

University of Groningen

Creatine homeostasis and protein energy wasting in hemodialysis patients

Post, Adrian; Schutten, Joëlle C; Kremer, Daan; van der Veen, Yvonne; Groothof, Dion; Sotomayor, Camilo G; Koops, Christa A; de Blaauw, Pim; Kema, Ido P; Westerhuis, Ralf

Published in:
Journal of translational medicine

DOI:
[10.1186/s12967-021-02780-y](https://doi.org/10.1186/s12967-021-02780-y)

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version
Publisher's PDF, also known as Version of record

Publication date:
2021

[Link to publication in University of Groningen/UMCG research database](#)

Citation for published version (APA):

Post, A., Schutten, J. C., Kremer, D., van der Veen, Y., Groothof, D., Sotomayor, C. G., Koops, C. A., de Blaauw, P., Kema, I. P., Westerhuis, R., Wallimann, T., Heiner-Fokkema, M. R., Bakker, S. J. L., & Franssen, C. F. M. (2021). Creatine homeostasis and protein energy wasting in hemodialysis patients. *Journal of translational medicine*, 19(1), [115]. <https://doi.org/10.1186/s12967-021-02780-y>

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: <https://www.rug.nl/library/open-access/self-archiving-pure/taverne-amendment>.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.


Downloaded from the University of Groningen/UMCG research database (Pure): <http://www.rug.nl/research/portal>. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

RESEARCH

Open Access



Creatine homeostasis and protein energy wasting in hemodialysis patients

Adrian Post^{1*} , Joëlle C. Schutten¹, Daan Kremer¹, Yvonne van der Veen¹, Dion Groothof¹, Camilo G. Sotomayor¹, Christa A. Koops², Pim de Blaauw², Ido P. Kema², Ralf Westerhuis³, Theo Wallimann⁴, M. Rebecca Heiner-Fokkema², Stephan J. L. Bakker¹ and Casper F. M. Franssen¹

Abstract

Muscle wasting, low protein intake, hypoalbuminemia, low body mass, and chronic fatigue are prevalent in hemodialysis patients. Impaired creatine status may be an often overlooked, potential contributor to these symptoms. However, little is known about creatine homeostasis in hemodialysis patients. We aimed to elucidate creatine homeostasis in hemodialysis patients by assessing intradialytic plasma changes as well as intra- and interdialytic losses of arginine, guanidinoacetate, creatine and creatinine. Additionally, we investigated associations of plasma creatine concentrations with low muscle mass, low protein intake, hypoalbuminemia, low body mass index, and chronic fatigue. Arginine, guanidinoacetate, creatine and creatinine were measured in plasma, dialysate, and urinary samples of 59 hemodialysis patients. Mean age was 65 ± 15 years and 63% were male. During hemodialysis, plasma concentrations of arginine (77 ± 22 to 60 ± 19 $\mu\text{mol/L}$), guanidinoacetate (1.8 ± 0.6 to 1.0 ± 0.3 $\mu\text{mol/L}$), creatine ($26 [16-41]$ to $21 [15-30]$ $\mu\text{mol/L}$) and creatinine (689 ± 207 to 257 ± 92 $\mu\text{mol/L}$) decreased (all $P < 0.001$). During a hemodialysis session, patients lost 1939 ± 871 μmol arginine, 37 ± 20 μmol guanidinoacetate, $719 [399-1070]$ μmol creatine and 15.5 ± 8.4 mmol creatinine. In sex-adjusted models, lower plasma creatine was associated with a higher odds of low muscle mass (OR per halving: 2.00 [1.05–4.14]; $P = 0.04$), low protein intake (OR: 2.13 [1.17–4.27]; $P = 0.02$), hypoalbuminemia (OR: 3.13 [1.46–8.02]; $P = 0.008$) and severe fatigue (OR: 3.20 [1.52–8.05]; $P = 0.006$). After adjustment for potential confounders, these associations remained materially unchanged. Creatine is iatrogenically removed during hemodialysis and lower plasma creatine concentrations were associated with higher odds of low muscle mass, low protein intake, hypoalbuminemia, and severe fatigue, indicating a potential role for creatine supplementation.

Keywords: Creatine, Guanidinoacetate, Arginine, Protein energy wasting, Muscle mass, Fatigue, Hemodialysis

Clinical perspectives

- Creatine homeostasis in hemodialysis patients is largely unknown, but impaired creatine homeostasis may contribute to protein energy wasting.
- Arginine, guanidinoacetate, creatine and creatinine all demonstrated an intradialytic decrease in plasma

concentrations. We quantified the intradialytic losses and demonstrated that the majority of creatine is removed from the intracellular compartment during hemodialysis.

- We demonstrated that lower plasma creatine concentrations are associated with higher odds of low muscle mass, low protein intake, hypoalbuminemia, and severe fatigue.

*Correspondence: a.post01@umcg.nl

¹ Department of Internal Medicine, University of Groningen, University Medical Center Groningen, 9713 GZ Groningen, The Netherlands
Full list of author information is available at the end of the article



© The Author(s) 2021. This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Introduction

Managing the nutritional aspects of chronic kidney disease (CKD) presents a wide variety of challenges. While overnutrition is a major problem in the general population, patients with CKD, especially those depending on hemodialysis treatment, are more prone to protein energy wasting (PEW), a state of decreased body stores of protein and energy fuels [1, 2]. Muscle wasting, low protein intake, hypoalbuminemia, low body mass and chronic fatigue are very prevalent in hemodialysis patients, with major impacts on the quality of life and longevity [1–3].

An impaired creatine status may be a generally overlooked potential contributor to symptoms related to PEW in hemodialysis patients. Creatine is a natural nitrogenous organic acid that is integral to energy metabolism and crucial for proper cell functioning [4]. In humans, roughly 90% of all creatine and its high-energy product phosphocreatine are located inside skeletal muscles where it serves as an energy buffer par excellence [5]. Yet, on a daily basis, roughly 1.6 to 1.7% of the total creatine pool is degraded to creatinine, necessitating a continuous replenishment of the pool with new creatine [6]. Generally, in omnivores, approximately 50% of the daily requirement of creatine is endogenously synthesized and the rest is taken up through the diet from alimentary sources, like fresh meat and fish [7]. Importantly, however, the first, and rate-limiting step of the endogenous synthesis of creatine is facilitated primarily by the kidneys, where the enzyme arginine:glycine amidinotransferase (AGAT) converts arginine into guanidinoacetate [7–14]. Subsequently, guanidinoacetate is converted into creatine in the liver by the enzyme guanidinoacetate *N*-methyltransferase (GAMT) [9]. Given the fact that the rate limiting step of creatine synthesis depends on kidney function, and provided that hemodialysis patients virtually have no kidney function, it has been hypothesized that low endogenous creatine biosynthesis contributes to low creatine status and development of PEW in hemodialysis patients [7, 8]. Additionally, there may be unopposed losses of creatine and its precursors to the dialysate during every hemodialysis session. However, literature on creatine homeostasis in hemodialysis patients is scarce and the intradialytic losses have not been quantified.

The goals of the current study were (1) to explore creatine homeostasis during hemodialysis treatment and (2) to investigate whether plasma creatine concentrations are associated with characteristics of the PEW phenotype. To achieve the first goal, we assessed intradialytic plasma changes as well as intra- and interdialytic losses of creatine, its precursors arginine and guanidinoacetate, and its metabolite creatinine. To achieve the second goal, we investigated whether plasma creatine concentrations

were associated low muscle mass, low protein intake, hypoalbuminemia, low body mass index, and chronic fatigue.

Material and methods

Design and study population

This observational study was performed according to ethical standards laid down in the 1964 Declaration of Helsinki and its later amendments and was approved by the Medical Ethical Committee of the University Medical Center Groningen, The Netherlands. All participating patients gave written informed consent. The methods of the study design have been described previously [15–17]. In short, inclusion criteria were twice or thrice weekly hemodialysis with 3–5 h per treatment, a hemodialysis vintage of ≥ 2 months, and absence of clinical signs of infection. Patients on a thrice weekly hemodialysis schedule dialyzed on either Monday–Wednesday–Friday or on Tuesday–Thursday–Saturday. In both cases the mid-week hemodialysis session was used in this study. For patients dialyzing twice weekly, the last hemodialysis session of the week was used.

In adherence to the Kidney Disease Outcomes Quality Initiative and national guidelines, our center applies a fistula first approach for vascular access creation, thereby starting with creation of a fistula as distally in the arm as possible [18]. The non-dominant arm was preferred to create the vascular access. In case of lower-arm arteriovenous fistulae, patients received a radiocephalic arteriovenous fistula. In case of upper-arm arteriovenous fistulae, patients received a brachiocephalic arteriovenous fistula or basilic vein transposition. In patients with previous maturation problems of fistulae, or patients with a subacute indication for dialysis (i.e. < 4 weeks), an arteriovenous graft was considered instead of the fistula first approach. The arteriovenous graft was a standard wall polytetrafluorethylene graft (Gore-Tex, WL Gore & Associates, Flagstaff, Arizona, USA), in either a loop or straight configuration. In patients without an arteriovenous fistula or an arteriovenous graft, a central venous catheter was used.

Hypertension was defined as predialysis systolic blood pressure ≥ 140 mmHg and/or diastolic blood pressure ≥ 90 mmHg. Medication usage and a history of cardiovascular disease and diabetes was obtained from the patients' medical records. Cardiovascular disease was defined as a history of ischemic heart disease, congestive heart failure, coronary artery bypass grafting, percutaneous coronary intervention, stroke, or peripheral vascular disease. Blood pressure and weight were measured before and after hemodialysis. Body mass index (BMI) was defined as body weight postdialysis divided by the square of body height. Body surface area (BSA)

was calculated using the formula of Du Bois and Du Bois [19]. All patients were asked to fill in the Checklist Individual Strength (CIS). The CIS is a self-reported multidimensional instrument to assess four qualitatively different aspects of fatigue (fatigue severity, concentration problems, reduced motivation, and reduced activity level). The CIS-questionnaire enquires about fatigue and fatigue-related behavioral aspects and consists of 20 statements for which the participant indicates on a 7-point Likert-scale to what extent the statement applies to the participant. CIS has been well-validated and is frequently used in research in patients with various illnesses [20–24], including dialysis patients [17, 25].

Hemodialysis settings

All studies were performed with the Fresenius 5008 hemodialysis apparatus with a low-flux dialyzer (Fresenius Medical Care, Bad Homburg, Germany) using smartbag dialysate concentrations (Fresenius Medical Care), as previously described [15–17]. Blood flow and dialysate flow were between 200 and 300 mL/min and between 500 and 700 mL/min, respectively. Dialysate temperature was 36.0 or 36.5 °C. Dialysis fluid sodium varied from 136 to 140 mmol/L, potassium from 1 to 3 mmol/L, depending on the plasma potassium concentration, calcium varied from 1.25 to 1.50 mmol/L and bicarbonate from 34 to 38 mmol/L.

Sample collection and laboratory measurements

During the hemodialysis session, all dialysate was collected in a 200-L tank. The total dialysate volume was measured by calculating the weight difference of the tank before and after the hemodialysis session. At the end of hemodialysis, all dialysate was homogenized, and samples were taken for analysis [15]. Blood was drawn directly from the dialysis line, at the start of hemodialysis and 5 min before the end of the hemodialysis session. Patients with significant residual diuresis, defined as a urine production of more than 200 ml/24-h, were asked to collect two 24-h urine collections before the hemodialysis session during which the dialysate was collected. For patients with a thrice-weekly hemodialysis schedule this was the complete interdialytic urine production.

Plasma and dialysate concentrations of arginine were measured using a LC–MS/MS method. In short, tandem mass spectrometry with hydrophilic interaction liquid chromatography (HILIC) was used to quantify the concentrations of arginine. Electrospray ionization (ESI) in positive mode has been used to monitor the transitions of arginine 175- > 116 and stable isotope labeled arginine (¹³C₆-¹⁵N₄-arginine) 185- > 122 as an internal standard.

Intra-assay and inter-assay coefficients of variation were 6.0% and 6.1% at 23 μmol/L and 3.9% and 5.3% at 492 μmol/L, respectively. Urinary arginine was measured using cation exchange chromatography with post column derivatisation (Biochrom Ltd.), separation was achieved by increasing the pH. Intra-assay and inter-assay coefficients of variation were 6.5% and 7.6% at 28 μmol/L. Plasma, dialysate, and urinary concentrations of guanidinoacetate and creatine were measured using a separate LC–MS/MS method. In short, tandem mass spectrometry reversed phase chromatography (RP-C18) was used to quantify the concentrations of creatine and guanidinoacetate. Electrospray ionization (ESI) in positive mode has been used to monitor the transitions of guanidinoacetate 118- > 76 and creatine 132- > 90, using stable isotope labeled guanidinoacetate (¹³C₂-guanidinoacetate) 120- > 78 and stable isotope labeled creatine (D3-creatinine) 135- > 93 as internal standards. For guanidinoacetate, intra-assay and inter-assay coefficients of variation were 2.9% and 6.1% at 2.8 μmol/L and 1.9% and 2.7% at 73 μmol/L, respectively. For creatine, intra-assay and inter-assay coefficients of variation were 3.9% and 6.5% at 17 μmol/L and 1.7% and 1.7% at 1065 μmol/L, respectively. Plasma creatinine, urea, albumin and high-sensitivity C-reactive protein were measured on Roche routine chemistry analyzers (Modular P/Cobas C, Roche Diagnostics, Mannheim, Germany). Interleukin-6 (IL-6) was measured in a subset of the participants (n=39) using a Human IL-6 Quantikine HS Elisa kit (R&D systems, Minneapolis, United States). Other laboratory measurements were performed with automated and validated routine methods (Roche Diagnostics, Mannheim, Germany). The Kt/V was used to express hemodialysis efficacy and was calculated according to formula of Daugirdas [26]:

$$Kt/V = -\ln(R - 0.008 * t + (4 - 3.5 * R) * UF/W)$$

in which R is the ratio between the post- and predialysis concentration of urea, t is duration of the hemodialysis session (h), UF is the ultrafiltration volume (L) and W the body weight after hemodialysis (kg).

Intradialytic plasma changes

Intradialytic plasma changes of arginine, guanidinoacetate, creatine and creatinine are assessed by calculating the absolute decrease, as well as the proportional decrease:

Absolute decrease (μmol/L) = Predialysis concentration (μmol/L) – Postdialysis concentration (μmol/L).

Proportional decrease (%) = Absolute decrease ($\mu\text{mol/L}$) / Predialysis concentration ($\mu\text{mol/L}$) * 100%.

Losses of arginine, guanidinoacetate, creatine and creatinine during hemodialysis

To determine the losses of arginine, guanidinoacetate, creatine and creatinine to the dialysis fluid, the concentration of individual analytes was multiplied by the total volume of the spent dialysate: Total removal (in μmol) = $V_{\text{Dialysate}} * D_X$. $V_{\text{Dialysate}}$ refers to the total volume of the spent dialysate (L) and D_X refers to the measured concentration of arginine, guanidinoacetate, creatine and creatinine in the collected dialysate ($\mu\text{mol/L}$). The total losses per dialysis of arginine, guanidinoacetate, creatine and creatinine can be split into the losses from the extracellular space and the losses from the intracellular space after being shifted towards the extracellular space during the hemodialysis. The estimated extracellular losses of arginine, guanidinoacetate, creatine or creatinine were calculated using the pre- and postdialysis plasma concentrations of arginine, guanidinoacetate, creatine and creatinine and estimates of the pre- and postdialysis extracellular volume (ECV): Estimated extracellular losses (μmol) = $P_{X \text{ predialysis}} * ECV_{\text{predialysis}} - P_{X \text{ postdialysis}} * ECV_{\text{postdialysis}}$. $P_{X \text{ predialysis}}$ and $P_{X \text{ postdialysis}}$ refer to predialysis and postdialysis plasma concentrations of arginine, guanidinoacetate, creatine and creatinine, respectively. In these calculations, the assumption is made that these small molecular solutes can freely diffuse throughout the extracellular fluid. $ECV_{\text{predialysis}}$ and $ECV_{\text{postdialysis}}$ were calculated using the formula by Abraham et al. (2011), defined as the square root of pre- or postdialysis body weight, multiplied by height [27]. Additionally, as a sensitivity analysis, the extracellular volume was also calculated using the formula by Bird et al. [28]: $ECV = \text{weight}^{0.649} * \text{height}^{0.7236} * 0.02154$.

The estimated intracellular losses of arginine, guanidinoacetate, creatine and creatinine during hemodialysis were calculated as the difference between the total losses of arginine, guanidinoacetate, creatine and creatinine and the estimated extracellular losses of arginine, guanidinoacetate, creatine and creatinine: Estimated intracellular losses (μmol) = total losses (μmol) – estimated extracellular losses (μmol).

Higher estimated intracellular losses indicate a larger shift from the intra- to extracellular space during the hemodialysis session. Besides losses to the dialysate, there may also be losses in urine in patients with residual diuresis. To determine the degree to which urinary losses contribute the daily losses in patients with residual diuresis, we compared the urinary and the dialysate losses of arginine, guanidinoacetate, creatine and creatinine in patients with residual diuresis. The daily dialysate

excretion rate of arginine, guanidinoacetate, creatine and creatinine are calculated as: Dialysate excretion rate ($\mu\text{mol}/24\text{-h}$) = $(V_{\text{Dialysate}} * D_X * n) / 7$. In this formula $V_{\text{Dialysate}}$ refers to the total volume of dialysate (L), D_X refers to the measured concentration of arginine, guanidinoacetate, creatine and creatinine in the collected dialysate ($\mu\text{mol/L}$), and n refers to the number of dialysis sessions per week. The daily urinary excretion rate was calculated as: Urinary excretion rate ($\mu\text{mol}/24\text{-h}$) = $(\text{Urinary excretion}_{\text{day 1}} + \text{Urinary excretion}_{\text{day 2}}) / 2$.

Characteristics of the protein energy wasting phenotype

The cross-sectional outcomes low muscle mass, low protein intake, hypoalbuminemia, low body mass index and severe fatigue were used as characteristics for the PEW phenotype.

Muscle mass was assessed by calculating the combined creatinine excretion rate (CER):

Creatinine excretion rate ($\text{mmol}/24\text{-h}$) = $((V_{\text{Dialysate}} * D_{\text{Creatinine}} * n) / 7 + \text{UCrE})$. In this formula $V_{\text{Dialysate}}$ refers to the total volume of dialysate (L), $D_{\text{Creatinine}}$ refers to measured creatinine concentration in the collected dialysate (mmol/L), n refers number of dialyses per week, and UCrE refers to 24-h urinary creatinine excretion ($\text{mmol}/24\text{-h}$), averaged from two 24-h urine collections. The creatinine excretion rate was converted to the skeletal muscle index by multiplying by the creatinine-equivalence (1 g of creatinine excretion per 24-h (8.84 mmol) = 22.73 kg of skeletal muscle mass) and dividing by height squared [29, 30]. A low muscle mass was defined as a SMI < 10.76 kg/m^2 in males and < 6.76 kg/m^2 in females [29, 31, 32].

Low protein intake was defined as a dietary protein intake < 0.8 $\text{g}/\text{kg}/24\text{-h}$ [2]. The dietary protein intake was calculated based on the combined (dialysate and urinary) excretion rate of urea, according to the Maroni formula and indexed to body weight [33]:

Protein intake ($\text{g}/\text{kg}/24\text{-h}$) = $(6.25 * (0.028 * \text{CUER} + 0.031 * \text{BW}) + \text{UPE}) / \text{BW}$.

In this formula CUER refers to combined excretion rate of urea ($\text{mmol}/24\text{-h}$), BW refers to body weight postdialysis (kg), and UPE refers to the 24-h urine protein excretion ($\text{g}/24\text{-h}$), averaged from two 24-h urine collections. CUER was calculated as follows: CUER (in $\text{mmol}/24\text{-h}$) = $(V_{\text{Dialysate}} * D_{\text{urea}} * n) / 7 + \text{UUE}$. In this formula, $V_{\text{Dialysate}}$ refers total volume of the spent dialysate (L), D_{urea} refers to the measured urea concentration in the collected dialysate (mmol/L), n refers to the number of hemodialysis sessions per week, and UUE refers to the 24-h urinary urea excretion ($\text{mmol}/24\text{-h}$), averaged from two 24-h urine collections. Hypoalbuminemia was defined as a plasma albumin concentration < 38 g/L [2]. Low body mass was defined as a body mass index < 23 $\text{kg}/$

m^2 [2]. Severe fatigue was defined as a subjective fatigue score of ≥ 35 on the checklist individual strength [23, 24]. An overview of all used formulas can be found in Additional file 1: Table S10.

Statistical analysis

Data analyses and computations were performed with R version 3.6.1 software (<http://cran.r-project.org/>). A two-sided $P < 0.05$ was considered to indicate statistical significance. Data were presented as mean \pm standard deviation for normally distributed data, median [interquartile range] for non-normally distributed data, and as numbers (percentages) for nominal data. Baseline characteristics, intradialytic plasma concentration changes, single hemodialysis removal and comparisons of urinary excretion rate and dialysate excretion rate were shown for all participants and stratified according to sex. Sex-stratified analyses were performed based on the known sex-based differences in creatine homeostasis [5, 34] as well as editorial statements calling for clinical trials and observational studies to report on sex-stratified results [35, 36]. The differences between males and females were tested using the independent sample t-test or the Mann–Whitney U test. Differences between pre- and postdialysis plasma concentrations, between extracellular removal and intracellular removal and between the urinary excretion rate and the dialysate excretion rate were tested using the paired sample t-test or the Wilcoxon signed-rank test. Correlations between predialysis plasma concentrations and the dialysate losses of arginine, guanidinoacetate, creatine and creatinine were assessed using the Pearson correlation coefficient. Associations of predialysis plasma concentrations of arginine, guanidinoacetate, creatine, and creatinine with low muscle mass, low protein intake, low plasma albumin, low BMI and severe fatigue were investigated using logistic regression analyses. Due to the small sample size, these analyses were not stratified according to sex, but performed with adjustment for sex. In addition, multivariable analyses were performed in which we additionally adjusted for the a priori selected potential confounders age, sex, body surface area, dialysis vintage and high-sensitivity C-reactive protein (hs-CRP). Due to a skewed distribution, plasma creatine concentrations were \log_2 -transformed prior to logistic regression analyses. To determine whether the found associations were driven by patients with high plasma creatine concentrations, we performed sensitivity analyses after prior exclusion of patients with predialysis plasma creatine concentrations above the 95th percentile. In addition, we also performed sensitivity analyses in which we adjusted for the type of vascular access. Also, we also performed sensitivity analyses in which we adjusted for the hemodialysis efficacy (Kt/V). Furthermore, we performed

sensitivity analyses in which we adjusted for the usage of erythropoietin-stimulating agents, vitamin D analogues and corticosteroids. Lastly, we performed sensitivity analyses in which we adjusted for the presence or absence of residual diuresis. To visualize the continuous associations of plasma creatine concentrations with low muscle mass, low protein intake, hypoalbuminemia and severe fatigue, plasma creatine concentration, as a continuous variable, was individually plotted against the probability of low muscle mass, low protein intake, hypoalbuminemia and severe fatigue using the *visreg* package in R. These analyses were adjusted for sex, age, body surface area, dialysis vintage and hs-CRP.

Results

Baseline characteristics

A total of 59 hemodialysis patients were included in the study, 37 (63%) of whom were male. Mean age at inclusion was 65 ± 15 years with a median hemodialysis vintage of 15 [6–41] months. Nearly all (95%) patients dialyzed thrice-weekly and most patients (81%) dialyzed four hours per session. A total of 32 (54%) patients had residual diuresis, with a mean urinary volume of 0.9 ± 0.6 L. Vascular access was a lower arm arteriovenous fistula in 13 (22%) patients, an upper arm arteriovenous fistula in 19 (32%) patients, a central venous line in 19 (32%) patients and an arteriovenous graft in 8 (14%) patients. The mean Kt/V was 1.4 ± 0.3 per hemodialysis session. Mean BMI was 25.5 ± 4.3 kg/m². Median hs-CRP was 4.9 [1.7–13.5] mg/L and median IL-6 was 5.6 [3.2–13.0] pg/mL. Hypertension, cardiovascular disease, and diabetes mellitus were prevalent in 33 (58%), 25 (42%) and 15 (25%) of the patients, respectively. Erythropoietin-stimulating agents, vitamin D analogues, corticosteroids and androgens were used by 49 (83%), 38 (64%), 12 (20%) and 2 (3%) patients, respectively. Differences in baseline characteristics stratified by sex are shown in Table 1. Compared to males, females had lower body weight, height and BSA (all $P < 0.05$). Compared to males, females had a higher Kt/V ($P = 0.01$). Other baseline characteristics were comparable between males and females.

Intradialytic changes in plasma concentrations.

Intradialytic changes in plasma concentrations of arginine, guanidinoacetate, creatine and creatinine are shown in Table 2. Plasma arginine concentrations decreased during dialysis from 77 ± 22 $\mu\text{mol/L}$ to 60 ± 19 $\mu\text{mol/L}$ ($P < 0.001$), corresponding to a proportional decrease of $18 \pm 25\%$. No significant sex-based differences were found. During hemodialysis, plasma guanidinoacetate concentrations decreased from 1.8 ± 0.6 $\mu\text{mol/L}$ to 1.0 ± 0.3 $\mu\text{mol/L}$ ($P < 0.001$), corresponding to a proportional decrease of $40 \pm 17\%$.

Table 1 Baseline characteristics of the hemodialysis patients

	Total cohort	Males	Females	P-value
Demographics				
Participants, n (%)	59 (100)	37 (63)	22 (37)	0.05
Age, years	65 ± 15	65 ± 14	64 ± 15	0.8
Race, n Caucasian (%)	52 (88)	31 (84)	21 (95)	0.4
Dialysis-related				
Dialysis vintage, months	15 [6–39]	15 [7–29]	16 [6–48]	0.7
Dialysis sessions, n (%)				
Two sessions per week	3 (5)	1 (3)	2 (9)	0.6
Three sessions per week	56 (95)	36 (97)	20 (91)	
Hours per dialysis, n (%)				
3 to 3.5 h	6 (10)	3 (8)	3 (14)	0.6
4 h	48 (81)	30 (81)	18 (82)	
4.5 to 5 h	5 (8)	4 (11)	1 (5)	
Dialysate volume, L	135 ± 27	140 ± 30	127 ± 23	0.08
Ultrafiltration volume, L	1.9 ± 0.9	1.9 ± 0.9	1.9 ± 0.8	0.8
Residual diuresis, n (%)	32 (54)	24 (65)	8 (36)	0.06
Urinary volume, L	0.9 ± 0.6	0.8 ± 0.6	1.0 ± 0.7	0.4
Kt/V	1.4 ± 0.3	1.3 ± 0.3	1.6 ± 0.3	0.01
Vascular access, n (%)				
Lower arm fistula	13 (22)	11 (30)	2 (9)	0.3
Upper arm fistula	19 (32)	11 (30)	8 (36)	
Central venous line	19 (32)	11 (30)	8 (36)	
Arteriovenous graft	8 (14)	4 (11)	4 (18)	
Body composition				
Weight predialysis, kg	80 ± 16	84 ± 17	74 ± 13	0.03
Weight postdialysis, kg	78 ± 16	82 ± 16	73 ± 13	0.04
Height, m	1.75 ± 0.09	1.80 ± 0.07	1.67 ± 0.05	<0.001
BMI ^a , kg/m ²	25.5 ± 4.3	25.2 ± 4.4	26.1 ± 4.2	0.4
BSA ^a , m ²	1.94 ± 0.21	2.01 ± 0.20	1.81 ± 0.16	<0.001
Inflammation				
Hs-CRP, mg/L	4.9 [1.7–13.5]	4.9 [1.6–12.0]	5.3 [2.0–17.5]	0.8
IL-6, pg/mL ^b	5.6 [3.2–13.0]	6.4 [3.2–15.0]	5.2 [3.6–7.2]	0.5
Pre-existing disease				
Hypertension, n (%)	33 (58)	24 (65)	9 (41)	0.1
Cardiovascular disease, n (%)	25 (42)	16 (43)	9 (41)	0.9
Diabetes, n (%)	15 (25)	10 (27)	5 (23)	0.9
Medication				
Erythropoietin-stimulating agents, n (%)	49 (83)	29 (78)	20 (90)	0.4
Vitamin D analogues, n (%)	38 (64)	23 (62)	15 (68)	0.9
Corticosteroids, n (%)	12 (20)	6 (10)	6 (27)	0.5
Androgens, n (%)	2 (3)	2 (5)	0 (0)	0.7

^a Calculated using postdialysis weight^b Data available in 39 participants

No significant sex-based differences were found. Plasma creatine concentrations decreased from 26 [16–41] μmol/L to 21 [15–30] μmol/L ($P < 0.001$), corresponding to a proportional decrease of $13 \pm 39\%$. Males had lower predialysis (19 [15–33] μmol/L vs 33

[24–56] μmol/L; $P = 0.01$) and postdialysis plasma creatine concentrations (20 [15–25] μmol/L vs 24 [15–37] μmol/L; $P = 0.01$), as compared to females. Additionally, in males, the proportional decrease in

Table 2 Intradialytic changes of plasma concentrations of arginine, guanidinoacetate, creatine and creatinine

Metabolite	Total cohort	Males	Females	P-value
Arginine				
Predialysis concentration, $\mu\text{mol/L}$	77 \pm 22 ^{***}	80 \pm 24 ^{***}	72 \pm 18 ^{***}	0.2
Postdialysis concentration, $\mu\text{mol/L}$	60 \pm 19 ^{***}	63 \pm 21 ^{***}	55 \pm 14 ^{***}	0.08
Absolute decrease, $\mu\text{mol/L}$	17 \pm 23 ^{***}	17 \pm 25 ^{***}	16 \pm 19 ^{***}	0.9
Proportional decrease, %	18 \pm 25 ^{***}	17 \pm 28 ^{***}	20 \pm 21 ^{***}	0.7
Guanidinoacetate				
Predialysis concentration, $\mu\text{mol/L}$	1.8 \pm 0.6 ^{***}	1.9 \pm 0.6 ^{***}	1.6 \pm 0.5	0.1
Postdialysis concentration, $\mu\text{mol/L}$	1.0 \pm 0.3 ^{***}	1.1 \pm 0.3 ^{***}	0.9 \pm 0.4	0.07
Absolute decrease, $\mu\text{mol/L}$	0.8 \pm 0.4 ^{***}	0.8 \pm 0.5 ^{***}	0.7 \pm 0.4 ^{***}	0.4
Proportional decrease, %	40 \pm 17 ^{***}	38 \pm 17 ^{***}	43 \pm 15 ^{***}	0.3
Creatine				
Predialysis concentration, $\mu\text{mol/L}$	26 [16–41] ^{***}	19 [15–33] [*]	33 [24–56] ^{***}	0.01
Postdialysis concentration, $\mu\text{mol/L}$	21 [15–30] ^{***}	20 [15–25] [*]	24 [15–37] ^{***}	0.01
Absolute decrease, $\mu\text{mol/L}$	5 [1–14] ^{***}	3 [0–8] [*]	10 [3–18] ^{***}	0.02
Proportional decrease, %	13 \pm 39 [*]	5 \pm 43	27 \pm 26 ^{***}	0.02
Creatinine				
Predialysis concentration, $\mu\text{mol/L}$	689 \pm 207 ^{***}	706 \pm 224 ^{***}	660 \pm 175 ^{***}	0.4
Postdialysis concentration, $\mu\text{mol/L}$	257 \pm 92 ^{***}	274 \pm 97 ^{***}	228 \pm 78 ^{***}	0.07
Absolute decrease, $\mu\text{mol/L}$	432 \pm 146 ^{***}	432 \pm 156 ^{***}	432 \pm 130 ^{***}	0.9
Proportional decrease, %	63 \pm 8 ^{***}	61 \pm 9 ^{***}	65 \pm 7 ^{***}	0.05

Differences between males and females are tested using an independent sample t-test. Differences between predialysis and postdialysis plasma concentrations are tested using paired sample t-test or the paired samples Wilcoxon test. Absolute decrease and proportional decrease are tested against no decrease by t-test or Wilcoxon test. Statistical significance is indicated with asterix (* $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$)

plasma creatine concentration was smaller ($5 \pm 43\%$ vs $27 \pm 26\%$; $P = 0.02$), as compared to females.

Plasma creatinine concentrations decreased from $689 \pm 207 \mu\text{mol/L}$ to $257 \pm 92 \mu\text{mol/L}$ ($P < 0.001$), corresponding to a proportional decrease of $63 \pm 8\%$. In contrast to creatine, predialysis plasma concentrations of creatinine ($706 \pm 224 \mu\text{mol/L}$ vs $660 \pm 175 \mu\text{mol/L}$; $P = 0.4$) and postdialysis plasma concentrations of creatinine ($274 \pm 97 \mu\text{mol/L}$ vs 228 ± 78 ; $P = 0.07$) were numerically higher in males, as compared to females, albeit not significant at the 5% level.

Losses during hemodialysis

The losses of arginine, guanidinoacetate, creatine and creatinine during a hemodialysis session are shown in Table 3. During hemodialysis, the total loss of arginine was $1939 \pm 871 \mu\text{mol}$, of which an estimated amount of $287 \pm 344 \mu\text{mol}$ (15%) was lost from the extracellular compartment and an estimated amount of $1655 \pm 859 \mu\text{mol}$ (85%) was lost from the intracellular compartment ($P < 0.001$). No significant sex-based differences were found for arginine removal. The total loss of guanidinoacetate was $37 \pm 20 \mu\text{mol}$, of which an estimated amount of $12 \pm 7 \mu\text{mol}$ (32%) was lost from the extracellular compartment and an estimated amount of $25 \pm 19 \mu\text{mol}$ (68%) was lost from the intracellular

compartment ($P < 0.001$). The total losses of guanidinoacetate were larger in males than in females ($41 \pm 20 \mu\text{mol}$ vs $31 \pm 18 \mu\text{mol}$; $P = 0.05$). The total loss of creatine was $719 [399–1070] \mu\text{mol}$, of which an estimated amount of $79 [19–196] \mu\text{mol}$ (18%) was lost from the extracellular compartment and an estimated amount of $612 [365–902] \mu\text{mol}$ (82%) was lost from the intracellular compartment ($P < 0.001$).

The total loss of creatinine was $15.5 \pm 8.4 \text{ mmol}$, of which an estimated amount of $6.8 \pm 2.5 \text{ mmol}$ (44%) was lost from the extracellular compartment and an estimated amount of $8.7 \pm 7.1 \text{ mmol}$ (56%) was lost from the intracellular compartment ($P < 0.05$). The total losses of creatinine were larger in males than in females ($17.5 \pm 9.6 \text{ mmol}$ vs $12.0 \pm 3.9 \text{ mmol}$; $P = 0.003$).

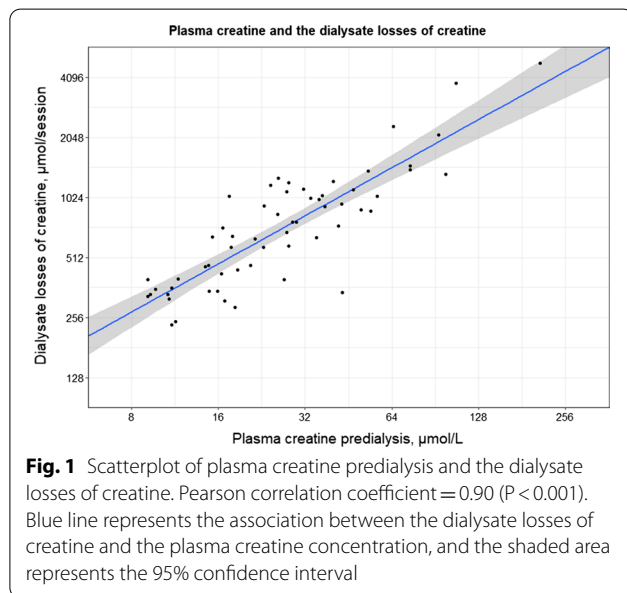
Correlation analyses of predialysis plasma values and dialysate losses are shown in Additional file 1: Table S2. Predialysis plasma concentrations were positively correlated with dialysate losses for arginine ($r = 0.51$; $P < 0.001$), guanidinoacetate ($r = 0.54$; $P < 0.001$), creatine ($r = 0.90$; $P < 0.001$) and creatinine ($r = 0.60$; $P < 0.001$). A graphical representation of the correlation between predialysis plasma creatine and the dialysate losses of creatine is shown in Fig. 1, showing that higher predialysis plasma creatine concentrations are associated with larger dialysate losses of creatine.

Table 3 Comparison of the dialysate losses of arginine, guanidinoacetate, creatine and creatinine per hemodialysis session

Metabolite	Total cohort	Males	Females	P-value
Arginine				
Total loss, μmol	1939 \pm 871	2053 \pm 921	1748 \pm 760	0.2
Estimated extracellular loss, μmol	287 \pm 344 ^{***}	318 \pm 390 ^{***}	235 \pm 248 ^{***}	0.3
Estimated intracellular loss, μmol	1655 \pm 859 ^{***}	1747 \pm 922 ^{***}	1497 \pm 733 ^{***}	0.3
Guanidinoacetate				
Total loss, μmol	37 \pm 20	41 \pm 20	31 \pm 18	0.05
Estimated extracellular loss, μmol	12 \pm 7 ^{***}	13 \pm 8 ^{***}	10 \pm 5 ^{**}	0.08
Estimated intracellular loss, μmol	25 \pm 19 ^{***}	28 \pm 20 ^{***}	21 \pm 16 ^{**}	0.2
Creatine				
Total loss, μmol	719 [399–1070]	655 [397–1015]	867 [464–1363]	0.2
Estimated extracellular loss, μmol	79 [19–196] ^{***}	58 [5–144] ^{***}	147 [42–231] ^{***}	0.04
Estimated intracellular loss, μmol	612 [365–902] ^{***}	545 [365–871] ^{***}	697 [413–1052] ^{***}	0.4
Creatinine				
Total loss, mmol	15.5 \pm 8.4	17.5 \pm 9.6	12.0 \pm 3.9	0.003
Estimated extracellular loss, mmol	6.8 \pm 2.5 [*]	7.2 \pm 2.7 [*]	6.2 \pm 1.9	0.2
Estimated intracellular loss, mmol	8.7 \pm 7.1 [*]	10.4 \pm 8.4 [*]	5.8 \pm 2.2	0.003

ECV is calculated according to the formula by Abraham et al. [27]

Differences between males and females are tested using an independent sample t-test or Mann–Whitney U test. Differences between extracellular and intracellular removal are tested using paired sample t-test or the paired samples Wilcoxon test, and statistical significance is indicated with asterix (^{*} P < 0.05; ^{**} P < 0.01; ^{***} P < 0.001)



Since hemodialysis patients with residual diuresis will have losses into urine in addition to losses to dialysate, we compared the urinary losses to the dialysate losses. An overview of the daily urinary and dialysate excretion rates of arginine, guanidinoacetate, creatine and creatinine is shown in Additional file 1: Table 3. In patients with residual diuresis, urinary excretion contributes 2.8% to the total daily excretion rate of arginine, 42% to

the total daily excretion rate of guanidinoacetate, 6.0% to the total daily excretion rate of creatine and 38% to the total daily excretion rate of creatinine. No sex-based differences were found for the proportion that daily urinary excretion rate contributes to the total daily excretion rate.

Characteristics of the protein energy wasting phenotype

Associations of predialysis plasma concentrations of arginine, guanidinoacetate, creatine and creatinine with low muscle mass, low protein intake, hypoalbuminemia, low BMI, and severe fatigue are shown in Table 4. In the sex-adjusted models, higher plasma arginine was inversely associated with low protein intake (OR per standard deviation decrease: 1.94 [1.08–3.94]; P = 0.04) and hypoalbuminemia (OR: 2.50 [1.35–5.33]; P = 0.007). In multivariable analyses, the association with low protein intake was lost, while the association with hypoalbuminemia remained materially unchanged. No associations were found between plasma arginine and low muscle mass, low BMI, or severe fatigue (all P > 0.05). For plasma guanidinoacetate, no associations were found with low muscle mass, low protein intake, hypoalbuminemia, low BMI, or severe fatigue (all P > 0.05).

In sex-adjusted models, lower plasma creatine was associated with higher odds of low muscle mass (OR per halving of plasma creatine: 2.00 [1.05–4.14]; P = 0.04), low protein intake (OR: 2.13 [1.17–4.27]; P = 0.02), hypoalbuminemia (OR: 3.13 [1.46–8.02]; P = 0.008) and severe fatigue (OR: 3.20 [1.52–8.05]; P = 0.006). After

Table 4 Associations of predialysis plasma concentrations of arginine, guanidinoacetate, creatine and creatinine with characteristics of the protein energy wasting phenotype

	Sex-adjusted analyses		Multivariable analyses	
	OR (95% CI)	P value	OR (95% CI)	P value
Plasma arginine (per 1-SD decrease)				
Low muscle mass	1.65 [0.84–3.89]	0.2	1.60 [0.74–4.27]	0.3
Low protein intake	1.94 [1.08–3.94]	0.04	1.91 [1.01–4.16]	0.07
Hypoalbuminemia	2.50 [1.35–5.33]	0.007	2.87 [1.39–7.06]	0.009
Low BMI	1.06 [0.59–1.86]	0.8	0.91 [0.32–2.89]	0.9
Severe fatigue	1.14 [0.63–2.08]	0.7	1.04 [0.50–2.15]	0.9
Plasma guanidinoacetate (per 1-SD decrease)				
Low muscle mass	1.08 [0.59–2.19]	0.8	0.94 [0.48–2.08]	0.9
Low protein intake	1.55 [0.89–2.94]	0.2	1.11 [0.60–2.24]	0.7
Hypoalbuminemia	1.45 [0.82–2.65]	0.2	1.35 [0.71–2.67]	0.3
Low BMI	1.17 [0.66–2.08]	0.6	1.61 [0.70–3.77]	0.2
Severe fatigue	1.20 [0.69–2.15]	0.5	0.96 [0.49–1.84]	0.9
Plasma creatine (per halving)				
Low muscle mass	2.00 [1.05–4.14]	0.04	2.32 [1.10–5.69]	0.04
Low protein intake	2.13 [1.17–4.27]	0.02	2.37 [1.20–5.41]	0.02
Hypoalbuminemia	3.13 [1.46–8.02]	0.008	6.20 [2.16–24.8]	0.003
Low BMI	0.74 [0.40–1.35]	0.3	0.51 [0.18–1.28]	0.2
Severe fatigue	3.20 [1.52–8.05]	0.006	3.74 [1.55–11.8]	0.009
Plasma creatinine (per 1-SD decrease)				
Low muscle mass	6.66 [2.14–28.6]	0.003	5.67 [1.59–37.3]	0.03
Low protein intake	1.35 [0.80–2.43]	0.3	3.15 [0.82–3.29]	0.2
Hypoalbuminemia	2.51 [1.35–5.41]	0.006	2.50 [1.25–5.72]	0.02
Low BMI	1.80 [1.02–3.45]	0.05	2.58 [0.94–9.66]	0.10
Severe fatigue	1.09 [0.62–1.90]	0.76	1.65 [0.80–3.51]	0.18

Plasma refers to plasma values predialysis. Analyses of plasma arginine, guanidinoacetate and creatinine are performed on scaled data, with the odds ratio presented per standard deviation decrease of the biomarker. Analyses of plasma creatine are performed on log₂ transformed data, with the odds ratio presented per halving of the biomarker

Low muscle mass is defined as a muscle mass < 10.76 kg/m² in males < 6.76 kg/m² in females. Low protein intake is defined as a protein intake < 0.8 g/kg/24-h. Low BMI is defined as a BMI < 23 kg/m². Hypoalbuminemia is defined as a serum albumin < 38 g/L. Severe fatigue is defined as a subjective fatigue score ≥ 35. Multivariable analyses are adjusted for sex, age, body surface area, dialysis vintage and hs-CRP

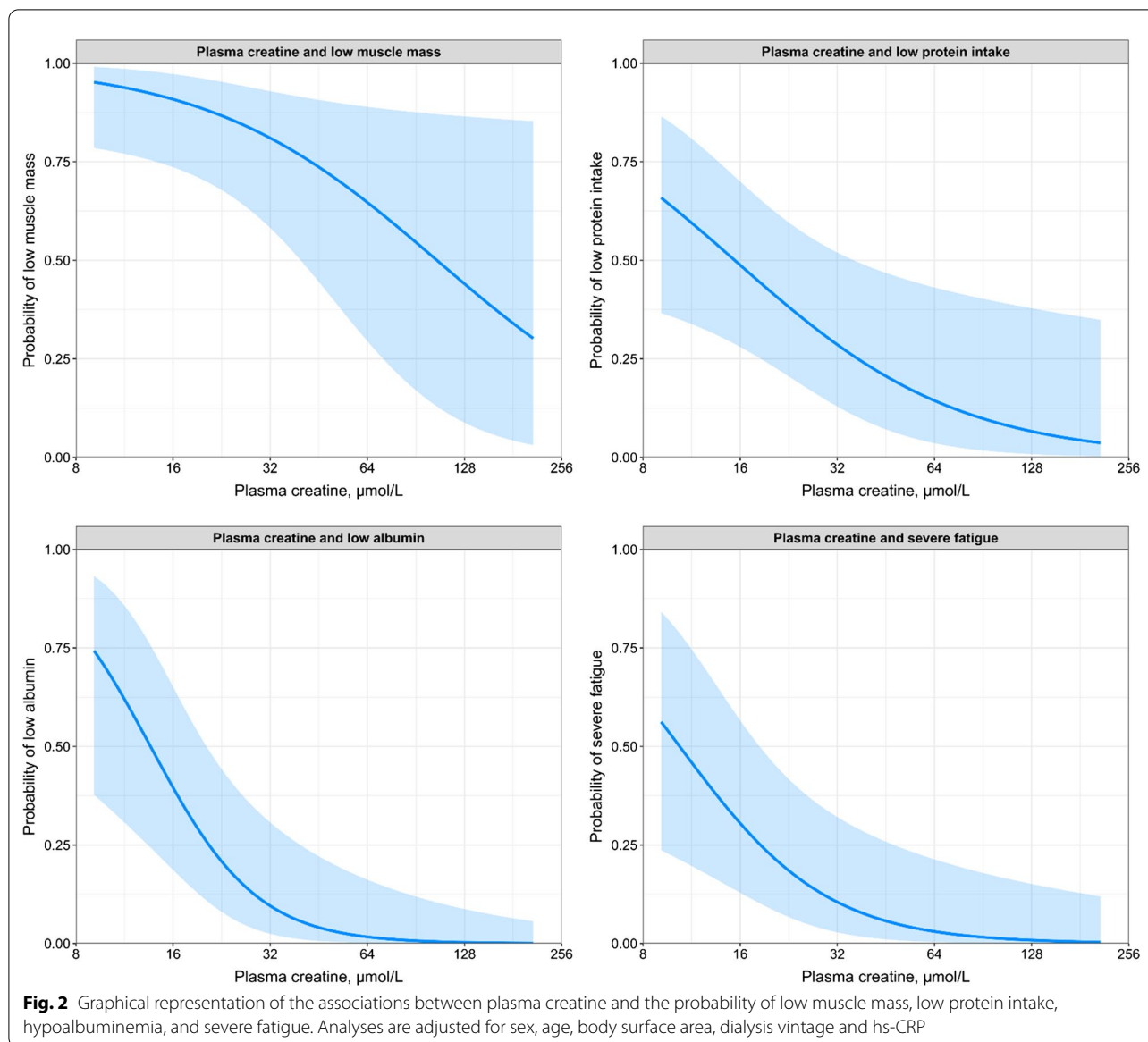
adjustment for potential confounders, these associations remained materially unchanged. No associations were found with low BMI. A graphical representation of the relationship between plasma creatine and low muscle mass, low protein intake, hypoalbuminemia and severe fatigue is shown in Fig. 2.

In sex-adjusted models, lower plasma creatinine was associated with low muscle mass (OR per standard deviation decrease: 6.66 [2.21–28.6]; P < 0.001) and hypoalbuminemia (OR: 2.51 [1.35–5.41]; P = 0.02). After adjustment for potential confounders, these associations remained materially unchanged. No associations were found with low protein intake, low BMI and severe fatigue.

Sensitivity analyses

To determine whether the associations of plasma creatine are driven by the patients with the highest creatine concentrations, we performed sensitivity analyses in which patients with predialysis plasma creatine values above the 95th percentile were excluded in Additional file 1: Table S4. In these analyses, the association of plasma creatine with low muscle mass was no longer significant (P = 0.20). However, the association between plasma creatine and low protein intake, hypoalbuminemia and severe fatigue remained significant and materially unchanged.

To investigate whether the associations of plasma creatine with low muscle mass, low protein intake, hypoalbuminemia and severe fatigue could be the consequence of differences in vascular access type, we performed sensitivity analyses in which we adjusted for the type of vascular access (Additional file 1: Table S5). The associations remained materially unchanged after adjustment for the type of vascular access. To investigate whether the associations of plasma creatine with low muscle mass, low protein intake, hypoalbuminemia and severe fatigue could be the consequence of differences in hemodialysis efficacy, we performed sensitivity analyses in which we adjusted for the Kt/V values (Additional file 1: Table S6). The associations remained materially unchanged after adjustment for the Kt/V. To investigate whether the associations of plasma creatine with low muscle mass, low protein intake, hypoalbuminemia and severe fatigue could be the consequence of differences in medication usage, we performed sensitivity analyses in which we adjusted for the usage of erythropoietin-stimulating agents, vitamin D analogues and corticosteroids (Additional file 1: Table S7). The associations remained materially unchanged after adjustment for these medications. To investigate whether the associations of plasma creatine with low muscle mass, low protein intake, hypoalbuminemia and severe fatigue could be the consequence of differences in residual diuresis, we performed sensitivity analyses in which we adjusted the associations for presence or absence of residual diuresis (Additional file 1: Table S8). The associations remained materially unchanged after adjustment for the presence or absence of residual diuresis. Associations of the dialysate losses



of arginine, guanidinoacetate, creatine and creatinine with low muscle mass, low protein intake, hypoalbuminemia, low BMI, and fatigue are shown in Additional file 1: Table S9. For the dialysate losses of arginine and guanidinoacetate no associations were found with low muscle mass, low protein intake, hypoalbuminemia, low BMI, or fatigue. Lower dialysate losses of creatine were associated with higher odds of low muscle mass (OR per halving: 2.61 [1.28–6.31]; $P=0.02$), hypoalbuminemia (OR: 4.77 [2.03–13.8]; $P=0.001$) and severe fatigue (OR: 2.80 [1.34–6.65]; $P=0.01$). After adjustment for potential confounders, these associations remained materially unchanged. No significant associations were found with low protein intake and low BMI.

Discussion

To our knowledge, this is the first study to comprehensively investigate creatine homeostasis in hemodialysis patients. Arginine, guanidinoacetate, creatine and creatinine all demonstrated an intradialytic decrease in plasma concentrations. The proportional decrease was greatest for creatinine and lowest for creatine. No sex-differences were found for arginine, guanidinoacetate and creatinine. By contrast, males had lower pre- and post-dialysis plasma creatine concentrations and had a lower proportional intradialytic decrease in plasma concentrations compared to females. In addition, we quantified the intradialytic losses and demonstrated that the majority of creatine is removed from the intracellular compartment,

thereby implicating a large intra- to extracellular shift of creatine during a hemodialysis session. For patients with residual diuresis we demonstrated that the intradialytic losses of arginine, guanidinoacetate, creatine and creatinine to the dialysis fluid are much larger than the interdialytic urinary losses. Lastly, we demonstrated that lower plasma creatine concentrations are associated with higher odds of low muscle mass, low protein intake, hypoalbuminemia, and severe fatigue. These findings were independent from potential confounders, including plasma levels of hs-CRP as a reflection of systemic inflammation.

Creatine and the energy-charged phosphocreatine play a crucial bioenergetic role in adenosine triphosphate turnover and are especially important as a cellular energy buffer and in energy transport, especially in tissues with high and fluctuating energetic demands, such as skeletal muscles and the brain [37, 38]. Creatine is therefore an essential metabolite for normal cell and body functioning, from embryonic life to adulthood [39]. The importance of creatine is highlighted by the genetic creatine deficiency syndromes, encompassing mutations in the genes of AGAT, GAMT or the gene coding for the creatine transporter (SLC6A8), where patients develop neuromuscular and neurological symptoms, including mental retardation, autism, and epilepsy [40]. In omnivorous humans, creatine is synthesized endogenously, but also ingested through the diet [7]. After being released into the circulation, creatine is transported into tissues by the Creatine Transporter 1 (CrT1), encoded by the SLC6A8 gene, located on the X-chromosome [41]. Throughout the years, several sex-based differences in creatine homeostasis have been identified, e.g. the overall creatine synthesis rate has been found to be lower in females than in males [5], which is also reflected by lower serum guanidinoacetate concentrations in females than in males [34]. A study in 66 healthy individuals found lower serum guanidinoacetate concentrations in females (2.0 ± 0.57 $\mu\text{mol/L}$) than in males (2.6 ± 0.52 $\mu\text{mol/L}$) [42]. Compared to the healthy individuals of the aforementioned study, the peak (predialysis) plasma concentrations in the hemodialysis patients of the current study were 27% and 20% lower in males and females, respectively. These findings support previous hypotheses that the first step of endogenous creatine synthesis is impaired in patients with end-stage kidney disease [8, 15]. Especially since generally plasma concentrations of small water-soluble molecules tend to increase with decreasing kidney function [43]. Since the conversion of AGAT is considered the rate limiting step in creatine synthesis [7–14], it follows that a reduction in AGAT activity in end-stage kidney disease also leads to reduced creatine concentrations, compatible with our findings. It is worth

noting that the concentrations of creatine in healthy individuals of the study by Marescau et al. [42] were nearly twice times higher in females, compared with males. Similarly, we also found higher plasma creatine concentrations in females as compared with males, both in pre- and postdialysis samples. Although the exact mechanism underlying this sex-based difference is not known, it has been reported before [44], and the same phenomenon is also seen in biopsies of the vastus lateralis muscle where females had 10% higher intracellular creatine concentrations compared to males [45]. Our analysis of the intradialytic changes in plasma concentrations demonstrated that arginine, guanidinoacetate, creatine and creatinine all significantly decreased during dialysis, in line with small molecules being removed to the dialysate. Interestingly, the proportional decrease in plasma concentrations was largest (63%) for creatinine and smallest for creatine (13%), implicating that plasma creatine concentrations are more prone to strict regulation. To elucidate losses to the dialysate, we quantified the losses to the dialysate during hemodialysis by collecting the entirety of the spent dialysate. In agreement with the lower plasma concentrations of guanidinoacetate compared to creatine, we also found less removal of guanidinoacetate as compared to creatine. By estimating the relative losses from the extracellular and intracellular compartments, we demonstrated that most creatine was lost from the intracellular compartment, indicative of a large intradialytic intra- to extracellular shift. A possibility is that plasma creatine is strictly regulated and that an intra- to extracellular shift is one of the mechanisms preventing too low plasma creatine concentrations. This hypothesis of a large intra- to extracellular shift during hemodialysis is congruent with previous imaging studies obtained by non-invasive *in vivo* ^{31}P -NMR demonstrating lower phosphocreatine concentrations in the hearts of patients on either hemodialysis treatment [46] or peritoneal dialysis treatment [47]. Additionally, lower phosphocreatine concentrations have also been measured in skeletal muscles of hemodialysis patients [48]. Combined, these findings indicate a wash-out of creatine from these organs during hemodialysis treatment.

Apart from losses to the dialysate, there may also be losses of creatine to the urine in patients with residual diuresis. Yet, in our supplementary analyses we demonstrated that urinary losses only contribute a minor 6.0% to the overall daily losses of creatine in hemodialysis patients.

Protein energy wasting is a major threat for patients on hemodialysis, as it is the strongest risk factor for morbidity and mortality [1, 3]. Clinically, the diagnostic criteria have been set up across several domains, including low muscle mass, low protein intake, hypoalbuminemia

and low body weight. A relatively simple, non-invasive and very reliable method to estimate muscle mass is to measure the creatinine excretion rate (CER). Creatinine is produced at a constant rate, depending on the quantity of muscle mass [7, 49]. Therefore, CER is an established method to study total body muscle mass in both healthy populations and a variety of patient populations, including patients with chronic kidney disease [15, 50–54]. An added benefit of CER over other techniques, such as dual-energy x-ray absorptiometry, is that CER by its biochemical nature is insensitive to hydration status, intramuscular fat, and edema and thereby provides a direct reflection of muscle mass [55, 56]. Lower plasma values of creatine, but not arginine and guanidinoacetate, were associated with higher odds for a low muscle mass, indicating that low plasma creatine concentrations may increase the risk of low muscle mass. Similarly, we found that lower plasma creatine concentrations were associated with higher odds of low protein intake and hypoalbuminemia. In contrast to low muscle mass, these associations remained materially unchanged after excluding the patients with highest plasma creatine concentrations. To assess protein intake, we used a biomarker-based method to assess dietary protein intake, thereby avoiding potential biases of classic dietary assessments, including under- and overreporting, changes in diet due to self-reflections, errors in portion size estimates, and socially desirable answers [15, 57, 58]. Additionally, we also found that higher plasma creatine values are associated with a reduced odds of severe fatigue. Although fatigue is not a part of the diagnostic criteria for PEW, it is an often-occurring symptom in hemodialysis patients, especially in those with PEW [59, 60]. We purposely included fatigue, as fatigue is an often under-recognized and under-treated symptom and is one of the most debilitating symptoms experienced during hemodialysis treatment from a patient's perspective. This is well-illustrated by a study demonstrating that 94% of the patients would accept more intense hemodialysis if it would increase their energy level, whereas merely 19% would accept this for an increase in longevity by 3 years [61].

The lower plasma values of guanidinoacetate and creatine compared to previous reports in healthy individuals and the found associations of plasma creatine with characteristics of PEW points toward a potential role for creatine supplementation in hemodialysis patients. Although the absolute decrease in plasma creatine from 26 to 21 $\mu\text{mol/L}$ seems rather small, the values should be interpreted in the context of the fact that creatine, like creatinine, is a small, water-soluble molecule, of which one would anticipate plasma concentrations to be high rather than low in the case of renal failure. Although

reported variation in fasting plasma creatine concentrations is high [62, 63], the median fasting plasma concentration of 26 $\mu\text{mol/L}$ observed in the hemodialysis patients in the current report is appreciably lower than reported mean values of 40.5 $\mu\text{mol/L}$ in 6 healthy subjects in a creatine supplementation study and 36.7 $\mu\text{mol/L}$ in 4,735 subjects of the general population [62, 63]. Interestingly, in the latter study, there was a significant inverse association between plasma creatine and plasma creatinine concentrations, which is in line with the low plasma creatine concentration that we observed in the current study, and signifies the likelihood of an important role of the kidney in endogenous creatine synthesis, which may be virtually absent in the context of dialysis-dependent chronic kidney disease [5, 7, 63]. Since hemodialysis is known to induce a generalized proteolytic response, particularly stimulating proteolysis from muscles [64], the falls in circulating concentrations of amino acids are lower and actual losses into the dialysate higher than one would expect in the absence of a proteolytic response [65]. Our data suggest that a similar scenario applies to creatine, with a relatively limited decrease in circulating concentrations and relatively large losses during hemodialysis. Creatine, other than amino acids, is important to be separately considered, because of its particular role in energy metabolism and bodily functions, including performance and cognitive functioning [4]. The hypothesis that dialysis treatment leads to lower intracellular creatine concentrations has also been demonstrated using imaging studies obtained by non-invasive *in vivo* ^{31}P -NMR demonstrating lower phosphocreatine concentrations in the hearts of patients on either hemodialysis treatment [46] or peritoneal dialysis treatment [47].

Creatine supplementation has been studied extensively in non-CKD settings, where it has been shown that creatine supplementation together with moderate resistance training will counteract loss of muscle mass, caused by either age-related sarcopenia or due to immobilization (recently reviewed in [66]). Furthermore, evidence indicates that creatine supplementation improves brain health, improves cognition, and is effective in alleviating brain ischemia and hypoxia (recently reviewed in [67]). Furthermore, the daily average intake of creatine via meat and fish by the general population of the U.S.A. is 1.38 g of creatine per day [68]. The same study demonstrated that 42.8% of the participants had an average intake below the recommended level of 1 g of creatine per day [68]. It is very likely that this percentage would be even higher if hemodialysis patients were studied, for they are generally advised to consume less meat and fish compared to the general population [69, 70] and thus they are likely to be even more creatine deficient. Nonetheless, to date, only a few creatine supplementation trials

have been conducted in hemodialysis patients. Nearly 20 years ago, Chang et al. [71] performed a small-scale randomized double-blind placebo-controlled trial of creatine supplementation to evaluate the potential of creatine to alleviate hemodialysis-associated muscle cramps in 10 hemodialysis patients. After 4 weeks of creatine supplementation, patients demonstrated a 60% reduction in muscle cramps, while no difference was found in the placebo group. After the wash-out period, the frequency of muscle cramps returned to the previous level [71]. More recently, Marini et al. (2019) performed a randomized double-blind placebo-controlled trial to evaluate whether oral creatine supplementation could attenuate loss of muscle mass and could reduce the malnutrition-inflammation score in 28 hemodialysis patients [72]. Compared to the placebo arm, the creatine treated arm demonstrated a significant increase in lean body mass after 4 weeks supplementation (effect size = 0.832 kg; $P = 0.03$). Additionally, the creatine treated arm demonstrated a sizable reduction of the malnutrition-inflammation score (effect size = 0.964; $P = 0.01$). In the latter study, after a loading phase with four times daily five grams of oral creatine for one week, the maintenance dosage was five grams of creatine per day. Besides potential problems with compliance, creatine also requires a large volume of water to be dissolved, which may negatively influence the fluid balance in patients already on a fluid restriction. In future studies, this may be circumvented by using intradialytic creatine supplementation [8], which also prevents the unopposed removal of creatine by hemodialysis treatment.

The strengths of this study are that we collected the entire dialysate production during the hemodialysis session instead of taking several samples, thereby increasing the accuracy of our assessments. In addition, we used a biomarker-based method to assess dietary protein intake, thereby avoiding potential biases of classic dietary assessments. However, we acknowledge that our study has limitations, primarily the relatively small sample size of this study. Our current study was too small to allow for analyses stratified for e.g. presence or absence of residual diuresis. Future larger studies are required to allow for investigation whether associations of plasma creatine with low muscle mass, low protein intake, hypoalbuminemia and severe fatigue differ in different strata of patients, e.g. strata of patients with or without residual diuresis. Additionally, we were unable to perform repeated measurements over time, to evaluate creatine homeostasis over a longer period of chronic hemodialysis treatment and to evaluate the associations between creatine status and loss of muscle mass and body weight over

a prolonged time. Lastly, statistical significance generally does not necessarily indicate a biologically relevant effect and the possibility of residual confounding remains.

In conclusion, we investigated pre- and postdialysis plasma concentrations of arginine, guanidinoacetate, creatine and creatinine. Compared to available literature, plasma concentrations of guanidinoacetate and creatine were lower than in healthy individuals. In addition, we demonstrated that arginine, guanidinoacetate, creatine and creatinine are removed by hemodialysis, with a large intradialytic shift of creatine from the intra- to the extracellular compartment. Lastly, we demonstrate that lower plasma creatine concentrations are associated with higher odds of low muscle mass, low protein intake, hypoalbuminemia, and severe fatigue. Future research is required to determine whether creatine supplementation can help to prevent or treat protein energy wasting and fatigue.

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s12967-021-02780-y>.

Additional file 1. Additional Tables S1–S10.

Acknowledgements

None

Authors' contributions

Conceptualization: AP, SJLB, RW, TW, and CFMF; Data curation: CAK, PdB and MRH-F; Formal analysis: AP, SJLB and CFMF; Investigation: AP, SJLB and CFMF; Methodology: AP, SJLB and CFMF; Project administration: YvdV; Roles/Writing—original draft: AP, SJLB and CFMF; Writing—review & editing: JCS, DK, YvdV, DG, CGS, IK, RW, T.W., and M.R.H-F.

Funding

None.

Data share statement

Data described in the manuscript, code book, and analytic code will be made available upon request of the editor.

Declarations

Ethics approval and consent to participate

This observational study was performed according to ethical standards laid down in the 1964 Declaration of Helsinki and its later amendments and was approved by the Medical Ethical Committee of the University Medical Center Groningen, The Netherlands. All participating patients gave written informed consent.

Consent for publication

Not applicable.

Competing interests

T.W. is engaged as a scientific consultant for a start-up company, "Crearene", Switzerland. The other authors declare no conflict of interest.

Author details

¹ Department of Internal Medicine, University of Groningen, University Medical Center Groningen, 9713 GZ Groningen, The Netherlands. ² Department

of Laboratory Medicine, University of Groningen, University Medical Center Groningen, Groningen 9713 GZ, the Netherlands. ³ Dialysis Center Groningen, 9713 GZ Groningen, The Netherlands. ⁴ Department of Biology, ETH Zurich, Zurich, Switzerland.

Received: 5 January 2021 Accepted: 8 March 2021
Published online: 20 March 2021

References

- Jadeja YP, Kher V. Protein energy wasting in chronic kidney disease: an update with focus on nutritional interventions to improve outcomes. *Indian J Endocrinol Metab.* 2012;16:246–51.
- Fouque D, Kalantar-Zadeh K, Kopple J, Cano N, Chauveau P, Cuppari L, Franch H, Guarnieri G, Ikizler TA, Kaysen G, Lindholm B, Massy Z, Mitch W, Pineda E, Stenvinkel P, Trevino-Becerra A, Wanner C. A proposed nomenclature and diagnostic criteria for protein-energy wasting in acute and chronic kidney disease. *Kidney Int.* 2008;73:391–8.
- Kopple JD. Pathophysiology of protein-energy wasting in chronic renal failure. *J Nutr.* 1999;129:247S–251S.
- Wallimann T, Tokarska-Schlattner M, Schlattner U. The creatine kinase system and pleiotropic effects of creatine. *Amino Acids.* 2011;40:1271–96.
- Brosnan JT, Brosnan ME. Creatine: endogenous metabolite, dietary, and therapeutic supplement. *Annu Rev Nutr.* 2007;27:241–61.
- Brosnan JT, da Silva RP, Brosnan ME. The metabolic burden of creatine synthesis. *Amino Acids.* 2011;40:1325–31.
- Post A, Tsikas D, Bakker SJL. Creatine is a conditionally essential nutrient in chronic kidney disease: a hypothesis and narrative literature review. *Nutrients.* 2019. <https://doi.org/10.3390/nu11051044>.
- Wallimann T, Riek U, Moddel M. Intradialytic creatine supplementation: a scientific rationale for improving the health and quality of life of dialysis patients. *Med Hypotheses.* 2017;99:1–14.
- Wyss M, Kaddurah-Daouk R. Creatine and creatinine metabolism. *Physiol Rev.* 2000;80:1107–213.
- Guthmiller P, Van Pilsom JF, Boen JR, McGuire DM. Cloning and sequencing of rat kidney L-arginine:glycine amidinotransferase. Studies on the mechanism of regulation by growth hormone and creatine. *J Biol Chem.* 1994;269:17556–60.
- Walker JB. Repression of arginine-glycine transaminase activity by dietary creatine. *Biochim Biophys Acta.* 1959;36:574–5.
- Walker JB. Creatine: biosynthesis, regulation, and function. *Adv Enzymol Relat Areas Mol Biol.* 1979;50:177–242.
- Garcia-Miranda P, Peral MJ, Ilundain AA. Effect of antidiuresis on renal creatine metabolism. *J Physiol Pharmacol.* 2010;61:83–8.
- McGuire DM, Gross MD, Van Pilsom JF, Towle HC. Repression of rat kidney L-arginine:glycine amidinotransferase synthesis by creatine at a pretranslational level. *J Biol Chem.* 1984;259:12034–8.
- Post A, Ozyilmaz A, Westerhuis R, Ipema KJR, Bakker SJL, Franssen CFM. Complementary biomarker assessment of components absorbed from diet and creatinine excretion rate reflecting muscle mass in dialysis patients. *Nutrients.* 2018. <https://doi.org/10.3390/nu10121827>.
- Post A, Huberts M, Poppe E, Faassen MV, Kema IP, Vogels S, Geleijnse JM, Westerhuis R, Ipema KJR, Bakker SJL, Franssen CFM. Tryptophan intake and tryptophan losses in hemodialysis patients: a balance study. *Nutrients.* 2019. <https://doi.org/10.3390/nu11122851>.
- Poppe ESJM, Polinder-Bos HA, Huberts M, Vogels S, Ipema KJR, Gansevoort RT, Westerhuis R, Bakker SJL, Gaillard CAJM, Franssen CFM. Creatinine synthesis rate and muscle strength and self-reported physical health in dialysis patients. *Clin Nutr.* 2019;39:1600–7.
- Vascular Access Work Group. Clinical practice guidelines for vascular access. *Am J Kidney Dis.* 2006;48(Suppl 1):S248–73.
- Du Bois D, Du Bois EF. A formula to estimate the approximate surface area if height and weight be known. *Nutrition.* 1989;5:303–11 (**discussion 312-3**).
- Goertz YMJ, Looijmans M, Prins JB, Janssen DJA, Thong MSY, Peters JB, Burtin C, Meertens-Kerris Y, Coors A, Muris JWM, Sprangers MAG, Wouters EFM, Vercoulen JH, Spruit MA. Fatigue in patients with chronic obstructive pulmonary disease: protocol of the Dutch multicentre, longitudinal, observational FANTasTIGUE study. *BMJ Open.* 2018;8:e021745.
- Vercoulen JH, Swanink CM, Fennis JF, Galama JM, van der Meer JW, Bleijenberg G. Dimensional assessment of chronic fatigue syndrome. *J Psychosom Res.* 1994;38:383–92.
- Van Herck M, Spruit MA, Burtin C, Djamin R, Antons J, Goertz YMJ, Ebadi Z, Janssen DJA, Vercoulen JH, Peters JB, Thong MSY, Otker J, Coors A, Sprangers MAG, Muris JWM, Wouters EFM, van 't Hul AJ. Fatigue is highly prevalent in patients with asthma and contributes to the burden of disease. *J Clin Med.* 2018. <https://doi.org/10.3390/jcm7120471>.
- Worm-Smeitink M, Gielissen M, Bloot L, van Laarhoven HWM, van Engelen BGM, van Riel P, Bleijenberg G, Nikolaus S, Knoop H. The assessment of fatigue: psychometric qualities and norms for the Checklist individual strength. *J Psychosom Res.* 2017;98:40–6.
- Bultmann U, de Vries M, Beurskens AJ, Bleijenberg G, Vercoulen JH, Kant I. Measurement of prolonged fatigue in the working population: determination of a cutoff point for the checklist individual strength. *J Occup Health Psychol.* 2000;5:411–6.
- van Sandwijk MS, Al Arashi D, van de Hare FM, van der Torren JMR, Kersten MJ, Bijlsma JA, Ten Berge IJM, Bemelman FJ. Fatigue, anxiety, depression and quality of life in kidney transplant recipients, haemodialysis patients, patients with a haematological malignancy and healthy controls. *Nephrol Dial Transplant.* 2019;34:833–8.
- Daugirdas JT. Second generation logarithmic estimates of single-pool variable volume Kt/V: an analysis of error. *J Am Soc Nephrol.* 1993;4:1205–13.
- Abraham AG, Munoz A, Furth SL, Warady B, Schwartz GJ. Extracellular volume and glomerular filtration rate in children with chronic kidney disease. *Clin J Am Soc Nephrol.* 2011;6:741–7.
- Bird NJ, Henderson BL, Lui D, Ballinger JR, Peters AM. Indexing glomerular filtration rate to suit children. *J Nucl Med.* 2003;44:1037–43.
- Janssen I, Heymsfield SB, Ross R. Low relative skeletal muscle mass (sarcopenia) in older persons is associated with functional impairment and physical disability. *J Am Geriatr Soc.* 2002;50:889–96.
- Iacone R, D'Elia L, Guida B, Barbato A, Scanzano C, Strazzullo P. Validation of daily urinary creatinine excretion measurement by muscle-creatinine equivalence. *J Clin Lab Anal.* 2018;32:e22407.
- Cruz-Jentoft AJ, Bahat G, Bauer J, Boirie Y, Bruyere O, Cederholm T, Cooper C, Landi F, Rolland Y, Sayer AA, Schneider SM, Sieber CC, Topinkova E, Vandewoude M, Visser M, Zamboni M. Writing Group for the European Working Group on Sarcopenia in Older People 2 (EWGSOP2), and the Extended Group for EWGSOP2: Sarcopenia: revised European consensus on definition and diagnosis. *Age Ageing.* 2019;48:16–31.
- Janssen I, Baumgartner RN, Ross R, Rosenberg IH, Roubenoff R. Skeletal muscle cutpoints associated with elevated physical disability risk in older men and women. *Am J Epidemiol.* 2004;159:413–21.
- Maroni BJ, Steinman TI, Mitch WE. A method for estimating nitrogen intake of patients with chronic renal failure. *Kidney Int.* 1985;27:58–65.
- Kalhan SC, Gruca L, Marczewski S, Bennett C, Kummitha C. Whole body creatine and protein kinetics in healthy men and women: effects of creatine and amino acid supplementation. *Amino Acids.* 2016;48:677–87.
- Lancet T. Taking sex into account in medicine. *The Lancet.* 1826;2011:378.
- Schiebinger L, Leopold SS, Miller VM. Editorial policies for sex and gender analysis. *Lancet.* 2016;388:2841–2.
- Wallimann T, Wyss M, Brdiczka D, Nicolay K, Eppenberger HM. Intracellular compartmentation, structure and function of creatine kinase isoenzymes in tissues with high and fluctuating energy demands: the "phosphocreatine circuit" for cellular energy homeostasis. *Biochem J.* 1992;281(Pt 1):21–40.
- Wallimann T, Tokarska-Schlattner M, Neumann D, Epanand RM, Epanand RF, Andres RH, Widmer HR, Hornemann T, Saks V, Agarkova I, Schlattner U. The phosphocreatine circuit: molecular and cellular physiology of creatine kinases, sensitivity to free radicals, and enhancement by creatine supplementation. In: *Molecular system bioenergetics*; 2007. p. 195–264.
- Wallimann T, Harris R. Creatine: a miserable life without it. *Amino Acids.* 2016;48:1739–50.
- Schulze A. Creatine deficiency syndromes. *Handb Clin Neurol.* 2013;113:1837–43.
- Guimbal C, Kilimann MW. A Na(+)-dependent creatine transporter in rabbit brain, muscle, heart, and kidney. cDNA cloning and functional expression. *J Biol Chem.* 1993;268:8418–21.
- Marescau B, Nagels G, Possemiers I, De Broe ME, Becaus I, Billioud JM, Lornoy W, De Deyn PP. Guanidino compounds in serum and urine of

- nondialyzed patients with chronic renal insufficiency. *Metabolism*. 1997;46:1024–31.
43. Lea-Henry TN, Carland JE, Stocker SL, Sevastos J, Roberts DM. Clinical pharmacokinetics in kidney disease: fundamental principles. *Clin J Am Soc Nephrol*. 2018;13:1085–95.
 44. Delanghe J, De Slypere JP, De Buyzere M, Robbrecht J, Wieme R, Vermeulen A. Normal reference values for creatine, creatinine, and carnitine are lower in vegetarians. *Clin Chem*. 1989;35:1802–3.
 45. Forsberg AM, Nilsson E, Werneman J, Bergstrom J, Hultman E. Muscle composition in relation to age and sex. *Clin Sci (Lond)*. 1991;81:249–56.
 46. Tagami T, Sakuma H, Matsumura K, Takeda K, Mori S, Takeuchi T, Nakano T. Evaluation of altered myocardial high energy phosphate metabolism in patients on maintenance dialysis using phosphorus-31 magnetic resonance spectroscopy. *Invest Radiol*. 1998;33:171–6.
 47. Ogimoto G, Sakurada T, Imamura K, Kuboshima S, Maeba T, Kimura K, Owada S. Alteration of energy production by the heart in CRF patients undergoing peritoneal dialysis. *Mol Cell Biochem*. 2003;244:135–8.
 48. Nishida A, Kubo K, Umeda M, Ando A, Nihei H, Sugino N. Clinical use of ³¹P-nuclear magnetic resonance spectroscopy for muscle damage in uremia. *Nephron*. 1991;57:489–90.
 49. Heymsfield SB, Arteaga C, McManus C, Smith J, Moffitt S. Measurement of muscle mass in humans: validity of the 24-hour urinary creatinine method. *Am J Clin Nutr*. 1983;37:478–94.
 50. Polinder-Bos HA, Nacak H, Dekker FW, Bakker SJL, Gaillard CAJM, Gansevoort RT. Low urinary creatinine excretion is associated with self-reported frailty in patients with advanced chronic kidney disease. *Kidney Int Rep*. 2017;2:676–85.
 51. Wang ZM, Gallagher D, Nelson ME, Matthews DE, Heymsfield SB. Total-body skeletal muscle mass: evaluation of 24-h urinary creatinine excretion by computerized axial tomography. *Am J Clin Nutr*. 1996;63:863–9.
 52. Proctor DN, O'Brien PC, Atkinson EJ, Nair KS. Comparison of techniques to estimate total body skeletal muscle mass in people of different age groups. *Am J Physiol*. 1999;277:E489–95.
 53. Welle S, Thornton C, Totterman S, Forbes G. Utility of creatinine excretion in body-composition studies of healthy men and women older than 60 y. *Am J Clin Nutr*. 1996;63:151–6.
 54. Stam SP, Oste MCJ, Eisenga MF, Blokzijl H, van den Berg AP, Bakker SJL, de Meijer VE. Posttransplant muscle mass measured by urinary creatinine excretion rate predicts long-term outcomes after liver transplantation. *Am J Transplant*. 2018;19:540–50.
 55. Abrahamsen B, Hansen TB, Hogsberg IM, Pedersen FB, Beck-Nielsen H. Impact of hemodialysis on dual X-ray absorptiometry, bioelectrical impedance measurements, and anthropometry. *Am J Clin Nutr*. 1996;63:80–6.
 56. Tosato M, Marzetti E, Cesari M, Saveria G, Miller RR, Bernabei R, Landi F, Calvani R. Measurement of muscle mass in sarcopenia: from imaging to biochemical markers. *Aging Clin Exp Res*. 2017;29:19–27.
 57. Shim JS, Oh K, Kim HC. Dietary assessment methods in epidemiologic studies. *Epidemiol Health*. 2014;36:e2014009.
 58. Naska A, Lagioui A, Lagioui P. Dietary assessment methods in epidemiological research: current state of the art and future prospects. *F1000Res*. 2017;6:926.
 59. Sgambat K, Matheson MB, Hooper SR, Warady B, Furth S, Moudgil A. Prevalence and outcomes of frailty: a frailty-inflammation phenotype in children with chronic kidney disease. *Pediatr Nephrol*. 2019;34:2563–9.
 60. Vanden Wyngaert K, Van Craenenbroeck AH, Eloot S, Calders P, Celie B, Holvoet E, Van Biesen W. Associations between the measures of physical function, risk of falls and the quality of life in haemodialysis patients: a cross-sectional study. *BMC Nephrol* 2020, 21:7-019-1671-9.
 61. Ramkumar N, Beddhu S, Eggers P, Pappas LM, Cheung AK. Patient preferences for in-center intense hemodialysis. *Hemodial Int*. 2005;9:281–95.
 62. Jager R, Harris RC, Purpura M, Francaux M. Comparison of new forms of creatine in raising plasma creatine levels. *J Int Soc Sports Nutr*. 2007;4:1–5.
 63. Post A, Groothof D, Schutten JC, Flores-Guerrero JL, Casper Swarte J, Douwes RM, Kema IP, de Boer RA, Garcia E, Connelly MA, Wallimann T, Dullaart RPF, Franssen CFM, Bakker SJL. Plasma Creatine and Incident Type 2 Diabetes in a General Population-Based Cohort: The PREVENT Study. *Clin Endocrinol (Oxf)* 2020.
 64. Ikizler TA, Pupim LB, Brouillette JR, Levenhagen DK, Farmer K, Hakim RM, Flakoll PJ. Hemodialysis stimulates muscle and whole body protein loss and alters substrate oxidation. *Am J Physiol Endocrinol Metab*. 2002;282:E107–16.
 65. Hendriks FK, Smeets JSJ, Broers NJH, van Kranenburg JMX, van der Sande FM, Kooman JP, van Loon LJC. End-stage renal disease patients lose a substantial amount of amino acids during hemodialysis. *J Nutr* 2020.
 66. Kreider RB, Stout JR. Creatine in health and disease. *Nutrients*. 2021. <https://doi.org/10.3390/nu13020447>.
 67. Roschel H, Gualano B, Ostojic MS, Rawson SE. Creatine Supplementation and Brain Health. *Nutrients*. 2021. <https://doi.org/10.3390/nu13020586>.
 68. Ostojic SM. Dietary creatine intake in U.S. population: NHANES 2017–2018. *Nutrition* 2021,111207.
 69. Kalantar-Zadeh K, Fouque D. Nutritional management of chronic kidney disease. *N Engl J Med*. 2017;377:1765–76.
 70. Carrero JJ, Gonzalez-Ortiz A, Avesani CM, Bakker SJL, Bellizzi V, Chauveau P, Clase CM, Cupisti A, Espinosa-Cuevas A, Molina P, Moreau K, Piccoli GB, Post A, Sezer S, Fouque D. Plant-based diets to manage the risks and complications of chronic kidney disease. *Nat Rev Nephrol*. 2020;16:525–42.
 71. Chang CT, Wu CH, Yang CW, Huang JY, Wu MS. Creatine monohydrate treatment alleviates muscle cramps associated with haemodialysis. *Nephrol Dial Transplant*. 2002;17:1978–81.
 72. Marini ACB, Motobu RD, Freitas ATV, Mota JF, Wall BT, Pichard C, Laviano A, Pimentel GD. Short-term creatine supplementation may alleviate the malnutrition-inflammation score and lean body mass loss in hemodialysis patients: a pilot randomized placebo-controlled trial. *JPEN J Parenter Enteral Nutr*. 2019;44:815–22.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

