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11-29-2021

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IgG4-Mediated Neurologic Autoimmunities

Understanding the Pathogenicity of IgG4, Ineffectiveness of IVIg, and Long-Lasting Benefits of Anti–B Cell Therapies

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Neurol Neuroimmunol Neuroinflamm 2022;9:e1116. doi:10.1212/NXI.00000000001116

Abstract

Background and Objectives

Describe the unique functions of immunoglobulin G4 (IgG4) in IgG4-neurologic disorders (IgG4-ND) and explain why, in contrast to their IgG1-counterparts, they respond poorly to intravenous immune globulin (IVIg) but effectively to anti–B cell therapies.

Methods

The IgG4 structure and isotype switch, B cells and plasmablasts relevant to IgG4 production, and IgG4-induced disruption of the targeted antigens are reviewed and compared with IgG1-mediated autoimmune ND, where IVIg inhibits IgG1-triggered inflammatory effects.

Results

The main IgG4-ND include muscle-specific kinase myasthenia; nodal/paranodal chronic inflammatory demyelinating polyradiculoneuropathy with antibodies to neurofascin-155, contactin-1/ caspr-1, or pan-neurofascins; antileucine-rich, glioma-inactivated-1 and contactin-associated proteinlike 2 associated-limbic encephalitis, Morvan syndrome, or neuromyotonia; and anti-IgLON5 disorder. The IgG4, because of its unique structural features in the hinge region, has noninflammatory properties being functionally monovalent and bispecific, unable to engage in cross-linking and internalization of the targeted antigen. In contrast to IgG1 subclass which is bivalent and monospecific, IgG4 does not activate complement and cannot bind to inhibitory $Fc\gamma$ receptor ($Fc\gamma$ RIIb) to activate cellular and complement-mediated immune responses, the key functions inhibited by IVIg. Because IVIg contains only 0.7%–2.6% IgG4, its idiotypes are of IgG1 subclass and cannot effectively neutralize IgG4 or sufficiently enhance IgG4 catabolism by saturating FcRn. In contrast, rituximab, by targeting memory B cells and IgG4-producing CD20-positive short-lived plasma cells, induces long-lasting clinical benefits.

Discussion

Rituximab is the preferred treatment in IgG4-ND patients with severe disease by effectively targeting the production of pathogenic IgG-4 antibodies. In contrast, IVIG is ineffective because it inhibits immunoinflammatory functions irrelevant to the mechanistic effects of IgG4 and contains IgG-1 idiotypes that cannot sufficiently neutralize or possibly catabolize IgG4. Controlled studies with anti-CD19/20 monoclonals that also activate $Fc\gamma$ RIIb may be more promising in treating IgG4-ND.

Go to Neurology.org/NN for full disclosures. Funding information is provided at the end of the article.

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The Article Processing Charge was funded by the author.

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Glossary

AChR = acetylcholine receptor; CASPR2 = contactin-associated protein-like 2; CIDP = chronic inflammatory demyelinating polyradiculoneuropathy; CTL = cytotoxic T lymphocytes; $Fc\gamma R = Fc\gamma$ receptor; IgG4 = immunoglobulin G4; IgG4-ND = IgG4-neurologic disorder; IgG4-RD = IgG4-related disease; IVIG = intravenous immune globulin; LGI1 = leucine-rich, glioma-inactivated-1; MAC = membranolytic attack complex; MG = myasthenia gravis; MuSK = muscle-specific kinase; PNS = peripheral nervous system.

Several autoimmune, multisystemic, or fibroinflammatory disorders have been recently identified based on their association with immunoglobulin G4 (IgG4) subclass of autoantibodies, referred to as IgG4-related diseases (IgG4-RD).¹⁻³ In contrast, however, to a broad IgG4-RD spectrum with nondiseasespecific pathogenic autoantibodies except for pemphigus vulgaris, membranous nephropathy, and thrombotic thrombocytopenic purpura, we are witnessing key IgG4-neurologic disorders (IgG4-ND) with pathogenic IgG4 antibodies targeting neural antigens highlighted by MuSK-myasthenia; nodal/paranodal chronic inflammatory demyelinating polyradiculoneuropathy (CIDP) with paranodal antibodies to neurofascin-155, contactin-1, contactin-associated protein-like 1 (CASPR1), and nodal/paranodal pan-neurofascins (NF140/ NF186/NF155); leucine-rich, glioma-inactivated-1 (LGI1) or the juxtaparanodal CASPR2-associated autoimmune encephalitis, Morvan syndrome, neuromyotonia, or autoimmune pain syndromes; and the rare anti-IgLON5 disorder.⁴⁻¹⁰ The unique feature of IgG4-ND is their significant disease severity that, in contrast to their IgG1 counterparts, exhibit poor response to intravenous immune globulin (IVIg) and inadequate response to steroids or plasmapheresis but excellent response to anti-B cell therapies, such as rituximab, that downregulate humoral immunity. Although some of these patients may have unique clinical phenotypes, most often present similarly to IgG1 counterparts and treated identically until recognized in retrospect that they are refractory to conventional immunotherapies. Their resistance to these therapies especially IVIg, which is the treatment of choice in their IgG1-counterparts based on controlled trials, is poorly understood, leading to therapeutic delays necessitating vigilance for proper therapy initiation.

Because IgG4-ND are now increasingly recognized, it has become imperative to understand the uniqueness of IgG4 pathogenicity and the rationale of the most effective immunotherapies. For the neurologists, the information is also relevant to IgG4-RD which, although present with autoimmune multisystemic, lymphoproliferative, or fibroinflammatory conditions, may also exhibit neurologic manifestations of meningeal and spinal cord disease, hypertrophic pachymeningitis, orbital myositis, or hypophysitis that may also need neurologic expertise.

The article addresses the uniqueness of IgG4 isotype; the role of regulatory B cells, cytokines, and plasmablasts in the IgG4 production; the mechanism by which IgG4 antibodies cause dysfunction of their targeted antigens; the reasoning of why IVIg, which is often the first-line therapy in their IgG1 counterparts, is ineffective; the currently successful anti–B cell therapy with rituximab, including practical issues on repeated infusions or IgG4 biomarkers; and promising future anti-IgG4-immunotherapies.

The Uniqueness of IgG4 Antibodies

IgG4 antibodies evolve as an anti-inflammatory response to chronic antigenic stimulation traditionally connected to peripheral tolerance because of high-dose allergen exposure, as occuring in beekeepers, cat owners, or helminth-infected subjects, alleviating allergic inflammation by interfering with the binding of allergen-specific IgE to the allergens.² In healthy adults, IgG4 is the least common IgG subclass, comprising only 5% of the total IgG with a concentration of 0.08-1.4 g/L.¹⁻³ Owing to its unique structural features in the hinge region, the IgG4 antibodies, although continuously undergo half antibody exchange with other IgG4 molecules, are considered immunologically inert and functionally monovalent because, in contrast to IgG1 which are bivalent and monospecific, they recognize the antigen essentially with only 1 Fab-arm of the IgG4; as a result, they are unable to engage in cross-linking and internalization of their target antigen or form immune complexes having noninflammatory properties.^{1,3,5,11} IgG4 functions differently from the other IgG subclasses by 2 key characteristics: first, cannot bind the first C1q complement component to activate the complement cascade, and second, they bind uniquely to Fc receptors with markedly reduced binding capacity to inhibitory Fcy receptor (Fc γ RIIb) but with enhanced binding to the activating FcyRI.¹⁻³ Collectively, IgG4-antibodies are inadequate in activating cellular or complement-mediated immune responses, which are directly targeted by IVIg and conventional immunotherapies; instead, they exert their pathogenicity by blocking protein-protein interactions and affecting signal transduction pathways. Whether genetic factors promote the development of IgG4 isotype as a response to certain antigens, as noted for certain paranodal antibodies mentioned later, is a possibility that needs to be further explored.

Role of Cytokines, T Cells, and B Cells in IgG4 Production and Isotype Switch

The IgG4 production is augmented by IL-10, which in IgG4-RD is overexpressed in affected tissues playing a key role in the isotype switching of B cells to IgG4 subclass.¹⁻³ IL-10-expressing T-follicular regulatory cells and follicular helper T (Tfh) cells that produce IL-4, IL-10, and IL-21 are also involved in class switch, contributing to the pathogenesis of IgG4-RD.^{1,3,11-14} CD4⁺ cytotoxic T lymphocytes (CTLs) are

prominent in IgG4-RD and decline after B cell-targeted therapy, suggesting that B cells present antigen to activate CD4⁺ CTLs.¹ Memory B cells and plasmablasts are also increased in the peripheral blood with the expansion of IgG4-producing CD19flCD20+CD27hiCD38hi plasmablasts, especially during relapses or active disease.¹⁻³

Insidious isotype switch can also occur late in the immune response because of maturation and hypermutation, as noted in nodal CIDP, from IgG3 against CNTN1/CASPR1 to IgG4 against CASPR1¹⁵ and in MuSK-MG from IgG4 to IgG1.⁵ Although overlooked, isotype switch can be clinically important in IgG4-ND when patients previously responding to IVIg become IVIgunresponsive after isotyped to IgG4.

IgG4-Neurologic Autoimmunity: The Effect of IgG4 Antibodies on Targeted Antigens

The IgG4 antibodies arise after chronic antigenic exposure late in the immune response and exhibit very high affinity for their antigen after undergoing several rounds of affinity maturation and somatic hypermutation.¹⁶ In contrast to most IgG4-RD, however, where the immunopathogenicity of IgG4 is poorly understood and their clinicopathology is broad-defined by increased serum IgG4 and tumefactive lesions with IgG4+ plasma cell infiltrates in multiple organs^{1,2}—the IgG4-neuroautoimmunity is distinct, characterized by antigen-specific disorders affecting only CNS or PNS. Most importantly, in IgG4-ND, the IgG4 antibodies exert a direct pathogenic effect by blocking enzymatic activity or disrupting protein-protein interactions of their target antigens affecting signal transduction pathways.⁵ Among the IgG4-RD, the main ones with antigen-specific IgG4 that share immunologic similarities with IgG4-ND are pemphigus vulgaris with antidesmoglein antibodies, membranous nephropathy with M-type phospholipase A2 receptor 1 or thrombospondin type-1 antibodies, thrombotic thrombocytopenic purpura with antibodies to metalloprotease ADAMTS13, and possibly Goodpasture syndrome with antitype IV collagen antibodies. The IgG4-ND include

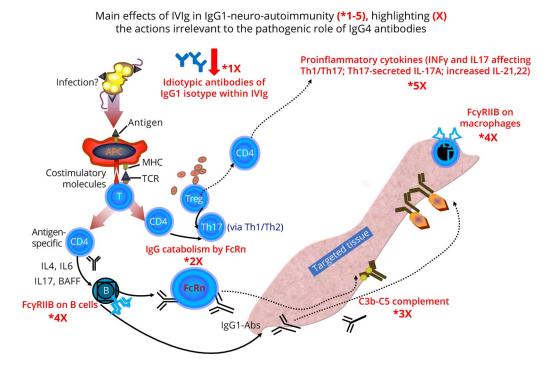
1. Anti-MuSK-MG. It comprises 7% of all patients with myasthenia gravis (MG) and is highlighted by IgG4 antibodies against muscle-specific kinase (MuSK), a transmembrane polypeptide expressed at the neuromuscular junction, that plays a fundamental role in acetylcholine receptor (AChR) clustering via interactions with agrin and rapsyn.^{4,5} Although MuSK-MG may have a phenotype similar to AChR-MG, many times has unique presentation with selective weakness and atrophy of the neck, tongue, shoulder, and bulbar muscles.^{17,18} MuSK-IgG4 antibodies are pathogenic, can passively transfer disease, and cause dysfunction of the neuromuscular junction by interfering with AChR clustering through the inhibition of Lrp4/MuSK signaling, but not by antigen cross-linking, internalization, or end-plate destruction as the IgG1-AChR antibodies do.^{4,19}

- 2. CIDP with nodal/paranodal antibodies. This clinicopathologically distinct CIDP subset, identified in the last 10 years, comprises 10% of patients with CIDP and is caused by IgG4 antibodies to paranodal neurofascin-155, CASPR1, and CNTN1^{6,20-22} and the nodal/paranodal pan-neurofascin (NF140/NF186/NF155 also called "nodal neurofascin"). These patients have distinct clinical phenotypes with distal weakness, tremor, and sensory (proprioceptive) ataxia. NF155 is a Schwann cell adhesion protein at the paranodal terminal myelin loops that binds to CNTN1, forming a complex critical for the maintenance of nodal structures ensuring rapid impulse propagation.²³ The antibodies are pathogenic, disrupting the NF155/CNTN1 complex by binding distinct epitopes associated with cell adhesion, resulting in the disadhesion of NF155/CNTN1 components by affecting glycosylation.^{6,20,22,23} The IgG4 NF155 antibodies are specifically deposited at the axoglial junction causing dissection at the paranode perturbing conduction in the absence of demyelination.^{23,24} These antibodies do not fix complement or internalize target antigens and do not cause inflammation; instead, they block proteinprotein interaction leading to conduction failure.^{5,23} Specific HLA-DRB1*15 alleles show strong association in anti-NF155 patients implicating a constitutive genetic risk susceptibility factor.²⁵
- LGI-1 and CASPR2-autoimmune syndromes presenting 3. with limbic encephalitis and seizures, Morvan syndrome and neuromyotonia, or severe neuropathic pain. These IgG4 antibodies are directed against the leucine-rich, glioma-inactivated-1 (LGI1) antigen or the juxtaparanodal contactin-associated protein-like 2 (CASPR2) that stabilize the voltage-gated potassium channel complex into the membrane.⁷⁻⁹ The LGI1 plays a role in bridging the presynaptic voltage-gated potassium channel protein Kv1.1 with the postsynaptic a-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptor through interaction with synaptic anchor molecules ADAM22/23; the anti-LGI1 antibodies alter the binding of LGI1 with ADAM22, decreasing the postsynaptic levels of a-amino-3-hydroxy-5methyl-4-isoxazolepropionic acid receptors.8

Although patients with anti-LGI-1 and CASPR2 antibodies may exhibit overlapping clinical symptomatology, the antibodies to LGI1 are most commonly associated with limbic encephalitis and epilepsy, whereas antibodies to CASPR2 with Morvan syndrome and neuromyotonia. Refractory epilepsy and mental and behavioral abnormalities are variably present. Pain, as seen in small fiber sensory neuropathy, and seizures involving ipsilateral face and limb dystonia-like seizures are also increasingly recognized.

4. Anti-IgLON5 disorder defines a complex syndrome of gait instability, craniofacial dyskinesias, sleep-disordered breathing, abnormal eye movements, and cognitive decline, characterized by antibodies, mostly of IgG4

Figure Immunopathogenic Network in IgG1-ND as Related to the Actions of IVIg (*1–5), Highlighting (*1–5 X) the Effects Irrelevant to IgG4-ND



In IgG1-ND, antigens presented via antigen presenting cells to CD4+T cells stimulate B cells to produce IgG1 antibodies which, by fixing complement at the target organ, lead to tissue damage. FcRn plays a role in catabolism of IgG1 antibodies, whereas the inhibitory FcyRIIB receptors on macrophages and B cells is effective in IgG1-ND collectively by supplying idiotypic antibodies of IgG1 subclass that exert neutralizing effects on circulating pathogenic IgG1 autoantibodies (*1); this function is irrelevant to IgG4-ND because IVIg contains only IgG1 isotypes that cannot neutralize IgG4 (*1-X); saturating the FcRn in endosomes resulting in increased catabolism of pathogenic IgG1(*2); this function is probably irrelevant to IgG4-ND because IgG1 sotypes may not sufficiently affect IgG4 catabolism (*2-X); binding to C3b, intercepting complement activation and the destructive effects or omacrophages and B cells (*4); this function is irrelevant to IgG4-ND because IgG4 does not fix complement (3*.X); upregulating FcyRIIB receptors on macrophages and B cells (*4); this function is irrelevant to IgG4-ND because IgG4 cannot bind to FcyRIIB(*4-X); and inhibiting proinflammatory cytokines (5*); this is irrelevant to IgG4-ND because IgG4 cannot bind to FcyRIIB (*4-X); and inhibiting proinflammatory cytokines (5*); this is irrelevant to IgG4-ND because IgG4 cannot bind to FcyRIIB (*4-X); and inhibiting proinflammatory cytokines (5*); this is irrelevant to IgG4-ND because IgG4 cannot bind to FcyRIIB (*4-X); and inhibiting proinflammatory cytokines (5*); this is irrelevant to IgG4-ND because IgG4 conto sufficiently induce such cytokines (*5-X). IgG1-ND = IgG1-neurologic disorder; FcyRIIb = Fcy receptor; IVIg = intravenous immune globulin.

isotype, against IgLON5, a neural cell adhesion molecule.¹⁰ IgLON5 antibodies disrupt the crosstalk between cell surface and cytoskeleton, leading to the abnormal accumulation of neurofilaments, providing a link between antibody-mediated autoimmunity and neurodegeneration.²⁶

Why IVIg Is Ineffective in IgG4-ND: Mechanisms of Actions of IVIg in Relevance to IgG4 Antibodies

In contrast to their autoimmune-ND counterparts that respond to IVIg in controlled studies, in IgG4-ND, there is no antigen cross-linking or immune complex formation, no recruitment of immune cells via Fc receptors, and no antigenic destruction via phagocytosis or antibody-dependent cellular cytotoxicity. Because these are the key functions inhibited by IVIg,^{27,28} all the hallmark anti-inflammatory effects of IVIg that define its success in IgG1-ND are irrelevant to the pathogenicity of IgG4, explaining not only the ineffectiveness of IVIg but also the suboptimal benefit of steroids. The key immunoregulatory actions of IVIg, contrasting with their irrelevance in IgG4-ND, include (Figure)

1. Supply idiotypic antibodies of IgG1 isotype capable of neutralizing pathogenic autoantibodies via $F(\alpha\beta')_2$ binding. IVIg has more than 95% IgG of IgG1 subclass but minimal IgG4 (0.7% - 2.6%). Derived from thousands of donors, IVIg contains anti-idiotypic IgG1 antibodies with a wide range of idiotypic and anti-idiotypic specificities, forming dimers connected by double-arm or single-arm binding between their $F(\alpha\beta)_2$ domains that bind to and neutralize pathogenic autoantibodies.^{27,29,30} The effect of idiotypes within the IVIg is highlighted in GBS and other demyelinating neuropathies with serum anti-GM1, anti-GQ1b, or other anti-glycolipid antibodies that block quantal release and impair neuromuscular transmission; the IgG1-idiotypic antibodies within the IVIg recognize these IgG glycoconjugates and inhibit or neutralize the blocking effects exerted by their sera, explaining the quick clinical benefits of IVIg.^{31,32} Because IVIg does not contain idiotypes of IgG4 subclass, it

cannot exert similar and sufficiently enough neutralizing or blocking effects on the circulating IgG4 antibodies (#1, Figure).

- 2. Acceleration of IgG1 catabolism by saturating the FcRn transport receptors. Normally, IgG antibodies bind to FcRn to return via endocytotic vesicles intact back into the circulation, being protected from degradation by the lysosomes.³³ The supraphysiological levels of IgG derived from IVIg administration saturate the FcRn, so a portion of endogenously produced pathogenic IgG antibodies are not recycled back to the circulation but degraded^{33,34}; as a result, the infused IVIg, by competing with pathogenic autoantibodies for FcRn binding, reduces the serum half-life of autoantibodies by approximately 40%, causing a reduction of the circulating IgG.³³⁻³⁵ This effect, although not studied, may not be applicable to IgG4 antibodies because IVIg has no IgG4 to affect sufficiently enough IgG4 recycling, adding to its ineffectiveness (#2, Figure).
- 3. Inhibition of complement binding and prevention of membranolytic attack complex (MAC) formation. IVIg inhibits complement uptake, intercepting the formation and deposition of MAC on the targeted tissues by activated complement or complement-fixing antibodies.^{36,37} On this basis, IVIg intercepts complement deposits on muscle or nerve fibers³⁶⁻³⁸ and prevents the cytotoxicity induced by dermatomyositis or GBS sera.^{37,38} These key anticomplement effects are not applicable to IgG4 antibodies because they do not mobilize or fix complement, as described earlier (#3, Figure).
- Modulation of $Fc\gamma R$. The $Fc\gamma Rs$ transduce either activating signaling via the FcyRIA, FcyRIIA, and FcyRIIIA or inhibitory signals via the FcyRIIB, regulating immune cell activation.^{39,40} IgG molecules bind through their Fc region to FcyR present on monocytes, macrophages, dendritic cells, and B cells and mediate inflammatory or immune effector functions by activating or inhibiting intracellular signaling, cellular activation, proliferation, and cytokine production.^{27,39,40} IVIg selectively upregulates the inhibitory FcyRIIB, a negative regulator of lymphoid cell activation, inhibiting phagocytosis and cytokine production and intercepting antibody-dependent cell-mediated cytotoxicity.³⁹⁻⁴¹ Patients with CIDP have lower than normal FcyRIIB expression on naïve B cells and monocytes but, after IVIG, the FcyRIIB expression is upregulated, coinciding with clinical improvement.^{27,39,41,42} This important effect, that also predicts response to IVIG in CIDP,³⁹ is not applicable to IgG4-ND because IgG4 antibodies cannot bind to inhibitory FcyRIIB receptor, as mentioned earlier (#4, Figure).
- 5. Suppression of pathogenic cytokines and immunoinflammatory molecules. IVIg effectively suppresses proinflammatory cytokines,^{27,28,41} but such cytokines are irrelevant in IgG4-ND because of the overall antiinflammatory effects of IgG4 that cannot recruit immune cells via Fc receptors or induce phagocytosis and trigger tissue inflammation (#5, Figure).

Rituximab Is the Preferred Treatment in IgG-4-ND

The expansion of activated B cells that correlate strongly with disease activity, the evidence of a pathogenic role for selfreactive B cells, and the clinical efficacy of B cell-depletion therapies¹⁻³ strongly support rituximab as the treatment of choice. Rituximab, a chimeric monoclonal antibody against CD20, a 297 AA membrane-associated phosphoprotein present on all B cells, except stem cells, pro-B cells, and plasma cells, depletes circulating B cells but not bone marrow and lymph node B cells.⁴³ Immunoglobulins are produced by long-lived plasma cells, but IgG4 are likely produced by CD20-positive short-lived plasma cells under the influence of IL-4 and IL-21.44 Rituximab eliminates B cells before they differentiate into plasma cells having no effect on non-CD20expressing plasma cells; it reduces, however, IgG4 levels by targeting IgG4-producing CD20-positive short-lived plasma cells and their related CD20⁺ precursors, resulting in steep decline of plasmablast counts.⁴⁵ In several nonrandomized trials in IgG4-RD, rituximab rapidly reduced serum IgG4 levels compared with serum IgG1.¹⁻³

In IgG4-ND, the success of rituximab is also impressive. In MuSK-MG, in a multicenter, blinded, prospective review, 58% (14/24) rituximab-receiving patients reached the primary outcome compared with 16% (5/31) of controls (p = 0.002), after a median follow-up of 3.5 years; furthernore, 29% of rituximab-treated patients required a mean-prednisone dose of 4.5 mg/d compared with 13 mg/d required by 74% of controls (p = 0.005).⁴⁶ IgG4-MuSK antibodies were markedly reduced 2–7 months after rituximab, being even undetectable within 2 years coinciding with clinical remission and sustained improvement for several years^{47,48}; in 1 patient who did not respond, MuSK-IgG4 antibodies remained unchanged, supporting the view that short-lived antibody-secreting CD20+- cells are the main producers of MuSK antibodies.⁴⁷

In neurofascin-155 and CASPR1/CNTN1 CIDP, the evidence of IVIg unresponsiveness is overwhelming with less than 10% partially and transiently responding to IVIg but more than 80% responding to rituximab, as confirmed in a recent large patient collection.²⁵ Similar is the experience in LGI1 and CASPR2-associated encephalitis, where in a large series the patients with worse outcome were those more frequently treated with both IVIg and steroids; however, the noted increased LGI1-specific plasmablasts/plasma cells in these patients' CSF fully justifies anti-CD19-specific immunotherapies.⁴⁹

Rituximab Maintenance

In IgG4-RD, the risk of relapse tended to be lower with rituximab maintenance, compared with 1 rituximab induction therapy with elevated serum IgG4 considered a risk factor for relapse.^{1,3,50} The value of circulating IgG4 as a biomarker in IgG4-RD is however unclear because IgG4 alone can nonspecifically increase in chronic immune activation.¹ The IgG4 levels also seem irrelevant in IgG4-ND; in 39 patients with CIDP, including several with IgG4-nodal antibodies, the IgG4 concentration was normal (<1,350 mg/L, the limit for active IgG4-RD [Dalakas et al. unpublished observations]). In contrast, IgG4-ND antibody-specific titers seem to correlate with disease activity as shown in MuSK-MG⁴⁷ but also in LGI1-encephalitis, where high CSF IgG4 titers strongly correlate with worse outcome.⁴⁹ The need for follow-up rituximab infusions remains still empirical. Some prefer a repeated infusion when clinical relapse occurs; others use 2 g every 6 months or 1 g every 3 months to ensure stability. 43,47,48,51 The most promising marker remains the re-emergence of CD27 + memory B cells^{43,52,53}; in 1 study, no MG relapses occurred when the CD27 + memory B cells were below the therapeutic target, whereas their resurgence was associated with clinical relapses.⁵² Because reduction of IgG4-MuSK antibodies coincides with clinical remission,⁴⁷ the value of follow-up antibody titers as a disease activity biomarker needs to be assessed in all IgG-4 ND, as discussed below, along with the reappearance of memory B cells and IgG4-producing CD20positive short-lived plasma cells.

Future Therapies in IgG4-ND

Some of the novel therapeutic approaches targeting B cells or CD4⁺ CTLs, currently evaluated in IgG4-RD, are also applicable to IgG4-ND and include (1) anti-CD19/20 including rituximab, inebilizumab—now approved for NMO-SD⁵⁴ and obexelimab (XmAb5871) all in phase-3 trials in IgG4-RD. Obexelimab, targeting CD19/FcyRIIB, is especially promising because it binds simultaneously to CD19 and FcyRIIb, promoting internalization of CD19 in the lipid rafts⁵⁵; obexelimab markedly enhances the inhibitory FcyRIIB and downregulates CD19, both effects tailored to IgG-4-ND by also activating FcyRIIB receptor; (2) dupilumab against IL-4Ra; (3) zanubrutinib and rilzabrutinib, both oral and well tolerated Bruton's tyrosine kinase inhibitors, now in phase 2 trials in pemphigus vulgaris and multiple sclerosis^{1,56} Because Bruton tyrosine kinase is an enzyme expressed on B lymphocytes and myeloid cells, its irreversible inhibition by these agents suppresses B cell activation, relevant to IgG4-ND; and (4) elotuzumab, a monoclonal anti-SLAMF7 antibody ready to begin¹; elotuzumab is a rational therapy for IgG-4 diseases because targets key cellular interactions between activated B cell subsets, plasmablasts, and CD4⁺ CTLs that all express SLAMF7.¹

Conclusions on the Clinical Importance of IgG4-ND, the Functions of IgG4 as Related to Immunotherapies, and the Potential Role of IgG4-Antibody Titers as Disease Biomarkers

The purpose of this article is to increase awareness and highlight the progress made in our understanding of the unique function of IgG4-antibodies and their associations with specific neurologic diseases which, in contrast to their IgG1 counterparts, have a more severe clinical phenotype and distinct response to immunotherapies. Unlike the other IgG isotypes, the IgG4 antibodies do not activate cellular or complement-mediated immune responses, which are directly targeted by IVIg and conventional immunotherapies, but they exert pathogenicity by blocking protein-protein interactions and signal transduction. As a result, IgG4-ND do not respond to IVIg like their IgG1 counterparts but respond impressively well to anti-B cell therapies which, if initiated early in the disease course, may ensure faster recovery preventing long-term disabilities. Importantly, there is convincing evidence that IgG-4 specific antibody titers are reduced in remissions and increased in exacerbations, having the potential to serve as reliable disease biomarkers not only in future controlled trials but also in current clinical practice. Because IgG4 antibody titers seem to correlate with the clinical status within the same patient,²⁵ it may serve as a monitoring tool not only in assessing disease activity and treatment response but also in guiding the need for the next anti–B cell infusion therapy. In some patients, IgG4 antibody titers decrease more than 90% or even become negative after rituximab, as noted in MuSK-MG47 and in some autoimmune nodopathy patients,²⁵ suggesting that antibody reappearance or significant titer increase may precede a relapse and guide the need for retreatment. If proven in controlled studies that IgG4 antibody titers are a solid biomarker for long-term therapy, it will be a remarkably novel observation in autoimmune neurotherapeutics. Currently, in IgG-1 antibody-mediated chronic autoimmune neurologic diseases, antibody titers are not reliable biomarkers, as observed with anti-MAG and antiganglioside antibodies in neuropathies,⁵³ anti-AChR antibodies in MG, anti-HMGCR or SRP antibodies in inflammatory myopathies, or anti-GAD antibodies in stiff-person syndrome⁵⁷; even anti-MOG and AQP-4 antibodies, although helpful, are not consistent predictors of disease relapse.

Study Funding

The author reports no targeted funding.

Disclosure

M. Dalakas is an Associate Editor for Neurology: Neuroimmunology & Neuroinflammation, Neurodiem, and Therapeutic Advances in Neurology; is on the editorial board for Acta Myolgica, Acta Neurologica Scandinavica, and Neuro-Therapeutics; is a commentator for Clinical Practice Elsevier; serves on the CIDP DSMB for Octapharma; has consulted for Grifols, Octapharma, Dysimmune Diseases Foundation, Takeda, Alexion and Argenx; has received Institutional support to Thomas Jefferson University, Neurology department or to Neuroimmunology Unit, University of Athens Medical School for research and education from: Merck-Serono, Novartis, Guillain-Barre/CIDP Foundation, Dysimmune Diseases Foundation, InfuCare, and Newfactor. Go to Neurology.org/NN for full disclosures.

Publication History

Received by Neurology: Neuroimmunology & Neuroinflammation August 23, 2021. Accepted in final form October 8, 2021.

Appendix Author

Name	Location	Contribution
Marinos	Thomas Jefferson University,	Drafting/revision of the
C.	Philadelphia, PA; National and	manuscript for content,
Dalakas,	Kapodistrian University of	including medical writing
MD	Athens Medical School, Greece	for content

References

- Perugino CA. Stone JH IgG4- related disease: an update on pathophysiology and implications for clinical care. Nat Rev Rheumatol. 2020;16(12):702-714.
- Koneczny IA. New classification system for igG4 autoantibodies front. ImmunolA new classification system for IgG4 autoantibodies. *Front Immunol.* 2018;9:97.
- Yamamoto M. B cell targeted therapy for immunoglobulin G4- related disease. Immunological Med. 2021:1-7. doi: 10.1080/25785826.2021.1886630.
- Koneczny I, Cossins J, Waters P, Beeson D, Vincent A. MuSK myasthenia gravis IgG4 disrupts the interaction of LRP4 with MuSK but both IgG4 and IgG1-3 can disperse preformed agrin-independent AChR clusters. *PLoS One*. 2013;8(11):e80695.
- Huijbers MG, Querol LA, Niks EH, et al. The expanding field of IgG4-mediated neurological autoimmune disorders. *Eur J Neurol.* 2015;22(8):1151-1161.
- Querol L, Nogales-Gadea G, Rojas-Garcia R, et al. Neurofascin IgG4 antibodies in CIDP associate with disabling tremor and poor response to IVIg. *Neurology*. 2014; 82(10):879-886.
- Lai M, Huijbers MG, Lancaster E, et al. Investigation of LGI1 as the antigen in limbic encephalitis previously attributed to potassium channels: a case series. *Lancet Neurol*. 2010;9(8):776-785.
- Dalmau J, Geis C, Graus F. Autoantibodies to synaptic receptors and neuronal cell surface proteins in autoimmune diseases of the central nervous system. *Physiol Rev.* 2017; 97(2):839-887.
- Bastiaansen AEM, van Sonderen A, Titulaer MJ. Autoimmune encephalitis with antileucine-rich glioma-inactivated 1 or anti-contactin-associated protein-like 2 antibodies (formerly called voltage-gated potassium channel-complex antibodies). *Curr Opin Neurol.* 2017;30(3):302-309.
- Gaig C, Graus F, Compta Y, et al. Clinical manifestations of the anti-IgLON5 disease. Neurology. 2017;88(18):1736-1743.
- van der Neut KM, Schuurman J, Losen M, et al. Antiinflammatory activity of human IgG4 antibodies by dynamic Fab arm exchange. *Science*. 2007;317(5844):1554-1557.
- Akiyama M, Suzuki K, Yamaoka K, et al. Number of circulating follicular helper 2 T cells correlates with IgG4 and interleukin-4 levels and plasmablast numbers in IgG4related disease. *Arthritis Rheumatol.* 2015;67(9):2476-2481.
- Morita R, Schmitt N, Bentebibel S, et al. Human blood CXCR5(fl)CD4(fl) T cells are counterparts of T follicular cells and contain specific subsets that differentially support antibody secretion. *Immunity*. 2011;34(1):108-121.
- Van De Veen W, Stanic B, Yaman G, et al. IgG4 production is confined to human IL-10-producing regulatory B cells that suppress antigen-specific immune responses. *J Allergy Clin Immunol.* 2013;131(4):1204-1212.
- Appeltshauser L, Brunder AM, Heinius A, et al. Antiparanodal antibodies and IgG subclasses in acute autoimmune neuropathy. *Neurol Neuroimmunol Neuroinflamm*. 2020;7(5):e817.
- Vidarsson G, Dekkers G, Rispens T. IgG subclasses and allotypes: from structure to effector functions. *Front Immunol.* 2014;5:520.
- Dalakas MC. Immunotherapy in myasthenia gravis in the era of biologics. Nat Rev Neurol. 2019;15(2):113-124.
- Evoli A, Tonali PA, Padua L, et al. Clinical correlates with anti-MuSK antibodies in generalized seronegative myasthenia gravis. *Brain*. 2003;126(pt 10):2304-2311.
- Phillips WD, Vincent A. Pathogenesis of myasthenia gravis: update on disease types, models, and mechanisms. F1000Res. 2016;5:1513.
- Querol L, Nogales-Gadea G, Rojas-Garcia R, et al. Antibodies to contactin-1 in chronic inflammatory demyelinating polyneuropathy. Ann Neurol. 2013;73(3):370-380.
- Cortese A, Lombardi R, Briani C, et al. Antibodies to neurofascin, contactin-1, and contactin-associated protein 1 in CIDP: clinical relevance of IgG isotype. *Neurol Neuroimmunol Neuroinflamm.* 2020;7(1):e639.
- Stathopoulos P, Alexopoulos H, Dalakas MC. Autoimmune antigenic targets at the node of Ranvier in demyelinating disorders. Nat Rev Neurol. 2015;11(3):143-156.
- Manso C, Querol L, Mekaouche M, Illa I, Devaux JJ. Contactin-1 IgG4 antibodies cause paranode dismantling and conduction defects. *Brain*. 2016;139(pt 6):1700-1712.
- 24. Manso C, Querol I, Lleixa C, et al. Anti-neurofascin-155 lgG4 antibodies prevent paranodal complex formation in vivo. *J Clin Invest*. 2019;129(6):2222-2236.
- Martín-Aguilar L, Lleixà C, Pascual-Goñi E, et al. Clinical and laboratory features in anti-NF155 autoimmune nodopathy. *Neurol Neuroimmunol Neuroinflamm* 2022;9(1): e1098.
- Landa J, Gaig C, Plagumà J, et al. Effects of IgLON5 antibodies on neuronal cytoskeleton: a link between autoimmunity and neurodegeneration. *Ann Neurol.* 2020; 88(5):1023-1027.
- Dalakas MC. Mechanism of action of IVIg and therapeutic considerations in the treatment of acute and chronic demyelinating neuropathies. *Neurology*. 2002;59(12 suppl 6):S13-S21.

- Dalakas MC. Intravenous immunoglobulin in autoimmune neuromuscular diseases. JAMA. 2004;291(19):2367-2375.
- Roux KH, Tankersley DL. A view of the human idiotypic repertoire. Electron microscopic and immunologic analyses of spontaneous idiotype-anti-idiotype dimers in pooled human IgG. J Immunol. 1990;144(4):1387-1395.
- Dietrich G, Kazatchkine MD. Normal immunoglobulin G (IgG) for therapeutic use (intravenous Ig) contains anti-idiotypic specificities against an immunodominant, disease-associated, cross-reactive idiotype of human anti-thyroglobulin autoantibodies. J Clin Invest. 1990;85(3):620-629.
- Buchwald B, Ahangari R, Weishaupt A, Toyka KV. Intravenous immunoglobulins neutralize blocking antibodies in Guillain-Barré syndrome. *Ann Neurol.* 2002;51(6): 673-680.
- Jacobs BC, O'Hanlon GM. Bullens RWM et al mmunoglobulins inhibit pathophysiological effects of anti-GQ1b-positive sera at motor nerve terminals through inhibition of antibody binding. *Brain*. 2003;126(pt 10):2220-2234.
- Dalakas MC, Spaeth PJ. The importance of FcRn in Neuro-immunotherapies: from IgG catabolism, FCGRT gene polymorphisms, IVIg dosing and efficiency to specific FcRn inhibitors. *Ther Adv Neurol Disord*. 2021;14:1756286421997381.
- Yu Z, Lennon VA. Mechanism of intravenous immune globulin therapy in antibodymediated autoimmune diseases. N Engl J Med. 1999;340(3):227-228.
- Li N, Zhao M, Hilario-Vargas J, et al. Complete FcRn dependence for intravenous Ig therapy in autoimmune skin blistering diseases. J Clin Invest. 2005;115(12):3440-3450.
- Basta M, Dalakas MC. High-dose intravenous immunoglobulin exerts its beneficial effect in patients with dermatomyositis by blocking endomysial deposition of activated complement fragments. J Clin Invest. 1994;94(5):1729-1735.
- Dalakas MC, Illa I, Dambrosia JM, et al. A controlled trial of high-dose intravenous immunoglobulin infusions as treatment for dermatomyositis. N Engl J Med. 1993; 329(27):1993-2000.
- Malik U, Oleksowicz L, Latov N, Cardo LJ. Intravenous y-globulin inhibits binding of anti-GMI to its target antigen. Ann Neurol. 1996;39(1):136-139.
- Tackenberg B, Jelcic I, Baerenwaldt A, et al. Impaired inhibitory Fcgamma receptor IIB expression on B cells in chronic inflammatory demyelinating polyneuropathy. Proc Natl Acad Sci U S A. 2009;106(12):4788-4792.
- Anthony RM, Nimmerjahn F, Ashline DJ, Reinhold VN, Paulson JC, Ravetch JV. Recapitulation of IVIG anti-inflammatory activity with a recombinant IgG Fc. Science. 2008;320(5874):373-376.
- Lunneman JD, Nimmerjahn F, Dalakas MC. Intravenous immunoglobulin in neurology: mode of action and clinical efficacy. Nat Rev Neurol. 2015;11(2):80-89.
- Quast I, Keller CW, Maurer MA, et al. Sialylation of IgG Fc domain impairs complement dependent cytotoxicity. J Clin Invest. 2015;125(11):4160-4170.
- Dalakas MC. B cells as therapeutic targets in autoimmune neurological disorders. Nat Clin Pract Neurol. 2008;4(10):557-567.
- Unger PA, Lighaam LC, Vermeulen E, et al. Divergent chemokine receptor expression and the consequence for human IgG4 B cell responses. *Eur J Immunol.* 2020;50(8):1113-1125.
- Wallace ZS, Mattoo H, Mahajan VS, et al. Predictors of disease relapse in IgG4-related disease following rituximab. *Rheumatology (Oxford)*. 2016;55(6):1000-1008.
- Hehir MK, Hobson-Webb LD, Benatar M. Rituximab as treatment for anti-MuSK myasthenia gravis: Multicenter blinded prospective review. *Neurology*. 2017;89(10): 1069-1077.
- Mariapaola Marino M, Basile U, Spagni G, et al. Long-lasting rituximab-induced reduction of specific—but not total—IgG4 in MuSK-positive myasthenia gravis. Front Immunol. 2020;11:613.
- Dalakas MC. Progress in the therapy of myasthenia gravis: getting closer to effective targeted immunotherapies. *Curr Opin Neurol.* 2020;33(5):545-552.
- Gadoth A, Zekeridou A, Klein CJ, et al. Elevated LGI1-IgG CSF index predicts worse neurological outcome. Ann Clin Transl Neurol. 2018;5(5):646-650.
- Majumder S, Mohapatra S, Lennon RJ, et al. Rituximab maintenance therapy reduces rate of relapse of pancreaticobiliary immunoglobulin G4-related disease. *Clin Gastroenterol Hepatol.* 2018;16(12):1947-1953.
- 51. Dalakas MC. Inhibition of B cell functions: implications for neurology. *Neurology*. 2008;70(23):2252-2260.
- Lebrun C, Bourg V, Bresch S, et al. Therapeutic target of memory B cells depletion helps to tailor administration frequency of rituximab in myasthenia gravis. J Neuroimmunol. 2016;298:79-81.
- Dalakas MC, Rakocevic G, Salajegheh M, et al. Placebo-controlled trial of rituximab in IgM anti-myelin-associated glycoprotein antibody demyelinating neuropathy. *Ann Neurol.* 2009;65(3):286-293.
- Cree BAC, BennettJL, KimHJ, et al. Inebilizumab for the treatment of neuromyelitis optica spectrum disorder(N- MOmentum): a double- blind, randomized placebocontrolled phase 2/3 trial. *Lancet.* 2019;394(10206):1352-1363.
- Alexopoulos H, Biba A, Dalakas MC. Anti–B-Cell therapies in autoimmune neurological diseases: rationale and efficacy trials. *Neurotherapeutics*. 2016;13(1):20-33.
- Reich DS, Arnold DL, Vermersch P, et al Safety and efficacy of tolebrutinib, an oral brain-penetrant BTK inhibitor, in relapsing multiple sclerosis: a phase 2b, randomised, double-blind, placebo-controlled trial. *Lancet Neurol.* 2021;20(9): 729-738.
- Dalakas MC, Fujii M, Li M, et al. High-dose intravenous immunoglobulin for stiffperson syndrome. N Engl J Med. 2001;345(26):1870-1876.
- Dalakas MC. Update on intravenous immunoglobulin in neurology: modulating neuro-autoimmunity, evolving factors on efficacy and dosing and challenges on stopping chronic IVIg therapy. *Neurotherapeutics*. Accessed November 21, 2021. https://doi.org/10.1007/s13311-021-01108-4.

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Marinos C. Dalakas Neurol Neuroimmunol Neuroinflamm 2022;9; DOI 10.1212/NXI.000000000001116

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