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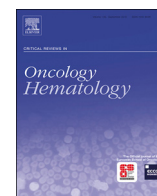
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# Differential redox-regulation and mitochondrial dynamics in normal and leukemic hematopoietic stem cells: A potential window for leukemia therapy



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## ABSTRACT

The prognosis for many patients with acute myeloid leukemia (AML) is poor, mainly due to disease relapse driven by leukemia stem cells (LSCs). Recent studies have highlighted the unique metabolic properties of LSCs, which might represent opportunities for LSC-selective targeting. LSCs characteristically have low levels of reactive oxygen species (ROS), which apparently result from a combination of low mitochondrial activity and high activity of ROS-removing pathways such as autophagy. Due to this low activity, LSCs are highly dependent on mitochondrial regulatory mechanisms. These include the anti-apoptotic protein BCL-2, which also has crucial roles in regulating the mitochondrial membrane potential, and proteins involved in mitophagy.

Here we review the different pathways that impact mitochondrial activity and redox-regulation, and highlight their relevance for the functionality of both HSCs and LSCs. Additionally, novel AML therapy strategies that are based on interference with those pathways, including the promising BCL-2 inhibitor Venetoclax, are summarized.

## 1. Introduction and scope of this review

Current treatment strategies for acute myeloid leukemia (AML) result in an initial reduction of leukemic blasts in the majority of patients. However, a small population of leukemia stem cells (LSCs) persists during therapy and leads to disease relapse. In order to improve therapy success, AML research has focused on studying the molecular mechanisms that underlie LSC properties, and highlighted common as well as distinct features of LSCs compared to normal hematopoietic stem cells (HSCs). The functionality of HSCs is closely connected to their ability to avoid accumulation of oxidative stress and maintain low levels of reactive oxygen species (ROS). With increasing levels of ROS, HSCs capability for long-term repopulation declines, suggesting that low levels of ROS might be an indicator of stem cell functionality (Hu et al., 2018; Singh et al., 2018). Several recent studies have supported the idea that a similar concept applies to LSCs (Jones et al., 2018; Lagadinou et al., 2013; Pei et al., 2018). LSCs are a relatively rare fraction of self-renewing malignant cells responsible for disease maintenance (Thomas and Majeti, 2017). Interestingly, even though a ROS-low state is indicative for both HSC and LSC function, the two cell populations seem to have an altered dependency on pathways

regulating ROS production and mitochondrial health. HSCs rely on glycolysis as their main energy source, which results in less oxidative burden compared to mitochondrial oxidative phosphorylation (OXPHOS). Mitochondrial activity and ROS production in HSCs are often induced by either growth factors or cytokines that promote cell differentiation or apoptosis. Although LSCs also have characteristically low levels of OXPHOS (Jones et al., 2018; Lagadinou et al., 2013; Pollyea et al., 2018a), they depend on this remaining activity for their survival. Consequently, they ensure low ROS levels and high mitochondrial integrity through mechanisms such as autophagy (Pei et al., 2018). Moreover, LSCs are highly dependent on pathways that counteract mitochondrial dysfunction and cell death. These LSC-specific characteristics led to the hypothesis that LSCs can be selectively targeted during AML treatment while sparing HSCs.

To address this hypothesis, we begin this review by describing mitochondrial ROS generation and removal, and discuss the relevance of mitochondrial activity for the maintenance and differentiation of stem cells. For both HSCs and LSCs we then summarize the current evidence suggesting that ROS levels are indicative for their functionality. Next, we describe the signaling pathways that have critical roles in regulating redox homeostasis and their potential impact on stem cell maintenance.

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This includes five modes of signaling – HIF-1 $\alpha$ , AMPK, mTOR, FOXO and SIRT – as well as a brief overview of crucial players in DNA damage response pathways. In the final part, we discuss vulnerabilities of LSCs and how interference with autophagy or the anti-apoptotic protein BCL-2, which is essential to mitochondrial health, could offer a strategy for targeting these proteins as part of AML treatment.

## 2. ROS generation and its relevance for stem cell maintenance and differentiation

ROS is a collective term for oxygen-containing molecules that are more reactive than molecular oxygen (O<sub>2</sub>), and mainly include hydroxyl radicals (OH $\cdot$ ), hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) and superoxide anion radicals (O<sub>2</sub> $\cdot^-$ ). Due their chemical characteristics, ROS can easily react with DNA or RNA bases, fatty acids in lipids or amino acids in proteins (Ray et al., 2012). For the majority of mammalian cells, mitochondrial energy production is the major source of ROS (Murphy, 2009). During ATP production by mitochondrial oxidative phosphorylation (OXPHOS), substrates are oxidized by a series of enzyme complexes (complex I-IV) located at the inner mitochondrial membrane. Reactions of this electron transport chain (ETC) lead to release of ROS. Besides ROS production by mitochondria, these molecules are also produced in enzymatic reactions by the family of nicotinamide adenine dinucleotide phosphate (NADPH) oxidases (NOXs) and other oxidases involved in inflammatory reactions and other processes. In this review we focus on the role of mitochondrial ROS.

The total amount of steady-state cellular ROS differs between cell types. In general, the primitive stem cell fraction has a lower ROS content during steady-state, while higher ROS levels are found in more differentiated cells. This difference mainly results from metabolic properties that are closely linked to the cellular function. (Bigarella et al., 2014). The low ROS content in the stem cell fraction might be protective for DNA damaging effects (van Galen et al., 2014; Yahata et al., 2011) and primitive stem cells use various strategies to avoid ROS generation. For instance, stem cells from different origins have been found to reside in defined anatomical compartments with low oxygen tension, which are called stem cell niches (Mohyeldin et al., 2010). These hypoxic conditions induce altered metabolic features, including characteristically low mitochondrial activity (Rafalski et al., 2012). Undifferentiated embryonic stem cells contain small, immature mitochondria, but during differentiation cells acquire a more elongated, mature mitochondrial phenotype, accompanied by higher copy numbers of mitochondrial DNA and elevated ATP production required for cell growth and proliferation (Facucho-Oliveira and St John, 2009). Similarly, HSCs were shown to have relatively low mitochondrial activity and low membrane potential, a feature that was shown to be essential for their stemness potential (Maryanovich et al., 2015; Norddahl et al., 2011; Rimmele et al., 2015; Simsek et al., 2010; Sukumar et al., 2016; Vannini et al., 2016). Quiescent HSCs rely primarily on anaerobic glycolysis for their energy generation (Suda et al., 2011). Although this is less efficient than OXPHOS, it also restricts the burden of ROS-mediated oxidative stress (Fig. 1). However, upregulation of mitochondrial activity in concert with increased ROS levels is crucial for HSC differentiation, since disruption of OXPHOS was shown to impair this process (Takubo et al., 2013; Yu et al., 2013).

ROS can drive cell proliferation and differentiation mechanistically via redox modification of proteins critically involved in these processes (Ray et al., 2012). In addition, ROS modulate redox-sensitive factors that regulate ROS production itself, such as forkhead homeobox type O family (FOXOs), ataxia telangiectasia mutated (ATM) or Sirtuins (SIRT). ROS also modulate molecules that have important functions in stem cell maintenance, differentiation or stress response, such as hypoxia-inducible factor 1 $\alpha$  (HIF-1 $\alpha$ ), p38 and p53. ROS-mediated activation of the p38-MAPK (mitogen-activated protein kinase) pathway was shown to have a crucial role in limiting the lifespan and functionality of HSCs (Ito et al., 2006; Jung et al., 2016; Miyamoto et al.,

2007; Zhang et al., 2016). P38 phosphorylation – and thus activation – promotes inhibitory programs such as cell-cycle arrest and apoptosis, or can activate purine metabolism, resulting in increased HSC cycling (Karigane et al., 2016). Furthermore, ROS-induced p53 was found to be important for controlling HSC survival (Abbas et al., 2010).

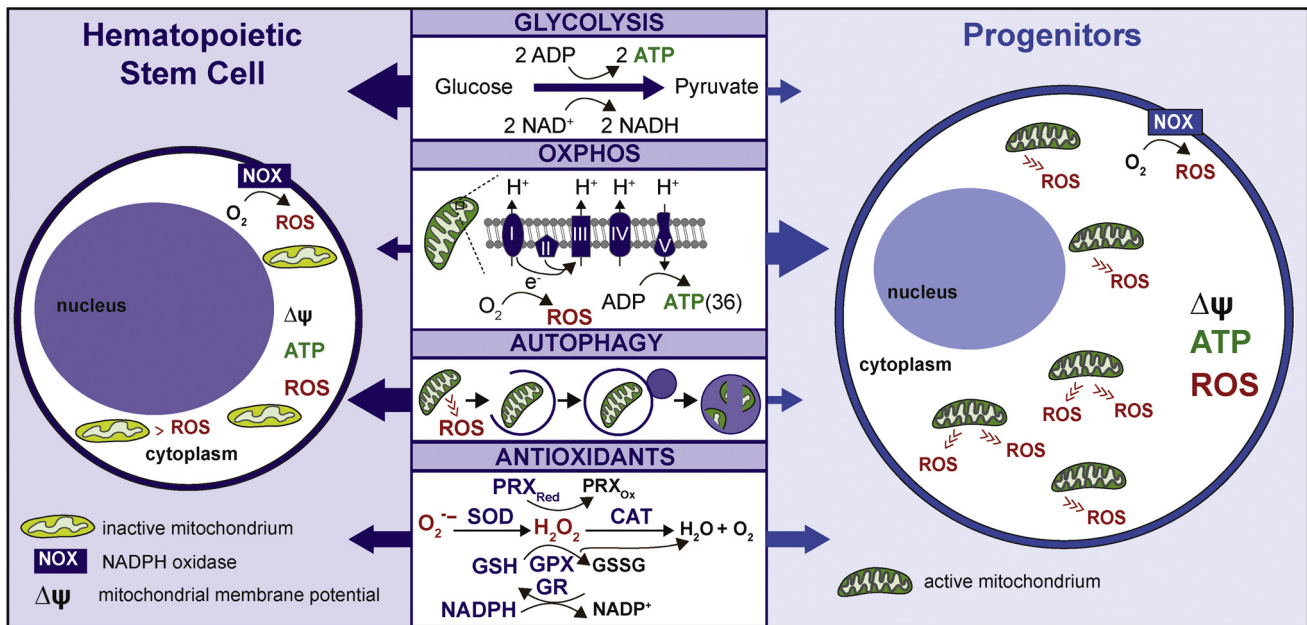
## 3. ROS-regulating mechanisms maintain HSC functionality

To counteract ROS production and prevent oxidative stress, HSCs have numerous ROS-scavenging strategies. These include low-molecular-weight reducing peptides (e.g. glutathione, thioredoxin, NADPH), peroxiredoxins and antioxidant enzymes (e.g. catalase, superoxide dismutase (SOD), glutathione peroxidase). They also have other ROS regulating mechanisms, such as mitophagy, that control mitochondrial quantity and quality (Fig. 1), or the recently discovered process of CoAlation (Aloum et al., 2019; Tsuchiya et al., 2017). Additionally, interactions of cells with their microenvironment can impact their ROS levels (Ludin et al., 2012; Taniguchi Ishikawa et al., 2012). In steady-state conditions, higher levels of antioxidant enzymes have not been found in HSCs than in more committed progenitors (Bagger et al., 2013), indicating that the higher availability of antioxidants does not explain the lower ROS levels in HSCs. However, transcription factors regulating the expression of anti-oxidative molecules are essential for HSC functioning (Jung et al., 2013; Miyamoto et al., 2007; Tothova et al., 2007; Yalcin et al., 2008). This suggests that stem cells could be more dependent on anti-oxidative molecules than more committed progenitors. This postulation is also supported by a recent study showing a correlation between the magnitude of ROS-mediated effects and cell size: in smaller cells ROS interact more easily with membranes (Molavian et al., 2016). Stem cells are characteristically smaller in size than their progeny (Li et al., 2015) and could thus rely more on ROS-scavenging mechanisms. In line with this possibility, HSCs were shown to depend on ROS-regulating pathways such as autophagy, in which cells break down dysfunctional organelles by lysosomal degradation (Mizushima et al., 2008).

Autophagy results in the clearance of ROS-producing mitochondria – a process known as mitophagy – which is especially important for HSC functioning (Ito et al., 2016; Joshi and Kundu, 2013). Autophagy is regulated by a number of autophagy-related genes (Atg); Atg5 and Atg7 knock-out mice show an increased number of mitochondria and ROS levels in hematopoietic stem and progenitor cells in conjunction with an impaired functionality (Gomez-Puerto et al., 2016; Mortensen et al., 2011). Similarly, low autophagy levels in aged HSCs are accompanied by high ROS levels and reduced functionality, whereas aged HSCs with high autophagy levels perform comparably to their younger counterparts (Ho et al., 2017). Efficient mitophagy is also closely linked to the dynamic mitochondrial fusion/fission process in which damaged mitochondria become segregated by fission (division) and healthy material is exchanged between mitochondria via fusion. Altered expression of fission-proteins DRP1 and FIS1 can impact stem cell maintenance (Cai et al., 2016; Pei et al., 2018).

## 4. Excessive mitochondrial ROS production triggers cell death pathways

Although physiological ROS levels are important for intracellular signaling, excessive ROS can induce cell death by triggering apoptotic pathways. Because they are the primary cellular location of ROS production, mitochondria are particularly vulnerable to ROS-induced damage. ROS can damage mitochondrial DNA, membrane lipids and proteins (reviewed in (Cline, 2012)), which can result in mitochondrial genomic instability and respiratory dysfunction. ROS can also oxidize and damage components in the inner and outer mitochondrial membrane, resulting in a dysfunctional respiratory chain, a drop in mitochondrial membrane potential and eventually triggering mitochondrial permeability transition (MPT) and cell death (Mantel et al., 2015).



**Fig. 1.** Steady-state ROS levels vary in the different types of hematopoietic cells. **Left panel:** Hematopoietic stem cells (HSCs) have characteristically low mitochondrial activity and low levels of reactive oxygen species (ROS), which are maintained by using glycolysis as their main energy source, removing stressed (and ROS producing) mitochondria via autophagy and neutralizing ROS via reactions of anti-oxidative enzymes. HSCs produce less ATP and have less active mitochondria compared to progenitors. **Right panel:** More differentiated progenitors characteristically have higher levels of mitochondrial activity and oxidative phosphorylation (OXPHOS), increased ATP production and lower levels of autophagy. They also make use of anti-oxidative enzymes to avoid oxidative stress. PRX: peroxiredoxin; SOD: superoxide dismutase; CAT: catalase; GSH: glutathione; GPX: glutathione peroxidase; GR: glutathione reductase; GSSG: glutathione disulfide; NADP: nicotinamide adenine dinucleotide phosphate.

Moreover, accumulation of ROS can induce structural changes in the mitochondrial outer membrane (MOM) and trigger MOM permeabilization (MOMP) mediated by BCL-2 family proteins. An important effector of ROS-induced cell death is the pro-apoptotic protein p53. Whereas mild oxidative stress leads to expression of p53-targets with antioxidant function, high ROS levels trigger p53-mediated cell death (Perri et al., 2016). Hyper-activation of p53 was shown to deplete HSCs (Abbas et al., 2010), whereas drug-induced mitochondrial dysfunction resulted in LSC-specific apoptosis in leukemia cells, accompanied by increased ROS levels and p53-activation. (Guzman et al., 2005) Furthermore, inhibition of autophagy – and thereby mitochondrial turnover – was shown to target p53-wildtype leukemias, while being ineffective on cells carrying p53 mutations (Folkerts et al., 2017). In line with these observations, several recent studies have shown the importance of mitochondrial health for the survival and function of both HSCs (Anso et al., 2017; Bejarano-Garcia et al., 2016; Ho et al., 2017; Hu et al., 2018; Ito et al., 2016; Luchsinger et al., 2016; Mohrin et al., 2015; Qian et al., 2016; Umemoto et al., 2018) and LSCs (Capala et al., 2016; Farge et al., 2017; Kuntz et al., 2017; Nguyen et al., 2019b; Pei et al., 2018; Seneviratne et al., 2019; Yehudai et al., 2018).

Mitochondrial ROS production also plays a central role in a different type of programmed cell death induced by oxidative stress, known as ferroptosis (Gao et al., 2019), which is the process of regulated necrosis induced by iron-mediated lipid peroxidation. In brief, if ROS interact with lipids and remove their free electrons, “lipid ROS” are formed. If lipid ROS accumulate, ferroptosis-mediated cell death is induced.

## 5. ROS levels in normal hematopoietic stem cells (HSCs)

It was initially assumed that HSCs have a lower number of mitochondria compared to more differentiated progenitors, which could contribute to the low rates of oxidative metabolism observed in HSCs (Mantel et al., 2012; Mohrin et al., 2015; Romero-Moya et al., 2013; Takubo et al., 2013; Xiao et al., 2012). However, more recent studies have challenged this idea by showing that the lower values of

mitochondrial mass observed when using dye-based methods are the result of an altered expression of efflux pumps in HSCs and not of a lower number of mitochondria. Instead, these researchers proposed that HSC contain a relatively high number of less active mitochondria (de Almeida et al., 2017; Norddahl et al., 2011). Notably, the accumulation of mitochondrial mutations in HSCs did not impair the maintenance of HSCs per se, but rather their capability to give rise to functional progenitors. This finding supports the notion that oxidative metabolism is less important for HSCs themselves, but is crucial for their repopulation capacity (Norddahl et al., 2011).

Studies that compared ROS levels between stem cells and progenitor cells found that HSCs and megakaryocyte-erythroid progenitors (MEPs) have the lowest ROS levels, whereas granulocyte-macrophage progenitors (GMPs) have the highest ROS levels (Khan et al., 2016; Shinohara et al., 2014). This observation is possibly in line with the results of several recent studies suggesting that the megakaryocytic lineage directly evolves from HSCs (Carrelha et al., 2018; Haas et al., 2015; Notta et al., 2016; Rodriguez-Fraticelli et al., 2018; Sanjuan-Pla et al., 2013; Yamamoto et al., 2013). Growth factors that promote myeloid proliferation and differentiation, as well as cytokines that are released upon tissue damage (Ishida et al., 2017), often induce mitochondrial biogenesis and ROS production. This correlates with HSC differentiation or exhaustion (Hu et al., 2018; Singh et al., 2018). However, there is still a dynamic range in levels of ROS within the phenotypically defined steady-state HSC population that is not exposed to any additional stressors, which is indicative of their functionality. In 2007, Jang and Sharkis separated viable murine Lin<sup>-</sup>CD45<sup>+</sup> bone marrow cells into a ROS-low and ROS-high fraction based on their signal intensity for the fluorescent ROS-dye DCFDA (2',7'-dichlorofluorescein diacetate) (Jang and Sharkis, 2007). ROS-low HSCs were shown to have higher self-renewal potential, whereas serial transplantation assays with ROS-high HSCs exhausted faster than their ROS-low counterparts due to increased activation of the p38-MAPK. More recently it was shown that TNF secretion by bone marrow cells and subsequent elevation of ROS can fluctuate between different times of

the day based on influences of light and darkness. This determines whether HSCs self-renew (when ROS levels are reduced) or differentiate (when ROS levels are higher) (Golan et al., 2018). Besides ROS-dependent variability in HSC function during steady state, numerous mouse gene knock-out studies reported that a stress-induced increase in ROS results in impaired HSC function or exhaustion (Ito et al., 2004; Liu et al., 2009; Miyamoto et al., 2007; Tothova et al., 2007). Furthermore, it was also shown that exposure of HSCs to ambient oxygen during the procedure of stem cell transplantation can lead to extra-physiologic oxygen shock/stress response (EPHOSS), resulting in ROS accumulation and decreased repopulation potential upon transplantation (Mantel et al., 2015; Woolthuis et al., 2013). In line with this finding, long-term engraftment of human HSCs in murine models could be enhanced by overexpression of catalase (a ROS-detoxifying agent) (Miao et al., 2013) or pre-treatment with valproic acid, which enhanced glycolytic potential and decreased mitochondrial activity, thereby counteracting ROS accumulation (Papa et al., 2018). In summary, these studies indicate that HSCs are best maintained under ROS-low conditions. During steady state, these conditions result from their glycolytic metabolism and location in the hypoxic bone marrow niche.

## 6. ROS levels in Leukemia stem cells (LSCs)

According to the prevailing LSC model, a rare population of malignant cells with properties similar to normal HSCs – such as self-renewal and quiescence – is capable of maintaining the disease. (reviewed in (Thomas and Majeti, 2017)). Moreover, LSCs can initiate leukemia when transplanted into immunodeficient mice (Jordan, 2007), and are therefore alternatively called leukemia initiating cells (LICs). LSCs are functionally defined, do not have a uniform phenotype (Eppert et al., 2011; Sarry et al., 2011) and exhibit great variability in molecular defects (Shlush et al., 2017). However, despite their heterogeneity, several recent studies have suggested that LSCs share metabolic features that are distinct from the total leukemia cell population and characteristically include low mitochondrial activity and low levels of cellular ROS (Hao et al., 2018; Lagadinou et al., 2013; Pei et al., 2018; Pollyea et al., 2018b).

Recently, Hao et al proposed that LSCs are more glycolytic compared to bulk AMLs cells (Hao et al., 2018), which would reflect the similar relation that HSCs have compared to their more differentiated progenitors. The investigators used a metabolic sensor called SoNar (sensor for NAD(H) redox) that has different fluorescent properties depending on whether it binds to NADH or NAD<sup>+</sup>, and thus reflects the cytosolic NADH/NAD<sup>+</sup> ratio of a cell. Glycolysis describes the conversion of glucose to pyruvate and leads to production of NADH, which may be further oxidized to NAD<sup>+</sup> by mitochondrial reactions or by the enzyme lactate dehydrogenase during lactate production. Hence, high levels of glycolysis accompany NADH accumulation, and SoNar-high cells are assumed to have higher glycolytic activity. SoNar-high cells were found to be highly enriched for LSCs, in conjunction with low levels of mitochondrial membrane potential and high expression of enzymes that block the entry of substrates in the TCA cycle such as the pyruvate dehydrogenase PDK2.

These findings are in line with other recent studies showing that AML LSCs have low levels of oxidative metabolism and reside in the “ROS-low” fraction of the AML mononuclear cell population (Lagadinou et al., 2013; Pei et al., 2018; Pollyea et al., 2018b). Additionally, our group recently demonstrated that even CD34<sup>+</sup> selected stem- and progenitor AML cells have a relatively broad range of ROS levels and that the CD34<sup>+</sup>/ROS-low fraction is enriched for phenotypic CD34<sup>+</sup>CD38<sup>-</sup> LSCs (unpublished results). Low levels of ATP were observed in both SoNar-high and ROS-low cells, but these cells also had opposing characteristics. In another study, ROS-low leukemia cells were described as having rather low levels of glycolysis and were enriched for quiescent cells (Lagadinou et al., 2013), whereas SoNar-high cells were described as non-quiescent (Hao et al., 2018). However, mice

transplantation studies have indicated that both groups of cells are enriched for LSCs (Lagadinou et al., 2013), (Hao et al., 2018).

In summary, the above studies suggest that LSCs tend to have low levels of oxidative metabolism and ROS, which coincides with other studies demonstrating that overexpression of glycolysis genes such as PDK2 and PDK3 is associated with poor prognosis in AML (Cui et al., 2018). Although these observations could lead to the assumption that LSCs are less dependent on oxidative metabolisms for their survival, the opposite seems to be the case. Multiple studies have demonstrated that functional mitochondria biology and OXPHOS are crucial for the survival and maintenance of leukemia cells (Samudio et al., 2010; Lagadinou et al., 2013; Cole et al., 2015; Sriskanthadevan et al., 2015). Inhibition of mitochondrial translation by Tigecycline resulted in decreased expression of the mitochondrial complex IV subunits COX-1 and COX-2 and impaired survival of AML LSCs (Skrtic et al., 2011). Similarly, Tigecycline-treatment efficiently targeted primitive CD34<sup>+</sup> CML LSCs, whereas it did not affect the colony-forming-capacity of normal CD34<sup>+</sup> cells (Kuntz et al., 2017). When mitochondrial activity is blocked, LSCs may also have decreased ability to switch to glycolysis as their main energy source (Lagadinou et al., 2013). Furthermore, treatment of primary AML blasts with a small molecule inhibitor of complex I of the mitochondrial electron transport chain (ICAS-010759) induced apoptosis, while treatment of normal bone marrow cells did not affect viability (Molina et al., 2018). Additionally, LSC survival was impaired not only by inhibition of mitochondria themselves, but also by pathways that generate substrates for the mitochondrial TCA cycle, including inhibition of fatty acid oxidation (Samudio et al., 2010), glutaminolysis (Goto et al., 2014) and amino acid metabolism (Jones et al., 2018).

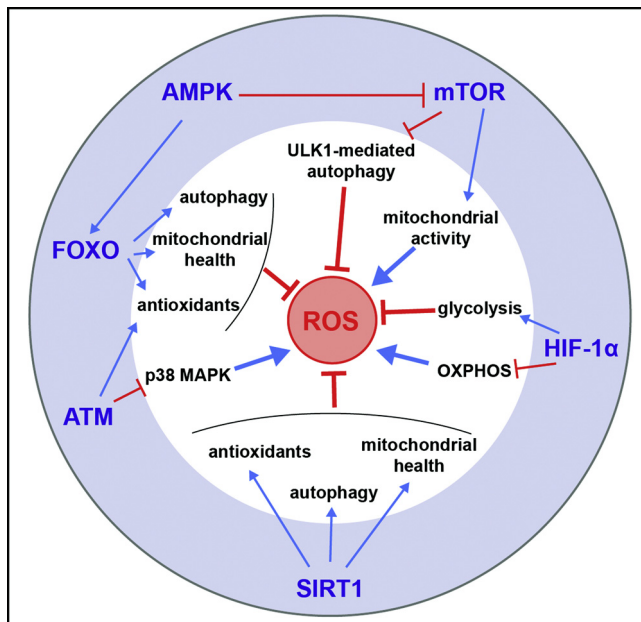
## 7. ROS-regulating pathways and their importance for stem cell functionality

As summarized above, low levels of ROS are indicative for both HSCs and LSCs. In line with this notion, pathways that contribute to a cellular ROS-low state are often found to function as guardians of stemness. These pathways are outlined in this section. Key players for controlling cellular ROS levels are involved not only in the regulation of metabolic pathways, but also in important stress response mechanisms such as DNA repair or autophagy. Furthermore, effective functioning of pathways that mediate mitochondrial health is essential for both HSC and LSC functionality (Fig. 2).

### 7.1. HIF signaling

The transcription factors hypoxia-inducible factor (HIF) 1 $\alpha$  and 2 $\alpha$  have key role in translating changes of the cellular environment and nutrient availability into a transcriptional response. HIF-1 $\alpha$  and HIF-2 $\alpha$  both form heterodimers composed of a constitutively-expressed  $\beta$ -subunit and an  $\alpha$ -subunit that is degraded by an oxygen-dependent hydrolase. Consequently, it is present only under hypoxic conditions (Ivan et al., 2001). HIF-1 $\alpha$  was shown to be highly expressed in HSCs with long-term repopulating capacity (Simsek et al., 2010) and has been described as a master regulator of metabolic pathways that contribute to a ROS-low environment (Gezer et al., 2014; Karigane and Takubo, 2017; Zhang and Sadek, 2014). In brief, HIF-1 $\alpha$  stimulates glycolysis by inducing expression of glycolytic enzymes (e.g. HK1, LDHA) and glucose transports (e.g. GLUT1), and inhibits mitochondrial OXPHOS by inducing expression of enzymes (e.g. PDK2, PDK4) that block entry of substrates into the TCA cycle.

Although no consensus has been reached on whether HIF expression is necessary for full functionality of HSCs, multiple studies have indicated that HIF plays a role in HSC function: Conditional deletion of *Hif-1 $\alpha$*  itself (Takubo et al., 2010) or its upstream regulator *Meis1* (Simsek et al., 2010) was shown to impair HSC quiescence. *Meis1*<sup>-/-</sup> HSCs showed increased ROS production, loss of HSC maintenance



**Fig. 2.** A crosstalk of signaling pathways determines the final ROS concentration and impacts stemness. Various pathways influence ROS levels by regulating mitochondrial activity, glycolysis, autophagy, expression of antioxidative enzymes or stress-responsive signaling cascades. AMPK: AMP-activated protein kinase; mTOR: mammalian target of rapamycin; FOXO: forkhead box class O family; ATM: ataxia telangiectasia mutated; hypoxia-inducible factor 1 $\alpha$  (HIF-1 $\alpha$ ); SIRT1: sirtuin 1; p38 MAPK: p38 mitogen activated protein kinase.

under stress conditions and increased HSC apoptosis, which is a phenotype that could be entirely rescued by treatment with NAC (Kocabas et al., 2012; Unnisa et al., 2012). Furthermore, depletion of the HIF-1 $\alpha$  target gene *Ldha*, a subunit of the enzyme lactate dehydrogenase (LDH) that regulates the last step of anaerobic glycolysis, increased ROS levels in mouse bone marrow and impaired HSC maintenance (Wang et al., 2014). Similarly, deletion of the HIF-1 $\alpha$  target genes *Pdk2* and *Pdk4* in murine HSCs impaired HSC transplantation capacity and resulted in increased levels of ROS and expression of senescence markers (Takubo et al., 2013). Also, knockdown of the HIF-1-inhibitor CITED2 (CBP/p300-interacting-transactivator-with-an-ED-rich-tail 2) was shown to affect HSC maintenance (Du et al., 2012; Korthuis et al., 2015; Kranc et al., 2009) and result in impaired glycolysis with elevated cellular ROS levels (Du et al., 2014). This suggests that tight regulation of HIF levels is crucial for maintaining HSC metabolism.

However, other recent studies reported that individual or combined deletion of HIF-1 and HIF-2 in HSCs does not affect their self-renewal and repopulation capacity, also in the context of serial transplantations or 5-fluorouracil treatment (Guitart et al., 2013; Vukovic et al., 2016, 2015). This disparity in results could be related to different experimental designs: Whereas *Hif-1 $\alpha$*  deletion in both murine HSCs and bone marrow environment affected HSC function (Takubo et al., 2010), *Hif-1 $\alpha$*  deletion only in the hematopoietic cells did not (Vukovic et al., 2016). In line with this explanation, a recent study concluded that stable expression of HIF proteins is less relevant for stem cell maintenance itself, but is instead important for effective mobilization of HSPCs from the bone marrow niche to the peripheral blood (Bisht et al., 2019). Moreover, other studies have shown that hypoxic conditions can also influence ROS levels and that the ROS-signaling network is in a certain manner independent of HIF (Fortenbery et al., 2018; Naranjo-Suarez et al., 2012). Therefore, HIF-mediated ROS regulation might not be fundamental for HSC functionality.

Similarly to HSCs, the role of HIF signaling for LSCs is also not yet fully understood. A recent study by Raffel et al. indicated that enhanced

activity of enzymes that target HIF-1 $\alpha$  for degradation can decrease AML stem cell maintenance (Raffel et al., 2017), thus supporting a role for HIF-1 in LSC function. Similarly, decreased expression of HIF-2 $\alpha$  and CITED2 induced by anti-diabetic drug treatment was suggested as being involved in CML-LSC elimination (Prost et al., 2015). In contrast, LSCs lacking both HIF-1 $\alpha$  and HIF-2 $\alpha$  are still capable of promoting AML development (Vukovic et al., 2015) and HIF-1 $\alpha$ -deleted leukemia cells showed an even faster disease progression after chemotherapy (Velasco-Hernandez et al., 2019). Consequently, the role of HIFs in LSCs and leukemia pathogenesis remains unclear.

## 7.2. FOXO signaling

Transcription factors of the forkhead box class O (FOXO) family are important to the oxidative defense machinery and stimulate the expression of genes coding for antioxidant proteins such as SOD, catalase and sestrin (reviewed in (Klotz et al., 2015)). FOXO proteins (including FOXO1a, FOXO3a, FOXO4 and FOXO6 in humans) are normally present in an active state in the cell nucleus, but are exported to the cytoplasm upon phosphorylation, frequently by the AKT kinase downstream of the PI3K-signaling pathway. However, other factors such as the NAD<sup>+</sup> dependent deacetylase SIRT1 can also impact the subcellular localization of FOXOs (Liang et al., 2016; Santo and Paik, 2016). The family member FOXO3a was shown to be essential for HSC maintenance, since *Foxo3a*<sup>-/-</sup> HSCs failed to support long-term reconstitution of hematopoiesis and were accompanied by increased ROS levels, activated p38 MAPK signaling and defective DNA damage repair (Bigarella et al., 2017; Miyamoto et al., 2007). FOXOs are also involved in glucose metabolism by regulating phosphoenol-pyruvate carboxykinase (PEPCK) and Glucose-6-phosphatase (G6Pase), two enzymes involved in gluconeogenesis (Zhang et al., 2018). Intriguingly, Rimmelé et al. demonstrated that elevated ROS levels observed in *Foxo3*<sup>-/-</sup> HSCs were not the result of a shift from glycolytic towards mitochondrial metabolism, nor did they have a causative role in impaired HSC functionality (Rimmelé et al., 2015). Besides high ROS levels, *Foxo3*<sup>-/-</sup> HSCs had increased glycolysis levels, decreased OXPHOS and were associated with abnormalities in mitochondrial membrane potential and mass, indicating an important role of FOXOs for mitochondrial metabolism.

A previous study showed that *in-vivo* treatment of *Foxo1/2/3*- triple knockout mice with NAC could rescue the impaired repopulation capacity of HSCs in mice transplantation studies when treatment was continued for 5 weeks (Tothova et al., 2007). However, Rimmelé et al. showed that a NAC-mediated rescue of dysfunctional *Foxo3a*<sup>-/-</sup> HSCs in repopulation studies was only short-term and was lost at 8 weeks after transplant, despite sustained lowering of ROS levels (Rimmelé et al., 2015). This indicates that elevated ROS levels in FOXO-deficient HSCs are not the main cause for their dysfunction; instead, this may reflect an abnormal mitochondrial function. This notion is supported by a study showing that FOXOs promote mitochondrial integrity by stimulating autophagy. FOXOs induce upregulation of the enzyme glutamine synthase, which consequently stimulates glutamine production (van der Vos et al., 2012). High glutamine levels block the mTOR-signaling pathway, which is a negative regulator of autophagy.

## 7.3. AMPK signaling pathway

The AMP-activated protein kinase (AMPK) is a master regulator of ROS production and elimination. Activated AMPK signaling inhibits the ROS-generating mammalian target of rapamycin (mTOR)- pathway and activates FOXO signaling, which promotes a ROS detoxifying cascade and stimulates ULK1-mediated autophagy that helps to remove damaged organelles. As its name suggests, AMPK is activated by the binding of AMP, which thus takes place in conditions where the AMP/ATP ratio is high. Upstream activators include the kinases LKB1 and CaMKK. Activation of AMPK by the diabetes drug metformin was shown to increase *ex vivo* maintenance of murine HSCs (Liu et al.,

2015). However, deletion of AMPK in murine HSC only moderately effected HSC function (Gan et al., 2010; Gurumurthy et al., 2010; Nakada et al., 2010), potentially highlighting the role of AMPK in HSC maintenance under certain stress conditions.

In contrast to its function in HSCs, AMPK seems to be more relevant for the survival of LSCs by protecting them from metabolic stress. AMPK deletion in AML LSCs was shown to result in increased ROS levels and DNA damage, as well as reduced glucose flux due to impaired glucose transporter expression, which significantly delayed leukemogenesis (Saito et al., 2015). Pei et al. (2018) showed that AMPK is intrinsically activated in LSCs and regulates LSC mitochondrial dynamics, thereby conferring LSCs with increased resistance to mitochondrial stress. Additionally, increased AMPK activation was shown to mediate resistance of AML cells to certain epigenetic agents by stimulating ULK1-mediated autophagy (Jang et al., 2017), which potentially protects cells from accumulation of ROS and oxidative damage and highlights elevated AMPK signaling as an important survival strategy of LSCs.

#### 7.4. mTOR signaling

AMPK indirectly inhibits the mTOR pathway by phosphorylating – and thereby activating – tuberous sclerosis complex 2 (TSC2), which is part of the TSC1-TSC2 complex that inhibits mTOR1. The mTOR signaling pathway is activated when the availability of nutrients and ATP is high; this pathway promotes anabolic processes like protein synthesis and mitochondrial activity, but inhibits autophagy. In opposition to AMPK, the pro-proliferative AKT pathway activates mTOR (reviewed in (Zhao et al., 2017)). Prevention of mTOR hyper-activation is essential for preservation of the HSC self-renewal capacity (Jang and Sharkis, 2007). *Tsc1* deletion in HSCs was shown to dramatically increase ROS production and to drive quiescent HSCs into rapid cycling (Chen et al., 2008), whereas treatment of cultured mouse bone marrow cells with the mTOR-inhibitor rapamycin preserved HSPCs (Huang et al., 2012). Furthermore, expression of miRNAs that target the mTOR pathway was shown to be critical for preserving long-term repopulating HSCs, while their loss resulted in enhanced mitochondrial biogenesis, metabolic activity and ROS production in HSCs (Qian et al., 2016).

mTOR promotes mitochondrial activity in multiple ways. First, it stimulates translation of mitochondria-related genes by phosphorylating eukaryotic translation initiation factor 4E (eIF4E)-binding proteins (4E-BPs), which leads to their dissociation from eIF4E and assembly of the translation initiation complex (Morita et al., 2013). Hypophosphorylation of 4E-BP is characteristic for HSCs and is important for their functionality (Signer et al., 2016). Second, mTOR stimulates activity of the transcriptional regulator PPAR $\gamma$  coactivator-1 $\alpha$  (PGC-1 $\alpha$ ), a master regulator of mitochondrial metabolism and activator of mitochondrial fatty acid oxidation genes (Cunningham et al., 2007; Hu et al., 2018). Of note, PGC-1 $\alpha$  also stimulates gluconeogenesis and is induced by stemness-associated genes such as CITED2, highlighting that different pathways can influence the impact of PGC-1 $\alpha$  signaling on ROS levels in an opposite way (Sakai et al., 2016, 2012). Finally, mTOR1 phosphorylates and inactivates the pro-autophagic kinase ULK1 (Kim et al., 2011) and thereby inhibits autophagy, resulting in the accumulation of mitochondria (Ho et al., 2017).

#### 7.5. DNA damage response pathways

Mouse models in which genes are deleted that participate in DNA damage response frequently show a similar phenotype of dysfunctional HSCs (reviewed in (Niedernhofer, 2008)). For the genes *ATM* (ataxia telangiectasia mutated) and *MLL5* (Mixed-Lineage-Leukemia-5), the phenotype of dysfunctional HSCs lacking their expression has been proposed to be causally connected to accumulation of ROS (Ito et al., 2004; Tasdogan et al., 2016). The ATM protein kinase is activated by DNA double strand breaks and modulates the activity of various targets to maintain genomic stability by initiating an effective DNA damage

response (Shiloh, 2003). These targets include numerous antioxidant enzymes that prevent increased ROS levels and oxidative stress (Barzilai et al., 2002). Ito et al. (2004) showed that increased ROS levels in *Atm*<sup>-/-</sup> HSCs led to an impaired self-renewal capacity, which could be restored by treatment with anti-oxidative agents. Mechanistically, high ROS levels in *Atm*<sup>-/-</sup> HSCs have been shown to activate the p38 MAPK pathway and upregulate p16INK4a, which is associated with induction of HSC senescence (Shao et al., 2011). A comparable observation regarding HSC dysfunctionality was made in *Mll5*<sup>-/-</sup> mice, which showed accumulation of DNA damage, ROS-mediated upregulation of p16INK4a and reversal of the phenotype when treated with the anti-oxidant NAC (Tasdogan et al., 2016). In summary, defective DNA damage response can result in increased ROS production or impaired ROS elimination, showing that DNA damage response pathways play a crucial role in redox homeostasis and HSC maintenance.

#### 7.6. SIRT1 signaling

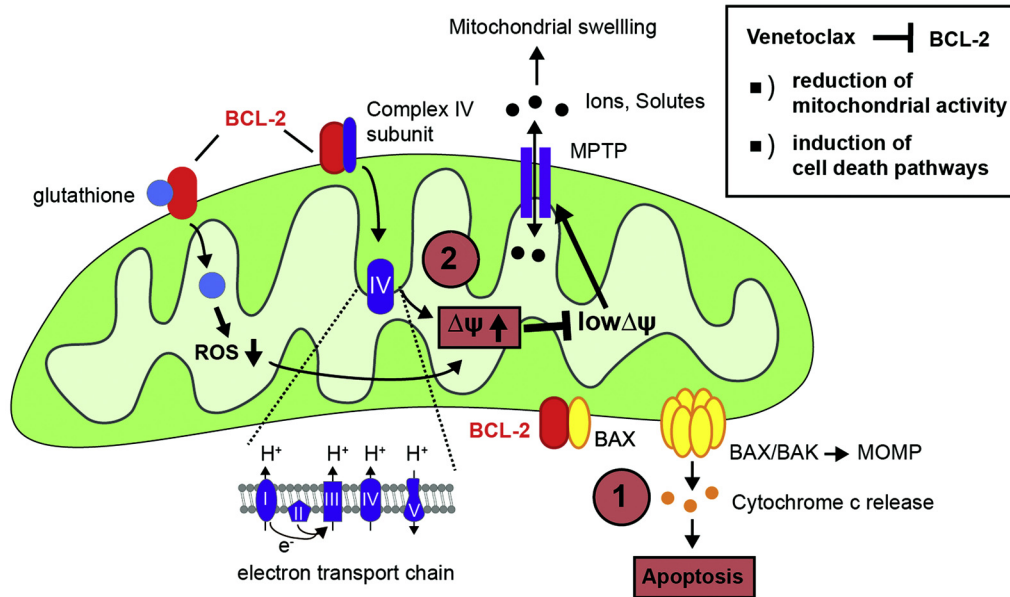
Sirtuins are a family of NAD<sup>+</sup>-dependent lysine deacetylases with seven members in mammals (SIRT1-7), of which SIRT3, SIRT4 and SIRT5 are localized exclusively within the mitochondria. The activity of sirtuins can be influenced by oxidative stress at multiple levels: ROS can induce posttranslational modification, change SIRT expression or protein-protein interactions, or affect cellular NAD levels (Santos et al., 2016). The enzymes can be viewed as metabolic sensors in a cell; they play an important role in mediating cellular stress responses or inducing metabolic changes by regulating the activity of targets such as p53, FOXOs, E2F1 or PGC-1 $\alpha$  (reviewed in (Chalkiadaki and Guarente, 2015)). Increased activation of SIRT3 eventually results in elevated expression of anti-oxidants such as SOD2 and catalase, as well as stimulation of autophagy, thereby reducing cellular ROS. Additionally, SIRT3 promote mitochondrial biogenesis and increased turnover of mitochondria, thereby stimulating removal of damaged mitochondria that otherwise would produce excessive ROS.

Several SIRT family members appear to play a crucial role in HSC functioning under stress conditions. For example, deletion of SIRT1 in HSCs has been shown to impair HSC homeostasis and to induce an aging-like phenotype by altered regulation of FOXO3 (Rimmele et al., 2014). Deletion of SIRT3 was shown not only to be indispensable for maintenance of young HSCs, but also to be essential under stress conditions or during ageing (Brown et al., 2013). SIRT6 was shown to regulate HSC function by suppressing Wnt target genes, and deletion of SIRT6 led to impaired HSC self-renewal ability (Wang et al., 2016). SIRT1 was found to be consistently overexpressed in AML (Bradbury et al., 2005), and was associated with increased leukemia cell survival, proliferation and drug resistance (Kim et al., 2015; Li and Bhatia, 2015), suggesting that SIRT1 also promotes maintenances of LSCs. Nevertheless, SIRT1 deficiency was shown to enhance – and not suppress – the maintenance of LSCs from MDS (myelodysplastic syndrome) patients by leading to decreased activity of the tumor suppressor Tet methylcytosine dioxygenase 2 (TET2) (Sun et al., 2018), indicating that SIRT1s have distinct, context-dependent functions in HSC/LSC maintenance. Furthermore, SIRT2 has been shown to be involved in metabolic reprogramming of leukemia cells by stimulating the pentose phosphate pathway (Xu et al., 2016), highlighting an important role of SIRT1s in both leukemia proliferation and maintenance via multiple routes.

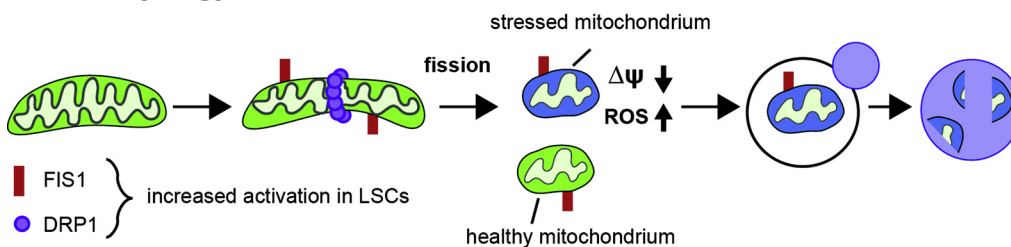
### 8. Strategies for targeting leukemia stem cells

HSCs and LSCs are both characterized by low ROS levels and impairment of various ROS-regulating pathways can influence both HSC and LSC function. However, the two populations appear to depend on different pathways to maintain their characteristic redox state or to prevent them from going into apoptosis. As mentioned earlier, LSCs rely on mitochondrial integrity and metabolism for their survival, and

## A BCL-2 functions



## B Mitophagy



mitochondrial ATP generation is crucial for leukemia progression. However, due to the altered metabolic properties of LSCs and their relatively low levels of mitochondrial membrane potential (Hao et al., 2018; Lagadinou et al., 2013; Lin et al., 2019), they are potentially more prone to undergo mitochondrial permeability transition (MPT) or apoptosis (Trotta et al., 2017). Therefore, it is likely that LSCs have an increased dependence on factors that counteract programmed cell death or protect them from stress-induced impairment (oxidative or otherwise) of mitochondrial function, which could provide a window for leukemia therapy. This concept is supported by multiple recent studies that reported increased dependency of LSCs on the anti-apoptotic BCL-2 or on stress-response mechanisms such as autophagy (Fig. 3), which are described below in more detail.

### 8.1. Compromising mitochondrial functionality of ROS-low LSCs by BCL-2 inhibition

BCL-2 can be seen as a master regulator of mitochondrial physiology and cellular stress response. In its canonical role, BCL-2 functions as an anti-apoptotic protein by preventing mitochondrial outer membrane permeabilization (MOMP) via its interaction with the pro-apoptotic proteins BAX and BAD. However, BCL-2 has also additional functions that influence mitochondrial activity and the cellular redox state by regulating both pro-oxidative and anti-oxidative processes (Chong et al., 2014; Gross and Katz, 2017). BCL-2 was shown to interact with the COX V $\alpha$ -subunit of complex IV that is part of the electron transport chain (ETC), thereby potentially facilitating the transport of this subunit to the mitochondria and increasing mitochondrial activity (Chen and Pervaiz, 2010). Furthermore, BCL-2 was found to promote the transport of the anti-oxidant glutathione to mitochondria via direct

Fig. 3. LSCs have increased dependence on BCL-2 mediated mitochondrial regulation and mitophagy. (A) BCL-2 regulates mitochondrial function mainly via 2 routes. 1: The canonical function of BCL-2 inhibits apoptosis by preventing oligomerization of the pro-apoptotic proteins BAX and BAK, which otherwise can induce mitochondrial outer membrane permeabilization (MOMP). 2: BCL-2 facilitates the import of the anti-oxidant glutathione and complex IV subunits into the mitochondria. This positively affects the mitochondrial membrane potential ( $\Delta\psi$ ) and counteracts its drop to prevent induction of the mitochondrial permeability transition pore (MPTP). (B) The processes of fission and mitophagy are tightly connected. Division of mitochondria by fission involves the GTPase DRP1 and the receptor FIS1, and enables separation of dysfunctional mitochondrial parts from healthy ones. Dysfunctional parts are further degraded by mitophagy. Interference with either BCL-2 functioning or mitophagy was shown to target LSCs.

interaction (Zimmermann et al., 2007). In line with this, high levels of BCL-2 were shown to provide increased resistance against a decline in membrane potential and MPT induction by maintaining reduced pyridine nucleotides (Kowaltowski et al., 2000). As early as 1993, BCL-2 was shown to function as an anti-oxidant (Hockenbery et al., 1993; Kane et al., 1993), and later studies demonstrated that BCL-2 overexpression shifts the redox-state of a cell to a more reduced state (Ellerby et al., 1996; Nguyen et al., 2019a). Consequently, BCL-2 also influences activation of redox-sensitive transcription factors such as NF- $\kappa$ B, p53, and AP-1, which have a crucial role in stress response. In summary, BCL-2 preserves mitochondrial stability by preventing a drop of the mitochondrial membrane potential by both stimulating mitochondrial activity and functioning as an anti-oxidant that counteracts the increased ROS production (Fig. 3A).

Campos et al. (1994) showed that BCL-2 inhibition affects the survival of leukemic stem cells and progenitor cells and improves the efficacy of chemotherapeutic drugs. More recently, several studies indicated that BCL-2 inhibition efficiently targets LSCs (Beurlet et al., 2013; Carter et al., 2016; Goff et al., 2013; Jones et al., 2018; Lagadinou et al., 2013; Pollyea et al., 2018b). Lagadinou et al. (2013) reported that LSCs – which they defined as the fraction of total mononuclear AML cells with the lowest ROS levels – are especially sensitive to BCL-2 inhibition due to their higher BCL-2 expression compared to their non-LSC counterparts (Lagadinou et al., 2013). Mechanistically, BCL-2 inhibition was found to impair OXPHOS, which is crucial for LSC survival (Lagadinou et al., 2013; Pollyea et al., 2018a). Additionally, results from our group indicate that ROS-low CD34<sup>+</sup> AML cells are more sensitive to the BCL-2 inhibitor Venetoclax (Souers et al., 2013) compared to ROS-high CD34<sup>+</sup> AML cells despite similar BCL-2 expression levels (unpublished results), highlighting their increased BCL-2



dependency. Interestingly, BCL-2 inhibition has been shown to impair the viability of leukemic cells with different backgrounds with regard to karyotype abnormalities and molecular mutations (Carter et al., 2016; Chan et al., 2015; Niu et al., 2014), indicating that LSCs have common intrinsic characteristics. Resistance to BCL-2 inhibition was observed in AMLs with high expression of the anti-apoptotic protein MCL-1, but these AMLs were shown to be targeted by combined treatment with BCL-2 and MCL-1 inhibitors (O' Reilly et al., 2018; Pan et al., 2015; Ramsey et al., 2018; Tron et al., 2018).

Paradoxically, certain leukemia-associated mutations can reinforce an increased dependency of LSCs on anti-apoptotic proteins. For instance, IDH1/2 mutants were shown to directly inhibit complex IV activity, thereby lowering the mitochondrial activity and increasing the dependency on apoptosis inhibitors such as BCL-2 in order to prevent induction of cell death pathways triggered by a very low mitochondrial membrane potential (Chan et al., 2015). A recent study aimed to identify pathways whose interference might confer increased sensitivity of AML cells to BCL-2 inhibition and identified several metabolic pathways including OXPHOS, nucleotide biosynthesis and the heme biosynthesis pathway (Lin et al., 2019). Mechanistically, heme depletion triggered impaired function of the mitochondrial ETC and caused lowering of mitochondrial membrane potential, thereby potentiating Venetoclax-induced apoptosis. This finding supports the concept that cells with low mitochondrial activity, such as LSCs, have an increased dependency on anti-apoptotic pathways. Clinical trials have shown promising results for treatment of relapsed/refractory AML when Venetoclax was used as monotherapy (Konopleva et al., 2016), as well as for newly-diagnosed, elderly AML patients when Venetoclax was used in combination with hypomethylating agents (DiNardo et al., 2019). However, Venetoclax-based treatment seems to work less efficiently in relapsed patients compared to previously untreated patients (DiNardo et al., 2018). Apparently, relapsed LSCs acquire different metabolic properties and fuel OXPHOS via different routes, for instance by up-regulation of fatty acid metabolism (Jones et al., 2018). Therefore, they seem to be less dependent on BCL-2 for maintaining sufficiently high mitochondrial activity to ensure their survival.

### 8.2. Targeting autophagy to eradicate LSCs

Autophagy is an important strategy for avoiding accumulation of damaged proteins and organelles that eventually can lead to cell death. For leukemia cells, functional autophagy was found to be crucial for leukemic transformation and for impairing cellular response to chemotherapy (reviewed in (Auberger and Puissant, 2017)). High expression of genes involved in autophagy regulation was shown to be associated with poor prognosis in AML (Nguyen et al., 2019b), and several recent studies have suggested autophagy inhibition as a potential strategy for targeting LSCs (Folkerts et al., 2017; Jang et al., 2017; Pei et al., 2018). ROS-low CD34<sup>+</sup> AMLs were shown to have higher basal levels of autophagy and increased sensitivity to autophagy compared to their ROS-high CD34<sup>+</sup> counterparts (Folkerts et al., 2017). Pei et al. (2018) identified factors that are expressed differently in ROS-low AML LSCs compared to the ROS-high non-LSC fraction and reported a crucial role of the mitophagy-related protein FIS1 (mitochondrial fission protein 1), which highlights the importance of mitochondrial dynamics for LSC maintenance (Fig. 3B). Knockdown of FIS1 was shown to severely impair the stem and progenitor potential of AML LSCs, but not of normal HSCs (Pei et al., 2018), and AML patients with a high FIS1 expression were less likely to respond to chemotherapy (Tian et al., 2014). Similarly, increased activation of the fission protein DRP1 in T-ALL cells by bone marrow-niche mediated stimuli was shown to lower ROS levels, change the mitochondrial phenotype to a more fragmented morphology and improve chemoresistance (Cai et al., 2016). Recently, it was shown that overexpression of the vacuole membrane protein (VMP1), a putative autophagy protein, results in reduced Venetoclax-sensitivity of CD34<sup>+</sup> AML cells (Folkerts et al., 2019). These data

indicate that high levels of autophagy can contribute to the characteristic features of LSCs. However, the regulation of autophagy is complex and its role in leukemia development and maintenance not yet fully understood, and might also depend on the mutational background. Inhibition of autophagy in normal HSCs was associated with accumulation of mitochondrial ROS, DNA damage and activation of the Notch signaling pathway that impairs normal HSC differentiation, which might be beneficial for AML development (Cao et al., 2015). In line with this, heterozygous loss of the key-autophagy gene *Atg5* in an MLL-ENL model for AML enhanced disease progression and aggressiveness (Watson et al., 2015). Additionally, the fusion oncoproteins PML-RARA and BCR-ABL were shown to be degraded by autophagy, and activation of autophagy is part of treatment-strategies for leukemias with these genetic alterations (Torgersen and Simonsen, 2013). In summary, these data highlight context-dependent roles of autophagy in leukemia.

### 9. Do ROS-low LSCs drive AML relapse after chemotherapy?

Chemotherapy of AML patients frequently includes treatment with the pyrimidine nucleoside analog cytarabine in combination with anthracyclines. While this strategy can lead to an initial reduction of leukemic blasts in the majority of patients, the 5-year overall survival rate of patients < 60 years has not significantly improved in recent decades and ranges from 35% to 40% (Dohner et al., 2015). Adverse outcomes of AML are mainly associated with leukemia relapse, which is assumed to be driven by LSCs that are not targeted by chemotherapy and retain their disease-initiating properties (Shlush et al., 2017). This model is supported by the increasing knowledge about characteristic ROS-low LSC properties, since features such as low metabolic activity, increased BCL-2 expression and high levels of autophagy can confer increased chemotherapy resistance as described above. Additionally, our group recently observed increased expression of the drug efflux transporter ABCB1 in the ROS-low defined LSC compartment (unpublished results), and high expression of ABC transporters in leukemia cells has been linked to poor treatment outcome in AML patients (de Jonge-Peeters et al., 2007; van der Kolk et al., 2000). Moreover, several longitudinal sequencing studies have strongly supported the concept that relapse is driven by a resistant cell population that is already present prior to chemotherapy treatment, and is not generated as a consequence of the mutagenic properties of the chemotherapeutic drugs (Bachas et al., 2012; Ding et al., 2012; Parkin et al., 2013; Shlush et al., 2017). However, it has not been shown that the ROS-low LSC population drives disease relapse. The promising results of clinical trials in which standard treatments were combined with Venetoclax are thought to be a result of efficient targeting of the ROS-low LSC population (Pollyea and Jordan, 2019), and therefore support the hypothesis that ROS-low LSCs drive disease recurrence after conventional chemotherapy. However, other studies reported that chemoresistant cells have features distinct from those of ROS-low cells: (Farge et al., 2017) showed that the AML cells that persist after chemotherapy treatment using cytarabine for 5 days have increased mitochondrial mass, mitochondrial membrane potential and ROS production – and thus have features somewhat opposite to ROS-low LSCs. Furthermore, other studies reported that treatment with cytarabine does not enrich for functional LSCs (Griessinger et al., 2014) and efficiently targets the CD34<sup>+</sup>CD38<sup>-</sup> cell population (Boyd et al., 2018). Notably, our group recently observed that ROS-low LSCs are enriched for CD34<sup>+</sup>CD38<sup>-</sup> cells (unpublished results), indicating that metabolic LSC characteristics do overlap with previously described phenotypic LSC features such as CD34/CD38 expression (Lapidot et al., 1994). These conflicting reports on the characteristics of chemoresistant cells can likely be explained by differences between steady-state LSCs and LSCs that were exposed to chemotherapy. LSCs have been shown to undergo phenotypic changes during the course of chemotherapy (Ho et al., 2016), and this might also apply to their metabolic characteristics. Consequently, LSCs with low ROS levels could be indeed the cell population that is

most resistant to chemotherapeutic drugs, but they might respond to the severe therapy-related stress with upregulation (transient or otherwise) of their mitochondrial activity.

## 10. Conclusion

Cellular ROS levels inversely correlate with the functionality of both normal stem cells and leukemic stem cells. Low ROS levels apparently reflect a metabolic state of relatively low mitochondrial activity, which coincides with the low mitochondrial membrane potential, low levels of cellular ATP and the small, fragmented mitochondria morphology found in both HSCs and LSCs. Avoiding accumulation of ROS and maintaining mitochondrial stability is essential for both types of stem cells in order to prevent ROS-mediated cell damage and induction of cell death pathways mediated by mitochondrial dysfunction. Multiple gene knockout studies have demonstrated that targeting metabolic pathways such as glycolysis and oxidative phosphorylation, or redox-regulating machineries such as autophagy and anti-oxidative enzymes, interferes with stem cell maintenance. In steady-state, however, HSCs and LSCs seem to rely on different strategies to maintain their characteristic ROS-low condition. HSCs reside in a hypoxic bone marrow niche and preferentially use glycolysis for their energy generation, which results in reduced ROS production compared to ATP production by mitochondrial oxidative phosphorylation. In contrast, LSCs rely strongly on their remaining mitochondrial activity, which they maintain through mitophagy and mechanisms dependent on the anti-apoptotic protein and master regulator of mitochondrial activity BCL-2. The strong dependency of LSCs on BCL-2 and certain key players in mitophagy seems to be a characteristic feature of LSCs, and is not observed to this extent in normal HSCs or in the total mononuclear AML cell population. Therefore, interference with those pathways is a promising strategy for eradicating LSCs, which are apparently not efficiently targeted by standard chemotherapy approaches.

An interesting question that has not been completely answered is whether the higher ROS levels observed in the total leukemia cell population compared to ROS-low LSCs have a causative role in reducing stemness potential, or if they are simply a reflection of a different metabolic state that is not compatible with stem cell function. A recent study showed that standard induction chemotherapy leads to elevation of ROS, but does not efficiently target LSCs, indicating that increased ROS levels alone are not sufficient to compromise LSC viability or function (Pollyea et al., 2018a). To improve current therapy protocols, future studies should continue to investigate the metabolic properties of LSCs and shed light on this question.

## CRedit authorship contribution statement

**Katharina Mattes:** Conceptualization, Writing - original draft. **Edo Vellenga:** Conceptualization, Writing - review & editing. **Hein Schepers:** Conceptualization, Writing - review & editing.

## Declaration of Competing Interest

The authors declare no competing financial interest.

## References

- Abbas, H.A., Maccio, D.R., Coskun, S., Jackson, J.G., Hazen, A.L., Sills, T.M., You, M.J., Hirschi, K.K., Lozano, G., 2010. Mdm2 is required for survival of hematopoietic stem cells/progenitors via dampening of ROS-induced p53 activity. *Cell Stem Cell* 7, 606–617. <https://doi.org/10.1016/j.stem.2010.09.013>.
- Aloum, L., Brimson, C.A., Zhyvoloup, A., Baines, R., Bakovic, J., Filonenko, V., Thompson, C.R.L., Gout, I., 2019. Coenzyme A and protein CoAlation levels are regulated in response to oxidative stress and during morphogenesis in *Dictyostelium discoideum*. *Biochem. Biophys. Res. Commun.* 511, 294–299. <https://doi.org/10.1016/j.bbrc.2019.02.031>.
- Anso, E., Weinberg, S.E., Diebold, L.P., Thompson, B.J., Malinge, S., Schumacker, P.T., Liu, X., Zhang, Y., Shao, Z., Steadman, M., Marsh, K.M., Xu, J., Crispino, J.D., Chandel, N.S., 2017. The mitochondrial respiratory chain is essential for haematopoietic stem cell function. *Nat. Cell Biol.* 19, 614–625. <https://doi.org/10.1038/ncb3529>.
- Auberger, P., Puissant, A., 2017. Autophagy, a key mechanism of oncogenesis and resistance in leukemia. *Blood* 129, 547–552. <https://doi.org/10.1182/blood-2016-07-692707>.
- Bachas, C., Schuurhuis, G.J., Assaraf, Y.G., Kwidama, Z.J., Kelder, A., Wouters, F., Snel, A.N., Kaspers, G.J.L., Cloos, J., 2012. The role of minor subpopulations within the leukemic blast compartment of AML patients at initial diagnosis in the development of relapse. *Leukemia* 26, 1313–1320. <https://doi.org/10.1038/leu.2011.383>.
- Bagger, F.O., Rapin, N., Theilgaard-Monch, K., Kaczkowski, B., Thoren, L.A., Jendholm, J., Winther, O., Porse, B.T., 2013. HemaExplorer: a database of mRNA expression profiles in normal and malignant haematopoiesis. *Nucleic Acids Res.* 41, D1034–9. <https://doi.org/10.1093/nar/gks1021>.
- Barzilai, A., Rotman, G., Shiloh, Y., 2002. ATM deficiency and oxidative stress: a new dimension of defective response to DNA damage. *DNA Repair (Amst.)* 1, 3–25.
- Bejarano-Garcia, J.A., Millan-Ucles, A., Rosado, I.V., Sanchez-Abarca, L.L., Caballero-Velazquez, T., Duran-Galvan, M.J., Perez-Simon, J.A., Piruat, J.I., 2016. Sensitivity of hematopoietic stem cells to mitochondrial dysfunction by Sdhb gene deletion. *Cell Death Dis.* 7, e2516. <https://doi.org/10.1038/cddis.2016.411>.
- Beurlet, S., Omidvar, N., Gorombe, P., Krief, P., Le Pogam, C., Setterblad, N., de la Grange, P., Leboeuf, C., Janin, A., Noguera, M.-E., Hervatin, F., Sarda-Mantel, L., Konopleva, M., Andreeff, M., Tu, A.W., Fan, A.C., Felsler, D.W., Whetton, A., Pla, M., West, R., Fenaux, P., Chomienne, C., Padua, R.A., 2013. BCL-2 inhibition with ABT-737 prolongs survival in an NRAS/BCL-2 mouse model of AML by targeting primitive LSK and progenitor cells. *Blood* 122, 2864–2876. <https://doi.org/10.1182/blood-2012-07-445635>.
- Bigarella, C.L., Li, J., Rimmel, P., Liang, R., Sobol, R.W., Ghaffari, S., 2017. FOXO3 transcription factor is essential for protecting hematopoietic stem and progenitor cells from oxidative DNA damage. *J. Biol. Chem.* 292, 3005–3015. <https://doi.org/10.1074/jbc.M116.769455>.
- Bigarella, C.L., Liang, R., Ghaffari, S., 2014. Stem cells and the impact of ROS signaling. *Development* 141, 4206–4218. <https://doi.org/10.1242/dev.107086>.
- Bisht, K., Brunck, M.E., Matsumoto, T., McGirr, C., Nowlan, B., Fleming, W., Keech, T., Magor, G., Perkins, A.C., Davies, J., Walkinshaw, G., Flippin, L., Winkler, I.G., Levesque, J.-P., 2019. HIF prolyl hydroxylase inhibitor FG-4497 enhances mouse hematopoietic stem cell mobilization via VEGFR2/KDR. *Blood Adv.* 3, 406–418. <https://doi.org/10.1182/bloodadvances.2018017566>.
- Boyd, A.L., Aslostovar, L., Reid, J., Ye, W., Tanasijevic, B., Porras, D.P., Shapovalova, Z., Almakadi, M., Foley, R., Leber, B., Xenocostas, A., Bhatia, M., 2018. Identification of chemotherapy-induced leukemic-regenerating cells reveals a transient vulnerability of human AML recurrence. *Cancer Cell* 34, 483–498. <https://doi.org/10.1016/j.ccell.2018.08.007>. e5.
- Bradbury, C.A., Khanim, F.L., Hayden, R., Bunce, C.M., White, D.A., Drayson, M.T., Craddock, C., Turner, B.M., 2005. Histone deacetylases in acute myeloid leukaemia show a distinctive pattern of expression that changes selectively in response to deacetylase inhibitors. *Leukemia* 19, 1751–1759. <https://doi.org/10.1038/sj.leu.2403910>.
- Brown, K., Xie, S., Qiu, X., Mohrin, M., Shin, J., Liu, Y., Zhang, D., Scadden, D.T., Chen, D., 2013. SIRT3 reverses aging-associated degeneration. *Cell Rep.* 3, 319–327. <https://doi.org/10.1016/j.celrep.2013.01.005>.
- Cai, J., Wang, J., Huang, Y., Wu, H., Xia, T., Xiao, J., Chen, X., Li, H., Qiu, Y., Wang, Y., Wang, T., Xia, H., Zhang, Q., Xiang, A.P., 2016. ERK/Drp1-dependent mitochondrial fission is involved in the MSC-induced drug resistance of T-cell acute lymphoblastic leukemia cells. *Cell Death Dis.* 7, e2459. <https://doi.org/10.1038/cddis.2016.370>.
- Campos, L., Sabido, O., Rouault, J.P., Guyotat, D., 1994. Effects of BCL-2 antisense oligodeoxynucleotides on *in vitro* proliferation and survival of normal marrow progenitors and leukemic cells. *Blood* 84, 595–600.
- Cao, Y., Cai, J., Zhang, S., Yuan, N., Fang, Y., Wang, Zhijian, Li, X., Cao, D., Xu, F., Lin, W., Song, L., Wang, Zhen, Wang, Jian, Xu, X., Zhang, Y., Zhao, W., Hu, S., Zhang, X., Wang, Jianrong, 2015. Autophagy sustains hematopoiesis through targeting notch. *Stem Cells Dev.* 24, 2660–2673. <https://doi.org/10.1089/scd.2015.0176>.
- Capala, M.E., Pruis, M., Vellenga, E., Schuringa, J.J., 2016. Depletion of SAMS50 specifically targets BCR-ABL-Expressing leukemic stem and progenitor cells by interfering with mitochondrial functions. *Stem Cells Dev.* 25, 427–437. <https://doi.org/10.1089/scd.2015.0151>.
- Carrelha, J., Meng, Y., Kettle, L.M., Luis, T.C., Norfo, R., Alcolea, V., Boukarabila, H., Grasso, F., Gambardella, A., Grover, A., Hogstrand, K., Lord, A.M., Sanjuan-Pla, A., Woll, P.S., Nerlov, C., Jacobsen, S.E.W., 2018. Hierarchically related lineage-restricted fates of multipotent haematopoietic stem cells. *Nature* 554, 106–111. <https://doi.org/10.1038/nature25455>.
- Carter, B.Z., Mak, P.Y., Mu, H., Zhou, H., Mak, D.H., Schober, W., Levenson, J.D., Zhang, B., Bhatia, R., Huang, X., Cortes, J., Kantarjian, H., Konopleva, M., Andreeff, M., 2016. Combined targeting of BCL-2 and BCR-ABL tyrosine kinase eradicates chronic myeloid leukemia stem cells. *Sci. Transl. Med.* 8, 355ra117. <https://doi.org/10.1126/scitranslmed.aag1180>.
- Chalkiadaki, A., Guarente, L., 2015. The multifaceted functions of sirtuins in cancer. *Nat. Rev. Cancer* 15, 608–624. <https://doi.org/10.1038/nrc3985>.
- Chan, S.M., Thomas, D., Corces-Zimmerman, M.R., Xavy, S., Rastogi, S., Hong, W.-J., Zhao, F., Medeiros, B.C., Tyvoll, D.A., Majeti, R., 2015. Isocitrate dehydrogenase 1 and 2 mutations induce BCL-2 dependence in acute myeloid leukemia. *Nat. Med.* 21, 178–184. <https://doi.org/10.1038/nm.3788>.
- Chen, C., Liu, Yu, Liu, R., Ikenoue, T., Guan, K.-L., Liu, Yang, Zheng, P., 2008. TSC-mTOR maintains quiescence and function of hematopoietic stem cells by repressing mitochondrial biogenesis and reactive oxygen species. *J. Exp. Med.* 205, 2397–2408. <https://doi.org/10.1084/jem.20081297>.

- Chen, Z.X., Pervaiz, S., 2010. Involvement of cytochrome c oxidase subunits Va and Vb in the regulation of cancer cell metabolism by Bcl-2. *Cell Death Differ.* 17, 408–420. <https://doi.org/10.1038/cdd.2009.132>.
- Chong, S.J.F., Low, I.C.C., Pervaiz, S., 2014. Mitochondrial ROS and involvement of Bcl-2 as a mitochondrial ROS regulator. *Mitochondrion* 19 (Pt A), 39–48. <https://doi.org/10.1016/j.mito.2014.06.002>.
- Cline, S.D., 2012. Mitochondrial DNA damage and its consequences for mitochondrial gene expression. *Biochim. Biophys. Acta* 1819, 979–991. <https://doi.org/10.1016/j.bbgrm.2012.06.002>.
- Cole, A., Wang, Z., Coyaud, E., Voisin, V., Gronda, M., Jitkova, Y., Mattson, R., Hurren, R., Babovic, S., Maclean, N., Restall, I., Wang, X., Jeyaraju, D.V., Sukhai, M.A., Prabha, S., Bashir, S., Ramakrishnan, A., Leung, E., Qia, Y.H., Zhang, N., Combes, K.R., Ketela, T., Lin, F., Houry, W.A., Aman, A., Al-Awar, R., Zheng, W., Wienholds, E., Xu, C.J., Dick, J., Wang, J.C.Y., Moffat, J., Minden, M.D., Eaves, C.J., Bader, G.D., Hao, Z.E., Kornblau, S.M., Raught, B., Schimmer, A.D., 2015. Inhibition of the mitochondrial protease ClpP as a therapeutic strategy for human acute myeloid leukemia. *Cancer Cell* 27, 864–876. <https://doi.org/10.1016/j.ccell.2015.05.004>.
- Cui, L., Cheng, Z., Liu, Y., Dai, Y., Pang, Y., Jiao, Y., Ke, X., Cui, W., Zhang, Q., Shi, J., Fu, L., 2018. Overexpression of PDK2 and PDK3 reflects poor prognosis in acute myeloid leukemia. *Cancer Gene Ther.* <https://doi.org/10.1038/s41417-018-0071-9>.
- Cunningham, J.T., Rodgers, J.T., Arlow, D.H., Vazquez, F., Mootha, V.K., Puigserver, P., 2007. mTOR controls mitochondrial oxidative function through a YY1-PGC-1alpha transcriptional complex. *Nature* 450, 736–740. <https://doi.org/10.1038/nature06322>.
- de Almeida, M.J., Luchsinger, L.L., Corrigan, D.J., Williams, L.J., Snoeck, H.-W., 2017. Dye-independent methods reveal elevated mitochondrial mass in hematopoietic stem cells. *Cell Stem Cell* 21, 725–729. <https://doi.org/10.1016/j.stem.2017.11.002> e4.
- de Jonge-Peeters, S.D.P.W.M., Kuipers, F., de Vries, E.G.E., Vellenga, E., 2007. ABC transporter expression in hematopoietic stem cells and the role in AML drug resistance. *Crit. Rev. Oncol. Hematol.* 62, 214–226. <https://doi.org/10.1016/j.critrevonc.2007.02.003>.
- DiNardo, C.D., Pratz, K., Pullarkat, V., Jonas, B.A., Arellano, M., Becker, P.S., Frankfurt, O., Konopleva, M., Wei, A.H., Kantarjian, H.M., Xu, T., Hong, W.-J., Chyla, B., Potluri, J., Pollyea, D.A., Letai, A., 2019. Venetoclax combined with decitabine or azacitidine in treatment-naïve, elderly patients with acute myeloid leukemia. *Blood* 133, 7–17. <https://doi.org/10.1182/blood-2018-08-868752>.
- DiNardo, C.D., Rausch, C.R., Benton, C., Kadia, T., Jain, N., Pemmaraju, N., Daver, N., Covert, W., Marx, K.R., Mace, M., Jabbour, E., Cortes, J., Garcia-Manero, G., Ravandi, F., Bhalla, K.N., Kantarjian, H., Konopleva, M., 2018. Clinical experience with the BCL2-inhibitor venetoclax in combination therapy for relapsed and refractory acute myeloid leukemia and related myeloid malignancies. *Am. J. Hematol.* 93, 401–407. <https://doi.org/10.1002/ajh.25000>.
- Ding, L., Ley, T.J., Larson, D.E., Miller, C.A., Koboldt, D.C., Welch, J.S., Ritchey, J.K., Young, M.A., Lamprecht, T., McLellan, M.D., McMichael, J.F., Wallis, J.W., Lu, C., Shen, D., Harris, C.C., Dooling, D.J., Fulton, R.S., Fulton, L.L., Chen, K., Schmidt, H., Kalicki-veizer, J., Magrini, V.J., Cook, L., McGrath, S.D., Vickery, T.L., Wendt, M.C., Heath, S., Watson, M.A., Link, D.C., Tomasson, M.H., Shannon, W.D., Payton, J.E., Kulkarni, S., Westervelt, P., Walter, M.J., Graubert, T.A., Mardis, E.R., Wilson, R.K., DiPersio, J.F., 2012. Clonal evolution in relapsed acute myeloid leukaemia revealed by whole-genome sequencing. *Nature* 481, 506–510. <https://doi.org/10.1038/nature10738>.
- Dohner, H., Weisdorf, D.J., Bloomfield, C.D., 2015. Acute myeloid leukemia. *N. Engl. J. Med.* 373, 1136–1152. <https://doi.org/10.1056/NEJMra1406184>.
- Du, J., Chen, Y., Li, Q., Han, X., Cheng, C., Wang, Z., Danielpour, D., Dunwoodie, S.L., Bunting, K.D., Yang, Y.-C., 2012. HIF-1alpha deletion partially rescues defects of hematopoietic stem cell quiescence caused by Cited2 deficiency. *Blood* 119, 2789–2798. <https://doi.org/10.1182/blood-2011-10-387902>.
- Du, J., Li, Q., Tang, F., Puchowitz, M.A., Fujioka, H., Dunwoodie, S.L., Danielpour, D., Yang, Y.-C., 2014. Cited2 is required for the maintenance of glycolytic metabolism in adult hematopoietic stem cells. *Stem Cells Dev.* 23, 83–94. <https://doi.org/10.1089/scd.2013.0370>.
- Ellerby, L.M., Ellerby, H.M., Park, S.M., Holleran, A.L., Murphy, A.N., Fiskum, G., Kane, D.J., Testa, M.P., Kayalar, C., Bredesen, D.E., 1996. Shift of the cellular oxidation-reduction potential in neural cells expressing Bcl-2. *J. Neurochem.* 67, 1259–1267.
- Eppert, K., Takenaka, K., Lechman, E.R., Waldron, L., Nilsson, B., van Galen, P., Metzeler, K.H., Poepl, A., Ling, V., Beyene, J., Canty, A.J., Danska, J.S., Bohlander, S.K., Buske, C., Minden, M.D., Golub, T.R., Jurisica, I., Ebert, B.L., Dick, J.E., 2011. Stem cell gene expression programs influencing clinical outcome in human leukemia. *Nat. Med.* 17, 1086–1093. <https://doi.org/10.1038/nm.2415>.
- Facucho-Oliveira, J.M., St John, J.C., 2009. The relationship between pluripotency and mitochondrial DNA proliferation during early embryo development and embryonic stem cell differentiation. *Stem Cell Rev.* 5, 140–158. <https://doi.org/10.1007/s12015-009-9058-0>.
- Farge, T., Saland, E., de Toni, F., Aroua, N., Hosseini, M., Perry, R., Bosc, C., Sugita, M., Stuani, L., Fraisse, M., Scotland, S., Larrue, C., Boutzen, H., Feliu, V., Nicolau-Travers, M.-L., Cassant-Sourdy, S., Broin, N., David, M., Serhan, N., Sarry, A., Tavitian, S., Kaoma, T., Vallar, L., Iacovoni, J., Linares, L.K., Montersino, C., Castellano, R., Griessinger, E., Collette, Y., Duchamp, O., Barreira, Y., Hirsch, P., Palama, T., Gales, L., Delhommeau, F., Garmy-Susini, B.H., Portais, J.-C., Vergez, F., Selak, M., Danet-Desnoyers, G., Carroll, M., Recher, C., Sarry, J.-E., 2017. Chemotherapy-resistant human acute myeloid leukemia cells are not enriched for leukemic stem cells but require oxidative metabolism. *Cancer Discov.* 7, 716–735. <https://doi.org/10.1158/2159-8290.CD-16-0441>.
- Folkerts, H., Hilgendorf, S., Wierenga, A.T.J., Jaques, J., Mulder, A.B., Coffey, P.J., Schuringa, J.J., Vellenga, E., 2017. Inhibition of autophagy as a treatment strategy for p53 wild-type acute myeloid leukemia. *Cell Death Dis.* 8, e2927. <https://doi.org/10.1038/cddis.2017.317>.
- Folkerts, H., Wierenga, A.T., van den Heuvel, F.A., Woldhuis, R.R., Kluit, D.S., Jaques, J., Schuringa, J.J., Vellenga, E., 2019. Elevated VMP1 expression in acute myeloid leukemia amplifies autophagy and is protective against venetoclax-induced apoptosis. *Cell Death Dis.* 10, 421. <https://doi.org/10.1038/s41419-019-1648-4>.
- Fortenberry, G.W., Sarathy, B., Carraway, K.R., Mansfield, K.D., 2018. Hypoxic stabilization of mRNA is HIF-independent but requires mtROS. *Cell. Mol. Biol. Lett.* 23, 48. <https://doi.org/10.1186/s11658-018-0112-2>.
- Gan, B., Hu, J., Jiang, S., Liu, Y., Sahin, E., Zhuang, L., Fletcher-Sananikone, E., Colla, S., Wang, Y.A., Chin, L., Depinho, R.A., 2010. Lkb1 regulates quiescence and metabolic homeostasis of haematopoietic stem cells. *Nature* 468, 701–704. <https://doi.org/10.1038/nature09595>.
- Gao, M., Yi, J., Zhu, J., Minikes, A.M., Monian, P., Thompson, C.B., Jiang, X., 2019. Role of mitochondria in Ferroptosis. *Mol. Cell* 73, 354–363. <https://doi.org/10.1016/j.molcel.2018.10.042> e3.
- Gezer, D., Vukovic, M., Soga, T., Pollard, P.J., Kranc, K.R., 2014. Concise Review: Genetic Dissection of Hypoxia Signaling Pathways in Normal and Leukemic Stem Cells. *Stem Cells* 32, 1390–1397. <https://doi.org/10.1002/stem.1657>.
- Goff, D.J., Court Recart, A., Sadarangani, A., Chun, H.-J., Barrett, C.L., Krajewska, M., Leu, H., et al., 2013. A Pan-BCL2 inhibitor renders bone-marrow-resident human leukemia stem cells sensitive to tyrosine kinase inhibition. *Cell Stem Cell* 12, 316–328. <https://doi.org/10.1016/j.stem.2012.12.011>.
- Golan, K., Kumari, A., Kollet, O., Khatib-Massalha, E., Subramaniam, M.D., Ferreira, Z.S., Avemaria, F., Rzeszotek, S., Garcia-Garcia, A., Xie, S., Flores-Figueroa, E., Gur-Cohen, S., Itkin, T., Ludin-Tal, A., Massalha, H., Bernshtein, B., Ciechanowicz, A.K., Brandis, A., Mehlman, T., Bhattacharya, S., Bertagna, M., Cheng, H., Petrovich-Kopitman, E., Janus, T., Kaushansky, N., Cheng, T., Sagi, I., Ratajczak, M.Z., Mendez-Ferrer, S., Dick, J.E., Markus, R.P., Lapidot, T., 2018. Daily onset of light and darkness differentially controls hematopoietic stem cell differentiation and maintenance. *Cell Stem Cell* 23, 572–585. <https://doi.org/10.1016/j.stem.2018.08.002> e7.
- Gomez-Puerto, M.C., Folkerts, H., Wierenga, A.T.J., Schepers, K., Schuringa, J.J., Coffey, P.J., Vellenga, E., 2016. Autophagy proteins ATG5 and ATG7 are essential for the maintenance of human CD34(+) hematopoietic stem-progenitor cells. *Stem Cells* 34, 1651–1663. <https://doi.org/10.1002/stem.2347>.
- Goto, M., Miwa, H., Shikami, M., Tsunekawa-Imai, N., Suganuma, K., Mizuno, S., Takahashi, M., Mizutani, M., Hanamura, I., Nitta, M., 2014. Importance of glutamine metabolism in leukemia cells by energy production through TCA cycle and by redox homeostasis. *Cancer Invest.* 32, 241–247. <https://doi.org/10.3109/07357907.2014.907419>.
- Griessinger, E., Anjos-Afonso, F., Pizzitola, I., Rouault-Pierre, K., Vargaftig, J., Taussig, D., Gribben, J., Lassailly, F., Bonnet, D., 2014. A niche-like culture system allowing the maintenance of primary human acute myeloid leukemia-initiating cells: a new tool to decipher their chemoresistance and self-renewal mechanisms. *Stem Cells Transl. Med.* 3, 520–529. <https://doi.org/10.5966/sctm.2013-0166>.
- Gross, A., Katz, S.G., 2017. Non-apoptotic functions of BCL-2 family proteins. *Cell Death Differ.* 24, 1348–1358. <https://doi.org/10.1038/cdd.2017.22>.
- Guitart, A.V., Subramani, C., Armesilla-Diaz, A., Smith, G., Sepulveda, C., Gezer, D., Vukovic, M., Dunn, K., Pollard, P., Holyoake, T.L., Enver, T., Ratcliffe, P.J., Kranc, K.R., 2013. Hif-2alpha is not essential for cell-autonomous hematopoietic stem cell maintenance. *Blood* 122, 1741–1745. <https://doi.org/10.1182/blood-2013-02-484923>.
- Gurumurthy, S., Xie, S.Z., Alagesan, B., Kim, J., Yusuf, R.Z., Saez, B., Tzatsos, A., Ozsolak, F., Milos, P., Ferrari, F., Park, P.J., Shirihai, O.S., Scadden, D.T., Bardeesy, N., 2010. The Lkb1 metabolic sensor maintains haematopoietic stem cell survival. *Nature* 468, 659–663. <https://doi.org/10.1038/nature09572>.
- Guzman, M.L., Rossi, R.M., Karnischky, L., Li, X., Peterson, D.R., Howard, D.S., Jordan, C.T., 2005. The sesquiterpene lactone parthenolide induces apoptosis of human acute myelogenous leukemia stem and progenitor cells. *Blood* 105, 4163–4169. <https://doi.org/10.1182/blood-2004-10-4135>.
- Haas, S., Hansson, J., Klimmeck, D., Loeffler, D., Velten, L., Uckelmann, H., Wurzer, S., Prendergast, A.M., Schnell, A., Hexel, K., Santarella-Mellwig, R., Blaszkiewicz, S., Kuck, A., Geiger, H., Milsom, M.D., Steinmetz, L.M., Schroeder, T., Trumpp, A., Krijgsvelde, J., Essers, M.A.G., 2015. Inflammation-induced emergency megakaryopoiesis driven by hematopoietic stem cell-like megakaryocyte progenitors. *Cell Stem Cell* 17, 422–434. <https://doi.org/10.1016/j.stem.2015.07.007>.
- Hao, X., Gu, H., Chen, C., Huang, D., Zhao, Y., Xie, L., Zou, Y., Shu, H.S., Zhang, Y., He, X., Lai, X., Zhang, X., Zhou, B.O., Zhang, C.C., Chen, G.-Q., Yu, Z., Yang, Y., Zheng, J., 2018. Metabolic imaging reveals a unique preference of symmetric cell division and homing of leukemia-initiating cells in an endosteal niche. *Cell Metab.* <https://doi.org/10.1016/j.cmet.2018.11.013>.
- Ho, T.-C., LaMere, M., Stevens, B.M., Ashton, J.M., Myers, J.R., O'Dwyer, K.M., Liesveld, J.L., Mender, J.H., Guzman, M., Morrisette, J.D., Zhao, J., Wang, E.S., Wetzler, M., Jordan, C.T., Becker, M.W., 2016. Evolution of acute myelogenous leukemia stem cell properties after treatment and progression. *Blood* 128, 1671–1678. <https://doi.org/10.1182/blood-2016-02-695312>.
- Ho, T.T., Warr, M.R., Adelman, E.R., Lansinger, O.M., Flach, J., Verovskaya, E.V., Figueroa, M.E., Passegue, E., 2017. Autophagy maintains the metabolism and function of young and old stem cells. *Nature* 543, 205–210. <https://doi.org/10.1038/nature21388>.
- Hockenbery, D.M., Oltvai, Z.N., Yin, X.M., Millman, C.L., Korsmeyer, S.J., 1993. Bcl-2 functions in an antioxidant pathway to prevent apoptosis. *Cell* 75, 241–251.
- Hu, M., Zeng, H., Chen, S., Xu, Y., Wang, S., Tang, Y., Wang, X., Du, C., Shen, M., Chen, F., Chen, M., Wang, C., Gao, J., Wang, F., Su, Y., Wang, J., 2018. SRC-3 is involved in maintaining hematopoietic stem cell quiescence by regulation of mitochondrial metabolism in mice. *Blood* 132, 911–923. <https://doi.org/10.1182/blood-2018-02-831669>.

- Huang, J., Nguyen-McCarty, M., Hexner, E.O., Danet-Desnoyers, G., Klein, P.S., 2012. Maintenance of hematopoietic stem cells through regulation of Wnt and mTOR pathways. *Nat. Med.* 18, 1778–1785. <https://doi.org/10.1038/nm.2984>.
- Shida, T., Suzuki, S., Lai, C.-Y., Yamazaki, S., Kakuta, S., Iwakura, Y., Nojima, M., Takeuchi, Y., Higashihara, M., Nakauchi, H., Otsu, M., 2017. Pre-Transplantation Blockade of TNF- $\alpha$ -Mediated Oxygen Species Accumulation Protects Hematopoietic Stem Cells. *Stem Cells* 35, 989–1002. <https://doi.org/10.1002/stem.2524>.
- Ito, K., Hirao, A., Arai, F., Matsuoka, S., Takubo, K., Hamaguchi, I., Nomiya, K., Hosokawa, K., Sakurada, K., Nakagata, N., Ikeda, Y., Mak, T.W., Suda, T., 2004. Regulation of oxidative stress by ATM is required for self-renewal of haematopoietic stem cells. *Nature* 431, 997–1002. <https://doi.org/10.1038/nature02989>.
- Ito, K., Hirao, A., Arai, F., Takubo, K., Matsuoka, S., Miyamoto, K., Ohmura, M., Naka, K., Hosokawa, K., Ikeda, Y., Suda, T., 2006. Reactive oxygen species act through p38 MAPK to limit the lifespan of hematopoietic stem cells. *Nat. Med.* 12, 446–451. <https://doi.org/10.1038/nm1388>.
- Ito, K., Turcotte, R., Cui, J., Zimmerman, S.E., Pinho, S., Mizoguchi, T., Arai, F., Runnels, J.M., Alt, C., Teruya-Feldstein, J., Mar, J.C., Singh, R., Suda, T., Lin, C.P., Frenette, P.S., Ito, Keisuke, 2016. Self-renewal of a purified Tie2+ hematopoietic stem cell population relies on mitochondrial clearance. *Science* 354, 1156–1160. <https://doi.org/10.1126/science.aaf5530>.
- Ivan, M., Kondo, K., Yang, H., Kim, W., Valiano, J., Ohh, M., Salic, A., Asara, J.M., Lane, W.S., Kaelin, W.G.J., 2001. HIF1 $\alpha$  targeted for VHL-mediated destruction by proline hydroxylation: implications for O<sub>2</sub> sensing. *Science* 292, 464–468. <https://doi.org/10.1126/science.1059817>.
- Jang, J.E., Eom, J.-I., Jeung, H.-K., Cheong, J.-W., Lee, J.Y., Kim, J.S., Min, Y.H., 2017. AMPK-ULK1-Mediated autophagy confers resistance to BET inhibitor JQ1 in acute myeloid leukemia stem cells. *Clin. Cancer Res.* 23, 2781–2794. <https://doi.org/10.1158/1078-0432.CCR-16-1903>.
- Jang, Y.-Y., Sharkis, S.J., 2007. A low level of reactive oxygen species selects for primitive hematopoietic stem cells that may reside in the low-oxygenic niche. *Blood* 110, 3056–3063. <https://doi.org/10.1182/blood-2007-05-087759>.
- Jones, C.L., Stevens, B.M., D'Alessandro, A., Reisz, J.A., Culp-Hill, R., Nemkov, T., Pei, S., Khan, N., Adane, B., Ye, H., Krug, A., Reinhold, D., Smith, C., DeGregori, J., Pollyea, D.A., Jordan, C.T., 2018. Inhibition of amino acid metabolism selectively targets human leukemia stem cells. *Cancer Cell* 34, 724–740. <https://doi.org/10.1016/j.ccell.2018.10.005>.
- Jordan, C.T., 2007. The leukemic stem cell. *Best Pract. Res. Clin. Haematol.* 20, 13–18. <https://doi.org/10.1016/j.beha.2006.10.005>.
- Joshi, A., Kundu, M., 2013. Mitophagy in hematopoietic stem cells: the case for exploration. *Autophagy* 9, 1737–1749. <https://doi.org/10.4161/auto.26681>.
- Jung, H., Kim, D.O., Byun, J.-E., Kim, W.S., Kim, M.J., Song, H.Y., Kim, Y.K., Kang, D.-K., Park, Y.-J., Kim, T.-D., Yoon, S.R., Lee, H.G., Choi, E.-J., Min, S.-H., Choi, I., 2016. Thioredoxin-interacting protein regulates haematopoietic stem cell ageing and rejuvenation by inhibiting p38 kinase activity. *Nat. Commun.* 7, 13674. <https://doi.org/10.1038/ncomms13674>.
- Jung, H., Kim, M.J., Kim, D.O., Kim, W.S., Yoon, S.-J., Park, Y.-J., Yoon, S.R., Kim, T.-D., Suh, H.-W., Yun, S., Min, J.-K., Lee, H.G., Lee, Y.H., Na, H.-J., Lee, D.C., Kim, H.-C., Choi, I., 2013. TXNIP maintains the hematopoietic cell pool by switching the function of p53 under oxidative stress. *Cell Metab.* 18, 75–85. <https://doi.org/10.1016/j.cmet.2013.06.002>.
- Kane, D.J., Sarafian, T.A., Anton, R., Hahn, H., Gralla, E.B., Valentine, J.S., Ord, T., Bredesen, D.E., 1993. Bcl-2 inhibition of neural death: decreased generation of reactive oxygen species. *Science* 262, 1274–1277.
- Karigane, D., Kobayashi, H., Morikawa, T., Ootomo, Y., Sakai, M., Nagamatsu, G., Kubota, Y., Goda, N., Matsumoto, M., Nishimura, E.K., Soga, T., Otsu, K., Suematsu, M., Okamoto, S., Suda, T., Takubo, K., 2016. p38 $\alpha$  activates purine metabolism to initiate hematopoietic Stem/Progenitor cell cycling in response to stress. *Cell Stem Cell* 19, 192–204. <https://doi.org/10.1016/j.stem.2016.05.013>.
- Karigane, D., Takubo, K., 2017. Metabolic regulation of hematopoietic and leukemic stem/progenitor cells under homeostatic and stress conditions. *Int. J. Hematol.* 106, 18–26. <https://doi.org/10.1007/s12185-017-2261-x>.
- Khan, N., Hills, R.K., Knapper, S., Steadman, L., Qureshi, U., Rector, J.L., Bradbury, C., Russell, N.H., Vyas, P., Burnett, A.K., Grimwade, D., Hole, P.S., Freeman, S.D., 2016. Normal hematopoietic progenitor subsets have distinct reactive oxygen species, BCL2 and cell-cycle profiles that are decoupled from maturation in acute myeloid leukemia. *PLoS One* 11, e0163291. <https://doi.org/10.1371/journal.pone.0163291>.
- Kim, H.-B., Lee, S.-H., Um, J.-H., Kim, M.-J., Hyun, S.-K., Gong, E.-J., Oh, W.K., Kang, C.-D., Kim, S.-H., 2015. Sensitization of chemo-resistant human chronic myeloid leukemia stem-like cells to Hsp90 inhibitor by SIRT1 inhibition. *Int. J. Biol. Sci.* 11, 923–934. <https://doi.org/10.7150/ijbs.10896>.
- Kim, J., Kundu, M., Viollet, B., Guan, K.-L., 2011. AMPK and mTOR regulate autophagy through direct phosphorylation of Ulk1. *Nat. Cell Biol.* 13, 132–141. <https://doi.org/10.1038/ncb2152>.
- Klotz, L.-O., Sanchez-Ramos, C., Prieto-Arroyo, I., Urbanek, P., Steinbrenner, H., Monsalve, M., 2015. Redox regulation of FoxO transcription factors. *Redox Biol.* 6, 51–72. <https://doi.org/10.1016/j.redox.2015.06.019>.
- Kocabas, F., Zheng, J., Thet, S., Copeland, N.G., Jenkins, N.A., DeBerardinis, R.J., Zhang, C., Sadek, H.A., 2012. Meis1 regulates the metabolic phenotype and oxidant defense of hematopoietic stem cells. *Blood* 120, 4963–4972. <https://doi.org/10.1182/blood-2012-05-432260>.
- Konopleva, M., Pollyea, D.A., Potluri, J., Chyla, B., Hogdal, L., Busman, T., McKeegan, E., Saleem, A.H., Zhu, M., Ricker, J.L., Blum, W., DiNardo, C.D., Kadia, T., Dunbar, M., Kirby, R., Falotico, N., Levenson, J., Humerickhouse, R., Mabry, M., Stone, R., Kantarjian, H., Letai, A., 2016. Efficacy and biological correlates of response in a phase II study of venetoclax monotherapy in patients with acute myelogenous leukemia. *Cancer Discov.* 6, 1106–1117. <https://doi.org/10.1158/2159-8290.CD-16-0313>.
- Korhuth, P.M., Berger, G., Bakker, B., Rozenveld-Geugien, M., Jaques, J., de Haan, G., Schuringa, J.J., Vellenga, E., Schepers, H., 2015. CITED2-mediated human hematopoietic stem cell maintenance is critical for acute myeloid leukemia. *Leukemia* 29, 625–635. <https://doi.org/10.1038/leu.2014.259>.
- Kowaltowski, A.J., Vercesi, A.E., Fiskum, G., 2000. Bcl-2 prevents mitochondrial permeability transition and cytochrome c release via maintenance of reduced pyridine nucleotides. *Cell Death Differ.* 7, 903–910. <https://doi.org/10.1038/sj.cdd.4400722>.
- Kranc, K.R., Schepers, H., Rodrigues, N.P., Bamforth, S., Villadsen, E., Ferry, H., Bouriez-Jones, T., Sigvardsson, M., Bhattacharya, S., Jacobsen, S.E., Enver, T., 2009. Cited2 is an essential regulator of adult hematopoietic stem cells. *Cell Stem Cell* 5, 659–665. <https://doi.org/10.1016/j.stem.2009.11.001>.
- Kuntz, E.M., Baquero, P., Michie, A.M., Dunn, K., Tardito, S., Holyoake, T.L., Helgason, G.V., Gottlieb, E., 2017. Targeting mitochondrial oxidative phosphorylation eradicates therapy-resistant chronic myeloid leukemia stem cells. *Nat. Med.* 23, 1234–1240. <https://doi.org/10.1038/nm.4399>.
- Lagadinou, E.D., Sach, A., Callahan, K., Rossi, R.M., Neering, S.J., Minhajuddin, M., Ashton, J.M., Pei, S., Grose, V., O'Dwyer, K.M., Liesveld, J.L., Brookes, P.S., Becker, M.W., Jordan, C.T., 2013. BCL-2 inhibition targets oxidative phosphorylation and selectively eradicates quiescent human leukemia stem cells. *Cell Stem Cell* 12, 329–341. <https://doi.org/10.1016/j.stem.2012.12.013>.
- Lapidot, T., Sirard, C., Vormoor, J., Murdoch, B., Hoang, T., Caceres-Cortes, J., Minden, M., Paterson, B., Caligiuri, M.A., Dick, J.E., 1994. A cell initiating human acute myeloid leukaemia after transplantation into SCID mice. *Nature* 367, 645–648. <https://doi.org/10.1038/367645a0>.
- Li, L., Bhatia, R., 2015. Role of SIRT1 in the growth and regulation of normal hematopoietic and leukemia stem cells. *Curr. Opin. Hematol.* 22, 324–329. <https://doi.org/10.1097/MOH.0000000000000152>.
- Li, Q., Rycak, K., Chen, X., Tang, D.G., 2015. Cancer stem cells and cell size: a causal link? *Semin. Cancer Biol.* 35, 191–199. <https://doi.org/10.1016/j.semcancer.2015.07.002>.
- Liang, R., Rimmele, P., Bigarella, C.L., Yalcin, S., Ghaffari, S., 2016. Evidence for AKT-independent regulation of FOXO1 and FOXO3 in haematopoietic stem and progenitor cells. *Cell Cycle* 15, 861–867. <https://doi.org/10.1080/15384101.2015.1123355>.
- Lin, K.H., Xie, A., Rutter, J.C., Ahn, Y.-R., Lloyd-Cowden, J.M., Nichols, A.G., Soderquist, R.S., Koves, T.R., Muoio, D.M., MacIver, N.J., Lamba, J.K., Pardee, T.S., McCall, C.M., Rizzieri, D.A., Wood, K.C., 2019. Systematic dissection of the metabolic-apoptotic interface in AML reveals heme biosynthesis to be a regulator of drug sensitivity. *Cell Metab.* <https://doi.org/10.1016/j.cmet.2019.01.011>.
- Liu, J., Cao, L., Chen, J., Song, S., Lee, I.H., Quijano, C., Liu, H., Keyvanfar, K., Chen, H., Cao, L.-Y., Ahn, B.-H., Kumar, N.G., Rovira, I.I., Xu, X.-L., van Lohuizen, M., Motoyama, N., Deng, C.-X., Finkel, T., 2009. Bmi1 regulates mitochondrial function and the DNA damage response pathway. *Nature* 459, 387–392. <https://doi.org/10.1038/nature08040>.
- Liu, X., Zheng, H., Yu, W.-M., Cooper, T.M., Bunting, K.D., Qu, C.-K., 2015. Maintenance of mouse hematopoietic stem cells ex vivo by reprogramming cellular metabolism. *Blood* 125, 1562–1565. <https://doi.org/10.1182/blood-2014-04-568949>.
- Luchsinger, L.L., de Almeida, M.J., Corrigan, D.J., Mumau, M., Snoeck, H.-W., 2016. Mitofusin 2 maintains haematopoietic stem cells with extensive lymphoid potential. *Nature* 529, 528–531. <https://doi.org/10.1038/nature16500>.
- Ludin, A., Itkin, T., Gur-Cohen, S., Mildner, A., Shezen, E., Golan, K., Kollet, O., Kalinkovich, A., Porat, Z., D'Uva, G., Schajnovitz, A., Voronov, E., Brenner, D.A., Apte, R.N., Jung, S., Lapidot, T., 2012. Monocytes-macrophages that express alpha-smooth muscle actin preserve primitive hematopoietic cells in the bone marrow. *Nat. Immunol.* 13, 1072–1082. <https://doi.org/10.1038/ni.2408>.
- Mantel, C., Messina-Graham, S., Moh, A., Cooper, S., Hangoc, G., Fu, X.-Y., Broxmeyer, H.E., 2012. Mouse hematopoietic cell-targeted STAT3 deletion: stem/progenitor cell defects, mitochondrial dysfunction, ROS overproduction, and a rapid aging-like phenotype. *Blood* 120, 2589–2599. <https://doi.org/10.1182/blood-2012-01-404004>.
- Mantel, C.R., O'Leary, H.A., Chitteti, B.R., Huang, X., Cooper, S., Hangoc, G., Brustovetsky, N., Srour, E.F., Lee, M.R., Messina-Graham, S., Haas, D.M., Falah, N., Kapur, R., Pelus, L.M., Bardeesy, N., Fitamant, J., Ivan, M., Kim, K.-S., Broxmeyer, H.E., 2015. Enhancing Hematopoietic Stem Cell Transplantation Efficacy by Mitigating Oxygen Shock. *Cell* 161, 1553–1565. <https://doi.org/10.1016/j.cell.2015.04.054>.
- Maryanovich, M., Zaltsman, Y., Ruggiero, A., Goldman, A., Shachnai, L., Zaidman, S.L., Porat, Z., Golan, K., Lapidot, T., Gross, A., 2015. An MTH2 pathway repressing mitochondrial metabolism regulates haematopoietic stem cell fate. *Nat. Commun.* 6, 7901. <https://doi.org/10.1038/ncomms8901>.
- Miao, W., Xufeng, R., Park, M.-R., Gu, H., Hu, L., Kang, J.W., Ma, S., Liang, P.H., Li, Y., Cheng, H., Yu, H., Epperly, M., Greenberger, J., Cheng, T., 2013. Hematopoietic stem cell regeneration enhanced by ectopic expression of ROS-detoxifying enzymes in transplant mice. *Mol. Ther.* 21, 423–432. <https://doi.org/10.1038/mt.2012.232>.
- Miyamoto, K., Araki, K.Y., Naka, K., Arai, F., Takubo, K., Yamazaki, S., Matsuoka, S., Miyamoto, T., Ito, K., Ohmura, M., Chen, C., Hosokawa, K., Nakauchi, H., Nakayama, K., Nakayama, K.I., Harada, M., Motoyama, N., Suda, T., Hirao, A., 2007. Foxo3a is essential for maintenance of the hematopoietic stem cell pool. *Cell Stem Cell* 1, 101–112. <https://doi.org/10.1016/j.stem.2007.02.001>.
- Mizushima, N., Levine, B., Cuervo, A.M., Klionsky, D.J., 2008. Autophagy fights disease through cellular self-digestion. *Nature* 451, 1069–1075. <https://doi.org/10.1038/nature06639>.
- Mohrin, M., Shin, J., Liu, Y., Brown, K., Luo, H., Xi, Y., Haynes, C.M., Chen, D., 2015. Stem cell aging. A mitochondrial UPR-mediated metabolic checkpoint regulates hematopoietic stem cell aging. *Science* 347, 1374–1377. <https://doi.org/10.1126/science.12361>.
- Mohyeldin, A., Garzon-Muvdi, T., Quinones-Hinojosa, A., 2010. Oxygen in stem cell

- biology: a critical component of the stem cell niche. *Cell Stem Cell* 7, 150–161. <https://doi.org/10.1016/j.stem.2010.07.007>.
- Molavian, H.R., Goldman, A., Phipps, C.J., Kohandel, M., Wouters, B.G., Sengupta, S., Sivaloganathan, S., 2016. Drug-induced reactive oxygen species (ROS) rely on cell membrane properties to exert anticancer effects. *Sci. Rep.* 6, 27439. <https://doi.org/10.1038/srep27439>.
- Molina, J.R., Sun, Y., Protopopova, M., Gera, S., Bandi, M., Bristow, C., McAfoos, T., Morlacchi, P., Ackroyd, J., Agip, A.-N.A., Al-Atrash, G., Asara, J., Bardenhagen, J., Carrillo, C.C., Carroll, C., Chang, E., Ciurea, S., Cross, J.B., Czako, B., Deem, A., Daver, N., de Groot, J.F., Dong, J.-W., Feng, N., Gao, G., Gay, J., Do, M.G., Greer, J., Giuliani, V., Han, J., Han, L., Henry, V.K., Hirst, J., Huang, S., Jiang, Y., Kang, Z., Khor, T., Konoplev, S., Lin, Y.-H., Liu, G., Lodi, A., Lofton, T., Ma, H., Mahendra, M., Matre, P., Mullinax, R., Peoples, M., Petrocchi, A., Rodriguez-Canale, J., Serreli, R., Shi, T., Smith, M., Tabe, Y., Theroff, J., Tiziani, S., Xu, Q., Zhang, Q., Muller, F., DePinho, R.A., Toniatti, C., Draetta, G.F., Heffernan, T.P., Konopleva, M., Jones, P., Di Francesco, M.E., Marszalek, J.R., 2018. An inhibitor of oxidative phosphorylation exploits cancer vulnerability. *Nat. Med.* 24, 1036–1046. <https://doi.org/10.1038/s41591-018-0052-4>.
- Morita, M., Gravel, S.-P., Chenard, V., Sikstrom, K., Zheng, L., Alain, T., Gandin, V., Avizonis, D., Arguello, M., Zakaria, C., McLaughlan, S., Nouet, Y., Pause, A., Pollak, M., Gottlieb, E., Larsson, O., St-Pierre, J., Topisirovic, I., Sonenberg, N., 2013. mTORC1 controls mitochondrial activity and biogenesis through 4E-BP-dependent translational regulation. *Cell Metab.* 18, 698–711. <https://doi.org/10.1016/j.cmet.2013.10.001>.
- Mortensen, M., Soilleux, E.J., Djordjevic, G., Tripp, R., Lutteropp, M., Sadighi-Akha, E., Stranks, A.J., Glanville, J., Knight, S., Jacobsen, S.-E.W., Kranc, K.R., Simon, A.K., 2011. The autophagy protein Atg7 is essential for hematopoietic stem cell maintenance. *J. Exp. Med.* 208, 455–467. <https://doi.org/10.1084/jem.20101145>.
- Murphy, M.P., 2009. How mitochondria produce reactive oxygen species. *Biochem. J.* 417, 1–13. <https://doi.org/10.1042/BJ20081386>.
- Nakada, D., Saunders, T.L., Morrison, S.J., 2010. Lkb1 regulates cell cycle and energy metabolism in haematopoietic stem cells. *Nature* 468, 653–658. <https://doi.org/10.1038/nature09571>.
- Naranjo-Suarez, S., Carlson, B.A., Tsuji, P.A., Yoo, M.-H., Gladyshev, V.N., Hatfield, D.L., 2012. HIF-independent regulation of thioredoxin reductase 1 contributes to the high levels of reactive oxygen species induced by hypoxia. *PLoS One* 7, e30470. <https://doi.org/10.1371/journal.pone.0030470>.
- Nguyen, L.X.T., Troadec, E., Kalvala, A., Kumar, B., Hoang, D.H., Viola, D., Zhang, B., Nguyen, D.Q., Aldoss, I., Ghoda, L., Budde, E., Pichiorri, F., Rosen, S., Forman, S.J., Marcucci, G., Pullarkat, V., 2019a. The Bcl-2 inhibitor venetoclax inhibits Nrf2 antioxidant pathway activation induced by hypomethylating agents in AML. *J. Cell. Physiol.* <https://doi.org/10.1002/jcp.28091>.
- Nguyen, T.D., Shaid, S., Vakhrusheva, O., Koschade, S.E., Klann, K., Tholken, M., Baker, F., Zhang, Y., Oellerich, T., Surun, D., Derlet, A., Haberbosch, I., Eimer, S., Osiewacz, H.D., Behrends, C., Munch, C., Dikic, I., Brandts, C.H., 2019b. Loss of the selective autophagy receptor p62 impairs murine myeloid leukemia progression and mitophagy. *Blood* 133, 168–179. <https://doi.org/10.1182/blood-2018-02-833475>.
- Niedernhofer, L.J., 2008. DNA repair is crucial for maintaining hematopoietic stem cell function. *DNA Repair (Amst)* 7, 523–529. <https://doi.org/10.1016/j.dnarep.2007.11.012>.
- Niu, X., Wang, G., Wang, Y., Caldwell, J.T., Edwards, H., Xie, C., Taub, J.W., Li, C., Lin, H., Ge, Y., 2014. Acute myeloid leukemia cells harboring MLL fusion genes or with the acute promyelocytic leukemia phenotype are sensitive to the Bcl-2-selective inhibitor ABT-199. *Leukemia*. <https://doi.org/10.1038/leu.2014.72>.
- Norddahl, G.L., Pronk, C.J., Wahlestedt, M., Sten, G., Nygren, J.M., Ugale, A., Sigvardsson, M., Bryder, D., 2011. Accumulating mitochondrial DNA mutations drive premature hematopoietic aging phenotypes distinct from physiological stem cell aging. *Cell Stem Cell* 8, 499–510. <https://doi.org/10.1016/j.stem.2011.03.009>.
- Notta, F., Zandi, S., Takayama, N., Dobson, S., Gan, O.I., Wilson, G., Kaufmann, K.B., McLeod, J., Laurenti, E., Dunant, C.F., McPherson, J.D., Stein, L.D., Dror, Y., Dick, J.E., 2016. Distinct routes of lineage development reshape the human blood hierarchy across ontogeny. *Science* 351, aab2116. <https://doi.org/10.1126/science.aab2116>.
- O'Reilly, E., Dhami, S.P.S., Baev, D.V., Ortutay, C., Halpin-McCormick, A., Morrell, R., Santocanale, C., Samali, A., Quinn, J., O'Dwyer, M.E., Szegezdi, E., 2018. Repression of Mcl-1 expression by the CDC7/CDK9 inhibitor PHA-767491 overcomes bone marrow stroma-mediated drug resistance in AML. *Sci. Rep.* 8, 15752. <https://doi.org/10.1038/s41598-018-33982-y>.
- Pan, R., Ruvolo, V.R., Wei, J., Konopleva, M., Reed, J.C., Pellicchia, M., Andreeff, M., Ruvolo, P.P., 2015. Inhibition of Mcl-1 with the pan-Bcl-2 family inhibitor (BI)97D6 overcomes ABT-737 resistance in acute myeloid leukemia. *Blood* 126, 363–372. <https://doi.org/10.1182/blood-2014-10-604975>.
- Papa, L., Zimran, E., Djedaini, M., Ge, Y., Ozbek, U., Sebra, R., Sealon, S.C., Hoffman, R., 2018. Ex vivo human HSC expansion requires coordination of cellular reprogramming with mitochondrial remodeling and p53 activation. *Blood Adv.* 2, 2766–2779. <https://doi.org/10.1182/bloodadvances.2018024273>.
- Parkin, B., Ouillette, P., Li, Y., Keller, J., Lam, C., Roulston, D., Li, C., Shedden, K., Malek, S.N., 2013. Clonal evolution and devolution after chemotherapy in adult acute myelogenous leukemia. *Blood* 121, 369–377. <https://doi.org/10.1182/blood-2012-04-427039>.
- Pei, S., Minhajuddin, M., Adane, B., Khan, N., Stevens, B.M., Mack, S.C., Lai, S., Rich, J.N., Inguva, A., Shannon, K.A., Kim, H., Tan, A.-C., Myers, J.R., Ashton, J.M., Neff, T., Pollyea, D.A., Smith, C.M., Jordan, C.T., 2018. AMPK/FIS1-mediated mitophagy is required for self-renewal of human AML stem cells. *Cell Stem Cell* 23, 86–100. <https://doi.org/10.1016/j.stem.2018.05.021>.
- Perri, F., Pisconti, S., Della Vittoria Scarpato, G., 2016. P53 mutations and cancer: a tight linkage. *Ann. Transl. Med.* 4, 522. <https://doi.org/10.21037/atm.2016.12.40>.
- Pollyea, D.A., Jordan, C.T., 2019. Why are hypomethylating agents or low-dose cytarabine and venetoclax so effective? *Curr. Opin. Hematol.* 26, 71–76. <https://doi.org/10.1097/MOH.0000000000000485>.
- Pollyea, D.A., Stevens, B.M., Jones, C.L., Winters, A., Pei, S., Minhajuddin, M., D'Alessandro, A., Culp-Hill, R., Riemondy, K.A., Gillen, A.E., Hesselberth, J.R., Abbott, D., Schatz, D., Gutman, J.A., Purev, E., Smith, C., Jordan, C.T., 2018a. Venetoclax with azacitidine disrupts energy metabolism and targets leukemia stem cells in patients with acute myeloid leukemia. *Nat. Med.* <https://doi.org/10.1038/s41591-018-0233-1>.
- Pollyea, D.A., Stevens, B.M., Jones, C.L., Winters, A., Pei, S., Minhajuddin, M., D'Alessandro, A., Culp-Hill, R., Riemondy, K.A., Gillen, A.E., Hesselberth, J.R., Abbott, D., Schatz, D., Gutman, J.A., Purev, E., Smith, C., Jordan, C.T., 2018b. Venetoclax with azacitidine disrupts energy metabolism and targets leukemia stem cells in patients with acute myeloid leukemia. *Nat. Med.* 24, 1859–1866. <https://doi.org/10.1038/s41591-018-0233-1>.
- Prost, S., Relouzat, F., Spentchian, M., Ouzegdough, Y., Saliba, J., Massonnet, G., Beressi, J.-P., Verhoeyen, E., Ragueneau, V., Maneglier, B., Castaigne, S., Chomienne, C., Chretien, S., Rousselot, P., Leboulch, P., 2015. Erosion of the chronic myeloid leukaemia stem cell pool by PPARG agonists. *Nature* 525, 380–383. <https://doi.org/10.1038/nature15248>.
- Qian, P., He, X.C., Paulson, A., Li, Z., Tao, F., Perry, J.M., Guo, F., Zhao, M., Zhi, L., Venkatraman, A., Haug, J.S., Parmely, T., Li, H., Dobrowsky, R.T., Ding, W.-X., Kono, T., Ferguson-Smith, A.C., Li, L., 2016. The Dlk1-Gtl2 locus preserves LT-HSC function by inhibiting the PI3K-mTOR pathway to restrict mitochondrial metabolism. *Cell Stem Cell* 18, 214–228. <https://doi.org/10.1016/j.stem.2015.11.001>.
- Rafalski, V.A., Mancini, E., Brunet, A., 2012. Energy metabolism and energy-sensing pathways in mammalian embryonic and adult stem cell fate. *J. Cell. Sci.* 125, 5597–5608. <https://doi.org/10.1242/jcs.114827>.
- Raffel, S., Falcone, M., Kneisel, N., Hansson, J., Wang, W., Lutz, C., Bullinger, L., Poschet, G., Nonnenmacher, Y., Barnert, A., Bahr, C., Zeisberger, P., Przybylla, A., Sohn, M., Tonjes, M., Erez, A., Adler, L., Jensen, P., Scholl, C., Frohling, S., Cocciaardi, S., Wuchter, P., Thiede, C., Florcken, A., Westermann, J., Ehninger, G., Lichter, P., Hiller, K., Hell, R., Herrmann, C., Ho, A.D., Krijgsvel, J., Radlwimmer, B., Trumpp, A., 2017. BCAT1 restricts alphaK levels in AML stem cells leading to IDHmut-like DNA hypermethylation. *Nature* 551, 384–388. <https://doi.org/10.1038/nature24294>.
- Ramsey, H.E., Fischer, M.A., Lee, T., Gorska, A.E., Arrate, M.P., Fuller, L., Boyd, K.L., Strickland, S.A., Sensintaffar, J., Hogdal, L.J., Ayers, G.D., Olejniczak, E.T., Fesik, S.W., Savona, M.R., 2018. A novel MCL1 inhibitor combined with venetoclax rescues venetoclax-resistant acute myelogenous leukemia. *Cancer Discov.* 8, 1566–1581. <https://doi.org/10.1158/2159-8290.CD-18-0140>.
- Ray, P.D., Huang, B.-W., Tsuji, Y., 2012. Reactive oxygen species (ROS) homeostasis and redox regulation in cellular signaling. *Cell. Signal.* 24, 981–990. <https://doi.org/10.1016/j.cellsig.2012.01.008>.
- Rimmele, P., Bigarella, C.L., Liang, R., Izac, B., Dieguez-Gonzalez, R., Barbet, G., Donovan, M., Brugnara, C., Blander, J.M., Sinclair, D.A., Ghaffari, S., 2014. Aging-like phenotype and defective lineage specification in SIRT1-deleted hematopoietic stem and progenitor cells. *Stem Cell Reports* 3, 44–59. <https://doi.org/10.1016/j.stemcr.2014.04.015>.
- Rimmele, P., Liang, R., Bigarella, C.L., Kocabas, F., Xie, J., Serasinghe, M.N., Chipuk, J., Sadek, H., Zhang, C.C., Ghaffari, S., 2015. Mitochondrial metabolism in hematopoietic stem cells requires functional FOXO3. *EMBO Rep.* 16, 1164–1176. <https://doi.org/10.15252/embr.201439704>.
- Rodriguez-Fraticelli, A.E., Wolock, S.L., Weinreb, C.S., Panero, R., Patel, S.H., Jankovic, M., Sun, J., Calogero, R.A., Klein, A.M., Camargo, F.D., 2018. Clonal analysis of lineage fate in native haematopoiesis. *Nature* 553, 212–216. <https://doi.org/10.1038/nature25168>.
- Romero-Moya, D., Bueno, C., Montes, R., Navarro-Montero, O., Iborra, F.J., Lopez, L.C., Martin, M., Menendez, P., 2013. Cord blood-derived CD34+ hematopoietic cells with low mitochondrial mass are enriched in hematopoietic repopulating stem cell function. *Haematologica* 98, 1022–1029. <https://doi.org/10.3324/haematol.2012.079244>.
- Saito, Y., Chapple, R.H., Lin, A., Kitano, A., Nakada, D., 2015. AMPK protects leukemia-initiating cells in myeloid leukemias from metabolic stress in the bone marrow. *Cell Stem Cell* 17, 585–596. <https://doi.org/10.1016/j.stem.2015.08.019>.
- Sakai, M., Matsumoto, M., Tujimura, T., Yongheng, C., Noguchi, T., Inagaki, K., Inoue, H., Hosooka, K., Takazawa, K., Kido, Y., Yasuda, K., Hiramatsu, R., Matsuki, Y., Kasuga, M., 2012. CITED2 links hormonal signaling to PGC-1alpha acetylation in the regulation of gluconeogenesis. *Nat. Med.* 18, 612–617. <https://doi.org/10.1038/nm.2691>.
- Sakai, M., Tujimura-Hayakawa, T., Yagi, T., Yano, H., Mitsuhashi, M., Unoki-Kubota, H., Kaburagi, Y., Inoue, H., Kido, Y., Kasuga, M., Matsumoto, M., 2016. The GCN5-CITED2-PKA signalling module controls hepatic glucose metabolism through a cAMP-induced substrate switch. *Nat. Commun.* 7, 13147. <https://doi.org/10.1038/ncomms13147>.
- Samudio, I., Harmancey, R., Fiegl, M., Kantarjian, H., Konopleva, M., Korchin, B., Kaluarachchi, K., Bormmann, W., Duvvuri, S., Taegtmeier, H., Andreeff, M., 2010. Pharmacologic inhibition of fatty acid oxidation sensitizes human leukemia cells to apoptosis induction. *J. Clin. Invest.* 120, 142–156. <https://doi.org/10.1172/JCI38942>.
- Sanjuan-Pla, A., Macaulay, I.C., Jensen, C.T., Woll, P.S., Luis, T.C., Mead, A., Moore, S., Carella, C., Matsuoka, S., Bouriez Jones, T., Chowdhury, O., Stenson, L., Lutteropp, M., Green, J.C.A., Facchini, R., Boukarabila, H., Grover, A., Gambardella, A., Thongjuea, S., Carrelha, J., Tarrant, P., Atkinson, D., Clark, S.-A., Nerlov, C., Jacobsen, S.E.W., 2013. Platelet-biased stem cells reside at the apex of the

- haematopoietic stem-cell hierarchy. *Nature* 502, 232–236. <https://doi.org/10.1038/nature12495>.
- Santo, E.E., Paik, J., 2016. FOXO3a & haematopoietic stem cells: goodbye PI3K, hello SIRT1? *Cell Cycle*. <https://doi.org/10.1080/15384101.2016.1151721>.
- Santos, L., Escande, C., Denicola, A., 2016. Potential modulation of sirtuins by oxidative stress. *Oxid. Med. Cell. Longev.* 2016, 9831825. <https://doi.org/10.1155/2016/9831825>.
- Sarry, J.-E., Murphy, K., Perry, R., Sanchez, P.V., Secreto, A., Keefer, C., Swider, C.R., Strzelecki, A.-C., Cavelier, C., Recher, C., Mansat-De Mas, V., Delabesse, E., Danet-Desnoyers, G., Carroll, M., 2011. Human acute myelogenous leukemia stem cells are rare and heterogeneous when assayed in NOD/SCID/IL2Rgammac-deficient mice. *J. Clin. Invest.* 121, 384–395. <https://doi.org/10.1172/JCI41495>.
- Seneviratne, A.K., Xu, M., Henao, J.J.A., Fajardo, V.A., Hao, Z., Voisin, V., Xu, G.W., Hurren, R., Kim, S., MacLean, N., Wang, X., Gronda, M., Jeyaraju, D., Jitkova, Y., Ketela, T., Mullokandov, M., Sharon, D., Thomas, G., Chouinard-Watkins, R., Hawley, J.R., Schafer, C., Yau, H.L., Khuchua, Z., Aman, A., Al-Awar, R., Gross, A., Claypool, S.M., Bazinet, R., Lupien, M., Chan, S., De Carvalho, D.D., Minden, M.D., Bader, G.D., Stark, K.D., LeBlanc, P., Schimmer, A.D., 2019. The mitochondrial transacylase, Tafazzin, regulates for AML stemness by modulating intracellular levels of phospholipids. *Cell Stem Cell* 24, 621–636. <https://doi.org/10.1016/j.stem.2019.02.020.e16>.
- Shao, L., Li, H., Pazhanisamy, S.K., Meng, A., Wang, Y., Zhou, D., 2011. Reactive oxygen species and haematopoietic stem cell senescence. *Int. J. Hematol.* 94, 24–32. <https://doi.org/10.1007/s12185-011-0872-1>.
- Shiloh, Y., 2003. ATM and related protein kinases: safeguarding genome integrity. *Nat. Rev. Cancer* 3, 155–168. <https://doi.org/10.1038/nrc1011>.
- Shinohara, A., Imai, Y., Nakagawa, M., Takahashi, T., Ichikawa, M., Kurokawa, M., 2014. Intracellular reactive oxygen species mark and influence the megakaryocyte-erythrocyte progenitor fate of common myeloid progenitors. *Stem Cells* 32, 548–557. <https://doi.org/10.1002/stem.1588>.
- Shlush, L.I., Mitchell, A., Heisler, L., Abelson, S., Ng, S.W.K., Trotman-Grant, A., Medeiros, J.J.F., Rao-Bhatia, A., Jaciw-Zurakowsky, I., Marke, R., McLeod, J.L., Doedens, M., Bader, G., Voisin, V., Xu, C., McPherson, J.D., Hudson, T.J., Wang, J.C.Y., Minden, M.D., Dick, J.E., 2017. Tracing the origins of relapse in acute myeloid leukaemia to stem cells. *Nature* 547, 104–108. <https://doi.org/10.1038/nature22993>.
- Signer, R.A.J., Qi, L., Zhao, Z., Thompson, D., Sigova, A.A., Fan, Z.P., DeMartino, G.N., Young, R.A., Sonenberg, N., Morrison, S.J., 2016. The rate of protein synthesis in hematopoietic stem cells is limited partly by 4E-BPs. *Genes Dev.* 30, 1698–1703. <https://doi.org/10.1101/gad.282756.116>.
- Simssek, T., Kocabas, F., Zheng, J., Deberardinis, R.J., Mahmoud, A.I., Olson, E.N., Schneider, J.W., Zhang, C.C., Sadek, H.A., 2010. The distinct metabolic profile of hematopoietic stem cells reflects their location in a hypoxic niche. *Cell Stem Cell* 7, 380–390. <https://doi.org/10.1016/j.stem.2010.07.011>.
- Singh, S.K., Singh, S., Gadamski, S., Sun, L., Pfannenstien, A., Magidson, V., Chen, X., Kozlov, S., Tassarollo, L., Klarman, K.D., Keller, J.R., 2018. Id1 ablation protects hematopoietic stem cells from stress-induced exhaustion and aging. *Cell Stem Cell* 23, 252–265. <https://doi.org/10.1016/j.stem.2018.06.001>. e8.
- Skrct, M., Sriskanthadevan, S., Jhas, B., Gebbia, M., Wang, X., Wang, Z., Hurren, R., Jitkova, Y., Gronda, M., Maclean, N., Lai, C.K., Eberhard, Y., Bartoszko, J., Spagnuolo, P., Rutledge, A.C., Datti, A., Ketela, T., Moffat, J., Robinson, B.H., Cameron, J.H., Wrana, J., Eaves, C.J., Minden, M.D., Wang, J.C.Y., Dick, J.E., Humphries, K., Nislow, C., Giaever, G., Schimmer, A.D., 2011. Inhibition of mitochondrial translation as a therapeutic strategy for human acute myeloid leukemia. *Cancer Cell* 20, 674–688. <https://doi.org/10.1016/j.ccr.2011.10.015>.
- Souers, A.J., Levenson, J.D., Boghaert, E.R., Ackler, S.L., Catron, N.D., Chen, J., Dayton, B.D., Ding, H., Enschede, S.H., Fairbrother, W.J., Huang, D.C.S., Hymowitz, S.G., Jin, S., Khaw, S.L., Kovar, P.J., Lam, L.T., Lee, J., Maecker, H.L., Marsh, K.C., Mason, K.D., Mitten, M.J., Nimmer, P.M., Oleksiejew, A., Park, C.H., Park, C.-M., Phillips, D.C., Roberts, A.W., Sampath, D., Seymour, J.F., Smith, M.L., Sullivan, G.M., Tahir, S.K., Tse, C., Wendt, M.D., Xiao, Y., Xue, J.C., Zhang, H., Humerickhouse, R.A., Rosenberg, S.H., Elmores, S.W., 2013. ABT-199, a potent and selective BCL-2 inhibitor, achieves antitumor activity while sparing platelets. *Nat. Med.* 19, 202–208. <https://doi.org/10.1038/nm.3048>.
- Srskanthadevan, S., Jeyaraju, D.V., Chung, T.E., Prabha, S., Xu, W., Skrct, M., Jhas, B., Hurren, R., Gronda, M., Wang, X., Jitkova, Y., Sukhai, M.A., Lin, F.-H., Maclean, N., Laister, R., Goard, C.A., Mullen, P.J., Xie, S., Penn, L.Z., Rogers, I.M., Dick, J.E., Minden, M.D., Schimmer, A.D., 2015. AML cells have low spare reserve capacity in their respiratory chain that renders them susceptible to oxidative metabolic stress. *Blood* 125, 2120–2130. <https://doi.org/10.1182/blood-2014-08-594408>.
- Suda, T., Takubo, K., Semenza, G.L., 2011. Metabolic regulation of hematopoietic stem cells in the hypoxic niche. *Cell Stem Cell* 9, 298–310. <https://doi.org/10.1016/j.stem.2011.09.010>.
- Sukumar, M., Liu, J., Mehta, G.U., Patel, S.J., Roychoudhuri, R., Crompton, J.G., Klebanoff, C.A., Ji, Y., Li, P., Yu, Z., Whitehill, G.D., Clever, D., Eil, R.L., Palmer, D.C., Mitra, S., Rao, M., Keyvanfar, K., Schrumpp, G.S., Wang, E., Marincola, F.M., Gattinoni, L., Leonard, W.J., Muranski, P., Finkel, T., Restifo, N.P., 2016. Mitochondrial membrane potential identifies cells with enhanced stemness for cellular therapy. *Cell Metab.* 23, 63–76. <https://doi.org/10.1016/j.cmet.2015.11.002>.
- Sun, J., He, X., Zhu, Y., Ding, Z., Dong, H., Feng, Y., Du, J., Wang, H., Wu, X., Zhang, L., Yu, X., Lin, A., McDonald, T., Zhao, D., Wu, H., Hua, W.-K., Zhang, B., Feng, L., Tohyama, K., Bhatia, R., Oberdoerfer, P., Chung, Y.J., Aplan, P.D., Boulton, J., Pellagatti, A., Khaled, S., Kortylewski, M., Pichiorri, F., Kuo, Y.-H., Carlesso, N., Marcucci, G., Jin, H., Li, L., 2018. SIRT1 activation disrupts maintenance of myelodysplastic syndrome stem and progenitor cells by restoring TET2 function. *Cell Stem Cell* 23, 355–369. <https://doi.org/10.1016/j.stem.2018.07.018>. e9.
- Takubo, K., Goda, N., Yamada, W., Iriuchishima, H., Ikeda, E., Kubota, Y., Shima, H., Johnson, R.S., Hirao, A., Suematsu, M., Suda, T., 2010. Regulation of the Hif-1alpha level is essential for hematopoietic stem cells. *Cell Stem Cell* 7, 391–402. <https://doi.org/10.1016/j.stem.2010.06.020>.
- Takubo, K., Nagamatsu, G., Kobayashi, C.I., Nakamura-Ishizu, A., Kobayashi, H., Ikeda, E., Goda, N., Rahimi, Y., Johnson, R.S., Soga, T., Hirao, A., Suematsu, M., Suda, T., 2013. Regulation of glycolysis by Pdk functions as a metabolic checkpoint for cell cycle quiescence in hematopoietic stem cells. *Cell Stem Cell* 12, 49–61. <https://doi.org/10.1016/j.stem.2012.10.011>.
- Taniguchi Ishikawa, E., Gonzalez-Nieto, D., Ghiaur, G., Dunn, S.K., Ficker, A.M., Murali, B., Madhu, M., Gutstein, D.E., Fishman, G.I., Barrio, L.C., Cancelas, J.A., 2012. Connexin-43 prevents hematopoietic stem cell senescence through transfer of reactive oxygen species to bone marrow stromal cells. *Proc. Natl. Acad. Sci. U. S. A.* 109, 9071–9076. <https://doi.org/10.1073/pnas.1120358110>.
- Tasdogan, A., Kumar, S., Allies, G., Bausinger, J., Beckel, F., Hofmeister, H., Mulaw, M., Madan, V., Scharfetter-Kochanek, K., Feuring-Buske, M., Doehner, K., Speit, G., Stewart, A.F., Fehling, H.J., 2016. DNA damage-induced HSPC malfunction depends on ROS accumulation downstream of IFN-1 signaling and bid mobilization. *Cell Stem Cell* 19, 752–767. <https://doi.org/10.1016/j.stem.2016.08.007>.
- Thomas, D., Majeti, R., 2017. Biology and relevance of human acute myeloid leukemia stem cells. *Blood* 129, 1577–1585. <https://doi.org/10.1182/blood-2016-10-696054>.
- Tian, Y., Huang, Z., Wang, Z., Yin, C., Zhou, L., Zhang, L., Huang, K., Zhou, H., Jiang, X., Li, J., Liao, L., Yang, M., Meng, F., 2014. Identification of novel molecular markers for prognosis estimation of acute myeloid leukemia: over-expression of PDCD7, FIS1 and Ang2 may indicate poor prognosis in pretreatment patients with acute myeloid leukemia. *PLoS One* 9, e84150. <https://doi.org/10.1371/journal.pone.0084150>.
- Torgersen, M.L., Simonsen, A., 2013. Autophagy: friend or foe in the treatment of fusion protein-associated leukemias? *Autophagy* 9, 2175–2177. <https://doi.org/10.4161/auto.26559>.
- Tothova, Z., Kollipara, R., Huntly, B.J., Lee, B.H., Castrillon, D.H., Cullen, D.E., McDowell, E.P., Lazo-Kallanian, S., Williams, I.R., Sears, C., Armstrong, S.A., Passgeue, E., DePinho, R.A., Gilliland, D.G., 2007. FoxOs are critical mediators of hematopoietic stem cell resistance to physiologic oxidative stress. *Cell* 128, 325–339. <https://doi.org/10.1016/j.cell.2007.01.003>.
- Tron, A.E., Belmonte, M.A., Adam, A., Aquila, B.M., Boise, L.H., Chiarparin, E., Cidado, J., Embrey, K.J., Gangl, E., Gibbons, F.D., Gregory, G.P., Hargreaves, D., Hendricks, J.A., Johannes, J.W., Johnstone, R.W., Kazmirski, S.L., Kettle, J.G., Lamb, M.L., Matulis, S.M., Nooka, A.K., Packer, M.J., Peng, B., Rawlins, P.B., Robbins, D.W., Schuller, A.G., Su, N., Yang, W., Ye, Q., Zheng, X., Secrist, J.P., Clark, E.A., Wilson, D.M., Fawell, S.E., Hird, A.W., 2018. Discovery of Mcl-1-specific inhibitor AZD5991 and preclinical activity in multiple myeloma and acute myeloid leukemia. *Nat. Commun.* 9, 5341. <https://doi.org/10.1038/s41467-018-07551-w>.
- Trotta, A.P., Gelles, J.D., Serasinghe, M.N., Loi, P., Arbiser, J.L., Chipuk, J.E., 2017. Disruption of mitochondrial electron transport chain function potentiates the pro-apoptotic effects of MAPK inhibition. *J. Biol. Chem.* 292, 11727–11739. <https://doi.org/10.1074/jbc.M117.786442>.
- Tsuchiya, Y., Peak-Chew, S.Y., Newell, C., Miller-Aidoo, S., Mangal, S., Zhyvolou, A., Bakovic, J., Malanchuk, O., Pereira, G.C., Kotiadis, V., Szabadkai, G., Duchon, M.R., Campbell, M., Cuenca, S.R., Vidal-Puig, A., James, A.M., Murphy, M.P., Filonenko, V., Skehel, M., Gout, I., 2017. Protein CoAlation: a redox-regulated protein modification by coenzyme A in mammalian cells. *Biochem. J.* 474, 2489–2508. <https://doi.org/10.1042/BCJ20170129>.
- Umemoto, T., Hashimoto, M., Matsumura, T., Nakamura-Ishizu, A., Suda, T., 2018. Ca<sup>2+</sup>-mitochondria axis drives cell division in hematopoietic stem cells. *J. Exp. Med.* 215, 2097–2113. <https://doi.org/10.1084/jem.20180421>.
- Unnisa, Z., Clark, J.P., Roychoudhuri, J., Thomas, E., Tassarollo, L., Copeland, N.G., Jenkins, N.A., Grimes, H.L., Kumar, A.R., 2012. Meis1 preserves hematopoietic stem cells in mice by limiting oxidative stress. *Blood* 120, 4973–4981. <https://doi.org/10.1182/blood-2012-06-435800>.
- van der Kolk, D.M., de Vries, E.G., van Putten, W.J., Verdonck, L.F., Ossenkoppele, G.J., Verhoef, G.E., Vellenga, E., 2000. P-glycoprotein and multidrug resistance protein activities in relation to treatment outcome in acute myeloid leukemia. *Clin. Cancer Res.* 6, 3205–3214.
- van der Vos, K.E., Eliasson, P., Proikas-Cezanne, T., Vervoort, S.J., van Bostel, R., Putker, M., van Zutphen, I.J., Mauthe, M., Zellmer, S., Pals, C., Verhagen, L.P., Groot Koerkamp, M.J.A., Braat, A.K., Dansen, T.B., Holstege, F.C., Gebhardt, R., Burgering, B.M., Coffey, P.J., 2012. Modulation of glutamine metabolism by the PI(3)K-PKB-FOXO network regulates autophagy. *Nat. Cell Biol.* 14, 829–837. <https://doi.org/10.1038/ncb2536>.
- van Galen, P., Kreso, A., Mbong, N., Kent, D.G., Fitzmaurice, T., Chambers, J.E., Xie, S., Laurenti, E., Hermans, K., Eppert, K., Marciniak, S.J., Goodall, J.C., Green, A.R., Wouters, B.G., Wienholds, E., Dick, J.E., 2014. The unfolded protein response governs integrity of the haematopoietic stem-cell pool during stress. *Nature* 510, 268–272. <https://doi.org/10.1038/nature13228>.
- Vannini, N., Girotra, M., Naveiras, O., Nikitin, G., Campos, V., Giger, S., Roch, A., Auwerx, J., Lutolf, M.P., 2016. Specification of haematopoietic stem cell fate via modulation of mitochondrial activity. *Nat. Commun.* 7, 13125. <https://doi.org/10.1038/ncb2536>.
- Velasco-Hernandez, T., Soneji, S., Hidalgo, I., Erlandsson, E., Cammenga, J., Bryder, D., 2019. Hif-1alpha deletion may lead to adverse treatment effect in a mouse model of MLL-AF9-Driven AML. *Stem Cell Reports* 12, 112–121. <https://doi.org/10.1016/j.stemcr.2018.11.023>.
- Vukovic, M., Guitart, A.V., Sepulveda, C., Villacreses, A., O'Duibhir, E., Panagopoulou, T.I., Ivens, A., Menendez-Gonzalez, J., Iglesias, J.M., Allen, L., Glykofrydis, F., Subramani, C., Armesilla-Diaz, A., Post, A.E.M., Schaak, K., Gezer, D., So, C.W.E., Holyoake, T.L., Wood, A., O'Carroll, D., Ratcliffe, P.J., Kranc, K.R., 2015. Hif-1alpha

- and Hif-2alpha synergize to suppress AML development but are dispensable for disease maintenance. *J. Exp. Med.* 212, 2223–2234. <https://doi.org/10.1084/jem.20150452>.
- Vukovic, M., Sepulveda, C., Subramani, C., Guitart, A.V., Mohr, J., Allen, L., Panagopoulou, T.I., Paris, J., Lawson, H., Villacreces, A., Armesilla-Diaz, A., Gezer, D., Holyoake, T.L., Ratcliffe, P.J., Kranc, K.R., 2016. Adult hematopoietic stem cells lacking Hif-1alpha self-renew normally. *Blood* 127, 2841–2846. <https://doi.org/10.1182/blood-2015-10-677138>.
- Wang, H., Diao, D., Shi, Z., Zhu, X., Gao, Y., Gao, S., Liu, X., Wu, Y., Rudolph, K.L., Liu, G., Li, T., Ju, Z., 2016. SIRT6 controls hematopoietic stem cell homeostasis through epigenetic regulation of wnt signaling. *Cell Stem Cell* 18, 495–507. <https://doi.org/10.1016/j.stem.2016.03.005>.
- Wang, Y.-H., Israelsen, W.J., Lee, D., Yu, V.W.C., Jeanson, N.T., Clish, C.B., Cantley, L.C., Vander Heiden, M.G., Scadden, D.T., 2014. Cell-state-specific metabolic dependency in hematopoiesis and leukemogenesis. *Cell* 158, 1309–1323. <https://doi.org/10.1016/j.cell.2014.07.048>.
- Watson, A.S., Riffelmacher, T., Stranks, A., Williams, O., De Boer, J., Cain, K., MacFarlane, M., McGouran, J., Kessler, B., Khandwala, S., Chowdhury, O., Puleston, D., Phadwal, K., Mortensen, M., Ferguson, D., Soilleux, E., Woll, P., Jacobsen, S.E.W., Simon, A.K., 2015. Autophagy limits proliferation and glycolytic metabolism in acute myeloid leukemia. *Cell Death Discov.* 1. <https://doi.org/10.1038/cddiscovery.2015.8>.
- Woolthuis, C.M., Brouwers-Vos, A.Z., Huls, G., de Wolf, J.T.M., Schuringa, J.J., Vellenga, E., 2013. Loss of quiescence and impaired function of CD34(+)/CD38(low) cells one year following autologous stem cell transplantation. *Haematologica* 98, 1964–1971. <https://doi.org/10.3324/haematol.2013.086744>.
- Xiao, N., Jani, K., Morgan, K., Okabe, R., Cullen, D.E., Jesneck, J.L., Raffel, G.D., 2012. Hematopoietic stem cells lacking Ott1 display aspects associated with aging and are unable to maintain quiescence during proliferative stress. *Blood* 119, 4898–4907. <https://doi.org/10.1182/blood-2012-01-403089>.
- Xu, S.-N., Wang, T.-S., Li, X., Wang, Y.-P., 2016. SIRT2 activates G6PD to enhance NADPH production and promote leukaemia cell proliferation. *Sci. Rep.* 6, 32734. <https://doi.org/10.1038/srep32734>.
- Yahata, T., Takanashi, T., Muguruma, Y., Ibrahim, A.A., Matsuzawa, H., Uno, T., Sheng, Y., Onizuka, M., Ito, M., Kato, S., Ando, K., 2011. Accumulation of oxidative DNA damage restricts the self-renewal capacity of human hematopoietic stem cells. *Blood* 118, 2941–2950. <https://doi.org/10.1182/blood-2011-01-330050>.
- Yalcin, S., Zhang, X., Luciano, J.P., Mungamuri, S.K., Marinkovic, D., Vercherat, C., Sarkar, A., Grisotto, M., Taneja, R., Ghaffari, S., 2008. Foxo3 is essential for the regulation of ataxia telangiectasia mutated and oxidative stress-mediated homeostasis of hematopoietic stem cells. *J. Biol. Chem.* 283, 25692–25705. <https://doi.org/10.1074/jbc.M800517200>.
- Yamamoto, R., Morita, Y., Ooehara, J., Hamanaka, S., Onodera, M., Rudolph, K.L., Ema, H., Nakauchi, H., 2013. Clonal analysis unveils self-renewing lineage-restricted progenitors generated directly from hematopoietic stem cells. *Cell* 154, 1112–1126. <https://doi.org/10.1016/j.cell.2013.08.007>.
- Yehudai, D., Liyanage, S.U., Hurren, R., Rizoska, B., Albertella, M., Gronda, M., Jeyaraju, D.V., Wang, X., Barghout, S.H., MacLean, N., Siriwardena, T.P., Jitkova, Y., Targett-Adams, P., Schimmer, A.D., 2018. The thymidine dideoxynucleoside analogue, alovudine, inhibits the mitochondrial DNA polymerase gamma, impairs oxidative phosphorylation and promotes monocytic differentiation in acute myeloid leukemia. *Haematologica*. <https://doi.org/10.3324/haematol.2018.195172>.
- Yu, W.-M., Liu, X., Shen, J., Jovanovic, O., Pohl, E.E., Gerson, S.L., Finkel, T., Broxmeyer, H.E., Qu, C.-K., 2013. Metabolic regulation by the mitochondrial phosphatase PTPMT1 is required for hematopoietic stem cell differentiation. *Cell Stem Cell* 12, 62–74. <https://doi.org/10.1016/j.stem.2012.11.022>.
- Zhang, C.-Y., Yin, H.-M., Wang, H., Su, D., Xia, Y., Yan, L.-F., Fang, B., Liu, W., Wang, Y.-M., Gu, A.-H., Zhou, Y., 2018. Transforming growth factor-beta1 regulates the nascent hematopoietic stem cell niche by promoting gluconeogenesis. *Leukemia* 32, 479–491. <https://doi.org/10.1038/leu.2017.198>.
- Zhang, C.C., Sadek, H.A., 2014. Hypoxia and metabolic properties of hematopoietic stem cells. *Antioxid. Redox Signal.* 20, 1891–1901. <https://doi.org/10.1089/ars.2012.5019>.
- Zhang, Y., Depond, M., He, L., Foudi, A., Kwarteng, E.O., Lauret, E., Plo, I., Desterke, C., Dessen, P., Fujii, N., Opolon, P., Herault, O., Solary, E., Vainchenker, W., Joulin, V., Louache, F., Wittner, M., 2016. CXCR4/CXCL12 axis counteracts hematopoietic stem cell exhaustion through selective protection against oxidative stress. *Sci. Rep.* 6, 37827. <https://doi.org/10.1038/srep37827>.
- Zhao, Y., Hu, X., Liu, Y., Dong, S., Wen, Z., He, W., Zhang, S., Huang, Q., Shi, M., 2017. ROS signaling under metabolic stress: cross-talk between AMPK and AKT pathway. *Mol. Cancer* 16, 79. <https://doi.org/10.1186/s12943-017-0648-1>.
- Zimmermann, A.K., Loucks, F.A., Schroeder, E.K., Bouchard, R.J., Tyler, K.L., Linseman, D.A., 2007. Glutathione binding to the Bcl-2 homology-3 domain groove: a molecular basis for Bcl-2 antioxidant function at mitochondria. *J. Biol. Chem.* 282, 29296–29304. <https://doi.org/10.1074/jbc.M702853200>.