

University of Groningen

## Quantifying the impact of an inference model in Bayesian phylogenetics

Bilderbeek, Richel J. C.; Laudanno, Giovanni; Etienne, Rampal S.

*Published in:*  
Methods in ecology and evolution

*DOI:*  
[10.1111/2041-210X.13514](https://doi.org/10.1111/2041-210X.13514)

**IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.**

*Document Version*  
Publisher's PDF, also known as Version of record

*Publication date:*  
2021

[Link to publication in University of Groningen/UMCG research database](#)

*Citation for published version (APA):*

Bilderbeek, R. J. C., Laudanno, G., & Etienne, R. S. (2021). Quantifying the impact of an inference model in Bayesian phylogenetics. *Methods in ecology and evolution*, 12(2), 351-358. <https://doi.org/10.1111/2041-210X.13514>

### Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: <https://www.rug.nl/library/open-access/self-archiving-pure/taverne-amendment>.

### Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

*Downloaded from the University of Groningen/UMCG research database (Pure): <http://www.rug.nl/research/portal>. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.*

# Quantifying the impact of an inference model in Bayesian phylogenetics

Richèl J. C. Bilderbeek  | Giovanni Laudanno  | Rampal S. Etienne 

Groningen Institute for Evolutionary Life Sciences, University of Groningen, Groningen, The Netherlands

**Correspondence**

Richèl J. C. Bilderbeek  
Email: r.j.c.bilderbeek@rug.nl

**Funding information**

Nederlandse Organisatie voor Wetenschappelijk Onderzoek

Handling Editor: Tiago Quental

**Abstract**

1. Phylogenetic trees are currently routinely reconstructed from an alignment of character sequences (usually nucleotide sequences). Bayesian tools, such as MrBayes, RevBayes and BEAST2, have gained much popularity over the last decade, as they allow joint estimation of the posterior distribution of the phylogenetic trees and the parameters of the underlying inference model. An important ingredient of these Bayesian approaches is the species tree prior. In principle, the Bayesian framework allows for comparing different tree priors, which may elucidate the macroevolutionary processes underlying the species tree. In practice, however, only macroevolutionary models that allow for fast computation of the prior probability are used. The question is how accurate the tree estimation is when the real macroevolutionary processes are substantially different from those assumed in the tree prior.
2. Here we present *pirouette*, a free and open-source R package that assesses the inference error made by Bayesian phylogenetics for a given macroevolutionary diversification model. *pirouette* makes use of BEAST2, but its philosophy applies to any Bayesian phylogenetic inference tool.
3. We describe *pirouette*'s usage providing full examples in which we interrogate a model for its power to describe another.
4. Last, we discuss the results obtained by the examples and their interpretation.

**KEYWORDS**

*babette*, Bayesian model selection, BEAST2, computational biology, evolution, phylogenetics, R, tree prior

## 1 | INTRODUCTION

The development of new powerful Bayesian phylogenetic inference tools, such as BEAST (Drummond & Rambaut, 2007), MrBayes (Huelsenbeck & Ronquist, 2001) or RevBayes (Höhna, Landis, et al., 2016) has been a major advance in constructing phylogenetic trees from character data (usually nucleotide sequences) extracted from organisms (usually extant, but extinction events and/or time-stamped

data can also be added), and hence in our understanding of the main drivers and modes of diversification.

BEAST (Drummond & Rambaut, 2007) is a typical Bayesian phylogenetics tool that needs both character data and priors to infer a posterior distribution of phylogenies. Specifically, for the species tree prior—which describes the process of diversification—BEAST has built-in priors such as the Yule (1925) and (constant-rate) birth–death (BD) (Nee et al., 1994) models as well as coalescent priors. These

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited and is not used for commercial purposes.

© 2020 The Authors. *Methods in Ecology and Evolution* published by John Wiley & Sons Ltd on behalf of British Ecological Society

simple tree priors are among the most commonly used, as they represent some biologically realistic processes (e.g. viewing diversification as a branching process), while being computationally fast.

To allow users to extend the functionalities of BEAST using plug-ins, BEAST2 was written (Bouckaert et al., 2019) (with BEAST and BEAST2 still independently being developed further). For example, one can add novel diversification models by writing a BEAST2 plugin that contains the likelihood formula of a phylogeny under the novel diversification model, that is, the prior probability of a species tree. Plugins have been provided, for instance, for the calibrated Yule model (Heled & Drummond, 2015), the BD model with incomplete sampling (Stadler, 2009), the BD model with serial sampling (Stadler et al., 2012), the BD serial skyline model (Stadler et al., 2013), the fossilized BD process (Gavryushkina et al., 2014) and the BD SIR model (Kühnert et al., 2014).

Many other diversification models (and their associated likelihood algorithms) have been developed, for example, models in which diversification is time-dependent (Nee et al., 1994; Rabosky & Lovette, 2008), or diversity-dependent (Etienne et al., 2012) or where diversification rates change for specific lineages and their descendants (Alfaro et al., 2009; Etienne & Haegeman, 2012; Laudanno et al., 2020; Rabosky, 2014). Other models treat speciation as a process that takes time (Etienne & Rosindell, 2012; Lambert et al., 2015; Rosindell et al., 2010), or where diversification rates depends on one or more traits (FitzJohn, 2012; Herrera-Alsina et al., 2019; Maddison et al., 2007).

These are, however, not yet available as tree priors in BEAST2, for reasons explained below. In this paper, we present methodology to determine whether such new plug-ins are needed, or whether currently available plug-ins are sufficient. We show this using the Yule and BD species tree priors, but our methods can be used with other built-in tree priors as well.

The rationale of our paper is as follows. When a novel diversification model is introduced, its performance in inference should be tested. Part of a model's performance is its ability to recover parameters from simulated data with known parameters (e.g. Etienne et al., 2014), where ideally the estimated parameter values closely match the known/true values. Even when a diversification model passes this test, it is not necessarily used as tree prior in Bayesian inference. Bayesian phylogenetic inference often requires that the prior probability of the phylogeny according to the diversification model has to be computed millions of times. Therefore, biologically interesting but computationally expensive tree priors are often not implemented, and simpler priors are used instead. This is not necessarily problematic, when the data are very informative or when the prior is truly uninformative, as this will reduce the influence of the tree prior. However, the assumption that tree prior choice is of low impact must first be verified.

There have been multiple attempts to investigate the impact of tree prior choice. For example, Sarver et al. (2019) showed that the choice of tree prior does not substantially affect phylogenetic inferences of diversification rates. However, they only compared current diversification models to one another, and thus this does not inform us on the impact of a new tree prior.

Similarly, Ritchie et al. (2016) showed that inference was accurate when birth–death or skyline coalescent priors were used, but

they simulated their trees with a Yule process only, as their focus was not so much on the diversification process but on the influence of inter- and intraspecific sampling.

Another way to benchmark a diversification model, is by doing a model comparison, in which the best model is determined from a set of models. A good early example is Goldman (1993) in which Goldman compared DNA substitution models. A recent approach to test the impact of tree prior choice, proposed by Duchene et al. (2018), allows to measure model adequacy for phylodynamic models that are mathematically described (i.e. have a known likelihood equation).

Here we introduce a method to quantify the impact of a novel tree prior, that is, a tree model, for which we can simulate phylogenies, but not yet calculate their likelihoods. This new method simultaneously assesses the substitution, clock and tree models (Duchêne et al., 2015). The method starts with a phylogeny generated by the new model. Next, nucleotide sequences are simulated that follow the evolutionary history of the given phylogeny. Then, using BEAST2's built-in tree priors, a Bayesian posterior distribution of phylogenies is inferred. We then compare the inferred with the original phylogenies. How to properly perform this comparison forms the heart of our method. Only new diversification models that result in a large discrepancy between inferred and simulated phylogenies will be worth the effort and computational burden to implement as a species tree prior for in a Bayesian framework.

Our method is programmed as an R package (R Core Team, 2013) called *pirouette*. *pirouette* is built on *babette* (Bilderbeek & Etienne, 2018), which calls BEAST2 (Bouckaert et al., 2019).

## 2 | DESCRIPTION

The goal of *pirouette* is to quantify the impact of a new tree prior. It does so by measuring the inference error made for a given reconstructed phylogeny, simulated under a (usually novel) diversification model. We refer to the model that has generated the given tree as the 'generative tree model'  $p_G$ . A 'generative tree model', in this paper, can be either the novel diversification model for which we are testing the impact of choosing standard tree priors for, or it is the model with which we generate the twin tree that is needed for comparison (see below). In the latter case, we also refer to it as the actual generative tree model, and it thus serves as a baseline model. This is done in the example, where the Yule model is the generative model.

The inference error we aim to quantify is not of stochastic nature. Stochastic errors are usually non-directional. We, instead, aim to expose the bias due to the mismatch between a generative model (that has generated the phylogeny) and the model(s) used in the actual inference. We define the birth–death (BD) model (Nee et al., 1994) as the standard tree model, as many (non-standard) tree models have a parameter setting such that it reduces to this model. One such example is the diversity-dependent (DD) diversification model (Etienne & Haegeman, 2020; Etienne et al., 2012) in which speciation or extinction rate depends on the number of species and a clade-level carrying capacity. The BD model can be seen as a special case of the DD model, because for an infinite carrying capacity, the DD model reduces to the

BD model. When benchmarking a novel tree model, one will typically construct phylogenies for different combinations of the diversification model's parameters, to assess under which scenarios the inference error cannot be neglected. While we recommend many replicate simulations when assessing a novel tree prior, our example contains only one replicate, as the goal is to show the workings of *pirouette*, instead of doing an extensive analysis. The Supporting Information includes results of replicated runs under multiple settings.

*pirouette* allows the user to specify a wide variety of custom settings. These settings can be grouped in macro-sections, according to how they operate in the pipeline. We summarize them in Tables 1 and 2.

**TABLE 1** Most important parameter options

Sub-argument	Description	Possible values
tree_prior	Macroevolutionary diversification model	BD, CBS, CCP, CEP, Yule
clock_model	Clock for the DNA mutation rates	RLN, strict
site_model	Nucleotide substitution model	GTR, HKY, JC, TN
mutation_rate	Pace at which substitutions occur	mutation_rate $\in R > 0$
root_sequence	DNA sequence at the root of the tree	any combination of a, c, g, t
model_type	Criterion to select an inference model	Generative, Candidate
run_if	Condition under which an inference model is used	Always, Best candidate
do_measure_evidence	Sets whether or not the evidence of the model must be computed	TRUE, FALSE
error_fun	Specifies how to measure the error	nLTT, $ \gamma $
burn_in_fraction	Specifies the percentage of initial posterior trees to discard	burn_in_fraction $\in [0, 1]$

Abbreviations: BD, birth–death (Nee et al., 1994); CBS, coalescent Bayesian skyline (Drummond et al., 2005); CCP, coalescent constant population; CEP, coalescent exponential population; Yule, pure birth model (Yule, 1925); RLN, relaxed log-normal clock model (Drummond et al., 2006); strict, strict clock model (Zuckermandl & Pauling, 1965); GTR, Generalized time-reversible model (Tavaré, 1986); HKY, Hasegawa, Kishino and Yano (Hasegawa et al., 1985); JC, Jukes and Cantor (Jukes et al., 1969); TN, Tamura and Nei (Tamura & Nei, 1993); nLTT, normalized lineage-through-time (Janzen et al., 2015);  $|\gamma|$ , absolute value of the gamma statistic (Pybus & Harvey, 2000).

**TABLE 2** Definitions of terms and relative symbols used in the main text and in Figure 1. To run the pipeline *A*, *X* and *E* must be specified

Symbol	Macro-argument	Description
<i>G</i>	Generative model	The full setting to produce BEAST2 input data. Its core features are the tree
$s_G$	Site model	prior $p_G$ , the clock model $c_G$ and the site model $s_G$
<i>A</i>	Alignment model	Both the substitution model and rate variation across sites
$X_i$	<i>i</i> -th candidate experiment	Specifies the alignment generation, such as the clock model $c_G$ , site model $s_G$ and root sequence
<i>I</i>	Inference model	Full setting for a Bayesian inference. It is made by a candidate inference model $I_i$ and its inference conditions $C_i$
<i>C</i>	Inference conditions	The assumed phylogenetic inference model, of which the main components are the tree prior $p_i$ , assumed clock model $C_i$ and assumed site model $s_i$ . Conditions under which <i>I</i> is used in the inference. They are composed of the model type, run condition and whether to measure the evidence
<i>E</i>	Error measure parameters	Errors measurement setup that can be specified providing an error function to measure the difference between the original phylogeny and the inferred posterior. The first iterations of the MCMC chain of the posterior may not be representative and can be discarded using a burn-in fraction

## 2.1 | Pirouette's pipeline

The pipeline to assess the error BEAST2 makes in inferring this phylogeny contains the following steps:

1. The user supplies one or (ideally) more phylogenies from a new diversification model.
2. From the given phylogeny an alignment is simulated under a known alignment model *A*.
3. From this alignment, according to the specified inference conditions *C*, an inference model *I* is chosen (which may or may not differ from the model that generated the tree).

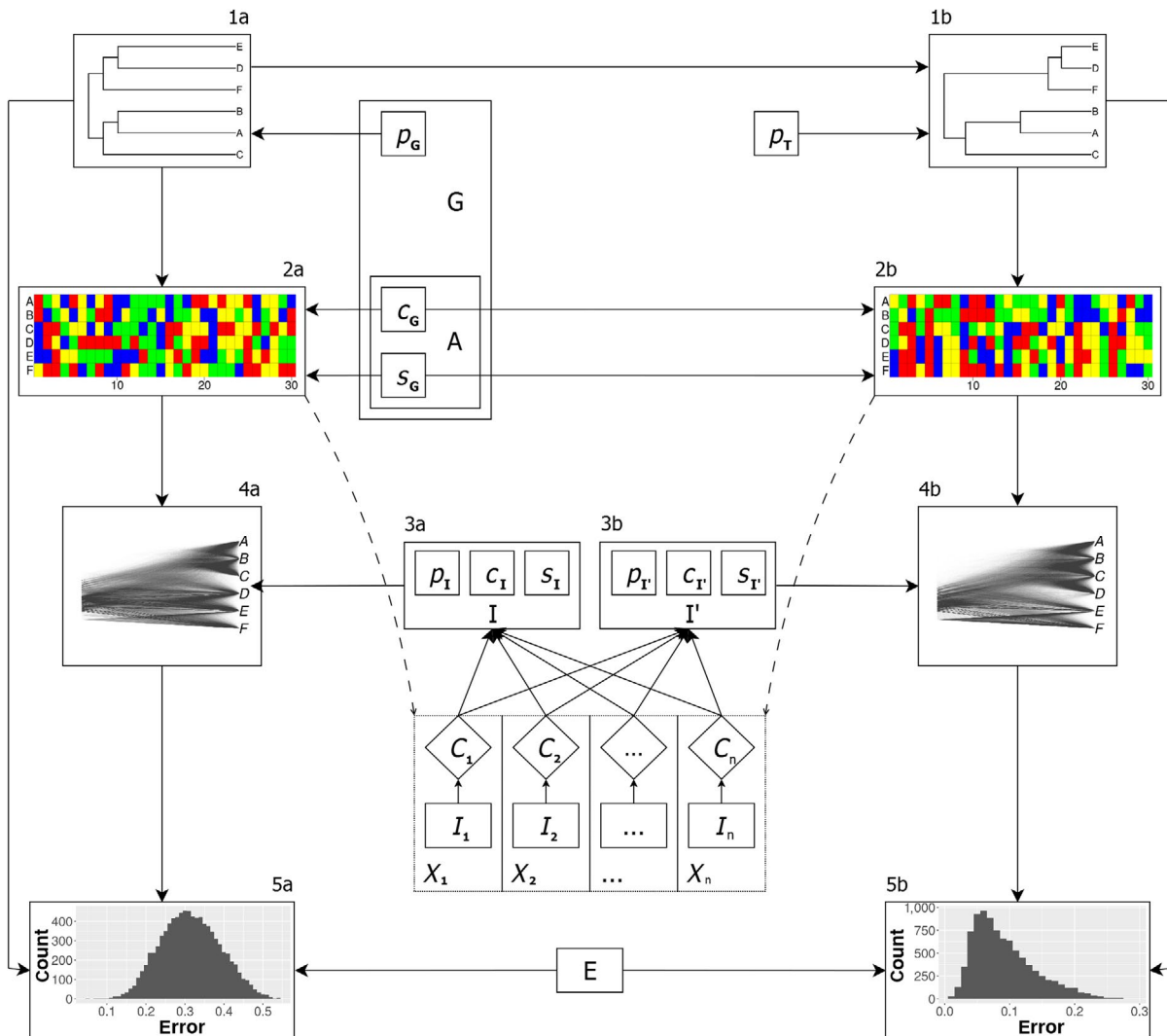
4. The inference model and the alignment are used to infer a posterior distribution of phylogenies.
5. The phylogenies in the posterior are compared with the given phylogeny to estimate the error made, according to the error measure  $E$  specified by the user.

The pipeline is visualized in Figure 1. There is also the option to generate a 'twin tree', that goes through the same pipeline (see supplementary subsection 9.5).

The first step simulates an alignment from the given phylogeny (Figure 1, 1a → 2a). For the sake of clarity, here, we will assume the

alignment consists of DNA sequences, but one can also use other heritable materials such as amino acids. The user must specify a root sequence (i.e. the DNA sequence of the shared common ancestor of all species), a mutation rate and a site model.

The second step (Figure 1, 3a) selects one or more inference model(s)  $I$  from a set of standard inference models  $I_1, \dots, I_n$ . For example, if the generative model is known and standard (which it is for the twin tree, see below), one can specify the inference model to be the same as the generative model. If the tree model is unknown or non-standard—which is the primary motivation for this paper—one can pick a standard inference model which is considered to be



**FIGURE 1** piourette pipeline. The pipeline starts from a phylogeny (1a) simulated by the generative tree model  $p_G$ . The phylogeny is converted to an alignment (2a) using the generative alignment model  $A = (c_G, s_G)$ , composed of a clock model and a site model. The user defines one or more experiments. For each candidate experiment  $X_i$  (a combination of inference model  $I_i$  and condition  $C_i$ ), if its condition  $C_i$  is satisfied (which can depend on the alignment), the corresponding inference model  $I = I_i$  is selected to be used in the next step. The inference models (3a) of the selected experiments use the alignment (2a) to each create a Bayesian posterior of (parameter estimates and) phylogenies (4a). Each of the posterior trees is compared to the true phylogeny (1a) using the error measure  $E$ , resulting in an error distribution (5a). Optionally, for each selected inference model a twin pipeline can be run. A twin phylogeny (1b) can be generated from the original phylogeny (1a) using the twin tree model  $p_T$ , selected among standard diversification models; the default option is the standard BD model, with parameters estimated from the original phylogeny. A twin alignment (2b) is then simulated from the twin phylogeny using clock model  $c_G$  and site model  $s_G$  used with the generative tree model (the novel tree model). The twin pipeline follows the procedure of the main pipeline, resulting in a twin error distribution (5b)

closest to the true tree model. Alternatively, if we want to run only the inference model that fits best to an alignment from a set of candidates (regardless of whether these generated the alignments), one can specify these inference models (see section 9.6).

The third step infers the posterior distributions, using the simulated alignment (Figure 1, 2a → 4a), and the inference models that were selected in the previous step (3a). For each selected experiment, a posterior distribution is inferred, using the *babette* (Bilderbeek & Etienne, 2018) R package which makes use of BEAST2.

The fourth step quantifies the new impact of choosing standard models for inference, that is, the inference error made. First the burn-in fraction is removed, that is, the first phase of the Markov chain Monte Carlo (MCMC) run, which samples an unrepresentative part of parameter and tree space. From the remaining posterior, *pirouette* creates an error distribution, by measuring the difference between the true tree and each of the posterior trees (Figure 1, 4a → 5a). The user can specify a function to quantify the differences between the true and posterior trees.

## 2.2 | Controls

*pirouette* allows for two types of control measurements. The first type of control is called ‘twinning’, which results in an error distribution that is the baseline error of the inference pipeline (see Supporting Information, subsection 9.5 for more details). This is the error that arises when the models used in inference are identical to the ones used in generating the alignments. The second type of control is the use of candidate models, which result in an error distribution for a generative model that is determined to be the best fit to the tree (see Supporting Information, section 9.6 for more details). The underlying idea is that using a substitution model in inference other than the one used in generating the alignment may partly compensate for choosing a standard tree model instead of the generative tree model as tree prior in inference, just because allowing more flexibility anywhere in the inference model, even if at the wrong place, may provide a better fit. This can happen if the effects of the models are similar; for example, allowing variation in diversification rates between branches or allowing variation in the clock rate between branches may result in similar inference of the phylogeny. Additionally, multiple *pirouette* runs are needed to reduce the influence of stochasticity (see Supporting Information, section 9.7 for more details).

## 3 | USAGE

We show the usage of *pirouette* on a tree generated by the non-standard diversity-dependent (DD) tree model (Etienne & Haegeman, 2020; Etienne et al., 2012), which is a BD model with a speciation rate that depends on the number of species.

The code to reproduce our results can be found at [https://github.com/richebilderbeek/pirouette\\_example\\_30](https://github.com/richebilderbeek/pirouette_example_30) and a simplified version is shown here for convenience:

```
library(pirouette)

# Create a DD phylogeny with 5 taxa and a crown age of 10
phylogeny <- create_exemplary_dd_tree()

# Use standard pirouette setup. This creates a list object with all
  settings for generating the alignment, the inference using
  BEAST2, the twinning parameters to generate the twin tree and
  infer it using BEAST2, and the error measure
pir_params <- create_std_pir_params()

# Do the runs
pir_out <- pir_run(
  phylogeny = phylogeny,
  pir_params = pir_params
)

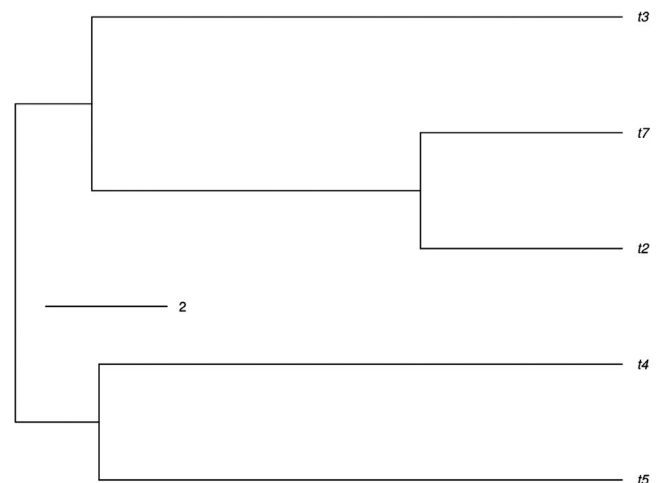
# Plot
pir_plot(pir_out)
```

The DD tree generated by this code is shown in Figure 2.

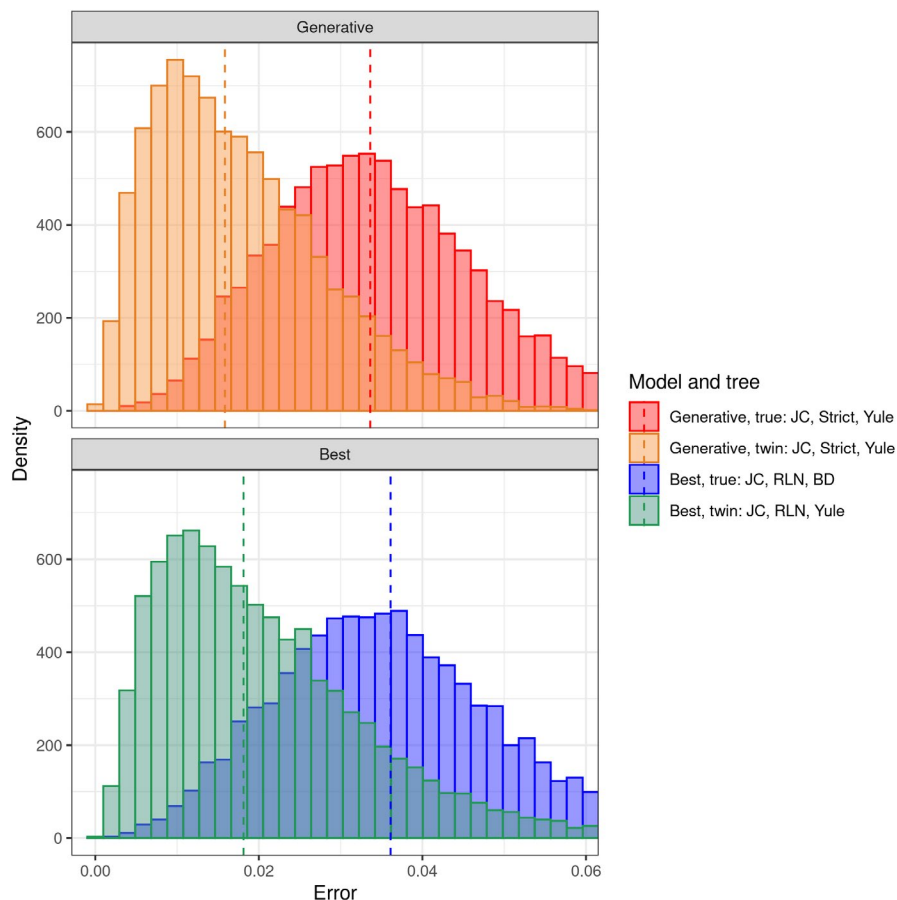
The error distribution shown in Figure 3 is produced, which uses the nLTT statistic (Janzen et al., 2015) to compare phylogenies (see section 9.8 for details regarding the nLTT statistic and its caveats).

In the upper panel of Figure 3, we can see that the error distributions of the (assumed) generative model (i.e. the known generative substitution and clock models, and the tree model that is assumed in inference of the true tree, and the tree model that is used for generating and inferring the twin tree) differ substantially between the true and twin tree. This difference shows the extent of the mismatch between the true tree model (which is DD) and the (Yule) tree prior used in inference. Because these distributions are distinctively different, the inference error made when using an incorrect tree prior on a DD tree is quite profound.

Comparing the upper and lower panel of Figure 3, we can see that the best candidate model is slightly worse at inferring the true



**FIGURE 2** The example tree resulting from a diversity-dependent (DD) simulation



**FIGURE 3** The impact of the tree prior for the example tree in Figure 2. The alignment for this true tree was generated using a JC substitution model and strict clock model. For inferring the tree from this alignment in the 'generative' scenario, the same substitution and clock models were used, and a Yule tree prior (this is the assumed generative model, because the real generative model is assumed to be unknown). For the twin tree, the same inference models were used. In the 'best' scenario, for the true tree, the best-fitting candidate models were JC substitution model, RLN clock model and BD tree prior, while for the twin tree, the best-fitting candidate models were JC substitution model, RLN clock model and Yule tree prior. The twin distributions show the baseline inference error. Vertical dashed lines show the median error value per distribution

tree, than the (assumed) generative model, indicating that the generative inference model we selected is a good choice.

The candidate model that had highest evidence given the simulated alignment, was JC, RLN and BD (see Table 1 for the meaning of these abbreviations). The RLN clock model is a surprising result: it assumes nucleotide substitutions occur at different rates between the taxa. The JC nucleotide substitution model matches the model used to simulate the alignment. The BD model is perhaps somewhat surprising for the true tree because the other alternative standard tree prior, Yule, is probably closest to the true DD model because it shows no pull-of-the-present (but also no slowdown).

## 4 | DISCUSSION

We showed how to use *pirouette* to quantify the impact of a tree prior in Bayesian phylogenetics, assuming—for illustrative purposes—the simplest standard substitution, clock and tree models, but also the models that would be selected among many different standard tree priors according to the highest marginal likelihood, as this would be a likely strategy for an empiricist. We recommend exploring different candidate models, but note that this is computationally highly demanding, particularly for large trees.

Figure 3 illustrates the primary result of our pipeline: it shows the error distributions for the true tree and the twin tree when

either the generative model (for substitution and clock models these are known, for the tree model, it must be assumed for the true tree and it is known for the twin tree) or the best-fitting set candidate model (i.e. combination of tree model, substitution model and clock model) is used in inference. The clear difference between the error distributions for the true tree and the twin tree suggests that the choice of tree prior matters. We note, however, that only one tree from a novel tree model is not enough to determine the impact of using an incorrect tree prior. Instead, a distribution of multiple trees, generated by the novel tree model, should be used. In the Supporting Information, we have provided some examples.

Like most phylogenetic experiments, the setup of *pirouette* involves many choices. A prime example is the length of the simulated DNA sequence. One expects that the inference error decreases for longer DNA sequences. We investigated this superficially and confirmed this prediction (see the Supporting Information). However, we note that for longer DNA sequences, the assumption of the same substitution rates across the entire sequence may become less realistic (different genes may experience different substitution rates) and hence longer sequences may require more parameters. Hence, simply getting longer sequences will not always lead to a drastic reduction of the influence of the species tree prior. Fortunately, *pirouette* provides a pipeline that works for all choices.

Interpreting the results of *pirouette* is up to the user; *pirouette* does not answer the question whether the inference error is too

large to trust the inferred tree. The user is encouraged to use different statistics to measure the error.

The nLTT statistic is a promising starting point, as it can compare any two trees and results in an error distribution of known range, but one may also explore other statistics, for example, statistics that depend on the topology of the tree. While *pirouette* allows for this in principle, in our example we used a diversification model (DD) that only deviates from the Yule and BD models in the temporal branching pattern, not in the topology. For models that make different predictions on topology, the twinning process should be modified.

As noted in the introduction, Duchene et al. (2018) also developed a method to assess the adequacy of a tree model on empirical trees. They simulated trees from the posterior distribution of the parameters and then compared this to the originally inferred tree using tree statistics, to determine whether the assumed tree model in inference indeed generates the tree as inferred. This is useful if these trees match, but when they do not, this does not mean that the inferred tree is incorrect; if sufficient data are available the species tree prior may not be important, and hence inference may be adequate even though the assumed species tree prior is not. In short, the approach is applied to empirical trees and compares the posterior and prior distribution of trees (with the latter generated with the posterior parameters!). By contrast, *pirouette* aims to expose when assuming standard priors for the species tree are a mis- or underparameterization. Hence, our approach applies to simulated trees and compares the posterior distributions of trees generated with a standard and non-standard model, but inferred with a standard one. The two methods therefore complement one another.

Furthermore, we note that the *pirouette* pipeline is not restricted to exploring the effects of a new species tree model. The pipeline can also be used to explore the effects of non-standard clock or site models, such as relaxed clock models with a non-standard distribution, correlated substitutions on sister lineages or elevated substitution rates during speciation events. It is, however, beyond the scope of this paper to discuss all these options in more detail.

In conclusion, *pirouette* can show the errors in phylogenetic reconstruction expected when the model assumed in inference is different from the actual generative model. The user can then judge whether or not this new model should be implemented in a Bayesian phylogenetic tool.

## ACKNOWLEDGEMENTS

We thank the Center for Information Technology of the University of Groningen for its support and for providing access to the Peregrine high performance computing cluster. We thank the Netherlands Organization for Scientific Research (NWO) for financial support through a VICI grant awarded to R.S.E.

## AUTHORS' CONTRIBUTIONS

R.J.C.B., G.L. and R.S.E. conceived the idea for the package; R.J.C.B. created, tested and revised the package; G.L. provided major contributions to the package; R.J.C.B. wrote the first draft of the manuscript; G.L. and R.S.E. contributed to revisions.

## PEER REVIEW

The peer review history for this article is available at <https://publons.com/publon/10.1111/2041-210X.13514>.

## DATA AVAILABILITY STATEMENT

All code for this manuscript is archived at [http://github.com/richelbilderbeek/pirouette\\_article](http://github.com/richelbilderbeek/pirouette_article), with <https://zenodo.org/record/3969845> (Bilderbeek, 2020a). The *pirouette* code used for the examples is archived at <https://doi.org/10.5281/zenodo.3969839> (Bilderbeek, 2020b). The *pirouette* examples (including intermediate data) are archived at <https://doi.org/10.5281/zenodo.3970000> (Bilderbeek, 2020c).

## ORCID

Richèl J. C. Bilderbeek  <https://orcid.org/0000-0003-1107-7049>

Giovanni Laudanno  <https://orcid.org/0000-0002-2952-3345>

Rampal S. Etienne  <https://orcid.org/0000-0003-2142-7612>

## REFERENCES

- Alfaro, M. E., Santini, F., Brock, C., Alamillo, H., Dornburg, A., Rabosky, D. L., Carnevale, G., & Harmon, L. J. (2009). Nine exceptional radiations plus high turnover explain species diversity in jawed vertebrates. *Proceedings of the National Academy of Sciences of the United States of America*, *106*, 13410–13414. <https://doi.org/10.1073/pnas.0811087106>
- Bilderbeek, R. J. (2020a). *pirouette\_article* v1.3. *Zenodo*, <https://doi.org/10.5281/zenodo.3969845>
- Bilderbeek, R. J. (2020b). *pirouette\_code* v1.6.4. *Zenodo*, <https://doi.org/10.5281/zenodo.3969839>
- Bilderbeek, R. J. (2020c). *pirouette\_examples*. *Zenodo*, <https://doi.org/10.5281/zenodo.3970000>
- Bilderbeek, R. J., & Etienne, R. S. (2018). *babette*: BEAUti 2, BEAST 2 and tracer for R. *Methods in Ecology and Evolution*, *9*(9), 2034–2040.
- Bouckaert, R., Vaughan, T. G., Barido-Sottani, J., Duchêne, S., Fourment, M., Gavryushkina, A., Heled, J., Jones, G., Kühnert, D., De Maio, N., & Matschiner, M. (2019). BEAST 2.5: An advanced software platform for Bayesian evolutionary analysis. *PLoS Computational Biology*, *15*, e1006650.
- Drummond, A. J., Ho, S. Y., Phillips, M. J., & Rambaut, A. (2006). Relaxed phylogenetics and dating with confidence. *PLoS Biology*, *4*, e88. <https://doi.org/10.1371/journal.pbio.0040088>
- Drummond, A. J., & Rambaut, A. (2007). BEAST: Bayesian evolutionary analysis by sampling trees. *BMC Evolutionary Biology*, *7*, 214. <https://doi.org/10.1186/1471-2148-7-214>
- Drummond, A. J., Rambaut, A., Shapiro, B., & Pybus, O. G. (2005). Bayesian coalescent inference of past population dynamics from molecular sequences. *Molecular Biology and Evolution*, *22*, 1185–1192. <https://doi.org/10.1093/molbev/msi103>
- Duchêne, D. A., Duchêne, S., Holmes, E. C., & Ho, S. Y. (2015). Evaluating the adequacy of molecular clock models using posterior predictive simulations. *Molecular Biology and Evolution*, *32*, 2986–2995. <https://doi.org/10.1093/molbev/msv154>
- Duchene, S., Bouckaert, R., Duchene, D. A., Stadler, T., & Drummond, A. J. (2018). Phylodynamic model adequacy using posterior predictive simulations. *Systematic Biology*, *68*, 358–364. <https://doi.org/10.1093/sysbio/syy048>
- Etienne, R. S., & Haegeman, B. (2012). A conceptual and statistical framework for adaptive radiations with a key role for diversity dependence. *The American Naturalist*, *180*, E75–E89. <https://doi.org/10.1086/667574>
- Etienne, R. S., & Haegeman, B. (2020). *DDD*. Retrieved from <https://CRAN.R-project.org/package=DDD>



- Etienne, R. S., Haegeman, B., Stadler, T., Aze, T., Pearson, P. N., Purvis, A., & Phillimore, A. B. (2012). Diversity-dependence brings molecular phylogenies closer to agreement with the fossil record. *Proceedings of the Royal Society B: Biological Sciences*, 279, 1300–1309. <https://doi.org/10.1098/rspb.2011.1439>
- Etienne, R. S., Morlon, H., & Lambert, A. (2014). Estimating the duration of speciation from phylogenies. *Evolution*, 68, 2430–2440. <https://doi.org/10.1111/evo.12433>
- Etienne, R. S., & Rosindell, J. (2012). Prolonging the past counteracts the pull of the present: Protracted speciation can explain observed slowdowns in diversification. *Systematic Biology*, 61, 204–213. <https://doi.org/10.1093/sysbio/syr091>
- FitzJohn, R. G. (2012). Diversitree: Comparative phylogenetic analyses of diversification in R. *Methods in Ecology and Evolution*, 3, 1084–1092.
- Gavryushkina, A., Welch, D., Stadler, T., & Drummond, A. J. (2014). Bayesian inference of sampled ancestor trees for epidemiology and fossil calibration. *PLoS Computational Biology*, 10, e1003919. <https://doi.org/10.1371/journal.pcbi.1003919>
- Goldman, N. (1993). Statistical tests of models of DNA substitution. *Journal of Molecular Evolution*, 36, 182–198. <https://doi.org/10.1007/BF00166252>
- Hasegawa, M., Kishino, H., & Yano, T. (1985). Dating of the human-ape splitting by a molecular clock of mitochondrial DNA. *Journal of Molecular Evolution*, 22, 160–174. <https://doi.org/10.1007/BF02101694>
- Heled, J., & Drummond, A. J. (2015). Calibrated birth–death phylogenetic time-tree priors for Bayesian inference. *Systematic Biology*, 64, 369–383. <https://doi.org/10.1093/sysbio/syu089>
- Herrera-Alsina, L., van Els, P., & Etienne, R. S. (2019). Detecting the dependence of diversification on multiple traits from phylogenetic trees and trait data. *Systematic Biology*, 68, 317–328. <https://doi.org/10.1093/sysbio/syy057>
- Höhna, S., Landis, M. J., Heath, T. A., Boussau, B., Lartillot, N., Moore, B. R., Huelsenbeck, J. P., & Ronquist, F. (2016). RevBayes: Bayesian phylogenetic inference using graphical models and an interactive model-specification language. *Systematic Biology*, 65, 726–736. <https://doi.org/10.1093/sysbio/syw021>
- Huelsenbeck, J. P., & Ronquist, F. (2001). MrBayes: Bayesian inference of phylogenetic trees. *Bioinformatics*, 17, 754–755. <https://doi.org/10.1093/bioinformatics/17.8.754>
- Janzen, T., Höhna, S., & Etienne, R. S. (2015). Approximate Bayesian computation of diversification rates from molecular phylogenies: Introducing a new efficient summary statistic, the nLTT. *Methods in Ecology and Evolution*, 6, 566–575.
- Jukes, T. H., Cantor, C. R. (1969). Evolution of protein molecules. *Mammalian Protein Metabolism*, 3, 132.
- Kühnert, D., Stadler, T., Vaughan, T. G., & Drummond, A. J. (2014). Simultaneous reconstruction of evolutionary history and epidemiological dynamics from viral sequences with the birth–death sir model. *Journal of the Royal Society Interface*, 11, 20131106. <https://doi.org/10.1098/rsif.2013.1106>
- Lambert, A., Morlon, H., & Etienne, R. S. (2015). The reconstructed tree in the lineage-based model of protracted speciation. *Journal of Mathematical Biology*, 70, 367–397. <https://doi.org/10.1007/s00285-014-0767-x>
- Laudanno, G., Haegeman, B., Rabosky, D. L., & Etienne, R. S. (2020). Detecting lineage-specific shifts in diversification: A proper likelihood approach. *Systematic Biology*. <https://doi.org/10.1093/sysbio/syaa048>
- Maddison, W. P., Midford, P. E., & Otto, S. P. (2007). Estimating a binary character's effect on speciation and extinction. *Systematic Biology*, 56, 701–710. <https://doi.org/10.1080/10635150701607033>
- Nee, S., May, R. M., & Harvey, P. H. (1994). The reconstructed evolutionary process. *Philosophical Transactions of the Royal Society of London B*, 344, 305–311.
- Pybus, O. G., & Harvey, P. H. (2000). Testing macro-evolutionary models using incomplete molecular phylogenies. *Proceedings of the Royal Society of London. Series B: Biological Sciences*, 267(1459), 2267–2272. <https://doi.org/10.1098/rspb.2000.1278>
- R Core Team. (2013). *R: A language and environment for statistical computing*. R Foundation for Statistical Computing.
- Rabosky, D. L. (2014). Automatic detection of key innovations, rate shifts, and diversity-dependence on phylogenetic trees. *PLoS ONE*, 9, e89543. <https://doi.org/10.1371/journal.pone.0089543>
- Rabosky, D. L., & Lovette, I. J. (2008). Explosive evolutionary radiations: Decreasing speciation or increasing extinction through time? *Evolution*, 62, 1866–1875. <https://doi.org/10.1111/j.1558-5646.2008.00409.x>
- Ritchie, A. M., Lo, N., & Ho, S. Y. W. (2016). The impact of the tree prior on molecular dating of data sets containing a mixture of inter- and intraspecies sampling. *Systematic Biology*, 66, 413–425. <https://doi.org/10.1093/sysbio/syw095>
- Rosindell, J., Cornell, S. J., Hubbell, S. P., & Etienne, R. S. (2010). Protracted speciation revitalizes the neutral theory of biodiversity. *Ecology Letters*, 13, 716–727. <https://doi.org/10.1111/j.1461-0248.2010.01463.x>
- Sarver, B. A., Pennell, M. W., Brown, J. W., Keeble, S., Hardwick, K. M., Sullivan, J., & Harmon, L. J. (2019). The choice of tree prior and molecular clock does not substantially affect phylogenetic inferences of diversification rates. *PeerJ*, 7, e6334. <https://doi.org/10.7717/peerj.6334>
- Stadler, T. (2009). On incomplete sampling under birth–death models and connections to the sampling-based coalescent. *Journal of Theoretical Biology*, 261, 58–66. <https://doi.org/10.1016/j.jtbi.2009.07.018>
- Stadler, T., Kouyos, R., von Wyl, V., Yerly, S., Böni, J., Bürgisser, P., Klimkait, T., Joos, B., Rieder, P., Xie, D., Günthard, H. F., Drummond, A. J., & Bonhoeffer, S. (2012). Estimating the basic reproductive number from viral sequence data. *Molecular Biology and Evolution*, 29, 347–357. <https://doi.org/10.1093/molbev/msr217>
- Stadler, T., Kühnert, D., Bonhoeffer, S., & Drummond, A. J. (2013). Birth–death skyline plot reveals temporal changes of epidemic spread in HIV and hepatitis C virus (HCV). *Proceedings of the National Academy of Sciences of United States of America*, 110, 228–233. <https://doi.org/10.1073/pnas.1207965110>
- Tamura, K., & Nei, M. (1993). Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees. *Molecular Biology and Evolution*, 10, 512–526.
- Tavaré, S. (1986). Some probabilistic and statistical problems in the analysis of DNA sequences. *Lectures on Mathematics in the Life Sciences*, 17, 57–86.
- Yule, G. U. (1925). A mathematical theory of evolution, based on the conclusions of Dr. JC Willis, FRS. *Philosophical Transactions of the Royal Society of London Series B, Containing Papers of a Biological Character*, 213, 21–87.
- Zuckerandl, E., & Pauling, L. (1965). Molecules as documents of evolutionary history. *Journal of Theoretical Biology*, 8, 357–366. [https://doi.org/10.1016/0022-5193\(65\)90083-4](https://doi.org/10.1016/0022-5193(65)90083-4)

## SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

**How to cite this article:** Bilderbeek RJC, Laudanno G, Etienne RS. Quantifying the impact of an inference model in Bayesian phylogenetics. *Methods Ecol Evol*. 2021;12:351–358. <https://doi.org/10.1111/2041-210X.13514>