

University of Groningen

An Overview of Hypoglycemia in Children Including a Comprehensive Practical Diagnostic Flowchart for Clinical Use

Casertano, Alberto; Rossi, Alessandro; Fecarotta, Simona; Rosanio, Francesco Maria; Moracas, Cristina; Di Candia, Francesca; Parenti, Giancarlo; Franzese, Adriana; Mozzillo, Enza

Published in:
Frontiers in endocrinology

DOI:
[10.3389/fendo.2021.684011](https://doi.org/10.3389/fendo.2021.684011)

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version
Publisher's PDF, also known as Version of record

Publication date:
2021

[Link to publication in University of Groningen/UMCG research database](#)

Citation for published version (APA):

Casertano, A., Rossi, A., Fecarotta, S., Rosanio, F. M., Moracas, C., Di Candia, F., Parenti, G., Franzese, A., & Mozzillo, E. (2021). An Overview of Hypoglycemia in Children Including a Comprehensive Practical Diagnostic Flowchart for Clinical Use. *Frontiers in endocrinology*, 12, [684011]. <https://doi.org/10.3389/fendo.2021.684011>

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: <https://www.rug.nl/library/open-access/self-archiving-pure/taverne-amendment>.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.



An Overview of Hypoglycemia in Children Including a Comprehensive Practical Diagnostic Flowchart for Clinical Use

Alberto Casertano^{1†}, Alessandro Rossi^{2,3†}, Simona Fecarotta^{2*},
Francesco Maria Rosanio¹, Cristina Moracas⁴, Francesca Di Candia¹,
Giancarlo Parenti^{2,5}, Adriana Franzese¹ and Enza Mozzillo^{1*}

OPEN ACCESS

Edited by:

Stefano Zucchini,
Sant'Orsola-Malpighi Polyclinic, Italy

Reviewed by:

Federico Baronio,
IRCCS S. Orsola-Malpighi, Italy
Vandana Jain,
All India Institute of Medical Sciences,
India

*Correspondence:

Enza Mozzillo
mozzilloenza@gmail.com
Simona Fecarotta
simona.fecarotta@unina.it;
simona.fecarotta@gmail.com

[†]These authors have contributed
equally to this work and
share first authorship

Specialty section:

This article was submitted to
Pediatric Endocrinology,
a section of the journal
Frontiers in Endocrinology

Received: 22 March 2021

Accepted: 31 May 2021

Published: 02 August 2021

Citation:

Casertano A, Rossi A, Fecarotta S,
Rosanio FM, Moracas C, Di Candia F,
Parenti G, Franzese A and Mozzillo E
(2021) An Overview of Hypoglycemia
in Children Including a Comprehensive
Practical Diagnostic
Flowchart for Clinical Use.
Front. Endocrinol. 12:684011.
doi: 10.3389/fendo.2021.684011

¹ Department of Translational Medical Science, Section of Pediatrics, Regional Center of Pediatric Diabetes, Federico II University of Naples, Naples, Italy, ² Department of Translational Medical Science, Section of Pediatrics, Metabolic Diseases Unit, Federico II University of Naples, Naples, Italy, ³ Section of Metabolic Diseases, Beatrix Children's Hospital, University Medical Centre Groningen, University of Groningen, Groningen, Netherlands, ⁴ Department of Translational Medical Science, Section of Pediatrics, Federico II University of Naples, Naples, Italy, ⁵ Telethon Institute of Genetics and Medicine, Pozzuoli, Italy

Hypoglycemia is the result of defects/impairment in glucose homeostasis. The main etiological causes are metabolic and/or endocrine and/or other congenital disorders. Despite hypoglycemia is one of the most common emergencies in neonatal age and childhood, no consensus on the definition and diagnostic work-up exists yet. Aims of this review are to present the current age-related definitions of hypoglycemia in neonatal-pediatric age, to offer a concise and practical overview of its main causes and management and to discuss the current diagnostic-therapeutic approaches. Since a systematic and prompt approach to diagnosis and therapy is essential to prevent hypoglycemic brain injury and long-term neurological complications in children, a comprehensive diagnostic flowchart is also proposed.

Keywords: neonatal hypoglycemia, childhood hypoglycemia, inborn errors of metabolism, endocrine hypoglycemia, glucose homeostasis, congenital hyperinsulinism

Abbreviations: AAP, American Academy of Pediatrics; ACTH, Adreno-CorticoTropic Hormone; AI, Adrenal Insufficiency; BAPM, British Association for Perinatal Medicine; CAH, Congenital Adrenal Hyperplasia; CAI, Central Adrenal Insufficiency; CDG, Congenital Disorders of Glycosylation; CH, Congenital Hyperinsulinism; CoA, Coenzyme A; CPT, Carnitine Palmitoyl-Transferase; CRH, Corticotropin-Releasing Hormone; DZX, Diazoxide; FAOD, Fatty Acid Oxidation Disorders; FBPase, Fructose 1,6 bisphosphatase; FFA, Free Fatty Acids; G6Pase, Glucose-6-Phosphatase; GALT, Galactose-1-phosphate uridylyltransferase; GDH, Glutamate Dehydrogenase Enzyme; GH, Growth Hormone; GHD, Growth Hormone Deficiency; GHRH, Growth Hormone-Releasing Hormone; GKD, Glycerol Kinase Deficiency; GLP-1, Glucagon-Like Peptide-1; GSD, Glycogen Storage Disease; HFI, Hereditary Fructose Intolerance; HY, Hypoglycemia; IGF1, Insulin Growth Factor1; IMD, Inborn Metabolic Disorder; IVA, Isovaleric Acidemia; KB, Ketone Bodies; LGA, Large for Gestational Age; MMA, Methylmalonic Acidemia; MRI, Magnetic Resonance Imaging; NBS, Newborn Screening; NGS, Next Generation Sequencing; OA, Organic Acidemias; OXPHOS, Oxidative Phosphorylation; PA, Propionic Acidemia; PAI, Primary Adrenal Insufficiency; PC, Pyruvate Carboxylase; PEPCCK, Phosphoenolpyruvate Carboxykinase; PES, Pediatric Endocrine Society; PET, Positron Emission Tomography; PG, Plasma Glucose; SGA, Small for Gestational Age; UCCS, Uncooked cornstarch; UDP, Uridine Di-Phosphate; UOA, Urine Organic Acids.

INTRODUCTION

Hypoglycemia (HY) in pediatric age shows some peculiarities regarding its diagnosis and management, mostly linked to age dependent features in glucose homeostasis and to the broad spectrum of causes. Such causes can initially present with the same unspecific picture, but they require different treatment (1). While being frequent but hard to detect in neonatal age, it is less common in infants and toddlers, even rarer in older children (1–3). In childhood, HY is a common metabolic-endocrine emergency possibly causing permanent neurological consequences. It is therefore essential to promptly detect and treat children with HY as well as those at risk. It is crucial to appropriately investigate its specific etiology for providing adequate and specific therapy (3, 4). In this review, we present current knowledge on management of HY in neonates and children including difficulties in establishing thresholds for both definition and therapeutic intervention and providing a comprehensive overall diagnostic approach through the use of a simple practical flowchart (5–8).

CONTROVERSIES ABOUT CLINICAL AND BIOCHEMICAL DEFINITION OF HYPOGLYCEMIA

Glucose is the primary energy source for central nervous system metabolism, independently from the feeding state (1). Several metabolic pathways cooperate to ensure normal blood glucose concentrations in the fasted state (**Figure 1**). Such pathways are tightly regulated by the hormonal (insulin, glucagon, cortisol, and growth hormone) and autonomic (catecholamines) response. In case of impaired metabolic pathways and/or altered hormonal regulation, glucose could become too low to satisfy neuronal demand, causing classical symptoms of HY. In pediatric age, both glucose homeostasis and clinical presentation of HY show peculiarities compared to adults. In newborns, the

adaptation to extrauterine life, characterized by immature hormonal and enzymatic pathways, and the higher glucose requirement of the brain, lead to a higher HY risk compared to older children and adults (9). Infants and children, have less glycogen storage and a higher substrates demand.

HY definition remains controversial in neonates and children. Some approaches define HY on the basis of symptoms, others on the plasma glucose value. In adolescents and adults, HY definition is based on the so-called “Whipple triad” [(I) symptoms of HY, (II) blood glucose level below 60 mg/dl, (III) resolutions of symptoms after glucose intake]. This definition appears inadequate for neonates and children in which symptoms are often subtle, and with the child being unable to communicate them (10). In addition, it is difficult to identify a single plasma glucose (PG) value below which symptoms of HY appear: in fact, symptoms appearance depends on additional factors, including the availability of alternative energy substrates (e.g. ketone bodies) and the severity, duration, and recurrence of low PG (7). Neurogenic symptoms are secondary to the neuroendocrine response, while neuroglycopenic symptoms are due to the low glucose availability to the brain. In neonates and infants, neurogenic and neuroglycopenic symptoms are not specific for HY. Therefore HY can be defined as the individualized condition in which PG concentration is low enough to cause symptoms and/or signs of impaired brain function (11).

Based on the above-mentioned considerations, three age-based different clinical scenarios exist:

- i. *Neonates <48 h of life*: signs or symptoms of neonatal HY may vary from severe (e.g. lethargy, tachypnea, hemodynamic instability, apnea, seizures, or even cardiac arrest) to milder (e.g. abnormal cry, decreased feeding, jitteriness, irritability, pallor, cyanosis, hypothermia, or diaphoresis) (12). In neonates showing specific symptoms, HY is diagnosed when PG is lower than a specific threshold: 47 mg/dl according to American Academy of Pediatrics (AAP) (13, 14), and 50 mg/dl according to Pediatric

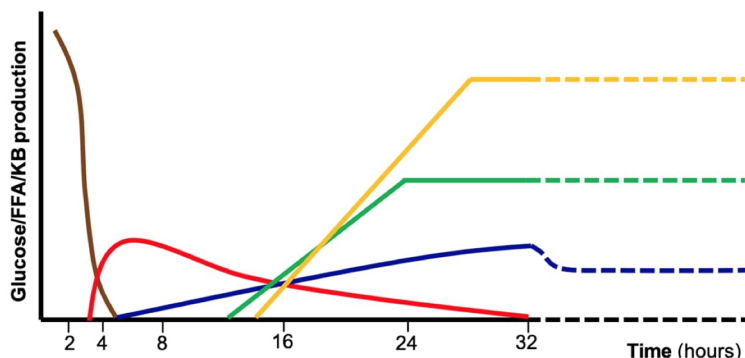


FIGURE 1 | Schematic representation of the major metabolic pathways involved in glucose homeostasis during absorptive phase and fasting including exogenous carbohydrates (brown), glycogenolysis (red), gluconeogenesis (blue), fatty acid oxidation (green), ketogenesis and ketolysis (yellow). These mechanisms are tightly controlled by hormonal regulation. Defects in specific enzymes or transporters involved in those pathways as well as endocrine disorders may result in fasting intolerance and hypoglycemia. FFA, free fatty acids, KB, ketone bodies.

Endocrine Society (PES) (7). A different glucose threshold has been proposed for pre-term newborns (15).

- ii. *Neonates >48 h of life, infants, and younger children unable to communicate*: HY is defined as PG <50–70 mg/dl (i.e. the normal threshold for neurogenic responses). Notably, recurrent PG levels in this range may cause the HY-associated autonomic failure, that in turn can attenuate HY autonomic symptoms (HY unawareness). Conflicting results on the definition of a safety glucose target level have emerged (16, 17). Currently, an acceptable threshold for this group is still considered 60 mg/dl (7).
- iii. *Older children able to communicate their symptoms*: for children who are able to communicate their symptoms, Whipple's triad could be adopted. In this age the clinical presentation is characterized by more specific symptoms as compared to neonates. In particular, the neuroglycopenic symptoms, due to the scarce availability of glucose for the central nervous system, and so indicative for lower PG level, are more clearly recognizable.

Signs and symptoms of HY are summarized and distinguished in neonatal and infant setting (**Table 1**). Neonatal HY cut off needing for intervention according with AAP, PES, BAPM are summarized in **Table 2**.

MANAGEMENT

During the first 48 h of life many healthy neonates could experience low PG, as a consequence of the physiologic adaptation to the extrauterine life. Sometimes it could last up to weeks being clinically relevant. So, it is crucial to identify as soon as possible neonates at risk of developing pathological low PG, and possibly distinguish between persistent and transient forms. Even though there is no consensus recommendation, HY persisting beyond the first 48 h, suggests a high risk of permanent pathology. Some authors consider the persistence over 7 days of HY and/or the need for more than 10 mg/kg/minute of

intravenous glucose infusion as indicative for persistent HY and anamnestic and clinical features could help to identify neonates at risk of persistent HY (18). Various glucose monitoring schemes have been proposed (**Table 3**) (18). A more practical and concise indication is that infants of diabetic mothers and LGA should be screened for 12 h after birth while SGA and preterm neonates should be screened for the first 24 h (14). Besides receiving a close glucose monitoring, at-risk neonates should be fed as soon as possible preferably with breast milk because it promotes ketogenesis, an alternate source of energy for the brain (19). Despite several interventional studies support the efficacy of the preventive use of dextrose oral gel, prior to the first hour (20–23), there is conflicting evidence regarding its ability to prevent the need for intravenous glucose infusion (24–27). Indeed, prompt treatment of HY is needed to quickly restore brain demand. Based on the severity of clinical manifestations, glucose should be administered intravenously or orally. Similarly to the diagnosis, several interventional thresholds for HY have been proposed (18, 28).

Two major age-based groups can be distinguished:

- i. *Neonates*: asymptomatic neonates who cannot maintain PG above 50 mg/dl (threshold for neuroglycopenic symptoms) after the first 48 h could be at risk of a persistent HY disorder. A structured intervention can only be indicated if the patient is symptomatic or has PG <60 mg/dl (threshold for neurogenic symptoms).
- ii. *Infants and children*: in case of mild to moderate HY, in infants/children able to take simple sugars orally, oral glucose has the quickest response (even in case of unknown etiology). Compared to sucrose, glucose leads to higher and earlier glycemic peak (29). Notably, in some metabolic conditions [e.g. hereditary fructose intolerance or defects in neoglucogenesis as Fructose 1,6 bisphosphatase (FBPase) deficiency] the administration of sugary drinks containing sucrose could considerably worsen the metabolic decompensation. In such conditions or in case of unknown etiology 10–20 grams of oral glucose are

TABLE 1 | Symptoms and signs of HY.

Age	Neuroglycopenic (PG < 30 mg/dl)	Neurogenic (PG < 55–65 mg/dl)
Newborn	Poor suck or poor feeding, weak or high-pitched cry, change in level of consciousness (lethargy, coma), seizures, hypotonia.	Jitteriness/tremors, pallor, sweating, irritability, tachypnea.
Infant–Child	Warmth, weakness, difficulty thinking, confusion, tiredness, drowsiness, coma, death.	<i>Cholinergic system</i> : sweating, hunger, tingling. <i>Adrenergic system</i> : shakiness, tremulousness, heart pounding, nervousness, anxiety.

TABLE 2 | Neonatal HY Cut Off needing for intervention according with AAP, PES, BAPM.

Guidelines	Asymptomatic	Symptomatic
American Academy of Pediatrics 2011	0–4 h: <25 mg/dl on two consecutive occasion 4–24 h: <35 mg/dl on two consecutive occasion	40 mg/dl
Pediatric Endocrine Society 2015	0–48 h: <50 mg/dl >48 h: >60 mg/dl	No clear cut off
British Association of Perinatal Medicine	Single value <18 mg/dl or two consecutive values <36 mg/dl	45 mg/dl

Modified from Kallern et al. (18).

TABLE 3 | Glucose monitoring for neonates at risk for HY.

Type of HY	Onset	Duration	HY degree	Response to glucose	Duration of monitoring	Examples
Early transitional adaptive	<6–12 h	12–24h	Mild	Good	24–48 h	Preterm infants, infant of diabetic mother, intrapartum glucose infusion, hypothermia
Secondary Classic transient neonatal	12–24 h 24–48 h	24–48 h 48–72 h or more	Mild Moderate to severe. About 80% are symptomatic	Good Requires often higher glucose infusion rate	24–48 h 48–72 h	Asphyxia, sepsis, intraventricular bleeding Small for gestation
Severe recurrent	Variable	>7 d	Severe	Requires higher glucose infusion rates >10–12	May be days to weeks	Congenital hyperinsulinism, metabolic and endocrine forms

Modified from Kallem et al. (18).

recommended, followed by a snack of starchy carbohydrates or a milk feed in infants (1, 30). In severe cases (when the patient is unconscious/unable to take anything orally) in an out of hospital setting and/or in the case of unavailable venous access, unless a diagnosis of a specific Inborn Metabolic Disorder (IMD) [e.g. Glycogen storage disease (GSD) type I] has been performed, glucagon (1 mg for children aged more than 12 years and/or weighing at least 25 kg, while 0.5 mg for younger/leaner) (1) should be used, due to its fast counter-insular action. Glucagon should be administered carefully because repeated/excessive doses may induce vomiting and so aggravate HY. Furthermore, it could be ineffective in case of long lasting HY or fasting, when liver glycogen stores may have already been depleted. In this case glucose must be infused starting with a bolus of 200–500 mg/kg (2–5 ml/kg of 10% glucose solution) followed by an infusion with 10% glucose adjusted to maintain euglycemia based on the age requirements (1).

ETIOLOGICAL DIAGNOSIS

Endocrine Causes

Congenital Hyperinsulinism

Congenital Hyperinsulinism (CH) represents the most common cause of persistent HY in infants and children, with an estimated incidence of 1:40,000–50,000 in general population. It is a heterogeneous and complex biochemical disorder characterized by the dysregulated insulin secretion from pancreatic β -cell causing random HY associated with low/normal ketones and absence of metabolic acidosis. Besides the classical neonatal onset, there are also late-onset forms that can appear in adolescence/adulthood (0.5–5.0% of cases) and could exhibit glycemic fluctuations from HY to hyperglycemia (31). In CH one or more steps of insulin secretion are disrupted due to a genetic defect, resulting into an insulin release that is independent from PG levels; sometimes, it is triggered by peculiar events, such as meal and exercise (31). CH genetic diagnosis could be achieved in about half of the patients. Besides syndromic conditions, currently about 14 genes are known to cause monogenic forms. Considering the known pathogenic mechanisms, currently CH could be grouped into four categories (32):

- i. *Channel Defects (ChD; genes ABCC8, KCNJ11, KCNQ1, CACNA1D)*: among these mutations, ABCC8 and KCNJ11 (KATP channel subunits Kir6.2 and SUR1, respectively) causes the most common and severe forms of CH especially in case of biallelic mutation, although there have been reported patients carrying ABCC8 biallelic mutations with optimal response to Diazoxide (DZX) (33, 34) even showing progressive resolution of hypoglycemia (35).
- ii. *Metabolic Defects (MeD; genes GLUD1, GCK, HADH, UCP2, HK1, PMM2, PGM1)*: this class includes enzyme defects causing abnormal intracellular levels of specific metabolites regulating insulin release. Among these mutations, GLUD1 activating result into increased glutamate dehydrogenase enzyme (GDH) activity and cause the Hyperinsulinism/Hyperammonemia syndrome (the second most common cause of CH). Since GDH is allosterically enhanced by Leucine, protein load can induce HY. HY due to dominantly inherited by GCK activating mutations is clinically heterogeneous with respect to severity and age of onset.
- iii. *Transcription factors Defects (TfD; genes HNF1 α , HNF4 α , FOXA2)*: this class includes molecular defects in the transcriptional factors that regulate the glucose-induced secretion of insulin. Patients affected by these mutations are subjected to transient hyperinsulinemic HY followed by the development of Maturity Onset Diabetes of the Young during adolescence (36, 37).
- iv. *Syndromic conditions*: CH could be a manifestation of several syndromic conditions (**Table 4**).

From the histopathological point of view, CH is classified into three variants (31, 38). In diffuse forms, all β -cells share the same molecular defect and show the same morphology. In focal forms, a β -cell cluster develops as a nodular adenomatous hyperplasia because of a confined molecular defect in the 11p15.1-11p15.5 imprinted region, that involve the ABCC8/KCNJ11 genes. These forms usually develop sporadically in a patient carrying a recessive ABCC8/KCNJ11 paternally inherited mutation, when a somatic loss of the maternal allele occurs (“double hit”). ABCC8 mutations can cause phenotypes who switch from HY in infancy to hyperglycemia in adolescence and even adulthood (39–41). Late onset forms are mostly linked to dominant mutations of ABCC8/KCNJ11 genes or to activating mutation of GCK gene (40). CH diagnosis may be suspected at any insulin

TABLE 4 | Syndromic causes of CH.

Overgrowth syndromes	Gene	Chromosome	Inheritance
Beckwith–Wiedemann syndrome	IGF2/H19 CDKN1C KCNQ1OT1	11p15.5–15.4	Autosomal dominant Sporadic Paternal uniparental disomy
Sotos syndrome	NSD1 NFIX	5q35.2–35.3 19p13.3	Autosomal dominant Sporadic
Simpson–Golabi–Behmel syndrome	GPC3	Xq26	X-linked
Perlman syndrome	DIS3L2	2q37	Autosomal recessive
Chromosomal abnormality syndromes			
Turner syndrome	KDM6A	Xp11.2	Sporadic
Trisomy 13	CDX2, IPF	Trisomy 13	Sporadic
Postnatal growth failure syndromes			
Kabuki syndrome	KMT2D KDM6A	12q13.12 Xp11.3	Autosomal recessive Sporadic
Costello syndrome	HRAS	11p15.5	Autosomal dominant Sporadic
Contiguous gene deletion affecting the ABCC8 gene			
Usher syndrome	USH1C	11p15.1	Autosomal recessive
Timothy syndrome	CACNA1C	3p21.1	Autosomal dominant Sporadic
Insulin receptor mutation			
Insulin resistance syndrome (Donohue syndrome)	INSR	19p13.2	Autosomal recessive
Congenital Disorders of Glycosylation (CDG)			
CDG Type Ia	PMM2	16p13.2–13.3	Autosomal recessive
CDG Type Ib	PMI	15q22–24	Autosomal recessive
CDG Type Ic	hALG3	3q27	Autosomal recessive
Other causes			
Poland syndrome	UCMA	10p13–14	Sporadic
CHARGE syndrome	CHD7	8q12	Autosomal dominant

Modified from Galcheva S, et al. (32).

concentration detectable in a hypoglycemic plasma sample thus as a marker of inappropriate insulin secretion (1, 42). Indirect signs of this phenomenon are the absence of ketonemia (excepted for HADH deficit) and fatty acidemia. Adjunctive diagnostic criteria for CH could be a positive response to glucagon or octreotide injection (glucose levels >1.5 mmol/L) and the need for more than 8 mg/kg/min of glucose infusion to maintain euglycemia (1, 32, 38, 40, 41). Some authors (42) have proposed a classification of CH diagnostic criteria as reported in the **Table 5**. Once the diagnosis of CH has been established, genetic test should be performed; as a general rule, ABCC8/KCNJ11 mutations must be investigated first. Concurrently to genetic tests, a prompt treatment should be started with diazoxide (DZX) and then, in case of unresponsiveness, with octreotide (38, 41, 42, 44). In fact, many ABCC8/KCNJ11 mutations cause refractoriness to DZX. These cases require an 18-Fluoro-DOPA-Positron Emission Tomography (PET) to search for any focal forms, and need to be treated with octreotide. Dosage adjustment could be required due to possible tachyphylaxis. Side effects include abdominal discomfort, diarrhea (rarely necrotizing enterocolitis), and in long term, bile sludge/gallstones and suppression of pituitary hormones. Long-acting release (LAR) octreotide analogues, administered monthly, have also been successfully tried in children (45, 46) and even preferable for better compliance and safety (47), however they take a long time to achieve the steady state (lanreotide needs 23–30 days and sandostatin 3 months to gain therapeutic blood concentration). They should initially be administered together with octreotide: the starting

TABLE 5 | Diagnostic criteria of CH.

Cardinal diagnostic criteria	Low plasma glucose (<3 mmol/L) with Detectable serum insulin Detectable C-peptide
Biochemical criteria	Suppressed/low beta-hydroxybutyrate and acetoacetate Suppressed/low serum free fatty acid
Clinical criteria	Increased requirement of glucose infusion rate (>8 mg/kg/min) Positive response to i.m./i.v. glucagon (glycemic response >1.5 mmol/L)
Supportive criteria	Positive response to s.c./i.v. octreotide Low serum levels of IGFBP-1 (suppressed by insulin) Suppressed branch chain amino acids Normal lactic acid Normal plasma hydroxybutyrylcarnitine Normal ammonia Appropriate counterregulatory hormone response (cortisol >20 mcg/dl, GH > 7 ng/ml) Provocation test (leucine loading or exercise testing) may be needed in some patients

Modified from Vora et al. (43).

dose is 30–60 mg for lanreotide (subcutaneously) while for sandostatin-LAR (intramuscularly) the dose is equivalent to the cumulative 31-day subcutaneous octreotide dose, calculated by multiplying the daily dose of octreotide (5–25 µg/kg) for 31. Other drugs proposed for the treatment of DZX un-responsive cases include Nifedipine, Sirolimus, and Glucagon-like peptide-1 (GLP-1) receptor antagonist “Exendin” (1, 42). In particular, Sirolimus, is an antiproliferative drug that reduces insulin secretion probably by lowering the mammalian target of rapamycin (mTOR), a serine/threonine kinase that is overexpressed in the diffuse variant of CHI and which enhances insulin secretion; moreover, Sirolimus causes depletion of intracellular Calcium decreasing insulin release (48). This drug has shown variable efficacy and safety being even able to avoid surgery up to 18 months, but sometimes ineffective both for duration and power to reduce hypoglycemic events (49). Although there haven't been reported major side effects, it should be used with caution for its immunosuppressive action and long-term follow-up studies are needed for chronic toxicity (42). Therapy effectiveness in CH patients is based on blood glucose monitoring. The tools used for this monitoring are the same used for the glycemic monitoring of type 1 diabetes patients (50) such as blood glucose sampling and continuous subcutaneous glucose monitoring (51). According with the most recent evidences, 18-Fluoro-DOPA PET should be offered not only in case of refractoriness to DZX, but also when no mutation is identified or in case of a single recessive paternal inherited mutation in *ABCC8/KCNJ11*, that could reveal focal forms, even if responsive to DZX. While surgery could be definitively curative for focal forms, near-total pancreatectomy, reserved for diffuse forms unresponsive to available drugs, could cause iatrogenic diabetes and allocate patients to life-long pancreatic enzyme replacement and insulin therapy. Moreover, the residual pancreatic tissue left near the common bile duct and along the duodenum could be responsible for persistent HY. Data from long-term follow-up show efficacy in prevention of severe hypoglycemic episodes but with only few cases of remission (52).

Adrenal Insufficiency

Adrenal insufficiency (AI) is a life-threatening condition in which the adrenal cortex is unable to adequately produce steroid hormones. AI can be distinguished in Primary (PAI) or Central (CAI), depending on the impairment of adrenal cortex or hypothalamus/pituitary gland respectively. The most common etiology of PAI in children is the Congenital Adrenal Hyperplasia (CAH) due to 21-hydroxylase deficiency. In PAI both glucocorticoids and mineralocorticoids synthesis are affected, while in CAI only glucocorticoid synthesis is compromised (53, 54).

Glucocorticoids, mostly cortisol, play an essential role in glucose metabolism (55): in the liver they promote glucose output by activating gluconeogenesis and triglycerides accumulation (56–59); in the muscle they suppress glucose uptake and glycogen synthesis, accelerate protein breakdown, and inhibit protein synthesis; in adipose tissue they promote lipolysis and so increase serum FFA and glycerol (60) thus, together with amino-acids coming from protein catabolism, they provide substrates for gluconeogenesis. Cortisol deficiency results in incapacity to raise up glucose levels in stressful

conditions, causing HY associated with low/normal ketones and absence of metabolic acidosis.

Clinical presentation of PAI may be non-specific with anorexia, weight loss, fatigue, abdominal pains, headache, nausea, arthralgia, myalgia, joint pain in chronic forms (61) or with an “adrenal crisis,” characterized by a cardiovascular decompensation due to massive impaired electrolyte and fluid balance, in case of acute onset (61). The critical sample collected during an adrenal crisis will show hyponatremia, metabolic acidosis (normal anion gap, increased serum chloride), HY, hyperkalemia, and low cortisol level.

Congenital Hypopituitarism and Growth Hormone Deficiency

Congenital Hypopituitarism is a pathologic condition characterized by a partial or a total deficiency in one or more pituitary hormones (62). Among those conditions, ACTH and GH deficiency (GHD) may present with HY associated with low/normal ketones and absence of metabolic acidosis. ACTH deficiency has been already described above.

Although the cause of HY in hypopituitarism is still debated, it is known that GH and cortisol, in physiologic doses, act synergistically to elevate the blood glucose, and that the replacement of both hormones is necessary to normalize insulin secretion and maintain normal glucose homeostasis in children with hypopituitarism; probably GHD is responsible of HY because of loss of amino acid mobilization to support gluconeogenesis (63, 64).

Congenital Hypopituitarism should be suspected in neonates carrying dysmorphic features with midline defects, ocular and craniofacial anomalies, and in males, micropenis often with undescended testes. Detection of persistent HY and jaundice reinforces the suspect (65–68). GHD could also appear in evolutive age mostly with its auxological consequences: short stature, delayed bone age, decrease in the growth rate (69).

Metabolic Disorders

Glycogen Storage Diseases (GSD)

All Metabolic disorders are resumed in **Table 6** and **Supplementary Table 1**.

GSD are secondary to defects of the enzymes and transporters involved in glycogen breakdown and synthesis. Their overall incidence is 1:25,000 births. More than 12 GSD types are recognized. Based on clinical presentation, they are classified as hepatic GSD (e.g. GSDI) and muscle GSD (e.g. GSDII, GSDV). HY and hepatomegaly are the primary manifestation of the hepatic GSD (GSD0a, GSDI, GSDIII, GSDVI, GSDIX, GSDXI). GSDIII is the only GSD presenting with concomitant liver and muscle involvement. Based on the ketone levels hepatic GSD are traditionally defined as ketotic (GSD0a, GSDIII, GSDVI, GSDIX, GSDXI) or non-ketotic (GSD I). Genetic studies are the preferred method for diagnosing hepatic GSD (enzyme tests are performed in selected cases). Dietary plan with frequent feedings and uncooked cornstarch (UCCS) and/or tube feeding are the cornerstone of the treatment for hepatic GSD; carbohydrates are given to maintain euglycemia, but excessive carbohydrate

TABLE 6 | Main clinical and biochemical features of major metabolic causes of childhood HY.

Disorder	Timing	Lactate	Ketones	Additional biochemical abnormalities	Clinical features
Ketotic hypoglycemia	Fasting >6 h	+/- M.A.	+	Low alanine	Fever Vomiting Diarrhea
Glycogen storage disease type I	Fasting (2–4 h)	+	(-)	Elevated lipids Elevated uric acid Elevated transaminases Neutropenia (GSDIb)	Hepatomegaly Doll-like face
Glycogen storage disease type III/VI/IX	Fasting (2–6 h)	-	+	Elevated lipids Elevated transaminases Elevated CK (GSDIIIa)	Hepatomegaly Cardiomyopathy (GSDIIIa)
Hereditary Fructose Intolerance	1–2 h after the ingestion of fructose, sucrose, sorbitol	+	+/-	Elevated transaminases	Vomiting Diarrhea Hepatomegaly Liver failure Fatty liver
Galactosemia	1–2 h after the ingestion of galactose, lactose	- M.A.	+/-	Elevated bilirubin Elevated transaminases Abnormal clotting tests	Vomiting Hepatomegaly Liver failure Cataract Sepsis
Fructose 1,6 bisphosphatase deficiency	Fasting >6 h	+	+	Elevated alanine	Intercurrent disease
		M.A.		Elevated pyruvate and glycerol 3-phosphate	Hepatomegaly
Pyruvate carboxylase deficiency	Variable	+	+	Hyperammonemia	Severe encephalopathy
		M.A.		Elevated citrulline	Seizures Movement disorders
Organic acidemias	Prolonged fasting or after an initial symptom-free period (neonatal)	+	+	Hyperammonemia Elevated branched chain amino acids and glycine Elevated acylcarnitines	Encephalopathy Movement disorders Renal disease
		M.A.		UOA abnormalities	Cardiomyopathy
Fatty acid oxidation disorders	Fasting >8 h	+	(-)	Elevated acylcarnitines	Exercise intolerance
		M.A.		FFA/KB > 2.5	Cardiomyopathy Arrhythmias
Ketogenesis defects	Prolonged fasting or after an initial symptom-free period (neonatal)	+	-	Hyperammonemia	Hepatomegaly
		M.A.		FFA/KB > 2.5	Seizures
Ketolysis defects	Prolonged fasting	-	+	UOA abnormalities	Cardiomyopathy
				FFA/KB < 0.3	Intercurrent disease Hepatomegaly
Disorders of Oxidative Phosphorylation	Variable	+	+/-	UOA abnormalities	Multisystem involvement
Congenital Disorders of Glycosylation	Variable	+/-	+/-	High insulin (mostly)	Psychomotor retardation Dysmorphic features Multisystem involvement

M.A., metabolic acidosis.

intake may result in hyperinsulinemia with consequent complications (70). Carbohydrates restriction with protein supplementation or ketogenic diets are recommended to avoid glycogen storage and to minimize insulin secretion in some forms (71). Restricted fructose and galactose intake aims at avoiding acidosis in GSDI. Major hepatic GSD are discussed.

GSD I is the most common and severe GSD (both glycogenolysis and gluconeogenesis are impaired). It is due to a defect of either the catalytic (GSDIa, 80% of cases) or the microsomal glucose 6-phosphate transporter (GSDIb, 20% of cases) of the G6Pase system. GSDI patients usually present at 3–6 months of age with fasting HY, lactic acidosis and hypoketosis

(usually 2–4 h after meal), hepatomegaly, doll-like face, failure to thrive, hyperlipidemia, and hyperuricemia. Additionally, GSDIb patients show neutropenia and recurrent infections (72). Long-term complications include liver neoplasms, renal disease, and increased risk of inflammatory bowel disease (73) and autoimmune (74, 75) or endocrine disorders (76–78).

GSDIII is due to glycogen debrancher enzyme deficiency. Two main subtypes are recognized: GSDIIIa (85% of the cases, mixed liver and muscle involvement) and GSDIIIb (15% of the cases, isolated liver involvement). As gluconeogenesis is intact, HY is usually less severe than GSDI showing prominent fasting ketosis without lactic acidosis. Transaminases concentrations are usually higher (may exceed 1,000 U/L) with less severe hyperlipidemia compared to GSDI. Bone disease (78) and benefit of a high-fat diet on muscle symptoms have been reported (79).

GSDVI and GSDIX are secondary to liver glycogen phosphorylase and glycogen phosphorylase kinase defect, respectively. They are generally mild disorders improving with age. However, they can also present with symptomatic fasting ketotic HY, hyperlipidemia, increased transaminases, hepatomegaly, growth retardation, and hypotonia (80).

GSD0 is caused by a deficiency of hepatic glycogen synthase resulting in inadequate production of hepatic glycogen. The clinical manifestations include fasting ketotic hypoglycemia accompanied by low levels of alanine and lactate and postprandial hyperglycemia and hyperlactatemia. Unlike other GSDs, patients with GSD0 usually do not develop hepatomegaly (81, 82).

GSDXI (Fanconi-Bickel syndrome) is caused by deficiency in a solute carrier family 2 protein (GLUT-2) that is expressed in hepatocytes and proximal renal tubule. Patients typically present at 3–10 months of age with hepatomegaly, Fanconi syndrome (e.g. severe glycosuria, polyuria, hyperaminoaciduria, hypophosphatemic rickets, acidosis, hypokalemia, hypochloremia), failure to thrive, fasting HY, and postprandial hyperglycemia. Only symptomatic treatment is available (frequent feeds with complex carbohydrates, electrolytes replacement, vitamin D) (5).

Hereditary Fructose Intolerance

HFI is caused by deficiency of Aldolase B, resulting into inhibition of gluconeogenesis (inhibition of Aldolase A) and glycogenolysis (inhibition of glycogen phosphorylase A) secondary to fructose 1-phosphate accumulation. Symptoms usually present at weaning, after the ingestion of food containing fructose, sucrose, or sorbitol (e.g. fruit, vegetables) and include post-prandial HY, with ketosis and lactic metabolic acidosis, hepatomegaly, vomiting, pallor, sweating, lethargy, failure to thrive, convulsions, and eventually coma. Acute liver failure and renal dysfunction (proteinuria, glycosuria, hyperaminoaciduria) are also observed. Most patients develop a natural aversion to fruit/sweets. Therapy includes avoidance of dietary fructose, sucrose, and sorbitol (5), although mild signs of liver injury, without progression on a long-term follow-up could be detected in patients on a FSS-free diet, particularly with specific genotypes (83).

Galactosemia

Classical galactosemia is caused by deficiency of galactose-1-phosphate uridyltransferase (GALT) (the enzyme converting lactose into glucose and galactose) resulting into accumulation of galactose 1-phosphate, galactitol, and galactonate in blood and tissues. Symptoms usually appear a few days after the ingestion of breast or formula milk and include vomiting, diarrhea, poor feeding, nuclear cataract, jaundice, hepatomegaly, and high transaminases evolving to liver failure (HY, bleeding tendency) and renal failure; *Escherichia coli* sepsis is common.

Inherited Disorders of Gluconeogenesis

The conversion of pyruvate into glucose is the central pathway of gluconeogenesis. Overall, disorders of gluconeogenesis present with recurrent HY and lactic acidosis with or without ketosis. Major inherited disorders of gluconeogenesis are described.

FBPase deficiency is a disorder of gluconeogenesis characterized by episodic acute crisis of HY, lactic acidosis (lactate may rise up to 25 mmol/L), and (usually) ketosis manifesting with hyperventilation, apneic spells, hepatomegaly (with normal transaminases), seizures, coma, and brain damage. The crises are likely to occur when glycogen reserves are limited (as in newborns or after ingestion of large amount of fructose) or exhausted (e.g. fasting, intercurrent illness) and are reversed by high glucose infusion rates (about 1.5 times maintenance). The frequency of the attacks decreases with age and patients are usually well between attacks. Treatment includes frequent feedings and avoidance of prolonged fasting (84).

Pyruvate carboxylase (PC) deficiency is a defect of both gluconeogenesis and Krebs cycle. Although fasting HY can occur, this disorder usually presents with severe encephalopathy, developmental delay, seizures, movement disorders, failure to thrive, and metabolic acidosis. A high lactate to pyruvate ratio with a low hydroxybutyrate to acetoacetate ratio is suggestive of the diagnosis. Treatments include intravenous glucose infusion, bicarbonate, dietary management, and supplementation with citrate, aspartate, dichloroacetate, biotin, and thiamine (85).

Phosphoenolpyruvate carboxykinase (PEPCK) deficiency affects gluconeogenesis and can cause HY, failure to thrive, lactic acidosis, and lipid accumulation in the kidney and liver. Only six patients have been reported in the literature and its clinical relevance is currently disputed (86).

Glycerol kinase deficiency (GKD) can present either isolated or together with congenital adrenal hypoplasia or Duchenne muscular dystrophy (partial deletion of Xp21). Patients with isolated GKD can develop episodic vomiting with HY, hyperketonemia, metabolic acidosis, and coma. Typically, high glycerol excretion in the urine is found by gas chromatography-mass spectrometry. Metabolic crises should be avoided by providing an adequate supply of fluid, calories, and glucose during intercurrent illness (87).

Congenital Disorders of Glycosylation

CDG constitute a group of conditions due to defects in the glycoprotein synthesis. Around 90 CDG types are currently recognized. Phosphomannomutase 2 Deficiency and Glucosyltransferase 1 Deficiency are the most common CDG.

A broad spectrum of symptoms including psychomotor retardation, failure to thrive, hypotonia, deafness, bleeding tendency, cerebral hemorrhage, cardiomyopathy, hypogonadism, and HY (hyper- or normoinsulinemic) is known (88).

Fatty Acids Oxidation Disorders

FAODs constitute a group of conditions characterized by hypoketotic HY and presenting with great variability. Three typical presentations are known for FAOD:

- i. Acute hypoketotic HY with lactic acidosis and encephalopathy with hepatomegaly and liver dysfunction (including hyperammonemia); symptoms usually present under catabolic circumstances (e.g. newborn, prolonged fasting, intercurrent illness)
- ii. (Hypertrophic) cardiomyopathy and arrhythmias
- iii. Myopathy presenting with weakness and/or acute rhabdomyolysis with symptoms precipitated by exercise or intercurrent illness (89).

Diagnosis can be suggested by acylcarnitine profile and confirmed by enzyme testing or gene sampling (90). Treatment includes a high carbohydrate diet to maintain euglycemia and to avoid prolonged fasting or stress induced states. Carnitine supplementation can be used for specific FAOD (91).

Disorders of Ketone Body Metabolism

Disorders of KB metabolism can present either in the first days of life or later in childhood. Similarly to FAOD, prolonged fasting and intercurrent illness are triggers to metabolic decompensation. Ketogenesis defects are characterized by hypoketotic HY with or without hyperammonemia, metabolic acidosis, and liver disease. Decompensations lead to encephalopathy, vomiting, and coma. Conversely, ketolysis defects present with episodes of hyperketotic HY and severe ketoacidosis in childhood; patients are healthy between episodes (92).

Disorders of Oxidative Phosphorylation

Disorders of Oxidative Phosphorylation (OXPHOS) are clinically, biochemically, and genetically heterogeneous. They are due to mutations in nuclear genes coding for respiratory complexes subunits and can present at any age with a wide range of possible symptoms, including fasting HY with lactic acidosis and variable ketone bodies levels. Children often suffer from encephalomyopathic disease (93).

Organic Acidemias

OA are disorders of intermediary metabolism due to defect of enzymes involved in branched-chain amino acid catabolism. They are characterized by the mitochondrial accumulation of CoA metabolites causing metabolic acidosis, elevated lactate, ketotic HY, and hyperammonemia. The most common OA are Methylmalonic acidemia (MMA), Propionic acidemia (PA), and Isovaleric acidemia (IVA).

Three clinical presentations are recognized:

- i. Neonatal (intoxication type): lethargy, poor feeding, encephalopathy, myoclonic jerks, multiorgan failure.
- ii. Chronic intermittent: episodes of ketoacidosis, lethargy, cerebral involvement.
- iii. Chronic progressive: vomiting, failure to thrive, psychomotor retardation, hypotonia, renal disease.

OA are diagnosed by their specific urinary organic acid profiles or abnormal plasma acylcarnitines.

The diagnosis is confirmed with enzymatic studies and/or molecular DNA testing (94). OA are included in NBS programs in several countries with an increasing number of patients diagnosed pre-symptomatically (95). Treatment of the acute phase is aimed at correcting hyperammonemia (by temporary stopping protein intake, promoting anabolism, and administering ammonia scavengers), metabolic acidosis, and HY. The cornerstone of chronic treatment are protein-restricted diet, long-term ammonia scavengers, vitamin cofactors, and carnitine supplementation (carnitine transforms toxic CoA esters into less toxic carnitine esters).

Idiopathic Ketotic HY

Ketotic HY is the most common cause of childhood HY. It usually presents between 18 months and 5 years and resolves spontaneously by the age of 9 years. Typically, the child presents with symptomatic HY in the morning after long fast often precipitated by an intercurrent illness. Glucose <55 mg/dl and massive ketosis are observed. Metabolic acidosis can also develop. The child improves dramatically on dextrose infusion (conversely glucagon injection elicits little or no increase in glucose concentrations) and is usually restored to normal health within hours. Despite being the most common cause of HY in childhood, there are no specific diagnostic tests for ketotic HY. Therefore, all possible causes of HY must be ruled out (diagnosis of exclusion). Treatment measures include avoidance of prolonged fasting, UCCS, and close monitoring of oral intake when in stressed states (such as illness) to avoid HY (2).

DIAGNOSTIC PATHWAY: PRACTICAL APPROACH TO HY IN CHILDHOOD

The diagnostic path of HY results from the combination of medical history and clinical, dietary, and biochemical data. A systematic approach is necessary to collect relevant information.

Personal History

The first step consists in collecting information on the timing of the hypoglycemic event, including:

- Age of onset (neonatal, infant, child)
- Fasting tolerance (e.g. feeding frequency, night snack, morning ketosis)
- Temporal relation with meals (fasting, post-prandial, random)
- Relation to/Avoidance of food [e.g. protein, fruit, fruit (juice), (ga)lactose]

- Associated conditions/triggers
- Recurrence (e.g. intercurrent disease, fatigue)

The following information should also be carefully detailed:

- Perinatal history: birth weight, gestational age, gestational diabetes, and any other form of perinatal distress and perinatal glucose requirements (e.g. >10 mg/kg/min glucose intravenously).
- Growth and developmental milestones (e.g. intellectual disability, movement disorders, epilepsy)
- Family history: relatives with HY/hyperglycemia or IMD, previous miscarriages or deaths, consanguinity, medications, and social history.

Physical Examination

Physical examination can reveal signs pointing to:

- Endocrine dysfunction, such as micropenis, short stature, midline anomalies (hypopituitarism), skin hyperpigmentation, abdominal pain, muscle pain, weight loss, signs of hyperandrogenism [Adrenal Insufficiency (AI)]
- Inherited metabolic disease, such as hepato(spleno)megaly (since the liver and spleen size become larger with age, patients' age and height should be considered for adequate assessment), jaundice, spider angiomas (e.g. GSD), cataract (e.g. galactosemia), absence of dental caries [e.g. Hereditary Fructose Intolerance (HFI)], arrhythmias and/or heart murmur [e.g. Fatty Acid Oxidation Disorders (FAOD)], movement disorders [e.g. Organic Acidemias (OA)], hypotonia and inverted nipples and/or bleeding tendency [e.g. Congenital Disorders of Glycosylation (CDG)], multisystem involvement (e.g. mitochondrial disorders)
- Dysmorphic features, macrosomia, hemihypertrophy (e.g. CH, genetic syndromes of overgrowth)

Laboratory Investigations

Biochemical tests can provide crucial information to reach a final diagnosis. Indeed, specific biochemical patterns can point to specific defects. In particular, low ketones at the time of hypoglycemia may immediately suggest a diagnosis of hyperinsulinism or FAOD. Contextual levels of Free Fatty Acids (FFA) could help to distinguish between endocrine or metabolic etiology, as low ketones together with low FFA suggest hyperinsulinism, while low ketones together with increased FFA suggest FAOD or defects of ketolysis (when FFA/KB ratio results below 0.3) (**Table 6**).

A "critical sample" (i.e. a sample obtained during HY) must be collected. Laboratory investigations should include: blood glucose, lactate, ketones (mainly beta-hydroxybutyrate), blood gases, FFA, insulin, C-peptide, cortisol, GH, Insulin, Growth Factor1 (IGF1), acylcarnitines, amino acids, as well as urinary organic acids (UOA). Some investigations may not be performed in all hospitals. Therefore, one or two spare tubes should also be collected for any additional tests to be performed afterwards. However, appropriate blood samples might be missed when an immediate treatment is required (e.g. severe HY). Still, collecting

(and store frozen) the first urine sample after HY might provide helpful information in such cases (e.g. increased/undetectable ketones, lactate, tricarboxylic acids).

Additional tests can be considered: ammonia, toxicology tests, urine reducing substances test (to assess fructosuria, galactosuria). Transferrin electrophoresis/isoelectric focusing should be required if a CDG is suspected. Over the past years, a number of minimally invasive continuous glucose monitoring systems have also become available, possibly providing additional information on the extent, timing, and duration of PG fluctuations.

Fasting test. Evaluation of metabolic changes after fasting may be helpful to reach the diagnosis and to assess patients' fasting tolerance to tailor the treatment. Since fasting can lead to the accumulation of toxic metabolites and sometimes fatal complications in some defects, a fasting challenge should only be performed in specialized metabolic units and only after less risky investigations have been performed without reaching a clear diagnosis (FAOD must be ruled out before a fasting test). The maximal duration of the fasting is based on the clinical suspicion and on the children's age (usually 12–16 h at 6–12 months, 18 h at 1–2 years, 20 h at 2–7 years, 24 h in children >7 years). The fasting should be stopped at any time if glucose concentration is below 2.6 mmol/L (47 mg/dl). Since newer diagnostic strategies (biochemistry and molecular biology) are rapidly becoming available, fasting test is not performed routinely; however, it can be helpful in selected cases (96).

Glucagon test explores the response of glucagon injection during HY to assess the availability of glycogen for compensation of low blood glucose. Typically, HY due to GSDI does not benefit from glucagon injection (with worsening hyperlactatemia) while an exaggerated glucose response to glucagon could be observed in case of CH. Due to its possible risks (prolonged HY) it has been largely superseded by enzyme or mutation analysis.

Alternative causes of abnormality of the test results should always be ruled out (e.g. lactate elevation secondary to laborious sampling or increased pCO₂ secondary to apnea during blood collection).

Additional Investigations

Imaging tests can provide additional information. Abdominal ultrasound and Magnetic Resonance Imaging (MRI)/Computed tomography/Scintigraphy scan can define liver, spleen, pancreas, and kidneys morphology and structure (e.g. liver steatosis, liver adenomas, focal hyperplasia). Left hand and wrist X-ray can be helpful in patients with growth retardation. Specific additional investigations may be performed based on the accompanying clinical features (e.g. cardiac ultrasound, brain MRI).

HY COMPREHENSIVE FLOWCHART

The combination of the aforementioned information enables reaching a working diagnosis in most of the children presenting with HY. To date no consensus exists on a standardized diagnostic flowchart. Several algorithms with various starting points and workflow have been proposed (5, 7, 8). Although such algorithms can provide metabolic or endocrine specialists with

specific pathophysiological insight, the two main groups of causes (namely endocrine and metabolic) may appear not clearly suited to the understanding and use of generalist pediatricians, which, on the other hand, are often the first level of observation of HY phenomena. Our center has a long-standing experience of cooperation between metabolic and endocrine experts in the management of childhood HY. In this respect, a comprehensive diagnostic flowchart is proposed (**Figure 2**). The major advantage of such flowchart is the ability to orient the diagnose, distinguishing both (main) metabolic and endocrine causes of childhood HY by using simple, routinely available tests such ketone bodies, emogas analysis, and lactate. As a matter of fact, data included in this flowchart can be easily implemented by physicians in a hospital setting, to obtain biochemical findings at the time of hypoglycemia, that should not be missed and that could be very useful to hypothesize the diagnosis. In order to provide adequate information, physicians should be aware that the flowchart applies to results collected on a “critical sample.” Possible limitations include the lack of rarer disorders (e.g. PEPCK deficiency, GKD, CDG) and the inability to diagnose uncommon presentations of common disorders.

Only the main diagnostic features that guide bedside diagnosis about the most common causes of pediatric HY are shown in the flowchart (e.g., hyperlactatemia is also found in OXPHOS defects and GSD I; hepatomegaly is also found in FBPase deficiency and GSD I). Firstly: the timing of HY is the starting point; the patients fasting tolerance can provide an essential clue to the diagnosis in children with fasting HY (e.g. HY after a short fast suggests hepatic GSD, HY after moderate to long fast suggests gluconeogenesis defects or FAOD/KB defects). Secondly: laboratory investigations play a pivotal role to reach a working diagnosis. In this respect, assessing the

presence of (un) detectable ketones (as well as metabolic acidosis, hyperlactatemia and, if possible, FFA) on a “critical sample” is of paramount importance. Thirdly: the presence of hepatomegaly can help differentiating disorders causing fasting ketotic HY.

So far, the resultant flowchart seems to facilitate the logical process leading to the diagnostic suspicion and help to address the biochemical and clinical elements that need to be sought. The subsequent diagnostic process is up to the specialists of the two endocrine and metabolic sectors.

In case of HY in otherwise healthy children and/or with no recognizable pattern, intoxications/factitious causes should always be ruled out by toxicological tests on blood and urine (most common drugs include insulin, sulfonylurea, beta-blockers, salicylates).

Diagnosis can be confirmed through enzymatic and/or molecular testing for IMD and CH and challenge tests for endocrine disorders. Enzyme diagnostics is generally performed on blood cells or skin fibroblast (e.g. debranching enzyme or very-long chain acetyl-CoA dehydrogenase activity). However, some enzymes (e.g. G6Pase) are not expressed in these mediums and require a liver biopsy. Since liver biopsy is invasive, it has been largely superseded by DNA analysis.

THE RECENT ROLE OF NGS

DNA analysis has become increasingly sophisticated and rapid in recent years. Various techniques are used to search for mutations in IMD/CH genes; single gene analysis (Sanger sequencing) has been traditionally used to confirm a specific diagnostic suspicion, after a traditional work-up. When a group of disease is considered, the traditional diagnostic approach would involve a long process with subsequent gene-by-gene molecular analyses.

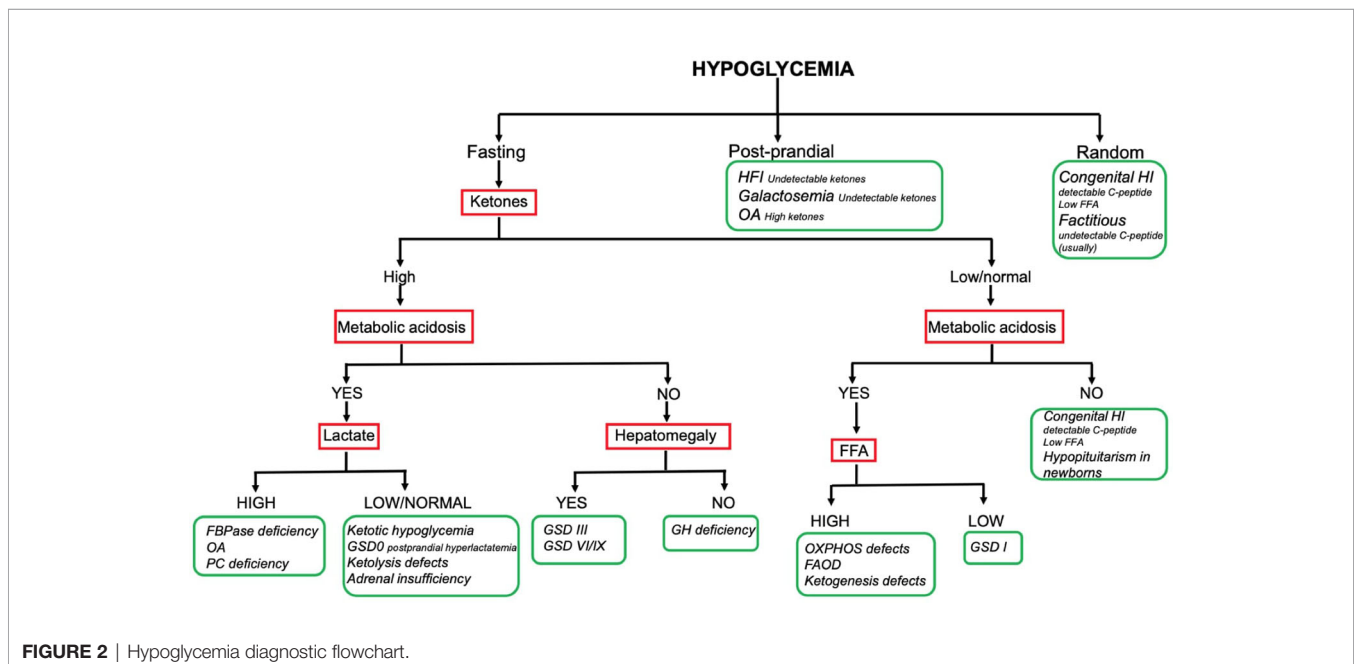


FIGURE 2 | Hypoglycemia diagnostic flowchart.

The gene-by-gene technique has now been superseded and replaced by the analysis of panels with NGS techniques. The introduction of NGS represents a major advancement in the diagnostic approach, allowing in parallel sequencing of millions of small fragments of DNA. Given the difficulties in the diagnostic workup in HY and due to the overlapping of clinical manifestations in several disorders of glucose metabolism, patients showing recurrent undiagnosed HY could be further investigated with an NGS-based approach. This modern technique has the potential to identify underrecognized rare disorders in the wide group of children with ketotic hypoglycemia, clinically diagnosed in the past as affected by benign idiopathic hypoglycemia. In addition to a targeted approach with gene panels, the NGS technology can be used through untargeted strategies based on whole-exome sequencing, having this approach also the potential to identify new genes involved in disorders of glucose metabolism (97).

DISCUSSION AND CONCLUSIVE REMARKS

Despite being a common emergency in pediatrics (3, 4), there are still controversies on the definition and management of HY in neonates and children. Neither the standard diagnostic PG threshold nor the operative threshold are defined. Also, no consensus on the definition of at-risk neonates exists. Such uncertainties together with the broad spectrum of causes, make the approach to HY in childhood complex and time consuming. Irrespective of its cause, prompt recognition and treatment of acute HY are critical to prevent its complications (namely brain damage). Bolus administration of dextrose (either intravenously or orally) is the cornerstone of the treatment. Glucose requirements may vary depending on the patient's age (e.g. higher in neonates) and disease (e.g. up to 10–15 mg/kg/min of glucose or more can be required in children with CH). Since additional treatments can be necessary for specific disorders (e.g. specific dietetic approach, drugs and cofactors in OA and FAOD or DZX in CH) a timely etiological diagnosis is crucial. Once acute HY has been managed, pending the results of confirmatory tests (e.g. enzyme/DNA diagnostics) specific actions should be taken in order to prevent HY relapse. As a general recommendation, fasting must be avoided and adequate carbohydrate intake must be maintained during any metabolic stress. Tailored dietary treatment plan with frequent feedings and UCCS and/or tube feeding are the most common interventions, such a plan aims at ensuring glucose concentrations as stable as possible and is generally sufficient in patients with ketotic HY, hepatic GSD, and disorders of KB metabolism. Additional dietary interventions may be required for specific IMD (e.g. life-long fructose-, sucrose- sorbitol-restricted diet regimen in HFI, or low-protein diet in OA). Irrespective of their final diagnosis, in acute situations (e.g. intercurrent illness, prolonged fasting) patients can become catabolic, due to (the combination of) high fever, a reduced intake, and/or increased losses. Therefore, it is important to know what to do in emergency situations. An

emergency protocol is designed at this purpose (98). Patients (and caregivers) should be encouraged to always carry an emergency protocol with them and follow its instructions. As HY can be secondary to a variety of different disorders, a systematic multidisciplinary approach is ideal in caring for neonates and children with HY. Interestingly, there is no consensus on standardized diagnostic algorithms for childhood HY. Therefore, a comprehensive practical diagnostic flowchart (including the main endocrine and metabolic causes) is proposed to guide the diagnostic suspicion, highlighting a minimal set of clue clinical and biochemical findings at the time of HY, that can be easily investigated in any hospital, at any time of the day. In fact, it is of paramount importance that samples are collected during HY (i.e. “critical sample”), otherwise the diagnosis can be missed (biochemical investigations might result normal when euglycemia has been reached). As shown in the proposed diagnostic flowchart, the minimal set of biochemical findings in children presenting with HY includes ketones, lactate (both in blood and urine), and blood gases (i.e. metabolic acidosis). Such findings can help reaching a provisional diagnosis, which can be confirmed with additional (biochemical and/or genetic) tests. In this respect, collecting (and store adequately) additional samples at the time of HY is crucial. Laboratory data must also be appropriately integrated with anamnestic, dietary, clinical, and imaging information. The proposed flowchart aims at guiding the diagnostic management of a such common manifestation in pediatric age that can be due to a wide spectrum of causes. A double level usefulness is expected for the proposed flowchart: the first one is addressed to general pediatrician by providing the clinical-anamnestic and laboratory findings to be sought in order to refer the patient to the most appropriate Tertiary Center (Endocrinological/Metabolic/Genetic disease), the second level is for specialists in pediatric endocrine-metabolic diseases in order to remind them the wider etiological spectrum of pediatric HY by giving the essential elements of the differential diagnosis involving different areas (genetic, endocrine, and metabolic). In other word our flowchart aims to be a quick scheme to help pediatricians of every setting in managing HY, attempting to be comprehensive of the main disorders and differential diagnosis. Of course, mostly compared to the current available flowcharts focused on peculiar fields (metabolic or endocrinological), some rare conditions or rare presentations cannot be included. Sometimes, clinicians are not able to reach a final diagnosis, despite multiple efforts, due to the lack of specific biochemical pattern or atypical presentation of some disorders. In such cases, innovative diagnostic techniques can be considered. Even if much progress has been made over past years, many things remain to be discovered and clarified for diseases causing HY in childhood. Advances in diagnostic techniques (e.g. NGS) will identify specific defects or even new entities in a subgroup of patients who have been diagnosed with ketotic HY, likely resulting in a change in the disease epidemiology or in the discovery of new conditions (97).

In conclusion, future studies are also needed to optimally define normal glucose thresholds in neonates and children (10). In addition, irrespective of the specific diagnosis prompt recognition and treatment of acute HY are critical to prevent its complications,

the diagnostic work-up should start at the emergency hospital, collecting critical sample at the time of HY and providing specialists the clued results from simple tests that are available at any hospital at any time and that are very useful to address the clinical suspicion.

Based on the recognized risks of some tests the traditional diagnostic process, including fasting or dynamic tests, is presently controversial and probably superseded by modern molecular diagnostic techniques. NGS approach has also the potential to diagnose disorders with mild biochemical abnormalities or atypical presentations or even to identify new diseases, changing the epidemiology of many disorders. In this respect, the development of extended collaboration networks for rare diseases is worthy (43).

AUTHOR CONTRIBUTIONS

AC, AR and EM wrote the manuscript. EM, AF, SF and GP reviewed the manuscript. FMR, CM and FDC edited the manuscript and collected data. EM and SF are the guarantors

of this work and, as such, had full access to all the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis. All authors were responsible for drafting the article and revising it critically for important intellectual content. All authors contributed to the article and approved the submitted version.

ACKNOWLEDGMENTS

The authors are grateful to the European Reference Network dedicated to Rare Endocrine Conditions (Endo-ERN) for useful discussions.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fendo.2021.684011/full#supplementary-material>

REFERENCES

- Ghosh A, Banerjee I, Morris AAM. Recognition, Assessment and Management of Hypoglycaemia in Childhood. *Arch Dis Child* (2016) 101(6):575–80. doi: 10.1136/archdischild-2015-308337
- Daly LP, Osterhoudt KC, Weinzimer SA. Presenting Features of Idiopathic Ketotic Hypoglycemia. *J Emerg Med* (2003) 25(1):39–43. doi: 10.1016/s0736-4679(03)00100-8
- Clarke W, Jones T, Rewers A, Dunger D, Klingensmith GJ. Assessment and Management of Hypoglycemia in Children and Adolescents With Diabetes. *Pediatr Diabetes* (2009) 10(Suppl 12):134–45. doi: 10.1111/j.1399-5448.2009.00583.x
- Kim SY. Endocrine and Metabolic Emergencies in Children: Hypocalcemia, Hypoglycemia, Adrenal Insufficiency, and Metabolic Acidosis Including Diabetic Ketoacidosis. *Ann Pediatr Endocrinol Metab* (2015) 20(4):179–86. doi: 10.6065/apem.2015.20.4.179
- JM Saudubray, G van den Berghe and J Walter eds. *Inherited Metabolic Diseases*. 6. Berlin, Germany: Springer (2016).
- Tas E, Garibaldi L, Muzumdar R. Glucose Homeostasis in Newborns: An Endocrinology Perspective. *Neoreviews* (2020) 21(1):e14–29. doi: 10.1542/neo.21-1-e14
- Thornton PS, Stanley CA, De Leon DD, Harris D, Haymond MW, Hussain K, et al. Recommendations From the Pediatric Endocrine Society for Evaluation and Management of Persistent Hypoglycemia in Neonates, Infants, and Children. *J Pediatr* (2015) 167(2):238–45. doi: 10.1016/j.jpeds.2015.03.057
- Gandhi K. Approach to Hypoglycemia in Infants and Children. *Transl Pediatr* (2017) 6(4):408–20. doi: 10.21037/tp.2017.10.05
- Hume R, Burchell A, Williams FL, Koh DK. Glucose Homeostasis in the Newborn. *Early Hum Dev* (2005) 81(1):95–101. doi: 10.1016/j.earlhumdev.2004.10.005
- Cornblath M, Hawdon JM, Williams AF, Aynsley-Green A, Ward-Platt MP, Schwartz R, et al. Controversies Regarding Definition of Neonatal Hypoglycemia: Suggested Operational Thresholds. *Pediatrics* (2000) 105(5):1141–5. doi: 10.1542/peds.105.5.1141
- Mitrakou A, Ryan C, Veneman T, Mogan M, Jenssen T, Kiss I, et al. Hierarchy of Glycemic Thresholds for Counterregulatory Hormone Secretion, Symptoms, and Cerebral Dysfunction. *Am J Physiol* (1991) 260(1 Pt 1):E67–74. doi: 10.1152/ajpendo.1991.260.1.E67
- Cowett RM, Loughead JL. Neonatal Glucose Metabolism: Differential Diagnoses, Evaluation, and Treatment of Hypoglycemia. *Neonatal Netw* (2002) 21(4):9–19. doi: 10.1891/0730-0832.21.4.9
- Stanley CA, Rozance PJ, Thornton PS, De Leon DD, Harris D, Haymond MW, et al. Re-Evaluating “Transitional Neonatal Hypoglycemia”: Mechanism and Implications for Management. *J Pediatr* (2015) 166(6):1520–5.e1. doi: 10.1016/j.jpeds.2015.02.045
- Adamkin DH Committee on Fetus and Newborn. Postnatal Glucose Homeostasis in Late-Preterm and Term Infants. *Pediatrics* (2011) 127(3):575–9. doi: 10.1542/peds.2010-3851
- Sharma A, Davis A, Shekhawat PS. Hypoglycemia in the Preterm Neonate: Etiopathogenesis, Diagnosis, Management and Long-Term Outcomes. *Transl Pediatr* (2017) 6(4):335–48. doi: 10.21037/tp.2017.10.06
- McKinlay CJ, Alsweller JM, Ansell JM, Anstice NS, Chase JG, Gamble GD, et al. Neonatal Glycemia and Neurodevelopmental Outcomes at 2 Years. *N Engl J Med* (2015) 373(16):1507–18. doi: 10.1056/NEJMoa1504909
- Goode RH, Rettiganti M, Li J, Lyle RE, Whiteside-Mansell L, Barrett KW, et al. Developmental Outcomes of Preterm Infants With Neonatal Hypoglycemia. *Pediatrics* (2016) 138(6):e20161424. doi: 10.1542/peds.2016-1424
- Kallem VR, Pandita A, Gupta G. Hypoglycemia: When to Treat? *Clin Med Insights Pediatr* (2017) 11:1–9. doi: 10.1177/1179556517748913
- Levene I, Wilkinson D. British Association of Perinatal Medicine. Identification and Management of Neonatal Hypoglycemia in the Full Term Infant, Framework for Practice. *Arch Dis Child Educ Pract Ed* (2019) 104(1):29–32. doi: 10.1136/archdischild-2017-314050
- Bennett C, Fagan E, Chaharbakshi E, Zamfirova I, Flicker J. Implementing a Protocol Using Glucose Gel to Treat Neonatal Hypoglycemia. *Nurs Womens Health* (2016) 20(1):64–74. doi: 10.1016/j.nwh.2015.11.001
- Scheans P, Bennett C, Harris D. Using Dextrose (Glucose) Gel to Reverse Neonatal Hypoglycemia. *Neonatal Netw* (2017) 36(4):233–8. doi: 10.1891/0730-0832.36.4.233
- Harris DL, Weston PJ, Signal M, Chase JG, Harding JE. Dextrose Gel for Neonatal Hypoglycaemia (the Sugar Babies Study): A Randomised, Double-Blind, Placebo-Controlled Trial. *Lancet* (2013) 382(9910):2077–83. doi: 10.1016/S0140-6736(13)61645-1
- Hegarty JE, Harding JE, Gamble GD, Crowther CA, Edlin R, Alsweller JM. Prophylactic Oral Dextrose Gel for Newborn Babies at Risk of Neonatal

- Hypoglycaemia: A Randomised Controlled Dose-Finding Trial (the Pre-POD Study). *PLoS Med* (2016) 13(10):e1002155. doi: 10.1371/journal.pmed.1002155
24. Weston PJ, Harris DL, Battin M, Brown J, Hegarty JE, Harding JE. Oral Dextrose Gel for the Treatment of Hypoglycaemia in Newborn Infants. *Cochrane Database Syst Rev* (2016) 5:CD011027. doi: 10.1002/14651858.CD011027.pub2
 25. Makker K, Alissa R, Dudek C, Travers L, Smotherman C, Hudak ML. Glucose Gel in Infants at Risk for Transitional Neonatal Hypoglycemia. *Am J Perinatol* (2018) 35(11):1050–6. doi: 10.1055/s-0038-1639338
 26. Ter M, Halibullah I, Leung L, Jacobs S. Implementation of Dextrose Gel in the Management of Neonatal Hypoglycaemia. *J Paediatr Child Health* (2017) 53(4):408–11. doi: 10.1111/jpc.13409
 27. Rawat M, Chandrasekharan P, Turkovich S, Barclay N, Perry K, Schroeder E, et al. Oral Dextrose Gel Reduces the Need for Intravenous Dextrose Therapy in Neonatal Hypoglycemia. *BioMed Hub* (2016) 1(3):1–9. doi: 10.1159/000448511
 28. Hussain K. Congenital Hyperinsulinism. *Semin Fetal Neonatal Med* (2005) 10(4):369–76. doi: 10.1016/j.siny.2005.03.001
 29. Lee BM, Wolever TM. Effect of Glucose, Sucrose and Fructose on Plasma Glucose and Insulin Responses in Normal Humans: Comparison With White Bread. *Eur J Clin Nutr* (1998) 52(12):924–8. doi: 10.1038/sj.ejcn.1600666
 30. Georgakopoulos K, Katsilambros N, Fragaki M, Pouloupoulou Z, Kimbouris J, Sfikakis P, et al. Recovery From Insulin-Induced Hypoglycemia After Saccharose or Glucose Administration. *Clin Physiol Biochem* (1990) 8(5):267–72.
 31. Arnoux JB, Verkarre V, Saint-Martin C, Montravers F, Brassier A, Valayannopoulos V, et al. Congenital Hyperinsulinism: Current Trends in Diagnosis and Therapy. *Orphanet J Rare Dis* (2011) 6:63. doi: 10.1186/1750-1172-6-63
 32. Galcheva S, Demirbilek H, Al-Khawaga S, Hussain K. The Genetic and Molecular Mechanisms of Congenital Hyperinsulinism. *Front Endocrinol (Lausanne)* (2019) 10:111. doi: 10.3389/fendo.2019.00111
 33. Arya VB. Congenital Hyperinsulinism: Clinical and Molecular Characterisation of Compound Heterozygous ABCC8 Mutation Responsive to Diazoxide Therapy. *Int J Pediatr Endocrinol* (2014) 2014(1):24. doi: 10.1186/1687-9856-2014-24
 34. Taylor-Miller T. Congenital Hyperinsulinism Due to Compound Heterozygous Mutations in ABCC8 Responsive to Diazoxide Therapy. *J Pediatr Endocrinol Metab* (2020) 33(5):671–4. doi: 10.1515/jpem-2019-0457
 35. Kumar A, Kapoor RR, Flanagan SE, Ellard S, Hussain K. Congenital Hyperinsulinism Due to a Compound Heterozygous ABCC8 Mutation With Spontaneous Resolution at Eight Weeks. *Hormone Res Paediatr* (2010) 73:287–92. doi: 10.1159/000284394
 36. Kapoor RR, Locke J, Colclough K, Wales J, Conn JJ, Hattersley AT, et al. Persistent Hyperinsulinemic Hypoglycemia and Maturity-Onset Diabetes of the Young Due to Heterozygous HNF4A Mutations. *Diabetes* (2008) 57(6):1659–63. doi: 10.2337/db07-1657
 37. Stanescu DE, Hughes N, Kaplan B, Stanley CA, De León DD. Novel Presentations of Congenital Hyperinsulinism Due to Mutations in the MODY Genes: HNF1A and HNF4A. *J Clin Endocrinol Metab* (2012) 97(10):E2026–30. doi: 10.1210/jc.2012-1356
 38. Mohamed Z, Arya VB, Hussain K. Hyperinsulinaemic Hypoglycaemia: Genetic Mechanisms, Diagnosis and Management. *J Clin Res Pediatr Endocrinol* (2012) 4(4):169–81. doi: 10.4274/jcrpe.821
 39. Abdulhadi-Atwan M, Bushman J, Tornovsky-Babaey S, Perry A, Abu-Libdeh A, Glaser B, et al. Novel De Novo Mutation in Sulfonylurea Receptor 1 Presenting as Hyperinsulinism in Infancy Followed by Overt Diabetes in Early Adolescence. *Diabetes* (2008) 57(7):1935–40. doi: 10.2337/db08-0159
 40. Casertano A, De Matteis A, Mozzillo E, Rosanio FM, Buono P, Fattorusso V, et al. Diagnosis of Congenital Hyperinsulinism Can Occur Not Only in Infancy But Also in Later Age: A New Flow Chart From a Single Center Experience. *Ital J Pediatr* (2020) 46(1):131. doi: 10.1186/s13052-020-00894-5
 41. Vieira TC, Bergamin CS, Gurgel LC, Moisés RS. Hyperinsulinemic Hypoglycemia Evolving to Gestational Diabetes and Diabetes Mellitus in a Family Carrying the Inactivating ABCC8 E1506K Mutation. *Pediatr Diabetes* (2010) 11(7):505–8. doi: 10.1111/j.1399-5448.2009.00626.x
 42. Demirbilek H, Hussain K. Congenital Hyperinsulinism: Diagnosis and Treatment Update. *J Clin Res Pediatr Endocrinol* (2017) 9(Suppl 2):69–87. doi: 10.4274/jcrpe.2017.S007
 43. Derks TGJ, Nemeth A, Adrian K, Arnell H, Roskjær AB, Beijer E, et al. Hepatic Glycogen Storage Diseases Toward One Global Collaborative Network. *J Inborn Errors Metab Screen* (2017) 52(12):1051–9. doi: 10.1177/2326409817733009 Porto Alegre.
 44. Maiorana A, Barbeti F, Boiani A, Rufini V, Pizzoferrero M, Francalanci P. Focal Congenital Hyperinsulinism Managed by Medical Treatment: A Diagnostic Algorithm Based on Molecular Genetic Screening. *Clin Endocrinol (Oxf)* (2014) 81(5):679–88. doi: 10.1111/cen.12400
 45. Van der Steen I. A Multicenter Experience With Long-Acting Somatostatin Analogues in Patients With Congenital Hyperinsulinism. *Horm Res Paediatr* (2018) 89(2):82–9. doi: 10.1159/000485184
 46. Haris B. Somatostatin Analogues for the Treatment of Hyperinsulinaemic Hypoglycaemia. *Ther Adv Endocrinol Metab* (2020) 11:1–23. doi: 10.1177/2042018820965068 2042018820965068.
 47. Shah P. Use of Long-Acting Somatostatin Analogue (Lanreotide) in an Adolescent With Diazoxide-Responsive Congenital Hyperinsulinism and Its Psychological Impact. *Horm Res Paediatr* (2015) 84:355–60. doi: 10.1159/000439131
 48. Yang SB. Rapamycin Induces Glucose Intolerance in Mice by Reducing Islet Mass, Insulin Content, and Insulin Sensitivity. *J Mol Med (Berl)* (2012) 90:575–85. doi: 10.1007/s00109-011-0834-3
 49. Hashemian S. Clinical Efficacy Evaluation of Sirolimus in Congenital Hyperinsulinism. *Int J Endocrinol* (2020) 2020:1–6. doi: 10.1155/2020/7250406 7250406.
 50. Scaramuzza A, Cherubini V, Tumini S, Bonfanti R, Buono P, Cardella F, et al. Recommendations for Self-Monitoring in Pediatric Diabetes: A Consensus Statement by the ISPED. *Acta Diabetol* (2014) 51(2):173–84. doi: 10.1007/s00592-013-0521-7
 51. Piona C, Marigliano M, Mozzillo E, Franzese A, Zanfardino A, Iafusco D, et al. Long-Term Glycemic Control and Glucose Variability Assessed With Continuous Glucose Monitoring in a Pediatric Population With Type 1 Diabetes: Determination of Optimal Sampling Duration. *Pediatr Diabetes* (2020) 21(8):1485–92. doi: 10.1111/pedi.13115
 52. Beltrand J. Glucose Metabolism in 105 Children and Adolescents After Pancreatectomy for Congenital Hyperinsulinism. *Diabetes Care* (2012) 35:198–203. doi: 10.2337/dc11-1296
 53. Shulman DI, Palmert MR, Kemp SFLawson Wilkins Drug and Therapeutics Committee. Adrenal Insufficiency: Still a Cause of Morbidity and Death in Childhood. *Pediatrics* (2007) 119(2):e484–94. doi: 10.1542/peds.2006-1612
 54. Bornstein SR, Allolio B, Arlt W, Barthel A, Don-Wauchope A, Hammer GD, et al. Diagnosis and Treatment of Primary Adrenal Insufficiency: An Endocrine Society Clinical Practice Guideline. *J Clin Endocrinol Metab* (2016) 101(2):364–89. doi: 10.1210/jc.2015-1710
 55. Ioakim KJ, Sydney GI, Paschou SA. Glucose Metabolism Disorders in Patients With Adrenal Gland Disorders: Pathophysiology and Management. *Hormones (Athens)* (2020) 19(2):135–43. doi: 10.1007/s42000-019-00147-z
 56. Kiousis D, Reshef L, Cohen H, Tilghman SM, Iynedjian PB, Ballard FJ, et al. Alterations in Translatable Messenger RNA Coding for Phosphoenolpyruvate Carboxykinase (GTP) in Rat Liver Cytosol During Deinduction. *J Biol Chem* (1978) 253(12):4327–32. doi: 10.1016/S0021-9258(17)34723-3
 57. Ballard FJ, Hopgood MF, Reshef L, Tilghman S, Hanson RW. Synthesis of Phosphoenolpyruvate Carboxykinase (Guanosine Triphosphate) by Isolated Liver Polyribosomes. *Biochem J* (1974) 144(2):199–207. doi: 10.1042/bj1440199
 58. Iynedjian PB, Ballard FJ, Hanson RW. The Regulation of Phosphoenolpyruvate Carboxykinase (GTP) Synthesis in Rat Kidney Cortex. The Role of Acid-Base Balance and Glucocorticoids. *J Biol Chem* (1975) 250(14):5596–603. doi: 10.1016/S0021-9258(19)41221-0
 59. Hanson RW, Reshef L. Regulation of Phosphoenolpyruvate Carboxykinase (GTP) Gene Expression. *Annu Rev Biochem* (1997) 66:581–611. doi: 10.1146/annurev.biochem.66.1.581
 60. Djurhuus CB, Gravholt CH, Nielsen S, Mengel A, Christiansen JS, Schmitz OE, et al. Effects of Cortisol on Lipolysis and Regional Interstitial Glycerol Levels in Humans. *Am J Physiol Endocrinol Metab* (2002) 283(1):E172–7. doi: 10.1152/ajpendo.00544.2001

61. Kirkgoz T, Guran T. Primary Adrenal Insufficiency in Children: Diagnosis and Management. *Best Pract Res Clin Endocrinol Metab* (2018) 32(4):397–424. doi: 10.1016/j.beem.2018.05.010
62. Ferrier PE. Congenital Absence of Hypoplasia of the Endocrine Glands. *J Genet Hum* (1969) 17(3):325–47.
63. Wajnrajch MP, Gertner JM, Harbison MD, Chua SCJr, Leibel RL. Nonsense Mutation in the Human Growth Hormone-Releasing Hormone Receptor Causes Growth Failure Analogous to the Little (Lit) Mouse. *Nat Genet* (1996) 12(1):88–90. doi: 10.1038/ng0196–88
64. Moseley CT, Mullis PE, Prince MA, Phillips JA3rd. An Exon Splice Enhancer Mutation Causes Autosomal Dominant GH Deficiency. *J Clin Endocrinol Metab* (2002) 87(2):847–52. doi: 10.1210/jcem.87.2.8236
65. Scotti G, Triulzi F, Chiumello G, Dinatale B. New Imaging Techniques in Endocrinology: Magnetic Resonance of the Pituitary Gland and Sella Turcica. *Acta Paediatr Scand Suppl* (1989) 356:5–14. doi: 10.1111/j.1651–2227.1989.tb11235.x
66. Hall JG. Pallister–Hall Syndrome has Gone the Way of Modern Medical Genetics. *Am J Med Genet C Semin Med Genet* (2014) 166C(4):414–8. doi: 10.1002/ajmg.c.31419
67. Di Iorgi N, Morana G, Allegri AE, Napoli F, Gastaldi R, Calcagno A, et al. Classical and non–Classical Causes of GH Deficiency in the Paediatric Age. *Best Pract Res Clin Endocrinol Metab* (2016) 30(6):705–36. doi: 10.1016/j.beem.2016.11.008
68. Phillips JA3rd, Hjelle BL, Seeburg PH, Zachmann M. Molecular Basis for Familial Isolated Growth Hormone Deficiency. *Proc Natl Acad Sci U S A* (1981) 78(10):6372–5. doi: 10.1073/pnas.78.10.6372
69. Growth Hormone Research Society. Consensus Guidelines for the Diagnosis and Treatment of Growth Hormone (GH) Deficiency in Childhood and Adolescence: Summary Statement of the GH Research Society. *J Clin Endocrinol Metab* (2000) 85(11):3990–3. doi: 10.1210/jcem.85.11.6984
70. Melis D, Rossi A, Pivonello R, Salerno M, Balivo F, Spadarella S, et al. Glycogen Storage Disease Type Ia (Gsdia) But Not Glycogen Storage Disease Type Ib (Gsdib) is Associated to an Increased Risk of Metabolic Syndrome: Possible Role of Microsomal Glucose 6–Phosphate Accumulation. *Orphanet J Rare Dis* (2015) 10:91. doi: 10.1186/s13023–015–0301–2
71. Ross KM, Ferrecchia IA, Dahlberg KR, Damska M, Ryan PT, Weinstein DA. Dietary Management of the Glycogen Storage Diseases: Evolution of Treatment and Ongoing Controversies. *Adv Nutr* (2020) 11(2):439–46. doi: 10.1093/advances/nmz092
72. Kishnani PS, Austin SL, Abdenur JE, Arn P, Bali DS, Boney A, et al. Diagnosis and Management of Glycogen Storage Disease Type I: A Practice Guideline of the American College of Medical Genetics and Genomics. *Genet Med* (2014) 16(11):e1. doi: 10.1038/gim.2014.128
73. Roe TF, Thomas DW, Gilsanz V, Isaacs HJr, Atkinson JB. Inflammatory Bowel Disease in Glycogen Storage Disease Type Ib. *J Pediatr* (1986) 109(1):55–9. doi: 10.1016/s0022–3476(86)80572–8
74. Melis D, Pivonello R, Parenti G, Della Casa R, Salerno M, Lombardi G, et al. Increased Prevalence of Thyroid Autoimmunity and Hypothyroidism in Patients With Glycogen Storage Disease Type I. *J Pediatr* (2007) 150(3):300–5. doi: 10.1016/j.jpeds.2006.11.056
75. Melis D, Della Casa R, Balivo F, Minopoli G, Rossi A, Salerno M, et al. Involvement of Endocrine System in a Patient Affected by Glycogen Storage Disease Ib: Speculation on the Role of Autoimmunity. *Ital J Pediatr* (2014) 40(1):30. doi: 10.1186/1824–7288–40–30
76. Rossi A, Ruoppolo M, Formisano P, Villani G, Albano L, Gallo G, et al. Insulin–Resistance in Glycogen Storage Disease Type Ia: Linking Carbohydrates and Mitochondria? *J Inherit Metab Dis* (2018) 41(6):985–95. doi: 10.1007/s10545–018–0149–4
77. Rossi A, Simeoli C, Salerno M, Ferrigno R, Della Casa R, Colao A, et al. Imbalanced Cortisol Concentrations in Glycogen Storage Disease Type I: Evidence for a Possible Link Between Endocrine Regulation and Metabolic Derangement. *Orphanet J Rare Dis* (2020) 15(1):99. doi: 10.1186/s13023–020–01377–w
78. Melis D, Rossi A, Pivonello R, Del Puente A, Pivonello C, Cangemi G, et al. Reduced Bone Mineral Density in Glycogen Storage Disease Type III: Evidence for a Possible Connection Between Metabolic Imbalance and Bone Homeostasis. *Bone* (2016) 86:79–85. doi: 10.1016/j.bone.2016.02.012
79. Rossi A, Hoogeveen IJ, Bastek VB, de Boer F, Montanari C, Meyer U, et al. Dietary Lipids in Glycogen Storage Disease Type III: A Systematic Literature Study, Case Studies, and Future Recommendations. *J Inherit Metab Dis* (2020) 43(4):770–7. doi: 10.1002/jimd.12224
80. Kishnani PS, Goldstein J, Austin SL, Arn P, Bachrach B, Bali DS, et al. Diagnosis and Management of Glycogen Storage Diseases Type VI and IX: A Clinical Practice Resource of the American College of Medical Genetics and Genomics (Acmg). *Genet Med* (2019) 21(4):772–89. doi: 10.1038/s41436–018–0364–2
81. Kamenets EA, Gusarova EA, Milovanova NV, Itkiss YS, Strokova TV, Melikyan MA, et al. Hepatic Glycogen Synthase (GYS2) Deficiency: Seven Novel Patients and Seven Novel Variants. *JIMD Rep* (2020) 53(1):39–44. doi: 10.1002/jimd.12082
82. Arko JJ, Debeljak M, Tansek MZ, Battelino T, Groselj U. A Patient With Glycogen Storage Disease Type 0 and a Novel Sequence Variant in GYS2: A Case Report and Literature Review. *J Int Med Res* (2020) 48(8):1–8. doi: 10.1177/0300060520936857
83. Di Dato F, Spadarella S, Puoti MG, Caprio MG, Pagliardini S, Zuppaldi C, et al. Daily Fructose Traces Intake and Liver Injury in Children With Hereditary Fructose Intolerance. *Nutrients* (2019) 11(10):2397. doi: 10.3390/nu11102397
84. Van Schaftingen LHE. Fructose 2,6–Bisphosphate. *Adv Enzymol Relat Areas Mol Biol* (1987) 59:315–95. doi: 10.1002/9780470123058.ch7
85. Marin–Valencia I, Roe CR, Pascual JM. Pyruvate Carboxylase Deficiency: Mechanisms, Mimics and Anaplerosis. *Mol Genet Metab* (2010) 101(1):9–17. doi: 10.1016/j.ymgme.2010.05.004
86. Weinstein DA, Steuerwald U, De Souza CFM, Derks TGJ. Inborn Errors of Metabolism With Hypoglycemia: Glycogen Storage Diseases and Inherited Disorders of Gluconeogenesis. *Pediatr Clin North Am* (2018) 65(2):247–65. doi: 10.1016/j.pcl.2017.11.005
87. Sjarif DR, Ploos van Amstel JK, Duran M, Beemer FA, Poll–The BT. Isolated and Contiguous Glycerol Kinase Gene Disorders: A Review. *J Inherit Metab Dis* (2000) 23(6):529–47. doi: 10.1023/a:1005660826652
88. Barone R, Fiumara A, Jaeken J. Congenital Disorders of Glycosylation With Emphasis on Cerebellar Involvement. *Semin Neurol* (2014) 34(3):357–66. doi: 10.1055/s–0034–1387197
89. Merritt JL, Norris M, Kanungo S. Fatty Acid Oxidation Disorders. *Ann Transl Med* (2018) 6(24):473. doi: 10.21037/atm.2018.10.57
90. Tucci S, Behringer S, Sturm M, Grünert SC, Spiekerkoetter U. Implementation of a Fast Method for the Measurement of Carnitine Palmitoyltransferase 2 Activity in Lymphocytes by Tandem Mass Spectrometry as Confirmation for Newborn Screening. *J Inherit Metab Dis* (2019) 42(5):850–6. doi: 10.1002/jimd.12098
91. Spiekerkoetter U, Lindner M, Santer R, Grotzke M, Baumgartner MR, Boehles H, et al. Management and Outcome in 75 Individuals With Long–Chain Fatty Acid Oxidation Defects: Results From a Workshop. *J Inherit Metab Dis* (2009) 32(4):488–97. doi: 10.1007/s10545–009–1125–9
92. Fukao T, Mitchell G, Sass JO, Hori T, Orii K, Aoyama Y. Ketone Body Metabolism and its Defects. *J Inherit Metab Dis* (2014) 37:541–51. doi: 10.1007/s10545–014–9704–9
93. Kanungo S, Morton J, Neelakantan M, Ching K, Saedian J, Goldstein A. Mitochondrial Disorders. *Ann Transl Med* (2018) 6(24):475. doi: 10.21037/atm.2018.12.13
94. Fraser JL, Venditti CP. Methylmalonic and Propionic Acidemias: Clinical Management Update. *Curr Opin Pediatr* (2016) 28(6):682–93. doi: 10.1097/MOP.0000000000000422
95. Hörster F, Kölker S, Loeber JG, Cornel MC, Hoffmann GF, Burgard P. Newborn Screening Programmes in Europe, Arguments and Efforts Regarding Harmonisation: Focus on Organic Acidurias. *JIMD Rep* (2017) 32:105–15. doi: 10.1007/8904_2016_537
96. Hoogeveen IJ, van der Ende RM, van Spronsen FJ, de Boer F, Heiner–Fokkema MR, Derks TG. Normoglycemic Ketonemia as Biochemical Presentation in Ketotic Glycogen Storage Disease. *JIMD Rep* (2016) 28:41–7. doi: 10.1007/8904_2015_511
97. Brown LM, Corrado MM, van der Ende RM, Derks TG, Chen MA, Siegel S, et al. Evaluation of Glycogen Storage Disease as a Cause of Ketotic Hypoglycemia in Children. *J Inherit Metab Dis* (2015) 38(3):489–93. doi: 10.1007/s10545–014–9744–1
98. Rossi A, Hoogeveen IJ, Lubout CMA, de Boer F, Fokkert–Wilts MJ, Rodenburg IL, et al. A Generic Emergency Protocol for Patients With

Inborn Errors of Metabolism Causing Fasting Intolerance: A Retrospective, Single-Center Study and the Generation of Www.Emergencyprotocol.Net. *J Inherit Metab Dis* (2021) 1–12. doi: 10.1002/jim.d.12386 Online ahead of print.

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of

the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Casertano, Rossi, Fecarotta, Rosanio, Moracas, Di Candia, Parenti, Franzese and Mozzillo. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.