



University of Groningen

Body surface temperature responses to food restriction in wild and captive great tits

Winders, L. A.; White, S. A.; Helm, Barbara; McCafferty, D. J.

Published in: Journal of Experimental Biology

DOI: 10.1242/jeb.220046

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version Early version, also known as pre-print

Publication date: 2020

Link to publication in University of Groningen/UMCG research database

Citation for published version (APA): Winders, L. A., White, S. A., Helm, B., & McCafferty, D. J. (2020). Body surface temperature responses to food restriction in wild and captive great tits. *Journal of Experimental Biology, 223*(8), [jeb220046]. https://doi.org/10.1242/jeb.220046

Copyright Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: https://www.rug.nl/library/open-access/self-archiving-pure/taverneamendment.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): http://www.rug.nl/research/portal. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

1	Title:
2	Body surface temperature responses to food restriction in wild and captive great tits (Parus major)
3	
4	Running title:
5	Body surface temperature in fasting great tits
6	
7	Winder, L.A. ^{1,2} , White, S.A. ¹ , Nord, A. ^{1,3} , Helm, B ^{1,4} . & McCafferty, D.J. ¹
8	
9	¹ Scottish Centre for Ecology and the Natural Environment, Institute of Biodiversity, Animal Health
10	& Comparative Medicine, College of Medical, Veterinary and Life Sciences, University of Glasgow,
11	Rowardennan, G63 0AW, Scotland, UK
12	
13	² Department of Animal and Plant Sciences, University of Sheffield, Sheffield S10 2TN, UK
14	
15	³ Department of Biology, Section for Evolutionary Ecology, Lund University, SE-223 62 Lund,
16	Sweden
17	
18	⁴ Groningen Institute for Evolutionary Life Sciences (GELIFES), University of Groningen,
19	Groningen, The Netherlands
20	
21	Email addresses:
22	
23	Lucy Winder: lwinder1@sheffield.ac.uk (ORCID iD: 0000-0002-8100-0568)
24	Stewart White: <u>Stewart.White@glasgow.ac.uk</u>
25	Andreas Nord: andreas.nord@biol.lu.se (ORCID iD: 0000-0001-6170-689X)
26	Barbara Helm: <u>Barbara.Helm@glasgow.ac.uk</u> (ORCID iD: <u>0000-0002-6648-1463</u>)
27	Dominic McCafferty: <u>Dominic.McCafferty@glasgow.ac.uk</u> (ORCID iD: <u>0000-0002-3079-3326</u>)
28	
29	Key words (3-6):
30	body temperature, food restriction, heterothermy; thermal imaging, winter
31	
32	Summary statement
33	We provide evidence that wild and captive great tits reduce temperature of the bill in response to
34	food restriction.

- 35 Abstract
- 36

37 During winter at temperate and high latitudes, low ambient temperatures, limited food supplies and 38 short foraging periods mean small passerines show behavioural, morphological and physiological 39 adaptations to reduce the risk of facing energy shortages. Peripheral tissues vasoconstrict in low 40 ambient temperatures to reduce heat loss and cold injury. Peripheral vasoconstriction has been 41 observed with food restriction in captivity but has yet to be explored in free-ranging animals. We 42 experimentally food restricted both wild and captive great tits during winter months and measured 43 surface temperatures of bill and eye-region using thermal imaging, to investigate if birds show rapid 44 local heterothermic responses, which may reduce thermoregulatory costs when facing a perceived 45 imminent food shortage. Our results of a continuously-filmed wild population showed that bill 46 temperature was immediately reduced in response to food restriction compared to when food was ad 47 *libitum*, an apparent autonomic response. Such immediacy implies a 'pre-emptive' response before 48 the bird experiences any shortfalls in energy reserves. We also demonstrate temporal variation in 49 vasoconstriction of the bill, with bill temperature gradually rising throughout the food restriction 50 after the initial drop. Eye-region temperature in the wild birds remained at similar levels throughout 51 the food restriction compared to unrestricted birds, possibly reflecting the need to maintain steady 52 circulation to the central nervous and visual systems. Our findings provide evidence that birds 53 selectively allow the bill to cool when a predictable food supply is suddenly disrupted, likely as a

54 means of minimising depletion of body reserves for a perceived future shortage in energy.

55 Introduction

56

Winter in seasonal habitats is often challenging for small endotherms as severe weather increases
thermoregulatory costs while limited food supply and short foraging periods potentially constrain
acquisition of resources to meet these increased costs. It follows that individuals must respond to

60 winter conditions, by morphological, behavioural and physiological adaptations, to avoid facing

61 energetic shortfalls. The thermoneutral zone (TNZ), where heat loss is offset by basal metabolic heat

62 production, for most passerines is 15-35 °C (Gavrilov and Dolnik, 1985). In winter at higher latitudes

63 small birds routinely experience environmental temperatures well below thermoneutrality and

64 therefore to maintain body temperature, metabolic heat production must increase (Scholander et al.,

65 1950; William et al., 1983). A first defence to minimise heat loss are morphological adaptations

66 (e.g., increased insulation from feathers) and behavioural responses (e.g., seeking shelter,

67 ptiloerection) (Nord et al., 2011; Shipley et al., 2019). Physiological adaptations in small endotherms

are directed to increasing heat production (Swanson and Vézina, 2015) and insulation *via* local or

69 global heterothermy (e.g. Johnsen et al., 1985; Ruf & Geiser 2015). These responses operate at

70 different temporal scales as seen by long term seasonal acclimatisation (Vezina & Swanson 2015) or

through instantaneous responses when there are sudden changes in weather (Marsh and Dawson,

72 1989).

73

74 Reduction in peripheral temperature by shunting blood flow to the core (local heterothermy) can lead 75 to significant energy savings in variable environments (Hagan and Heath, 1980; Steen and Steen, 76 1965; Tattersall et al., 2016). In birds, the legs, bill and eyes are usually unfeathered and are, 77 therefore, key regions of heat transfer. Counter-current vascular arrangements, and sphincteric 78 contractions in major vessels in and around birds' legs, allow the normally uninsulated region to 79 remain at, or close to, ambient temperature (Johansen and Bech, 1983; Midtgård, 1981; Steen and 80 Steen, 1965). This reduces heat loss and prevents cold injury. The bill is highly vascularised but 81 uninsulated, and is known to play a role in thermoregulation particularly in large-billed species in hot 82 climates, though recent work highlights the role of the bill also in cold environments and in small-83 billed species (Schraft et al., 2019; reviewed by Tattersall et al., 2017). In line with this, bill size 84 declines with decreasing minimum winter temperature (Danner and Greenberg, 2015; Friedman et 85 al., 2017; Symonds and Tattersall, 2010). It is, therefore, a realistic expectation that there will be 86 thermoregulatory responses in the bill (as well as in other peripheral tissues) to manage energetically 87 challenging situations, such as cold snaps and food shortage. Additionally, reduced circulation to the 88 head region might lower evaporative heat loss through uninsulated regions such as the eyes and

89 respiratory heat loss through the nasal passages (Midtgård, 1984). However, while local

- 90 heterothermic responses carry energetic benefits, the resultant lower tissue temperature in
- 91 appendages such as the legs and bill, and other peripherally located structures such as the eyes, may
- 92 reduce ease of locomotion, foraging or sensory perception. Therefore, the use of local heterothermy
- may be subject to a trade-off between environmental conditions, energetic state and food availability.
- 94 For example, a study of Muscovy ducklings (*Cairina moschata*) showed cold-acclimated birds had a
- 95 more stable bill temperature, with evidence of vasoconstriction of the bill, when fasting for relatively
- 96 long periods, than birds that were kept in thermoneutrality (Tattersall et al., 2016). A recent study on
- 97 blue tits (*Cyanistes caeruleus*) found that low periorbital temperature was correlated with low body
- 98 condition (Jerem et al., 2018). Local heterothermy has also been shown to be a response to fasting in
- 99 several other bird species, and likely explains why in some studies core body temperature remains
- 100 constant but, nevertheless, energy savings are made (Hohtola, 2012). There is now a need to
- 101 experimentally test predictions from this work on wild models in their natural environment.
- 102

103 In this study, we experimentally tested the effects of environmental conditions on peripheral body

- 104 temperature of wild and captive great tits (*Parus major*) in winter, using thermal imaging. In both
- settings, we temporarily manipulated access to food and recorded the dynamics of the birds' eye and
- 106 bill temperatures before, during, and after food restriction. We predicted that peripheral body
- 107 temperatures would decrease in response to the food restriction, and more so when ambient
- 108 temperature was lower. We expected to reliably record body surface temperature in uninsulated areas
- 109 of the body, specifically the bill and eye-region, which are likely key areas of heat-exchange. We did
- 110 not record responses to food restriction in the uninsulated legs, because previous work in our
- 111 population has shown that wild parids (including great tits) maintain stable low leg temperatures in
- 112 winter, even when fed *ad libitum*. By contrast, bill temperature is consistently maintained well above
- ambient (Nord, A., Huxtable, A., Reilly, H., McCafferty, D. J., in prep.).
- 114
- 115

116 Material and methods

117

118 The study used great tits in two populations of separate subspecies; one captive (ssp. *newtoni*) and

- 119 one wild (ssp. *major*). In both populations we compared food-restricted birds to unrestricted control
- 120 birds. The wild study consisted of continuous filming on days with and without a food restriction
- 121 experiment (treatment or control days). For the captive study, filming occurred before and after a
- 122 food restriction event and two consecutive days before the food restriction day. The air temperature

range was between -10 and +2°C in the captive study, and +2 to +13°C in the wild study, below the
thermoneutral zone of great tits (Broggi et al., 2005).

125

126 *Captive study*

127

128 Fourteen wild great tits were captured near Vomb, Sweden (55°39'N, 13°33'E) and were 129 immediately transferred to four outdoor aviaries $(6.0 \times 3.0 \times 2.5 \text{ m}; \text{width} \times \text{length} \times \text{height})$ at 130 Stensoffa Ecological Field Station, Sweden (55°42'N, 13°27'E), where they were kept in mixed sex 131 groups from October 2012 to January 2013 and handled as described in Nord et al., (2016). The 132 aviaries contained both a covered and non-covered area, perches and nest boxes for the number of 133 individuals in each aviary. The birds were left for two weeks to acclimate to the aviaries before the 134 start of the experiment. All procedures on the captive birds were approved by the Malmö/Lund 135 Animal Ethics Committee (permit no. M236-10). Catching and ringing of birds was licensed by the 136 Swedish Ringing Centre (license no. 475), and the use of radio transmitters was permitted by the

137 Swedish Post and Telecom Authority (permit no. 12-9096).

138

139

140 Thermal videos were taken at 3 Hz of birds at the feeders at 1.4 m distance using a SC640 FLIR 141 camera (FLIR® Systems, Inc), FOL 76mm lens on three consecutive days (1-3 December). On days 142 1 and 2, food remained *ad libitum* throughout the day (including while filming). On day 3, the food 143 was restricted for three hours (mean: $3hr17min \pm 8min$) staggered by an hour between aviaries, with 144 the first restriction beginning in the first aviary at 9:00 h (local time) and beginning in the last aviary 145 at 13:00 h. Water was freely available in heated trays (that prevented freezing) throughout the 146 experiment. Thermal imaging took place before the food restriction (data also include the two days 147 prior to the food restriction) and after the food restriction period and lasted for one hour (mean: 148 54mins \pm 14mins) at each aviary (for day 2, aviary 4, filming lasted for 4hrs 29mins). A video 149 camera (Panasonic Model: HC-V720, Hamburg, Germany) was used to film the feeder so individual 150 birds could be identified from unique colour ring combinations (birds were also fitted with 151 subcutaneous PIT tags and radio transmitters for other research projects, see Nord et al., 2016).

152

153 Air temperature (accuracy $\pm 0.5^{\circ}$ C, resolution 0.0625°C) was recorded continuously from the centre

154 of the aviary (iButton DS1922-L, Maxim Integrated Products, CA, USA; accuracy $\pm 0.5^{\circ}$ C). Relative

155 humidity was recorded by a weather station at Lund University, 17 km from the study site.

157 Wild study

158 Data for the wild study was collected in an oak (Quercus robur) woodland surrounding the Scottish

159 Centre for Ecology and the Natural Environment on Loch Lomond, Scotland UK (56°3'N, 04°33'W)

160 between January and March 2017. A bird feeder containing peanut granules (Haith's, Grimsby, UK)

161 was provided two months prior to the start of the experiment to attract resident birds.

162

163 Nineteen great tits were then caught by mist netting around the feeder from January to February

164 2017, and were fitted with a British Trust for Ornithology (BTO) ring on the right leg and a passive

165 integrated transponder (PIT) tag (EM4102 PIT Tag, Eccel Technology, Leicester, UK), used for

166 identification, on the left leg. A custom-built PIT tag recorder (University of Glasgow Bioelectronics

167 Unit, Glasgow) was attached to the feeder in order to identify birds visiting at a given time. All

168 procedures were approved by BTO ringing permits, and by a UK Home Office Licence.

169

170

171 Thermal video was collected from food-restricted and control birds at 7.5 Hz using a FLIR AX5 172 thermal camera from 0.7 m distance, on nine days between 10 February and 2 March 2017. Food 173 was restricted on five of those days (14, 16, 21, 23 February and 2 March 2017) for three hours 174 (mean: $2hrs 43mins \pm 6mins$) between 10:00 and 13:20. On these days, thermal videos were taken 175 for one hour before the food restriction, three hours during the food restriction and an hour and a half 176 after the food restriction (with the exception of 16 February, when due to equipment failure filming 177 occurred only after food restriction). Each food restriction was considered as a stand-alone event as 178 at least one control day separated each day of food restriction. For the remaining four control days 179 (10, 13, 15 and 20 February 2017), where there was no food restriction, filming occurred 180 continuously at the feeder. A dummy camera was deployed five days prior to filming to habituate 181 birds to the presence of the camera and was subsequently returned each day after thermal imaging 182 was completed. Air temperature was measured using a thermocouple attached to the feeder (Tinytag 183 Talk 2, Gemini Data Loggers, Chichester, England). Relative humidity data were available from a 184 MiniMet Automatic Weather Station (Skye Instruments, Powys, UK), within 200 m of the thermal 185 camera. 186 187 Thermal image analysis

188

189

190 Individual thermal images (sample sizes shown in Table 1) were extracted and analysed from the 191 thermal videos using FLIR Tools 4.1. Images were selected where a clear lateral view of the head

- was shown. When a bird visited the feeder, a unique PIT tag code was recorded with the time of visit. The time could be compared to the thermal imaging video to identify individuals in the wild study. We only analysed one image per bird within a 10 min period so each image could be considered as an independent visit to the feeder. As many birds in the wild study could not be identified when visiting the feeder, we used 41 images from unknown birds. To prevent repeated measurements of the same bird, we only used images of unknown individuals that were ≥ 10 min in time apart. For the wild experiment, the entire video was used. For the captive study, we randomly selected an aviary to be filmed for an hour at the feeder from 8:00-12:00 (before food restriction) and 12:30-15:30 (after food restriction), so that despite a single camera, all aviaries were filmed on each day.

- Table 1. Sample sizes in the experiment. The number of individual birds and images used in the
- experiment. Unidentified individuals were used on control days as equipment failure limited our sample size
- (see thermal imaging analysis in methods).

		Individuals	Images
Wild	Food restricted days	19 (6 female, 8 male, 5 unknown sex)	126
	Control days	46 (41 unknown IDs, of known: 3 female, 2 male)	55
		known: 5 female, 2 male)	55
Captive	Before food restriction	15 (4 female, 11 male)	99
-	After food restriction	17 (5 female, 12 male)	52
atmospheric	and reflected temperatu	et as 0.98 (Best and Fowler, 19 res during image analysis were	set as the hourl
		her station during recording. Re	lative humidity
mean for eac	ch recording session.		
Fig. 1. Data e	extraction from thermal in	mage of bird at feeder. Lateral in	mage of a great ti
temperature v	vas extracted by drawing a	a line from the base of the nostril t	o the tip of the bil
temperature v	vas extracted by drawing a	a box around the head to select th	e hottest pixel ins
was consister	ntly found on the unfeather	ed periorbital ring.	

-1/	
220	Mean bill temperature (hereafter referred to as "bill temperature") was measured from the mean
221	surface temperature of a straight line fitted from the base of the nostril to the tip of the bill (Fig. 1).
222	Maximum eye region temperature (hereafter referred to as "eye temperature") was taken by fitting a
223	rectangle across the head which was large enough to encompass the periorbital ring, where the
224	maximum temperature of the head is typically recorded (see Jerem et al., 2015). Image focus was
225	recorded as a three-level factor. Each image was ranked as "Good" when all edges of the bill were
226	clearly defined in the image, "Medium" when either the tip or base of the bill was not clearly
227	defined, and "Poor" when the edges of the entire bill were undefined. Though images were selected
228	for quality and lateral view of the head, in some images, the head of the bird was slightly turned to
229	one side. As the length of the line along the bill varies depending on the angle of the head, distance
230	from the camera, as well as the individual size of the bird, the pixel length of the bill was recorded as
231	a continuous variable as a proxy of position of the bird (hereafter referred to as "position index").
232	
233	
234	Statistical analyses
235	
236	All statistical analyses were conducted using R version 3.3.2 (R Development Core Team, 2009).
237	Generalised linear mixed effect models (GLMM) were used to analyse bill and eye region
238	temperatures for both datasets using the <i>lme4</i> package (Bates et al., 2015).
239	
240	Captive
241	
242	Bill temperature and eye region temperature were both modelled using air temperature, the position
243	index, treatment (factorial: before/after food restriction). Bird ID with a first order autoregressive
244	(AR1) covariance structure and the aviary ID were tested as random effects in separate models.
245	However, aviary ID did not improve model fit in any case and was removed from all models.
246	Predicted means (\pm standard error) of the bill and eye region temperatures for each treatment in the
247	model described were calculated using the <i>predictmeans</i> package (version 1.0.1, Luo et al., 2018).
248	
249	Wild
250	We tested effects of food restriction in two ways. Firstly, we tested treatment effects in a model with
251	surface temperatures as the dependent variables and "time" (i.e., before, during, or after food-
252	restriction) as a categorical explanatory variable. We calculated predicted means (\pm standard error)

253	of surface temperature from the described model for each of these "times" using the predictmeans
254	package (version 1.0.1, Luo et al., 2018). Tukey HSD post hoc tests were used to compare
255	differences between food restriction treatments in both wild and captive birds, using the stats
256	package (version 3.5.2, R Development Core Team, 2009). In both tests, we confined the after food-
257	restriction to 1.5 hours from the end of the food restriction to mirror the timings of the captive
258	experiment.
259	
260	Secondly, we also used continuous body surface temperature data from before, during and after food
261	restriction. Bill temperature and eye region temperature were both modelled using, as fixed effects,
262	air temperature, the position index, and the interaction between treatment/control day and time of
263	day both as linear and quadratic terms along with their main effects. Bird ID with a covariance
264	structure (AR1 covariance structures) and focus level were random factors. Focus level did not
265	improve fit and was removed from the model.
266	
267	
268	
269	Results
270	
271	Bill and eye region were linearly related to air temperature in both experiments (Bill: Captive:
272	p<0.0001, Fig. 2A; Wild: p<0.0001, Fig. 2B; Table 2. Eye region: Captive: p<0.0001, Fig. 2C; Wild:
273	p = 0.03, Fig. 2D; Table 2).
274	
275	The position index also accounted for significant variation in the observed bill temperature for
276	captive (p<0.0001, Table 2) and wild great tits (p<0.0001, Table 2).
277	
278	
279	
280	
281	Fig. 2. The relationship between bill and eye region temperatures and air temperature for captive and
282	wild great tits. Captive ($n = 151$ images of 18 birds [15 before, 17 after food restriction]), and wild ($n = 181$
283	images of 60 (incl. 41 unknown) birds [19 on food restricted days and 46 on control days]). Lines are slopes
284 285	from linear models of bill and eye region temperatures against air temperature. Shaded regions are 95% confidence intervals.
286	

207	
288	In the captive study, bill temperature was 1.8 ± 0.5 greater after food restriction (p = 0.0008, Fig.
289	3A, Table 2). In the wild study, bill temperature was significantly lower during the food restriction
290	than both before and after (Before: 14.0 (mean) \pm 0.3 (SE), During: 12.7 \pm 0.2, After: 13.9 \pm 0.3;
291	combined effect: p < 0.0001; Fig. 3B, Table 2). Eye region temperature in captive birds was higher
292	after the food restriction compared to before (Before: $20.0 \pm 0.3^{\circ}$ C; After: $20.8 \pm 0.3^{\circ}$ C, p = 0.04652,
293	Fig. 3C, Table 2). For the wild study, eye region temperature was significantly lower after the food
294	restriction compared to before (Before: 27.6 ± 0.3 , During: 27.0 ± 0.2 , After: 26.7 ± 0.2 ; combined
295	effect: p = 0.0023; Fig. 3D, Table2).
296	
297	
298	
299	
300	
301	Fig. 3. Bill and eye region temperature before, during and after food restriction for wild and captive
302	great tits. Only food-restricted days are shown. The wild study is confined to 1.5 hours from the end of the
303	food restriction to maintain a similar timeframe as in the captive study. Boxes are first and third quartiles and
304	whiskers extend to lowest and highest observation within 1.5 times the interquartile range. Observations
305	outside of this range are shown as solid circles. The mean value is indicated by a cross on each box.
306	Significance values are from Tukey HSD. Significance is indicated by brackets with asterisks indicating
307	significance level (* = p<0.05, *** = p<0.0001). Sample size above each plot indicates the number of images
308	used. The number of individual birds in the treatment groups for the wild were, 11 before food-restriction, 17
309	during food-restriction and 9 after food-restriction. In the captive experiment, 15 individuals were measured
310	before food-restriction and 17 after food-restriction.
311	
312	
313	In the wild study, bill temperature was measured continuously from the start of recording and was
314	found to vary temporally between food restricted and food available days (Fig. 4, Table 2). During
315	food restriction, bill temperature was 1.3 ± 0.3 °C below bill temperature on food available days at
316	the corresponding time period when ambient temperature was accounted for (Fig. 4). After the initial
317	decrease, however, the bill temperature of food restricted birds increased throughout the food
318	restriction period and was similar to that in birds on food available days at the end of the observation
319	period, unlike in the captive birds. Before and after food restriction temperatures were, thus, similar
320	for both food restricted and food available days.

\sim	-	0
~	• ,	')
.,	\angle	_

 324 325 326 327 328 329 330 	Fig. 4. Effects of food restriction on bill temperature for wild great tits. Food restricted days are shown in blue (n = 126 images, 19 birds) and days where food was available are shown in orange (n = 55 images, 46 birds). The smooth curve line and 95% confidence intervals are fitted from locally estimated scatterplot smoothing. The grey shaded region indicates the food restriction period (variation in start and end time between days was < 15 min).
331	Eye region temperature in the wild study was not significantly influenced by food restriction (Fig. 5,
332	Table 2), and the 95% confidence intervals overlapped between food restricted and food available
333	days throughout the experiment. There was a general decrease in eye temperature throughout the
334	experiment, however, as this was true for both food restricted and food available days, this trend was
335	not driven by the food restriction event.
336 337	
338	
339	
 340 341 342 343 344 345 346 	Fig. 5. Effects of food restriction on eye temperature for wild great tits. Food restricted days are shown in blue (n = 126 images, 19 birds) and days where food was available are shown in orange (n = 55 images, 46 birds). The smooth curve line and the 95% confidence intervals are fitted from locally estimated scatterplot smoothing. The grey shaded region indicates the food restriction period (variation in start and end time between days was < 15 min).
347	
348	Table 2. Model outputs of bill temperature for wild and captive great tits. For the captive study, filming
349	occurred before and after a food restriction event and two consecutive days before the food restriction day
350	(included in the control group) (see methods section). The models used are described in the table with the
351	response variable and fixed effects (all models were mixed effects and details of random effect can be found
352	in the methods). Interactions are represented by "x" between variables. Estimates are the change in the
353 354	response variable (i.e., surface temperature) per unit increase in the parameter, or for categorical variables, per unit increase when the baseline equals zero. Baseline levels for categorical variables are indicated by ^a .
355	For interactions, the estimates give the change in slope from the regression of the response for each
356	treatment level compared to the baseline treatment level.
357	
358	

	Model	Parameter	Estimate	SE	F-value	d.f.	Р
	Captive T _{bill} ~ T _{air} + treatment category + position index	Intercept	-0.12	1.42	220.51	1, 130	<0.0001
		Air temperature	0.83	0.08	142.83	1, 130	<0.0001
		Treatment: Before ^a / after food restriction	Before: 4.32 ± 0.39 1.79 After: 6.11 ± 0.45	0.50	14.69	1, 130	0.0008
		Position index	0.32	0.06	30.39	1, 130	<0.0001
-	Wild						
	T _{bill} ~ T _{air} + treatment category + position index	Intercept	7.26	0.88	5055.80	1, 61	<0.0001
ature		Air temperature	0.62 <mark>Before:</mark>	0.09	106.38	1, 61	<0.0001
Bill temperature		Treatment: Before ^a / during/ after food	<i>14.01 ± 0.28</i> During: (During <i>12.71 ± 0.22</i> -1.20	(During) 0.31	20.64	1,61	<0.0001
		restriction	After: (After) 13.92 ± 0.27 -0.09	(After) 0.35			
		Position index	0.17	0.05	9.69	1, 61	0.0028
-	Wild	Intercept	24.67	7.43	6708.43	68	<0.0001
	<mark>time +</mark>	Air temperature	0.42	0.05	107.25	1, 68	<0.0001
	position index + treatment category x time + treatment category x time ²	Treatment: food restricted ^a / food available day	-6.88	15.43	3.31	1, 68	0.0731

		Time of day	-3.31		1.29	0.01	68	0.9177
		Position index	0.23		0.05	24.31	1,68	<0.0001
		Treatment x Time of day	1.63		2.72	3.11	1,68	0.0823
		Treatment x Time of day ²	(Food restrict) (Food availat	,	0.06 0.1	3.78	2,68	0.0279
	Captive	Intercept	19.42	2	1.07	6117.29	1, 107	<0.0001
	T _{eye} ~ T _{air} + treatment category + position index	Air temperature	0.49		0.06	78.66	1, 107	<0.0001
		Treatment: Before ^a / after food restriction	Before: 20.03 ± 0.29 After: 20.81 ± 0.34	0.78	0.37	5.52	1, 107	0.04652
rature		Position index	0.10		0.04	5.08	1, 107	0.02868
n tempe	Wild	Intercept	22.25	5	0.90	40586.53	1, 61	<0.0001
Eye region temperature	$\frac{T_{eye}}{treatment}$	Air temperature	0.44		0.08	42.31	1, 61	<0.0001
	category + position index	Treatment: Before ^a / during/ after food restriction	Before: 27.61 ± 0.26 During: 26.97 ± 0.18 After: 26.69 ± 0.24	(During) -0.64 (After) -0.92	(During) 0.32 (After) 0.36	6.74	1,61	0.0023

	Position index	0.16	0.06	7.67	1, 61	0.0074
Wild	Intercept	20.97	7.5	38927.14	1, 68	<0.0001
	Air temperature	0.1	0.05	5	1, 68	0.0286
T _{eye} ∼ T _{air} + treatment	Treatment: food restricted ^a / food available day	31.66	14.78	1.53	1, 68	0.22
<mark>category +</mark> <mark>time +</mark>	Time of day	0.35	1.3	2.19	1, 68	0.1434
position index + treatment category x time + treatment category x time ²	Position index	0.25	0.05	24.15	1, 68	<0.0001
	Treatment x Time of day	-5.5	2.61	0.27	1, 68	0.6062
	Treatment x Time of day ²	(Food restricted) -0.02 (Food available) 0.22	0.06 0.1	2.52	2, 68	0.088

Discussion We found that the bill temperature of free-ranging great tits decreased significantly during periods of food restriction compared to periods when supplemented food was available to birds. As bill temperature returned to before-food-restriction temperature (or higher, in the case of the captive birds) on food available days, we are confident that the reduction in bill temperature was a direct response to the removal of a reliable food source. The relative immediacy (the lowest temperatures

369 occurs in less than an hour from the beginning of the restriction) of the reduction in bill temperature

- indicates control of vasoconstriction by the bird, rather than reductions in temperature due to lower
- 371 metabolic heat production as a result of the lack of food. This is suggestive of a cautionary measure,
- as an autonomic response, to minimize subsequent energetic shortfalls, should the lack of food
- 373 persist. The putative mechanism, constriction of the blood vessels that supply the bill (cf., Midtgård,
- 1984), reduces the tissue-skin gradient and hence heat loss rate. Tattersall et al., (2017) suggest that
- 375 small birds are disproportionately more affected by heat loss from uninsulated regions compared to
- 376 larger birds. Therefore, vasoconstriction of the bill is likely an important energy-saving response for
- 377 small passerines in cold environments.
- 378

379 Conversely, we found no difference in eye region temperature when wild birds were food restricted 380 compared to periods when food was available. This suggests that the bill temperature response was 381 caused by local vasoconstriction, and not by reduced circulation to the entire head region. A possible 382 cause for maintaining eye region temperature could be the close proximity of the eye to the brain, 383 which must receive a continuous supply of warm blood to maintain function. Likewise, steady, high, 384 temperature in the eye region is likely of value for visual acuity, and hence beneficial for maintained 385 foraging efficiency in a visually guided bird such as the great tit. The relatively long duration the bill 386 was at a lower temperature on food restricted days compared to food available days indicates that 387 vasoconstriction of the bill was not driven by an acute stress response triggered by the experiment. If 388 so, we would have expected to see a considerably faster return to before-food restriction values than 389 in this study, based on the timeline of the thermal response to an acute stressor in periorbital skin in 390 the closely related blue tit (*Cyanistes caeruleus*) (Jerem et al., 2019). This provides evidence for 391 selective vasoconstriction of the bill as opposed to a global drop in peripheral temperature as is 392 expected in response to an acute stressor (e.g., Herborn et al., 2015; Nord and Folkow, 2019; 393 Robertson et al., 2020).

394

The blood supply to the bill must also serve some purpose in functionality, or else it would remain permanently low when the bird is below the thermoneutral zone, even when food is plentiful. It follows that even though vasoconstriction of the bill is likely reflecting a first major defence against energetic shortfalls, it is conceivable that the bird will act to minimise periods of reduced bill function. This could explain why, in the wild, bill temperature gradually increased throughout the food restriction period following the initial drop. This gradual increase in temperature throughout the food restriction may, in part, be through increased activity as birds tried to locate, and potentially

402 ingested, alternative food sources. This is supported by surface temperature increases seen in non-

403 manipulated wild birds throughout the morning, likely from activity-generated heat. Though no

404 filming occurred during the food restriction in the captive study, the significantly higher bill and eye

- 405 temperatures in these birds after the food restriction, compared to before, is likely due to increased
- 406 activity and/or metabolic heat production when re-fed (Zhou and Yamamoto, 1997).
- 407

408 Bill and eye temperature of wild and captive great tits decreased with air temperature, which we 409 believe was largely due to greater heat loss to the environment. Similar trends have been observed in 410 other studies of birds at varying environmental temperatures (McCafferty et al., 2011; Robinson et 411 al., 1976; Tattersall et al., 2016). It is important to note the effect of air temperature on body surface 412 temperature occurred regardless of whether food was being restricted at the time or not. Our data, 413 and those of other studies, highlight the role of the bill in thermoregulation. Under low ambient 414 temperatures, heat loss through the bill is reduced by vasoconstriction; conversely, at high ambient 415 temperatures there is increased circulation to the bill to facilitate heat loss (Tattersall et al., 2009; 416 Wolf and Walsberg, 1996). This thermoregulatory role of the bill, consolidated by our data, should 417 be taken into account when interpreting recently described adaptive changes in bill size, notably in 418 great tits (Bosse et al., 2017; Danner and Greenberg, 2015; Friedman et al., 2017; Symonds and

- 419 Tattersall, 2010; Tattersall et al., 2017).
- 420
- 421 Conclusion
- 422 We have shown the bill plays a key role in the thermoregulatory response to a sudden drop in food availability in wild passerines. This is probably a pre-emptive response by the bird to prevent future 423 424 energetic shortfalls by immediately reducing thermoregulatory costs. In addition, our results also 425 suggest that the level of vasoconstriction is flexible, as bill temperature increased throughout the 426 food restriction, possibly through active control to allow resumed functionality of the bill, or through 427 increased activity to locate alternate food sources. This study gives novel insight into the 428 thermoregulatory responses of birds to meet immediate changes to prospects of energy acquisition. 429 430 Acknowledgements
- 431
- 432

⁴³³ We thank Ruedi Nager, Marina Lehmann, Ross MacLeod and Jan-Åke Nilsson for assistance in data 434 collection and feedback throughout the project, Paul Jerem for crucial advice on experimental setup

435	and comments on the manuscript, and Adam Wynne, Fanny Maillard, Güney Guüvenç and Jean
436	Brustel for assistance in data collection. We would also like to thank staff of both Stensoffa and
437	SCENE field-stations for support throughout this study.
438	
439	Competing interests
440	
441	The authors declare no competing or financial interests.
442	
443	Funding
444	
445	AN was supported by the Birgit and Hellmuth Hertz Foundation / The Royal Physiographic Society
446	of Lund (grant no. 2017-39034) and the Swedish Research Council (grant no. 637-2013-7442). Data
447	collection in Sweden was enabled by an ERASMUS Training Mobility Grant. LAW was supported
448	by a SCENE research bursary for the MRes Ecology and Environmental Biology at the University of
449	Glasgow.
450	
451	
452	
453	
454	
455	
456	
457	
458	
459	
460	
461	
462	
463	
464	
465	
466	
467	

References

- Bates, D., Maechler, M., Bolker, B. and Walker, S. (2015). Fitting Linear Mixed-Effects Models Using lme4. J. Stat. Softw. 67, 1–48.
- Best, R. G. and Fowler, R. (1981). Infrared emissivity and radiant surface temperatures of canada and snow geese. *J. Wildl. Manage.* **45209157**, 1026–1029.
- Bosse, M., Spurgin, L. G., Laine, V. N., Cole, E. F., Firth, J. A., Gienapp, P., Gosler, A. G., McMahon, K., Poissant, J., Verhagen, I., et al. (2017). Recent natural selection causes adaptive evolution of an avian polygenic trait. *Science* (80-.). 358, 365–368.
- **Broggi, J., Hohtola, E., Orell, M. and Nilsson, J. Å.** (2005). Local adaptation to winter conditions in a passerine spreading north: A common-garden approach. *Evolution (N. Y).* **59**, 1600–1603.
- **Danner, R. M. and Greenberg, R.** (2015). A critical season approach to Allen's rule: Bill size declines with winter temperature in a cold temperate environment. *J. Biogeogr.* **42**, 114–120.
- Friedman, N. R., Harmáčková, L., Economo, E. P. and Remeš, V. (2017). Smaller beaks for colder winters: Thermoregulation drives beak size evolution in Australasian songbirds. *Evolution (N. Y).* 1–10.
- Gavrilov, V. M. and Dolnik, V. R. (1985). Basal metabolic rate, thermoregulation and existence energy in birds: World data. *Acta XVIII Congr. Int. Ornithol.* **1**, 421–466.
- Hagan, A. A. and Heath, J. E. (1980). Regulation of heat loss in the duck by vasomotion in the bill.*J. Therm. Biol.* 5, 95–101.
- Herborn, K. A., Graves, J. L., Jerem, P., Evans, N. P., Nager, R., McCafferty, D. J. and McKeegan, D. E. F. (2015). Skin Temperature Reveals the Intensity of Acute Stress. *Physiol. Behav.* 152, 225–230.
- Hohtola, E. (2012). Thermoregulatory Adaptations to Starvation in Birds. In *Comparative Physiology of Fasting, Starvation, and Food Limitation* (ed. McCue, M. D.), pp. 155–170. Berlin, Heidelberg: Springer Berlin Heidelberg.
- Jerem, P., Herborn, K., McCafferty, D., McKeegan, D. and Nager, R. (2015). Thermal Imaging to Study Stress Non-invasively in Unrestrained Birds. *J. Vis. Exp.* e53184.
- Jerem, P., Jenni-Eiermann, S., Herborn, K., McKeegan, D., McCafferty, D. J. and Nager, R. G. (2018). Eye region surface temperature reflects both energy reserves and circulating glucocorticoids in a wild bird. *Sci. Rep.* 8, 1–10.

Jerem, P., Jenni-Eiermann, S., McKeegan, D., McCafferty, D. J. and Nager, R. G. (2019). Eye

region surface temperature dynamics during acute stress relate to baseline glucocorticoids independently of environmental conditions. *Physiol. Behav.* **210**, 112627.

- Johansen, K. and Bech, C. (1983). Heat conservation during cold exposure in birds (vasomotor and respiratory implications). *Polar Res.* **1**, 259–268.
- Johnsen, H. K., Blix, A. S., Jorgensen, L. and Mercer, J. B. (1985). Vascular basis for regulation of nasal heat exchange in reindeer. *Am. J. Physiol.* 617–623.
- Luo, D., Ganesh, S. and Koolaard, J. (2018). predictmeans: Calculate Predicted Means for Linear Models. R package version 1.0.1. https://CRAN.R-project.org/package=predictmeans.
- Marsh, R. L. and Dawson, W. R. (1989). Avian Adjustments to Cold. In Animal Adaptation to Cold (ed. Wang, L. C. H.), pp. 205–253. Berlin, Heidelberg: Springer Berlin Heidelberg.
- McCafferty, D. J. (2013). Applications of thermal imaging in avian science. *Ibis (Lond. 1859).* **155**, 4–15.
- McCafferty, D. J., Gilbert, C., Paterson, W., Pomeroy, P., Thompson, D., Currie, J. I. and Ancel, A. (2011). Estimating metabolic heat loss in birds and mammals by combining infrared thermography with biophysical modelling. *Comp. Biochem. Physiol. A Mol. Integr. Physiol.* 158, 337–345.
- Midtgård, U. (1981). The Rete tibiotarsale and Arterio-venous association in the hind limb of birds: a comparative morphological study on counter-current heat exchange systems. *Acta Zool.* 62, 67–87.
- **Midtgård, U.** (1984). Blood vessels and the occurrence of arteriovenous anastomoses in cephalic heat loss areas of mallards, Anas platyrhynchos (Aves). *Zoomorphology* 323–335.
- Nord, A. and Folkow, L. P. (2019). Ambient temperature effects on stress-induced hyperthermia in Svalbard ptarmigan. *Biol. Open* **8**, 1–5.
- Nord, A., Nilsson, J. F. and Nilsson, J.-Å. (2011). Nocturnal body temperature in wintering blue tits is affected by roost-site temperature and body reserves. *Oecologia* **167**, 21–25.
- Nord, A., Lehmann, M., MacLeod, R., McCafferty, D. J., Nager, R. G., Nilsson, J.-Å. and Helm, B. (2016). Evaluation of two methods for minimally invasive peripheral body temperature measurement in birds. J. Avian Biol. 47, 417–427.

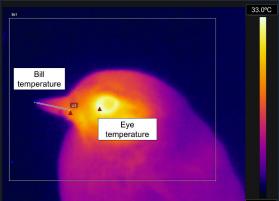
R Development Core Team (2009). R: A Language and Environment for Statistical Computing.

- **Robertson, J. K., Mastromonaco, G. and Burness, G.** (2020). Evidence that stress-induced changes in surface temperature serve a thermoregulatory function. *J. Exp. Biol.* 788182.
- Robinson, D. E., Campbell, G. S. and King, J. R. (1976). An evaluation of heat exchange in small birds. *J. Comp. Physiol. B* **105**, 153–166.

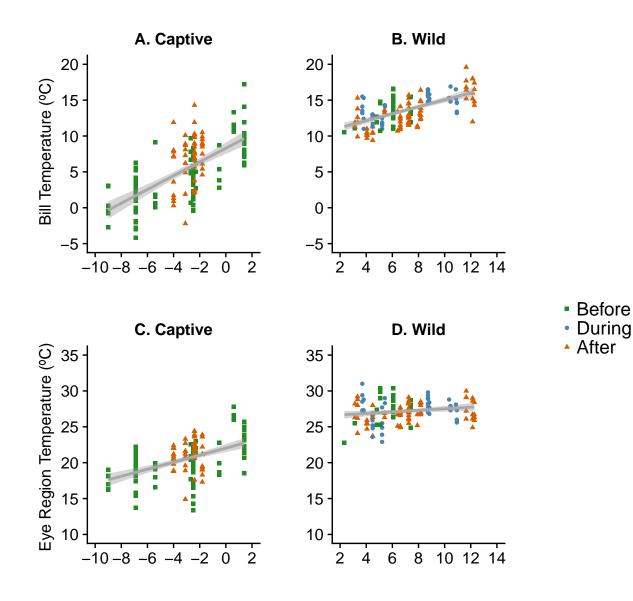
Scholander, P. F., Hock, R., Walters, V. and Irving, L. (1950). Adaption To Cold In Arctic And

Tropical Mammals And Birds In Relation To Body Temperature, Insulation, And Basal Metabolic Rate. *Biol. Bull.* **99**, 259–271.

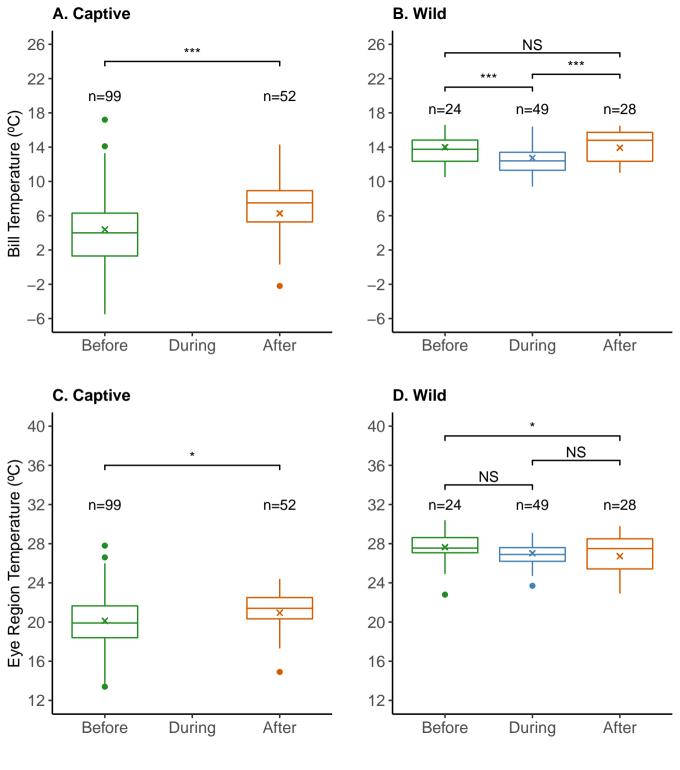
- Schraft, H. A., Whelan, S. and Elliott, K. H. (2019). Huffin' and puffin: Seabirds use large bills to dissipate heat from energetically demanding flight. J. Exp. Biol. 222, 2017–2019.
- Shipley, A. A., Sheriff, M. J., Pauli, J. N. and Zuckerberg, B. (2019). Snow roosting reduces temperature-associated stress in a wintering bird. *Oecologia* 190, 309–321.
- Steen, I. and Steen, J. B. (1965). The Importance of the Legs in the Thermoregulation of Birds. *Acta Physiol. Scand.* 63, 285–291.
- Swanson, D. L. and Vézina, F. (2015). Environmental, ecological and mechanistic drivers of avian seasonal metabolic flexibility in response to cold winters. J. Ornithol. 156, 377–388.
- Symonds, M. R. E. and Tattersall, G. J. (2010). Geographical Variation in Bill Size across Bird Species Provides Evidence for Allen's Rule. *Am. Nat.* 176, 188–197.
- Tattersall, G. J., Andrade, D. V. and Abe, A. S. (2009). Heat Exchange from the Toucan Bill Reveals a Controllable Vascular Thermal Radiator. *Science (80-.).* 325, 468–470.
- Tattersall, G. J., Roussel, D., Voituron, Y. and Teulier, L. (2016). Novel energy-saving strategies to multiple stressors in birds: the ultradian regulation of body temperature. *Proc. R. Soc. London B Biol. Sci.* 283,.
- Tattersall, G. J., Arnaout, B. and Symonds, M. R. E. (2017). The evolution of the avian bill as a thermoregulatory organ. *Biol. Rev.* 92, 1630–1656.
- Tattersall, G. J., Chaves, J. A. and Danner, R. M. (2018). Thermoregulatory windows in Darwin's finches. *Funct. Ecol.* **32**, 358–368.
- William, R., Marsh, R. L. and Yacoe, M. E. (1983). Metabolic adjustments of small passerine birds for migration and cold. Am. J. Physiol. 245, 755–767.
- Wolf, B. O. and Walsberg, G. E. (1996). Respiratory and cutaneous evaporative water loss at high environmental temperatures in a small bird. *J. Exp. Biol.* **199**, 451–457.
- Zhou, W. T. and Yamamoto, S. (1997). Effects of environmental temperature and heat production due to food intake on abdominal temperature, shank skin temperature and respiration rate of broilers. *Br. Poult. Sci.* 38, 107–114.



12.0°C



Air Temperature (°C)



Food Restriction Treatment

