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Published in: Archives of Physiology and Biochemistry

DOI: 10.3109/13813455.2010.543136

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Document Version Publisher's PDF, also known as Version of record

Publication date: 2011

Link to publication in University of Groningen/UMCG research database

Citation for published version (APA): Koonen, D. P. Y., Jensen, M. K., & Handberg, A. (2011). Soluble CD36-a marker of the (pathophysiological) role of CD36 in the metabolic syndrome? *Archives of Physiology and Biochemistry*, 117(2), 57-63. https://doi.org/10.3109/13813455.2010.543136

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**Archives Of Physiology And Biochemistry** 

ISSN: 1381-3455 (Print) 1744-4160 (Online) Journal homepage: https://www.tandfonline.com/loi/iarp20

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To link to this article: https://doi.org/10.3109/13813455.2010.543136



Published online: 21 Jan 2011.



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#### **REVIEW ARTICLE**

# Soluble CD36– a marker of the (pathophysiological) role of CD36 in the metabolic syndrome?

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#### Abstract

CD36 is a class B scavenger receptor observed in many cell types and tissues throughout the body. Recent literature has implicated CD36 in the pathogenesis of metabolic dysregulation such as found in obesity, insulin resistance, and atherosclerosis. Genetic variation at the *CD36* loci have been associated with obesity and lipid components of the metabolic syndrome, with risk of heart disease and type 2 diabetes. Recently, non-cell bound CD36 was identified in human plasma and was termed soluble CD36 (sCD36). In this review we will describe the functions of CD36 in tissues and address the role of sCD36 in the context of the metabolic syndrome. We will also highlight recent findings from human genetic studies looking at the *CD36* locus in relation to metabolic profile in the general population. Finally, we present a model in which insulin resistance, oxLDL, low-grade inflammation and liver steatosis may contribute to elevated levels of sCD36.

Keywords: Obesity; insulin resistance; type 2 diabetes; inflammation; ox-LDL; NAFLD

#### Introduction

Type 2 Diabetes (T2D) is one of the most costly and burdensome chronic diseases of our time. It is associated with the development of devastating complications and a significantly higher risk for cardiovascular disease. Unfortunately, T2D is becoming gradually more common as life expectancy is increasing and obesity rates are rising. Increased expression of the class B scavenger receptor CD36 has been observed in many cell types and tissues throughout the body and has been implicated in the pathogenesis of atherosclerosis (Collot-Teixeira et al., 2007; Silverstein, 2009) and metabolic disease (Glatz et al., 2010). In addition, a region along chromosome 7q, containing the CD36 gene, has been linked to components of the metabolic syndrome in several genome-wide linkage studies (An et al., 2005; Arya et al., 2002; Malhotra et al., 2007). Variants in the CD36 gene have now been shown to influence the susceptibility for the metabolic syndrome, and associate with risk of heart disease and Type 2 Diabetes (T2D) (Love-Gregory et al., 2008, 2010).

Recently, non-cell bound CD36 was identified in human plasma and was termed soluble CD36 (sCD36) (Handberg et al., 2006). sCD36 clusters with markers of insulin resistance and is progressively related to the severity of insulin resistance and atherosclerosis in the human population (Handberg et al., 2006, 2008, 2010). Interestingly, sCD36 parallels the increased CD36 expression observed in multiple cell types and tissues in human and rodent models of insulin resistance and T2D (Aguer et al., 2010, Bonen et al., 2004; Griffin et al., 2001; Koonen et al., 2007; Luiken et al., 2001; Sampson et al., 2003). Indeed, results from association studies have led to the hypothesis that sCD36 reflects tissue CD36 expression level, and in particular monocyte and macrophage expression level (Handberg et al., 2006, 2008, 2009). As elevated sCD36 may be a marker of increased CD36 expression derived from a number of tissues associated with the metabolic syndrome, this review will discuss the proposed (patho)-physiological role of CD36 in atherosclerosis and metabolic disease. We will highlight recent findings from human genetic studies that examined the CD36 locus in relation to metabolic profile

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<sup>(</sup>Received 31 October 2010; revised 20 November 2010; accepted 22 November 2010)

in the general population, and propose a model in which insulin resistance, oxLDL, low-grade inflammation and liver steatosis may contribute to increased soluble CD36 in the circulation.

## Role of CD36 in foam cell formation, platelet activation and fatty acid uptake

CD36 is a heavily glycosylated membrane protein containing two predicted transmembrane areas, two short intracellular domains and a large extra-cellular loop (Rac et al., 2007; Su and Abumrad, 2009). CD36 is involved in many different functions depending on cell-type and ligand-specific binding. Membrane CD36 in monocytes and macrophages bind and internalize oxLDL (Kunjathoor et al., 2002; Podrez et al., 2002). In addition, CD36 expression is up-regulated by oxLDL, and thus CD36 plays an important role in formation of foam-cells, which may eventually turn into atherosclerotic plaques in the arterial wall (Febbraio et al., 2000; Podrez et al., 2000). In platelets and endothelial cells CD36 functions as an adhesion molecule, and through interaction with anionic phospholipids present in, for example, oxLDL, CD36 is involved in platelet activation (Podrez et al., 2007, Valiyaveettil and Podrez, 2009). In metabolically active tissues CD36 has been shown to bind long-chain fatty acids (LCFA) and facilitate LCFA uptake across the plasma membrane (for review, see Glatz et al., 2010). Increased recruitment of CD36 from intracellular storage sites towards the plasma membrane has been shown to regulate fatty acid (FA) metabolism under various physiological conditions, however, is also associated with enhanced efficiency of FA uptake and lipid accumulation in rodent and human muscle (Aguer et al., 2010, Bonen et al., 2004, Holloway et al., 2009, Smith et al., 2007), and may therefore contribute to the aetiology of insulin resistance.

### Obesity, low-grade inflammation, and the metabolic syndrome

The metabolic syndrome describes a cluster of factors associated with an increased risk of T2D and atherosclerosis. Among many definitions, the newly published IDF criteria include: abdominal obesity and two out of the following four components: elevated triglyceride, reduced HDL-cholesterol, elevated systolic blood pressure or diastolic blood pressure, and increased fasting glucose (Alberti *et al.*, 2006).

Increased adiposity and lipid accumulation in liver and skeletal muscle have strongly been associated with the development of insulin resistance and T2D. Intracellular lipid metabolites, including fatty acyl-CoA and diacylglycerol, have been shown to interfere with normal insulin signalling at the level of insulin receptormediated tyrosine phosphorylation and at downstream sites within the insulin signalling cascade (Hegarty *et al.*, 2003).

Insulin resistance and T2D are, like atherosclerosis, associated with systemic low-grade inflammation (for review, see Shoelson et al., 2007 and Rocha and Libby, 2009). Abdominal and visceral obesity in particular has been proposed to induce a low-grade inflammatory condition. The prevailing understanding is that distension of fat cells above a critical level influence vascularization of fat tissue through an adipokine dependent mechanism. The resulting fat cell necrosis is followed by infiltration of macrophages and a local low-grade inflammation, which may become systemic (Kloting et al., 2010; Shoelson et al., 2006). Cytokines like TNFalpha induce insulin resistance in muscle, fat and liver, and inflammation and insulin resistance stimulate atherosclerosis development through associated dyslipidaemia, per-oxidation of lipoproteins, local influences on the vessel wall among other mechanisms (Holvoet et al., 2008; Rocha and Libby, 2009; Shoelson et al., 2007).

## Elevated expression of CD36 associated with T2D and metabolic syndrome

Over the years evidence has accumulated that implicate CD36 in the development of atherosclerosis and metabolic disease. Lipid accumulation in human obesity and T2D is associated with increased rates of skeletal muscle FA transport and increased CD36 expression due to enhanced membrane recruitment of CD36 (Aguer et al., 2010; Bonen et al., 2004). CD36 expression is increased in adipose tissue and skeletal muscle in human obesity and T2D (Bonen et al., 2004, 2006), and in liver biopsies from non-alcoholic liver disease (NAFLD) patients correlating with the degree of steatosis (Greco et al., 2008). CD36 is also upregulated in vascular lesions derived from hyperglycaemic patients and in human monocytederived macrophages differentiated in the presence of high glucose, providing a mechanism for accelerated atherosclerosis in diabetic patients (Griffin et al., 2001). In addition, recent data provided new evidence to support the atherogenic property of CD36 by demonstrating that lipid-induced insulin resistance following a 24-h Intralipid/heparin infusion is associated with increased monocyte expression of CD36 and internalization of oxidized LDL in healthy subjects (Kashyap et al., 2009). Consistent with this idea, a delay in atherosclerosis development was observed in CD36-deficient mice bred on an ApoE-/-background compared to Apo-E-/- mice, whereas re-introduction of CD36 into the double-knockout mice resulted in a significant increase in the degree of atherosclerosis (Febbraio et al., 2000).

Increased CD36 expression, enhanced efficiency in FA uptake and increased FA esterification has also been observed in animal models of insulin resistance and T2D, including high-fat feeding and obesity (Coort *et al.*, 2004a; Han *et al.*, 2007; Hegarty *et al.*, 2002; Holloway *et al.*, 2009; Luiken *et al.*, 2001). In heart and skeletal muscle, CD36 was found in excess on the cell surface, which was demonstrated to be due to a permanent relocation of CD36

from its intracellular storage pool towards the membrane (for review, see Glatz *et al.*, 2010). This permanent relocation towards the cell membrane was shown to be an early event in the development of insulin resistance (Coort *et al.*, 2004b; Ouwens *et al.*, 2007). Interestingly, CD36 turnover is abnormally slow in macrophages from insulin resistant ob/ob mice reflecting a defect in CD36 receptor trafficking in response to altered insulin signalling (Liang *et al.*, 2004). In liver, increased CD36 expression in response to diet-induced obesity was shown to be sufficient to exacerbate hepatic triglyceride storage and secretion, confirming a role for CD36 in the pathogenesis of metabolic disease (Koonen *et al.*, 2007).

Interestingly, a combination of metformin and exercise was shown to reduce muscle CD36 expression and lipid accumulation and blunt the progression of high-fat diet induced hyperglycaemia in rats (Smith *et al.*, 2007). Moreover, green tea polyphenols and cinnamon extract had a similar reducing effect on CD36 expression in heart and adipose tissue from high fructose-fed rats (Qin *et al.*, 2010a, b), suggesting that CD36 might represent a potential therapeutic target for the prevention and/or treatment of insulin resistance.

Although the mechanisms are still under investigation, not one single factor is likely to explain the altered CD36 expression seen in atherosclerosis and metabolic disease. Some of those factors are inter-related, such as glucose, plasma lipids and insulin resistance, inflammation and oxidative stress, most of which are predisposing for atherosclerosis. Therefore, defective CD36 expression might reflect an intricate mechanism involving the interaction of multiple factors at different stages in time.

### Genetic variation at the CD36 locus and metabolic profile in humans

Other strategies for the investigation of CD36 in metabolic diseases in humans include the study of naturally occurring genetic variants in population-based data. Several genome-wide linkage studies identified a region along chromosome 7 (7q11.2-7q21.11), containing the CD36 gene, that was linked to components of the metabolic syndrome (An et al., 2005; Arya et al., 2002; Malhotra et al., 2007). In other, more focused, candidate gene studies of the CD36 locus, investigators gather information on the metabolic and cardiovascular consequences of lifelong exposure to genetic variants that may produce CD36 gene-products with differential functionality. Already, several single nucleotide polymorphisms (SNPs) at the CD36 locus have been found to be associated with parameters of lipid metabolism, insulin sensitivity, and cardiovascular disease. Studies in Europeans found CD36 SNPs associated with adiponectin (Lepretre et al., 2004a, b), serum free FA and triglyceride levels (Ma et al., 2004), fasting plasma glucose, insulin resistance, risk of T2DM (Corpeleijn et al., 2010), and levels of highdensity and low-density lipoprotein cholesterol (HDL-C and LDL-C) (Goyenechea et al., 2008). Recently a group of publications have explored the role of genetic variants in CD36 in relation to adiposity. One study suggested that four SNPs, known from the previous studies of lipid traits, were associated with adolescent obesity without any evidence for an association with plasma lipids (Bokor et al., 2009). However, in the largest study to date, a metaanalysis of almost 10,000 participants neither supported the evidence for the previously indicated SNPs, nor suggested any other SNPs captured on their genome-wide arrays, to play a role in early onset obesity in European adolescents (Choquet et al., 2010). Conversely, a simultaneous publication of six genotyped CD36 SNPs in 1790 non-diabetic Germans found associations with measures of body-mass index and waist circumference, and no associations with detailed measures of glucose tolerance, insulin sensitivity, triglycerides, HDL-C, and LDL-C (Heni et al., 2010). Some studies have indicated that the impact of CD36 variants may differ across population characteristics (Corpeleijn et al., 2010). So far, there has been no detailed investigation of context-dependent effects of the CD36 SNPs on the metabolic traits of interest but certainly the discrepancy in the findings between studies conducted within populations with differences in population-characteristics that might play a role for the observed association between genotype and metabolic outcomes (such as age, gender, obesity and smoking), highlight that exploration of gene-environment interacts will be of interest for future studies.

CD36 is also a receptor for Plasmodium falciparum infected erythrocytes and therefore linked to malaria susceptibility. Thus, the polymorphisms' in CD36 differ in frequency and position between European and non-European populations due to selection. Investigations of a CD36 SNP that encodes a truncated protein (rs3211938) found in African and Asian populations, suggests that decreased CD36 expression as found in heterozygotes is more advantageous for metabolic traits as compared to complete CD36 deficiency as observed in the homozygous carriers of this variant (Love-Gregory et al., 2008). In addition, the HyperGen population sample of 2020 African-Americans found 15 other CD36 SNPs that were also associated with HDL-cholesterol. Recently this investigator group followed up their initial HDL-related CD36 SNPs with detailed assessments of CD36 expression on monocytes and platelets (Love-Gregory et al., 2010). Here they observed that the SNPs that were associated with an advantageous metabolic profile were also associated with less CD36 expression. It is of equally great interest to address whether such kind of functionality can also be established for the above-mentioned metabolically relevant SNPs that were identified in European populations.

### sCD36, a novel marker for insulin resistance, atherosclerosis and inflammation

Recently, non-cell bound CD36 (soluble CD36; sCD36) was identified in the human population and a simple

sandwich ELISA suitable to measure sCD36 in plasma was developed (Handberg et al., 2006). This is convenient as compared to tissue CD36- both in terms of sample accessibility and the methodological procedure. This new measure of plasma CD36 has been investigated in studies of sCD36 in T2D, insulin resistance, and atherosclerosis where altered expression levels of CD36 could play an important pathophysiological role. In particular enhanced CD36 in monocytes and macrophages induced by hyper-glycaemia and insulin resistance is believed to be an important link between T2D and atherosclerosis. Consistent with this hypothesis, sCD36 is up to 4-fold higher in plasma from obese T2D-patients compared with lean healthy control subjects and tightly associated with insulin resistance (Handberg et al., 2006). Likewise, in chronic kidney disease (CKD) elevated sCD36 in serum was associated with the presence of T2D (Chmielewski et al., 2010). Furthermore, insulin resistance in nondiabetic obese individuals as well as in women with polycystic ovarian syndrome (PCOS) was associated with increased sCD36 (Glintborg et al., 2008; Handberg et al., 2006). In addition to the consistent correlation between insulin resistance and sCD36, pioglitazone treatment reduced sCD36 while improving insulin-sensitivity in PCOS (Glintborg et al., 2008). Interestingly, the correlation between sCD36 and insulin resistance was independent of BMI or alternatively, abdominal obesity (Glintborg et al., 2008; Handberg et al., 2006). Factor analysis of data from a diabetes prediction study propose that sCD36 is associated with the insulin resistance component in a model of the metabolic syndrome, comprising blood pressure, lipids, glucose, inflammation, and obesity/ insulin resistance, and is associated with elevated risk of diabetes (Handberg et al., 2010).

In line with the important role of CD36 in atherosclerosis development sCD36 in serum was found to predict cardiovascular mortality in a cohort of CKD stage 5 patients (Chmielewski et al., 2010). Moreover, the use of HMG-CoA reductase inhibitors (statins) reduced serum concentrations of sCD36 (Chmielewski et al., 2010). sCD36 is associated with triglyceride, LDL, and inversely with HDL but not oxLDL in several patient populations (Glintborg et al., 2008; Handberg et al., 2006, 2010). In addition, a modest correlation between intima-mediathickness determined by ultra-sound, and sCD36, independent of gender and age, was reported in healthy individuals (Handberg, submitted). We found that in patients with severe carotid artery atherosclerosis, sCD36 level was higher in patients that reported recent cerebral symptoms, compared to those who did not have symptoms in the previous two months (Handberg et al., 2008) In contrast, levels of hsCRP were not different between these two group of patients. In the same patients, elevated CD36 expression found in unstable (symptomatic) atherosclerotic plaques was correlated with higher plasma sCD36 levels.

In terms of the relationship between inflammatory markers and sCD36 this was reported in glucose intolerant men, where fat-free mass and IL-6 independently contributed to sCD36, possibly through decreased insulin action (Handberg *et al.*, 2010). sCD36 has also been identified in rat plasma where is correlates with adverse lipid profiles and increased TNFalpha and IL-6 cytokine levels in plasma in rats fed a high-fructose diet (Qin *et al.*, 2010b). Moreover, addition of green tea polyphenols to the high-fructose diet completely restored sCD36 levels and alleviated the insulin-resistant cardiac phenotype in these rats (Qin *et al.*, 2010b). Ameliorations in plasma lipid profiles, sCD36 expression and insulin sensitivity were also observed in high-fructose fed rats following supplementation with cinnamon extract for 8 weeks (Qin *et al.*, 2010a).

So far, the relationship between the degree of liver fat accumulation and inflammation and sCD36 has not been investigated. However, sCD36 correlates positively with liver amino-transferase activity in subjects with glucose intolerance, indicating that sCD36 might also be a marker of liver injury (Fernandez-Real *et al.*, 2009). Indeed, even in healthy individuals, risk of non-alcoholic liver disease as scored by the fatty liver index was associated with an increase in sCD36 (Handberg, submitted). The validation of sCD36 as a risk marker of insulin resistance, inflammation and atherosclerosis is only in its beginning. However, the introduction of this easily accessible biomarker represents a promising tool for future risk stratification and monitoring of components of the metabolic syndrome.

### Proposed model of CD36 release into the circulation

Despite the fact that sCD36 has been shown to cluster with markers of the metabolic syndrome, the mechanism(s) of CD36 release into the circulation are not known.

As elevated sCD36 may be a marker of increased CD36 expression known from a number of tissues that are associated with the metabolic syndrome, single factors (hyperglycemia, oxLDL) or a combination of factors (insulin resistance, low grade inflammation) that induce CD36 expression are likely to promote CD36 release into the circulation (Figure 1). Therefore, CD36 could be released into the circulation as part of the low-grade inflammatory state commonly seen in atherosclerosis and insulin resistance (Handberg et al., 2006). Alternatively, increased plasma concentration of sCD36 could be directly related to apoptosis following cholesterol accumulation in foam-cells or to ectopic fat accumulation in general (Fernandez-Real et al., 2009; Handberg et al., 2006). However, given the wide-spread tissue expression of CD36 and its broad range of functions it is difficult to foresee which specific pathological processes may reflect alterations in sCD36, or alternatively, may induce. In addition, limited information is available about the structure of sCD36 in the circulation. To date, it has not been studied whether sCD36 consists of full-length CD36 or represents a proteolytic fragment of CD36 (i.e. the



Figure 1. Proposed model of the pathophysiology behind elevated sCD36 in plasma in the metabolic syndrome. sCD36 reflects tissue CD36 expression level, and in particular monocyte and macrophage expression level. Elevated sCD36 may be a marker of increased CD36 expression known from a number of tissues that are associated with the metabolic syndrome; macrophage infiltration and low-grade inflammation in abdominal obesity, which may lead to dyslipidemia and peroxidation of lipoproteins, and liver steatosis. A combination of these factors influence atherosclerosis development and induces increased CD36 in the vessel wall. Insulin resistance, oxLDL, low-grade inflammation and liver steatosis stimulate CD36 expression in monocytes and macrophages in fat, liver, and arteries, which lead to elevated plasma sCD36. In addition, the involvement of CD36 in ectopic fat accumulation associated with insulin resistance, dyslipidemia and inflammation in liver and skeletal muscle may result in elevated circulating sCD36.

extracellular part of CD36). Detection and measurement of sCD36 in plasma have been based on immunological approaches, and all of the antibodies used for these experiments were directed against the predicted extracellular part of CD36. Since only a minor part of CD36 is located in the intracellular or trans-membraneous areas, and since the extracellular part of CD36 is heavily glycosylated, the size of the extracellular part and that of full length CD36 can not be discriminated by Western blotting. Therefore, more research is warranted to unravel the mechanism(s) involved.

### Conclusion

The scavenger receptor and FA transport protein CD36 has been shown to play an important role in early atherosclerosis and insulin resistance. Non-cell bound CD36 (sCD36) has recently been identified in human plasma and is tightly related to risk factors of accelerated atherosclerosis in T2D such as insulin resistance and glycaemic control. Results from association studies suggest that sCD36 reflects tissue CD36 expression level, and in particular monocyte and macrophage expression level. Insulin resistance, oxLDL, low-grade inflammation and liver steatosis stimulate CD36 expression in monocytes and macrophages in fat, liver, and arteries, which lead to elevated plasma sCD36. While genetic variation at the CD36 locus has also been associated with lipid levels, free fatty acids, insulin resistance, obesity, and risk of coronary heart disease, so far evidence linking the genetic variants to plasma concentrations of sCD36 is lacking. sCD36 may thus in future studies have a potential as marker of processes central in the development of insulin

resistance and atherosclerosis related to the metabolic syndrome.

### Acknowledgements

The authors are greatly indebted to Jens K. Kjær and Morten H. Nielsen for preparation of Figure 1.

### **Declaration of interest**

DK is supported within the framework of CTMM, the Center for Translational Molecular Medicine (www. ctmm.nl), project PREDICCt (grant 01C-104), and supported by the Netherlands Heart Foundation, Dutch Diabetes Research Foundation and Dutch Kidney Foundation. MKJ is supported by a stipend from the Villum Kan Rasmussen Foundation. AH is supported by the Novo Nordisk Foundation, the Danish Diabetes Foundation, the Danish Heart Association, the Danish Research Council and Juelsgaards Mindefond.

#### References

- Aguer C, Mercier J, Man CY, Metz L, Bordenave S, Lambert K, Jean E, Lantier L, Bounoua L, Brun JF, et al. (2010). Intramyocellular lipid accumulation is associated with permanent relocation ex vivo and in vitro of fatty acid translocase (FAT)/CD36 in obese patients. *Diabetologia* 53:1151–63.
- Alberti KG, Zimmet P, SHAW J. (2006). Metabolic syndrome– a new world-wide definition. A consensus statement from the International Diabetes Federation. *Diabet Med* 23:469–80.
- An P, Freedman BI, Hanis CL, Chen YD, Weder AB., Schork NJ, Boerwinkle E, Province MA., Hsiung CA, Wu X, et al. (2005). Genome-wide linkage scans for fasting glucose, insulin, and insulin resistance in the National Heart, Lung, and Blood Institute Family Blood Pressure Program: Evidence of linkages

to chromosome 7q36 and 19q13 from meta-analysis. *Diabetes* 54:909–14.

- Arya R, Blangero J, Williams K, Almasy L, Dyer TD, Leach RJ, O'Connell P, Stern MP, Duggirala R. (2002). Factors of insulin resistance syndrome– related phenotypes are linked to genetic locations on chromosomes 6 and 7 in nondiabetic Mexican-Americans. *Diabetes* 51:841–7.
- Bokor S, Legry V, Meirhaeghe A, Ruiz JR, Mauro B, Widhalm K, Manios Y, Amouyel P, Moreno LA, Molnar D, Dallongeville J. (2009). Single-nucleotide polymorphism of CD36 locus and obesity in European adolescents. *Obesity (Silver Spring)* 18:1398-403.
- Bonen A, Parolin ML, Steinberg GR, Calles-Escandon J, Tandon NN, Glatz JF, Luiken JJ, Heigenhauser GJ, Dyck DJ. (2004). Triacylglycerol accumulation in human obesity and type 2 diabetes is associated with increased rates of skeletal muscle fatty acid transport and increased sarcolemmal FAT/CD36. *FASEB J* 18:1144–6.
- Bonen A, Tandon NN, Glatz JF, Luiken JJ, Heigenhauser GJ. (2006). The fatty acid transporter FAT/CD36 is upregulated in subcutaneous and visceral adipose tissues in human obesity and type 2 diabetes. *Int J Obes (Lond)* 30:877–83.
- Chmielewski M, Bragfors-Helin AC, Stenvinkel P, Lindholm B, Anderstam B. (2010). Serum soluble CD36, assessed by a novel monoclonal antibody-based sandwich ELISA, predicts cardiovascular mortality in dialysis patients. *Clin Chim Acta* 411:2079-82.
- Choquet H, Labrune Y, de Graeve F, Hinney A, Hebebrand J, Scherag A, Lecoer C, Tauber M, Balkau B, Elliot P, et al. (2010).Lack of association of CD36 SNPs with early onset obesity: A meta-analysis in 9973 European subjects. *Obesity (Silver Spring)*. doi:10.1038/ oby.2010.226 epub ahead of print.
- Collot-Teixeira S, Martin J, McDermott-Roe C, Poston R, McGregor JL. (2007). CD36 and macrophages in atherosclerosis. *Cardiovasc Res* 75:468-77.
- Coort SL, Hasselbaink DM, Koonen DP, Willems J, Coumans WA, Chabowski A, van der Vusse GJ, Bonen A, Glatz JF, Luiken JJ. (2004a). Enhanced sarcolemmal FAT/CD36 content and triacylglycerol storage in cardiac myocytes from obese zucker rats. *Diabetes* 53:1655-63.
- Coort SL, Luiken JJ, van der Vusse GJ, Bonen A, Glatz, JF. (2004b). Increased FAT (fatty acid translocase)/CD36-mediated long-chain fatty acid uptake in cardiac myocytes from obese Zucker rats. *Biochem Soc Trans* 32:83-5.
- Corpeleijn E, Petersen L, Holst C, Saris WH, Astrup A, Langin D, MacDonald I, Martinez JA, Oppert JM, Polak J, et al. (2010). Obesityrelated polymorphisms and their associations with the ability to regulate fat oxidation in obese Europeans: The NUGENOB study. *Obesity (Silver Spring)* 18:1369–77.
- Febbraio M, Podrez EA, Smith JD, Hajjar DP., Hazen SL, Hoff HF, Sharma K, Silverstein RL. (2000). Targeted disruption of the class B scavenger receptor CD36 protects against atherosclerotic lesion development in mice. *J Clin Invest* 105:1049–56.
- Fernandez-Real, J. M., Handberg, A., Ortega, F., Hojlund, K., Vendrell J, Ricart W. (2009). Circulating soluble CD36 is a novel marker of liver injury in subjects with altered glucose tolerance. J Nutr Biochem 20:477-84.
- Glatz F, Luiken J, Bonen, A. (2010), Membrane fatty acid transporters as regulators of lipid metabolism: implications for metabolic disease. *Physiol Rev* 90:367–417.
- Glintborg D, Hojlund K, Andersen M, Henriksen JE, Beck-Nielsen H, Handberg A. (2008). Soluble CD36 and risk markers of insulin resistance and atherosclerosis are elevated in polycystic ovary syndrome and significantly reduced during pioglitazone treatment. *Diabetes Care* 31:328–34.
- Goyenechea E, Collins LJ, Parra D, Liu G, Snieder H, Swaminathan R, Spector TD., Martinez JA, O'Dell SD. (2008). CD36 gene promoter polymorphisms are associated with low density lipoproteincholesterol in normal twins and after a low-calorie diet in obese subjects. *Twin Res Hum Genet* 11:621–8.

- Greco D, Kotronen A, Westerbacka J, Puig O, Arkkila P, Kiviluoto T, Laitinen S, Kolak M, Fisher RM, Hamsten A, et al. (2008). Gene expression in human NAFLD. Am J Physiol Gastrointest Liver Physiol 294:G1281-7.
- Griffen E, Re A, Hamel N, Fu C, Bush H, McCaffrey T, Asch AS. (2001). A link between diabetes and atherosclerosis: Glucose regulates expression of CD36 at the level of translation. *Nat Med* 7: 840-6.
- Han XX, Chabowski A, Tandon NN, Calles-Escandon J, Glatz JF, Luiken JJ, Bonen A. (2007). Metabolic challenges reveal impaired fatty acid metabolism and translocation of FAT/CD36 but not FABPpm in obese Zucker rat muscle. *Am J Physiol Endocrinol Metab* 293:E566-75.
- Handberg A, Levin K, Hojlund K, Beck-Nielsen H. (2006). Identification of the oxidized low-density lipoprotein scavenger receptor CD36 in plasma: A novel marker of insulin resistance. *Circulation* 114:1169–76.
- Handberg A, Lopez-Bermejo A, Bassols J, Vendrell J, Ricart W, Fernandez-Real JM. (2009). Circulating soluble CD36 is associated with glucose metabolism and interleukin-6 in glucose-intolerant men. *Diab Vasc Dis Res* 6:15–20.
- Handberg A, Norberg M, Stenlund H, Hallmans G, Attermann J, Eriksson JW. (2010). Soluble CD36 (sCD36) clusters with markers of insulin resistance, and high sCD36 is associated with increased type 2 diabetes risk. *J Clin Endocrinol Metab* 95:1939-46.
- Handberg, A., Skjelland, M., Michelsen, A. E., Sagen, E. L., Krohg-Sorensen, K., Russell, D., Dahl, A., Ueland, T., Oie, E., Aukrust, P, et al. (2008). Soluble CD36 in plasma is increased in patients with symptomatic atherosclerotic carotid plaques and is related to plaque instability. *Stroke* 39:3092–5.
- Hegarty BD, Cooney GJ, Kraegen EW, Furler SM. (2002). Increased efficiency of fatty acid uptake contributes to lipid accumulation in skeletal muscle of high fat-fed insulin-resistant rats. *Diabetes* 51:1477-84.
- Hegarty BD, Furler SM, Ye J, Cooney GJ, Kraegen EW. (2003). The role of intramuscular lipid in insulin resistance. *Acta Physiol Scand* 178:373–83.
- Heni M, Mussig K, Machicao F, Machann J, Schick F, Claussen CD, Stefan N, Fritsche A, Haring HU, Staiger H. (2010). Variants in the CD36 gene locus determine whole-body adiposity, but have no independent effect on insulin sensitivity. *Obesity (Silver Spring)*. doi:10.1038/oby.2010.251 epub ahead of print.
- Holloway GP, Benton CR, Mullen KL, Yoshida Y, Snook LA, Han XX, Glatz JF, Luiken JJ, Lally J, Dyck DJ, et al. (2009). In obese rat muscle transport of palmitate is increased and is channeled to triacylglycerol storage despite an increase in mitochondrial palmitate oxidation. *Am J Physiol Endocrinol Metab* 296:E738-47.
- Holvoet P, de Keyzer D, Jacobs DR. (2008). Oxidized LDL and the metabolic syndrome. *Future Lipidol* 3:637–649.
- Kashyap SR, Ioachimescu AG, Gornik HL, Gopan T, Davidson MB, Makdissi A, Major J, Febbraio M, Silverstein RL. (2009). Lipidinduced insulin resistance is associated with increased monocyte expression of scavenger receptor CD36 and internalization of oxidized LDL. Obesity (Silver Spring) 17:2142–8.
- Kloting N, Fasshauer M, Dietrich A, Kovacs P, Schon MR, Kern M, Stumvoll M, Bluher M. (2010). Insulin-sensitive obesity. Am J Physiol Endocrinol Metab 299:E506-15.
- Koonen DP, Jacobs RL, Febbraio M, Young ME, Soltys CL, Ong H, Vance DE, Dyck JR. (2007). Increased hepatic CD36 expression contributes to dyslipidemia associated with diet-induced obesity. *Diabetes* 56:2863–71.
- Kunjathoor VV, Febbraio M, Podrez EA, Moore KJ, Andersson L, Koehn S, Rhee JS, Silverstein R, Hoff HF, Freeman MW. (2002). Scavenger receptors class A-I/II and CD36 are the principal receptors responsible for the uptake of modified low density lipoprotein leading to lipid loading in macrophages. J Biol Chem 277:49982–8.
- Lepretre F, Linton KJ, Lacquemant C, Vatin V, Samson C, Dina C, Chikri M, Ali S, Scherer P, Seron K, et al. (2004a). Genetic study

of the CD36 gene in a French diabetic population. *Diabetes Metab* 30:459–63.

- Lepretre F, Vasseur F, Vaxillaire M, Scherer PE, Ali S, Linton K, Aitman T, Froguel P. (2004b). A CD36 nonsense mutation associated with insulin resistance and familial type 2 diabetes. *Hum Mutat* 24:104.
- Liang CP, Han S, Okamoto H, Carnemolla R, Tabas I, Accili D, Tall AR. (2004). Increased CD36 protein as a response to defective insulin signaling in macrophages. J Clin Invest 113:764–73.
- Love-Gregory L, Sherva R, Schappe T, Qi JS, McCrea J, Klein S, Connelly MA, Abumrad N. A. (2010). Common CD36 SNPs reduce protein expression and may contribute to a protective atherogenic profile. *Hum Mol Genet* 20:193–201.
- Love-Gregory L, Sherva R, Sun L, Wasson J, Schappe T, Doria A, Rao DC, Hunt SC, Klein S, Neuman RJ, et al. (2008). Variants in the CD36 gene associate with the metabolic syndrome and high-density lipoprotein cholesterol. *Hum Mol Genet* 17:1695-704.
- Luiken JJ, Arumugam Y, Dyck DJ, Bell RC, Pelsers MM, Turcotte LP, Tandon NN, Glatz JF, Bonen A. (2001). Increased rates of fatty acid uptake and plasmalemmal fatty acid transporters in obese Zucker rats. *J Biol Chem* 276:40567-73.
- Ma X, Bacci S, Mlynarski W, Gottardo L, Soccio T, Menzaghi C, Iori E, Lager RA, Shroff AR, Gervino EV, et al. (2004). A common haplotype at the CD36 locus is associated with high free fatty acid levels and increased cardiovascular risk in Caucasians. *Hum Mol Genet* 13:2197–205.
- Malhotra A, Elbein SC, Ng MC, Duggirala R, Arya R, Imperatore G, Adeyemo A, Pollin TI, Hsueh WC, Chan JC, et al. (2007). Meta-analysis of genome-wide linkage studies of quantitative lipid traits in families ascertained for type 2 diabetes. *Diabetes* 56:890-6.
- Ouwens DM, Diamant M, Fodor M, Habets DD, Pelsers MM, El Hasnaoui M, Dang ZC, van den Brom CE, Vlasblom R, Rietdijk A, et al. (2007). Cardiac contractile dysfunction in insulin-resistant rats fed a high-fat diet is associated with elevated CD36-mediated fatty acid uptake and esterification. *Diabetologia* 50:1938–48.
- Podrez EA, Byzova TV, Febbraio M, Salomon RG, Ma Y, Valiyaveettil M, Poliakov E, Sun M, Finton PJ, Curtis BR, et al. (2007). Platelet CD36 links hyperlipidemia, oxidant stress and a prothrombotic phenotype. *Nat Med* 13:1086-95.
- Podrez EA, Febbraio M, Sheibani N, Schmitt D, Silverstein RL, Hajjar DP, Cohen PA, Frazier WA, Hoff HF, Hazen SL. (2000). Macrophage scavenger receptor CD36 is the major receptor for LDL modified

by monocyte-generated reactive nitrogen species. J Clin Invest 105:1095-108.

- Podrez, E. A., Poliakov, E., Shen, Z., Zhang, R., Deng, Y., Sun, M., Finton, P. J., Shan, L., Febbraio M, Hajjar DP, et al. (2002). A novel family of atherogenic oxidized phospholipids promotes macrophage foam cell formation via the scavenger receptor CD36 and is enriched in atherosclerotic lesions. J Biol Chem 277:38517-23.
- Qin B, Polansky MM, Anderson RA. (2010a). Cinnamon extract regulates plasma levels of adipose-derived factors and expression of multiple genes related to carbohydrate metabolism and lipogenesis in adipose tissue of fructose-fed rats. *Horm Metab Res* 42:187–93.
- Qin B, Polansky MM, Harry D, Anderson RA. (2010b). Green tea polyphenols improve cardiac muscle mRNA and protein levels of signal pathways related to insulin and lipid metabolism and inflammation in insulin-resistant rats. *Mol Nutr Food Res* 54 Suppl 1:S14–23.
- Rac ME, Safranow K, Poncyljusz W. (2007). Molecular basis of human CD36 gene mutations. *Mol Med* 13:288–96.
- Rocha VZ, Libby P. (2009). Obesity, inflammation, and atherosclerosis. Nat Rev Cardiol 6:399-409.
- Sampson MJ, Davies IR, Braschi S, Ivory K, Hughes DA. (2003). Increased expression of a scavenger receptor (CD36) in monocytes from subjects with Type 2 diabetes. *Atherosclerosis* 167:129-34.
- Shoelson SE, Herrero L, Naaz A. (2007). Obesity, inflammation, and insulin resistance. *Gastroenterology* 132:2169–80.
- Shoelson SE, Lee J, Goldfine AB. (2006). Inflammation and insulin resistance. J Clin Invest 116:1793–801.
- Silverstein RL. (2009). Inflammation, atherosclerosis, and arterial thrombosis: role of the scavenger receptor CD36. *Cleve Clin J Med* 76 Suppl 2:S27–30.
- Smith AC, Mullen KL, Junkin KA, Nickerson J, Chabowsky A, Bonen A, Dyck DJ. (2007). Metformin and exercise reduce muscle FAT/ CD36 and lipid accumulation and blunt the progression of highfat diet-induced hyperglycemia. *Am J Physiol Endocrinol Metab* 293:E172–81.
- Su X, Abumrad NA. (2009). Cellular fatty acid uptake: A pathway under construction. *Trends Endocrinol Metab* 20:72-7.
- Valiyaveettil M, Podrez EA. (2009). Platelet hyperreactivity, scavenger receptors and atherothrombosis. *J Thromb Haemost 7 Suppl* 1:218–21.