# University of Windsor Scholarship at UWindsor

**Chemistry and Biochemistry Publications** 

Department of Chemistry and Biochemistry

12-22-2021

# Sharing the salt bowl: counterion identity drives N-alkyl resorcinarene affinity for pyrophosphate in water

Kwaku Twum Department of Chemistry, Oakland University

Seyed Iraj Sadraei Department of Chemistry and Biochemistry

Jordan Feder Department of Chemistry, Oakland University

S. Maryamdokht Taimoory Department of Chemistry and Biochemistry, University of Windsor

Kari Rissanen Department of Chemistry, University of Jyvaskyla

See next page for additional authors

Follow this and additional works at: https://scholar.uwindsor.ca/chemistrybiochemistrypub

Part of the Biochemistry, Biophysics, and Structural Biology Commons, and the Chemistry Commons

#### **Recommended Citation**

Twum, Kwaku; Sadraei, Seyed Iraj; Feder, Jordan; Taimoory, S. Maryamdokht; Rissanen, Kari; Trant, John F.; and Beyeh, Ngong Kodiah. (2021). Sharing the salt bowl: counterion identity drives N-alkyl resorcinarene affinity for pyrophosphate in water. *Organic Chemistry Frontiers*, 9, 1267-1275. https://scholar.uwindsor.ca/chemistrybiochemistrypub/225

This Article is brought to you for free and open access by the Department of Chemistry and Biochemistry at Scholarship at UWindsor. It has been accepted for inclusion in Chemistry and Biochemistry Publications by an authorized administrator of Scholarship at UWindsor. For more information, please contact scholarship@uwindsor.ca.

#### Authors

Kwaku Twum, Seyed Iraj Sadraei, Jordan Feder, S. Maryamdokht Taimoory, Kari Rissanen, John F. Trant, and Ngong Kodiah Beyeh

# ORGANIC CHEMISTRY

FRONTIERS

# **RESEARCH ARTICLE**

View Article Online View Journal | View Issue

ROYAL SOCIETY

CHEMISTRY

Check for updates

Cite this: Org. Chem. Front., 2022, 9, 1267

# Sharing the salt bowl: counterion identity drives *N*-alkyl resorcinarene affinity for pyrophosphate in water†

HINESE

CHEMICAL

Kwaku Twum, 🝺 <sup>a</sup> Seyed Iraj Sadraei,<sup>b</sup> Jordan Feder,<sup>a</sup> S. Maryamdokht Taimoory, 🝺 \*<sup>b,c</sup> Kari Rissanen, 🝺 \*<sup>d</sup> John F. Trant 🕩 \*<sup>b</sup> and Ngong Kodiah Beyeh 🕩 \*<sup>a</sup>

*N*-Alkyl ammonium resorcinarene chloride receptors, NARX<sub>4</sub>, have been shown to act as high-sensitivity detectors of pyrophosphate (PPi), a biomarker of disease, in aqueous media through the chloride-to-PPi exchange [NAR(Cl)<sub>4</sub> to NARPPi]. The nature of the anion of the macrocyclic NARX<sub>4</sub> (X = Cl<sup>-</sup>, Br<sup>-</sup>, triflate OTf<sup>-</sup>) receptor greatly influences the PPi-affinity in aqueous media. The binding affinity for [NAR (Cl)<sub>4</sub>] is  $3.61 \times 10^5 \text{ M}^{-1}$ , while the NAR (Br)<sub>4</sub> and NAR (OTf)<sub>4</sub> show stronger binding of  $5.30 \times 10^5 \text{ M}^{-1}$ , and  $6.10 \times 10^5 \text{ M}^{-1}$ , respectively. The effects of upper rim ammonium cation,  $-N^+H_2R$  substituents (R = 3-hydroxy-propyl, cyclohexyl, benzyl, or napththalen-1-ylmethyl), of the macrocyclic resorcinarene hosts have also been evaluated. The highest affinity was obtained using 3-hydroxypropyl groups due to the additional hydrogen bonds and the naphthyl upper-rim group that provides a larger hydrophobic surface area and favorable stacking interaction (*i.e.*,  $\pi - \pi$  and CH $-\pi$ ). We note that two PPi molecules can bind to the more selective receptors through an additional interaction with the lower rim hydroxyls, making the resorcinarene a divalent binder. Comparing PPi with other phosphate anions (PO<sub>4</sub><sup>3-</sup>, AMP, ADP, and ATP) shows that the receptors are more selective for PPi due to the size and charge complementarity. Experimental (<sup>1</sup>H, <sup>31</sup>P NMR, and isothermal titration calorimetry), and computational analyses support the reported trends for PPi selectivity even in highly competing aqueous media.

Received 22nd December 2021, Accepted 12th January 2022 DOI: 10.1039/d1qo01877a

rsc.li/frontiers-organic

# Introduction

A significant challenge in supramolecular chemistry is developing high-affinity receptors for anions in biologically relevant solvents,<sup>1–5</sup> and the design of such receptors has elicited considerable effort from the research community.<sup>6–15</sup> Primarily, this challenge has been addressed by leveraging the cooperative effect of non-covalent attractions.<sup>16–18</sup> One such anion is pyrophosphate (PPi), produced as a side product during ATP metabolism.<sup>14</sup> Physiological levels of PPi are used as a clinical indicator for disease diagnosis and prognosis.<sup>19–22</sup> For example, low PPi levels are common in hemodialysis patients<sup>23</sup> as it correlates inversely with the levels of vascular calcification in patients with chronic kidney diseases.<sup>24</sup> As a result, considerable effort has gone into developing chemosensors for analytical detection of anions. Most of these sensors are based on metal complexes of europium,<sup>25</sup> palladium,<sup>26</sup> iron,<sup>27</sup> copper,<sup>28</sup> cadmium,<sup>29</sup> and iron.<sup>30</sup> Other classical receptors based on polyammoniums are also used for the anion recognition in water. A naphthalimide-based receptor has been reported for pyrophosphate anion sensing in buffered solutions with around 10<sup>3</sup>–10<sup>5</sup> M<sup>-1</sup> affinities.<sup>31</sup>

Our preliminary contributions reported on *N*-alkyl ammonium resorcinarene salts (NARX<sub>4</sub>), consisting of a macrocyclic resorcinarene tetra-ammonium cation and four anions. Chloride, *viz*. NAR (Cl)<sub>4</sub>, is preferred for resorcinarene conformational stability in organic media over the bromide, nitrate, triflate, or picrate.<sup>32–34</sup> This binding preference is due to the chloride's suitable size, the snug fit between two adjacent ammonium moieties, and the strong hydrogen bond circular seam  $[NH_2^+...-Cl^-...NH_2^+...-Cl^-...]_2$  along the upper rim of the macrocycle that holds it in place. The tetra-cationic NARX<sub>4</sub>s are, on even a cursory inspection, excellent potential

<sup>&</sup>lt;sup>a</sup>Department of Chemistry, Oakland University, 146 Library Drive, Rochester,

MI 48309-4479, USA. E-mail: beyeh@oakland.edu

<sup>&</sup>lt;sup>b</sup>Department of Chemistry and Biochemistry, University of Windsor,

<sup>401</sup> Sunset Avenue, Windsor, ON, N9B 3P4, Canada.

E-mail: Nazanin.Taimoory@uwindsor.ca, j.trant@uwindsor.ca

<sup>&</sup>lt;sup>c</sup>University of Michigan, Department of Chemistry, Ann Arbor, MI, USA

<sup>&</sup>lt;sup>d</sup>University of Jyvaskyla, Department of Chemistry, P. O. Box 35, 40014 Jyvaskyla, Finland E-mail: kari t rissanen@ivv fi

Finland. E-mail: kari.t.rissanen@jyu.fi

<sup>†</sup>Electronic supplementary information (ESI) available: Synthesis, analytical methods, experimental details, NMR, ITC and UV-Vis/fluorescence data and detailed DFT. See DOI: 10.1039/d1qo01877a

hosts for tetra-anionic pyrophosphate due to size-charge complementarity and chelate cooperativity. In 2018, we reported that an *N*-alkyl ammonium resorcinarene chloride receptor is capable of selective high-affinity  $(10^7 \text{ M}^{-1})$  binding of PPi in pure water.<sup>35</sup> Binding is driven by the entropically favorable displacement of the four chloride counter ions upon complexation of PPi. Based on the mechanism, we recognized the potential to improve the receptor's affinity by employing more weakly coordinating counter ions. The upper rim of the ammonium salt receptor is available for structural modification. In our previous study, a seemingly minor change, incorporating an additional methylene group converting the chain from ethanol to propanol, greatly improved PPi affinity.<sup>35</sup> In these cooperative systems, even minor differences can drastically affect the thermodynamics of complexation.

We now report the application of less strongly coordinating counter anions, intending to significantly increase NARX<sub>4</sub>'s affinity towards PPi. We synthesized ten new R-NARX<sub>4</sub>'s (R = upper rim substituent, X = counter anion) receptors with three different counter anions: chloride, bromide, and triflate (OTf); and four different upper rim substituents: one with a flexible terminal hydroxyl propyl group (C3OHNARX<sub>4</sub>, X = Cl, Br, and OTf), the second with a rigid cyclohexyl group at the upper rim (CyNARX<sub>4</sub>, X = Cl and Br), the third with a flexible benzyl group (BnNARX, X = Cl and Br), and the fourth with a flexible and fluorescent napththalen-1-ylmethyl group (NpNARX<sub>4</sub>, X = Cl, Br and OTf, Fig. 1). In addition to PPi, the binding properties of the receptors towards a tribasic monophosphate (K<sub>3</sub>PO<sub>4</sub>), and a dibasic monophosphate (AMP), diphosphate (ADP), and triphosphate (ATP) are also investigated (Fig. 1).



Fig. 1 The resorcinarene salt receptors C3OHNARX<sub>4</sub>, CyNARX<sub>4</sub>, BnNARX<sub>4</sub>, NpNARX<sub>4</sub> (X = Cl<sup>-</sup>, Br<sup>-</sup> and CF<sub>3</sub>SO<sub>3</sub><sup>-</sup>), and the phosphate guests K<sub>3</sub>PO<sub>4</sub>, Na<sub>4</sub>PPi, Na<sub>2</sub>AMP, Na<sub>2</sub>ADP, and Na<sub>2</sub>ATP.

Binding was confirmed through <sup>1</sup>H and <sup>31</sup>P NMR experiments. Quantifying the thermodynamics of binding was accomplished using a series of isothermal titration calorimetry (ITC) and computational studies. The binding modes were justified using density functional theory (DFT) at the (wB96XD/6-311-G (d,p)) level of theory which was supported by molecular dynamic (MD) simulations.

### **Results and discussion**

The Mannich condensation between aliphatic amines and C-<sub>hydroxyl</sub>-resorcinarene in the presence of excess formaldehyde generates tetrabenzoxazines.<sup>36</sup> Ring-opening of the six-membered tetrabenzoxazine ring is effected by refluxing in the presence of an acid (HCl, HBr, and triflic acid) to provide crude R-NARX<sub>4</sub>s (Fig. 1); these are purified through recrystallization to give the final products in 50–85% yields (see ESI<sup>†</sup> for details).

#### Isothermal titration calorimetry (ITC)

Thermodynamic parameters (K,  $\Delta H$ ,  $\Delta S$ , and  $\Delta G$ ) between the R-NARX<sub>4</sub> receptors and the different phosphate anions were measured by a series of ITC experiments in 90% H<sub>2</sub>O/10% DMSO. A mixed solvent system was necessary to ensure the complete dissolution of the variable hydrophobic groups on the different R-NARX<sub>4</sub> receptors at the concentrations used for the study (0.25 mM).

Probing different functionalization of the upper rim alkyl (R) substituent was necessary to underscore its importance in defining phosphate anion affinity (Table 1). The switch to a combination of solvent systems is a likely explanation for lower K<sub>a</sub> values than we earlier reported.<sup>35</sup> The NARX<sub>4</sub> with naphthalene groups show higher binding when compared to benzyl or cyclohexyl analogs, potentially due to the presence of a series of stacking interactions (Table 1). However, our previous C3OH chains provided the best affinity constant due to the extra hydrogen bond potential of the hydroxyl groups. On all scaffolds, counter anion identity showed the same trend; the  $K_1$  value for the complexation of PPi to Np-NARX<sub>4</sub> increased by 52.5% and 68.2% (Table 1) upon switching the chloride counterion to bromide and triflate, respectively. The same trend remained when the water was replaced with pH 7.4 Tris buffer, confirming that the observed isotherms do not arise from (de)protonation but host-guest binding processes. Negative  $\Delta G$  values in all cases confirm that association is spontaneous at 298 K. The  $\Delta H$  and  $T\Delta S$  results also indicate the first binding event is both enthalpically and entropically favorable. This is slightly unexpected; entropically-driven complexation is the norm as solvent and counterions are liberated upon binding, but the energy of dehydrating pyrophosphate was expected to be substantial. The favorable  $\Delta H$  term hints at the significant strength of the salt bridges formed in the binding pocket.37,38 Affinity is maintained in these new receptors: the  $K_1$  values for PPi are generally more than for AMP, ADP, and ATP (Fig. S23–S33 and Tables S1–S11<sup>†</sup>).

Complex	$K_1 (\times 10^5)  \mathrm{M}^{-1}$	$\Delta H_1$ kcal mol <sup>-1</sup>	$T\Delta S_1$ kcal mol <sup>-1</sup>	$\Delta G_1$ kcal mol <sup>-1</sup>	Complex	$K_1 (\times 10^5) \mathrm{M}^{-1}$	$\Delta H_1$ kcal mol <sup>-1</sup>	$T\Delta S_1$ kcal mol <sup>-1</sup>	$\Delta G_1$ kcal mol <sup>-1</sup>
PPi@C3OHNAR(Br) <sub>4</sub> PPi@CyNAR(Br) <sub>4</sub> PPi@BnNAR(Br) <sub>4</sub> PPi@NpNAR(Br) <sub>4</sub>	$5.3 \pm 1.1 \\ 2.6 \pm 0.5 \\ 3.1 \pm 0.2 \\ 3.0 \pm 0.2 \\ 3.0 \pm 0.2 \\$	$\begin{array}{c} -6.7 \pm 2.2 \\ -1.1 \pm 0.5 \\ -0.4 \pm 0.1 \\ -4.9 \pm 0.6 \end{array}$	1.09 6.29 2.55	-7.81 -7.38 -7.48 -7.47	PPi@NpNAR(Cl) <sub>4</sub> PPi@NpNAR(Br) <sub>4</sub> PPi@NpNAR(OTf) <sub>4</sub>	$2.0 \pm 0.5$ $3.0 \pm 0.2$ $3.3 \pm 0.9$	$-3.3 \pm 2.0 \\ -4.9 \pm 0.6 \\ 7.0 \pm 0.7$	3.97 2.55 14.5	-7.23 -7.47 -7.51
Complex	$K_2 (\times 10^5)  \mathrm{M}^{-1}$	$\Delta H_2$ kcal mol <sup>-1</sup>	$T\Delta S_2$ kcal mol <sup>-1</sup>	$\Delta G_2$ kcal mol <sup>-1</sup>	Complex	$K_2 (\times 10^5) { m M}^{-1}$	$\Delta H_2$ kcal mol <sup>-1</sup>	$T\Delta S_2$ kcal mol <sup>-1</sup>	$\Delta G_2$ kcal mol <sup>-1</sup>
PPi@C3OHNAR(Br) <sub>4</sub> PPi@CyNAR(Br) <sub>4</sub> PPi@BnNAR(Br) <sub>4</sub> PPi@NpNAR(Br) <sub>4</sub>	$\begin{array}{c} 1.40 \pm 0.03 \\ 0.18 \pm 0.01 \\ 0.29 \pm 0.03 \\ 1.13 \pm 0.01 \end{array}$	$8.4 \pm 1.7$ $26.7 \pm 0.8$ $16.0 \pm 1.5$ $32.6 \pm 2.1$	15.3 32.2 22.1 39.6	-6.94 -5.60 -6.10 -7.00	PPi@NpNAR(Cl) <sub>4</sub> PPi@NpNAR(Br) <sub>4</sub> PPi@NpNAR(OTf) <sub>4</sub>	$0.59 \pm 0.08$ 1.13 $\pm 0.01$ 4.6 $\pm 2.1$	$\begin{array}{c} 11.1 \pm 1.8\\ 32.6 \pm 2.1\\ -1.4 \pm 0.2 \end{array}$	17.6 39.6 7.72	-6.50 -7.00 -9.09
<sup><i>a</i></sup> ITC was done in H <sub>2</sub> C	(90%) /DMSO (1	$(0\%)$ at 298 K. $K_1$ an	Id $K_2$ represent the f	first and second bi	nding constants.				

All the data, with few exceptions, were fitted to a two-set-ofsites binding model. As the cavity is committed to binding the first PPi molecule, the second interaction in C3OH-NARX may be allosteric exo binding with the hydroxyls of the top rim, as we previously speculated.<sup>35</sup> However, as the other hydrophobic R-NARXs, lacking these hydroxyl hydrogen bond participants, present a similar two binding site event in this solvent mixture, this seems unlikely. There is, however, another allosteric binding site formed from the four dangling hydroxyl chains of the lower rim. To investigate this possibility, we turned to NMR spectroscopy.

#### NMR spectroscopy

To probe these receptors' binding capability, we performed a series of <sup>1</sup>H NMR measurements on the phosphate guests alone. The methylene (Ar-CH<sub>2</sub>N) signals of the host alone confirm that a host-guest interaction between the NARX<sub>4</sub>s and the PPi occurs when the two compounds are mixed (Fig. 2 and Fig. S35–43<sup>†</sup>). We used the same solvent mixture of  $D_2O/[D_6]$ DMSO (90%/10%) at 298 K. The binding processes are fast on the NMR timescale; however, clear changes in the hosts' signals demonstrate that binding occurs. Hydrogen/Deuterium exchange in this solvent mixture prevents the -OH and -NH2 signals from being monitored. However, we observed changes in the chemical environment of the non-exchangeable methylene protons (Ar-CH<sub>2</sub>N) closest to the -OH and -NH<sub>2</sub> groups. We also observed, by NMR, that the resorcinarene core distorts its bowl conformation as observed from the Ar-H signals to accommodate PPi better. We first performed a representative NMR titration with C3OH-NAR(OTf)<sub>4</sub> and C3OH-NAR(Br)<sub>4</sub> in D<sub>2</sub>O to confirm the binding event. A Job's plot of the Aryl-H and R-CH<sub>2</sub>-NH<sub>2</sub> proton signals that participate in the cavitand binding pocket reveals a 1:2 stoichiometry (Fig. S34†).39 Moreover, a careful examination of the proton R-C(H<sub>2</sub>)OH signals of the lower rim reveal a gradual shielding due to this secondary binding event (Fig. S35†). From the NMRs, the -OH and -NH<sub>2</sub> protons of the cationic core of the receptors form hydrogen bonds with the counter anions. When the R-NARX<sub>4</sub> receptors bind to PPi, the counter anions are displaced to accommodate PPi. This reorganization produces observable differences in the <sup>1</sup>H resonances of the -OH, -NH<sub>2</sub>, and neighboring -CH<sub>2</sub>- protons.

Taking the binding of PPi by C3OHNARX<sub>4</sub>s as an example, up to 0.09 ppm upfield shifts are realized by the methylene protons, and up to 0.22 ppm upfield shifts are observed for the aromatic protons of the resorcinarene core. The changes in the host's signals support the host re-organizing the cavity during the binding process. No significant differences in the <sup>1</sup>H NMR of the receptor in PPi@R-NAR(X)<sub>4</sub> complex was observed between the chloride, bromide, or triflate counter ions once the counter anions are displaced to accommodate pyrophosphate. The final assembly is expected to be the same PPi@C3OHNAR(X)<sub>4</sub>, where X represents the initial counterion that has now been fully displaced (Fig. 2 and Fig. S36-S43<sup>†</sup>). An even better way to access the binding process is to monitor the <sup>31</sup>P NMR signals of the phosphate guests upon complexa-

 $[able\ 1]$  Thermodynamic binding parameters of formed complexes between PPi and the receptor R-NAR(X) $_4$  in mixed water system by ITC $^5$ 



**Fig. 2** Sections of the <sup>1</sup>H NMR spectra in  $D_2O/[D_6]DMSO$  (90/10 v/v) at 298 K of (a) C3OHNARCl<sub>4</sub> and (b) the equimolar mixture C3OHNARCl<sub>4</sub> and PPi, (c) C3OHNARBr<sub>4</sub>, and (d) the equimolar mixture C3OHNARBr<sub>4</sub> and PPi, (e) C3OHNAROTf<sub>4</sub> and (f) the equimolar mixture C3OHNAROTf and PPi. The dashed lines indicate the signal changes in ppm. Star represents the residual  $D_2O$  solvent.



**Fig. 3** Sections of the <sup>31</sup>P NMR spectra in  $D_2O/[D_6]DMSO$  (90/10 v/v) at 298 K. (a) Equimolar mixtures of PPi and NpNARX<sub>4</sub>s (X = Cl, Br, and OTf) as compared to pure PPi. (b) Equimolar mixtures of PPi and CyNARBr<sub>4</sub>, PPi and C3OHNARBr<sub>4</sub>, PPi and BnNARBr<sub>4</sub>, PPi and NpNARBr<sub>4</sub> compared to pure PPi. The dashed lines indicate the signal changes in ppm. Star represents the residual H<sub>3</sub>PO<sub>4</sub> as an external standard. Note on nomenclature: for clarity, the original counter anion is indicated in parenthesis.

tion with the NARX<sub>4</sub> receptors. A series of <sup>31</sup>P NMR experiments of the phosphate anions and equimolar concentrations of pure phosphates and receptors were used to qualitatively probe the binding processes (Fig. 3 and Fig. S44–S56†). The magnitude of the upfield shift of the phosphorus isotopic signals upon binding the R-NARX<sub>4</sub> receptors qualitatively infers how deep the guests' sit in the host's binding pocket. Of the different R-NARBr<sub>4</sub> receptors, PPi experienced the most significant upfield movement with R = naphthalene (Fig. 3). This was followed closely by the benzyl (Bn)–NARBr<sub>4</sub> receptor and 1-propanol (C3OH)–NARBr<sub>4</sub>. Qualitatively, the absence of an aromatic environment or a hydrogen bond potential of hydroxyl groups in the cyclohexyl upper rim modification may explain why it has the smallest effect on the PPi resonance (Fig. 3, Table 1 and Fig. S44–S56†).

Additionally, it is evident how the different counter ions contribute to the shielding of the pyrophosphate anions. The PPi signal was shielded by 1.14 ppm with C3OHNAR(Cl)<sub>4</sub>, 1.76 ppm with C3OHNAR(Br)<sub>4</sub>, and 1.85 ppm with C3OHNAR (OTf)<sub>4</sub>. Larger upfield shifts of 2.42, 2.50, and 2.72 ppm were observed upon switching from chloride to bromide or triflate with the NpNARX<sub>4</sub> receptors. The magnitude of these shifts is a qualitative representation of the different affinities. As the product is identical in each case, the changes are better interpreted as a measurement of the population ratios between bound (upfield shifted) and free (unchanged) PPi. This is further complicated by the second binding interaction of the guest (NaPPi) with the lower rim. Computational results suggest that PPi phosphorous atoms in the cavity have a higher point-charge electron density (Natural charge = 2.53e)

than a PPi bound to the lower rim (Natural charge = 2.48e, Fig. S57<sup>†</sup>). The weaker the coordinating anion, the greater the binding affinity for the upper rim. Thus, one would expect the instantaneous relative population of PPi in the upper vs. lower binding sites to grow as the anion coordinating ability drops. The increased shielding observed for phosphorus in the triflate pro-receptor compared to the chloride or bromide strongly supports this inference. The most significant shifts are observed for largely dissociative triflate as this counterion is easiest to displace. This same differential effect is not immediately apparent in the <sup>1</sup>H NMR spectra, where we note that the absolute shifts are similar. But they start from different starting points: the chemical shift of the indicated resonances in the parent resorcinarenes are not identical (although the peak shapes are), but upon binding, we do see differences in peak broadness, suggesting that although the PPi-bound complexes are expected to be all identical, we might be observing indicators of the dynamism of the binding process.

Similar <sup>31</sup>P experiments were used to probe the binding of the receptors towards other phosphates:  $PO_4^{3-}$ , AMP, ADP, and ATP.  $PO_4^{3-}$  showed similar but much weaker upfield movement than PPi, while AMP showed greater field changes than PPi. Moderate but measurable deshielding was observed for ADP and ATP. The interaction of ADP and ATP with the host might be through an *exo*-binding mode, explaining the weaker effects; their larger size could be the main factor in this behavior. Table 1 summarizes the <sup>31</sup>P signal changes upon complexation with the different R-NARX<sub>4</sub>s receptors.

#### **Computational studies**

The structural features of selected host-guest complexes, NpNARX<sub>4</sub>, C3OHNARX<sub>4</sub>, PPi@NpNAR, PPi@C3OHNAR, were then computationally evaluated in a 90% H<sub>2</sub>O/10% DMSO (SCRF = SMD solvation model) using  $\omega$ B97X-D/6-311-G(d,p) method.<sup>40</sup> For details of the methods, please see the ESI.† When X = Cl, the four chlorides position between the fourupper rim NH<sub>2</sub><sup>+</sup> groups, forming a plane parallel to the resorcinarene base (Fig. 4) as expected. Both C3OHNARCl<sub>4</sub> and NpNARCl<sub>4</sub> adopt superficially similar conformations but with different energetic behavior. The calculated binding energy of the host-counterion complex in this solvent system shows that the Cl in NpNARCl<sub>4</sub> ( $\Delta E_{\rm b}$  (1DMSO: 9H<sub>2</sub>O) = -62.4 kcal mol<sup>-1</sup>) is far less tightly bound than those in C3OHNARCl<sub>4</sub> (( $\Delta E_b$  $(1DMSO: 9H_2O) = -88.1$  kcal mol<sup>-1</sup>). This would imply that NpNARX should more readily lose its counterions, allowing for easier binding with the incoming phosphate, all else being equal. In the OTf-coordinated complexes, the bulky counterions distort the host geometry, forcing it further open, a decidedly unfavorable conformation. The C3OHNAROTf<sub>4</sub> complex has a positive host-anion binding energy ( $\Delta E_{\rm b}$  (1DMSO: 9H<sub>2</sub>O) = 7.6 kcal  $mol^{-1}$ ), suggesting that it should spontaneously dissociate. This unfavorable form was highlighted for the NpNAROTf<sub>4</sub> complex, where all attempts to optimize the structure failed: the ions simply do not form a defined, stable salt complex. This can be due to the high hydration energy in the



Fig. 4 The optimized geometries and the calculated binding energies of three representative complexes of the chloride and triflate salts. NpNARTf<sub>4</sub> could not be converged.

1DMSO/9H<sub>2</sub>O solvent system and strong competition between the host and the solvent for the counterions' attention. To better understand this relationship, a molecular dynamics (MD) simulation was performed on NpNAROTf<sub>4</sub> using OPLS-2005<sup>28,29</sup> for the counterions and the receptor to reflect the experimental conditions.41 The optimized geometry of the counterion-coordinated host system was initially relaxed, then the relaxed structure was immersed within the solvent box using the disordered system builder implemented in Schrödinger's Materials module.30 This solvent coordinated system was then subjected to MD simulations, with an initial production run of 5 ns (see ESI† for more details). The dynamic behavior of the counterion was then traced through four replicate MD simulations of 5 to 10 ns, each with different starting vectors. Due to this observed dynamic behavior of the counterion in the solvent system, the MD sampling was further extended with four more replicates of 20 ns each. The starting point and end result of representative MD trajectories are shown in Fig. 5. In all the simulated trajectories, two of the four OTf anions displace from their original positions between the pendant upper arms of host, leaving all the ions to be stabilized by solvent instead of each other. This explains why we were unable to optimize the structure using DFT method. This also prophesizes an incredibly favorable dissociation of triflate and helps explain the high affinity for PPi.

This arrangement is maintained throughout the full 20 ns of our MD sampling. This bulky and more weakly coordinating counterion, by readily dissociating, should greatly increase the binding affinity of incoming PPi for the host. With this initial



Fig. 5 MD simulation cell of the system containing disordered DMSO,  $H_2O$ , and two Np-8OTf systems, showing the position and displacement of the OTf counteranions in the beginning (left) and at the end of the equilibration state (right). The 2Np-8OTf complexes are shown in space-filling mode, and the solvent systems are shown in lines.

view of host-counteranions, we next investigated the complex formation with PPi.

The relative value of the predicted solvated binding energy of complex formation for PPi@C3OHNAR was found to be 1.8 kcal mol<sup>-1</sup> ( $\Delta\Delta G$ ) more favorable than for the PPi@NpNAR complex. The very strong hydrogen bonds in the former are met by the formation of new intra-host upper rim  $\pi$ - $\pi$  interactions between the naphthyl groups. It can be seen from the calculated host-guest and intra-host bond lengths (Fig. 7) that in PPi@NpNAR, four of PPi's oxygen atoms sit deep in the NpNAR cavity, forming strong interactions with the trimethyl ammonium and phenols, leaving only two PPi oxygen atoms to face the solvent. In PPi@C3OHNAR, the oxygen atoms of PPi are also immersed in the C3OHNAR host cavity, but less deeply and potentially more easily interrupted by the solvent. The deeper positioning of the guest in the PPi@NpNAR complex facilitates the intra-host interactions between upper rim OH…OH and the  $\pi$ - $\pi$  interactions between the Np groups, which are obviously not available to the PPi@C3OHNAR complex. The interactions block solvent from almost half of the circumference of the complex, protecting the host-guest interactions from interference.

However, all these calculations simply look at the 1:1 system. With it becoming increasingly clear that a 2:1



**Fig. 6** The optimized geometry of PPi@NpNAR, and PPi@C3OHNAR complexes. BE is binding energy, Ezpe is the zero-point energy; these terms together equate to  $\Delta H$ .



Fig. 7 The predicted structural parameters of the optimized PPi@NpNAR, and PPi@C3OHNAR complexes.

PPi: host stoichiometry exists, we reoptimized the trimeric complexes (Fig. 8). As we had considered, the optimal structure does have the second PPi unit, in the form of  $Na_4PPi$ , localized to the bottom rim of the cavitand. It does not adopt the same conformation for both systems; with C3OHNAR, it sits perpendicular to the axis of the cavitand, roughly parallel with the PPi in the upper cavity, while for NpNAR it sits aligned



Fig. 8 The optimized geometry of 2PPi@NpNAR and 2PPi@C3OHNAR complexes. Side view, top-down view, bottom-up view. BE is binding energy, Ezpe is the zero-point energy; these terms together equate to  $\Delta H$ .

with the cavitand's axis. This difference is likely because these are both low-lying interactions, and in neither case can all four rim hydroxyls engage with the PPi. However, it would be sensible that the degree of entropic contribution (provided by PPi and cavitand desolvation) would drive this forward. The overall  $\Delta G$  of binding to form this ternary complex, -60.6 kcal mol<sup>-1</sup> for C3OHNAR and -61.6 kcal mol<sup>-1</sup> for NpNAR, is roughly double that of the dimer above; binding to the lower rim is less favorable than the initial interaction with the cavity but is still highly exogenic. The reaction actually becomes endothermic on the  $\Delta H$  term (sum of the binding energy and the zeropoint correction energy), with the dimer (Fig. 6) being more favorable than the trimer; trimer formation is driven entirely by entropy. The entropy terms are overestimated in these calculations. This binding could likely be further improved through cooperative anion-cation interactions.

In all, the computational data parallels the relative results of the experimental ITC data: binding is favorable for a twosite model, and the NpNAR forms the stronger interactions with PPi.

# Conclusions

In conclusion, an extensive study in solution of ten *N*-alkyl ammonium resorcinarene salts with varying counter anions using <sup>1</sup>H and <sup>31</sup>P NMR, and ITC show the resorcinarene salts to be high-affinity receptors for PPi in aqueous media. The

receptor with the terminal propyl hydroxyl groups C3OHNAR  $(X)_4$  and the napththalen-1-ylmethyl group at the upper rim, both with weakly coordinating triflate counter anions, gave the best results with PPi. The results show that both the nature of the upper rim substituent and the coordinating strength of the counter anions play crucial roles in determining binding affinity. The C3OHNAR $(X)_4$  receptors provide an extra hydrogen bond for enhanced binding, while the NpNAR $(X)_4$  provides a larger surface area for binding. Weakly coordinating counter anions such as triflate enhanced the binding affinity over more coordinating for AMP while PO<sub>4</sub><sup>3-</sup> due to its smaller size was only weakly bound. The larger sizes of ADP and ATP suggest the binding to the receptor to be *exo*-cavity.

A detailed series of molecular dynamic simulations (MD) and the density functional theory (DFT) study highlighted the significance of the upper rim substituents and the counter anion as crucial factors for establishing the high binding affinity. These results show that modifying the upper rim and counter anions of the R-NAR(X)<sub>4</sub> enhances their affinity and sensor ability towards PPi. Similarly, they support the contention that the second binding interaction with PPi is competitive and occurs with a specific conformation with the lower rim hydroxyls rather than as an *exo*-interaction with the cavity, as we had previously speculated. These results pave the way to using a supramolecular approach using cavity containing resorcinarene salts receptors as qualitative sensors for PPi in biological media.

# Author contributions

Conceptualization, NKB, JFT, SMT, KR; methodology, KT, SIS, JF, SMT; software, SMT, validation, KT, SIS, JF, SMT; writing original draft preparation, KT, NKB, JFT, SIS, JF, SMT; writing —review and editing, KT, SMT, JFT, NKB, supervision, NKB, JFT, KR, SMT, project administration, NKB, JFT, KR; funding acquisition, NKB, KT, JFT, KR All authors have read and agreed to the published version of the manuscript.

# Conflicts of interest

The authors declare no conflict of interest.

# Acknowledgements

The authors gratefully acknowledge financial support from the Provost Graduate Research Scholarship (KT, JF), Oakland University, MI, USA, and the University of Windsor, ON, Canada. This work was made possible by the facilities of the Shared Hierarchical Academic Research Computing Network (SHARCNET: http://www.sharcnet.ca) and Compute/Calcul Canada. This work was funded in part by the Natural Sciences and Engineering Research Council of Canada (Grant # 2018-06338 to JFT), the Windsor Cancer Centre Foundation Seeds4Hope Seed Grant Funding Program (Grant # 2017-03 to JFT), and an Ontario Early Researcher Award (Grant # ER18-14-114 to JFT).

# Notes and references

- 1 M. A. Palacios, R. Nishiyabu, M. Marquez and P. Anzenbacher, Supramolecular chemistry approach to the design of a high-resolution sensor array for multianion detection in water, *J. Am. Chem. Soc.*, 2007, **129**(24), 7538– 7544.
- 2 A. E. Hargrove, S. Nieto, T. Zhang, J. L. Sessler and E. V. Anslyn, Artificial Receptors for the Recognition of Phosphorylated Molecules, *Chem. Rev.*, 2011, **111**(11), 6603–6782.
- 3 A. Rostami, C. J. Wei, G. Guérin and M. S. Taylor, Anion Detection by a Fluorescent Poly(squaramide): Self-Assembly of Anion-Binding Sites by Polymer Aggregation, *Angew. Chem., Int. Ed.*, 2011, **50**, 2059–2062.
- 4 T. Egawa, K. Hirabayashi, Y. Koide, C. Kobayashi, N. Takahashi, T. Mineno, T. Terai, T. Ueno, T. Komatsu, Y. Ikegaya, N. Matsuki, T. Nagano and K. Hanaoka, Red fluorescent probe for monitoring the dynamics of cytoplasmic calcium ions, *Angew. Chem., Int. Ed.*, 2013, **52**, 3874–3877.
- 5 F. Zapata, A. Caballero, N. G. White, T. D. W. Claridge, P. J. Costa, V. Félix and P. D. Beer, Fluorescent chargeassisted halogen-bonding macrocyclic halo-imidazolium receptors for anion recognition and sensing in aqueous media, *J. Am. Chem. Soc.*, 2012, 134(28), 11533–11541.
- 6 X. Ji, R. T. Wu, L. Long, C. Guo, N. M. Khashab, F. Huang and J. L. Sessler, Physical Removal of Anions from Aqueous Media by Means of a Macrocycle-Containing Polymeric Network, *J. Am. Chem. Soc.*, 2018, **140**(8), 2777–2780.
- 7 F. Pan, N. K. Beyeh and K. Rissanen, Dimeric resorcinarene salt capsules with very tight encapsulation of anions and guest molecules, *RSC Adv.*, 2015, **5**, 57912–57916.
- 8 A. Borissov, I. Marques, J. Y. C. Lim, V. Félix, M. D. Smith and P. D. Beer, Anion Recognition in Water by Charge-Neutral Halogen and Chalcogen Bonding Foldamer Receptors, *J. Am. Chem. Soc.*, 2019, **141**(9), 4119–4129.
- 9 D. H. Lee, S. Y. Kim and J. I. Hong, A fluorescent pyrophosphate sensor with high selectivity over ATP in water, *Angew. Chem., Int. Ed.*, 2004, **43**(36), 4777–4780.
- 10 N. G. White, S. Carvalho, V. Félix and P. D. Beer, Anion binding in aqueous media by a tetra-triazolium macrocycle, *Org. Biomol. Chem.*, 2012, **10**, 6951–6959.
- 11 P. A. Gale, J. T. Davis and R. Quesada, Anion transport and supramolecular medicinal chemistry, *Chem. Soc. Rev.*, 2017, 46, 2497–2519.
- 12 N. Busschaert, C. Caltagirone, W. Van Rossom and P. A. Gale, Applications of Supramolecular Anion Recognition, *Chem. Rev.*, 2015, **115**(15), 8038–8155.
- 13 M. J. Langton, S. W. Robinson, I. Marques, V. Félix and P. D. Beer, Halogen bonding in water results in enhanced

anion recognition in acyclic and rotaxane hosts, *Nat. Chem.*, 2014, **6**, 1039–1043.

- 14 S. K. Kim, D. H. Lee, J.-I. Hong and J. Yoon, Chemosensors for pyrophosphate, *Acc. Chem. Res.*, 2009, 42(1), 23–31.
- 15 V. Amendola, L. Fabbrizzi and L. Mosca, Anion recognition by hydrogen bonding: urea-based receptors, *Chem. Soc. Rev.*, 2010, **39**, 3889–3915.
- 16 R. A. Tromans, T. S. Carter, L. Chabanne, M. P. Crump, H. Li, J. V. Matlock, M. G. Orchard and A. P. Davis, A biomimetic receptor for glucose, *Nat. Chem.*, 2019, **11**, 52–56.
- 17 Z. Zhang and P. R. Schreiner, (Thio)urea organocatalysis what can be learnt from anion recognition?, *Chem. Soc. Rev.*, 2009, **38**, 1187–1198.
- S. Kubik, Anion recognition in water, *Chem. Soc. Rev.*, 2010, 39, 3648–3663.
- 19 L. Li Tong, Z. Z. Chen, Z. Y. Jiang, M. M. Sun, L. Li, J. Liu and B. Tang, Fluorescent sensing of pyrophosphate anion in synovial fluid based on DNA-attached magnetic nanoparticles, *Biosens. Bioelectron.*, 2015, 72, 51–55.
- 20 A. Micheli, J. Po and G. H. Fallet, Measurement of soluble pyrophosphate in plasma and synovial fluid of patients with various rheumatic diseases, *Scand. J. Rheumatol.*, 1981, **10**, 237–240.
- 21 R. G. Russell, S. Bisaz, A. Donath, D. B. Morgan and H. Fleisch, Inorganic pyrophosphate in plasma in normal persons and in patients with hypophosphatasia, osteogenesis imperfecta, and other disorders of bone, *J. Clin. Invest.*, 1971, **50**, 961–969.
- 22 M. Doherty, C. Belcher, M. Regan, A. Jones and J. Ledingham, Association between synovial fluid levels of inorganic pyrophosphate and short term radiographic outcome of knee osteoarthritis, *Ann. Rheum. Dis.*, 1996, **55**, 432–436.
- 23 K. A. Lomashvili, W. Khawandi and W. C. O'Neill, Reduced Plasma Pyrophosphate Levels in Hemodialysis Patients, *J. Am. Soc. Nephrol.*, 2005, **16**(8), 2495–2500.
- 24 W. C. O'Neill, M. K. Sigrist and C. W. McIntyre, Plasma pyrophosphate and vascular calcification in chronic kidney disease, *Nephrol., Dial., Transplant.*, 2010, **25**(1), 187–191.
- 25 S.-H. Li, C.-W. Yu, W.-T. Yuan and J.-G. Xu, A Lanthanide Hybrid Cluster as a Selective Optical Chemosensor for Phosphate-containing Anions in Aqueous Solution, *Anal. Sci.*, 2004, **20**(10), 1375–1377.
- 26 J. Gao, T. Riis-Johannessen, R. Scopelliti, X. Qian and K. Severin, A fluorescent sensor for pyrophosphate based on a Pd(ii) complex, *Dalton Trans.*, 2010, **39**, 7114– 7118.
- 27 W. Wang, H. Zhao, B. Zhao, H. Liu, Q. Liu and Y. Gao, Highly Selective Recognition of Pyrophosphate by a Novel Coumarin-Iron(III) Complex and the Application in Living Cells, *Chemosensors*, 2021, **9**(3), 48.
- 28 J. Qiang, C. Chang, Z. Zhu, T. Wei, W. Yu, F. Wang, J. Yin, Y. Wang, W. Zhang, J. Xie and X. Chen, A dinuclearcopper(II) complex-based sensor for pyrophosphate and its applications to detecting pyrophosphatase activity and monitoring polymerase chain reaction, *Sens. Actuators, B*, 2016, 233, 591–598.

- 29 J. X. Liu and S. N. Ding, Monitoring pyrophosphate anions via cobalt(II)-modulated fluorescence of cadmium sulfide quantum dots, *Anal. Methods*, 2016, **8**, 2170–2175.
- 30 N. Kumari, H. Huang, H. Chao, G. Gasser and F. Zelder, A Disassembly Strategy for Imaging Endogenous Pyrophosphate in Mitochondria by Using an FeIII-salen Complex, *ChemBioChem*, 2016, 17(13), 1211–1215.
- 31 A. S. Oshchepkov, R. R. Mittapalli, O. A. Fedorova and E. A. Kataev, Naphthalimide-Based Polyammonium Chemosensors for Anions: Study of Binding Properties and Sensing Mechanisms, *Chem. – Eur. J.*, 2017, 23(40), 9657–9665.
- 32 N. K. Beyeh, F. Pan, S. Bhowmik, T. Mäkelä, R. H. A. Ras and K. Rissanen, N-Alkyl Ammonium Resorcinarene Salts as High-Affinity Tetravalent Chloride Receptors, *Chem. – Eur. J.*, 2016, 22(4), 1355–1361.
- 33 N. K. Beyeh, A. Valkonen, S. Bhowmik, F. Pan and K. Rissanen, N-Alkyl ammonium resorcinarene salts: Multivalent halogen-bonded deep-cavity cavitands, *Org. Chem. Front.*, 2015, 2, 340–345.
- 34 N. K. Beyeh, M. Cetina and K. Rissanen, Halogen bonded analogues of deep cavity cavitands, *Chem. Commun.*, 2014, 50, 1959–1961.
- 35 N. K. Beyeh, I. Díez, S. M. Taimoory, D. Meister, A. I. Feig, J. F. Trant, R. H. A. Ras and K. Rissanen, High-affinity and

selective detection of pyrophosphate in water by a resorcinarene salt receptor, *Chem. Sci.*, 2018, **9**, 1358–1367.

- 36 N. K. Beyeh and K. Rissanen, N-Alkyl Ammonium Resorcinarene Salts: A Versatile Family of Calixarene-Related Host Molecules, in *Calixarenes and Beyond*, P. Neri, J. Sessler and M. X. Wang, 2016, pp. 255–284.
- 37 D. H. Williams, E. Stephens, D. P. O'Brien and M. Zhou, Understanding Noncovalent Interactions: Ligand Binding Energy and Catalytic Efficiency from Ligand-Induced Reductions in Motion within Receptors and Enzymes, *Angew. Chem., Int. Ed.*, 2004, 43(48), 6596–6616.
- 38 H. Saint-Martin, I. Ortega-Blake, A. Leś and L. Adamowicz, The role of hydration in the hydrolysis of pyrophosphate. A Monte Carlo simulation with polarizable-type interaction potentials, *Biochim. Biophys. Acta, Protein Struct. Mol. Enzymol.*, 1994, **1207**(1), 12–23.
- 39 K. A. Connors, *Binding constants: the measurement of molecular complex stability*, Wiley, 1987.
- 40 J. da Chai and M. Head-Gordon, Long-range corrected hybrid density functionals with damped atom–atom dispersion corrections, *Phys. Chem. Chem. Phys.*, 2008, **10**, 6615–6620.
- 41 D. E. Shaw, *Desmond Molecular Dynamics System*, Schrödinger, New York, 2019.