⁵³ Presence of bridging or multilobular necrosis on liver biopsy also connotes severity of necrotic inflammation, and therefore motivates initiation of treatment.^{28,61} Serious clinical deterioration with development of ascites, endogenous encephalopathy, or other features of liver failure warrant therapeutic intervention.⁸⁹ The decision to initiate treatment in patients with mild to moderately active hepatitis (CAHmin and CAHa) is difficult. That disease of mild to moderate severity can progress to cirrhosis and liver failure is unquestioned: 33% of HBsAg-negative patients with CAHa progress to cirrhosis, and the 7-year mortality of these patients is 14%.^{68,69} In patients not fulfilling biochemical, histological or clinical criteria for severe IAI-CAH the potential benefits of treatment have to balanced against the risks of therapy and the likelihood of spontaneous resolving of the disease in the individual patient. Controlled clinical trials have not yet been conducted in this area. Immunosuppressive treatment of CAH of subgroups other than IAI-CAH, like anti-HBe positive patients, sometimes can be justified as has been discussed already.

4. Drug toxicity

Development of major steroid-induced side-effects is dose-related: Only 11% of patients receiving 10 mg/day of prednisolone developed major side effects after 24 months of therapy, while this percentage approximated 75% when the dose was higher than 20 mg/day.²⁴⁰ Azathioprine enables dose-reduction of corticosteroids.^{50,86} Corticosteroid-related cosmetic changes (truncal obesity, facial rounding, dorsal hump formation, acne or hirsutism) are frequent, weight gain in excess of 20 pounds, hypertension, diabetes mellitus, cataracts, osteoporosis²⁴² and severe skeletal complications (vertebral compression fractures and aseptic necrosis of the hip) can occur, while psychosis or serious infections are rare.⁸⁷ Peptic ulcer disease is no more frequent in patients with CAH treated with prednisone than in those receiving placebo,²⁴³ although peptic ulcer and gastritis are more frequent in CAH than in the general population, especially in those with portal hypertension.²⁴⁴ Diabetes mellitus is not only a complication of treatment, but is also more frequent in patients with untreated CAH than in the general population.^{245,246,247} Of all the complications, about 70% developed during the first

12 months.⁸⁷ Antacids significantly reduce prednisone bioavailability, and therefore should not be ingested simultaneously.²⁴⁸ Oral antidiabetic therapy should be used with caution in CAH, because these drugs can induce chronic active hepatitis.²⁴⁹ Calcium supplementation of 1 gram/day can reduce steroid-induced bone resorption without detectable suppression of bone formation.²⁵⁰ 25-hydroxy(OH)vitamin D administration can improve parameters of mineral and bone metabolism in steroid-induced osteoporosis,²⁵¹ but 1,25 (OH)² vitamin D appears ineffective.²⁵² Another treatment option for corticoid-induced osteoporosis may be the use of salmon calcitonin,²⁵³ while other investigators recently reported that administration of 150 mg 3-amino-1-hydroxy-propylidene-1,1-biphosphonate (APD) daily in 'enteric coated' form, in combination with calcium supplementation 1 gram/day effectively prevented development of steroid-induced osteoporosis, and even reversed it after development.²⁵⁴ Further studies would establish whether this therapy is effective in the long term prevention and therapy of steroid-induced osteoporosis, and whether it can be used safely in CAH.

Azathioprine toxicity occurs in fewer than 10% of patients receiving low-dose therapy (50 mg daily). It usually manifests early with hepatotoxicity, gastrointestinal upset, skin rashes, leukopaenia, and thrombocytopaenia and is reversed by reduction in dosage or discontinuation of medication.^{89,50} The risks of oncogeneity after low-dose azathioprine therapy are low but probably increased. What has been reported underlines the efficacy and safety of low-dose azathioprine, the benefits outweighing the long-term risks of extrahepatic neoplasm.^{87,255-257}

In patients for whom azathioprine has increased potential toxicity (children, patients with severe leukopaenia or severe thrombocytopaenia (e.g. $<50x10^9$ /l), during pregnancy) an initially high dose of steroids with a rapid taper to 20 mg daily has been recommended,²⁵⁸ although during pregnancy in CAH-patients no important side-effects could be attributed to azathioprine, as will be discussed.

5. Complications of CAH

The frequent development of cirrhosis has already been mentioned. Following institution of treatment development of ascites or encepahalopathy rarely occurs, and if it does this is an ominous prognostic sign.²⁵⁹ This probably also applies to progression of liver insufficiency despite therapy. The presence or development of oesofageal varices is less threatening in treated patients, however: Oesofageal varices bleed infrequently in treated CAH, and their presence appeared to be not associated with mortality. Bleeding from gastritis or gastric ulcer, however, is more likely in patients with varices than in others.²⁶⁰

If a sudden decompensation occurs despite compliance with the treatment regimen after long-standing stable disease, without recent changes in other medication, the possibility of presence of primary hepatocellular carcinoma should be considered.^{66,67}

Serious infection is an uncommon occurrence during standard maintenance therapy.⁸⁷ However, systemic mycosis can complicate high dose corticosteroid therapy given for treatment-failure,²⁶¹ and therefore this possibility should be considered if a patient does not improve despite such therapy.

6. Pregnancy in CAH

Many patients with IAI-CAH are in the child-bearing age. One retrospective analysis of pregnancy in CAH compared to pregnancy in normal (Australian) controls has been reported.²⁶² Pregnancy did not adversely influence the behaviour of maternal liver disease. A higher than expected rate of fetal loss and obstetric complications occurred, but the majority of deliveries were successful, and the babies normal. These findings suggest that conception is not absolutely contraindicated by the presence of CAH, but that close monitoring (especially obstetric) is necessary during pregnancy. Therapy during pregnancy has been discussed above.

7. When and how to stop therapy?

Criteria for discontinuing therapy in patients after induction of remission are arbitrarily defined. The current practice is to confirm histological remission by liver biopsy after a fixed period of sustained remission during therapy, e.g. two years, and then try to withdraw therapy in a period of variable length. Remission is defined arbitrarily as absence of clinical symptoms, biochemical resolution of AST to less than twofold normal, with normal bilirubin and gamma globulin, and histologic remission to a histology of chronic persistent hepatitis or complete resolution of the inflammatory changes, with or without cirrhosis. In some studies even minimal parenchymal inflammatory activity (grade 1 piecemeal necrosis) did not preclude classification as remission.^{50,86,263} Relapse occurs in 49-87 percent of patients within 6 months after remission and withdrawal or discontinuation of treatment, and the majority of patients who experience relapse do so within the first 3 months after termination of treatment.^{263,264,265} As will be discussed, it therefore will be necessary to find better endpoints of therapy and early indicators of relapse.

Intolerable cosmetic changes, weight gain of at least 40 pounds, inability to satisfactorily control diabetes mellitus or hypertension, osteoporosis with vertebral collapse, persistent severe cytopaenia, psychosis, or hypersensitivity skin eruption are current indications for dose modification or drug withdrawal. Some consider pregnancy, upper gastrointestinal bleeding of unknown origin or severe gastrointestinal upset also as indications for modification of therapy.¹⁴³

8. Managing the sub-optimal response of CAH to therapy.

Sub-optimal responses of CAH to therapy can be divided into:

- treatment failure
- slow response
- multiple relapses

Treatment failure

Treatment failure was defined as death or deterioration characterized by the development of hepatic encephalopathy or increasing ascites with a twofold increase in serum bilirubin to greater than 12 mg/dl.^{50,86} After 1971, the criteria were revised to include a 67% increase in bilirubin (if earlier >4mg%) or of SGOT (if earlier >59 IU/l).²⁵⁹ At present, patients requiring reassignment of therapy because of complications, or with a fatal outcome due to complications from therapy, are also classified as treatment failures.⁸⁷

Patients who fail standard treatment usually do so within 3 months after institution of therapy. They are more likely to have evidence of hepatic insufficiency (ascites or hypoprothrombinaemia)²⁵⁹ and histologic evidence of confluent necrosis and cirrhosis at presentation.⁶¹ Although the mean serum AST level is higher in patients who fail treatment than in those who do not,²⁵⁹ treatment failure is not more common in individual patients with marked serum aminotransferase elevations (>1000 IU/I).²⁶⁶ For patients meeting criteria for treatment failure, initial therapy with high-dose prednisolone (60 mg per day) or a combination of prednisolone 30 mg daily in conjunction with azathioprine 150 mg daily has been recommended, and the results after this high-dose treatment (81 percent survival) were better than previously observed.⁷⁹ The likelihood of subsequent sustained remission in these patients is low (25 percent) however,⁸⁸ and prolonged administration of high-dose corticosteroid therapy may carry an increased risk of side-effects. Liver transplantation may be the ultimate option for some of these patients.¹⁴³

Slow or incomplete response

Within 3 years of therapy 65% of patients enter remission, and 20% fail treatment. The other 15% of patients improve during therapy, but fail to satisfy remission criteria after this period of treatment.²⁶⁷ Continuation of treatment beyond 3 years in these patients is associated with a per annum probability of remission of only 7 percent, while the risk of side effects accrues at a similar rate.²⁶⁷ The current policy in these patients is to administer conventional maintenance doses of medication until early evidence of drug intolerance.

Relapse

Relapse is defined as return of symptoms, a threefold or greater elevation of serum AST level or increase in serum gamma globulin concentration to at least 2 gram/100 ml, and histologic evidence of moderate to severe CAH (CAHa or CAHb).44,50,263 Relapse occurs in 49-87 percent of patients within 6 months after remission and during or after discontinuation of treatment. The majority of patients who experience relapse do so within the first 3 months after termination of treatment,²⁶³⁻²⁶⁵ Although relapse is usually accompanied by symptoms like fatigue, arthralgias, pruritus, anorexia, and abdominal pain, one-quarter of patients will have no clinical symptoms to signal recurrence of inflammatory activity.²⁶³ It has been found that elevation of the AST level to at least threefold the upper reference limit during or after drug-withdrawal is invariably associated with histologic findings of CAH and indicates relapse.²⁶⁸ The histologic pattern and degree of inflammation are related to the length of period between the onset of relapse and diagnosis, and may differ from the initial findings before therapy.⁴⁴ Patients who develop cirrhosis during therapy regularly relapse (85 to 90 percent), but patients with cirrhosis prior to treatment are as eligible for sustained remission after treatment as patients without cirrhosis before institution of therapy.^{143,144,263} At present no other features predict the likelihood of relapse after treatment. Recently in a trial of treatment withdrawal the appearance of antibodies to liver-specific membrane lipoprotein (LSP) (with the asialoglycoprotein-receptor as a major antigenic determinant) preceded (in 15 out of 22 patients) or occurred concurrent with (in one patient) biochemical and histologic relapse. All 6 patients who sustained their remission remained anti-LSP negative.²⁶⁹ If these results can be confirmed, testing for the presence of anti-LSP antibodies might (partially) obviate the need for sequential liver biopsies, and provide a more appropriate endpoint of therapy. In patients experiencing relapse, the pathogenetic mechanisms of the disease are probably not, or not permanently, disrupted and/or the inflammatory activity inadequately suppressed. Indeed, relapse occurs in only 28 percent of patients who revert to normal liver tissue as compared to a relapse frequency of 76 percent in patients with residual portal hepatitis at the time of drug-withdrawal.²⁷⁰ In patients with cirrhosis whose inflammatory activity ceased, the frequency of relapse is still 79 percent, consistent with our knowledge that many patients develop cirrhosis after diagnosis of CAH, especially during the first two years of treatment when hepatic inflammation is most severe.^{143,144} Once relapse has occurred, the probability of re-entering remission with successive courses of corticosteroid treatment remains similar to that experienced during initial therapy.²⁶³ The likelihood of relapse within 6 months after a second course of treatment, however, increases from 49 percent to 60 percent, and after a third course becomes 86 percent.^{143,263} 27 percent of patients followed longitudinally after remission will experience multiple (three or more) relapses.²⁶³ Currently, therapy is reinstituted after each relapse until remission, treatment failure, or drug-intolerance necessitates treatment adjustments and improvisations.¹⁴³

Liver transplantation in CAH

Orthotopic liver transplantation (OLT) may be the ultimate option for patients failing to respond to treatment of IAI-CAH.¹⁴³ Current criteria for determination of prognosis may not be entirely adequate, however.²⁷¹⁻²⁷⁴ Development of new survival models for decision making in these patients may be required (similar to PBC).^{275,275a} It is felt that transplantation should be offered before the patient develops preterminal liver insufficiency, severe coagulopathy, variceal haemorrhage or hepatorenal syndrome and before the patient becomes severely catabolic. The odds of surviving transplantation have to be weighed against the probability of surviving the liver disease in every individual patient.²⁷⁶ As has been mentioned already, most patients with IAI-CAH respond to immunosuppressive therapy, and do not need liver transplantation. The patient who, despite optimal immunosuppressive therapy, develops decompensated liver disease with spontaneous encephalopathy, progressive jaundice, diuretic-resistant ascites, uncorrectable and worsening coagulopathy, repeated uncontrollable variceal haemorrhage, or recurrent septicaemia ought to be considered as a transplantation candidate.¹²¹ Recurrence of IAI-CAH after OLT has been reported once.²⁷⁷ Partial heterotopic 'auxiliary' liver transplantation has been reported to be an alternative option in high-risk patients with end-stage chronic liver disease.²⁷⁸

§11 MONITORING ACTIVITY OF DISEASE AND LIVER FUNCTION IN CAH

Liver biopsy

In a liver biopsy of sufficient size^{279,280} the observer variability in scoring the degree of hepatitis in CAH is small with a 94% consistency rate for overall morphologic interpretation. Inter- and intraobserver variability in grading the degree of piecemeal necrosis in other forms of liver disease is greater than 50 and 40%, respectively²⁸¹.

The diagnosis of cirrhosis by needle biopsy is dependant on the different criteria for diagnosing cirrhosis histologically. When complete regeneration nodules are required for diagnosis, false negative results are common. In many instances only a tentative diagnosis of cirrhosis can be made on the base of fragmentation, liver cell appearance, parenchymal regeneration, and fibrosis.^{40,41} The combination of liver biopsy with laparoscopy increases the accuracy of diagnosing cirrhosis.²⁸²

However, CAH is defined principally by its activity rather than by structural changes. The classifications of the degree of severity of CAH are variable. The recommendations by an international group are still widely used.^{31,40,42} Activity may be variable from location to location in one and the same biopsy, and the most severely affected area will

determine the degree of activity reported. Sample error in CAH can be less than 10 percent when scoring the severity of inflammation.²⁸³ Recently, Knodell *et al.* defined a numerical scoring system to assess histological activity in asymptomatic chronic active hepatitis.²⁸⁴

Laboratory testing

Liver morphology in CAH has to be considered in combination with clinical and biochemical findings in order to enable therapeutic decision-making.²⁸⁵ Although it is a minor procedure, a liver biopsy is still invasive and has its own -low- morbidity and mortality.²⁸⁶ This precludes its use for frequent routine monitoring of activity of disease. Furthermore, morphology does not reflect hepatic functional capacity.

Standard tests

Aspartate aminotransferase (AST) and gamma globulin (GG) to some extent reflect hepatocellular necro-inflammation.²⁸⁷⁻²⁹³ Currently, these two routine tests are used to determine sufficient severity of disease to require treatment,⁸⁹ help to assess response to treatment,^{89,268} define treatment failure,^{259,266} resolution of disease,^{44,263,270} and disease relapse.^{44,89,95,96,263} However, the accuracy in predicting morphologic activity of CAH is limited, especially for normal values of AST and/or GG when using standard reference ranges.²⁶⁸ In addition, these tests do not reflect liver function, and they are of no great help in differential diagnosis of CAH.

Data is limited on changes in functional routine laboratory parameters like albumin, coagulation parameters, and water- and salt homeostasis, in relation to activity of disease and hepatic function.

No single clinical or standard laboratory parameter such as serum albumin, prothrombin time or platelet count, was able to predict the presence of cirrhosis. The combination of such parameters, however, increased sensitivity and specificity.²⁹⁴

Specialized tests

Serum amino acids

Specific plasma acid patterns have been described in patients with chronic liver disease,²⁹⁵⁻³⁰² including CAH.^{299,303,304} Elevation of the aromatic amino acids (AAA) phenylalanine, tyrosine, and methionine, with concomitant reduction of the branched-chain amino acids (BCAA) valine, leucine, and isoleucine appear in cirrhosis. These

abnormalities have been explained by imbalances between hepatic clearance and peripheral utilization of the amino acids.^{305,306} The BCAA are decreased in chronic liver disease due to increased peripheral oxidation,³⁰⁵ increased ketogenesis,³⁰⁷ increased renal gluconeogenesis,³⁰⁸ increased distribution volume,³⁰⁹ and/or increased intracellular distribution, ^{310,311} while AAA are increased due to enhanced proteolysis ^{312,313} and altered hepatic metabolism.³¹⁴⁻³¹⁶ In several studies the molar ratio of BCAA to AAA levels has been shown to correlate with severity of inflammation in hepatic histology,^{298,300,318,319} liver function,^{297,302,320,321} and prognosis.^{298,301,319} However, prospective studies did not confirm the high predictive value of the BCAA/AAA ratio for these parameters.²⁹⁷ These differences in results may in part be explained by the many other conditions that influence the BCAA/AAA ratio, like portal systemic shunting,³²² nutritional status,³²³⁻³²⁵ alcohol use,³²⁵ and gender.³²⁶ A normal plasma molar ratio, however, is of value in excluding the presence of cirrhosis.^{295,298,300,317}

Serum bile acids

The same problems in interpretation are encountered when assessing the usefulness of serum bile acids (SBA) in the evaluation of liver disease. Results are influenced by the analytical method used, gallbladder contraction, digestive and interdigestive gastrointestinal motility, enterocyte absorption, hepatocyte clearance, and the function of the sphincter of Oddi and iliocecal valve. Only competitive binding radioimmuno-assays^{327,328} and heterogeneous³²⁹ and homogeneous³³⁰ enzyme assays have been shown to be sufficiently sensitive to accurately measure SBA concentrations.

Some studies suggest that SBA determinations correlate well with severity of histologically active liver disease in CAH-patients.^{331,332} Fasting SBA levels have been reported to discriminate active from inactive CAH equally or even better than did postprandial SBA-levels³³³⁻³³⁵ and were shown to be superior to conventional liver tests in this respect.³³⁴⁻³³⁹ Hepatic clearance of SBA in liver disease is dependant upon portosystemic shunting as well as hepatic function.³⁴⁰ SBA levels appear to correlate better with quantitative liver function tests such as bromosulfophtalein and indocyanine green clearance tests³⁴¹ and the aminopyrine breath tests,³⁴² than with standard liver tests. These correlations explain the described ability of SBA levels to predict mortality risks in cirrhosis.³⁴³ SBA levels might thus prove useful as one of the parameters in the selection of patients for liver transplantation,³⁴⁴ despite the many factors that influence SBA levels. However, these studies need further confirmation.

Quantitative measurements of hepatic function

Aminopyrine breath test

Like SBA, the aminopyrine breath test (ABT)³⁴⁵ is an infrequently used quantitative measure of hepatic function.³⁴⁶⁻³⁴⁸ It measures the combination of elimination and clearance rates,³⁴⁹ and as such is influenced by the functional microsomal mass, and

complex biochemical reactions that affect CO₂ formation and elimination. These are influenced by alterations in levels of folic acid, vitamin B₁₂, reduced glutathione, and NAD/NADH redox status. The ABT is difficult to standardize, because each of these factors may or may not be independantly impaired during liver disease, and the ABT is also influenced by such factors as gastric emptying³⁵⁰ and patient age.³⁵¹ It is a relatively easy and non-invasive test, however.^{350,352} The ABT has been shown to be abnormal in non-cholestatic liver diseases, particularly cirrhosis.^{346,350-356} The ABT reflects the severity of inflammation in chronic hepatitis, but a significant overlap between mild and severe CAH and between controls and patients with mild liver disease limits its use in this respect.^{342,356,357} In addition some investigators reported that ABT was no better than^{352,356} or worse than³⁵⁸ standard liver tests in predicting histologic severity. ABT was reported to discriminate between CAH and PBC,^{359,360} which might be of use in CAH with prominent cholestatic features. There is some evidence that the ABT may have prognostic value in predicting mortality³⁶¹ prior to surgery and it may aid in the selection of patients for liver transplantation.

Other tests of function

The hepatic extraction fraction of many substances can be used as a measure of liver function. Galactose elimination clearance (GEC) has the advantage that it is less influenced by disturbing variables than some of the other tests. However, especially the second phase of galactose elimination varies with variations in the hepatic flow rate. The place of GEC in diagnosis and follow-up of liver diseases like CAH remains to be established.^{361a}

Portal pressure measurements

Portal pressure measurements can probably only give insight into the degree of portal hypertension, but there is no reason to assume that portal pressure reflects histologic activity.²⁸⁵

Procollagen peptides

Fibrosis and formation of cirrhosis are sequelae of untreated CAH,^{362,363} and can develop during therapy.^{364,365} About 70% of liver collagens consist of collagen types I and III.³⁶⁶ Collagen type III is predominant in early fibrosis; in late cirrhosis, type I, hard collagen" prevails.³⁶⁶ Type III collagen has a more rapid turnover than type I collagen.³⁶⁷ The C- and the N-terminal procollagen-III-peptide (PIIINP) are split from the procollagen-III molecules before extracellular assemblage into fibers in the extracellular matrix. The amount of free PIIINP that circulates predominantly stochiometrically reflects deposition of collagen-III.³⁶⁸⁻³⁷⁰ Some of the serum PIIINP is derived from PIIINP which was retained in the deposited fibers.^{371,372} Decreased degradation of PIIINP due to impaired liver function can account for a rise in serum

PIIINP in the anhepatic situation during liver transplantation.³⁷³ However, these two factors probably do not materially affect the relationship of serum PIIINP with deposition of collagen type III in the liver during CAH. Some studies have indicated that serum PIIINP parallels the activity of inflammation in acute hepatitis, and returns to normal with transaminases and bilirubin³⁷⁴ and in alcoholic hepatitis PIIINP correlated well with periportal and intralobular inflammation.³⁷⁵⁻³⁷⁹ Several studies suggested a relation between activity of CAH and PIIINP,³⁸⁰⁻³⁸³ although this could be masked in children with CAH by elevated PIIINP due to growth.³⁸⁴ Persistent elevation of PIIINP in serum of patients with viral hepatitis for more than 6.5 months correlated with chronicity.^{374,385} Therapy with corticosteroids can lower an elevated PIIINP in CAH,³⁸⁶⁻³⁹⁰ and recently it was shown that PIIINP changed in accordance with other liver tests and normalized when remission of CAH had been achieved.^{391,392}

Autoimmune phenomena

Anti-LSP antibodies

It was reported that patients with IAI-CAH requiring immunosuppressive therapy invariably have anti-LSP antibodies,^{393,394} as already discussed above. In several studies, titres of anti-LSP antibodies were shown to correlate with presence and severity of periportal inflammation in CAH.^{393,395,396} Moreover, in a trial of treatment withdrawal, patients without anti-LSP who remained negative did not relapse, while patients with these antibodies or in whom anti-LSP reappeared, relapsed.²⁶⁹ Therefore it appears that testing for anti-LSP could be important in monitoring of CAH.

Other immunological phenomena

Several autoantibodies, like antinuclear antibodies (ANA), antibodies to liver membrane antigens (LMA), anti smooth muscle antibodies (SMA), anti soluble liver antigen (SLA), anti liver-kidney microsomal antibodies (LKMI), and other antibodies, including those against exogeneous agents like measles and cytomegalovirus, have been identified in IAI-CAH and CAH of other aetiologies. The value of many other tests like the concentration of circulating interleukin-2 (IL-2) receptors,³⁹⁷ and tests for measuring cytotoxicity and T-suppressor function for routine monitoring of IAI-CAH remains to be established.

To conclude, there is great need for non-invasive parameters for following the response to therapy, determining the optimum duration of treatment, for monitoring and predicting disease activity after cessation of therapy, and for help in differential diagnosis of CAH.

More information on long-term prognosis in CAH is needed, and data on accurate assessment of early response to treatment are scarce.¹⁴⁶ Elucidation of the aetiologies of idiopathic 'autoimmune' CAH may lead to more specific therapies.

Section 1

Patients and Methods

CHAPTER 1

PATIENTS

Between January 1969 and January 1988, a histological diagnosis of chronic active hepatitis (CAH)³⁹⁸ was made in 186 patients referred to the division of Hepatology at Groningen University Hospital, a tertiary care center. These are the patients studied in this thesis. During this period sera were frozen and stored at -20°C on all examinations and admissions, and the records of all liver biopsies and revisions of liver biopsies were kept systematically. The 186 CAH-patients were found by reviewing all pathological descriptions of liver biopsies from this period, with continued follow-up, and by reviewing our own patient database and the computerarchives of the hospital. Patient records were reviewed and, where necessary, liver biopsies were reviewed. Patients with chronic persistent hepatitis (CPH), but no biopsy with CAH, were excluded, as were patients where a probable diagnosis of CAH was made without taking a liver biopsy. All patients with cryptogenic cirrhosis and no biopsy with CAH were excluded,

Actiology*	Number of patients	S N	ex 1:F	
IAI-CAH	69	12:	57	
anti-HBsCAH	13	3:	10	
HBsAgCAH	51	37:	14	
HCV-CAH	10	4:	6	
DI-CAH	20	3:	17	
Other CAH	23	6:	17	
Total	186	65:	121	
ad *)				
IAI-CAH	= idiopathic	'autoimmune' (CAH	
HBsAg CAH	= CAH with	circulating HB	lsAg	

Table 1.1	CAH-patients	grouped	according	to aetiology	y.
-----------	---------------------	---------	-----------	--------------	----

IAI-CAH	=	idiopathic 'autoimmune' CAH
HBsAg CAH	=	CAH with circulating HBsAg
Anti-HBs CAH	=	CAH with circulating anti-HBs
HCV-CAH	=	CAH due to parenteral non-A, non-B hepatitis (probably due to
		hepatitis C virus)
DI-CAH	=	drug-induced 'CAH'

Other CAH:

9x CAH/PBC= 'cholestatic CAH' or 'mixed form'CAH/PBC

5x CAH/PSC='small-duct' PSC and/or CAH

3x Wilson-CAH= 'CAH' associated with Wilson's disease

5x A1AT-CAH= 'CAH' related to α_1 -antitrypsin deficiency

1x 'CAH' in multicystic liver

even if serology, clinical findings or reaction to corticosteroids suggested CAH. Furthermore, patients with acute hepatitis B and a histology of CAH were only included if hepatitis perpetuated for more than six months after the onset of disease. Patients with "CAH-related disorders" and a histology indistinguishable from CAH, e.g. α_1 antitrypsin deficiency + CAH (see introduction), were included. Classification as to aetiology and age, sex and therapy of the patients is shown in tables 1.1, 1.2 and 1.3, and age is also shown in figure 1.1.

Therapy	IAI	HBsA	Ag anti-H	Bs HCV	CAH/P	BCCAH	I/PSC A1	AT Wils	on cys	ts drug	s TOTALS
·P+A 1	25(21)	0	5(3)	1(1)	0	1(0)	0	0	0	1(0)	33(25)
P+A ₂	7(6)	0	0	0	0	0	2(1)	0	0	0	9(7)
P+A ₃	4(4)	0	0	0	1(1)	2(1)	2(1)	0	1(1)	0	10(8)
P ₄	1(1)	0	0	0	0	0	0	0	0	0	1(1)
N 5	32(25)	51(17)	8(7)	9(5)	8(7)	2(0)	1(1)	3(3)	0	19(17)	133(82)
TOTAL	.S6 9(57)	51(17)	13(10)	10(6)	9(8)	5(1)	5(3)	3(3)	1(1)	20(17)	186(123)

Table 1.2	Classification of CAH patients into etiological and therapeutic
	categories.

Numbers denote therapy groups and number of patients, numbers between brackets denote number of women, P=prednisolone, A=azathioprine, N=(group 5)no immunosuppressive therapy Therapy-groups 1 and 2: standardized immunosuppression, with resp. without prior low-dose immunosuppression, therapy-groups 3 and 4: not standardized immunosuppression. For further explanation :see text.

Age(yrs) Id	iop HI	BsAg an	tiHBs H	CV CAI	H/PBC (CAH/PS	C A1A	[Wilson	1 cysts (drugs T	OTALS
0-9	0	0	0	0	0	0	0	0	0	0	0
10-19	9	2	1	1	0	1	1	1	0	0	16
20-29	5	15	3	3	0	1	2	2	0	3	34
30-39	11	8	1	1	1	2	1	0	1	1	27
40-49	8	12	2	1	1	0	1	0	0	1	26
50-59	19	9	3	3	5	0	0	0	0	3	42
60-69	13	4	2	0	1	1	0	0	0	7	28
70-79	4	2	0	1	1	0	0	0	0	5	13
80-89	0	0	1	0	0	0	0	0	0	0	1
TOTALS	69	51	13	10	9	5	5	3	1	20	187

Table 1.3	Age	at onset	of CAH.
-----------	-----	----------	---------



Figure 1.1. Age of 186 patients with CAH. Bars show age at 5-year intervals. Lines give cumulated percentages of categories of these patients: o = IAI-CAH without anti-HBs, x = HBsAg-positive CAH, + = other types of CAH.

Some patients (shown in table 1.4) took drugs during the course of CAH that could themselves induce CAH, but the drug was assumed to be the cause of CAH only if a rechallenge proved an aetiological role, or if relation in time to the appearance and disappearance of CAH were very suspect in this regard.

A diagnosis of non-A, non-B CAH, probably due to the hepatitis C virus, (HCV-CAH) was only made if the onset of disease was within half a year of documented blood-transfusion (9 patients) or intravenous drug-abuse (1) without evidence of hepatitis B infection; otherwise the CAH was classified as idiopathic.

Documented risk factors for transmission of hepatitis B and non-A,non-B virus(es) in several patients are shown in table 1.5, which also shows that 2 patients abused alcohol in addition to the other aetiological factors.

All patients were tested for HBsAg, anti-HBs and anti-HBc-IgG. Where tested, HBsAg and HBcAg in liver biopsies were only positive in the group with circulating HBsAg. 42 out of 51 HBsAg-positive patients were positive for serum-HBeAg, 9 had anti-HBe antibodies.

In 9 patients (table 1.2), histological distinction between CAH and primary biliary cirrhosis (PBC) was impossible; 6 of these patients had high titres of antimitochondrial antibodies (AMA), but in 3 no AMA was detected. A diagnosis of PBC was later made in one of these AMA-positive patients when the liver was examined after orthotopic liver transplantation (OLT). One other patient showed a favorable reaction to therapy with prednisolone and azathioprine.

CAH-classification*	Therapy*	Additional factors		
IAI-CAH	1	sulphonamides		
HBsAg CAH	5	MZ-A1AT deficiency		
HBsAg CAH	5	oxyfenisatin		
HBsAg CAH	5	sulphonamides		
Anti-HBs CAH	5	nitrofurantoin		
HCV-CAH	5	salicylates		
HCV-CAH	5	nitrofurantoin		
САН/РВС	5	sulfonamides		
CAH/PSC	2	MZ-A1AT deficiency		
CAH/PSC	2	oxyfenisatin+sulfonamides		
CAH/PSC	2	methyldopa		
A1AT-CAH	5	nitrofurantoin		
DI-CAH (oxyphenisatin)	5	sulfonamides		

Table 1.4Factors that might have played an additional etiological role in 13
patients.

Table 1.5Documented risk factors in patients with 'viral' CAH.

Risk factor(s)	Number of patients
iv drug abuse	6
iv drug abuse, alcohol abuse	2
transfusion	3
renal Tx, transfusion	3#
haemodialysis, transfusion	1
haemodialysis, transfusion, renal TX	3
homosexual+changing contacts	1
homosexual+changing contacts and alcohol abuse	e 1
iv drug abuse	2
iv drug abuse and homosexual+changing contacts	s 1
haemodialysis, transfusion	1
transfusion	9\$,&
iv drug abuse	1
	Risk factor(s) iv drug abuse iv drug abuse, alcohol abuse transfusion renal Tx, transfusion haemodialysis, transfusion, renal TX homosexual+changing contacts homosexual+changing contacts and alcohol abuse iv drug abuse iv drug abuse and homosexual+changing contacts haemodialysis, transfusion transfusion iv drug abuse

ad *)see table 1.2	Tx=transplantation	iv=intravenous	
#=one of these patie	nts also used sulphona	mides	(see table 1.4).
\$=one of these patie	nts also used salicylate	es	(see table 1.4).
&=one of these pati	ents also used nitrofura	antoin	(see table 1.4).

In 3 patients (table 1.2) distinction between CAH and small- duct primary sclerosing cholangitis (PSC) could not be made with a combination of liver biopsy and endoscopic retrograde cholangiopancreaticography (ERCP). One reacted favorably to standardized immunosuppression, one remained stable and well while untreated, and in one patient histology and serum parameters did not improve despite all kinds of immunosuppression (including cyclosporin) and finally at liver transplantation a diagnosis of PSC was made. Two other patients were classified as CAH/PSC because of the presence of colitis ulcerosa and the impossibility to exclude PSC in the era before endoscopie retrograde cholangiopancreaticography (ERCP).

A diagnosis of Wilson's disease was established in 3 patients with CAH. One of these patients is alive and well with tri-ethylene-tetra-amine therapy (TETA), which was given because she was allergic to d-penicillamine. The other 2 patients died: one patient with disturbed swallowing (related to Wilson's disease) choked on food and died, despite previous improvement of the disease with TETA. The other patient had very severe therapy-resistant neurological disturbances and - in the pre-OLT-era - died of aspiration pneumonia.

Seven patients with α_1 -antitrypsin (A1AT) deficiency were detected by phenotyping using immunoelectrophoresis. In two of these patients with MZ-phenotype, the A1AT deficiency was probably not the main cause of CAH; one HBsAg-positive patient had an acute hepatitis B, followed by CAH; the other had high AMA-titres and CAH/PBC histology. These two patients were classified accordingly. In the other 5 patients the A1AT deficiency was the only abnormality that could be aetiologically associated with the CAH in these patients (table 1.1). The phenotypes were ZZ in one, MZ in three, and MS in one patient. One patient with phenotype MS and one with MZ were only detected after A1AT phenotyping of the whole group of idiopatic CAH patients for reasons of this study, while the other patients were detected before then by phenotyping. The patient with ZZ phenotype received an orthotopic liver graft and is doing well.

One patient had a multicystic liver and histology of CAH, reacting favorably to shortterm prednisolone and she is in remission without therapy for years now.

Drug-induced CAH was due to oxyphenisatin in 14, nitrofurantoin in 1, methyldopa in 3, sulphonamides in 1, and diclophenac in 1 patient(s).

From 176 patients stored sera were available. These are the patients investigated in the chapters on anti-cardiolipin and on anti-cytomegaloviral antibodies.

From the patients with IAI-CAH (with or without anti-HBs) CAH, 21 patients receiving standardized immunosuppressive therapy fulfilled inclusion criteria for the studies on effects of therapy.

CHAPTER 2

METHODS

All methods will be described or referred to in the chapter where they are used.

Some methods were investigated in detail to select or establish the best method for the purpose under investigation. These are described in chapters 3, 4 and 5.

CHAPTER 3

NUMERICAL VERSUS CONVENTIONAL HISTOLOGICAL SCORING OF ACTIVITY IN 'AUTOIMMUNE' CHRONIC ACTIVE HEPATITIS

SUMMARY

A histological activity index (HAI) generating a numerical score facilitates analysis of serial changes in liver histology of patients with chronic active hepatitis (CAH). In the present study, we compare the HAI as designed by Knodell et al. for asymptomatic CAH, with conventional pathological descriptions in the study of 164 liver biopsies from 38 patients with 'autoimmune' CAH requiring immunosuppressive therapy. In numerical scoring, periportal(PP), lobular(L), and portal(P) inflammation and fibrosis/cirrhosis (HFS=histological fibrosis score) were scored both separately and in combination (HAS=histological activity score=PP+L+P) (Knodell's HAI=histological activity index=HAS+HFS). HAS, HAI and PP correlated best with conventional scores (Spearman R .88,.86, and .92 respectively). Between conventional activity categories there is more overlap in HAI than in HAS, due to relative staticity of HFS. 10% of biopsies with chronic persistent hepatitis (CPH) when scored conventionally and 64% of biopsies with minimal CAH (CAHmin.) had a HAS>3. We conclude that HAS most accurately reflected activity of CAH in this patient group, and appeared to be a better parameter for monitoring activity during immunosuppression than conventional description. On the basis of the current experience we consider CAH histologically to be in 'partial remission' with a HAS ≤ 3 , and propose to reserve the term 'complete histological remission' for HAS ≤ 1 .

INTRODUCTION

Histological scoring of inflammation in CAH

Chronic active hepatitis (CAH) is defined principally by its activity rather than by structural changes. In diagnosis and follow-up of CAH, an accurate and reproducible scoring system would be of great help. The original definition of CAH by de Groote *e t al.*, revised ³⁹⁹ and extended by Bianchi *et al.*,⁴⁰⁰ is widely used. In this classic conventional pathological description, three degrees of activity of CAH are discerned: (1) Minimal activity (CAH-min): In these borderline cases periportal piecemeal necrosis may not be present *in all* portal tracts, or in a given portal tract may be seen only in a segment of the perimeter of the portal tract. (2) Moderate activity (CAHa) is characterized by dense portal infiltration with extensive piecemeal necrosis involving

substantial parts of the perimeter of almost every portal tract, but still restricted to periportal areas. (3) Severe activity (CAHb) includes cases with piecemeal necrosis in the periportal region and along fibrous septa (periseptal piecemeal necrosis), or cases with confluent bridging hepatic necrosis in addition to features of periportal piecemeal necrosis. To establish a more objective assessment of histological activity in asymptomatic CAH, Knodell *et al.* recently proposed a numerical scoring system, the Histological Activity Index (HAI).⁴⁰¹ In the current study, we applied this Histological Activity Index in symptomatic 'autoimmune' CAH and compared the data with conventional scoring of activity.

MATERIALS AND METHODS

Patients and liver biopsies

164 liver biopsies from 38 patients with CAH requiring immunosuppressive therapy according to accepted criteria⁴⁰²⁻⁴⁰⁴ were reviewed. 28 patients were classified as primary 'autoimmune' CAH, 10 patients belonged to other aetiological CAH-categories with predominant secondary autoimmune features (table 3.1). All patients in groups A and B (table 3.1) had been maintained on standardized immunosuppression: 15 mg. prednisolone and 75 mg. azathioprine for two months, followed by 10 mg. prednisolone and 50 mg. of azathioprine for two years.⁴⁰³ Low-dose immunosuppression preceded standardized therapy in therapy-group B, while patients in group A did not receive immunosuppressive therapy on referral. Patients in group C (table 3.1) had received non-standardized therapy with prednisolone alone (1 patient) or combined with azathioprine (4 patients). Needle biopsies obtained before and during immunosuppressive therapy with the Menghini technique were fixed in formalin and embedded in paraffin. Four micron sections were routinely stained with hematoxylin-eosin, periodic acid-Schiff with previous diastase digestion, Perl's iron stain, Gomori's reticulin stain, and Azan collagen stain.

Scoring activity

For the purpose of this study, all biopsies were re-examined under code by two of us (J.G. and B.v.H.), without knowledge of the patient's identity or previous histological diagnosis. In case of persisting disagreement the scores of the liver pathologist were used.

It is well-known that in a biopsy of sufficient size,^{404a,404b} the observer variability in scoring the degree of hepatitis in CAH is small with a 94% consistency rate for overall morphologic interpretation.^{404c}

In order to further minimalize intra-observer error we scored the biopsies in a short period of time using standardized forms.

	AETIOLOGY:	IAI	OTHER	TOTAL	
THERAPY:					
Α	patients	21	6	27	
	biopsies	92	24	116	
В	patients	5	1	6	
	biopsies	24	4	28	
С	patients	2 *	3	5	
	biopsies	8	12	20	
TOTAL	patients	28	10	38	
	biopsies	124	40	164	

Table 3.1Patients (37) and biopsies (164)

ad Therapy:

A= standardized (see text) immunosuppression without prior therapy,

B= standardized immunosuppression with prior low-dose corticosteroid therapy,

C= non-standardized therapy with prednisolone (2.5-30 mg) and azathioprine (25-100 mg).

ad*) 1 patient (2 biopsies) with IAI-CAH in group C received prednisolone without azathioprine

ad Aetiology: IAI: idiopathic 'autoimmune' CAH

Other:	A:	(a)	3 patients (16 biopsies): CAH with anti-HBs or/and anti-HBc in
			serum without signs of viral replication.
		(b)	1 patient (5 biopsies): like (a), combined with use of oxyphenisatin.
		(c)	1 patient (1 biopsy): HCV-CAH.
		(d)	1 patient (2 biopsies): probably IAI-CAH, but 'small-duct' primary
			sclerosing cholangitis cannot be excluded (CAH/PSC).
	B:		1 patient (4 biopsies) MZ- α_1 -antitrypsin deficiency.
	C:	(a)	1 patient (7 biopsies) MS- α_1 -antitrypsin deficiency.
		(b)	1 patient (3 biopsies) 'cholestatic CAH' (CAH/PBC).
		(c)	1 patient (2 biopsies) CAH/PSC.

The conventional description of histology according to Bianchi ⁴⁰⁰ was applied on all biopsies. To enable Spearman rank correlation between classical and numerical scoring, the conventional score of severity of CAH was assigned a number as follows: 0=(almost) no inflammation, scored normal=N, 1=chronic persistent hepatitis (CPH), 2=CAHmin., 3=CAHa, 4=CAHb. Conventional descriptions of the presence of fibrosis/cirrhosis were scored as follows : 0=no fibrosis/cirrhosis detected, 1=fibrosis/cirrhosis probable, 2=fibrosis/cirrhosis present with certainty.

Table 3.2 HAS, HFS and HAI for numerical scoring of liver biopsy specimens.

				HAS				_		HFS	
1.1	Periportal +/- bridg- ing necrosia	Score	II. ti	Intralobular degenera- on and focal necrosis*	Score	111	Portal inflammation	Score		IV. Fibrosis	-00
Α.	None	0	Α.	None	0	Α.	No portal inflam- mation	0	A.	No fibrosis	0
В.	Mild piecemeal necrosis	1	B.	Mild (acidophilic bodies, ballooning degeneration and/ or scattered foci of hepatocellular ne- crosis in <'s of lobules or podules)	1	B.	Mild (sprinkling of inflammatory cells in < ¹ /2 of portal tracts)	1	B.	Fibrous portal expansion	1
2.	Moderate piece- meal necrosis (in- volves <i>less</i> than 50% of the cir- cumference of most portal tracted	3	C.	Moderate (in- volvement of ½-½ of lobules or nod- ules)	3	C.	Moderate (in- creased inflamma- tory cells in ½-⅔ of portal tracts)	3	C.	Bridging fibrosis (portal-portal or portal-central linkage)	3
).	Marked piecemeal necrosis (involves more than 50% of the circumference of most portal tracts)	4	D.	Marked (involve- ment of > ³ /2 of lob- ules or nodules)	4	D.	Marked (dense packing of inflam- matory cells in >% of portal tracts)	4	D.	Cirrhosia ^e	4
E.	Moderate piece- meal necrosis <i>plus</i> bridging necrosis ^d	5									
F.	Marked piecemeal necrosis plus bridging necrosis	6									
G.	Multilobular ne- crosis'	10									

HAIa

is the

* Degeneration-acidophilic bodies, ballooning; focal necrosis-scattered foci of hepatocellular necrosis

Loss of normal hepatic lobular architecture with fibrous septae separating and surrounding nodules. "Bridging is defined as ≥2 bridges in the liver biopsy specimen; no distinction is made between portal-portal and portal-central linka

' Two or more contiguous lobules with panlobular necrosis.

Numerical scoring was performed exactly as described by Knodell *et al.* (table 3.2).⁴⁰¹ This system uses four histological categories: (I)periportal inflammation (piecemeal necrosis-PMN-) with or without bridging necrosis; (II) intralobular degeneration and focal necrosis; (III) portal inflammation; and (IV) fibrosis/cirrhosis. A numerical score was assigned ranging from 0 (no alteration) to 10 for caregory I, and from 0 to 4 for categories II to IV. As the score for inflammation (Categories I to III) and the score for fibrosis/cirrhosis (=HFS) (Category IV) might vary in the opposite direction during follow-up, the four categories were monitored separately in the individual patient, and the addition of categories I, II, and III was termed "Histological Activity Score" (HAS). In doing so the combined score for piecemeal necrosis and bridging necrosis weighed more heavily than the separate scores for the other three categories.

Statistical analysis

Statistical analysis was performed with Spearman's rank correlation, Mann Whitney U test and analysis of variance where appropriate, with p < 0.05 as the level of significance. If a feature could not be scored it was assigned a missing value code in the statistical analysis.

RESULTS

Results of comparison of conventional and numerical scoring of activity of CAH and relation with fibrosis/cirrhosis are shown in figures 3.1 and 3.2 and in tables 3.3 and 3.5. Analysis of variance revealed no differences in the measured parameters between primary 'autoimmune' CAH and the other forms of CAH with autoimmune features meeting requirements for immunosuppressive therapy. The combined data are presented. In 6 biopsies, fibrosis/cirrhosis could not be scored and here fibrosis/cirrhosis was assigned a missing value code. Scoring of inflammation was possible in all biopsies.



Figure 3.1 Comparison of conventional with numerical scoring of activity of CAH and its relation with fibrosis/cirrhosis. Conventional CAH-categories: 1 = normal histology, 2 = CPH, 3 = CAHmin, 4 = CAHa, 5 = CAHb.

Within each conventional category, there is a wide range of numerical scores of periportal, portal and lobular inflammation (figures 3.1 a,b,c) and fibrosis/cirrhosis (figure 3.1d), resulting in wide ranges for HAI and HAS in each of the conventional categories (figures 3.2a and b). In general, HAS is different between all conventional categories of severity while HAI does not differ significantly between CAHa and CAHb and between N and CPH. This is due to lack of significant difference in HFS (fibrosis/cirrhosis), a parameter included in the HAI, between subsequent conventional categories of CAH (fig. 3.2d). A higher degree of inflammation generally coincides with a higher HFS, indicating more fibrosis/cirrhosis when more inflammation was present. 78% of the biopsies contained some degree of fibrosis/cirrhosis (HFS \geq 1).

Significant differences in lobular inflammation could only be detected between CAHa and CAHb and not between less severe categories or between less severe categories and CAHa. Portal inflammation shows an increase between N and CPH, and between CPH



Figure 3.2 Comparison of conventional with numerical scoring of activity of CAH: a) histological activity index, b) histological activity score.

Table 3.3Correlation (R) between conventional and numerical scoring of
histological activity of CAH (Spearman rank) with p-values;
1=p<0.05, 2=p<0.001, 3=p<0.0005, 4=p<0.0001.</th>

	CONVENTIONAL			
	ACTI	VITY	FIBROSIS/0	CIRRHOSIS
	R	р	R	р
NUMERICAL periportal (PP)	.92	3	.28	2
lobular (L)	.69	4	.20	1
portal (P)	.55	4		NS
HAS (=PP+L+P)	.88	4	-	NS
HIFS	.32	3	.78	4
HAI (=HAS+HFS)	.86	4	.37	4
CONVENTIONAL cirrhosis	.30	2		

n=164 (for HFS and HAI and fibrosis n=158)

and CAHmin., but not between more severe conventional categories. Scores of periportal inflammation obtained for each conventional category of severity were different (except between N and CPH, as could be expected), although scores covered a wide range of values within each of these categories (figure 3.1a).

Spearman rank correlation coefficients between HAI and HAS and its components on one side, and conventional scoring of severity of inflammation and cirrhosis on the other side are depicted in table 3.3. Spearman rank correlation coefficients between the components of the numerical scoring system versus each other and versus the resulting HAS and HAI are shown in table 3.4.

	PERIPORTAL (PP)		LOBULAR		PORTAL		HAS		HFS	
	R	р	R	р	R	р	R	р	R	р
LOBULAR (L)	.75	2								
PORTAL (P)	.51	2	.44	2						
HAS (=PP+L+P)	.91	2	.79	2	.72	2				
HFS	.34	2	.27	1	.07	ns	.22	1		
HAI (=HAS+HFS)	.90	2	.74	2	.59	2	.91	2	.54	2

 Table 3.4.
 Correlation (R) between the components of numerical scores of histology in CAH (Spearman rank).

Significance level: 1=p<0.01, 2=p<0.0001. n=164 (for HFS and HAI n=158)

Table 3.5 shows that 10% of biopsies with chronic persistent hepatitis (CPH) and 64% of biopsies with minimal CAH (CAHmin.) had a HAS >3 when scored conventionally.

Table 3.5. Co	nventional versus i	numerical (HAS)	score of activity	(n=164).
---------------	---------------------	-----------------	-------------------	----------

conventional					
		HAS≤3	Н	AS>3	
N	14	(100%)	0	(0%)	
СРН	57	(90%)	6	(10%)	
CAHmin.	10	(36%)	18	(64%)	
CAHa	2	(5%)	40	(95%)	
CAHb	0	(0%)	17	(100%)	

DISCUSSION

A histological activity index generating a numerical score was proposed by Knodell and coworkers in 1981 to facilitate the analysis of serial changes in liver histology of patients with asymptomatic CAH of various aetiologies.⁴⁰¹ Very recently, Lindh *et al.* demonstrated that this new way of histological activity scoring was also a valuable tool in monitoring activity of chronic hepatitis B.⁴⁰⁵ Our current data in liver biopsies of patients with symptomatic 'autoimmune' CAH and CAH of other aetiologies with predominant autoimmune features and requiring immunosuppression, demonstrates the value of such a numerical score in this category of chronic liver disease. The histological activity score (HAS), which is an addition of periportal, portal and parenchymal (lobular) inflammation, and excludes the more static parameter of fibrosis (which is included in the HAI), correlated best with conventional scoring, and reflects most exactly the degree of inflammation in a liver biopsy in these patients.

Periportal inflammation may not be the only component expressing the severity of CAH. Patients with dense portal infiltrates, but only slight or no piecemeal necrosis are considered in remission in classical scoring systems. One could question whether all conventionally scored 'CAH in remission' really indicates remission. Investigators from the Mayo Clinic observed relapse rates of 28% in patients who reverted to normal liver biopsies, but 76% if residual portal hepatitis was present at the time of drug-withdrawal.^{406,407} In our patients with CAH there was a significant overlap in HAS between conventional histologic categories of severity of CAH (figure 3.2b). A number of 6/63 (10%) and 18/28 (64%) out of biopsies scored as CPH and CAHmin conventionally had a HAS >3, indicating significant inflammation in portal - and sometimes lobular- areas. As will be discussed in chapter 11, almost all of the patients with HAS >3 display elevated serum collagen-split products predominantly indicating ongoing increased deposition of collagen type III in the liver. Therefore, we propose to term HAS \leq 3 'partial remission', and HAS \leq 1 'complete remission'.

Although CAHb tended to have a lower score of portal inflammation than CAHa, this was not a significant difference. This observation may indicate a tendency to underscore portal inflammation in the presence of severe periportal inflammation.

In conclusion, the advantages of the numerical scoring system are that the HAS more accurately reflects activity of CAH when compared to conventional scoring of activity in following the natural history and treatment responses of CAH. It facilitates the comparison of histological and serological assessments of severity of disease. On the basis of the current experience we consider CAH histologically to be in 'partial remission' with a HAS \leq 3, and propose to reserve the term 'complete histological remission' for HAS \leq 1. In future studies these new criteria for remission must be validated by testing them against outcome.
CHAPTER 4

SERUM N-TERMINAL PROPEPTIDE OF COLLAGEN TYPE III IN 'AUTOIMMUNE' CHRONIC ACTIVE HEPATITIS

Comparison of three radioimmunoassays and relation with histology.

SUMMARY

We compared three radioimmunoassays (RIAs) for determination of PIIINP in autoimmune chronic active hepatitis (AI-CAH), and the relation between serum PIIINP values and numerical scores of histological activity of CAH.

90 random sera with synchronous coded biopsies in 31 AI-CAH patients were investigated. Histological activity was numerically scored (0-18). Sera, obtained during all stages of activity of disease at the time of these liver biopsies, were tested in 1) the 'PIIIP RIA-gnost®' and 2) 'PIIIP Fab assay®'(Behring) which are based on a reference bovine PIIINP antigen (col 1-3) and antibodies raised in rabbits, and 3) a recently described rapid equilibrium type RIA which is based on the human propeptide ('human PIIINP RIA'). This rapid equilibrium type of assay is not sensitive to smaller degradation products. Therefore, unlike the other two assays, no serial dilutions of the sera and no 50% intercept method for calculating the results were necessary, since the standards and the serum samples give parallel inhibition curves.

Correlation between the 'human PIIINP RIA' and the 'PIIIP RIA-gnost®' was excellent: r=.96. Between the 'PIIIP RIA-gnost®' and the 'PIIIP Fab assay' this was r=.78, between the 'human PIIINP RIA' and the 'PIIIP Fab assay' r=.76. The correlation between PIIINP in the 'human PIIINP RIA' and histological activity score (HAS) was r=.67. Correlation of all three tests with fibrosis was only weak (.27<r<.33).

With the human PIIINP RIA, a rapid non-invasive way for measuring deposition of collagen III has become available, which circumvents the problems of the assays based on the bovine PIIINP. The role for determination of PIIINP in CAH requires further study.

INTRODUCTION

Although its exact place in monitoring diseases has not yet been established, measuring the serum N-terminal propeptide of collagen type III (PIIINP) as a parameter of deposition of collagen type III appeared to be useful in several diseases,⁴⁰⁸ including chronic active hepatitis. Now that new therapeutic strategies for e.g. hepatic fibrosis

may become available in the near future, the need for a rapid and accurate PIIINP determination increases.

Type III collagen is a major constituent of most dense and loose connective tissues in the body, although bone, tendon and cartilage are devoid of it. Type III collagen is synthesized as a procollagen which contains propeptide extensions at both ends of the molecule.⁴⁰⁹ Most of the aminoterminal propeptides are set free during the synthesis and deposition of type III collagen, while some are retained in the molecules which remain on the surface of the collagen fibrils.⁴¹⁰ This antigen, when found in serum, can thus be derived from the posttranslational modification after synthesis of new type III collagen or from the degradation of PIIINP retained in the collagen fibrils.^{411,412} Smaller degradation products related to PIIINP are found in serum and excreted in urine.⁴¹³ The accurate analysis of serum antigens related to PIIINP has been difficult because of non-parallelism between the inhibition curves obtained with standard and serum samples, which is mainly due to the fact that the 'PIIIP RIA-Gnost' is sensitive to smaller degradation products.^{411,412}

The 'PIIIP Fab assay' (Behring) is also based on a reference bovine PIIINP antigen (col 1-3) and antibodies raised in rabbits. This assay uses Fab fragments with identical affinity for col 1-3 and col 1,^{413,414} 'PIIIP Fab assay®'(Behring). Therefore there is the need for serial dilutions in this assay. However, since the 'PIIIP Fab assay®' measures both antigen and degradation products, results do not necessarily parallel those obtained with the 'PIIIP RIA-Gnost'.

Recently a new radioimmunoassay (RIA) for the determination of PIIINP was developed by Risteli and Risteli.⁴¹⁵ This rapid equilibrium type of assay is based on the human propeptide and is not sensitive to smaller degradation products.⁴¹⁶ Thus, unlike the 'PIIIP RIA-gnost®',⁴¹⁷ in this new PIIINP-assay no serial dilutions of the sera and no 50% intercept method⁴¹⁸ for calculating the results are necessary, since the standards and the serum samples give parallel inhibition curves.

MATERIALS AND METHODS

Patient sera

90 sera from 31 patients with autoimmune CAH with and without immunosuppressive therapy had been frozen and stored (-20°C) until use. The only criterium for selection of these sera for this chapter was the availability of a liver biopsy performed at the same time.

Liver biopsies

Liver biopsies were processed and examined under code and assigned a Histological Activity Score (HAS), ranging from 0 to 18, which correlates well with conventional ways of scoring histology in CAH, as described.⁴¹⁹ In contrast to the original system,⁴²⁰ presence or absence of fibrosis/cirrhosis is not included in the HAS, but expreved as a different score (0-4), the histological fibrosis score (HFS) - which was also measured -.

The HAS is expressed as a numerical score (0-18), and is the summation of portal (score 0-4), periportal (score 0-10) and lobular (score 0-4) inflammation and necrosis.

Serum PIIINP

The 'PIIIP RIA-gnost®'⁴¹⁷, the 'PIIIP Fab assay' (Behring),^{413,414} and the new radioimmunoassay based on the human propeptide, developed by Risteli and Risteli,⁴¹⁵ were performed as described. Storage of sera at -20°C,⁴²¹ and bilirubin- and prednisolone-levels ^{422,423} in serum do not affect results of PIIINP-determination. PIIINP can be elevated in renal insufficiency.⁴²³ One patient had a 40% lowered glomerular filtration rate due to membrano-proliferative glomerulonephritis, but PIIINP values in this patient paralleled those found in the other CAH-patients. The age of all the patients was above 16 years, so that the normal reference values for adults (upper limit 4.2 $\mu g/l$)⁴¹⁵ could be applied. The intra-assay variation of the new equilibrium-type RIA, which is based on the human propeptide, was 4.1%, and PIIINP values were determined in one assay.

Statistical analysis

Since the populations showed similar (near-)normal distributions, statistical analysis was performed with Pearson's correlation with p<0.001 as the level of significance.

RESULTS

Correlations are shown in table 4.1. Between the 'human PIIINP RIA' and the 'PIIIP RIA-Gnost®' there was an excellent correlation of r=.96, as shown in figure 4.1A. Correlation between the 'human PIIINP RIA' and the 'PIIIP Fab RIA' was r=.78 (figure 4.1B). Between the PIIIP RIA-Gnost® and the 'PIIIP Fab RIA' this was r=.76 (figure 4.1C). The correlation between the new 'human PIIINP RIA' and histological activity (HAS) was r=.67 (figure 4.2). In all but 3 cases a PIIINP within the reference range was related to HAS \leq 3. All three tests had only a week correlation with HFS.

Table 4.1 Correlation between serum PIIINP-values obtained with three
different assays, and between these values, histological activity scores
(HAS) and histological fibrosis scores (HFS) in 'autoimmune' CAH
(n=90) (Pearson's correlation, p<.001) (p<.05 for HFS).</th>

	Human RIA	RIA-Gnost	Fab-assay	
RIA-Gnost	.96			
Fab-assay	.78	.76		
HAS	.67	.64	.55	
HFS	.31	.33	.27	



Figure 4.1 Correlation (Pearson) between serum PIIINP-values ($\mu g/l$) obtained with three different assays in 'autoimmune' CAH (n=90) (p<.001). A)Human PIIINP RIA vs. PIIIP RIA-Gnost. R=0.96, y=-0.376 + 0.370x, B)Human PIIINP RIA vs. PIIIP Fab-assay. R=0.78, y=-37.17 + 1.24x, C)PIIIP RIA-Gnost vs. PIIIP Fab-assay. R=0.76, y=39.73 + 3.21x.



Figure 4.2 Correlation between PIIINP (human PIIINP RIA) and histological activity (HAS).(Pearson's correlation R=0.67, p<.001, n=90).

DISCUSSION

The current study shows that results obtained with the new PIIINP-assay, which is based on the human propeptide, correlate very well with the standard 'PIIIP RIA-gnost®'. The 'PIIIP Fab assay' measures both antigen and degradation products, which probably explains why results do not parallel those obtained with the two other assays. The fact that PIIINP is only weakly correlated to HFS probably reflects the fact that serum PIIINP reflects (dynamic) fibrogenesis rather than (static) fibrosis.

Serum PIIINP, predominantly reflecting deposition of collagen type III in the liver in CAH, correlated to some extent with activity of hepatic inflammation. Some studies indicated that serum PIIINP values parallel the activity of inflammation in acute hepatitis, and return to normal with transaminases and bilirubin.⁴²⁴ In alcoholic hepatitis, serum PIIINP correlated well with periportal and intralobular inflammation.⁴²⁵⁻⁴²⁹ Several studies suggested a relation between activity of CAH and PIIINP,⁴³⁰⁻⁴³³ although this could be masked in children with CAH by elevated PIIINP due to growth.⁴³⁴ Persistent elevation of PIIINP in serum of patients with viral hepatitis for more than 6.5 months correlated with chronicity.^{424,435} Therapy with corticosteroids can lower an elevated PIIINP in CAH,^{422,436-439} and in a recent study PIIINP changed in accordance with other liver tests and normalized when remission of CAH had been

achieved.^{421,440} In the current transversal study, the correlation between serum PIIINP and HAS was only r=.67. The correlations between PIIINP and seperate scores of periportal inflammation were similar (data not shown). Many factors influence fibrogenesis, which may explain why this correlation is not better than 0.67. Another explanation may be a lay-time in the decline, of the inflammatry infiltrate. This will be discussed in chapter 11 as well. Some of the data represent several tests done in the same patients, so that we might have missed a correlation in the region with higher HAS and PIIINP.

Human PIIINP values above 20 μ g/l were related to HAS>3 (active disease), while PIIINP values below 2 μ g/l were related to HAS≤3 (partial histological remission). Serum PIIINP might be helpful in monitoring activity of CAH. Between 2 and 20 μ g/l, however, there was a 'gray zone' where the serum PIIINP was less predictive for the HAS. Apart from a few exceptions, however, it can be stated that a PIIINP within the reference range is related to histological remission (HAS≤3).

With the 'human PIIINP RIA', a rapid non-invasive way of measuring deposition of collagen III has become available, which circumvents the problems⁴⁴¹ of the assays based on the bovine PIIINP. The role for determination of PIIINP in monitoring CAH and possible future therapies for increased hepatic fibrogenesis and collagen-deposition⁴⁴² requires further study. Further studies aimed at delining the value of testing for PIIINP will follow in chapters 11 and 12.

CHAPTER 5

ESTABLISHMENT OF A MODIFIED QUANTITATIVE RADIOIMMUNOASSAY FOR THE DETERMINATION OF ANTIBODIES AGAINST 'LIVER-SPECIFIC PROTEIN' (LSP)

SUMMARY

Antibodies against 'liver specific membrane lipoprotein' (anti-LSP) play an important role in pathogenesis of idiopathic 'autoimmune' chronic active hepatitis (IAI-CAH), and may be useful for immune monitoring this disease. Quantitative determination of anti-LSP, however, required dilution in several steps, which limited routine use in IAI-CAH. We here describe a modification of the original anti-LSP assay. Binding percentages as determined in the previously described assay and in our modified assay did not differ. The inter-assay variation was intra-labelling <10%, inter-labelling 15%, and the intra assay variation <10%. Using 3 dilutions calculated anti-LSP titres did not differ from measured titres. The normal binding range was 5-24%. We developed a modified quantitative, reproducible radioimmunoassay for determination of anti-LSP, facilitating studies on monitoring IAI-CAH.

INTRODUCTION

'Liver-specific protein' (LSP) is a macromolecular, lipid-associated complex containing a number (as yet unidentified) of hepatocyte plasma membrane-derived antigens, some of which are non-organ-specific and others are liver-specific (including at least one that is species-specific and at least one other that is species cross-reactive).⁴⁴³ The high incidence of autoantibodies against LSP in chronic active hepatitis (CAH), especially in idiopathic 'autoimmune' CAH (IAI-CAH) and its role in antibody-dependant cytotoxicity have been discussed in the introduction. In order to assess the value of anti-LSP in our population of standardized treated IAI-CAH patients, we established a modified radioimmunoassay for the determination of anti-LSP.

METHODS

Preparation of LSP has been described.⁴⁴⁴ To establish a radioimmunoassay for determination of anti-LSP antibodies, rabbit-LSP prepared as described (with slight modifications) was used. A modification in preparation had been the dialysis of LSP

against sucrose for rabbit-LSP, and acetazolamide for human LSP, for concentration of the material. This was an alternative to ultrafiltration through an Amicon PM 30 membrane. The rabbit-LSP (1 mg/ml) used for this study was a gift from B.M. McFarlane-Wojcicka and I.G. MacFarlane (King's College Hospital -KCH-, London). This facilitated comparison with the original assay. Recently human LSP was prepared in the department of hepatology, University Hospital Groningen, and the following also applies to the assay using human LSP as an antigen.

The essentials of the radioimmunoassay (RIA) have been described.⁴⁴⁵ Our modifications concern both the labelling and the assay itself:

Labelling

Instead of older labelling techniques⁴⁴⁶ or labelling with ¹²⁵I coupled to Sephadex,⁴⁴⁷ we labelled the LSP with Iodobeads® (Pierce Chemical Company, Rockford III, U.S.A.). These are N-chloro-benzenesulfonamide (sodium salt) derivatized uniform 2.8 mm diameter nonporous polystyrene spheres, with the structure as shown in figure 5.1. These beads facilitate smooth and reproducible iodination without the preparation of



Figure 5.1 Structure of Todobeads'.

additional solid phase chemical agents, and without the harsh effects of chloramine T. They also enable labelling over a broad pH and temperature range.⁴⁴⁸ After washing the beads twice with borate-buffer 0.1 M, with 1mM EDTA with or without azide (no difference in results), 200 μ l BSA-borate buffer was added to the beads. Then 5 μ l Na-¹²⁵I solution (Radiochemical Centre, Amersham), was added (0.5-1 mCi), and after 5 minutes at room temperature, 20 μ l of LSP was added. After 10 minutes, the reaction volume was separated from the Iodobeads to terminate iodination. Then, to remove free iodide, the reaction contents were directly transferred to an equilibrated Sephadex G-25 column, and the radiolabelled LSP was recovered in the usual way in the void volume with the help of a fraction collector. Every fraction was counted in the gamma counter as depicted in figure 5.2. The first (LP1-) peak contains the LSP as has been previously described. In contrast to the original assay, where labelled LSP was diluted 1:1 with unlabelled LSP, we pooled the fractions that contained LSP and added buffer with BSA 1% and azide up to 5 ml. In the 10 μ l fractions with LSP, the percentage LSP-bound ¹²⁵I (*LSP) was determined by the trichloric acid (TCA) precipitation technique as



Figure 5.2 Recovery of labelled LSP from a Sephadex G-25 column after iodination. Counts per minute minus background (CPM) as determined by a gammacounter in the fractions collected are shown.

ad *) Fractions 9,10,11 and 12 form the LPI-peak that contain LSP.

described,⁴⁴⁹ and the usual pattern obtained is shown in fig 5.2. TCA-precipitation was done each time before an assay was performed, and the *LSP only used if the precipitatable *LSP was >75%. Before use, 1 ml of *LSP was purified on a Sepharose 6B column, and the LSP-peak determined as described above. Again, the first peak was collected and 0.1% buffer with BSA and buffer with azide added up to 5 ml. In the calculations after performance of an assay, the activity of this *LSP was set at 100%.

Assay

Essentially, the assay was performed as described earlier,⁴⁴⁵ but we made some modifications: Patient sera were thawed at 4-7°C, and 50-100 μ l aliquots were inactivated for 30 minutes at 56°C using glass tubes. Sera were diluted 1:50 with 1% BSA-borate (pH 8.5) in duplicate. If quantitation was required, additional doubling dilutions 1:100 and 1:200 were made. Pooled normal serum was used as a negative control, blanks were added, and a titrated serum containing high anti-LSP titres (binding percentages also determined in KCH, London) was used as a positive control. 25 μ l of diluted sera and 25 μ l of *LSP (without further dilution with unlabelled LSP) were put into plastic tubes. Two tubes for the precipitation test contained only 25 μ l *LSP. After mixing and incubating overnight at 4-7°C 100 μ l of dried staphylococcal cells suspended in 1% BSA-borate pH 8.5 (concentration 10 mg/ml) was added to each tube. To the tubes containing only *LSP and no serum, 475 μ l 1% BSA-borate and 500 μ l 40% TCA was added. The contents were mixed and incubated for 1 hour at 4-7°C with

additional mixing at the half hour. 850 μ l 0.25 M sucrose (pH 8.5) was added to the tubes with sera, and all tubes centrifuged for 10 minutes at 3500 rpm. Instead of counting one tube with the infranatant and half of the supernatant, and one tube with the other half of the supranatant, we removed all supranatant and counted only the infranatant. We counted 4 minutes/tube in an LKB gammacounter linked to a computer to calculate titres (according to the formulas given in the results), and binding-percentages (%B) as follows:

%B= [cpm(patient serum) - cpm(blank)] %B= *100% [cpm(*LSP) - cpm(background activity)]

(cpm=counts per minute)

The effect of reducing the counting time to 2 minutes per tube was assessed. Calculated titres were compared with measured titres. For titration of the positive control, we determined dilutions from 1:50, 150, 200, 250, 300, 350, 400, 450, 500 and 1:1000. The consequences of our modifications for the results were assessed, by simultaneous comparison with the results obtained in the original method as described.⁴⁴⁵ Statistical analysis was performed with the Wilcoxon two-sample test and Spearman's rank correlation where appropriate, and p-values <0.05 were considered significant.

RESULTS

There was no significant difference betweem binding percentages (%B) as determined in London (KCH) with the original assay, and those determined in our assay.

The correlation between %B in these two assays was r=.80. The inter-assay coefficient of variation in our assay was <10%, inter-labelling this was 15%, and the intra-assay variation was <10%. Our modifications in labelling and in the assay (dilution with buffer instead of unlabelled LSP, and counting only the infranatant) did not affect the results. Binding percentages obtained after 2 minutes of counting did not differ from those obtained after 4 minutes counting per tube (although we usually count 4 minutes). Curves indicating %B obtained in increasing dilutions of sera could be linearized by log_{10} -transformation of %B, as shown in figure 5.3. For dilutions above 1:50 the correlation between dilution and log_{10} (%B) was median r=0.98 (range 0.95-0.99).

This enabled us to calculate titres with only three doubling dilutions of a serum (1:50, 1:100, 1:200). The upper limit of normal was set at $\log_{10}(\%B_0 + 2SD)$ (SD=standard deviation, $\%B_0$ =mean %B of 32 negative controls). The best fitting straight line through the %B obtained in 1:50, 1:100 and 1:200 dilutions was determined, and the abscissa at



Figure 5.3 Calculation of anti-LSP titres. (N+2SD = median + 2 times standard deviation of reference values). See text for further explanation.

its intercept with the upper reference limit yielded the anti-LSP titre. Only on the rare occasion that the correlation between dilution and %B was lower than r=0.95 was the measurement repeated. In formulae:

These formulae were entered into a computer linked to the gamma-counter, so that results were obtained immediately in %B and titres. Anti-LSP titres calculated in this way did not differ significantly from measured titres: the correlation between measured titres and calculated titres was r=.98. The reference range of normal binding was 5-24%, and titres >1:50 were considered positive.

DISCUSSION

The importance of a reproducible anti-LSP assay lies in the predictive value of anti-LSP antibodies for the outcome of therapy withdrawal in IAI-CAH,⁴⁵⁰ and in its help in distinguishing AI-CAH from non-A,non-B CAH (HCV-CAH).⁴⁵¹ Determination by means of a radioimmunoassay appeared to be superior to measurement with a micro-ELISA.⁴⁵²

We established a modified, reproducible radioimmunoassay for the determination of anti-LSP that yields quantitative results with 3 dilutions. Modifications did not affect the results. The method used labelling, and the dilution of *LSP with buffer instead of unlabelled LSP gave higher cpm, enabling labelling with less radioactivity and/or counting for a shorter time. Having to count only the infranatant, the linkage of a computer to the gamma-counter to calculate %B and titres, and the use of a pipetting-automate were further time-saving steps.

The difference in gradients of the regression line connecting binding percentages of the dilutions 1:50, 1:100 and 1:200 between patients, indicates that among patients different antibodies directed at more than one antigenic site within the LSP-complex may be present. Although it appears that the asialoglycoprotein receptor (hepatic lectin) is one of the important antigenic determinants within LSP,⁴⁵³⁻⁴⁵⁵ other antigenic sites may play a role. For immune monitoring of IAI-CAH this is not a problem, and it may even be preferable to measure all anti-LSP antibodies involved, to assess 'immunological activity' of IAI-CAH.

The present assay enables rapid, reproducible and quantitative determination of anti-LSP antibodies, facilitating studies on monitoring IAI-CAH. This assay was used for the studies described in chapters 13 and 14. For further details on immunology in CAH the reader is referred to the introduction of this thesis.

SECTION 2

Analysis of survival in chronic active hepatitis of various aetiologies

CHAPTER 6

LONG-TERM PROGNOSIS IN CHRONIC ACTIVE HEPATITIS

SUMMARY

We assessed long-term survival and prognostic factors in chronic active hepatitis (CAH). Methods: Survival (Kaplan-Meier) and prognostic factors, using Cox's proportional hazards estimation were evaluated, in 186 patients with CAH, consecutively referred from 1969 through 1988. End of follow-up was November 1988. Information was retrieved from charts, other physicians and public registry offices. All HBsAgpositive patients and 3/4 of anti-HBs-positive patients received no therapy. Results: Ages among therapy- and aetiological groups did not differ. Kaplan-Meier probability of survival was:

All patients 18 AETIOLOGY: HCV 1	36				
AETIOLOGY: HCV 1		85(±3)	70(±4)	55(±5)	40(±10)
HCV 1					
	0	100	100	100	100
IAI (no anti-HBs) 6	9	90(±4)	76(±7)	60(±11)	-
Anti-HBs-positive IAI 1	3	50(±17)	15(±13)	15(±13)	15(±13)
HBsAg-positive 5	1	80(±6)	72(±7)	65(±9)	1
other aetiology 4	3	85(±6)	67(±8)	45(±10)	22(±12)
THERAPY:					
standardized therapy 4	2		85(±8)		85(±8)
none/non-standardized 14	4		67(±5)		35(±9)
BOTH:					
no/n-s therapy + anti-HBs	3	17(±15)	0		
standard + anti-HBs	5		50(±35)	50(±35)	
standard, no anti-HBs 3 none/non-standard +	9	97(±3)	90(±7)		90(±7)
no anti-HBs 13	86	84(±3)	70(±5)	55(±6)	

Most causes of deaths registrated in patients with CAH were not related to the liver disease. Remarkably, there was a high incidence of plasmocytoma. In Cox's regression analysis independant riskfactors for mortality were: age (increase in risk-IR- x1.3/10 years), presence of anti-HBs (IR x6.7), inability to institute standard immunosuppressive therapy (IR x3.6), and abnormal values of serum alanineaminotransferase (ALT) (IR x3.6). Sex, year of diagnosis, duration of disease until the end of follow-up, serum values of gamma globulin, cholinesterase, albumin, and presence or absence of ANA and SMA at the time of diagnosis were not risk factors.

INTRODUCTION

Early assessments of natural history and prognosis in CAH have been summarized recently.⁴⁵⁶ Mortality ranged from 20% in 3 years,⁴⁵⁷ 40% in 4 years,⁴⁵⁸ 80% within 5 years⁴⁵⁹ or 70% in 6 years,⁴⁶⁰ to 50% within 10 years.⁴⁶¹ Prospective studies have indicated that mortality approximates 40 percent within 6 months from diagnosis if corticosteroids are not administered.⁴⁶² In the era before the introduction of immunosuppressive therapy, cirrhosis, bridging or multilobular necrosis at presentation was associated with the highest mortality.463-466 Late mortality was related to the consequences of portal hypertension rather than liver failure, which is the main cause of early death in CAH.⁴⁶⁷ Survival beyond 3 years in untreated CAH was associated with a spontaneous reduction in inflammatory activity, and with transition to an inactive macronodular cirrhosis.⁴⁶⁷ The abrupt onset of symptoms, persistent jaundice, hepatic encephalopathy, ascites, and ulcerative colitis were associated with poor prognosis,⁴⁶³ Serum aminotransferase elevation of greater than tenfold normal, or fivefold normal in conjunction with twofold elevation of gamma globulin level, was associated with mortality of greater than 50% within 3 years of diagnosis.⁴⁶⁸ Corticosteroid therapy with or without azathioprine has been shown to induce remission in 60 percent of patients with severe autoimmune CAH within 3 years, with an increase of 5-year survival to approximately 90 percent.^{469,470} Treated IAI-CAH patients without cirrhosis on admission had the best prognosis, while development of cirrhosis during therapy -which occurred in 40% of patients without cirrhosis on entry- did not adversely affect the probability of survival. Other results indicate that prognosis in transfusion-related non-Anon-B CAH -probably due to hepatitis C virus (HCV-CAH)- appeared best, followed by that of treated IAI-CAH, with the worst prognosis in HBsAg-positive CAH.⁴⁵⁶

In the present study, we analyzed survival in 186 consecutive patients with liver biopsy proven chronic active hepatitis. The analysis was in relation to aetiology of CAH, instituted therapy, age, sex, year of diagnosis, duration of disease until the end of follow-up, cause of death, serum values of alanineaminotransferase, gamma globulin, cholinesterase, albumin, and presence or absence of ANA and SMA at the time of diagnosis.

MATERIALS AND METHODS

Patients

From November 1969 to November 1988, a total of 186 consecutive patients referred to the division of hepatology, State University Hospital Groningen, were diagnosed as having liver biopsy proven chronic active hepatitis. During this period liver biopsy records were kept systematically, and from 175 patients sera were frozen (-20°C). Patient-characteristics were described in chapter 1. Where necessary, biopsies (sometimes from other hospitals) and diagnoses were reviewed, hepatitis B serology redone, and in all patients alpha-1-antitrypsin phenotyping was performed. For the purpose of this study, we discern five aetiological groups: 1) idiopathic 'autoimmune' CAH without anti-HBs (IAI-CAH), 2) hepatitis B surface antigen-positive CAH (HBsAg-CAH), 3) CAH with circulating antibodies to HbsAg (anti-HBs CAH), 4) transfusion- or needlestick- and iv drug-abuse related non-A,non-B CAH (HCV-CAH), 5) other aetiologies (table 6.1).

Actiology*	Number of patients	Sex M:F		Age (years) median range		
1)IAI-CAH	69	12:	57	51	11-78	
2)HBsAg-CAH	51	37:	14	42	18-74	
3)anti-HBs-CAH	13	3:	10	46	17-81	
4)HCV-CAH	10	4:	6	37	18-77	
5}DI-CAH	20	3:	17	61	24-78	
other CAH	23	6:	17	34	15-72	
Total	186	65:	121	46	11-81	

Table 6.1CA	H-patients	grouped	according	to aetiology.
-------------	------------	---------	-----------	---------------

ad *)		
IAI-CAH	=	idiopathic 'autoimmune' CAH without anti-HBs
HBsAg CAH	=	CAH with circulating HBsAg
Anti-HBs CAH	I=	CAH with circulating anti-HBs
HCV-CAH	=	CAH due to parenteral non-A, non-B hepatitis (probably due to hepatitis C virus)
DI-CAH	=	drug-induced 'CAH' due to: oxyphenisatin(12), α-methyl-dopa(3), nitrofurantoin(1), diclofenac(1).
Other CAH:		
9x CAH/PBC		= 'cholestatic CAH' or 'mixed form'CAH/PBC
5x CAH/PSC		= 'small-duct' PSC and/or CAH 3x Wilson-CAH=CAH associated with Wilson's disease
5x A1AT-CAH	ł	= CAH related to α_1 -antitryps in deficiency 1X CAH in multicystic liver

Five therapeutic groups were present: standardized immunosuppression with prednisolone (P) and azathioprine (A) as described in chapter 1, a) without and b) with preceding low-dose (2.5-10 mg prednisolone) immunosuppression, c) P in conjunction with A, but not according to a standardized regimen, d) no immunosuppressive therapy, e) P only incidentally during exacerbations.

Methods

In this analysis of survival we recorded: age (in years) at the time of the diagnostic liver biopsy, sex of the patient, aetiology of CAH, the kind of therapy for CAH which was instituted, the serum values of alanine aminotransferase (ALT), gamma globulin (GG), cholinesterase (CHE), albumin (ALB), and presence or absence of antinuclear antibodies (ANA) and anti-smooth muscle antibodies (SMA) at the time of diagnosis, the year of diagnosis, duration (in days) between diagnosis and the end of follow-up (November 1988, or time of death or liver transplantation), and cause of death. Duration of survival was related to these characteristics. In our study we assumed an equal distribution of cirrhosis over the several groups, since sufficient data to exclude the presence of cirrhosis were lacking. If serum ALT or total bilirubin were continuously elevated more than twofold for more than a month preceding the diagnostic liver biopsy, the onset of these abnormalities in liver tests were regarded as the moment of diagnosis. The family practitioners and specialists were asked whether a patient was still alive, and, if not, what the cause of death had been. Where necessary, data were derived from public registry offices.

Statistical analysis

Survival curves were plotted by the Kaplan-Meier productlimit method.⁴⁷¹ Results are expressed as probabilities \pm standard error. Differences between subsets of patients were analysed by means of the log rank test. For identification of individual prognostic factors we used Cox's proportional hazards estimation.⁴⁷²

RESULTS

We assessed survival probability in 186 consecutive patients with chronic active hepatitis. This assessment was in relation to aetiology of CAH, therapy undertaken, age, sex, year of diagnosis, duration of disease until the end of follow-up, cause of death, serum values of alanineaminotransferase, gamma globulin, cholinesterase, albumin, and presence or absence of ANA and SMA at the time of diagnosis. $85(\pm 3)$ percent of all 186 patients with CAH survived 5 years, $70(\pm 4)$ percent survived 10 years, $55(\pm 5)$ survived 15 years, and $40(\pm 10)$ survived 20 years after diagnosis (figure 6.1). Ages between therapygroups and aetiological groups did not differ significantly.



Figure 6.1 Probability of survival in all 186 patients with CAH.

On stratification of the patients according to aetiologies (figure 6.2), survival markedly differed between groups (p=.001): None of the ten HCV-CAH patients in group 4) died,





leading to a probability of survival of 100% after 20 years. The probability of survival in groups 1) and 2) after 5 years in IAI-CAH (treated and untreated) was $90(\pm 4)$ percent and in HbsAg-CAH (always untreated) $80(\pm 6)$ percent. After 10 years this was $76(\pm 7)$ percent and in HbsAg-CAH $72(\pm 7)$ percent, and after 15 years $60(\pm 11)$ and $65(\pm 9)$ percent. In the anti-Hbs positive patients (75 percent of them untreated) of group 3), the Kaplan-Meier (K-M) probability of survival was worst of all patients: $50(\pm 17)$ percent after 5 years, $15(\pm 13)$ percent after 10 and 20 years. The group 5) with CAH of other aetiology had a probability of survival of $85(\pm 6)$ percent after 5 years, $67(\pm 8)$ percent after 10 years, $45(\pm 10)$ percent after 15 years, and $22(\pm 12)$ percent after 20 years.

When stratifying the patients according to therapies (figures 6.3 and 6.4), survival between therapy groups a and b versus therapy groups c,d and e differed significantly. There was a $85(\pm 8)$ percent 10 and 20-year survival in the first group versus $67(\pm 5)$ percent 10-year survival and $35(\pm 9)$ percent 20-year survival in the latter group (p=.01). No differences were detected between groups a and b, nor between groups c,d, and e.



Figure 6.3 Probability of survival stratified on therapy. a=standard therapy prednisolone+azathioprine without pretreatment b=standard therapy prednisolone+azathioprine with pretreatment c=non-standard therapy prednisolone+azathioprine d=untreated e=prednisolone only during exacerbations of CAH

Stratification of the patients according to both aetiological groups and therapy groups also yielded significant differences (figure 6.5): Anti-HBs positive patients in therapy groups c,d, or e had a survival probability of $17(\pm 15)$ percent after 5 years and zero after 10 years.



Figure 6.4 Probability of survival stratified on therapy. (a and b combined, c + d + e combined). See also legend of figure 3.



Figure 6.5 Probability of survival stratified on both aetiology and therapy. A= no anti-HBs, standardized therapy

- B= anti-HBs, standardized therapy
- C= no anti-HBs, no standardized therapy
- D= anti-HBs, no standardized therapy

aetiolo	ogy therapy	cause of death	n=
1	a	lung metastases of adenocarcinoma	1
1	d	pulmonary embolism + liver metastases	1
1	d	myocardial infarction	2
1	d	hepatic coma	1
1	e	3' years after OLT, cause unknown	1
1	d	digestive tract bleeding 1 1 4 cause unknown	4
1	е	cause unknown	5
1			total 16
2	d	bleeding from oesophageal varices	2
2	d	cardiac death	1
2	d	plasmocytoma (Kahler's disease)	1
2	d	HCC + plasmocytoma (Kahler's disease)	1
2	d	cause unknown	6
2	е	cause unknown	3
2			total 14
3	8	myocardial infarction + tamponade	1
3	d	traumatic subdural haematoma	1
3	d	during OLT (coogulation disorders)	1
3	d	during OLT (intraoperative problems)	1
3	d	6 days after OLT bleeding from ulcus ventriculi	1
3	c	cause unknown	1
3	d	cause unknown	1
3	-		total 7
4			total 0
54	ρ	nlasmocutoma (Kahler's disease)	1
54	d	cause unknown	3
54	e		1
5R	c	acute liver failure	1
5B	0	bleeding from accombageal varices. HCC with portal thromby	neie 1
5B	d	cause unknown 1	515 1
5 <u>C</u>	d	choking on food caused by abnormal swallowing in	
50	4	CAH' related to Wilson's disease	1
5C	d	nneumonitis in 'CAH' related to Wilson's disease	1
50	4	with severe neurological abnormalities	1
5D	9	perforated sigmoiditis	1
50	ď	acute liver failure and nulmonary carcinoma	1
50	d	urosensis	2
50	d	myocardial inferction + rhythm disorder	1
5D	d	cause unknown	1 <u>1</u>
-		enuse diffiguri	1 10
Э			total 19
			Total 56

 Table 6.2
 Causes of death in CAH and CAH-related disorders (n=186).

Actiology:

1) IAI-CAH, 2) HBsAg-positive CAH, 3) anti-HBs positive CAH, 4) non-A, non-B (HCV) CAH 5) other CAH: 5A) CAH/PBC, 5B) CAH/PSC, 5C) 'CAH' related to Wilson's disease, 5D) oxyphenisatin-induced CAH.

Therapy: a) standardized prednisolone (P) and azathioprine (A), no pretreatment, b) idem with pretreatment (2.5-10 mg P, one with 25 mg A), c) non-standardized P+A, d) untreated, e) P only during exacerbations.

Five anti-HBs positive patients in therapy-groups a and b, however, had a probability of survival of $50(\pm 35)$ percent after 10 and 15 years. Patients with CAH, but without anti-HBs, had the best survival probabilities: in therapy-groups a and b this was $97(\pm 3)$ percent after 5 years and $90(\pm 7)$ percent after 10 and 20 years. In therapy-groups c, d, and e this was $84(\pm 3)$ percent after 5 years, $70(\pm 5)$ percent after 10 years, and $55(\pm 6)$ percent after 15 years. Most causes of deaths registered in patients with CAH were not related to the liver disease. Remarkably, there was a high incidence of plasmocytoma (table 6.2).

Patients with ALT above the upper reference limit had a worse long-term and short-time prognosis than patients with an ALT within the reference range at the time of diagnosis (p=<0.05).



Figure 6.6 Probability of survival in patients with ALT above the upper reference limit (solid line) and patients with ALT within the reference range (dashed line) at the time of diagnosis (p<0.05) (n=127)

Cox's proportional hazards estimation for identification of individual prognostic factors yielded a model with age, presence/absence of anti-HBs, possibility to institute standardized immunosuppressive therapy and elevated ALT as independant risk factors for mortality (table 6.2). No other prognostic factors were identified among the other parameters mentioned above. Using these data individual prognosis in CAH can be determined.

12 CAH patients underwent a liver transplantation (LT), as shown in table 6.3. The results in anti-HBs positive patients contrasted with survival probability in patients from the same aetiological group receiving immunosuppression, as stated above. One patient

actiology of CAH	therapy L before LT	T performed after diagno	status at end of follow-up (days)	
1	d	778	1105	alive and well
1	d	259	3163	alive and well
1	d	152	1326	alive and well
1	e	3504	1356	died after OLT
2	d	46	2388	alive and well
3	d	341	0	died during OLT
3	d	1547	0	died during OLT
3	d	934	6	died after OLT
4	d	1241	6	alive and well
5A	С	3597	3487	alive and well
5B	С	30 *	537	alive and well
5E	d	176	1281	alive and well

 Table 6.3
 CAH-patients undergoing liver transplantation (LT).

ad *) This patient underwent an auxilary heterotopic transplantation in Rotterdam, and histology from the acceptor-liver changed the diagnosis from CAH/PSC to PSC lateron. The other patients underwent orthotopic liver transplantation (OLT) in Groningen. Causes of death: see table 2. For aetiology- and therapy-numbers: see table 2.

with 'CAH' in relation to the homozygous ZZ-phenotype α_1 -antitrypsin deficiency needed a liver graft soon after diagnosis, despite a short course of non-standardized immunosuppression. This contrasts with three patients with partial α_1 -antitrypsin deficiency who all responded to immunosuppressive therapy with prednisolone and azathioprine. LT in patients without anti-HBs was successful. The incidence of LT in anti-

Table 6.4Final model of long-term prognostic factors for chronic active
hepatitis of various aetiology.

Higher age, presence of anti-HBs in serum without HBsAg, no institution of standardized immunosuppressive therapy and elevated ALT increased the risk of mortality.

	Relative risk	95% co			
Co-variate	of mortality	lower	upper	T-stat	p-value
Age	x 1.3/10 years	1.04	1.5	2.420	<0.001
anti-HBs	x 6.7	1.3	34.2	2.277	< 0.001
No St-therapy	x 3.6	1.03	13.6	1.910	<0.015
ALT elevated	x 2.0	1.04	3.9	-2.083	<0.010

HBs-negative IAI-CAH was low (only 4 patients, of whom 3 without therapy and 1 on non-standardized immunosuppression). This probably reflects the excellent survival probability in those patients selected for immunosuppressive therapy.

DISCUSSION

From the present study it follows that in chronic active hepatitis, long-term prognosis is determined by aetiology of CAH -especially the presence of anti-HBs-, the decision whether to administer (fixed-dose) immunosuppressive therapy or not, as well as the age of the patient and the ALT at diagnosis. The difference in survival between patients with high and low ALT became clear early, because of liver failure in some of the patients with higher ALT. Remarkably, CHE at diagnosis, and presence/absence of ANA and SMA were found not to be risk-factors. Patients with putative non-A,non-B CAH -probably due to HCV- had the best prognosis. One of these patients had fulfilled requirements for standardized immunosuppression and had been treated accordingly. Untreated anti-HBs positive patients with CAH had the worst prognosis. Besides causes of death unrelated to liver disease, this was due to a high incidence of OLT with high per- and postoperative mortality in the subgroup of untreated anti-HBs positive patients. These patients had more advanced disease. CAH in these patients may also have been caused by HCV.

Intermediate between the likelihood of survival of these 'aetiological groups' lies the survival of (untreated) HBsAg-positive CAH and (both treated and untreated) IAI-CAH. Patients with IAI-CAH without anti-HBs and anti-HBs-positive CAH maintained on standardized immunosuppressive therapy with fixed doses of prednisolone (P) and azathioprine (A) had a much better survival probability than untreated patients from the same 'aetiological groups'. Their outcome was also better than that of patients receiving non-standardized immunosuppressive therapy, e.g. only given during exacerbations of the disease. Survival was as good as in treated IAI-CAH in two 'CAH'-patients with partial α_1 -antitrypsin deficiency of the MZ-phenotype on standardized therapy with P and A, and in one patient with partial α_1 -antitrypsin deficiency of the A.

In more than half of the deceased patients with known cause of death, fatality was not related to the liver disease, and death from gastrointestinal bleeding was uncommon. The incidence of plasmocytoma was quite high, while HCC occurred infrequently.

In the era before the introduction of immunosuppressive therapy, cirrhosis,bridging or multilobular necrosis at presentation was associated with the highest mortality.⁴⁶³⁻⁴⁶⁶ Late mortality was related to the consequences of portal hypertension rather than liver failure, which is the main cause of early death in CAH.⁴⁶⁷ Survival beyond 3 years in untreated CAH was associated with a spontaneous reduction in inflammatory activity, and with transition to an inactive macronodular cirrhosis.⁴⁶⁷ Other studies indicated that

patients that improved without corticosteroids could not be distinguished from others on accession by disease duration, age, sex, initial histologic findings, associated autoimmune disease (with exception of colitis ulcerosa), immunoserologic studies (presence of LE-cell phenomenon, antinuclear antibody, smooth muscle antibody), or biochemical abnormalities (AST, bilirubin, gamma globulin, albumin).^{456,474} It was shown that patients with mild disease improve without therapy as frequently as those with severe disease (13 percent versus 20 percent).⁴⁷⁴ In a study of 80 asymptomatic patients, including 34 with HBsAg, 20% developed cirrhosis and 9% died of liver failure, while the 5- and 10-year survival probabilities were 91 and 81 percent.⁴⁷⁵ The abrupt onset of symptoms, persistent jaundice, hepatic encephalopathy, ascites, and ulcerative colitis were associated with poor prognosis.⁴⁶⁵ Serum aminotransferase elevation of gamma globulin level, was associated with mortality of greater than 50% within 3 years of diagnosis.⁴⁶⁸

Some other studies indicate that aetiology is an important prognostic factor in CAH,⁴⁵⁶ and that non-A,non-B (or HCV)-CAH usually is a self-limiting process. During the initial episode of HCV-hepatitis most patients are anicteric and 35% asymptomatic. Nevertheless, the majority (55-65%) of these patients develop progressive liver disease with CAH, with or without cirrhosis.^{476,477} In transfusion-related HCV-CAH however, spontaneous improvement is usual, a complete resolution having been described in half of the patients within 3 years after diagnosis.^{478,479} Cirrhosis developed in 10-60 percent, immediate survival was excellent and HCC was absent.^{477,479,480}

Our finding that aetiology is an important determinant influencing survival may be explained by several factors. A different propensity to develop hepatocellular carcinoma (HCC) among several aetiologies is one of these. Although it has been described, HCC in patients with IAI-CAH^{481,482} and HCV-CAH^{483,484} is rare. However, 70% of HBsAgpositive patients develop cirrhosis and 30 percent died within 10 years.^{485,486} These HBsAgpositive patients have an increased risk of the development of hepatocellular carcinoma (HCC),⁴⁸⁷ even if a patient has become an asymptomatic carrier.⁴⁸⁸ During the current study HCC was detected in two of the 186 CAH-patients: one with HBsAgpositive CAH, the other with CAH/PSC.

Other studies showed that corticosteroids in severe autoimmune CAH induce remission in 60 percent of the patients within 3 years, while 5-year survival is approximately 90 percent.⁴⁷¹ The cumulative remission rate for therapy of IAI-CAH with a combination of fixed dose prednisolone and azathioprine is 90%.⁴⁷⁰ From 151 treated HBsAgnegative patients, 92% survived 5 years and 86% survived 10 years, while in 22 HBsAg-positive (65% also HBeAg-positive) patients, survival was lower: 73% and 65% at the same intervals (p<0.01). While the survival curve in treated HBsAg-negative CAH declines gradually, highest mortality in HBsAg-positive CAH occurred within the first 2 to 3 years and was quite low thereafter.⁴⁸⁹ Our findings in untreated HBsAgpositive CAH and treated IAI-CAH were similar. Untreated IAI-CAH patients however, followed a course similar to HBsAg-positive CAH. Immunosuppressive therapy in IAI-CAH leads to a marked enhancement of survival in non-viral (idiopathic 'autoimmune') CAH: In the Mayo experience, the 10-year probability of survival without hepatic failure is 98 percent for patients without cirrhosis at presentation. For patients with cirrhosis prior to treatment, the likelihood of survival without hepatic failure is lower: 80 percent after 5 years and 65 percent after 10 years.^{490,491} Interestingly, patients who develop cirrhosis during therapy have a 5-year survival after the development of cirrhosis that is similar to that of patients who do not develop cirrhosis,492 indicating that the propensity to develop cirrhosis in IAI-CAH depends more on the rapidity and completeness of the response to treatment than on any histologic feature at presentation.⁴⁹² However, the presence of cirrhosis can not be excluded by a liver biopsy,^{493,494} so that prospective studies using other methods, are required to assess the prognostic value of the presence of cirrhosis. The ability to suppress inflammatory activity with subsequent repair and restoration of liver function may be a more important prognostic factor, since in the current study the decision to institute immunosuppressive therapy is among the most important prognostic factors.

The fact that autoimmune features had no prognostic significance, neither in our findings nor in a previous report⁴⁹⁵, may be due to selection of patients with the greatest abnormality of these factors for therapy. The same applies to CHE levels in the current study. Indeed, Page et al. observed that patients with an acute onset of disease, evidence of multisystemic involvement, the LE cell phenomenon, and extreme hypergammaglobulinaemia were more likely to respond to corticosteroids than others.⁴⁶⁵ Later on it was demonstrated, however, that the presence or absence of autoimmune markers does not influence the response to corticosteroids in patients with severe CAH, screened for drug-induced or virus-related disease. The frequency of remission, treatment failure, hepatic death, and progression to cirrhosis were similar in patients with and without the LE cell phenomenon or antinuclear antibody (ANA). Our study supports the policy of applying the same program of management to all patients with idiopathic, presumably autoimmune, CAH of sufficient severity, regardless of immunologic findings.⁴⁹⁵ It also favors immunosuppressive treatment of patients with other aetiology, as long as no replicating pathogenetically involved virus like HBV or HCV is present. The current study also suggests that anti-HBs positive patients should be treated like IAI-CAH patients.

To conclude, the degree of inflammation will determine the extent of liver failure by influencing hepatocyte function via cytokines, and by cell necrosis and hence reduction of the number of functional hepatocytes, and is connected with the induction of cirrhosis and its complications. Hepatitis will especially influence prognosis via these mechanisms. The ability to institute immunosuppressive therapy is the most important prognostic factor. This means that in idiopathic 'autoimmune' CAH, the initial reaction

to immunosuppressive therapy must be the most important prognostic factor. The same applies to anti-HBs positive patients, and probably also to other CAH-patients without replicating HBV or HCV, who fulfill requirements for treatment. An accurate analysis of the early effects of immunosuppressive therapy -e.g. on liver function- has been published recently, confirming the importance of initial reaction to immunosuppressive treatment for prognosis.⁴⁹⁶ The combination of that study and the current data on long-term survival can hopefully lead to improved decision making in CAH. The long-term prognostic model we derived from the current study has the advantage over a different recently published model⁴⁹⁷ in that it is confined to CAH and does not need liver biopsy data. Our survival model has to be cross-validated on an independant data set from other patients, however. Therefore, further studies in this field are warranted.

Section 3

'Autoimmune' chronic active hepatitis: Activity of disease and repair during standarized treatment

CHAPTER 7

ROUTINE BLOOD TESTS IN STANDARDIZED IMMUNOSUPPRESSIVE THERAPY OF 'AUTOIMMUNE' CHRONIC ACTIVE HEPATITIS

§7.1

ROUTINE 'LIVER BIOCHEMISTRY' IN STANDARDIZED IMMUNOSUPPRESSIVE THERAPY OF 'AUTOIMMUNE' CHRONIC ACTIVE HEPATITIS

SUMMARY

We analyzed routine liver biochemistry from 21 patients with idiopathic autoimmunechronic active hepatitis (IAICAH), before and during standardized immunosuppression. ALT, AST and gammaglobulin (GG) (medians 337 U/l, 359 U/l, and 28 g/l at start respectively) improved considerably (median 20 U/l, 20 U/l and 12 g/l after two years), while conjugated bilirubin (median 22.5 μ mol/l) normalized within two months to median 2.5 μ mol/l. Median unconjugated bilirubin was normal (14 μ mol/l), but declined to 9 μ mol/l in the first two months of therapy. Results from the current study indicate that standardized immunosuppression with a fixed dose schedule was able to improve biochemistry within 2 months, with further improvement thereafter.

INTRODUCTION

Idiopathic 'autoimmune' chronic active hepatitis (IAI-CAH) usually reacts favorably to immunosuppressive therapy,⁴⁹⁸ and immunosuppression has been shown to enhance survival in this type of CAH.⁴⁹⁹⁻⁵⁰³ Remission has been induced in 60% of patients within 2.5 years of treatment in the Mayo Clinic studies.^{501,504} A combination of fixed dose prednisone and azathioprine daily proved superior to fixed dose, titrad or alternate day therapy with prednisone alone, when regarding efficacy and adverse effects of therapy.⁴⁹⁹ Azathioprine alone cannot induce remission in IAI-CAH,⁴⁹⁹ but can maintain remission without prednisone in a selected group of patients.⁵⁰⁵ In order to assess the effects of a combination of fixed dose prednisolone and azathioprine on histology and routine liver tests we analysed protocol liver biopsies and biochemistry at 0,2,14 and 26 months after commencement of accordingly standardized immunosuppression in 21 patients with symptomatic 'autoimmune' CAH.

MATERIALS AND METHODS

Patients

We studied 21 untreated patients with idiopathic 'autoimmune' chronic active hepatitis (IAI-CAH)- 3 of them with antibodies to hepatitis B surface antigen: 4 men and 17 women. These patients required immunosuppressive therapy according to accepted criteria.⁴⁹⁹ Their median age was 52 years (range 16-71), and median duration of disease 75 days (range 0 months to 3 years). All were subsequently treated with oral prednisolone 15 mg and azathioprine 75 mg daily for two months, followed by at least one year (2 patients) or two years (19 patients) of therapy with 10 mg prednisolone and 50 mg azathioprine daily. Serial routine liver biopsies and sera were studied at commencement and at two months, one and two years after the beginning of therapy. One biopsy at 2 months, one at 14 months and 6 at 26 months were missing, although biochemistry was available.

Serum biochemistry

Sera were obtained on the same day on which the liver biopsies were performed. Alanine aminotransferase (ALT) and aspartate aminotransferase (upper reference limit 30 U/l), total and conjugated bilirubin (UB and CB, with upper reference limits of 26 and 5 μ mol/l respectively), and total protein (TP)(g/l) were determined automatically (Technicon SMA-C). Unconjugated bilirubin (UB) values were calculated by subtracting CB from TB (upper reference value 21 μ g/l). The summation of gammaglobulin (GG) was determined in percentage of TP by electrophoresis, and values were calculated by multiplication with TP (upper reference limit 20 g/l).

Statistical analysis

Non-parametric statistical analysis was performed with Friedman's analysis of variance (ANOVA), Wilcoxon's matched pairs test (p-values shown in figures), and Mann-Whitney U test where appropriate, with p<0.05 as the level of significance.

RESULTS

Before therapy, ALT levels were markedly elevated in all patients (median 337 U/l). These declined during the first two months of therapy to median 32 U/l (p<.001), with stabilization thereafter at median 20 U/l, both at 14 and 26 months (figure 7.1.1A). AST values at commencement were median 359 U/l, with reduction to 32 U/l after two months (p<.001), and a trend to further improvement over the next two years (p=.06), with median values of 24 and 20 at 14 and 26 months respectively (figure 7.1.1B).



Figure 7.1.1. A) ALT, B) AST and C) gammaglobulin (GG) in 'autoimmune' CAH and changes during immunosuppression. Dotted lines indicates upper reference limits (N), upper dotted lines in ALT and AST indicate 2xN.



Figure 7.1.2 A) total, B) conjugated and C) unconjugated and bilirubin in 'autoimmune' CAH and changes during immunosuppression. Dotted lines indicate upper reference limits.

Gammaglobulin levels (n=17) were elevated at the start with a median value of 28 mg/l, falling to median 13.5 at two months (p<.005), and stabilization thereafter, with median values of 15 and 12 at 14 and 26 months respectively (figure 7.1.1C).

Direct serum bilirubin (CB) was elevated at commencement in 15 (70%) of the 21 patients, with a median value of 22.5 μ g/l, which is 4.5 times the upper reference limit. Levels were reduced to median 2.5 μ g/l after two months of therapy (p<.001), and to normal values in all patients after 14 months (median 2 μ g/l) (p<.05), with stabilization during the next year (median 2) (figure 7.1.2B).

Indirect bilirubin (UB) was elevated in only 6 (30%) of the 21 patients, with a median of 14 μ g/l initially, which is within the reference range. These values fell to 9 μ g/l after two months, and stabilized during the next two years (median 9 at 14 and 26 months) (figure 7.1.2.C).

This resulted in a reduction in total bilirubin (TB) levels from median 34.5 μ g/l at commencement to median 12 μ g/l after two months (p<.001), with stabilization thereafter (median 11 at both 14 and 26 months) (figure 7.1.2A).

Correlations

For correlations the reader is referred to chapter 8.

DISCUSSION

With standardized immunosuppression routine serum liver biochemistry improved considerably in patients with previously untreated idiopathic 'autoimmune' IAI-CAH who had required such therapy.

During active disease the increase in serum conjugated bilirubin values was more pronounced and frequent than that of unconjugated bilirubin levels. This may indicate that the uptake of bilirubin by the hepatocytes is less disturbed than the excretion of conjugated bilirubin in IAI-CAH. Both normalized rapidly during therapy, even in the patient whose histology failed to improve.

Often values of AST and ALT did not normalize completely. It is well-known that serum ALT, gammaglobulin and bilirubin, using standard reference ranges, do not accurately predict histological activity.⁵⁰⁶ Follow-up biopsies to monitor inflammatory activity are therefore needed as long as no better serological parameters of activity are available. In a biopsy of sufficient size,⁵⁰⁷ sampling error for determination of inflammatory activity in CAH is less than 10%,⁵⁰⁸ and intra-observer error less than 6%.⁵⁰⁸ In other forms of liver disease the inter- and intra observer variability in grading the degree of piecemeal necrosis is greater than 50 and 40%, respectively.⁵⁰⁹ In the next chapter we will therefore assess histology and relate this to serum biochemistry.

SERUM SODIUM, DEPRESSED IN 'AUTOIMMUNE' CHRONIC ACTIVE HEPATITIS, NORMALIZES DURING IMMUNOSUPPRESSIVE THERAPY

Relation to Histology, Synthesizing Capacity of the Liver and Routine Serum Biochemistry

SUMMARY

Although idiopathic 'autoimmune' chronic active hepatitis (IAI-CAH) usually reacts favorably to immunosuppressive therapy, it is unknown whether functional renal failure as the onset of hepatorenal syndrome (HRS) complicating IAI-CAH, is reversible without liver transplantation.

We studied serum sodium values (Na⁺) in IAI-CAH before and during standardized immunosuppressive therapy with prednisolone and azathioprine. Na⁺ in IAI-CAH (n=17) was lower than in healthy controls (n=105), and in 12 (70%) of the patients Na⁺ was below the 99% lower limit of the reference range. Na⁺ increased during therapy in all patients tested, from median 137 mmol/l at start to median 141 mmol/l after two months (p<.01), with a trend to further increase in the next year (p=.09), although the median remained 141 mmol/l, with finally normal values in all patients tested. The median Na⁺ after two years was 141 mmol/l. Serum creatinin (Creat)increased from median 63 µmol/l at start of therapy to median 68 µmol/l after two months (p<.05), with medians of 72 and 62 µmol/l after 14 and 26 months respectively. Blood urea nitrogen (BUN) values increased from median 4.2 mmol/l at start to median 5.1 mmol/l after 14 months of therapy (<.001). These increases in Creat and BUN paralleled those in serum parameters of hepatic protein synthesizing capacity.

In untreated IAI-CAH Na⁺ was negatively correlated to ALT (R=-.46) and gammaglobulin values (R=-.67). BUN positively correlated with antithrombin III values (R=.56).

In conclusion, serum sodium in IAI-CAH is below normal, probably as a result of reduced free water clearance, representing the onset of HRS. In IAI-CAH reacting to immunosuppression, this condition is reversible.

INTRODUCTION

Hepatorenal syndrome (HRS) is a condition associated with end-stage liver disease. The onset of this condition is usually termed 'functional renal failure' in liver disease. The kidneys from patients with HRS are normal, the function of these kidneys recovers if
they are used for transplantation,⁵¹⁰ and renal function usually normalizes after succesful liver transplantation in patients with HRS,^{511,512} indicating the functional nature of the renal disorder in HRS. We recently showed that in patients with idiopathic 'autoimmune' chronic active hepatitis (IAI-CAH) reacting favorably to immunosuppressive therapy, hepatic protein-synthesizing capacity (HPSC) can normalize despite the presence of cirrhosis.⁵¹³ Whether this therapy can reverse HRS accompanying IAI-CAH is unknown.

Hyponatraemia in HRS is a consequence of reduced free water clearance (FWC) and it is a constant finding in patients with a FWC of 1 ml/min or less, while it is often considered an infrequent finding if patients excrete free water above this level.⁵¹⁴ However, if one determines Na⁺ in a healthy population the 99% reference range is small (between 138 and 146 mmol/l), and Na⁺ in a heterogenous population of patients with various liver diseases is lower than normal, even in the absence of cirrhosis.⁵¹⁵

In the current study we investigated serum sodium values (Na⁺) in patients with idiopathic 'autoimmune' chronic active hepatitis (IAI-CAH) and no signs of renal disease before and during standardized immunosuppressive therapy with prednisolone and azathioprine. Values were related to histology, serum creatinin, blood urea nitrogen, alanine aminotransferase, gammaglobulin, and parameters of protein synthesizing capacity.

MATERIALS AND METHODS

Patients

17 patients with IAI-CAH, 5 men and 12 women, all requiring immunosuppressive therapy according to accepted criteria, 516 were studied to evaluate therapy effects on serum sodium (Na⁺), creatinin (Creat) and urea nitrogen (BUN). Their median age was 52 years (range 17-63), and median duration of disease 189 days (range 0-1841). Cirrhosis was present in 16 of these patients as assessed by laparoscopy and liver biopsy. All were subsequently treated with standardized immunosuppression: oral prednisolone 15 mg and azathioprine 75 mg daily for two months, followed by at least one year (14 patients) or two years (6 patients) of therapy with 10 mg prednisolone and 50 mg azathioprine daily. Serial routine liver biopsies and sera were obtained at the start and at two months, one and two years of therapy.

Liver histology

Liver biopsies were processed and examined under code and assigned a Histological Activity Score (HAS), ranging from 0 to 18, which correlates well with conventional ways of scoring histology in CAH, as described.⁵¹⁷ In contrast to the original system,⁵¹⁸ fibrosis/cirhhosis is not included in the HAS. The HAS is expressed as a numerical score (0-18), and is the summation of portal (score 0-4), periportal (score 0-10) and lobular (score 0-4) inflammation and necrosis.

Serum biochemistry

In the routine protocol sera Na⁺, Creat, BUN, Alanine aminotransferase (ALT) (upper reference limit 30 U/l), and total protein (TP) were determined automatically (Technicon SMA-C) in blood taken between 7 and 10 A.M. The summation of gammaglobulin (GG) was determined as a percentage of TP by electrophoresis, and values were calculated by multiplication with TP (upper reference limit 20 g/l). Parameters of hepatic protein synthesizing capacity were determined as described in chapter 9.

Statistical analysis

Statistical analysis was performed with analysis of variance (ANOVA) with repeated measurements, the Mann-Whitney U test, the Wilcoxon matched pairs test and Spearman's rank correlation where appropriate, with p<0.05 as the level of significance.

RESULTS

Na⁺ in IAI-CAH (n=17) was lower than in healthy controls (n=105), and in 12 (70%) of the patients Na⁺ was below the 99% lower limit of the reference range. Na⁺ increased during therapy in all patients tested, from median 137 mmol/l at start to median 141 mmol/l after two months (p<.01), with a trend to further increases in the next year to median 141 mmol/l (p=.09), with finally normal values in all patients. The median Na⁺ after two years was 141 mmol/l (figure 7.2.1A).

Serum creatinin increased from median 63 μ mol/l at start to median 68 μ mol/l after two months (p<.05), with medians of 72 and 62 μ mol/l after 14 and 26 months respectively (figure 7.2.1B).

Blood urea nitrogen values increased from median 4.2 mmol/l at start to median 5.1 mmol/l after 14 months of therapy (p<.001) (figure 7.2.1C).

For results of serum- and plasmabiochemistry and histology the reader is referred to chapter 7.1, 8, 9, 10 and 11.

In untreated IAI-CAH, Na⁺ was negatively correlated to ALT (R=-.46) and gammaglobulin values (R=-.67). BUN positively correlated with antithrombin III values (R=.56). No other correlations were detected.



Figure 7.2.1 A) Serum sodium, B) creatinin and C) blood urea nitrogen in 'autoimmune' CAH and changes during immunosuppression. Lower (A,C) and higher (A,B,C) dotted lines are the lower and higher 99% limits of the reference range.

DISCUSSION

In the current study, we demonstrated that serum sodium levels in IAI-CAH with cirrhosis are below the 99% reference range for a healthy controlpopulation and that these serum Na⁺levels all normalized with standardized immunosuppressive therapy with prednisolone and azathioprine. There was an inverse relation of serum Na⁺ levels with serum ALT and GG levels.

Low Creat and BUN in untreated IAI-CAH probably indicate malnutrition and impaired hepatic protein synthesis and the increase in Creat and BUN with therapy probably indicate an ameliorated state of nutrition and protein synthesis.

Usually reference ranges for serum sodium (Na⁺) are chosen with wide margins, but the use of more strict criteria based on a healthy reference population allows detection of hyponatraemia in many unselected liver patients and IAI-CAH patients. In the absence of other overt signs of hepatorenal failure, low Na⁺ levels in IAI-CAH may reflect the almost subclinical onset of impaired free water clearance, marking the onset of HRS. The current knowledge of pathophysiology of ascites and functional renal failure in cirrhosis has been reviewed recently.^{519,520} Hyponatraemia in HRS is a consequence of reduced free water clearance (FWC),⁵¹⁴ which, in part appears due to peripheral vasodilatation with an increase in antidiuretic hormone (ADH),⁵²¹⁻⁵²³ which is insufficiently counteracted by an increased renal prostaglandin E₂ (PGE₂) production.⁵²⁴ This concept is supported by reduction of free water clearance by administration of nonsteroidal antiinflammatory drugs that inhibit renal PGE₂ production.⁵²⁵

The normalization of Na⁺ values in IAI-CAH without LT but with immunosuppressive therapy, probably indicates that repair and restoration of functional hepatic capacity can reverse HRS. This confirms again the functional nature of the renal disorder in HRS. Furthermore it indicates that not only liver transplantation, but also immunosuppressive therapy in IAI-CAH is able to reverse functional renal failure or maybe even more advanced HRS in corticosteroid responsive patients. The initial response to such therapy an important determinant in both early⁵²⁶ and longterm⁵²⁷ prognosis in this disease.

Currently, we are investigating the effect of immunosuppression in IAI-CAH on serum ADH values.

In conclusion, serum sodium in IAI-CAH is below normal, probably as a result of reduced free water clearance, representing the onset of functional hepatorenal impairment. In IAI-CAH reacting to immunosuppression, this condition is, at least in its early stages, reversible without liver transplantation.

Further studies, including measurements of urine Na⁺, are warranted.

ROUTINE HAEMATOLOGICAL PARAMETERS IN 'AUTOIMMUNE' CHRONIC ACTIVE HEPATITIS, AND CHANGES DURING IMMUNOSUPPRESSION

SUMMARY

We looked at routine haematological parameters in IAI-CAH, and changes in these parameters during fixed-dose immunosuppression with prednisolone and azathioprine. This was done to evaluate the likelihood of induction of cytopenia by azathioprine.

Leucocytes increased from median 5.3×10^9 at the start to 7.8×10^9 at two months (p<.001), with stabilization thereafter at median 7.35×10^9 at one year and 6.3×10^9 at two years. (Friedman 12.30, p<.01).

Thrombocytes did not change overall (Friedman ANOVA), although there was an increase from median 151×10^3 at start to median 173×10^3 at 2 months (p<.05). Values at 14 and 26 months (median 158 and 162) did not differ from start values, although values at 26 months tended to be higher (p=.06).

Haemoglobin increased from median 128 g/l at start to 136 g/l after 2 months (p<.05), further increased to median 145 g/l at 14 months (p<.01), but slightly decreased in the second year of therapy to median 140 g/l (p<.05). Hb at one and two years was higher than at start (p<.001 and p<.01 respectively). (Friedman 15.73, p<.001)).

Haematocrit increased between two months (median 39.7 %) and 14 months (median 41.9 %) (p<.005). Values at 14 and 26 months (median 40.9) were higher than those at start (median 37.7 %), and Ht at 26 months was higher than after 2 months (p<.05). (Friedman 16.23, p<.001).

Mean cell volume of erythrocytes only tended to increase from median 93.0 flat start to median 97.2 fl at two months (p=.06), but no significant changes were observed (medians at 14 and 26 months were 92.8 fl and 94.8 fl respectively) (Friedman 5.10, p=.165).

The present study shows that standardized immunosuppressive therapy with prednisolone and azathioprine in a fixed dose combination schedule in 'autoimmune' CAH can result in improval of routine haematological parameters.

INTRODUCTION

Hypersplenism associated with cirrhosis and portal hypertension in chronic active hepatitis (CAH) is able to induce pancytopaenia.⁵²⁸ Therapy with azathioprine may further reduce the number of blood cells. If cytopaenia is already marked, it has been

recommended to withhold azathioprine and to administer only corticosteroids as a therapy for idiopathic 'autoimmune' (IAI-) CAH.⁵²⁹

To assess the justification for fear of induction of cytopaenia by azathioprine in IAI-CAH, we looked at routine haematological parameters in IAI-CAH, and changes in these parameters during fixed dose immunosuppression with prednisolone and azathioprine.

MATERIALS AND METHODS

Patients

17 patients with IAI-CAH, 5 men and 12 women, all requiring immunosuppressive therapy according to accepted criteria,⁵³⁰ were studied. Their median age was 52 years (range 17-63), and median duration of disease 189 days (range 0-1841). Cirrhosis was present in 16 of these patients as assessed by laparoscopy and liver biopsy. All were subsequently treated with standardized immunosuppression: oral prednisolone 15 mg and azathioprine 75 mg daily for two months, followed by at least one year (14 patients) or two years 6 patients) of therapy with 10 mg prednisolone and 50 mg azathioprine daily. Serial routine liver biopsies and sera were obtained at the start and at two months, one and two years of therapy.

We evaluated therapy effects on the number of peripheral leucocytes (LEU), thrombocytes (THR), haemoglobin content (Hb) of red blood cells, haematocrit (Ht) and mean cell volume (MCV) of erythrocytes.

Liver histology

Liver biopsies were processed and examined under code and assigned a Histological Activity Score (HAS), ranging from 0 to 18, which correlates well with conventional ways of scoring histology in CAH, as described.⁵³¹ In contrast to the original system,⁵³² fibrosis/cirhhosis is not included in the HAS. The HAS is expressed as a numerical score (0-18), and is the summation of portal (score 0-4), periportal (score 010) and lobular (score 0-4) inflammation and necrosis.

Haematology

LEU, THR, Hb, Ht and MCV were determined in blood (taken between 7 and 10 A.M.) by Coulter counter.

Statistical analysis

Statistical analysis was performed with the Wilcoxon's matched pairs tests with p<0.05 as the level of significance.

RESULTS

Results for histology and routine serum biochemistry are shown in chapters 7.1, 7.2 and 8. LEU increased from median 5.3×10^9 at start to 7.8×10^9 at two months (p<.001), with stabilization thereafter at median 7.35×10^9 at one and 6.3×10^9 at two years. (Friedman 12.30, p<.01) (figure 7.3.1A).

THR did not change overall (Friedman ANOVA), although there was an increase from median 151×10^3 at start to median 173×10^3 at 2 months (p<.05). In one patient THR dropped initially, then stabilized after 1 year. Another patient returned to baseline THR after an initial increase. Values at 14 and 26 months (median 158 and 162) did not differ from start values, although values at 26 months tended to be higher (p=.06) (figure 7.3.1B).



Figure 7.3.1 Numbers of A) leucocytes and B) thrombocytes C) Haemoglobin content and D) haematocrit in peripheral blood in 'autoimmune' CAH before and during immunosuppression. Dotted lines indicate upper and lower limits of the reference range.



Hb increased from median 128 g/l at start to 136 g/l after 2 months (p<.05), further increased to median 145 g/l at 14 months (p<.01), but slightly decreased in the second year of therapy to median 140 g/l (p<.05). Hb at one and two years was higher than at start (p<.001 and p<.01 respectively). (Friedman 15.73, p<.001) (figure 7.3.1C).

Ht increased between two months (median 39.7 %) and 14 months (median 41.9 %) (p<.005). Values at 14 and 26 months (median 40.9) were higher than those at start (median 37.7 %), and Ht at 26 months was higher than after 2 months (p<.05). (Friedman 16.23, p<.001) (figure 7.3.1D).

MCV only tended to increase from median 93.0 fl at start to median 97.2 fl at two months (p=.06), but no significant changes were observed (medians at 14 and 26 months were 92.8 fl and 94.8 fl respectively) (Friedman 5.10, p=.165).

DISCUSSION

The present study shows that standardized immunosuppressive therapy with prednisolone and azathioprine in a fixed dose combination schedule in 'autoimmune' CAH can result in improval of routine haematological parameters. This contrasts with the common fear that azathioprine might induce cytopaenia, either through a depressive effect on the bonemarrow or by induction of liverfibrosis/cirrhosis causing worsening of portal hypertension and hypersplenism.

The peripheral cytopenia that is encountered in hypersplenic states can mainly be ascribed to an increased reversible pooling of blood cells in the enlarged spleen. These cells are in dynamic equilibrium with circulating cells and not permanently sequestered. For instance, the splenic platelet pool will only be dependent on two factors: the splenic blood flow on the one hand and intrasplenic transit time on the other. The splenic blood flow is dependent on spleen size and adrenergic activity.⁵³³

The observed beneficial effect of immunosuppressive therapy on leucocytes, Hb, Ht and, at least, the absence of a deleterious effect on the number of thrombocytes, may be due to recruitment of cells from the spleen. This may follow reduction in splenic blood flow and spleen size, when portal hypertension diminishes in relation to a reduction of hepatic inflammation.

Steroid administration has been shown to initially (first 8-12 hours) reduce the number of uncommitted circulating pluripotential haematopoietic cells (CFU-GEMM).^{534,535} An initial depletion also equally affects all subclasses of lymphocytes.⁵³⁶ A rise in CFU-GEMM occurs 24-36 hours after administration of steroids to levels above pre-treatment levels. Steroids reset the normal diurnal variation in numbers of blood cells. Initial high neutrophil counts drop 36 hours post steroid administration, and return to baseline values at about 60 hours. The number of monocytes drops after steroids, but then normalizes without rebound. Committed precursor levels (erythroid burst-forming units (BFU-E) and granulocyte/monocyte colony-forming units (CFU-GM)) showed diurnal variations similar to CFU-GEMM.^{534,535} Unlike these studies, where one dose of steroids was administered, steroid therapy is continued in patients with IAI-CAH, so that the latter effect may prevail. This could be an additional explanation for the improvement in peripheral blood cell counts after commencement of immunosuppressive therapy in IAI-CAH.

Hb may also rise through enhancement of hepatic protein synthesizing capacity.⁵³⁷ No variceal bleedings were observed before or during the treatment period in these patients, although minor gastrointestinal blood loss before therapy cannot be completely ruled **out**.

Addition of azathioprine to the therapeutic regimen can allow lower doses of prednisolone in order to achieve the same therapeutic results with lower rates of adverse effects. We feel that mild and moderate cytopaenia should no longer be considered as an absolute contraindication to the use of azathioprine. In severe cytopaenia in a patient with IAI-CAH it might be wise, however, to start induction-therapy with prednisolone alone and to evaluate the possibility (dependent on improval of routine haematology) to add azathioprine after e.g. one or two months. In all patients it is, of course, advisable to monitor blood cell counts frequently especially in the first weeks of immunosuppressive therapy for IAI-CAH, especially if azathioprine is included in such therapy. This is particularly important to identify the occasional patient susceptible to azathioprine-induced cytopaenia.⁵³⁷a-^{537d}

Peripheral blood cell counts will improve in the majority of patients with IAI-CAH during therapy with prednisolone and azathioprine.

CHAPTER 8

SERIAL HISTOLOGY AND ROUTINE LIVER BIOCHEMISTRY IN STANDARDIZED IMMUNOSUPPRESSIVE THERAPY OF IDIOPATHIC 'AUTOIMMUNE' CHRONIC ACTIVE HEPATITIS.

SUMMARY

We analyzed histology and routine liver biochemistry from 21 patients with idiopathic autoimmune chronic active hepatitis (CAH) before and during standardized immunosuppression. Portal (P), periportal (PP) and lobular (L) inflammation were scored, and the summation of these produced a histological activity score (HAS) (0-18). A Histological Fibrosis/Cirrhosis Score (HFS) (0-4) was produced, and the numbers of four cell types scored in a 0-3 scale. Values before and changes during therapy were correlated to each other and with routine serum liver biochemistry.

At two months, 67 percent of the patients had already improved to a HAS \leq 3. A HAS \leq 1 ('complete histological remission') was achieved in 80 percent of the patients and 1<HAS \leq 3 ('partial histological remission') in 10 percent of the patients within two years. Periportal and lobular inflammation had disappeared in all of these 19 (90%) out of 21 patients in remission.

Continuing decrease of portal and periportal inflammatory activity with disappearance of plasma cells and decrease in numbers of lymphocytes was observed during the two years of immunosuppressive therapy.

HFS was high (median score 3) and did not change during therapy.

ALT, AST and gammaglobulin (GG) (medians 337 U/l, 359 U/l, and 28 g/l at start respectively) improved considerably (median 20 U/l, 20 U/l and 12 g/l after two years), while conjugated bilirubin (median 22.5 μ mol/l) normalized within two months to median 2.5 μ mol/l. Median unconjugated bilirubin was normal (14 μ mol/l), but declined to 9 μ mol/l in the first two months of therapy.

In untreated IAI-CAH the HAS, PP and number of plasma cells were weakly correlated to ALT (R=.41, .41, and 48 respectively) and did not correlate to the other serum biochemical parameters. L correlated only with GG (R=.50). ALT, AST, GG, CB and TB did correlate to the number of (peri)portal lymphocytes (R=.51, .52, .63, .43, and 43). Correlations among histological parameters and among serum biochemical parameters are shown as well.

In conclusion, results from the current study indicate that standardized immunosuppression with a fixed dose schedule was able to improve histology in 95%, with induction of 'complete histological remission' (HAS \leq 1) in 80% and 'partial histological remission' (HAS \leq 3, but >1) in 10% of the patients with moderate to severe 'autoimmune' CAH. We propose that HAS \leq 3 is an objective of therapy, while a HAS \leq 1 and the absence of plasma cells in the biopsy may be required before an attempt at discontinuation of therapy can be successful. However, determination of satisfactory endpoints for treatment and establishing optimal guidelines for discontinuation of therapy remains a nagging clinical problem.

INTRODUCTION

Idiopathic 'autoimmune' chronic active hepatitis (CAH) usually reacts favorably to immunosuppressive therapy,⁵³⁸ and immunosuppression has been shown to enhance survival in this type of CAH.⁵³⁹⁻⁵⁴³ Remission has been induced in 60% of patients within 2.5 years of treatment in the Mayo Clinic studies.^{541,544} A combination of fixed dose prednisolone and azathioprine daily proved superior to fixed dose, titrad or alternate day therapy with prednisolone alone, with regard to efficacy and adverse effects of therapy.⁵³⁹ Azathioprine alone cannot induce remission in CAH,⁵³⁹ but can maintain remission without prednisolone in a selected group of patients.⁵⁴⁵ The descriptive nature of conventional systems for scoring histological activity of CAH^{546,547} hampered the systematic study of changes in histology during therapy. Recently a numerical scoring system for asymptomatic CAH was introduced,⁵⁴⁸ and a modified 'Histological Activity Score' correlated well with the conventional way of scoring activity in CAH.⁵⁴⁹

To assess the effects of a combination of fixed dose prednisolone and azathioprine on histology and routine liver tests, we analysed protocol liver biopsies and biochemistry at 0,2,14 and 26 months of such standardized immunosuppression in 21 patients with symptomatic 'autoimmune' CAH.

MATERIALS AND METHODS

Patients

We studied 21 untreated patients with idiopathic 'autoimmune' chronic active hepatitis (CAH) -3 of them with antibodies to hepatitis B surface antigen: 4 men and 17 women. These patients required immunosuppressive therapy according to accepted criteria.⁵³⁹ Their median age was 52 years (range 16-71), and median duration of disease 75 days (range 0 months to 3 years). All were subsequently treated with oral prednisolone 15 mg and azathioprine 75 mg daily for two months, followed by at least one year (2 patients) or two years (19 patients) of therapy with 10 mg prednisolone and 50 mg azathioprine daily. Serial routine liver biopsies and sera were studied at the start and at two months, one and two years of therapy. One biopsy at 2 months, one at 14 months and 6 at 26 months were missing, although biochemistry was available.

Histology

Routine liver biopsies were obtained by using the Menghini technique. After fixation with formalin, biopsies were embedded in paraffin. Four micron sections were stained with hematoxylin-eosin, periodic acid-Schiff with previous diastase digestion, Perl's iron stain, Gomori's reticulin stain and Azan collagen stain. The biopsies were reviewed under code without knowledge of the patient's identity or previous histological diagnosis (by J.G. and B.v.H.). The numerical scoring system as described was used.548 This system uses four histological categories: I)periportal inflammation (PP) (piecemeal necrosis) with or without bridging necrosis; II) intralobular degeneration and focal necrosis (L); III) portal inflammation (P); and IV) fibrosis/cirrhosis. A numerical score was assigned ranging from 0 (no alteration) to 10 for category I, and from 0 to 4 for categories II to IV. As the score for inflammation (Categories I to III) and the "Histologic Fibrosis/cirrhosis Score" (HFS) (Category IV) might vary in opposite directions during follow-up, the four categories were monitored separately in the individual patient, and the summation of categories I, II, and III was termed "Histological Activity Score" (HAS).549 In doing so, the combined score for piecemeal necrosis and bridging necrosis weighed more heavily than the separate scores for the other three categories. We showed that this system provides more accurate information of histological activity than conventional scoring does, while it correlates well with conventional grading of CAH.549 'Partial histological remission' was defined as HAS≤3, and HAS≤1 was termed 'complete histological remission'.

In addition, the composition of the inflammatory infiltrate was studied by semiquantitative grading of the numbers of lymphocytes, plasma cells, eosinophils and polymorphonuclear leucocytes in a 0-3 scale, before and during immunosuppression.

Serum biochemistry

Sera were obtained on the same day on which the liver biopsies were performed. Alanine aminotransferase (ALT) and aspartate aminotransferase (upper reference limit 30 U/l), total and conjugated bilirubin (UB and CB, with upper reference limits of 26 and 5 μ mol/l respectively), and total protein (TP)(g/l) were determined automatically (Technicon SMA-C). Unconjugated bilirubin (UB) values were calculated by sub-tracting CB from TB (upper reference value 21 μ g/l). The summation of gammaglobulin (GG) was determined as a percentage of TP by electrophoresis, and values were calculated by multiplication with TP (upper reference limit 20 g/l).

Statistical analysis

Non-parametric statistical analysis was performed with Friedman's analysis of variance (ANOVA), Wilcoxon's matched pairs test (p-values shown in figures), and Mann-Whitney U test where appropriate, with p<0.05 as the level of significance.

RESULTS

Histology

- Fibrosis/cirrhosis:

As shown in figure 8.1, initial scores (0-4) of hepatic fibrosis/cirrhosis (HFS) were high (median 3), and did not change during the 26 months of immunosuppressive therapy (median 3.5, 3, and 3 at 2, 14 and 26 months respectively) (p=.938 in ANOVA).



Figure 8.1 Histological Fibrosis Score (HFS) in 'autoimmune' CAH before and during immunosuppressive therapy.

- Inflammation/necrosis:

Results and the sequence of changes are shown in figures 8.2A-C and figure 8.3. Portal inflammation scores (0-4) (P) declined from median 3 at start of therapy to median 1 within the first 14 months of therapy (p<.05), and stabilized thereafter. Median values at 2 months and two years of therapy were both 1.

Periportal inflammation scores (0-10) (PP) fell from median 5 at start of therapy to median 0.5 at two months of therapy (p<.001), with further reduction to median 0 after 26 months (p<.05). The median PP score at 14 months was also 0.

Lobular inflammation scores (0-4) (L) were quite low at the start of therapy (median score 1). Lobular inflammation almost disappeared within two months of therapy (median 0) (p<.005), with a trend to even further reduction in the next two years (median 0 at both 14 and 26 months)(p=.09).



Figure 8.2 Scores of A) portal (P), B) periportal (PP) and C) lobular (L) inflammation in 'autoimmune' CAH before and during immunosuppressive therapy.

As shown in figure 8.3, the 'Histological Activity Score' (HAS), which is the summation of P, PP and L, fell from median 9 at start of therapy to median 3 after two months (p<.001). There was further more gradual improvement over the next two years of therapy with median values of 1 both at 14 and 26 months (p<.05). Eventually 17 (80%) of the 21 patients were in complete histological remission (HAS \leq 1), and 2 (10%) in partial histological remission (HAS \leq 3). CAH remained active in two (10%) of these patients during therapy: HAS only declined from 12 at start to 10 after 14 months in one of these two patients, while it declined from 10 at start to 4 after two years in the other patient. One of the patients with HAS \leq 1 at two months relapsed (HAS 9) at one year during unaltered immunosuppression, but reentered complete histological remission during the second year of unchanged therapy. After a HAS \leq 1 was reached, no other patients showed an increase in HAS to scores above 1 during this therapy. Mild to moderate cholestasis was only present in two patients with severe CAH with impaired synthetic function. This cholestasis disappeared within two months of immunosuppressive therapy.



Figure 8.3 Histological Activity Score (=P+PP+L) in 'autoinumune' CAH before and during immunosuppressive therapy.

-Composition of the inflammatory infiltrate:

With respect to the composition of the inflammatory infiltrate, the results of scoring four different cell types in the inflammatory infiltrate of the liver on a 0-3 scale are shown in figure 8.4.

Lymphocytes dominated the infiltrate, followed by plasma cells, especially in the biopsies prior to therapy. During immunosuppression, the few granulocytes (eosinophils



Figure 8.4 Composition of the inflammatory infiltrate in 'autoimmune' CAH before and during immunosuppressive therapy. Median scores are shown. o: lymphocytes *: plasma cells +: polymorphonuclear leucocytes x: eosinophils

and polymorphonuclear leucocytes) disappeared first, and the reduction in the number of plasma cells was more rapid than the reduction in the number of lymphocytes. After 26 months of therapy, portal lymphocytes remained.

Lymphocytes were invariably present at start (median 2), with a gradual decline over all 26 months of therapy (p<.005), especially in the first 14 months. Median scores of the number of lymphocytes were 2 at two and 1 at 14 months (p<.05), and 1 at 26 months (p=.50), when these were virtually the only inflammatory cells remaining.

The median score of plasma cells at start was 2, with a rapid decline in the first two months to median 0.5 (p<.01), stabilization in the next year of therapy at median 0 (p=.80), and a trend towards further improvement in the second year of therapy in some patients (p=.07), with a median score of 0 after two years.

Few polymorphonuclear leucocytes were present in the infiltrate before therapy (median score 1). They were reduced in number in the first two months of therapy (median score 0) (p<.01), and remained absent in the next two years of therapy (medians 0) (p=.78 and p=.59 respectively).

The few eosinophils (median score 0) at start disappeared in the first two months (median score 0 at 2 months) (p<.001) and did not reappear (median 0 at 14 and 26 months) (p=.18 and .32 respectively).

Serum biochemistry

For these results the reader is referred to chapter 7. For ALT the results are also shown in figure 8.5.



Figure 8.5 ALT in 'autoimmune' CAH before and during immunosuppressive therapy. Dotted lines indicate upper reference limit (N) and 2 N.

Correlations

ALT-, AST-, bilirubin-, and gammaglobulin values were related to each other and to parameters of liver histology in the 21 patients before institution of therapy. In untreated IAI-CAH the correlations between histological parameters, between serum biochemical parameters, and between histological and biochemical parameters are shown in tables 8.1, 8.2 and 8.3 respectively. No other correlations were detected.

		PP	L	lympho	pmn	eo	plasma	
HAS	R p	.92 5 ·	.77 4	.62 2	.54 1		.56 1	
Р	R p				.37 0			
PP	R p		.64 3	.66 3	.49 1		.45 0	
L	R p			.55 1	.39 0		.60 2	
lympho	R p					.45 1	.46 1	

 Table 8.1.
 Correlations among histological parameters in untreated IAI-CAH.

R=Spearman's rank correlation

p-values: 0:<.10, 1:<.05, 2:<.01, 3:<.005, 4:<.001, 5:<.0005, 6:<.0001.

Abbreviations: (see also text). Lympho=lymphocytes; pmn=polymorphonuclear leucocytes; eo=eosinophils; plasma=plasma cells.

Table 8.2.	Correlations between histological and serum biochemical parameters in
	untreated IAI-CAH.

		ALT	AST	GG	CB	TB	
HAS	R	.41					
	p	0					
PP	R	.41					
	р	0					
L	R			.50			
	р			1			
lympho	R	.51	.52	.63	.43	.43	
	р	1	1	2	0	0	
plasma	R	.48					
-	р	1					

R=Spearman's rank correlation

p-values: 0:<.10, 1:<.05, 2:<.01, 3:<.005, 4:<.001, 5:<.0005, 6:<.0001.

For abbreviations: see text and table 8.1.

		ALT	СВ	UB	TB
AST	R	.82	.80	.66	.81
	p	5	4	3	4
ALT	R		.58	.51	.60
	p		1	1	2
СВ	R			.84	.99
•	p			5	6
ID	D				80
UB	p R				.09

 Table 8.3.
 Correlations among serum biochemical parameters in untreated IAI-CAH.

R=Spearman's rank correlation

p-values: 0:<.10, 1:<.05, 2:<.01, 3:<.005, 4:<.001, 5:<.0005, 6:<.0001.

Abbreviations: see text.

DISCUSSION

With standardized immunosuppression complete (HAS \leq 1), partial histological remission (HAS \leq 3) was achieved in 90% of patients with previously untreated idiopathic 'autoimmune' CAH requiring such therapy.

Our results therefore confirm the benefits of standardized immunosuppressive therapy in this category of CAH, the anti-HBs positive patients included.⁵³⁸ However, a close follow-up is mandatory, since the current study also confirms that some patients are - partially- resistant to immunosuppressive therapy,⁵⁵⁰ as illustrated by the two patients with no or only partial improvement in histology.

On light microscopy, the inflammatory infiltrate consisted predominantly of cells with the morphology of lymphocytes and plasma cells (IgG-type). Eggink recently showed that the former are predominantly CD8-positive cytotoxic/suppressor cells or killer/ natural killer (OKM1,2+ and CD1-,CD3-) cells.⁵⁵¹

Histology continued to improve during the 26 month period of immunosuppression, with progressive reduction of inflammation due to disappearance of the many plasma cells and of the few granulocytes, and with reduction in the number of lymphocytes. This observation supports the policy of continued immunosuppression for two years before considering a change in therapy.

It is well known that relapse rates can be high, especially after discontinuation of therapy.^{544,552} Investigators from the Mayo Clinic observed relapse rates of 30% in patients who reverted to normal liver biopsies, but 80% if residual portal hepatitis was present at the time of drug withdrawal^{553,554}. We therefore distinguished complete (HAS \leq 1) from partial (HAS \leq 3) histological remission. Interestingly, in 80% of the patients, such a complete histological remission was achieved with the standardized therapeutic regimen. HAS \leq 1 may be one of the requirements that have to be fulfilled before attempting to discontinue therapy. Although this is speculative, another requirement may be the absence of plasma cells in the liver biopsy, since their presence might reflect the potential for antibody-dependent cellular cytotoxicity, which plays a role in 'autoimmune' CAH.⁵⁵⁵

CAH is defined principally by its histological activity rather than by structural changes, such as fibrosis and cirrhosis. In the current study, histological scores of cirrhosis/ fibrosis (HFS) did not differ between 0 and 2, 2 and 12, and 12 and 24 months of immunosuppression. This could be the result of the decrease in inflammation and hence fibrogenesis after the institution of immunosuppression. Another explanation may be the considerable degree of fibrosis/cirrhosis in most patients before immunosuppression -this was confirmed laparoscopically, as biopsies show a considerable sampling error in this respect.⁵⁵⁶

During active disease, the increase in serum conjugated bilirubin values was more pronounced and frequent than that of unconjugated bilirubin levels. This may indicate that the uptake of bilirubin by the hepatocytes is less disturbed than the excretion of conjugated bilirubin in CAH. Both normalized rapidly during therapy, even in the patient whose histology failed to improve.

In accordance with the literature,⁵⁵⁷ serum ALT, gammaglobulin and bilirubin, using standard reference ranges, did not accurately predict histological activity (table 8.2). This may be partly due to differences in long-time after the start of immunosuppression. Follow-up biopsies to monitor inflammatory activity are therefore needed as long as no better serological parameters of activity are available. In a biopsy of sufficient size,⁵⁵⁸ sampling error for determination of inflammatory activity in CAH is less than 10%,⁵⁵⁹ and intra-observer error less than 6%,⁵⁵⁹ while in other forms of liver disease the inter-and intra-observer variability in grading the degree of piecemeal necrosis is greater than 50% and 40%, respectively.⁵⁶⁰

The patients in this study underwent a control biopsy after 2 months of standardized therapy, with subsequent biopsies after one and two years of therapy. As long as no non-invasive means for assessing histological activity have been established, it may be advisable to continue induction-therapy with 15 mg prednisolone and 75 mg azathio-prine for at least two months, with a control liver biopsy after two months of therapy. The two month period is appropriate, since major reductions in inflammatory activity

take place in the first two months of therapy in responding patients. This allows therapy failure to be identified.

If after two months, the HAS is still more than 3 (despite improvement), one might continue induction-therapy for another two months, with a further biopsy at the end. If, however, there is worsening despite therapy, especially if there is a ≥ 67 percent increase in serum bilirubin and/or AST (ALT) level, the first step is usually to double the dose to 30 mg prednisolone in conjunction with 150 mg azathioprine daily.⁵⁵⁰

Further liver biopsies in patients with a HAS \leq 3 at two months can probably be limited to cases of major worsening of serum biochemistry, or before attempting to discontinue therapy. This is justified, since relapse during unchanged therapy was unusual after induction of remission.

Many of these control liver biopsies may become superfluous if new serological markers of disease-activity can be found.

In conclusion, results from the current study indicate that standardized immunosuppression with a fixed dose schedule improved histology in 95%, with induction of 'complete histological remission' (HAS \leq 1) in 80% and 'partial histological remission' (HAS \leq 3, but >1) in 10% of the patients with moderate to severe 'autoimmune' CAH. We propose that HAS \leq 3 is an objective of therapy, while a HAS \leq 1 and the absence of plasma cells in the biopsy may be required before an attempt at discontinuation of therapy can be successful. However, determination of satisfactory endpoints for treatment and establishing of optimal guidelines for discontinuation of therapy remains a nagging clinical problem.⁵⁶¹

CHAPTER 9

REPAIR AND NORMALIZATION OF HEPATIC FUNCTIONAL CAPACITY OF PROTEIN SYNTHESIS, DESPITE PRESENCE OF CIRRHOSIS IN 'AUTOIMMUNE' CHRONIC ACTIVE HEPATITIS

SUMMARY

We evaluated the effect of immunosuppressive therapy on serum parameters of hepatic protein synthesizing capacity in 'autoimmune' chronic active hepatitis (CAH) in relation to histological activity of disease. Serum levels of albumin (ALB) were abnormal in 71%, of cholinesterase (CHE) in 76%, and antithrombin III (AT III) in 82%. Prothrombin time (PT) was abnormal in 42% and activated partial thromboplastin time (APTT) in 74% of the patients before therapy. All parameters improved during the first 14 months of therapy and normalized in all patients during two years of standardized immunosuppressive therapy with prednisolone and azathioprine. This was despite the presence of advanced fibrosis/cirrhosis in 20/21 of the patients at the start. CHE levels even increased during the second year of therapy.

Hepatic protein synthesizing capacity normalized in all patients with 'autoimmune' CAH in the course of two years of immunosuppressive therapy. CHE appears to be the most useful parameter for the routine follow-up of hepatic protein synthesizing capacity in CAH, while the other parameters can provide additional information. Normalization of hepatic protein synthesizing capacity is an important endpoint of therapy. Its normalization can take two years, probably reflecting ongoing repair and regeneration. This supports our current practice of continuing standardized immunosuppression for two years before considering an attempt to withdraw therapy.

INTRODUCTION

In HBsAg-negative chronic active hepatitis (CAH) protein synthesizing capacity of the liver can be severely impaired, as reflected in a low serum albumin (ALB) and a prolonged prothrombin time (PT),⁵⁶²⁻⁵⁶⁵ especially if cirrhosis is present.⁵⁶⁶ Hypo-albuminaemia and a prolonged PT were recognized with similar frequency in patients with and without extreme (>1000 IU/l) aminotransferase elevation.⁵⁶⁷ Patients who sustain remission cannot be discerned from those who relapse after first treatment on the basis of serum albumin values.⁵⁶⁸

Serum cholinesterase (CHE) is a glycoprotein with a molecular weight of 350 kD, predominantly derived from pseudocholinesterase, which is exclusively synthesized in

the liver.^{569,570} Its hypothesized function is to prevent inhibition of acetyl-cholinesterase.⁵⁷¹⁻⁵⁷³ Reduced serum concentrations of CHE have been observed frequently in patients with CAH⁵⁷⁴ and in other forms of hepatocellular damage,^{575,576} almost paralleling reductions in albumin,^{577,578} while a normal CHE virtually excludes severe impairment of liver function.⁵⁷⁹ A sudden drop in CHE can precede fulminant hepatic failure with hepatic coma in acute and chronic hepatitis.⁵⁷⁴ The preoperative serum CHE level correlates with outcome of portocaval surgery in liver disease,⁵⁷⁶ and monitoring of CHE serum levels proved useful in the follow-up after liver transplantation.^{578,580} CHE levels, like the prothrombin index, correlated highly with the Child-Turcotte score,⁵⁸¹ and are of prognostic significance.^{582,583}

Antithrombin III (ATIII) is a glycoprotein, a protease inhibitor, exclusively synthesized in the liver. Its levels, like those of CHE, are prognostic factors for blood loss during liver transplantation,⁵⁸⁴ and ATIII levels have been shown to be reduced in liver cirrhosis in parallel to serum albumin values.⁵⁸⁵⁻⁵⁸⁹ These reductions in ATIII levels contribute to an increased fibrinogen turnover, which might reflect low-grade intravascular coagulation.⁵⁹⁰

Early death in untreated CAH is usually caused by liver failure, while late mortality is more frequently due to complications of cirrhosis.⁵⁹¹ In idiopathic 'autoimmune' CAH, immunosuppressive therapy markedly enhances remission rates and survival, ⁵⁹²⁻⁵⁹⁴ although cirrhosis is frequently present at the time of diagnosis.⁵⁹⁵ Indeed, the presence of cirrhosis before the institution of therapy has been shown to adversely affect the response to treatment⁵⁹⁶ and the likelihood of survival without hepatic failure,⁵⁹⁴ although the development of cirrhosis during immunosuppression does not affect survival.⁵⁹⁷ The conclusion is that the gain in survival appears predominantly to be the result of improved liver function. Our knowledge about changes in liver function during treatment of 'autoimmune' CAH is limited, however.

Elevated serum bile acid levels (SBA) in CAH were reported to normalize with achievement of remission,⁵⁹⁸ and SBA levels appeared to correlate with tests such as bromosulfophtalein, and indocyanine green and the aminopyrine breath test.^{599,600} However, the value of these tests in assessing liver function in clinical practice remains to be established.

With regard to the hepatic protein synthesizing function, Soloway *et al.* showed that an abnormal PT occurs in about 40% of the CAH-patients before institution of therapy with prednisone and azathioprine, with normalization within three months of therapy. Albumin levels were reduced in 80% of these patients, with normal values in half of the 14 patients after one year of therapy.⁵⁹²

The purpose of the current study was to evaluate the effect of standardized immunosuppressive therapy on serum levels of albumin (ALB), cholinesterase (CHE) and antithrombin III (AT III), plasma prothrombin time (PT) and activated partial thromboplastin time (APTT), as parameters of hepatic protein synthesizing capacity in patients with 'autoimmune' CAH.

MATERIALS AND METHODS

Patients

We studied 21 untreated patients with idiopathic 'autoimmune' chronic active hepatitis (IAI-CAH) - 3 of them with antibodies to hepatitis B surface antigen: 4 men and 17 women. These patients required immunosuppressive therapy according to accepted criteria.⁵⁹² Their median age was 52 years (range 16-71), and median duration of disease 75 days (range 0 months to 3 years). All were subsequently treated with oral prednisolone 15 mg and azathioprine 75 mg daily for two months, followed by at least one year (2 patients) or two years (19 patients) of therapy with 10 mg prednisolone and 50 mg azathioprine daily. Serial routine liver biopsies and serum and plasma biochemistry were studied at the start and at two months, one and two years of therapy. One biopsy after 2 months, one at 14 months and 6 at 26 months were missing, although biochemistry was available. Two of the patients received a sublingual vitamin K analogon (10 mg daily) before and during the first three weeks of therapy, and analysis of start values of these two patients were performed after two and three weeks respectively of such therapy.

Liver histology

Liver biopsies were processed and examined under code and assigned a Histological Activity Score (HAS), ranging from 0 to 18, which correlates well with conventional ways of scoring histology in CAH, as described.⁶⁰¹ In contrast to the original system,⁶⁰² fibrosis/cirrhosis is not included in the HAS. The HAS is expressed as a numerical score (0-18), and is the summation of portal (score 0-4), periportal (score 0-10) and lobular (score 0-4) inflammation and necrosis.

Serum and plasma biochemistry

Serum and plasma samples were obtained on the same day on which the liver biopsies were performed. Alanine aminotransferase (ALT) (upper reference limit 30 U/l), total and conjugated bilirubin (UB and CB, with upper reference limits of 26 and 5 μ mol/l respectively), and albumin (ALB)(lower reference value 40 g/l) were determined automatically (Technicon sequential multiple analyzer plus computer [SMA-C]). Unconjugated bilirubin (UB) values were calculated by subtracting CB from TB (upper reference value for UB was 21 μ g/l).

Prothrombin time (PT) (upper reference limit 13 seconds), and activated partial thromboplastin time (APTT) (upper reference limit 13 seconds) were determined in plasma by Coagalyzer (Sherwood) using automated APTT (General Diagnostics) and Activated Thromboplastin (Dale) reagents, while antithrombin III (AT III) (lower reference limit 80%), was measured in plasma using a chromogenic substrate (S-2238 Kabri) as described.⁶⁰³

The serum cholinesterase concentration (CHE) was determined by measuring its catalytic activity, using acetylthiocholine iodide as a substrate⁶⁰⁴ in a commercially available kinetic test (Boehringer Mannheim GmbH). When not determined immediately, stored (-20°C) and coded sera were used. Repeated freezing and thawing does not influence results.⁶⁰⁵ Results of this assay are comparable to those of a more recently described assay.^{606,607} For the reference values of CHE, we used the sera of 77 controls (41 women and 36 men) with apparently normal health: the lower reference limit of CHE is 1600 U/l for women and 1800 U/l for men.⁶⁰⁵ We also compared values obtained before and during immunosuppression in patients to CHE values in 68 sexmatched controls out of this reference population.

Apart from giving absolute results, for reasons of comparison, values of ALB, CHE and AT III are also expressed as a percentage of the lower limit of the normal range. PT and APTT are always expressed as percentages of the upper limit of the normal range of standard control sera determined on the same day.

Statistical analysis

Non-parametric statistical analysis was performed with Friedman's analysis of variance (ANOVA), Wilcoxon's matched pairs test, Mann-Whitney U test and Spearman's rank correlation where appropriate, with p<0.05 as level of significance.

RESULTS

All parameters under investigation, except HFS, improved during immunosuppressive therapy (ANOVA p<.01).

Liver histology

As shown in chapter 8, initial scores (0-4) of hepatic fibrosis/cirrhosis (HFS) were high (median 3), and did not change during the 26 months of immunosuppressive therapy (median 3.5, 3, and 3 at 2, 14 and 26 months respectively) (p=.938 in ANOVA) (figure 9.1).

As shown in figure 9.2, the histologic activity diminished significantly from median 9 to 3 within 2 months (p<.001), with further improvement to median 1 in the next two years of therapy (p<.05), and stabilization thereafter. 19 (90%) of the patients achieved a HAS \leq 3, while 80% achieved a HAS \leq 1. Regarding histology, one patient was therapy-resistant, while a persisting HAS=10, and another patient improved to HAS=4.



Figure 9.1 HFS in IAI-CAH before and during immunosuppression.



Figure 9.2 HAS in 'autoimmune' CAH before and during immunosuppression.

Serum and plasma biochemistry

For results of routine biochemistry the reader is referred to chapter 7.1. Results for ALT and UB are shown in figure 9.3A and 9.3B respectively.



Figure 9.3 A) ALT and B) UB in 'autoimmune' CAH before and during immunosuppression.

ALB serum levels increased from median 32 to 37 g/l after the first two months (p<.05), to median 42 g/l after the next year (p<.001), stabilizing at 42 g/l after the second year (p=.18). Normal values were found in all but three patients (with near-nomal values of 38, 38 and 39 g/l) at the end of this study (figure 9.4A).

CHE serum levels increased from median 1185 to 1698 U/l after the first two months (p<.01) and to median 2323 U/l after the next year (p<.001). Levels increased to 2514 U/l after the second year (p<.05), with normal values in all patients within two years of therapy when compared to the 77 controls (figure 9.4B).



Figure 9.4 A) ALB, B) CHE and C) ATIII in 'autoimmune' CAH before and during immunosuppression. Dotted line indicates lower limit of reference range.



Figure 9.4 D) PT and E) APTT in 'autoimmune' CAH before and during immunosuppression. Dotted line indicates upper limit of reference range.

Compared to the 68 sex-matched controls CHE values in the IAI-CAH patients were below reference values at the start of therapy (p<.0001), after 2 months (p<.0001), and after one year (p<.01). However, CHE values did not differ significantly from reference values at two years of immunosuppression, although values were on the lower side of the reference range (p=.06).

ATIII plasma levels (n=17) increased from median 60% at the start to 69% after the first two months (p<.005), to median 87% after the next year (p<.01), and a median value of 96% after the second year (p=.23). Normal values were found in 16 (94%) out of 17 patients we could evaluate for this parameter within two years of therapy, while one patient had improved from an ATIII of 42% at start to 72% after two years (figure 9.4C).

There is a trend towards worsening of PT after the institution of therapy (p=.07), with a peak between 15 and 30 days after start. PT improved from median 100% at start to median 98% during the first 14 months (p<.05), and from median 105.5 at two months to median 98.5 at 26 months (p<.05). However, there were no major changes between values at start and after two years (p=.14), and Friedman ANOVA did not show changes during therapy either (figure 9.4D).

The APTT fell from median 119% to 110% after two months (p<.05), with further improvement in the next year to median 89% (p<.05), and a trend to further change in the next year (median 91%) (p=.06) (figure 9.4E).

Correlations

All parameters were related to each other and to parameters of liver histology.

In untreated IAI-CAH, the correlations between histological parameters, and between serum and plasma biochemical parameters, and between biochemical parameters is shown in tables 9.1 and 9.2 respectively. No other correlations were detected.

Table 9.1Correlations in untreated IAI-CAH between parameters of serum/plasmabiochemistry and histology (Spearman rank)(p<0.05).</td>

	ALT	UB	CB	TB	ALB	CHE	ATIII	PT	APTT	
Р										
PP	.41									
L								.51		
HAS	.41									
HFS									.56	

	AST	ALT	UB	СВ	TB	ALB	CHE	ATIII	PT	
AST		.82	.66	.80	.81		54			
ALT	.82		.51	.58	.60					
UB	.66	.51		.84	.89	42	31	54		
CB	.80	.58	.84		.99	62	56	56		
TB	.81	.60	.89	.99		58	55	56		
ALB			42	62	58		.74	.78	57	
CHE	54		31	56	55	.74		.65		
ATIII			54	56	56	.78	.65		81	
PT						57		81		
APTT				.44	.42	70	53	75	.55	

 Table 9.2
 Correlations in untreated IAI-CAH between parameters of serum/plasma biochemistry (Spearman rank)(p<0.05). (open spaces indicate lack of significant correlation)</th>

DISCUSSION

Despite the presence of advanced stages of fibrosis/cirrhosis in all of these CAHpatients, standardized immunosuppression was not only able to induce remission, but induced restoration of functional hepatic protein synthesizing capacity. In this regard the results of such therapy in these patients are comparable to those obtained with liver transplantation.⁵⁷⁸

Prothrombin time (PT) tends to worsen immediately after institution of therapy, then improves (data not shown). This may be due to direct inhibitory effect of corticosteroids on synthesis of some clotting factors. Alternatively, there may be an imbalance among clotting factors during restoration of protein synthesis with, for instance, the disappearance of the stimulation of synthesis of coagulation factors like fibrinogen which may be present during the acute phase response. Glucocorticosteroids also have a direct effect on some coagulation tests, resulting in an increase in PT and ATIII, and a decrease in APTT.^{607a} A relative shortage of vitamin K immediately after improvement of protein synthesis may also play a role. However, this would probably have affected the APTT more than PT. It may be worth considering routinely adding sublingual vitamin K (e.g. 10 mg daily) to the medication in the first two months of therapy.

The synthesis of ALB can be inhibited by several cytokines like interleukin-1 β (IL-1 β), interleukin-6 (IL-6), and tumor necrosis factor α (TNF α).⁴¹⁻⁴⁵ Experiments with primary cultures of human hepatocytes and human hepatoma cells showed that II-1 β , IL-6 and TNF- α inhibit liver synthesis of albumin at the transcriptional level in an

additive manner, with IL-6 as the principal inhibitor, 610,611 although monocytes can also produce inhibitory activity to IL-1B. 613,614

The normalization of hepatic functional protein-synthesizing capacity in severe CAH usually takes about two years, and therefore appears to be predominantly the result of repair and regeneration of liver tissue. Because CHE, like albumin,⁶¹⁵ is a negative acute phase reactant, the increment (particularly initially) in CHE levels during therapy may also result from a decline in titres of e.g. IL-1 β , IL-6 and TNF α . The same applies to the coagulation proteins involved in the APTT. Glucocorticoids can also selectively inhibit the transcription of the IL-1 β gene and decrease the stability of IL-1 β mRNA.⁶¹⁶ Furthermore, the rise in ALB, CHE and coagulation proteins may partly result from a direct stimulation of protein synthesis by corticosteroids, as is also observed in hepatocyte cultures.⁶¹⁵

Determination of the PT appeared to be of limited value in CAH. On the other hand, CHE continues to improve, and APTT tends to improve, even during the second year of immunosuppressive therapy, while the other parameters of protein synthesis stabilize during this second year. This may indicate that CHE is a very sensitive parameter of protein synthesis, and is therefore ideal for the follow-up of hepatic protein synthesizing capacity in CAH.

There are well known conditions with elevation or highnormal levels of CHE, such as fatty liver, functional hyperbilirubinaemia, diabetes mellitus, hyperlipidaemia, nephrotic syndrome, protein losing enteropathy, and sometimes in thyrotoxicosis. These elevated serum levels can also be present in patients with familial hypercholinesterasaemia.⁵⁶⁹ In the current study, no such influences were detected. Serum bilirubin levels, if elevated at all, normalized within two months, excluding an influence on CHE levels from the second month of therapy onwards. No elevation of CHE during remission from CAH was detected, not even in the one patient with corticosteroidinduced diabetes mellitus after two months of therapy. A lowering influence of oral contraceptives on CHE levels,^{617,618} AT III (as much as 60% of its normal value),⁶¹⁹ and albumin⁶¹⁷ was described in healthy women. We did not detect such an influence on CHE values.⁶⁰⁵ Since patients with CAH are usually advised not to use such medication, this is not a common problem when interpreting ATIII levels. Higher CHE levels during pregnancy were reported from the third month until six weeks after delivery.⁵⁷²

Serum albumin levels can be influenced by many other factors, as recently decribed.^{620,621} High levels of albumin synthesis can be observed in the presence of ascites. In these patients, the serum albumin level is usually not a reliable index of the hepatic albumin synthesizing capacity. In non-ascitic cirrhotic subjects, however, a depressed serum albumin reflects decreased hepatic protein synthesis.⁶²²

Malnutrition is common in patients with severe liver disease. This can cause serum concentrations of albumin to be low.⁶²² Unpublished observations have found parallel changes in CHE and AT III. The improval of nutritional status during immunosuppressive therapy can also account for part of the increase in hepatic protein synthesis.

CHE levels vary considerably between individuals, but in any one person the level

remains remarkably constant from the third month after birth onwards, so that slight variations, even in the 'normal' area, may indicate changes in liver function^{572,580}. This, combined with its short plasma half-life of 10 days, and with the ease, low cost and rapidity of the assay make CHE an essential parameter in the follow-up of CAH. The other parameters of protein synthesis provide complemetary information, and may be determined at larger intervals during follow-up of CAH.

To conclude, the protein synthesizing capacity of the liver of patients with 'autoimmune' CAH can normalize despite the presence of advanced fibrosis/cirrhosis. In this respect, the standardized immunosuppression mentioned yields results equivalent to those of a succesful liver transplantation. This is in accordance with the knowledge that patients with 'autoimmune' CAH do not usually require liver transplantation if therapy failure (especially if combined with endogenous hepatic encephalopathy, severe hepato-renal syndrome with ascites or recurrent variceal hemorrhage) does not complicate the disease.^{623,624} Follow-up of hepatic protein synthesizing capacity in CAH is of major importance. CHE appeared to be very useful for routine follow-up determinations. Normalization of protein synthesizing capacity is an important endpoint of immunosuppressive therapy in 'autoimmune' CAH. It may require two years of therapy to achieve this goal, indicating that repair and restoration of liver function may take this long, but can eventually be achieved in all patients. This supports our current practice of continuing standardized immunosuppression for two years before considering an attempt to withdraw therapy.

CHAPTER 10

ACUTE PHASE REACTANTS, C-REACTIVE PROTEIN AND SERUM AMYLOID A, IN CHRONIC ACTIVE HEPATITIS

Relation to Histology and Synthesizing Capacity of the Liver and Influence of Immunosuppressive Therapy.

ABSTRACT

The liver is the major site of production of acute phase proteins. We studied the acute phase proteins C-reactive protein (CRP) and serum amyloid A (SAA) in patients with active chronic inflammation of the liver, the main organ producing CRP and SAA. In 81 untreated patients with histologically proven chronic active hepatitis (CAH), the CRP serum levels were only elevated slightly in 54 (67%) and SAA in only 12 (15%) patients. The CRP values were higher than the basal values of healthy controls (p<.0001); this was true for all CAH patients irrespective of aetiology. SAA values in CAH did not differ from basal control values. CRP serum levels correlated with periportal (R=.39, p<.05) and overall (R=.40, p<.05) inflammation, but not to portal or lobular inflammation (n=29). SAA values , however, were correlated to serum cholinesterase (CHE) levels (R=0.61, p<.005), but not to inflammation (n=29). The effects of prednisolone and azathioprine on CRP and SAA in relation to CHE and histology were evaluated in 21 patients during two years of standardized treatment. Histologic activity as measured by the Histological Activity Score diminished significantly from median 9 to 3 within 2 months (p<.001), with further improvement to median 1 in the next two years of therapy (p<.05), and stabilization thereafter. CHE serum levels increased from median 1185 to 1698 U/l after the first two months (p<.01) and to median 2323 U/l after the next year (p<.001), and 2514 U/l after the second year (p<.05). In all patients, CHE values normalized within two years of therapy. CRP levels, median 7.1 mg/l at start, fell to normal (median 0.9 mg/l) within two months (p<0.0005), and stabilized thereafter. SAA increased from median 1.1 mg/l before therapy to 2.1 mg/l after one year of therapy (p<0.05), with stabilization thereafter.

To conclude, CRP and SAA responses appear to be markedly depressed in CAH. CRP did reflect histological inflammation to a minor degree, while SAA did not. Impairment of hepatic capacity to synthesize proteins apparently does affect SAA more severely than CRP. Immunosuppressive therapy resulted in a histology of 'CAH in remission' and normal CHE levels, thus leading to a downward regulation of the CRP-response, and a slight increase in SAA values.

INTRODUCTION

Monitoring the acute phase response (APR) is widely accepted as a valuable clinical tool in evaluating the activity of acute and chronic inflammatory diseases. The onset, magnitude and duration of the response varies considerably among the various acute phase proteins. Notably C-reactive protein (CRP) and serum amyloid A (SAA) can increase several hundredfold, thereby exceeding all other acute phase plasma proteins .^{625,626}

CRP is synthesized exclusively in the liver,⁶²⁷ although one report presents evidence for synthesis and secretion by cultured human T-lymphocytes.⁶²⁸ The function of CRP may be bactericidal, by causing bacterial capsular swelling, promoting agglutination and complement fixation, and enhancing the phagocytosing capacity of neutrophils.⁶²⁵ Degradation of CRP by lysosomes of neutrophils results in tuftsin-like peptides modulating the function of the neutrophils.^{629,630}

The major site of SAA production is the liver,⁶³¹ although SAA-specific mRNAs have also been detected in a variety of extrahepatic tissues.⁶³²⁻⁶³⁴ Although there is some speculation, the function of SAA remains unclear. An immunosuppressive role in the level of the T-lymphocyte macrophage interaction has been suggested,⁶³⁵ while others showed that the SAA-HDL₃ complex binds to neutrophils in vitro and may modulate the function of these cells.^{636,637} About 4-18 hours after tissue injury, both SAA and CRP plasma levels start to rise, reaching a peak value after 24-72 hours, with a subsequent rapid fall when the provoking stimulus has disappeared.^{625,638,639} In chronic inflammatory conditions like rheumatoid arthritis and Crohn's disease, however, SAA and CRP levels remain elevated and fluctuate with disease activity.⁶²⁵

Although CRP measurements are very practical for routine use, considerable differences may exist in the response of CRP and SAA among diseases.⁶⁴⁰ In some diseases SAA may be a more sensitive -although perhaps less specific- index of disease activity than CRP.⁶⁴¹⁻⁶⁴³

A complicated situation exists in hepatitis, for here the major producing organ of acute phase proteins is affected. Most data deal with acute hepatitis, where the acute phase response (APR) appears to be depressed.⁶⁴⁵⁻⁶⁴⁷ Low levels of haptoglobulin and increased serum levels of α_2 -macroglobulin have been described in chronic hepatitis.^{648,649} Elevated CRP levels were reported in only 0-24%,⁶⁴⁹⁻⁶⁵¹ and increased SAA values were found in about 35% of patients with CAH.⁶⁵¹

Serum cholinesterase (CHE) is a direct marker for hepatic protein synthesizing capacity. In patients with hepatocellular damage, CHE is frequently reduced, almost paralleling reductions in albumin, while a normal CHE virtually excludes severe impairment of liver function.⁶⁵²⁻⁶⁵⁶

In order that we may investigate CRP and SAA responses in CAH, we studied these acute phase reactants in patients with clinical, biochemical and histologic features of CAH, but with differences in aetiology (n=80). In 23 of these untreated patients with
idiopathic 'autoimmune' CAH without anti-HBs, and 5 with anti-HBs, the overall grade of histologic activity and the degree of inflammation in different zones of the liver were related to the serum levels of both CRP and SAA. Both acute phase proteins were also related to serum CHE. In 20 of these CAH patients, the effect of two years of immunosuppression was evaluated by serial study of histology, CHE, CRP and SAA.

MATERIALS AND METHODS

Patients

We studied 81 untreated patients (32 men and 49 women) with histological, clinical and biochemical features of chronic active hepatitis (CAH), with a median age of 49 years (range 11 to 81), and aetiologies as shown (table 10.1).

Aetiology of CAH	all	fig 10.2 [*]	fig 10.3 ^{**}	
IAI, anti-HBs negative	36	24	18	
IAI, anti-HBs positive	6	5	3	
HBsAg-positive	19			
Hepatitis C (non-A,non-B)	5			
CAH/PBC 'overlap'	4			
CAH/PSC 'overlap'	1			
AAT-CAH	1			
'CAH' due to Wilson's disease	1			
drug-induced 'CAH' -oxyphenisatin	5			
-methyldopa	3			
Total	81	29	21	

Table 10.1.CAH-patients: Aetiology.

ad *) fig 10.2 denotes patients where CRP and SAA are related to HAS and CHE, as shown in figure 10.2A-D.

ad **) fig 10.3 denotes standardized treated patients with serum samples and biopsies at 0,2 and 12 and/or 24 months, as shown in figure 10.3A-D.

CAH/PBC : 'overlap' with primary biliary cirrhosis ('cholestatic CAH').

CAH/PSC : 'overlap' with primary sclerosing cholangitis.

AAT-'CAH': 'CAH' due to α 1-antitrypsin deficiency.

IAI: idiopathic 'autoimmune'.

In 29 out of these patients with idiopathic 'autoimmune' CAH, 5 of them with anti-HBs (5 men and 23 women, median age 53 years, range 11-81), localization and degree of inflammation and fibrosis/cirrhosis in coded routine liver biopsies were numerically

scored, and serum cholinesterase (CHE), was measured. Both were compared with CRP and SAA.

21 out of these 29 patients, 4 men and 17 women, requiring immunosuppressive therapy according to accepted criteria,⁶⁵⁷ were studied to evaluate therapy effects. Their median age was 52 years (range 16-71), and median duration of disease 75 days (range 0 weeks to 3 years). All were subsequently treated with oral prednisolone 15 mg and azathioprine 75 mg daily for two months, followed by at least one year (14 patients) or two years (6 patients) of therapy with 10 resp. 50 mg daily. Serial routine liver biopsies and sera (CHE, CRP and SAA) were studied at the start and at two months, one and two years of therapy. Two patients had a follow-up of 14 months.

Liver histology

Liver biopsies were processed and examined under code and assigned a Histological Activity Score (HAS), ranging from 0 to 18, which correlates well with conventional ways of scoring histology in CAH, as described.⁶⁵⁸ In contrast to the original system,⁶⁵⁹ fibrosis/cirhosis is not included in the HAS. The HAS is expressed as a numerical score (0-18), and is the summation of portal (score 0-4), periportal (score 0-10) and lobular (score 0-4) inflammation and necrosis.

Serology

All serum samples were immediately frozen and stored (-20°C) until analysis. Freezing and thawing did not affect the results of CRP, SAA and CHE,^{660,661} and this is one of the reasons why we preferred determination of CHE to that of albumin. Basal control values for CRP and SAA were determined in sera from a group of 50 apparently healthy persons. For the reference values of CHE, we used the sera of 41 women and 36 men with apparently normal health.⁶⁶¹

Cholinesterase (CHE) concentration was determined by measuring its catalytic activity, using acetylthiocholine iodide as a substrate in a commercially available kinetic test (Boehringer Mannheim GmbH).⁶⁶² The lower reference limit of CHE is 1600 U/l for women and 1800 U/l for men.⁶⁶¹

C-reactive protein (CRP) was quantified by a sandwich ELISA.⁶⁶³ The intra-assay coefficient of variation (CV) was 4.9% and the inter-assay CV was 5.1%. The basal control values provided a median and mean value of 0.5 mg/l, with a 95 percent upper limit of 2.0 mg/l.

Serum amyloid A (SAA) was quantified by a recently developed sandwich ELISA, with monoclonal anti-human-SAA as a second antibody.⁶⁶⁰ The intra-assay CV was 6.0% and the inter-assay CV was 13%. The basal control values provided a median value of 0.7 mg/l, and a mean value of 0.8 mg/l, with a 95% upper limit of 2.6 mg/l.

Statistical analysis

Statistical analysis was performed with analysis of variance (ANOVA) with repeated measurements, the MannWhitney U test, the Wilcoxon matched pairs test and Spearman's rank correlation where appropriate, with p<0.05 as the level of significance.

RESULTS

CRP and SAA in untreated CAH in relation to aetiology

In 81 untreated patients with CAH, the CRP serum levels were only slightly elevated in 54 (67%) and SAA in only 12 (15%) of the patients.

Figure 10.1A shows the CRP values and figure 10.1B those of SAA for the different



Figure 10.1 A) CRP and B) SAA in CAH and controls. Dotted lines indicate upper reference limits.

M=median. N=upper reference limit.

aetiologic groups. The CRP values in idiopathic 'autoimmune' CAH (IAI-CAH) without anti-HBs (median 6.6 mg/l) and CAH of other aetiology (median 2.9 mg/l) were higher than the basal control values (median 0.5 mg/l) (p<0.0001). SAA levels in both CAH groups (medians 0.7 and 0.5 mg/l), however, were not higher than the control values (median 0.7 mg/l) (p=0.88 and p=0.44).

The CRP-values in patients with anti-HBs negative IAI-CAH did not differ from those of the other CAH patients (p=0.16). The SAA values in these patients with IAI-CAH also did not differ from those in the other CAH patients (p=0.44).

CRP and SAA in relation to histologic activity and CHE

In 29 of these patients, correlations were made between CRP, SAA and CHE levels and the grade of inflammation in different localizations of the liver (portal, periportal, lobular, and the summation of these), as shown in table 10.2. The only correlations found were between CRP and overall, especially periportal, inflammation (R=.40, p<.05 and R=.39, p<0.05 respectively), and between SAA and CHE (R=.61, p<0.005) as shown in figures 10.2A-D.

		CRP		SAA		CHE	
		R	p<	R	p<	R	p<
portal	(P)	.24	.20	.25	.19	05	.78
periporta	l(PP)	.39	.05	.09	.65	06	.75
lobular	(L)	.25	.19	.21	.28	.09	.65
HAS(=P	+PP+L)	.40	.05	.06	.76	10	.62
CHE		06	.75	.61	.005		
SAA		.22	.24				

Table 10.2.	CRP, SAA and	d CHE versus	Histological	Activity in	I CAH

R= Spearman rank correlation. n=29 p= level of significance ns=not significant





Figure 10.2A-D. CRP and SAA versus periportal inflammation and CHE in CAH.

121

Effects of standardized immunosuppressive therapy

21 patients (18 IAI-CAH without anti-HBs, 3 anti-HBs positive) maintained on standardized immunosuppressive therapy were fully evaluated with regard to CHE, SAA, and CRP levels and a liver biopsy at the start, after two months and after one and/ or two years of therapy.

As shown in figure 10.3A, the HAS diminished from median 9 to median 3 within 2 months of standardized therapy (p<.001) and a further improvement in the next two years of therapy to median HAS=1 (p<.05). Reductions in periportal (p<.001) and lobular (p<.005) inflammation were observed in the first two months of therapy, with a further decrease in periportal and a trend to improvement in lobular inflammation in the next two years (p<.05 and p=.09 respectively). A gradual reduction in portal inflammation occurred in the first 14 months of therapy (p<.05).⁶⁵⁶ A histology similar to chronic persistent hepatitis ('CAH in histological remission') usually remained, while periportal and lobular inflammation had diminished and (almost) disappeared in patients with HAS \leq 3.

As shown in figure 10.3B, CHE steadily increased during standardized immunosuppression,⁶⁵⁶ from a median value of 1185 U/l at start to 1698 U/l after two months (p<.01), to 2323 U/l at one year (p<.001) and stabilization at median 2514 U/l after two years (p=.36). At the start of therapy, 5 (25%) patients had a normal CHE, while CHE became normal in all 21 patients within two years of therapy.

The CRP values fell from a median value of 7.1 mg/l at the start to 0.9 mg/l at two months (p<.0005) and stabilization at a median level of 1.3 mg/l at one (p=0.11) and 1.2 mg/l at two years of therapy (p=1.0), as shown in figure 10.3C. At the start of therapy, the CRP values were higher than basal control values (p<.0001). Compared to basal control values, CRP values did not differ at two months (p=.24) and 14 months (p=.11), and became slightly higher after two years of therapy (p<.05).

SAA values at start (median 1.1 mg/l) and after two months of therapy (median 2.4 mg/l) did not differ (p=.25), but there was a slight increase between SAA values at the start and after one year of therapy (median 2.1 mg/l) (p<.05), with no change in the second year (median 2.4 mg/l) (p=.16) (figure 10.3D). SAA values did not differ from basal control values (p=0.08), but they became higher than these values at two months, one and two years of therapy (p<0.0005).





Figure 10.3 A) HAS, B) CHE, C) CRP and D) SAA in CAH before and during standardized immunosuppression. Dotted lines indicate upper (CRP, SAA) or lower (CHE) limits of reference range.

DISCUSSION

Evaluation of the acute phase response in liver disease is complicated by the fact that the liver is the major producing organ of most acute phase proteins (APP), including CRP and SAA.

No differences in CRP serum values in relation to the aetiology of CAH were observed. During more active disease, serum CRP values rise. However, the correlation with periportal hepatitis and piecemeal necrosis is weak and insufficient for clinical purposes. Although CRP acts as an APP in CAH, values lie in a range about two to three times lower than in a randomly selected group of patients with Crohn's disease or rheumatoid arthritis.⁶⁶⁰ During immunosuppressive therapy, CRP values normalize in parallel to the decrease in HAS.

The response of SAA is remarkable in contrast to CRP. In untreated CAH, SAA levels do not differ from those in the controls, and there is no correlation with histologic activity.

The normal SAA in CAH may be due to impaired protein synthesis as the result of impaired liver function, as reflected in serum CHE levels. It has been demonstrated that the capacity of the liver to produce most acute phase proteins is well retained, even when the hepatic synthetic capacity is rapidly declining.⁶⁴⁶ However, our results show that SAA is an exception to this rule and its reaction in CAH resembles those of serum amyloid P-component (SAP) and haptoglobin.^{649,664,665}

Several explanations are possible for the observed profiles in CAH. Apart from the impaired liver function as a cause of an unresponsiveness of SAA, the difference between CRP- and SAA response could also result from different preferential areas of production for CRP and SAA, and therefore a different reaction of local protein synthesis to local, (peri)portal production of cytokines. CRP production starts in zone 1 and shifts towards zone 3 during the APR.⁶²⁷ Studies regarding the site of SAA production in the liver are not conclusive.⁶⁶⁶⁻⁶⁶⁸

SAA is associated with high density lipoprotein (HDL_3) .⁶⁶⁹ Since both bound and unbound SAA are measured in our ELISA, the fact that HDL_3 is a negative acute phase reactant⁶⁷⁰ is unlikely to influence results.

The synthesis of CRP and SAA can be stimulated by several cytokines like interleukin-1ß (IL-1ß),^{671,672} interleukin-6 (IL-6)⁶⁷³⁻⁶⁷⁶ and tumor necrosis factor α (TNF α).⁶⁷⁷ Experiments with primary cultures of human hepatocytes and human hepatoma cells showed that II-1ß, IL-6 and TNF- α stimulate liver synthesis of SAA and CRP at the transcriptional level in an additive manner, with IL-6 as the principal stimulator.⁶⁷⁴⁻⁶⁷⁶ However, monocytes may also produce substances that inhibit SAA synthesis,⁶⁷⁸ and they can produce inhibitory activity to IL-18.^{679,680} The degree of stimulation of synthesis can lead to different levels among various APP's as the result of stimulation by one or more of these cytokines. However, the difference in CRP and SAA responses in CAH cannot be entirely explained by these differences in stimulation. The normalization of hepatic functional protein synthesizing capacity (CHE) in severe CAH usually takes about two years, and therefore appears to be predominantly the result of repair and regeneration of liver tissue. Because CHE, like albumin,⁶⁷⁴ is a negative acute phase reactant, the increment in CHE levels may also be the result of a decline in titres of e.g. IL-1B, IL-6 and TNF α . Glucocorticoids can also selectively inhibit the transcription of the IL-1B gene and decrease the stability of IL-1B mRNA.⁶⁸¹ Furthermore, the rise in CHE and SAA may partly result from a direct stimulation of protein synthesis by corticosteroids, as is also observed in hepatocyte cultures.^{675,682}

In conclusion, our results show that in chronic active hepatitis of all aetiologies studied, the acute phase response is depressed with regard to CRP and especially to SAA. SAA serum levels are usually not elevated in CAH, CRP is elevated, but not to a degree that reflects the high degree of inflammation in a voluminous organ like the liver. SAA is correlated with the serum cholinesterase, while CRP is not. CRP modestly reflected the degree of histological (periportal) inflammation, while SAA did not at all. Impairment of the liver capacity to synthesize proteins probably affected SAA more severely than CRP, resulting in this different behaviour of both proteins, as summarized in table 10.3.

	CRP	SAA	
Local inflammation	$\uparrow \uparrow \uparrow$	$\uparrow\uparrow$	
Depressed protein synthesis	\downarrow	$\downarrow\downarrow\downarrow$	
Net result	$\uparrow \uparrow$	N	

Table 10.3. Mechanisms responsible for responses of CRP and SAA in CAH.

Effects of immunosuppression on CRP and SAA in CAH.

	CRP	SAA
Decline in inflammation	$\downarrow \downarrow \downarrow$	$\downarrow\downarrow$
Increased protein synthesis	1	$\uparrow \uparrow \uparrow$
Direct effect of steroids	\downarrow ?	N ?
Net result	$\downarrow \downarrow (\downarrow ?)$	↑/N

 \uparrow = increase \downarrow = decrease (number of arrows denotes degree of increase/decrease). N=no change.

CHAPTER 11

SERUM N-TERMINAL PROPEPTIDE OF COLLAGEN TYPE III IN 'AUTOIMMUNE' CHRONIC ACTIVE HEPATITIS

Relation to Histology and Functional Capacity of the Liver and Effects of Standardized Immunosuppression.

SUMMARY

The serum N-terminal propeptide of collagen type III (PIIINP) predominantly reflects deposition of collagen type III in the liver. We studied this in untreated idiopathic 'auto-immune' chronic active hepatitis (IAI-CAH) (n=25, 5 anti-HBS positive), during complete histological remission under immunosuppressive therapy (n=23). In 21 patients, the effects of standardized therapy were related to histology and serum cholinesterase, reflecting liver function.

PIIINP was measured with a new rapid equilibrium type of PIIINP assay which is based on the human propeptide. We found that this correlated very well (R=.96) with the standard assay which is based on bovine PIIINP.

PIIINP was positively correlated to periportal inflammation (R=.45), but not to portal or lobular inflammation and necrosis, resulting in a correlation between PIIINP and overall histologic activity of R=.43. Very low PIIINP values (<2.5 μ g/l) were related to histological remission, and very high values ($\geq 18 \mu$ g/l) correlated to CAH. However, for values in between, serum PIIINP measurements could not replace a liver biopsy for assessment of histological activity. Static measurement of histological fibrosis/cirrhosis did not correlate with serum PIIINP levels. Despite a negative correlation between PIIINP and CHE in untreated CAH (R= -.62) and during therapy induced remission (R= -.44), the influence of liver functional capacity on PIIINP appeared minimal.

To conclude, elevated serum PIIINP levels in IAI-CAH reflect increased deposition of collagen type III in the liver, and are correlated with periportal inflammation, while impairment of liver function only leads to a slight increase in PIIINP. Although standardized immunosuppression led to normalization of PIIINP in half of the patients, PIIINP remained slightly elevated in the other 50%, despite induction of histological remission and normalization of CHE. This may (partly) explain ongoing formation of cirrhosis during therapy, as has been observed in some studies.

INTRODUCTION

Idiopathic 'autoimmune' chronic active hepatitis (CAH) usually reacts favorably to immunosuppressive therapy immunosuppression was shown to enhance survival in this type of CAH⁶⁸³⁻⁶⁸⁷. Fibrosis and formation of cirrhosis are sequelae of untreated CAH, and can even develop during therapy intended to suppress inflammation^{687,688}. About 70% of liver collagens consist of collagen types I and III ⁶⁸⁹. Collagen type III is predominant in early fibrosis; in late cirrhosis, type I, "hard collagen" prevails ⁶⁸⁹. Type III collagen has a more rapid turnover than type I collagen ⁶⁹⁰. The C- and the N-terminal procollagen-III-peptide (PIIINP) are split from the procollagen-III molecules before extracellular assemblage into fibers in the extracellular matrix. Generally, the amount of free PIIINP that circulates stochiometrically reflects deposition of collagen-III.⁶⁹¹⁻⁶⁹³

Some of the serum PIIINP is derived from PIIINP which was retained in the deposited fibers.^{694,695} Decreased degradation of PIIINP due to absent liver function can account for a rise in serum PIIINP in the anhepatic situation during liver transplantation⁶⁹⁶. However, it is unknown whether a decrease in hepatic functional capacity in itself, as can occur in CAH, can affect serum PIIINP levels.

Some studies indicated that serum PIIINP parallels the activity of inflammation in acute hepatitis, and returns to normal along with transaminases and bilirubin.⁶⁹⁷ In alcoholic hepatitis, PIIINP correlated well with periportal and intralobular inflammation⁶⁹⁸⁻⁷⁰². Several studies suggested a relationship between activity of CAH and PIIINP⁷⁰³⁻⁷⁰⁶, although this could be masked in children with CAH by elevation of PIIINP due to growth⁷⁰⁷. Elevation of PIIINP in serum of patients with viral hepatitis persisting for more than 6.5 months, correlated with chronicity^{697,708}. Therapy with corticosteroids can lower an elevated PIIINP in CAH.^{704,709-712}

Recently it has been shown that PIIINP changed in accordance with other liver tests and normalized when remission of CAH had been achieved^{713,714}.

A recent study on liver transplantation in pigs indicated that PIIINP serum levels can rise in the anhepatic situation⁶⁹⁶. Serum cholinesterase (CHE) is a direct marker for hepatic protein synthesizing capacity. In patients with hepatocellular damage, CHE is frequently reduced, almost paralleling reductions in albumin, while a normal CHE virtually excludes severe impairment of liver function.⁷¹⁵⁻⁷¹⁹

In the present study, we analysed the serum PIIINP levels in relation to histology and liver function in untreated idiopathic 'autoimmune' CAH before and during two years of standardized immunosuppression, and we related these to histology and CHE.

MATERIALS AND METHODS

Patients

We studied 25 untreated patients (4 men and 21 women) with histological, clinical and biochemical features of chronic active hepatitis (CAH). They had a median age of 53 years (range 16 to 81) and a median duration of disease (between the tested serum sample and the diagnostic biopsy) of 72 days (range -21 days to + 3 years). 20 of these patients had idiopathic 'autoimmune' CAH, and 5 had circulating anti-HBs antibodies. In these untreated CAH patients, localization and degree of inflammation and fibrosis/cirrhosis in coded routine liver biopsies were numerically scored, and serum cholinesterase (CHE), reflecting the hepatic protein synthesizing capacity, was measured. Both were compared with PIIINP. In 23 of the 25 patients, PIIINP was related to CHE on 44 occasions during complete histological remission (HAS \leq 1) under therapy with prednisolone (5-15 mg), in 21 patients in combination with azathioprine (50-75 mg).

21 out of these untreated CAH patients (3 of them with antiHBs: 4 men and 17 women), requiring immunosuppressive therapy according to accepted criteria ⁵⁰, were studied to evaluate therapy-effects. Their median age was 52 years (range 16-71), and median duration of disease 75 days (range -21 days to +3 years). All were subsequently treated with oral prednisolone 15 mg and azathioprine 75 mg daily for two months, followed by at least one year (14 patients) or two years (6 patients) of therapy with 10 resp. 50 mg daily. Serial routine liver biopsies and sera (CHE, PIIINP) were studied at the start and at two months, one and two years of therapy. 16 of the patients were followed up for 26 months, the other 5 patients for 14 months. One biopsy at 2 months and one at 14 months was missing. One serum measured at 14 months failed in a patient followed up for 26 months.

Liver histology

Liver biopsies were processed and examined under code and assigned a Histological Activity Score (HAS), ranging from 0 to 18, which correlates well with conventional ways of scoring histology in CAH, as described.⁷¹⁹ In contrast to the original system,⁷¹⁹ fibrosis/cirhhosis is not included in the HAS, but is presented as a separate histologic fibrosis score (HFS) ranging from 0 to 4. The HAS is expressed as a numerical score (0-18), and is an addition of portal (score 0-4), periportal (score 0-10) and lobular (score 0-4) inflammation and necrosis.

Serum cholinesterase

Cholinesterase (CHE) concentration was determined by measuring its catalytic activity, using acetylthiocholine iodide as a substrate in a commercially available kinetic test (Boehringer Mannheim GmbH).⁷²⁰ For the reference values of CHE we used the sera of 41 women and 36 men with apparently normal health. The lower reference limit of CHE is 1600 U/l for women and 1800 U/l for men.⁷²¹

Serum PIIINP

A new radioimmunoassay (RIA) developed by Risteli and Risteli⁷²² for the determination of PIIINP was applied. This rapid equilibrium type of assay is based on the human propeptide and is not sensitive to smaller degradation products⁷²¹. Therefore, in this new PIIINP-assay no serial dilutions of the sera and no 50% intercept method for calculating the results are necessary, since the standards and the serum samples give parallel inhibition curves. This is unlike the PIIINP RIA-gnost® (Behring).722,723,724 This solves the most important problems that have been inherent in the determination of PIIINP⁷²⁵. Results of serum PIIINP obtained with the new equilibrium-RIA, based on the human propeptide, showed excellent correlation (r=.96, $r^2=.92$) with those of the RIA-gnost' in CAH-sera (see chapter 4) and showed similar changes during immunosuppression. The intra-assay variation was 4.1%. Sera had been frozen and stored (-20°C) until use. Storage of sera at -20°C ⁷¹³, and bilirubin- and prednisolone-levels ^{704,727} in serum do not affect results of PIIINPdetermination. PIIINP can be elevated in renal insufficiency⁷²⁸. One patient had a 40% lowered glomerular filtration rate due to membrano-proliferative glomerulonephritis, but PIIINP values in this patient paralleled those of the other CAH patients. The age of all patients was above 16 years, allowing normal reference values for adults (upper limit 4.2 μ g/l)⁷²² to be applied.

Statistical analysis

Statistical analysis was performed with Friedman's analysis of variance (ANOVA) with repeated measurements, MannWhitney's U test, Wilcoxon's matched pairs test, and Spearman's rank correlation test where appropriate, with p<0.05 as level of significance.

RESULTS

During untreated CAH

The correlations in 25 untreated patients, between PIIINP on one side, and CHE and histology on the other side, are shown in table 11.1. PIIINP was positively correlated to periportal (R=.45), but not to portal or lobular inflammation and necrosis, resulting in a correlation between PIIINP and HAS of R=.43. Static measurement of histological fibrosis/cirrhosis did not correlate with serum PIIINP levels. A negative correlation between PIIINP and CHE was detected (R=-.62) (figure 11.1A).

During histological remission under immunosuppression

On 44 occasions in 23 out of the 25 patients, we related PIIINP to CHE during complete histological remission (HAS \leq 1) under immunosuppressive therapy. There was no correlation between PIIINP and histological inflammation/necrosis or fibrosis/cirrhosis. However, a negative correlation still existed between PIIINP and CHE (R=-.44, p<0.01) during histological remission, although the PIIINP values that went with the lower CHE values were usually still below 6 µg/l (figure 11.1B), indicating that impaired hepatic functional capacity leads to only minimal increases in serum PIIINP values.

Correlation with PIIINP		R	р	
Histology Infla	mmation/necrosis:			
portal	(P)	.10	.13	
periportal	(PP)	.45	<.05	
lobular	(L)	.28	.15	
HAS	(=P+PP+L)	.43	<.05	
Fibrosis/cirrhos	sis (HFS)	.19	.34	
CHE		62	<.005	

Table 11.1.PIIINP, CHE and histology in untreated CAH (n=25).
(Spearman's rank correlation).



Figure 11.1. PIIINP versus CHE (A) in untreated IAI-CAH and (B) during therapy-induced histological remission (with HAS≤1).

Effects of standardized immunosuppressive therapy

It was possible to evaluate 21 patients maintained on standardized immunosuppressive therapy with regard to PIIINP- and CHE levels and a liver biopsy at the start, after two months and after one and/or two years of therapy. As shown in figure 11.2A, the HAS diminished from median 9 to median 3 within 2 months of standardized therapy (p<0.001), with a trend to further improvement in the next year of therapy to median HAS=1 (p=0.08), and stabilization thereafter (median HAS=1 at two years)(p=0.295). In the first two months of therapy, reduction in the periportal inflammation score (0-4) from median 5 to a median score of 0.5 (p<0.001) took place, while the lobular





Figure 11.2. A) HAS, B) HFS, C) CHE, D) ALT and E) PIIINP in 'autoimmune' CAH before and during immunosuppressive therapy (n=21).

Dotted lines indicate upper (ALT, PIIINP) or lower (CHE) limits of reference range.

inflammation score (0-4) was reduced from median 1 to a median score of 0 (p<0.005), with further reduction and trend to reduction in the next two years of therapy (median scores of periportal and lobular inflammation at one and two years: 0)(p<0.05 and p=0.09 respectively). Portal inflammation scores diminished in the initial 14 months of therapy from median 3 to median 1 (p<0.05), with stabilization thereafter. A histology similar to chronic persistent hepatitis ('CAH in histological remission') usually remained (median scores of portal inflammation were 1 at one and two years), while periportal and lobular inflammation had diminished and (almost) disappeared. The fibrosis/cirrhosis score (HFS) (0-4) was high at the start (median 3) and did not change during therapy (median 3.5, 3 and 3 at two months, one and two years) (p=.29, p=.92, p=.20 respectively) (figure 11.2B). These data, together with data on routine serum biochemistry have been shown in chapters 7 and 8.

CHE steadily increased during standardized immunosuppression, from a median value of 1185 U/l at the start to 1698 U/l after two months (p<0.01), to 2323 U/l at one year (p<0.001) and to median 2514 U/l after two years (p<0.05). At the start of therapy 5(25%) patients had a normal CHE, while CHE was normal in all 17 patients with one or two years of observation during therapy. This was frequently observed, despite the presence of cirrhosis as the HFS indicated. This is shown in figure 11.2C. For more details on serum biochemistry (including ALT and CHE) data shown in figure 11.2D) the reader is referred to chapters 7,8 and 9.

PIIINP values fell from a median value of $19.1 \,\mu$ g/l at start to $4.7 \,\mu$ g/l at two months (p<0.001) with a further reduction to median 4.6 μ g/l at one (p<0.05) and 4.1 μ g/l at two years of therapy (p0.005), as shown in figure 11.2E. HAS or PIIINP values before therapy (data not shown) did not differ between patients with HAS>3 at 14 or 26 months and those patients with HAS>3 at both 14 and 26 months.

DISCUSSION

Fibrogenesis is closely associated with activity of CAH. The mechanisms underlying fibrogenesis in the liver are not completely clear. The increased production of collagen leading to hepatic fibrosis is the result of either increased net production of collagen per cell, or increased number of collagen-producing cells. The current knowledge of the many levels known at which this process is regulated was recently reviewed⁷²⁹. It has been suspected that cytokines such as interleukin-1, which is present in high levels during inflammation, stimulate Kupffer cells to induce fibrogenesis in fat-storing cells, myofibroblasts, and parenchymal cells in the liver. Kuppfer cell stimulation is augmented by fibroblast-activating and fibroblast-growth factors released from T-cells in the inflammatory infiltrate⁷³⁰⁻⁷³⁴. Tumor necrosis factor-alpha (TNF- α) and a release of transformation growth factor B-1 (TGF-B1) derived from Kupffer cells and Ito-cells, may play a role in stimulation of fibrogenesis⁷³⁵⁻⁷³⁸. In vitro experiments show that TGF-B1 gene expression is inhibited by dexamethason.⁷³⁸ This is probably one of the mechanisms whereby corticosteroids inhibit collagen synthesis in the liver, in addition to reducing inflammation.

Although some studies^{700,701,704} suggest the contrary, the current study shows that PIIINP levels do not quantitatively reflect static measurement of hepatic fibrosis. As shown, histological scores of cirrhosis/fibrosis (HFS) were high and did not change during immunosuppressive therapy. An explanation may be the considerable degree of fibrosis/cirrhosis present in most of the patients before immunosuppression, and a decrease of inflammation, hence fibrogenesis, after the institution of immunosuppression. In our results, correlation of static measurements of histological fibrosis/cirrhosis (HFS) with PIIINP was weak, as could be expected, since PIIINP predominantly

reflects the dynamics of deposition of collagen III in the liver in CAH. This weak correlation probably results from the fact that patients with the most active deposition of collagen III will usually have the most advanced degree of cirrhosis. It has been demonstrated before, that PIIINP values are less affected than AST and ALT by the presence or absence of cirrhosis^{739,740} and necrosis^{704,741,742}.

The correlation between PIIINP and periportal inflammation found in the current study appears insufficient to replace a liver biopsy for the assessment of histological inflammation in the liver in all cases. Many explanations may exist for this weak correlation:

Firstly, levels of cytokines, degree of fibrogenesis, and deposition of collagen may fluctuate quite rapidly, while the mounting or disappearance of a hepatic inflammatory infiltrate takes more time.

Secondly, a reduction in degradation of PIIINP may exist during severe CAH, when the liver function is impaired, as will be discussed.

Only high and very low PIIINP levels accurately reflect histological CAH or remission respectively. In some of these cases serum PIIINP levels might replace a liver biopsy. Serum PIIINP may be a useful parameter during follow-up of CAH. Especially as PIIINP predominantly reflects deposition of collagen type III in the liver in CAH, which fluctuates with immunological state of activation of the disease. Similarly, PIIINP levels during 'CAH in remission' in patients with cirrhosis are usually within a much smaller range than, for instance, those of aminotransferases.

Therefore, a slight elevation of serum PIIINP titres (up to 6 μ g/l) in patients with some reduction in serum CHE levels does not necessarily reflect increased deposition of collagen type III in the liver, but may be due to impaired liver function. However, during histological remission in CAH patients with an impaired hepatic functional capacity, PIIINP titres above 6 μ g/l probably indicate increased deposition of collagen type III. In severe CAH the hepatic functional capacity is usually impaired. In such a situation increased synthesis and deposition of collagen in the liver excist, but some of the elevation of PIIINP levels must be due to decreased breakdown of PIIINP in the liver⁶⁹⁶.

Results of the present study indicate that, although PIIINP levels changed in concert with standard liver tests and were abnormal in severe CAH, PIIINP levels frequently did not completely normalize when remission of disease had been achieved after two years of standardized immunosuppression in CAH. This is in contradiction with a recent study, which showed that PIIINP values not only changed in accordance with other liver tests, but also normalized when remission of CAH had been achieved^{713,714}. In that study, however, patients were selected to have either active severe disease, or complete remission.

Although we found that PIIINP levels can be slightly influenced by a reduction in liver function, even in the virtual absence of inflammation (HAS \leq 1), CHE normalized in all

patients receiving standardized immunosuppression. Therefore, the persistent, usually slight, elevation of PIIINP levels despite induction of histological remission probably reflects increased deposition of collagen type III in the liver of these patients.

This process, together with the increased levels of PIIINP during the induction of remission, may account for the observed development of fibrosis. In addition, fibrosis in combination with regeneration may lead to cirrhosis in many patients, during therapy of IAI-CAH.

Normalization of serum PIIINP and hepatic functional capacity could be important endpoints of therapy in IAI-CAH. However, it is questionable whether it is always necessary to prevent development of cirrhosis in all patients, although in the absence of cirrhosis there is probably a lower risk of development of hepatocellular carcinoma. Presence of fibrosis/cirrhosis at presentation is a frequent finding in IAI-CAH^{708,743}, and patients who develop cirrhosis during therapy have a 5-year survival after the development of cirrhosis that is similar to that of patients who do not develop this complication^{687,688,744}. Furthermore, the risks of increased dosages of immunosuppressive medication may outweigh the potential benefits of lowering the rate of deposition of collagen III in many patients. In the near future, additional antifibrotic therapies will probably become available.^{731, 747} This may prove useful as an additional therapy in some CAH patients, e.g. those with partial α_1 -antitrypsin deficiency -phenotype MZwhere PIIINP remained markedly elevated despite the induction of histological remission (our unpublished observation).

In conclusion, serum PIIINP levels are elevated in CAH and show a continuous decline during two years of immunosuppressive therapy, with eventual normalization in about half of the patients. Serum PIIINP levels are not only related to periportal inflammation, but are also negatively correlated to serum CHE levels. However, in the absence of histological inflammation this influence of hepatic functional capacity on PIIINP levels is weak. The persistent, usually slight, elevation of PIIINP levels, despite induction of histological remission and normalized hepatic functional capacity in half of the IAI-CAH patients, probably reflects increased deposition of collagen type III in the liver of these patients, leading to progression of fibrosis/cirrhosis.

This may (partly) explain ongoing formation of cirrhosis during immunosuppressive therapy for IAI-CAH, as has been observed in the quoted studies.^{687,688}

CHAPTER 12

THE SERUM N-TERMINAL PROPEPTIDE OF COLLAGEN TYPE III AS A PREDICTOR OF RELAPSE IN 'AUTOIMMUNE' CHRONIC ACTIVE HEPATITIS

SUMMARY

We studied the serum N-terminal propeptide of collagen type III (PIIINP) and the alanine aminotransferase (ALT) in serial sera of 16 histologically documented relapses of idiopathic 'autoimmune' chronic active hepatitis (IAI-CAH). The relapses occurred during tapering of immunosuppressive therapy in 9 patients with previous histological and biochemical remission.

PIIINP was above the upper reference limit (>N) before ALT was >3N (p<0.005) or >2N (p<0.005). However, the time from 'diagnostic moment' (i.e. the moment the value lies obove the upper reference limit) until peak-ALT did not differ if the upper reference limit for both PIIINP and ALT was defined as 1.25 times the 'baseline value'; despite these very strict criteria, still some patients showed elevation of PIIINP preceding that of ALT. ALT>1.25 times 'baseline value' occurred before ALT>90 or ALT>60 (p<0.001), but the moment that PIIINP was >N equalled the moment that PIIINP was >1.25 times 'baseline value'. Nevertheless, if we chose either PIIINP>N or PIIINP>1.25 times 'baseline value' as the 'diagnostic moment' (whichever of the two exceeded the limit first), the moment one of these two limits was exceeded preceded ALT>1.25 times 'baseline value'(p<0.05), it preceded PIIINP>N (p<0.005), and it preceded PIIINP>1.25 times 'baseline value' (p<0.001) as limits.

We conclude that PIIINP is useful in early detection of relapse in IAI-CAH. Unlike ALT, when using PIIINP no individual reference ranges have to be used.

INTRODUCTION

Remission from CAH is usually defined as inflammation and necrosis confined to the portal tracts, serum aspartate aminotransferase (AST) less than twice the upper limit of normal, normalization of serum bilirubin and gammaglobulin, and absence of clinical symptoms^{748,749}. Relapse with a histologic evidence of moderate to severe CAH has been reported to be invariably present during a threefold or greater elevation of the serum AST level with return of symptoms ^{748,750}.

Relapse occurs in 49-87 percent of patients within 6 months after remission - during or after discontinuation of treatment. The majority of patients who experience relapse do

so within the first 3 months after termination of treatment,^{749,751,752} one quarter of the patients having no clinical symptoms.⁷⁴⁹

Despite the widespread use of AST and gammaglobulin (GG) values during follow-up of CAH, biochemical values of AST and GG are unreliable predictors of morphologic activity: negative predictive values are only 44% and 33% for AST and GG respectively⁷⁵³, while a two-fold or greater elevation in AST does not always identify histological CAH. 19% of normal AST determinations can occur in association with CAH after treatment.⁷⁵³

Although initial studies appeared to indicate that serum concentrations of bile acids^{754,755} or amino acids⁷⁵⁶⁻⁷⁵⁹ would provide a more sensitive indication of disease activity in CAH, subsequent publications proved that these parameters cannot replace a liver biopsy in the assessment of inflammatory activity, because they are influenced by many other variables.⁷⁶⁰⁻⁷⁶² In a biopsy of sufficient size^{762,763}, sampling error for determination of inflammatory activity in CAH is less than 10%.⁷⁶⁴ Unlike other types of liver disease, intra-observer error is 6-7 percent in chronic hepatitis^{764,765}. From these data it follows that at present a liver biopsy is required, not only for diagnosis, but also for monitoring inflammatory activity^{766,767}. Although it is a minor procedure, a liver biopsy is still an invasive procedure with its own, albeit low, morbidity and mortality.⁷⁶⁸ This precludes its use for frequent routine monitoring of activity of disease. Therefore, the search for a non-invasive parameter of histological inflammation and an early detector of relapse during and after diminishing therapy, continues.

Fibrosis and formation of cirrhosis are sequelae of untreated CAH, and can even develop during therapy intended to suppress inflammation^{769,770}. About 70% of liver collagens consist of collagen types I and III 771. Collagen type III is predominant in early fibrosis; in late cirrhosis, type I, "hard collagen" prevails ⁷⁷¹. Type III collagen has a more rapid turnover than type I collagen 772. The C- and the N-terminal procollagen-III-peptide (PIIINP) are split from the procollagen-III molecules before extracellular assemblage into fibers in the extracellular matrix. Generally, the amount of free PIIINP that circulates stochiometrically reflects deposition of collagen-III.⁷⁷³⁻⁷⁷⁵ Some of the serum PIIINP is derived from PIIINP which was retained in the deposited fibers.776,777 Some studies indicated that serum PIIINP parallels the activity of inflammation in acute hepatitis, and returns to normal with transaminases and bilirubin ⁷⁷⁸, and that in alcoholic hepatitis PIIINP correlated well with periportal and intralobular inflammation⁷⁷⁹⁻⁷⁸³. Several studies suggested a relationship between activity of CAH and PIIINP⁷⁸⁴⁻⁷⁸⁷, although this could be masked in children with CAH by elevated PIIINP due to growth⁷⁸⁸. Persistent elevation for more than 6.5 months of PIIINP in the serum of patients with viral hepatitis correlated with chronicity778,789. Therapy with corticosteroids can lower an elevated PIIINP in CAH785.790-793, and recently it was shown that PIIINP changed in accordance with other liver test and normalized when remission of CAH had been achieved.794,795 However, we found that normalization often is incomplete and requires prolonged therapy (Chapter 11), which might facilitate the reported^{796,797} development of fibrosis/cirrhosis during immunosuppressive therapy for IAI-CAH. A recent study on liver transplantation in pigs indicated that PIIINP serum levels can rise in the anhepatic situation⁷⁹⁸. However, only severe liver function abnormalities influence serum PIIINP levels significantly (Chapter 11).

This study observed serum N-terminal propeptide of collagen type III (PIIINP) and the alanine aminotransferase (ALT) in serial sera of 16 histologically documented relapses of idiopathic 'autoimmune' chronic active hepatitis (IAI-CAH). These relapses occurred during withdrawal of immunosuppressive therapy in 9 patients with previous histological, clinical and biochemical remission.

MATERIALS AND METHODS

Patients

Serial sera from 16 IAI-CAH relapses in 9 patients were studied from the time of remission at the point where immunosuppression was diminished, until peak-ALT during subsequent relapse of IAI-CAH. At least two years earlier, before immunosuppression, all patients had fulfilled accepted criteria for the diagnosis and immunosuppressive treatment of IAI-CAH.⁷⁹⁹ Histological (Histological Activity Score -HAS- \leq 3), clinical and biochemical remission had been confirmed before reducing maintenance immunosuppressive therapy. During the 16 histologically documented relapses (HAS>3), the serum ALT was above three times normal (>3N). In part of the patients relapse was also documented by a live biopsy. At the peak-ALT, immunosuppression was increased, resulting in return to histological, clinical and biochemical remission in all patients.

Routine liver biochemistry

Determination of alanine aminotransferase (ALT) (upper reference limit 30 U/l) was performed automatically (Technicon SMA-C) on the day of serum sampling.

The upper reference limit for remission was set as an ALT level twice (2N) or three times (3N) that for a healthy population, or 1.25 times the 'baseline value' at start of therapy withdrawal (ALT=1.25 x B). The 'diagnostic moment' relapse was defined as that time when values became elevated above this upper reference limit, and remained elevated until the peak-ALT.

ΡΠΝΡ

A new radioimmunoassay (RIA), developed by Risteli and Risteli was applied for the determination of PIIINP.⁸⁰⁰ This rapid equilibrium type of assay is based on the human propeptide and is not sensitive to smaller degradation products⁸⁰¹. Therefore, no serial dilutions of the sera and no 50% intercept method for calculating the results are necessary in this new PIIINP-assay, since the standards and the serum samples give

parallel inhibition curves. This is unlike the PIIINP RIA-gnost[®] (Behring).^{802,803} This solves the most important problems that were inherent in the determination of PIIINP until now⁸⁰⁴. Results of serum PIIINP obtained with the new equilibrium-RIA, based on the human propeptide, showed excellent correlation (r=.96, r²=.92) with those of the 'RIA-gnost' in CAH-sera (see chapter 4) and showed similar changes during immunosuppression (data not shown). The intra-assay variation was 4.1%.

Sera had been frozen and stored (-20°C) until use. Storage of sera at -20°C ⁸⁰⁵, and bilirubin- and prednisolone-levels ^{806,807} in serum do not affect results of PIIINP determination. Creatinine clearances were normal (PIIINP can be elevated in renal insufficiency⁸⁰⁷). The age of all patients was above 16 years, so the normal reference values for adults (upper limit 4.2 μ g/l)⁸⁰⁸ could be applied.

The upper reference limit for ALT was set at 2N, 3N or 1.25 times the 'baseline value' $(1.25 \times B)$. For PIIINP N or 1.25 x B was considered the upper reference limit. 'Baseline value' indicates the ALT or PIIINP at the start of tapering of immunosuppression. The 'diagnostic moment' is defined as the moment when values are elevated above the upper reference limit, and remain elevated until the peak-ALT.

Statistical analysis

Statistical analysis was performed with Wilcoxon's matched pairs test and Chi-square test where appropriate with p<0.05 as the level of significance.

RESULTS

Figure 12.1 shows PIIINP and ALT at the commencement of therapy withdrawal and during the 16 relapses at peak-ALT. Both significantly increase during progression of relapse (p<0.01).

In figure 12.2, median ALT and PIIINP are shown from start of therapy withdrawal until peak-ALT for 10 relapses. PIIINP was above the upper reference limit (N) before ALT >3N (p<0.005) and before ALT>2N (p<0.005). However, the time from diagnostic moment until peak-ALT did not differ, if the upper reference limit for both PIIINP and ALT was defined as 1.25 x B. Despite these strict criteria elevation of PIIINP preceded that of ALT in some patients. ALT>1.25 x B occurred before ALT>90 or ALT>60 (p<0.001), with no difference evident between using ALT>2N and ALT>3N as a limit. The moment that PIIINP was >N coincided with PIIINP>1.25 x B. Nevertheless, if we chose PIIINP>N and/or PIIINP>1.25 x B as the 'diagnostic moment', this preceded ALT>1.25 x B (p<0.05). It also preceded PIIINP>N (p<0.005) and PIIINP>1.25 x B (p<0.001) alone (figure 12.3).



Figure 12.1. A) PIIINP and B) ALT during remission (REN) and relapse (REL) in idiopathic 'autoimmune' CAH. N=upper reference limit.



Figure 12.2. PIIINP and ALT before and during progression of relapse of idiopathic 'autoimmune' CAH during therapy withdrawal. N=the upper limit of the reference range: N=4.2 for PIIINP, and N=30 for ALT.



Figure 12.3. Number of days from 'diagnostic moment' until peak-ALT. (N=upper reference limit, B= baseline value at start of therapy withdrawal). Single-hatched bars: 50% above median number of days. Cross-hatched bars: 50% below median number of days.

DISCUSSION

From the current study, it follows that serum PIIINP is superior to serum ALT in serological detection of relapse in IAI-CAH, and that results largely depend on the chosen upper reference limit. Clearly, this limit should be set at a point where positive and negative predictive values are highest. In accordance with the mentioned serological definitions of relapse and remission in CAH, we chose 3N and 2N as upper reference limits for ALT. In doing so, it was clear that elevation of PIIINP preceded that of ALT. The positive predictive value of ALT could be improved by setting the upper reference limit at 1.25 x B. However, our research confirms that ALT and AST are unreliable predictors of morphologic activity in CAH. Even using a cut off point of 2N or 3N, the number of false positive diagnoses of relapse will undoubtedly increase if the upper reference limit is lowered to 1.25 x B. The combination of PIIINP>N and/or PIIINP>1.25 x B proved superior to all other possibilities mentioned. It also became clear from several relapses in the same patients in this study, that the elevation of PIIINP systematically precedes that of ALT more noticeably in some patients than in others.

We recently demonstrated that PIIINP can remain elevated despite induction of remission (HAS \leq 3). Fibrogenesis is probably directly stimulated by e.g. tumor necrosis factor-alpha and a release of transformation growth factor β -1 derived from Kupffer

cells and Ito-cells.^{808a-808d} Elevated serum ALT is the result of damage to hepatocytes resulting from the buildup of an inflammatory infiltrate, following immunological induction. Since such a buildup takes time, this could explain the earlier rise in PIIINP when compared to ALT. Moreover, serum ALT levels can be influenced by the degree of cirrhosis, while PIIINP is not; instead, PIIINP predominantly reflects the dynamic process of deposition of collagen III.

It may be better not to reduce immunosuppression in patients with elevated PIIINP. Such patients may even need additional therapy in order to prevent formation or progression of cirrhosis.

While such points require further elucidation, it is clear from this study that serum PIIINP is useful in early detection of relapse in IAI-CAH.

Section 4

Humoral immunology in chronic active hepatitis 'Autoimmune" chronic active hepatitis

CHAPTER 13

ANTIBODIES AGAINST 'LIVER-SPECIFIC MEMBRANE LIPOPROTEIN' (LSP) IN RELATION TO HISTOLOGY IN 'AUTOIMMUNE' CHRONIC ACTIVE HEPATITIS, BEFORE AND DURING STANDARDIZED IMMUNOSUPPRESSION.

SUMMARY

Anti-LSP titres have been shown to correlate closely with the severity of histological damage in untreated patients with idiopathic 'autoimmune' chronic active hepatitis (IAICAH).

To further study the relationship between activity of disease and presence of anti-LSP antibodies in CAH, we assessed the presence of anti-LSP in relation to histological activity score -HAS (0-18)- in 'autoimmune' CAH, before and during two years of standardized immunosuppression.

Before therapy, all 21 patients tested had detectable antiLSP antibodies (titre median 150, range 50 to 750). In untreated IAI-CAH, there was no correlation between anti-LSP titres on the one side and HAS or periportal inflammation on the other side. Anti-LSP titres were, however, correlated to alanine aminotransferase (ALT) levels (R=.78, p<.005). At two months of therapy, anti-LSP was detectable in 15 (71%) of the 21 patients (p<0.001). After one and two years of standard therapy 6/21 (29%) (p<0.01) and 6/16 (38%) (p=0.28) patients respectively had detectable anti-LSP. During therapy, anti-LSP titres decreased in all patients under investigation. After one year of therapy, no changes from detectable to undetectable anti-LSP were observed. In the first two months of therapy the decrease in anti-LSP titres paralleled the decrease in HAS. Patients most frequently had a HAS \leq 1 and no detectable anti-LSP antibodies after one year of therapy. If HAS>1, anti-LSP was invariably detectable. Presence of anti-LSP, however, did not always indicate a HAS>1.

The results of the present study demonstrate that high titres of anti-LSP were invariably present in patients with IAI-CAH requiring immunosuppressive therapy according to accepted criteria. Despite the induction of histological remission, anti-LSP remained detectable in about half of these patients, although the titres declined. This finding probably indicates ongoing 'immunological activity' despite induction of histological remission.

INTRODUCTION

In IAI-CAH, there is considerable evidence that cytotoxic reactions of the antibody dependent (ADCC-)type are directed at hepatocyte plasma membrane-derived epitopes in the macromolecular, lipid-associated complex called 'liver specific antigen' (LSP).^{809,810} In IAI-CAH, there is a defect in the LSP-specific suppressor T cell function.^{811,812} Recent studies indicate that the basal defect probably lies in the inducers of these suppressor cells.⁸¹³

Anti-LSP titres have been shown to correlate closely with the severity of histological damage in untreated patients with autoimmune chronic active hepatitis IAI-CAH.^{814,815} One study reports that presence or re-appearance of anti-LSP during treatment withdrawal in patients who are apparently in complete remission invariably predicts relapse in IAI-CAH. It was suggested that regular determination of anti-LSP could provide a sensitive method for achieving optimum control in IAI-CAH.⁸¹⁶

To study further the relationship between activity of disease and presence of anti-LSP antibodies in IAI-CAH, we assessed the presence of anti-LSP in relation to histology in 'autoimmune' CAH, before and during two years of standardized immunosuppression.

MATERIALS AND METHODS

Patients

We studied 21 untreated IAI-CAH patients (3 of them with anti-HBs, 4 men and 17 women), requiring immunosuppressive therapy according to accepted criteria,⁸¹⁷ to evaluate therapy effects. Their median age was 52 years (range 16 to 71), and median duration of disease 75 days (range -21 days to +3 years). All were subsequently treated with oral prednisolone 15 mg and azathioprine 75 mg daily for two months, followed by at least one year (14 patients) or two years (6 patients) of therapy with 10 resp. 50 mg daily. Serial routine liver biopsies and sera were studied at the start and at two months, one and two years of therapy. 16 of the patients were followed for 26 months, the other 5 patients for 14 months. One biopsy at 2 months and one at 14 months were missing. One serum at 14 months failed in a patient followed for 26 months.

Liver histology

Liver biopsies were processed and examined under code and assigned a Histological Activity Score (HAS), ranging from 0 to 18, which correlates well with conventional ways of scoring histology in CAH, as described.⁸¹⁸ In contrast to the original system,⁸¹⁹ fibrosis/cirhosis is not included in the HAS, but is presented as a separate histologic fibrosis score (HFS) ranging from 0 to 4. The HAS is expressed as a numerical score (0-18), and is an addition of portal (score 0-4), periportal (score 0-10) and lobular (score 0-4) inflammation and necrosis.

Routine liver biochemistry

Sera were obtained on the same day on which the liver biopsies were performed. Immediate determination of alanine aminotransferase (ALT) (upper reference limit 30 U/l) and total protein (TP) was performed automatically (Technicon SMA-C).

Anti-LSP

All serum samples were immediately frozen and stored (-20°C) until analysis. We found that freezing and thawing did not affect the results of anti-LSP determination.⁸²⁰ Basal control values for anti-LSP were determined in sera from a group of 20 apparently healthy persons. Anti-LSP antibodies were measured in coded, heat-inactivated sera (30 min. at 56°C) with a staphylococcal protein A radioimmunoprecipitation assay, derived from the assay as described,⁸¹⁴ and modified as described.⁸²⁰

Statistical analysis

Statistical analysis was performed with Friedman's twotailed analysis of variance (ANOVA) with repeated measurements, Mann-Whitney's U test, Wilcoxon's matched pairs test and Spearman's rank correlation where appropriate, with p<0.05 as the level of significance.

RESULTS

Histology

As shown in figure 13.1A, the HAS diminished from median 9 to median 3 within 2 months of standardized therapy (p<.001) with a further improvement in the next two years of therapy to median HAS=1 (p<.05). Reductions in periportal (p<.001) and lobular (p<.005) inflammation were observed in the first two months of therapy, with a further decrease in periportal inflammation and a trend to improvement in lobular inflammation in the next two years (p<.05 and p=.09 respectively). A gradual reduction in portal inflammation occurred in the first 14 months of therapy (p<.05).⁸²¹ A histology similar to chronic persistent hepatitis ('CAH in histological remission') usually remained, while periportal and lobular inflammation had diminished and (almost) disappeared in patients with HAS≤3.

Serum biochemistry

Before therapy, ALT levels were markedly elevated in all patients (median 337 U/l). The levels declined during the first two months of therapy to median 32 U/l (p<.001), with stabilization thereafter at median 20 U/l, both at 14 and 26 months (figure 13.1B). Gammaglobulin levels (n=17) were elevated at start of therapy with a median value of 28 mg/l, falling to median 13.5 at two months (p<.005), and stabilizing thereafter, with median values of 15 and 12, at 14 and 26 months respectively (figure 13.1C).





Figure 13.1. A) HAS, B) ALT, C) GG and D) anti-LSP in 'autoimmune' CAH before and during immunosuppressive therapy. Upper reference limit N for anti-LSP is 1:50, for ALT 30 (2N=60).

Anti-LSP

Before therapy, all 21 patients tested had detectable antiLSP antibodies (titre median 150, range 50 to 750).

At two months of therapy, anti-LSP was detectable in 15 (71%) of the 21 patients (p<0.001). After one and two years of standard therapy, 6/21 patients (29%) (p<0.01) and 6/16 patients (38%) (p=0.28) respectively had detectable anti-LSP (figure 13.1D). During therapy, anti-LSP titres decreased in all patients under investigation. In all patients tested both after one and two years of therapy, no changes from detectable to undetectable anti-LSP were observed after one year. As indicated in figures 13.1A and 13.1D, for the first two months of therapy the decrease in anti-LSP titres paralleled the decrease in HAS. Patients most frequently had a HAS \leq 1 and no detectable anti-LSP antibodies after one year of therapy. If HAS>1, anti-LSP was invariably detectable. Presence of anti-LSP, however, did not always indicate a HAS>1.

In untreated IAI-CAH, there was no correlation between anti-LSP titres on the one side and HAS or periportal inflammation on the other side. However, anti-LSP titres were correlated to alanine aminotransferase (ALT) levels (R=.78, p<.005). In both treated and



Figure 13.2. Correlation between anti-LSP binding percentages (radioimmunoassay) and HAS in treated and untreated IAI-CAH.

DISCUSSION

The results of the present study demonstrate that high titres of anti-LSP were invariably present in patients with IAI-CAH requiring immunosuppressive therapy according to accepted criteria. Despite the induction of histological remission, anti-LSP remained detectable in about half of these patients, although the titres declined. This finding probably indicates ongoing 'immunological activity', despite induction of histological remission. Our (uncontrolled) experience in some patients is, that higher levels of immunosuppressive drugs can sometimes suppress such a persistent manifestation of anti-LSP.

Corticosteroids improve the non-antigen specific T-suppressor cell functions in IAI-CAH, resulting in reversal of histological and biochemical manifestations of disease and sufficient to induce 'remission' according to currently accepted criteria.⁸²² However, corticosteroids have no effect on the genetically determined LSP-specific T-suppressor cell defect. It was previously postulated that this latter defect could allow continued stimulation of anti-LSP producing B lymphocytes. This obviously requires help of cytotoxic lymphocytes for maintenance of the antibody-dependent cellular cytotoxicity (ADCC) that plays an important role in the pathogenesis of IAI-CAH. The ADCC can probably be stopped, while the anti-LSP production still goes on at a lower level.^{809-813,823,824}

The liver-specific, species cross-reactive hepatic asialoglycoprotein receptor (ASGR), also known as hepatic lectin, is an important constituent of LSP.⁸²⁵ Anti-LSP antibodies appear to be predominantly directed at ASGR, although a role for anti-LSP directed at other antigenic specificities has not yet been excluded. In a recent study, anti-ASGR was found in 83% of patients with IAI-CAH.⁸²⁶

After one year of therapy, patients in remission most commonly had a HAS \leq 1 and no detectable anti-LSP. Therefore, we may assume that this combination reflects 'complete remission', and is an important endpoint of therapy in IAI-CAH. It is well known that relapse rates can be high, especially after discontinuation of therapy.^{827,828} Investigators from the Mayo Clinic observed relapse rates of 30% in patients who reverted to normal liver biopsies, but 80% if residual portal hepatitis was present at the time of drug-withdrawal.^{829,830} We therefore distinguished complete histological remission (HAS \leq 1) from partial histological remission (HAS \leq 3).⁸¹⁸ It is interesting to note that in 80% of the patients, such a complete histological remission was achieved with the standardized therapeutic regimen. HAS \leq 1 and negative anti-LSP may be mandatory requirements before attempting to discontinue therapy.

The finding that there was no change in levels of detectable anti-LSP after one year of standardized therapy, probably indicated that within one year of therapy the maximum immunosuppressive effect has been reached. Thus, further reduction in inflammatory
activity thereafter appears unlikely. Continuation of therapy may be warranted in most patients, however, to secure repair and full restoration of liver function, as has been shown in chapter 9.

CHAPTER 14

ANTIBODIES AGAINST 'LIVER SPECIFIC MEMBRANE LIPOPROTEIN' (LSP) AS AN EARLY PREDICTOR OF RELAPSE OF 'AUTOIMMUNE' CHRONIC ACTIVE HEPATITIS DURING THERAPY WITHDRAWAL

SUMMARY

Relapse is frequent during or after withdrawal of immunosuppressive therapy in idiopathic 'autoimmune' chronic active hepatitis (IAI-CAH). Serum aminotransferase levels, using standard reference ranges, insufficiently reflect histological activity, as do most other serological parameters. We determined serial alanineaminotransferase (ALT) and antibodies to the macromolecular, lipid-associated complex called 'liver specific membrane lipoprotein' (antiLSP), using a modified radioimmunoassay, in 12 relapses in 9 women during therapy withdrawal after histologically proven remission. *Results*: Anti-LSP was positive or became so in 11 (92%) out of the 12 cases of relapse during therapy withdrawal, while in 11 (92%) out of the 12 cases it had been undetectable at the start of reduction of therapy. In 2 cases, anti-LSP was positive at the start of reduction of therapy, and remained detectable when relapse occurred (with rising titres). In one patient, anti-LSP was not detected both before and during development of relapse. However, relapse was also not diagnosed using ALT values as a serological detector before the time of peak-ALT in this patient. In 9 (82%) of the 12 relapses, anti-LSP was negative at the start of diminishing therapy, while it was positive at the time of relapse (peak-ALT) (Chi-square 16.667, p<.005). In these nine cases, anti-LSP became positive before ALT>3N in 7 cases, at the same time as ALT>3N in 1 case, and later than ALT>3N in 1 case; for ALT>2N, these numbers were 6, 2, and 1; for ALT>1.25xB, it was 4, 2 and 3 respectively. For these 9 patients, the 'diagnostic moment' for detection of relapse was median 0 days before peak-ALT (range 0-49 days) with ALT>3N or ALT>2N as a criterium for relapse; for ALT>1.25xB, this was median 49 days (range 0-427); for presence of anti-LSP, it was median 133 (range 0-249) days before peak-ALT. Using anti-LSP, relapse was detected earlier than with the use of ALT>3N and ALT>2N as criteria (p<0.005 and p<0.01 respectively). With ALT>1.25xB as cut-off point, serological detection of relapse was also earlier than with ALT>3N or ALT>2N (both p<0.05) as diagnosticum. Between ALT>3N and ALT>2N there was no difference in timing of 'diagnostic moment'(p=0.63). Neither was this different between anti-LSP and ALT>1.25xB (p=0.54).

The current study shows that anti-LSP was, or became, positive in 92% of the cases of relapse of idiopathic 'autoimmune' CAH that we studied. When compared to the current

serological criterium for relapse (ALT>3N), or even when compared to ALT>2N as a criterium, relapse was detected earlier using positivity of anti-LSP or ALT>1.25xB as a criterium.

INTRODUCTION

Remission from chronic active hepatitis (CAH) has been defined as inflammation and necrosis confined to the portal tracts, serum aspartate aminotransferase (AST) less than twice the upper limit of normal, normalization of serum bilirubin and gammaglobulin, and absence of clinical symptoms^{831,832}. Relapse with histologic evidence of moderate to severe CAH has been reported to be invariably present during a threefold or greater elevation of serum aminotransferase levels, with return of symptoms.^{831,832,833}

Relapse occurs in 49-87 percent of patients within 6 months after remission and during or after discontinuation of treatment. The majority of patients who experience relapse do so within the first 3 months after termination of treatment.^{832,834,835} In one quarter of these patients there are no clinical symptoms when a relapse occurs.⁸³²

Despite the widespread use of AST and gammaglobulin (GG) values during follow-up of CAH, biochemical values of AST and GG are unreliable predictors of morphologic activity. Negative predictive values are only 44% and 33% for AST and GG respectively⁸³². 19% of normal AST determinations can occur in association with CAH after treatment.⁸³².

Titres of antibodies against 'liver-specific membrane lipoproteins' (anti-LSP) have been shown to correlate closely with the severity of histological damage in untreated patients with idiopathic 'autoimmune' chronic active hepatitis (IAI-CAH)^{836,837} and can become negative in 60% of patients after two years of standardized immunosuppressive therapy.⁸³⁸ McFarlane *et al.* demonstrated that presence/absence of anti-LSP predicted outcome of treatment withdrawal in IAI-CAH,⁸³⁹ but these findings have never been confirmed. There is good reason for determining anti-LSP during therapy withdrawal, since considerable evidence exists that in CAH, cytotoxic reactions of the antibody dependent (ADCC-)type are directed at hepatocyte plasma membrane-derived epitopes in LSP.^{840,841} In IAI-CAH, a defect in the LSP-specific suppressor T cell function exists.^{842,843} Recent studies indicate that the basal defect probably lies in the inducers of these suppressor cells.⁸⁴⁴

In order to evaluate the clinical value of anti-LSP determinations, we studied serum anti-LSP in relation to ALT in 9 patients during 12 relapses of IAI-CAH, occurring during withdrawal of immunosuppressive therapy.

MATERIALS AND METHODS

Patients

Serial sera were studied from 12 randomly selected IAI-CAH relapses occurring during withdrawal of immunosuppression in 9 patients (all women, age median 51 -range 16 to 58 years). They were studied from histologically confirmed remission at the start of reduction of immunosuppression, until peak-ALT during subsequent relapse of CAH. At least two years earlier, all patients had fulfilled accepted criteria for diagnosis and immunosuppressive treatment of IAI-CAH⁸³¹. Histological, biochemical and clinical remission had been confirmed before diminishing maintenance immunosuppressive therapy. Dosage of prednisolone and azathioprine at the start of therapy withdrawal and during relapse (peak-ALT) are shown in table 14.1. During the histologically documented relapses, the serum ALT was more than three times the upper reference limit (>3N). At the time of peak-ALT, immunosuppression was increased, resulting in a return to histological and biochemical remission in all patients.

patient	prednis	olone (mg)	azathioj	azathioprine (mg)		
	start	relapse	start	relapse		
1	10	0	50	50		
2	10	0	0	0		
3	10	5	50	25		
4	10	10	50	0		
5	7.5	0	50	0		
6	10	1	0	0		
7	5	2	0	0		
8	10	0	50	0		
9	10	5	0	0		
10	7.5	6.5	50	50		
11	10	10	75	50		
12	7.5	5	0	0		

Table 14.1.Dosage of prednisolone and azathioprine at start of diminishing therapy
and at the time of peak-ALT during subsequent relapse.

Routine liver biochemistry

Determination of alanine aminotransferase (ALT) (upper reference limit 30 U/l) was performed automatically (Technicon SMA-C) on the day of serum sampling. 2N, 3N, or 1.25 times the 'baseline value' at start of therapy withdrawal (ALT=1.25xB) was considered the upper reference limit for ALT. The 'diagnostic moment' reference limit, and remained so until the peak-ALT.

Anti-LSP

All serum samples were immediately frozen and stored (-20°C) until analysis. We found that freezing and thawing did not affect the results of anti-LSP.⁸³⁸ Basal control values for anti-LSP were determined in sera from a group of 20 apparently healthy persons, and binding percentages above mean+2SD of this group were considered positive results. Anti-LSP antibodies were measured in coded, heat-inactivated sera (30 min. at 56°C) with a staphylococcal protein A radioimmunoprecipitation assay, derived from the assay as described,⁸³⁶ and modified as described.⁸³⁸

Statistical analysis

Statistical analysis was performed with Mann-Whitney U test and Chi-square test where appropriate, with p<0.05 as the level of significance.

RESULTS

ALT and anti-LSP titres as a function of time before peakALT are shown in figure 14.1. As shown in table 14.2, anti-LSP was positive or became so in 11 (92%) of the 12 cases of relapse during therapy withdrawal. In two cases, anti-LSP was positive at the start of diminishing therapy, and remained detectable when relapse developed (with rising titres). In one patient, anti-LSP was not detected either before or during development of relapse. However, relapse was also not diagnosed with ALT values as a serological



Figure 14.1. ALT and anti-LSP (x=positive, o=negative) from start of diminishing therapy (remission) to peak-ALT during subsequent relapse.

Table 14.2.Number of cases with and without anti-LSP during remission at start of
diminishing immunosuppression and at the time of CAH relapse
(peak ALT) (n=14).

anti-LSP	no	anti-LSP	TOTAL
anti-LSP	2	0	2
REMISSION			
no anti-LSP	9	1	10
TOTAL	11	1	12

RELAPSE

Chi-square = 16.667 (p<0.005)

detector, before the time of peak-ALT in this patient. In 9 (82%) of the 12 relapses, anti-LSP was negative at the start of diminishing therapy, while it was positive at the time of relapse (peak-ALT) (Chi-square 16.667, p<.005).

As shown in table 14.3, in these nine cases, anti-LSP became positive before ALT>3N in 7 cases, at the same time as ALT>3N in 1 case, and later than ALT>3N in 1 case; for ALT>2N these numbers were 6, 2, and 1; for ALT>1.25xB it was 4, 2 and 3 respectively. For these 9 patients, the 'diagnostic moment' for detection of relapse was median 0 days before peak-ALT (range 0-49 days) with ALT>3N or ALT>2N as a criterium for relapse; for ALT>1.25xB, this was median 49 days (range 0-427); for presence of anti-LSP, it was median 133 (range 0-249) days before peak-ALT (table 14.3). Using anti-LSP, relapse was detected earlier than with the use of ALT>3N and ALT>2N as criteria (p<0.005 and p<0.01 respectively). With ALT>1.25xB as cut-off point, serological detection of relapse was also earlier than with ALT>3N or ALT>2N (both p<0.05) as diagnosticum. Between ALT>3N and ALT>2N there was no difference in timing of 'diagnostic moment'(p=0.63). Neither was this different between presence of anti-LSP and ALT>1.25xB (p=0.54).

patient	ALT>3N	ALT>2N	ALT>1.25xB	anti-LSP+
1	0	0	0	181
2	14	14	427	133
3	0	0	132	175
4	0	0	0	28
5	49	49	49	21
6	0	0	203	203
7	0	49	49	49
8	0	0	84	0
9	4	4	4	249
median number of days.	0	0	49	133
patient				
10	0	0	0	*
11	0	105	105	**
12	28	28	126	**

Table 14.3.Number of days between diagnostic moment and peak-ALT using
different criteria for serological diagnosis relapse, in 14 cases of CAH-
relapse during therapy withdrawal.

N=upper reference limit. B='baseline value' of ALT at start of therapy withdrawal during histologically documented 'CAH in remisssion'.

*:anti-LSP negative both during remission and relapse. **:anti-LSP positive both during remission and relapse.

DISCUSSION

The current study shows that anti-LSP was or became positive in 92% of the cases of relapse of idiopathic 'autoimmune' CAH we studied. When compared to the current serological criterium for relapse (ALT>3N), or even when compared to ALT>2N as a criterium, relapse was detected earlier with positivity of anti-LSP or ALT>1.25xB as a criterium. In the only case where anti-LSP was and remained undetectable during therapy withdrawal, early diagnosis was not made with the help of ALT values either. If ALT>1.25xB was used as the 'cut-off point', detection of relapse as early as with anti-LSP was possible. If we assume that the findings are correct in that anti-LSP is

pathogenetically involved in IAI-CAH, this early rise in ALT could mean that cell damage starts early after immunological activation of the disease occurs.

The sensitivity of a positive anti-LSP or ALT>1.25xB for detection of relapse is better than the current serological criterium of aminotransferase levels above three times normal. However, for determination of the specificity of a change from undetectable to detectable anti-LSP, or for a rise of ALT above 1.25 times individual baseline values during remission a prospective study with repeated liver biopsies during therapy withdrawal would be required. However such a study might have ethical drawbacks. Our additional experience also seems to indicate that patients without anti-LSP at the time of therapy withdrawal with remaining absence of anti-LSP thereafter tend to relapse less than patients with anti-LSP before or during therapy withdrawal.

Therefore, extrapolating the results of our study, we think it is safest not to diminish therapy in an IAI-CAH patient with detectable anti-LSP. In our opinion, other requirements to be fulfilled before reducing therapy (in a patient without complications) include the completion of at least two years of continuous immunosuppressive therapy in adequate dosage for at least two years, preferably concurrent with normalized synthetic capacity of the liver.⁸⁴⁵ Determination of an individual 'baseline' ALT (B) in a patient without detectable anti-LSP at the time of a biopsy with 'CAH in remission' can help in follow-up during and after diminishing therapy. During this follow-up determination of anti-LSP (or ALT with ALT>1.25xB as cut-off) on a regular basis can enable early detection of relapse, requiring immediate reinstitution of the previous therapy.

CHAPTER 15

CLINICAL SIGNIFICANCE OF SUBGROUPS OF IDIOPATHIC 'AUTOIMMUNE' CHRONIC ACTIVE HEPATITIS, SEROLOGICALLY DEFINED BY PRESENCE/ABSENCE OF ANTI SOLUBLE LIVER ANTIGEN (SLA) AND OTHER AUTOANTIBODIES.

SUMMARY

A classification of idiopathic 'autoimmune' CAH, based on the presence or absence of autoantibodies against LKM-1, SLA, ANA, SMA and AMA was recently proposed (type 1= ANA-positive, 2=LKM-positive, 3= SLA-positive, 4= SMA-positive but ANA-negative, 5= unclassified: no autoantibodies or only AMA).

The aim of the present study was to investigate the clinical significance of such a classification in terms of survival, independent prognostic factors, and requirement for immunosuppressive therapy. We studied 79 patients with CAH, split into two overlapping groups: group I with IAI-CAH (n=69) and group II requiring standardized therapy with prednisolone and azathioprine (n=42, of which 32 had IAI-CAH). In addition, the prevalence of two AMA-subtypes and antibodies against liver membrane antigen (LMA) was evaluated.

The antibody profiles of IAI-CAH types 1,3 and 4 were frequently encountered in our group of Dutch patients with IAI-CAH, or other types of CAH and CAH-like liver disorders, while the aggressive type 2 was not seen among our patients.

Survival did not differ among patients with IAI-CAH type 1, 3, 4 or 'unclassified'. Similar results were obtained when this classification was applied to group II. Among these 'types' of CAH, no differences were found regarding the necessity to apply immunosuppressive therapy, although a subgroup of patients with IAI-CAH type 1 without SMA required immunosuppression less often than other patients.

We recently showed that immunosuppressed patients with IAI-CAH do better than those who do not fulfill treatment criteria and are left untreated, and that age, aetiology of CAH (especially presence or absence of anti-HBs), and the administration of standardized immunosuppression -versus other or no therapy- are independent risk factors for mortality in CAH. We now show that presence or absence of ANA, SMA, SLA, LMA, AMA are not independent prognostic variables in IAI-CAH or CAH of other aetiology, requiring immunosuppression.

To conclude, there is no apparent clinical reason for distinguishing, by presence or absence of ANA, SMA or SLA, more types of IAI-CAH than 'IAI-CAH type 2' and 'other IAI-CAH'. Pathogenetic differences between CAH with and without these autoantibodies cannot be excluded, however.

INTRODUCTION

Chronic active hepatitis (CAH) is a histologically and clinically defined syndrome of viral (hepatitis B, hepatitis C, or hepatitis D virus) or unknown aetiology (idiopathic 'autoimmune' CAH - IAI-CAH). Several 'CAH-related liver disorders' have been described with similar manifestations.⁸⁴⁶⁻⁸⁴⁸ Although not pathognomonic for CAH or for one of its aetiologies, antinuclear antibodies (ANA) and anti smooth-muscle antibodies (SMA) -usually directed against actin-, antibodies directed at liver membrane antigen (LMA) or against epitopes in 'liver-specific membrane lipoprotein'(LSP) have been detected frequently and in high titres in IAI-CAH, while antimitochondrial antibodies (AMA) are infrequent in this disease. The outcome in patients requiring immunosuppressive therapy in IAI-CAH did not differ, however, between patients with 'classic' CAH with ANA or SMA, or those without these antibodies.⁸⁴⁹

Recently, apart from the 'classic (lupoid)' form of IAI-CAH, a second form of IAI-CAH was recognized with a more aggressive behaviour, worse prognosis, and frequently an acute (even fulminant) onset, requiring immunosuppressive treatment. Serologically, this 'autoimmune hepatitis type 2' (IAI-CAH type 2) was characterized by the presence of -previously described⁸⁵⁰- anti liver/kidney microsome antibodies of type 1 (anti-LKM1, directed at a 50 kD cytoskeletal protein in rough and smooth endoplasmatic reticulum) and the absence of antiactin, ANA, and AMA - although organ-specific autoantibodies, such as antithyroid antibodies, were often present.⁸⁵¹ It was found that an antibody to liver cytosol antigen(s) (anti-LC1) is a second marker of IAI-CAH type 2, while anti-LKM1 and anti-LC1 were not found in patients with other disorders.⁸⁵²

Manns *et al.* described autoantibodies to another soluble liver antigen (SLA) in 23 of 60 IAI-CAH patients (6 without detection of other autoantibodies), that were not detectable in viral CAH or other liver disorders.⁸⁵³ It was proposed that the presence of SLA (with or without SMA, LMA or AMA) serologically defined a third type of IAI-CAH, responding well to immunosuppressive therapy. Furthermore, patients with SMA but without ANA might form a fourth subgroup where the necessity to institute therapy was more variable.⁸⁵³ However, in a recent review of IAI-CAH, the significance of such a classification was questioned, and it was stated that more research in this field was needed.⁸⁵⁴

The aim of the present study was to investigate the clinical significance of a classification of IAI-CAH based on the presence or absence of SLA, anti-LKM1, ANA, SMA, and/or AMA in terms of patient survival, recognition of independent prognostic factors and requirement for immunosuppressive therapy. In addition, the prevalance of two AMA subtypes and anti liver membrane antibody (LMA) was evaluated.

MATERIALS AND METHODS

Patients

We studied 79 patients with histological and clinical features of CAH,⁸⁴⁸ referred between 1970 and 1988, and divided into two overlapping groups (table 15.1).

Chronic active hepatitis	(CAH)	group I	group II	
1-idiopathic 'autoimmune'	(IAI-CAH)	69	32	
2-anti-HBs positive	(antiHBs CAH)		5	
3-hepatitis C (non-A,non-B)	(HCV-CAH)		1	
'CAH-related liver disease'	('CAH')			
4-'overlap CAH/small-duct PSC'	(CAH/PSC)		1	
5-oxyphenisatin induced CAH	(oxy-CAH)		1	
$6-\alpha_1$ -antitrypsin deficiency	(A1AT-CAH)		2	
total number of patients		69	42	

Table 13.1. I allow and activity of CATTOL CATTOLACCE INCLUSION
--

In group I, consisting of 69 consecutive IAI-CAH patients, presently known causes of CAH and related liver disease (like viral CAH, α_1 -antitrypsin deficiency, Wilson's disease, or drug-induced CAH) had been excluded.

Group II was formed by 32 out of the 69 patients with IAI-CAH from group I, and 10 other patients with CAH and related disorders of other aetiology (table 15.1). In group II, the common feature was that all patients required immunosuppressive therapy according to accepted criteria,^{855,856} and were subsequently treated with standardized immunosuppression (prednisolone(Pred) 15 mg and azathioprine(Aza) 75 mg for two months, followed by 10 mg Pred and 50 mg Aza).

Methods

Coded sera, stored at -20°C, were inactivated by heating at 56°C for 30 minutes, then tested. Positive and negative control sera were added to all assays.

SLA was determined by radioimmunoassay (RIA) according to the blocking principle, as described.⁸⁵³ Positive/negative ratios above 2.1 were regarded as diagnostic.

The AMA subtype antibodies M2 and Mx were determined by RIA.857

LKM-1 was determined by RIA according to a similar procedure⁸⁵⁸ as well as by indirect immunofluorescence on cryostat rat kidney sections.

SMA and AMA, were also determined by indirect immunofluorescence on cryostat rat kidney sections.

LMA was detected by immunofluorescence on viable rabbit hepatocytes, as described. 859

Analysis of survival

Data on survival had been retrieved from patient records, public registry offices, or family practitioners and specialists currently seeing the patients. We analyzed the interval between diagnosis of CAH by liver biopsy and November 1988, or until death or loss to follow-up. The probability of survival was calculated according to the Kaplan-Meier product limit method.⁸⁶⁰

Statistical analysis

Where appropriate, statistical analysis was performed with Chi-square analysis with p<0.05 as level of significance. Differences between subsets of patients in the analysis of survival were analyzed statistically by means of the log-rank test with p<0.05 as level of significance.⁸⁶¹ The prognostic significance of type of IAI-CAH as determined by presence or absence of ANA, SMA, SLA or LKM₁, as recently described, was determined by adding these variables to those we recently used for determining long-term prognostic variables in CAH.⁸⁶² Recalculation was performed using Cox's proportional hazards estimation of independent risk factors.⁸⁶³

RESULTS

The autoantibody profiles for SLA, anti-LKM1, ANA, SMA and AMA in patient groups I and II are shown in tables 15.2 and 15.3 respectively. Anti-LKM1, serologically defining IAI-CAH type 2, was not present in any of these patients.

As shown in table 15.2, the 'classic' type 1 IAI-CAH pattern, with detectable ANA with or without SMA, was found in 33 (48%) out of the 69 patients of group I, IAI-CAH type 3 (with SLA) was diagnosed in 6 (9%), and IAI-CAH type 4 (SMA but no ANA) in 17 (25%) of these patients. 13 (19%) out of the 69 patients remained unclassified when using this system and may be referred to as having 'cryptogenic' CAH. 45 (65%) out of the 69 IAI-CAH patients required some form of immunosuppressive treatment: such treatment was administered in 19 (58%) out of the 33 'type 1' patients, in five (83%) out of the six patients with type 3 IAI-CAH (SLA positive), in 15 (79%) out of the 19 patients with IAI-CAH patients. Observed occurrence of patients requiring immunosuppression did not differ from expected frequencies for any of the types of IAI-CAH, when compared to the whole group of IAI-CAH patients. Inclusion or exclusion of patients who were only given therapy during exacerbations did not affect

IAI-CAH	LKM-1	SLA	ANA	SMA	AMA		Т	herap	y			
type:						1	2	3	4	5	tota	ıl
						n	n	n	n	n	n	
1	-		+	+	-	9	3	2	7	3		_
1	-	~	+	-	-	1	0	0	7	1	33	
3	-	+	+	+	-	2	1	0	0	0		
3	-	+	-	+	-	1	0	0	1	0		
3	-	+	+	+	+	0	0	0	0	1	6	
4	-	-	-	+	-	9	2	2	2	2	17	
x	-	-	+	-	+	0	0	0	5	0		
х	-	-	-	-	+	0	0	0	0	1		
x	-	-	-	-	-	3	1	0	2	1	13	
Total nun	nber of p	atients:				25	7	4	24	9	69	

Table 15.2.Autoantibody profiles and therapy of 65 patients with IAI-CAH
(group I).

ad therapy: 1=Standardized therapy with prednisolone and azathioprine without preceding immunosuppression, 2= as 1, but with preceding immunosuppression, 3=non-standardized therapy with prednisolone and azathioprine, 4=untreated, 5=prednisolone during exacerbations alone.

x=unclassified (also called: type 5).

n denotes number of patients with this autoantibody pattern and this therapy.

Table 15.3. Number of patients in group I requiring immunosuppressive therapy; there was no significant difference between 'IAI-CAH types', although fewer patients in the subgroup of type 1 IAI-CAH without SMA required such therapy.

IAI-CAH type	patient continu	patients on immunosuppres continuous		kind	
1	15/32	(47%)*	19/32	(59%)**	
3	4/6	(67%)	5/6	(83%)	
4	13/17	(76%)	15/17	(88%)	
x	4/13	(31%)	6/13	(46%)	
Total	36/69	(52%)	45/69	(65%)	

ad *) 1/7 patients without SMA (Chi-square=11.2, p<0.005)

ad **)2/7 patients without SMA (Chi-square= 6.3, p<0.05)

results. The only exception was that the subgroup of patients with IAI-CAH 'type 1' with ANA but without SMA, required therapy significantly less often than the other patients (table 15.3).

As indicated in table 15.4, in group II, consisting of 42 patients requiring standardized immunosuppressive therapy according to accepted criteria, 16 patients had the autoantibody profile of IAI-CAH type 1; 12 of these patients had IAI-CAH without anti-HBs, while in 4 of these 16 CAH-patients antibodies to hepatitis B surface antigen (anti-HBs) were detected.

type	:LKM-1	SLA	ANA	SMA	AMA	aetiology	Numb n	er of patients total n
1	-	-	+	+		1	12	
1	÷	-	+	+	1	2	4	16
3	-	+	+	+	-	1	3	
3	(-)	+	-	+		1	1	4
4				+		1	11	
4	-	-	-	+	-	4	1	
4	-	-	-	+	-	5	1	
4	-	-	i n ti:	+	3=0	6	2	15
x		(H)	+	-	-	1	1	
x) ((- .)		-		1	4	
x	-	120	- 10 C	14	3 <u>1</u> 2	2	1	
x	-	17.1	-	: T .:	3 - 5	3	1	7

Table 15.4.Autoantibody-profiles in 42 patients in group II(CAH requiring immunosuppression).

Actiology: numbers indicating actiology of CAH correspond with those used in table 1. Type denotes antibody profiles as described for IAI-CAH types 1-4. x=unclassified (also called:type 5).

All 4 patients with SLA (the autoantibody pattern of IAI-CAH type 3) requiring standardized therapy had IAI-CAH without anti-HBs, and therefore also belonged to group I described above. SLA was not detected in the ten patients of group II with CAH or 'CAH-related liver disease' of other aetiologies.

The antibody profile of IAI-CAH type 4 (only SMA) was detected in 15 (36%) out of the 42 patients; 11 had IAI-CAH without anti-HBs, 1 an overlap syndrome between

CAH and 'small-duct' primary sclerosing cholangitis (CAH/PSC), two patients had partial MZ-phenotype α_1 -antitrypsin deficiency and one patient had been abusing oxyphenisatin before and during immunosuppressive therapy.

Seven patients in group II remained unclassified, when using the classification designed for IAI-CAH: one patient (2%) with IAI-CAH had only ANA, and in six patients (14%) of the 42 patients, none of the antibodies under investigation were detectable: 4 of these 6 patients had IAI-CAH, in one patient anti-HBs was detectable, and in one patient the onset of CAH had -in retrospect- followed a blood-transfusion, suggesting CAH due to hepatitis C.

The AMA-subtype antibodies against M2 and Mx were detected in none of the patients of group II (tested before therapy). In group II, LMA was detected in 7 (25%) out of 28 IAI-CAH patients tested. All of these patients required standardized immunosuppressive therapy. Out of these 7 patients, 6 (86%) had IAI-CAH type 1 (5 patients with both ANA and SMA, one with ANA but without SMA), and one patient had IAI-CAH type 4 (only SMA positive).

As shown in previous chapters for IAI-CAH, standardized immunosuppressive therapy improved the histological activity score, alanine aminotransferase (ALT) and bilirubin, and hepatic protein synthesizing capacity, reflected in serum cholinesterase. Analysis of variance of results of this therapy in group II revealed no difference between treated patients with antibody profiles of IAI-CAH types 1,3 and 4. However, in three patients out of the group with antibody profile type 4, serum cholinesterase showed less improvement after two years of therapy (p<0.05). In these patients, however, this was related to aetiology of CAH: two patients had partial MZ-phenotype α_1 -antitrypsin deficiency and one patient had been abusing oxyphenisatin during therapy.

Although ANA and SMA frequently became undetectable after institution of immunosuppressive therapy, titres of SLA diminished, but with the antibody remaining detectable during immunosuppression. No new ANA or SMA was detected in patients tested serially before and during immunosuppression (data not shown).

Analysis of survival showed no differences in the log-rank test among patients with autoantibody patterns of type 1,3, 4 or the unclassified group ('type 5'), neither in group I (figure 15.1), nor in group II.

When presence/absence of ANA, SMA, SLA or AMA, were added to the variables we had recently evaluated for their long-term prognosis of survival in CAH,⁸⁶² recalculation showed that presence or absence of none of these antibodies was detected as an independent risk factor in the Cox's proportional hazards analysis.



Figure 15.1. Probability of survival (Kaplan-Meier) of patients with IAI-CAH (group I). 1= ANA-positive, 3= SLA-positive, 4= SMA-positive but ANA-negative, 5= unclassified (no autoantibodies or only AMA). Dashed line: probability of survival in all patients. No patients with IAI-CAH type 2 were detected. Probability of survival between groups 1, 3, 4 and 5 did not differ significantly.

DISCUSSION

Recently, an aggressive IAI-CAH type 2 was described with a worse prognosis than other types of IAI-CAH, characterized by the presence of antibodies against LKM-1 and/or LC1.^{851,852} Further subdivision of IAI-CAH into four subtypes based on presence or absence of ANA, SMA, LKM-1 and SLA was proposed after detection of autoantibodies against SLA in a subgroup of patients with IAI-CAH,⁸⁵³ but the clinical significance of such a classification was unknown. Using the proposed classification, the present study shows that outcome in terms of survival does not differ among patient groups with IAI-CAH type 1, 3, 4 or 'unclassified'. Similar results were obtained when this classification was applied to patients with CAH of various aetiologies who had fulfilled requirements for standardized immunosuppressive therapy. No differences among these 'types' of CAH were found regarding the necessity to apply immunosuppressive therapy, although a subgroup of patients with IAI-CAH type 1 without SMA required immunosuppression less often than other patients.

The antibody profiles of IAI-CAH types 1,3 and 4 were frequently encountered in our group of Dutch patients with IAI-CAH or other types of CAH and CAH-like liver disorders, while the aggressive type 2 was not seen among our patients.

We recently showed that immunosuppressed patients with IAI-CAH do better than those who do not fulfill treatment criteria and are left untreated. We also showed that age, aetiology of CAH (especially presence or absence of anti-HBs), and the administration of standardized immunosuppression versus other therapy or no therapy are independent risk factors for mortality in CAH.⁸⁶² We have now demonstrated that presence or absence of ANA, SMA, SLA or AMA are not independent prognostic variables in either IAI-CAH, or CAH requiring immunosuppressive therapy.

The question remains as to whether these antibodies have a pathogenetic role, or are related to different aetiologies of IAI-CAH, and can justify classification on such a basis.

Recently, it has been shown that antibodies against LMA are not specifically reactive to polypeptides or glycolipids,⁸⁶⁴ and are directed against submembraneous constituents of the hepatocyte, i.e. cytoskeletal components. This means that LMA might be directed at neoantigens, presented on the cell membrane in reaction to CAH, questioning their pathogenetic role.⁸⁶⁵ Presence of ANA and SMA in CAH are generally also regarded as such epiphenomena.

There is considerable evidence, however, confirming initial suggestions for a pathogenetic role of autoantibodies directed against 'liver-specific membrane lipoprotein' (LSP),⁸⁶⁶⁻⁸⁶⁸ especially for an important constituent of LSP, the asialoglycoprotein receptor, also known as 'hepatic lectin':^{869,870} An LSP-specific T-suppressor lymphocyte defect probably remains present when the aspecific T-suppressor defect in IAI-CAH is reversed by corticosteroids,^{871,872} and a defect in the inducers T-lymphocytes of these suppressor cells probably plays an important role in 'classic' IAI-CAH, and possibly IAI-CAH generally.⁸⁷³

It has been shown that antibodies to SLA and LSP are not cross-reactive.⁸⁵³ This, of course does not exclude a possible pathogenetic role of antibodies against SLA, for instance in antibody dependent cellular cytotoxicity, in a subgroup of patients with IAI-CAH. Neither is such a pathogenetic role for SLA excluded by the cytoplasmic nature of the antigens, since cytoplasmic antigens can be present at the surface of the cell membrane.⁸⁷⁴

Anti-LKM-1 has been extensively studied.⁸⁷⁵⁻⁸⁷⁶ Recently Manns *et al.* cloned the LKM-1 gene,⁸⁷⁷ and part of the gene showed homology with part of the genetic code of the Epstein-Barr virus(EBV) (Manns: personal communication). At present, it is unknown if this homology has an aetiological implication, although it is tempting to speculate that EBV might trigger autoimmunity in anti-LKM1 positive CAH. Other evidence for a viral 'trigger' of IAI-CAH is related to the measles virus. Antibodies to this virus are frequent in IAI-CAH,^{878,879} and the persisting measles virus genome was detected in a high percentage of IAI-CAH patients, in contrast to a control group.⁸⁸⁰

Antiviral antibodies to the cytomegalovirus (CMV) are also frequently detected in CAH,^{881,882} but we recently demonstrated that an aetiological role in IAI-CAH is unlikely, at least by molecular mimicry between CMV-induced antigens and epitopes in LSP.⁸⁸³

In conclusion, other studies indicate that the prognosis of patients with IAI-CAH type 2 is worse than that of patients without anti-LKM-1 or anti-LC1 antibodies. However, the present study indicates that there is no apparent clinical reason for distinguishing subtypes of IAI-CAH based on presence or absence of ANA, SMA or SLA. There might however be pathogenetic considerations for such classifications. Cloning of autoantigens such as SLA may provide more insight into the possible pathogenetic importance of presence of antibodies against other autoantigens than those represented in LSP.

Chronic active hepatitis of various aetiologies

CHAPTER 16

ANTI-CARDIOLIPIN ANTIBODIES IN CHRONIC ACTIVE HEPATITIS.

Prevalence in Subgroups and Lack of Correlation with Anti-DNA Antibodies or Extrahepatic Disease.

SUMMARY

We assessed the prevalence, clinical associations and interrelations of anti-cardiolipin antibodies (ACA), anti-DNA antibodies and anti-nuclear antibodies (ANA) in 176 patients with histologically proven chronic active hepatitis (CAH) of various aetiologies. ACA were present in 30 (17%) of all 176 patients, and in 17 (26%) of the patients with idiopathic or 'autoimmune' CAH (IAI-CAH). ANA were present in 57 (32%) of the 176 CAH-patients, and were in most cases (n=52) of the homogeneous type. In 2 out of 3 ANA-positive patients with 'cholestatic CAH', anticentromere antibodies were detected. Anti-'nuclear lamina' antibodies (ANLA) were present in 96 (55%) of the patients and were equally distributed over the several subgroups.

Anti-DNA as assessed by immunofluorescence with Crithidia Luciliae as a substrate, were absent in 98% of the 176 CAH-patients. In 2 idiopathic or 'autoimmune'-CAH patients without anti-HBs, and one patient with 'anti-HBs positive CAH' these anti-bodies were detectable. By ELISA, 10% of IAI-CAH patients were positive for IgG-anti-DNA and 18% for IgM-class anti-DNA.

No clear relationship was detected in CAH between ACA and thrombosis or habitual abortion. Neither with regard to ANA, nor to anti-DNA or ACA could a clear relationship be detected with associated extrahepatic autoimmune disease.

INTRODUCTION

Anti-phospholipid antibodies (anti-PL) have been detected in infectious and autoimmune diseases since the introduction of the Wasserman reaction.⁸⁸⁴ The finding of a biological false positive syphilis test serology (BFP-STS),⁸⁸⁵ which is considered to be primarily an IgM response,⁸⁸⁶ was noted in 16% of patients with systemic lupus erythematosus (SLE), ⁸⁸⁷ and in several other diseases such as rheumatoid arthritis,⁸⁸⁸ autoimmune thyroiditis,⁸⁸⁸ and chronic active hepatitis.⁸⁸⁹⁻⁸⁹¹ In the 1950's, several reports of an (in vitro) coagulation inhibitor in association with the BFP-STS led to the demonstration of the anti-PL antibody nature of the 'lupus anticoagulant' (LAC), which in some cases could be absorbed by cardiolipin and Kahn antigen as used in syphilis test reactions.⁸⁹²

Recently, sensitive assays have been developed to quantify these anti-cardiolipin antibodies (ACA).^{893,894} The association between a BFP-STS and the presence of LAC, and between the presence of ACA and LAC has been demonstrated.^{892,893,895-897} IgG-ACA have been detected in the sera of 19-82% of patients with SLE,^{893,894,898-901} and also in 33-50% of patients with rheumatic disease,^{899,901} 28% of patients with psoriatic arthritis,⁸⁹⁹ 71% of patients with syphilis,⁹⁰¹ 61% of patients with malaria.⁹⁰¹ It has also been demonstrated in ANA-positive Felty's syndrome, essential type II mixed cryoglobulinaemia, mixed connective tissue disease, systemic vasculitis, primary sicca syndrome,^{893,899,901} and even with a prevalence of 52% in an elderly normal population.⁹⁰²

A relationship between both ACA and LAC with thrombosis,^{893,903-906} habitual abortion,⁹⁰⁷⁻⁹¹¹ and thrombopaenia^{895,897} has been suggested in several studies in highly selected groups of (mostly) SLE patients.⁹¹²

Cross-reactivity between ACA and anti-DNA antibodies in SLE may exist⁹¹³⁻⁹¹⁶ and may be more frequent with single stranded DNA than with double stranded DNA.⁹¹⁷ The epitopes involved in this cross-reactivity appear to be the phosphodiester-linked phosphate groups present in all polynucleotides and in cardiolipin.^{918,919} Smeenk *et al.* have found that low-avidity serum anti-DNA antibodies cross-react with cardiolipin, while high-avidity antibodies do not.⁹²⁰

In chronic active hepatitis (CAH), anti-DNA antibodies have been recognized since the earlier descriptions of the disease, usually assessed by the Farr assay.⁹²¹⁻⁹²⁵ However, no data are available on the prevalence of ACA, its relationship with ANA and anti-DNA, and its possible clinical significance in CAH.

To assess the prevalence and clinical associations of ACA in a disease different from SLE, we studied ACA and its relation to ANA and anti-DNA as assessed by two different methods in a well-defined population of 176 consecutive patients with histologically proven chronic active hepatitis of various aetiology. We evaluated whether the presence of ACA was associated with extrahepatic and/or thrombotic disease in this group of patients.

MATERIALS AND METHODS

Patients

We studied 176 consecutive patients with a histology of CAH,^{926,927} referred to the division of Hepatology of the University Hospital of Groningen between 1969 and 1988 and belonging to several subgroups (table 16.1). Patients were classified according to

the (presumed) aetiology of CAH into CAH with circulating hepatitis B surface antigen (HBsAg-CAH); CAH with circulating antibodies to HBsAg but without HBsAg (anti-HBs-CAH); idiopathic 'autoimmune' CAH (IAI-CAH) without anti-HBs; 'non-A,non-B' CAH without HBsAg or anti-HBs but related to blood-transfusion or intravenous drugabuse - probably related to the hepatitis C virus in most of these patients (HCV-CAH)-; CAH related to α 1-antitrypsin deficiency (A1AT-CAH) (all patients underwent phenotyping); and "other CAH". The group with "other CAH" consisted of the CAH-related disorders: 'overlap' between CAH and primary biliary cirrhosis (CAH/PBC); 'overlap' between CAH and primary sclerosing cholangitis (CAH/PSC); drug-induced CAH (DI-CAH); and CAH related to Wilson's disease (Wilson-CAH). Age, sex and classification of the patients are shown in table 1. Sera from 32 apparently healthy adults served as controls.

Methods

Patient records were reviewed according to a protocol for the presence of venous or arterial thrombosis, habitual abortion and extrahepatic autoimmune disease. Sera obtained during active hepatitis at the time of the diagnostic biopsy had been frozen at -20°C and were thawed at room temperature (without heat inactivation) for determination of ACA, anti-DNA, and ANA. Positive and negative controls were added to all measurement series. All sera were tested for ACA, ANA and anti-DNA (immunofluorescence). Testing for anti-DNA of the IgG and the IgM class by enzyme-linked immunosorbent assay (ELISA) was performed in all ACA positive sera, and in all sera from patients with idiopathic or 'autoimmune' (IAI-)CAH without anti-HBs. In addition, ANA-positive IAI-CAH sera were tested for the presence of antibodies to extractable nuclear antigens (ENA).

Anti-cardiolipin ELISA

ACA were quantified by ELISA according to the descriptions of Loizou *et al.*⁸⁹⁴ The upper reference limit, defined as the 95th percentile of the control population, was 22.5 Units (median 5 U) (figure 16.1).

Anti-DNA ELISA and immunofluorescence

An immunofluorescence assay for the detection of anti-DNA antibodies, using the kinetoplast of Crithidia Luciliae as a substrate, was performed as described.⁹²⁸ As a conjugate, FITC-labelled anti-total IgG (Sigma) was used. Sera were tested in doubling dilutions from 1:10 through to 1:360. For anti-DNA by immunofluorescence, a positive fluorescence in a dilution of 1:40 or more was considered a positive result.

Where anti-DNA were also assessed by ELISA, we used calf thymus DNA (Sigma) as a substrate (after precoating with protamine sulphate). Horse radish peroxidase labelled goat anti-human IgG or IgM (Kallestad) was used as a conjugate. The upper reference limit for anti-DNA antibodies by ELISA, defined as the 95th percentile of the control population, was 10 Units.

ANA immunofluorescence

ANA fluorescence with cultured human fibroblasts as a substrate, was performed as described.⁹²⁹ Sera were tested in doubling dilutions from 1:10 through to 1:360. A positive fluorescence in a dilution of 1:40 or more was considered a positive result.

Antibodies against extractable nuclear antigens (a-ENA)

Determination of a-ENA was performed by counter immunoelectrophoresis using an extract of rabbit thymus acetone powder (Pel-Freeze) (for SS-B (La), nRNP and Sm) and an extract of human spleen (for SS-A (Ro)) as antigenic sources, together with the corresponding reference sera (CDC, Atlanta, USA).

Statistical analysis

Statistical analysis was performed with the Mann-Whitney U test, and Spearman's rank correlation where appropriate, with p<0.05 as the level of significance.

RESULTS

Patients

The ages of the patients with oxy-CAH were higher (median 61 years) than in all other groups (p<.005-p<.05). Ages between other groups did not differ (table 16.1).

ACA

The prevalence of ACA in the several subgroups is shown in table 16.2. In 30 of 176 CAH-patients (17%), ACA were detected during active disease (figure 16.1). There was no significant difference between subgroups of CAH regarding positivity for ACA. Although the median values (8.0-13.0 U) were within the reference range for all CAH groups, ACA values in all groups were higher than reference ACA-levels (median 3.9 U) (p<.05-p<.0001).

ANA

57/176 (32%) patients tested had anti-nuclear antibodies during active CAH (figure 16.2). ANA-titres in IAI-CAH (median 40)(p<.0001), in anti-HBs CAH (median 40) (p<.005), and in 'other' CAH (median 5) (p<.01), were higher than in HBsAg-CAH (median 0). ANA titres in anti-HBs CAH were higher than those in NANB-CAH (median 0) (p<.05). In 53/59 (90%) positive cases, ANA were of the homogeneous type. A discretely speckled fluorescence pattern was seen in 2 IAI-CAH patients and 2 CAH/PBC patients (1 of these in combination with a faint homogeneous fluorescence). A coarsely speckled pattern, was observed in one patient with IAI-CAH. A ring-like fluorescence of the nuclear lamina was observed in 55% of the patients, with equal distribution over the subgroups (table 16.2).

Aetiology*	Number of	Sex	Age (years)		
	patients	M: F	median	range	
IAI, no anti-HBs	65	11: 54	51	11-78	
HbsAg	49	36: 13	42	18-74	
IAI, with anti-HBs	13	3: 10	46	17-81	
HCV	10	4: 6	37	18-77	
DI	17	3: 14	61	24-78	
other	22	6: 16	34	15-72	
Total	176	63:113	46	11-81	

 Table 16.1.
 CAH patients grouped according to aetiology.

IAI = idiopathic or 'autoimmune' CAH

HBsAg = CAH with circulating hepatitis B surface antigen (HBsAg) Anti-HBs CAH= CAH with circulating antibodies to HbsAg

HCV= CAH due to parenteral non-A, non-B hepatitis (probably due to hepatitis C virus)

DI = drug-induced 'CAH' due to: oxyphenisatin(12), α-methyl-dopa(3), nitrofurantoin(1), diclofenac(1).

Other CAH:

9x CAH/PBC=	'cholestatic CAH' or 'mixed form'CAH/PBC
5x CAH/PSC=	'small-duct' PSC and/or CAH 3x Wilson-CAH= CAH associated
	with Wilson's disease
5x A1AT-CAH=	CAH related to α_1 -antimypsin deficiency

(PBC= primary biliary cirrhosis) (PSC= primary sclerosing cholangitis)

aetiology	patients (n)	ANA (n,%)	ANLA (n,%)	ACA (n,%)	anti-DNA(IF) (n,%)
IAI-CAH	65	36(55)*	34(52)	17(26)	2(3)&
HBsAg CAH	49	4 (8)	26(53)	5(10)	0
anti-HBs CAH	13	7(54)	8(62)	0	1(8)
HCV-CAH	10	0	6(60)	1(10)	0
DI-CAH	17	4(24)	6(35)	3(18)	0
CAH/PBC	9	3(33)#	7(78)	1(11)	0
CAH/PSC	5	1(20)	3(60)	3(60)	0
Wilson-CAH	3	0	2(67)	0	0
AAT-CAH	5	2(40)	4(80)	0	0
Total	176	57(32)	96(55)	30(17)	3(1.7)

	Table 16.2.	Prevalence of several	autoantibodies i	n CAH subgroups.
--	-------------	-----------------------	------------------	------------------

numbers denote number of patients with positive results (percentages between brackets).

ad*) 2 sera discretely speckled, 1 serum coarsely speckled + homogeneous.

ad #) 1 serum discretely speckled, 1 serum discretely speckled + homogeneous.

All other ANA-fluorescence patterns were homogeneous.

ad &) 6/61(10%) IgG anti-DNA by ELISA, and 14/61(23%) IgM anti-DNA by ELISA.



Figure 16.1. Anti-cardiolipin antibodies (ELISA) in CAH (n=176) and controls (n=32). M= median. Dotted line=95% upper reference limit (22.5U). IAI=IAI-CAH (with or without anti-HBs) (n=78), HBsAg=HBsAg-positive CAH (n=49), other=other types of CAH, as shown in table 16.1 (n=49). *:p<0.05, **:p<0.005, ***:p<0.001 versus controls.



Figure 16.2. Anti-nuclear antibodies in CAH (n=176). Fluorescence pattern: Δ =homogeneous, *=homogeneous+discretely speckled, x=discretely speckled, +=coarsely speckled. IAI*=IAI-CAH without anti-HBs.

For further abbreviations: see table 16.1.

ENA

No antibodies to ENA were detected in sera from the ANA-positive IAI-CAH patients.

Anti-DNA

Only 3/176 patients showed anti-DNA antibodies by immunofluorescence with the kinetoplast of Crithidia Luciliae as a substrate, as shown in table 16.2. All three patients (2 with IAI-CAH, 1 with anti-HBs CAH) had a homogeneous ANA fluorescence pattern with a titre \geq 320. ACA, ANA and anti-DNA (both immunofluorescence and ELISA) of these 3 patients are shown in table 16.3.

Patient a number	nti-DNA(IF) Crithidia (titre)	anti-DNA ELISA(IgG) (Units)	anti-DNA ELISA(IgM) (Units)	ACA(IgG) ELISA (Units)	ANA (IF) (titre)	CAH- subgroup
1	160	3	19	6	640	IAI
2	320	9	2	19	640	IAI
3	160	15	52	7	640	anti-HBs
normal	<10	<10	<10	<22.5	<40	

Table 1	6.3. Antibody	titres in	CAH with	positive	anti-DNA(IF).
---------	---------------	-----------	-----------------	----------	---------------

By ELISA, anti-DNA antibodies of the IgG-class were detected in 6/61 (10%) IAI-CAH patients, and anti-DNA antibodies of the IgM-class in 11/61 (18%) IAI-CAH patients, as shown in figure 16.3. In 3 of these patients, IgG and IgM anti-DNA were both present (table 16.4).

The total prevalence of anti-DNA antibodies was 17(28%) out of 61 patients with IAI-CAH, as determined by ELISA.

Table 16.4 .	ACA and anti-DNA	(ELISA) in IAI-CAH (n=61)
---------------------	------------------	----------------------	-------

		IgG a	IgG anti-DNA		inti-DNA	
		+	-	+	-	
ACA	+	2	13	5	10	
	8 — 2	4	42	9	37	

numbers denote number of patients.



Figure 16.3. Anti-DNA antibodies (ELISA) in IAI-CAH (n=61).

Serological relation between ACA, ANA, and anti-DNA

In the IAI-CAH patients, co-occurrence of anti-DNA IgG with ACA was rare, while cooccurrence of ACA with anti-DNA IgM was present in only 5/61 IAI-CAH patients tested (table 16.4). None of 9 ACA-positive patients with CAH of other aetiologies had detectable anti-DNA IgG and IgM anti-DNA was found by ELISA in only one of these patients. A weak correlation was present between the titres of ACA and that of ANA (R=.19, p<.05).

Correlations with extrahepatic disease

One patient had had thrombosis just before the start of standardized therapy. No ACA were present when tested on several occasions before and during therapy with anticoagulants and immunosuppressive medication.

14 (25%) of 55 women in the child-bearing age (17 to 45 years of age) had detectable ACA. In our retrospective analysis, no cases of habitual abortion were detected. From two patients who had had a spontaneous abortion (one patient before and one patient after the time of diagnosis of CAH), one had ACA while the other lacked this autoantibody. Two patients (one with and one without ACA) had each delivered three healthy children during remission from CAH during maintainance therapy consisting of 10 mg prednisolone and 50 mg azathioprine daily; the patient without ACA experienced one spontaneous abortion after the second uncomplicated delivery of a healthy baby.

	n=	number of patients with			
symptoms		ACA	ANA	anti-DNA(IF)	
Venous thrombosis	1	0	0	0	
Recurrent abortion (out of 55 women)	0				
Spontaneous abortion (only once)	2	1	0	0	
Hemolytic anaemia	3	1	0	0	
Arthritis	17	5	9	1	
Pleuritis	1	1	1	1	
Pericarditis	2	1	1	1	
Hashimoto's thyroiditis					
with hypothyroidism	2	0	1	0	
Goiter (hypothyroidism after thyroidectomy)	1	1	0	0	
Diabetes mellitus	6	2	2	0	
Bronchial asthma	7	1	2	0	
Interstitial pulmonary fibrosis	1	0	0	0	
Raynaud's phenomenon	1	1	0	0	
Oesophageal hypomotility	1	1	1	0	
Transient lymphadenopathy	2	1	0	0	
Relapsing iridocyclitis	1	0	0	0	
Ulcerative colitis	2	2	0	0	
Achlorhydria	1	0	1	1	
Chronic gastritis	2	0	1	0	
Glomerulonephritis					
-Membranoproliferative	1	0	1	0	
-Focal	1	0	1	1	
Proteinuria, cause unknown	3	0	1	1	
Pancreatic insufficiency	1	0	1	1	
Malabsorption	1	0	1	1	
Polyneuritis + tremors	1	0	1	0	
Vitiligo					
Stiff painful muscles (myositis?)	3	0	1	0	
Arteritis	1	0	1	0	
Allergic vasculitis	1	0	1	0	
Sjögren's syndrome	1	0	1	0	
Alzheimer's disease	1	0	1	0	
TOTAL	69	18	31	8	

Table 16.5.Associated (autoimmune) disease in 176 patients with CAH.

One or more extrahepatic (autoimmune) disease manifestations were detected in association with CAH 69 times in 47 (27%) out of the 176 patients, as shown in table 16.5. No clear associations between associated (autoimmune) disease and the presence of ANA, ACA or anti-DNA (ELISA) could be detected, although two of the three patients with anti-DNA (as detected by the Crithidia assay) had associated autoimmune disease (table 16.5).

DISCUSSION

Prevalence

The present study indicates that ACA is not infrequent in CAH. Cross-reactivity between ACA and anti-DNA in CAH appears to be limited, and no relation of ACA with thrombotic events, recurrent abortion or associated extra-hepatic autoimmune diseases became apparent.

Anti-double-stranded DNA antibodies were absent in 98% of our 176 CAH patients as assessed by indirect immunofluorescence with Crithidia Luciliae as a substrate, while these antibodies were detected more frequently by ELISA. Others, however, have reported a higher prevalence of anti-DNA positivity in CAH by the Crithidia assay;^{931,932} however, most of these cases of CAH were drug-related. Our findings support the suggestion from other studies^{933,934,935} that anti-DNA, as detected by the Crithidia assay, allows an immunological distinction between CAH and SLE.

ANA were detected frequently in our CAH patients, the highest frequency being found in IAI-CAH. The fluorescence pattern was usually of the homogeneous type, in accordance with the literature.⁹³⁶ ANA in autoimmune liver disease can be directed to a nuclear protein of 54 kD, to a nuclear matrix protein of 150 kD, a U1-RNP protein of 70 kD, and to a nuclear 25 kD doublet.⁹³⁷ ANA against histones, as frequently present in PBC, are rare in hepatitis B virus-associated CAH.⁹³⁸ Characterization of these antigens is still incomplete.

A ring-like perinuclear lamina fluorescence pattern was detected in 55% of our CAH patients. Such a pattern has recently been described both in acute hepatitis, granulomatous hepatitis, CAH and PBC. The antibodies responsible for the ring-like fluorescence in CAH and acute hepatitis appeared to be predominantly directed at lamins A and C of 70 and 60 kD (anti-nuclear lamina antibodies, or ANLA),^{939,940} and not at the nuclear envelope-associated proteins with a molecular weight of approximately 200 kD, which is specific for PBC.⁹⁴⁰⁻⁹⁴³

The absence of antibodies to ENA in our IAI-CAH patients, as assessed by counter immunoelectrophoresis, is in agreement with the absence of anti-SS-B (La) in CAH,⁹³³ when detected by immunodiffusion. However, Kurki *et al.* found anti-SS-B (La) in 38%

of CAH patients when using a sensitive ELISA.⁹³³ Presence of anti-SS-B would be in agreement with the concept that the sicca syndrome associated with CAH is of the primary type, but the sicca syndrome is of PBC of the secondary type.⁹⁴⁴ We detected the sicca syndrome in only one patient in our population. This could explain why we could not find anti-SS-B (La). Using sensitive methods of detection, Penner also found that anti-La antibodies (50 kD) and circulating immune complexes containing histones (especially anti-H1) were invariably present in patients with IAI-CAH associated with the sicca syndrome. IAI-CAH associated with membranous glomerulonephritis has been described to be associated with U1-RNP-containing immune complexes.⁹⁴⁵ Sjoegren's syndrome in PBC can also be associated with sensitisation to salivary antigens and deposition of Ro/anti-Ro immune complexes.⁹⁴⁶

Serological correlations

As has already been mentioned, low-avidity anti-DNA antibodies cross-react extensively with cardiolipin, while high-avidity antibodies do not.⁹²⁰ Our study indicates that cross-reactivity between ACA and anti-DNA detected by immuno-fluorescence is limited in CAH. In most patients, anti-DNA antibodies detected by ELISA remained undetected in the immunofluorescence assay with Crithidia Luciliae as a substrate. This may be due to the fact that the latter assay detects predominantly high avidity anti-DNA, while the ELISA probably binds also anti-DNA of low-avidity. This indicates that low-avidity anti-DNA antibodies can occur in CAH, while high-avidity anti-DNA is very uncommon.

Clinical correlations

Despite the limitations of a retrospective study, our findings confirm the frequent occurrence of multisystem involvement in CAH and its lack of association with ANA,⁹⁴⁷ while no correlations with ACA could be detected.

We observed a relatively high prevalence of a discretely speckled ANA-fluorescence in cholestatic CAH ('CAH/PBC'). In ANA-positive PBC-patients,⁹⁴⁸⁻⁹⁵⁰ antibodies to a chromatin-associated centromere-protein of 19.5 kD, at present known to be part of protein(s) with a larger molecular weight, are associated with the presence of one or more features of the CREST-syndrome (calcinosis, Raynaud's phenomenon, oesofageal hypomotility, sclerodactyly, telangiectasia),^{935,949-952} the sicca syndrome9^{53,954} and kerato-conjunctivitis sicca.^{955,956} Despite the presence of a fluorescence pattern consistent with the presence of anti-centromere antibodies in CAH/PBC 'overlap' patients, no signs of the CREST syndrome or the sicca syndrome were detected in our 'CAH/PBC overlap' patients. However, no 'anti-centromere fluorescence pattern' was detected in patients with such features.

As mentioned, the correlation between ACA and thrombotic events or habitual abortion has been detected predominantly in highly selected groups of patients, usually with SLE. In an unselected population of patients, such correlations may be less clear. For instance, in an unselected population of patients with recurrent foetal loss Petri *et al.* found no association with the presence of LAC and ACA.⁹⁵⁶ In our CAH patients, no correlation between the presence of ACA and thrombosis or habitual abortion could be detected. Unlike SLE, in CAH the occurrence of spontaneous abortion (10 percent) is not higher than expected, although a higher than expected rate of perinatal fetal loss and obstetric complications occurs.⁹⁵⁷ ACA may be an epiphenomenon delineating a subgroup of patients at risk for habitual abortion and thrombosis in some diseases, -for instance SLE-, but not in other diseases such as CAH.

To conclude, ACA were present in 17% (in IAI-CAH 26%) of patients with CAH. Anti-DNA of high avidity is uncommon in CAH, while low avidity anti-DNA can be present. Detection of anti-DNA by indirect immunofluorescence on the kinetoplast of Crithidia Luciliae, favors a diagnosis of SLE instead of CAH. Cross-reactions between ACA and anti-DNA appear to be limited in CAH. ANA are frequently present and usually of the homogeneous type, except in CAH/PBC 'overlap syndrome' where a discrete-speckled ANA-pattern is more common. A ring-like anti nuclear lamina fluorescence pattern was frequent, probably reflecting the presence of anti-lamin A and C antibodies. No relationship between ACA and thrombosis or habitual abortion was detected. Neither were relationships detected with frequently occurring associated (autoimmune) disease.

CHAPTER 17

HUMORAL RESPONSES AGAINST DEFINED CYTOMEGALOVIRUS ANTIGENS AND 'LIVER-SPECIFIC MEMBRANE LIPOPROTEIN' IN CHRONIC ACTIVE HEPATITIS

Lack of evidence for aetiologic involvement of cytomegalovirus by molecular mimicry.

SUMMARY

There is evidence that antibodies to 'liver-specific membrane lipoprotein' (anti-LSP) are aetiologically involved in idiopathic or 'autoimmune'(IAI) chronic active hepatitis (CAH), but the trigger for development of IAI-CAH remains unknown. The prevalence of anti-cytomegalovirus antibodies (aCMV) in CAH has been reported to be higher than in the normal population. We tested the hypothesis that CMV is aetiologically involved in IAI-CAH.

Methods: Sera taken during active disease from 178 patients with CAH (82 IAI, 48 HBsAg-positive, 48 'other' CAH) were stored at -20°C and then tested by ELISA for aCMV of both IgG- and IgM-class to late antigen (LA). 34 IAI-CAH patients were also tested for presence of anti-LSP by radioimmunoassay (RIA), and the effect of preabsorption of anti-LSP positive IAI-CAH serum with LA was tested. 16 IAI-CAH sera - 4 anti-LSP negative but anti-LA-IgG-positive (group I), 6 anti-LSP-positive but anti-LA-negative (group II), and 6 positive for both anti-LSP and anti-LA-IgG (group III)- were tested for presence of defined anti-CMV-membrane antigens with a molecular weight of 58 kD (58-MA) and CMV-immediate early antigen of 67 kD (67-IEA), by newly developed sandwich ELISA's using monoclonal antibodies.

Results: 122 (69%) of the 178 patients had anti-LA-IgG (more in IAI-CAH and HBsAgpositive CAH than in 'other CAH'). 69 (39%) had anti-LA-IgM (lower in IAI-CAH than in HBsAg-CAH or 'other' CAH). Age- and sex-matched prevalence of IgG-class aCMV in CAH was higher than reported in 1731 healthy blood donors (HBD) in a recent German study, but similar to prevalence in 1251 HBD in a recent Swedish study. Anti-LA-IgG titres were higher in IAI-CAH and HBsAg-positive CAH than in 'other CAH'. 22/34 IAI-CAH patients tested had both anti-LA-IgG and anti-LSP, 2/34 had none of these antibodies, 8/34 had anti-LSP but no anti-LA-IgG, 2/34 had no anti-LSP but had anti-LA-IgG. Preabsorption of anti-LSP positive IAI-CAH serum with LA before RIA did not lower anti-LSP titres. In group I, 4/4 patients had a67-IEA and 3/4 had a58-MA. None of the 6 patients in group II, but all in group III had a58-MA or a67-IEA. Conclusions: Prevalence of anti-CMV-LA IgG in CAH is similar to or higher than in HBD. Prevalence and titres are higher in IAI-CAH and HBsAg-positive CAH than in 'other' CAH. There is no correlation between presence/absence of anti-LA-IgG and anti-LSP, despite the fact that both are often present. It is unlikely that there is an aetiological role for CMV in inducing autoimmunity in IAI-CAH by molecular mimicry between LSP and defined CMV-induced membrane antigens.

INTRODUCTION

Chronic active hepatitis (CAH) is characterized by a chronic portal and periportal inflammation of the liver, with piecemeal necrosis and peripolesis,⁹⁵⁸ and with many aetiologies resulting in the same morphology. Frequently, however, no aetiology can be detected, and in these patients the CAH is termed 'idiopathic' or 'cryptogenic'. Since many of these patients display autoantibodies, hypergammaglobulinaemia, and other characteristics attributed to autoimmunity, idiopathic CAH in such patients is termed idiopathic 'autoimmune' CAH (IAI-CAH). It is this group of patients that usually reacts favorably to immunosuppressive therapy.⁹⁵⁹ More specific treatment modalities, however, await elucidation of aetiological factors.

Besides the development of autoantibodies, increased antibody titres to exogenous antigens were reported, notably bacterial and viral.^{960,961} High antibody titres can occur against gut-derived bacteria such as E. Coli, Bacteroides, and Salmonella.^{960,962} The production of antibodies to Haemophilus influenzae B -which usually targets the respiratory tract- is, however, seldom increased in patients with acute and chronic liver disease. If increased, it is due to a non-specific hyperreactive state.⁹⁶⁰ With regard to antiviral antibodies in CAH, elevated antibody titres were described for measles, rubella^{963,966} and cytomegalovirus (CMV),⁹⁶⁶ while no increased antibody titres against Coxsackie B virus were detected.⁹⁶⁵ Elevated antibody titres to CMV early antigen⁹⁶⁷ were also described in oxyphenisatin-induced CAH,⁹⁶⁸ and in acute hepatitis B.⁹⁶⁹ A higher prevalence of antibody titres to adenovirus was also found in acute hepatitis.⁹⁷⁰

The presence of antiviral antibodies was found to be uncommon in chronic persistent hepatitis, and in cirrhosis without hepatitis.^{963,971} In a patient with CAH, significantly rising antibody titres to rubella and measles virus were described, following the onset of liver disease.⁹⁶⁵ Anti-measles antibodies in CAH can belong to the IgM-class of antibodies,⁹⁷² suggesting persistent production of measles virus antigens.

In contrast to the situation in the immunocompromised host (ICH),⁹⁷³ hepatitis especially of the chronic type- due to CMV in the non-ICH is less common. Histological and immunological differences between these two situations exist.⁹⁷⁴ Before intranuclear early and late antigens are induced by CMV-infection, the cell surface already expresses CMV-specific membrane neoantigens, CMV-MA.⁹⁷⁵ CMV-MA may contribute to early recognition and elimination of infected cells by cell-mediated immunity. Analysis of serological responses against defined CMV-antigens is now feasible by antigen capture ELISA's using monoclonal antibodies.^{976,977}
There is considerable evidence that in CAH, cytotoxic reactions of the antibody dependent (ADCC-)type are directed at hepatocyte plasma membrane-derived epitopes in the macromolecular, lipid-associated complex called 'liver specific antigen' (LSP).978,979 The liver-specific, species cross-reactive hepatic receptor for desialylated glycoproteins, the asialoglycoprotein receptor (ASGR), also known as hepatic lectin, is an important constituent of LSP.980 Anti-LSP antibodies appear to be predominantly directed at the ASGR, although a role for anti-LSP directed at other antigenic sites is not yet excluded. Anti-LSP is usually present in IAI-CAH patients requiring immunosuppressive therapy according to accepted criteria, and anti-ASGR is found in most of these anti-LSP positive CAH patients.981 It has also been shown that anti-ASGR antibodies preferentially coat periportal hepatocytes in the antegradely and retrogradely perfused rat liver.⁹⁸² An LSP-specific T-suppressor lymphocyte defect remains present when the aspecific T-suppressor defect in IAI-CAH is reversed by corticosteroids.983,984 A defect in the inducer T-lymphocytes of these suppressor cells probably plays an important role in IAI-CAH.985 The triggers for the induction of autoimmunity remain unknown, however.

It is not clear whether the increased antibody titres in CAH to viral and bacterial antigens are epiphenomena, or whether these agents are involved in the aetiology of IAI-CAH.

Another autoimmune disease, diabetes mellitus type I, recently appeared to be associated with the cytomegalovirus. CMV genome was found in the lymphocytes of 13/59 (22%) of diabetic patients, in contrast to 1/38 (2.6%) of the control subjects, and there was a strong correlation between CMV genome and presence of islet cell autoantibodies.⁹⁸⁶

To further investigate the relationship between CAH and CMV, we quantified antibodies of the IgG and IgM class to CMV late antigen (LA) in a population of 178 CAH patients with various aetiologies. This was to compare qualitative and quantitative differences between IAI-CAH patients and patients with CAH of known aetiology, with respect to these antibodies. The prevalence of anti-CMV antibodies was compared to studies of healthy blood donors. In the IAI-CAH patients we investigated the relationship between the presence of anti-LSP, anti-CMV-LA, anti-CMV-specific immediate early antigen with a molecular weight of 67 kiloDalton (67-IEA) and anti-CMV-specific membrane antigen of 58 kD (58-MA). The ability of CMV-LA antigens to inhibit binding of IAI-CAH serum containing anti-LSP in the radioimmunoassay for detection of anti-LSP was tested for, in order that we may study a possible molecular mimicry between CMV-encoded and auto-antigens. Such a mimicry might provide a clue to the induction of IAI-CAH.

MATERIALS AND METHODS

Patients

From a series of 178 consecutive patients with histologically confirmed CAH, referred to the division of Hepatology of the University Hospital Groningen between 1969 and 1988, sera had been frozen and stored (-20°C) until use. 82 patients had IAI- CAH (12 with anti-HBs), 48 had hepatitis B-surface antigen positive CAH (HbsAg CAH), and 48 had 'other' CAH (table 17.1).

AETIOLOGY OF CAH		n=	TOTAL
'AUTOIMMUNE' -idiopathic	(IAI-CAH)	70	
-anti-HBs positive	(anti-HBs CAH)	12	82
-HBsAg POSITIVE	(HBsAg CAH)	48	48
-OTHER			
-hepatitis C ('non-A,non-B') CAH	(HCV-CAH)	11	
-'cholestatic CAH'	(CAH/PBC)	9	
-'small-duct' PSC and/or CAH	(CAH/PSC)	5	
-drug-induced 'CAH'*	(DI-CAH)	16	
-Wilson's disease 'CAH	'(Wilson-CAH)	3	
$-\alpha_1$ -antitrypsin deficiency	(A1AT-CAH)	3	
-IAI-CAH in multicystic liver	(cyst-CAH)	1	
-			48
Total			178

Table 17.1. CAH patients stratified on aetiologies

n denotes number of patients in each group.

ad *) Drug-induced cases were due to: oxyphenisatin(12), α -methyl-dopa(3), nitrofurantoin(1), diclofenac(1).

Sera from all 178 patients were investigated for the presence of antibodies to CMV- late antigen (CMV-LA) of the IgG and IgM classes. Assessment for anti-LSP was carried out in 34 untreated patients with IAI-CAH (4 patients with circulating antibodies to hepatitis B surface antigen (anti-HBs CAH), and 30 patients without such antibodies). We tested for inhibition of anti-LSP binding by LA-antigen. Newly developed sandwich ELISA's for detection of anti CMV 67-IEA and anti CMV 58-MA antibodies were performed on sera from 16 IAI-CAH patients: 6 with anti-LSP but no anti-LA IgG, 4 with anti-LA IgG but no anti-LSP, and 6 with both antibodies.

To provide enough age-matched controls with which to compare prevalence of anti CMV antibodies in CAH, we compared prevalence of anti CMV antibodies in our patient groups with that in North European healthy blood donors from two recently published studies that used similar sensitive detection techniques.^{987,988}

Anti-LSP radioimmunoassay

We slightly modified the original methods for radiolabelling LSP and performing the radioimmunoassay (RIA)⁹⁸⁹, and the assay was performed as described.⁹⁹⁰ Positive and negative control sera were added to all runs.

Anti-CMV-LA ELISA

IgG and IgM antibodies to CMV-LA were determined by enzyme-linked immunosorbent assay (ELISA) as described.⁹⁹¹ CMV and control antigens used as a substrate were prepared from fibroblasts, harvested 6-7 days after CMV infection. As controls in each assay, and for standardization between different assays, a CMV antibody-positive reference serum and a CMV-antibody negative reference serum pool were always included. All sera were tested at several dilutions. The difference in concentration of antibodies in the tested serum and the CMV antibody-positive reference serum was determined essentially by the so-called effective dose method.^{991a} The concentration of antibodies to CMV-induced antigens was expressed as a percentage of the CMV antibody-positive reference sample, which was designated as 100%, after logit transformation of the sigmoidal titration curve into a linear response curve. Then these percentages were converted into arbitrary units, and a concentration of more than 1 U/ml was considered to be a positive result.

Inhibition experiments

Inhibition of anti-LSP binding by LA-antigen was tested by adding LA in three doubling dilutions (1:50, 1:100, 1:200), diluted with the 0.1% BSA-borate-EDTA buffer used in the anti-LSP RIA, to an IAI-CAH serum containing both anti-LSP and anti-CMV-LA of the IgG class, and to another IAI-CAH serum containg anti-LSP but not anti-CMV LA antibodies. As controls in the same run, the anti-LSP titre was determined in these two sera in the same dilution, but without adding LA. This was done in buffer with LA in the three doubling dilutions mentioned before, and in buffer with control antigen (uninfected fibroblast material) in a 1:50 dilution, which is used in the anti-CMV-LA ELISA. Other anti-LSP positive and negative reference sera were included in the same run.

Antigen capture ELISA for 67-IEA and 58-MA

Newly developed monoclonal antibody based antigen capture immunoassays,⁹⁹² were used for detection of anti-CMV antibodies against the CMV-specific immediate early antigen with a molecular weight of 67 kiloDalton (67-IEA) and the CMV-specific membrane antigen of 58 kD (58-MA). The cell-free human CMV strain AD-169,

prepared as decribed,⁹⁹¹ was used to infect monolayers of human embryonic fibroblasts. Antigens from late-infected fibroblasts (CMV-antigen, 4-5 days post infection) were prepared by detergent extraction and purified with low- and high-speed centrifuging. Monoclonal antibodies against these antigens were raised and isolated from mouse ascites using standard ammonium sulphate precipitation and DEAE chromatography techniques. Monoclonal antibody C10 reacted with a CMV-antigen present in the nucleus of CMV infected cells from about 1h post-infection,992a and in both nucleus and cytoplasmic granules of late-infected cells. Another monoclonal antibody (C40), also prepared with these techniques, was reactive with a CMV glycoprotein which has homology with herpes simplex glycoprotein B (58-MA) (unpublished results, courtesy of M.P. Cranage, Cambridge; Cranage et al., 1986).993 C40 is reactive with the perinuclear region and the plasma membranes of late infected cells. These monoclonal antibodies were used for coating the ELISA plates (Greiner) in Na₂CO₃ pH 9.6 for 40 hrs at 4°C. After incubating with either CMV- or control antigen, diluted 1:75 in incubation buffer (10 mM Tris-HCl pH 8.0, 0.3 M NaCl, 1% bovine serum albumin (BSA) and 0.05% Tween 20) each serum was tested in several dilutions with starting from 1:100 in duplicate. After coating, and between incubations (45 min at 20°C), plates were washed five times with 'washing buffer' (10 mM Tris-HCl pH 8.0, 0.15 M NaCl and 0.05% Tween 20) using an automatic washing device (Titertek). Peroxidase labelled conjugate (goat anti-human IgG, Kallestad) was added in a dilution of 1:100 in incubation buffer containing 1% normal mouse serum. After the final wash, the plates were treated with a substrate solution (ortho-phenylene-diamine dihydrochloride (Kodak), 0.2 mg/ml in 50 mM phosphate buffer pH 5.6 containing 0.006% H₂SO₄. Extinctions were measured at 492 nm using a Titertek Multiscan. The antigen-specific extinction was calculated for each serum dilution by subtracting the optical density (OD) of the control antigen from the OD of the CMV-LA. The amount of antibody present in a serum to antigens captured in the assay, was expressed as a percentage of the positive reference serum, as described for anti CMV-LA antibodies (see above). It was established that the apparent molecular weights of the captured antigens were similar to the apparent molecular weights obtained in immunoprecipitation experiments with C10 and C40 monoclonal antibodies.

Statistical analysis

Statistical analysis was performed with the Mann Whitney U test, Chi-square test and Spearman rank correlation where appropriate, and with p<0.05 as the level of significance. All analyses were performed in duplo, with a Pearson correlation coefficient between both results of >0.97.

RESULTS

Anti-CMV-LA IgG and IgM ELISA

IgG class antibodies to CMV-LA were detected in 122 (69%) of the 178 CAH patients. The prevalence of anti-CMV-LA-IgG in IAI-CAH and HBsAg-CAH was higher than in 'other' CAH (Chi-square=23.111, p<0.0001) (table 17.2), and the titres were higher in IAI-CAH and HBsAg-CAH than in 'other' CAH (figure 17.1A).

AETIOLOGY OF CAH	Anti-CMV-LA IgG-class		antibodies of IgM-class		
'Autoimmune' CAH	61/82	(74%)	26/82	(32%)**	
HBsAg-CAH	35/48	(73%)	22/48	(46%)	
'Other' CAH	26/48	(54%)*	21/48	(44%)	
Total	122/178	(69%)	69/178	(39%)	
	0.0001				-

Table 17.2. Prevalence of anti-CMV-LA antibodies in CAH. (n=178)

ad *) Chi-square=23.111, p<0.0001

ad **) Chi-square= 5.992, p<0.02



Figure 17.1. Anti-CMV-LA of A) IgG class and B) IgM class in IAI-CAH (n=82), HBsAgpositive CAH (n=48), and 'other' CAH (n=48).

Age	Observed	Ex based o	pected n reference
		988	987
1-9	0/0	0/0	0/0
10-20	5/13	7/13	2/13
21-25	3/3	2/3	1/3
26-30	5/5	3/5	2/5
31-35	3/5	4/5	2/5
36-40	5/6	5/6	3/6
41-45	1/4	3/4	2/4
46-50	7/7	6/7	3/7
51-55	8/12	10/12	8/12
56-60	10/11	10/11	6/11
61-65	6/7	6/7	4/7
≥66	8/9	9/9	5/9
Total	61/82	65/82 *	38/82 **

Table 17.3.	Prevalence of anti-CMV-LA-IgG A) in 'autoimmune' CAH (n=82)
	compared to anti-CMV antibodies of age-matched healthy blood donors,
	B) in all CAH patients (n=178).

A) 'Autoimmune' CAH (n=82)

Observed versus expected frequences: ad *) Chi-square= 1.187, p=0.2753 ad **) Chi-square=25.944, p<0.0001

B) All CAH patients (n=178).

	Observed	Expected based on reference		
		988	987	
Total1	22/178	131/178 #	66/178 ##	

Observed versus expected frequences: ad #) Chi-square= 2.342, p=0.1218 ad ##) Chi-square=75.515, p<0.0001

IgM-class antibodies to CMV-LA were detected in 69 (39%) of the 178 CAH patients, usually in low titres, as shown in figure 17.1B. There was no difference in titres of anti-CMV-LA-IgM among IAI-CAH, HBsAg-CAH or 'other' CAH, but the prevalence of anti-CMV-LA-IgM was lower in IAI-CAH than in HBsAg-CAH or 'other' CAH (Chi-square=5.992, p<0.02) (table 17.2).

Prevalence of IgG-class anti-CMV-LA among our 178 patients with CAH was higher than expected when compared to the prevalence of anti-CMV antibodies in 1731 healthy German blood donors (Chi-square=75.515, p<0.0001). However, it was similar to prevalence in 1251 Swedish healthy blood donors (Chi-square=2.342, p=0.1218). Also, when IAI-CAH or HBsAg-CAH were considered separately, the observed number of patients with anti-CMV-LA-IgG was higher than expected based on the German study, but not different from expected values when based on the Swedish study of healthy blood donors. When matching our CAH patients for age with the blood donors from both studies, similar results were obtained (table 17.3 shows the results for IAI-CAH).

Anti-LSP in relation to anti-CMV-LA antibodies

Out of the 34 IAI-CAH patients tested for both antibodies, 22 had both anti-LSP and anti-LA-IgG, 2 had none of these antibodies, 8 had anti-LSP but no anti-CMV-LA-IgG, and 2 had anti-CMV-LA-IgG but no anti-LSP. All 4 anti-HBs positive patients of this group had anti-LSP and 3 of them had anti-CMV-LA of the IgG-class (table 17.4). With (Chi-square=0.926, p=0.338) or without (Chi-square=0.78, p=0.381) these anti-HBs positive patients, presence/absence of one or the other antibody did not predict presence/absence of the other (table 17.4), although the group of patients with both antibodies was bigger than the other groups (Chi-square=31.412 or 23.552 respectively, both with p<0.001).

	IAI or anti-HBs		IAI (no anti-HBs)	
	aLA	no aLA	aLA	no aLA
aLSP	22	8	18	7
no aLSP	2	2	2	2
Chi-square= p =	C C	0.926 0.338		0.78 0.381

Table 17.4. Presence/absence of anti-LSP and anti-CMV-LA-IgG in 34 patients with 'autoimmune' CAH did not correlate.

numbers denote number of patients.

All of the anti-LSP positive patients, and the one anti-HBs-positive but anti-LSP negative patient, fulfilled requirements for immunosuppressive therapy. In contrast, the anti-LSP negative and anti-HBs negative IAI-CAH patients did not.

Inhibition experiment

Addition of LA-antigen to an anti-LSP positive, anti-LA IgG positive serum did not inhibit binding of anti-LSP in the anti-LSP RIA. The same finding applied to addition of LA to an anti-LSP positive, but anti-LA IgG negative serum (table 17.5).

sample	aLA	aLSP	added LA-antigen	anti-LSP titre
patient 1	+	+	none	100
serum	+	+	1: 50	100
	+	+	1:100	100
	+	+	1:200	100
patient 2		+	none	200
serum	-	+	1: 50	200
	-	+	1:100	150
	-	+	1:200	200
buffer		-	1: 50	neg
	-		1:100	neg
	а С		1:200	neg
buffer			control antigen 1:50	neg

 Table 17.5.
 No inhibition of anti-LSP binding with LSP by CMV-LA in idiopathic 'autoimmune' CAH.

aLSP denotes anti-LSP antibodies.

aLA denotes anti-CMV antibodies of IgG-class to late antigen.

67-IEA and 58-MA sandwich ELISA

To test for coexpression of two defined CMV-induced antigens on the one hand (immediate early antigen of 67 kD molecular weight, and membrane antigen of 58 kD) and epitopes of LSP on the other hand, we tested 16 sera from patients during untreated IAI-CAH with anti-LSP and/or anti-CMV-LA IgG. Antibodies against 67-IEA were present in all 10 anti-CMV-LA-IgG-positive sera. Anti-58-MA was present in all sera with both anti-LSP, and in three of the four anti-LA IgG positive but anti-LSP negative sera. Anti-67-IEA or anti-58-MA were not present in any of the 6 anti-LSP-positive but anti-CMV-LA-negative sera (table 17.6).

The conclusion from these experiments is that there are no arguments for expression of CMV membrane antigens as an inducer for development of anti-LSP, neither by coexpression of antigens nor by molecular mimicry.

]	Patient	aLSP	aLA-IgG	aLA-IgM	a67-IEA	a58-MA
	1	neg	82	1	212	10
aLA-IgG +	2	neg	69	1	60	45
aLSP -	3	neg	62	1	109	26
	4	neg	55	1	131	neg
	5	250	neg	neg	neg	neg
	6	200	neg	2	neg	neg
aLA-IgG -	7	550	neg	neg	neg	neg
aLSP +	8	150	neg	neg	neg	neg
	9	100	neg	neg	neg	neg
	10	100	neg	neg	neg	neg
	11	50	95	5	65	20
	12	100	300	neg	570	30
aLA-IgG +	13	200	180	1	366	28
aLSP +	14	100	300	9	229	181
	15	150	500	1	985	191
	16	150	145	1	309	115

 Table 17.6.
 Antibodies against LSP and defined CMV antigens in 16 patients with IAI-CAH.

aLSP denotes anti-LSP antibodies.

aLA denotes anti-CMV antibodies to late antigen (IgG or IgM class).

a67-IEA denotes antibodies against CMV immediate early antigen with a molecular weight of 67 kDalton.

a58-MA denotes antibodies against CMV induced membrane antigen with a molecular weight of 58 kDalton.

DISCUSSION

The results from the current study demonstrate that in idiopathic 'autoimmune' chronic active hepatitis (IAI-CAH), induction of autoimmunity, either by coexpression of LSP with CMV membrane antigens or by molecular mimicry between these antigens, is highly unlikely. Anti-LSP did not cross-react with LA, 67-IEA or 58-MA, the main membrane antigens induced by CMV.

Anti-CMV-LA, anti-LSP, or both antibodies were frequently present in IAI-CAH, and in patients with CAH of other aetiology. Prevalence of anti-CMV-LA IgG in IAI-CAH and in HBsAg-CAH is similar to or higher than in healthy blood donors. Titres of IgG-class anti-CMV-LA were higher in IAI-CAH and HBsAg-CAH than in 'other' CAH. An

explanation for this might be that these patients had more active disease. For HBsAg-CAH and anti-HBs-positive IAI-CAH, another possibility is that these patients have a higher risk of becoming a CMV-carrier.

As demonstrated before, antibodies against LSP or its main antigen, the asialoglycoprotein receptor (ASGR) (=hepatic lectin),⁹⁸¹ are usually present in IAI-CAH patients requiring immunosuppressive therapy. Anti-LSP titres have been shown to correlate with the severity of histological damage in untreated patients with IAI-CAH. Changes in titres more or less parallel those in histological inflammation immediately after the institution of immunosuppressive therapy.⁹⁹⁰

Cytotoxic reactions of the ADCC-type are directed at antigenic sites of LSP in CAH and some other liver diseases: CD8+ lymphocytes and OKM+ (natural killer/killer -NK/K-) cells predominated the periportal area in IAI-CAH.⁹⁹⁴ Lymphocytes from patients with a variety of acute and chronic liver diseases are able to kill' heterologous⁹⁹⁴⁻⁹⁹⁹ or autologous¹⁰⁰⁰⁻¹⁰⁰³ hepatocytes, or LSP-coated avian erythrocytes¹⁰⁰⁴ *in vitro*. These cytotoxic reactions are apparently directed at antigenic sites of LSP -cytotoxicity is blocked by addition of LSP¹⁰⁰³-, with lymphocyte cytotoxicity predominantly mediated by a non-T cell population bearing receptors for complement.

In IAI-CAH, a genetically determined defect exists in T-cell inducers of antigen-specific T suppressor-cells^{984,985}. It is possible that these patients are provoked into ADCC against LSP, or more specifically, the ASGR. Corticosteroids can improve the non-antigenspecific T-suppressor cell functions in AI-CAH, resulting in reversal of histological and biochemical manifestations of disease and sufficient to induce 'remission' according to currently accepted criteria. However, corticosteroids have no effect on the genetically determined LSP-specific T-suppressor cell defect.^{983,984} This latter defect could allow for continued stimulation of anti-LSP producing B lymphocytes.

The high titres of IgG-class anti-CMV-LA in untreated IAI-CAH and HBsAg-CAH may be due to a state of polyclonal stimulation of B-lymphocytes as a result of the nonantigen specific T-suppressor cell defects. This is consistent with the literature, indicating that no increased anti-CMV antibody titres are detected in chronic persistent hepatitis.

Results from the present study do not exclude induction of LSP-autoantigens on the hepatocyte membrane and subsequent anti-LSP formation by CMV membrane antigens not tested for. However this is unlikely, since the antigens tested for are almost invariably present after infection with CMV. Therefore, in patients with anti-LSP but not anti-CMV-LA, such a mechanism would be less likely. The same applies to the hypothesis of diminished sequestration of CMV-derived antigens in the diseased liver, with subsequent antibody formation.¹⁰⁰⁵

Recently, persistent measles virus genome was identified in the lymphocytes from 12/18 patients with IAI-CAH, in contrast to 1/45 controls.¹⁰⁰⁶ This finding correlated strongly with the presence of high antibody titres to measles. This finding raises the possibility

that measles virus may be involved in the aetiology and pathogenesis of IAI-CAH. However, there is no direct proof of aetiological involvement, and age-matched comparison with the healthy population may be required to validate these data. Other viruses might also play a role in IAI-CAH. Recently the autoantigen of the severe form of IAI-CAH , the liver-kidney microsomal antigen type 1, was cloned.¹⁰⁰⁷ It appeared to show homology with nucleic acid sequences of the Epstein Barr virus (Manns: personal communication).

From the present study, it follows that a role for CMV in inducing IAI-CAH is unlikely to take place by molecular mimicry or coexpression of epitopes in LSP and in CMV-induced membrane antigens. If CMV indeed plays an aetiological role in (some of) the patients with IAI-CAH, it might be by induction of other cell membrane antigens, for instance by inducing HLA class II antigens, or by encoding a glycoprotein homologous to MHC class-I antigens, as recently reported,¹⁰⁰⁸ making the cell a target for T-lymphocyte cytotoxicity. Further investigation is warranted into a possible aetiological role for CMV by such other mechanisms, and of the role of other environmental factors and their interplay with genetic susceptibility, in the induction of IAI-CAH.

GENERAL DISCUSSION

The aim of the present study was to analyze the progression of a small group of patients with idiopathic 'autoimmune' CAH (IAI-CAH), treated according to the same standardized protocol, against the background of a larger group of patients with CAH of various etiologies and referred in the same period of time. Guidelines were derived for monitoring this disease, with or without therapy.

The present study shows that:

Treatment of IAI-CAH according to a standardized protocol with fixed dose immunosuppressive therapy induces a rapid reduction in inflammation, apparent after two months, resulting in an excellent prognosis. It enables clinicians to limit the number of follow-up visits for patients fulfilling certain criteria. Serial monitoring of histological inflammation in CAH is facilitated by the help of a numerical scoring system for histological activity. An existing scoring system has been modified, and proved effective. From the current work, it also can be derived that histology is not only important for obtaining a diagnosis of CAH, but also for monitoring disease activity thereafter. Simple parameters of activity are required for this follow-up. Among such parameters, serum ALT (using individual reference ranges) and PIIINP appear to be the most useful.

Among circulating autoantibodies in IAI-CAH, at least antibodies to antigens in the 'liver-specific membrane lipoprotein' (LSP) (including the asialoglycoprotein receptor (ASGR)) appears to have pathogenetic importance. Relapse in IAI-CAH is invariably preceded by presence of anti-LSP antibodies, which means that this antibody can also have clinical significance in the guidance of therapy-withdrawal, along with PIIINP and ALT (using individual reference values).

Other autoantibodies have some relative significance in diagnosis of IAI-CAH, although most of these are probably epiphenomena. In autoimmune CAH, the incidence of anti-DNAis very low (unlike SLE) and, if present, is of low avidity. This may enable an immunological distinction from SLE, a disease that seldom co-exists with IAI-CAH. Until now, the clinician treating a patient with IAI-CAH has been most interested in parameters for activity of disease and inflammation.

However, the present work illustrates that, as the result of reduction/disappearance of inflammation, the hepatic protein synthesizing capacity can increase and normalize, even if cirrhosis is present. This applied, not only to more common parameters like serum albumin or cholinesterase, but also to SAA and haemoglobin. The significance is that during therapy of IAI-CAH (and possibly other kinds of liver disease), not only should inflammation or cholestasis be monitored, but also cholinesterase (and e.g. ATIII and albumin).

Furthermore, immunosuppressive therapy resulted in an increase in, and normalization of, the serum sodium concentration, indicating an apparently subclinical derangement of the hepatorenal axis in IAI-CAH which can be reversed with therapy. Until now, the literature has only described worsening of the hepatorenal syndrome during the course of several types of liver disease, only reversible by liver transplantation. This also applies to hepatic protein synthesizing capacity: although scattered data indicated restoration of serum albumin during therapy of CAH, the present study clearly demonstrates that normalization can occur despite the presence of cirrhosis, and that continuation of therapy is usually necessary beyond the point of reduction or disappearance of inflammation. The hallmark of therapy has been the suppression of inflammation. Although prednisolone and azathioprine can have considerable side effects, these were not severe or frequent. Some studies reported that azathioprine might enhance the formation of fibrosis and could induce cytopenia, but our data does not support these fears.

Future research

Most patients with IAI-CAH need immunosuppressive treatment for more than our standard period of two years. Further investigation is warranted into simple parameters to aid early identification or prediction of CAH relapse, and into factors that play a role in relapse and therapy-failure. The proposed role for especially PIIINP, ALT, CHE, and anti-LSP in monitoring IAI-CAH should be confirmed in other studies. Further study is justified in the field of hepatic protein synthesis and in water- and salt-homeostasis in CAH, and other diseases with a component of hepatitis. Research into the pathogenesis and etiology of IAI-CAH should be extended, to design more specific therapies.

Model

The cause of 'idiopathic autoimmune' CAH remains unknown. We closely examined hepatitis B and the cytomegalovirus. Patients with IAI-CAH who previously lost the hepatitis B virus and are anti-HBs positive can have the same favorable response to immunosuppressive therapy as other IAI-CAH patients. Induction of autoimmune CAH in these patients might be caused by persistence of LSP on the hepatocytes, induced by the previous HBV-infection, with antibody-formation and antibody-dependant cellular cytotoxicity. With regard to the cytomegalovirus (CMV), no relationship could be detected between presence of the virus and autoimmune CAH. Molecular mimicry between CMV-LA antigens and LSP-antigens was excluded. However, neither for HBV nor for CMV has an etiological relationship with autoimmune CAH been excluded in patients with circulating antibodies to these viruses. Although it is well known that HBV can induce formation of anti-LSP (which can persist after the disappearance of

IAI-CAH patients with antibodies to both CMV membrane antigens and LSP-antigens, expression of LSP with subsequent antibody formation may have been induced by CMV-infection of hepatocytes. In this thesis, the model which assumes genetic factors cooperating with environmental factors is supported. These environmental factors can be of viral, bacterial, chemical or other origin. Further research is warranted into viral, bacterial and other possible etiological factors. The inflammation which progressively results from the host/environment interaction has been followed by measurements relatively early (like anti-LSP) or late (serum albumin, ALT) in this progression. Inbetween lies the acute-phase reaction (CRP, SAA) and fibrogenesis (PIIINP). These parameters can be useful in monitoring the disease. The ideal parameter for monitoring reflects both the present and the past status of inflammation and hepatic functional reserve. By combining several of the mentioned parameters, like PIIINP and CHE, almost ideal monitoring is feasable. The behaviour of parameters and their application in monitoring of IAI-CAH require further investigation, especially the early indicators such as interleukins and circulating interleukin-receptors.

SUMMARY

This thesis describes the outcome -survival- of a large group of 186 consecutive patients with chronic active hepatitis of various etiologies, and describes in detail the progress of 21 patients from this group with 'autoimmune' chronic active hepatitis maintained on standardized immunosuppressive therapy.

Idiopathic 'autoimmune' chronic active hepatitis (IAI-CAH) is generally considered as a primarily hepatocytic liver disease of unknown cause, characterized by (episodes of) periportal and portal inflammation and piecemeal necrosis, with the potential to progress to cirrhosis or associated with cirrhosis. It lasts 6 months or longer.

The introduction briefly reviews the current knowledge on IAI-CAH. This is put into a historical perspective, definitions are summarized, clinical symptoms and differential diagnoses are mentioned, data on dismal prognosis in untreated disease and improved prognosis in treated IAI-CAH are reviewed. Treatment options are summarized, problems in the diagnosis and management of the disease are mentioned, and an overview regarding parameters of inflammation and liver function useful in monitoring IAI-CAH is provided. Furthermore, the present knowledge on pathogenesis and possible aetiologies of the disease is put in an immunological context. Untreated IAI-CAH has a high morbidity and mortality. Early mortality is typically due to liver failure, while late mortality usually results from the complications related to cirrhosis. Immunosuppressive therapy can reduce inflammation and enhance survival. Formation of cirrhosis can continue despite induction of 'remission' from CAH. Clinical, histological and biochemical findings must all be considered in concert for decisions concerning therapy. Data are scarce on early changes in inflammation and liver function during immunosuppressive therapy. No optimal tests for the follow-up of IAI-CAH are available. Histology forms the basis for the diagnosis of CAH, but may be insufficient and impractical for follow-up once therapy has been started. Standard laboratory bloodtests do not accurately reflect morphology. Until now, limited information was available regarding changes in hepatic functional capacity during immunosuppressive therapy of IAI-CAH. This warranted a search into parameters of activity and hepatic functional capacity in IAI-CAH.

Section 1 deals with patients and methods.

Chapter 1 provides data on the patients included in these studies. Chapter 2 refers to the methods used in the studies.

In Chapter 3, we compared numerical histological scoring of activity with conventional pathological descriptions of treated and untreated CAH. The proposed histological activity score (HAS) correlated very well with conventional scores. In contrast to the original method of scoring, we propose to leave fibrosis/cirrhosis out of this score of

activity. Maximum HAS is 18. HAS \leq 1 denotes the (virtual) absence of inflammatory infiltrate. A HAS of 2 or 3 usually denotes a histology similar to chronic persistent hepatitis (CPH); 90% of biopsies with a histology similar to CPH when scored conventionally, and 36% of biopsies with minimal CAH (CAHmin.), showed a HAS \leq 3. We therefore termed 1<HAS \leq 3 "partial histological remission" and propose to reserve the term "complete histological remission" for HAS \leq 1. We conclude that HAS accurately reflected histological activity of CAH, and is more accurate than conventional descriptions for serial measurement of morphological activity of disease. Furthermore, it enables comparison with other parameters.

In Chapter 4, three radioimmunoassays for the determination of the serum N-terminal propeptide of collagen type III were compared in chronic active hepatitis: a new rapid equilibrium type of assay based on the human propeptide, developed by Risteli and Risteli, was compared with the PIIINP RIA-gnost® (Behring), and the 'PIIINP Fab assay®'(Behring), both based on a reference bovine PIIINP antigen (col 1-3) and antibodies raised in rabbits. The correlation between the human PIIINP RIA and the PIIINP RIA-gnost® was excellent, with less correlation with the 'PIIINP Fab assay®'. The new human assay is therefore as accurate as the standard PIIINP RIA-gnost® used thus far, but it is not sensitive to smaller degradation products. Therefore, unlike in the PIIINP RIA-gnost® (Behring), no serial dilutions of the sera and no 50% intercept method for calculating the results are necessary in this new PIIINP assay, since the standards and the serum samples give parallel inhibition curves. This solves the most important problems that have been inherent in the determination of PIIINP, and provides a rapid reliable test. We used this tests in studies on PIIINP that will follow.

In Chapter 5, the establishment of a modified radioimmunoassay for the determination of autoantibodies against 'liver-specific membrane lipoprotein (LSP)' is described. This modification of the original procedure yielded a rapid, reproducible and quantitative assay for determination of anti-LSP antibodies.

In section 2 Chapter 6, we performed an analysis of survival of all 186 consecutive patients with CAH of various aetiologies included in the current study. $85(\pm 3)$ percent of all 186 patients with CAH survived 5 years, $70(\pm 4)$ percent survived 10 years, $55\%(\pm 5)$ survived 15 years, and $40\%(\pm 10)$ survived 20 years after diagnosis. Ages between therapy groups and aetiological groups did not differ significantly. Cox's proportional hazards estimation identified increasing age, presence of antiHBs, and no therapy (patients receiving standard immunosuppression had the best prognosis) as independant risk factors. Sex, presence or absence of antinuclear antibodies, antismooth muscle antibodies, level of serum alanine aminotransferase or serum cholinesterase did not influence survival. Most causes of deaths registered in patients with CAH were not directly related to the liver disease. Remarkably, there was a high incidence of plasmocytoma.

In Section 3, several parameters of inflammation and liver function, their relations, and the changes during immunosuppressive therapy are evaluated in 21 patients with 'autoimmune' CAH.

In Chapter 7, routine liver biochemistry in IAI-CAH before and during standardized immunosuppression was serially analyzed. Serum alanine aminotransferase (ALT), aspartate aminotransferase (AST) and gammaglobulin (GG) were elevated at the start and improved considerably to normal median values with therapy. However, ALT and AST remained elevated between one and three times the upper reference limit in about half of the patients. Conjugated bilirubin was elevated, then normalized within two months to normal values in all patients. Median unconjugated bilirubin was normal at the start, nevertheless values declined within the first two months of therapy.

Chapter 8 describes histology and its changes during immunosuppressive therapy (0, 2, 14 and 26 months). The high median HAS declined considerably within the first two months of therapy, with further improvement during the next two years of therapy. At two months 67 percent of the patients had already improved to a HAS \leq 3. The fixed-dose therapy schedule was able to improve histology in 95% of the patients. A HAS \leq 1 ('complete histological remission') was achieved in 80 percent of the patients and 1<HAS \leq 3 ('partial histological remission') in 10 percent of the patients within two years of treatment. Periportal and lobular inflammation had disappeared in all of these patients in remission. Continuing decrease in portal and periportal inflammatory activity, with disappearance of plasma cells and decrease in numbers of lymphocytes, was observed during the two years of immunosuppressive therapy. Usually, after two years only lymphocytes remained. HFS was high (median score 3) and did not change during therapy.

Chapter 9 shows that hepatic protein synthesizing capacity, impaired in the majority of the 21 AI-CAH patients, normalized in the course of two years in all patients under investigation, despite the presence of cirrhosis in all but one patient. This was reflected in increasing and normalizing serum albumin (ALB), (pseudo-)cholinesterase (CHE), plasma antithrombin III (AT III), and decreasing and normalizing activated partial thromboplastin time (APTT), usually already in the first 14 months of therapy. The median prothrombin time (PT) was slightly elevated but did not change during therapy, although there was a trend to a transient further increase into the abnormal range within the first four weeks of therapy. CHE, unlike ALB and AT III was still increasing within the second year of therapy. Hence, CHE may be the most sensitive of the parameters of hepatic protein synthesizing capacity under investigation.

Chapter 10 shows that the acute-phase response is depressed in CAH: serum values of C-reactive protein (CRP) are only slightly elevated during this inflammation of its producing organ, and these levels still correlated to periportal inflammation scores. In

contrast to CRP, serum amyloid precursor A protein (SAA) levels are normal in CAH, and slightly increase during therapy. SAA values are related to CHE: its levels reflect hepatic protein synthesizing capacity and not inflammation in CAH.

Chapter 11 describes that the serum N-terminal propeptide of collagen type III (PIIINP) is elevated in 'autoimmune' CAH. PIIINP values rapidly decrease in the first two months of therapy. PIIINP levels finally normalize, or nearly normalize, but this can take 14 months of therapy despite a more rapid improvement in histology. This delay in normalization, and the incompleteness of normalization in half of the patients, may clarify why cirrhosis can develop during immunosuppressive therapy. PIIINP is predominantly correlated to histological inflammation and to ALT, but its levels also show a weak inverse correlation to CHE, probably reflecting reduced degradation of PIIINP in the liver during severely impaired liver function.

In Chapter 12, serum ALT and PIIINP were studied in 16 CAH relapses occurring during therapy withdrawal in 9 patients. During remission, ALT was below twice the upper reference limit, and PIIINP was below the reference limit in all cases. Using two or three times the upper reference limit of ALT as 'cut-off' for diagnosis of CAH-relapse, as recommended in the literature, ALT only rose above this limit median one week before the peak-ALT. An increase of PIIINP above the reference limit occurred weeks to months before the peak-ALT was reached. Using ALT, it was only possible to detect relapse as early as with PIIINP if each patient served as his/her own control for ALT, with the (arbitrarily chosen) upper reference limit at 1.25 times the ALT value obtained during histological remission (HAS \leq 3 and preferably HAS \leq 1).

Section 4 describes some immunological abnormalities in chronic active hepatitis, and their clinical relevance. The results in Chapter 13 show that autoantibodies to 'liver-specific membrane lipoprotein (LSP) were invariably present in patients with 'autoimmune' CAH requiring standardized immunosuppressive therapy. Titres declined in all patients during such therapy. In the course of two years anti-LSP became undetectable in half of the patients, while it remained detectable in the other half. As shown by others, presence of anti-LSP probably indicates ongoing stimulation of B-lymphocytes as the result of a defect in inducers of LSP-specific T-suppressor lymphocytes.

In Chapter 14, serum anti-LSP antibodies in 12 CAH relapses occurring during therapy withdrawal in 9 patients were studied. At the start of therapy withdrawal, all patients were in histological remission, and in only one case anti-LSP was detectable. Weeks to months before the peak-ALT was reached, anti-LSP became detectable where previously undetectable (with one exception). A change from undetectable to detectable anti-LSP preceded relapse, and it preceded its detection by the conventional serological definition of relapse (ALT>3x the upper reference limit). It also preceded "non-

remission" based on ALT exceeding two times the upper reference limit. This supports the theory that in IAI-CAH anti-LSP should be undetectable before considering to withdraw immunosuppressive therapy.

Chapter 15 describes that autoantibodies to a soluble liver antigen (SLA) are present in about 10 percent of our population of patients with idiopathic 'autoimmune' CAH. No patients with anti liver-kidney-microsomal antibodies type 1 (anti-LKM1) -denoting a more severe type of IAI-CAH- were detected in this population. Therefore, our patients with IAI could be divided into three of the recently proposed four subtypes of IAI-CAH with the help of ANA, SMA and SLA. Survival in these three groups did not differ, and presence or absence of the antibodies mentioned were not independant prognostic factors. This questions the clinical significance of this classification of IAI-CAH in four subtypes. However, the groups may have different aetiologies of IAI-CAH.

Chapter 16 shows that anti-cardiolipin antibodies (ACA), as detected by ELISA, were not infrequent in CAH and 'CAH-related disorders' of various aetiologies, with the highest prevalence and titres in IAI-CAH. In contrast, anti-DNA antibodies, detected by immunofluorescence with the kinetoplast of Crithidia Luciliae as a substrate, were absent in 98 percent of CAH patients, allowing the possibility of an immunological distinction from SLE. The prevalence of anti-DNA measured by ELISA was higher than with the immunofluorescence mentioned, especially for anti-DNA antibodies of the IgM class. This difference may be due to a difference in avidity of the anti-DNA antibodies measured using both methods. From the results, it also followed that cross-reactivity between ACA and anti-DNA was limited in CAH and, if present, probably only applies to anti-DNA of low avidity.

Chapter 17 shows the high incidence and titres of IgG-class antibodies to cytomegalovirus-induced late antigens (CMV-LA) in IAI-CAH and HBsAg-positive CAH. Frequently, anti-CMV-LA and anti-LSP were both present in IAI-CAH. However, there was no significant correlation between presence of these antibodies. Binding of anti-LSP in the radioimmunoassay for anti-LSP was not inhibited by addition of CMV-LA. Antibodies to the defined CMV-induced antigens 'immediate early antigen (IEA)' of 67 kD and to 'membrane antigen (MA)' of 58 kD molecular weight were only positive in patients with anti-CMV-LA IgG, not in patients with anti-LSP but without anti-CMV-LA IgG. From these results, it can be deduced that an aetiological role of CMV in IAI-CAH by molecular mimicry between CMV-induced membrane antigens and LSP is highly unlikely. This does not exclude an aetiological role for CMV by means of other pathways.

SAMENVATTING

Chronisch actieve hepatitis (CAH) is een chronische ontsteking van de lever -6 maanden of langer-, waarbij met name de zogenaamde portale driehoekjes en omringende gebieden door ontstekingscellen 'aangevreten' worden ('piecemeal necrosis'). De ziekte kan snel ontstaan en in korte tijd fataal zijn, maar meestal is het een ontsteking die aktieve en minder aktieve perioden kent. Bij een aantal vormen van CAH is er een af weerreactie van het lichaam tegen de eigen lever gericht. Een dergelijke autoimmuunreactie kan ontstaan op basis van bekende (als hepatitis B virus) maar ook onbekende ('idiopathische') oorzaken. In het laatste geval spreken we van idiopathische CAH, welke veelal autoimmune kenmerken heeft (idiopatische 'autoimmune' CAH, IAI-CAH).

In dit proefschrift wordt een groep van 186 patiënten beschreven met, onder andere door leverbiopsie bewezen, CAH met verschillende onderliggende oorzaken. De langetermijn overleving van deze patienten, en factoren die daarbij een rol spelen, werden geanalyseerd. Uit deze groep wordt een kleine groep van 21 patiënten met 'autoimmune' CAH in detail beschreven; deze groep werd op gestandaardiseerde wijze behandeld met medicatie die immuunreacties onderdrukt (immunosuppressie).

In de inleiding wordt een kort overzicht van de huidige kennis omtrent IAI-CAH gegeven. De geschiedenis van onze kennis van deze aandoening, definities, klinische verschijnselen, uit te sluiten andere ziekten welke op IAI-CAH kunnen lijken, kennis omtrent slechte prognose zonder behandeling en verbetering van overlevingskansen met behandeling van IAI-CAH worden behandeld, mogelijke behandelingen samengevat, en een overzicht wordt verschaft van diverse parameters om ontsteking en leverfunctie te meten in IAI-CAH (van belang voor 'monitoring' van zowel ziekte als resultaat van behandeling). Voorts wordt de huidige kennis van pathogenese en mogelijke inducerende factoren die leiden tot IAI-CAH samengevat in immunologische context. Onbehandelde IAI-CAH leidt veelal tot morbiditeit en sterfte. Een vroeg overlijden is in het algemeen het gevolg van een falen van de lever om de normale functies uit te oefenen, terwijl late sterfte veelal het gevolg is van complicaties verbonden met het ontstaan van levercirrose door CAH. Immuunsuppressieve behandeling kan de chronische leverontsteking doen afnemen, en kan de overlevingskans aanmerkelijk verbeteren in IAI-CAH. Door oorzaken die tot nu toe onbekend waren kan het ontstaan van cirrose echter voortgaan, ondanks het induceren van 'CAH in remissie' met een dergelijke therapie. Klinische (klachten, symptomen), biochemische (bloedonderzoek), en histologische (leverbiopsie) gegevens moeten in combinatie beoordeeld worden om beslissingen omtrent therapie te kunnen nemen. Gegevens omtrent vroege veranderingen in ontsteking en leverfunctie na het starten van immunosuppressieve behandeling zijn schaars. Voor het vervolgen van deze patiënten zijn geen optimale testen beschikbaar. Een leverbiopsie vormt de basis voor de diagnose CAH, maar is onvoldoende en onpraktisch voor het vervolgen van een patiënt nadat therapie gestart is. De absolute uitkomsten van routine bloedonderzoek vormen geen nauwkeurige weerspiegeling van de morfologie van de lever. Tot nu

toe was slechts weinig bekend van veranderingen in functionele capaciteit van de lever onder invloed van immunosuppressieve therapie in IAI-CAH. Daarom was het gewenst parameters van ontstekingsactiviteit en leverfunctie in IAI-CAH te onderzoeken.

Sectie 1 behandelt patiënten en methoden.

Hoofdstuk 1 beschrijft de patiënten die in de diverse studies zijn opgenomen. Hoofdstuk 2 refereert aan de methoden die in de diverse hoofdstukken zijn toegepast. In Hoofdstuk 3 vergeleken we een scoringsmethode waarbij de ernst van de ontsteking en van de verbindweefseling/cirrose in de lever in getallen wordt uitgedrukt, met de klassieke pathologische beschrijvingen, zowel in behandelde als onbehandelde CAH. De eerste, numerieke, methode leverde voor wat betreft de ontsteking een histologische activiteitsscore (HAS) op van nul tot achttien. In tegenstelling tot de originele methode scoorden wij fibrose/cirrose apart. De HAS correleerde zeer goed met de conventionele scoringsmethode. HAS≤1 betekent (vrijwel) afwezigheid van ontstekingsinfiltraat. Een HAS van 2 of 3 houdt meestal een histologie als van chronisch persisterende hepatitis (CPH) in. 90% van de biopten met een histologie als van CPH en 36% van de biopten met 'minimal CAH (CAHmin.)' volgens de oude methode, had een HAS≤3. We stellen daarom voor 1<HAS<3 'partiële histologische remissie' te noemen, en HAS<1 met 'complete histologische remissie' aan te duiden. De conclusie luidt dat HAS de histologische activiteit van CAH goed weerspiegelt, op een meer precieze wijze dan met klassieke scoringsmethoden mogelijk is, en dat de HAS vergelijking van opeenvolgende leverbiopten vergemakkelijkt. Bovendien maakt het een statistisch verantwoorde vergelijking met andere parameters (bijvoorbeeld bloedonderzoek) mogelijk.

In Hoofdstuk 4 worden drie radioimmunoassays vergeleken waarmee het N-terminale propeptide van collageen type III (PIIINP) gemeten werd bij patiënten met CAH. Bij het 'afzetten' van bindweefsel in de lever komt een klein stukje van het 'procollageen III' in het bloed vrij, en er zijn aanwijzingen dat dit PIIINP voornamelijk de depositie van collageen type III in de lever weergeeft bij patiënten met CAH. Aangezien verbindweefseling en cirrosevorming in de lever complicaties van CAH zijn lijkt een dergelijke test van belang voor dynamische 'monitoring' van depositie van collageen III in de lever bij CAH. Een van de drie tests, de zogenaamde PIIINP RIA-gnost® (Behring), is de standaardtest. Helaas is deze test omslachtig en een berekening is vereist om te compenseren voor het feit dat kleinere degradatieprodukten ook gemeten worden (waardoor standaarden en te testen sera geen parallelle inhibitiecurves geven). Een nieuwe 'equilibrium-type' test is gebaseerd op het humane propeptide waardoor dit laatste probleem vervalt. De correlatie van resultaten van deze nieuwe 'humane PIIINP test' met genoemde standaardtest is uitstekend, terwijl de uitkomsten in de 'PIIINP Fab assay' slechts matig met beide andere tests correleerde. De humane PIIINP test geeft dus resultaten die vergelijkbaar zijn met de standaard PIIINP RIA-gnost®, maar heeft een aantal nadelen van de standaardtest niet. De humane PIIINP test werd dan ook in de latere studies gebruikt.

In Hoofdstuk 5 wordt onze modificatie van een radioimmunoassay voor het bepalen van autoantistoffen gericht tegen 'lever-specifiek membraan lipoproteine (LSP)' beschreven. Zowel labellingsmethode als de assay zelf werden gewijzigd. Dit leverde een snelle, reproduceerbare en kwantitatieve testmethode op voor de meting van anti-LSP.

In sectie 2, Hoofdstuk 6 verrichtten we een survival-analyse van alle 186 patiënten met CAH die in de huidige studie waren begrepen. Met behulp van de Kaplan-Meier product-limit methode werd de overleving van patiënten met CAH berekend voor alle patiënten, en uitgesplitst naar etiologie en therapie. Van de gehele groep van 186 patiënten was de 5-jaars overlevingskans na stellen van de diagnose 85(±3)%, terwijl deze kans voor 10 jaar $70(\pm 4)\%$ was, voor 15 jaar $55(\pm 5)\%$, en voor 20 jaar $40(\pm 10)\%$. De leeftijd van de patiënten in de verschillende etiologische- en therapie-groepen verschilde niet. Met behulp van de Cox's 'proportional hazards estimation' identificeerden wij hogere leeftijd, aanwezigheid van anti-HBs antistoffen in het bloed, het niet (kunnen) geven van standaard immunosuppressieve therapie en verhoogd serum alanine aminotransferase (ALT) als onafhankelijke risicofactoren voor overlijden, terwijl geslacht, aanwezigheid van antistoffen tegen kernantigenen (ANA) of gericht tegen gladde-spier antigenen (SMA), en serum cholinesterase (CHE) de kans om te overleven niet beïnvloedden. In de meeste gevallen waarin de oorzak van overlijden achterhaald kon worden, stond deze niet in direkt verband met de leverziekte. Wel was het opvallend dat meer patiënten dan verwacht de ziekte van Kahler (plasmocytoom) kregen.

In sectie 3 worden diverse parameters van ontsteking en leverfunctie apart en in relatie tot elkaar bestudeerd, en worden de veranderingen in deze parameters onder invloed van gestandaardiseerde immunosuppressieve behandeling geëvalueerd bij 21 patiënten met 'autoimmune' CAH.

In Hoof dstuk 7 wordt de routine 'lever-biochemie' in het bloed voor en tijdens deze behandeling bij deze 21 patiënten serieel vervolgd. Serum alanine aminotransferase (ALT), aspartaat aminotransferase (AST) en gamma globuline (GG) in onbehandelde IAI-CAH waren hoger dan de referentiewaarden, en deze parameters verbeterden aanzienlijk -met medianen binnen het referentiegebied- onder invloed van therapie, hoewel ALT en AST in ongeveer de helft van de patiënten tussen één en drie maal de bovenste grens van het referentiegebied bleven. Geconjugeerd bilirubine was verhoogd, maar normaliseerde in alle patiënten binnen twee maanden behandeling. Het mediane ongeconjugeerde bilirubine was normaal bij aanvang van therapie, maar de waarden daalden binnen het referentiegebied in de eerste twee maanden van behandeling.

Hoofdstuk 8 beschrijft de histologie voor en tijdens behandeling (0, 2, 14 en 26 maanden). De hoge mediane HAS (ontsteking in de lever, zie boven) aan het begin van de behandeling nam aanzienlijk af in de eerste twee maanden van behandelen, met verdere verbetering in de daaropvolgende twee jaar behandeling. Na twee maanden behandelen had tweederde van de patiënten al een HAS \leq 3. Uiteindelijk was het gebruikte behande-

lingsschema, met standaarddoses prednisolon en azathioprine, in staat de ontsteking in de leverbiopsie bij 95% van de patiënten terug te dringen. Hierbij werd in 80% van de patiënten een HAS \leq 1 ('complete histologische remissie') bereikt, en bij 10% een 'partië-le histologische remissie' (1<HAS \leq 3) binnen twee jaar behandeling, waarbij periportale en lobulaire ontsteking in alle patiënten bij wie histologische remissie bereikt werd verdwenen was. Gedurende de twee jaar immuunsuppressieve medicatie werd een voortgaande afname van portale en periportale ontsteking (driehoekjes en daaromheen) in de leverbiopsie gezien, waarbij, in het algemeen, na twee jaar slechts lymfocyten resteerden. De histologische fibrose score (HFS) was hoog (mediaan 3) en veranderde niet tijdens therapie.

De gegevens in Hoofdstuk 9 tonen aan dat de capaciteit voor het aanmaken van nieuwe eiwitten, die te laag is bij de 21 patiënten met onbehandelde IAI-CAH, normaliseert in de loop van twee jaar immunosuppressie bij al deze patiënten, ondanks de aanwezigheid van cirrose bij 20 van de 21 patiënten (mede met laparoscopie vastgesteld). Dit bleek uit een toename in bloedspiegels van albumine (ALB), (pseudo-) cholinesterase (CHE), antithrombine III (ATIII), en een afname en normaliseren van de partiëel geactiveerde thromboplastine tijd (APTT), meestal reeds binnen de eerste 14 maanden van de behandeling. De mediane prothrombinetijd (PT) was licht verhoogd voor behandeling, maar veranderde niet tijdens therapie, hoewel er een trend tot passagière verslechtering was in de eerste vier weken van de behandeling. Aangezien CHE bleef stijgen in het tweede jaar therapie, terwijl dat niet gold voor ALB en ATIII, is CHE mogelijk de meest sensitieve van de onderzochte parameters voor capaciteit van eiwitsynthese door de lever.

In Hoofdstuk 10 blijkt dat de zogenaamde 'acute fase respons' verlaagd is bij patiënten met CAH: in het serum van patiënten met CAH zijn de waarden van het C-reactieve eiwit (CRP) slechts licht verhoogd tijdens deze ontsteking van het belangrijkste CRP-producerende orgaan. De serum CRP-waarden zijn wel gecorreleerd met periportale ontsteking in het leverbiopt. In tegenstelling tot CRP, zijn de serumspiegels van een ander 'acute fase eiwit', het 'serum amyloid precursor A eiwit' (SAA), normaal in CAH, en de SAA-spiegels nemen in geringe mate toe tijdens immunosuppressie. De SAA-waarden blijken met CHE (eiwitsynthese) gecorreleerd te zijn en niet met histologische parameters van ontsteking.

Hoofdstuk 11 beschrijft dat de serumwaarden van het N-terminale propeptide van collageen type III (PIIINP) abnormaal hoog zijn bij patiënten met IAI-CAH. PIIINP-spiegels worden normaal of bijna normaal onder invloed van gestandaardiseerde immunosuppressie, hoewel dit 14 maanden kan duren ondanks het feit dat de histologie al eerder verbeterd is. De traagheid waarmee de verbetering plaatsvindt -na een aanvankelijk snelle verbetering in de eerste twee maanden-, en het feit dat PIIINP niet geheel normaliseert in de helft van de patiënten, zijn mogelijk de verklaring voor het feit dat cirrose kan ontstaan tijdens immunosuppressieve therapie. Serum PIIINP spiegels zijn voornamelijk gecorreleerd met ontsteking in de leverbiopsie en met alanine aminotransferase (ALT), hoewel PIIINP ook een zwakke omgekeerde correlatie met CHE vertoont, hetgeen waarschijnlijk een afname van afbraak van PIIINP in de lever tijdens ernstig gestoorde leverfunctie weergeeft.

In Hoofdstuk 12 werden 16 situaties (bij 9 patiënten) waarbij IAI-CAH terugkeerde ('relapse') na verminderen van immuunsuppressie, terwijl tevoren de CAH 'in remissie' was, bestudeerd. Tijdens remissie was de ALT onder twee maal de bovenste grens van het referentiegebied, terwijl PIIINP in alle gevallen binnen het referentiegebied lag. Als twee of drie maal de bovengrens van het referentiegebied als grens werd genomen, waarboven van 'relapse' gesproken werd -zoals in de literatuur aangegeven-, dan kwam ALT pas boven deze grens vlak (een week) voor het maximum van de maximale ALT tijdens 'relapse'. Een stijging van PIIINP boven referentiewaarden echter, trad weken tot maanden voor de maximale ALT bij 'relapse' op. Als ALT gebruikt werd als indicator was een even vroege detectie van relapse mogelijk als met PIIINP indien iedere patiënt diende als zijn/haar eigen controle voor ALT waarden, en we tevens de (arbitrair gekozen) bovenste referentie-limiet op 1.25 maal de ALT-waarde tijdens histologische remissie (HAS≤3 en liefst HAS≤1) stelden.

Sectie 4 beschrijft enkele immunologische afwijkingen bij chronisch actieve hepatitis, en hun klinische relevantie.

In Hoofdstuk 13 laten we zien dat autoantistoffen tegen het zogenaamde 'lever-specifieke membraan lipoproteïne (LSP)' steeds aanwezig waren bij patiënten met 'autoimmuun' CAH die voldeden aan de criteria voor behandeling met gestandaardiseerde immunosuppressieve therapie. De titers van anti-LSP namen af tijdens een dergelijke behandeling, en in de loop van twee jaar behandelen werd anti-LSP in de helft van de patiënten 'niet detecteerbaar'. Zoals anderen aantoonden, geeft de aanwezigheid van anti-LSP waarschijnlijk een voortgaande stimulatie van B-lymfocyten weer die het gevolg is van een defect in de 'inducers' van LSP-specifieke 'T-suppressor' cellen, met andere woorden: LSP geeft waarschijnlijk de 'immunologische activiteit' van de ziekte weer.

In Hoofdstuk 14 worden anti-LSP antistoffen bestudeerd in 9 patiënten die tijdens behandeling in histologische remissie waren tijdens 12 situaties waarin CAH terugkeerde na vermindering van immunosuppressieve medicatie. De periode van begin van vermindering van medicatie tot maximale ALT tijdens 'relapse' werd bestudeerd. Aan het begin van de vermindering van therapie was anti-LSP in slecht één van de 12 gevallen aantoonbaar, terwijl dit tijdens de piek-ALT bij 'relapse' in 11 van de 12 gevallen (92%) het geval was. Weken tot maanden voor de piek-ALT was anti-LSP al te detecteren in de 11 gevallen waarin anti-LSP eerst niet aantoonbaar was. Een verandering van niet detecteerbaar zijn van anti-LSP naar detecteerbaar anti-LSP ging vooraf aan 'relapse' van CAH, en ging vooraf aan de diagnose daarvan op basis van huidige serologische definities van relapse (ALT boven drie maal de hoogste referentiewaarde), en ging ook vooraf aan het niet in remissie zijn gebaseerd op een ALT boven twee maal de hoogste referentiewaarde.

De gegevens in Hoofdstuk 15 laten zien dat autoantistoffen tegen een oplosbaar (soluble) lever antigen (SLA) aanwezig waren bij 10% van de geteste patiënten met IAI-CAH. Wij vonden géén anti lever-nier-microsomale antistoffen van type 1 (anti-LKM1) -geassocieerd met een ernstiger vorm van IAI-CAH- in deze groep. Derhalve konden onze IAI-CAH patiënten ingedeeld worden in drie van de vier recentelijk voorgestelde subtypes van IAI-CAH met behulp van ANA, SMA en SLA. Het bleek echter dat de overlevingskansen tussen deze groepen patiënten niet verschilde, en dat de aan- of afwezigheid van genoemde autoantistoffen geen onafhankelijke prognostische factor is. Het is dus de vraag of de recent voorgestelde classificatie van IAI-CAH zinvol is voor de kliniek. Het zou echter zo kunnen zijn dat de etiologie in deze groepen verschillend is.

Hoofdstuk 16 laat zien dat aanwezigheid van anti-cardiolipine antistoffen (ACA), bepaald met behulp van ELISA, geen zeldzaamheid is in CAH en 'CAH-related disorders' met verschillende etiologieën. De hoogste titers van ACA kwamen voor bij IAI-CAH. Daarentegen kwamen anti-DNA antistoffen, bepaald door middel van immunof luorescentie met de kinetoplast van Crithidia Luciliae als substraat, niet voor bij 98% van de patiënten met CAH. Dit zou gebruikt kunnen worden als hulp bij de differentiaal diagnose tussen CAH en systemische lupus erythematosus (SLE) in de zeldzame gevallen waar het onderscheid moeilijk is. De prevalentie van anti-DNA gemeten met een ELISA was hoger dan met voorgenoemde immunofluorescentie methode, vooral wat betreft anti-DNA van de IgM-klasse. Dit verschil kan het gevolg zijn van het feit dat deze twee tests anti-DNA en ACA slechts in beperkte mate voorkomen in CAH, en, indien aanwezig, waarschijnlijk vooral anti-DNA van lage aviditeit betreffen.

In Hoofdstuk 17 wordt aangetoond dat er met name in IAI-CAH en HBsAg-CAH een hoge incidentie van door cytomegalovirus geïnduceerde late antigenen (CMV-LA) is. In IAI-CAH kwam zowel anti-CMV-LA als anti-LSP veelvuldig voor, maar het vóórkomen van deze antistoffen was niet gecorreleerd. Binding van anti-LSP in de RIA voor anti-LSP werd niet geremd door toevoeging van CMV-LA. Antistoffen tegen de goed gedefinieerde, door CMV geïnduceerde, antigenen 'immediate early antigen (IEA)', met een molecuulgewicht van 67 kiloDalton (kD), en 'membrane antigen (MA)' met een molecuulgewicht van 58 kD werden slechts gevonden bij die patiënten met IAI-CAH die anti-CMV-LA-IgG hadden. Bij patiënten met anti-LSP, maar zonder anti-CMV-LA-IgG, werden deze antistoffen niet gevonden. Hieruit volgt dat CMV geen etiologische rol speelt in IAI-CAH via 'molecular mimicry' (het op elkaar lijken) van door CMV geïnduceerde antigenen in LSP. Dit sluit echter een oorzakelijke rol voor CMV via andere mechanismen niet uit.

REFERENCES

- 1. Terrasse J, Moinade S, Rigal D. Introduction a la thérapeutique par les antimétaboliques des cirrhoses hépatiques. *Presse Méd* 1967; 75: 653-654.
- 2. Cullinan ER. Idiopathic jaundice (often recurrent) associated with subacute necrosis of the liver. St Barth Hosp Rep 1936; 69: 55-142.
- 3. Amberg S. Hyperproteinemia associated with severe liver damage. Proc Staff Meet Mayo Clin 1942; 17: 360-362.
- 4. Waldenström J. Leber, Blutproteine und Nahrungseiweiss. Dtsch Gesellsch Z Verdau Stoffwechselkr 1950; 15: 113-121.
- Kunkel HG, Ahrens EH Jr, Eisenmenger WJ, Bongiovanni AM, Slater RJ. Extreme hypergammaglobulinemia in young women with liver disease of unknown etiology (Abstract). J Clin Invest 1951; 30: 654.
- 6. Bongiovanni AM, Eisenmenger WJ. Adrenal cortical metabolism in chronic liver disease. J Clin Endocrinol 1951; 11: 152-172.
- 7. Zimmerman HJ, Heller P, Hill RP. Extreme hypergammaglobulinemia in subacute hepatic necrosis. N Engl J Med 1951; 244: 245-249.
- 8. Bearn AG, Kunkel HG, Slater RJ. The problem of chronic liver disease in young women. Am J Med 1956; 21: 3-15.
- 9. Good RA. Plasma-cell hepatitis and extreme hypergammaglobulinemia in adolescent females (Abstract). Am J Dis Child 1956; 92: 508-509.
- Saint EG, King WE, Joske RA, Finckh ES. The course of infectious hepatitis with special reference to prognosis and the chronic stage. *Austral Ann Med* 1953; 2: 113-127.
- 11. Mackay JR, Taft LI, Cowling DC. Lupoid hepatitis. Lancet 1956; 2: 1323-1326.
- 12. Joske RA, King WE. The "L.E.-cell" phenomenenon in active chronic viral hepatitis. Lancet 1955; ii: 477-479.
- Bartholomew LG, Hagedorn AB, Cain JC, Baggenstoss AH. Hepatitis and cirrhosis in women with positive clot tests for lupus erythematosus. N Engl J Med 1958; 259: 947-956.
- 14. Aronson AR, Montgomery MM. Chronic liver disease with a "lupus erythematosus-like syndrome". Arch Intern Med 1959; 104: 544-552.
- 15. Mackay IR, Taft LI, Cowling DC. Lupoid hepatitis and the hepatic lesions of systemic lupus erythematosus. *Lancet* 1959; i: 65-69.
- 16. Bartholomew LG, Cain JC, Baggenstoss AH, Hagedorn AB. Further observations on hepatitis in young women with positive clot tests for lupus erythematosus. *Gastroenterology* 1960; 39: 730-736.
- 17. Taft LI, Mackay IR, Cowling DC. Autoclasia: a perpetuating mechanism in hepatitis. Gastroenterology 1960; 38: 563-566.
- Mackay IR, Wood IJ. Lupoid hepatitis: a comparison of 22 cases with other types of liver disease. QJ Med 1962; 31: 485-507.
- 19. Whittingham S, Mackay IR, Irwin J. Autoimmune hepatitis: immunofluorescence reactions with cytoplasm of smooth muscle and renal glomerular cells. *Lancet* 1966; i: 1333-1335.
- 20. Whittingham S, Irwin J, Mackay IR, Smalley M. Smooth muscle autoantibody in "autoimmune" hepatitis. *Gastroenterology* 1966; 51: 499-505.

- 21. Doniach D, Roitt IM, Walker JG, Sherlock S. Tissue antibodies in primary biliary cirrhosis, active chronic (lupoid) hepatitis, cryptogenic cirrhosis and other liver diseases and their clinical implications. *Clin Exp Immunol* 1966; 1: 237-262.
- 22. Bulkley BH, Heizer WD, Goldfinger SE, Isselbacher KJ. Distinctions in chronic active hepatitis based on circulating hepatitis-associated antigen. *Lancet* 1970; ii: 1323-1326.
- 23. Dudley FJ, Scheuer PJ, Sherlock S. Natural history of hepatitis-associated antigenpositive chronic liver disease. *Lancet* 1972; ii: 1388-1393.
- 24. Sherlock S. Chronic hepatitis. Gut 1974; 15: 581-597.
- Golding PL, Smith M, Williams R. Multisytem involvement in chronic liver disease: studies on the incidence and pathogenesis. Am J Med 1973; 55: 772-782.
- 26. Olsson R, Hultén K. Concurrence of ulcerative colitis and chronic active hepatitis: clinical courses and results of colectomy. *Scand J Gastroenterol* 1975; 10: 331-335.
- 27. Copenhagen Study Group for Liver Diseases. Sex, ascites and alcoholism in survival of patients with cirrhosis: effect of prednisolone. N Engl J Med 1974; 291: 271-273.
- 28. Baggenstoss AH, Soloway RD, Summerskill WHJ, Elveback LR, Schoenfield LJ. Chronic active liver disease: the range of histologic lesions, their response to treatment and evolution. *Hum Pathol* 1972; 3: 183-198.
- 29. De Groote J, Desmet VJ, Gedigk P, et al. A classification of chronic hepatitis. Lancet 1968; i: 626-628.
- 30. Geall MG, Schoenfield LJ, Summerskill WHJ. Classification and treatment of chronic active liver disease. *Gastroenterology* 1968; 55: 724-729.
- 31. Bianchi L, de Groote J, Desmet VJ, *et al.* Acute and chronic hepatitis revisited. Review by an international group. *Lancet* 1977; ii: 914-919.

- 32. Kalk H. Die chronischen Verlaufsformen der Hepatitis epidemica in Beziehung zu ihren anatomischen Grundlagen. *Deutsch Med Wschr* 1947; 72: 308-313.
- 33. Schmid M. Die chronische Hepatitis. In: R Hegglin, F Leuthardt, R Schoen, H Schwiegk, A Studer, HU Zollinger, eds. *Experimentelle Medizin, Pathologie und Klinik*. Berlin: Springer Verlag 1966, vol. 18.
- Leevy CM, Tygstrup N. Standardization of nomenclature, diagnostic criteria and diagnostic methodology for diseases of the liver and biliary tract. In: Leevy CM, Newark NJ, eds. Diseases of the liver and biliary tract. Basel: S. Karger, 1976.
- 35. Ludwig J. Morphology of chronic active hepatitis: differential diagnosis and therapeutic implications. In: Czaja AJ, Dickson ER, eds. Chronic active hepatitis- The Mayo Clinic experience. New York: Marcel Dekker 1986, 83-104.
- Ludwig J. Liver biopsy diagnoses and reports: SNOMED Codes, ICD-9-CM Codes, Nomenclature and Terminology. Basel: S. Karger Publ 1984.

- 37. Mackay IR, Taft LI, Cowling DC. Lupoid hepatitis. Lancet 1956; ii: 1323-1326.
- Popper H, Paronetto F, Schaffner F. Immune processes in the pathogenesis of liver disease. Ann N Y Acad Sci 1965; 124: 781-799.
- 39. Kerr JFR, Cooksley WGE, Searle J, et al. The nature of piecemeal necrosis in chronic active hepatitis. Lancet 1979; ii: 827-828.

- 40. Bianchi L, Spichtin HP, Gudat F. Chronic hepatitis. In: RNM MacSween, PP Anthony, PJ Scheuer, eds. Pathology of the Liver. 2nd ed. Edinburgh, London, Melbourne, New York 1987, 310-341.
- 41. Scheuer PJ. Liver Biopsy Interpretation, 3rd ed. London: Baillière Tindall 1980
- 42. Bianchi L, De Groote J, Desmet VJ, *et al.* Morphologic criteria in viral hepatitis: review by an international group. *Lancet* 1971: i: 333-337.
- 43. Gerber MA, Hadziyannis S, Vissoulis C, Schaffner F, Paronetto F, Popper H. Electron microscopy and immunoelectronmicroscopy of cytoplasmic hepatitis B antigen in hepatocytes. *Am J Pathol* 1974; 75: 489-502.
- 44. Czaja AJ, Ludwig J, Baggenstoss AH, Wolf A. Corticosteroid-treated chronic active hepatitis in remission: uncertain prognosis of chronic persistent hepatitis. N Engl J Med 1981; 304: 5-9.
- 45. Popper H, Schaffner F. Chronic hepatitis: taxonomic, etiologic and therapeutic problems. In: PopperH, Schaffner F (eds.). Progress in Liver Disease, vol V. New York: Annals of the New York Academy of Sciences, 1976, 531-558.
- 46. Popper H, Schaffner F. The vocabularity of chronic hepatitis. N Engl J Med 1971; 284: 1154-1156.

47. Seeff LB, Koff RS. Therapy for chronic active hepatitis. Adv Intern Med 1984; 29: 109-145.

INTRODUCTION §5

- 48. Mistilis SP, Skyring AP, Blackburn CRB. Natural history of chronic active hepatitis. I. Clinical features, course, diagnostic criteria, morbidity, mortality and survival. Austr Ann Med 1968; 17: 214-223.
- Sherlock S. Waldenström's chronic active hepatitis. Acta Med Scand 1966; 179(Suppl.): 426-433.
- 50. Soloway RD, Summerskill WHJ, Baggenstoss AH, Geall MG, Gitnick GL, Elveback LR, Schoenfield LJ. Clinical, biochemical, and histological remission of severe chronic active liver disease: a controlled study of treatments and early prognosis. *Gastroenterology* 1972; 63: 820-833.
- Czaja AJ. Autoimmune chronic active hepatitis. In: In: Czaja AJ, Dickson ER, eds. Chronic active hepatitis- The Mayo Clinic experience. New York: Marcel Dekker 1986, 105-126.
- 52. Chwalinska-Sadowska H, Milewski B, Maldyk H. Diagnostic troubles connected with differentiation of systemic lupus erythematosus against chronic active hepatitis. *Mater Med Pol* 1977; 30: 60-64.

- 53. Czaja AJ. Natural history of chronic active hepatitis. In: Czaja AJ, Dickson ER, eds. Chronic active hepatitis- The Mayo Clinic experience. New York: Marcel Dekker 1986, 9-24.
- 54. Harvald B, Madsen S. Long-term treatment of cirrhosis of the liver with prednisolone. Acta Med Scand 1961; 169: 381-387.

- 55. Copenhagen Study Group for Liver Diseases. Effect of prednisone on the survival of patients with cirrhosis of the liver. *Lancet* 1969; i: 119-121.
- 56. Wilcox RG, Isselbacher KJ. Chronic liver disease in young people. Clinical features and course of thirty-three patients. *Am J Med* 1961; 30: 185-195.
- 57. Read AE, Sherlock S, Harrison CV. Active "juvenile" cirrhosis considered as part of a systemic disease and the effect of corticosteroid therapy. *Gut* 1963; 4: 378-393.
- 58. Mistilis SP, Blackburn CRB. Active chronic hepatitis. Am J Med 1970; 48: 484-495.
- 59. Mistilis SP. Natural history of active chronic hepatitis. II. Pathology, pathogenesis and clinico-pathological correlation. Austr Ann Med 1968; 17: 277-288.
- Page AR, Condie RM, Good RA. Suppression of plasma cell hepatitis with 6mercaptopurine. Am J Med 1964; 36: 200-213.
- 61. Schalm SW, Korman MG, Summerskill WHJ, Czaja AJ, Baggenstoss AH. Severe chronic active liver disease. Prognostic significance of initial morphologic patterns. A m J Dig Dis 1977; 22: 973-980.
- 62. Soloway RD, Summerskill WHJ, Baggenstoss AH, Schoenfield LJ. "Lupoid" hepatitis, a nonentity in the spectrum of chronic active liver disease. *Gastroenterology* 1972; 63: 458-465.
- 63. Thaler H. The natural history of chronic active hepatitis. In: Schaffner F, Sherlock S, Leevy CM, eds. The liver and its diseases. New York: Stratton Intercontinental Medical Books 1974, 207-215.
- 64. Kemeny MJ, O'Hanlon G, Gregory PB. Asymptomatic chronic active hepatitisprognosis and treatment (Abstract). *Gastroenterology* 1984; 86: 1325.
- 65. Poulsen H, Christoffersen P. Abnormal bile duct epithelium in chronic aggressive hepatitis and cirrhosis: a review of morphology and clinical, biochemical, and immunologic features. *Hum Pathol* 1972; 3: 217-225.
- 66. Burroughs AK, Bassendine MF, Thomas HC, Sherlock S. Primary liver cell cancer in autoimmune chronic liver disease. Br Med J 1981; 282: 273.
- 67. Jakobovits AW, Gibson PR, Dudley FJ. Primary liver cell carcinoma complicating autoimmune chronic active hepatitis. *Dig Dis Sci* 1981; 26: 694-699.
- De Groote J, Fevery J, Lepoutre L. Long-term follow-up of chronic active hepatitis of moderate severity. Gut 1978; 19: 510-513.
- 69. Fevery J, Desmet VJ, De Groote J. Long-term follow-up and management of asymptomatic chronic active hepatitis. In: Cohen S, Soloway RD, eds. Chronic active liver disease. New York: Churchill Livingstone 1983, 51-64.
- 70. Omata M, Ashcacai M, Liew C-T, Peters RL. Hepatocellular carcinoma in the USA, etiologic considerations. *Gastroenterology* 1979; 76: 279-287.
- 71. Tong MJ, Sun S-C, Schaeffer BT, Chang N-K, Lo K-J, Peters RL. Hepatitis-associated antigen and hepatocellular carcinoma in Taiwan. *Ann Intern Med* 1971; 75: 687-691.
- 72. Koretz RL, Stone O, Mousa M, Gitnick GL. Non-A, non-B posttransfusion hepatitis- a decade later. *Gastroenterology* 1985; 88: 1251-1254.
- 73. Mattson L, Weiland O, Glaumann H. Long-term follow-up of chronic post-transfusion non-A,non-B hepatitis: clinical and histological outcome. *Liver* 1988; 8: 184-188.
- 74. Berman M, Alter HJ, Ishak KG, Purcell RH, Jones EA. The chronic sequelae of non-A, non-B hepatitis. Ann Intern Med 1979; 91: 1-6.
- 75. Koretz RL, Stone O, Gitnick GL. The long-term course of non-A, non-B post-transfusion hepatitis. *Gastroenterology* 1980; 79: 893-898.

- 76. Koretz RL, Suffin SC, Gitnick GL. Post-transfusion chronic liver disease. Gastroenterology 1976; 71: 797-803.
- 77. Resnick RH, Stone K, Antonioli D. Primary hepatocellular carcinoma following non-A,non-B post-transfusion hepatitis. *Dig Dis Sci* 1983; 28: 908-911.
- Gilliam JH, Geisinger KR, Richter JE. Primary hepatocellular carcinoma after chronic non-A,non-B post-transfusion hepatitis. Ann Intern Med 1984; 101: 794-795.
- 79. Schalm SW, Ammon HV, Summerskill WHJ. Failure of customary treatment in chronic active liver disease: causes and management. Ann Clin Res 1976; 8: 221-227.

- Cook GC, Mulligan R, Sherlock S. Controlled prospective trial of corticosteroid therapy in chronic active hepatitis. *Quart J Med* 1971; 158: 159-185.
- 81. Kirk AP, Jain S, Pocock S, Thomas HC, Sherlock S. Late results of the Royal Free Hospital prospective controlled trial of prednisolone therapy in hepatitis B surface antigen negative chronic active hepatitis. *Gut* 1980; 21: 78-83.
- 82. Vogten AJM. Chronische Hepatitis. De klinische en morfologische diagnose en de resultaten van een geïndividualiseerde behandeling. Thesis. Catholic University Nijmegen, The Netherlands 1974. [English summary].
- Copenhagen Study Group for Liver Diseases. Effect of prednisone on the survival of patients with cirrhosis of the liver. *Lancet* 1969; i: 119-121.
- 84. Copenhagen Study Group for Liver Diseases. Sex, ascites, and alcoholism in survival of patients with cirrhosis. Effect of prednisone. *N Engl J Med* 1974; 291: 271-273.
- Murray-Lyon IM, Stern RB, Williams R. Controlled trial of prednisone and azathioprine in active chronic hepatitis. *Lancet* 1973; i: 735-737.
- 86. Summerskill WHJ, Korman MG, Ammon HV, Baggenstoss AH. Prednisone for chronic liver disease: dose titration, standard dose, and combination with azathioprine. *Gut* 1975; 16: 876-883.
- 87. Ammon HV. Assessment of treatment regimens. In: Czaja AJ, Dickson ER, eds. Chronic active hepatitis- The Mayo Clinic experience. New York: Marcel Dekker 1986, 33-46.
- 88. Czaja AJ. Current problems in the diagnosis and management of chronic active hepatitis. Mayo Clin Proc 1981; 56: 311-323.
- Czaja AJ, Summerskill WHJ. Chronic hepatitis. To treat or not to treat? Med Clin N Am 1978; 62: 71-85.
- Schalm SW, Summerskill WHJ, Go VLW. Prednisone for chronic active liver disease: pharmacokinetics including conversion to prednisolone. *Gastroenterology* 1977; 72: 910-913.
- 91. Uribe M, Summerskill WHJ, Go VLW. Comparative serum prednisone and prednisolone concentrations following administration to patients with chronic active liver disease. *Clin Pharmacokinetics* 1982; 7: 452-459.
- 92. Brunner G, Hopf U. Relapse after azathioprine withdrawal in autoimmune chronic active hepatitis. *Lancet* 1985; i: 1216.
- 93. Brunner G, Perings E, Creutzfeldt W. Ergebnisse eines Auslassversuches einer Azathioprin-Langzeit-behandlung bei Patienten mit chronisch agressiver Hepatitis. Zeitschr Gastroenterol 1973; 11: 637-639.
- 94. Stellon AJ, Hegarty JE, Portmann B, Williams R. Randomized controlled trial of azathioprine withdrawal in autoimmune chronic active hepatitis. *Lancet* 1985; i: 668-670.

- 95. Hegarty JE, Nouri Aria KT, Portmann B, Eddleston ALWF, Williams R. Relapse following treatment withdrawal in patients with autoimmune chronic active hepatitis. *Hepatology* 1983; 3: 685-689.
- 96. McCullough AJ, Czaja AJ. Relapse following treatment withdrawal in autoimmune chronic active hepatitis (Letter). *Hepatology* 1984; 4: 747-748.
- 97. Czaja AJ, Beaver SJ, Shiels MT. Sustained remission after corticosteroid therapy of severe hepatitis B surface antigen-negative chronic active hepatitis. *Gastroenterology* 1987; 92: 215-219.
- Sagnelli E, Piccinino F, Manzillo G, et al. Effect of immunosuppressive therapy on HBsAg-positive chronic active hepatitis in relation to presence or absence of HBeAg and antiHBe. *Hepatology* 1983; 3: 690-695.
- 99. European Association for the Study of the Liver. A multicentre randomized clinical trial of low-dose steroid treatment in chronic active HBsAg-positive liver disease (Abstract). *Gastroenterology* 1984; 86: 1317.
- 100. Wu PC, Lai CL, Lam KC, Ho J. Prednisone in HBsAg-positive chronic active hepatitis: histologic evaluation in a controlled prospective study. *Hepatology* 1982; 2: 777-783.
- Schalm SW, Heijtink RA, Masurel N. Temporary disappearance of viral replication in hepatitis B surface antigen-positive chronic active hepatitis. J Infect Dis 1981; 144: 282.
- 102. Hoofnagle JH, Dusheiko GM, Schafer DF, et al.Reactivation of chronic hepatitis B virus infection by cancer chemotherapy. Ann Int Med 1982; 96: 447-449.
- Scullard GH, Robinson WS, Merigan TC, Gregory PB. The effect of immunosuppressive therapy on viral markers in patients with chronic active hepatitis B. *Gastroenterology* 1981; 81: 987-991.
- Rakela J, Redeker AG, Welicky B. Effect of short-term prednisone therapy on aminotransferase levels and hepatitis B virus markers in chronic type B hepatitis. *Gastroenterology* 1983; 84: 956-960.
- 105. Sagnelli E, Manzillo G, Maio G, *et al.* Serum levels of hepatitis B surface and core antigens during immunosuppressive treatment of HBsAg-positive chronic active hepatitis. *Lancet* 1980; ii: 395-397.
- 106. Galbraith GH, Eddleston ALWF, Williams R, et al. Fulminant hepatic failure in leukemia and chronic carcinoma related to withdrawal of cytotoxic drug therapy. Lancet 1975; ii: 105-112.
- 107. Hoofnagle JH, Davis GL, Hanson RG, et al. Randomized double-blind placebocontrolled trial of a one-month course of prednisone in chronic type B hepatitis (Abstract). Gastroenterology 1984; 86: 1324.
- 108. Lam KC, Lai CL, Ng RP, et al. Deleterious effect of prednisolone in HBsAg-positive chronic active hepatitis. N Engl J Med 1981; 304: 380-386.
- Davis GL, Czaja AJ, Taswell HF, Ludwig J, Go VLW. Hepatitis B virus replication in steroid-treated severe HBsAg-positive chronic active hepatitis. *Dig Dis Sci* 1985; 30: 97-103.
- Schalm S, Davis GL, Shiels MT. Chronic active hepatitis B. In: Czaja AJ, Dickson ER, eds. Chronic active hepatitis- The Mayo Clinic experience. New York: Marcel Dekker 1986, 127-151.
- Conn HO, Maddrey WC, Soloway RD. The detrimental effects of adrenocorticosteroid therapy in HBsAg-positive chronic active hepatitis: Facts or artefact? *Hepatology* 1982; 2: 885-887.
- 112. Maier KP, Scholmerich J, Volk B, Gerok W. Liver histology in patients with and without immunosuppressive therapy due to non-A,non-B chronic active hepatitis (Abstract). *Hepatology* 1981; 1: 529.

- 113. Realdi G, Alberti A, Rugge M, et al. Long-term follow-up of acute and chronic non-A,non-B posttransfusion hepatitis: evidence of progression to liver cirrhosis. Gut 1982; 23: 270-275.
- 114. Bradley DW, McCaustland KA, Cook EH, Ebert JW, Maynard JE. Non-A,non-B hepatitis in chimpanzees: effects of immunosuppression on course of disease and recovery of tubule-forming agent from infected liver. In: Vyas GN, Dienstag JL, Hoofnagle JH, eds. Viral Hepatitis and Liver Disease. New York: Grune and Stratton 1984, 451-458.
- 115. Hoofnagle JH, Mullen KD, Jones B, *et al*. Treatment of chronic non-A,non-B hepatitis with recombinant human alpha interferon. *N Engl J Med* 1986; 315: 1575-1578.
- 116. Ideo G, Bellati G, Alfieri G, Boncinelli L, Colombo A. A prospective controlled trial of recombinant alpha interferon versus no treatment in non A non B chronic active hepatitis (Abstract). *Hepatology* 1988; 8: 1269.
- 117. Choo Q-L, Kuo G, Weiner AJ, Overby LR, Bradley DW, Houghton M. Isolation of a cDNA clone derived from a bloodborne non-A,non-B viral hepatitis genome. *Science* 1989; 244: 359-362.
- 118. Kuo G, Choo Q-L, Alter HJ, et al. An assay for circulating antibodies to a major etiologic virus of human non-A, non-B hepatitis. Science 1989; 244: 362-364.
- 119. Thomas HC, Path MRC. Hepatitis B viral infection. Am J Med 1988; 85(Suppl. 2A): 135-140.
- 120. Alexander GJM, Williams R. Natural history and therapy of chronic hepatitis B virus infection. Am J Med 1988; 85(Suppl. 2A): 143-146.
- 121. Czaja AJ, Rakela J, Soloway RD, Davis GL, Ammon HV. Chronic active hepatitis: Controversies and future directions. In: Czaja AJ, Dickson ER, eds. Chronic active hepatitis- The Mayo Clinic experience. New York: Marcel Dekker 1986, 285-314.
- 122. Chase WF, Winn RE, Mayes GR. Oral pulse prednisone therapy in the treatment of HBsAg-negative chronic active hepatitis. *Gastroenterology* 1982; 83: 1292-1296.
- 123. Wang KK, Czaja AJ, Shiels MT, Rakela J. Pulse prednisone in the treatment of severe HBsAg-negative chronic active hepatitis in relapse: a prospective randomized treatment trial (Abstract). *Gastroenterology* 1989; 96: A671.
- 124. Mistilis SP, Vickers CR, Darroch MH, McCarthy SW. Cyclosporin, a new treatment for autoimmune chronic active hepatitis. *Med J Austr* 1985; 143: 463-465.
- 125. Hyams JS, Ballow M, Leichtner AM. Cyclosporine treatment of autoimmune chronic active hepatitis. *Gastroenterology* 1987; 93: 890-893.
- 126. Gilmore IT, Cowan RE, Axon ATR, Thompson RPH. Controlled trial of cyclophosphamide in active chronic hepatitis. *Br Med J* 1979; 1: 1120-1121.
- 127. Jenkins PJ, Portmann BP, Eddleston ALWF, Williams R. Use of phosphatidyl choline in HBsAg negative chronic active hepatitis: results of prospective double-blind controlled trial. *Liver* 1982; 2: 77-81.
- 128. Miracco A, Iodice G, Peluso C, *et al.* Arginine thiazolidinecarboxylate in the treatment of chronic active hepatitis: double-blind comparison with placebo. *J Int Med Res* 1984; 12: 35-39.
- 129. Stern RB, Wilkinson SP, Howorth PJN, Williams R. Controlled trial of synthetic Dpenicillamine and prednisone in maintenance therapy for active chronic hepatitis. *Gut* 1977; 18: 19-22.
- 130. Fuller RK, Jones PK. Colchicine in the treatment of cirrhosis. N Engl J Med 1988; 319: 1285

- 131. Bodenheimer H, Schafner F, Pezzulo J. Evaluation of colchicine therapy in primary biliary cirrhosis. *Gastroenterology* 1988; 95: 124-129.
- Chojkier M, Brenner DA. Therapeutic strategies for hepatic fibrosis. *Hepatology* 1988; 8: 176-182.
- 133. Poupon R, Poupon RE, Calmus Y, Chretien Y, Ballet F, Darnis F. Is ursodeoxycholic acid an effective treatment for primary biliary cirrhosis ? Lancet 1987; i: 834-836.
- 134. Geubel AP, Baggenstoss AH, Summerskill WHJ. Responses to treatment can differentiate chronic active liver disease with cholangitic features from primary biliary cirrhosis syndrome. *Gastroenterology* 1976; 71: 444-449.
- 135. Graubaum H-J, Metzner Chr, Ziesenheim K. Körperliche Belastung und Hepatitisverlauf. Deutsch Med Wchschr 1987; 112: 47-49.
- 136. Ritland S, Foss NE, Skrede S. The effect of a standardized work load on 'liver tests' in patients with chronic active hepatitis. *Scand J Gastroenterol* 1982; 17: 1013-1016.
- 137. Franken FH, Wiechers B. Ergometrische Untersuchungen zur Körperlichen Belastbarkeit chronischer Leberkranker. *Deutsch Med Wchschr* 1973; 98: 528-531.
- Linneke P, Hoffmann H, Ehrke D, Lazarus P. Plasmaenzymaktivitäten bei Lebererkrankungen unter normierter körperlichen Belastung. Z Gesamte Inn Med 1973; 28: 11-14.
- 139. Anando P, Tamburro CH, Leevy CM. Detrimental effect of exercise in hepatitis. *Gastroenterology* 1971; 60: 739.
- Massarat S, Lang N. Änderungen der Aktivitäten von SGOT, SGPT und anderen Serumenzymen sowie des Bromsulphaleintestes unter körperlicher Belastung bei entzündlichen aktiven und nichtaktiven Lebererkrankungen. Gastroenterologia (Basel) 1962; 97(Suppl.): 231-237.
- 141. Zwirner K. Über das Verhalten der Serumtransaminasen nach körperlichen Belastung bei Leberkranken. Acta Hepatosplenol 1970; 7: 97-103.

- 142. Czaja AJ. Natural history, clinical features, and treatment of autoimmune hepatitis. Semin Liver Dis 1984; 4: 1-12.
- 143. Davis GL, Czaja AJ. Immediate and long-term results of corticosteroid therapy for severe idiopathic chronic active hepatitis. In: Czaja AJ, Dickson ER, eds. Chronic active hepatitis- The Mayo Clinic experience. New York: Marcel Dekker 1986, 269-283.
- 144. Davis GL, Czaja AJ, Ludwig J. Development and prognosis of histologic cirrhosis in corticosteroid-treated hepatitis B surface antigen-negative chronic active hepatitis. *Gastroenterology* 1984; 87: 1222-1227.
- 145. Czaja AJ, Davis GL, Ludwig J, Baggenstoss AH, Taswell HF. Autoimmune features as determinants of prognosis in steroid-treated chronic active hepatitis of uncertain etiology. *Gastroenterology* 1983; 85: 713-717.
- Czaja AJ, Rakela J, Ludwig J. Features reflective of early prognosis in corticosteroidtreated severe autoimmune chronic active hepatitis. *Gastroenterology* 1988; 95: 448-453.
INTRODUCTION §9

- 147. Vento S, Eddleston ALWF. Immunological aspects of chronic active hepatitis. Clin Exp Immunol 1987; 68: 225-232.
- 148. Eddleston ALWF. Immunology of chronic active hepatitis. *Quart J Med* 1985; 55: 191-198.
- 149. Vento S, Nouri-Aria KT, Eddleston ALWF. Immune mechanisms in autoimmune chronic active hepatitis. Scand J Gastroenterol 1985; 114: 91-103.
- 150. Meyer zum Büschenfelde K-H, Kossling FK, Miescher PA. Experimantal chronic active hepatitis in rabbits following immunization with human liver proteins. *Clin Exp Immunol* 1972; 11: 99-108.
- 151. Meyer zum Büschenfelde K-H, Miescher PA. Liver specific antigens, purification and characterization. *Clin Exp Immunol* 1972; 10: 89-102.
- 152. Thomson AD, Cochrane MAG, McFarlane IG, Eddleston ALWF, Williams R. Lymphocyte cytotoxicity to isolated hepatocytes in chronic active hepatitis. *Nature* (London) 1974; 252: 721-722.
- 153. Cochrane MAG, Moussouros A, Thomson AD, Eddleston ALWF, Williams R. Antibody-dependent cell-mediated (K-cell) cytotoxicity against isolated hepatocytes in chronic active hepatitis. *Lancet* 1976; i: 441-444.
- 154. Kakumu S, Tateki H, Goji H, Sakamoto N. Lymphocyte cytotoxicity against Chang liver cells in chronic active hepatitis. *Cellular Immunol* 1978; 36: 46-53.
- Facchini A, Stefanini GF, Bernardi M, Miglio F, Gassbarrini G, Labo G. Lymphocytotoxicity test against rabbit hepatocytes in chronic liver diseases. *Gut* 1978; 19: 189-193.
- 156. Fernandez-Cruz E, Escartin P, Bootello A, Kreisler M, Segovia de Arara JH. Hepatic damage induced by lymphocytes from patients with chronic liver diseases, as detected by LDH release. *Clin Exp Immunol* 1978; 31: 436-442.
- 157. Paronetto F, Vernace S. Immunological studies in patients with chronic active hepatitis. Cytotoxic activity of lymphocytes to autochtonous liver cells grown in tissue culture. *Clin Exp Immunol* 1975; 19: 99-104.
- 158. Wands JR, Isselbacher KJ. Lymphocyte cytotoxicity to autologous liver cells in chronic active hepatitis. *Proc Natl Acad Sci* 1975; 72: 1301-1303.
- 159. Mieli-Vergani D, Vergani D, Jenkins PJ, Portmann B, Mowat AP, Eddleston ALWF, Williams R. Lymphocyte cytotoxicity to autologous hepatocytes in HBsAg-negative chronic active hepatitis. *Clin Exp Immunol* 1979; 38: 16-21.
- Mieli-Vergani D, Vergani D, Portmann B, White Y, Murray-Lyon I, Marigold JH, Woolf I, Eddleston ALWF, Williams R. Lymphocyte cytotoxicity to autologous hepatocytes in HBsAg-positive chronic liver disease. *Gut* 1982; 23: 1029-1036.
- 161. Vogten AJM, Hadzic N, Shorter RG, Summerskill WHJ, Taylor WF. Cell-mediated cytotoxucity in chronic liver disease: a new test system. *Gastroenterology* 1978; 74: 883-889.
- McFarlane IG, McFarlane BM, Major GN, Tolley P, Williams R. Identification of the hepatic asialoglycoprotein receptor (hepatic lectin) as a component of liver specific membrane lipoprotein (LSP). *Clin Exp Immunol* 1984; 55: 347-354.
- 163. McFarlane BM, McSorley CG, Vergani IG, McFarlane IG, Williams R. Serum autoantibodies reacting with the hepatic asialoglycoprotein receptor protein (hepatic lectin) in acute and chronic liver disorders. J Hepatol 1986; 3: 196-205.

- 164. Sipos J, McFarlane BM, Gove CD, McSorley CG, McFarlane IG, Williams R. Antibodies against the hepatic asialoglycoprotein receptor preferentially coat periportal hepatocytes in the in situ perfused rat liver. J Hepatol 1988; 7: S177.
- McFarlane IG. Autoimmunity in liver disease. Editorial review. Clin Sci 1984; 67: 569-578.
- 166. De Kretser TA, McFarlane IG, Eddleston ALWF, Williams R. A species non-specific liver plasma membrane antigen and its involvement in chronic active hepatitis. *Biochem J* 1980; 186: 679-685.
- 167. Lebwohl N, Gerber MA. Characterization and demonstration of human liver-specific protein (LSP) and apo-LSP. Clin Exp Immunol 1981; 46: 435-442.
- Gerber MA, Lebwohl N, Thung SW. Liver-specific protein. How specific is it ? In: Berk PD, Chalmers TC, eds. Frontiers in Liver Disease. New York: Thieme-Stratton 1981, 139-143.
- 169. Manns M, Meyer zum Büschenfelde K-H. Fractionation of the liver membrane lipoprotein (LSP) and characterization of its antigenic determinants by autoantibodies and a heterologous anti-serum. Gut 1982; 23: 14-20.
- 170. Lambert KJ, Major GN, Welsh CJR, Ryall JE, McFarlane IG, Williams R. Production and preliminary characterization of monoclonal antibodies to human liver-specific lipoprotein (LSP). *Liver* 1984; 4: 122-127.
- 171. Poralla T, Dienes HP, Dippold W, Manns M, Meyer zum Büschenfelde K-H. A monoclonal antibody directed against an organ-specific liver cell membrane antigen in rabbits.(Abstract). *Hepatology* 1983; 3: 842.
- 172. Wiedmann KH, Trejdosiewcz LK, Goodall AH. Thomas HC. Analysis of the antigenic composition of liver-specific lipoprotein using murine monoclonal antibodies. *Gut* 1985; 26: 510-517.
- 173. Kenna JG, Major GN, Lambert KJ, McFarlane IG, Williams R. Murine monoclonal antibodies against "liver specific lipoprotein" (LSP) defining three antigenic sites which differ in tissue- and species-distribution and subcellular location. *Liver* 1985; 5: 13-20.
- Murakami H, Kuriki J, Kakunu S, Fukui K, Sakamoto N. The specificity of human liver membrane lipoprotein: studies with monoclonal antibodies. *Hepatology* 1984; 4: 192-198.
- 175. Riisom K, Diederichsen H. Demonstration of organ-non-specific antigens in liverspecific protein. Gastroenterology 1983; 85: 1271-1276.
- 176. Behrens U, Paronetto F. Studies on 'liver-specific' antigens. I. Evaluation of the liver specificity of 'LSP' and 'LP2'. *Gastroenterology* 1979; 77: 1045-1052.
- 177. Kakumu S, Kazuaki Y, Kashio T. Immunoregulatory T-cell function in acute and chronic liver disease. *Gastroenterology* 1980; 79: 613-619.
- 178. Hotta R, Kuriki J, Kakumu S. Loss of suppressor T-cell function and circulating immune complexes in chronic active liver diseases. *Clin Exp Immunol* 1981; 44: 459-466.
- Abdou NI, Sagawa A, Pascual E, Hebert J, Sadeghee S. Suppressor T cell abnormality in idiopathic systemic lupus erythematosus. *Clin Immunol Immunopathol* 1976; 6: 192-199.
- Keystone EC, Gladman DD, Buchanan R, Cane D, Poplonski L. Impaired antigenspecific suppressor cell activity in patients with rheumatoid arthritis. Arthritis Rheum 1980; 23: 1246-1250.
- 181. Frazer IH, Mackay IR. Antibodies to liver cell membrane antigens in chronic active hepatitis (CAH). III. Partial characterization of the liver cell membrane antigens and comparison of reactivities in sera from patients with various liver diseases. Clin Exp Immunol 1984; 57: 429-437.

- 181a. Ikeda T, Uchihara M, Daiguji Y, Hasamura Y, Takeuchi J. Immunological mechanisms of corticosteroid therapy in chronic active hepatitis: analysis of peripheral blood suppressor T-cell and interleukin 2 activities. *Clin Immunol Immunopathol* 1988; 48: 371-379.
- Nouri-Aria KT, Hegarty JE, Alexander GJM, Eddleston ALWF. Effect of corticosteroids on suppressor-cell activity in "autoimmune" and viral chronic active hepatitis. N Engl J Med 1982; 307: 1301-1304.
- Vento S, Hegarty J, Botazzo G, Macchia E, Williams R, Eddleston ALWF. Antigen specific suppressor cell function in autoimmune chronic active hepatitis. *Lancet* 1984; i: 1200-1204.
- 184. Vento S, O'Brien CJ, McFarlane IG, et al. T-cell inducers of suppressor lymphocytes control liver-directed autoreactivity. *Lancet* 1987; i: 886-887.
- 185. Watanabe Y, Kawakami H, Ikemoto Y, et al. Effect of neonatal thymectomy on experimental autoimmune hepatitis in mice. Clin Exp Immunol 1987; 67: 105-113.
- 186. Lindberg J, Lindholm A, Lundin P, Iwarson S. Trigger factors and HL-A antigens in chronic active hepatitis. Br Med J 1975; 4: 77-79.
- Mackay IR, Morris PJ. Association of autoimmune active chronic hepatitis with HL-A 1. Lancet 1972; ii: 793-795.
- Galbraith RM, Eddleston ALWF, Smith MGM, Williams R, et al. Histocompatibility antigens in active chronic hepatitis and primary biliary cirrhosis. Br Med J 1974; 3: 604-605.
- 189. Page AR, Sharp HL, Greenberg LJ. Genetic analysis of patients with chronic active hepatitis. J Clin Invest 1975; 56: 530-535.
- 190. Galbraith RM, Smith M, Mackenzie RM. High prevalence of seroimmunolic abnormalities in relatives of patients with chronic active hepatitis or primary biliary cirrhosis. N Engl J Med 1974; 190: 63-69.
- 191. Mackay IR, Whittingham S, Matthew JD. Genetic determinations of autoimmune chronic active hepatitis. Springer Semin Immunopathol1980; 3: 285-296.
- 192. Vento S, Nouri-Aria KT, Eddleston ALWF. Immune mechanisms in autoimmune chronic active hepatitis. Scand J Gastroenterol 1984; 20(Suppl.114); 91-103.
- 193. O'Brien C, Eddleston ALWF. Immunology of autoimmune and viral chronic active hepatitis. In: Wright R, Hodgson HJF, eds. Gastrointestinal and Liver Immunology. Ballières Clinical Gastroenterology. London, Philadelphia, Toronto, Sydney, Tokyo: Ballière Tindall, 1987: 1: 647-674.
- 194. Kato Y, Nakogawa H, Kobayashi K, Hattori N, Hatano K. Interferon production by peripheral lymphocytes in HBsAg positive liver diseases. *Hepatology* 1982; 2: 789-790.
- 195. Abb J, Zachoval R, Eisenberg J, et al. Production of interferon alpha and interferon gamma by peripheral blood leukocytes from patients with chronic hepatitis B virus infection. J Med Virol 1985; 16: 171-176.
- 196. Saxena S, Nouri-Aria KT, Anderson MG, Williams R, Eddleston ALWF. In vitro alphainterferon treatment of peripheral blood mononuclear cells improves interleukin-2 activity in HBV related chronic liver disease. J Hepatol 1985; 4: 385.
- 197. Chisari FV. Regulation of human lymphocyte function by a soluble extract from normal liver. J Immunol 1978; 121: 1279-1286.
- 198. Grol M, Schumacher K. Purification and biochemical characterization of human liverderived inhibitory protein (LIP). J Immunol 1983; 130: 323-326.
- Schrempf-Delker GE, Baron DP, Brattig DP, Bockhorn H, Berg PA. Biological and immunological characterization of a human liver immunoregulatory protein. *Hepato*logy 1983; 3: 939-946.

- 200. Chisari FV, Edgington TS. Lymphocyte E-rosette inhibitory factor: a regulatory serum lipoprotein. J Exp Med 1975; 142: 1092-1107.
- 201. Chisari FV, Routenberg JA, Edgington TS. Mechanisms responsible for defective human T lymphocyte sheep erythrocyte rosette function associated with hepatitis B virus infection. *J Clin Invest* 1976; 57: 1227-1238.
- 202. Chisari FV, Routenberg JA, Fiala M, Edgington TS. Extrinsic modulation of human Tlymphocyte E rosette function associated with prolonged hepatocellular injury after viral hepatitis. J Clin Invest 1976; 59: 134-142.
- 203. Sanders G, Perrillo RP. Rosette inhibitory factor: T-lymphocyte subpopulation specificity and potential immunoregulatory role in hepatitis B viral infection. *Hepatology* 1982; 2: 547-552.
- 204. Brattig N, Berg PA. Serum inhibitory factors (SIF) in patients with acute and chronic hepatitis and their clinical significance. *Clin Exp Immunol* 1976; 25: 40-44.
- 205. Nakao M, Mizoguchi Y, Monna T, Yamamoto S, Morisawa S. Studies on an inhibitory factor to phytohemagglutinin-induced lymphocyte transformation found in the serum of patients with various liver diseases. *Acta Hepatol Scand* 1978; 25: 335-343.
- Brattig N, Berg PA. Immunosuppressive serum factors in viral hepatitis. I. Characterization of serum inhibition factors as lymphocyte antiactivators. *Hepatology* 1983; 3: 638-646.
- Curtiss LK, Edgington TS. Regulatory serum lipoproteins: regulation of lymphocyte stimulation by a species of low-density lipoprotein. J Immunol 1976; 116: 1452-1458.
- 208. Curtiss LK, Edgington TS. Differential sensitivity of lymphocyte subpopulations to suppression by low density lipoprotein inhibitor, an immunoregulatory human serum low density lipoprotein. J Clin Invest 1979; 63: 193-201.
- 209. Sanders GE, Perrillo RP. Suppression of T helper function: an immunoregulatory effect of rosette inhibitory factor in hepatitis B virus infection. *Hepatology* 1985; 5: 392-396.
- Vogten AL, Shorter RG. The immunologic features of chronic active hepatitis. In: Czaja AJ, Dickson ER, eds. Chronic Active Hepatitis, the Mayo Clinic experience. New York: Marcel Dekker, Inc. 1986, 189-204.
- 211. Eggink HF, Houthoff HJ, Huitema S, Gips CH, Poppema S. Cellular and humoral immune reactions in chronic active liver disease. I. Lymphocyte subsets in liver biopsies of patients with untreated idiopathic autoimmune hepatitis, chronic active hepatitis B and primary biliary cirrhosis. *Clin Exp Immunol* 1982; 50: 17-24.
- 212. Si L, Whiteside TL, Schade RS, Van Thiel DH. Studies on lymphocyte subpopulations in the liver tissue and blood of patients with chronic active hepatitis. J Clin Immunol 1983; 3: 408-419.
- 213. Kaneda K, Kurioka N, Seki S, Wake K, Yamamoto S. Pit-cell hepatocyte contact in autoimmune hepatitis. *Hepatology* 1984; 4: 955-958.
- 214. Husby G, Blomhoff JP, Elgjo K, Williams RC Jr. Immunohistochemical characterization of hepatic tissue lymphocyte subpopulations in liver disease. Scand J Gastroentero l 1982; 17: 855-860.
- 215. Cochrane AMG, Tsantoulas DC, Moussouros A, McFarlane IG, Eddleston ALWF, Williams R. Lymphocyte cytotoxicity for kidney cells in the renal tubular acidosis of auto-immune liver disease. Br Med J 1976; ii: 276.
- 216. Vento S, Hegarty JE, Alberti A, O'Brien C, Alexander GM, Eddleston ALWF, Williams R. T-lymphocyte sensitization to HBcAg and T cell-mediated unresponsiveness to HBsAg in hepatitis B virus-related chronic liver disease. *Hepatology* 1985; 5: 192-197.
- 217. Wands JR, Isselbacher KJ: Lymphocyte cytotoxicity to autologous liver cells in chronic active hepatitis. Proc Natl Acad Sci 1975; 72: 1301-1303.

- 218. Wands JR, Perrotto JL, Alpert R, Isselbacher KJ. Cell-mediated immunity in acute and chronic hepatitis. J Clin Invest 1975; 55: 921-929.
- Geubel AP, Keller RH, Summerskill WHJ, Dickson ER, Tomasi TB, Shorter RG. Lymphocyte cytotoxicity and inhibition studied with autologous liver cells: observations in chronic active liver disease and the primary biliary cirrhosis syndrome. *Gastroenterology* 1976; 71: 450-456.
- 220. Vierling JM, Nelson DL, Strober W. In vitro cell-mediated cytotoxicity in primary biliary cirrhosis and chronic active hepatitis: dysfunction of spontaneous cell-mediated cytotoxicity in primary biliary cirrhosis. J Clin Invest 1977; 60: 1116-1128.
- 221. El Sheikh H, Osman CG, Cullens H, Eddlestom ALWF, Williams R. T-lymphocyte mediated cytotoxicity in HBsAg positive liver disease. *Clin Exp Immunol* 1978; 31: 158-165.
- 222. Dienstag JL, Bhan AK. Enhanced in vitro cell-mediated cytotoxicity in chronic hepatitis B virus infection: absence of specificity for virus-expressed antigen on target cell membranes. *J Immunol* 1980; 123: 2269-2276.
- 223. Chisari FV, Bieger MJ, Josepho CA, Xavier C, Anderson DS. Functional properties of lymphocyte subpopulations in hepatitis B virus infection. II. Cytotoxic effector killing of targets that naturally express hepatitis B surface antigen and liver-specific lipoprotein. J Immunol 1981; 126: 45-49.
- 224. Poralla T, Hütteroth TH, Meyer zum Büschenfelde K-H. Cellular cytotoxicity against autologous hepatocytes in acute and chronic non-A, non-B hepatitis. *Gut* 1984; 25: 114-120.
- 225. Naumov NV, Mondelli M, Alexander GJM, Tedder RS, Eddleston ALWF, Williams R. Relationship between expression of hepatitis B virus antigens in isolated hepatocytes and autologous lymphocyte cytotoxicity in patients with chronic hepatitis B virus infection, *Hepatology* 1984; 4: 63-68.
- 226. Mondelli M, Eddleston ALWF. Lymphocyte cytotoxicity for autologous hepatocytes. Gut 1984; 25: 109-113.
- 227. Triger DR, Alp MH, Wright R. Bacterial and dietary antibodies in liver disease. Lancet 1972; i: 60-63.
- 228. Triger DR. Bacterial, viral and auto antibodies in acute and chronic liver disease. Ann Clin Res 1976; 8: 174-181.
- 229. Protell RL, Soloway RD, Martin WJ, Schoenfield LJ, Summerskill WHJ. Antisalmonella agglutinins in chronic active liver disease. *Lancet* 1971; ii: 330-331.
- 230. Closs O, Haukenes G, Gjone E, Blomhoff JP. Raised antibody titres in chronic disease. Lancet 1971; ii: 1202-1203.
- 231. Thomas HC, Holden R, Ironside J, Somerville RG. Viral and bacterial antibodies in primary biliary cirrhosis and other chronic liver disease (Abstract). Gut 1974; 15: 826.
- 232. Triger DR, Kurtz JB, MacCallum FO, Wright R. Raised antibody titres to measles and rubella viruses in chronic active hepatitis. *Lancet* 1972; i: 665-667.
- 233. Triger DR, Kurtz JB, Wright R. Viral antibodies and auto-antibodies in chronic liver disease. *Gut* 1974; 15: 94-98.
- 234. The TH, Klein G, Langenhuysen MMAC. Antibody reactions to virus-specific early antigens (EA) in patients with cytomegalovirus (CMV) infection. *Clin Exp Immunol* 1974; 16: 1-12.
- 235. The TH, Gips CH. Activation of antibodies against cytomegalovirus early antigen in a population of oxyphenisatin users (Abstract). *Digestion* 1974; 10: 308.
- 236. Christie KE, Haukenes G. Measles virus-specific IgM antibodies in sera from patients with chronic active hepatitis. *J Med Virol* 1983; 12: 267-272.

237. Robertson DAF, Thang SL, Guy EC, Wright R. Persistent measles virus genome in autoimmune chronic active hepatitis. Lancet 1987; ii: 9-11.

INTRODUCTION §10

- Czaja AJ. Current problems in the diagnosis and management of chronic active hepatitis. Mayo Clin Proc 1981; 56: 311-323.
- 239. Scheuer PJ. Liver biopsy in the diagnosis of cirrhosis. Gut 1970; 11: 275-278.
- Boyer JL. Chronic hepatitis: a perspective on classification and determinants of prognosis. Gastroenterology 1976; 70: 1161-1171.
- 241. Uribe M, Go VLW. Prednisone pharmacokinetics and toxicity in chronic active liver disease and health. In: Czaja AJ, Dickson ER, eds. Chronic Active Hepatitis, the Mayo Clinic experience. New York: *Marcel Dekker, Inc.* 1986, 47-67.
- Stellon AJ, Davies A, Compston J, Williams R. Bone loss in autoimmune chronic active hepatitis on maintenance corticosteroid therapy. *Gastroenterology* 1985; 89: 1078-1083.
- 243. Uribe M, Go VLW, Summerskill WHJ. Prednisone and peptic ulcer in chronic active liver disease. N Engl J Med 1977; 296: 173-174.
- 244. Czaja AJ, Wolf AM, Summerskill WHJ. Development and early prognosis of esophageal varices in severe chronic active liver disease (CALD) treated with prednisolone. *Gastroenterology* 1979; 77: 629-633.
- 245. Kingston ME, Ashraf Ali M, Atiyeh M, Donneley RJ. Diabetes mellitus in chronic active hepatitis and cirrhosis. *Gastroenterology* 1984; 87: 688-694.
- 246. Cacciatore L, Cozzolino G, Giardina MG, et al. Liver cirrhosis as a diabetogenic condition. Dig Dis Sci 1986; 31: 111.
- 247. Gentile S, Marmo R, Coltori M, Del Vecchio Blanco C. Diabetes mellitus associated with chronic liver disease. *Dig Dis Sci* 1987; 32: 947-948.
- 248. Uribe M, Casian C, Rojas S, Sierra JG, Go VLW. Decreased bioavailability of prednisone due to antacids in patients with chronic active liver disease. *Gastroenterology* 1981; 80: 661-665.
- 249. Ludwig J, Axelsen R. Drug effects on the liver: an updated tabular compilation of drugs and drug-related hepatic diseases. *Dig Dis Sci* 1983; 28: 651-666.
- 250. Reid IR, Ibbertson HK. Calcium supplements in the prevention of steroid-induced osteoporosis. Am J Clin Nutr 1986; 44: 287-290.
- Hahn TJ, Halstead LR, Teitelbaum SL. Altered mineral metabolism in glucocorticoidinduced osteopenia. Effect of 25-hydroxyvitamin D administration. J Clin Invest 1979; 64: 655-665.
- 252. Dijkman TR, Haralson KM, Gluck OS, Murphy WA, Teitelbaum SL, Hahn TJ, Hahn BV. Effect of oral 1,25-dihydroxyvitamin D and calcium on glucocorticoid-induced osteopenia in patients with rheumatic diseases. *Arthritis Rheum* 1984; 27: 1336-1343.
- 253. Ringe JD, Wetzel D. Salmon calcitonin in the therapy of corticoid-induced osteoporosis. Europ J Clin Pharmacol 1987; 33: 35-39.
- 254. Reid IR, King AR, Alexander CJ, Ibbertson HK. Prevention of steroid-induced osteoporosis with (3-amino-1-hydroxy-propylidene)-1,1-biphosphonate (APD). Lancet 1988; i: 143-146.
- 255. Czaja AJ, Summerskill WHJ. Malignancy in chronic liver disease (letter to the editor). *Gastroenterology* 1977; 73: 192.
- 255a. Wang KK, Czaja AJ, Beaver SJ, Go VLW. Extrahepatic malignancy following long-term immunosuppressive therapy of severe hepatitis B surface antigen-negative chronic active hepatitis. *Hepatology* 1989; 10: 39-43.

- 256. Tage-Jensen U, Schlichting P, Thomsen HF, Hoybye G, Thomsen AAC and The Copenhagen Study Group for Liver Diseases. Malignancies following long-term azathioprine treatment in chronic liver disease. *Liver* 1987; 7: 81-83.
- 257. Schaffner F. The oncogenicity of azathioprine? Hepatology 1988; 8: 693.
- 258. Dabezies MA, Soloway RD. Interpretation, application, and limitations of Mayo Clinic results. In:In: Czaja AJ, Dickson ER, eds. Chronic Active Hepatitis, the Mayo Clinic experience. New York: *Marcel Dekker, Inc.* 1986, 25-32.
- 259. Schalm SW, Ammon HV, Summerskill WHJ. Failure of customary treatment in chronic active liver disease: causes and management. Ann Clin Res 1976; 8: 221-227.
- Czaja AJ, Wolf AM, Summerskill WHJ. Development and early prognosis of esophageal varices in severe chronic active liver disease (CALD) treated with prednisone. *Gastroenterology* 1979; 77: 629-633.
- Wright SH, Czaja AJ, Katz RS, Soloway RD. Systemic mycosis complicating high dose corticosteroid treatment of chronic active liver disease. Am J Gastroenterol 1980; 74: 428-432.
- 262. Steven MM, Buckly JD, Mackay IR. Pregnancy in chronic active hepatitis. Quart J Med 1979; 192: 519-531.
- Czaja AJ, Ammon HV, Summerskill WHJ. Clinical features and prognosis of severe chronic active liver disease (CALD) after corticosteroid-induced remission. *Gastroenterology* 1980; 78: 518-523.
- 264. Hegarty JE, Nouri Aria KT, Portmann B, Eddleston ALWF, Williams R. Relapse following treatment withdrawal in patients with autoimmune chronic active hepatitis. *Hepatology* 1983; 3: 685-689.
- McCullough AJ, Czaja AJ. Relapse following treatment withdrawal in patients with autoimmune chronic active hepatitis (Letter to the editor). *Hepatology* 1984; 4: 747-748.
- Davis GL, Czaja AJ, Baggenstoss AH, Taswell HF. Prognostic and therapeutic implications of extreme serum aminotransferase elevation in chronic active hepatitis. Mayo Clin Proc 1982; 57: 303-309.
- 267. Davis GL, Czaja AJ. Prolonged steroid therapy for severe chronic active liver disease (CALD): a diminishing return (Abstract). Gastroenterology 1980; 78: 1153.
- Czaja AJ, Wolf AM, Baggenstoss AH. Laboratory assessment of severe chronic active liver disease during and after corticisteroid therapy: correlation of serum transaminase and gamma globulin levels with histologic features. *Gastroenterology* 1981; 80: 687-692.
- McFarlane IG, Hegarty JE, McSorley CG, McFarlane BM, Williams R. Antibodies to liver specific protein predict outcome of treatment withdrawal in autoimmune chronic active hepatitis. *Lancet* 1984; ii: 954-956.
- Czaja AJ, Davis GL, Ludwig J, Taswell HF. Complete resolution of inflammatory activity following corticosteroid treatment of HBsAg-negative chronic hepatitis. *Hepatology* 1984; 4: 622-627.
- 271. Haagsma EB, Gips CH, Wesenhagen H, de Jong GMTh, van Imhoff GW, Krom RAF. Child and Turcotte's classification of hepatic functional reserve and its modifications by Campbell and by Pugh in predicting blood loss during liver transplantation. In: Gips CH and Krom RAF, eds. Progress in liver transplantation. Dordrecht, Boston, Lancaster: Martinus Nijhoff Publ. [Hingham, MA: Kluwer Academic Publishers], 1985; 209-220.
- 272. Owen CA, Rettke SR, Bowie EJW, et al. Hemostatic evaluation of patients undergoing liver transplantation. Mayo Clin Proc 1987; 62: 761-772.
- 273. Cuervas-Mons V., Millan I, Gavaler JS, et al. Prognostic value of preoperatively obtained clinical and laboratory data in predicting survival following orthotopic liver transplantation. *Hepatology* 1986; 6: 922-927.

- 274. Arroyo V, Bosch J, Gaya-Beltran J, et al. Plasma renin activity and urinary sodium excretion as prognostic indicators in non-azotemic cirrhosis with ascites. Ann Int Med 1981; 94: 198-201.
- 275. Dickson ER, Grambsch PM, Fleming TR, Fisher LD, Langworthy A. Prognosis in primary biliary cirrhosis: model for decision making. *Hepatology* 1989; 10: 1-7.
- 275a. Christensen E. Prognostication in primary biliary cirrhosis: relevance to the individual patient. *Hepatology* 1989; 10: 111-113.
- 276. Van der Putten ABMM, Bijleveld CMA, Slooff MJH, et al. Selection criteria and decisions in 375 patients considered for liver transplantation during 1977-1985. Liver 1987; 7: 84-90.
- 277. Neuberger J, Portmann B, Calne R, Williams R. Recurrence of autoimmune chronic active hepatitis following orthotopic liver grafting. *Transplantation* 1984; 37: 363-365.
- 278. Terpstra OT, Schalm SW, Weimar W, et al. Auxilliary partial liver transplantation for end-stage chronic liver disease. N Engl J Med 1988; 319: 1507-1511.

INTRODUCTION §11

- 279. Ludwig J, Czaja AJ. The role of liver biopsy interpretation in the management of chronic active hepatitis. In: S Cohen, RD Soloway, eds. *Chronic active liver disease*. New York: Churchill Livingstone 1983, 171-187.
- Schlichting P, Holund B, Poulsen H. Liver biopsy in chronic aggressive hepatitis. Diagnostic reproducibility in relation to size of specimen. Scand J Gastroenterol 1983; 18: 27-32.
- 281. Theodossi A, Skene AM, Portmann B, et al. Observor variation in assessment of liver biopsies, including analysis by Kappa statistics. Gastroenterology 1980; 79: 232-241.
- 282. Pagliaro L, Rinaldi F, Craxi A, DiPiazza S, et al. Percutaneous blind biopsy versus laparoscopy with guided biopsy in diagnosis of cirrhosis. Dig Dis Sci 1983; 28: 39-43.
- Soloway RD, Baggenstoss AH, Schoenfield LJ, Summerskill WHJ. Observer error and sampling variability tested in evaluation of hepatitis and cirrhosis by liver biopsy. Am J Dig Dis 1971; 16: 1082-1086.
- 284. Knodell RG, Ishak KG, Black WC, *et al.* Formulation and application of a numerical scoring system for assessing histological activity in asymptomatic chronic active hepatitis. *Hepatology* 1981; 1: 431-435.
- 285. McCullough AJ. Laboratory assessment of liver function and inflammatory activity in chronic active hepatitis. In: Czaja AJ, Dickson ER, eds. Chronic active hepatitis- The Mayo Clinic experience. New York: Marcel Dekker 1986, 205-246.
- Zamcheck N, Klausenstock O. Liver biopsy. II. The risk of needle biopsy. N Engl J Med 1953; 249: 1062-1069.
- 286a. Perrault J, McGill DB, Ott BJ, Taylor WF. Liver biopsy: complications in 1000 inpatients and outpatients. *Gastroenterology* 1978; 74: 103-106.
- 287. McIntyre N. The limitations of conventional liver function tests. Sem Liver Dis 1983; 3: 265-274.
- 288. Clermont RJ, Chalmers TC. The transaminase tests in liver disease. *Medicine* 1967; 46: 197-207.
- 289. Zimmerman HJ, West M. Serum enzyme levels in the diagnosis of hepatic disease. A m J Gastroenterol 1963; 40: 387-404.
- 290. Owen JA, Robertson RF. Paper electrophoresis of serum proteins in hepatobiliary disease. Lancet 1956; ii: 1125-1129.

- 291. Wroblewski F. The clinical significance of transaminase activities of serum. Am J Med 1959; 27: 911-923.
- 292. See reference 270 above.
- 293. Kallai L, Hahn H, Roder V. Correlation between histological findings and serum transaminase values in chronic disease of the liver. Acta Med Scand 1964; 175: 49-56.
- 294. Czaja AJ, Wolf AM, Baggenstoss AH. Clinical assessment of cirrhosis in severe chronic active liver disease. Specificity and sensitivity of physical and laboratory findings. *Mayo Clin Proc* 1980; 55: 360-364.
- McCullough AJ, Czaja AJ, Jones JD, Go VLW. The nature and prognostic significance of serial amino acid determinations in severe chronic active liver disease. *Gastro*enterology 1981; 81: 645-652.
- 296. Bernardini P, Fischer JE. Amino acid imbalance and hepatic encephalopathy. Ann Rev Nutr 1982; 2: 419-454.
- 297. Shiels MT, Cza ja AJ, Ludwig J, McCullough AJ, Jones JD, Go VLW. Diagnostic and prognostic implications of plasma amino acid determinations in chronic active hepatitis. *Dig Dis Sci* 1985; 30: 819-823.
- 298. Steigmann F, Szanto PB, Poulos A, Lim PE, Dubin A. Significance of serum aminograms in diagnosis and prognosis of liver diseases. J Clin Gastroenterol 1984; 6: 453-460.
- 299. Zoli M, Marchesini G, Angiolini A, Dondi C, Bianchi FB, Pisi E. Plasma amino acids as markers of liver dysfunction in cirrhotics. *Scand J Gastroenterol* 1981; 16: 689-692.
- 300. Morgan MY, Marshall AW, Milsom JP, Sherlock S. Plasma amino-acid patterns in liver disease. Gut 1982; 23: 362-370.
- Reilly JJ, Halow GM, Gerhardt AL, Ritter PS, Gavaler JS, Van Thiel D. Plasma amino acids in liver transplantation: correlation with clinical outcome. Surgery 1985; 97: 263-269.
- 302. Shiota T, Nakatsukasa H, Fujiwara M, et al. Plasma amino acid imbalance in alcoholic liver cirrhosis. Biochemical Med 1984; 32: 181-188.
- 303. Levine RJ, Conn HO. Tyrosine metabolism in patients with liver disease. J Clin Invest 1967; 46: 2012-2020.
- 304. Freund H, James JH, Brenner W, Fischer JE. Plasma amino acid analysis in the differential diagnosis of jaundice. Am J Surg 1980; 139: 142-146.
- 305. Soeters PB, Fischer JE. Insulin, glucagon, amino acid imbalance and hepatic encephalopathy. *Lancet* 1976; ii: 880-882.
- 306. Munro HN, Fernstrom JD, Wurtman RJ. Insulin, plasma amino acid imbalance and hepatic coma. *Lancet* 1975; i: 722-724.
- 307. Reichle FA, Owen OE, Golsorkhi M, Kreulen T. Hepatic metabolism in patients with alcoholic ciπhosis. Surgery 1978; 84: 33-36.
- Fisher JE. Porta systemic encephalopathy. In: Wright R, Alberti KGGM, Karran S, Millward-Sadler GH, eds. Liver and Biliary Disease. London: W.B. Saunders 1979, 973-1001.
- 309. Elia M, Farell R, Ilic V, Smith R, Williamson DH. The removal of infused leucine after injury, starvation and other conditions in man. *Clin Sci* 1980; 59: 275-283.
- Roth E, Funovics J, Karner J, et al. In: Kleinberger G, Ferenci P, Riedered P, Thaler H, eds. Muscle amino acid levels in patients with liver diseases. Basel: Karger 1094, 527-537.
- 311. Furst P, Alvestrand A, Bergstrom J, Askanazi J, Elwyn D, Kinney J. In: Johnston IDA, ed. Branched-Chain Amino Acids. Lancaster (England): MTP Press Ltd. 1983, 23-34.

- 312. Marchesini G, Zoli M, Dondi C, Bianchi G, Cirulli M, Pisi E. Anti-catabolic effect of branched-chain amino acid-enriched solutions in patients with liver cirrhosis. *Hepatology* 1982; 2: 420-425.
- 313. O'Keefe SJD, Abraham RR, El-Zayadi A, Davis M, Williams R. Increased plasma tyrosine concentrations in patients with cirrhosis and fulminant hepatic failure associated with increased plasma tyrosine flux and reduced hepatic oxidation capacity. *Gastroenterology* 1981; 81: 1017-1024.
- 314. Nordlinger BM, Fulenwider JT, Ivey GL, et al. Tyrosine metabolism in cirrhosis. J Lab Clin Med 1979; 94: 832-840.
- 315. Faraj BA, Fulenwider JT, Rypins EB, et al. Tyramine kinetics and metabolism in cirrhosis. J Clin Invest 1979; 64: 413-420.
- 316. Jagenburg R, Olsson R, Regardh CG, Rodjer S. Kinetics of intravenous administered Lphenylalanine in patients with cirrhosis of the liver. *Clin Cim Acta* 1977; 78: 453-463.
- 317. Morgan MY, Milsom JP, Sherlock S. Plasma ratio of valine, leucine and isoleucine to phenylalanine and tyrosine in liver disease. Gut 1978; 19: 1068-1073.
- 318. Freund H, James JH, Brenner W, Fischer JE. Plasma amino acid analysis in the differential diagnosis of jaundice. Am J Surg 1980; 139: 142-146.
- 319. Krawitt EL, Clifton JA. Serum concentrations and renal clearance of amino acids in patients with chronic active hepatitis. *Gastroenterology* 1968; 54: 866-871.
- 320. Miller LL. The role of the liver and non-hepatic tissue in the regulation of free amino acid levels in the blood. In: Hloden JT, ed. Amino Acid Pools. Amsterdam: Elsevier 1962, 708-721.
- 321. McMenamy R, Shoemaker W, Richmond J, Elwyn D. Uptake and metabolism of amino acids by the dog liver perfused in situ. Am J Physiol 1962; 202: 407-414.
- 322. Rosen HM, Soeters PB, James JH, Hodgman J, Fischer JE. Influences of exogeneous intake and nitrogen balance on plasma and brain aromatic amino acid concentrations. *Metabolism* 1978; 27: 393-404.
- 323. Swendseid ME, Tuttle SG, Figueroa WS, Mulcare D, Clark AJ, Massey J. Plasma amino acid levels of men fed diets differing in protein content, some observations with valine-deficient diets. J Nutr 1966; 88: 239-248.
- 324. Holt LE, Syndermann SE, Norton PM. The plasma aminogram as affected by protein intake. In: Leathem JH, ed. Protein Nutrition and Free Amino Acid Patterns. New Brunswick (N.J.): Rutgers University Press 1968, 32-39.
- 325. Shaw S, Lieber CS. Plasma amino acid abnormalities in the alcoholic. Respective role of alcohol, nutrition and liver injury. *Gastroenterology* 1978; 74: 677-682.
- 326. Milson JP, Morgan MY, Sherlock S. Factors affecting plasma amino acid concentrations in control subjects. *Metabolism* 1979; 28: 313-319.
- 327. Simmonds WJ, Korman MG, Go VLW, Hofmann AF. Radioimmunoassay of conjugated cholyl bile acids in serum. *Gastroenterology* 1973; 65: 705-711.
- 328. Spenney JG, Johnson BJ, Hirschowitz BI, Mihas AA, Gibson R. An ¹²⁵I radioimmunoassay for primary conjugated bile acids. *Gastroenterology* 1977; 72: 305-311.
- 329. Matern S, Tietjen K, Matern H, Gerok W. Enzyme labelled immunoassay for a bile acid in human serum. In: Pal SB, ed. Enzyme Labelled Immunoassay of Hormones and Drugs. Berlin: Walter DeGruyter & Co. 1978, 457.
- 330. Baqir YA, Ross PE, Borchier IAD. Homogeneous enzyme immunoassay of chenodeoxycholate conjugates in serum *Anal Biochem* 1979; 93: 361-365.

- 331. Korman MG, Hofman AF, Summerskill WHJ. Assessment of activity in chronic liver disease. Serum bile acids compared with conventional tests and histology. N Engl J Med 1974; 290: 1399-1402.
- 332. Annoni G, Barbi G, Donati C. Total serum bile acids sensitivity during therapy of chronic active liver disease. Acta Gastroenterol Belg 1978; 41: 643-652.
- 333. Linnet K, Kelbaek, Bahnsen M. Diagnostic values of fasting and postprandial concentrations in serum of 3 ‡-hydroxy-bile acids and gamma glutarnyl transferase in hepatobiliary disease. Scand J Gastroenterol 1983; 18: 49-56.
- 334. Festi D, Morselli Labate AM, Roda A, *et al.* Diagnostic effectiveness of serum bile acids in liver diseases as evaluated by multivariate statistical methods. *Hepatology* 1983; 3: 707-713.
- 335. Mannes GA, Stellaard F, Paumgartner G. Increased serum bile acids in cirrhosis with normal transaminases. *Digestion* 1982; 25: 217-221.
- 336. Einarsson K, Angelin B, Bjorkhem I, Glauman H. The diagnostic value of fasting individual serum bile acids in anicteric alcoholic liver disease: relation to liver morphology. *Hepatology* 1985; 5: 108-111.
- 337. Rickers H, Christensen M, Amfred T, Dige V, Hess Thaysen E. The diagnostic value of fasting serum total bile acid concentration in patients with suspected liver disease. Scand J Gastroenetrol 1982; 17: 565-570.
- 338. Tobiasson P, Boeryd B. Serum cholic acid and chenodeoxycholic acid conjugates and standard liver function tests in various morphological stages of alcoholic liver disease. *Scand J Gastroenterol* 1980; 15: 657-663.
- 339. Milstein HJ, Bloomer JR, Klatskin G. Serum bile acids in alcoholic liver disease. Comparison with histological features of the disease. *Dig Dis Sci* 1976; 21: 281-285.
- 340. Ohkubo H, Okuda K, Iida S, Ohnishi S, Makino I. Role of portal and splenic vein shunts and impaired hepatic extraction in the elevated serum bile acids in liver cirrhosis. *Gastroenterology* 1983; 86: 514-520.
- Bircher J. Quantitative assessment of deranged hepatic functions: A missed opportunity? Sem Liver Dis 1983; 3: 275-284.
- 342. Monroe PS, Baker AL, Schneider JE, Krager PS, Klein PD, Schoeller D. The aminopyrine breath test and bile acids reflect histologic severity in chronic hepatitis. *Hepatology* 1982; 2: 317-322.
- 343. Paumgartner G. Serum bile acids: physiological determinants and results in liver disease. In: Advances in Diagnostic Disease Mechanisms and Liver Transplantation, AASLD Post-graduate Course. Thorofare (New Jersey): Charles Slack, Inc. 1984, 141-152.
- 344. Sherlock S. Chronic hepatitis and cirrhosis. Hepatology 1984; 4: 255-285.
- 345. Lauterburg BH, Bircher J. Expiratory measurement of maximal aminopyrine demethylation in vivo: effect of phenobarbital, partial hepatectomy, portocaval shunt and bile duct ligation in the rat. J Pharm Exp Ther 1976; 196: 501-509.
- 346. Bircher J, Kupfer A, Gikalov I, Preisig R. Aminopyrine demethylation measured by breath analysis in cirrhosis. *ClinPharmTherapeut* 1976; 20: 484-492.
- 347. Hepner GW, Vessell ES. Assessment of aminopyrine metabolism in man by breath analysis after oral administration of ¹⁴C-aminopyrine. Effects of phenobarbital, disulfiram and portal cirrhosis. N Engl J Med 1974; 291: 1384-1388.
- 348. Baker AL, Kotake AN, Scholler DA. Clinical utility of breath tests for the assessment of hepatic function. Sem Liv Dis 1983; 3: 318-329.
- 349. Branch RA. Drugs as indicators of hepatic function. Hepatology 1982; 2: 97-105.

- Schoeller DA, Baker AL, Monroe PS, Krager PS, Schneider JF. Comparison of different methods of expressing results of the aminopyrine breath test. *Hepatology* 1982; 2: 455-462.
- 351. Pirotte J, El Allaf D. Effect of age and sex on the N-demethylation rate of ¹⁴Caminopyrine, studied by the breath test. *Digestion* 1983; 28: 210-215.
- 352. Galizzi J, Long RG, Billing BH, Sherlock S. Assessment of the (¹⁴C)aminopyrine breath test in liver disease. *Gut* 1978; 19: 40-45.
- 353. Rhodes JC, Aarons LJ, Houston JB. Interpretation of CO² exhalation rate data from demethylation of aminopyrine and its metabolite monomethylamino anti-pyrine. Br J Clin Pharmacol 1982; 14: 409-414.
- Hepner G, Vessell ES. Quantitative assessment of hepatic function by breath analysis after oral administration of ¹⁴C aminopyrine. Ann Int Med 1975; 83: 632-638.
- 355. Saunders JB, Lewis KO, Paton A. Early diagnosis of alcoholic cirrhosis by the aminopyrine breath test. *Gastroenterology* 1980; 79: 112-114.
- 356. Carlisle R, Galambos JT, Dean Warren W. The relationship between conventional liver tests, quantitative function tests and histopathology in cirrhosis. *Dig Dis Sci* 1979; 24: 358-362.
- 357. Pauwels S, Geubel AP, Dive C, Beckers C. Breath ¹⁴CO² after intraveneous administration of ¹⁴C aminopyrine in liver disease. *Dig Dis Sci* 1982; 27: 49-56.
- 358. Baker AL, Krager PS, Glagov S, Schoeller D. Aminopyrine breath test; Prospective comparison with liver histology and liver chemistry tests following jejunoileal bypass performed for refractory obesity. *Dig Dis Sci* 1983; 28: 405-410.
- 359. Hepner GW, Vessell ES. Aminopyrine metabolism in the presence of hyperbilirubinemia due to cholestasis or hepatocellular disease. Combined use of laboratory tests to study disease-induced alterations in drug disposition. *Clin Pharmacol Ther* 1977; 21: 620-626.
- Burnstein AV, Galambos JT. [¹⁴C] aminopyrine breath test in chronic liver disease. Dig Dis Sci 1981; 26: 1078-1083.
- 361. Gill RA, Goodman MW, Golfus GR, Onstad GR, Bubrick MP. Aminopyrine breath test predicts surgical risk for patients with liver disease. *Ann Surg* 1983; 198: 701-704.
- 361a. Keiding S. Hepatic clearance and liver blood flow. J Hepatol 1987; 4: 393-398.
- 362. See reference 270 above.
- 363. McCullough AJ, Czaja AJ. Relapse following withdrawal in patients with autoimmune chronic active hepatitis. *Hepatology* 1984; 4: 747-748.
- 364. Mistilis SP, Skyring AP, Blackburn CRB. Natural history of chronic active hepatitis. I. Clinical features, course, diagnostic criteria, morbidity, and survival. Aust Ann Med 1968; 17: 214-223.
- 365. Mistilis SP. Natural history of active chronic hepatitis. II. Pathology, pathogenesis, and clinico-pathological correlation. *Aust Ann Med* 1968; 17: 277-288.
- 366. Rojkind M, Giambrone MA, Biempica L. Collagen types in normal and cirrhotic liver. *Gastroenterology* 1979; 76: 710-719.
- 367. Rojkind M, Kershenobich D. Hepatic fibrosis. Clin Gastroenterol 1981; 10: 737-754.
- 368. Risteli L, Risteli J. Noninvasive methods for detection of organ fibrosis. In: M. Rojkind, ed. Focus on connective tissue in health and disease. Boca Ratton: CRC Press Inc. 1988.
- 369. Schuppan D, Dumont JM, Kim KY, Hennings G, Hahn EG. Serum concentration of the aminoterminal procollagen type III peptide in the rat reflects early formation of connective tissue in experimental liver cirrhosis. J Hepatol 1986; 3: 27-37.

- Pérez Tamayo R. Is cirrhosis of the liver experimentally produced by CCl⁴ an adequate model of human cirrhosis? *Hepatology* 1984; 3: 112-120.
- Gay S, Fietzek PP, Remberger K, Eder M, Kühn K. Liver cirrhosis: immunofluorescence and biochemical studies demonstrate two types of collagen. *Klin Wochenschr* 1975; 53: 205-208.
- Wick G, Brunner H, Penner E, Timpl R. The diagnostic application of specific antiprocollagen sera. II. Analysis of liver biopsies. Int Arch Allergy Appl Immunol 1978; 56: 316-324.
- 373. Bentsen KD, Boesby S, Kirkegaard P et al. Is the aminoterminal propeptide of type III procollagen degraded in the liver? A study of type III procollagen peptide in serum during liver transplantation in pigs. J Hepatol 1988; 6: 144-150.
- 374. Bentsen KD, Hörslev-Petersen K, Junker P, Juhl E, Lorenzen I. The Copenhagen Hepatitis Acuta Programme. Serum aminoterminal procollagen type III peptide in acute viral hepatitis. A long-term follow-up study. *Liver* 1987; 7: 96-105.
- 375. Savolainen E-R, Goldberg B, Leo MA, Velez M, Lieber CS. Diagnostic value of serum procollagen peptide measurements in alcoholic liver disease. *Alcoholism: Clin Exp Res* 1984; 8: 384-389.
- 376. Niemelä O, Risteli L, Sotaniemi EA, Risteli J. Aminoterminal propeptide of type III procollagen in serum in alcoholic liver disease. *Gastroenterology* 1983; 85: 254-259.
- 377. Nouchi T, Worner T, Sato S, Lieber CS. Fab-radioimmunoassay for serum procollagen-III-peptide as a marker of fibrosis in alcoholics (Abstract). *Hepatology* 1985; 5: 1048.
- 378. Torres-Salinas M, Parés A, Caballería J, Jiménez W, Heredia D, Bruguera M, Rodés J. Serum procollagen type III peptide as a marker of hepatic fibrogenesis in alcoholic hepatitis. *Gastroenterology* 1986; 90: 1241-1246.
- 379. Tanaka Y, Minato Y, Hasamura Y, Takeuchi J. Evaluation of hepatic fibrosis by serum proline and amino-terminal type III procollagen peptide levels in alcoholic patients. *D ig Dis Sci* 1986; 31: 712-717.
- 380. Weigand K, Zaugg P-E, Frei A, Zimmerman A. Long-term follow-up of serum Nterminal propeptide of collagen type III levels in patients with chronic liver disease. *Hepatology* 1984; 4: 835-838.
- Trinchet JC, Hartmannn DJ, Pateron D, et al. Could seric C1 and PIIIP take place of liver biopsy in the follow-up of patients with chronic hepatitis? (Abstract). J Hepatol 1986; 3: S82.
- 382. Raedsch R, Stiehl A, Waldherr R, et al. Procollagen-type III-peptide in chronic persistent and chronic active hepatitis and in cirrhosis of the liver and their diagnostic value. Z Gastroenterol 1982; 20: 738-743.
- 383. Ackermann W, Pott G, Voss B, Müller K-M, Gerlach U. Serum concentration of procollagen-III-peptide in comparison with the serum activity N-acetyl-ß-glucosaminidase for diagnosis of the activity of liver fibrosis in patients with chronic active liver diseases. *Clin Chim Acta* 1981; 112: 365-369.
- 384. Trivedi P, Cheeseman P, Portmann B, et al. Variation in serum type III procollagen peptide with age in healthy subjects and its comparative value in the assessment of disease activity in children and adults with chronic active hepatitis. Eur J Clin Invest 1985; 15: 69-74.
- 385. Annoni G, Cargnel A, Colombo M, Hahn EG. Persistent elevation of the aminoterminal peptide of procollagen type III in serum of patients with acute viral hepatitis distinguishes chronic active hepatitis from resolving or chronic persistent hepatitis. J Hepatol 1986; 2: 379-388.

- Weiner FR, Czaja MJ, Glambrone M-A, Takahashi S, Zern MA. Transcriptional and posttranscriptional effects of dexamethasone on an in vivo model of fibrinogenesis (Abstract). *Hepatology* 1985; 5: 1013.
- 387. Annoni G, Cargnel A,Colombo M,Hahn EG. Serum procollagen in the development of chronic viral liver disease:diagnostic value for chronic active hepatitis and response to immunosuppressive therapy (Abstract). *Liver* 1982; 2: 298.
- 388. Ballardini,G,Faccani,A,Bianchi,FB,Fallani,M,Patrono,D,Capelli,M,Pisi,E. Steroid treatment lowers hepatic fibroplasia, as explored by serum aminoterminal procollagen 3 peptide, in chronic liver disease *Liver* 1984; 4: 348-352.
- Zaugg,P-Y,Weigand,K,Zimmermann,A,Preisig,R. Procollagen peptide levels in chronic liver disease: comparison of antibody and Fab-RIA for assessment of liver fibrosis (Abstract). *Hepatology* 1985; 5: 973.
- 390. Frei,A,Zimmermann,A,Weigand,K. The N-terminal propeptide of collagen type 3 in serum reflects activity and degree of fibrosis in patients with chronic liver disease. *Hepatology* 1984; 4: 830-834.
- McCullough AJ, Stassen WN, Wiesner RH, Czaja AJ. Serum type III procollagen peptide concentrations in severe chronic active hepatitis: relationship to cirrhosis and disease activity. *Hepatology* 1987; 7: 49-54.
- 392. McCullough,AJ,Stassen,WN,Wiesner,RH,Czaja,AJ. Serial determinations of the aminoterminal peptide of type III procollagen in severe chronic active hepatitis. J Lab Clin Med 1987; 109: 55-61.
- 393. Jensen DM, McFarlane IG, Portmann B, Eddleston ALWF, Williams R. Detection of antibodies directed against a liver-specific membrane lipoprotein in patients with acute and chronic active hepatitis. N Engl J Med 1978; 299: 1-7.
- 394. McFarlane IG, Williams R. Liver membrane antibodies. J Hepatol 1985; 1: 313-319.
- 395. Meliconi R, Miglio F, Stancari MV, Baraldini M, Stefanini GF, Gasbarrini G. Hepatocyte membrane-bound IgG and circulating liver-specific autoantibodies in chronic liver disease: relation to hepatitis B virus serum markers and liver histology. *Hepatology* 1983; 3: 155-161.
- 396. Tsantoulas D, Perperas A, Portmann B, Eddleston ALWF, Williams R. Antibodies to human liver membrane lipoprotein (LSP) in primary biliary cirrhosis. *Gut* 1980; 21: 557-560.
- Lobo-Yeo A, Mieli-Vergani G, Mowat AP, Vergani D. Soluble interleukin 2 receptors in serum of children with autoimmune chronic active hepatitis (Abstract). J Hepatol 1987; 5: S40.

CHAPTER 1

398. See reference 29 above.

- 399. See reference 31 above.
- 400. See reference 40 above.
- 401. See reference 284 above.
- 402. See reference 50 above.

- 403. See reference 86 above.
- 404. See reference 143 above.
- 404a. See reference 279 above.
- 404b. See reference 280 above.
- 404c. See reference 281 above.
- 405. Lindh G, Weiland O, Glauman H. The application of a numerical scoring system for evaluating the histological outcome in patients with chronic hepatitis B followed in long term. *Hepatology* 1988; 8: 98-103.
- 406. See reference 270 above.
- 407. See reference 363 above.

- 408. See reference 368 above.
- 409. Fessler JH, Fessler LI. Biosynthesis of collagen. Ann Rev Biochem 1978; 47: 129-162.
- 410. Fleischmajer R, Timpl R, Tuderman L, Raisher L, Wiestner M, Perlish JS, Graves PN. Ultrastructural identification of extension aminopropeptides of type I and III collagens in human skin. *Proc Natl Acad Sci USA* 1981; 78: 7360-7364.
- 411. Risteli L, Risteli J. Radioimmunoassays for monitoring connective tissue metabolism. *Rheumatology* 1986; 10: 216-245.
- 412. Risteli L, Risteli J. Analysis of extracellular proteins in biological fluids. Meth Enzymol 1987; 145: 391-411.
- 413. Rohde H, Langer I, Krieg T, Timpl R. Serum and urine analysis of the aminoterminal procollagen type III by radioimmunoassay with antibody Fab fragments. *Coll Relat Res* 1983; 3: 371-379.
- 414. Niemelä O. Radioimmunoassays for type III procollagen amino-terminal peptides in humans. Clin Chem 1985; 1301-1304.
- 415. Risteli J, Niemi S, Trivedi P, Mäentausta O, Mowat AP, Risteli L. Rapid equilibrium radioimmunoassay for the amino-terminal propeptide of human type III procollagen. *Clin Chem* 1988; 34: 715-718.
- 416. Niemelä O, Risteli L, Sotaniemi EA, *et al.* Heterogeneity of the antigens related to the aminoterminal propeptide of type III procollagen in the serum. *Clin Chim Acta* 1982; 124: 39-44.
- 417. Rohde H, Vargas L, Hahn E, Kalbfleish H, Bruguera M, Timpl R. Radioimmunoassay for type III procollagen and its application to human liver disease. Eur J Clin Invest 1979; 9: 451-457.
- 418. Hahn EG. Blood analysis for liver fibrosis. J Hepatol 1985; 1: 67-73.
- 419. van Hoek B, Grond J, Gips CH. Numerical versus conventional histological scoring of activity in 'autoimmune' CAH (Abstract). *Hepatology* 1989 (in press).
- 420. See reference 284 above.
- 421. See reference 392 above.
- 422. See reference 390 above.
- 423. Bolarin DM, Savolainen E-R, Kivirikko KI. Three serum markers of collagen biosynthesis in Nigerians with cirrhosis and various infectious diseases. Eur J Clin Invest 1984; 14: 90-5.

- 424. See reference 374 above.
- 425. See reference 375 above.
- 426. See reference 376 above.
- 427. See reference 377 above.
- 428. See reference 378 above.
- 429. See reference 379 above.
- 430. See reference 380 above.
- 431. See reference 381 above.
- 432. See reference 382 above.
- 433. See reference 383 above.
- 434. See reference 384 above.
- 435. See reference 385 above.
- 436. See reference 386 above.
- 437. See reference 387 above.
- 438. See reference 388 above.
- 439. See reference 389 above.
- 440. See reference 392 above.
- 441. Rojkind M. The blue glass and the predictive value of serum amino-terminal propeptide of type III procollagen as a marker of liver fibrosis. *Hepatology* 1984; 4: 977-978.
- 442. See reference 132 above.

- 443. See reference 165 above.
- 444. McFarlane IG, Wojcicka BM, Zucker GM, Eddleston ALWF, Williams R. Purification and characterization of human liver-specific membrane lipoprotein (LSP). *Clin Exp Immunol* 1977; 27: 381-390.
- 445. See reference 393 above.
- 446. Hunter WM, Greenwood FC. Preparation of iodine-131 labelled human growth hormone of high specific activity. *Nature* 1962; 194: 495-496.
- 447. David GS, Reisfeld RA. Protein iodination with solid state lactoperoxidase. Biochemistry 1974; 13: 1014-1021.
- 448. Markwell MAK. A new solid-state reagent to iodinate proteins. Annal Biochem 1982; 125: 427-432.
- 449. See reference 448 above.
- 450. See reference 269 above.
- 451. Manns M, Arnold W, Meyer zum Büschenfelde K-H, Nagai S, Hoffmann H. Studies on anti LSP autoantibodies in acute and chronic non-B hepatitis- Evidence for the lack of anti LSP in non-A,non-B (NANB) viral hepatitis. *Klin Wochenschr* 1981; 59: 685-689.
- 452. McFarlane IG, Tolley P, Major G, McFarlane BM, Williams R. Development of a microenzyme-linked immunosorbent assay for antibodies against liver-specific membrane lipoprotein. J Immunol Meth 1983; 215-225.

- 453. See reference 162 above.
- 454. McFarlane BM, McSorley CG, McFarlane IG, Williams R. A radioimmunoassay for detection of circulating antibodies reacting with the hepatic asialoglycoprotein receptor protein. J Immunol Meth 1985; 77: 219-228.
- 455. See reference 163 above.

- 456. See reference 53 above.
- 457. See reference 54 above.
- 458. See reference 55 above.
- 459. See reference 56 above.
- 460. See reference 57 above.
- 461. See reference 8 above.
- 462. See reference 50 above.
- 463. See reference 58 above.
- 464. See reference 59 above.
- 465. See reference 60 above.
- 466. See reference 61 above.
- 467. See reference 48 above.
- 468. See reference 30 above.
- 469. See reference 51 above.
- 470. See reference 87 above.
- 471. Kaplan EL, Meier P. Nonparametric estimation from incomplete observations. J Am Stat Assoc 1958; 53: 457-481.
- 472. Cox DR. Regression models and life-tables. J R Stat Soc [B] 1972; 34: 187-202.
- 473. See reference 62 above.
- 474. See reference 63 above.
- 475. See reference 64 above.
- 476. See reference 72 above.
- 477. See reference 73 above.
- 478. See reference 74 above.
- 479. See reference 75 above.
- 480. See reference 76 above.
- 481. See reference 66 above.
- 482. See reference 67 above.
- 483. See reference 77 above.
- 484. See reference 78 above.

- 485. See reference 68 above.
- 486. See reference 69 above.
- 487. See reference 70 above.
- 488. See reference 71 above.
- 489. See reference 110 above.
- 490. See reference 142 above.
- 491. See reference 143 above.
- 492. See reference 144 above.
- 493. See reference 283 above.
- 494. Bruguera M, Bordas JM, Mas P, Rodés J. A comparison of the accuracy of peritoneoscopy and liver biopsy in the diagnosis of cirrhosis. Gut 1974; 15: 799-800.
- 495. See reference 145 above.
- 496. See reference 146 above.
- 497. Christensen E. Individual therapy-dependent prognosis based on data from controlled clinical trials in chronic liver disease. *Dan Med Bull* 1988; 35: 167-182.

CHAPTER 1

§7.1

- 498. See reference 86 above.
- 499. See reference 50 above.
- 500. See reference 57 above.
- 501. See reference 48 above.
- 502. See reference 142 above.
- 503. See reference 143 above.
- 504. See reference 263 above.
- 505. Stellon AJ, Keating JJ, Johnson PJ, McFarlane IG, Williams R. Maintenance of remission in autoimmune chronic active hepatitis with azathioprine after corticosteroid withdrawal. *Hepatology* 1988; 8: 781-784.
- 506. See reference 268 above.
- 507. See reference 280 above.
- 508. See reference 283 above.
- 509. See reference 281 above.

§7.2

510. Koppel MH, Coburn JW, Mirns MM, *et al.* Transplantation of cadaveric kidneys from patients with hepatorenal syndrome. Evidence for the functional nature of renal failure in advanced liver disease. *N Engl J Med* 1969; 280: 1367-1371.

- 511. Iwatsuki S, Popovtzer MM, Corman JL, et al. Recovery from hepatorenal syndrome after orthotopic liver transplantation. N Engl J Med 1973; 289: 1155-1159.
- 512. Wood RP, Ellis D, Starzl TE. The reversal of the hepatorenal syndrome in four patients following successful orthotopic liver transplantation. *Ann Surg* 1987; 205: 415-419.
- 513. van Hoek B, Huizenga JR, Grond J, Gips CH. Repair and normalization of hepatic protein synthesis despite presence of cirrhosis in 'autoimmune' chronic active hepatitis. (Abstract) *Hepatology* 1989 (in press).
- 514. Arroyo V, Rodés J, Gutierrez-Lizárraga MA. Prognostic value of spontaneous hyponatremia in cirrhosis with ascites. *Dig Dis* 1976; 21: 249-256.
- 515. de Jong GMTh, Huizenga JR, Gips CH. Evaluation of routine data on sodium- and water homeostasis, and renal function in 247 patients with liver disease: support for the new defenition of the hepatorenal syndrome. In: de Jong GMTH. Sodium and water homeostasis in liver impairment. Thesis. State University Groningen, The Netherlands 1988.
- 516. See reference 50 above.
- 517. See reference 419 above.
- 518. See reference 284 above.
- 519. Arroyo V, Bernardi M, Epstein M, Henriksen JH, Schrier RW, Rodés J. Pathophysiology of ascites and functional renal failure in cirrhosis. *J Hepatol* 1988; 6: 239-257.
- 520. Schrier RW. Pathogenesis of sodium and water retention in high-output and low-output cardiac failure, nephrotic syndrome, cirrhosis, and pregnancy (Second of two parts). N Engl J Med 1988; 319; 1127-1134.
- 521. Epstein M, Weitzman RE, Preston S, *et al.* The relationship between plasma AVP and renal water handling in decompensated cirrhosis. *Miner Electrolyte Metab* 1984; 10: 155-165.
- 522. Reznick RK, Langer B, Taylor BR, et al. Hyponatremia and arginine vasopressin secretion in patients with refractory ascites undergoing peritovenous shunting. *Gastroenterology* 1983; 84: 713-718.
- 523. Bichet D, Szatalowicz V, Chaimovitz C, Schrier RW. Role of vasopressin in abnormal water excretion in cirrhotic patients. *Ann Intern Med* 1982; 96: 413-417.
- 524. Pérez Ayuso RM, Arroyo V, Camps J, *et al.* Evidence that renal prostaglandins are involved in renal water metabolism in cirrhosis. *Kidney Int* 1984; 26: 72-80.
- 525. Arroyo V, Planas R, Gaya J, et al. Sympathetic nervous activity, renin-angiotensin system, and renal excretion of prostaglandin E² in cirrhosis. Relationship to functional renal failure and sodium and water excretion. Eur J Clin Invest 1983; 13: 271-287.
- 526. See reference 146 above.
- 527. van Hoek B, Glade G, Fidler V, Gips CH. Long-term prognosis in chronic active hepatitis (Abstract). *Hepatology* 1989 (in press).

§7.3

- 528. See reference 294 above.
- 529. Czaja AJ. Treatment strategies in chronic active hepatitis. In: Czaja AJ, Dickson ER, eds. Chronic active hepatitis- The Mayo Clinic experience. New York: Marcel Dekker 1986, 247-267.
- 530. See reference 50 above.
- 531. See reference 419 above.

- 532. See reference 284 above.
- 533. Wadenvik H, Kutti J. The spleen and pooling of blood cells. Eur J Haematol 1988; 41: 1-5.
- Lasky LC, Ascensao J, McCullough J, Zanjani ED. Steroid modulation of naturally occurring diurnal variation in circulating pluripotential haematopoietic cells (CFU-GEMM). Br J Haematol 1983; 55: 615-622.
- 535. Heal JM, Abboud CN, Brightman A, Rubery P, Brennan JK, Nusbacher J. The effect of steroids and filtration leukapheresis on circulating hematopoietic progenitor cells. Int J Cell Cloning 1983; 1: 464-477.
- 536. Schuyler M, Gerblich A, Urda G. Prednisone and T-cell subpopulations. Ann Intern Med 1984; 144: 973-975.
- 537. See reference 513 above.
- 537a. De Clerck YA, Ettenger RB, Ortega JA, et al. Macrocytosis and pure RBC anemia caused by azathioprine. Am J Dis Child 1980; 134: 377-379.
- 537b. Bacon BR, Treuhaft WH, Goodman AM. Azathioprine-induced pancytopenia. Arc h Intern Med 1981; 141: 223-226.
- 537c. Maddock JL, Lennard L, Amess J, et al. Azathioprine and severe bone marrow depression. Lancet 1986; i: 156.
- 537d. Verhelst JA, Van der Enden E, Mathys R. Rapidly evolving azthioprine induced pancytopenia. J Rheumatol 1987; 14: 862.
- 538. See reference 86 above.

- 539. See reference 50 above.
- 540. See reference 57 above.
- 541. See reference 48 above.
- 542. See reference 142 above.
- 543. See reference 143 above.
- 544. See reference 263 above.
- 545. See reference 505 above.
- 546. See reference 31 above.
- 547. See reference 40 above.
- 548. See reference 284 above.
- 549. See reference 419 above.
- 550. See reference 79 above.
- 551. See reference 211 above.
- 552. See reference 44 above.
- 553. See reference 270 above.
- 554. See reference 265 above.
- 555. See reference 193 above.

- 557. See reference 268 above.
- 558. See reference 280 above.
- 559. See reference 283 above.
- 560. See reference 281 above.
- 561. See reference 88 above.

- 562. See reference 13 above.
- 563. See reference 59 above.
- 564. Page AR, Good RA, Pollara B. Long-term results of therapy in patients with chronic liver disease associated with hypergammaglobulinemia. *Am J Med* 1970; 47: 765-774.
- 565. Rakela J, Czaja AJ. Clinical, biochemical, and histologic features of HBsAg-negative chronic active hepatitis. In Czaja AJ, Dickson ER (eds.) Chronic active hepatitis. The Mayo Clinic experience. New York: Marcel Dekker, Inc., 1986, 69-82.
- 566. See reference 294 above.
- 567. See reference 266 above.
- 568. See reference 97 above.
- 569. Yamamoto K, Morito F, I N, Setoguchi Y, Fujii S, Kariya T, Sakai T. Characterization of serum cholinesterase in familial hyper-cholesterasemia associated with an isozyme variant band. *Gastroenterol Jpn* 1987; 22: 187-193.
- 570. Svensmark O. Precursors of serum cholinesterase in human liver. Acta Physiol Scand 1963; 59: Suppl. 148.
- 571. Weber H. Rasche und einfache Ultramikromethode zur Bestimmung der Serumcholinesterase. Dtsch Med Wschr 1966; 91: 1927.
- 572. Wiesman M. Klinische Bedeutung der Plasmacholinesterase. Schweiz Med Wschr 1967; 97: 422.
- 573. Silk E, King J, Whittaker M. Assay of cholinesterase in clinical chemistry. Ann Clin Biochem 1979; 16: 57-75.
- 574. Fintelman V., Lindner H. Diagnostische Bedeutung der Serum-Cholinesterase bei Lebererkrankungen. Dtsch Med Wschr 1970; 9: 469-470.
- 575. McArdle B. The serum choline esterase in jaundice and diseases of the liver. Quart J Med 1940; 9: 107-119.
- 576. Hunt AH, Lehmann H. Serum albumin, pseudocholinesterase and transaminases in the assessment of liver function before and after shunt operations. *Gut* 1960; i: 303-311.
- 577. Gentz HO, Schlicht J, Wiederholt W. Pseudocholinesterase bei hepatischen und nichthepatischen Erkrankungen. *Med Klinik* 1978; 73: 1422.
- 578. Gips CH. Liver transplantation in Groningen- Long-term survivors in the first cohort. Neth J Med 1986; 29: 357-358.
- 579. Adolph L. Diagnostische Bedeutung der Cholinesterase-Bestimmung im menschlicher Serum. *Münch Med Wsch* 1979; 121: 1527-1530.
- 580. Evans DB, Lehmann H. Pseudocholinesterase activity in liver transplantation. Lancet 1971; i: 1040-1044.
- 581. Schlichting P, Christensen E, Andersen PK, et al. Prognostic factors in cirrhosis identified by Cox's regression model. *Hepatology* 1983; 3: 889-895.

- Tygstrup N, Christensen E. Prognostic estimates in cirrhosis. In:Tygstrup N, Orlandi F, eds. Cirrhosis of the liver: Methods and fields of research. Amsterdam: Elsevier 1987, 385-401.
- 583. See reference 497 above.
- 584. See reference 271 above.
- 585. Hallen A, Nilsson IM. Coagulation studies in liver disease. Thrombosis et Diathesis Haemorrhagica 1964; 11: 51-63.
- 586. Abilsgaard U, Fagerhol MK, Egeberg O. Comparison of progressive antithrombin and the concentration of three thrombin inhibitors in human plasma. Scand J Clin Lab Invest 1970; 26: 349-354.
- 587. Chan V, Lai CL, Chan TK. Metabolism of antithrombin III in cirrhosis and carcinoma of the liver. *Clin Sci* 1981; 60: 681-688.
- 588. Knot EAR, ten Cate JW, Drijfhout HR, Kahlé LH, Tytgat GN. Antithrombin III in patients with liver disease. *J Clin Pathol* 1984; 37: 523-530.
- 589. Sinclair TS, Booth NA, Penman SM, Brunt PW, Mowat NAG, Bennett NB. Protease inhibitors in liver disease. Scand J Gastroenterol 1988; 23: 620-624.
- 590. Schipper HG, ten Cate JW. Antithrombin III transfusion in patients with hepatic cirrhosis. Br J Haematol 1982; 52: 25-33.
- 591. See reference 364 above.
- 592. See reference 50 above.
- 593. See reference 85 above.
- 594. See reference 142 above.
- 595. See reference 61 above.
- 596. See reference 60 above.
- 597. See reference 144 above.
- 598. See reference 331 above.
- 599. See reference 342 above.
- 600. See reference 341 above.
- 601. See reference 419 above.
- 602. See reference 284 above.
- 603. Bick RL, Kovacs I, Fekeke LF. A new two-stage functional assay for antithrombin III (heparin cofactor): Clinical and laboratory evaluation. *Thromb Res* 1976; 8: 745.
- 604. Ellman GL, Courtney KD, Andres V, Featherstone RM. A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochem. Pharmacol* 1961; 7: 88-95.
- 605. Huizenga JR, Van der Belt K, Gips CH. The effect of storage at different temperatures on cholinesterase activity in human serum. J Clin Chem Clin Biochem 1985; 23: 283-285.
- 606. Ashihara Y, Kasahara Y, Sugiyama M, Harada T. Rate assay for determination of serum pseudo-cholinesterase activity. *J Biochem* 1983; 94: 11-15.
- 607. Huizenga JR, Gips CH. Evaluation of the UV-340 spectrophotometric determination for pseudocholinesterase activity (EC 3.1.1.8) in human serum. J Clin Chem Clin Biochem 1987; 25: 161-165.

- 607a. Jörgensen K, Sörensen P, Freund L. Effect of glucocorticosteroids on some coagulation tests. Acta Haematol 1982; 68: 39-42.
- 608. Moshage HJ, Janssen JAM, Franssen JH, Hafkenscheid JCM, Yap SH. Study of the molecular mechanism of decreased liver synthesis of albumin in inflammation. J Clin Invest 1987; 79: 1635-1641.
- 609. Dinarello CA. Interleukin-1 and the pathogenesis of the acute-phase response. N Engl J Med 1984; 22:1413-1418.
- 610. Ramadori G, Van Damme J, Rieder H, Meyer zum Büschenfelde K-H, Interleukin 6, the third mediator of acute-phase reaction, modulates hepatic protein synthesis in human and mouse. Comparison with interleukin 1 β and tumor necrosis factor- α Eur J Immunol 1988; 1259-1264.
- 611. Castell JV, Gómez-Lechón MJ, David M, Hirano T, Kishimoto T, Heinrich PC. Recombinant human interleukin-6 (IL-6/BSF-2/HSF) regulates the synthesis of acute phase proteins in human hepatocytes. *FEBS Letters* 1988; 232: 347-350.
- 612. Beutler B, Cerami A. Cachectin, More than tumor necrosis factor. N Engl J Med 1987; 316: 379.
- 613. Prieur A-M, Kaufmann M-T, Griscelli C, Dayer J-M. Specific interleukin-1 inhibitor in serum and urine of children with systemic juvenile chronic arthritis. *Lancet* 1987; ii: 1240-1242.
- Westacott CI, Whicher JT, Hutton CW, Dieppe PA. Increased spontaneous production of interleukin-1 together with inhibitory activity in systemic sclerosis. *Clin Sci* 1988; 75: 561-567.
- 615. Ganapathi MK, Schultz D, Mackiewicz A, Samols D, Hu S-I, Brabenec A, Macintyre SS, Kushner I. Heterogeneous nature of the acute phase response. Differential regulation of human serum amyloid A, C-reactive protein, and other acute phase proteins by cytokines in Hep 3B cells. *J Immunol* 1988; 141: 564-569.
- 616. Lee SW, Tsou A-P, Chan H, Thomas J, Petrie K, Eugui EM, Allison AC. Glucocorticoids selectively inhibit the transcription of the interleukin 1β gene and decrease the stability of interleukin 1β mRNA. Proc Natl Acad Sci USA 1988; 85: 1204-1208.
- 617. Robertson GS. Serum protein and cholinesterase changes in association with contraceptive pills. *Lancet* 1967; i: 232-235.
- 618. Sidell FR, Kaminskis A. Influence of age, sex, and oral contraceptives on human blood cholinesterase activity. *Clin Chem* 1975; 21: 1393-1395.
- 619. Fagerhol MK, Abildgaard U, Bergsjö P, Jacobsen JH. Oral contraceptives and low antithrombin III concentration. *Lancet* 1970; i: 1175.
- 620. Rothschild MA, Oratz M, Schreiber SS. Serum albumin. Hepatology 1988; 8: 385-401.
- 621. Rothschild MA, Oratz M, Zimmon D, et al. Albumin synthesis in cirrhotic subjects with ascites studied with carbonate-C14. J Clin Invest 1969; 48: 344-350.
- 622. Rothschild MA, Oratz M, Schreiber SS. Alcohol, amino acids, and albumin synthesis. *Gastroenterology* 1974; 67: 1200-1213.
- 623. Bismuth H, Ericzon BG, Rolles K, Castaing D, Otte JB, Ringe B, Sloof M. Hepatic transplantation in Europe. First report of the European Liver Transplant RegistryLancet 1987; ii: 674-676.
- 624. See reference 276 above.

- 625. Pepys MB, Baltz ML. Acute phase proteins with special reference to C-reactive protein and related proteins (pentaxins) and serum amyloid A protein. Adv Immunol 1983; 34: 141-212.
- 626. Dowton SB, Colten HR. Acute phase reactants in inflammation and infection. Seminars in Hematology 1988; 25: 84-90.
- 627. Kushner I, Feldmann G. Control of the acute phase response. Demonstration of Creactive protein synthesis and secretion by hepatocytes during acute inflammation in the rabbit. J Exp Med 1978; 148: 466-477.
- 628. Ikuta T, Okubo H, Ishibashi H, Okumura Y, Hayashida K. Human Lymphocytes synthesize C-reactive protein. *Inflammation* 1986; 10: 223-232.
- 629. Robey FA, Ohura K, Futaki S, Fujii N, Yajima H, Goldman N, Jones KD, Wahl S. Proteolysis of human C-reactive protein produces peptides with potent immunomudulating activity. J Biol Chem 1987; 262: 7053-7057.
- 630. Shephard EG, Anderson R, Beer M, Jansen van Rensburg CE, De Beer FC. Neutrophil lysosomal degradation of human CRP: CRP-derived peptides modulate neutrophil function. *Clin Exp Immunol* 1988; 73: 139-145.
- Dowton SB, Colten HR. Sites and regulation of biosynthesis of SAA. In: Marrink J, van Rijswijk MH, eds. Amyloidosis. Dordrecht, Boston, Lancester: Martinus Nijhoff, 1986; 107-113.
- 632. Ramadori G, Sipe JD, Colten HR. Expression and regulation of the murine serum amyloid A (SAA) gene in extrahepatic sites. *J Immunol* 1985; 135: 3645-3647.
- 633. Sipe J, Ramadori G. Sites of SAA/AA synthesis. In: Marrink J, van Rijswijk MH, eds. *Amyloidosis*. Dordrecht, Boston, Lancester: Martinus Nijhoff, 1986; 319-327.
- 634. Meek RL, Benditt EP. Amyloid A gene family expression in different mouse tissues. J Exp Med 1986; 164: 2006-2017.
- 635. Aldo-Benson MA, Benson MD. A A suppression of immune response in vitro: evidence for an effect on T cell-macrophage interaction. J Immunol 1982; 128: 2390-2392.
- 636. Shepherd EG, De Beer FC, De Beer MC, Jeenah MS, Coetzee GA, Van der Westhuyzen DR. Neutrophil association and degradation of normal and acute-phase high-density lipoprotein 3. *Biochem J* 1987; 248: 919-926.
- 637. Nel AE, De Beer MC, Shephard EG, Strachan AF, Vandenplas ML, De Beer FC. Phosphorilation of human serum amyloid A protein by protein kinase C. *Biochem J* 1988; 255: 29-34.
- 638. Hutton CW, Collins AJ, Chambers RE, Whicher J, Dieppe PA. Systemic response to local urate crystal induced inflammation in man: a possible model to study the acute phase response. *Ann Rheum Dis* 1985; 44: 533-536.
- 639. Marhaug G, Hårklau L, Olsen B, Husby G, Husebekk A, Wang H. Serum amyloid A protein in acute myocardial infarction. *Acta Med Scand* 1986; 220: 303-306.
- 640. Maury CPJ. Comparative study of serum amyloid A protein and C-reactive protein in disease. *Clin Sci* 1985; 68: 233-238.
- 641. Chambers RE, MacFarlane DG, Whicher JT, Dieppe PA. Serum amyloid-A protein concentration in rheumatoid arthritis and its role in monitoring disease activity. Ann Rheum Dis 1983; 42: 665-667.
- 642. Grindulis KA, Scott DL, Robinson MW, Bacon PA, McConkey B. Serum amyloid A protein during the treatment of rheumatoid arthritis with second-line drugs. Br J Rheumatol 1985; 24: 158-163.

- 643. Chambers RE, Stross P, Barry RE, Whicher JT. Serum amyloid A protein compared with C-reactive protein, alpha 1-antichymotrypsin and alpha 1-acid glycoprotein as a monitor of inflammatory bowel disease. Eur J Clin Invest 1987; 17: 460-467.
- 644. Murray-Lyon JM, Williams R. Quantitive immunoelectro- phoresis of plasma proteins in acute viral hepatitis, extrahepatic biliary obstruction, primary biliary cirrhosis and idiopathic haemochromatosis. *Clin Chim Acta* 1974; 51:303-308.
- 645. Carlson J, Eriksson S. α_1 -Antitrypsin and other acute phase reactants in liver disease. Acta Med Scand 1980; 207:79-83.
- 646. Imanishi T. Clinical and experimental studies on the profiles of serum proteins in acute hepatic injury *Gastroenterol Jpn* 1981; 16:493-505.
- 647. van Gool J. Profiles of acute-phase reactants and clinical significance of α_2 -macroglobulin in acute hepatitis B. Inflammation 1983; 7: 277-289.
- 648. Ricciardi R, Crisafi A. The behaviour of α_1 -antitrypsin, haptoglobulin, ceruloplasmin, α_2 -macroglobulin and transferrin in chronic hepatitis. G Mal Infett Parawitt 1978; 30:562-565.
- 649. Meliconi R, Parracino O, Facchini A, Moreselli-Labate AM, Flavia Bortolotti, Trembolacta F, Martuzzi M, Miglio F, Gasbarrini G. Acute phase proteins in chronic and malignant liver diseases. *Liver* 1988; 8; 65-74.
- 650. Pepys MB, Dash AC, Markham RE, Thomas HC, Williams BD, Petrie A. Comparative clinical study of protein SAP (amyloid P component) and C-reactive protein in serum. *Clin Exp Immunol* 1978; 32: 119-124.
- 651. Raynes JG, Cooper EH. Comparison of serum amyloid A protein and C-reactive protein concentrations in cancer and non-malignant disease. *J Clin Pathol* 1983; 36: 798-803.
- 652. See reference 580 above.
- 653. Schmidt E, Schmidt FW. Enzym-Diagnostik von Leber-Erkrankungen in der Praxis. Diagnostik 1977; 10: 348.
- 654. See reference 577 above.
- 655. See reference 579 above.
- 656. van Hoek B, Grond J, Huizenga JR, van Zanten A, Gips CH. Parameters of protein synthesis, fibrinogenesis, fibrosis, and inflammatory activity in standardized treatment of autoimmune chronic active hepatitis (Abstract). J Hepatology 1987; 5(suppl 1): S216.
- 657. See reference 50 above.
- 658. See reference 419 above.
- 659. See reference 284 above.
- 660. Hazenberg BPC, Limburg PC, Bijzet J, van Rijswijk MH. SAA versus CRP serum levels in different inflammatory conditions, studied by ELISA using polyclonal anti-AA and monoclonal anti-SAA antibodies. In: Isobe T, Araki S, Uchino F, Kito S, Tsubara E, eds. Amyloid and Amyloidosis. New York: Plenum Press 1988, 229-233.
- 661. See reference 605 above.
- 662. See reference 604 above.
- 663. Janssen S, Limburg PC, Bijzet J, de Jong HJ, Marrink J, van Leeuwen MA, van Rijswijk MH. SAA versus CRP in chronic inflammatory diseases. In: Peeters H, ed. XXXIVth Colloquium Protides of the Biological Fluids. Oxford: Pergamon Press Ltd. 1986; 34: 347-350.

- 664. Pepys MB, Landhamm JG, de Beer F. C-reactive protein in systemic lupus erythematosus. In: Hughes GRV ed. Systemic Lupus Erythematosus. Clinics in rheumatic disease. Eastbourne: W.B. Saunders, 1982:91-103.
- 665. Levo Y, Shalit M, Tur-Kaspa R. Serum amyloid P-component as a marker of liver disease. Am J Gastroenterol 1982; 77:427-430.
- 666. Benson MD, Kleiner E. Synthesis and secretion of serum amyloid protein A (SAA) by hepatocytes in mice treated with casein. J Immunol 1980; 124: 495-499.
- 667. Miura K, Takahashi Y, Shirasawa H. Immunohistochemical detection of serum amyloid A protein in the liver and the kidney after casein injection. Lab Invest 1985; 53: 453-463.
- 668. Shirahama T, Skinner M, Cohen AS. Heterogenous participation of the hepatocyte population in amyloid protein AA synthesis. *Cell Biology International Reports* 1984; 8: 849-856.
- 669. Hoffman JS, Benditt EP. Changes in high density lipoprotein content following endotoxin administration in the mouse. J Biol Chem 1982; 257: 10510-10517.
- 670. Seidel D. Lipoproteins in liver disease. Clin Chem Clin Biochem 1987; 25: 541-551.
- 671. See reference 609 above.
- 672. Sipe JD, Colten HR, Goldberger G, et al. Human serum amyloid A (SAA): biosynthesis and postsynthetic processing of preSAA and structural variants defined by complementary DNA. Biochem 1985; 24: 2931-2916.
- 673. Gauldie J, Richards C, Harnish D, Lansdorp P, Baumann H. Interferon β₂/B-cell stimulatory factor type 2 shares identity with monocyte-derived hepatocyte-stimulating factor and regulates the major acute phase protein response in liver cells. *Proc Natl Acad Sci USA* 1987; 84: 7251-7255.
- 674. See reference 610 above.
- 675. Moshage HJ, Roelofs HMJ, van Pelt JF, Hazenberg BPC, van Leeuwen MA, Limburg PC, Aarden LA, Yap SH. The effect of interleukin-1, interleukin-6 and its interrelationship on the synthesis of serum amyloid A and C-reactive protein in primary cultures of adult human hepatocytes. *Biochemical and biophysical Research Communications* 1988; 155: 112-117.
- 676. See reference 611 above.
- 677. See reference 612 above.
- 678. Moshage HJ, Roelofs HMJ, Hazenberg BPC, van Leeuwen MA, Limburg PC, Aarden LA, Yap SH. The effect of interleukin-1 (IL-1), interleukin -6 (IL-6) and crude monocytic product on the synthesis of C-reactive protein (CRP) and serum amyloid A (SAA) in primary cultures of human hepatocytes (Abstract). J Hepatol 1988; 7 (Suppl.1):S 153.
- 679. See reference 613 above.
- 680. See reference 614 above.
- 681. See reference 616 above.
- 682. See reference 615 above.

- 683. See reference 86 above.
- 684. See reference 50 above.

- 685. See reference 57 above.686. See reference 142 above.
- 687. See reference 143 above.
- 688. See reference 144 above.
- 689. See reference 366 above.
- 690. See reference 367 above.
- 691. See reference 368 above.
- 692. See reference 369 above.
- 693. See reference 370 above.
- 694. See reference 371 above.
- 695. See reference 372 above.
- 696. See reference 373 above.
- 697. See reference 374 above.
- 698. See reference 375 above.
- 699. See reference 376 above.
- 700. See reference 377 above.
- 701. See reference 378 above.
- 702. See reference 379 above.
- 703. See reference 380 above.
- 704. See reference 390 above.
- 705. See reference 382 above.
- 706. See reference 383 above.
- 707. See reference 384 above.
- 708. See reference 385 above.
- 709. See reference 386 above.
- 710. See reference 387 above.
- 711. See reference 388 above.
- 712. See reference 389 above.
- 713. See reference 391 above.
- 714. See reference 392 above.
- 715. See reference 580 above.
- 716. See reference 653 above.
- 717. See reference 577 above.
- 718. See reference 579 above.
- 719. See reference 656 above.
- 719a. See reference 419 above.
- 719b. See reference 284 above.

- 720. See reference 604 above.
- 721. See reference 605 above.
- 722. See reference 415 above.
- 723. See reference 416 above.
- 724. See reference 417 above.
- 725. See reference 413 above.
- 726. See reference 418 above.
- 727. See reference 441 above.
- 728. See reference 414 above.
- 729. See reference 423 above.
- 730. See reference 423 above.
- 731. See reference 442 above.
- 732. Tsutsumi M, Takada A, Takase S, Ooshima A. Connective tissue components in cultured parenchymal and nonparenchymal cells of rat liver. Immunohistochemical studies. *Hepatology* 1988; 58: 88-92.
- 733. Johnson RL, Ziff M. Lymphokine stimulation of collagen accumulation. J Clin Invest 1976; 58: 240-252.
- 734. Wahl SM, Wahl LM, McCarty JB. Lymphocyte-mediated activation of fibroblast proliferation and collagen production. *J Immunol* 1978; 121: 942-946.
- 735. Postlethwaite AE, Kang AH. Induction of fibroblsat proliferation by human mononuclear leukocyte-derived proteins. *Arthritis and Rheum* 1983; 26: 22-27.
- 736. Casini A, Ricci OE, Paoletti F, Surrenti C. Immune mechanisms for hepatic fibrinogenesis, T-lymphocyte-mediated stimulation of fibroblast collagen production in chronic active hepatitis. *Liver* 1985; 5: 134-141.
- 737. Annoni G, Czaja MJ, Weiner FR, Zern MA. Increased transforming growth factor-B1 (TGF-B1) gene expression in human liver disease. *Hepatology* 1988; 8: 1227 (abstract).
- 738. Weiner FR, Giambrone MA, Thompson E, Czaja MJ, Zern MA. Characterization of the molecular phenotype of Ito cells (Abstract). *Hepatology* 1988; 8: 1307.
- 739. Matsuoka M, Pham N-T, Tsukamoto H. Release of transforming growth factor beta1 (TGF beta1) like activity by Kupffer cells in alcoholi liver fibrogenesis and its stimulation of lipocyte collagen formation (Abstract). *Hepatology* 1988; 8: 1231.
- 740. Weiner FR, Giambrone MA, Takahashi S, Annoni G, Czaja MJ, Zern MA. Modulators of Ito cell collagen synthesis. *Hepatology* 1988; 8: 1253 (abstract).
- 741. Korn JH, Torres D, Downie E. Clonal heterogeneity in the fibroblast response to mononuclear cell derived mediators. Arthr Rheum 1984; 27: 174-179.
- 742. Surrenti C, Casini A, Milani S, Ambu S, Ceccatelli P, D'Agata A. Is determination of serum N-terminal procollagen type III peptide (sPIIIP) a marker of hepatic fibrosis? *Dig Dis Sci* 1987; 7: 705-709.
- 743. Pencev D, Pittner P, Hahn EG, et al. Discriminant analysis of laboratory, histological, and serum procollagen peptide data in patients with acute and chronic liver disease. In: U. Gerlach, G. Pott, J. Rauterberg and B. Voss (eds.). Connective tissue of the normal and fibrotic human liver. Stuttgart, New York: Thieme 1981, 212-214.
- 744. See reference 61 above.
- 745. See reference 28 above.

- 746. Davis GL, Czaja AJ, Ludwig J. Development and prognosis of cirrhosis in steroidtreated HBsAg-negative severe chronic active hepatitis (CAH) (Abstract). *Gastroenterology* 1982; 82: 1040.
- Ruwart MJ, Rush BD, Snyder KF, Peters KM, Appelman HD, Henley KS. 16,16dimethyl prostaglandin E₂ delays collagen formation in nutritional injury in rat liver. *Hepatology* 1988; 8: 61-64.

- 748. See reference 52 above.
- 749. See reference 263 above.
- 750. See reference 46 above.
- 751. See reference 254 above.
- 752. See reference 255 above.
- 753. See reference 258 above.
- 754. See reference 331 above.
- 755. See reference 332 above.
- 756. See reference 298 above.
- 757. See reference 300 above.
- 758. See reference 318 above.
- 759. See reference 319 above.
- 760. See reference 297 above.
- 761. See reference 341 above.
- 762. See reference 342 above.
- 762. See reference 279 above.
- 763. See reference 280 above.
- 764. See reference 283 above.
- 765. See reference 281 above.
- 766. See reference 268 above.
- 767. See reference 279 above.
- 768. See reference 286 above.
- 769. See reference 143 above.
- 770. See reference 144 above.
- 771. See reference 366 above.
- 772. See reference 367 above.
- 773. See reference 368 above.
- 774. See reference 369 above.
- 775. See reference 370 above.
- 776. See reference 371 above.

- 777. See reference 372 above.
- 778. See reference 374 above.
- 779. See reference 375 above.
- 780. See reference 376 above.
- 781. See reference 377 above.
- 782. See reference 378 above.
- 783. See reference 379 above.
- 784. See reference 380 above.
- 785. See reference 390 above.
- 786. See reference 382 above.
- 787. See reference 383 above.
- 788. See reference 384 above.
- 789. See reference 385 above.
- 790. See reference 386 above.
- 791. See reference 387 above.
- 792. See reference 388 above.
- 793. See reference 389 above.
- 794. See reference 391 above.
- 795. See reference 392 above.
- 796. See reference 143 above.
- 797. See reference 144 above.
- 798. See reference 373 above.
- 799. See reference 50 above.
- 800. See reference 415 above.
- 801. See reference 416 above.
- 802. See reference 417 above.
- 803. See reference 418 above.
- 804. See reference 48 above.
- 805. See reference 421 above.
- 806. See reference 422 above.
- 807. See reference 423 above.
- 808. See reference 415 above.
- 808a. See reference 735 above.
- 808b. See reference 736 above.
- 808c. See reference 737 above.
- 808d. See reference 738 above.

CHAPTER 13

- 809. See reference 148 above.
- 810. See reference 193 above.
- 811. See reference 192 above.
- 812. See reference 183 above.
- 813. See reference 184 above.
- 814. See reference 393 above.
- 815. See reference 395 above.
- 816. See reference 269 above.
- 817. See reference 50 above.
- 818. See reference 419 above.
- 819. See reference 284 above.
- 820. van Hoek B, Lijnema TH, van Zanten AK, Huizenga JR, Grond AJK, Gips CH. A modified anti-liver specific protein (LSP)-RIA for immunemonitoring of autoimmune chronic active hepatitis (AI-CAH): a longitudinal study (Abstract). J Hepatol 1988; 7 (Suppl 1): S192.
- 821. See reference 656 above.
- 822. See reference 52 above.
- 823. See reference 178 above.
- 824. See reference 182 above.
- 825. See reference 162 above.
- 826. See reference 163 above.
- 827. See reference 263 above.
- 828. See reference 44 above.
- 829. See reference 270 above.
- 830. See reference 363 above.

- 831. See reference 50 above.
- 832. See reference 263 above.
- 833. See reference 44 above.
- 834. See reference 95 above.
- 835. See reference 96 above.
- 836. See reference 393 above.
- 837. See reference 395 above.
- 838. See reference 820 above.

- 839. See reference 269 above.
- 840. See reference 148 above.
- 841. See reference 193 above.
- 842. See reference 192 above.
- 843. See reference 183 above.
- 844. See reference 184 above.
- 845. See reference 656 above.

- 846. See reference 29 above.
- 847. See reference 31 above.
- 848. See reference 40 above.
- 849. See reference 145 above.
- 850. Smith MGM, Williams R, Walker R, Rizetto M, Doniach D. Hepatic disorders associated with liver-kidney microsomal antibodies. *Br Med J* 1974; ii: 80-84.
- 851. Homberg J-C, Abuaf N, Bernard O, et al. Chronic active hepatitis associated with antiliver/kidney microsome antibody type 1: a second type of "autoimmune" hepatitis. *Hepatology* 1987; 7: 1333-1339.
- 852. Martini E, Abuaf N, Cavalli F, Durand V, Johanet C, Homberg J-C. Antibody to liver cytosol (anti-LC1) in patients with autoimmune chronic active hepatitis type 2. *Hepatology* 1988; 8: 1662-1666.
- 853. Manns M, Gerken G, Kyriatsoulis A, Staritz M, Meyer zum Büschenfelde K-H. Characterisation of a new subgroup of autoimmune chronic active hepatitis by autoantibodies against a soluble liver antigen. *Lancet* 1987; i: 292-294.
- 854. Maddrey WC. Subdivisions of idiopathic autoimmune chronic active hepatitis. Hepatology 1987; 7: 1372-1375.
- 855. See reference 50 above.
- 856. See reference 86 above.
- 857. Manns M, Meyer zum Büschenfelde K-H. A mitochondrial antigen-antibody system in cholestatic liver disease detected by radioimmunoassay. *Hepatology* 1982; 2: 1-7.
- 858. Manns M, Meyer zum Büschenfelde K-H, Slusarczyk J, Dienes HP. Detection of liverkidney microsomal antibodies by radioimmunoassay and their relation to antimitochondrial antibodies in inflammatory liver diseases. *Clin Exp Immunol* 1984; 57: 600-608.
- Hopf U, Meyer zum Büschenfelde K-H, Arnold W. Detection of liver membrane autoantibody in HBsAg negative chronic active hepatitis. N Engl J Med 1976; 294: 574-582.
- 860. See reference 471 above.
- Peto R, Pike MC, Armitage P, et al. Design and analysis of randomized clinical trials requiring prolonged observation of each patient.II.Analysis and examples. Br J Cancer 1977; 35: 1-39.
- 862. See reference 527 above.
- 863. See reference 472 above.

- 864. Frazer IH, Jordan TW, Collins EC, Andrews P, Mackay IR. Antibody to liver membrane antigens in chronic active hepatitis. IV. Exclusion of specific reactivity to polypeptides and glycolipids by immunoblotting. Hepatology 1987; 7: 4-10.
- 865. Gerken G, Manns M, Ramadori G, Poralla T, Dienes HP, Meyer zum Büschenfelde K-H. Role of liver membrane autoantibodies in autoimmune hepatitis. (Abstract). J Hepatol 1987; 5 (suppl.1): S135.
- Meyer zum Büschenfelde K-H, Miescher PA. Liver specific antigens. Purification and characterization. *Clin Exp Immunol* 1972; 10: 89-102.
- 867. See reference 444 above.
- 868. See reference 393 above.
- 869. See reference 162 above.
- 870. See reference 163 above.
- 871. See reference 182 above.
- 872. See reference 183 above.
- 873. See reference 184 above.
- 874. Khoury ED, Hammond L, Botazzo GF, Doniach D. Presence of organ-specific 'microsomal' autoantigen on the surface of human thyroid cells in culture: its involvement in the complement-mediated cytotoxicity. *Clin Exp Immunol* 1981; 45: 316-328.
- 875. Odievre M, Maggiore G, Homberg JC, et al. Seroimmunologic classification of chronic hepatitis in 57 children. *Hepatology* 1983; 3: 407-409.
- 875a. Manns M, Meyer zum Büschenfelde K-H, Dienes HP, Dormeyer HH. Chronisch active Hepatitis: Recidiv nach langjähriger immunsuppressiver Therapie trotz normaler Leberhistologie. Z Gastoenterol 1983; 21: 700-708.
- 875b. Pealeman M, Lobo-Yeo A, Mieli-Vergani G, Davies ET, Mowat AP, Vergani D. Characterization of anti-liver kidney microsomal antibody in childhood autoimmune chronic active hepatitis: evidence for IgG1 subclass restriction, polyclonality and non cross-reactivity with hepatocyte surface antigens. *Clin Exp Immunol* 1987; 69: 543-549.
- 876. Manns M, Kyriatsoulis A, Amelizad Z, Gerken G, Lohse A, Reske K, Meyer zum Büschenfelde K-H, Osch F. The target antigen of liver-kidney-microsomal (LKM)antibodies is shared by cytochrome P-450 isozymes (Abstract). J Hepatol 1987; 5 (suppl.1): S42.
- 877. Manns M, Johnson EF, Griffin KJ, Tan EM, Sullivan KF. Molecular cloning of liverkidney-microsomal (LKM-1) autoantigen. J Hepatol 1988; 7 (suppl.1): S56. 878. See reference 232 above.
- 879. See reference 228 above.
- 880. See reference 237 above.
- 881. See reference 233 above.
- 882. See reference 235 above.
- 883. van Hoek B, Kallenberg CGM, Lijnema TK, Huizenga JR, The TH, Gips CH. Lack of evidence for etiologic involvement of cytomegalovirus in idiopathic or 'autoimmune' chronic active hepatitis (Abstract). *Hepatology* 1989 (in press).

- 884. Wassermann A, Neisser A, Bruck C. A serodiagnostic reaction for syphilis. Deutsch Med Wschr 1906; 32: 745.
- 885. Moore JE, Mohr CF. Biologically false positive serologic tests for syphilis; type, incidence and cause. JAMA 1952; 150: 467-473.
- 886. Aho K. Studies of syphilic antibodies. II. Substances responsible for biological false positive sero-reactions. Br J Vener Dis 1968; 44: 49-54.
- Kraus SJ, Haserick JR, Lantz MA. Fluorescent treponemal antibody-absorption test reactions in lupus erythematosus. Atypical binding pattern and probable false-positive reactions. N Engl J Med 1970; 282: 1287-1290.
- 888. Peter CR, Thompson MA, Wilson DL. False-positive reactions in the rapid plasma reagin-card, fluorescent treponemal antibody-absorbed, and hemagglutination treponemal syphilis serology tests. J Clin Microbiol 1979; 9: 369-372.
- 889. See reference 5 above.
- 890. See reference 4 above.
- 891. See reference 8 above.
- 892. Laurell AB, Nilsson IM. Hypergammaglobulinemia, circulating anticoagulant and biological false positive Wasserman reaction. J Lab Clin Med 1957; 49: 694-707.
- 893. Harris EN, Gharavi AE, Boey ML, Patel BM, Mackworth-Young CG, Loizou S, Hughes GRV. Anticardiolipin antibodies: detection by radioimmunoassay and association with thrombosis in systemic lupus erythematosus. *Lancet* 1983; ii: 1211-1214.
- 894. Loizou S, McCrea JD, Rudge AC, Reynolds R, Boyle CC, Harris EN. Measurement of anti-cardiolipin antibodies by an enzyme-linked immunosorbent assay (ELISA): standardization and quantitation of results. *Clin Exp Immunol* 1985; 62: 738-745.
- 895. Schleider MA, Nachman RL, Jaffe EA, Coleman M. A clinical study of the lupus anticoagulant. *Blood* 1976; 48: 499-509.
- 896. Lechner K. Acquired inhibitors in nonhemophilic patients. Haemostasis 1974; 3: 65-93.
- 897. Lee SL, Miotti AB. Disorders of hemostatic function in patients with systemic lupus erythematosus. Semin Arthritis Rheum 1975; 4: 241-252.
- McHugh NJ, Maymo J, Skinner RP, James I, Maddison PJ. Anticardiolipin antibodies, levido reticularis, and major cerebrovascular and renal disease in systemic lupus erythematosus. Ann Rheum Dis 1988; 47: 110-115.
- 899. Fort JG, Cowchock FS, Abruzzo JL, Smith JB. Anticardiolipin antibodies in patients with rheumatic diseases. Arthritis and Rheumatism 1987; 30: 752-755.
- 900. Koike T, Sueishi M, Funaki H, Tomioka H, Yoshida S. Anti-phospholipid antibodies and biological false positive serological test for syphilis in patients with systemic lupus erythematosus. *Clin Exp Immunol* 1984; 56: 193-199.
- 901. Colaço CB, Male DK. Anti-phospholipid antibodies in syphilis and a thrombotic subset of SLE: distinct profiles of epitope specificity. *Clin Exp Immunol* 1985; 59: 449-456.
- 902. Manoussakis MN, Tzioufas AG, Silis MP, Pange PJE, Goudevenos J, Moutsopoulos HM. High prevalence of anti-cardiolipin and other autoantibodies in a healthy elderly population. *Clin Exp Immunol* 1987; 69: 557-65.
- 903. Mueh JR, Herbst KD, Rapaport SI. Thrombosis in patients with systemic lupus crythematosus. Ann Intern Med 1980; 92: 156-159.

- 904. Boey ML, Colaçao CB, Gharavi AE, Elkoru KB, Loizou S, Hughes GRV. Thrombosis in SLE: striking association with the presence of circulating "lupus anticoagulant". B r Med J 1983; 287: 1021-1023.
- 905. Byron MA. The clotting defect in SLE. Clin Rheum Dis 1982; 8: 137-151.
- 906. Elias M, Eldor A. Thromboembolism in patients with the 'lupus'-type circulating anticoagulant. Arch Intern Med 1984; 144: 510-515.
- 907. Firkin BG, Howard MA, Radford N. Possible relationship between the lupus inhibitor and recurrent abortion in young women. *Lancet* 1980; ii: 366.
- 908. Carreras LO, Defreyn G, Machin SJ, et al. Arterial thrombosis, intrauterine death and "lupus" anticoagulant: detection of immunoglobulin interfering with prostacyclin formation. *Lancet* 1981; i: 244-246.
- 909. Nilsson IM, Astedt B, Hedner U, Berezin D. Intrauterine death and circulating anticoagulant "antithromboplastin". Acta Med Scand 1975; 197: 153-159.
- 910. Soulier RP, Boffa MC. Avortements à répétition, thromboses et anticoagulant circulant antithromboplastine. *Nouv Presse Med* 1980; 9: 859-864.
- 911. Lubbe WF, Butler WS, Palmer SJ, Liggins GC. Lupus anticoagulant in pregnancy. Br J Obstetr Gynaecol 1984; 91: 357-363.
- 912. Editorial. Lupus anticoagulant. Lancet 1984; i: 1157-1158.
- 913. Koike T, Homioka H, Kumagi A. Antibodies cross-reactive with DNA and cardiolipin in patients with systemic lupus erythematosus *Clin Exp Imunol* 1982; 50: 298-302.
- 914. Isenberg DA, Shoenfeld Y, Schwartz RS. Multiple serologic reactions and their relationship to clinical activity in systemic lupus erythematosus. *Arthritis Rheum* 1984; 27: 132-138.
- 915. Harris EN, Gharavi AE, Loizou S, et al. Crossreactivity of antiphospholipid antibodies. J Clin Lab Immunol 1985; 1: 1-6.
- 916. Eilat D, Zlotnick AY, Fishel R. Evaluation of the cross-reactivion between anti-DNA and anti-cardiolipin antibodies in SLE and experimental animals. *Clin Exp Immunol* 1986; 65: 269-278.
- '917. Valesini G, Tincani A, Harris EN, et al. Use of monoclonal antibodies to identify shared idiotypes on anticardiolipin and anti-DNA antibodies in human sera. Clin Exp Immunol 1987; 70: 18-25.
- 918. Lafer EM, Rausch J, Andrzejewski C, et al. Polyspecific antibodies reactive with both polynucleotides and phospholipids. J Exp Med 1981; 153: 897-909.
- 919. Shoenfield Y, Rausch J, Massicotte H, et al. Polyspecificity of monoclonal lupus autoantibodies produced by human-human hybridomas. N Engl J Med 1983; 308: 414-420.
- 920. Smeenk RJT, Lucassen WAM, Swaak TJG. Is anticardiolipin activity a cross-reaction of anti-DNA or a separate entity ? Arthritis and Rheum 1987; 30: 607-17.
- 921. Wood JR, Czaja AJ, Beaver SJ, et al. Frequency and significance of antibody to doublestranded DNA in chronic active hepatitis. *Hepatology* 1986; 6: 976-980.
- 922. Hall S, Czaja AJ, Kaufman DK, Markowitz H, Ginsburg WW. How lupoid is lupoid hepatitis? J Rheumatol 1986; 13: 95-98.
- 923. Davis P, Read AE. Antibodies to double-stranded (native) DNA in active chronic hepatitis. Gut 1975; 16: 413-415.
- 924. Kingham JGC, Rassam S, Ganguly NK, et al. DNA-binding antibodies and hepatitis B markers in acute and chronic liver disease. Clin Exp Immunol 1978; 33: 204-210.

- Jain S, Markham R, Thomas HC, Sherlock S. Double-stranded DNA-binding capacity of serum in acute and chronic liver disease. *Clin Exp Immunol* 1976; 26: 35-41.
 See reference 31 above.
- 927. See reference 40 above.
- 928. Somerfield SD, Roberts MW, Booth RJ. Double-stranded DNA antibodies: a comparison of four methods of detection. J Clin Pathol 1982; 34: 1032-1035.
- 929. Kallenberg CGM, van der Meulen J, Pastoor GW, Snijder JAM, Feltkamp TEW, The TH. Human fibroblast, a convenient nuclear substrate for detection of antinuclear antibodies including anti-centromere antibodies. Scand J Rheumatol 1983; 12: 193-200.
- 930. Isenberg DA, Maddison PJ. Detection of antibodies to double stranded DNA and extractable nuclear antigen. Association of Clinical Pathologists 1987; Broadsheet 117: 1374-1381.
- 931. Caruana A, André C, Zafrani E-S, Saint-Marc Girardin M-F, Metreau J-M, Dhumeaux D. Serum antibodies against double-stranded DNA in chronic active hepatitis: an index of drug-induced liver disease ? (Letter to the editor). *Lancet* 1983; i: 776.
- 932. Metreau J-M, André C, Zufrani E-S, Saint-Marc Girardin MF, Caruana A, Dhumeaux D. Hépatites chroniques actives associées à des anticorps anti-ADN natif: fréquence de l'étiologie médicamenteuse. *Gastroenterol Clin Biol* 1984; 8: 833-837.
- 933. Kurki P, Gripenberg M, Teppo A-M, Salaspuro M. Profiles of antinuclear antibodies in chronic active hepatitis, primary biliary cirrhosis and alcoholic liver disease. *Liver* 1984; 4: 134-138.
- 934. Gurian LE, Rogoff TM, Ware AJ, Jordan RE, Combes B, Gilliam JN. The immunologic diagnosis of chronic active "autoimmune" hepatitis: distinction from systemic lupus erythematosus. *Hepatology* 1985; 5: 397-402.
- 935. Smeenk R, van der Lelij G, Swaak T, Groenwold J, Aarden L. Specificity in systemic lupus erythematosus of antibodies to double-stranded DNA measured with the polyethylene glycol precipitation assay. *Arthritis Rheum* 1982; 25: 631-638.
- 936. Mackay IR. Immunological aspects of chronic active hepatitis. *Hepatology* 1983; 5: 724-728.
- 937. Penner E, Kindas-Mügge I, Hitchman E, Sauermann G. Nuclear antigens recognized by antibodies present in liver disease sera. *Clin Exp Immunol* 1986; 63: 428-433.
- 938. Penner E, Muller S, Zimmerman D, Van Regenmortel MHV. High prevalence of antibodies to histones among patients with primary biliary cirrhosis. *Clin Exp Immunol* 1987; 70: 47-52.
- 939. Penner E, Wesierska-Gadek J, Nitchman E, Sauermann G. Proteins of the nuclear envelope as antigens in lupoid hepatitis (Abstract). *Hepatology* 1987; 7: 1115.
- 940. Lassoued K, Danon F, André C, Guilly MN, Brouet JC, Dhumeaux D, Courvalin JC. Liver diseases associated with antinuclear antibodies displaying a fluorescent ring-like pattern (Abstract). J Hepatol 1988; 7 (Suppl.1): S47.
- 941. Lozano F, Pares A, Borche L, Gallart T, Rodes J. Nuclear lamina antibodies in primary biliary cirrhosis (Abstract). J Hepatol 1986; 3 (Suppl.1): S28.
- 942. Lozano F, Pares A, Borche L, Gallart T, Rodes J, Vives J. Autoantibodies against nuclear envelope-associated protein in primary biliary cirrhosis. *Hepatology* 1988; 8: 930-938.
- 943. Lassoued K, Danon F, André C, Guilly MN, Brouet JC, Dhumeaux D, Courvalin JC. Antibodies directed to 200 kD polypeptide(s) of the nuclear envelope: a new serological marker associated with primary biliary cirrhosis (Abstract). *Hepatology* 1987; 7: 1115.
- 944. Hamlyn AN, Adams D, Sherlock S. Primary or secondary sicca complex? Investigation in primary biliary cirrhosis by histocompatibility testing. Br Med J 1980; 281: 425-426.
- 945. Penner E. Nature of immune complexes in autoimmune chronic active hepatitis. Gastroenterology 1987; 92: 304-308.
- 946. Penner E, Reichlin M. Primary biliary cirrhosis associated with Sjögren's syndrome: evidence for circulating and tissue-deposited Ro/anti-Ro immune complexes. Arthritis Rheum 1982; 25: 1250-1253.
- 947. Golding PL, Smith M, Williams R. Multisystem involvement in chronic liver disease. Studies on the incidence and pathogenesis. *Am J Med* 1973; 55: 772-782.
- 948. Bernstein RM, Steigwald JC, Tan EM. Association of antinuclear and antinucleolar antibodies in progressive systemic sclerosis. *Clin Exp Immunol* 1982; 48: 43.
- 949. Makinen D, Fritzler M, Davis P, Sherlock S. Anticentromere antibodies in primary biliary cirrhosis. Arthritis Rheum 1983; 26: 914-917.
- Cassani F, Biancho Bianchi F, Lenzi M, Volta U, Pisi E. Immunomorphological characterization of antinuclear antibodies in chronic liver disease. J Clin Pathol 1985; 38: 801-805.
- 951. Moroi Y, Peebles C, Fritzler M, et al. Autoantibody to centromere (kinetochore) in scleroderma sera. Proc Natl Acad USA 1980; 77: 1627-1631.
- 952. Powell FC, Winkelman RK, Venencie-Lemarchand F, Spurbeck JL, Schroeter AL. The anticentromere antibody; disease specificity and clinical significance. *Mayo Clin Pro c* 1984; 59: 700-706.
- 953. Bernstein RM, Neuberger JM, Bunn CC, et al. Diversity of autoantibodies in primary biliary cirrhosis and chronic active hepatitis. Clin Exp Immunol 1984; 55: 553-560.
- 954. Powell FC, Schroeter AL, Dickson ER. Primary biliary cirrhosis and the CREST syndrome: a report of 22 cases. *Quart J Med* 1987; 62: 75-82.
- 955. Epstein O. Primary biliary cirrhosis and the CREST syndrome: new terminology?. *Hepatology* 1988; 8: 189.
- 956. Petri M, Golbus M, Anderson R, Whiting-O'Keefe Q, Corash L, Hellmann D. Antinuclear antibody, lupus anticoagulant, and anticardiolipin antibody in women with idiopathic habitual abortion. A controlled, prospective study of forty-four women. Arthritis and Rheum 1987; 30: 601-606.
- 957. Steven MM, Buckley JD, Mackay IR. Pregnancy in chronic active hepatitis. Quart J Med 1979; 192: 519-531.

CHAPTER 17

- 958. See reference 40 above.
- 959. See reference 50 above.
- 960. See reference 227 above.
- 961. See reference 228 above.
- 962. See reference 229 above.
- 963. See reference 230 above.
- 964. See reference 231 above.
- 965. See reference 232 above.
- 966. See reference 233 above.
- 967. See reference 234 above.
- 968. See reference 235 above.

- 969. Baals H, Bulow B, Freisenhausen HD, Mai K. Nachweis von Cytomegalievirus Antikorpern bei der Au(SH)-Ag positiven and der Au(SH)-Ag negativen Hepatitis. Verh Deutsch Ges Inn Med 1972; 78: 928-933.
- 970. Alwen J, Emmerson AM. Antibodies against adeno-, cytomegalo- and rubella viruses in Australia-antigen-negative sera from patients with infectious hepatitis. J Hyg Camb 1974; 72: 433-439.
- 971. Laitinen O, Vaheri A. Very high measles and rubella virus antibody titres associsated with hepatitis, systemic lupus erythematosus, and infectious mononucleosis. *Lancet* 1974; i: 194-197.
- 972. See reference 236 above.
- 973. Ware AJ, Luby JP, Eigenbrodt EH, et al. Spectrum of liver diseases in renal transplant recipients. Gastroenterology 1975; 68: 755-764.
- 974. Ten Napel CHH, Houthoff HJ, The TH. Cytomegalovirus hepatitis in normal and immune compromised hosts. *Liver* 1984; 4: 184-194.
- 975. The TH, Langenhuysen MMAC. Antibodies against membrane antigens of cytomegalovirus infected cells in sera of patients with a cytomegalovirus infection. Clin Exp Immunol 1972; 11: 475-482.
- 976. Cremer N, Cossen CK, Shell GR, Pereira L. Antibody response to cytomegalovirus polypeptides captured by monoclonal antibodies on the solid phase in enzyme immunoassays. J Clin Microbiol 1985; 21: 517-521.
- 977. Nielsen SL, Ronholm E, Sorensen I, *et al.* Detection of immunoglobulin G antibodies to cytomegalovirus antigens by antibody capture enzyme-linked immunosorbent assay. *J Clin Microbiol* 1986; 24: 998-1003.
- 978. See reference 148 above.
- 979. See reference 149 above.
- 980. See reference 162 above.
- 981. See reference 163 above.
- 982. See reference 164 above.
- 983. See reference 182 above.
- 984. See reference 183 above.
- 985. See reference 184 above.
- 986. Pak CY, Eun H-M, McArthur RG, Yoon J-W. Association of cytomegalovirus infection with autoimmune type I diabetes. *Lancet* 1988; ii: 1-4.
- 987. Koerner K, Freudenberg J, Kubanek B. Blutspenderauswahl zur Bereitstellung von anti-Cytomegalievirus-negativen Blutcomponenten. Dtsch Med Wschr 1989; 114: 203-207.
- 988. Ahlfors K. IgG antibodies to cytomegalovirus in a normal urban Swedish population. Scand J Infect Dis 1984; 16: 335-337.
- 989. See reference 393 above.
- 990. See reference 820 above.
- 991. Middeldorp JM, Jongsma J, Haar A ter, Schirm J, The TH. Detection of antibodies against cytomegalovirus early and late antigens by enzyme-linked immunosorbent assay. *J Clin Microbiol* 1984; 20: 763-771.
- 992. van der Voort LHM, van Zanten J, de Leij LFMH, The TH. Detection of human antibodies to specific antigens of cytomegalovirus using an antigen capture immunoassay. J Immunol Meth 1989; 121: 95-103.

- 992a. Van der Bij B, Torensma R, Van Son WJ, Anema J, Schirm J, Tegzess AM, The TH. Rapid immunodiagnosis of active cytomegalovirus infection by monoclonal antibody staining of blood leukocytes. *J Med Virol* 1988; 12: 179-188.
 - 993. Cranage MP, Couzarides T, Bankier AT, Satchwell S, Weston K, Tomlinson P, Barrell B, Hart H, Bell SE, Minson AC, Smith GL. Identification of the human cytomegalovirus glycoprotein B gene and induction of neutralizing antibodies via its expression in recombinant vaccinia virus. *EMBO J* 1986; 5: 3057-3063.
- 994. Eggink HF, Houthoff HJ, Huitema S, Gips CH, Poppema S. Cellular and humoral immune reactions in chronic active liver disease. I. Lymphocyte subsets in liver biopsies of patients with untreated idiopathic autoimmune hepatitis, chronic active hepatitis B and primary biliary cirrhosis. *Clin Exp Immunol* 1982; 50: 17-24.
- 995. See reference 152 above.
- 996. See reference 153 above.
- 997. See reference 154 above.
- 998. See reference 155 above.
- 999. See reference 156 above.
- 1000. See reference 157 above.
- 1001. See reference 158 above.
- 1002. See reference 159 above.
- 1003. See reference 160 above.
- 1004. See reference 161 above.
- 1005. Triger DR, Cynamon MH, Wright R. Studies on the hepatic uptake of antigen. I. Comparison of inferior vena cava and portal vein routes of immunization. *Immunology* 1973; 25: 941-950.
- 1006. See reference 237 above.
- 1007. See reference 877 above.
- 1008. Beck S, Barrell BC. Human cytomegalovirus encodes a glycoprotein homologous to MHC class-I antigens. Nature 1988; 331: 269-272.

CURRICULUM VITAE

The author of this thesis was born on June 14, 1956 at Voorburg, The Netherlands. In 1962 he moved to Arnhem, The Netherlands, where he attended highschool from 1968-1974 (gymnasium β, Thorbecke Lyceum). From 1974-1979 he attended medical school at the State University of Groningen, The Netherlands, where he also assisted in educational programs in obstetrics/gynaecology and did a special project in nephropathology. In 1979 he married Beryl Flikweert. After internships and a special project on lung disease in the Groningen University Hospital (Academisch Ziekenhuis Groningen) he obtained his M.D. degree in February 1982. January 1983 he entered a residency in internal medicine in the same hospital, which was completed in December 1987. He developed a special interest in hepatology, gastroenterology and immunology. During his residency he passed the American Visa Qualifying Exam (VQE or FMGEMS) in 1984. In august 1986 his interests in hepatology and immunology urged him to start research in chronic active hepatitis. As a specialist in internal medicine he worked in the Division of Hepatology of Groningen University Hospital, where he also continued research, resulting in this PhD-thesis. For this purpose he visited the Liver Unit in King's College Hospital, London (G.-B.) (director: Roger Williams) in 1986 to work in the laboratory and attend the liver clinical course. In 1987 he visited and worked in the laboratory of prof. dr. K.-H. Meyer zum Büschenfelde and prof. dr. M. Manns, in the Clinic of the Johannes Gutenberg Universität in Mainz (F.R.G.).

Bart van Hoek is currently employed as a fellow in the advanced hepatology training program at Mayo Clinic, Rochester, Minnesota, U.S.A., Department of Gastroenterology and Internal Medicine. He is involved in immunological research in liver disease and is supported by the Netherlands Organization for Scientific Research. He is a member of the European Association for the Study of the Liver, The Dutch Society for Gastroenterology, The Dutch Society for Hepatology, the Flemish Society for Gastroenterology and the American Association for the Study of Liver Diseases.