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Published in:
Heredity

DOI:
[10.1038/hdy.2014.43](https://doi.org/10.1038/hdy.2014.43)

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Document Version
Publisher's PDF, also known as Version of record

Publication date:
2014

[Link to publication in University of Groningen/UMCG research database](#)

Citation for published version (APA):

Ma, W-J., Pannebakker, B. A., Beukeboom, L. W., Schwander, T., & van de Zande, L. (2014). Genetics of decayed sexual traits in a parasitoid wasp with endosymbiont-induced asexuality. *Heredity*, 113(5), 424-431. <https://doi.org/10.1038/hdy.2014.43>

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ORIGINAL ARTICLE

Genetics of decayed sexual traits in a parasitoid wasp with endosymbiont-induced asexuality

W-J Ma¹, BA Pannebakker², LW Beukeboom¹, T Schwander^{1,3,4} and L van de Zande^{1,4}

Trait decay may occur when selective pressures shift, owing to changes in environment or life style, rendering formerly adaptive traits non-functional or even maladaptive. It remains largely unknown if such decay would stem from multiple mutations with small effects or rather involve few loci with major phenotypic effects. Here, we investigate the decay of female sexual traits, and the genetic causes thereof, in a transition from haplodiploid sexual reproduction to endosymbiont-induced asexual reproduction in the parasitoid wasp *Asobara japonica*. We take advantage of the fact that asexual females cured of their endosymbionts produce sons instead of daughters, and that these sons can be crossed with sexual females. By combining behavioral experiments with crosses designed to introgress alleles from the asexual into the sexual genome, we found that sexual attractiveness, mating, egg fertilization and plastic adjustment of offspring sex ratio (in response to variation in local mate competition) are decayed in asexual *A. japonica* females. Furthermore, introgression experiments revealed that the propensity for cured asexual females to produce only sons (because of decayed sexual attractiveness, mating behavior and/or egg fertilization) is likely caused by recessive genetic effects at a single locus. Recessive effects were also found to cause decay of plastic sex-ratio adjustment under variable levels of local mate competition. Our results suggest that few recessive mutations drive decay of female sexual traits, at least in asexual species deriving from haplodiploid sexual ancestors. *Heredity* (2014) **113**, 424–431; doi:10.1038/hdy.2014.43; published online 30 April 2014

INTRODUCTION

Owing to environmental or life style changes and associated shifts in selective pressures, formerly adaptive traits may become non-functional or even maladaptive, and as a consequence they might decay (Fong *et al.*, 1995; Wiens, 2001; Ellers *et al.*, 2012). Trait decay has been observed for morphological, behavioral and physiological features. Examples include reduced wings in flightless birds (McNab, 1994), loss of eye function, decay of pigmentation in cave-dwelling animals (Jeffery, 2009; Protas *et al.*, 2011) and loss of lipid synthesis pathways in parasitoid wasps (Visser *et al.*, 2010). Such trait decay, especially when occurring in parallel in independent lineages, highlights the importance of natural selection for the maintenance of adaptations (Fong *et al.*, 1995; Wiens, 2001; Lahti *et al.*, 2009). Despite the fact that trait decay is a common evolutionary phenomenon, its underlying genetic mechanisms are poorly understood. It is largely unknown if trait decay typically stems from multiple mutations with small effects, or rather has a simple genetic architecture involving few loci with major phenotypic effects (Jeffery, 2009; Lahti *et al.*, 2009).

In particular, many traits are expected to decay following a transition from sexual to asexual reproduction, making this transition of special interest for studies of trait decay and the genetic causes thereof (for example, Carson *et al.*, 1982; Pannebakker *et al.*, 2004a; Jeong and Stouthamer, 2005; Kraaijeveld *et al.*, 2009; Russell and Stouthamer, 2011; Schwander *et al.*, 2013). For example, any trait

specific to the male sex is useless under asexual reproduction in all-female species. The same holds for sexual traits expressed in females, such as those involved in attracting mates, mating behavior and egg fertilization.

Here, we investigate the decay of female sexual traits and its genetic basis in the asexual wasp *Asobara japonica*. *A. japonica* is a parasitoid wasp that uses *Drosophila* larvae as host. It consists of both sexual and all-female asexual strains (Murata *et al.*, 2009). Genetic analyses can be difficult in asexual organisms because of the inability to perform crosses. An exception to this constraint applies to those species in which asexuality is induced by infection with bacterial endosymbionts, such as *A. japonica* (Kremer *et al.*, 2009; Reumer *et al.*, 2012). Endosymbiont-induced asexuality is mainly found among wasps and among other groups in which sexual species are characterized by haplodiploid sex determination (Werren, 1997; Werren *et al.*, 2008; Mateo Leach *et al.*, 2009; Giorgini *et al.*, 2010; Kageyama *et al.*, 2012; Ma *et al.*, 2014), although it has also been suggested to occur in species with other sex determination systems (for example, Pike and Kingcombe, 2009). Under haplodiploidy, females develop from fertilized, diploid eggs, whereas males develop from unfertilized, haploid eggs (Whiting, 1933). However, unfertilized haploid eggs laid by endosymbiont-infected females undergo diploidization in the absence of fertilization with sperm (Suomalainen *et al.*, 1987; Stouthamer *et al.*, 1990; Werren, 1997; Werren and O'Neil, 1997; Gottlieb and Zchori-Fein 2001; Pannebakker *et al.*, 2004b). In species

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Received 26 November 2013; revised 23 March 2014; accepted 27 March 2014; published online 30 April 2014

with endosymbiont-induced asexuality, the genetics of traits involved in sexual reproduction can be studied because infected asexual females can often be cured of their endosymbionts via treatment with antibiotics. Such cured asexual females produce males, and these males can be crossed with females from related sexual strains (Jeong and Stouthamer, 2005; Pannebakker *et al.*, 2005; Russell and Stouthamer, 2011). In addition, asexual females occasionally produce males under natural conditions and in the absence of antibiotic treatments. This is possibly caused by incomplete endosymbiont transmission or by the occasional inability of endosymbionts to manipulate host reproduction (Heath *et al.*, 1999; Reumer *et al.*, 2012). Although these males usually do not have any mating opportunities or success in the asexual populations, they can be mated with females of sexual strains, similar to the males induced by antibiotic treatment.

We investigated four female sexual traits for signs of decay in asexual *A. japonica* and studied the genetic architecture of decayed traits via introgression of alleles from the asexual strain into the sexual one. As the expressed level of decay should depend on the degree of introgression of the asexual genome, we can make inferences about the genetic architecture underlying sexual trait decay, using simple genetic models to estimate the number of loci involved. Specifically, we investigated attractiveness to males, mating behavior and egg fertilization in two different contexts specific to sexual reproduction in haplodiploid parasitoids. We tested which fraction of females with different levels of sexual–asexual admixture would still fertilize eggs, and to what extent these females would adjust the proportion of fertilized eggs (that is, the proportion of females among their offspring) to different conditions. Females of many parasitoid species adjust the sex ratio of their offspring adaptively when mating occurs among the offspring of a few females in isolated patches (local mate competition (LMC); Hamilton, 1967). If only a single female is present in a patch, her sons will compete with each other to mate with their sisters. Hence, it is in the female's interest to produce the minimum number of sons required to fertilize all her daughters and allocate more energy into the production of daughters (Hamilton, 1967). When several females are present in a patch, sons from different females will compete, favoring a larger investment in sons. Sexual females are therefore predicted to produce a smaller proportion of sons when ovipositing alone than when other females are present in the same patch (Hamilton, 1967; Werren, 1980), a pattern we also found in a sexual *A. japonica* strain in a pilot experiment. This plasticity in sex-ratio adjustment makes it an interesting sexual trait to consider in the context of decaying sexual functions under asexual reproduction in parasitoid wasps, as there is no selection for the maintenance of plasticity under asexuality. Our results point to a surprisingly simple genetic architecture underlying the decay of female sexual traits, which most likely facilitated the spread of reduced sexual traits in the asexual population.

MATERIALS AND METHODS

Wasp culturing and collection of individuals

A sexual and a *Wolbachia*-induced, asexual strain of *A. japonica* were used in this study. Both strains were collected from Japan and have been cultured in the laboratory since 2009; the sexual strain originated from the island of Amami-oshima and the asexual strain from Kagoshima on the mainland of Japan. These two strains are closely related (Murata *et al.*, 2009; Reumer *et al.*, 2012), which minimizes the probability of genetic incompatibility in crosses. *A. japonica* was cultured on second-instar *Drosophila melanogaster* larvae as hosts at 25 °C, with a 16L:8D light–dark cycle and 60% relative humidity (for details see Ma *et al.*, 2013).

We used five different classes of virgin individuals: sexual females, sexual males, asexual females cured of their *Wolbachia* infection via antibiotic treatment, untreated asexual females and males produced by asexual females. The males produced by asexual females were either induced via antibiotic treatment or directly collected from our mass culture with the highest incidence of accidental males (we collected 40–70 males among ~6000 individuals). After using these males in our experiments, we verified, via flow cytometry, that they were haploid (as expected under normal haplodiploid sex determination in Hymenoptera; for details, see Ma *et al.*, 2013). Virgin sexual males and females were also collected directly from standard laboratory cultures. Sexual males were collected first because they emerge one or two days earlier than females. After the emergence of males, sexual females were collected by individually isolating wasp pupae in plastic vials (diameter 2.4 cm, height 7.5 cm) containing a layer of agar to control humidity (Ma *et al.*, 2013).

Wolbachia-infected asexual females were cured of their bacteria with antibiotics applied to the *Drosophila* host larvae. In total, 10 mg of rifampicin was added to 1 g yeast powder, which was mixed with water to feed the second-instar *Drosophila* larvae. Rifampicin treatment has been shown to have little impact on the development of *Asobara* wasps (Dedeine *et al.*, 2001), and no effects on life-history traits, such as brood size and pupal mortality, were found in our experiments (data not shown). Female wasps that emerged from rifampicin-fed *Drosophila* hosts were individually collected in plastic vials containing a layer of agar. To confirm complete removal of *Wolbachia* by antibiotic treatment, 23 emerged females were allowed to lay eggs in antibiotic-free host larvae, and the production of only male offspring was verified.

Observation of courtship behavior and spermatheca dissections

To investigate sexual attractiveness and mating behavior of females, we set up 109 no-choice mating trials. For each trial, asexual females ($n = 20$ cured, 40 untreated) or sexual females ($n = 49$) were individually paired with a sexual male for 20 min. We evaluated female attractiveness to sexual males by scoring if the male attempted to court the females (that is, displayed wing vibration and actively approached the female). For female mating behavior, we scored if females responded to the copulation attempts of sexual males with escape behavior or acceptance. A successful copulation was scored when it lasted at least 7 s. As these experiments revealed that cured asexual females were relatively unattractive to sexual males (see results), we evaluated if decreased attractiveness could stem from lineage divergence that is unrelated to the asexual mode of reproduction (that is, as could be observed between two sexual species). To this end, we repeated similar tests with males produced by asexual females, as there is no lineage divergence between these males and asexual females.

To investigate if cured asexual females fertilize their eggs and produce daughters when mated with sexual males, two experiments were performed. First, we paired each of 20 cured asexual females with a sexual male for 24 h. Second, to increase the chance of obtaining any mated asexual females, we paired each of 13 additional cured asexual females with a group of at least 50 sexual males in one mating vial for 24 h. Females were then offered 50–100 second-instar host larvae for egg laying for ~24 h (both experiments), to test if they produce any daughters, indicative of successful egg fertilization. Next, the spermathecae of all asexual females were dissected to check for the presence of sperm, as an indication of successful mating and sperm transfer.

For spermatheca dissections, an individual female wasp was placed in a drop of *Drosophila* Ringer's solution (Rajaram *et al.*, 2005) on a microscope slide. The wasp abdomen was first separated from the rest of the body using a very fine needle. The spermatheca was then carefully separated from the rest of the abdomen. The spermatheca was isolated under a Zeiss Stemi SV 6 stereo microscope (with 25 × 2.4 magnification, Carl Zeiss AG, Oberkochen, Germany), and a cover slide was gently put on top of it. Motile sperm were scored as present or absent under a Zeiss Axio Lab.A1 binocular microscope (with 10 × 40 magnification, Carl Zeiss GmbH, Göttingen, Germany).

Introgression experiment

To determine the genetic architecture of decayed female sexual traits, we introgressed alleles from the asexual into the sexual strain. Upon emergence of virgin sexual females, they were individually paired with a male from the

asexual strain for 24 h. Each female was then offered ~100 second-instar *D. melanogaster* larvae for oviposition during 36 h. After 12–14 days, wasp pupae were isolated from parasitized hosts to prevent matings between individuals upon emergence. Females emerging from these pupae (the F1 hybrid generation) were collected and individually paired with a male from the asexual strain to produce the offspring for the next generation (for further details, see Ma *et al.*, 2013). This experimental procedure was continued for successive generations of introgressions. Because almost no daughters were produced after the 4th generation ('G4') of introgression (because females with high proportions of their genome stemming from the asexual strain do not produce daughters, see results), the introgression experiment was stopped after the 4th generation of introgression. We repeated the same introgression experiment three times independently over the course of 3 months, because only a few asexually produced males were available on specific days for crosses. Because there was no difference for brood size or offspring sex ratio or any other parameter between different experiments (analyses not shown), all data were pooled for further analysis. Given our crossing design, the proportion of asexual alleles in each generation increased from 50% in F1 hybrid females to 75% in females of G2, 87.5% in G3 and a final 93.8% in females of G4. For each generation of introgression, the emerging wasps were anaesthetized with CO₂, counted and sexed (Ma *et al.*, 2013).

The combination of decayed sexual attractiveness, mating behavior and egg-fertilization results in cured asexual females producing only sons. Therefore, we monitored the proportion of admixed females producing only sons across successive generations of introgression. For each generation of introgression, a subset of hybrid females (those not used to continue the introgression crosses) were paired with a sexual male for 24 h followed by oviposition for 36 h. The sex ratio of offspring produced during these 36 h was then determined for each female to infer the proportion of females producing only sons. Using the same experimental conditions, we also determined the sex ratio produced by sexual females and cured asexual females.

Local mate competition experiment

To investigate if *A. japonica* females produce different offspring sex ratios (proportion of male offspring) in response to different situations of LMC, we investigated offspring sex ratios produced by females that oviposited alone or in groups of three females per patch. LMC theory predicts that the proportion of sons increases with increasing numbers of females in a patch. A pilot experiment (data not shown) showed that sexual females do indeed change the sex ratio among their offspring depending on the number of females (one or three) per patch. Thus we tested if sexual females, cured asexual females and females from different introgression generations would produce different offspring sex ratios when alone in a vial relative to when in a group of three. To avoid any interactions between females in different vials (Shuker *et al.*, 2007), the vials were spaced 5 cm apart. As not all hybrid females mate, which would constrain their sex allocation, females were observed while paired with a sexual male and only those females that copulated were used to test for sex-ratio adjustments. Prior to this experiment, each female was offered ~50 host

larvae to get oviposition experience in order to minimize super-parasitism (oviposition in already-parasitized hosts; van Alphen and Nell, 1982; Ma *et al.*, 2013). To control for effects of host larva density on wasp offspring sex ratio, we adjusted the number of the host larvae to the number of females, that is, 50 larvae were offered to a single and 150 to a group of three females for ~15 h. Fifty larvae per female are an excess as a single female can handle a maximum of 30–40 larvae in 15 h (W-J Ma, personal observation). Offspring were counted and sexed upon their emergence from the host pupae.

Statistical analyses

Fisher's exact tests were used to compare the proportion of courting attempts by males, rejection rates of males and the frequency of successful copulations of sexual and cured asexual females. Offspring sex ratios of females from different generations of introgression were compared with logistic regressions specified in generalized linear models (glm). The proportions of females producing only sons between different introgression generations were tested using glm models with a logit link function, and a quasi-binomial error structure to correct for over-dispersion (Crawley, 2007). The proportion of asexual genome in hybrid females was used as a quantitative explanatory variable and the progeny type (either only sons or at least one daughter) as the response. To pinpoint significant differences between introgression generations, a sequential multiple comparison was used. To conduct the multiple comparison, generation had to be used as a categorical variable. For comparison of offspring sex ratios of females producing at least one daughter among different generations of introgression, a glm model was used with a logit link function and a quasi-binomial error structure to correct for over-dispersion (Crawley, 2007). In this model, the proportion of males was used as the response variable (weighted by brood size via the logit link function), and generation as the explanatory variable. A similar glm model was used for comparing the proportion of male offspring in the LMC experiments, that is, between single and triple ovipositing females. All statistical analyses were performed with R 2.13.0 (R Development Core Team, 2011), multiple comparisons of traits among generations were done using the Tukey test as implemented in the R package multcomp for general linear hypotheses (Hothorn *et al.*, 2008).

RESULTS

Sexual traits in asexual females

First, we evaluated whether sexual traits were functional in *Wolbachia*-cured asexual *A. japonica* females. Specifically, we tested whether cured asexual females were able to attract sexual males, mate, and store sperm in their spermathecae, and, if so, whether they fertilize eggs and display plastic sex-allocation behavior under different levels of LMC.

No-choice mating trials (20 min) revealed that cured asexual females were less attractive to sexual males than were sexual females. Sexual males courted a significantly smaller percentage of asexual (55%; 11 out of 20) than sexual females (88%; 43 out of 49; Fisher's

Table 1 Summary of sample sizes and results for the different sexual traits evaluated in asexual *A. japonica* females

Sexual trait evaluated	Female sexual attractiveness		Female copulation behaviour		Female mating success		Spermatheca status (with or without sperm) and proportion of females producing only male offspring		
	Repl.	Court (mean ± s.e.)	Repl.	Accept (mean ± s.e.)	Repl.	Mate (mean ± s.e.)	Repl.	No. of females with sperm	Prop. male offspring
Untreated asexual female	40	0.08 ± 0.04	3	0	40	0	—	—	—
Cured asexual female	20	0.55 ± 0.11	11	0	20	0	20 ^a	0	1
							13 ^b	3	1
Sexual female	49	0.88 ± 0.13	43	0.77 ± 0.07	49	0.67 ± 0.07	—	—	—

Abbreviations: Accept, proportion of females accepting to copulate; Court, proportion of females courted by sexual males; Mate, proportion successfully mated females; Prop., proportion; Repl., number of replicates.

^aTwenty cured females were individually paired with a single sexual male for 24 h.

^bThirteen cured females were individually paired with a group of more than 50 sexual males for 24 h.

exact test, $P=0.008$, Table 1). In addition, among the females that were courted by sexual males, cured asexual females rejected all mounting and copulation attempts of males (100%, $n=11$), whereas rejections were significantly less frequent for sexual females (23%; 10 out of 43, Fisher's exact test, $P<0.0001$, Table 1). As a consequence, not a single 1 out of 20 cured asexual females successfully copulated with a sexual male during the 20 min observation period, as compared with over 67% (33 out of 49) sexual females (Fisher's exact test, $P=0.0003$, Table 1). The reduced attractiveness and copulation propensity of asexual females as compared with sexual females are unlikely to be caused by the treatment with antibiotics, as untreated asexual females were also unattractive to sexual males; only 3 out of 40 untreated asexual females (8%) were courted, compared with 43 out of 49 sexual females (88%; Fisher's exact test, $P=0.0001$, Table 1). The comparison of cured and untreated asexual females further revealed that sexual males are more inclined to court the former (Fisher's exact test, $P=0.0001$). The reduced attractiveness of asexual females to sexual males is unlikely to stem from divergence of functional sexual signals between sexual and asexual strains (as could be expected for comparisons between two diverged sexual strains). This is revealed by the fact that males produced by asexual females also courted sexual females more often (79%; 50 out of 63) than cured asexual females (52%; 11 out of 21; Fisher's exact test, $P=0.024$).

To investigate if cured asexual females were able to fertilize their eggs and produce daughters, we conducted two experiments to increase the chances of finding mated asexual females, given the above-described 20 min no-choice trials did not result in any copulations. In the first experiment, 20 cured asexual females were each paired with a sexual male for 24 h, but all 11 females that produced offspring had only sons (40–60 offspring per female, over 400 offspring in total). The remaining nine females produced no offspring at all (for unknown reasons, subsequent dissections revealed a normal egg load and apparently developed ovaries in all females). In the second experiment, we paired each of 13 cured asexual females with groups of at least 50 sexual males for 24 h. Again, nine females produced only sons (40–60 offspring per female, over 300 offspring in total), even though dissecting them after the experiment revealed that three of these contained sperm in their spermathecae. The remaining four females, of which one contained sperm in her spermatheca, did not produce any offspring (despite having active ovaries). Thus, even in the rare cases where asexual females do mate and store sperm, sperm is apparently not used to fertilize eggs.

Patterns of sexual trait change across different levels of introgression

The combination of low attractiveness to males, low copulation propensity and the absence of egg fertilization, leads to cured asexual females producing only sons under conditions where the majority of sexual females (89.3%; Figure 1) produce daughters in addition to sons. Hence, we monitored the proportion of females producing only sons, as a measure of sexual trait decay, across increasing levels of introgression of the asexual genome into the sexual one. As expected, given the phenotype of the sexual and asexual strains for this trait, the proportion of females producing only sons increased with increasing levels of introgression (Figure 1, glm, $F_{1,405}=72.3$, $P<0.0001$). Importantly, the first significant increase was in the second generation (glm, $t=-4.5$, $P<0.0001$); no significant increase was observed in the first generation (glm, $t=1.0$, $P=0.340$; Figure 1). This indicates that recessive genetic effects cause females to produce only sons. More precisely, the quantitative increase of the proportion of females

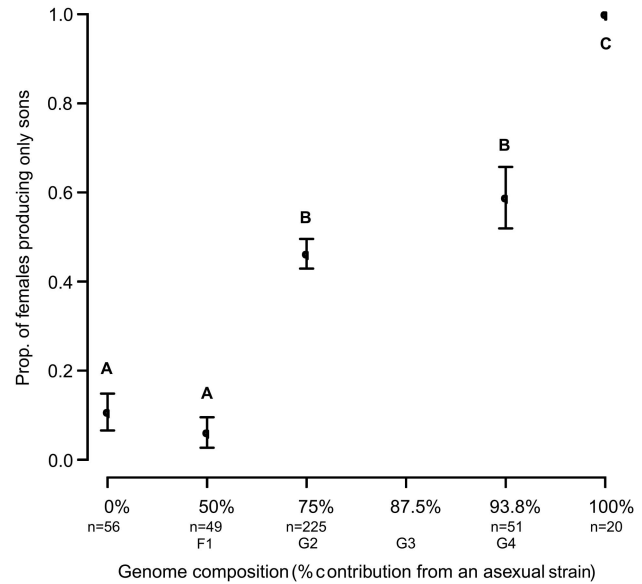


Figure 1 Proportion of females producing only sons when provided with a sexual male mating partner, for different categories of sexual–asexual hybrid females, and *Wolbachia*-cured asexual females (100% asexual genome). Bars indicate standard errors. The x axis represents the proportion of alleles stemming from the asexual genome for each consecutive generation. Different capital letters indicate statistical difference ($P<0.01$), n indicates the sample size for each generation and G1–4 refers to the different introgression generations. For logistic reasons, phenotyping of the G3 generation was not possible and therefore no data are provided.

producing only sons in the second generation is consistent with a simple genetic architecture of this trait (Table 2). Indeed, for hybrid females with 75% of their genome stemming from asexual strains, 46% (104 out of 225) produced only sons, which is in agreement with the expected 50% under a single-locus model, but differs significantly from the expected frequencies under models with two or more loci (Table 2).

Given that in addition to the females producing only sons, there was a (decreasing) fraction of females producing daughters in addition to sons, we also monitored how the offspring sex ratios of these females changed across increasing levels of introgression. However, in contrast to the proportion of females producing only sons, we had no specific prediction for how this trait would change with an increased representation of the asexual genome, given that this trait would derive from the sexual rather than the asexual genome. Indeed, as revealed by the experiments described above, cured asexual females do not fertilize any of their eggs, even if they have copulated. In our introgression experiment, we needed hybrid females for initiating each successive generation, hence we indirectly selected for females that copulated and fertilized at least some of their eggs. Following this indirect selection, we found that the sex ratio among offspring of females with at least some daughters fluctuated across increasing levels of introgression (Figure 2). Although females from all different introgression levels produced significantly more female-biased sex ratios than 'pure' sexual females, the most female-biased offspring sex ratios were found for the F1 hybrid females (Figure 2, glm, Tukey contrasts, all $P<0.001$). Even though these females also produced significantly fewer offspring (76.6 ± 2.6) than 'pure' sexual females (88 ± 3.1 ; Welch t -test, $t_{91,6}=2.9$, $P=0.004$), the low sex ratios are unlikely to stem from high mortality of males as a consequence of hybrid breakdown. Hybrid breakdown should cause

Table 2 Inferring the number of loci involved in the decay of a female sexual trait in asexuals

Number of loci Parental genotypes (F1 hybrid female × asexual male)	1 locus $Aa \times a$	2 loci $AaBb \times ab$	3 loci $AaBbCc \times abc$
Expected genotypes of offspring females (f = expected frequency); in bold: the genotype producing only sons	Aa ; aa ($f = 0.50$ each)	$AaBb$; $Aabb$; $aaBb$; $aabb$ ($f = 0.25$ each)	$AaBbCc$; $AaBbcc$; $AabbCc$; $Aabbc$; $aaBbCc$; $aaBbcc$; $aabbCc$; $aabccc$ ($f = 0.125$ each)
Binomial test	$P = 0.286$	$P < 0.00001$	$P < 0.00001$

Predicted proportion of sexual–asexual hybrid females (75% of the genome from the asexual strain) producing only sons for one to three unlinked loci, and the binomial test values for the comparison of each model with the observed data (0.46; 104 out of 225 females producing only sons, data from Figure 1).

Note: The predictions are based on two alleles at each locus: A and a, B and b, C and c, respectively. Capital letters denote dominant alleles, lower-case letters recessive alleles. Fixed homozygosity at each locus is assumed for the parental populations (for the dominant alleles in the sexual, the recessive alleles in the asexual population), and there is no epistasis among different loci. The predictions of recessive allele frequency from two and three-loci genetic model are significantly lower than the observed data, and higher number of loci would predict an even lower frequency.

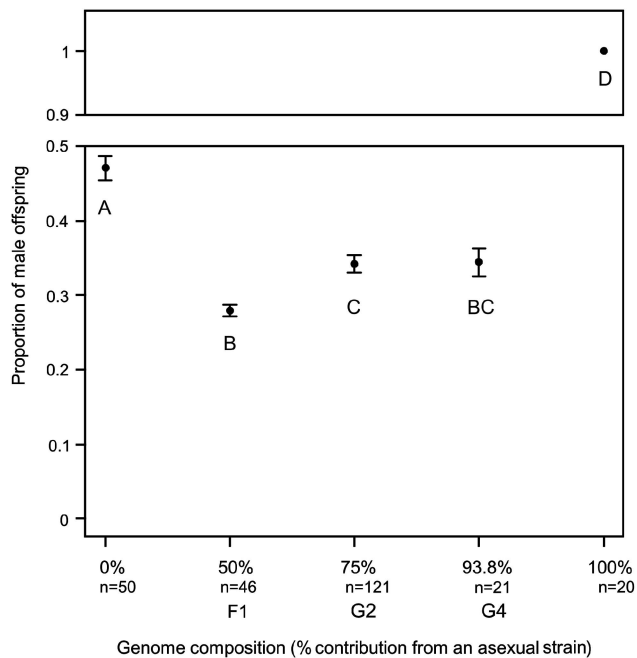


Figure 2 Offspring sex ratios produced by females with at least one daughter (that is, females producing only sons are excluded) for different categories of sexual–asexual hybrid females and *Wolbachia*-cured asexual females (100% asexual genome), when mated with sexual males. Bars indicate standard errors. Different capital letters indicate statistical difference ($P < 0.01$), n indicates the sample size for each generation and F1, G2, G4 refers to the different introgression generations.

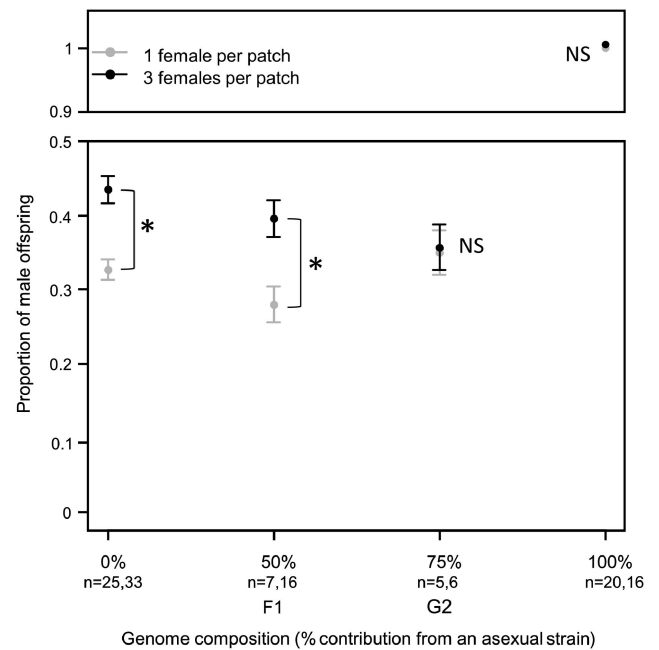


Figure 3 Offspring sex ratios produced by different sexual–asexual hybrid females and *Wolbachia*-cured asexual females (100% asexual genome), when alone (in gray) or in groups of three (in dark). Bars indicate standard errors. Proportions labeled with stars differ significantly ($P < 0.01$), NS indicates non-significant differences, n with two numbers indicates the sample sizes for the single females and the groups of three females per patch, respectively, and F1, G2 refers to the different introgression generations.

increased mortality of sons produced by F1 hybrid females independently of rearing conditions, yet we found no indication of hybrid breakdown among the females producing only sons (described above), and there was also no evidence for it in the LMC experiment (see the next paragraph). Hence, the reason for the initial drop in sex ratio and offspring number of F1 hybrid females remains unknown.

As asexual females produce only daughters, there is no selection for plastic sex allocation in response to different levels of LMC in asexual strains. We first verified that sexual *A. japonica* females indeed displayed plastic sex allocation. This was the case, as revealed by sexual females producing a significantly larger proportion of sons when in groups of three than when alone (Figure 3, glm, $F_{1,57} = 13.9$, $P < 0.001$). A similar response was also displayed by sexual–asexual F1 hybrid females (Figure 3, glm, $F_{1,22} = 5.5$, $P = 0.029$). However, in the second generation (females with 75% of their genome stemming

from asexual strains), females did not adjust their offspring sex ratio to different levels of LMC (all of these females produced at least one daughter; Figure 3, glm, $F_{1,10} = 0.2$, $P = 0.66$). We did not perform the LMC experiment for G3 and G4 introgression generations because plastic sex allocation was already completely lost in generation G2. In combination, these patterns suggest that there is decay of sex-allocation plasticity in asexual females, and that this decay is due to recessive genetic effects, similar to the decay of attractiveness and copulation propensity. However, an estimation of the number of loci affecting this trait cannot be provided, given that the presence vs absence of sex-allocation plasticity is evaluated qualitatively at the group level.

DISCUSSION

In this study, we used the parasitoid wasp *A. japonica* with *Wolbachia*-induced asexuality to investigate if female sexual traits decay under

asexuality, and if so, whether trait decay is caused by many loci with small effects or by only a few major-effect loci. A combination of behavioral experiments with crosses designed to introgress alleles from the asexual into sexual genome revealed decay of all investigated sexual traits and suggested a surprisingly simple genetic architecture underlying trait decay.

We found evidence for moderate decay of sexual attractiveness and extensive decay of mating and egg-fertilization behavior in asexual *A. japonica*. Asexual females were always less likely to be courted by males than sexual females, but the percentage of asexual females that were courted depended on whether these females were infected with *Wolbachia* (8%) or cured of their infection by antibiotic treatment (55%). This suggests that *Wolbachia* negatively affects the sexual attractiveness of infected females via an unknown mechanism. *Wolbachia* may, for example, modify sexual signals expressed in asexual females. Bacteria are known to alter pheromone production in some insects, such as commensal bacteria in *D. melanogaster* (Sharon *et al.*, 2010), and gut bacteria in the desert locust *Schistocerca gregaria* (Dillon and Charnley, 2002). In *D. melanogaster*, curing individuals from endosymbiont infection decreases levels of mate discrimination between populations by about 50%, an effect likely mediated by the effect of endosymbionts on female pheromone production (Koukou *et al.*, 2006; see also Sharon *et al.*, 2010). *Wolbachia* could also affect female attractiveness by altering cuticular hydrocarbon profiles, which function as mating cues in many insect species (for example, Ivy *et al.*, 2005; Yew *et al.*, 2009).

Mate attraction is often associated with significant costs in sexual species (Daly, 1978), because of resource investment in the production of mate attraction signals and/or because it can increase the risk of predation (for example, Zuk *et al.*, 2006). As a consequence, mate attraction is expected to be under strong negative selection in asexual species where it is superfluous, a prediction largely supported by empirical data (for example, Lehmann *et al.*, 2011; reviewed by van der Kooi and Schwander, 2014). However, once mate attraction is low, for example as a consequence of endosymbiont infection as we found for asexual *A. japonica*, the strength of selection for decreased expression of signals involved in mating interactions might be reduced. This may explain why asexual *A. japonica* females cured of their endosymbionts were still somewhat attractive to sexual males, although less than sexual females.

In contrast to the moderate decay of sexual attractiveness, mating behavior is largely disrupted in asexual females. This disruption is not caused by *Wolbachia* infection. Both infected and cured asexual females rejected all mating attempts by males under conditions where the majority of sexual females (88%) would accept mates and copulate. The mechanisms underlying disrupted mating interactions in asexual females remain to be investigated.

We also aimed at investigating if asexual females fertilize their eggs upon mating with a sexual male. In order to obtain any mated asexual females, we had to pair cured asexual females individually with groups of over 50 sexual males. The four asexual females that contained sperm in their spermathecae either produced no offspring at all (one female) or did not fertilize their eggs and produced only sons (three females). Although the sample is small, this pattern suggests that the ability to produce fertilized embryos decayed in asexual *A. japonica* females. Decay of egg fertilization can have different causes, for example egg modifications such as impermeability to sperm, and/or a lack of structures for sperm maintenance in the spermatheca. In the asexual parasitoid wasp *Muscidifurax uniraptor*, an essential spermatheca-associated muscle is completely absent (Gottlieb and Zchori-Fein, 2001). Similarly, non-reproductive

workers of many ant species have degenerated spermathecae with a flattened reservoir epithelium with few organelles (Gobin *et al.*, 2008).

Different mechanisms have been suggested to drive the decay of female sexual traits. In *A. japonica* and other asexual species deriving from haplodiploid sexual ancestors, low copulation propensity and the absence of egg fertilization lead to cured asexual females producing only sons even when presented with mating opportunities, a pattern referred to as 'functional virginity' (Jeong and Stouthamer, 2005; Stouthamer *et al.*, 2010; Russell and Stouthamer, 2011). It has been suggested that such 'functional virginity' would be favored selectively in sexual populations where only a fraction of females are asexual as a consequence of endosymbiont infection (Stouthamer *et al.* 2010). Because 'functional virginity' would enhance male production in populations with female-biased sex ratios, uninfected females might benefit from such a trait, assuming their sons could reproduce sexually with the majority of females in that population. 'Functional virginity' may therefore become fixed in the population before the fixation of endosymbiont-induced asexuality. This could explain why female mating and egg-fertilization behaviors decay rapidly in species with endosymbiont-induced asexuality (Stouthamer *et al.*, 2010), a pattern reported for many haplodiploid species (Pijls *et al.*, 1996; Arakaki *et al.*, 2000; Pannebakker *et al.*, 2004a, 2005; Jeong and Stouthamer, 2005; Russell and Stouthamer, 2011; this study).

The spread of 'functional virginity' mutations along with endosymbiont-induced asexuality is, however, not the only possible cause leading to the decay of mating behavior and egg fertilization in asexual females. A more general driving force underlying trait decay under asexuality might be strong negative selection on sexual traits that are expressed in females (Pijls *et al.*, 1996; van der Kooi and Schwander, 2014). Indeed, reduced female sexual attractiveness, lower copulation propensity and the absence of egg fertilization are 'the norm' in asexual lineages (van der Kooi and Schwander, 2014). The majority of known asexual lineages derive from sexual ancestors with genetic sex determination systems other than haplodiploidy. In these cases, virgin sexual females produce no offspring at all (rather than producing sons as under haplodiploidy), such that the decay of their mating behavior and lack of egg fertilization cannot be explained by the spread of 'functional virginity' mutations.

To gain insights into the genetics of cured asexual females producing only sons, we monitored the proportion of such females across increasing levels of introgression of alleles from the asexual into the sexual strain. The proportion of females producing only sons increased gradually with an increasing representation of the asexual genome, indicating that alleles underlying asexuality-specific traits can indeed be introgressed into a sexual genome. The first significant increase was observed in the second generation, indicating that recessive genetic effects underlie an increased propensity of females to produce only sons (Figure 1). In addition, the proportion of second-generation (G2) hybrid females producing only sons matched the predicted proportion for a single-locus model (50%, Table 2). Cured asexual females produce only sons because of the combination of decreased sexual attractiveness, low copulation propensity and the absence of egg fertilization. Because we did not monitor each of these traits separately across the introgression generations, we cannot infer whether the single-locus effect that we found stems from changes at only one or some combination of these traits. Independently of whether the locus acts on a single or multiple traits, our study points to a rather simple genetic architecture underlying trait decay, possibly a single locus. A simple genetic architecture for other decayed sexual traits was also uncovered in three previous studies of independently

derived wasp lineages with endosymbiont-induced asexuality. Pannebakker *et al.* (2004a) found a single major locus underlying reduced male fertility using quantitative trait locus mapping. Two other studies investigated the decay of egg fertilization and found evidence for a single mutation with recessive effects in one case and dominant effects in the other (Jeong and Stouthamer, 2005; Russell and Stouthamer, 2011).

Two of the three studies investigating the genetic basis of female sexual trait decay in endosymbiont-induced asexuals found trait decay to be caused by recessive alleles (Jeong and Stouthamer, 2005; this study). This raises the question of how recessive alleles could spread and fix within a short evolutionary time; asexuality in *A. japonica* evolved very recently, as indicated by little or no molecular genetic differentiation between sexual and asexual populations for mitochondrial DNA (Murata *et al.*, 2009; Reumer *et al.*, 2012). One possible explanation may lie in the cytological mechanism of *Wolbachia*-induced parthenogenesis. Diploidy of the embryo is frequently restored through gamete duplication, which results in complete homozygosity (for example, Stouthamer and Kazmer, 1994; Gottlieb *et al.*, 2002; Pannebakker *et al.*, 2004b). Such homozygosity would facilitate spread of adaptive recessive mutations, similar to dominance favoring the spread of adaptive mutations in diploid species (Otto and Goldstein, 1992; Orr and Otto, 1994). It remains to be investigated whether recessive mutations may also frequently cause trait decay in species with forms of asexuality that maintain heterozygosity or whether trait decay in these cases is mostly caused by dominant mutations.

In addition to the females producing only sons, there was a (decreasing) fraction of females producing daughters in addition to sons across successive introgression generations. Copulation and egg-fertilization behaviors required for production of daughters most likely stem from the sexual rather than the asexual genome, given that cured asexual females almost never copulate, and do not fertilize their eggs in the rare event of copulation. As we needed daughters for initiating each successive introgression generation, we may have indirectly selected for a portion of the sexual genome favoring copulation and egg fertilization. Importantly, however, such putative indirect selection does not affect our conclusions for simple genetic effects underlying the increased tendency of females to produce only sons across successive introgression generations. Indeed, given the recessive effects underlying this tendency are only expressed in the second introgression generation, indirect selection would not have occurred prior to this generation, and we only used the first two generations to make inferences on genetic architecture. Following indirect selection for copulation and fertilization, we found that sex ratios among offspring of females with at least one daughter fluctuated across increasing levels of introgression, but were always more female-biased than sex ratios produced by 'pure' sexual females (Figure 2).

We found that sexual *A. japonica* females change the sex ratio of their offspring according to the level of LMC their sons are exposed to (Figure 3), and we therefore monitored sex allocation in single vs multiple female set-ups across successive introgression generations. Plastic sex allocation was displayed by first generation sexual–asexual F1 hybrid females, but not by females with a higher level of introgression of the asexual genome. This indicates that plastic sex-allocation behavior has decayed in asexual females. Similar to the traits discussed above, the decay of plasticity is most likely also caused by recessive genetic effects, given that plasticity was still expressed in the first, but not in the second or any of the later introgression generations. The mechanisms underlying this decay remain a matter of speculation, but might be due to a loss of control over egg fertilization or to a loss of perception of levels of competition.

The decay of plastic sex-allocation response under asexuality has not been investigated previously, hence it is impossible to evaluate if it represents a general trend in asexuals derived from haplodiploid ancestors or if such decay is specific to asexual *A. japonica*. Nevertheless, selection experiments in the (sexual) spider mite *Tetranychus urticae* have revealed that plastic sex-allocation behavior can decay rapidly (within 54 generations) under situations of relaxed selection (Macke *et al.*, 2011). Such rapid decay, whatever the causes underlying it, would likely result in decay of plastic sex allocation in most if not all asexual species.

In conclusion, we have investigated the potential decay of four female sexual traits in asexual *A. japonica*: sexual attractiveness, mating behavior, egg fertilization and plastic sex allocation under different levels of LMC, and we have found evidence for decay of all four traits. We have shown that the propensity for females to produce only sons is likely caused by a recessive allele at a single locus. Recessive genetic effects also caused the decay of sex-allocation plasticity in asexuals. Whether trait decay frequently stems from recessive genetic effects, or whether recessive effects may be specific to the decay of sexual traits in asexuals characterized by gamete duplication remains to be investigated. Genetic mapping studies in progress, facilitated by next-generation sequencing techniques, will provide insights into this question, as well as into the molecular causes of sexual trait decay.

DATA ARCHIVING

Data available from the Dryad Digital Repository: doi:10.5061/dryad.rf0bh.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

ACKNOWLEDGEMENTS

We thank Rogier Houwerzijl and Peter Hes for assistance with wasp culturing; Ken Kraaijeveld and Barbara Reumer for supplying *A. japonica* strains. We thank Eric Wajnberg and three anonymous reviewers for constructive comments on a previous version of this manuscript. This work was supported by funding from the Netherlands Organization for Scientific Research to TS (Veni grant no. 863.09.001), a NWO/ALW TOP grant (no. 854.10.001) to LWB and by funding from the Netherlands Genomics Initiative to BAP (NGI Horizon Breakthrough no. 935.19.006 and NGI Zenith no. 935.11.04). TS was also funded by the Swiss National Science Foundation (FNS grant PP00P3_139013).

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- Arakaki N, Noda H, Yamagishi K (2000). *Wolbachia*-induced parthenogenesis in the egg parasitoid *Telenomus nawai*. *Entomol Exp Appl* **96**: 177–184.
- Carson HL, Chang LS, Lyttle TW (1982). Decay of female sexual behavior under parthenogenesis. *Science* **218**: 68–70.
- Crawley MJ (2007). *The R Book*, 1st edn. John Wiley & Sons Ltd: Chichester, UK.
- Daly M (1978). The cost of mating. *Am Nat* **112**: 771–774.
- Dedine F, Vavre F, Fleury F, Loppin B, Hochberg ME, Boulétreau M (2001). Removing symbiotic *Wolbachia* bacteria specifically inhibits oogenesis in a parasitic wasp. *Proc Natl Acad Sci USA* **98**: 6247–6252.
- Dillon R, Charnley K (2002). Mutualism between the desert locust *Schistocerca gerrardi* and its gut microbiota. *Res Microbiol* **153**: 503–509.
- Ellers J, Kiers ET, Currie CR, McDonald BR, Visser B (2012). Ecological interactions drive evolutionary loss of traits. *Ecol Lett* **15**: 1071–1082.
- Fong DW, Kane TC, Culver DC (1995). Vestigialization and loss of nonfunctional characters. *Annu Rev Ecol Syst* **26**: 249–268.
- Giorgini M, Bernardo U, Monti MM, Nappo AG, Gebiela M (2010). *Rickettsia* symbionts cause parthenogenetic reproduction in the parasitoid wasp *Pnigalio soemius* (Hymenoptera: Eulophidae). *Appl Environ Microbiol* **76**: 2589–2599.
- Gobin B, Ito F, Billen J, Peeters C (2008). Degeneration of sperm reservoir and the loss of mating ability in worker ants. *Naturwissenschaften* **95**: 1041–1048.

- Gottlieb Y, Zchori-Fein E (2001). Irreversible thelytokous reproduction in *Muscidifurax uniraptor*. *Entomol. Exp Appl* **100**: 271–278.
- Gottlieb Y, Zchori-Fein E, Werren JH, Karr TL (2002). Diploidy restoration in *Wolbachia*-infected *Muscidifurax uniraptor* (Hymenoptera: Pteromalidae). *J Invert Pathol* **81**: 166–174.
- Hamilton WD (1967). Extraordinary sex ratios. *Science* **156**: 477–488.
- Heath BD, Butcher RDJ, Whitfield WGF, Hubbard SF (1999). Horizontal transfer of *Wolbachia* between phylogenetically distant insect species by a naturally occurring mechanism. *Curr Biol* **9**: 313–316.
- Hothorn T, Bretz F, Westfall P (2008). Simultaneous inference in general parametric models. *Biometrical J* **50**: 346–363.
- Ivy TM, Weddle CB, Sakaluk SK (2005). Females use self-referent cues to avoid mating with previous mates. *Proc Roy Soc Lond B* **272**: 2475–2478.
- Jeffery WR (2009). Regressive evolution in *Astyanax* cavefish. *Annu Rev Genet* **43**: 25–47.
- Jeong G, Stouthamer R (2005). Genetics of female functional virginity in the parthenogenesis-*Wolbachia* infected parasitoid wasp *Telenomus nawai* (Hymenoptera: Scelionidae). *Heredity* **94**: 402–407.
- Kageyama D, Narita S, Watanabe M (2012). Insect sex determination manipulated by their endosymbionts: incidences, mechanisms and implications. *Insects* **3**: 161–199.
- Koukou K, Pavlikaki H, Kiliass G, Werren JH, Bourtzis K, Alahiotis SN (2006). Influence of antibiotic treatment and *Wolbachia* curing on sexual isolation among *Drosophila melanogaster* cage populations. *Evolution* **60**: 87–96.
- Kraaijeveld K, Franco P, Reumer BM, van Alphen JJM (2009). Effects of parthenogenesis and geographic isolation on female sexual traits in a parasitoid wasp. *Evolution* **63**: 3085–3096.
- Kremer N, Charif D, Henri H, Bataille M, Prévost G, Kraaijeveld K *et al.* (2009). A new case of *Wolbachia* dependence in the genus *Asobara*: evidence for parthenogenesis induction in *Asobara japonica*. *Heredity* **103**: 248–256.
- Lahti DC, Johnson NA, Ajie BC, Otto SP, Hendry AP, Blumstein DT *et al.* (2009). Relaxed selection in the wild. *Trends Ecol Evol* **24**: 487–496.
- Lehmann GUC, Siozios S, Bourtzis K, Reinhold K, Lehmann AW (2011). Thelytokous parthenogenesis and the heterogeneous decay of mating behaviours in a bushcricket (Orthoptera). *J Zool Syst Evol Res* **49**: 102–109.
- Ma W-J, Kuijper B, de Boer JG, van de Zande L, Beukeboom LW, Wertheim B *et al.* (2013). Absence of complementary sex determination in the parasitoid wasp genus *Asobara* (Hymenoptera: Braconidae). *PLoS ONE* **8**: e60459.
- Ma W-J, Vavre F, Beukeboom LW (2014). Manipulation of arthropod sex determination by endosymbionts: diversity and molecular mechanisms. *Sex Dev* **8**: 59–73.
- Macke E, Magalhães S, Bach F, Olivieri I (2011). Experimental evolution of reduced sex ratio adjustment under local mate competition. *Science* **334**: 1127–1129.
- McNab BK (1994). Energy conservation and the evolution of flightlessness in birds. *Am Nat* **144**: 628–642.
- Mateo Leach I, Pannebakker BA, Schneider MV, Driessen G, van de Zande L, Beukeboom LW (2009). Thelytoky in Hymenoptera with *Venturia canescens* and *Leptopilina clavipes* as case studies. In: Schön I, Martens K, van Dijk P (eds) *Lost Sex-The Evolutionary Biology of Parthenogenesis*. Springer: Dordrecht, pp 347–375.
- Murata Y, Ideo S, Watada M, Mitsui H, Kimura MT (2009). Genetic and physiological variation among sexual and parthenogenetic populations of *Asobara japonica* (Hymenoptera: Braconidae), a larval parasitoid of drosophilid flies. *Eur J Entomol* **106**: 171–178.
- Orr AH, Otto SP (1994). Does diploidy increase the rate of adaptation? *Genetics* **136**: 1475–1480.
- Otto SP, Goldstein DB (1992). Recombination and the evolution of diploidy. *Genetics* **131**: 745–751.
- Pannebakker BA, Beukeboom LW, van Alphen JJM, Brakefield PM, Zwaan BJ (2004a). The genetic basis of male fertility in relation to haplodiploid reproduction in *Leptopilina clavipes* (Hymenoptera: Figitidae). *Genetics* **168**: 341–349.
- Pannebakker BA, Pijnacker LP, Zwaan BJ, Beukeboom LW (2004b). Cytology of *Wolbachia*-induced parthenogenesis in *Leptopilina clavipes* (Hymenoptera: Figitidae). *Genome* **47**: 299–303.
- Pannebakker BA, Schidlo NS, Boskamp GJF, Dekker L, van Dooren TJM, Beukeboom LW *et al.* (2005). Sexual functionality of *Leptopilina clavipes* (Hymenoptera: Figitidae) after reversing *Wolbachia*-induced parthenogenesis. *J Evol Biol* **18**: 1019–1028.
- Pijls JWAM, van Steenberg HJ, van Alphen JJM (1996). Asexuality cured: the relations and differences between sexual and asexual *Apoanagyrus diversicornis*. *Heredity* **76**: 506–513.
- Pike N, Kingcombe R (2009). Antibiotic treatment leads to the elimination of *Wolbachia* endosymbionts and sterility in the diploid collembolan *Folsomia candida*. *BMC Biol* **7**: 54.
- Protas ME, Trontelj P, Patel NH (2011). Genetic basis of eye and pigment loss in the cave crustacean, *Asellus aquaticus*. *Proc Natl Acad Sci USA* **108**: 5702–5707.
- R Development Core Team (2011). *R: A Language and Environment for Statistical Computing*. Vienna, Austria.
- Rajaram S, Scott RL, Nash HA (2005). Retrograde signaling from the brain to the retina modulates the termination of the light response in *Drosophila*. *Proc Natl Acad Sci USA* **102**: 17840–17845.
- Reumer BM, van Alphen JJM, Kraaijeveld K (2012). Occasional males in parthenogenetic populations of *Asobara japonica* (Hymenoptera: Braconidae): low *Wolbachia* titer or incomplete coadaptation? *Heredity* **108**: 341–346.
- Russell JE, Stouthamer R (2011). The genetics and evolution of obligate reproductive parasitism in *Trichogramma pretiosum* infected with parthenogenesis-inducing *Wolbachia*. *Heredity* **106**: 58–67.
- Schwander T, Crespi BJ, Gries R, Gries G (2013). Neutral and selection-driven decay of sexual traits in asexual stick insects. *Proc Roy Soc Lond B* **280**: 20130823.
- Sharon G, Segal D, Ringo JM, Hefetz A, Zilber-Rosenberg I, Rosenberg E (2010). Commensal bacteria play a role in mating preference of *Drosophila melanogaster*. *Proc Natl Acad Sci USA* **107**: 20051–20056.
- Shuker DM, Reece SE, Lee A, Graham A, Duncan AB, West SA (2007). Information use in space and time: sex allocation behaviour in the parasitoid wasp *Nasonia vitripennis*. *Anim Behav* **73**: 971–977.
- Stouthamer R, Luck RF, Hamilton WD (1990). Antibiotics cause parthenogenetic *Trichogramma* (Hymenoptera: Trichogrammatidae) to revert to sex. *Proc Natl Acad Sci USA* **87**: 2424–2427.
- Stouthamer R, Kazmer DJ (1994). Cytogenetics of microbe-associated parthenogenesis and its consequences for gene flow in *Trichogramma* wasps. *Heredity* **73**: 317–327.
- Stouthamer R, Russell JE, Vavre F, Nunney L (2010). Intra-genomic conflict in populations infected by parthenogenesis inducing *Wolbachia* ends with irreversible loss of sexual reproduction. *BMC Evol Biol* **10**: 229.
- Suomalainen E, Saura A, Lokki J (1987). *Cytology and Evolution in Parthenogenesis*. CRC Press: Boca Raton, FL.
- van Alphen JJM, Nell HW (1982). Superparasitism and host discrimination by *Asobara tabida* Nees (Braconidae, Alysiinae), a larval parasitoid of *Drosophila*. *Neth J Zool* **32**: 232–260.
- van der Kooi C, Schwander T (2014). On the fate of sexual traits under asexuality. *Biol Rev*. (e-pub ahead of print 21 January 2014; doi:10.1111/brv.12078).
- Visser B, Le Lann C, den Blanken FJ, Harvey JA, van Alphen JJM, Eilers J (2010). Loss of lipid synthesis as an evolutionary consequence of a parasitic lifestyle. *Proc Natl Acad Sci USA* **107**: 8677–8682.
- Werren JH (1980). Sex ratio adaptations to local mate competition in a parasitoid wasp. *Science* **208**: 1157–1159.
- Werren JH (1997). Biology of *Wolbachia*. *Annu Rev Entomol* **42**: 587–609.
- Werren JH, Baldo L, Clark M (2008). *Wolbachia*: master manipulators of invertebrate biology. *Nat Rev Microbiol* **6**: 741–751.
- Werren JH, O'Neill SL (1997). The evolution of heritable symbionts. In: O'Neill SL, Hoffmann AA, Werren JH (eds) *Influential Passengers Inherited Microorganisms and Arthropod Reproduction*. Oxford University Press: New York, NY, USA, pp 1–41.
- Whiting PW (1933). Selective fertilization and sex-determination in Hymenoptera. *Science* **78**: 537–538.
- Wiens JJ (2001). Widespread loss of sexually selected traits: how the peacock lost its spots. *Trends Ecol Evol* **16**: 517–523.
- Yew JY, Dreisewerd K, Luftmann H, Muthing J, Pohlentz G, Kravitz EA (2009). A new male sex pheromone and novel cuticular cues for chemical communication in *Drosophila*. *Curr Biol* **19**: 1245–1254.
- Zuk M, Rothenberry JT, Tinghitella RM (2006). Silent night: adaptive disappearance of a sexual signal in a parasitized population of field crickets. *Biol Lett* **2**: 521–524.



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