

University of Groningen

Oxidative stress is associated with suspected non-alcoholic fatty liver disease and all-cause mortality in the general population

Damba, Turtushikh; Bourgonje, Arno R.; Abdulle, Amaal E.; Pasch, Andreas; Sydor, Svenja; van den Berg, Eline H.; Gansevoort, Ron T.; Bakker, Stephan J. L.; Blokzijl, Hans; Dullaart, Robin P. F.

Published in:
Liver International

DOI:
[10.1111/liv.14562](https://doi.org/10.1111/liv.14562)

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version
Publisher's PDF, also known as Version of record

Publication date:
2020

[Link to publication in University of Groningen/UMCG research database](#)

Citation for published version (APA):

Damba, T., Bourgonje, A. R., Abdulle, A. E., Pasch, A., Sydor, S., van den Berg, E. H., Gansevoort, R. T., Bakker, S. J. L., Blokzijl, H., Dullaart, R. P. F., van Goor, H., & Moshage, H. (2020). Oxidative stress is associated with suspected non-alcoholic fatty liver disease and all-cause mortality in the general population. *Liver International*, 40(9), 2148-2159. <https://doi.org/10.1111/liv.14562>

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: <https://www.rug.nl/library/open-access/self-archiving-pure/taverne-amendment>.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.



Oxidative stress is associated with suspected non-alcoholic fatty liver disease and all-cause mortality in the general population

Turtushikh Damba^{1,2} | Arno R. Bourgonje¹ | Amaal E. Abdulle³ | Andreas Pasch⁴ | Svenja Sydor⁵ | Eline H. van den Berg¹ | Ron T. Gansevoort⁶ | Stephan J. L. Bakker⁶ | Hans Blokzijl¹ | Robin P. F. Dullaart⁷ | Harry van Goor⁸ | Han Moshage^{1,9}

¹Department of Gastroenterology and Hepatology, University Medical Center Groningen, University of Groningen, Groningen, the Netherlands

²School of Pharmacy, Mongolian National University of Medical Sciences, University of Groningen, Ulaanbaatar, Mongolia

³Department of Internal Medicine, Division Vascular Medicine, University Medical Center Groningen, University of Groningen, Groningen, the Netherlands

⁴Institute for Physiology and Pathophysiology, Johannes Kepler University Linz, Linz, Austria

⁵Department of Gastroenterology, Hepatology, and Infectious Diseases, Otto von Guericke University Hospital Magdeburg, Magdeburg, Germany

⁶Department of Internal Medicine, Division Nephrology, University Medical Center Groningen, University of Groningen, Groningen, the Netherlands

⁷Department of Endocrinology, University Medical Center Groningen, University of Groningen, Groningen, the Netherlands

⁸Department of Pathology and Medical Biology, University Medical Center Groningen, University of Groningen, Groningen, the Netherlands

⁹Department of Laboratory Medicine, University Medical Center Groningen,

Abstract

Background & Aims: Non-alcoholic fatty liver disease (NAFLD) is characterized by excessive lipid accumulation, inflammation and an imbalanced redox homeostasis. We hypothesized that systemic free thiol levels, as a proxy of systemic oxidative stress, are associated with NAFLD.

Methods: Protein-adjusted serum free thiol concentrations were determined in participants from the Prevention of Renal and Vascular End-Stage Disease (PREVEND) cohort study (n = 5562). Suspected NAFLD was defined by the Fatty Liver Index (FLI \geq 60) and Hepatic Steatosis Index (HSI > 36).

Results: Protein-adjusted serum free thiols were significantly reduced in subjects with FLI \geq 60 (n = 1651). In multivariable logistic regression analyses, protein-adjusted serum free thiols were associated with NAFLD (FLI \geq 60) (OR per doubling of concentration: 0.78 [95% CI 0.64-0.96], P = .016) even when adjusted for potential confounding factors, including systolic blood pressure, diabetes, current smoking, use of alcohol and total cholesterol (OR 0.80 [95% CI 0.65-0.99], P = .04). This association lost its significance (OR 0.94 [95% CI 0.73-1.21], P = .65) after additional adjustment for high-sensitive C-reactive protein. Stratified analyses showed significantly differential associations of protein-adjusted serum free thiol concentrations with suspected NAFLD for gender (P < .02), hypertension (P < .001) and hypercholesterolemia (P < .003). Longitudinally, protein-adjusted serum free thiols were significantly

Abbreviations: ALT, alanine aminotransferase; AST, aspartate aminotransferase; BMI, body mass index; CVD, cardiovascular disease; FLI, fatty liver index; GGT, gamma-glutamyl-transferase; HCC, hepatocellular carcinoma; HIS, Hepatic Steatosis Index; hs-CRP, high-sensitive C-reactive protein; IBD, inflammatory bowel disease; NAFLD, non-alcoholic fatty liver disease; NASH, non-alcoholic steatohepatitis; PREVEND, Prevention of Renal and Vascular End-Stage Disease; RNS, reactive nitrogen species; ROS, reactive oxygen species; RSS, reactive sulphur species; SAAs, sulphur-based amino acids; T2D, type 2 diabetes; TG, triglycerides.

Turtushikh Damba and Arno R. Bourgonje contributed equally to this study.

Harry van Goor and Han Moshage contributed equally to this study.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2020 The Authors. Liver International published by John Wiley & Sons Ltd

University of Groningen, Groningen, the Netherlands

Correspondence

Han Moshage, Department of Gastroenterology and Hepatology, University Medical Center Groningen, University of Groningen, Hanzeplein 1, 9713 GZ, Groningen, The Netherlands. Email: a.j.moshage@umcg.nl

Funding information

The Dutch Kidney Foundation supported the infrastructure of the PREVENT program from 1997 to 2003 (Grant E.033). The University Medical Center Groningen supported the infrastructure from 2003 to 2006.

Handling Editor: Michelle Long

associated with the risk of all-cause mortality in subjects with NAFLD (FLI \geq 60) (HR 0.27 [95% CI 0.17-0.45], $P < .001$).

Conclusion: Protein-adjusted serum free thiol levels are reduced and significantly associated with all-cause mortality in subjects with suspected NAFLD. Quantification of free thiols may be a promising, minimally invasive strategy to improve detection of NAFLD and associated risk of all-cause mortality in the general population.

KEYWORDS

fatty liver index FLI, free thiols, NAFLD, oxidative stress

1 | INTRODUCTION

Non-alcoholic fatty liver disease (NAFLD) is defined as an abnormal accumulation of triglycerides (TG) in hepatocytes in the absence of excessive alcohol consumption. NAFLD is emerging as the most prevalent chronic liver disease in Western countries. NAFLD encompasses a spectrum of diseases that ranges from simple steatosis to non-alcoholic steatohepatitis (NASH), in combination with fibrosis. NASH can subsequently lead to cirrhosis with its known complications, such as hepatocellular carcinoma (HCC).¹ Many co-morbidities coincide with the development of NAFLD, such as obesity, insulin resistance and metabolic syndrome, including type 2 diabetes (T2D).^{2,3} In the general population, suspected NAFLD can be estimated by calculating proxies of the disease, including the Fatty Liver Index (FLI) or the Hepatic Steatosis Index (HSI). Both of these scoring systems are considered to be potential predictors for NAFLD and are based on prominent risk factors, including obesity indices, plasma triglycerides, gamma-glutamyl-transferase (GGT), body mass index (BMI) and liver transaminases.^{4,5}

A number of previous studies demonstrated that inflammation significantly contributes to the progression of NAFLD. During NAFLD, hepatocytes no longer tolerate the toxicity of accumulated fatty acids, resulting in dysfunction of cellular homeostasis, including mitochondrial β -oxidation and endoplasmic reticulum stress. Following this, an overproduction of endogenous reactive species (consisting of reactive oxygen species [ROS], reactive nitrogen species [RNS] and reactive sulphur species [RSS]) as well as an inflammatory signalling cascade in the liver is being generated.^{6,7} An increased production of reactive species subsequently leads to hepatocellular injury, which in turn results in secretion of inflammatory cytokines (TNF- α , IL-6, IL-10) and cellular death. The pro-inflammatory signalling pathways, increased β -oxidation in mitochondria and

peroxisomes involved in this process lead to dysregulation of antioxidant homeostasis.⁸

Thiols (R-SH) comprise a group of organosulphur compounds that can be found mainly in proteins (e.g. albumin) that contain sulphur-based amino acids (SAAs) as well as in low-molecular-weight (LMW) molecules like cysteine, homocysteine and glutathione. Thiols are known to be involved in various biological processes, such as enzymatic catalysis, cell signalling and metal complexing in the body.⁹ Most importantly, plasma or serum thiols are considered as a global marker of the systemic load of reactive species and as potent anti-oxidants because of their high reducing activity.¹⁰ According to recently proposed terminology, reactive species can be identified as ROS, as well as RNS and RSS, which are collectively referred to as the 'Reactive Species Interactome' (RSI).⁹ Depending on their redox state, thiols are classified as reduced or "free" thiols (R-SH) and oxidized or "bound" thiols, in which case a thiol is bound to another thiol via a disulfide bridge (R-SS-R'). In the circulation, the largest share of free thiols is embedded within the single cysteine residue (Cys³⁴) of albumin

Key Points

- Protein-adjusted serum free thiol levels are reduced and significantly associated with all-cause mortality in subjects with suspected Non-Alcoholic Fatty Liver Disease (NAFLD) (FLI \geq 60).
- Quantification of systemic free thiols may be a promising, minimally invasive strategy to improve detection of NAFLD and associated risk of all-cause mortality in the general population.

(HSA-SH) which exerts its antioxidant capacity. Remaining free thiols are classified as LMW free thiols, and the sum of protein free thiols and LMW free thiols is defined as *total free thiols*. Free thiols are able to scavenge reactive species and form disulphide bonds. Generally, total free thiol levels in serum could be interpreted as a direct and reliable reflection of the systemic redox system since they are readily oxidized by reactive species.^{11,12} Typically, high concentrations of serum free thiols are representative of a more beneficial or 'healthy' redox status. Changes in serum free thiol levels have been reported for many risk factors in which reactive species are known to play a prominent role, such as ageing, smoking, alcohol consumption, as well as for several diseases including inflammatory bowel disease (IBD), cardiovascular disease (CVD), obesity and ischaemia-reperfusion injury.¹³⁻¹⁵ Only one study reported that total serum thiol concentration is reduced while thiol-disulphide level is increased in NASH patients, compared to healthy controls.¹⁶

In this study, we determined systemic levels of serum free thiols in 5562 participants included in the Prevention of Renal and Vascular End-stage Disease (PREVEND) cohort, a large population-based cohort study from the Northern part of the Netherlands. Firstly, we compared protein-adjusted serum free thiol levels between subjects with $FLI < 60$ and $FLI \geq 60$ values and established associations between free thiol levels and multiple clinical, biochemical and NAFLD-specific parameters. Secondly, we investigated the association between baseline protein-adjusted serum free thiol concentrations and the risk of all-cause mortality during a follow-up of 10 years.

2 | MATERIALS AND METHODS

2.1 | Study population

This study used data from the Prevention of Renal and Vascular End-stage Disease (PREVEND) cohort study.¹⁷ This is a large, prospective population-based cohort study with participants from the Northern part of the Netherlands. The PREVEND study was set up to investigate cardiovascular and renal disease outcomes. From 1997 to 1998, 85 421 inhabitants aged 28-75 years from the Northern part of the Netherlands, received a questionnaire asking information about demographics, medication use, cardiovascular disease and pregnancy, including a request to supply an early morning urine sample. Participants who had a previous diagnosis of type 1 diabetes mellitus, insulin-treated type 2 diabetes mellitus and pregnant women were excluded from the study. In total, 40 856 subjects responded to the questionnaire and were analyzed for urinary albumin concentrations. Subjects with a urinary albumin concentrations ≥ 10 mg/L ($n = 6000$) were invited to visit the outpatient research clinic, as well as a random selection of participants with urinary albumin concentrations < 10 mg/L ($n = 2592$). The PREVEND study consisted of a total of 8592 participants who completed the full study program.¹⁸ However, for the current study we excluded

subjects ($n = 3030$) of which data on serum levels of free thiols and clinical and biochemical variables to calculate the Fatty Liver Index (FLI), as a proxy of NAFLD, were not available. This study was approved by the Institutional Review Board (IRB) of the University Medical Center Groningen (UMCG). The study was conducted in accordance with the principles of the Declaration of Helsinki (2013). All study participants provided written informed consent.

2.2 | Data collection

All study participants visited the outpatient research clinic of the UMCG, Groningen, the Netherlands. During the first visit, participants were requested to complete a questionnaire that contained information about demographics, health status, history of cardiovascular diseases (CVD), use of medications and lifestyle (e.g. self-reported smoking and alcohol consumption). Smoking was categorized as either current smoking or never or previous smoking. Alcohol consumption was documented with the assumption of one alcoholic drink to contain 10 grams of alcohol. History of cardiovascular disease included the following: hospitalization for myocardial ischaemia, obstructive coronary artery disease or revascularization procedures. Subsequently, anthropometric measurements were performed, including height (meters), weight (kilograms), body-mass index (BMI, weight divided by squared height), waist circumference (cm, defined as the smallest girth between rib cage and iliac crest), and waist/hip ratio (waist circumference divided by the largest girth between waist and thigh).^{19,20} During the second visit, systolic and diastolic blood pressure was measured automatically every minute until 8 minutes in supine position (Dinamap XL Model 9300 series device, Johnson & Johnson Medical). Blood pressure was defined as the average of the last two measurements in this procedure. Next, venous serum samples were withdrawn after an overnight fast while the participants had rested for 15 minutes. In addition, patients were asked to collect 24-hours urine specimens after they were provided with both oral and written instructions. In the current study, data were used of participants who completed the second screening evaluation in the PREVEND study.

2.3 | Laboratory measurements

Urinary albumin excretion (UAE) and high-sensitive C-reactive protein (hs-CRP) were measured by nephelometry (Dade Behring Diagnostics). UAE was measured twice in two different 24-hour urine specimens and the average of these was used in further analyses. Serum total cholesterol and serum glucose levels were measured by dry chemistry (Eastman Kodak). Low-density lipoprotein (LDL) cholesterol was determined by the Friedewald formula (if triglycerides ≤ 4.5 mmol/L). High-density lipoprotein (HDL) cholesterol was measured using a homogeneous method (direct HDL, Aeroset™ System, Abbott Laboratories).

Triglycerides were measured using an enzymatic method. Serum creatinine was measured with an enzymatic method as well (Roche Modular, Roche Diagnostics). Serum cystatin C was measured using the Gentian Cystatin C Immunoassay (Gentian AS) on a modular analyzer (Roche Diagnostics). Cystatin C was directly calibrated using a standard from the manufacturer (according to the International Federation of Clinical Chemistry Working Group for Standardization of Serum Cystatin C).²¹ Serum ALT and AST were measured using the standardized kinetic method with pyridoxal phosphate activation (Roche Modular P, Roche Diagnostics). Serum GGT was assayed by an enzymatic colorimetric method (Roche Modular P, Roche Diagnostics).

2.4 | Measurement of serum free thiols

Serum samples were stored at -80°C until analysis to avoid significant alterations in free thiol stability. Serum free thiol concentrations were measured as previously described, with minor modifications.^{22,23} After thawing, serum samples were diluted four-fold using 0.1 mol/L Tris buffer (pH 8.2). Using the Varioskan microplate reader (Thermo Scientific, Breda, the Netherlands), background absorption was measured at 412 nm, together with a reference measurement at 630 nm. Subsequently, 20 μL 1.9 mmol/L 5,5'-dithio-bis(2-nitrobenzoic acid) (DTNB, Ellman's Reagent, CAS-number 69-78-3, Sigma Aldrich Corporation) in 0.1 mol/L phosphate buffer (pH 7.0) was added to the samples and absorbance was measured again after the samples were incubated for 20 min at room temperature. Final concentrations of serum free thiols were established by parallel measurement of an L-cysteine (CAS-number 52-90-4, Fluka Biochemika) calibration curve (concentration range from 15.625 to 1000 $\mu\text{mol/L}$) in 0.1 mol/L Tris/10 mmol/L EDTA (pH 8.2). Intra- and interday coefficients of variation (CV) of all measurement values were below 10%. Ultimately, serum free thiol concentrations were adjusted to total serum protein levels (measured according to standard procedures) by calculating the free thiol/total protein ratio ($\mu\text{mol/g}$ of protein). This adjustment was performed since serum proteins harbour the largest amount of free thiols and therefore largely determine the amount of potentially detectable free thiols.²⁴

2.5 | Study outcomes and definitions

The estimated glomerular filtration rate (eGFR) was calculated using the combined creatinine cystatin C-based Chronic Kidney Disease Epidemiology Collaboration (CKD-EPI) equation.²⁵ Type 2 diabetes (T2D) was defined as a fasting glucose level ≥ 7.0 mmol/L, a random glucose level ≥ 11.1 mmol/L, self-report of a physician's diagnosis or the use of antidiabetic medications according to the guidelines of the American Diabetic Association (ADA). The algorithm of the Fatty Liver Index (FLI) was used as a proxy for the diagnosis of suspected NAFLD.⁵ The FLI was calculated according to the following formula: $\text{FLI} = [e^{(0.953 \times \log_e(\text{triglycerides}) + 0.139 \times \text{BMI} + 0.718 \times \log_e$

$(\text{GGT}) + 0.053 \times \text{waist circumference} - 15.745)]/[1 + e^{(0.953 \times \log_e(\text{triglycerides}) + 0.139 \times \text{BMI} + 0.718 \times \log_e(\text{GGT}) + 0.053 \times \text{waist circumference} - 15.745)}] \times 100$. The optimal cut-off value of the FLI for detecting NAFLD is established as 60 with a corresponding sensitivity of 61%, specificity of 86% and an accuracy of 84% as determined by ultrasonography.⁵ Therefore, $\text{FLI} \geq 60$ was used as a definition of suspected NAFLD, which is used nowadays as one of the best-validated steatosis scores for large scale screening studies.²⁶ Alternatively, we used the Hepatic Steatosis Index (HSI), which has been used previously in predominantly Asian populations and is defined as follows⁴: $\text{HSI} = 8 \times \text{ALT/AST ratio} + \text{BMI} (+2, \text{ if diabetes; } +2, \text{ if female})$. The optimal cut-off value of the HSI for detecting NAFLD is a score of 36. In the above equations, BMI was expressed as kg/m^2 , triglycerides as mmol/L and gamma-glutamyltransferase (GGT), alanine aminotransferase (ALT) and aspartate aminotransferase (AST) as U/L.

Metabolic syndrome (MetS) was defined according to the revised National Cholesterol Education Program Adult Treatment Panel (NCEP-ATP) III criteria. Study participants were assigned to have MetS when at least three of the following five criteria were fulfilled: (a) waist circumference >102 cm for men and >88 cm for women; (b) plasma triglycerides ≥ 1.7 mmol/L; (c) HDL cholesterol <1.0 mmol/L for men and <1.3 mmol/L for women; (d) hypertension (blood pressure $\geq 130/85$ mmHg or the use of antihypertensive drugs); (e) hyperglycemia (fasting glucose ≥ 5.6 mmol/L or the use of glucose lowering drugs).

Information on death (all-cause mortality) was obtained from the Dutch national registry of all hospital discharge diagnoses (Prismant). This information was classified in accordance with the International Statistical Classification of Diseases (ICD-10) and the International Classification of Health Interventions.²⁷

2.6 | Statistical analyses

Demographic, clinical and biochemical characteristics of the study population were presented as means \pm standard deviations (SD), proportions n with corresponding percentages (%) or medians [interquartile range (IQR)] in case of non-normal distributions. Assessment of normality was performed using histograms and normal probability plots (Q-Q plots). Between-group comparisons were performed using independent sample t -tests or Mann-Whitney U -tests in case of continuous variables, while chi-square tests were used in case of nominal variables. Protein-adjusted serum free thiol concentrations were ²log-transformed prior to analysis to facilitate interpretation of the results (expressed as per doubling). Univariable and multivariable logistic regression analyses were performed to evaluate associations between serum free thiol concentrations and NAFLD parameters expressed as odds ratios (OR) (per doubling) with corresponding 95% confidence intervals (CI). Stratified analyses were performed to examine the association between serum free thiols and NAFLD across various subgroups. Survival distributions for subjects with and without NAFLD were assessed according to tertiles of protein-adjusted

serum free thiol concentrations using Kaplan-Meier curves and compared to each other using log-rank tests. Survival time was defined from baseline (time of serum sample withdrawal) until the date of the last examination participants attended, either death or January 1, 2010 (end of follow-up period). Subsequently, Cox proportional hazards regression analyses were performed to assess associations between protein-adjusted serum free thiol concentrations and the risk of all-cause mortality, expressed as hazard ratios (HRs) (per doubling) with corresponding 95% CIs. Univariable associations were followed by multivariable models to adjust for potential confounding factors. Data analysis was performed using SPSS Statistics 25.0 software package (SPSS Inc) and data visualization using GraphPad Prism 5.0 (GraphPad software). Two-tailed P -values $\leq .05$ were considered statistically significant.

3 | RESULTS

3.1 | Baseline characteristics of the study population

Baseline characteristics of the study population are presented in Table 1. The study population consisted of 5562 participants, of whom 1651 (29.7%) subjects were classified with a FLI ≥ 60 . Participants classified with a FLI ≥ 60 were significantly older, as compared to subjects with a FLI < 60 (56.0 years vs 49.8 years, $P < .001$). In addition, subjects with a FLI ≥ 60 more frequently had a history of cardiovascular disease ($P < .001$), MetS ($P < .001$) and more often used antihypertensive medication ($P < .001$) and lipid-lowering drugs ($P < .001$). Moreover, anthropometric tests (ie BMI, waist circumference, waist/hip ratio), cholesterol levels and liver transaminase levels were higher in subjects with a FLI ≥ 60 ($P < .001$ for all). Conversely, LDL-cholesterol levels were not found to be significantly different between groups. With regard to serum levels of protein-adjusted free thiols, we observed significantly reduced concentrations in subjects with a FLI ≥ 60 , as compared to subjects with a FLI < 60 (4.91 $\mu\text{mol/L/g}$ vs 5.05 $\mu\text{mol/L/g}$, $P < .001$).

3.2 | Associations between protein-adjusted serum free thiol levels and FLI and HSI scores

Multivariable logistic regression analyses were subsequently performed in order to establish the extent to which serum levels of free thiols were associated with a FLI ≥ 60 (Table 2). In the age- and gender-adjusted analysis, we found a significant association between protein-adjusted free thiols ($^2\log$ -transformed, per doubling of concentration) and FLI (Model 2: OR 0.78 [95% CI 0.64-0.96], $P = .016$). This association remained statistically significant after additional adjustment for systolic blood pressure, diabetes, current smoking, use of alcohol and total cholesterol (Model 3: OR 0.80 [95% CI 0.65-0.99], $P = .04$). After additional adjustment for hs-CRP, this association lost significance (Model 4: OR 0.94 [95% CI 0.73-1.21], $P = .65$).

Similar results were observed in the analysis for HSI (Table S1). For instance, the association between HSI and serum levels of protein-adjusted free thiols ($^2\log$ -transformed, per doubling of concentration) only lost its significance after additional adjustment for hs-CRP (OR 0.87 [95% CI 0.68-1.10], $P = .24$). Stratified analyses for the association between protein-adjusted serum free thiols (per doubling) and FLI scores are presented in Table 3. Stratification by gender, the presence of hypertension and the presence of hypercholesterolemia showed significant differences between groups. Corresponding HRs were lower for female subjects ($P_{\text{interaction}} = 0.02$), subjects without hypertension ($P_{\text{interaction}} = 0.001$) and subjects without hypercholesterolemia ($P_{\text{interaction}} = 0.003$). Comparable results were obtained in stratified analyses when using the HSI instead of the FLI (Table S2).

3.3 | Protein-adjusted serum free thiols and risk of all-cause mortality

During follow-up, 291 (5.2%) subjects died (FLI < 60 , $n = 162$ (4.1%), FLI ≥ 60 , $n = 129$ (7.8%)). Kaplan-Meier survival analysis showed a significantly differential survival distribution between tertiles of protein-adjusted serum free thiols among subjects with a FLI < 60 and FLI ≥ 60 (Figure 1, $P < .001$, log-rank test). Cox proportional hazards regression analyses showed a significant inverse predictive association between $^2\log$ -transformed protein-adjusted serum free thiol concentrations and the risk of all-cause mortality for subjects with a FLI < 60 (Table 4A, model 1, HR per doubling of concentration 0.33 [0.22-0.50], $P < .001$) and subjects with a FLI ≥ 60 (Table 4B, model 1, HR per doubling of concentration 0.27 [0.17-0.45], $P < .001$). This association lost its significance after adjustment for potential confounders in subjects with a FLI < 60 (Table 4B, model 4, HR per doubling of concentration 0.78 [0.44-1.39], $P = .41$), while it remained statistically significant in subjects with a FLI ≥ 60 (Table 4B, model 4, HR per doubling of concentration 0.50 [0.27-0.95], $P = .03$). Similar results were obtained in Cox proportional hazard regression analyses using HSI instead of FLI, showing a statistically significant inverse association between $^2\log$ -transformed protein-adjusted serum free thiol concentrations and the risk of all-cause mortality for subjects with both an HSI < 36 and HSI ≥ 36 (Table S3). However, statistical significance vanished after adjustment for potential confounders in subjects of both subgroups, with the exception of the highest tertile of protein-adjusted serum free thiol concentrations in the group with HSI ≥ 36 (Table S3B, model 4, HR per doubling of concentration 0.39 [0.16-0.94], $P = .04$).

4 | DISCUSSION

In this study, we reported that protein-adjusted serum free thiol concentrations, as a marker of the systemic redox status, were lowered in subjects with suspected NAFLD (FLI ≥ 60). In addition, protein-adjusted serum free thiols were significantly associated with an increased risk of all-cause mortality in subjects with suspected NAFLD

TABLE 1 Clinical and laboratory characteristics including protein-adjusted serum free thiols in 3911 subjects with a fatty liver index (FLI) < 60 and 1651 subjects with a FLI ≥ 60

	FLI < 60 n = 3911	FLI ≥ 60 n = 1651	P-value
Age year, median (IQR)	49.84 (42.11-59.43)	55.99 (47.96-65.78)	<.001
Gender (men), n (%)	1599 (40.9)	1096 (66.4)	<.001
Ethnicity			
Caucasian, n (%)	3727 (95.3)	1575 (95.4)	.176
Asian, n (%)	85 (2.2)	26 (1.6)	
Black, n (%)	31 (0.8)	21 (1.3)	
Other, n (%)	41 (1.0)	17 (1.0)	
Unknown, n (%)	27 (0.7)	12 (0.7)	
Current smokers, n (%)	1093 (28.3)	440 (26.9)	.312
Use of alcohol, n (%)	2975 (76.7)	1190 (72.5)	<.001
BMI (kg/m ²), median (IQR)	24.62 (22.83-26.71)	30.15 (25.02-32.91)	<.001
Waist circumference (cm), median (IQR)	86 (79-93)	104 (99-110)	<.001
Waist/hip ratio, mean ± SD	0.86 ± 0.07	0.96 ± 0.07	<.001
Systolic blood pressure (mm Hg), median (IQR)	120 (110-133)	135 (123-147)	<.001
Diastolic blood pressure (mm Hg), median (IQR)	71 (65-77)	78 (71-84)	<.001
Antihypertensive medication, n (%)	471 (12.4)	485 (30.1)	<.001
Lipid-lowering drugs, n (%)	171 (4.5)	161 (10)	<.001
History of cardiovascular disease, n (%)	97 (2.5)	87 (5.3)	<.001
MetS, n (%)	309 (7.9)	958 (58.1)	<.001
Glucose (mmol/L), median (IQR)	4.70 (4.40-5.10)	5.10 (4.60-5.60)	<.001
Insulin (mU/L), median (IQR)	6.80 (5.10-9.20)	12.70 (9.40-18.40)	<.001
HOMA-IR (mU × mmol/L ² /22.5), median (IQR)	1.43 (1.03-2.00)	2.88 (2.04-4.35)	<.001
HOMA-β (%), median (IQR)	25.38 (18.31-35.13)	46.15 (33.02-66.64)	<.001
Urinary albumin excretion (mg/24 h), median (IQR)	7.78 (5.76-12.55)	11.03 (7.08-23.47)	<.001
eGFR (mL/min/1.73m ²), median (IQR)	96.30 (84.86-106.07)	89.20 (77.26-100.63)	<.001
hs-CRP (mg/L), median (IQR)	1.01 (0.48-2.27)	2.36 (1.17-4.22)	<.001
ALT (U/L), median (IQR)	15 (12-20)	23 (17-32)	<.001
AST (U/L), median (IQR)	21 (19-25)	25 (21-30)	<.001
ALP (U/L), median (IQR)	59 (49-71)	58 (57-79)	<.001
GGT (U/L), median (IQR)	19 (14-27)	41 (29-62)	<.001
Total cholesterol (mmol/L), mean ± SD	5.31 ± 0.99	5.78 ± 1.05	<.001
Non-HDL cholesterol (mmol/L), median (IQR)	3.91 (3.30-4.58)	4.64 (3.98-5.36)	<.001
LDL cholesterol (mmol/L), median (IQR)	3.32 (2.70-4.08)	3.48 (2.69-4.22)	.446
HDL cholesterol (mmol/L), median (IQR)	1.30 (1.11-1.51)	1.06 (0.9232-1.23)	<.001
Triglycerides (mmol/L), median (IQR)	0.94 (0.71-1.25)	1.72 (1.29-2.33)	<.001
Free thiols (protein-adjusted) (μmol/L/g), mean ± SD	5.05 ± 0.99	4.91 ± 1.02	<.001

Bold P-values indicate statistical significance.

Data are presented as mean ± standard deviation (SD) for normally distributed data or median with interquartile ranges (IQR) for non-normally distributed data.

Abbreviations: BMI, Body Mass Index; MetS, Metabolic syndrome; HOMA-IR, Homeostatic Model Assessment of Insulin Resistance; HOMA-β, Homeostatic Model Assessment of β cell function; hs-CRP, high sensitive C reactive protein; ALT, Alanine Aminotransferase; AST, Aspartate aminotransferase; GGT, gamma-glutamyltransferase; HDL, High-Density Lipoprotein; LDL, Low-Density Lipoprotein.

in this population-based cohort. Multivariable regression analyses showed maintenance of this significant association after adjustment for potential confounding factors, including the adjustment

for systolic blood pressure, diabetes, current smoking, use of alcohol and total cholesterol in subjects with FLI ≥ 60. As expected, this association lost its significance after additional adjustment for

TABLE 2 Multivariable logistic regression analysis to test the relationship between FLI and serum levels of protein-adjusted serum free thiols (²log-transformed)

	Model 1		Model 2		Model 3		Model 4	
	OR [95% CI]	P-value	OR [95% CI]	P-value	OR [95% CI]	P-value	OR [95% CI]	P-value
Free thiols (² log)	0.65 [0.54-0.78]	<.001	0.78 [0.64-0.96]	.016	0.80 [0.65-0.99]	.04	0.94 [0.73-1.21]	.65
Age			1.03 [1.03-1.04]	<.001	1.00 [0.99-1.01]	.26	1.00 [0.99-1.01]	.62
Gender (reference = male)			0.35 [0.31-0.40]	<.001	0.35 [0.31-0.40]	<.001	0.33 [0.28-0.39]	<.001
Diabetes (no = reference)					3.92 [2.63-5.82]	<.001	4.02 [2.35-6.90]	<.001
Current smoking (reference = no)					1.01 [0.88-0.1.17]	.87	0.98 [0.83-1.16]	.82
Use of alcohol (reference = no)					0.69 [0.59-0.80]	<.001	0.69 [0.58-0.82]	<.001
Systolic blood pressure					1.03 [1.03-1.04]	<.001	1.03 [1.03-1.04]	<.001
Total cholesterol					1.54 [1.44-1.64]	<.001	1.54 [1.44-1.66]	<.001
hs-CRP							1.08 [1.06-1.10]	<.001

Note: Model 1: crude.

Model 2: model 1 + additional correction for age and gender.

Model 3: model 2 + additional correction for systolic blood pressure, diabetes, current smoking, use of alcohol and total cholesterol.

Model 4: model 3 + additional correction for hs-CRP.

high-sensitive C-reactive protein (hs-CRP), indicating that inflammation and oxidative stress are both associated with NAFLD and not independent of each other.²⁸ Stratified analyses showed that there were significantly differential associations of protein-adjusted serum free thiol concentrations (per doubling) by gender, hypertension and hypercholesterolemia. Our results were further confirmed by comparable associations with the Hepatic Steatosis Index (HSI > 36), which is also a widely applied and recommended proxy to determine NAFLD in large population-based cohort studies.^{4,5} Taken together, the current study demonstrated that protein-adjusted serum free thiols could be a prominent minimally invasive marker of reactive species-driven development of NAFLD and are associated with the risk of all-cause mortality in subjects with suspected NAFLD.

NAFLD, thought to be caused by an imbalanced influx of free fatty acids (FFAs) and excessive accumulation of triglycerides in hepatocytes, is strongly associated with insulin resistance and metabolic syndrome (MetS). During the development of NAFLD, FFA governing transcription regulators are disrupted (e.g. the transcription factors peroxisome proliferator-activated receptor alpha [PPAR α], or sterol regulatory element-binding proteins [SREBPs]) causing inappropriate activation of pro-inflammatory signalling pathways (via protein-kinase B [AKT] or AMP-activated protein kinase [AMPK]) that contribute to the production of pro-inflammatory cytokines such as IL-6, TNF- α or IL-1 β and increased hepatocellular damage.^{8,29} Concurrently, a shift in redox balance occurs through the combined sequence of mitochondrial dysfunction, impaired oxidation of free fatty acids (FFAs) and toxicity of excessively accumulated triglycerides. In our study, subjects with FLI \geq 60 had a

significantly higher frequency of previous cardiovascular disease and MetS as well as significantly increased plasma concentrations of triglycerides, alanine aminotransferase (ALT) and aspartate aminotransferase (AST), as compared to subjects without suspected NAFLD (FLI < 60). Most importantly, protein-adjusted free thiol concentrations were significantly lower in subjects with FLI \geq 60. These results were consistent in subjects having an HSI > 36. Altered serum thiol balance in NAFLD has been reported in only one study before. Asil *et al* reported that serum *total* thiols were reduced in patients with NASH and simple steatosis as compared to healthy controls (n = 90).¹⁶ In comparison to our data, that study focused on total/native thiol ratios, included relatively low numbers of patients and applied liver biopsy to define NAFLD. Several other studies reported that there were no significant differences with regard to total serum thiol concentrations in subjects with insulin resistance (IR), type 2 diabetes (T2D).³⁰⁻³² Additionally, in paediatric subjects, increased serum thiols such as cysteine and homocysteine were observed in patients with NAFLD, while they were reduced in patients with NASH or liver fibrosis.³³ However, these studies focused on thiol/disulphide homeostasis using different measurement protocols i.e. distinct thiol-reactive reagents which compromises comparability of results between studies as measurements of either free thiols or total thiols lead to different classifications and terminology.³⁴ In addition, all these studies were based on datasets with relatively low numbers of study participants or they focused on different types of populations, e.g. solely on paediatric or female subjects.

Oxidative stress is referred to as an imbalance between oxidant and anti-oxidant substances. In NAFLD, the antioxidant system is

Variable	Total (n)	OR [*]	95% CI	P-value (interaction)
Overall	5562	0.80	0.65-0.99	.042
Gender				
Female	2815	0.63	0.46-0.87	.020
Male	2639	0.96	0.72-1.27	
BMI				
<25.0	2167	3.30	1.11-9.79	.902
>25.0	3275	0.81	0.63-1.05	
Albuminuria				
No	4813	0.83	0.66-1.05	.060
Yes	639	0.86	0.51-1.45	
Hypertension				
No	3765	0.78	0.59-1.03	.001
Yes	1690	0.88	0.63-1.22	
CVD history				
No	5273	0.83	0.66-1.03	.200
Yes	181	0.42	0.15-1.19	
Diabetes				
No	5322	0.82	0.66-1.02	.386
Yes	132	0.62	0.21-1.88	
Smoking				
No	3936	0.81	0.63-1.04	.611
Yes	1518	0.77	0.52-1.15	
Alcohol consumption				
No	1336	0.64	0.43-0.96	.209
Yes	4118	0.88	0.68-1.14	
Hypercholesterolemia				
No	3904	0.77	0.59-1.00	.003
Yes	1579	0.86	0.60-1.25	

Abbreviations: BMI, body-mass index; CI, confidence interval; CV, cardiovascular; CVD, cardiovascular disease; OR, odds ratio.

*Adjusted for potential confounding factors (gender, age, history of diabetes, current smoking, alcohol consumption, blood pressure and hypercholesterolemia).

Bold P-values indicate statistical significance.

disrupted because of excessive fat accumulation-mediated endoplasmic reticulum (ER) stress and mitochondrial β -oxidation dysfunction, leading to oxidative stress-induced complications caused by endogenous production of reactive species.^{6,7} It should be noted that serum free thiols have been considered a prominent antioxidant marker in serum because of their potent capacity to scavenge reactive species.^{9,14,31} High-sensitive C-reactive protein (hs-CRP) has been reported to be a prominent ROS-induced inflammatory marker in NAFLD.³⁵ A diminished antioxidant capacity is significantly associated with hs-CRP during disrupted redox homeostasis in multiple oxidative stress-related diseases.³⁶⁻³⁹ Similarly, in our study, serum hs-CRP levels were significantly increased in subjects with FLI ≥ 60 . In addition, in multivariable regression analyses, the persistent statistically significant associations of ²log-transformed

TABLE 3 Stratified analyses for the association between ²log-transformed protein-adjusted serum free thiols and the fatty liver index (FLI) across various subgroups. Stratifications by gender, hypertension and hypercholesterolemia showed significant interactions

protein-adjusted serum free thiols and systolic blood pressure, diabetes, current smoking, use of alcohol and total cholesterol with FLI ≥ 60 lost their significances after adjustment for hs-CRP. The same results were obtained in the analysis of the HSI > 36 group. This similar association of hs-CRP and thiols has been observed in several studies related to antioxidant homeostasis. For instance, one study found a negative correlation between hs-CRP levels and thiol/disulphide ratio and a positive correlation with total thiols during acute appendicitis in children (n = 80).⁴⁰ In addition, in patients with inflammatory bowel disease (IBD), hs-CRP was also significantly inversely associated with free thiols.^{14,41} These results further underscore that systemic free thiols are significantly associated with hs-CRP as oxidative stress-induced acute inflammation marker. Interestingly, in our stratified analyses, women with

FIGURE 1 Kaplan-Meier survival distributions for tertiles of protein-adjusted serum free thiol concentrations ($\mu\text{mol/L/g}$). Kaplan-Meier curve representing survival with the highest mortality rate occurring in the lowest tertile of protein-adjusted serum free thiols in both groups (log-rank test, $P < .001$)

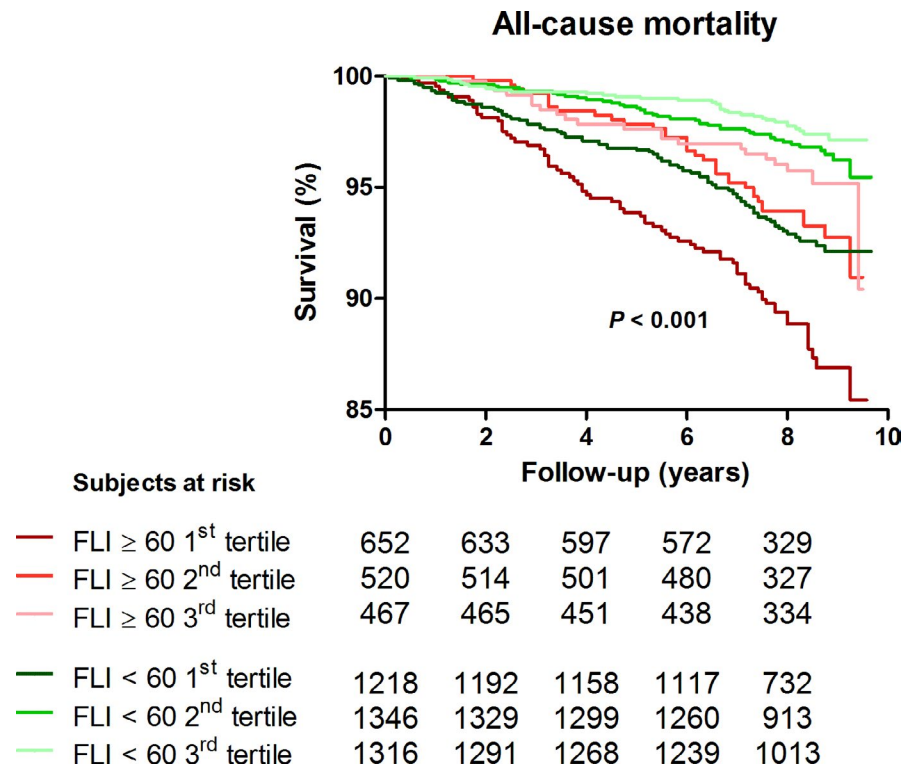


TABLE 4 Cox proportional hazards regression models of the association between $^2\log$ -transformed protein-adjusted serum free thiols and potential confounding factors with all-cause mortality, for patients with FLI < 60 (A) and FLI \geq 60 (B)

	HR per doubling	Tertiles of protein-adjusted serum free thiols		
		<4.65 $\mu\text{mol/g}$	4.65-5.46 $\mu\text{mol/g}$	>5.46 $\mu\text{mol/g}$
(A) FLI < 60				
Model 1	0.33 [0.22-0.50] $P < .001$	1.00 (Reference)	0.44 [0.31-0.64] $P < .001$	0.33 [0.22-0.49] $P < .001$
Model 2	0.75 [0.45-1.24] $P = .26$	1.00 (Reference)	0.72 [0.50-1.05] $P = .09$	0.72 [0.47-1.11] $P = .14$
Model 3	0.77 [0.46-1.27] $P = .30$	1.00 (Reference)	0.73 [0.50-1.07] $P = .11$	0.71 [0.46-1.09] $P = .12$
Model 4	0.78 [0.44-1.39] $P = .41$	1.00 (Reference)	0.85 [0.57-1.28] $P = .44$	0.68 [0.42-1.11] $P = .13$
(B) FLI \geq 60				
Model 1	0.27 [0.17-0.45] $P < .001$	1.00 (Reference)	0.54 [0.36-0.81] $P = .003$	0.37 [0.23-0.60] $P < .001$
Model 2	0.62 [0.36-1.06] $P = .08$	1.00 (Reference)	0.80 [0.53-1.21] $P = .29$	0.66 [0.40-1.09] $P = .11$
Model 3	0.65 [0.38-1.12] $P = .12$	1.00 (Reference)	0.84 [0.55-1.27] $P = .41$	0.69 [0.42-1.15] $P = .15$
Model 4	0.50 [0.27-0.95] $P = .03$	1.00 (Reference)	0.87 [0.55-1.36] $P = .53$	0.64 [0.36-1.14] $P = .13$

Note: Model 1: crude.

Model 2: model 1, age- and gender-adjusted.

Model 3: model 2, adjusted for systolic blood pressure, diabetes, current smoking, use of alcohol and total cholesterol.

Model 4: model 3, additionally adjusted for hs-CRP.

Bold P -values indicate statistical significance.

Abbreviation: HR, hazard ratio.

suspected NAFLD had a higher risk of impaired free thiol status. In agreement, Ates *et al* found that iron and the antioxidant enzyme ferroxidase activity was higher, while total plasma native thiol level was lower in women ($n = 95$) with obesity and insulin resistance (IR).³² Furthermore, our results support the fact that dysregulation of redox homeostasis is a crucial indicator in the presence of NAFLD.

In the future, free thiols could be further investigated for their potential to be implemented as a diagnostic or monitoring tool in NAFLD. Recent studies reported that systemic free thiol levels were significantly associated with heart failure, inflammatory bowel disease and levels of triglycerides and VLDL.^{13,14,30} Furthermore, dynamics of free thiols in serum could be a useful characteristic to determine the severity of disease. For instance, rapidly increased systemic free thiol levels were observed during the recovery phase of systemic sclerosis patients⁴² indicating that hypoxia elicits up-regulation of the antioxidant status. In the present study, serum free thiol levels were significantly lowered in subjects with $FLI \geq 60$ compared to $FLI < 60$. Since systemic free thiols (R-SH) are considered to be amenable to therapeutic manipulation, it could also become a beneficial treatment target in NAFLD. In this regard, hydrogen sulphide (H_2S) or precursors like N-acetylcysteine (NAC) and glutathione as low molecular weight thiol-containing compounds (and many other antioxidant supplementations) are considered to be potential treatment options to correct an imbalanced redox status in diseases like NAFLD.^{43,44} Endogenous production of H_2S is reduced in the cirrhotic liver, while exogenous H_2S supplementation prevents NASH in an animal experimental model via abating oxidative stress and suppressing inflammation.⁴⁵ In addition, antioxidant supplementation with riboflavin (vitamin B_2) significantly decreased inflammatory markers, while it increased systemic levels of free thiols in patients with Crohn's disease, demonstrating that antioxidant therapy holds promise in diseases which are characterized by overproduction of reactive species.⁴⁴

A recent meta-analysis study reported a significant positive association between NAFLD and all-cause mortality.⁴⁶ Thus, there is importance for an early and non-invasive screening method to enable prediction for all-cause mortality in NAFLD.⁴⁷ Of note, measuring free thiols in serum is relatively minimally invasive. In this study, using Cox proportional hazard regression analysis, we showed a significant predictive association between protein-adjusted serum free thiols and the risk of all-cause mortality for subjects with $FLI \geq 60$. This association lost its significance after adjustment for potential confounders in subjects with a $FLI < 60$ and remained significant in subjects with a $FLI \geq 60$ (Table 4). Since serum free thiols could be a potential therapeutic target in NAFLD, interventions targeted to increase the free thiol pool could also potentially predict the risk of all-cause mortality. Taken together, protein-adjusted serum free thiols could be a prominent predictor of all-cause mortality in NAFLD. However, it is important to further investigate the association between serum free thiol levels and different stages of NAFLD.

Our study has several strengths and limitations that need to be acknowledged. For example, to the best of our knowledge,

this is the first large study to report a significant association between serum free thiols - as a minimally invasive method to quantify systemic oxidative stress - and NAFLD. Most importantly, the protein-adjusted serum free thiol level was significantly associated with the risk of all-cause mortality in patients with identified NAFLD. We were able to establish this association in a population-based cohort study with a large sample size ($n = 5562$) that enabled us to properly adjust for potential confounding variables with sufficient study power. Furthermore, the association of serum free thiols with suspected NAFLD individuals in the general population were determined using two different, but accurate proxies of NAFLD: the FLI and HSI indices. However, FLI cannot identify absolute clinical NAFLD because of the lack of discrimination between severe steatosis levels and liver fat, but it is considered to be an acceptable method to indicate NAFLD in large-population based studies.⁴⁸ Although the HSI has only been validated in Asian populations, results were comparable in our cohort. Indeed, both methods are widely accepted and recommended to characterize NAFLD in large population-based cohort studies.^{4,5,26} However, several study limitations need to be addressed as well. For instance, the PREVENT cohort study mainly comprises individuals of European descent, which are predominantly derived from Caucasian populations, limiting the external applicability of our results to other ethnic populations. In addition, in the PREVENT cohort, it was not feasible to determine NAFLD by other diagnostic methods like liver ultrasound or liver biopsy. Lastly, the association between redox homeostasis and the severity of NAFLD might be important.⁴⁹ However, it was not possible to correlate free thiols with the different stages of NAFLD, e.g. NASH, fibrosis or cirrhosis because of the lack of necessary data to enable this characterization. Similarly, it was not possible to exclude other potential causes of liver disease as these data were not available in the present cohort.

In conclusion, protein-adjusted serum free thiol concentrations were significantly reduced in subjects with suspected NAFLD, even after adjustment for known risk factors for NAFLD. Furthermore, protein-adjusted serum free thiols were significantly associated with the risk of all-cause mortality in subjects with suspected NAFLD. Future studies are warranted that focus on the clinical utility of systemic free thiols in patients with NAFLD and the detailed discovery of potential associations with therapeutic outcome, disease course and overall prognosis. As free thiols are known to be receptive for therapeutic manipulation, future thiol-targeted therapy should be investigated as well to ameliorate disease outcome in NAFLD.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

ORCID

Turtushikh Damba  <https://orcid.org/0000-0003-3141-249X>

Arno R. Bourgonje  <https://orcid.org/0000-0001-5754-3821>

Svenja Sydor  <https://orcid.org/0000-0002-1349-8309>

Han Moshage  <https://orcid.org/0000-0002-4764-0246>

REFERENCES

1. Loomba R, Sanyal AJ. The global NAFLD epidemic. *Nat Rev Gastroenterol Hepatol*. 2013;10:686-690.
2. Ballestri S, Zona S, Targher G, et al. Nonalcoholic fatty liver disease is associated with an almost twofold increased risk of incident type 2 diabetes and metabolic syndrome. Evidence from a systematic review and meta-analysis. *J Gastroenterol Hepatol*. 2016;31:936-944.
3. van den Berg EH, Amini M, Schreuder TCMA, et al. Prevalence and determinants of non-alcoholic fatty liver disease in lifelines: a large Dutch population cohort. *PLoS One*. 2017;12:1-15.
4. Lee J-H, Kim D, Kim HJ, et al. Hepatic steatosis index: A simple screening tool reflecting nonalcoholic fatty liver disease. *Dig Liver Dis*. 2010;42:503-508.
5. Bedogni G, Bellentani S, Miglioli L, et al. The fatty liver index: a simple and accurate predictor of hepatic steatosis in the general population. *BMC Gastroenterol*. 2006;6:1-7.
6. Gambino R, Musso G, Cassader M. Redox balance in the pathogenesis of nonalcoholic fatty liver disease: mechanisms and therapeutic opportunities. *Antioxidants Redox Signal*. 2011;15:1325-1365.
7. Podrini C, Borghesan M, Greco A, Paziienza V, Mazzoccoli G, Vinciguerra M. Redox homeostasis and epigenetics in Non-alcoholic Fatty Liver Disease (NAFLD). *Curr Pharm Des*. 2013;19:2737-2746.
8. Asrih M, Jornayvaz FR. Inflammation as a potential link between nonalcoholic fatty liver disease and insulin resistance. *J Endocrinol*. 2013;218(3):R25-R36.
9. Cortese-Krott MM, Koning A, Kuhnle GGC, et al. The reactive species interactome: evolutionary emergence, biological significance, and opportunities for redox metabolomics and personalized medicine. *Antioxidants Redox Signal*. 2017;27:684-712.
10. Sen CK, Packer L. Thiol homeostasis and supplements in physical exercise. *Am J Clin Nutr*. 2000;72(2):653S-669S.
11. Banne AF, Amiri A, Pero RW. Reduced level of serum thiols in patients with a diagnosis of active disease. *J Anti Aging Med*. 2003;6:327-334.
12. Chung HS, Wang SB, Venkatraman V, Murray CI, Van Eyk JE. Cysteine oxidative posttranslational modifications: emerging regulation in the cardiovascular system. *Circ Res*. 2013;112:382-392.
13. Koning AM, Meijers WC, Pasch A, et al. Serum free thiols in chronic heart failure. *Pharmacol Res*. 2016;111:452-458.
14. Bourgonje AR, Gabriëls RY, de Borst MH, et al. Serum free thiols are superior to fecal calprotectin in reflecting endoscopic disease activity in inflammatory bowel disease. *Antioxidants (Basel)*. 2019;8:351.
15. Prakash M, Shetty MS, Tilak P, Anwar N. Total Thiols: biomedical importance and their alteration in various disorders. *Online J Heal Allied Sci*. 2009;8:1-9.
16. Asil M, Dertli R, Biyik M, et al. Dynamic thiol-disulfide homeostasis is disturbed in patients with non-alcoholic fatty liver disease. *J Lab Med*. 2018;42:31-38.
17. Hillege HL, Janssen WMT, Bak AAA, et al. Microalbuminuria is common, also in a nondiabetic, nonhypertensive population, and an independent indicator of cardiovascular risk factors and cardiovascular morbidity. *J Intern Med*. 2001;249:519-526.
18. Kieneker LM, Gansevoort RT, de Boer RA, et al. Urinary potassium excretion and risk of cardiovascular events. *Am J Clin Nutr*. 2016;103:1204-1212.
19. Borggreve SE, Hillege HL, Wolffenbuttel BHR, et al. The effect of cholesteryl ester transfer protein -629C→A promoter polymorphism on high-density lipoprotein cholesterol is dependent on serum triglycerides. *J Clin Endocrinol Metab*. 2005;90:4198-4204.
20. Kappelle PJWH, Gansevoort RT, Hillege JL, Wolffenbuttel BHR, Dullaart RPF. Apolipoprotein B/A-I and total cholesterol/high-density lipoprotein cholesterol ratios both predict cardiovascular events in the general population independently of nonlipid risk factors, albuminuria and C-reactive protein. *J Intern Med*. 2011;269:232-242.
21. Grubb A, Blirup-Jensen S, Lindström V, Schmidt C, Althaus H, Zegers I. First certified reference material for Cystatin C in human serum ERM-DA471/IFCC. *Clin Chem Lab Med*. 2010;48:1619-1621.
22. Ellman GL. Tissue Sulfhydryl Groups. *Arch Biochem Biophys*. 1959;82:70-77.
23. Hu ML, Louie S, Cross CE, Motchnik PHB. Antioxidant protection against hypochlorous acid in human plasma. *J Lab Clin Med*. 1993;121:257-262.
24. Turell L, Radi R, Alvarez B. The thiol pool in human plasma: the central contribution of albumin to redox processes. *Free Radic Biol Med*. 2013;65:244-253.
25. Inker LA, Schmid CH, Tighiouart H, et al. Estimating glomerular filtration rate from serum creatinine and cystatin C. *N Engl J Med*. 2012;367:20-29.
26. European Association for the Study of the Liver (EASL). EASL-EASD-EASO Clinical Practice Guidelines for the management of non-alcoholic fatty liver disease. *Diabetologia*. 2016;59:1121-1140.
27. WHO. International Statistical Classification of Diseases and Related Health Problems - 10th revision. World Heal Organ. 2011; 2.
28. Lee J, Yoon K, Ryu S, Chang Y, Kim HR. High-normal levels of hs-CRP predict the development of non-alcoholic fatty liver in healthy men. *PLoS One*. 2017;12:1-12.
29. Masarone M, Rosato V, Dallio M, et al. Role of oxidative stress in pathophysiology of nonalcoholic fatty liver disease. *Oxid Med Cell Longev*. 2018;2018:1-14.
30. van Dijk P, Abdulle AE, Bulthuis MLC, et al. The systemic redox status is maintained in non-smoking type 2 diabetic subjects without cardiovascular disease : association with elevated triglycerides and large VLDL. *J Clin Med*. 2020;9:49.
31. Schillern EEM, Pasch A, Feelisch M, et al. Serum free thiols in type 2 diabetes mellitus: a prospective study. *J Clin Transl Endocrinol*. 2019;16:100182.
32. Ates E, Set T, Karahan SC, Biçer C, Erel Ö. Thiol/Disulphide homeostasis, ischemia modified albumin, and ferroxidase as oxidative stress markers in women with obesity with insulin resistance. *J Med Biochem*. 2019;38:445-451.
33. Pastore A, Alisi A, di Giovamberardino G, et al. Plasma levels of homocysteine and cysteine increased in pediatric NAFLD and strongly correlated with severity of liver damage. *Int J Mol Sci*. 2014;15:21202-21214.
34. Sutton TR, Minnion M, Barbarino F, et al. A robust and versatile mass spectrometry platform for comprehensive assessment of the thiol redox metabolome. *Redox Biol*. 2018;16:359-380.
35. Foroughi M, Maghsoudi Z, Khayatzadeh S, Ghiasvand R, Askari G, Iraj B. Relationship between non-alcoholic fatty liver disease and inflammation in patients with non-alcoholic fatty liver. *Adv Biomed Res*. 2016;5:28.
36. Bozic MA, Subbarao G, Molleston JP. Pediatric nonalcoholic fatty liver disease. *Nutr Clin Pract*. 2013;28:448-458.
37. Mortensen C, Andersen O, Krag A, Bendtsen F, Møller S. High-sensitivity C-reactive protein levels predict survival and are related to haemodynamics in alcoholic cirrhosis. *Eur J Gastroenterol Hepatol*. 2012;24:619-626.
38. Noren Hooten N, Ejiogu N, Zonderman AB, Evans MK. Association of oxidative DNA damage and C-reactive protein in women at risk for cardiovascular disease. *Arterioscler Thromb Vasc Biol*. 2012;32:2776-2784.
39. Kim JH, Baik HW, Yoon YS, et al. Measurement of antioxidant capacity using the biological antioxidant potential test and its role as a predictive marker of metabolic syndrome. *Korean J Intern Med*. 2014;29:31-39.
40. Elmas B, Yildiz T, Yazar H, et al. New oxidative stress markers useful in the diagnosis of acute appendicitis in children. *Pediatr Emerg Care*. 2017;00:1-6.
41. Bourgonje AR, von Martels JZH, Bulthuis MLC, et al. Crohn's disease in clinical remission is marked by systemic oxidative stress. *Front Physiol*. 2019;10:1-10.

42. Abdulle AE, van Roon AM, Smit AJ, et al. Rapid free thiol rebound is a physiological response following cold-induced vasoconstriction in healthy humans, primary Raynaud and systemic sclerosis. *Physiol Rep*. 2019;7:1-12.
43. Wang P, Wu L. Hydrogen sulfide and nonalcoholic fatty liver disease. *Hepatobiliary Surg Nutr*. 2018;7:122-124.
44. von Martels JZH, Bourgonje AR, Klaassen MAY, et al. Riboflavin supplementation in patients with Crohn ' s disease [the RISE-UP study]. *J Crohns Colitis*. 2020;1:1-13.
45. Wu D, Zheng N, Qi K, et al. Exogenous hydrogen sulfide mitigates the fatty liver in obese mice through improving lipid metabolism and antioxidant potential. *Med Gas Res*. 2015;5:1-8.
46. Liu Y, Zhong GC, Tan HY, Hao FB, Hu JJ. Nonalcoholic fatty liver disease and mortality from all causes, cardiovascular disease, and cancer: a meta-analysis. *Sci Rep*. 2019;9:11124.
47. Stefan N. Nonalcoholic fatty liver disease and mortality. *Clin Gastroenterol Hepatol*. 2018;16:1043-1045.
48. Keating SE, Parker HM, Hickman IJ, et al. NAFLD in clinical practice: can simple blood and anthropometric markers be used to detect change in liver fat measured by 1H-MRS? *Liver Int*. 2017;37:1907-1915.
49. Ore A, Akinloye OA. Oxidative stress and antioxidant biomarkers in clinical and experimental models of non-alcoholic fatty liver disease. *Medicina (Kaunas)*. 2019;55:26.

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

How to cite this article: Damba T, Bourgonje AR, Abdulle AE, et al. Oxidative stress is associated with suspected non-alcoholic fatty liver disease and all-cause mortality in the general population. *Liver Int*. 2020;00:1-12. <https://doi.org/10.1111/liv.14562>