

University of Groningen

Photochemistry and organic complexation of iron

Rijkenberg, Michaël Johannes Adrianus

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version

Publisher's PDF, also known as Version of record

Publication date:

2005

[Link to publication in University of Groningen/UMCG research database](#)

Citation for published version (APA):

Rijkenberg, M. J. A. (2005). *Photochemistry and organic complexation of iron: Interactions in the Southern Ocean*. s.n.

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: <https://www.rug.nl/library/open-access/self-archiving-pure/taverne-amendment>.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): <http://www.rug.nl/research/portal>. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

Chapter 8

Iron-binding ligands in Dutch estuaries are not affected by UV induced photochemical degradation

Micha J.A. Rijkenberg, Loes J.A. Gerringa, Ilona Velzeboer, Klaas R. Timmermans, Anita G.J. Buma, Hein J.W. de Baar

Abstract

This study shows that ultraviolet B (UV-B: 280-315 nm) and ultraviolet A (UVA: 315-400 nm) have no significant influence on the photo-degradation of organic Fe-binding ligands in estuarine waters from Marsdiep and Scheldt (The Netherlands). High salinity estuarine seawater from the Marsdiep and Scheldt contain concentrations of organic Fe-binding ligands as high as 24.4 equivalents of nM Fe (eq nM Fe) and ~ 4.6 eq nM Fe respectively with conditional stability constants (K') of $10^{21.0}$ and $10^{20.1}$ respectively. Investigation of the relation between the organically bound iron fraction and the photo-production of Fe(II) in the estuarine Marsdiep and Scheldt water showed that the concentration of Fe(II) produced was very low (< 240 pM). This demonstrates that the major part of the organically complexed Fe is not involved in photo-induced Fe redox cycling. Consequently UV has no influence on the transport of dissolved organically complexed Fe from the estuarine environment to the coastal zone.

1. Introduction

The overall dissolved iron (Fe) input of rivers in the world oceans is estimated to be 26×10^9 mol y^{-1} (de Baar and de Jong, 2001). The majority of dissolved Fe in river water exist as small colloid particles (Fox, 1988). Flocculation of these colloids, due to the change in ionic strength upon mixing of fresh river water with salt seawater, causes a massive removal of the freshly formed particulate Fe (Sholkovitz, 1978; Sholkovitz et al., 1978). In the Scheldt estuary flocculation occurs between salinities of 1-5 (Wollast and Peters, 1980).

Organic complexation is an important factor in the biogeochemistry of Fe in estuarine waters, as it maintains iron in the dissolved phase at high salinities after the flocculation zone. The dissolved phase will be flushed from the estuary while the non-organically complexed fraction tends to aggregate and adsorb to particles, thereby residing within the internal cycle of the estuary for a longer time (Morris et al., 1986). Thus organically complexed iron of estuarine systems can act as a source of dissolved iron for coastal zone waters (Powell and Wilson-Finelli, 2003). Despite its importance in the geochemical Fe cycle and despite the fact

that primary production can be limited by Fe availability in coastal environments (Glover, 1978; Hutchins and Bruland, 1998; Hutchins et al., 1998; Kirchman et al., 2000; Bruland et al., 2001), data on the organic complexation of Fe in estuaries is scarce.

The chemistry of Fe in the more saline estuarine waters is complex. Dissolved Fe can exist in two different oxidation states, Fe(II) and Fe(III). The Fe(III) is the thermodynamically stable form in oxygenated waters and has a very low solubility in high saline water (Millero, 1998; Liu and Millero, 2002). Fe(III) becomes rapidly hydrolyzed into various Fe(III) oxyhydroxides. The low solubility of Fe oxyhydroxides and the tendency to form colloids contributes to the scarcity of directly bioavailable Fe species in the marine environment (Martin et al., 1995; Powell et al., 1996; Sanudo-Wilhelmy et al., 1996; Wen et al., 1999).

Dissolved Fe(III) occurs for a major part as complexes (FeL) with strong organic ligands (Gledhill and van den Berg, 1994; Hutchins et al., 1999; Waite, 2001). This organic complexation is normally described in terms of concentration and conditional stability constants (K'_{FeL}) for various oceans (Gledhill and van den Berg, 1994; Rue and Bruland, 1995; Wu and Luther, 1995; Rue and Bruland, 1997; Gledhill et al., 1998; Nolting et al., 1998; Witter and Luther, 1998; Witter et al., 2000) and coastal seas (Gledhill and van den Berg, 1994; Croot and Johansson, 2000; Macrellis et al., 2001). The conditional stability constants of FeL complexes range in magnitude between 10^{18} and 10^{23} (Witter et al., 2000). Although the K'_{FeL} of organic complexes between different oceans may range over 5 orders of magnitude it is not possible to use the K'_{FeL} to identify organic ligands using the K'_{FeL} of known model ligands (Witter et al., 2000). Instead Witter et al. (2000) suggested that, based on a comparison of formation- and dissociation rate constants between model ligands and field samples, most unknown ligands in seawater originate from porphyrin and siderophore-like compounds. Porphyrins are molecules that relate to the photosynthetic pigments of autotrophic organisms, e.g. chlorophyll-*a*. Siderophores are low-molecular weight, high affinity Fe(III) binding ligands secreted by e.g. marine cyanobacteria and heterotrophic bacteria under Fe limiting circumstances to bind and transport Fe for uptake (Wilhelm and Trick, 1994; Martinez et al., 2000). Furthermore, Macrellis et al. (2001) found hydroxamate or catecholate Fe-binding functional groups, characteristic for siderophores, present in different size classes of samples taken along the Californian coast. The presence of Fe-binding compounds in the low molecular weight classes (<300 Da) was suggested to be caused by UV induced degradation of larger Fe-binding siderophores in the surface waters.

Photochemical (-induced) reactions of Fe(III)-siderophore complexes can result in the decomposition of the siderophores and the production of Fe(II) in sunlit waters (Barbeau et al., 2001). Furthermore, organic Fe-binding ligands can increase or decrease Fe photoreducibility by affecting Fe aggregation and by binding of Fe resulting in photostable complexes (Chapter 5). Iron(II), although well soluble in seawater, becomes rapidly oxidized

by O₂ and H₂O₂ (Millero et al., 1987; Millero and Izaguirre, 1989; Millero and Sotolongo, 1989; King et al., 1991; King et al., 1995; King, 1998) but at the same time Fe(II) is thought to be an important biological available species (Anderson and Morel, 1980; Takeda and Kamatani, 1989; Auclair, 1995; Croot et al., 2001). The redox cycling of (colloidal) iron hydroxides and iron chelates initiated by photochemical processes is mentioned as an important mechanism by which iron bioavailability for phytoplankton is enhanced by increasing the concentration of reactive inorganic species of Fe(II) and Fe(III) (Anderson and Morel, 1982; Finden et al., 1984; Rich and Morel, 1990; Wells and Mayer, 1991; Johnson et al., 1994; Miller and Kester, 1994; Sunda and Huntsman, 1995; Barbeau et al., 2001).

There is still much controversy about the importance of photodegradation and subsequent Fe(II) production of organically complexed Fe in the natural environment. Siderophores may be photodegradable, but until recently none of the depth profiles of Fe(III) chelators reported (Gledhill and van den Berg, 1994; Rue and Bruland, 1995; Wu and Luther, 1995) exhibit shallow mixed layer minima or any other features which would suggest a surface/photochemical sink (Moffett, 2001). Yet, Boye et al. (2001) recently reported depth profiles, where lower concentrations of organic Fe-binding ligands in Southern Ocean surface waters suggest photodegradation. Furthermore, Powell and Wilson-Finelli (2003) reported photodegradation of organic Fe-binding ligands at the timescale of hours in the Gulf of Mexico.

Photodegradation of organically bound Fe will influence the biogeochemical cycle of iron and could increase its biological availability and reactivity. Here we report experiments performed with high saline estuarine Marsdiep and Scheldt waters to investigate the influence of light on the presence of organic Fe-binding ligands. Furthermore, we show that although high concentrations organic Fe-binding ligands are present in the Marsdiep and Scheldt water, upon irradiation the Fe(II) production is relatively low as compared with the fraction of Fe bound to organic ligands.

2. Materials and methods

2.1 Samples

Marsdiep water (0.2 µm filtered) was collected at the 8th of May 2003 at 11:50 from the NIOZ jetty during high tide (Figure 1). The salinity was 29.1 versus the practical salinity

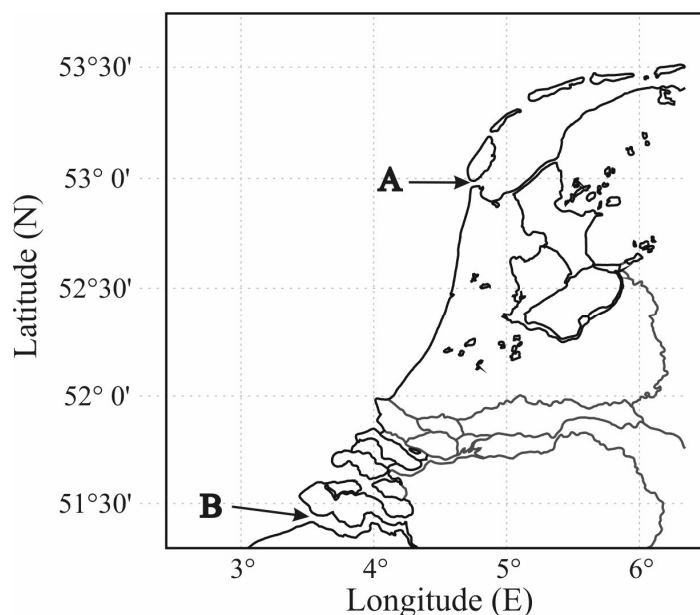


Figure 1. The coastline of the Netherlands. Samples were taken in the Marsdiep (A) and in the Westerschelde (Scheldt) (B).

scale, and the DOC concentration was $298 \pm 14.6 \mu\text{M C}$. The Marsdiep water was collected in an acid cleaned 20 liter carboy using a portable flow bench (Interflow) and acid cleaned tubing. The carboy was stored in the dark at 4°C . The collected Marsdiep water was used within a week after collection.

Scheldt water ($0.2 \mu\text{m}$ filtered) was collected in the Westerschelde near Vlissingen at the 5th of April 2001 (Figure 1). Surface seawater was pumped into an over-pressurized class 100 clean air container using a Teflon diaphragm pump (Almatec A-15, Germany) driven by a compressor (Jun-Air, Denmark, model 600-4B) connected via acid-washed braided PVC tubing to a torpedo towed at approximately 3 m depth alongside the ship (Navicula, Royal NIOZ). The seawater was filtered in-line by a (Sartorius Sartobran filter capsule 5231307H8) with a cut-off of $0.2 \mu\text{m}$. The $<1 \text{ kDa}$ fraction was filtered using an acid-cleaned Amicon SP60 cartridge and a peristaltic pump of Watson Marlow (604S/R). The salinity of the Scheldt water was 26 and the DOC concentration was $217.6 \mu\text{M}$. The collected Scheldt water was used the next day for the deck-incubations.

2.2 Experimental

The influence of irradiance on the organic Fe-binding ligands in Marsdiep water was investigated in the laboratory. Before use, acid-cleaned UV transparent polymethylmetacrylate (PMMA) bottles (Steeneken et al., 1995) were conditioned overnight with Marsdiep water ($0.2 \mu\text{m}$ filtered) at 15°C , the temperature of the water in the field at the

time of collection. The experiments were performed in a temperature controlled class 100 clean container. The pH of the Marsdiep samples was 8.14 +/- 0.006 (Metrohm 713 pH meter). To investigate the influence of UV on the photodegradation of organic Fe-binding ligands a PMMA bottle containing Marsdiep water was placed in the light and another PMMA bottle containing Marsdiep water was placed in the dark. Samples were taken at $t = 0$ and $t = 18.5$ h. The samples were frozen and used within a week for competitive ligand exchange-adsorptive cathodic stripping voltammetry (CLE-ACSV) to determine the concentration and the log K' of the organic Fe-binding ligands (equivalents of Fe-binding places).

The influence of irradiance on the organic Fe-binding ligands in Scheldt water was performed by means of deck incubations. The incubations were performed using acid cleaned PMMA and pre-conditioned bottles in UV transparent PMMA incubators. The bottles, incubated between 8:45 and 22:40 at the 7th of April 2001, were kept at a constant ambient temperature of 12°C using running seawater from the ship's pumping system. The pH of the Scheldt water was 8.8 (Metrohm 713 pH meter). Sampling was performed in a laminar flow bench in a class 100 clean container. Samples for the determination of the concentration organic iron binding ligands and their log K' were immediately frozen. Stored samples for dissolved iron were acidified with 100 μ l 3x quartz distilled HCl per 100 ml sample (pH = 2).

Investigation of the photoproduction of Fe(II) was performed in a temperature controlled class 100 clean container. The PMMA bottles were pre-conditioned with the water as used during the experiments and at the temperatures as used during the experiments. Water was continuously stirred using a magnetic induced, Teflon stirring bean during the experiment. An experiment typically started with measuring the background signal in the dark, after which the lamps were switched on. During 90-120 minutes the production of Fe(II) was followed. When enough sample volume was available, the oxidation of Fe(II) was followed for 30 minutes after having turned off the light.

2.3 Light

Philips UVB (TL-12), UVA (TL' 40W/05) and VIS (TL'D 36W/33) lamps were used to simulate the solar spectrum in the experiments with Marsdiep water. Spectral conditions were measured using a MACAM Spectroradiometer SR9910 with a small spherical 4π sensor. All sides of the box shaped PMMA bottle, except the top, were covered with black plastic to prevent focussing effects. Spectra were recorded from 280-700 nm.

Fe(III) siderophore complexes have been shown to absorb light in the high UVB and low UVA range (Barbeau et al., 2001; Barbeau et al., 2003). Therefore, we used mainly UVB

containing 3 M HCl and a counter electrode of glassy carbon. The titration was performed using 0.01 M stock solution of TAC in 3x quartz distilled (QD) methanol, 1 M boric acid (Suprapur, Merck) in 0.3 M ammonia (Suprapur, Merck) (extra cleaning by the addition of TAC after which TAC and Fe(TAC)₂ was removed with a C18 SepPak column) to buffer the samples to a pH of 8.05 and a 10⁻⁶ M Fe(III) stock solution acidified with 0.012 M HCl (3xQD). Aliquots of 15 ml were spiked with Fe(III) up to final concentrations between 0 and 20 nM and allowed to equilibrate overnight (> 15 hours) with 5 mM borate buffer and 10 μM TAC. The concentration Fe(TAC)₂ in the samples was measured using the following procedure: i) removal of oxygen from the samples for 200 seconds with dry nitrogen gas, after which a fresh Hg drop was formed, ii) a deposition potential of -0.40 V was applied for 30-60 seconds according to the sample measured, the solution was stirred to facilitate the adsorption of the Fe(TAC)₂ to the Hg drop, iii) at the end of the adsorption period the stirrer was stopped and the potential was scanned using the differential pulse method from -0.40 to -0.90 V at 19.5 mV s⁻¹ and the stripping current from the adsorbed Fe(TAC)₂ recorded.

The ligand concentration and the conditional stability constants were calculated using the non-linear fit of the Langmuir isotherm (Gerringa et al., 1995).

2.5 Iron(II) analysis

Concentrations of Fe(II) were followed using an automated flow injection analysis system employing a luminol-based chemiluminescence detection of Fe(II) (Seitz and Hercules, 1972; King et al., 1995; O'Sullivan et al., 1995). An alkaline luminol solution is mixed with the Fe(II) sample in a spiral shaped flow cell in front of a Hamamatsu HC135 photon counter. At pH 10, Fe(II) is rapidly oxidized by oxygen on a millisecond time scale causing the oxidation of luminol and producing blue light (Xiao et al., 2002).

Samples were transported in-line from the PMMA bottle to a sample loop. Then the sample was, by introducing it in a pure water carrier (18.2 MΩ nanopure water), transported into the flow cell every 93 seconds.

The complete analytic system was built inside a light-tight wooden box. The luminol reagent and the carrier were kept in light-tight bags (as used for storage of photographic films). The tubing was covered by aluminum foil and the tubing of the peristaltic pump was shaded by black plastic.

A 15 mM 5-amino-2,3-dihydro-1,4-phthal-azinedione (Luminol) (SIGMA) stock solution was prepared weekly in 20 mM Na₂CO₃. The 50 μM luminol reagent solution was made in 0.5 M NH₃ (Suprapur, Merck) and 0.1 M HCl (Suprapur, Merck). The luminol reagent solution was stored in the dark for at least 24 hours before use to ensure that the reagent properties had stabilized. An 0.01 M Fe(II) stock was prepared monthly by dissolving ferrous ammonium sulfate hexahydrate (Fe^(II)(NH₄SO₄)₂·6H₂O) (Baker Analyzed, reagent

Table 1. The doses of UVB, UVA and VIS irradiance received by the coastal Marsdiep water during the incubation of 18.5 hours. As comparison the doses of solar UVB and UVA irradiance detected nearby (within 300 m) at the Royal NIOZ during a whole day (14.75 hours of daylight) are given.

wavelength region	Incubation (kJ m ⁻²)	solar irradiance 8 th of May (kJ m ⁻²)
UVB	204.39	48.02
UVA	377.04	1170.71
VIS	919.24	1.98 10 ⁶

and UVA lamps during the incubation, but VIS lamps were included as well. Table 1 shows the doses of UVB, UVA and VIS received by the Marsdiep water during the incubation. The doses of solar UVB and UVA as measured by a hyperspectral radiometer (280-680 nm) containing a cosine collector (TriOS Optical Sensors) at the roof of the Royal NIOZ at 300 m from the Marsdiep sample station, and the dose of solar VIS as measured with TriOS Ramses ACC irradiation meter with cosine collector at the Marsdiep sample station, for the 8th of May 2003, are included for comparison. During the experiments an UVB irradiance dose 4 times higher, an UVA dose 3 times lower and a VIS dose 2200 times lower than the dose which estuarine seawater could receive during a day at the water surface (8th May 2003) were used (Table 1). Because we hereby ignored the effect of mixing and the strong light attenuation in these estuarine waters the applied UVB dose and probably also UVA irradiance are likely higher than *in situ* UVB and UVA doses.

A PUV 500 surface sensor (Biospherical Instruments) was used to measure the irradiance during the deck-incubation of the Scheldt seawater. The PUV measured irradiance between 9:15 and 16:05 (local time, 7th of April 2001). The irradiance values, given as dose, were measured at 305 nm (0.14 kJ m⁻² nm⁻¹), 320 nm (3.09 kJ m⁻² nm⁻¹), 340 nm (6.04 kJ m⁻² nm⁻¹), 380 nm (7.91 kJ m⁻² nm⁻¹) and VIS (400-700 nm, 20.49 E m⁻²). The deck-incubations of the Scheldt seawater were performed between 8:45 and 22:40 so the actual dose of irradiance received by the Scheldt sample was somewhat higher than measured with the PUV.

2.4 CLE-ACSV

Determination of the organic speciation of iron in the Marsdiep and Scheldt water was performed using CLE-ACSV. The 2-(2-Thiazolylazo)-p-cresol (TAC) (Aldrich, used as received) reagent was used as competing ligand (Croot and Johansson, 2000). All solutions were prepared using 18.2 MΩ nanopure water. The equipment consisted of a μAutolab voltammeter (Ecochemie, Netherlands), a static mercury drop electrode (Metrohm Model VA663), a double-junction Ag/saturated AgCl reference electrode with a salt bridge

grade) in 0.012 M (3xQD) HCl. Working solutions were prepared daily. All Fe(II) stock solutions were refrigerated in the dark at 4°C when not in use.

Calibration was performed by the standard addition of known concentrations of Fe(II) to the sample matrix. The standard addition for the experiments with Scheldt water was performed in Southern Ocean seawater because of a lack of sample. The time delay between Fe(II) addition and measurement caused an oxidation effect. This oxidation effect was accounted for by extrapolating the data back to time zero using the fact that Fe(II) oxidation in seawater very closely approximates pseudo-first-order kinetics.

2.6 Fe(III) analysis

Dissolved iron, defined as the Fe fraction passing an 0.2 µm filter, was determined using flow injection analysis with luminol chemiluminescence and H₂O₂ (de Jong et al., 1998).

3. Results and discussion

3.1 Iron speciation of the Marsdiep and Scheldt estuary samples

Marsdiep water was sampled during the last stage of a phytoplankton springbloom with low cell numbers caused by an early depletion of the nutrients phosphate and nitrate. The bloom developed from the 16th of March until the 15th of May, 2003. The Marsdiep water contained 25.4 nM dissolved iron and 24.4 eq nM Fe of dissolved ligands with a log K'_{FeL} of 21.0 ± 0.18 (n=4). Some 4 nM Fe is present as TAC-labile Fe (TAC-labile Fe being the Fe bound by 10 µM TAC after >12 hours equilibration) (Table 2). Samples from the Scheldt estuary contained concentrations dissolved Fe of 12-15 nM and a relatively high concentration of TAC-labile Fe: 11-12 nM. The Fe-binding ligand concentration was 4.6 eq nM Fe with a log K'_{FeL} of 20.1 (n=1) (Table 2).

Similar values for the log K'_{FeL} in coastal waters were reported by (Rose, 2003) using a kinetic approach and by Powell and Wilson-Finelli (2003) using CLE-ACSV. Croot and Johansson (2000) found similar values for the log K' and the initial concentration TAC-labile Fe during an algal bloom at Valö, Sweden. However, (Gobler et al., 2002) found extremely high conditional stability constants, log $K' \sim 23$, for organic iron binding ligands at similar salinities in the Peconic Estuary (Long Island, USA).

Table 2. Dissolved Fe, TAC-labile Fe, total ligand concentration and the logarithm of the conditional stability constant (K') for the Marsdiep-, Scheldt samples and the UV destruction of Marsdiep water. The $[Fe]_{dissolved}$ is the fraction iron determined in 0.2 μm filtered seawater as measured by flow injection analysis using luminol chemiluminescence (de Jong et al., 1998). The TAC-labile Fe is the labile iron concentration recoverable by adding 10 μM TAC, allowing the sample to equilibrate overnight, this value is an average of the first few values of the titration. The $[L]$, the concentration of Fe-binding ligand, and $\log K'_{FeL}$, the conditional stability constant of the iron binding ligand expressed with respect to Fe^{3+} , and their 95% confidence interval were estimated using non-linear least squares analysis (Systat) (Gerringa et al., 1995).

<i>samples</i>	$[Fe]_{dissolved}$ (nM)	[TAC-labile Fe](nM)	$[L]_{total}$ (nM)	$\pm 95\%$ confidence interval	$\log K'_{FeL}$	$\pm 95\%$ confidence interval
<i>Marsdiep samples</i>						
dark (t = 0)	25.39	4.30	22.89	0.78	21.23	0.27
light (t = 0)	25.42	4.46	25.89	0.82	20.83	0.11
dark (t = 18.5 h)	25.21	3.49	23.49	0.55	21.37	0.22
light (t = 18.5 h)	25.59	3.52	25.07	0.84	21.17	0.19
<i>Scheldt samples</i>						
dark	12.34	11.04	4.19	2.62	20.09	1.04
light	14.68	11.62	5.01	1.36	20.56	1.10
<i>UV destruction</i>						
A (t=0)	26.09	6.01	23.62	1.32	20.89	0.26
B (t=0)	26.61	6.15	22.73	1.41	21.03	0.40
A (t=10 h)	18.55	2.75	18.01	0.79	21.42	0.34
B (t=10 h)	18.38	2.39	17.92	0.51	21.41	0.21

3.2 TAC-labile Fe concentrations and their response to irradiance

In open ocean waters the concentration of ligands are always higher than the concentration of dissolved Fe (Gledhill and van den Berg, 1994; Rue and Bruland, 1995; van den Berg, 1995). This means that after equilibration (>12 hours) all dissolved iron is complexed by organic ligands and not available for TAC. Titration with Fe result typically in low TAC-labile Fe concentrations for the first few Fe additions in open ocean water (Figure 2).

The curvature in the titration curves of Marsdiep and Scheldt water (Figures 2, 3) indicating competition between the natural ligands and the added concentration TAC revealed the presence of strong ligands. The concentration of TAC-labile Fe of titrations with estuarine seawater was higher then observed for titrations of open ocean water.

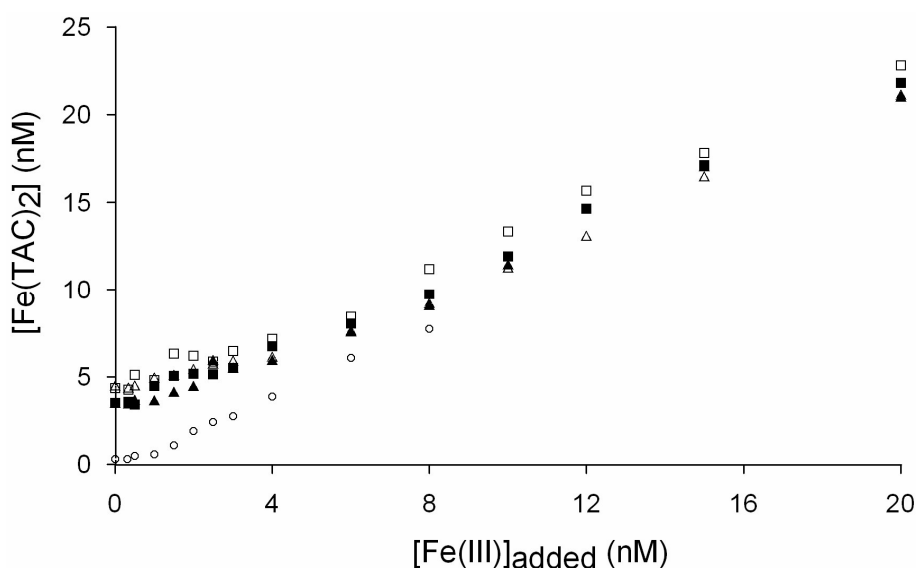


Figure 2. The CLE-ACSV Fe titration data from the Marsdiep and from Southern Ocean seawater (○). The titration curves for the coastal Marsdiep water were determined before and after the sample received 18.5 hours irradiance or darkness: $t = 0$ light (\triangle), $t = 0$ dark (\square), $t = 18.5$ h light (\blacktriangle) and $t = 18.5$ dark (\blacksquare).

Croot and Johansson (2000) reported TAC-labile Fe values of ~ 6.2 nM during a bloom and ~ 4.6 nM after a bloom (10 μ M TAC, overnight equilibration) at Valö, Sweden. The TAC-labile Fe concentrations determined during this study were between 3.5 and 4.5 nM for Marsdiep water and between 11 and 12 nM for water sampled in the Scheldt estuary.

The high TAC-labile Fe concentrations are explained by the existence of weak Fe-binding compounds in the coastal seawater samples. These weak Fe-binding ligands cannot compete with TAC and lose their Fe to TAC (Chapter 7). Furthermore, in competition with the strong Fe-binding ligands, TAC is able to catch some of the Fe initially bound by the natural strong ligands and so creating a pool of iron free ligands which were subsequently titrated (Chapter 7). This explains why the titration shows a curve in the presence of Fe bound by weaker ligands.

Comparing 1 kDa filtered Scheldt samples with 0.2 μ m filtered Scheldt samples showed us that the initial TAC-labile Fe fraction decreased with filter pore size, indicating that at least part of the TAC-labile Fe fraction had a relationship with the colloidal iron fraction (Figure 4).

The TAC-labile iron concentration in Marsdiep water decreased in time independent of the irradiance treatment (Figure 5). Marsdiep water incubated in the dark for 18.5 hours showed the same decrease in TAC-labile Fe as Marsdiep water incubated during 18.5 hours in the light. This time-dependent decrease was probably caused by degradation of weak organic iron binding ligands or by Fe-binding places related to colloidal material subject to

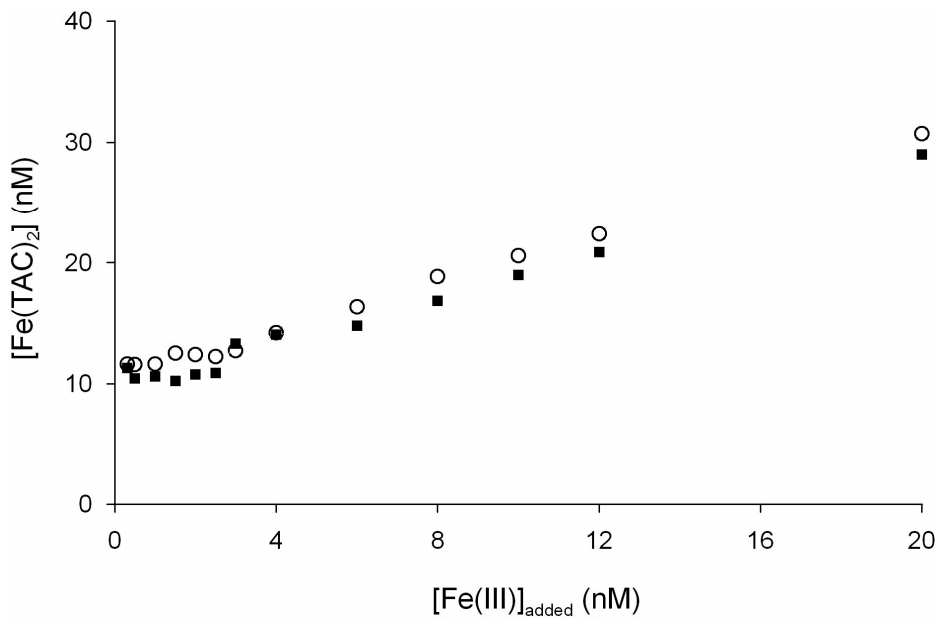


Figure 3. The CLE-ACSV Fe titration data from the Westerschelde. The titration curves for the Scheldt water were determined after a sample received 12 hours of daylight in spring (7th of April, 2001) (○) and after being incubated for 12 hours in the dark (■).

processes as ageing (Cornell and Schwertmann, 1996), adsorption of organic material (Kreller et al., 2003) and aggregation (Wells and Goldberg, 1993; Hunter et al., 1997).

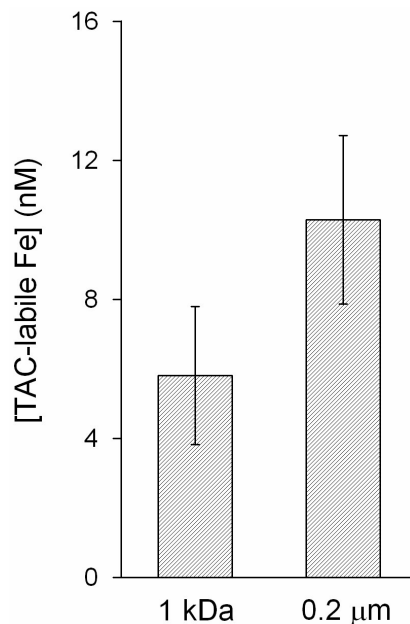


Figure 4. The TAC-labile Fe concentrations for two size fractions of Scheldt water; <1000 Da filtered (n=3) and <0.2 μm filtered (n=4).

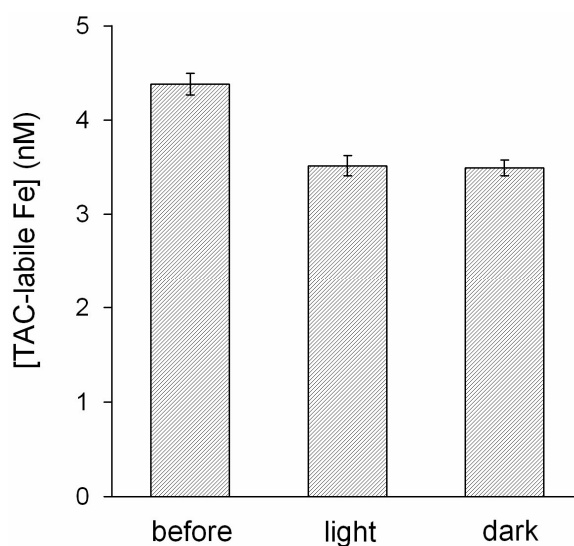


Figure 5. The TAC-labile Fe values determined for Marsdiep water, before and after incubation during 18.5 hours in the light or in the dark.

3.3 The influence of UV on the organic Fe-binding ligands

Irradiation with UVC (254 nm) during UV-destruction of the Marsdiep water resulted in a 22% decrease in the concentration of Fe(III) chelators. Also a decrease in the dissolved Fe concentration was observed (Figure 6A, Table 2). The high UV irradiance inducing the degradation of the Fe(III) chelators and increasing the concentration of free iron in the Marsdiep water probably caused an increase in inorganic Fe resulting in the precipitation of particulate iron to the bottom of the quartz tubes. Also the concentration TAC-labile Fe decreased during the UV-exposure (Figure 6B, Table 2). This indicates that particularly the weaker ligand class is UV degraded. The use of a high, unnatural UVC irradiance level, shows the Fe chemical effects of photo-induced degradation of Fe-binding ligands.

However, exposure of estuarine water sampled in the Marsdiep with simulated solar irradiance showed no significant decrease in the concentration organic Fe-complexing ligands or change in the $\log K'_{\text{FeL}}$, (Figure 2, 7). Additionally, organic Fe-complexing ligands in Scheldt water (S=26) showed no decrease in concentration after incubation under natural sunlight for a whole day as compared to incubation in the dark (Figure 3, 8).

Our results are consistent with the absence of vertical gradients of Fe(III) chelators in open ocean waters, i.e. the absence of minima within the shallow mixed layer minima, or any other feature which would suggest a surface/photochemical sink (Gledhill and van den Berg, 1994; Rue and Bruland, 1995; Wu and Luther, 1995; Moffett, 2001). On the other hand, it was reported recently that a weak Fe-binding ligand class ($\log K'$ 20-22, <1 kDa) in the Gulf

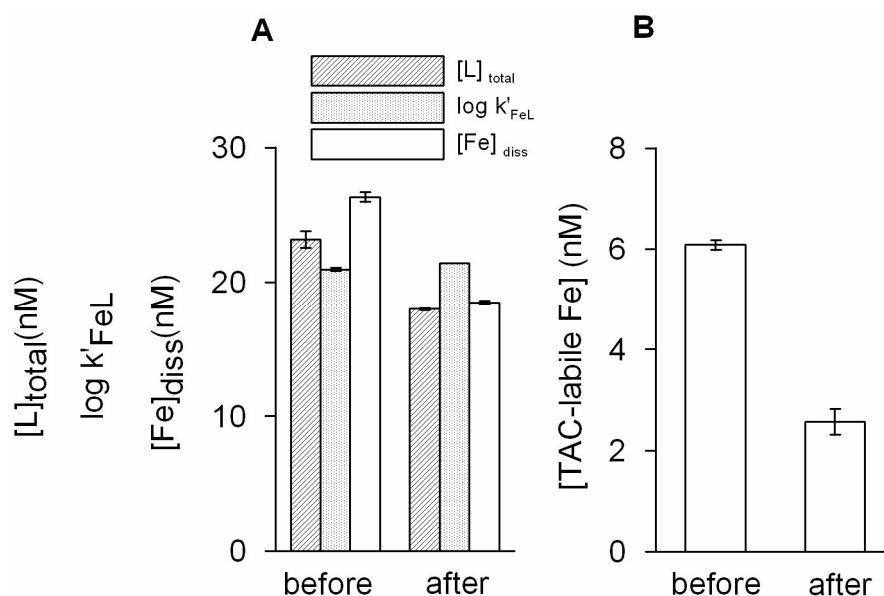


Figure 6. (A) The total concentration of Fe-binding ligand, $\log K'_{FeL}$ and the concentration dissolved Fe of Marsdiep water before and after 10 hours of UV destruction. (B) The TAC-labile Fe concentration before and after 10 hours of UV destruction of the Marsdiep water. The error bars give the standard errors ($n = 2$).

of Mexico is being photodegraded (Powell and Wilson-Finelli, 2003). Powell and Wilson-Finelli (2003) demonstrated that organic Fe-complexing ligands change little in the surface samples of the stratified Mississippi plume. Yet they explained the steady presence of the ligands with the continuous production of new ligands replacing the photo-degraded organic Fe-binding ligands. Production of organic Fe-binding ligands to compensate for the photo-degraded ligands can not explain the constant Fe-binding capacity in our experiments because phytoplankton and the majority of bacteria were filtered out.

Macrellis et al. (2001) had suggested larger iron binding siderophores to be photo-degraded to smaller Fe-binding molecules. This has been postulated to explain the detection of molecules with siderophore-like characteristics in a size class smaller than the defined size range for siderophores (300-1000 Da) in surface waters (Macrellis et al., 2001). Reduction of Fe(III) with photo-degradation of the siderophore and a significant drop in Fe(III) binding strength was also shown by Barbeau et al. (2001). The reason that we did not measure photo-degradation of the Fe-binding capacity could be that the overall input of photo-sensitive Fe(III) chelators was low. Related to this the photo-reactive fraction could already have been quickly photo-degraded during river transport or in the shallow well mixed coastal seawaters. Estuarine waters continuously receive new organic material originating from river input and

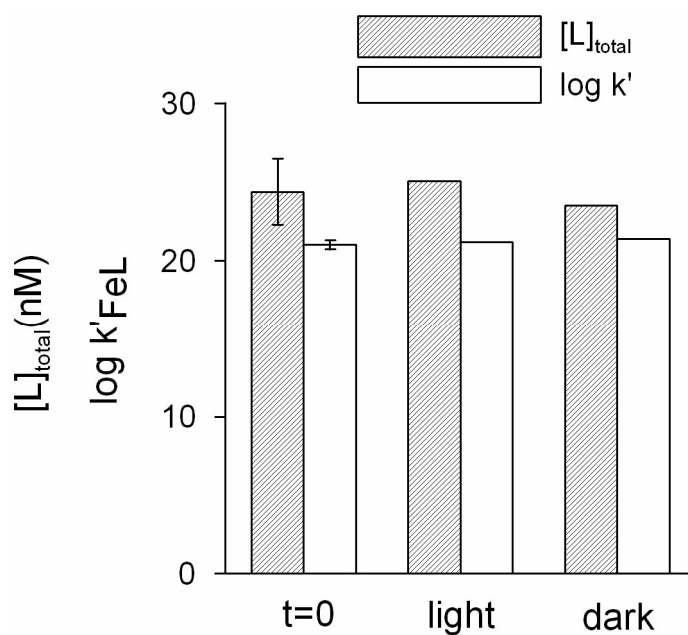


Figure 7. The total concentration of Fe-binding ligand and $\log K'_{FeL}$ for Marsdiep water at $t=0$ ($n=2$) and before and after 18.5 hours in the light ($n=1$) or in the dark ($n=1$).

biological activity. The fraction of photo-degradable Fe(III) chelators within this organic material appears to be low.

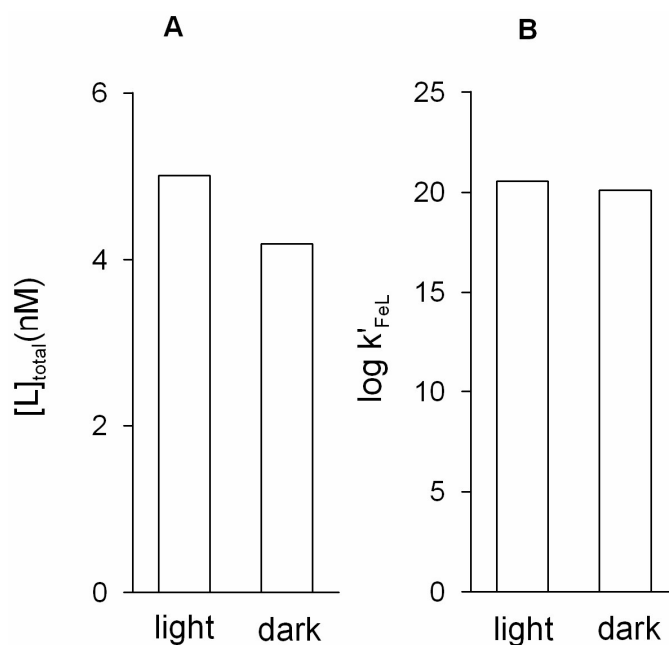


Figure 8. (A) The total concentration of Fe-binding ligand and (B) $\log K'_{FeL}$ of Scheldt water after 12 hours of natural sunlight or 12 hours in the dark.

3.4 Fe(II) photo-production in Marsdiep and Scheldt water

Although there was no significant influence of light on the photo degradation of the Fe(III) chelators, the Fe(II) was still photoproduced (Figure 9). The photo-production of Fe(II) in Marsdiep water was investigated under three different optical treatments (Figure 9). The 0.2 μm and 1 kDa filtered Scheldt water was only irradiated with the total spectrum (280-700 nm) (Figure 10).

The Fe(II) concentration in the dark for Marsdiep water was 43 ± 9 pM Fe(II) and for the Scheldt water 57 ± 4 pM Fe(II). Comparison of the Fe(II) production of the Marsdiep and Scheldt water receiving the same optical treatment showed a “steady state” value of 177 ± 8 pM for the 0.2 μm Marsdiep water and 239 ± 4 pM and 131 ± 8 pM for 0.2 μm and 1 kDa filtered Scheldt water respectively.

We have no knowledge about the nature and concentrations of the photo-reactive Fe(III) species but these experiments show that the concentrations Fe(II) formed during irradiation experiments were very low. The determined concentrations of FeL were a factor 100 higher than the concentration Fe(II) for the Marsdiep water and a factor of 20 higher than

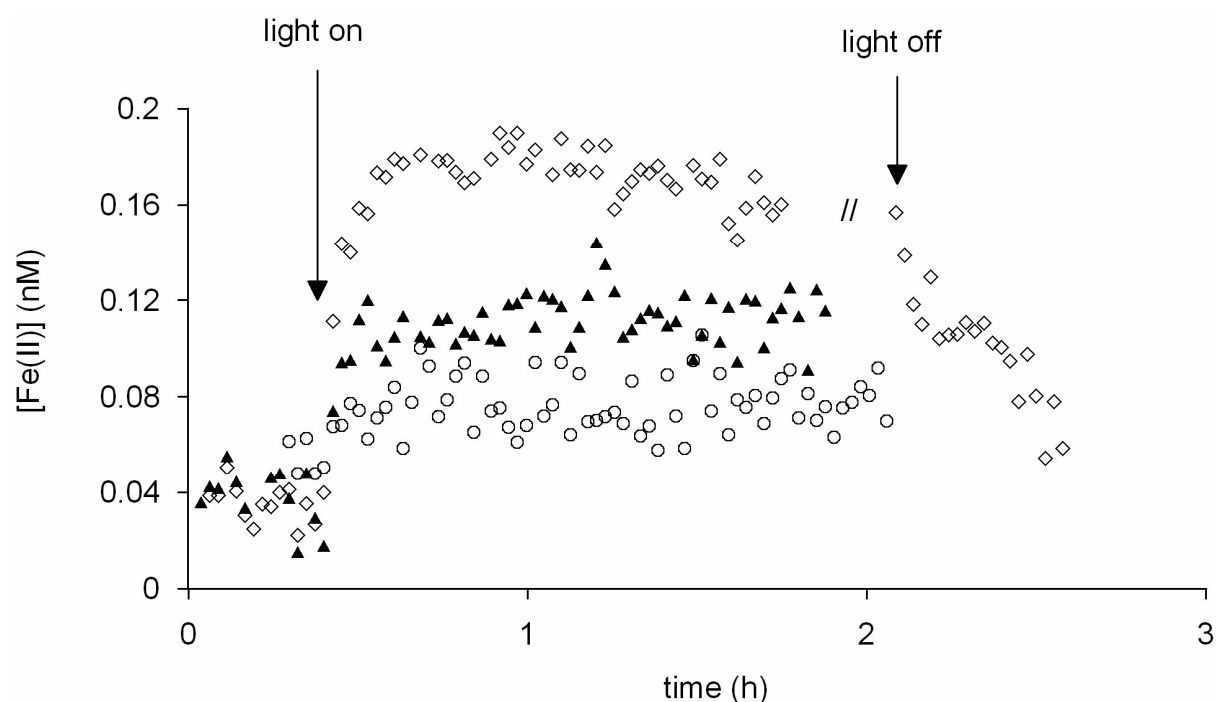


Figure 9. The concentration of Fe(II) formed during irradiation of Marsdiep water with three different wavelength regions: 280-700 nm (◇), 320-700 nm (▲) and 360-700 nm (○).

the concentration of Fe(II) in the Scheldt water. This means that the majority of the organic complexed iron is not involved in the photo-production of Fe(II).

The Fe(II) photoproduction in Marsdiep water showed the typical pattern seen during experiments with inorganic iron colloids (Waite and Morel, 1984; Wells and Mayer, 1991). The pattern observed when the light is on, namely a rapid increase until a maximum, followed by a slow decrease in the Fe(II) concentration during further continued irradiance, has been reported before in experiments using colloidal Fe (Wells and Mayer, 1991; Miller et al., 1995; Emmenegger et al., 2001). Wells and Mayer (1991) found that the photoconversion rates diminished with continued irradiation when ferrihydrite was irradiated. Waite and Morel (1984) found the same effect for the photodissolution rate of lepidocrocite. This pattern was explained by the presence of a limited amount of chromophoric colloidal Fe species (Wells and Mayer, 1991; Kimball et al., 1992; Chapter 3, Rijkenberg et al., 2005; Chapter 5). The occurrence of this similar pattern in the production of Fe(II) with time suggests that the produced Fe(II) could originate from a colloidal Fe fraction. Our results support this suggestion clearly when UVB was included in the irradiance spectra offered.

Moreover, we found that the production of Fe(II) was lower in the truly dissolved fraction (1 kDa filtered) of Scheldt water, 131 ± 8 pM Fe(II) compared to the Fe(II) produced in the dissolved fraction (0.2 μm filtered), 239 ± 4 pM Fe(II). This means that part of the Fe(II) produced results from a photo-reactive colloidal Fe fraction. We cannot conclude that the Fe(II) formed in the truly dissolved Scheldt water fraction is the result of a photo-reactive

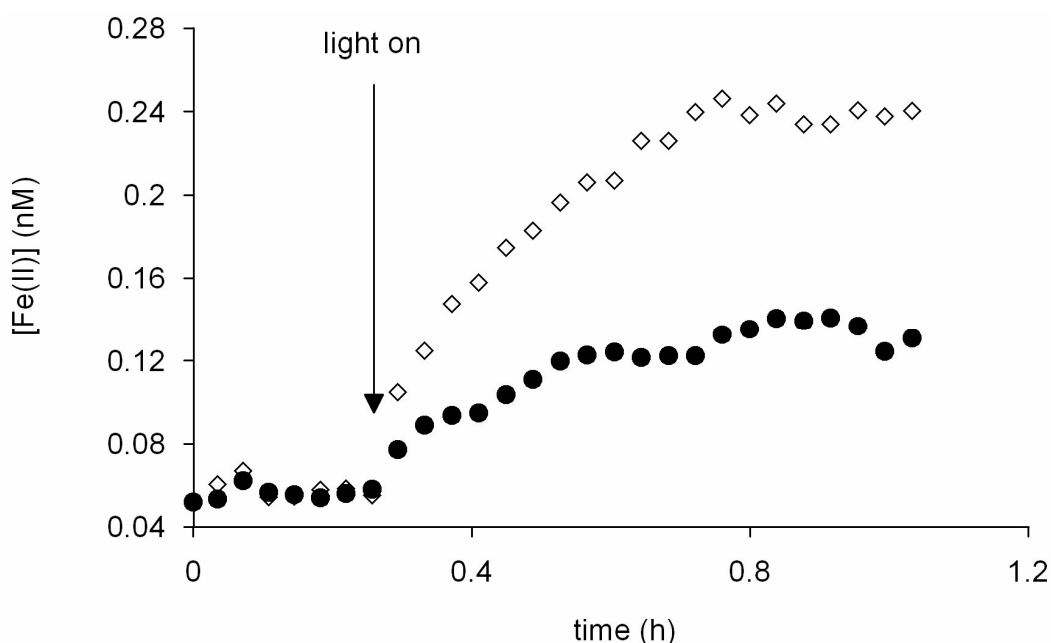


Figure 10. The concentration Fe(II) in Scheldt water formed upon irradiation of 0.2 μm filtered (\diamond) and 1 kDa filtered (\bullet) water with irradiance between the wavelengths 280 and 700 nm.

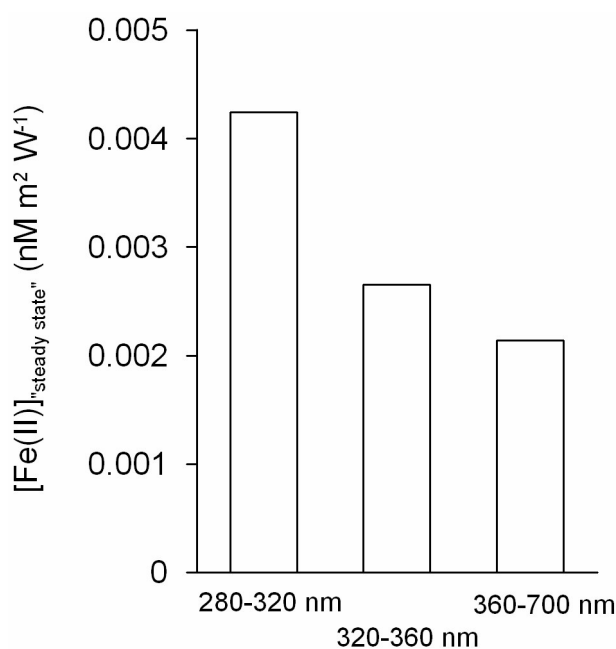


Figure 11. The mean Fe(II) concentration in Marsdiep at steady state/highest point due to the wavelength regions 280-320 nm, 320-360 nm and 360-700 nm normalised to 1 W m^{-2} .

organic complexed iron or of an inorganic truly soluble fraction, because colloids could have formed in this sample during sample handling in the laboratory. Enclosing a sample of (coastal) seawater in a container causes a decrease in the concentration of soluble iron and an increase in the concentration of particulate iron (Lewin and Chen, 1973). Another effect possibly increasing the colloidal iron fraction is the observation that iron is less soluble in warmer waters (Kuma et al., 1996). The colloidal iron fraction could increase when the estuarine seawater was kept in a warmer laboratory environment during various stages of the process.

Changing the wavelength range of the optical treatments of the Marsdiep water showed that less Fe(II) was formed upon excluding the lower wavelength regions UVB and UVA from the optical treatment. Comparing the values for the steady state normalized to 1 W m^{-2} between the different wavelength regions shows us that UVB produces most Fe(II), followed by UVA and VIS, respectively (Figure 11).

4. Summary and conclusion

We conclude from the present study that UV does not significantly photo-degrade the organic Fe-binding ligands in the waters of the Marsdiep and the Scheldt estuary. The UV light has no influence on the organically complexed Fe fraction and will not influence the transport of the dissolved organically complexed Fe phase to the coastal zone.

The concentrations of Fe(II) formed during irradiation experiments were very low (<240 pM). The concentrations of organic Fe-binding ligands are in the order of 24.4 nM for the Marsdiep and 4.6 nM for the Scheldt. Thus, the majority of the organically complexed iron in the Marsdiep water is not involved in the photo-production of Fe(II) and is not being photo-degraded during the process of photo-reduction of Fe bound to organic ligands. The pattern of Fe(II) production resembled the pattern observed during irradiation experiments with inorganic colloidal iron like ferrihydrite and goethite (Waite and Morel, 1984; Wells and Mayer, 1991) or amorphous iron oxyhydroxides (Chapter 5), suggesting that it might be a colloidal Fe fraction responsible for the photoproducted Fe(II).

Acknowledgements

We want to thank Patrick Laan (Royal NIOZ) and Jurjen Kramer (Royal NIOZ) for analysis of dissolved iron. Marcel Wernand (Royal NIOZ) is thanked for the irradiance data of the 8th of May 2003. Thomas Reinthaler (Royal NIOZ) and Rainer Amon (Alfred Wegener Institut für Polar und Meeresforschung, Germany; presently working at the Texas A&M University, Galveston, USA) are thanked for the 1 kDa filtrated seawater samples. The advice of Peter Croot (IfM, Kiel) is appreciated. This research is funded by NWO/NAAP grant number 85120004.

References

- Anderson, M.A. and Morel, F.M.M., 1980. Uptake of Fe(II) by a diatom in oxic culture medium. *Mar. Biol. Lett.*, 1: 263-268.
- Anderson, M.A. and Morel, F.M.M., 1982. The influence of aqueous iron chemistry on the uptake of iron by the coastal diatom *Thalassiosira weissflogii*. *Limnol. Oceanogr.*, 27(5): 789-813.
- Auclair, J.C., 1995. Implications of increased UV-B induced photoreduction - Iron(II) enrichment stimulates picocyanobacterial growth and the microbial food-web in clear-water acidic Canadian shield lakes. *Can. J. Fish. Aquat. Sci.*, 52(8): 1782-1788.
- Barbeau, K., Rue, E.L., Bruland, K.W. and Butler, A., 2001. Photochemical cycling of iron in the surface ocean mediated by microbial iron(III)-binding ligands. *Nature*, 413(6854): 409-413.
- Barbeau, K., Rue, E.L., Trick, C.G., Bruland, K.T. and Butler, A., 2003. Photochemical reactivity of siderophores produced by marine heterotrophic bacteria and cyanobacteria based on characteristic Fe(III) binding groups. *Limnol. Oceanogr.*, 48(3): 1069-1078.
- Boye, M., van den Berg, C.M.G., de Jong, J.T.M., Leach, H., Croot, P.L. and de Baar, H.J.W., 2001. Organic complexation of iron in the Southern Ocean. *Deep Sea Res. I*, 48(6): 1477-1497.
- Bruland, K.W., Rue, E.L. and Smith, G.J., 2001. Iron and macronutrients in California coastal upwelling regimes: Implications for diatom blooms. *Limnol. Oceanogr.*, 46(7): 1661-1674.
- Cornell, R.M. and Schwertmann, U., 1996. *The Iron Oxides*. VCH Publishers, New York.

- Croot, P.L., Bowie, A.R., Frew, R.D., Maldonado, M.T., Hall, J.A., Safi, K.A., la Roche, J., Boyd, P.W. and Law, C.S., 2001. Retention of dissolved iron and Fe-II in an iron induced Southern Ocean phytoplankton bloom. *Geophys. Res. Lett.*, 28(18): 3425-3428.
- Croot, P.L. and Johansson, M., 2000. Determination of iron speciation by cathodic stripping voltammetry in seawater using the competing ligand 2-(2-thiazolylazo)-p-cresol (TAC). *Electroanalysis*, 12(8): 565-576.
- de Baar, H.J.W. and de Jong, J.T.M., 2001. Distributions, sources and sinks of iron in seawater. In: D.R. Turner and K.A. Hunter (Editors), *The biogeochemistry of iron in seawater*. John Wiley & sons, New York, pp. 123-253.
- de Jong, J.T.M., den Das, J., Bathmann, U., Stoll, M.H.C., Kattner, G., Nolting, R.F. and de Baar, H.J.W., 1998. Dissolved iron at subnanomolar levels in the Southern Ocean as determined by ship-board analysis. *Anal. Chim. Acta*, 377(2-3): 113-124.
- Emmenegger, L., Schonenberger, R.R., Sigg, L. and Sulzberger, B., 2001. Light-induced redox cycling of iron in circumneutral lakes. *Limnol. Oceanogr.*, 46(1): 49-61.
- Finden, D.A.S., Tipping, E., Jaworski, G.H.M. and Reynolds, C.S., 1984. Light-induced reduction of natural iron(III) oxide and its relevance to phytoplankton. *Nature*, 309(5971): 783-784.
- Fox, L.E., 1988. The solubility of colloidal ferric hydroxide and its relevance to iron concentrations in river water. *Geochim. Cosmochim. Acta*, 52(3): 771-777.
- Gerringa, L.J.A., Herman, P.M.J. and Poortvliet, T.C.W., 1995. Comparison of the linear van den Berg Ruzic transformation and a nonlinear fit of the Langmuir isotherm applied to Cu speciation data in the estuarine environment. *Mar. Chem.*, 48(2): 131-142.
- Gledhill, M. and van den Berg, C.M.G., 1994. Determination of complexation of iron(III) with natural organic complexing ligands in seawater using cathodic stripping voltammetry. *Mar. Chem.*, 47(1): 41-54.
- Gledhill, M., van den Berg, C.M.G., Nolting, R.F. and Timmermans, K.R., 1998. Variability in the speciation of iron in the northern North Sea. *Mar. Chem.*, 59(3-4): 283-300.
- Glover, H.E., 1978. Iron in Maine coastal waters; seasonal variation and its apparent correlation with a dinoflagellate bloom. *Limnol. Oceanogr.*, 23: 534-537.
- Gobler, C.J., Donat, J.R., Consolvo, J.A. and Sanudo-Wilhelmy, S.A., 2002. Physicochemical speciation of iron during coastal algal blooms. *Mar. Chem.*, 77(1): 71-89.
- Hunter, K.A., Leonard, M.R., Carpenter, P.D. and Smith, J.D., 1997. Aggregation of iron colloids in estuaries: A heterogeneous kinetics study using continuous mixing of river and sea waters. *Colloid Surf. A-Physicochem. Eng. Asp.*, 120(1-3): 111-121.
- Hutchins, D.A. and Bruland, K.W., 1998. Iron-limited diatom growth and Si: N uptake ratios in a coastal upwelling regime. *Nature*, 393(6685): 561-564.
- Hutchins, D.A., DiTullio, G.R., Zhang, Y. and Bruland, K.W., 1998. An iron limitation mosaic in the California upwelling regime. *Limnol. Oceanogr.*, 43(6): 1037-1054.
- Hutchins, D.A., Witter, A.E., Butler, A. and Luther, G.W., 1999. Competition among marine phytoplankton for different chelated iron species. *Nature*, 400(6747): 858-861.
- Johnson, K.S., Coale, K.H., Elrod, V.A. and Tindale, N.W., 1994. Iron photochemistry in seawater from the equatorial Pacific. *Mar. Chem.*, 46(4): 319-334.

- Kimball, B.A., McKnight, D.M., Wetherbee, G.A. and Harnish, R.A., 1992. Mechanisms of iron photoreduction in a metal-rich, acidic stream (St-Kevin Gulch, Colorado, USA). *Chem. Geol.*, 96(1-2): 227-239.
- King, D.W., 1998. Role of carbonate speciation on the oxidation rate of Fe(II) in aquatic systems. *Environ. Sci. Technol.*, 32(19): 2997-3003.
- King, D.W., Lin, J. and Kester, D.R., 1991. Spectrophotometric determination of iron(II) in seawater at nanomolar concentrations. *Anal. Chim. Acta*, 247(1): 125-132.
- King, D.W., Lounsbury, H.A. and Millero, F.J., 1995. Rates and mechanism of Fe(II) oxidation at nanomolar total iron concentrations. *Environ. Sci. Technol.*, 29(3): 818-824.
- Kirchman, D.L., Meon, B., Cottrell, M.T., Hutchins, D.A., Weeks, D. and Bruland, K.W., 2000. Carbon versus iron limitation of bacterial growth in the California upwelling regime. *Limnol. Oceanogr.*, 45(8): 1681-1688.
- Kreller, D.I., Gibson, G., Novak, W., Van Loon, G.W. and Horton, J.H., 2003. Competitive adsorption of phosphate and carboxylate with natural organic matter on hydrous iron oxides as investigated by chemical force microscopy. *Colloid Surf. A-Physicochem. Eng. Asp.*, 212(2-3): 249-264.
- Kuma, K., Nishioka, J. and Matsunaga, K., 1996. Controls on iron(III) hydroxide solubility in seawater: The influence of pH and natural organic chelators. *Limnol. Oceanogr.*, 41(3): 396-407.
- Lewin, J. and Chen, C., 1973. Changes in the concentration of soluble and particulate iron in seawater enclosed in containers. *Limnol. Oceanogr.*, 18: 590-596.
- Liu, X.W. and Millero, F.J., 2002. The solubility of iron in seawater. *Mar. Chem.*, 77(1): 43-54.
- Macrellis, H.M., Trick, C.G., Rue, E.L., Smith, G. and Bruland, K.W., 2001. Collection and detection of natural iron-binding ligands from seawater. *Mar. Chem.*, 76(3): 175-187.
- Martin, J.M., Dai, M.H. and Cauwet, G., 1995. Significance of colloids in the biogeochemical cycling of organic-carbon and trace-metals in the Venice lagoon (Italy). *Limnol. Oceanogr.*, 40(1): 119-131.
- Martinez, J.S., Barbeau, K. and Butler, A., 2000. Marine bacterial siderophores. *Abstracts of Papers of the American Chemical Society*, 219: 522-INOR.
- Miller, W.L. and Kester, D., 1994. Photochemical iron reduction and iron bioavailability in seawater. *J. Mar. Res.*, 52: 325-343.
- Miller, W.L., King, D.W., Lin, J. and Kester, D.R., 1995. Photochemical redox cycling of iron in coastal seawater. *Mar. Chem.*, 50(1-4): 63-77.
- Millero, F.J., 1998. Solubility of Fe(III) in seawater. *Earth. Planet. Sci. Lett.*, 154(1-4): 323-329.
- Millero, F.J. and Izaguirre, M., 1989. Effect of ionic-strength and ionic interactions on the oxidation of Fe(II). *J. Solut. Chem.*, 18(6): 585-599.
- Millero, F.J. and Sotolongo, S., 1989. The oxidation of Fe(II) with H₂O₂ in seawater. *Geochim. Cosmochim. Acta*, 53(8): 1867-1873.
- Millero, F.J., Sotolongo, S. and Izaguirre, M., 1987. The oxidation-kinetics of Fe(II) in seawater. *Geochim. Cosmochim. Acta*, 51(4): 793-801.
- Moffett, J.W., 2001. Transformations among different forms of iron in the ocean. In: D.R. Turner and K.A. Hunter (Editors), *The Biogeochemistry of Iron in Seawater*. IUPAC series on analytical

- and physical chemistry of environmental systems. John Wiley & Sons, LTD, New York, pp. 343-372.
- Morris, A.W., Bale, A.J., Howland, R.J.M., Millward, G.E., Ackroyd, D.R., Loring, D.H. and Rantala, R.T.T., 1986. Sediment mobility and its contribution to trace-metal cycling and retention in a macrotidal estuary. *Water Sci. Technol.*, 18(4-5): 111-119.
- Nolting, R.F., Gerringa, L.J.A., Swagerman, M.J.W., Timmermans, K.R. and de Baar, H.J.W., 1998. Fe(III) speciation in the high nutrient, low chlorophyll Pacific region of the Southern Ocean. *Mar. Chem.*, 62(3-4): 335-352.
- O'Sullivan, D.W., Hanson, A.K. and Kester, D.R., 1995. Stopped-flow luminol chemiluminescence determination of Fe(II) and reducible iron in seawater at subnanomolar levels. *Mar. Chem.*, 49(1): 65-77.
- Powell, R.T., Landing, W.M. and Bauer, J.E., 1996. Colloidal trace metals, organic carbon and nitrogen in a southeastern US estuary. *Mar. Chem.*, 55(1-2): 165-176.
- Powell, R.T. and Wilson-Finelli, A., 2003. Importance of organic Fe complexing ligands in the Mississippi River plume. *Est. Coast. Shelf Sci.*, 58(4): 757-763.
- Powell, R.T. and Wilson-Finelli, A., 2003. Photochemical degradation of organic iron complexing ligands in seawater. *Aquat. Sci.*, 65(4): 367-374.
- Rich, H.W. and Morel, F.M.M., 1990. Availability of well-defined iron colloids to the marine diatom *Thalassiosira weissflogii*. *Limnol. Oceanogr.*, 35(3): 652-662.
- Rijkenberg, M.J.A., Fischer, A.C., Kroon, J.J., Gerringa, L.J.A., Timmermans, K.R., Wolterbeek, H.T. and de Baar, H.J.W., 2005. The influence of UV irradiation on the photoreduction of iron in the Southern Ocean. *Mar. Chem.*, 93: 119-129.
- Rose, A.L., Waite, T.D., 2003. Kinetics of iron complexation by dissolved natural organic matter in coastal waters. *Mar. Chem.*, xx: xx-xx.
- Rue, E.L. and Bruland, K.W., 1995. Complexation of iron(III) by natural organic-ligands in the central north Pacific as determined by a new competitive ligand equilibration adsorptive cathodic stripping voltammetric method. *Mar. Chem.*, 50(1-4): 117-138.
- Rue, E.L. and Bruland, K.W., 1997. The role of organic complexation on ambient iron chemistry in the equatorial Pacific Ocean and the response of a mesoscale iron addition experiment. *Limnol. Oceanogr.*, 42(5): 901-910.
- Sanudo-Wilhelmy, S.A., RiveraDuarte, I. and Flegal, A.R., 1996. Distribution of colloidal trace metals in the San Francisco Bay estuary. *Geochim. Cosmochim. Acta*, 60(24): 4933-4944.
- Seitz, W.R. and Hercules, D.M., 1972. Determination of trace amounts of iron (II) using chemiluminescence analysis. *Anal. Chem.*, 44: 2143-2149.
- Sholkovitz, E.R., 1978. Flocculation of dissolved Fe, Mn, Al, Cu, Ni, Co and Cd during estuarine mixing. *Earth. Planet. Sci. Lett.*, 41(1): 77-86.
- Sholkovitz, E.R., Boyle, E.A. and Price, N.B., 1978. Removal of dissolved humic acids and iron during estuarine mixing. *Earth. Planet. Sci. Lett.*, 40(1): 130-136.
- Steeneken, S.F., Buma, A.G.J. and Gieskes, W.W.C., 1995. Changes in transmission characteristics of polymethylmethacrylate and cellulose-(III) acetate during exposure to ultraviolet-light. *Photochem. Photobiol.*, 61(3): 276-280.
- Sunda, W.G. and Huntsman, S.A., 1995. Iron uptake and growth limitation in oceanic and coastal phytoplankton. *Mar. Chem.*, 50(1-4): 189-206.

- Takeda, S. and Kamatani, A., 1989. Photoreduction of Fe(III)-EDTA complex and its availability to the coastal diatom *Thalassiosira weissflogii*. Red Tides: Biol., Environ. Sci., Toxicol.: 349-352.
- van den Berg, C.M.G., 1995. Evidence for organic complexation of iron in seawater. Mar. Chem., 50(1-4): 139-157.
- Waite, T.D., 2001. Thermodynamics of the iron system in seawater. In: D.R. Turner and K.A. Hunter (Editors), The biogeochemistry of iron in seawater. IUPAC series on Analytical and Physical Chemistry of Environmental Systems. John Wiley & Sons, LTD, pp. 291-342.
- Waite, T.D. and Morel, F.M.M., 1984. Photoreductive dissolution of colloidal iron-oxide - effect of citrate. J. Colloid Interface Sci., 102(1): 121-137.
- Wells, M.L. and Goldberg, E.D., 1993. Colloid aggregation in seawater. Mar. Chem., 41(4): 353-358.
- Wells, M.L. and Mayer, L.M., 1991. The photoconversion of colloidal iron oxyhydroxides in seawater. Deep Sea Res. I, 38(11): 1379-1395.
- Wen, L.S., Santschi, P., Gill, G. and Paternostro, C., 1999. Estuarine trace metal distributions in Galveston Bay: importance of colloidal forms in the speciation of the dissolved phase. Mar. Chem., 63(3-4): 185-212.
- Wilhelm, S.W. and Trick, C.G., 1994. Iron-limited growth of cyanobacteria: multiple siderophore production is a common response. Limnol. Oceanogr., 39(8): 1979-1984.
- Witter, A.E., Hutchins, D.A., Butler, A. and Luther, G.W., 2000. Determination of conditional stability constants and kinetic constants for strong model Fe-binding ligands in seawater. Mar. Chem., 69(1-2): 1-17.
- Witter, A.E., Lewis, B.L. and Luther, G.W., 2000. Iron speciation in the Arabian Sea. Deep Sea Res. II, 47(7-8): 1517-1539.
- Witter, A.E. and Luther, G.W., 1998. Variation in Fe-organic complexation with depth in the northwestern Atlantic Ocean as determined using a kinetic approach. Mar. Chem., 62(3-4): 241-258.
- Wollast, R. and Peters, J.J., 1980. Transfer of materials in estuarine zones, Symposium on transport processes in estuarine and near-shore zones. International Council for the Exploration of the Sea. 68th Statutory Meeting, Copenhagen, Denmark, pp. 20 pp.
- Wu, J.F. and Luther, G.W., 1995. Complexation of Fe(III) by natural organic-ligands in the northwest Atlantic Ocean by a competitive ligand equilibration method and a kinetic approach. Mar. Chem., 50(1-4): 159-177.
- Xiao, C.B., Palmer, D.A., Wesolowski, D.J., Lovitz, S.B. and King, D.W., 2002. Carbon dioxide effects on luminol and 1,10-phenanthroline chemiluminescence. Anal. Chem., 74(9): 2210-2216.