



University of Groningen

ALK immunohistochemistry positive, FISH negative NSCLC is infrequent, but associated with impaired survival following treatment with crizotinib

Thunnissen, E; Lissenberg-Witte, B I; van den Heuvel, M M; Monkhorst, K; Skov, B G; Sørensen, J B; Mellemgaard, A; Dingemans, A M C; Speel, E J M; de Langen, A J

Published in: Lung Cancer

DOI:

10.1016/j.lungcan.2019.09.023

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version
Publisher's PDF, also known as Version of record

Publication date: 2019

Link to publication in University of Groningen/UMCG research database

Citation for published version (APA):

Thunnissen, E., Lissenberg-Witte, B. I., van den Heuvel, M. M., Monkhorst, K., Skov, B. G., Sørensen, J. B., Mellemgaard, A., Dingemans, A. M. C., Speel, E. J. M., de Langen, A. J., Hashemi, S. M. S., Bahce, I., van der Drift, M. A., Looijen-Salamon, M. G., Gosney, J., Postmus, P. E., Samii, S. M. S., Duplaquet, F., Weynand, B., ... Smit, E. F. (2019). ALK immunohistochemistry positive, FISH negative NSCLC is infrequent, but associated with impaired survival following treatment with crizotinib. *Lung Cancer*, *138*, 13-18. https://doi.org/10.1016/j.lungcan.2019.09.023

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: https://www.rug.nl/library/open-access/self-archiving-pure/taverne-amendment.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): http://www.rug.nl/research/portal. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

ELSEVIER

Contents lists available at ScienceDirect

Lung Cancer

journal homepage: www.elsevier.com/locate/lungcan



ALK immunohistochemistry positive, FISH negative NSCLC is infrequent, but associated with impaired survival following treatment with crizotinib*



E. Thunnissen^{a,*}, B.I. Lissenberg-Witte^b, M.M. van den Heuvel^c, K. Monkhorst^d, B.G. Skov^e, J.B. Sørensen^f, A. Mellemgaard^g, A.M.C. Dingemans^h, E.J.M. Speelⁱ, A.J. de Langen^j, S.M.S. Hashemi^j, I. Bahce^j, M.A. van der Drift^k, M.G. Looijen-Salamon^l, J. Gosney^m, P.E. Postmusⁿ, S.M.S. Samii^o, F Duplaquet^p, B. Weynand^q, X. Durando^r, F. Penault-Llorca^s, S. Finn^t, A.O Grady^u, B. Oz^v, N. Akyurek^w, R. Buettner^x, J. Wolf^y, L. Bubendorf^z, S. Duin^a, I. Marondel^A, L.C. Heukamp^B, W. Timens^C, E.M.D. Schuuring^C, P. Pauwels^D, E.F. Smit^{c,j}

- ^a Department of Pathology, Amsterdam UMC, Location VU University Medical Center, the Netherlands
- ^b Epidemiology and Biostatistics, Amsterdam UMC, Location VU University Medical Center, the Netherlands
- ^c Department of Thoracic Oncology Netherlands Cancer Institute Antoni van Leeuwenhoek, Amsterdam, the Netherlands
- ^d Pathology, Netherlands Cancer Institute Antoni van Leeuwenhoek, Amsterdam, the Netherlands
- e Department of Pathology, Copenhagen University Hospital, Rigshospitalet, Copenhagen, Denmark
- f Oncology, Copenhagen University Hospital, Rigshospitalet, Copenhagen, Denmark
- g Department of Oncology, Herlev University Hospital, Copenhagen, Denmark
- h Department of Pulmonary Diseases, Maastricht University Medical Center, Maastricht, the Netherlands
- ⁱ Pathology, Maastricht University Medical Center, Maastricht, the Netherlands
- Department of Pulmonary Diseases, Amsterdam UMC, VU University Medical Center, Amsterdam, the Netherlands
- k Department of Pulmonary Diseases, Radboud University Medical Center, Nijmegen, the Netherlands
- ¹ Pathology, Radboud University Medical Center, Nijmegen, the Netherlands
- ^m Department of Pathology, Royal Liverpool University Hospital, Liverpool, United Kingdom
- ⁿ Department of medical oncology, Clatterbridge Cancer Centre, Bebington, Cheshire, United Kingdom
- ^o Department of Pulmonary Diseases, Deventer Hospital, Deventer, the Netherlands
- ^p Department of Pneumology, CHU Mont-Godinne, Catholic University of Louvain (UCL), Yvoir, Belgium
- ^q Pathology, CHU Mont-Godinne, Catholic University of Louvain (UCL), Yvoir, Belgium
- ^r Department of Medical Oncology, Centre Jean Perrin, Clermont-Ferrand, France
- ^s Pathology, Centre Jean Perrin, Clermont-Ferrand, France
- ^t Department of Pathology, Trinity College Dublin, Dublin, Ireland
- ^u Royal College of Surgeons in Ireland, Department of Pathology, Dublin, Ireland
- ^v Istanbul University Cerrahpasa, Cerrahpasa Medical Faculty, Istanbul, Turkey
- w Department of Pathology, Gazi University Medical Faculty, Ankara, Turkey
- ^x Institute for Pathology, University Hospital Cologne, Köln, Germany
- ^y Department of Internal Medicine, University of Cologne, Center for Integrated Oncology Köln Bonn, Germany
- ² Institute of Pathology, University Hospital Basel, Basel, Switzerland
- ^A Pfizer Innovative Health, Oncology International Developed Markets, Berlin, Germany
- ^B Institute of Haematopathology Hamburg, Hamburg, and Lung Cancer Network NOWEL.org, Oldenburg, Germany
- ^C Pathology and Medical Biology, University of Groningen, University Medical Center Groningen, Groningen, the Netherlands
- ^D Department of Pathology, Antwerp University Hospital, Edegem, Belgium

ARTICLE INFO

ABSTRACT

Keywords: non-small cell lung cancer alk immunohistochemistry Objective: Metastasized non-small cell lung cancer (NSCLC) with an anaplastic lymphoma kinase (ALK) rearrangement is usually sensitive to a range of ALK-tyrosine kinase inhibitors. ALK-positive NSCLC have been identified in pivotal phase III trials with fluorescence in situ hybridization (ALK FISH+). These tumors are also

E-mail address: e.thunnissen@amsterdamumc.nl (E. Thunnissen).

^{*} Key message: This study is the first comparative analysis of metastasized discordant ALK IHC positive, FISH negative with concordant ALK and FISH positive non-small cell lung cancer. The prognosis of the discordant cases is worse than of concordant cases. A suitable predictive testing strategy may be to screen first with IHC and then confirm with FISH instead of considering ALK IHC equivalent to ALK FISH according to the current ESMO and CAP/AMP/ IASLC guidelines.

^{*} Corresponding author at: Department of Pathology, Amsterdam University Medical Centers, Location VUmc, Amsterdam De Boelelaan 1117 1081HV Amsterdam the Netherlands.

fluorescence in situ hybridisation treatment prognosis expressing the fusion product (ALK immunohistochemistry (IHC) +). However, discrepant cases occur, including ALK IHC + FISH-. The aim of this study was to collect ALK IHC + cases and compare within this group response to crizotinib treatment of ALK FISH + cases with ALK FISH- cases.

Materials and methods: In this European prospective multicenter research study patients with Stage IV ALK IHC + NSCLC treated with crizotinib were enrolled. Tumor slides were validated centrally for ALK IHC and ALK FISH

Results: Registration of 3523 ALK IHC tests revealed a prevalence of 2.7% (n = 94) ALK IHC + cases. Local ALK FISH analysis resulted in 48 concordant (ALK IHC+/FISH+) and 16 discordant (ALK IHC+/FISH-) cases. Central validation revealed 37 concordant and 7 discordant cases, 5 of which had follow-up. Validation was hampered by limited amount of tissue in biopsy samples. The PFS at 1 year for ALK concordant and discordant was 58% and 20%, respectively (HR = 2.4; 95% CI: 0.78-7.3; p=0.11). Overall survival was significantly better for concordant cases than discordant cases after central validation (HR = 4.5; 95% CI = 1.2-15.9; p=0.010. Conclusion: ALK IHC + FISH- NSCLC is infrequent and associated with a worse outcome on personalized treatment. A suitable predictive testing strategy may be to screen first with IHC and then confirm with FISH instead of considering ALK IHC equivalent to ALK FISH according to the current guidelines.

1. Introduction

In 2007, the first anaplastic lymphoma kinase (ALK) fusion was described in non-small cell lung cancer (NSCLC) [1]. In 2013, a phase 3 study demonstrated a significant improvement in progression free survival (PFS) and overall survival (OS) in patients with metastasized ALK positive lung cancer treated with crizotinib compared to chemotherapy [2]. Subsequently, testing for ALK aberrations in patients with metastasized adenocarcinoma of the lung was recommended by international guidelines [3,4].

When testing for ALK rearrangements, both ALK fluorescence in situ hybridization (FISH) and ALK immunohistochemistry (IHC) may be used. Although in many studies a high association has been shown between immunohistochemistry positive (IHC+) and ALK FISH positivity (FISH+) [5], occasional discrepant cases may occur [6–9]. Cases with positive ALK FISH and negative ALK IHC do not seem to respond on treatment with ALK tyrosine kinase inhibitor [10,11]. As testing with IHC is preferred over testing by FISH for ALK fusions, it is likely that discordant cases with ALK IHC positivity and negative ALK FISH (ALK IHC + FISH-) will occur in practice.

Case of patients reports with discordant ALK IHC + FISH- tests show response to crizotinib [7,12–16]. However, a comparative study with treatment outcome is lacking.

The aim of this study was to prospectively collect a cohort of ALK IHC + NSCLC cases and after validation compare within this group response to crizotinib treatment of ALK FISH + cases with ALK FISH-cases.

2. Materials and methods

A prospective multicenter investigator initiated study on ALK IHC + metastasized (M+) NSCLC was started across Europe on April 1, 2014. Monthly, the number of ALK IHC tests on M + NSCLC and number ALK IHC + was recorded per center until June 2016, providing prevalence. Entry of individual ALK IHC + cases in central database with clinical information was possible until November 2017. The ALK antibodies 5A4 or D5F3 were allowed for local testing in NSCLC. The study required local a) ALK IHC + metastatic NSCLC, b) ALK FISH was optional for local testing; c) central validation for ALK IHC and FISH testing, d) treatment with crizotinib and minimal follow-up at 12 weeks. As the outcome of ALK FISH could be positive or negative, patients were stratified into ALK IHC positive and FISH positive (IHC + FISH+) and ALK IHC positive and FISH negative (IHC + FISH-). This study was approved by the VU University Medical Center (VUmc) institutional review board. Patient informed consent was locally arranged. Entry into the study was possible by the treating physician (oncologist/ pulmonologist, who was not always aware of availability of tissue sections for validation) or via the pathologist (who was not always aware of the treatment details). Therefore, two data sets were initially compiled and subsequently merged. During final analysis March 2019 from most, but not all patients all required information was available.

2.1. Clinical data

Collection of clinical data and validation data was performed in parallel. The clinical database contained 66 NSCLC cases with local data on testing, of which 5 with unknown IHC status and one without follow-up information. The following parameters were recorded: age, gender, smoking history, WHO performance status, clinical-stage at start of crizotinib treatment, resonse assessment according to Response Evaluation Criteria in Solid Tumors (RECIST) at 12 weeks after start on crizotinib, site primary lung cancer, date of first NSCLC diagnosis, comorbidities, other malignancy, sample type, sample site, histological diagnosis, local ALK IHC test used, local outcome IHC test, ALK FISH test used, local outcome FISH test, testing for EGFR, KRAS, HER2, PI3KCA, RET, BRAF, ROS1, progression free survival (PFS) and overall survival (OS).

2.2. Validation data

For validation of ALK testing blank histological sections were submitted to VUmc Amsterdam for validation with two ALK IHC assays and an ALK FISH assay. The ALK D5F3 antibody was performed according assay of supplier (Roche Ventana, land) in Groningen, NL (ES). The ALK 5A4 was done according a previously described protocol [17], performed in Amsterdam, NL (ET). The ALK FISH assay was performed in Antwerp (PP) with the Vysis ALK test (Abbott Molecular Inc. Des Plaines, IL, USA). In time 5 batches of sections were distributed to Groningen and Antwerp. Testing evaluation was performed blinded for clinical data. In case of limited number of slides, the order of ALK validation was i) 5A4, ii) D5F3 and iii) FISH. Upon receipt in Amsterdam, slides were sent within 3 months in batches to Groningen, Netherlands, and Antwerp, Belgium.

2.3. Statistics

The prevalence of ALK IHC + was calculated based on the number of monthly recorded ALK IHC tests per laboratory, and ALK IHC + outcome. Clinicopathologic parameters were summarized for local test outcome and after central validation. Overall survival (OS) was defined as start of treatment with crizotinib until death, and patients alive at their last follow-up time were censored. PFS was defined as start of treatment with crizotinib until progressive disease or death. OS and PFS were compared with Kaplan-Meier curves and the log-rank test. Statistical analyses (BW [18]) were carried out by SPSS for Windows and Mac version 22 (IBM Corp., Armonk, NY, USA). The significance level was set at 0.05

3. Results

In total 3523 ALK IHC tests were recorded in a period of 25 months, of which 94 were ALK IHC+, resulting in a prevalence of 2.7%.

In total 72 ALK IHC + M + NSCLC cases were signed up in the central database, see supplemental figure for consort diagram in Fig. 1.

3.1. Validation

After initial registration, blank slides were centrally received for validation of 72 cases in which the original (i.e. local test) diagnosis was ALK IHC + M + NSCLC. The outcome of the local ALK FISH analysis resulted in 48 condordant (ALK IHC + FISH+) and 16 discordant (ALK IHC + FISH-) cases. In 8 cases the ALK FISH was unknown/ uninformative.

The results of central validation for all 3 assays is shown in supplemental table S1. Note that due to limited availability of tumor in the remaining of the formalin fixed and paraffin embedded samples, not all cases could be adequately examined for validation purposes. In 54 of the 62 cases (87%) ALK IHC + was confirmed with 5A4 IHC and in 41 of the 55 cases (75%) with the D5F3 IHC.

The comparison of 5A4 and D5F3 ALK IHC is shown in supplemental table S2. Of the 55 cases with a test outcome, slightly more cases were positive for 5A4 than D5F3.

The distribution of cases with outcome of IHC and FISH validation is shown in supplemental table S3. For this analysis, a case was considered ALK IHC+, if at least one of the IHC validation assays was positive. In total 37 of the 48 cases were concordant ALK IHC+ FISH+ and 7 discordant ALK IHC+ FISH-. In 4 out of 48 cases (10%) the initial ALK IHC+ status could not be confirmed.

3.2. Clinical data and treatment

The clinicopathological data for locally and central validated ALK testing performed ALK tests are shown in Table 1. All patients were stage IV. There are no major differences between the clinicopathological variables (gender, age, performance status, treatment).

Information about crizotinib treatment and ALK test results in the local institution was available for 58 IHC + cases. ALK FISH was positive in 44 cases (76%), negative in 8 cases (14%), 'uninformative' in 2 cases (3%) and 'missing' in 4 cases (7%). Of the 52 cases with ALK FISH test result, RECIST determined response at 12 weeks was missing in 1 case. Forty-five out of 52 patients were still on treatment after 12 weeks

After central testing the median follow-up time for concordant cases was 54 weeks [6–188], and for discordant cases 40 weeks [4–125].

The overall survival between patients with ALK IHC + FISH + and ALK IHC + FISH- tumors did not differ significantly according to local testing: 1 year OS were 89% and 71% for ALK concordant and discordant cases, respectively (HR = 1.7; 95% CI = 0.45–6.3; p=0.42). OS, however, was significantly better for concordant cases than discordant cases, 85% versus 40% at 1 year, after central validation (HR=4.3; 95% CI=1.2–15.4; p=0.012, Fig. 1A).

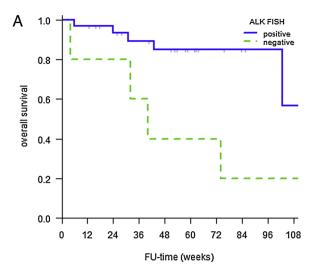
The PFS at 1 year by local ALK testing for ALK concordant and discordant was 68% and 50%, respectively (HR = 0.75; 95% CI: 0.30-2.6; p=0.83). For centrally ALK validated cases, the PFS at 1 year for ALK concordant and discordant was 58% and 20%, respectively (HR = 2.4; 95% CI: 0.78-7.3; p=0.11, Fig. 1B).

4. Discussion

This study showed a better overall survival for ALK IHC and FISH

Table 1 Clinicopathological data are shown for ALK IHC + metastasized NSCLC tested in local center for IHC and FISH.

		Local $(n = 52)$					Central validation $(n = 38)$				
		Concordant N		Discordant N		p-value	Concordant		Discordant		p-value
							N		N		
Gender	Male	22		3		0.52	15		3		0.65
	Female	22		5			18		2		
Age	mean (SD)	55.0	13.7	59.0	13.8	0.45	55.1	14.2	55.8	7.3	0.91
Smoking	Never smoker	29		0		0.002	19		1		0.29
	Former smoker	12		4			11		3		
	Current smoker	3		3			3		1		
	Unknown	0		1			0		0		
WHO performance status	0	17		3		0.92	15		2		0.27
	1	23		4			14		1		
	2	3		1			3		2		
	3	1		0			1		0		
Prior systemic therapy	No	23		2		0.25	15		1		0.37
	Yes	21		6			18		4		
Sample	Small biopsy	25		4		0.95	21		2		0.40
	Incisional biopsy	3		1			3		1		
	Excision biopsy	6		1			4		0		
	Resection	10		2			5		2		
Site	Lung	20		5			16		3		
	Mediastinal lymph node	5		2			3		0		
	Cervical lymph node	6		0			5		0		
	Liver	1		0			0		1		
	Pleura	4		1			5		1		
	Bone	4		0			2		0		
	Adrenal gland	1		0			_		Ü		
	Brain	1		0							
	Other	2		0			2		0		
Histology	Adenocarcinoma	39		7			28		4		
	NSCLC-NOS	3		0			3		1		
	Large cell carcinoma	1		0			1		0		
	Other	1		1			1		0		
	Other	1		1			1		U		



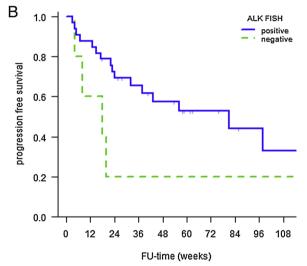


Fig. 1. Overall survival (A) of ALK IHC + NSCLC is shown after central validation for FISH+ (blue) and FISH- (green) cases (HR = 4.3; 95% CI = 1.2–15.4; p = 0.012) and (B) progression free survival (HR = 2.4; 95% CI: 0.78–7.3; p = 0.11) (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article).

concordant cases as compared to discordant cases after central validation, but not according to local testing.

The 1 year PFS for ALK IHC and FISH concordant cases treated with crizotinib (68% median) is similar as reported in the literature [19–22]. Although in our study the number of discordant ALK IHC + FISH- cases is low, their 1-year overall survival was significantly lower than in concordant cases. In a post-hoc analysis of the ALEX phase 3 trial, where patients with ALK-IHC positive NSCLC, assessed with D5F3 assay, showed better efficacy for alectinib than for crizotinib, [20] a subset of cases with discordant ALK IHC + FISH- also revealed a lower response rate than in the concordant cases [23]. This was in accordance with our findings. The difference between these two studies (Alex post-hoc analysis and our study) on the one hand and the case reports on ALK IHC + FISH- NSCLC showing a treatment response on the other hand can be explained by publication bias for the latter.

The prevalence of ALK IHC + NSCLC of 2.6% in this study by multiple institutions in Europe is in line with that reported in the literature. In a meta-analysis of 27 studies comparing clinicopathological characteristics of patients with NSCLC having a EML4-ALK fusion gene the frequency of ALK positive lung cancer was 6.8% (range 2.4%–32.6%) [24]. In consecutively tested pulmonary adenocarcinomas series ranging from 1.9%–5% [21,25,26] and in a series of

consecutive resection specimen ranging from 4.4 to 8.6% [17,27–29].

Literature comparison of ALK IHC and FISH testing reveals an impressive high concordance [30,31]. However, the discordant ALK IHC + FISH- are in this context at population level (metastasized adenocarcinomas of the lung) hidden in the specificity, ranging for 5A4 from 96 to 100% and for D5F3 from 95 to 100% with one outlier of 82% [31]. A recent review [5], comprised 18 studies with 5.5% ALK IHC positivity out of 10404 NSCLC cases, of which 0.7% discordant IHC + FISH- of the tested NSCLC. Remarkably, when expressed on test outcome level (as a fraction of ALK IHC + positive cases), the number of discordant ALK IHC + FISH- is 13%. In our study, at population level, the frequency of discordant ALK IHC + FISH- cases in stage IV NSCLC is lower (0.1%).

To understand the nature of the ALK IHC + FISH- discordancy, analysis with an orthogonal method is useful. In most cases not enough tumor material was available for further analysis. Explanations for ALK IHC + FISH- include (1) false-negative interpretation of FISH results, especially for results that are close to the threshold of 15% sections [32]; (2) counting in FISH normal cells as tumor cells; (3) double rearrangement involving ALK, reducing the visible distance of the two FISH probes [33]; (4) amplification of the ALK gene (which has been associated with ALK protein expression in some but not all cases), possibly leading to 1+ or 2+ staining [34,35]; (5) false-positive IHC staining with less specific antibodies (e.g. 1A4 [36]) (6) false positive interpretation of ALK IHC results due to high signal enhancement [37]; (7) Infrequently, ALK IHC may be positive in high grade neuroendocrine carcinomas of e.g. lung [38–40] and Merkel cell carcinomas [IASLC atlas [31] chapter 4] and (8) an indeterminate mechanism.

The central validation of the assays revealed surprising discordances with local testing in a small number of cases with respect to false positive IHC and false negative FISH. In daily practice these discordances may be addressed by participation in external quality assessment schemes [37]. However, these schemes do not always have a sufficient amount of material from the informative cases for distribution to a large number of laboratories.

The fact that the remaining tumor material was often not sufficient for the validation process of ALK IHC and FISH testing is a major limitation of this paper. For a portion of the cases, sufficient blank histological slides were only available for validation of one or two of the three assays. This is explained by the small biopsies, where most of the sample was used for primary diagnostic and predictive testing and very little or no tumor was left in the remaining of the block. This also prevented inclusion of several local ALK IHC + cases into the study. A selection bias by tissue sample size is not excluded, as larger samples are likely to be overrepresented (see table 3). The use of the remaining archival part of the small biopsies may, at least in part, be circumvented by better tissue management, where during the first cutting of the small biopsy sample, blank slides are set aside. These can be used, depending of the histological diagnosis, for future diagnostic, predictive and research purposes [41].

In conclusion, ALK IHC + FISH- NSCLC is infrequent and associated with a worse prognosis on personalized treatment. In combination with a similar trend in ALK FISH + IHC- discordant cases [10], a suitable predictive testing strategy may be to screen first with IHC and then confirm with FISH instead of considering ALK IHC equivalent to ALK FISH according to the current ESMO [42,43] and CAP, AMP, IASLC [44] guidelines.

Funding

This work was supported by an 'investor initiated research' grant by Pfizer.

Declaration of competing interest

Dr. Thunnissen reports Inverstigator Initiated Study grant from

Pfizer to VUmc, Amsterdam for the conduct of the study.

Dr. De Langen reports grants from AstraZeneca, BMS, MSD, Boehringer, non-financial support from Roche, Merck-Serono, outside the submitted work.

Dr. Monkhorst reports grants from AstraZeneca, MSD, Roche, personal fees from MSD, Roche, AstraZeneca, Pfizer, BMS, Roche, MSD, Abbvie, AstraZeneca, Diaceutics, outside the submitted work.

Dr. Postmus reports personal fees from Novartis, MSD, Celgene, Abbvie, AstraZeneca, Roche, BMS, Eli Lilly, Precision Oncology, outside the submitted work.

Dr. Gosney reports personal fees, non-financial support from Astra Zeneca, Boehringer Ingelheim, Bristol-Meyers Squibb, Merck Sharp and Dome, Roche, grants, personal fees, non-financial support from Pfizer outside the submitted work.

Dr. Dingemans reports grants and personal fees from BMS, personal fees from Roche, MSD, Eli Lily, Takeda, Pfizer, Boehringer Ingelheim, outside the submitted work.

Dr. Speel reports personal fees from Pfizer, MSD, Roche, AbbVie, Novartis, BMS, outside the submitted work.

Dr. Buettner reports personal fees from RB is a co-owner and Chief Scientific Officer of Targos Molecular Pathology, Inc., outside the submitted work.

Dr. Bubendorf reports personal fees from Pfizer, grants and personal fees from Roche, during the conduct of the study; personal fees from Bayer, MSD, BMS, outside the submitted work.

Dr. Schuuring reports grants, personal fees and non-financial support from Biocartis, personal fees and non-financial support from BMS, Astrazeneca, Pfizer, Roche, Bayer, Illumina, Jansen Cilag, Agena Biosciences, grants from CC Diagnostics, Boehringer Ingelheim, outside the submitted work.

Dr. Pauwels reports grants and other from Pfizer, other from Roche, outside the submitted work.

Dr. Ivonne E. Marondel, Ph.D. is an employee of and owns stock in Pfizer.

Dr. Finn reports personal fees from Pfizer, Roche, MSD, Sanofi, outside the submitted work.

Dr. Heukamp reports and Advisory Role for NEO New Oncology GmbH.

Dr. Timens reports personal fees from Pfizer, personal fees from GSK, Chiesi, Roche Diagnostics / Ventana, Biotest, Merck Sharp Dohme, Novartis, Lilly Oncology, Boehringer Ingelheim, Astra-Zeneca, Bristol-Myers-Squibb, AbbVie, grants from Dutch Asthma Fund, outside the submitted work.

Dr. PENAULT-LLORCA reports personal fees and non-financial support from PFIZER, NOVARTIS, grants, personal fees and non-financial support from ROCHE, outside the submitted work.

Dr. van der Drift, Lissenberg-Witte, Skov, Duin, Sørensen, Looijen-Salamon, Weynand, Hashemi, O Grady, Akyurek, Mellemgaard, Samii, Bahce, van den Heuvel, *Durando*, Duplaquet, *Oz, Wolf*, and Smit have nothing to disclose.

Acknowledgements

This study was funded by an unrestricted research grant by Pfizer, the Netherlands. The support of Annemieke Hiemstra during the administrative initiation of the study is greatly appreciated.

Appendix A. Supplementary data

Supplementary material related to this article can be found, in the online version, at doi:https://doi.org/10.1016/j.lungcan.2019.09.023.

References

 M. Soda, Y.L. Choi, M. Enomoto, et al., Identification of the transforming EML4-ALK fusion gene in non-small-cell lung cancer, Nature 448 (7153) (2007) 561–566. [2] B.J. Solomon, D.W. Kim, Y.L. Wu, et al., Final overall survival analysis from a study comparing first-line crizotinib versus chemotherapy in alk-mutation-positive non-small-cell lung cancer, J. Clin. Oncol. 36 (22) (2018) 2251–2258.

- [3] N.I. Lindeman, P.T. Cagle, M.B. Beasley, et al., Molecular testing guideline for selection of lung Cancer patients for EGFR and ALK tyrosine kinase inhibitors: guideline from the college of american pathologists, international association for the study of lung cancer, and association for molecular patho, J. Mol. Diagn. 15 (4) (2013) 415–453.
- [4] S. Novello, F. Barlesi, R. Califano, et al., Metastatic non-small-cell lung cancer: ESMO clinical practice guidelines for diagnosis, treatment and follow-up, Ann. Oncol. Off. J. Eur. Soc. Med. Oncol. 27 (suppl. 5) (2016) v1–v27.
- [5] E. Thunnissen, T.C. Allen, J. Adam, et al., Immunohistochemistry of pulmonary biomarkers: a perspective from members of the pulmonary pathology society, Arch. Pathol. Lab. Med. 142 (2018) 408–419, https://doi.org/10.5858/arpa.2017-0106-SA
- [6] A. McLeer-Florin, M. Duruisseaux, J. Pinsolle, et al., ALK fusion variants detection by targeted RNA-next generation sequencing and clinical responses to crizotinib in ALK-positive non-small cell lung cancer, Lung Cancer 116 (September 2017) (2018) 15–24
- [7] A. Marchetti, A. Di Lorito, M.V. Pace, et al., ALK protein analysis by IHC staining after recent regulatory changes: a comparison of two widely used approaches, revision of the literature, and a new testing algorithm, J. Thorac. Oncol. 11 (4) (2016) 487–495.
- [8] I. Letovanec, S. Finn, P. Zygoura, et al., Evaluation of NGS and RT-PCR methods for ALK rearrangement in european NSCLC patients: results from the european thoracic oncology platform lungscape project, J. Thorac. Oncol. 13 (3) (2018) e33–e34.
- [9] J.S. Jang, X. Wang, P.T. Vedell, et al., Custom gene capture and next-generation sequencing to resolve discordant ALK status by FISH and IHC in lung adenocarcinoma, J. Thorac. Oncol. 11 (11) (2016) 1891–1900.
- [10] A.J. van der Wekken, R. Pelgrim, N. 't Hart, et al., Dichotomous ALK-IHC is a better predictor for ALK inhibition outcome than traditional ALK-FISH in advanced nonsmall cell lung Cancer, Clin. Cancer Res. (2017), https://doi.org/10.1158/1078-0432.CCR-16-1631.
- [11] J.-C. Soria, S.N. Ho, M. Varella-Garcia, et al., Correlation of extent of ALK FISH positivity and crizotinib efficacy in three prospective studies of ALK-positive patients with non-small-cell lung cancer, Ann. Oncol. Off. J. Eur. Soc. Med. Oncol. 29 (9) (2018) 1964–1971.
- [12] A. Rosoux, P. Pauwels, F. Duplaquet, et al., Effectiveness of crizotinib in a patient with ALK IHC-positive/FISH-negative metastatic lung adenocarcinoma, Lung Cancer 98 (2016) 118–121.
- [13] F. Cabillic, A. Gros, F. Dugay, et al., Parallel FISH and immunohistochemical studies of ALK status in 3244 non-small-cell lung cancers reveal major discordances, J. Thorac. Oncol. 9 (3) (2014) 295–306.
- [14] M.I. Ilie, C. Bence, V. Hofman, et al., Discrepancies between FISH and immunohistochemistry for assessment of the ALK status are associated with ALK' borderline'-positive rearrangements or a high copy number: a potential major issue for anti-ALK therapeutic strategies, Ann. Oncol. 26 (1) (2015) 238–244.
- [15] M. Pekar-Zlotin, F.R. Hirsch, L. Soussan-Gutman, et al., Fluorescence in situ hybridization, immunohistochemistry, and next-generation sequencing for detection of EML4-ALK rearrangement in lung cancer, Oncologist 20 (3) (2015) 316–322.
- [16] N. Peled, G. Palmer, F.R. Hirsch, et al., Next-generation sequencing identifies and immunohistochemistry confirms a novel crizotinib-sensitive ALK rearrangement in a patient with metastatic non-small-cell lung cancer, J. Thorac. Oncol. 7 (9) (2012) 14–16.
- [17] Blackhall FHFH, S. Peters, L. Bubendorf, et al., Prevalence and clinical outcomes for patients with ALK-positive resected stage I to III adenocarcinoma: results from the European Thoracic Oncology Platform Lungscape Project, J. Clin. Oncol. 32 (25) (2014) 2780–2787.
- [18] M. ter Wee, B. Lissenberg-Witte, A Quick Guide on How to Conduct Medical Research, From set-up to publication, Houten: Bohn Stafleu van Loghem, 2019, https://doi.org/10.1007/978-368-2248-0.
- [19] A.T. Shaw, D.-W. Kim, K. Nakagawa, et al., Crizotinib versus chemotherapy in advanced ALK -positive lung cancer, N. Engl. J. Med. 368 (25) (2013) 2385–2394.
- [20] S. Peters, D.R. Camidge, A.T. Shaw, et al., Alectinib versus Crizotinib in untreated ALK-positive non-small-cell lung cancer, N. Engl. J. Med. (2017) NEJMoa1704795.
- [21] F. Barlesi, J. Mazieres, J.-P. Merlio, et al., Routine molecular profiling of patients with advanced non-small-cell lung cancer: results of a 1-year nationwide programme of the French Cooperative Thoracic Intergroup (IFCT), Lancet (London, England) 387 (10026) (2016) 1415–1426.
- [22] D.R. Camidge, H.R. Kim, M.-J. Ahn, et al., Brigatinib versus Crizotinib in ALK -positive non-small-cell lung cancer, N. Engl. J. Med. (2018) NEJMoa1810171.
- [23] T. Mok, S. Peters, D. Camidge, et al., JCES 01. 27 Patients with ALK IHC-Positive/ FISH-Negative NSCLC Benefit from ALK TKI Treatment: Response Data from the Global ALEX Trial, J. Thorac. Oncol. 12 (S2(11)) (2017) S1739–S1740.
- [24] F. Zhao, M. Xu, H. Lei, et al., Clinicopathological characteristics of patients with non-small-cell lung cancer who harbor EML4-ALK fusion gene: a meta-analysis, PLoS One 10 (2) (2015) e0117333.
- [25] B.G. Skov, P. Clementsen, K.R. Larsen, et al., The prevalence of ALK rearrangement in pulmonary adenocarcinomas in an unselected Caucasian population from a defined catchment area: impact of smoking, Histopathology 70 (6) (2017) 889–895.
- [26] J.H. Paik, C.-M. Choi, H. Kim, et al., Clinicopathologic implication of ALK rearrangement in surgically resected lung cancer: a proposal of diagnostic algorithm for ALK-rearranged adenocarcinoma, Lung Cancer 76 (3) (2012) 403–409.
- [27] J.H. Paik, G. Choe, H. Kim, et al., Screening of anaplastic lymphoma kinase rearrangement by immunohistochemistry in non-small cell lung cancer: correlation with fluorescence in situ hybridization, J. Thorac. Oncol. 6 (3) (2011) 466–472.

[28] A.S. Williams, W. Greer, D. Bethune, et al., ALK+ lung adenocarcinoma in never smokers and long-term ex-smokers: prevalence and detection by immunohistochemistry and fluorescence in situ hybridization, Virchows Arch. 469 (5) (2016) 533–540.

- [29] K. Takeuchi, Y.L. Choi, M. Soda, et al., Multiplex reverse transcription-PCR screening for EML4-ALK fusion transcripts, Clin. Cancer Res. 14 (20) (2008) 6618–6624.
- [30] L. Jiang, H. Yang, P. He, et al., Improving selection criteria for ALK inhibitor therapy in non-small cell lung cancer: a pooled-data analysis on diagnostic operating characteristics of immunohistochemistry, Am. J. Surg. Pathol. 40 (5) (2016) 697–703.
- [31] IASLC Atlas of ALK and ROS1 Testing in Lung Cancer, 2nd ed., Editorial Rx Press, 2016.
- [32] M. von Laffert, A. Stenzinger, M. Hummel, et al., ALK-FISH borderline cases in non-small cell lung cancer: implications for diagnostics and clinical decision making, Lung Cancer 90 (2015) 465–471.
- [33] J. Heuckmann, P. Pauwels, E. Thunnissen, Comprehensive hybrid capture–Based next- generation sequencing identifies a double ALK gene fusion in a patient previously identified to Be false-negative by FISH, J. Thorac. Oncol. 12 (3) (2017) e22–24.
- [34] H. Kim, X. Xu, S.-B. Yoo, et al., Discordance between anaplastic lymphoma kinase status in primary non-small-cell lung cancers and their corresponding metastases, Histopathology 62 (2) (2013) 305–314.
- [35] M. Salido, L. Pijuan, L. Martínez-Avilés, et al., Increased ALK gene copy number and amplification are frequent in non-small cell lung cancer, J. Thorac. Oncol. 6 (1) (2011) 21–27.
- [36] Q. Wang, L. Zhao, X. Yang, et al., Antibody 1A4 with routine immunohistochemistry demonstrates high sensitivity for ALK rearrangement

- screening of Chinese lung adenocarcinoma patients: a single-center large-scale study, Lung Cancer 95 (2016) 39-43.
- [37] M. Ibrahim, S. Parry, D. Wilkinson, et al., ALK immunohistochemistry in NSCLC: discordant staining can impact patient treatment regimen, J. Thorac. Oncol. 11 (12) (2016) 2241–2247.
- [38] Y. Murakami, T. Mitsudomi, Y. Yatabe, A screening method for the ALK fusion gene in NSCLC, Front. Oncol. 2 (March) (2012) 24.
- [39] H. Nakamura, K. Tsuta, A. Yoshida, et al., Aberrant anaplastic lymphoma kinase expression in high-grade pulmonary neuroendocrine carcinoma, J. Clin. Pathol. 66 (8) (2013) 705–707.
- [40] K. Takeuchi, Interpretation of Anti-ALK immunohistochemistry results, J. Thorac. Oncol. 8 (7) (2013) e67–e68.
- [41] L. Bubendorf, S. Lantuejoul, A.J. de Langen, E. Thunnissen, Nonsmall cell lung carcinoma: diagnostic difficulties in small biopsies and cytological specimens, Eur. Respir. Rev. 26 (144) (2017) 170007.
- [42] D. Planchard, S. Popat, K. Kerr, et al., Metastatic non-small cell lung cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up, Ann. Oncol. Off. J. Eur. Soc. Med. Oncol. 29 (Supplement_4) (2018) iv192–iv237.
- [43] Y.L. Wu, D. Planchard, S. Lu, et al., Pan-asian adapted clinical practice guidelines for the management of patients with metastatic non-small-cell lung cancer: a escoesmo initiative endorsed by jsmo,ksmo, mos, sso and tos, Ann. Oncol. 30 (2) (2019) 171–210
- [44] N.I. Lindeman, P.T. Cagle, D.L. Aisner, et al., Updated molecular testing guideline for the selection of lung cancer patients for treatment with targeted tyrosine kinase inhibitors: guideline from the college of american pathologists, the international association for the study of lung cancer, and the association for molecular pathology, Arch. Pathol. Lab. Med. 142 (3) (2018) 321–346.