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# E-waste polycyclic aromatic hydrocarbon (PAH) exposure leads to child gut-mucosal inflammation and adaptive immune response

Guangcan Chen<sup>1</sup> · Xia Huo<sup>2</sup> · Xiuli Luo<sup>1</sup> · Zhiheng Cheng<sup>1,3</sup> · Yuling Zhang<sup>1</sup> · Xijin Xu<sup>1,4</sup> 

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## Abstract

Polycyclic aromatic hydrocarbon (PAH) exposure alters immunological responses. Research concerning PAH exposure on intestinal immunity of children in electronic waste (e-waste) areas is scarce. The aim of this study was to evaluate the effects of polycyclic aromatic hydrocarbon (PAH) pollutants on intestinal mucosal immunity of children in e-waste areas. Results showed higher hydroxylated PAH (OH-PAH) concentrations in e-waste-exposed children, accompanied with higher sialyl Lewis A (SLA) level, absolute lymphocyte and monocyte counts, decreased of percentage of CD4<sup>+</sup> T cells, and had a higher risk of diarrhea. OH-PAH concentrations were negative with child growth. 1-OHNap mediated through WBCs, along with 1-OHPyr, was correlated with an increase SLA concentration. 2-OHFlu, 1-OHPhe, 2-OHPhe, 1-OHPyr, and 6-OHChr were positively correlated with secretory immunoglobulin A (sIgA) concentration. Our results indicated that PAH pollutants caused inflammation, affected the intestinal epithelium, and led to transformation of microfold cell (M cell). M cells initiating mucosal immune responses and the subsequent increasing sIgA production might be an adaptive immune respond of children in the e-waste areas. To our knowledge, this is the first study of PAH exposure on children intestinal immunity in e-waste area, showing that PAH exposure plays a negative role in child growth and impairs the intestinal immune function.

**Keywords** Polycyclic aromatic hydrocarbon · E-waste · Secretory immunoglobulin A · Sialyl Lewis A · Intestinal immunity · Adaptive immunity

## Introduction

Polycyclic aromatic hydrocarbons (PAHs) are listed as a kind of priority environmental pollutants with public health concerns

(Ramirez et al. 2011). Existing studies show that diabetes, metabolic syndrome, cardiovascular disease, asthma, dyslipidemia, hematology, neurobehavioral disorders, and cancer are related to exposure to PAHs (Hu et al. 2018; Karimi et al. 2015; Kim et al. 2021; Manoli et al. 2016; Rengarajan et al. 2015; Roshandel et al. 2012; Yang et al. 2014; Yilmaz et al. 2007; Wang et al. 2020a; Zhang et al. 2020a). Studies also reveal a link between PAHs and gastrointestinal (GI) symptoms and diseases and even GI tumors (Bansal and Kim 2015; Diggs et al. 2011; Gunter et al. 2007; Henkler et al. 2012; Poirier et al. 2019; Prince 2015; Roshandel et al. 2012). These findings strongly suggest an association between PAH exposure and impairment of the GI tract.

The GI tract is the main organ providing an internal barrier against environmental exposure and plays an important role in the physical and immune barriers to entry of harmful compounds in the body (Arnal and Lalles 2016; Ghosh et al. 2020). Breaking the epithelial barrier or even a minor disorder can lead to serious pathological consequences, including infection and inflammation (Citi 2018). Mounting studies imply an association between PAHs and microflora. By altering bacterial

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communities and interrupting the function of the intestinal microflora, PAHs can cause intestinal inflammatory disorders and immune respond (Defois et al. 2018; Mantey et al. 2014; Roslund et al. 2019; Roslund et al. 2020). In vitro and in vivo studies have also shown that PAHs can affect the immune system (Abdel-Shafy and Mansour 2016; Kim et al. 2013). However, very few studies have investigated alterations of the GI immune system (Abdel-Shafy and Mansour 2016).

To maintain gut immune homeostasis, there are multiple layers of defense in the intestinal mucosa, including innate and adaptive defenses (Ren et al. 2016). M cells mediate antigen uptake and specific secretory immunoglobulin A (sIgA) production which play a vital role in adaptive defenses (Kobayashi et al. 2019). M cells initiate mucosal immune responses by active phagocytosis and transcytosis of luminal bacteria and antigen presentation to dendritic cells (DCs) in the underlying lymphoid follicles (Ohno 2016). T cells and B cells are activated when DCs present antigen. Mucosal B cells then undergo IgA class switch recombination (CSR), migrate into the lamina propria and mature into IgA-producing plasma cells to produce IgA (Li et al. 2020; Liu et al. 2013). IgA binds to the polymeric immunoglobulin receptor (pIgR) and is transported across the cell to the lumen to form the molecule sIgA (Li et al. , 2020).

SLA is a lectin largely restricted to M cells within epithelial tissues (Giannasca et al. 1999; Ragupathi et al. 2009). During inflammation, M cells are induced in the intestinal tract and their number increases in inflamed mucosa (Lugering et al. 2004). In vitro, epithelial cells are transformed to cells with an M cell-like morphology and upregulate SLA antigen production (Gullberg et al. 2000). It has been observed that an increase in IgA level is associated with the increase in SLA level in colon disease (Iarumov et al. 1998; Jasim et al. 2008). All these results suggest a link between M cell differentiation and sIgA production in intestinal inflammation.

Although epidemiological studies have revealed a close link between PAH exposure and human digestive disease, the underlying mechanisms remain unexplored (Mantey et al. , 2014; Shiue 2016). Previous studies on PAH exposure and serum IgA expression are not consistent (Gao et al. 2014; Jeng et al. 2011; Karakaya et al. 1999; Szczeklik et al. 1994). Currently, limited studies of PAH exposure and human intestinal immunity have been about occupational exposure, and most of those studies have focused on IgA levels. Since serum IgA is monomolecular and sIgA is multimolecular, serum IgA cannot fully reflect the mucosal immunity (Li et al. , 2020). According to the literature, most serum sIgA probably originated from the digestive tract, and its levels of determination can be the most direct way to assess the amount of sIgA secretion in digestive tract (Pérez-Griera et al. 2017). The association between PAH exposure and serum sIgA level has not been studied, especially for the children in e-waste areas.

Guiyu is an e-waste recycling town located in Guangdong province, in southeast China, and has a more than 40-year

history of e-waste disposal (Zeng et al. 2018). It has been reported that environmental medias surrounding e-waste dismantling areas extremely contaminated by PAHs from thermal recycling activities (Liu et al. 2020; Wang et al. 2020a). Our previous studies have shown that local residents in Guiyu are exposed to PAHs and have health problems (Guo et al. 2012; Huang et al. 2020; Wang et al. 2020a; Xu et al. 2013; Xu et al. 2015; Zeng et al. 2020; Zheng et al. 2019). Previous studies shows PAH exposure affects immune system (Abdel-Shafy and Mansour 2016; Burchiel and Luster 2001; Dupuy et al. 2014; Ekhtator et al. 2018; Gou et al. 2017; Kim et al. , 2013). To better understand the relationship between PAH exposure and intestinal immunity, we recruited children from Guiyu and Haojiang (as a reference area located 31.6 km to the east of Guiyu) for the current study. We hypothesize that PAH exposure may cause GI tract inflammation and lead to M cell differentiation, which may consequently alter intestinal immunity.

## Materials and methods

### Study population

A total of 232 children (2-7 years old), all residents in Guiyu and Haojiang for more than 1 year, were included in this study (exposed group  $n = 119$  vs. reference group  $n = 113$ ). All children were recruited from two kindergartens during November to December 2018 and were free from general medical conditions and diseases. Apart from e-waste pollution, the two regions are very similar in ethnicity, cultural background, and population. Informed consent with a questionnaire on general characteristics, dwelling environment, children's living habits, family history, monthly household income, and parental educational level was obtained from the parents or guardians of all participants. This study was approved by the Human Ethics Committee of Shantou University Medical College, China. As previously described, fasting venous blood was collected by a nurse. The whole blood was used to measure immune cells and serum was used to measure SLA and sIgA. The rest of the serum was aliquoted and stored at  $-80\text{ }^{\circ}\text{C}$  until analysis (Zheng et al. 2019). A 15 mL urine were collected into a polypropylene conical centrifuge tube from children after getting up in the morning and preserved in  $-20\text{ }^{\circ}\text{C}$  until PAH metabolite measurement (Dai et al. 2019).

### Measurement of PAH metabolites in urine

Eleven urinary PAH metabolites (1-hydroxynaphthalene (1-OHNaP), 2-hydroxynaphthalene (2-OHNaP), 2-hydroxyfluorene (2-OHFlu), 9-hydroxyfluorene (9-OHFlu), 1-hydroxyphenanthrene (1-OHPhe), 2-hydroxyphenanthrene (2-OHPhe), 3-hydroxyphenanthrene (3-OHPhe), 4-

hydroxyphenanthrene (4-OHPhe), 9-hydroxyphenanthrene (9-OHPhe), 1-hydroxypyrene (1-OHPyr), and 6-hydroxychrysene (6-OHChr)) were measured by gas chromatography/mass spectrometry (GC/MS, 7890A-5975C Agilent Technologies) according to previous studies, with electron ionization used in selected ion monitoring mode (Campo et al. 2008; Cheng et al. 2020; Dai et al. 2019; Huang et al. 2020; Huo et al. 2019; Wang et al. 2020b; Zheng et al. 2019). Methods for QA/QC were based on our previously published methods with minor modifications (Cheng et al. 2020; Dai et al. 2019).  $\Sigma$ OH-PAHs was defined as the sum of the eleven congeners in urine.  $\Sigma$ OHNap was defined as the sum of 1-OHNap and 2-OHNap.  $\Sigma$ OHFlu was defined as the sum of 2-OHFlu and 9-OHFlu.  $\Sigma$ OHPhe was defined as the sum of five OHPhe congeners in urine.

### General physical tests and biological measurements

General physical examinations, including height, weight, and chest circumference, were performed by trained physician as described previously (Dai et al. 2019; Wang et al. 2020b; Xu et al. 2015; Zeng et al. 2020). A Sysmex XE-2100 automatic hematology analyzer was used for determining the white blood cell count in peripheral blood. The serum levels of SLA and sIgA were measured with a CA19-9/Sialyl Lewis A (Human) ELISA Kit (KA0207, Abnova, Taiwan) and a Secretary IgA (Human) ELISA Kit (KA3980, Abnova, Taiwan). Sensitivity was 10 U/mL and 0.6  $\mu$ g/mL, respectively. ELISAs were performed following the manufacturer's instructions.

### Flow cytometry

To determine the B lymphocytes (CD3<sup>-</sup>CD19<sup>+</sup>) and CD3<sup>+</sup>CD4<sup>+</sup>CD8<sup>-</sup>T (CD4<sup>+</sup>T cells) cell phenotype, 100  $\mu$ L whole blood was mixed with appropriate volume of the following monoclonal antibodies: CD3-APC-Cy7, CD4-PE-CF594, CD8-FITC, and CD19-PE-Cy7 (BD Bioscience, USA) and incubated for 15 min away from light at room temperature, then 2 mL of 1 $\times$  lysing solution (BD Bioscience, USA) was added and vortexed gently and incubated for 10 min away from light at room temperature. After centrifugation at 500g for 5 min, the supernatant was discarded, the cells were washed twice with 2 mL of 1 $\times$  PBS, followed by resuspension in 500  $\mu$ L of 1 $\times$  PBS. Cells were analyzed by FACS using an CYTEK Aurora flowcytometer (CYTEK Biosciences inc., USA). Data was analyzed with Spectro Flo (CYTEK Biosciences inc., USA).

### Statistical analysis

Data were analyzed by SPSS (Statistical Package for Social Sciences) version 22. All data were expressed as median and ranges or mean and standard deviation. We used the Pearson

chi-square test, independent sample *t* test, or Mann-Whitney *U* test to assess demographic and other characteristic differences between the two groups. Spearman's correlation analysis was performed to assess the relevant factors contributing to urinary PAH metabolites and the effects of PAH metabolites on the other indicators. Variables with skewed distributions were ln-transformed prior to regression and mediation analysis. *P* < 0.05 was set as the significance level in a two-tailed test.

## Results

### Basic characteristics of the study population

A total of 232 children were enrolled in the study (Table 1). Children in the two groups have no significant differences in gender and age (*P* > 0.05). The weight, height, chest circumference, and BMI of children are lower in the exposed group (*P* < 0.05). Children in the exposed group reside in a poorer residential environment, and the majority of the children reside in a residence near a road or an e-waste site or live in family workshops and have a family member who smoking (all *P* < 0.05). All these provide potential exposure to environmental pollution. The education level of the parents and monthly household income are lower for exposed children. Moreover, children in the exposed group more commonly display irritable bowel symptoms and diarrhea (more than three movements of loose stools a day) compared to the reference (odds ratio (OR) = 2.21; 95% CI: 1.16, 4.21; *P* = 0.014).

### Urinary PAH metabolite concentrations and factors influencing OH-PAHs

Except for 9-OHFlu, 2-OHPhe, 3-OHPhe, and 9-OHPhe, the other seven urinary OH-PAH concentrations are significantly higher in the exposed than the reference group (Table 2). Spearman correlation analysis showed that most urinary PAH metabolites are highly correlated with BMI, height, weight, and chest circumference (Table 3). In order to investigate the potential influencing factors of PAH exposure, a Spearman correlation analysis is performed (Table 4). Results show that urinary 1-OHNap concentration is positively correlated with e-waste contact, family workshops, residence within 50 m from an e-waste site and family member smoking. Urinary 1-OHNap,  $\Sigma$ OHNap, and  $\Sigma$ OHPAHs concentrations are negatively correlated with distance between residence and road, and urinary 1-OHNap, 1-OHPyr,  $\Sigma$ OHNap,  $\Sigma$ OHPhe, and  $\Sigma$ OHPAH concentrations are negatively correlated with the educational level of children's parents.

**Table 1** General characteristics of the study population

	<i>N</i>	Reference group	<i>N</i>	Exposed group	<i>P</i>
Gender (boys/girls)	113	68 (60.2%)/45 (39.8%)	119	66 (55.5%)/53 (44.5%)	0.467 <sup>a</sup>
Age (median (IQR), years)	113	4.88 (4.37, 5.80)	119	5.11 (4.39, 5.76)	0.431 <sup>b</sup>
Height (mean ± SD, cm)	113	109.29 ± 7.24	119	107.13 ± 7.52	0.027* <sup>b</sup>
Weight (median (IQR), kg)	113	18.50 (16.50, 20.25)	119	16.50 (15.00, 19.00)	0.000* <sup>b</sup>
BMI (median (IQR), kg/m <sup>2</sup> )	113	15.49 (14.71, 16.30)	118	14.93 (13.87, 15.75)	0.000* <sup>b</sup>
Chest circumference (median (IQR), cm)	113	52.50 (50.40, 54.95)	116	51.25 (49.63, 53.58)	0.026* <sup>b</sup>
Contact with electronic waste (yes/no)	113	12 (11.65%)/101 (89.38%)	119	33 (27.73%)/86 (72.27%)	0.002* <sup>a</sup>
Diarrhea (never/1~2 times monthly)	109	91 (83.5%)/18 (16.5%)	115	80 (70.7%)/35 (29.3%)	0.020* <sup>a</sup>
Distance between residence and road ( <i>n</i> (%), m)	111		119		0.000* <sup>a</sup>
< 10		15 (13.52%)		48 (40.3%)	
~ 50		30 (27.03%)		34 (28.6%)	
~ 100		24 (21.62%)		22 (18.5%)	
> 100		42 (37.83%)		15 (12.6%)	
Residence within 50 m from an e-waste site (yes/no)	112	3 (2.68%)/109 (97.32%)	116	27 (23.28%)/89 (76.72%)	0.000* <sup>a</sup>
Residence as a workshop (yes/no)	110	6 (5.5%)/104 (94.5%)	119	32 (26.9%)/87 (73.1%)	0.000* <sup>a</sup>
Family member daily cigarette consumption ( <i>n</i> (%))	112		118		0.032* <sup>a</sup>
Non-smoking		56 (50.0%)		40 (33.9%)	
~ 2 cigarettes		12 (10.7%)		24 (20.3%)	
~ 10 cigarettes		15 (13.4%)		20 (17.0%)	
~ 20 cigarettes		25 (22.3%)		23 (19.5%)	
> 20 cigarettes		4 (3.6%)		11 (9.3%)	
Father's educational level ( <i>n</i> (%))	113		119		0.000* <sup>a</sup>
Middle school or lower		23 (20.4%)		90 (75.6%)	
Secondary school		19 (16.8%)		8 (6.7%)	
High school		17 (15.0%)		12 (10.1%)	
College/university		54 (47.8%)		9 (7.6%)	
Mother's educational level ( <i>n</i> (%))	113		118		0.000* <sup>a</sup>
Middle school or lower		31 (27.4%)		87 (73.7%)	
Secondary school		21 (18.6%)		10 (8.5%)	
High school		15 (13.3%)		9 (7.6%)	
College/university		46 (40.7%)		12 (10.2%)	
Monthly household income ( <i>n</i> (%), Yuan)	111		112		0.000* <sup>a</sup>
< 3000		12 (10.8%)		11 (9.8%)	
~ 4500		27 (24.3%)		26 (23.2%)	
~ 6000		19 (17.1%)		46 (41.1%)	
> 6000		53 (47.8%)		29 (25.9%)	

BMI, body mass index; SD, standard deviation. Statistical significance, \* $P < 0.05$

<sup>a</sup> Analysis by the Pearson chi-square test

<sup>b</sup> Analysis by the independent-sample *t* test

### Peripheral leukocyte count and associations between urinary OH-PAHs

As shown in Fig. 1, the absolute lymphocyte and monocyte counts in the exposed group children are significantly higher than the reference group (both  $P < 0.05$ ). Both white blood cells (WBCs) and absolute neutrophil counts of children in the

exposed group tended to be higher than the reference group, but there is no significant difference (both  $P > 0.05$ ). Spearman correlation analysis show that urinary 1-OHNAp is positively correlated with WBCs and the absolute lymphocyte and monocyte counts ( $r_s = 0.142$ ,  $r_s = 0.147$ , and  $r_s = 0.206$ , respectively, all  $P < 0.05$ ); urinary 1-OHPyr is positively correlated with the absolute lymphocyte counts ( $r_s = 0.132$ ,  $P < 0.05$ )

**Table 2** Urinary PAH metabolite concentrations in e-waste-exposed and reference groups

	Reference group (N = 113)	Exposed group (N = 119)	P
OH-PAH (µmol/mmol Cr)/median (25th, 75th)			
Urine-Cre	12.48 (7.25, 26.07)	12.27 (6.89, 24.12)	0.777
1-OHNap	0.18 (0.08, 0.34)	0.85 (0.47, 1.51)	0.000*
2-OHNap	3.16 (1.58, 5.48)	4.49 (2.57, 6.99)	0.008*
2-OHFlu	0.54 (0.26, 0.93)	0.64 (0.34, 1.11)	0.046*
9-OHFlu	1.86 (0.82, 4.59)	2.31 (1.13, 5.25)	0.167
1-OHPhe	0.77 (0.38, 1.40)	1.18 (0.59, 2.05)	0.001*
2-OHPhe	0.89 (0.46, 1.53)	1.04 (0.59, 1.82)	0.067
3-OHPhe	1.66 (1.12, 2.53)	1.85 (1.22, 3.17)	0.081
4-OHPhe	0.87 (0.46, 1.50)	1.06 (0.63, 1.85)	0.015*
9-OHPhe	0.79 (0.36, 1.37)	0.87 (0.48, 1.73)	0.117
1-OHPyr	1.26 (0.60, 3.40)	2.49 (1.27, 5.49)	0.000*
6-OHChr	0.44 (0.23, 0.81)	0.56 (0.33, 0.95)	0.033*
ΣOHNap	3.46 (1.72, 5.82)	5.43 (3.14, 7.91)	0.000*
ΣOHFlu	2.50 (1.11, 5.64)	2.93 (1.47, 6.06)	0.136
ΣOHPhe	4.98 (2.75, 8.32)	6.15 (3.55, 10.80)	0.027*
ΣPHAs	13.77 (6.65, 23.01)	19.20 (11.18, 32.16)	0.002*

Cre, creatinine

Analysis by independent-sample *t* test

\**P* < 0.05

\*\**P* < 0.01

**Comparison of SLA, sIgA concentration, and immune cells**

The SLA concentration of children in the exposed group is significantly higher compared with the reference group (Fig. 2,

*P* < 0.05). The percentage of CD4<sup>+</sup> T cells is lower in the exposed group than the reference group (*P* < 0.05). The percentage of B cells tended to be higher in the exposed children, but no significance difference is obtained when compared with the reference (49.83% vs. 46.58%, *P* > 0.05).

**Table 3** Spearman analysis of the association between urinary PAH metabolites and characteristics of children

	BMI	High	Weight	Head circumference	Chest circumference
1-OHNap	− 0.219**	− 0.211**	− 0.307**	0.002	− 0.199**
2-OHNap	− 0.090	− 0.245**	− 0.250**	− 0.091	− 0.222**
2-OHFlu	− 0.078	− 0.178**	− 0.182**	− 0.036	− 0.162*
9-OHFlu	− 0.077	− 0.083	− 0.099	0.025	− 0.084
1-OHPhe	− 0.043	− 0.232**	− 0.219**	− 0.061	− 0.180**
2-OHPhe	− 0.067	− 0.228**	− 0.221**	− 0.090	− 0.200**
3-OHPhe	− 0.075	− 0.196**	− 0.201**	− 0.051	− 0.167*
4-OHPhe	− 0.092	− 0.205**	− 0.216**	− 0.051	− 0.187**
9-OHPhe	− 0.060	− 0.204**	− 0.190**	− 0.068	− 0.175**
1-OHPyr	− 0.075	− 0.237**	− 0.254**	− 0.092	− 0.187**
6-OHChr	− 0.077	− 0.223**	− 0.220**	− 0.091	− 0.186**
ΣOHNap	− 0.115	− 0.253**	− 0.272**	− 0.083	− 0.230**
ΣOHFlu	− 0.074	− 0.096	− 0.107	0.022	− 0.092
ΣOHPhe	− 0.070	− 0.221**	− 0.218**	− 0.064	− 0.190**
ΣOHPHAs	− 0.085	− 0.213**	− 0.219**	− 0.048	− 0.182**

\**P* < 0.05

\*\**P* < 0.01



**Table 4** Spearman correlation analysis between urinary metabolites of PAHs and related factors

Related factors	1-OHNap	1-OHPyr	ΣOHNap	ΣOHPh	ΣOHPAHs
	$r_s$	$r_s$	$r_s$	$r_s$	$r_s$
Electronic waste contact	0.207**	0.095	0.105	0.058	0.095
Residence as a workshop	0.237**	0.119	0.126	0.078	0.119
Distance between residence and road	− 0.271**	− 0.129	− 0.138*	− 0.093	− 0.133*
Residence within 50 m from an e-waste site	0.161*	0.092	0.063	0.065	0.070
Family member cigarette smoker	0.136*	0.057	0.079	0.041	0.061
Father's educational level	− 0.416**	− 0.217**	− 0.214**	− 0.149*	− 0.201**
Mother's educational level	− 0.343**	− 0.183**	− 0.210**	− 0.133**	− 0.186**
Monthly household income	0.073	0.095	0.095	0.101	0.097

\* $P < 0.05$ \*\* $P < 0.01$ 

### SLA and sIgA concentration of the 4- and 5-year-old children

Due to the fact that children included in this study had sample bias, we only analyzed the subgroup of 4- and 5-year-old children from the two areas (Fig. 3). Results show that for the 4-year-old children, both SLA and sIgA are both higher in the exposed group than reference (both  $P < 0.05$ ), but no significance difference in the 5-year-old children. The sIgA concentration of the 5-year-old children is lower than that in 4-year-old in the exposed group ( $P < 0.05$ ), but no significance difference in the reference group.

### Urinary PAH metabolite concentrations of the 4- and 5-year-old children

In the reference group, urinary ΣOHPAHs, 2-OHNap, 2-OHFlu, 1-OHPhe, 2-OHPhe, 4-OHPhe, 9-OHPh, 1-OHPyr, 6-OHChr, ΣOHNap, and ΣOHPh concentrations are higher in the 4-year-old group compared with that in the 5-year-old group (all  $P < 0.05$ ). In the exposed area, the urinary OH-PAH concentrations between the two age groups show no significant difference (Fig. 4).

### Associations between urinary OH-PAHs with sIgA, SLA, and B cell percentage

A multivariable linear regression model was performed to identify the contributions of OH-PAHs to sIgA, SLA, and B cell percentage in children (Fig. 5). Unadjusted regression analysis shows that 2-OHFlu, 1-OHPhe, 2-OHPhe, 1-OHPyr, and 6-OHChr are positively correlated with sIgA concentration; 1-OHPyr is positively correlated with SLA concentration, and 1-OHNap, 2-OHNap, 1-OHPyr, ΣOHNap, and ΣOH-PAHs are positively correlated with the percentage of B cells. The correlations between 1-OHPhe, 2-OHPhe, 1-

OHPyr, 6-OHChr, and sIgA concentration; 1-OHPyr level and SLA concentration; and 1-OHNap and B cell percentage remain significant after further adjustment for gender, age, BMI, contact with e-waste, parental educational level, and monthly household income.

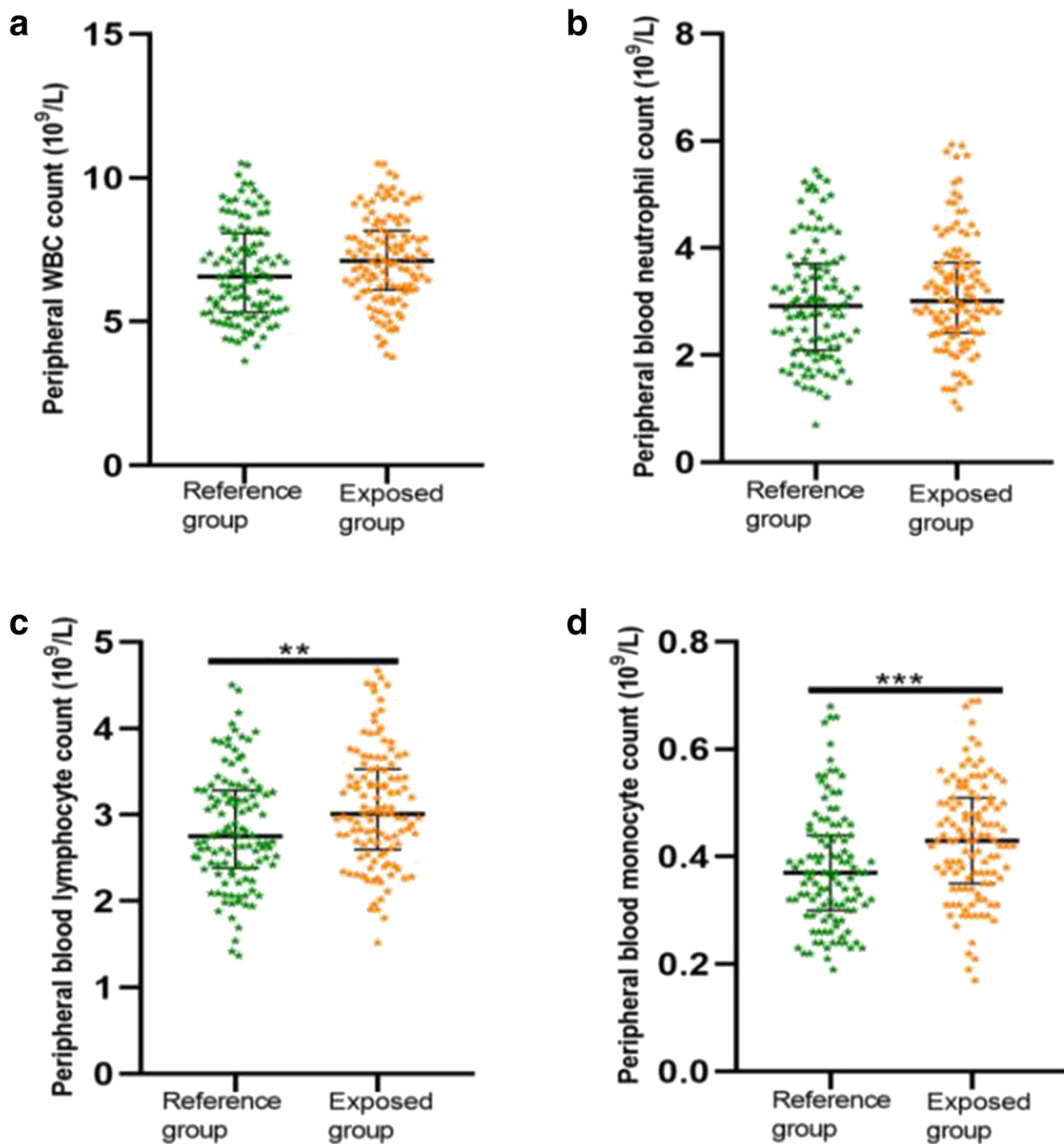
### Mediation effect analysis

Mediation model analysis shows that WBC concentration is a mediator in the correlation of 1-OHNap and SLA level (Fig. 6). In increasing WBC concentration, each 1-unit increase in 1-OHNap concentration is estimated to be correlated with a 0.020 μg/mL increase in SLA level. However, the direct effect of 1-OHNap on SLA level is not statically significant in mediation effect analysis, indicating that WBC concentration is completely responsible for the mediation.

### Discussion

In this study, we find that exposure group children had higher risk for diarrhea (odds ratio (OR) = 2.21) compared to the reference children. PAH exposure is negatively correlated with BMI, height, weight, and chest circumference of the children. Most of hydroxylated polycyclic aromatic hydrocarbon (OH-PAH) concentrations of children were higher in the exposed group and positively correlates with inflammation cells and intestinal immune biomarkers. Our results demonstrate that PAH exposure may be associated with gastrointestinal inflammation and immune responses. To our knowledge, this is the first study to provide evidence about the relationship between PAH exposure and intestinal immunity.

Urinary levels of OH-PAHs are widely used as a biomarker for estimating human exposure to PAHs from all routes of exogenous compounds (Lu et al. 2016; Yang et al. 2016). In



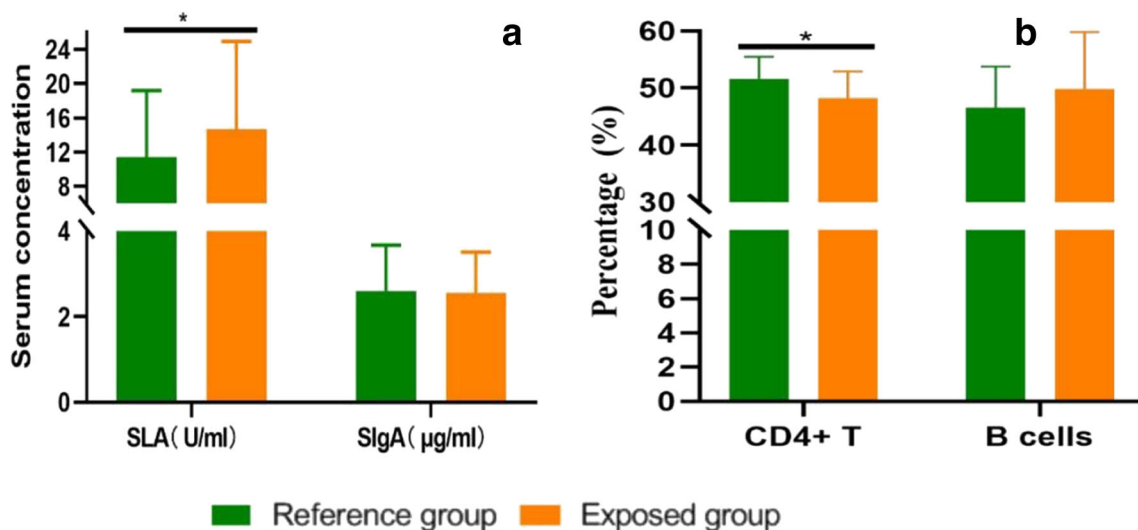
**Fig. 1** White blood cells, neutrophils, lymphocytes, and monocytes between the two groups. Reference group,  $n = 113$ ; exposed group,  $n = 119$ . **a** Results are presented as mean  $\pm$  standard deviation, analyzed by

independent-sample  $t$  test. **b–d** Results are presented as median (interquartile range), analyzed by the Mann-Whitney  $U$  test. \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$

this study, eight of eleven urinary OH-PAH concentrations of children are significantly higher in the exposed group than in the reference group, which is consistent with our previous studies indicating that children from e-waste recycling areas have elevated OH-PAH levels (Dai et al. 2019; Wang et al. 2020b; Xu et al. 2015; Zeng et al. 2020; Zheng et al. 2019). We find that 1-OHNap is negative with BMI, whereas most of the OH-PAHs negatively correlate with height, weight, and chest circumference, suggesting that PAH exposure has adverse effects on the development of children. As the recent studies described, we used biomonitoring studies to reveal the

relationships between urinary PAH metabolite levels and several lifestyle and/or demographic variables (Keir et al. 2020; Oliveira et al. 2020). The results show that urinary 1-OHNap metabolites are positively correlated with e-waste contact, family workshops, residence within 50 m from an e-waste site and family member smoking. We also find that OH-PAHs are negatively correlated with parental education levels, which corroborates previous studies suggesting that child health is associated with parental educational attainment reflecting knowledge-related assets, as well as other health-related characteristics (Carozza et al. 2010; Faught et al. 2019). In total,





**Fig. 2** Biomarkers in exposed and reference groups. **a** Serum SLA and sIgA concentration in the two groups (SLA: exposed group,  $n = 108$ ; reference group,  $n = 105$ ; sIgA: exposed group,  $n = 113$ ; reference group,  $n = 113$ ). **b** Percentage of CD4<sup>+</sup> and B cells between the two

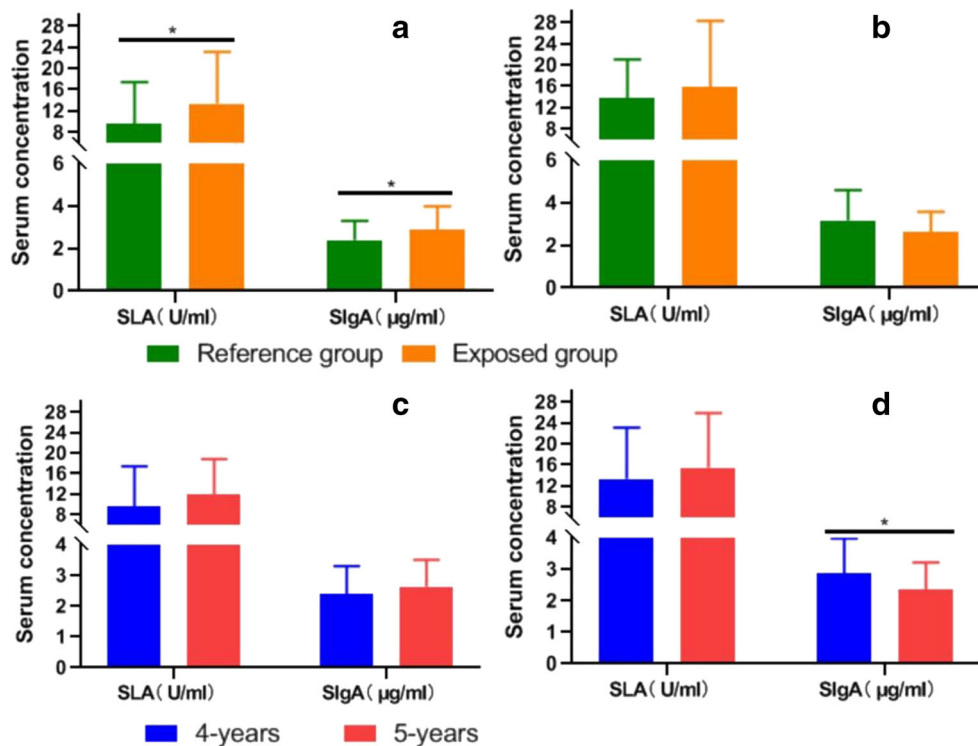
groups (exposed group,  $n = 113$ ; reference group,  $n = 119$ ). Results are presented as mean  $\pm$  standard deviation (median interquartile range), obtained with an independent-sample  $t$  test. \* $P < 0.05$

our findings indicate that pollution in e-waste areas and living habits affect children's urinary OH-PAH levels and have adverse effects on child growth.

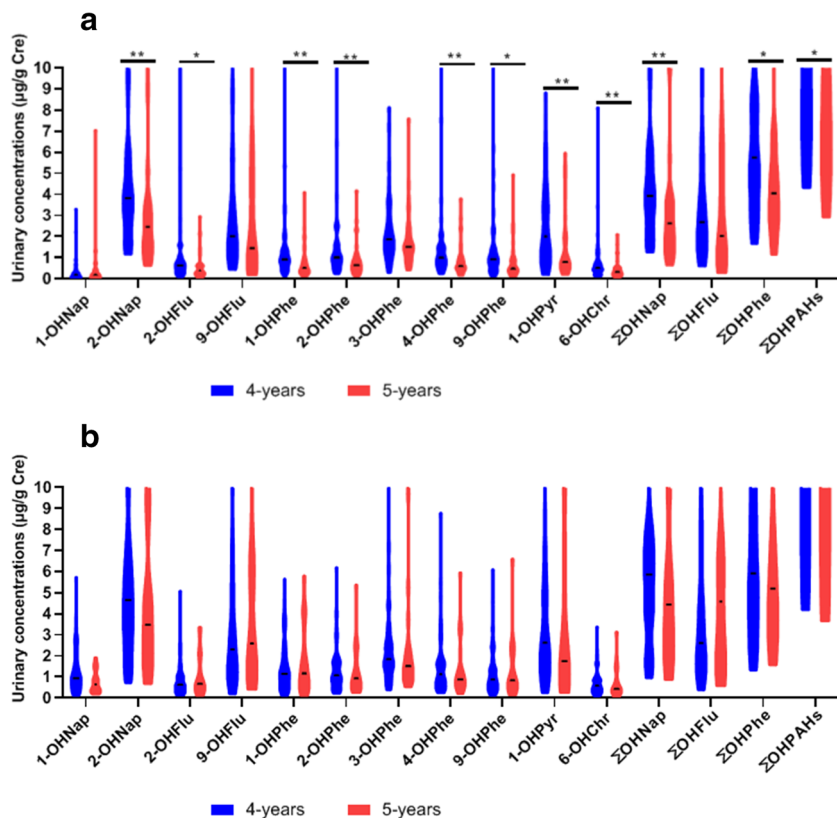
Many studies have reported associations of PAH exposure and inflammation (Alshaarawy et al. 2013; Kamal et al. 2014; Zhang et al. 2020b). PAH exposure is known to induce oxidative stress and promote the production of

reactive oxygen species (Dupuy et al. 2014; Huang et al. 2018; Jeng et al. 2011; Lu et al. 2016; Yang et al. 2014; Yilmaz et al. 2007; Zhang et al. 2020b). The results of the present study showed that 1-OHNap and 1-OHPyr correlate with inflammatory cells, and both compounds are elevated in the exposed group, consistent with our prior studies demonstrating that PAH exposure is associated with

**Fig. 3** Subgroup analysis serum SLA and sIgA concentration of 4- and 5-year-old children. **a** The 4-year-old group (SLA: exposed group,  $n = 31$ ; reference group,  $n = 42$ ; sIgA: exposed group,  $n = 38$ ; reference group,  $n = 45$ ). **b** The 5-year-old group (SLA: exposed group,  $n = 46$ ; reference group,  $n = 31$ ; sIgA: exposed group,  $n = 40$ ; reference group,  $n = 34$ ). **c** Reference group (SLA: 4-year-old group,  $n = 42$ ; 5-year-old group,  $n = 31$ ; sIgA: 4-year-old group,  $n = 45$ ; 5-year-old group,  $n = 34$ ). **d** Exposed group (SLA: 4-year-old group,  $n = 31$ ; 5-year-old group,  $n = 46$ ; sIgA: 4-year-old group,  $n = 38$ ; 5-year-old group,  $n = 40$ ). Results are presented as median (interquartile range) and analyzed by the Mann-Whitney  $U$  test. \* $P < 0.05$



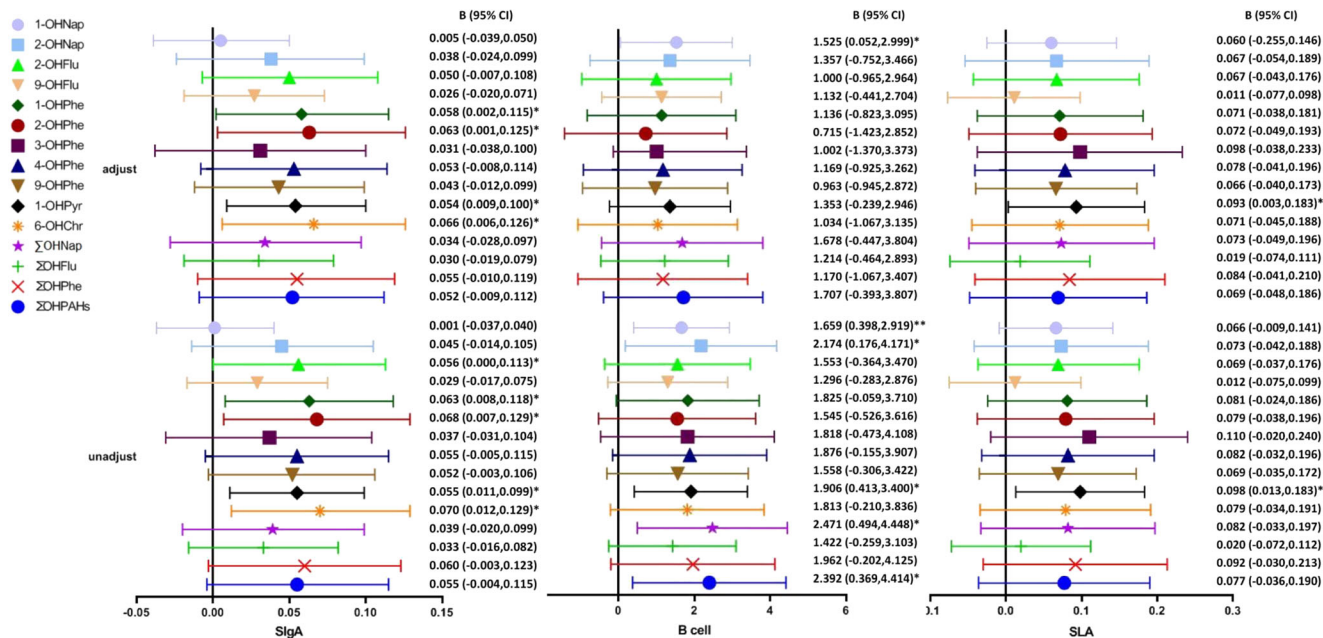
**Fig. 4** Subgroup analysis of urinary PAH metabolite concentrations ( $\mu\text{g/g Cre}$ ) of the 4- and 5-year-old children. **a** Reference group (4-year-old group,  $n = 45$ ; 5-year-old group,  $n = 36$ ). **b** Exposed group (4-year-old group,  $n = 38$ ; 5-year-old group,  $n = 46$ ). Analysis by independent-sample  $t$  test. \* $P < 0.05$ , \*\* $P < 0.01$



inflammation (Cheng et al. 2020; Dai et al. 2019; Zheng et al. 2019).

Relationship between PAH exposure and GI inflammation has been poorly studied in human. An animal study has

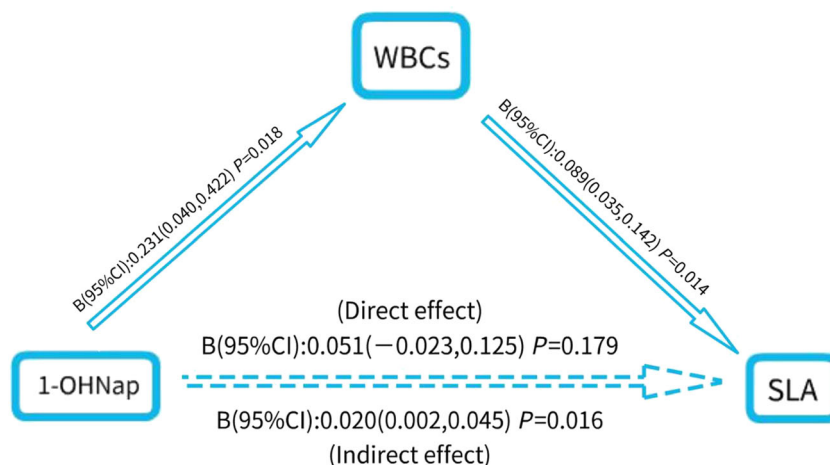
indicated that PAHs can be metabolized, and the metabolic products secreted into the GI tract to cause toxicity to epithelial cells (Mantey et al. 2014). Except for the direct impact on the digestive tract, PAHs can also affect intestinal flora to



**Fig. 5** Effect estimates and 95% confidence intervals for OH-PAHs with sIgA, SLA, and B cells. Adjusted model adjusting for gender, age, BMI, contact with e-waste, parental educational level, and monthly household

income. BMI, body mass index;  $B$ , unstandardized coefficient; CI, confidence interval. \* $P < 0.05$ , \*\* $P < 0.01$

**Fig. 6** Mediation effect of WBCs on the relationship between 1-OHNap and SLA. *B*, unstandardized coefficient; CI, confidence interval, BML, body mass index; 5000 bootstrap samples;  $n = 203$ .  $P < 0.05$  were considered statistically significant.



cause intestinal inflammatory disorders by secreting toxic metabolic products, altering bacterial communities and interrupting the functions of the intestinal microflora (Defois et al. 2018; Mantey et al. 2014; Roslund et al. 2019; Roslund et al. 2020). Under inflammatory conditions, intestinal epithelial cells may be converted to M cells (Gullberg and Soderholm 2006; Lugerling et al. 2004). We applied a linear mediation model with adjustment for factors to quantify the association between PAH exposure and SLA expression. Results show that 1-OHPyr affects the SLA level directly while 1-OHNap affects the SLA level through WBC mediation. For each 1-unit increase in concentration of 1-OHPyr and 1-OHNap, the SLA concentration increases 0.093  $\mu\text{g}/\text{mL}$  and 0.020  $\mu\text{g}/\text{mL}$ , respectively. Collectively, our results suggest that PAH exposure may be linked to GI inflammation and leads to M cell differentiation. Previous studies indicated that 1-OHNap present almost exclusively in vapor phase and is associated with inhalation exposure while 1-OHPyr as particulate matter with dietary exposure (Kim et al. 2021; Lao et al. 2018; Manoli et al. 2016; Nethery et al. 2012; Onyemauwa et al. 2009). We speculate that atmospheric PAH exposure causes a systemic inflammatory reaction and impairs the epithelium of the GI tract, whereas dietary PAH exposure directly modulates the gastrointestinal immune response, and both lead to M cell differentiation in the intestinal epithelium, as manifested by increased SLA concentrations in children.

Diarrhea is the manifestation of a disturbed gut environment as a symptom of an intestinal tract infection usually caused by a host of pathogens, which most likely results from disturbances in antigen-specific mucosal immune responses (Dong et al. 2017; Nagai et al. 2019; Yaya et al. 2018). We find that children in exposure group had higher risk for diarrhea (odds ratio (OR) = 2.21) compared to the reference. Previous studies have shown that PAH exposure was associated with suppression of T cell proliferation and decreased the percentage of  $\text{CD4}^+$  T cells, while lower numbers of  $\text{CD4}^+$  T cells are predictive of chronic diarrhea (Gou et al. 2017; Lauer et al. 2019; Navin et al.

1999). In this study, the percentage of  $\text{CD4}^+$  T cell decrease in the exposed group may associate with PAH exposure, which may be the reason for the diarrhea in exposed children. The correlation and regression analysis for OHPAHs and B cells is also consistent with a prior study reporting that exposure to PAHs might affect the differentiation of B cells (Huang et al. 2018). Together, our results suggest that PAH exposure may impair intestinal immune function, raise the risk of GI tract pathogen infection, lead children diarrhea, and favor B cell differentiation as an adaptive response.

Previous studies indicate that PAH exposure alters immunological responses and changes the expression of serum IgA. However, those results are not consistent. An increase of serum IgA level has been suggested in bitumen workers exposed to PAHs compared to the control group, but this disparity was not significant (Karakaya et al. 1999). By contrast, Jeng et al. (2011) found an inverse association between levels of PAHs and IgA (Jeng et al. 2011). Szczeklik et al. (1994) also found that workers chronically exposed to PAHs had depression of mean IgA levels (Szczeklik et al. 1994). Gao et al. (2014) showed that individuals exposed to high levels of PAHs had significantly lower mean IgA level (Gao et al. 2014). Limited studies about associations between PAH exposure and serum IgA expression have yielded inconsistent findings. All of those studies focused on adult occupational exposure, by analyzing the association between PAH exposure with serum IgA, which cannot totally reflect the mucosal immunity. According to the literature, serum sIgA is presumed to be a reliable indicator of mucosal immunity and the increase levels are evidence of subclinical intestinal compromise (Arias et al. 2020; Pérez-Griera et al. 2017). Here, we explored the relationship of PAH exposure and serum sIgA in the children in an e-waste area, which has not been studied. Results show that 1-OHPhe, 2-OHPhe, 1-OHPyr, and 6-OHChr are estimated to be correlated with an increase in sIgA level, suggesting that PAH exposure might affect child mucosal immunity and elevate the level of serum sIgA.

Younger children are more seriously exposed to PAHs because they prefer to play and crawl around on the floor and ground and display hand-to-mouth behavior (Huang et al. 2019; Oliveira et al. 2019). Our results showing that the concentrations of most OHPAHs decrease in the 5-year-old children than 4-year-old children in the reference group supports this suggestion. However, in the exposed group, no significant difference of OHPAH concentration was observed between the 4- and 5-year-old children. We speculate that the high urine OHPAH levels of children in e-waste area are associated with the high concentration of PAHs in the environment. Even though behavioral changes with age can reduce PAH exposure, the urinary OH-PAH levels of older children remain high, indicating that environmental PAH pollution continues to pose a long-term serious threat to local children.

M cells have critical roles in intestinal sIgA production (Ren et al. 2016). For the 4-year-old, both SLA and sIgA are significantly elevated in the exposed group. Depending on various parameters, PAHs exert complex effects on the immune system resulting in immune suppression or immune potentiation (Abdel-Shafy and Mansour 2016). Low levels of PAH exposure may lead to immune enhancement or an adjuvant effect (Burchiel and Luster 2001). In the current study, we found that PAH exposure increases children's sIgA levels. The reason may be that occupational PAH exposure is more serious than lifestyle exposure of children in e-waste areas. In addition, children enrolled in this study are all healthy individuals, and the concentrations of PAHs are estimated to be low, even in the exposed group. The effects of low levels of PAHs on human health, particularly in children, are unknown (Ekhtor et al. 2018). We speculate that the increase level of sIgA might be an adaptive protective response of children to the external PAHs related to toxic intestinal inflammation, suggesting that mucosal immunity strengthens, to some degree, the protective mechanism against environmental irritants.

There are more toxic substances in e-waste site than reference area, which also had impact on children mucosal immunity (Fitch et al. 2020; Kish et al. 2013; Woodby et al. 2020). We hypothesize that except the PAH pollutants, other toxic substances may irritate and lead to initiate the mucosal immunity, with an increase of children's sIgA levels. Our result shows that compared with the 4-year-old children, the sIgA levels of the exposure group were decreased in the 5-year-old children (Fig. 3d). According to the literature, toddlers aged 2–4 years were estimated to have the highest exposure to contaminants and gradually decreased with the increase of chronic exposure duration (Rodriguez et al. 2008; Song et al. 2017). The decrease of sIgA levels of the 5-year-old children in the exposure group may be associated with reduced exposure to other e-waste pollutants. But in the reference group, both SLA and sIgA levels of the 4- and 5-year-old group showed no significance (Fig. 3c). Further analyses of the urine OH-

PAH levels show that most of them decrease in the 5-year-old group. We speculate that children urine OH-PAHs were estimated to be low and had less effect of mucosal immunity.

The limitations of this study are as follows. Firstly, though our study provides an association between PAHs and intestinal immune responses, this may not necessarily indicate a cause-and-effect relationship between PAH exposure and intestinal immune-mediated inflammation. Secondly, children included in this study have sample bias, due to the insufficient number of samples of the 2-, 3-, and 6-year-old children, we only compare the subgroups of the 4- and 5-year-old children for some parameters. We are not able to determine the trend of the influence of PAHs on mucosal immunity, so a large-sample follow-up observation is necessary. Thirdly, there is a wide variety of toxic substances in e-waste site. We only analyze the effect of PAH metabolites on mucosal immunity and only collected morning urine samples at one point. To confirm our findings, more samples of contaminants and multiple measurements of PAH metabolites are needed for analyses.

## Conclusion

In summary, this is the first study to identify the relationship between mucosal immune response and PAH exposure of children from an e-waste area. The results show diarrhea occurs more often in e-waste-exposed children, and PAH exposure has adverse effects on child growth. We also find that PAH exposure causes inflammation and leads to M cell differentiation, with subsequently initiating an adaptive immune response by secreting sIgA. Younger children are more susceptible to PAH exposure, especially in e-waste areas. These available data support the hypothesis that for young children in e-waste areas, low level of PAH exposure may lead to intestinal inflammation and alter the intestinal immune response, which may raise the risk of GI tract pathogen infection, lead children diarrhea, and affect development. The elevation of sIgA levels may be a protective immune response to PAH exposure. Even though behavioral changes with age can reduce PAH exposure, urinary OH-PAH levels remain high in older children in e-waste areas, suggesting PAH exposure poses a long-term health threat to the local children. It is necessary to take more preventive measures to further reduce organic pollutant exposure in e-waste areas and pay more attention to protect children from e-waste contamination.

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**Author contribution** GC: conceptualization, investigation, formal analysis, and writing original draft. XL: investigation and formal analysis. ZC:



data curation and investigation. YZ: investigation and project administration. XX and XH: review and editing, supervision, project administration, funding acquisition, and formal analysis. All authors read and approved the final manuscript.

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**Data availability** The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

## Declarations

**Ethics approval** This study was approved by the Human Ethics Committee of Shantou University Medical College, China. Informed consent was obtained from each child's parents or guardians.

**Consent to participate** All authors were participated in this work

**Consent for publication** All authors agree to publish.

**Conflict of interest** The authors declare that they have no conflict of interest

## References

- Abdel-Shafy H, Mansour M (2016) A review on polycyclic aromatic hydrocarbons: source, environmental impact, effect on human health and remediation. *Egypt J Pet* 25:107–123. <https://doi.org/10.1016/j.ejpe.2015.03.011>
- Alshaarawy O, Zhu MT, Ducatman A, Conway B, Andrew ME (2013) Polycyclic aromatic hydrocarbon biomarkers and serum markers of inflammation. A positive association that is more evident in men. *Environ Res* 126:98–104. <https://doi.org/10.1016/j.envres.2013.07.006>
- Arias I, Herrera D, Bautista-Molano W, Bello-Gualtero JM, De Avila J, Salas-Cuestas F, Romero-Sánchez C (2020) Increasing of SIgA serum levels may reflect subclinical intestinal involvement in non-radiographic axial and peripheral spondyloarthritis. *Clin Rheumatol* 40:1343–1351. <https://doi.org/10.1007/s10067-020-05369-w>
- Amal ME, Lalles JP (2016) Gut epithelial inducible heat-shock proteins and their modulation by diet and the microbiota. *Nutr Rev* 74:181–197. <https://doi.org/10.1093/nutrit/nuv104>
- Bansal V, Kim KH (2015) Review of PAH contamination in food products and their health hazards. *Environ Int* 84:26–38. <https://doi.org/10.1016/j.envint.2015.06.016>
- Burchiel SW, Luster MI (2001) Signaling by environmental polycyclic aromatic hydrocarbons in human lymphocytes. *Clin Immunol* 98:2–10. <https://doi.org/10.1006/clim.2000.4934>
- Campo L, Rossella F, Fustinoni S (2008) Development of a gas chromatography/mass spectrometry method to quantify several urinary monohydroxy metabolites of polycyclic aromatic hydrocarbons in occupationally exposed subjects. *J Chromatogr B Anal Technol Biomed Life Sci* 875:531–540. <https://doi.org/10.1016/j.jchromb.2008.10.017>
- Carozza SE, Puumala SE, Chow EJ, Fox EE, Horel S, Johnson KJ, McLaughlin CC, Reynolds P, Von Behren J, Mueller BA, Spector LG (2010) Parental educational attainment as an indicator of socioeconomic status and risk of childhood cancers. *Brit J Cancer* 103:136–142. <https://doi.org/10.1038/sj.bjc.6605732>
- Cheng Z, Huo X, Dai Y, Lu X, Hylkema MN, Xu X (2020) Elevated expression of AhR and NLRP3 link polycyclic aromatic hydrocarbon exposure to cytokine storm in preschool children. *Environ Int* 139:105720. <https://doi.org/10.1016/j.envint.2020.105720>
- Citi S (2018) Intestinal barriers protect against disease. *Science*. 359:1097–1098. <https://doi.org/10.1126/science.aat0835>
- Dai Y, Huo X, Cheng Z, Wang Q, Zhang Y, Xu X (2019) Alterations in platelet indices link polycyclic aromatic hydrocarbons toxicity to low-grade inflammation in preschool children. *Environ Int* 131:105043. <https://doi.org/10.1016/j.envint.2019.105043>
- Defois C, Ratel J, Garrait G, Denis S, Le Goff O, Talvas J, Mosoni P, Engel E, Peyret P (2018) Food chemicals disrupt human gut microbiota activity and impact intestinal homeostasis as revealed by in vitro systems. *Sci Rep* 8:11006. <https://doi.org/10.1038/s41598-018-29376-9>
- Diggs DL, Huderson AC, Harris KL, Myers JN, Banks LD, Rekhadevi PV, Niaz MS, Ramesh A (2011) Polycyclic aromatic hydrocarbons and digestive tract cancers: a perspective. *J Environ Sci Health C Environ Carcinog Ecotoxicol Rev* 29:324–357. <https://doi.org/10.1080/10590501.2011.629974>
- Dong YL, Han YN, Wang ZX, Qin ZM, Yang CY, Cao J, Chen YX (2017) Role of serotonin on the intestinal mucosal immune response to stress-induced diarrhea in weaning mice. *BMC Gastroenterol* 17:17. <https://doi.org/10.1186/s12876-017-0634-5>
- Dupuy C, Galland C, Devaux A, Bony S, Loizeau V, Danion M, Pichereau V, Fournier M, Laroche J (2014) Responses of the European flounder (*Platichthys flesus*) to a mixture of PAHs and PCBs in experimental conditions. *Environ Sci Pollut Res Int* 21:13789–13803. <https://doi.org/10.1007/s11356-014-2563-y>
- Ekhator OC, Udowelle NA, Igbiri S, Asomugha RN, Frazzoli C, Orisakwe OE (2018) Street foods exacerbate effects of the environmental burden of polycyclic aromatic hydrocarbons (PAHs) in Nigeria. *Environ Sci Pollut Res Int* 25:5529–5538. <https://doi.org/10.1007/s11356-017-0894-1>
- Faught EL, McLaren L, Kirkpatrick SI, Hammond D, Minaker LM, Raine KD, Olstad DL (2019) Socioeconomic disadvantage across the life course is associated with diet quality in young adulthood. *Nutrients*. 11. <https://doi.org/10.3390/nu11020242>
- Fitch MN, Phillippi D, Zhang Y, Lucero J, Pandey RS, Liu J, Brower J, Allen MS, Campen MJ, McDonald JD, Lund AK (2020) Effects of inhaled air pollution on markers of integrity, inflammation, and microbiota profiles of the intestines in Apolipoprotein E knockout mice. *Environ Res* 181:108913. <https://doi.org/10.1016/j.envres.2019.108913>
- Gao M, Li Y, Zheng A, Xue X, Chen L, Kong Y (2014) Lymphocyte oxidative stress/genotoxic effects are related to serum IgG and IgA levels in coke oven workers. *ScientificWorldJournal*. 2014:801346–801310. <https://doi.org/10.1155/2014/801346>
- Ghosh SS, Wang J, Yannie PJ, Ghosh S (2020) Intestinal barrier dysfunction, LPS translocation, and disease development. *J Endocr Soc* 4:bvz039. <https://doi.org/10.1210/jendso/bvz039>
- Giannasca PJ, Giannasca KT, Leichtner AM, Neutra MR (1999) Human intestinal M cells display the sialyl Lewis A antigen. *Infect Immun* 67:946–953. <https://doi.org/10.1128/IAI.67.2.946-953.1999>
- Gou P, Chang X, Ye Z, Yao Y, Nguyen PK, Hammond SK, Wang J, Liu S (2017) A pilot study comparing T-regulatory cell function among healthy children in different areas of Gansu, China. *Environ Sci Pollut Res Int* 24:22579–22586. <https://doi.org/10.1007/s11356-017-9907-3>
- Gullberg E, Soderholm JD (2006) Peyer's patches and M cells as potential sites of the inflammatory onset in Crohn's disease. *Ann N Y Acad Sci* 1072:218–232. <https://doi.org/10.1196/annals.1326.028>
- Gullberg E, Leonard M, Karlsson J, Hopkiss AM, Brayden D, Baird AW, Artursson P (2000) Expression of specific markers and particle transport in a new human intestinal M-cell model. *Biochem*



- Biophys Res Commun 279:808–813. <https://doi.org/10.1006/bbrc.2000.4038>
- Gunter MJ, Divi RL, Kulldorff M, Vermeulen R, Haverkos KJ, Kuo MM, Strickland P, Poirier MC, Rothman N, Sinha R (2007) Leukocyte polycyclic aromatic hydrocarbon-DNA adduct formation and colorectal adenoma. *Carcinogenesis*. 28:1426–1429. <https://doi.org/10.1093/carcin/bgm022>
- Guo YY, Huo X, Wu KS, Liu JX, Zhang YL, Xu XJ (2012) Carcinogenic polycyclic aromatic hydrocarbons in umbilical cord blood of human neonates from Guiyu, China. *Sci Total Environ* 427:35–40. <https://doi.org/10.1016/j.scitotenv.2012.04.007>
- Henkler F, Stolpmann K, Luch A (2012) Exposure to polycyclic aromatic hydrocarbons: bulky DNA adducts and cellular responses. *Exp Suppl* 101:107–131. [https://doi.org/10.1007/978-3-7643-8340-4\\_5](https://doi.org/10.1007/978-3-7643-8340-4_5)
- Hu C, Hou J, Zhou Y, Sun H, Yin W, Zhang Y, Wang X, Wang G, Chen W, Yuan J (2018) Association of polycyclic aromatic hydrocarbons exposure with atherosclerotic cardiovascular disease risk: a role of mean platelet volume or club cell secretory protein. *Environ Pollut* 233:45–53. <https://doi.org/10.1016/j.envpol.2017.10.042>
- Huang XJ, Zhou Y, Cui XQ, Wu XJ, Yuan J, Xie JG, Chen WH (2018) Urinary polycyclic aromatic hydrocarbon metabolites and adult asthma: a case-control study. *Sci Rep-Uk* 8:7658. <https://doi.org/10.1038/s41598-018-26021-3>
- Huang X, Deng X, Li W, Liu S, Chen Y, Yang B, Liu Q (2019) Internal exposure levels of polycyclic aromatic hydrocarbons in children and adolescents: a systematic review and meta-analysis. *Environ Health Prev Med* 24:50. <https://doi.org/10.1186/s12199-019-0805-9>
- Huang XF, Xu XJ, Dai YF, Cheng ZH, Zheng XB, Huo X (2020) Association of prenatal exposure to PAHs with anti-Mullerian hormone (AMH) levels and birth outcomes of newborns. *Sci Total Environ* 723:138009. <https://doi.org/10.1016/j.scitotenv.2020.138009>
- Huo X, Wu Y, Xu L, Zeng X, Qin Q, Xu X (2019) Maternal urinary metabolites of PAHs and its association with adverse birth outcomes in an intensive e-waste recycling area. *Environ Pollut* 245:453–461. <https://doi.org/10.1016/j.envpol.2018.10.098>
- Iarumov N, Ignatov A, Viiachki I (1998) The pre- and postoperative monitoring of the immunological indices and tumor markers in colorectal carcinoma. *Khirurgiia (Sofia)* 51:42–48
- Jasim SM, Salim SA-d, Reda MM (2008) A study of carbohydrate antigen 19-9 level in patients with benign and malignant colorectal tumors in relation to the level of immunoglobulins. *Iraqi J Med Sci* 6:104–111
- Jeng HA, Pan CH, Diawara N, Chang-Chien GP, Lin WY, Huang CT, Ho CK, Wu MT (2011) Polycyclic aromatic hydrocarbon-induced oxidative stress and lipid peroxidation in relation to immunological alteration. *Occup Environ Med* 68:653–658. <https://doi.org/10.1136/oem.2010.055020>
- Kamal A, Malik RN, Martellini T, Cincinelli A (2014) PAH exposure biomarkers are associated with clinico-chemical changes in the brick kiln workers in Pakistan. *Sci Total Environ* 490:521–527. <https://doi.org/10.1016/j.scitotenv.2014.05.033>
- Karakaya A, Yücesoy B, Turhan A, Erdem O, Burgaz S, Karakaya AE (1999) Investigation of some immunological functions in a group of asphalt workers exposed to polycyclic aromatic hydrocarbons. *Toxicology*. 135:43–47. [https://doi.org/10.1016/S0300-483X\(99\)00048-7](https://doi.org/10.1016/S0300-483X(99)00048-7)
- Karimi P, Peters KO, Bidad K, Strickland PT (2015) Polycyclic aromatic hydrocarbons and childhood asthma. *Eur J Epidemiol* 30:91–101. <https://doi.org/10.1007/s10654-015-9988-6>
- Keir JLA, Cakmak S, Blais JM, White PA (2020) The influence of demographic and lifestyle factors on urinary levels of PAH metabolites—empirical analyses of Cycle 2 (2009–2011) CHMS data. *J Expo Sci Env Epid* 31:386–397. <https://doi.org/10.1038/s41370-020-0208-4>
- Kim KH, Jahan SA, Kabir E, Brown RJ (2013) A review of airborne polycyclic aromatic hydrocarbons (PAHs) and their human health effects. *Environ Int* 60:71–80. <https://doi.org/10.1016/j.envint.2013.07.019>
- Kim SS, Vuong AM, Dietrich KN, Chen A (2021) Proximity to traffic and exposure to polycyclic aromatic hydrocarbons in relation to Attention Deficit Hyperactivity Disorder and conduct disorder in U.S. children. *Int J Hyg Environ Health* 232:113686. <https://doi.org/10.1016/j.ijheh.2020.113686>
- Kish L, Hotte N, Kaplan GG, Vincent R, Tso R, Gänzle M, Rioux KP, Thiesen A, Barkema HW, Wine E, Madsen KL (2013) Environmental particulate matter induces murine intestinal inflammatory responses and alters the gut microbiome. *PLoS One* 8:e62220. <https://doi.org/10.1371/journal.pone.0062220>
- Kobayashi N, Takahashi D, Takano S, Kimura S, Hase K (2019) The roles of Peyer's patches and microfold cells in the gut immune system: relevance to autoimmune diseases. *Front Immunol* 10:2345. <https://doi.org/10.3389/fimmu.2019.02345>
- Lao JY, Xie SY, Wu CC, Bao LJ, Tao S, Zeng EY (2018) Importance of dermal absorption of polycyclic aromatic hydrocarbons derived from barbecue fumes. *Environ Sci Technol* 52:8330–8338. <https://doi.org/10.1021/acs.est.8b01689>
- Lauer FT, Parvez F, Factor-Litvak P, Liu X, Santella RM, Islam T, Eunus M, Alam N, Hasan A, Rahman M, Ahsan H, Graziano J, Burchiel SW (2019) Changes in human peripheral blood mononuclear cell (HPBMC) populations and T-cell subsets associated with arsenic and polycyclic aromatic hydrocarbon exposures in a Bangladesh cohort. *PLoS One* 14:e0220451. <https://doi.org/10.1371/journal.pone.0220451>
- Li Y, Jin L, Chen T (2020) The effects of secretory IgA in the mucosal immune system. *Biomed Res Int* 2020:2032057. <https://doi.org/10.1155/2020/2032057>
- Liu J, Cui H, Peng X, Fang J, Zuo Z, Deng J, Wang H, Wu B, Deng Y, Wang K (2013) Decreased IgA+ B cells population and IgA, IgG, IgM contents of the cecal tonsil induced by dietary high fluorine in broilers. *Int J Environ Res Public Health* 10:1775–1785. <https://doi.org/10.3390/ijerph10051775>
- Liu R, Ma S, Yu Y, Li G, Yu Y, An T (2020) Field study of PAHs with their derivatives emitted from e-waste dismantling processes and their comprehensive human exposure implications. *Environ Int* 144:106059. <https://doi.org/10.1016/j.envint.2020.106059>
- Lu SY, Li YX, Zhang JQ, Zhang T, Liu GH, Huang MZ, Li X, Ruan JJ, Kannan K, Qiu RL (2016) Associations between polycyclic aromatic hydrocarbon (PAH) exposure and oxidative stress in people living near e-waste recycling facilities in China. *Environ Int* 94:161–169. <https://doi.org/10.1016/j.envint.2016.05.021>
- Lugering A, Floer M, Lugering N, Cichon C, Schmidt MA, Domschke W, Kucharzik T (2004) Characterization of M cell formation and associated mononuclear cells during indomethacin-induced intestinal inflammation. *Clin Exp Immunol* 136:232–238. <https://doi.org/10.1111/j.1365-2249.2004.02438.x>
- Manoli E, Kouras A, Karagkiozidou O, Argyropoulos G, Voutsas D, Samara C (2016) Polycyclic aromatic hydrocarbons (PAHs) at traffic and urban background sites of northern Greece: source apportionment of ambient PAH levels and PAH-induced lung cancer risk. *Environ Sci Pollut Res Int* 23:3556–3568. <https://doi.org/10.1007/s11356-015-5573-5>
- Mantey JA, Rekhadevi PV, Diggs DL, Ramesh A (2014) Metabolism of benzo(a)pyrene by subcellular fractions of gastrointestinal (GI) tract and liver in Apc(Min) mouse model of colon cancer. *Tumour Biol* 35:4929–4935. <https://doi.org/10.1007/s13277-014-1647-0>
- Nagai M, Noguchi R, Takahashi D, Morikawa T, Koshida K, Komiyama S, Ishihara N, Yamada T, Kawamura YI, Muroi K, Hattori K, Kobayashi N, Fujimura Y, Hirota M, Matsumoto R, Aoki R, Tamura-Nakano M, Sugiyama M, Katakai T, Sato S, Takubo K, Dohi T, Hase K (2019) Fasting-refeeding impacts immune cell

- dynamics and mucosal immune responses. *Cell* 178:1072–1087.e14. <https://doi.org/10.1016/j.cell.2019.07.047>
- Navin T, Weber R, Vugia D, Rimland D, Roberts J, Addiss D, Visvesvara G, Wahlquist S, Hogan S, Gallagher L, Juranek D, Schwartz D, Wilcox C, Stewart J, Thompson S, Bryan R (1999) Declining CD4+ T-lymphocyte counts are associated with increased risk of enteric parasitosis and chronic diarrhea: results of a 3-year longitudinal study. *J Acquir Immune Defic Syndr Human Retrovirology* 20: 154–159. <https://doi.org/10.1097/00042560-199902010-00007>
- Nethery E, Wheeler AJ, Fisher M, Sjödin A, Li Z, Romanoff LC, Foster W, Arbuckle TE (2012) Urinary polycyclic aromatic hydrocarbons as a biomarker of exposure to PAHs in air: a pilot study among pregnant women. *J Expos Sci Environ Epidemiol* 22:70–81. <https://doi.org/10.1038/jes.2011.32>
- Ohno H (2016) Intestinal M cells. *J Biochem* 159:151–160. <https://doi.org/10.1093/jb/mvv121>
- Oliveira M, Slezakova K, Delerue-Matos C, Pereira MC, Morais S (2019) Children environmental exposure to particulate matter and polycyclic aromatic hydrocarbons and biomonitoring in school environments: a review on indoor and outdoor exposure levels, major sources and health impacts. *Environ Int* 124:180–204. <https://doi.org/10.1016/j.envint.2018.12.052>
- Oliveira M, Capelas S, Delerue-Matos C, Morais S (2020) Grill workers exposure to polycyclic aromatic hydrocarbons: levels and excretion profiles of the urinary biomarkers. *Int J Environ Res Public Health* 18. <https://doi.org/10.3390/ijerph18010230>
- Onyemauwa F, Rappaport SM, Sobus JR, Gajdosova D, Wu R, Waidyanatha S (2009) Using liquid chromatography-tandem mass spectrometry to quantify monohydroxylated metabolites of polycyclic aromatic hydrocarbons in urine. *J Chromatogr B Anal Technol Biomed Life Sci* 877:1117–1125. <https://doi.org/10.1016/j.jchromb.2009.02.067>
- Pérez-Griera J, Andreu-Ballester JC, Hueso Zarándieta A, García de la Asunción J, Masquefa Bondia S (2017) A quantitative enzyme-linked immunosorbent assay for quantification of secretory immunoglobulin A in serum. *J Immunoassay Immunochem* 38:67–71. <https://doi.org/10.1080/15321819.2016.1216443>
- Poirier MC, Lair S, Michaud R, Hernandez-Ramon EE, Divi KV, Dwyer JE, Ester CD, Si NN, Ali M, Loseto LL, Raverty SA, St Leger JA, Van Bonn WG, Colegrove K, Burek-Huntington KA, Suydam R, Stimmelmayer R, Wise JP, Wise SS, Beauchamp G, Martineau D (2019) Intestinal polycyclic aromatic hydrocarbon-DNA adducts in a population of beluga whales with high levels of gastrointestinal cancers. *Environ Mol Mutagen* 60:29–41. <https://doi.org/10.1002/em.22251>
- Prince U (2015) Environmental effects of polycyclic aromatic hydrocarbons. *J Nat Sci Res* 5(7):117–131
- Ragupathi G, Damani P, Srivastava G, Srivastava O, Sucheck SJ, Ichikawa Y, Livingston PO (2009) Synthesis of sialyl Lewis(x) (sLe(x)), CA19-9 and construction of an immunogenic sLe(x) vaccine. *Cancer Immunol Immunother* 58:1397–1405. <https://doi.org/10.1007/s00262-008-0654-7>
- Ramirez N, Cuadras A, Rovira E, Marce RM, Borrull F (2011) Risk assessment related to atmospheric polycyclic aromatic hydrocarbons in gas and particle phases near industrial sites. *Environ Health Perspect* 119:1110–1116. <https://doi.org/10.1289/ehp.1002855>
- Ren W, Wang K, Yin J, Chen S, Liu G, Tan B, Wu G, Bazer FW, Peng Y, Yin Y (2016) Glutamine-induced secretion of intestinal secretory immunoglobulin A: a mechanistic perspective. *Front Immunol* 7: 503. <https://doi.org/10.3389/fimmu.2016.00503>
- Rengarajan T, Rajendran P, Nandakumar N, Lokeshkumar B, Rajendran P, Nishigaki I (2015) Exposure to polycyclic aromatic hydrocarbons with special focus on cancer. *Asian Pac J Trop Biomed* 5:182–189. [https://doi.org/10.1016/S2221-1691\(15\)30003-4](https://doi.org/10.1016/S2221-1691(15)30003-4)
- Rodriguez C, Cook A, Devine B, Van Buynder P, Lugg R, Linge K, Weinstein P (2008) Dioxins, furans and PCBs in recycled water for indirect potable reuse. *Int J Environ Res Public Health* 5:356–367. <https://doi.org/10.3390/ijerph5050356>
- Roshandel G, Semmani S, Malekzadeh R, Dawsey SM (2012) Polycyclic aromatic hydrocarbons and esophageal squamous cell carcinoma. *Arch Iran Med* 15:713–722. <https://doi.org/10.12151/AIM.0013>
- Roslund MI, Rantala S, Oikarinen S, Puhakka R, Hui N, Parajuli A, Laitinen OH, Hyöty H, Rantalainen AL, Sinkkonen A, team, A (2019) Endocrine disruption and commensal bacteria alteration associated with gaseous and soil PAH contamination among daycare children. *Environ Int* 130:104894. <https://doi.org/10.1016/j.envint.2019.06.004>
- Roslund MI, Puhakka R, Grönroos M, Nurminen N, Oikarinen S, Gazzali AM, Cinek O, Kramná L, Siter N, Vari HK, Soinen L, Parajuli A, Rajaniemi J, Kinnunen T, Laitinen OH, Hyöty H, Sinkkonen A (2020) Biodiversity intervention enhances immune regulation and health-associated commensal microbiota among daycare children. *Sci Adv* 6:6. <https://doi.org/10.1126/sciadv.aba2578>
- Shiue I (2016) Urinary polyaromatic hydrocarbons are associated with adult celiac disease and kidney stones: USA NHANES, 2011–2012. *Environ Sci Pollut Res Int* 23:3971–3977. <https://doi.org/10.1007/s11356-015-5980-7>
- Song Y, Wang Y, Mao W, Sui H, Yong L, Yang D, Jiang D, Zhang L, Gong Y (2017) Dietary cadmium exposure assessment among the Chinese population. *PLoS One* 12:e0177978. <https://doi.org/10.1371/journal.pone.0177978>
- Szczeklik A, Szczeklik J, Galuszka Z, Musial J, Kolarzyk E, Targosz D (1994) Humoral immunosuppression in men exposed to polycyclic aromatic hydrocarbons and related carcinogens in polluted environments. *Environ Health Perspect* 102:302–304. <https://doi.org/10.1289/ehp.94102302>
- Wang Q, Xu X, Zeng Z, Hylkema MN, Cai Z, Huo X (2020a) PAH exposure is associated with enhanced risk for pediatric dyslipidemia through serum SOD reduction. *Environ Int* 145:106132. <https://doi.org/10.1016/j.envint.2020.106132>
- Wang Q, Xu X, Zeng Z, Zheng X, Ye K, Huo X (2020b) Antioxidant alterations link polycyclic aromatic hydrocarbons to blood pressure in children. *Sci Total Environ* 732:138944. <https://doi.org/10.1016/j.scitotenv.2020.138944>
- Woodby B, Schiavone ML, Pambianchi E, Mastaloudis A, Hester S N, Wood S M, Pecorelli A, Valacchi G (2020) Particulate matter decreases intestinal barrier-associated proteins levels in 3D human intestinal model. *Int J Environ Res Public Health* 17:3234. <https://doi.org/10.3390/ijerph17093234>
- Xu X, Yekeen TA, Xiao Q, Wang Y, Lu F, Huo X (2013) Placental IGF-1 and IGFBP-3 expression correlate with umbilical cord blood PAH and PBDE levels from prenatal exposure to electronic waste. *Environ Pollut* 182:63–69. <https://doi.org/10.1016/j.envpol.2013.07.005>
- Xu XJ, Liu JX, Huang CY, Lu FF, Chiung YM, Huo X (2015) Association of polycyclic aromatic hydrocarbons (PAHs) and lead co-exposure with child physical growth and development in an e-waste recycling town. *Chemosphere*. 139:295–302. <https://doi.org/10.1016/j.chemosphere.2015.05.080>
- Yang LL, Zhou Y, Sun HZ, Lai HP, Liu CY, Yan K, Yuan J, Wu TC, Chen WH, Zhang XM (2014) Dose-response relationship between polycyclic aromatic hydrocarbon metabolites and risk of diabetes in the general Chinese population. *Environ Pollut* 195:24–30. <https://doi.org/10.1016/j.envpol.2014.08.012>
- Yang B, Deng Q, Zhang W, Feng Y, Dai X, Feng W, He X, Huang S, Zhang X, Li X, Lin D, He M, Guo H, Sun H, Yuan J, Lu J, Hu FB, Zhang X, Wu T (2016) Exposure to polycyclic aromatic hydrocarbons, plasma cytokines, and heart rate variability. *Sci Rep* 6:19272. <https://doi.org/10.1038/srep19272>
- Yaya S, Hudani A, Udenigwe O, Shah V, Ekholuenetale M, Bishwajit G (2018) Improving water, sanitation and hygiene practices, and housing quality to prevent diarrhea among under-five children in Nigeria.

- Trop Med Infect Dis 3:41. <https://doi.org/10.3390/tropicalmed3020041>
- Yilmaz B, Ssempebwa J, Mackerer CR, Arcaro KF, Carpenter DO (2007) Effects of polycyclic aromatic hydrocarbon-containing oil mixtures on generation of reactive oxygen species and cell viability in MCF-7 breast cancer cells. *J Toxicol Environ Health Part A* 70:1108–1115. <https://doi.org/10.1080/15287390701208545>
- Zeng Z, Huo X, Zhang Y, Xiao Z, Zhang Y, Xu X (2018) Lead exposure is associated with risk of impaired coagulation in preschool children from an e-waste recycling area. *Environ Sci Pollut Res Int* 25:20670–20679. <https://doi.org/10.1007/s11356-018-2206-9>
- Zeng ZJ, Huo X, Wang QH, Wang CY, Hylkema MN, Xu XJ (2020) PM2.5-bound PAHs exposure linked with low plasma insulin-like growth factor 1 levels and reduced child height. *Environ Int* 138:105660. <https://doi.org/10.1016/j.envint.2020.105660>
- Zhang B, Pan BL, Zhao XY, Fu Y, Li XJ, Yang AM, Li Q, Dong J, Nie JS, Yang J (2020a) The interaction effects of smoking and polycyclic aromatic hydrocarbons exposure on the prevalence of metabolic syndrome in coke oven workers. *Chemosphere*. 247:125880. <https://doi.org/10.1016/j.chemosphere.2020.125880>
- Zhang H, Han Y, Qiu X, Wang Y, Li W, Liu J, Chen X, Li R, Xu F, Chen W, Yang Q, Fang Y, Fan Y, Wang J, Zhang H, Zhu T (2020b) Association of internal exposure to polycyclic aromatic hydrocarbons with inflammation and oxidative stress in prediabetic and healthy individuals. *Chemosphere*. 253:126748. <https://doi.org/10.1016/j.chemosphere.2020.126748>
- Zheng XB, Huo X, Zhang Y, Wang QH, Zhang YL, Xu XJ (2019) Cardiovascular endothelial inflammation by chronic coexposure to lead (Pb) and polycyclic aromatic hydrocarbons from preschool children in an e-waste recycling area. *Environ Pollut* 246:587–596. <https://doi.org/10.1016/j.envpol.2018.12.055>

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