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Dietary *Sargassum fusiforme* improves memory and reduces amyloid plaque load in an Alzheimer's disease mouse model

Jeroen Bogie¹, Cindy Hoeks¹, Melissa Schepers^{1,9}, Assia Tiane^{1,9}, Ann Cuypers², Frank Leijten³, Yupyn Chintapakorn⁴, Thiti Suttiyut⁴, Surachai Pornpakakul⁵, Dicky Struik⁶, Anja Kerksiek⁷, Hong-Bing Liu⁸, Niels Hellings¹, Pilar Martinez-Martinez⁹, Johan W. Jonker⁶, Ilse Dewachter¹, Eric Sijbrands¹⁰, Jochen Walter¹⁰, Jerome Hendriks¹, Albert Groen¹¹, Bart Staels¹², Dieter Lütjohann⁷, Tim Vanmierlo^{1,9} & Monique Mulder³

Activation of liver X receptors (LXRs) by synthetic agonists was found to improve cognition in Alzheimer's disease (AD) mice. However, these LXR agonists induce hypertriglyceridemia and hepatic steatosis, hampering their use in the clinic. We hypothesized that phytosterols as LXR agonists enhance cognition in AD without affecting plasma and hepatic triglycerides. Phytosterols previously reported to activate LXRs were tested in a luciferase-based LXR reporter assay. Using this assay, we found that phytosterols commonly present in a Western type diet in physiological concentrations do not activate LXRs. However, a lipid extract of the 24(S)-Saringosterol-containing seaweed *Sargassum fusiforme* did potentially activate LXR β . Dietary supplementation of crude *Sargassum fusiforme* or a *Sargassum fusiforme*-derived lipid extract to AD mice significantly improved short-term memory and reduced hippocampal A β plaque load by 81%. Notably, none of the side effects typically induced by full synthetic LXR agonists were observed. In contrast, administration of the synthetic LXR α activator, AZ876, did not improve cognition and resulted in the accumulation of lipid droplets in the liver. Administration of *Sargassum fusiforme*-derived 24(S)-Saringosterol to cultured neurons reduced the secretion of A β_{42} . Moreover, conditioned medium from 24(S)-Saringosterol-treated astrocytes added to microglia increased phagocytosis of A β . Our data show that *Sargassum fusiforme* improves cognition and alleviates AD pathology. This may be explained at least partly by 24(S)-Saringosterol-mediated LXR β activation.

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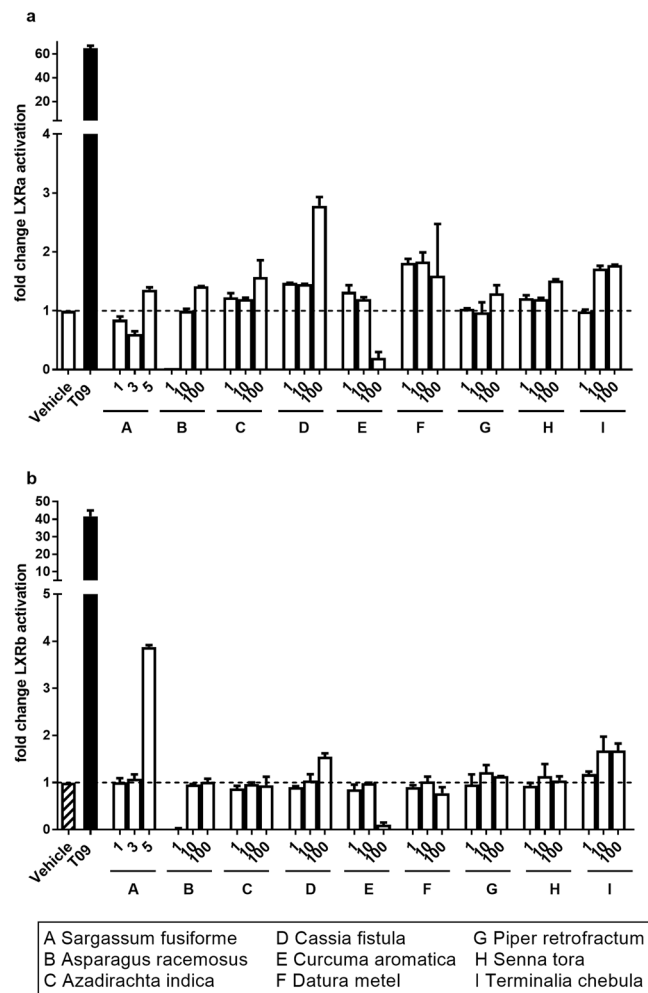


Figure 1. A crude lipid extract of *Sargassum fusiforme* activates LXR β . LXR α (a) and LXR β (b) activation was assessed with a luciferase-based reporter assay. CHME3 cells were stimulated for 18 hours with vehicle (striped bar/dotted line), 1, 3, or 5 $\mu\text{g}/\mu\text{l}$ *Sargassum fusiforme* or 1, 10 or 100 $\mu\text{g}/\mu\text{l}$ of the specified plant extracts. No difference was found between groups (LXR α $\chi^2(7) = 11.55$, $p = 0.1165$, LXR β $\chi^2(7) = 7.689$, $p = 0.3608$; all datasets analysed using Kruskal-Wallis test). All results are displayed as fold change compared to vehicle control (striped bar/dotted line). Bars represent mean \pm SEM ($n \geq 3$).

Alzheimer's disease (AD) is a progressive neurological disorder characterized by an accumulation of extracellular amyloid- β (A β), intracellular neurofibrillary tangles, loss of synapses, neuroinflammation, and by a gradual progression of memory loss¹. Accumulating evidence suggests a role for a disturbed cholesterol turnover in the central nervous system (CNS) in AD pathogenesis^{2–11}. In line with this, stimulation of cholesterol turnover improves disease outcome in animal models of AD^{2,12–18}. Liver X receptors (LXR) are master regulators of cholesterol and triglyceride turnover and suppress an inflammatory transcriptional profile via trans-repression of NF κ B signaling¹⁹. Therefore, LXRs are promising well-studied therapeutic targets for increasing cholesterol turnover and decreasing neuroinflammation in AD^{20–24}. We and others have reported that synthetic pan LXR agonists improve the cognitive phenotype in animal models of AD, decrease synaptic compensatory mechanisms, and stimulate the proteolytic degradation of A β by microglia^{2,13,14,24–26}. However, synthetic full LXR agonists systematically cause adverse side effects, such as hypertriglyceridemia and hepatic steatosis, hampering their translation to the clinic^{27–30}.

Phytosterols are structurally similar to cholesterol. However, in contrast to cholesterol, they can cross the blood-brain barrier (BBB) and accumulate in brain parenchyma^{11,31–33}. Several of the more than 260 identified phytosterols, such as β -sitosterol, fucosterol, stigmasterol, schottenol, 24(S)-Saringosterol, and spinasterol, have been reported to activate LXRs *in vitro*^{34–39}. Moreover, β -sitosterol and stigmasterol modulate AD pathology in *in vitro* models for AD^{40–42}. Phytosterols do not induce hypertriglyceridemia and hepatic steatosis, which may be a consequence of their ABCG5/G8-mediated hepatic excretion into the bile^{11,43,44}. The absence of unwanted side effects renders phytosterols interesting therapeutic candidates for inducing LXR activation in the CNS.

We aim to identify phytosterols and phytosterol-containing extracts that activate LXRs *in vitro*, to test their effect on memory performance and A β plaque pathology in an animal model of AD. We found that phytosterols typically present in a Western type diet or extracts from a range of Eastern plants hardly activate LXR α

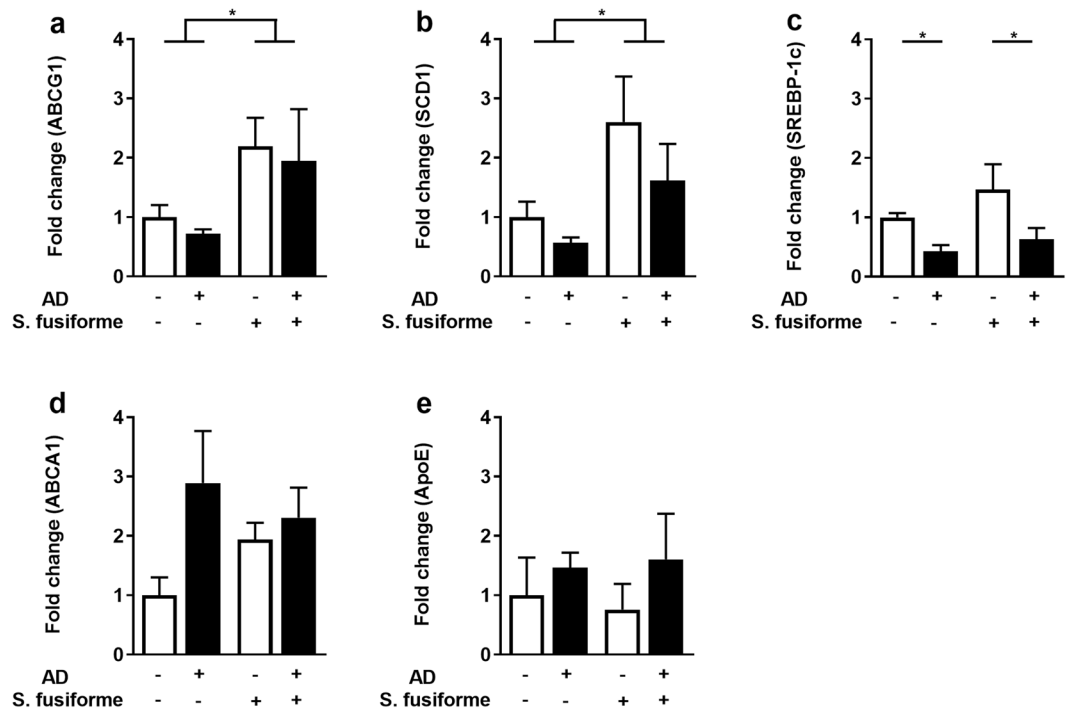


Figure 2. Dietary supplementation with *Sargassum fusiforme* activates LXRs in the brain. Gene expression of *Abcg1* (a), *Scd1* (b), *Srebp-1c* (c), *Abca1* (d), and *ApoE* (e) was measured in the brain of WT and APPswPS1ΔE9 mice (AD) fed normal chow or chow supplemented with *Sargassum fusiforme*. Two-way ANOVA revealed a diet effect for *Abcg1* ($F(1, 17) = 6.244, p = 0.0230$), and *Scd1* ($F(1, 15) = 7.673, p = 0.0143$), and a genotype effect for *Srebp-1c* ($F(1, 16) = 8.446, p = 0.0103$). Gene expression was normalized to *CycA* and *Hmbs*, and expressed as fold change compared to WT mice fed the control diet. Bars represent mean \pm SEM ($n \geq 5$).

or LXR β . In contrast, a lipid extract of the edible brown seaweed *Sargassum fusiforme* which contains large amounts of 24(S)-Saringosterol potentially activated LXR β . In a mouse model for AD, dietary supplementation with *Sargassum fusiforme* or its lipid extract not only increased the expression of LXR response genes in the CNS, but also improved cognition without inducing hepatic steatosis. Improved memory performance in these mice was paralleled by a strong reduction in CNS A β plaque load. In contrast, the selective LXR α agonist AZ876 did not counteract cognitive decline in AD mice or reduce A β plaque load, and induced liver steatosis. These findings indicate that *Sargassum fusiforme* is an attractive option for add-on treatments in the emerging field of nutritional neuroscience.

Results

Phytosterols present in a Western diet do not activate LXRs. First, we determined the capacity of phytosterols typically present in a Western diet to activate LXRs. To define cellular specificity in LXR activation *in vitro*, cells derived from peripheral tissues (HEK293.T and COS7) and the CNS (CHME3, MO3.13, N2a/APPswE) were tested. Physiologically relevant concentrations of stigmasterol, fucosterol, brassicasterol, β -sitosterol, or a mix of phytosterols did not activate LXR α or LXR β in any of the cell lines used (Fig. S2). An increased incubation period promoted the capacity of the synthetic LXR agonist T0901317, but not of phytosterols, to activate LXR α and LXR β (Fig. S3a,b). Although cellular uptake of phytosterols and their capacity to activate nuclear receptors improves when complexed to proteins^{45,46}, pre-incubation of phytosterols with BSA did not significantly increase their ability to activate LXR α and LXR β (Fig. S3c,d).

An extract of *Sargassum fusiforme* activates LXR β *in vitro*. In addition to phytosterols present in a Western diet, crude extracts of Eastern plants were screened for their capacity to activate LXR α or LXR β . Plants were chosen based on their application in Asian traditional medicine and their presence in an Eastern diet. Extracts from *Asparagus racemosus*, *Azadirachta indica*, *Cassia fistula*, *Curcuma aromatica*, *Datura metel*, *Piper retrofractum*, *Senna tora*, and *Terminalia chebula* did not significantly activate either LXR α or LXR β in microglial CHME3 cells (Fig. 1). However, at a dose of 5 μ g/ml an extract of *Sargassum fusiforme* containing the recently identified LXR β agonist 24(S)-Saringosterol³⁴ significantly activated LXR β but not LXR α (Fig. 1). Higher doses of *Sargassum fusiforme* induced cell death (data not shown). As *Sargassum fusiforme* showed the highest capacity to activate LXR β , it was selected for further *in vivo* testing.

Dietary supplementation with *Sargassum fusiforme* results in 24(S)-Saringosterol accumulation in the cerebellum and activation of LXR-response genes in AD mice. To assess *in vivo* effects

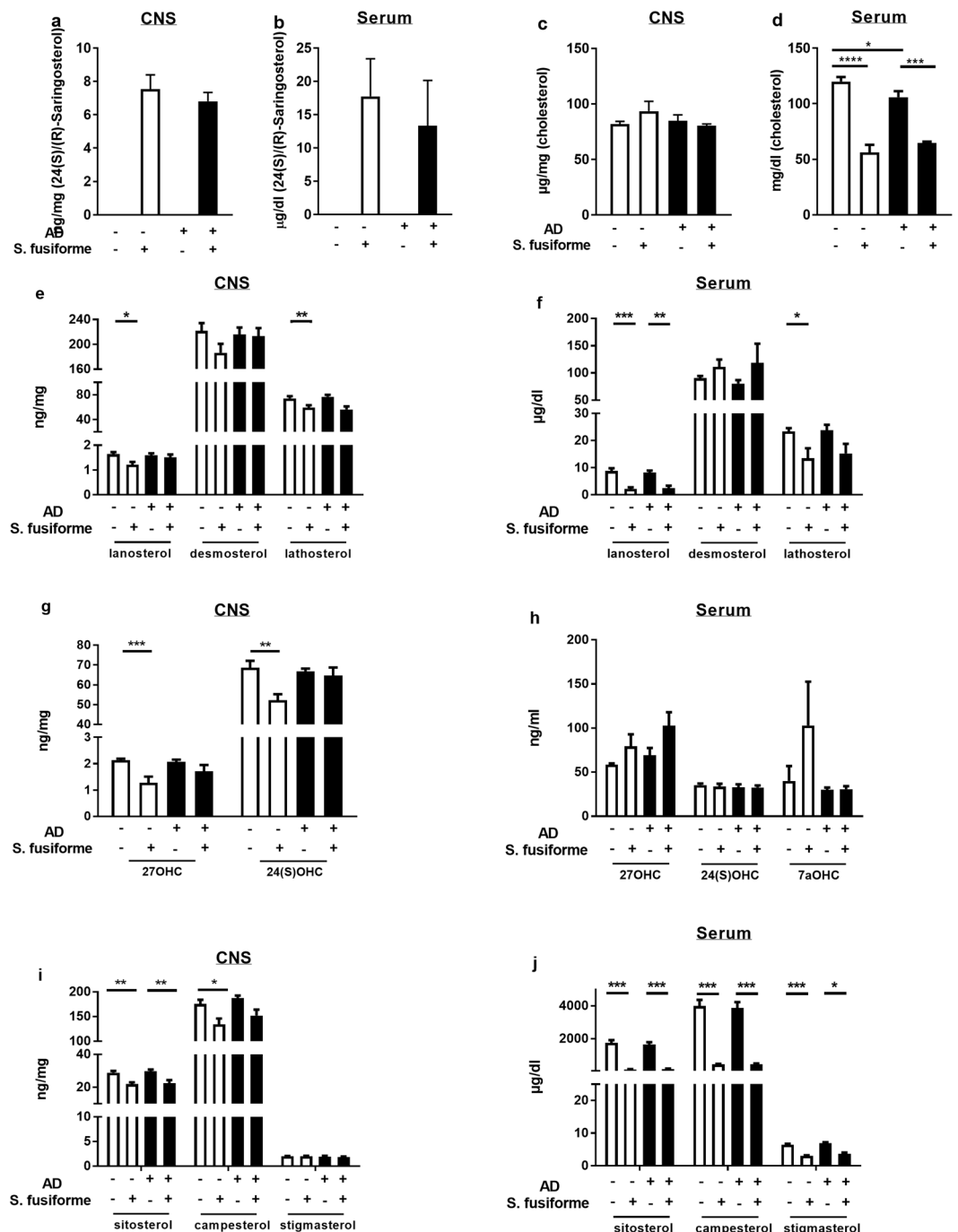


Figure 3. Dietary supplementation with *Sargassum fusiforme* reduces serum cholesterol, and increases 24(S)/(R)-Saringosterol in serum and cerebellum. Concentrations of 24(S)/(R)-Saringosterol (a,b), cholesterol (c,d), cholesterol precursors (e,f), cholesterol metabolites (g,h), and phytosterols (i,j) were determined in serum and cerebellum samples of WT and APPswePS1ΔE9 mice (AD) fed either control chow or chow supplemented with *Sargassum fusiforme*. 24(S)/(R)-Saringosterol concentration was affected by diet in CNS ($F(1, 31) = 438.8, p < 0.0001$) and serum ($F(1, 31) = 30.05, p < 0.0001$) as determined by ANOVA, and post-hoc Sidak's test showed that this effect existed in both genotypes (WT CNS: $p < 0.0001$; WT serum: $p = 0.0005$; AD CNS: $p < 0.0001$; AD serum: $p = 0.0157$). Cholesterol concentration in serum was affected by diet ($F(3, 31) = 80.59, p < 0.0001$) for both genotypes (WT: $p < 0.001$; AD: $p = 0.0002$). Regarding the cholesterol precursors, concentration of lanosterol was affected by diet in CNS ($F(1, 32) = 6.463, p = 0.0161$) and serum ($F(1, 31) = 38.28, p < 0.0001$) of WT animals (CNS: $p = 0.0265$; serum: $p = 0.0002$) and AD animals (serum: $p = 0.0024$). Concentration of lathosterol was affected by diet in CNS ($F(1, 32) = 19.32, p = 0.0001$) and serum ($F(1, 31) = 13.42, p = 0.0009$) of WT animals (CNS: $p = 0.0076$; serum: $p = 0.0474$). Regarding the cholesterol metabolites, concentration of 27-hydroxycholesterol (27OHC) was affected by diet in CNS ($F(1, 32) = 20.04, p = 0.0009$) of WT animals ($p = 0.0004$). Concentration of 24(S)-hydroxycholesterol (24(S)OHC) was affected by diet in CNS ($F(1, 31) = 8.071, p = 0.0079$) of WT animals ($p = 0.0079$). Regarding the

plant sterols, concentration of sitosterol was affected by diet in CNS ($F(1, 32) = 30.62, p < 0.0001$) and serum ($F(1, 31) = 96.88, p < 0.0001$) of WT animals (CNS: $p = 0.0026$; serum: $p < 0.0001$) and AD animals (CNS: $p = 0.0028$; serum: $p < 0.0001$). Concentration of campesterol was affected by diet in CNS ($F(1, 32) = 17.02, p = 0.0002$) and serum ($F(1, 31) = 76.4, p < 0.0001$) of WT animals (CNS: $p = 0.0175$; serum: $p < 0.0001$) and AD animals (serum: $p < 0.0001$). Concentration of stigmasterol was affected by diet in serum ($F(1, 31) = 99.89, p < 0.0001$) of WT animals ($p < 0.0001$) and AD animals ($p < 0.0001$). Bars represent mean \pm SEM ($n = 5$). Abbreviations: 7aOHC 7a-hydroxycholesterol.

of *Sargassum fusiforme* APPswePS1 Δ E9 mice were used as a model of AD⁴⁷. These mice begin to develop A β plaques at 4 months of age and cognitive decline occurs from 6 months onwards^{48,49}. From the age of 5 months APPswePS1 Δ E9 mice and WT littermates were fed either standard chow or chow supplemented with 50% (w/w) dried crude *Sargassum fusiforme*. Ten weeks of *Sargassum fusiforme* dietary supplementation resulted in LXR activation in the CNS, evidenced by a cerebral induction of LXR response genes (*Abcg1, Scd1*) (Fig. 2a,b). *ApoE, Srebp-1c*, and *Abcg1* expression was not altered in animals treated with *Sargassum fusiforme* (Fig. 2c–e). Interestingly, 24(S)/(R)-Saringosterol was detectable in serum and in the cerebellum of animals that were fed *Sargassum fusiforme*, but not in those fed normal chow (Fig. 3a,b). As *Sargassum fusiforme* contains predominantly 24(S)-Saringosterol³⁴, we postulate that this isoform is the most abundant isoform present in animals fed *Sargassum fusiforme*. Animals fed a diet supplemented with *Sargassum fusiforme* did not show differences in total cholesterol content in the cerebellum (Fig. 3c), but showed a marked reduction in circulating cholesterol levels compared to chow fed animals (Fig. 3d). Levels of cholesterol precursors, cholesterol metabolites, and phytosterols were decreased in serum and cerebellum of animals supplemented with *Sargassum fusiforme* (Fig. 3e–j). On a standard chow diet, WT mice displayed significantly higher serum cholesterol levels than AD mice (Fig. 3d), as described previously^{10,50}.

***Sargassum fusiforme* improves memory and reduced A β plaque load in AD mice.** Next, we investigated the impact of dietary supplementation with crude *Sargassum fusiforme* on cognition and neuropathology in APPswePS1 Δ E9 mice. To assess the influence of selective LXR α activation on cognition, animals were fed with the synthetic selective LXR α agonist AZ876. The open field test revealed no effect of genotype or diet on general locomotor activity, and anxiety (Table 1). Dietary supplementation with *Sargassum fusiforme* significantly improved object memory in APPswePS1 Δ E9 mice in the object recognition task (ORT) at a 1 h but not a 24 h inter-trial interval (Fig. 4a,b). In contrast, selective LXR α activation did not impact object memory at either the 1 h or 24 h inter-trial interval. *Sargassum fusiforme*, but not AZ876, markedly decreased A β plaque load in the cortex (70% decrease; Fig. 4c,e) and hippocampus (81% decrease; Fig. 4d,e) in APPswePS1 Δ E9 mice. In concordance, *Sargassum fusiforme* supplementation reduced A β 40 protein and APP mRNA expression in the CNS of AD animals (Fig. 4f,h). *Sargassum fusiforme* did not significantly reduce insoluble A β 42 (Fig. 4g). Reduced A β plaque load was not associated with an increased expression of phagocytic receptors such as Axl and MerTK (Fig. S4a,b)⁵¹. Yet, the reduced A β plaque load was associated with a reduced TREM2 expression, reflecting the expelled need for further A β clearance (Fig. S4c).

To determine whether the lipid moiety of *Sargassum fusiforme* was sufficient to modulate the AD-related pathology, a purified lipid extract of *Sargassum fusiforme* was administered daily by oral gavage to APPswePS1 Δ E9 and WT littermates for 45 days. Working memory was significantly improved in APPswePS1 Δ E9 mice, resulting in the prevention of the spatial working deficit (Fig. 5a,b). In line with these findings, a significant reduction in insoluble A β ₄₂ was observed in the cortex (99% decrease; Fig. 5c) and hippocampus (57% decrease; Fig. 5d) of the treated APPswePS1 Δ E9 mice. Notably, treatment with the *Sargassum fusiforme* lipid extract resulted in an increased expression of ApoE in the CNS (Fig. 5e–g), which indicates active LXR signaling.

***Sargassum fusiforme* feeding does not lead to liver steatosis.** While synthetic LXR agonists are well-known to induce liver steatosis in mice⁵², *Sargassum fusiforme* supplementation did not induce liver steatosis in APPswePS1 Δ E9 mice, as evidenced by the absence of lipid droplets within the liver of these animals (Fig. 6a,b). Supplementation with AZ876 resulted in a marked increase in lipid droplets in the liver (Fig. 6a,b). Likewise, while AZ876 increased circulating triglyceride levels, *Sargassum fusiforme* did not impact the level of triglycerides (Fig. 6c). Similar findings were observed in WT mice fed chow supplemented with *Sargassum fusiforme* or AZ876 (data not shown). *Sargassum fusiforme* is known to contain relatively high levels of (in)organic arsenic that can accumulate in the body⁵³. Although relatively high levels of arsenic were measured in dried *Sargassum fusiforme* (40.4 mg/kg), no arsenic accumulation was detectable in lung tissue of the animals that consumed *Sargassum fusiforme* (Fig. 6d).

24(S)-Saringosterol increases microglial A β clearance and reduces neuronal A β release. To obtain insight into the mechanisms by which *Sargassum fusiforme*-derived 24(S)-Saringosterol impacts AD pathogenesis we used well-established *in vitro* models that mimic AD-related pathological processes. Supplementation of astrocytes with 24(S)-Saringosterol resulted in an increased ApoE secretion (Fig. 7a,b). Furthermore, supplementation of microglia with conditioned medium of 24(S)-Saringosterol-treated astrocytes promoted microglial clearance of A β _{1–42} (Fig. 7c). 24(S)-Saringosterol did not directly impact the capacity of microglia to internalize A β _{1–42} (Fig. 7d). Finally, 24(S)-Saringosterol was found to reduce the release of A β ₄₂ using neuronal N2a cells overexpressing APP (Fig. 7e).

Genotype	Diet	TIZ centre (s)	TIZ walls and corners (s)	DM (m)	N
WT	Control	29.5 ± 7.7	570 ± 7.7	36.1 ± 1.6	12
AD	Control	29.4 ± 4.2	569.6 ± 4.3	39.0 ± 1.9	13
WT	<i>S. fusiforme</i>	30.9 ± 3.4	564.9 ± 4.3	42.0 ± 1.9	9
AD	<i>S. fusiforme</i>	33.5 ± 4.1	564.9 ± 5.2	47.6 ± 2.0	7
WT	AZ876	28.3 ± 4.3	569.2 ± 4.9	37.3 ± 2.4	11
AD	AZ876	31.7 ± 6.4	567.2 ± 6.4	39.9 ± 2.7	13

Table 1. Genotype and diet do not impact general locomotor activity or anxiety levels. Results of open field test. Values are displayed as mean ± SEM. Abbreviations: TIZ, time in zone; DM, distance moved.

Discussion

Our data show that dietary supplementation with the brown seaweed *Sargassum fusiforme* improves cognitive function and reduces A β load in APPswePS1 Δ E9 mice without inducing liver steatosis. Our data further suggest the involvement of the stereospecific oxyphytosterol 24(S)-Saringosterol, a selective LXR β activator, in the observed effects on cognition and A β plaque load. Considering that selective LXR α activation did not improve cognitive decline or A β plaque deposition in APPswePS1 Δ E9 mice, our findings suggest that natural and possibly also synthetic LXR β agonists are attractive options for the treatment of neurodegenerative disorders such as AD.

Phytosterols, which can cross the blood-brain barrier, have been reported to activate LXRs without inducing hypertriglyceridemia and hepatic steatosis, rendering them potentially suitable candidates for treatment of neurodegenerative disorders¹¹. Yet, in contrast with reported data on LXR activation by phytosterols common in a Western diet^{34–39}, we observed very little if any effect of stigmasterol, brassicasterol, β -sitosterol, fucosterol, or a commercial mix of phytosterols, on activation of LXR α and LXR β *in vitro*. The discrepancy between our data and the literature may be explained by differences in the experimental design, such as phytosterol concentration, incubation time, and cell type and assay used to measure LXR activation. For instance, we used a cell-based luciferase reporter assay and CNS-derived cell lines, as well as HEK293.T and COS-7 cells, to define LXR activation and cellular specificity in LXR activation, while other studies used cell-free assays³⁸. Furthermore, we used phytosterol concentrations in a physiological range that can be reached through dietary supplementation (1–25 μ M), whereas other studies used concentrations far exceeding this physiological range up to 200 μ M³⁶. Of note, compared to synthetic LXR agonists, the reported ability of phytosterols commonly present in a Western diet to activate LXRs is relatively limited^{34–39}. In line with this, no alterations were observed in the expression of LXR-response genes in the brain of ABCG5-knockout mice despite high cerebral levels of common phytosterols²⁶. In summary, our data indicate that phytosterols present in a Western type diet do not activate LXRs at concentrations that can be reached through dietary supplementation¹¹.

Out of the tested extracts of Asian plants, the extract of *Sargassum fusiforme* most potently activated LXR β *in vitro*. This is in line with the recently reported LXR β -activating effects of *Sargassum fusiforme*-derived 24(S)-Saringosterol³⁴. In *Sargassum fusiforme*-fed animals, 24(S)-Saringosterol was detected in the cerebellum in concentrations equal to common oxysterols, suggesting a good CNS bioavailability. The amount of 24(S)-Saringosterol detected in the CNS was comparable to previously reported concentrations of other dietary phytosterols in the CNS^{26,32,33}. With respect to these findings, we found that cerebral expression of LXR-response genes was upregulated in animals fed *Sargassum fusiforme*-enriched chow, further supporting the ability of *Sargassum fusiforme* to activate LXR β . The route of entry of 24(S)-Saringosterol to the CNS remains unclear. Further research is warranted to define the role of ApoE and SR-B1 in facilitating the transport of 24(S)-Saringosterol into the CNS^{11,32}. Moreover, since 24(S)-Saringosterol is oxydated plant sterol, we hypothesize that 24(S)-Saringosterol can enter the CNS via transcellular diffusion, comparable to other oxidized cholesterol derivatives such as 24(S)-Hydroxycholesterol⁵⁴. Interestingly, in contrast to 24(S)-Saringosterol, all plant sterol concentrations decreased in serum and CNS upon feeding *Sargassum fusiforme*. It can be hypothesized that 24(S)-Saringosterol activates intestinal and hepatic Abcg5/g8 transporters and thereby drives the enterohepatic secretion of plant sterols, thereby lowering circulating levels of sterols⁵⁵. The latter hypothesis is in line with the strong reduction in circulating cholesterol levels we found in this study.

Our data indicate that the LXR α selective agonist AZ876 does not significantly counteract cognitive decline or reduce A β plaque load in APPswePS1 Δ E9 mice. However, no active LXR signaling was found in the CNS of AZ876 treated animals, which suggest poor CNS bioavailability of AZ876 and consequently does not allow us to conclude that LXR α activation does not impact AD pathology. On the other hand, while both LXR α and LXR β are expressed in the murine CNS, expression of LXR β is 15-fold higher compared to LXR α ^{56,57}. Hence, the relative contribution of LXR α activation upon full-LXR agonist treatment is therefore expected to be limited. The latter studies may suggest that the beneficial impact of pan-LXR agonists on cognitive decline in AD models likely relies on LXR β activation.

Synthetic LXR agonists that reduce AD pathology and improve cognitive performance induce hypertriglyceridemia and liver steatosis^{2,13,14,24,25,27–30}. *Sargassum fusiforme* supplementation counteracted cognitive decline of APPswePS1 Δ E9 transgenic mice without inducing hepatic steatosis. This may be explained by selective hepatic LXR β activation, since animals fed AZ876 (2 mg/kg body weight) displayed a marked increase of liver lipid droplets, indicative of liver steatosis. Alternatively, the presence of other protective compounds present in *Sargassum fusiforme* such as fucosterol may prevent hepatic steatosis¹¹. This is supported by the observation that activation of LXR β by synthetic agonists was also reported to induce plasma and hepatic hypertriglyceridemia⁵⁸.

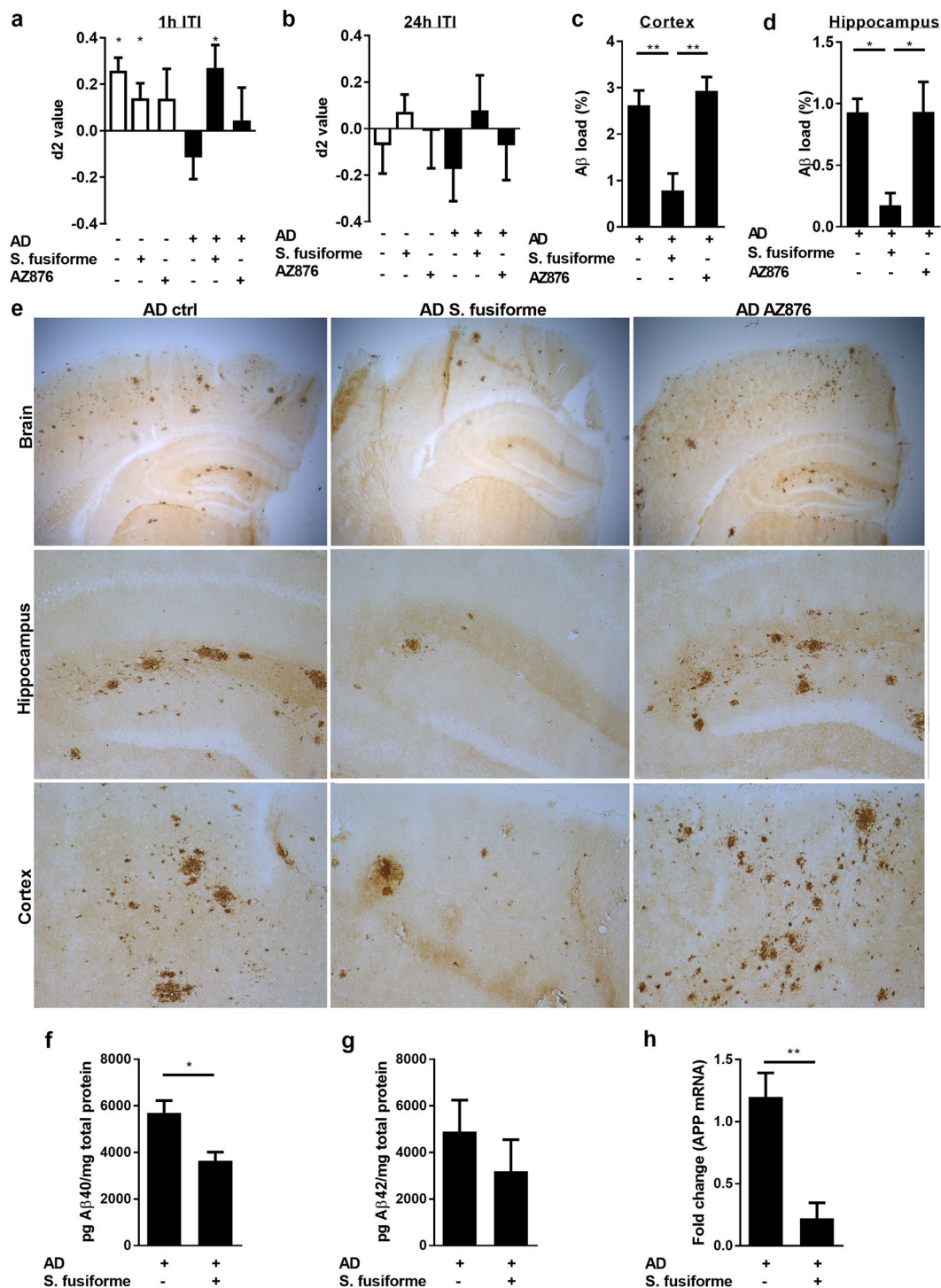


Figure 4. Dietary supplementation with *Sargassum fusiforme* reduces cognitive decline and A β plaque load in APPswePS1 Δ E9 mice. **(a,b)** Cognitive functioning was determined using the object recognition task. The interval between first and second trial was set at 1 hour (1 h ITI) or 24 hours (24 h ITI). D2 value is calculated as the ratio between exploration time spent at the new object and the familiar object in the second trial, with d2 value > 0 indicating intact object memory. At 1 h ITI the object memory was found to be intact in WT animals on control diet ($t(9) = 4.71$, $p = 0.0011$, one sample t-test), in WT animals on *Sargassum fusiforme*-supplemented diet ($t(9) = 2.166$, $p = 0.0585$, one sample t-test), and in AD animals on *Sargassum fusiforme*-supplemented diet ($t(8) = 2.77$, $p = 0.0243$, one sample t-test). Bars represent mean \pm SEM from two independent experiments ($n \geq 9$ per treatment). **(c–e)** A β plaque load was quantified in cortex **(d)** and hippocampus **(e)** of APPswePS1 Δ E9 mice using immunohistochemistry ($n \geq 5$ per treatment). A β load is calculated as percentage of surface coverage, and was found to be decreased in AD animals fed *Sargassum fusiforme*-enriched chow in cortex ($F(3, 24) = 25.79$, $p < 0.0001$, ANOVA; Tukey's post-hoc for diet effect in AD genotype: $p = 0.0005$) and hippocampus ($F(3, 24) = 32.13$, $p < 0.0001$, ANOVA; Tukey's post-hoc for diet

effect in AD genotype: $p < 0.0001$). Representative IHC staining of all groups is shown (e). (f,g) *Sargassum fusiforme* treated APP^{swe}PS1 Δ E9 mice show a significant decrease in A β_{40} ((f); $U = 6$, $n_{\text{ctrl}} = 11$, $n_{\text{extract}} = 5$, $p = 0.0133$, Mann-Whitney) but not A β_{42} levels (g). (h) *Sargassum fusiforme* treated APP^{swe}PS1 Δ E9 mice show a significant decrease in the mRNA expression of APP ($U = 0$, $n_{\text{ctrl}} = 8$, $n_{\text{extract}} = 4$, $p = 0.0040$, Mann-Whitney).

We show that dietary supplementation with crude or lipid extract of *Sargassum fusiforme* reduces A β load in AD mice. Our *in vitro* data indicate that factors released by 24(S)-Saringosterol-treated astrocytes promote microglial clearance of A β_{1-42} . Within the brain, astrocytes are the major source of the LXR response gene *ApoE*. ApoE is well-known to increase microglial clearance of A β^{59} , possibly via inhibiting cellular uptake of A β^{60} . Of interest, 24(S)-Saringosterol and a lipid extract of *Sargassum fusiforme* increased ApoE protein expression in astrocytes and the CNS respectively, which suggests a role for ApoE in the observed increase in microglial A β clearance. In addition, we provide evidence that 24(S)-Saringosterol decreases the release of A β by neurons *in vitro*. Together, these findings indicate that 24(S)-Saringosterol impacts both the clearance and generation of A β_{42} . These findings are of particular interest, as beneficial effects of LXR agonists on cognition are not necessarily accompanied by a reduced A β plaque load^{14,41}. Recently, it has been reported that other constituents of *Sargassum fusiforme*, such as fucosterol, fucoidan, and fucoxanthin improve learning and memory deficiencies in pharmacological models for cognitive impairment⁶¹⁻⁶³. Although synergism among constituents of *Sargassum fusiforme* is likely, future studies should determine whether 24(S)-Saringosterol is sufficient to improve cognitive performance and AD pathology.

Although relatively high levels of arsenic were measured in extracts of *Sargassum fusiforme*, no arsenic was detectable in lung tissue, which generally represents a major reservoir for this element⁶⁴. Arsenic present in *Sargassum fusiforme* may therefore be rapidly excreted, degraded or absorbed in low amounts in the intestine.

In summary, our data indicate that phytosterols present in a Western type diet do not activate LXRs at concentrations that can be reached through dietary supplementation. However, a crude lipid extract of the edible seaweed *Sargassum fusiforme* did selectively activate LXR β *in vitro*. Here, we show for the first time that dietary supplementation with *Sargassum fusiforme* significantly improves memory performance and reduces A β plaque load in a well-established AD model. The accumulation of the natural LXR β agonist 24(S)-Saringosterol in the CNS suggests its involvement in the neuroprotective impact of *Sargassum fusiforme*. Collectively, our findings point to 24(S)-Saringosterol-containing *Sargassum fusiforme* as being a novel candidate for dietary supplementation to prevent or modulate neurodegenerative disorders such as Alzheimer's disease.

Materials and Methods

Preparation of plant extracts. Indigenous Asian plants (*Asparagus racemosus*, *Azadirachta indica*, *Cassia fistula*, *Curcuma aromatica*, *Datura metel*, *Piper retrofractum*, *Sargassum fusiforme*, *Senna tora*, and *Terminalia chebula*), were selected based on their use in Asian traditional medicine as cognition enhancers. Extracts from all plants, with exception of *Sargassum fusiforme*, were prepared by the maceration method. All plants were dried in a hot air oven at 50 °C. The dried samples were finely powdered and soaked in 95% ethanol overnight at room temperature and filtered with Whatman filter paper No. 3 (Sigma-Aldrich, Bornem, Belgium). Three consecutive filtrates were pooled and evaporated in a vacuum rotary evaporator at 40 °C. The crude lipid fraction of *Sargassum fusiforme* was extracted using an adaptation of the Folch method⁶⁵. Briefly, *Sargassum fusiforme* was harvested by hand in early spring, washed in sea water, boiled for three hours and dried in a hot air oven. Next, dried *Sargassum fusiforme* (Clearspring Ltd, London, UK) was soaked overnight at room temperature in a 2:1 (v/v) chloroform and methanol (both VWR, Leuven, Belgium) mixture. The chloroform/methanol extract was evaporated under a N₂ stream and the remaining lipid fraction was dissolved in ethanol. Using gas chromatography/mass spectrometry, 24(S)-Saringosterol content of dried *Sargassum fusiforme* was determined to be 69.4 ng/mg.

Cell culture. Immortalized human embryonic kidney cells (HEK293.T), human microglia (CHME3; a kind gift from prof. dr. M. Tardieu, Universite Paris-Sud, France⁶⁶), human oligodendrocytes (MO3.13), mouse neuroblastoma expressing APP^{swe} (N2a/APP^{swe}; a kind gift from prof. dr. T.W. Kim, Columbia University, USA⁶⁷), and monkey kidney cells (COS7) were used for *in vitro* experiments. All cell lines were cultured in DMEM (Sigma-Aldrich) containing 10% heat-inactivated FCS (Invitrogen, Merelbeke, Belgium) and 100 U penicillin/100 μ g streptomycin/ml (Invitrogen), at 37 °C/5% CO₂. For phytosterol treatment, cells were incubated for 18 hours in culture medium without FCS containing the Eastern plants extracts, brassicasterol (Sigma-Aldrich), β -sitosterol, (Sigma-Aldrich), fucosterol (Sigma-Aldrich), stigmasterol (analytic confirmed purity of 99.9%), phytosterol mix (containing 60% β -sitosterol, 25% campesterol, and 15% stigmasterol; kindly provided by Ingmar Wester Raisio, Finland), T0901317 (Cayman Chemicals, Huissen, the Netherlands), ethanol (VWR), or DMSO (Sigma-Aldrich).

Luciferase-based nuclear receptor reporter assay. To determine the ability of plant extracts and phytosterols to bind LXR α and LXR β , a luciferase-based reporter assay was performed using the ONE-GloTM Luciferase Assay System kit (Promega, Leiden, the Netherlands), according to manufacturer's instructions. Cell lines were transfected with bacterial plasmid constructs expressing luciferase under control of the promoter region of the ligand-binding domain for LXR α or LXR β ⁶⁸. Cells were grown to 50–60% confluency in 60 mm plates, transfected with 1.8 μ g of plasmid DNA including 0.2 μ g pGAL4hLXR α or pGAL4hLXR β , 1 μ g pG5-TK-GL3, and 0.6 μ g of pCMV- β -galactosidase, using JetPEI (Polyplus-transfection SA, Illkirch, France) as transfection reagent. Following treatment, cells were lysed in lysis buffer (25 mM Glycyl-Glycine, 15 mM MgSO₄, 4 mM EGTA, and 1x Triton; all from Sigma). To correct for transfection efficacy, β -galactosidase activity was measured using lysate diluted 1:10 in B-gal buffer, consisting of 20% 2-Nitrophenyl β -D-galactopyranoside (ONPG; Sigma) and 80%

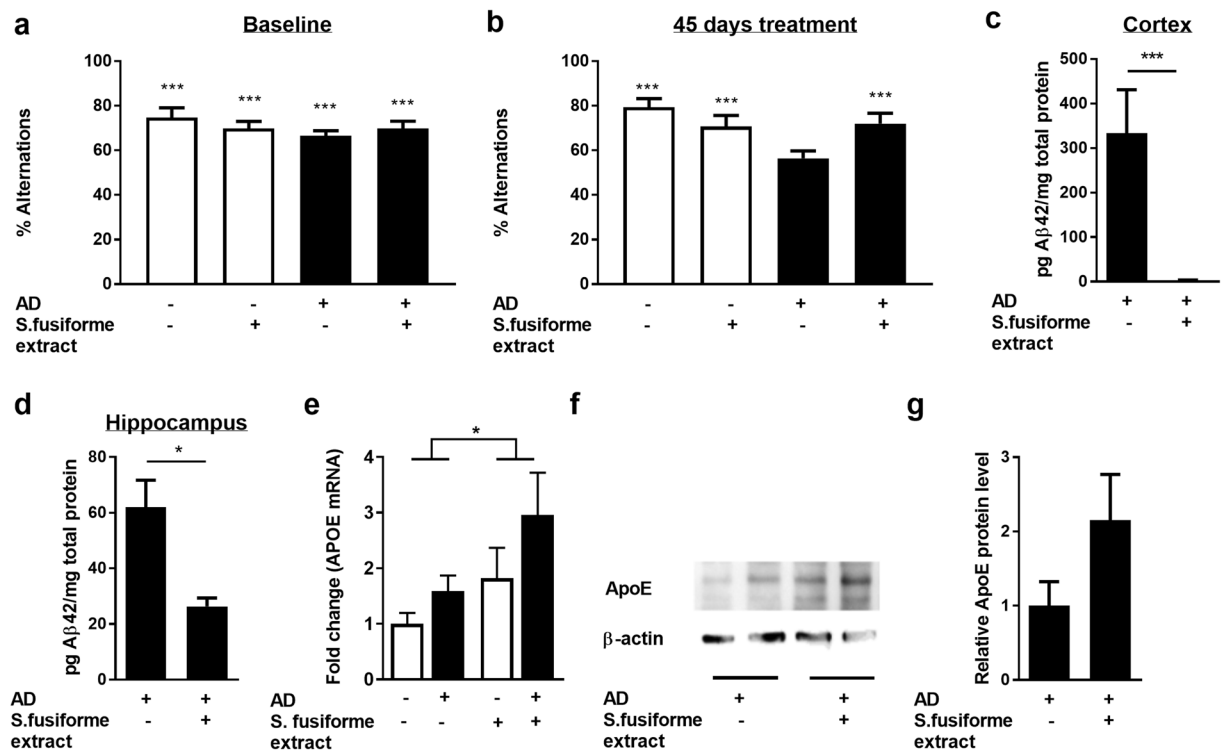


Figure 5. Dietary supplementation with a lipid extract of *Sargassum fusiforme* reduces cognitive decline and A β plaque load in APPswePS1 Δ E9 mice. (a,b) A *Sargassum fusiforme* lipid extract was administered by gavage to determine the impact of *Sargassum fusiforme*-derived lipids on working memory in the spatial alteration Y maze. All mice showed an intact working memory at baseline ((a), spatial alteration > 50% (chance level); WT control $p = 0.0002$; WT *Sargassum fusiforme* $p = 0.0008$; AD control $p < 0.0001$, AD *Sargassum fusiforme* $p = 0.0006$, one-sample t-test). Upon six weeks of daily treatment (b), the untreated AD group displayed an impaired working memory ($p = 0.085$) while wild type mice and *Sargassum fusiforme* extract-treated mice retained their working memory (WT control $p < 0.0001$, WT *Sargassum fusiforme* $p = 0.0018$, AD *Sargassum fusiforme* $p = 0.0008$, one-sample t-test). (c,d) The extract-treated APPswePS1 Δ E9 mice show a significant decrease in formic acid-extracted insoluble A β_{42} levels in the cortex ((c); $U = 0$, $n_{\text{ctrl}} = 8$, $n_{\text{extract}} = 8$, $p = 0.0045$, Mann-Whitney) and in the hippocampus ((d); $U = 3$; $n_{\text{ctrl}} = 7$, $n_{\text{extract}} = 5$, $p = 0.0177$, Mann-Whitney). (e-g) Mice treated with a lipid extract of *Sargassum fusiforme* show an increased mRNA (e) expression of *ApoE* in their CNS ($F(1, 18) = 4.885$, $p = 0.0403$, two-way ANOVA), although the increase in protein expression (f,g) does not reach significance. Data are shown as mean \pm SEM.

(v/v) Buffer Z (0.1 M Na $_2$ HPO $_4$, 10 mM KCl, 1 mM MgSO $_4$, and 3.4 μ l/ml β -mercaptoethanol; all from Sigma). Luminescence and absorbance (410 nM) was measured using the FLUOstar Optima (BMG Labtech, Ortenberg, Germany).

Animals and diet. Male APPswePS1 Δ E9 mice (Radboud University Medical Center, Nijmegen⁴⁷) were backcrossed with female C57Bl6/J mice (Harlan Netherlands B.V., Horst, The Netherlands) to obtain male APPswePS1 Δ E9 and wild-type (WT) littermates. Animals were housed in a conventional animal facility at Hasselt University. Two series of animal experiments were conducted, the first using dietary supplementation of dried *Sargassum fusiforme*, and the second using gavage of a lipid extract of *Sargassum fusiforme*. A timeline of the animal experiments is depicted in Supplemental Fig. 1.

For the first series, experimental diets consisted of either powdered standard chow (Teklad 2018; Harlan Netherlands B.V.) or chow supplemented with 50% (w/w) pulverized dried *Sargassum fusiforme* (Clearspring Ltd, London, UK) or the synthetic LXR α agonist AZ876 (2 mg/kg body weight, kindly provided by AstraZeneca, Mölndal, Sweden⁶⁹). The amount of 24(S)-Saringosterol used was 69.4 ng/mg dried *Sargassum fusiforme*. Based on an estimated intake of 7 g dry food/day, the amount of daily 24(S)-Saringosterol corresponds to 242 μ g/day. Animals were divided into three groups and received either the *Sargassum fusiforme* containing diet (APPswePS1 Δ E9 $n = 13$; C57Bl6/J $n = 10$), AZ876 containing diet (APPswePS1 Δ E9 $n = 14$; C57Bl6/J $n = 13$), or the control diet (APPswePS1 Δ E9 $n = 13$; C57Bl6/J $n = 12$) from 5 until 7.5 months of age.

The second batch of animals was divided into two groups and received either the *Sargassum fusiforme*-derived lipid extract (APPswePS1 Δ E9 $n = 9$; C57Bl6/J $n = 8$), or the vehicle (APPswePS1 Δ E9 $n = 11$; C57Bl6/J $n = 12$) from 6 until 7.5 months of age. Lipids of *Sargassum fusiforme* were extracted using the Folch-method⁶⁵. Briefly, dried *Sargassum fusiforme* was pulverized and soaked in a 2:1 (v/v) chloroform:methanol (VWR, Leuven,

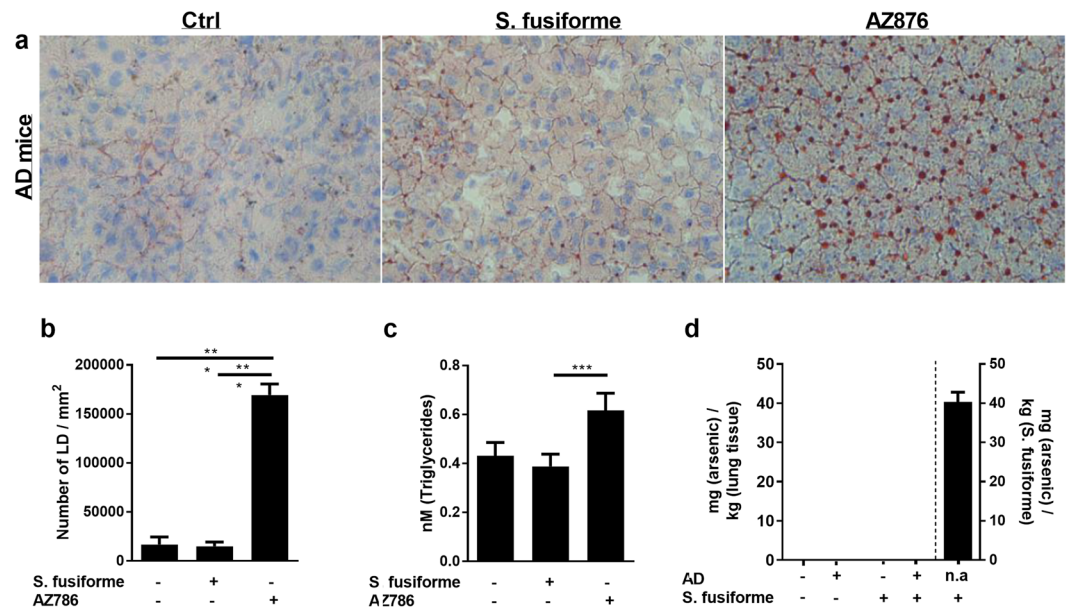


Figure 6. *Sargassum fusiforme* supplementation does not result in liver steatosis, elevated circulating triglyceride levels, or arsenic accumulation in the lungs. (a) An ORO staining was performed to define lipid droplets in liver samples of APPswePS1 Δ E9 mice fed normal chow or *Sargassum fusiforme*/AZ876-enriched chow. (b) Quantification of the ORO staining shown in (1) calculated as number of lipid droplets per mm² of liver tissue. (c) AZ876 but not *Sargassum fusiforme* increases the circulating levels of triglycerides in and APPswePS1 Δ E9 mice (F (2, 31) = 4.056, $p = 0.0272$, ANOVA; Tukey's post-hoc for diet effect: $p = 0.0305$). (d) Arsenic was assessed in dried *Sargassum fusiforme* and lung samples of WT and APPswePS1 Δ E9 mice (AD) fed normal chow or chow supplemented with *Sargassum fusiforme*.

Belgium) mixture for 16 hours at room temperature. Next, the mixture was sonicated (10 min), filtered, and subsequently evaporated by heating the extract until 60 °C. The remaining extract was washed in ethanol, sonicated (10 min), and evaporated. Finally, the extract was dissolved in corn oil (Vita D'or, the Netherlands) as vehicle and sonicated prior to *in vivo* supplementation. Using gas chromatography/mass spectrometry, 24(S)/(R)-Saringosterol content was measured to be 332 mg/dl (7,744 mM). The extract was administered by daily gavage (200 μ l/25 g body weight) for a period of 45 days, corresponding to a daily lipid intake of 3.5 g seaweed/day (664 μ g 24(S)/(R)-Saringosterol/day). In a previous study we defined that *Sargassum fusiforme* contains predominantly 24(S)-Saringosterol³⁴. Hence, we expect that this isoform is the most abundant isoform present in our *in vivo* experiments.

One week prior to the start of the behavioral experiments all animals were housed individually. Animals were kept at an inverse 12 h light/12 h dark cycle, and behavioral experiments were performed during the dark phase of the cycle. Cognitive performance was scored blind. All animal procedures were performed in accordance with institutional guidelines and approved by the ethical committee for animal experiments of Hasselt University.

Behavioral tasks. The object recognition task (ORT) was conducted after 9 weeks of *Sargassum fusiforme* supplementation when the mice were ~8 months old, as described previously^{70,71}. In brief, all animals were habituated to the arena over 2 days, in one trial of 4 min per day. In each habituation trial, two different objects were placed symmetrically in the center of the arena, about 10 cm from the wall. Four different objects were available and at the end of the habituation period each animal had encountered all four objects once. After one resting day, the experiment started. During the first trial (T1) two identical objects (samples) were placed in the arena. After exploring the samples for 4 min, the animal was placed back in its home cage. Subsequently, after a predetermined delay interval (1 or 24 hours), the second trial of 4 minutes (T2) was performed using one familiar object from trial 1 and one new object. Exploration time for each object during T1 and T2 was recorded manually using a personal computer. Sitting on the object or biting in the object was not considered exploratory behavior. Discrimination index d_2 in T2 ($d_2 = [(exploration\ time\ for\ the\ novel\ object) - (exploration\ time\ for\ the\ familiar\ object)] / (total\ exploration\ time\ in\ T2)$) was calculated as measure for object memory.

The Y-maze spontaneous alternation test⁷² was conducted at baseline and after 6 weeks of extract treatment. The maze used for the Y-maze spatial alteration task consisted of three arms of grey Perspex of equal length (40 \times 10 \times 15 cm) (labeled A, B and C), separated at an angle of 120°. The animals were free to explore the maze for 6 minutes. Afterwards, the working memory is calculated in order of percentage of alternations defined as: (number of triads/(total number of entries - 2)) * 100. The number of triads was recorded manually and was defined as subsequently entering the three different arms, while entering an arm was defined as placing both hind paws in that arm. After each trial, the maze was cleaned with 70% ethanol to prevent olfactory cues.

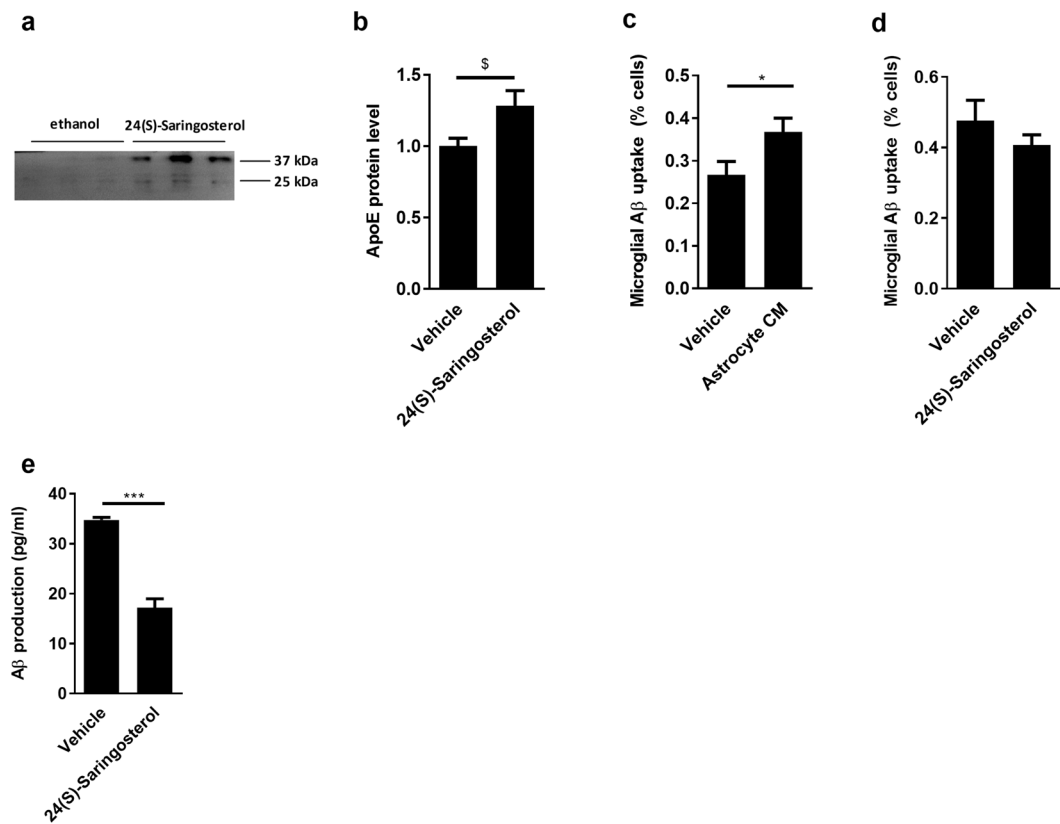


Figure 7. 24(S)-Saringosterol increases astrocytic ApoE production and microglial A β clearance and reduces neuronal A β release *in vitro*. **(a,b)** Primary astrocytes were treated with 10 μ M 24(S)-Saringosterol for 2 consecutive days and medium was collected 48 h after the last treatment. Western Blot was used to measure secreted ApoE in the cellular supernatant. 24(S)-Saringosterol increased ApoE secretion ($t(5) = 2.531$, $p = 0.0524$, unpaired t-test). Representative WB bands are displayed in the figure. **(c)** Primary murine microglia were examined for their capacity to internalize FAM-labeled A β_{1-42} . Microglia were incubated for 1 h with fluorescently-labeled FAM-A β_{1-42} and 40% conditioned medium derived from astrocytes treated with vehicle (ethanol) or 24(S)-Saringosterol (10 μ M). Data are depicted as the percentage of cells that phagocytosed FAM-A β_{1-42} . 24(S)-Saringosterol increased microglial uptake of FAM-A β_{1-42} ($t(14) = 2.218$, $p = 0.0436$, unpaired t-test). **(d)** To define a direct effect on microglial uptake of A β_{1-42} , primary murine microglia were exposed to 24(S)-Saringosterol prior to defining FAM-labeled A β_{1-42} uptake. **(e)** N2aAPPsw cells were stimulated with vehicle (ethanol) or 24(S)-Saringosterol (10 μ M). ELISA was used to define the release of A β_{42} . 24(S)-Saringosterol decreased A β_{42} production ($t(5) = 8.126$, $p = 0.0005$, unpaired t-test).

The open field test (OF) was conducted as described previously⁷³, after 9.5 weeks of *Sargassum fusiforme* treatment. In short, animals were placed in a square, Perspex box (50 cm \times 50 cm \times 30 cm), with an open top, grey walls, and a white floor. Video tracking with a computerized system (Etho VisionTM, Noldus, Wageningen, The Netherlands) was used to record and analyse movements and position of the animals, in order to determine time in zone (TIZ) and distance moved (DM) over the 10-minute trial¹⁴.

Tissue sample preparation. Mice were anaesthetized with Nembutal (100 mg/kg i.p., CEVA Logistics, Brussels, Belgium) prior to transcardial perfusion with Ringer's solution. Blood was collected and centrifuged for 10 min at 200 g to obtain serum, which was snap-frozen in liquid nitrogen. Liver samples were divided into two parts and either directly snap-frozen or frozen in Tissue-Tek O.C.T embedding compound (Sakura Finetek, Berchem, Belgium). Brains were divided into three parts prior to snap-freezing; the cerebellum and two hemispheres. The right hemisphere was preserved for immunohistochemistry and frozen in Tissue-Tek O.C.T. embedding compound. The rostral half of the left hemisphere was preserved for mRNA expression analyses with quantitative PCR. The cerebellum was preserved for sterol analyses.

A β quantification. The right hemisphere of the crude *Sargassum fusiforme*-treated mice was cut in the coronal plane with a Leica CM1900UV cryostat (Leica Microsystems, Wetzlar, Germany) to obtain 10 μ m sections. Sections were mounted on glass slides, air-dried overnight, and stored at room temperature until used. After blocking of endogenous peroxidases by incubation for 30 min with 3% H₂O₂ in methanol, the sections were incubated for 30 min with blocking solution (5% bovine serum albumin in 1x TBS) followed by overnight 4 $^{\circ}$ C primary antibody incubation in blocking solution (clone 3D6, Amyloid- β Antibody 1:8000). The sections were then incubated in

Gene symbol	Gene name	Forward and reverse primer
<i>Cyca</i>	Cyclin-A	F: TGGGATTGTACCACAGCTCCA R: CTCATGATGACTGCAGCAAACC
<i>Hmbs</i>	Hydroxymethylbilane synthase	F: GATGAAGCCATTGCTGAACCTTG R: GTCTCCTTGGGTATCCGATGTC
<i>Abca1</i>	ATP-binding cassette, sub-family A, member 1	F: CCCAGAGCAAAAAGCGACTC R: GGTCAATCATCACTTTGGTCCTTG
<i>Abcg1</i>	ATP-binding cassette, sub-family G, member 1	F: CAAGACCCTTTTGAAGGGATCT R: GCCAGAATATTCATGAGTGTGGAC
<i>ApoE</i>	Apolipoprotein E	F: CCTGAACCGCTTCTGGGATF R: GCTCTTCTGGACCTGGTCA
<i>App</i>	Amyloid precursor protein	F: CGA ACC CTA CGA AGA AGC CAC R: GCT TTC TGG AAA TGG GCA TGT TC
<i>Axl</i>	Axl receptor tyrosine kinase	F: GGA ACC CAG GGA ATA TCA CAG G R: AGT TCT AGG ATC TGT CCA TCT CG
<i>Mertk</i>	Proto-oncogene tyrosine-protein kinase Mer	F: TGC GTT TAA TCA CAC CAT TGG A R: TGC CCC GAG CAA TTC CTT TC
<i>Scd1</i>	Stearoyl-CoA desaturase 1	F: TGCGATACACTCTGGTGCTCA R: CTCAGAAGCCCAAAGCTCAGC
<i>Srebp-1c</i>	Sterol regulatory element binding protein 1c	F: GGAGCCATGGATTGCACATT R: GCTTCCAGAGAGGAGCCAG
<i>Trem2</i>	Triggering receptor expressed on myeloid cells 2	F: CTG GAA CCG TCA CCA TCA CTC R: CGA AAC TCG ATG ACT CCT CGG

Table 2. Quantitative PCR primers. Nucleotide sequence of primers used for quantitative PCR. F denotes forward primer, R denotes reverse primer.

biotinylated goat anti-human IgG (BA-3000, 1:1000, Vector Laboratories, Burlingame, CA), and the immunoreactivity was developed by the ABC avidin–biotin peroxidase KIT (PK-6100, Vector) for 1 hour, and 5 min with diaminobenzidine (DAB, ImmPACT SK-4105, Vector). Between every other step, the slides were rinsed (3×) with TBS/0.3% Triton-X 100. The sections were counterstained with haematoxylin for 30 seconds and coverslipped. Digital images of the sections were obtained using a Leica DM 2000 LED microscope (Leica Microsystems, Diegem, Belgium) equipped with hardware and software from Leica Application suite software (Leica Microsystems). The A β plaque load was quantified in 3 sections per brain in 5 animals per group using Fiji ImageJ, by defining the pixel intensity of A β plaques in the total cortical or hippocampal area at Bregma –1.5 to –2.5.

Next, left hippocampus and cortex were dissected from the brains of the *Sargassum fusiforme* extract-treated APPswPS1 Δ E9 mice and homogenated in a 2% SDS TBS-T buffer. A β_{42} levels were quantified and related to the protein content in the starting homogenate using an A β_{42} ELISA (Invitrogen, USA), according to the manufacturer's instructions⁷⁴. In brief, the homogenates were sonicated twice and mixture centrifuged to generate an SDS-insoluble fraction (21000 g, 10 min). The SDS-insoluble pellet was sonicated twice in 70% formic acid (FA), yielding the FA-extracted insoluble A β_{42} fraction.

Mouse neuroblastoma (N2a) cells, stably overexpressing APP, were cultured in high glucose DMEM medium supplemented with 10%FCS and 100 U pen/100 μ g strep/ml (37 °C, 5% CO₂) Next, the cells were incubated for 24 h with ethanol (vehicle control) or 10 μ M 24(S)-Saringosterol, purified from *Sargassum fusiforme* as described earlier³⁴. A β_{42} levels were detected in supernatant using an A β_{42} ELISA (Invitrogen, USA), according to the manufacturer's instructions.

Determination of astrocytic ApoE secretion. To generate 24(S)-Saringosterol-astrocyte conditioned medium, mouse primary astrocytes were seeded in a 24-well plate with a density of 250000 cells/well. Next, astrocytes were treated with either 10 μ M purified 24(S)-Saringosterol or ethanol for 2 consecutive days after which medium was replenished and collected 2 days afterwards. Next, ApoE secretion was determined using western blot. Equal amounts of cellular supernatant (40 μ l) were loaded and separated via electrophoresis through a 10% sodium dodecyl sulfate polyacrylamide gel (SDS-PAGE). Afterwards, proteins were transferred onto a PVDF membrane (VWR, Belgium), and blots were blocked (5% non-fat dry milk TBS-Tween solution) for 1 h at room temperature. Next, membranes were incubated with rabbit anti-ApoE primary antibody (ABBIOTEC, USA). After washing the membranes with TBS-Tween, the blots were incubated with swine anti-rabbit HRP conjugated antibody (WAKO, Japan). Finally, membranes were washed in TBS and protein bands were detected using Pierce™ ECL Plus Western Blotting Substrated and ImageQuant™ LAS 4000 mini. ImageJ (<http://imagej.nih.gov/ij/>) was used to quantify the protein bands.

A β phagocytosis assay. Primary murine cells were isolated and cultured as described previously⁷⁵. In short, mixed glial cultures were prepared from postnatal d0 mouse cerebral cortices of C57Bl6/J mice (Harlan Netherlands B.V.) and cultured in high glucose DMEM medium supplemented with 10% FCS and 100 U pen/100 μ g strep/ml. Mixed glial cultures were used to generate microglia-enriched glial cultures by separating the microglia from the astrocyte monolayer using orbital shaking followed by purification via differential adhesion to plastic after 14 days in culture. Purified microglia were seeded on poly-L-lysine (5 μ g/ml; Sigma-Aldrich) coated 96-well plates with a density of 50000 cells/well. Primary microglia were cultured in high glucose DMEM medium supplemented with 10% FCS, 100 U pen/100 μ g strep/ml and 15% L929 conditioned medium.

The capacity of primary microglia to phagocytose FAM-labeled A β_{1-42} (Eurogentec, Seraing Belgium) was assessed with a plate-based assay. Twenty hours after seeding the microglia, cells were treated with 24(S)-Saringosterol-astrocyte conditioned medium (40%) and FAM-labeled A β_{1-42} was added for 1 h at a final concentration of 500 nM. Next, medium was removed and 100 μ l 0.2% trypan blue in PBS (pH 4.4) was added to quench extracellular A β . After aspiration, fluorescence was measured at 485/535 nm excitation/emission. Finally, to normalize for cell number, 100 μ l DAPI in PBS was added, incubated for 10 min and medium was aspirated. Fluorescence was subsequently measured in a plate reader using 360 nm/465 nm excitation/emission wavelengths.

Sterol profile and triglyceride content determination. Sterol profiles were determined in serum and cerebellum. Preceding the analysis, samples were spun in a speed vacuum dryer (Savant AES 1000) at 12 mbar for 24 h to relate individual sterol concentrations to their dry weight. The sterols were extracted from the dried tissue by placing them in a mixture of chloroform:methanol (2:1) for 24 h at 4 °C. Sterol levels were determined by gas chromatography/mass spectrometry (GC/MS) as described previously^{32,76}. Triglyceride contents of serum were determined using the GOD-PAP method with enzymatic reagent kits according to manufacturer's instructions (DiaSys Diagnostic Systems, Holzheim, Germany).

Determination of arsenic concentration. Arsenic halogenide concentrations in lung tissues were analyzed using inductively coupled plasma-optical emission spectrometry (ICP-OES, Agilent Technologies, 700 Series, Belgium). Tissues were oven-dried and digested with 70–71% HNO₃ in a heat block. Prior to analysis, lung samples and dried *Sargassum fusiforme* were diluted 1:10 in 1% nitric acid. Concentrations were measured using a standard calibration curve (NIST Spinach 1570a).

RNA isolation and RT-Q-PCR. Brain tissue was homogenized in Qiazol (Qiagen, Venlo, The Netherlands) preceding RNA isolation, and total RNA was prepared using the RNeasy mini kit (Qiagen), according to manufacturer's instructions. RNA concentration and quality was determined with a NanoDrop spectrophotometer (Isogen Life Science, IJsselstein, The Netherlands). RNA was reverse transcribed to cDNA using the qScript™ cDNA synthesis kit (Quanta Biosciences, Gaithersburg, USA). As previously described^{77,78}, quantitative PCR was subsequently conducted on a StepOnePlus detection system (Applied biosystems, Gaasbeek, Belgium). Relative quantification of gene expression was performed using the comparative Ct method. Data were normalized to the most stable reference genes (Cyca and Hmbs). Primers were chosen according to literature or designed using Primer-Express (<http://www.ncbi.nlm.nih.gov/tools/primer-blast>), and details of primers used are shown in Table 2.

Experimental design and statistical analyses. All statistical analyses were performed using GraphPad Prism 7™ and are reported as mean \pm SEM. D'Agostino and Pearson omnibus normality test was used to test normal distribution. Unless specified differently, one-way ANOVA (post-hoc: Tukey) or two-tailed unpaired Student t-test was used for normally distributed data sets. The Kruskal-Wallis (Dunns post hoc comparison) or Mann-Whitney analysis was used for data sets assessed not to be normally distributed. The ORT discrimination index d2 (compared to the 0) and spatial alteration Y-maze performance (compared to the 50% chance level) was analyzed using the one-sample t-test. Animals that did not reach the minimum of 4 s exploration in T1 or T2 were excluded from further analysis^{71,79}. Data from RT-Q-PCR was analyzed using two-way ANOVA with treatment and genotype as independent variables. In all data sets, extreme values were excluded by means of Dixon's principles of exclusion of extreme values^{80,81}. Significance levels are denoted as follows: \$p < 0.10 *p < 0.05, **p < 0.01, ***p < 0.001 or ****p < 0.0001.

Ethical approval. All animal procedures were performed in accordance with institutional guidelines and approved by the ethical committee for animal experiments of Hasselt University.

Data Availability

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

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Author Contributions

J.B., C.H., M.S., A.T., A.C., F.L., D.S., A.K., P.M.M., J.W.J., I.D., S.P., D.L. and T.V. performed the experiments and analyzed the data. J.B., C.H., T.V. and M.M. wrote the manuscript. Y.P., T.S., H.B.L. and B.S. provided experimental materials. All authors revised the manuscript. J.B., C.H., N.H., J.W., P.M.M., E.S., J.W., J.H., A.G., B.S., D.L., T.V. and M.M. participated in the design and coordination of the project.

Additional Information

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