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Effective Dose Fractionation Schemes of Radiotherapy for Prostate Cancer

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Abstract

Radiation therapy remains as one of the main cancer treatment modalities. Typical regimens for radiotherapy comprise a constant dose administered on weekdays, and no radiation on weekends. In this paper, we examine adaptive dosages of radiation treatment strategies for heterogeneous tumors using a dynamical system model that consist of radiation-resistant and parental populations with unique interactive properties, namely, PC3 and DU145 prostate cancer cell lines. We show that stronger doses of radiation given in longer time intervals, while keeping the overall dosage the same, are effective in PC3 cell lines, but not in DU145 cell lines. In addition, we tested an adaptive dosing schedule by administering a stronger dosage on Friday to compensate for the treatment-off period during the weekend, which was effective in decreasing the final tumor volume of both cell lines. This result creates interesting possibilities for new radiotherapy strategies at clinics that cannot provide treatment on weekends.

Keywords: mathematical oncology, tumor growth, radiotherapy, dose fractionation, lotka-volterra model

1 Introduction

Radiotherapy is one of the predominant cancer treatment modalities. Approximately 50% of all cancer patients receive radiotherapy treatments during the course of their illness [11, 3], and it is estimated that radiation therapy contributes around 40% towards curative treatment [2]. It is also a highly cost effective single modality treatment, accounting for only about 5% of the total cost of cancer care [28]. Furthermore, advances in imaging techniques, computerized treatment planning systems, radiation treatment machines with improved X-ray production and treatment delivery, as well as improved understanding of the radiobiology are all increasing the impact and importance of radiotherapy [4].

In clinics, the dosing schedule of radiotherapy is standardized using daily doses of 1.8 to 2.0 for 39–45 fractions [33]. This is called fractionation, where the total dosage is divided into smaller doses that are given over a period of one to two months. The specific dose for each patient depends on the location and severity of the tumor, as well as the radiation-induced toxicity of normal tissues surrounding the tumor. Various studies focus on dose alteration to improve radiotherapy outcome, including hypofractionation and hyperfractionation [26]. Hypofractionation is a treatment regimen that uses higher doses of radiation in fewer visits to lower the effects of accelerated tumor

growth that typically occurs during the later stages of radiotherapy. On the other hand, hyperfractionation is a strategy dividing the same total dose into more frequent deliveries, for instance, radiation doses given more than once a day. A recent study in [37] shows that increasing the total dosage to 80 Gy, that corresponds to a biologically equivalent dose of 200 Gy for prostate cancer is associated with improved outcome, but doses above that level did not result in additional clinical benefit. Moreover, population-based research revealed an association between overall survival of prostate cancer patients in doses over 75.6 Gy [33]. However, the dosage schemes that can be tested in clinical trials are very limited. Simulation using in-silico models can help address this limitation to test various dosing schedules without the concern of toxicity to patients.

A large number of mathematical and computational models have been developed to study tumor growth and cancer treatments, including differential equations [24, 19, 8], multiphase models based on mixture theory [6, 31] or phase field theory [30], and multiscale models that couple subcellular, cellular, and tissue scale phenomena [18]. The availability of detailed information about tumors has undoubtedly stimulated this field to more complex models, although as the model complexity increases, it becomes more challenging to uniquely identify the model parameter values [7]. In particular, often in the clinical setting, it is not possible to collect appropriate amounts of patients' data to calibrate complex models. Hence, simple

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models such as ordinary differential equations (ODEs) are commonly used when dealing with clinical data. See [12, 9, 36] for recent literature using ODE models to calibrate data of radiotherapy treated tumors as in our study.

Radiotherapy also has a long history of mathematical modeling. Radiation dosing is typically modeled using the linear-quadratic (L-Q) model [32, 13]. Several recent studies have applied the L-Q model to patient-specific data, in an effort to evaluate and predict individual responses to radiotherapy [29, 10, 31, 12]. Logistic type of radiotherapy response has been proposed with a concept of proliferation saturation index, defined as the ratio of tumor volume to the host-influenced tumor carrying capacity, that correlates inversely with radiotherapy response [27, 25]. Other radiotherapy models, including those assuming a dynamic carrying capacity, have been developed to more accurately calibrate and predict individual patient response to radiotherapy [36].

Here, we investigate the effect of different radiotherapy regimens on the growth of two types of heterogeneous prostate tumors comprising radiation-sensitive (or parental) populations and resistant populations that interact with one another. Although adaptive radiotherapy and its potential clinical benefits have been proposed in the clinical community since 1997 [35], it has not been a common practice in the clinics until now [5]. However, recent improvements in imaging technologies along with mathematical modeling have been proclaimed to allow us new opportunities toward patient-specific adaptive radiation therapy [12]. Our work is in accordance with this idea of using mathematical models to guide radiation dose planning, which has been done for various types of tumors, such as glioblastomas [22] and lung cancer [17]. The contribution of our study is to test the potential of adaptive radiotherapy *in silico* for two types of prostate cancer studied in [18]. The paper is structured as follows. Section 2 summarizes the mathematical model that is used to describe the growth of heterogeneous cancer and its response to radiotherapy with biological interpretations of the model parameters. We also describe how radiotherapy fractionation treatments are incorporated into the model in order to properly simulate cell death due to radiotherapy. In Section 3 we explore two different scenarios on dose fractionation. Section 3.1 studies a constant dosage treatment, while we change the dosage and time interval between the administration of radiation. The overall dosage is kept constant. In section 3.2, we study a strategy to overcome the clinical radiotherapy schedule with no radiotherapy treatments being administered on weekends. We compare the strategy of changing the dosage to be more heavily concentrated on Fridays at the end of the week to a dosage that is spread out evenly. Section 4 concludes our paper and summarizes our key results. In addition, it includes our proposed fractiona-

tion scheme that incorporates our findings into a possible dosage plan.

2 Mathematical Model of Cancer Growth and Radiotherapy

The Lotka-Volterra model is one of the typical approaches to describe the interactions between multiple types of cancer cells [15, 38, 14, 23, 18, 20]. We also employ the Lotka-Volterra model to describe the growth of mixtures consisting of parental and radioresistant tumor cell populations. The equation tracks the dynamics of $V_c(t)$, which is the volume of the parental tumor population, or the “control”, in other words, the radiation-sensitive population, and $V_r(t)$, which is the volume of the radiation-resistant tumor population.

$$\begin{aligned}\frac{dV_c}{dt} &= p_c V_c \left(1 - \frac{V_c}{K_c} - \lambda_r \frac{V_r}{K_c} \right), \\ \frac{dV_r}{dt} &= p_r V_r \left(1 - \frac{V_r}{K_r} - \lambda_c \frac{V_c}{K_r} \right).\end{aligned}\tag{1}$$

The parameters used in the Lotka-Volterra model each have their own biological function. The rate of growth of the control population is p_c , and the rate of growth of the radio-resistant population is p_r . K_c is the carrying capacity of the control population, the maximum volume to which the radio-sensitive or parental population is limited to, and K_r is the carrying capacity of the radioresistant population. In addition, λ_c and λ_r model the interaction between the two populations, where λ_c describes the effect that the parental cell population has on radioresistant cells, and λ_r describes the effect that the radioresistant cell population has on the parental cells. We note that $-1 \leq \lambda_c, \lambda_r \leq 1$. The signs of the interaction parameters λ_c and λ_r need not be equivalent. A positive λ_c parameter represents a detrimental effect on V_r , or the volume of the resistant tumor population, and a negative λ_c parameter represents a beneficial effect on V_r . The same is true for the effects of λ_r on V_c . If both interaction parameters are positive then this represents a competitive interaction between the two tumor populations, and if both are negative then this represents a mutualistic interaction. One positive and one negative interaction parameters signify that one tumor population exerts a positive effect on the other while the latter exerts a negative effect on the former tumor population, a so-called antagonistic interaction. An interaction parameter of 0 represents no effect on the other tumor population. All model parameters and their biological interpretations are summarized in Table 1.

We now explain how we incorporate treatment with radiotherapy in the cancer model Eq. (1). We consider

Table 1: Model parameters and their biological interpretation.

Parameter	Biological meaning
$V_c(t)$	Volume of the control (radiation-sensitive/parental) tumor population at time t
$V_r(t)$	Volume of the radiation-resistant tumor population at time t
p_c	Rate of growth of the control population
p_r	Rate of growth of the radio-resistant population
K_c	Carrying capacity of the control population
K_r	Carrying capacity of the radio-resistant population
λ_c	Effect of the control cell population on the radio-resistant cells
λ_r	Effect of the radio-resistant cell population on the control cells
α_c, β_c	Radio-sensitivity parameter of control cells
α_r, β_r	Radio-sensitivity parameter of radio-resistant cells

Table 2: A summary of the parameter values used in the simulation for prostate cancer cell lines, PC3 and DU145. The pre-treatment parameters p , K , and λ are taken from [18], provided in supplementary Table S1 and Figure 2d of [18]. The radiotherapy parameters α and β are estimated from data in supplementary Figure S1b in [18].

Parameter	Cell line		Units
	PC3	DU145	
p_c	0.36	0.6	day ⁻¹
p_r	0.48	0.36	day ⁻¹
K_c	0.85	0.75	mm ³
K_r	2.0	1.4	mm ³
λ_c	0.2	0.25	day ⁻¹ mm ⁻³
λ_r	0.0	-0.5	day ⁻¹ mm ⁻³
α_c	0.43	0.2843	Gy ⁻¹
α_r	0.3	0.23	Gy ⁻¹
β_c	0.0407	0.0161	Gy ⁻²
β_r	0.0402	0.0124	Gy ⁻²

a typical tumor treatment regimen in which daily doses of d Gy are administered Monday through Friday for 6 weeks. We use the linear-quadratic model [16] to account for the effect of radiotherapy. This model assumes that the fraction of cells that survive exposure to a dose d of radiotherapy is given by

$$\text{Survival fraction} = e^{-\alpha d - \beta d^2}, \tag{2}$$

where α and β are tissue specific radiosensitivity parameters that model single and double strand breaks of the DNA [21]. We assume that the effect of radiotherapy is instantaneous, with the non-surviving cell fraction immediately removed when therapy is administered. In particular, we denote the dosing schedule by $u(t)$ as the following summation of indicator functions,

$$u(t) = \sum_{i=1}^N \delta(t - t_i), \tag{3}$$

where $\delta(t)$ is a Dirac-delta function that is $\delta(0) = 1$ and zero elsewhere. This makes $u(t)$ to be one only at t_i (for $i = 1, 2, \dots, N$), the times at which radiotherapy is delivered. As we combine the radiotherapy model with the radiosensitivity cancer growth model, we have

$$\frac{dV_c}{dt} = p_c V_c \left(1 - \frac{V_c}{K_c} - \lambda_r \frac{V_r}{K_c} \right) - (1 - e^{-\alpha_c d - \beta_c d^2}) u(t) V_c, \tag{4}$$

$$\frac{dV_r}{dt} = p_r V_r \left(1 - \frac{V_r}{K_r} - \lambda_c \frac{V_c}{K_r} \right) - \underbrace{(1 - e^{-\alpha_r d - \beta_r d^2}) u(t) V_r}_{\text{cell death due to RT}}. \tag{5}$$

Here, α_c and β_c are radiosensitivity parameters for the control cells, and α_r and β_r are for the resistant cells.

3 Simulation of Radiotherapy Dosage Fractionation

These simulations will investigate dosage strategies in two separate prostate cancer cell lines, PC3 and DU145. Important to note when discussing each cell lines' response to radiation, carrying capacity, and in the interaction parameters between control and radio-resistant cell populations. For instance, regarding the interaction between the control and radio-resistant population, PC3 has competitive interaction, while the control population of DU145 has antagonistic relation to the radio-resistant population and only the control population benefits from coexistence. Also, regarding sensitivity to radiotherapy, PC3 is more sensitive to the treatment compared to DU145. See Table 2 for the parameter values.

In our simulations, we consider a period of 6 weeks of radiotherapy treatment with an initial 2 week cancer growth period. The dosage plans that we study are described in each section in detail, but see Figure 1 for some examples and a brief summary. The initial condition for the model is taken as $V_c(0) = 0.5\text{mm}^3$, and $V_r(0) = 0.5\text{mm}^3$, assuming the parental and resistant tumor cell lines initially in a 1:1 ratio consistent with conventional experimental methods [18]. Although we test only for 1:1 ratio, we report on the control and resistant volume separately to examine the efficacy of treatment on each population.

In order to best measure the efficacy of radiation treatment, we decided to quantify two basic features of each simulation as a metric to determine the magnitude of influence each of our dosage strategies had on the growth of the simulated tumor population. Those are tumor volume and the tumor volume integrated in time, or area under the curve. While tumor volume is the most obvious metric to follow in order to measure the efficacy of simulated treatment, the area under the curve is also an important metric to follow because it represents the tumor volume integrated over the treatment period, which gives you an idea of the overall burden placed on the patient during that period of time.

3.1 Comparison of constant dosage strategies with different dose levels

In this section, we study radiotherapy schedules in which the dosage value and time interval remain constant over the six-week treatment period. In particular, we compare different dosage levels, from 1 Gy to 4 Gy, while increasing the time interval between each administered dose for stronger dosages to keep the total dose at the end of 6 weeks constant, at 42 Gy in total. The case of

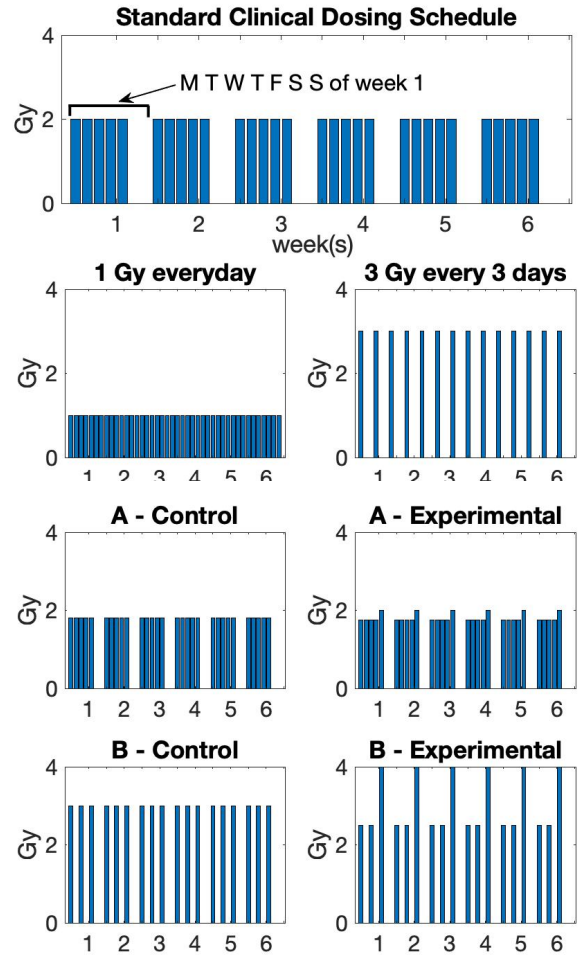


Figure 1: Examples of radiotherapy dosage schedules tested in this study. The clinical standard dosage plan for prostate cancer is 2 Gy on weekdays, Monday to Friday, for 6 weeks (top). The middle figures show examples of constant dosage schedules studied in section 3.1, e.g., 1 Gy every day plan (middle, left) and 3 Gy every 3 days plan (middle, right). The bottom figures show the dosage schedules tested in section 3.2, e.g., Control schedules using constant dosage (bottom, left) and Experimental schedules using stronger dosage on Friday (bottom, right).

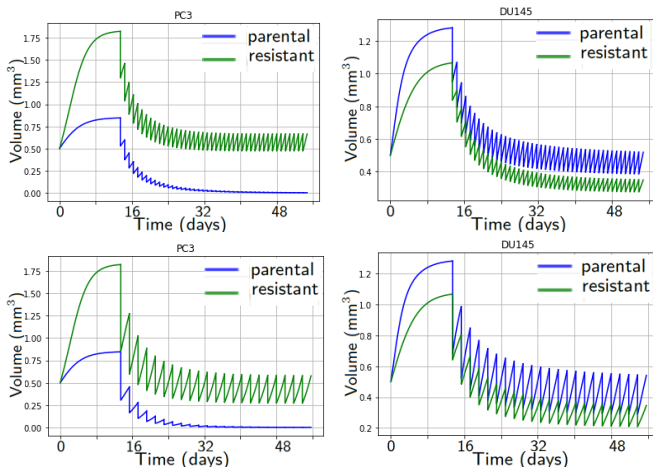


Figure 2: Comparison of 1 Gy administered every 1 day (top) versus 2 Gy administered every 2 days (bottom) to a tumor model initiated as a 1:1 mixture of parental and resistant tumor cell populations. The results show the volumes for the individual parental (blue) and resistant (green) populations from the PC3 cell line (left) and DU145 cell line (right). When subjected to this change, a 30% decrease in average tumor volume can be seen in the PC3 cell line when comparing the 1 Gy schedule and 2 Gy schedule. This change in average tumor size is not as noticeable in the DU145 cell line, as average tumor size remains unaffected, but a larger variation of values is noticed. See Tables 3–4 and Figure 3 for further comparison.

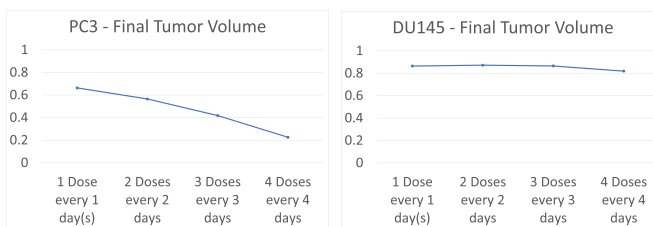


Figure 3: A line graph representing total final tumor volume for the PC3 cell line (left) based on values from Table 3 as well as the DU145 cell line (right) based on values in Table 4. Increasing both dosage values and the time intervals in between dosages will not change the overall dosage, so results are dependent on hypofractionation and not an increased overall dosage. This strategy provides promising results for the PC3 cell line, but not so much for the DU145 cell line.

4 Gy has a lower total dosage due to the 6 week time constraint, but still continues the observed trend even at 40 Gy total. This strategy is called hypofractionation.

In Tables 3 and 4, we can compare the results generated by ramping up dosage values as well as increasing the time in between each administered dose. By organizing these events according to their respective dosage and time interval increase, we can show a clear trend in the data for the PC3 cell line, as shown in Table 3, in which a higher dose administered over a greater period of time gives better results than a lower dose administered over a shorter length of time. For example, 2 Gy administered every 2 days yields better results than 1 Gy administered every 1 day. As an example, in Figure 2, we show the tumor trajectory for the case of comparing a single (1 Gy) dose administered every 1 day versus a 2 Gy dose administered every 2 days in both cell lines. Figure 2 follows the basic parameters listed in Table 2. It is important to note that the PC3 and DU145 cell lines are defined by their own separate parameters and therefore present with varying changes after dosage administration. For example, the resistant population of tumor cells in the PC3 cell line have a higher carrying capacity, growth rate, and higher levels of radio-sensitivity than do resistant cells in the DU145 cell line. The same is true for parental cells in the DU145 cell line vs. parental cells in the PC3 cell line. This is why, despite it being seemingly counterintuitive, we see resistant cells end with a smaller volume than the more radiation-sensitive parental cells in the DU145 cell line. In this instance, one should consider the growth rate of the parental population and how it far exceeds that of the resistant cells.

In Figure 2 we can see significant decay for the PC3 cell line in both final tumor size and the area under the curve. In particular, the average tumor volume shows a 30% decrease when comparing the 1 Gy schedule and 2 Gy schedule. However, results did not prove to be as effective for the DU145 cell line, as shown in Table 4 and Figure 3, since only a slight decrease in the area under the curve is observable, and hardly any change can be noted from the final volume of the tumor. In particular, the parental DU145 cell line shows increasing final tumor volume as dosages become stronger. Therefore, out of the two cell lines, it would be safe to assume that this dosage strategy proves effective only for the PC3 cell line and not the DU145 cell line. We will further investigate the underlying mechanism that makes the contrast between the two cell lines in the future.

3.2 Comparison of uniform dosage versus stronger dosage on Fridays

Next, we study various scenarios in which no radiation would be administered over the weekend, to more closely

Table 3: PC3 Cell line: Final tumor volume after treatment and area under the curve following an increase in time interval with stronger dosage, while maintaining a constant overall dosage (i.e., under different hypofractionation strategies). A steep decrease in final tumor volume was observed, as well as a decrease in the area under the curve.

		Dosage (Gy)	1	2	3	4
		Time Interval (days)	1	2	3	4
Final Tumor Volume	Parental		0.0020287	0.00043197	0.000076458	0.0000069054
	Resistant		0.66080	0.56423	0.41890	0.22569
	Total		0.662823	0.56466	0.41808	0.22570
Area Under Curve	Parental		298.16	277.67	262.53	251.33
	Resistant		1046.4	911.99	791.61	694.12
	Total		1344.5	1189.7	1054.1	945.45

Table 4: DU145 Cell line: Final tumor volume after treatment and area under the curve following an increase in time interval with stronger dosage, while maintaining a constant overall dosage (i.e., under different hypofractionation strategies). While a decrease in the area under the curve can be noted, the final tumor volumes of parental and resistant populations yielded results with no obvious downward trend.

		Dosage (Gy)	1	2	3	4
		Time interval (days)	1	2	3	4
Final Tumor Volume	Parental		0.5162	0.53188	0.53996	0.526363
	Resistant		0.34821	0.34030	0.32530	0.29257
	Total		0.86444	0.87218	0.86526	0.81893
Area Under Curve	Parental		836.98	794.75	757.06	742.58
	Resistant		661.44	624.52	590.47	569.93
	Total		1498.4	1419.3	1347.5	1312.5

resemble a real-life scenario in which the radiotherapy clinic would be closed on the weekends, therefore unable to administer any radiation to a patient during this time frame. This weekend time frame allows for unchecked tumor growth for approximately 3 days until the next dose of radiation is administered. The standard radiation dosage strategy for most radiotherapy clinics would be a consistent administration of 2 Gy of radiation every weekday, and 0 Gy on weekends. However, instead of administering a constant radiation dose of 2 Gy every weekday, we decided to allocate more radiation at the end of the week (on Friday) at the expense of lowering the radiation dose for all of the days prior. The idea is to maintain the same total amount of radiation at the end of each week so as to not compromise the total amount of radiation the patient receives, however instead of maintaining a consistent dosage throughout, we can modify the administration of radiation so that more of it is administered at the end of each week in order to combat weekend tumor growth.

We considered two different scenarios (Scenario A and Scenario B) to test how a simulated tumor model might shift as we vary the dosage values and their respective time intervals. We also provide a variation on either scenario (Scenario A' and Scenario B') in order to con-

firm the consistency of a stronger Friday dose by changing the Friday dosage value. We compared the results from dosing every weekday (Scenarios A and A') versus dosing every Monday, Wednesday, and Friday (Scenarios B and B'). However, we integrated the aforementioned guiding principle in this investigation, i.e. stronger doses given at the end of each week on Fridays, to compensate for the weekend time intervals spent without radiotherapy. For example, in this investigation, we administered a higher dosage on Fridays than on any other weekday, and then we allowed the tumor to “re-grow” on the weekends to represent a lack of radiotherapy treatment given within that time frame. We used “end-of-week” values as our main metric for determining the effectiveness of each treatment modification, and each of these values are measured every Friday after radiation administration. Each scenario is as follows. We remark that the dosage schedule of scenarios A and B are plotted as bar graph in the bottom of Figure 1.

- Scenario A involved the simulated administration of 1.8 Gy every day for five days to total 9 Gy at the end of every week as our control group. In our experimental group this weekly 9 Gy value was maintained while varying daily dosage values. The dosage

value in our experimental group would increase from 1.75 Gy every day Monday–Thursday, to a slightly higher dose of 2 Gy every Friday. This sums to 9 Gy at the end of every week, same as the control group. Figures 5 and 6 depict the results of these changes in either of the PC3 or DU145 cell lines, which shows only a slight decrease in total tumor volume for the experimental group as a result of these changes. Tables A1 and A2 in the Appendix list parental and resistant end-of-week tumor volume sizes for the PC3 and DU145 cell lines in Scenario A, respectively.

- Scenario A' involved a similar process, administering 2 Gy every day for five days as our control, and 1.75 Gy Monday–Thursday and 3 Gy on Friday as our experimental schedule. We remark that the control case is one of the standard clinical treatment schedules. This totals 10 Gy at the end of every week. Similar results to Scenario A were observed, with a steady decrease in end-of-week tumor volume values. This change resulted in a much larger difference between experimental and control values than Scenario A, however, and A' more clearly exhibits the benefits of a stronger Friday dosage. See tables A5 and A6 in the Appendix for numerical results as well as Figure A1 for a graphical representation.

Scenarios B and B' consist of a variation from scenarios A and A', in which a dose is administered every Monday, Wednesday and Friday (skipping Tuesday and Thursday), still incorporating a stronger Friday dosage. The motivations behind this were to incorporate some of the findings from earlier on in the paper to this dosage strategy as well, that being the effectiveness of increasing the time interval in between doses while maintaining a constant overall dosage.

- Scenario B involved administering 3 Gy every Monday, Wednesday and Friday for our control. Our experimental regime included 2.5 Gy on Monday and Wednesday, and 4 Gy on Friday. This totals 9 Gy at the end of each week. These values show a promising decrease in end-of-week tumor volume size when concentrating a greater dose on Fridays. Figure 4 depicts the first two weeks of this scenario in both the control and experimental groups for either cell line as a side-by-side comparison. Figures 5 and 6 show the difference between control and experimental tumor volumes as a result of these changes in dosage for the PC3 and DU145 cell lines, respectively. See tables A3 and A4 in the Appendix for end-of-week values.
- Similarly to Scenario B, Scenario B' involved administering a certain dosage, 2.5 Gy, every Monday, Wednesday and Friday for our control. Our experimental schedule consists of 2 Gy on Monday and

Wednesday, and 3.5 Gy on Friday. This totals 7.5 Gy at the end of the week each week. Similar results to Scenario B were observed, with a steady decrease in end-of-week tumor volume values. See tables A7 and A8 in the Appendix for numerical results as well as Figure A2 for a graphical representation.

Scenarios A and A' show us that by concentrating a higher dosage at the end of the week while maintaining a consistent weekly dosage, we end up with a smaller tumor volume at the end of each week for both cell lines. Scenarios B and B' combined this idea with the previously discussed idea of allowing for more time in between larger doses. We reconfirm the effectiveness of concentrating a higher dosage on Friday compared to Monday and Wednesday in these scenarios. Smaller final tumor volumes are obtained in both cell lines. However, when we compare scenario A versus B, that is, smaller dosages every five days versus larger dosages on every other day, scenario B is more effective than scenario A only in the PC3 cell line, not in the DU145 cell line. This is consistent with the results obtained in section 3.1, hypofractionation is effective only in PC3 cell line, but not in DU145 cell line, even when combined with the stronger dosage on Friday strategy. We remark that increasing the total dosage using the same strategy always results in smaller tumor size. For instance, A' with total dosage 10 Gy is more effective than A with total dosage 9 Gy and B with total dosage 9 Gy is more effective than B' with total dosage 7.5 Gy.

4 Conclusion and Future Outlook

In this study, we sought to study the effect of different radiation dosing strategies on the growth of prostate tumor spheroids consisting of a heterogeneous mixture of parental and radiation-resistant populations. Our results show that the administration of a higher dose with a longer period of time in between doses gives more promising results for the PC3 cell line, but not necessarily for the DU145 cell line. However, shifting dosage values so that more of it is administered on Friday and less during the week yields a moderate decrease in tumor volume at the end of each week for both cell lines. Our results showed us that by simply shifting dosage administration values so that more of it is administered on Fridays, this will yield lower overall tumor volume outcomes the more you shift the dosage values, as seen in both cell lines in Scenarios A and B. Combining this shift in dosage values with hypofractionation strategies yields much more promising results in PC3 cell line, as noted in Scenarios B and B'. It is important to note that all tests were run in a simulated 1:1 ratio environment, and it would be insightful to look into other heterogeneous mixtures of

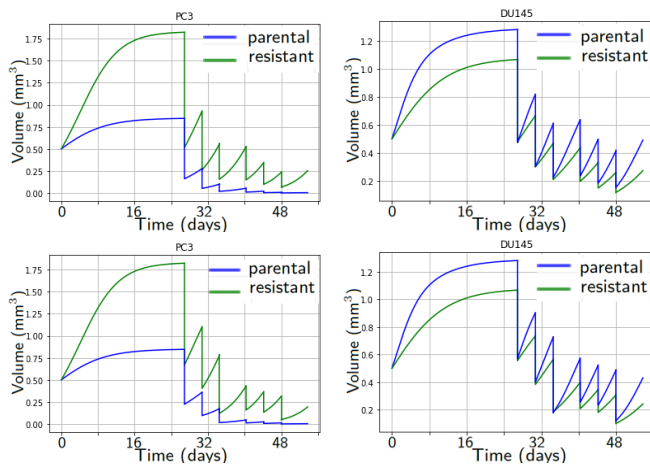


Figure 4: Scenario B - A comparison of 3 Gy administered every Monday, Wednesday, and Friday (Control, top) vs. 2.5 Gy given on Monday and Wednesday, and 4 Gy every Friday (Experimental, bottom), in a tumor initiated as a 1:1 mixture of parental (blue) and resistant green tumor cell populations. The results show the volumes for the individual parental and resistant populations from the PC3 cell line (left) and DU145 cell line (right). Only the first 2 weeks are depicted to show more detail. A modest decrease in values at the end of each week can be noted for both cell lines. A consistent dose will allow for more growth on the weekend, however a stronger Friday dose will yield a larger dip in the graph, increasing overall dosing effectiveness.

parental and radiation resistant tumor populations. It is also important to note that the significant difference in results from the PC3 and DU145 cell lines shows us the impact of the diversity of the tumor population itself on radiation treatment.

We conclude from our results that we can improve the treatment for a 1:1 PC3 cell line mixture using a combination of both methods tested. Many different types of treatments are possible in this manner, but one such strategy is proposed in Figure 7. In this scheme, 3 Gy of radiation is administered daily for the first week, with a 4 Gy dose administered on the first Friday. Afterward a constant dose of 2 Gy is administered every Monday, Wednesday, and Friday for 5 weeks, with a final dose of 3 Gy administered at the end of the 6-week time period. No radiation is administered on any weekend. This sample combination strategy assimilates both ideas described in this paper; such that when the tumor is at its largest we may give a stronger overall dose at the first week with a stronger dose on the first Friday, and then afterward apply hypofractionation strategies from weeks 2–6. Because a standard clinical radiotherapy procedure would involve applying 10 Gy per week, and in this scenario the tumor undergoes standard radiotherapy procedures for a 6-week time frame, then this proposal strategy follows that 60 Gy limit while providing a higher dose of radiation at the first week. While this greatly slows future growth from weeks 2–6 in the model (along with the effectively proven implemented hypofractionation strategies), the effects of increasing the radiation dosage in the first week by about 60% at the outset may have adverse biological effects that would need to be observed in a laboratory setting.

In essence, this proposal offers a starting point to what may be a simulation-based approach to radiotherapy that would need to be balanced with the biological limitations of living cells. A more balanced approach would only be possible after testing various simulation-based radiotherapy dosage strategies in a laboratory setting, and then possibly using a feedback approach to integrate the results of each tested strategy into a simulation that can better optimize each strategy with the use of more data.

When it comes to a 1:1 PC3 cell line mixture, comparison of this dosage strategy to our previous findings shows us that implementation of both the usage of constant dosage strategies with adaptive dose levels, as well as a stronger dose on Fridays, proves to be more effective than using just one of the two strategies by itself. Table 5 compares values between our proposed dosage strategy and results from Table 3, and Table 6 compares values between Scenario A, defined in Table A5, and our proposed dosage plan. Both data sets show a clear drop in tumor volumes, proving the effectiveness of combining both strategies together.

Clinicians may be able to propose varying dosage

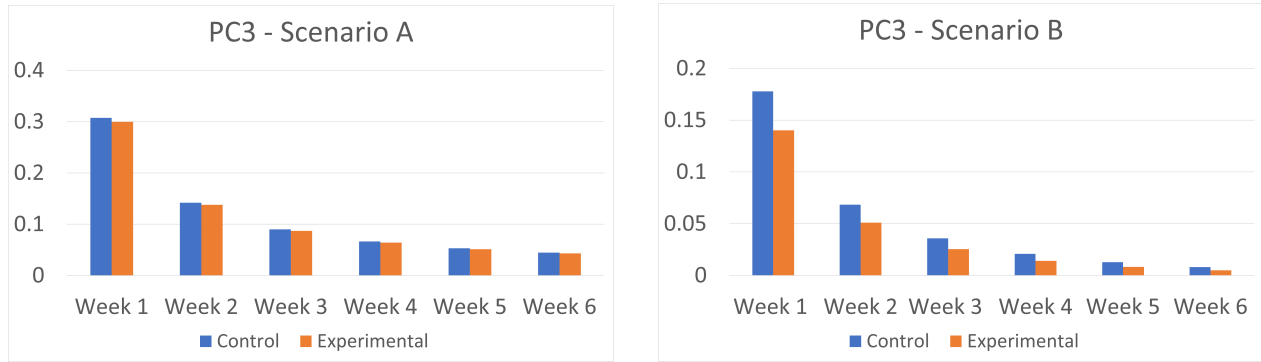


Figure 5: PC3 cell line for Scenario A (top) and Scenario B (bottom). Each bar represents the total tumor volume at the end of each week for either the control population (blue) or the experimental population (orange). 9 Gy are administered every week for a period of 6 weeks for either scenario. Scenario A presents a slight decrease between experimental and control populations, but by combining the two strategies, Scenario B presents itself as a much more promising approach in decreasing overall tumor volume at the end of each week and across all 6 weeks of treatment.

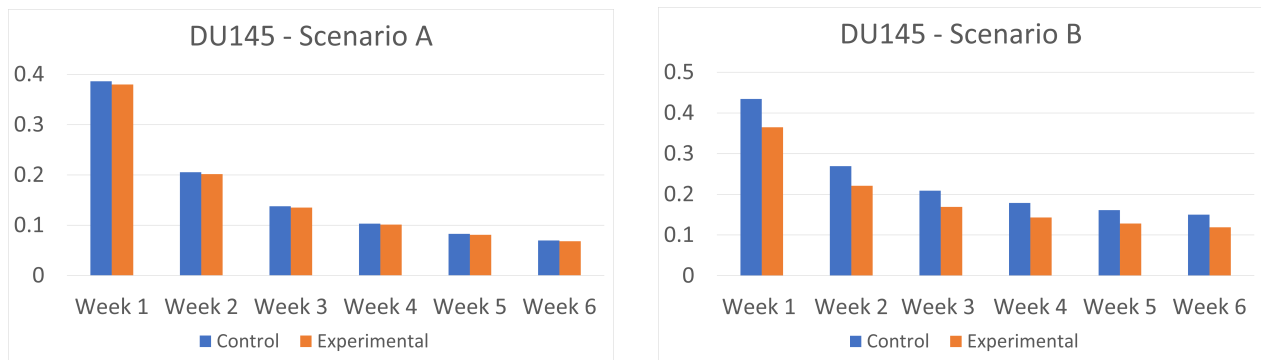


Figure 6: DU145 cell line for Scenario A (top) and Scenario B (bottom). Each bar represents total tumor volume at the end of each week. Control - blue, experimental - orange. 9 Gy are administered in a single week. DU145 presents with similar results as the PC3 cell line when following the same dosage protocol described in Scenario A, however as per the previous section, implementing increasing time intervals in between tumor fractionation is not as effective for the DU145 cell line as it is for the PC3 cell line, therefore it presents with worse overall results than Scenario A for DU145, confirming the ineffectiveness of hypofractionation on DU145.

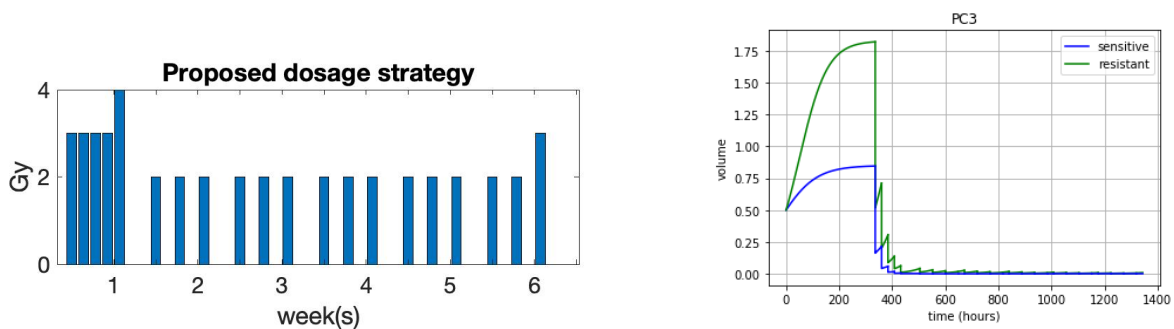


Figure 7: Proposed dosage strategy (top) applied to a tumor initiated as a 1:1 mixture of parental and resistant PC3 cell line (bottom). In the first week, 3 Gy administered daily, with 4 Gy administered on the first Friday. Afterward a constant dose of 2 Gy is administered every Monday, Wednesday, and Friday for 5 weeks, with a final dose of 3 Gy administered at the end of the 6 week time period. The results show the volumes for the individual parental and resistant populations. As detailed in Tables 5 and 6, our proposed dosage strategy results in the smallest final tumor volume compared to all dosage plans tested in the previous sections and the standard clinical schedule.

Table 5: Comparison of values between data collected from our simulation of the standard clinical radiotherapy schedule (a constant 2 Gy dose every weekday with a break on weekends) and results from our proposed dosage strategy. Both models used PC3 cell line parameters. A clear decrease in final tumor volume and total area can be observed in both parental and resistant populations after implementation of the proposed dosage strategy.

		Standard Clinical Radiotherapy Schedule	Proposed Dosage Strategy
Final Tumor Volume	Parental	1.0739×10^{-7}	1.0790×10^{-8}
	Resistant	0.030486	0.0074191
	Total	0.030486	0.0074191
Area Under Curve	Parental	19.910	6.1733
	Resistant	140.41	33.693
	Total	160.32	39.866

Table 6: Comparison of values between the standard clinical radiotherapy model, results referenced in Table A5, and our proposed dosage strategy. Scenario A involved administering 1.75 Gy every day from Monday through Thursday and then 3 Gy every Friday, as well as withholding any radiation on weekends. We mirrored this scheme more effectively in our proposed strategy, as exhibited by the decrease in end-of-week values for all six weeks. Our proposed strategy proved to be most effective.

Tumor Volume		week 1	week 2	week 3	week 4	week 5	week 6
Clinical Standard	Parental	0.051106	0.0077640	0.0013167	0.00022713	3.9305×10^{-5}	6.8229×10^{-6}
	Resistant	0.15534	0.064823	0.034116	0.019652	0.011837	0.0073082
	Total	0.20645	0.072587	0.035433	0.019879	0.011876	0.0073150
Scenario A	Parental	0.044009	0.0063787	0.0010296	0.00016888	2.7777×10^{-5}	4.5829×10^{-6}
	Resistant	0.13129	0.052691	0.026704	0.014785	0.0085415	0.0050479
	Total	0.17530	0.059070	0.027734	0.014954	0.0085693	0.0050525
Proposed Strategy	Parental	0.00039753	3.4299×10^{-5}	2.9631×10^{-6}	2.5612×10^{-7}	2.2186×10^{-8}	3.6478×10^{-9}
	Resistant	0.0097035	0.0061374	0.0039301	0.0025354	0.0016432	0.0017628
	Total	0.010101	0.0061717	0.0039331	0.0025357	0.0016432	0.0017628

strategies prior to the treatment cycle depending on the cell mixture, cell line, size, and growth rate of the tumor in order to fit the needs of each individual patient. This proposal would be most effective in theory on this particular cell line (PC3) within a 1:1 mixture of parental to radio-resistant cells. Due to the particularity of each tumor in any individual patient, the importance of math modeling in clinical radiotherapy remains well-grounded, as ODE models can be used to predict the most effective course of treatment in any particular case, as opposed to following standard procedure that does not take into account all of the variables involved in each individual tumor population, such as tumor heterogeneity. Our hope is that in the future, with the help of ever-advancing medical technology, patients' tumors may be able to be parameterized in a similar manner to this study, and then those parameters can be modeled using ODE's that can extrapolate the approximate growth behavior of the tumor. Using this extrapolation, the best possible dosage strategy can be approximated using much more advanced

optimization strategies than those implemented here, but also one that builds upon the framework proposed earlier in this paper.

Of note to point out is that all experiments in this simulation study were done *in silico*, and although parameters were derived from experiments, the natural world contains many other complex parameters that cannot yet be replicated within a computer simulation. Therefore, further means of study and real-life investigations are necessary before we can apply the findings of this study to a clinical application. In addition, side effects and toxicity of using higher dosage has to be addressed [1, 34]. That being said, our work underscores the potential of adaptive cancer treatments, which is of recent interest in the field. In particular, this concept can be applied to those patients that have heterogeneous mixtures of cancer, with both temporal and spatial variability in their cancer microenvironment and even therapy-induced perturbations.

In our future work, we propose to do further analysis into different prostate cancer cell lines, such as the DU145

cell line to examine the efficacy of adaptive dosages. While the PC3 cell line showed promising results using the hypofractionation strategy, the DU145 cell line did not. This could be due to various factors, including the differences in the proliferation rate and interaction types of the populations. We aim to further identify the key parameters and underlying mechanisms that yield such differences to identify the cancer types and patients that hypofractionation strategy should be applied to. Our final goal is to develop strategies for cancer cell lines with different interaction properties to improve treatment outcome. Comparing other radiotherapy models with our current choice, the linear-quadratic model with instant response, will be our future work as well. It will be interesting to validate our results across different radiotherapy models including a logistic type of radiotherapy model [27, 25], dynamic carrying capacity [36], and delayed response from necrotic population [7]. In the future, we also hope to do prospective experiments to examine the efficacy of proposed dosage strategies, in addition to studying other initial mixture ratios.

Code Availability

Code is available at <https://github.com/josedualv/Effective-dose-fractionation-schemes-of-radiotherapy-on-prostate-cancer>.

Author Contributions

Conceptualization, H. C.; Methodology, J. A., H. C.; Software, J. A., H. C., K. S.; Data Curation, P. K.; Formal Analysis, J. A.; Investigation, J. A.; Writing – Original Draft, J. A.; Writing – Review & Editing, J. A., H. C., P. K., K. S.; Visualization, J. A., H. C.

Appendix

Tables and bar graphs for Scenarios A, B, A', and B' can be found on the following pages 27–29.

References

- [1] Kamran A Ahmed et al. “Altered fractionation schedules in radiation treatment: a review”. In: *Seminars in oncology*. Vol. 41. 6. Elsevier. 2014, pp. 730–750.
- [2] G. C. Barnett et al. “Normal tissue reactions to radiotherapy: towards tailoring treatment dose by genotype”. In: *Nat. Rev. Cancer* 9 (2009), pp. 134–142.
- [3] A. C. Begg, F. A. Stewart, and C. Vens. “Strategies to improve radiotherapy with targeted drugs”. In: *Nat. Rev. Cancer* 11 (2011), pp. 239–253.
- [4] J. Bernier, E. J. Hall, and A. Giaccia. “Radiation oncology: a century of achievements”. In: *Nature* 4 (2004), pp. 737–747.
- [5] Kristy K Brock. “Adaptive radiotherapy: moving into the future”. In: *Seminars in radiation oncology*. Vol. 29. 3. NIH Public Access. 2019, p. 181.
- [6] Helen M. Byrne. “Dissecting cancer through mathematics: From the cell to the animal model”. In: *Nature Reviews Cancer* 10 (2010), pp. 221–230.
- [7] H. Cho et al. “A framework for performing data-driven modeling of tumor growth with radiotherapy treatment”. In: *Springer Special Issue: Using Mathematics to Understand Biological Complexity, Women in Mathematical Biology 22* (2020), pp. 179–216.
- [8] Heyrim Cho and Doron Levy. “Modeling the chemotherapy-induced selection of drug-resistant traits during tumor growth”. In: *J. Theor. Biol.* 436.7 (2018), pp. 120–134.
- [9] Heyrim Cho, Allison L Lewis, and Kathleen M Storey. “Bayesian Information-Theoretic Calibration of Radiotherapy Sensitivity Parameters for Informing Effective Scanning Protocols in Cancer”. In: *Journal of clinical medicine* 9.10 (2020), p. 3208.
- [10] David Corwin et al. “Toward patient-specific, biologically optimized radiation therapy plans for the treatment of glioblastoma”. In: *PLoS ONE* 8.e79115 (2013).
- [11] G. Delaney et al. “The role of radiotherapy in cancer treatment: estimating optimal utilization from a review of evidence-based clinical guidelines.” In: *Cancer* 104 (2005), pp. 1129–1137.
- [12] Heiko Enderling et al. “Integrating Mathematical Modeling into the Roadmap for Personalized Adaptive Radiation Therapy”. In: *Trends in Cancer* 5.8 (2019), pp. 467–474.
- [13] J. F. Fowler. “The linear-quadratic formula and progress in fractionated radiotherapy”. In: *British Journal of Radiology* 62 (1989), pp. 679–694.
- [14] Audrey R Freischel et al. “Frequency-dependent interactions determine outcome of competition between two breast cancer cell lines”. In: *Scientific reports* 11.1 (2021), pp. 1–18.
- [15] Robert A Gatenby. “Models of tumor-host interaction as competing populations: implications for tumor biology and treatment”. In: *Journal of theoretical biology* 176.4 (1995), pp. 447–455.

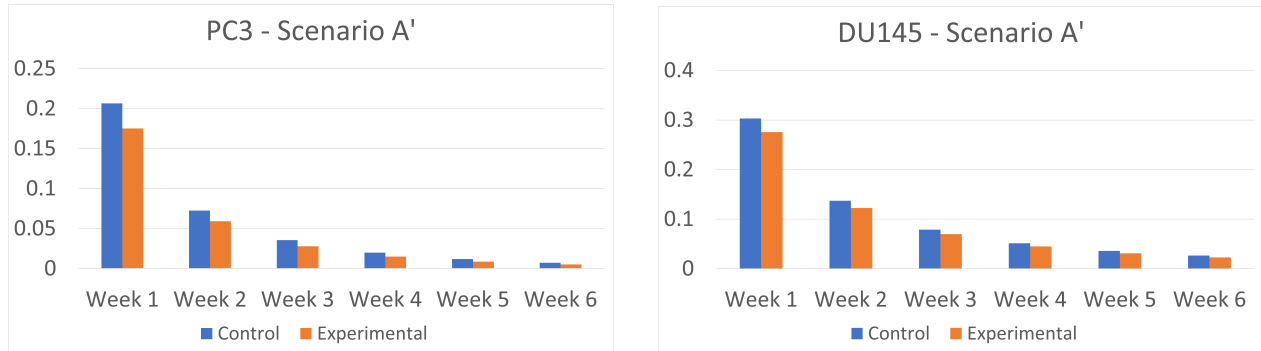


Figure A1: End-of-week tumor volumes in graphical format for Scenario A' on the PC3 cell line (top) and on the DU145 cell line (bottom). Scenario A' shows the effectiveness of a higher Friday dosage without the implementation of hypofractionation strategies on either cell line (as in Scenarios B and B'). While weekly dosage is higher than that of Scenarios A and B (10 Gy vs 9 Gy), this value is consistent between control and experimental simulations within this scenario, meaning the difference in end-of-week tumor volumes seen between the control and experimental simulations is entirely due to a stronger Friday dosage.

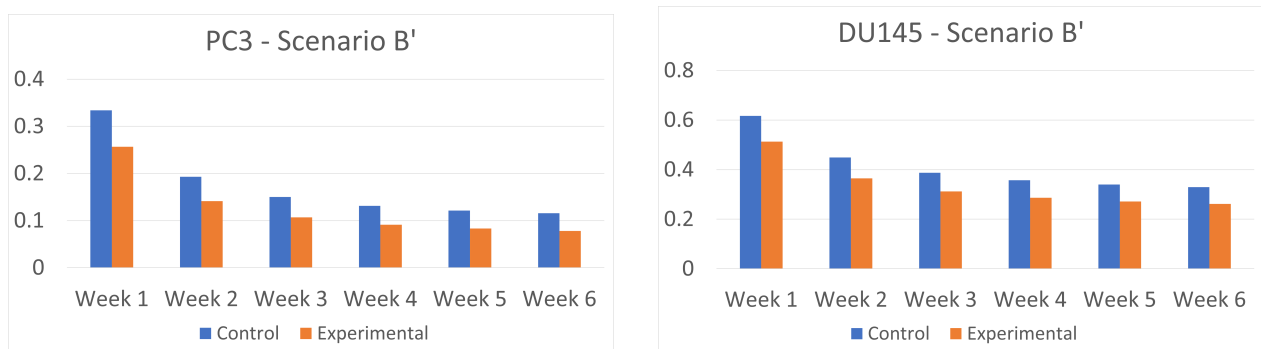


Figure A2: End-of-week tumor volumes in graphical format for Scenario B' on the PC3 cell line (top) and on the DU145 cell line (bottom). This scenario involves administration of only 7.5 Gy every week, and implements a stronger Friday dosage as well as hypofractionation strategies discussed earlier. While a stronger Friday dosage is still effective for either cell line, hypofractionation strategies remain minimally effective to the overall tumor volume of DU145 cell line.

Table A1: PC3 cell line for Scenario A. Values represent tumor volume at the end of each week. Control: 1.8 Gy administered Monday through Friday; Experimental: 1.75 Gy administered Monday through Thursday, 2 Gy administered on Friday. 9 Gy administered in a single week.

Tumor Volume		week 1	week 2	week 3	week 4	week 5	week 6
Control	Parental	0.084901	0.021027	0.0061529	0.0018777	0.00058006	0.00017988
	Resistant	0.22261	0.12080	0.083701	0.064383	0.052479	0.044384
	Total	0.30751	0.14183	0.089854	0.066261	0.053059	0.044564
Experimental	Parental	0.082921	0.020487	0.0059828	0.0018221	0.00056178	0.00017386
	Resistant	0.21667	0.11725	0.081114	0.062311	0.050729	0.042854
	Total	0.29959	0.13774	0.087097	0.064133	0.051291	0.043028

Table A2: DU145 cell line for Scenario A. Values represent tumor volume at the end of each week. Control: 1.8 Gy administered Monday through Friday; Experimental: 1.75 Gy administered Monday through Thursday, 2 Gy administered on Friday. 9 Gy administered in a single week.

Tumor Volume		week 1	week 2	week 3	week 4	week 5	week 6
Control	Parental	0.15188	0.066600	0.036114	0.021379	0.013240	0.0084134
	Resistant	0.23423	0.13887	0.10169	0.081892	0.069616	0.061283
	Total	0.38611	0.20547	0.13780	0.10327	0.082856	0.069696
Experimental	Parental	0.14927	0.065328	0.035387	0.020933	0.012954	0.0082259
	Resistant	0.23038	0.13631	0.099727	0.080260	0.068197	0.060011
	Total	0.37965	0.201638	0.13511	0.10119	0.081151	0.068237

Table A3: PC3 cell line for Scenario B. Values represent tumor volume at the end of each week. Control: 3 Gy administered Monday, Wednesday, and Friday; Experimental: 2.5 Gy administered Monday and Wednesday, 4 Gy on Friday. 9 Gy administered in a single week.

Tumor Volume		week 1	week 2	week 3	week 4	week 5	week 6
Control	Parental	0.019411	0.0015771	0.00013555	1.1705×10^{-5}	1.0112×10^{-6}	8.7391×10^{-8}
	Resistant	0.15839	0.066878	0.035654	0.020827	0.012734	0.0079866
	Total	0.17780	0.068455	0.035790	0.020839	0.012735	0.0079867
Experimental	Parental	0.016345	0.0012511	0.00010121	8.2218×10^{-6}	6.6823×10^{-7}	5.4331×10^{-8}
	Resistant	0.12392	0.049846	0.025389	0.014144	0.0082270	0.0048978
	Total	0.14027	0.051097	0.025490	0.014152	0.0082277	0.0048979

Table A4: DU145 cell line for Scenario B. Values represent tumor volume at the end of each week. Control: 3 Gy administered Monday, Wednesday, and Friday; Experimental: 2.5 Gy administered Monday and Wednesday, 4 Gy on Friday. 9 Gy administered in a single week.

Tumor Volume		week 1	week 2	week 3	week 4	week 5	week 6
Control	Parental	0.22442	0.15275	0.12874	0.11798	0.11256	0.10967
	Resistant	0.20989	0.11644	0.080110	0.060718	0.048637	0.040383
	Total	0.43431	0.26919	0.20881	0.17870	0.16120	0.15005
Experimental	Parental	0.18009	0.12056	0.10093	0.092219	0.087874	0.085586
	Resistant	0.18451	0.10023	0.068033	0.050978	0.040404	0.033206
	Total	0.36460	0.22079	0.16896	0.14320	0.12828	0.11879

Table A5: PC3 cell line for Scenario A'. Control: 2 Gy administered Monday through Friday; Experimental: 1.75 Gy administered Monday through Thursday, 3 Gy administered on Friday. 10 Gy administered in a single week.

Tumor Volume		week 1	week 2	week 3	week 4	week 5	week 6
Control	Parental	0.051106	0.0077640	0.0013167	0.00022712	0.000039305	0.0000068229
	Resistant	0.15534	0.064823	0.034116	0.019652	0.011837	0.0073082
	Total	0.20645	0.072587	0.035433	0.019879	0.011876	0.0073150
Experimental	Parental	0.044009	0.0063787	0.0010296	0.00016888	0.000027777	0.0000045829
	Resistant	0.13129	0.052691	0.026704	0.014785	0.0085415	0.0050479
	Total	0.17530	0.059070	0.027734	0.014954	0.0085693	0.0050525

Table A6: DU145 cell line for Scenario A'. Control: 2 Gy administered Monday through Friday; Experimental: 1.75 Gy administered Monday through Thursday, 3 Gy administered on Friday. 10 Gy administered in a single week.

Tumor Volume		week 1	week 2	week 3	week 4	week 5	week 6
Control	Parental	0.11442	0.040030	0.017021	0.0077624	0.0036484	0.0017389
	Resistant	0.18871	0.096959	0.061894	0.043493	0.032255	0.024753
	Total	0.30313	0.13699	0.078915	0.051255	0.035903	0.026492
Experimental	Parental	0.10364	0.035558	0.014866	0.0066652	0.0030785	0.0014413
	Resistant	0.17204	0.086988	0.054900	0.038185	0.028037	0.021301
	Total	0.27568	0.12255	0.069766	0.044850	0.031116	0.022742

Table A7: PC3 cell line for Scenario B'. Values represent tumor volume at the end of each week. Control: 2.5 Gy administered Monday, Wednesday, and Friday; Experimental: 2 Gy administered Monday and Wednesday, 3.5 Gy on Friday. 7.5 Gy administered in a single week.

Tumor Volume		week 1	week 2	week 3	week 4	week 5	week 6
Control	Parental	0.046328	0.0090815	0.0020226	0.00046238	0.00010633	0.000024483
	Resistant	0.28760	0.18373	0.14793	0.13083	0.12139	0.11577
	Total	0.33393	0.19281	0.14995	0.13129	0.12150	0.11579
Experimental	Parental	0.037598	0.0069530	0.0014604	0.00031438	0.000068035	0.000014740
	Resistant	0.21896	0.13435	0.10526	0.091107	0.083055	0.078059
	Total	0.25656	0.14130	0.10672	0.091421	0.083123	0.078074

Table A8: DU145 cell line for Scenario B'. Values represent tumor volume at the end of each week. Control: 2.5 Gy administered Monday, Wednesday, and Friday; Experimental: 2 Gy administered Monday and Wednesday, 3.5 Gy on Friday. 7.5 Gy administered in a single week.

Tumor Volume		week 1	week 2	week 3	week 4	week 5	week 6
Control	Parental	0.32275	0.24864	0.22279	0.21006	0.20257	0.19766
	Resistant	0.29400	0.20008	0.16489	0.14745	0.13762	0.13169
	Total	0.61675	0.44872	0.38768	0.35751	0.34019	0.32935
Experimental	Parental	0.25674	0.19434	0.17308	0.16272	0.15668	0.15274
	Resistant	0.25608	0.17060	0.13906	0.12344	0.11460	0.10922
	Total	0.51282	0.36494	0.31214	0.28616	0.27128	0.26196

- [16] Eric J. Hall. *Radiobiology for the radiologist*. J.B. Lippincott, Philadelphia, 1994.
- [17] Wen-song Hong, Shun-guan Wang, and Gang-qing Zhang. “Lung Cancer Radiotherapy: Simulation and Analysis Based on a Multicomponent Mathematical Model”. In: *Computational and Mathematical Methods in Medicine 2021* (2021).
- [18] Pavitra Kannan et al. “Radiation resistant cancer cells enhance the survival and resistance of sensitive cells in prostate spheroids”. In: *bioRxiv* (2019).
- [19] James A. Koziol, Theresa J. Falls, and Jan E. Schnitzer. “Different ODE models of tumor growth can deliver similar results”. In: *BMC Cancer* 20.226 (2020), pp. 1–10.
- [20] Yang Kuang, John D Nagy, and Steffen E Eikenberry. *Introduction to mathematical oncology*. CRC Press, 2018.
- [21] D. E. Lea and D. G. Catcheside. “The mechanism of the induction by radiation of chromosome aberrations in *Tradescantia*”. In: *Journal of Genetics* 44 (1942), pp. 216–245.
- [22] Kevin Leder et al. “Mathematical modeling of PDGF-driven glioblastoma reveals optimized radiation dosing schedules”. In: *Cell* 156.3 (2014), pp. 603–616.
- [23] Yingting Liu et al. “A multiscale computational approach to dissect early events in the erb family receptor mediated activation, differential signaling, and relevance to oncogenic transformations”. In: *Annals of Biomedical Engineering* 35 (2007), pp. 1012–1025.
- [24] Hope Murphy, Hana Jaafari, and Hana M. Dobrovoly. “Differences in predictions of ODE models of tumor growth: A cautionary example”. In: *BMC Cancer* 16.163 (2016), pp. 1–10.
- [25] Jan Poleszczuk et al. “Predicting Patient-Specific Radiotherapy Protocols Based on Mathematical Model Choice for Proliferation Saturation Index”. In: *Bulletin of Mathematical Biology* 80.5 (2018), pp. 1195–1206.
- [26] Alan Pollack and Ahmed Mansoor. *Hypofractionation: Scientific Concepts and Clinical Experiences*. Ellicott City: LimiText Publishing, 2011.
- [27] Sotiris Prokopiou et al. “A proliferation saturation index to predict radiation response and personalize radiotherapy fractionation”. In: *Radiation Oncology* 10.159 (2015), pp. 1–8.
- [28] U. Ringborg et al. “The Swedish Council on Technology Assessment in Health Care: systematic overview of radiotherapy for cancer including a prospective survey of radiotherapy practice in Sweden 2001-summary and conclusions.” In: *Acta. Oncol.* 42 (2003), pp. 357–365.
- [29] R. Rockne et al. “Predicting the efficacy of radiotherapy in individual glioblastoma patients in vivo: A mathematical modeling approach”. In: *Physics in Medicine and Biology* 55 (2010), pp. 3271–3285.
- [30] C. E. Shannon. “A Mathematical Theory of Communication”. In: *Bell System Technical Journal* 27 (1948), pp. 379–423.
- [31] Enakshi D. Sunassee et al. “Proliferation saturation index in an adaptive Bayesian approach to predict patient-specific radiotherapy responses”. In: *International Journal of Radiation Biology* 95.10 (2019), pp. 1421–1426.
- [32] Howard D. Thames et al. “Changes in early and late radiation responses with altered dose fractionation: Implications for dose-survival relationships”. In: *International Journal of Radiation Oncology, Biology, Physics* 8 (1982), pp. 219–226.
- [33] Ben G. L. Vanneste et al. “Prostate Cancer Radiation Therapy: What Do Clinicians Have to Know?” In: *Biomed. Res. Int.* 6829875 (2016), pp. 1–14.
- [34] Anders Widmark et al. “Ultra-hypofractionated versus conventionally fractionated radiotherapy for prostate cancer: 5-year outcomes of the HYPO-RT-PC randomised, non-inferiority, phase 3 trial”. In: *The Lancet* 394.10196 (2019), pp. 385–395.
- [35] Di Yan et al. “Adaptive radiation therapy”. In: *Physics in Medicine & Biology* 42.1 (1997), p. 123.
- [36] Mohammad U Zahid et al. “Forecasting Individual Patient Response to Radiotherapy in Head and Neck Cancer with a Dynamic Carrying Capacity Model”. In: *Int. J. Radiat. Oncol. Biol. Phys.* (2021), in press.
- [37] Nicholas G. Zaorsky et al. “What is the ideal radiotherapy dose to treat prostate cancer? A meta-analysis of biologically equivalent dose escalation”. In: *Radiation Oncology* 115.3 (2015), pp. 295–300.
- [38] Jingsong Zhang et al. “Integrating evolutionary dynamics into treatment of metastatic castrate-resistant prostate cancer”. In: *Nature communications* 8.1 (2017), pp. 1–9.