DOI: 10.1111/1365-2745.13823

RESEARCH ARTICLE

Polyploidy promotes divergent evolution across the leaf economics spectrum and plant edaphic niche in the *Dianthus broteri* complex

Javier López-Jurado 💿	Enrique Mateos-Naranio 💿	Francisco Balao
Javici Lopez Julauo 🧹	Enrique Mateus Maranjo	

Departamento de Biología Vegetal y Ecología, Facultad de Biología, Universidad de Sevilla, Sevilla, Spain

Correspondence Javier López-Jurado Email: javlopez@us.es

Funding information

Áridos La Melera S.L., Grant/Award Number: 2234/0724; Fifth Research Plan from the University of Seville, Grant/Award Number: USE-19606-X; Sixth Research and Transfer Plan from the University of Seville, Grant/Award Number: USE-21358-Z; Ministerio de Ciencia e Innovación, Grant/Award Number: PGC2018-098358-B-100

Handling Editor: Stephen Bonser

Abstract

- The evolution of the leaf economics spectrum (LES) is known to be constrained by genetic relatedness but also promoted at small geographical and phylogenetic scales. In those cases, we hypothesized that polyploidy would play a prominent role as an outstanding source of functional divergence and adaptive potential.
- 2. We registered leaf-level nutrient, water- and light-economy-related traits from the LES as well as edaphic properties in the four cytotypes of the autopolyploid *Dianthus broteri* complex (2×, 4×, 6× and 12×). We analysed the effect of ploidy level on the integration of the LES network, checked if concerted evolution occurred between LES and soil niche and tested the influence of phylogeny on the variables. Alternative evolutionary models for both sets of traits were compared.
- 3. We found higher divergence of polyploids (especially 6× and 12×) compared to diploids in the LES and soil niche, but these traits are not coevolving. 6× and 12× showed opposite ecological strategies regarding resource use and higher uncoupling of the LES network. Early divergence of traits prevailed in both LES and edaphic niche (supported by better fitted evolutionary models with one optimum per cytotype), but post-polyploidization processes played an important role for the photochemical behaviour.
- 4. *Synthesis.* Our results indicated shifts in ecological strategies across *Dianthus broteri* cytotypes and suggested a powerful role of polyploidy in overcoming constraints for the evolution of plant functional traits.

KEYWORDS

adaptive divergence, ecological strategies, functional traits, leaf economics spectrum, polyploidy, soil niche

1 | INTRODUCTION

Plant functional traits are those impacting their demographic fitness, that is, fecundity, growth, development or survival (Violle et al., 2007; Volaire et al., 2020). Notably, major functional traits in vascular plants are concentrated and coordinated in a multidimensional space explaining plant form and function at a global scale (Díaz et al., 2016). Within the global plant trait variation, there is a fully differentiated axis corresponding to the leaf economics spectrum (LES; Wright et al., 2004), which accounts for the return on investments in

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made. © 2021 The Authors. *Journal of Ecology* published by John Wiley & Sons Ltd on behalf of British Ecological Society. leaves regarding carbon and nutrient acquisition and processing. The coordination among LES traits, especially on a mass basis, causes this spectrum to be defined by a trade-off between plant longevity and growth potential (Wright et al., 2004). According to this, ecological strategies would be reflected by the positioning of a given plant species along the LES (Reich, 2014), varying from nutrient-acquisitive strategies, also known as fast (with high photosynthetic capacity and short-lived leaves), to conservative ones (i.e. slow, with low photosynthetic capacity and long-lived leaves).

Beyond nutrient-associated traits, this spectrum seems to be coordinated with the essential resources of water and light (Reich, 2014). Previous studies have already demonstrated the correlation between plant nutrient and water economy (e.g. Pérez-Ramos et al., 2012; Prieto et al., 2018), being the species with slow strategies also conservative about water and therefore more tolerant to drought (Hallik et al., 2009). Furthermore, the coordination of light economy with the purely LES traits has also been indirectly addressed by linking shade tolerance with less thick and dense leaf blades, probably to achieve larger harvesting surfaces in light-limited environments (e.g. Hallik et al., 2009; Lusk et al., 2008). On the basis of these findings and the results obtained by Hallik et al. (2012) about chlorophyll fluorescence characteristics in woody and herbaceous species under different growth irradiance conditions, we hypothesized that light collection at the photosystem level would be enhanced for less light-exposed leaves while PSII photochemical efficiency would be diminished as a result of photoinhibition. In any case, the joint study of variables related to nutrient, water and light use are needed to delve into the interdependence of functional traits that assure plant success in a given environment, following the respective recommendations of Poorter et al. (2014) and Reich (2014).

However, global-scale trait covariation in the LES is challenged by non-global scale predictions (Mason et al., 2016). Locally, natural selection can generate trait correlations that are relatively easy to escape (soft constraint) by drivers of phenotypic variation, altering the LES trade-off. These drivers include plasticity, ecological filtering, sexual reproduction, adaptation, genetic drift or ontogenetic mechanisms (Messier et al., 2017) and may lead to a diversity of functional trait associations within phylogenetically closely related groups (Carvalho et al., 2020; Niinemets, 2015) and even at the intra-individual level (between leaf cohorts; Puglielli, 2019; Puglielli & Varone, 2018). Noticeably, a significant relationship occurs between environmental conditions and LES traits, as the influence of soil properties (Lambers & Poorter, 1992; Lambers et al., 2008), but also the interaction between soil and climate (Luo et al., 2015, 2016) has been addressed on plant functional traits and resourceuse strategies. This supports the interpretation of trait-environment associations as a result of adaptive evolution (Reich et al., 2003), highlighting the role of resource availability or climatic conditions in functional divergence along the LES (Brouillette et al., 2014; Sartori et al., 2019).

Apart from natural selection, harder constraints (i.e. those that are almost impossible to escape) to novel trait associations shape the expression of the LES. These are of biophysical and genetic nature (Messier et al., 2017), being the former the most restrictive ones and consistent within and across ecological scales. Among biophysical constraints, the link between photosynthetic capacity and leaf life span is of paramount importance since there are unavoidable limitations to construct long-lasting leaves with high productivity. Furthermore, the evolution of the LES is expected to be limited to some extent by a genetic component, although this topic deserves further investigation (Donovan et al., 2011). Specifically, the referenced study suggested that both selection and genetic constraints need to be measured jointly to test their relative roles on the LES evolution. To date, it is known that a few pleiotropic genes maintain the strong correlations between LES traits (Vasseur et al., 2012), which would have a major role in the phenotypic integration within this complex network (Ji et al., 2020).

In the light of this evidence, the release of genetic constraints would be the baseline for promoting functional divergence in the LES. It would cause evolutionary novelties such as adaptive phenotypic plasticity or epigenetic variation (Kalisz & Kramer, 2008). Thus, polyploidy (i.e. the state of having more than two paired sets of chromosomes) may play a pivotal role on LES evolution over low phylogenetic scales given that it is considered an outstanding driver of these sources of adaptive modifications (Chen, 2007; Hahn et al., 2012). Allopolyploids (those originated from interspecific hybridization) are prone to produce numerous combinations of traits due to their diverse origin (Seehausen, 2004), but genome duplication itself and subsequent adaptation have also demonstrated the capacity to cause phenotypic changes in autopolyploids (those originated from a single species; Martínez et al., 2018; Visger et al., 2016; Wei et al., 2020). Moreover, autopolyploidy per se can lead to novelty by modifying the phenotypic covariance matrix, meaning the disintegration or uncoupling of those traits (Oswald & Nuismer, 2011). This evolutionary innovation in polyploids can be caused either by relaxed selection on duplicate genes due to their higher functional redundancy (Flagel & Wendel, 2009) or increased variation in dosage-regulated gene expression levels (Osborn et al., 2003).

Using a recently radiated autopolyploid complex, we can not only assess the effect of polyploidization and rapid adaptive processes on functional divergence, but we also would provide new insights into the neglected ecological context of non-crop polyploid systems (Soltis et al., 2016). An approach like the one used in this work would facilitate underlying explanations to several widely studied ecological processes in polyploids, such as competitive interactions (see e.g. Čertner et al., 2019), fitness advantage (see e.g. Wei et al., 2019) or niche differentiation (see e.g. Rojas-Andrés et al., 2020).

Considering this background, we studied LES variation across the autopolyploid complex *Dianthus broteri* (Caryophyllaceae). This group of perennial herbaceous plants consists of four different ploidy levels (2×, 4×, 6× and 12×; Balao et al., 2009) that diverged about 0.9–2.1 Mya (Balao et al., 2010) and have different evolutionary histories (the 4× cytotype has two independent origins while 6× and 12× have single origins; Balao et al., 2010). Moreover, *D. broteri* has a Mediterranean distribution (exclusively in the Iberian Peninsula), where its ploidies do not coexist and occupy disjunct geographical ranges with markedly different environmental conditions. Comparatively, diploids and tetraploids inhabit temperate and broad niches with nutrient-rich soils, whereas hexaploids and dodecaploids are distributed in warm and arid habitats with infertile soils (López-Jurado et al., 2019). The inter-cytotype differentiation pattern in this complex also encompasses global methylation levels (Alonso et al., 2016) and photochemical responses (López-Jurado et al., 2020). Interestingly, morphological features (vegetative and reproductive organs) not only showed divergence among D. broteri cytotypes, but also a loss of phenotypic integration with increasing ploidy (i.e. lower covariation among functionally related traits; Balao et al., 2011). The shared conclusions from the mentioned studies suggest adaptations of polyploids (particularly 6× and 12×) to specific biotic and abiotic factors. Besides that, a focus on the 12× has been established caused by unique properties as its floral scent emission (Balao et al., 2011), high tolerance to recurrent drought (López-Jurado et al., 2016) and alleged key role in ecosystem functioning (López-Jurado et al., 2019).

Based on the mentioned previous results obtained for this complex, our starting hypotheses were that (a) *D. broteri* has a variety of LES trait combinations and hence ecological strategies, which would be acquisitive in low-order ploidy levels (2x and 4x) and conservative in high-order ones (6x and 12x; in response to the stressful niches that they inhabit) and (b) polyploidy is enhancing the uncoupling of functional traits in this complex, especially in the 6x and 12x cytotypes. In this study, we aimed to uncover polyploidy-mediated shifts in the LES and the edaphic niche, as well as resource (nutrient, water and light) acquisition and resource-use strategies that enable the *D. broteri* polyploid complex to occupy heterogeneous environments.

2 | MATERIALS AND METHODS

2.1 | Characterization of study sites

During July 2017, three natural populations of each *Dianthus broteri* cytotype encompassing their corresponding geographical distributions were sampled (between 10:00 and 14:00 hr; Figure S1). All these locations were characterized by considerably low *D. broteri* density, ranging between 30 and 100 individuals, which were accompanied mostly by xerophytic Mediterranean shrubland. In each location, the main climatic conditions were recorded (Table S1), besides performing measurement and sampling procedures described in the following sections.

As previously shown in López-Jurado et al. (2019), populations of lower cytotypes ($2 \times$ and $4 \times$) had milder climatic conditions compared to those of higher ones ($6 \times$ and $12 \times$), and showed intermediate values in relative humidity and the lowest temperatures. On average, the niche of hexaploids was the most humid during sampling time (summer) and dodecaploids were exposed to the warmest and driest conditions (Table S1).

2.2 | Soil composition and properties

In each population, soil samples (n = 3) from the upper 15 cm were collected throughout the area occupied by D. broteri individuals, placed in plastic containers and conserved at 4°C until their processing in the laboratory. Soil granulometric composition was determined and chemical characterization considered, for example, soil electrical conductivity (EC), which is an indirect indicator of the amount of water and water-soluble nutrients available for plant uptake (Smith & Doran, 1997). Organic matter content (OM; by the Walkley-Black method), total carbonates (CaCO₃), active limestone and nitrogen (N, by the Kjeldahl method) were also included in the analysis. Finally, we quantified assimilable phosphorus (P) by ultraviolet-visible spectrophotometry, as well as exchangeable potassium (K), calcium (Ca), magnesium (Mg), sodium (Na), iron (Fe), manganese (Mn), zinc (Zn) and copper (Cu) by atomic absorption spectroscopy (Fernandez-Hernandez et al., 2010; Usman et al., 2017).

2.3 | Leaf nutrient concentrations and isotopic composition

Completely developed mature distal leaves (n = 5 per individual, n = 50 per population) were collected from the middle canopy layer of plants. Leaves from the same population were blended, dried at 60°C for 48 hr and finely ground using a mixer mill (Retsch GmbH-MM 301). Carbon and nitrogen contents of each leaf pool replicate (n = 3 per population) were quantified using a LECO TruSpec CN elemental analyzer. Additionally, Inductively Coupled Plasma Optical Emission Spectrometry (ICP-OES) analysis was carried out in a Thermo Scientific iCAP 6500 spectrometer to determine levels of macro- and micro-nutrients in leaf pools (n = 3 per population; Table S3). Among the numerous leaf nutrients, we focused on N, P and K levels because of their tight association with plant productivity (Niinemets & Kull, 2005; Sardans & Peñuelas, 2015). N and P influence photosynthesis by having structural and functional roles in the Calvin cycle and the thylakoids (Evans, 1989; Vance et al., 2003) and K is involved in a broad range of plant physiological functions (e.g. enzyme activation and regulation of osmotic potential or stomatal opening; Wang et al., 2013).

The calculation of C and N isotopic signature on dry and pulverized leaf samples (n = 3 per population) was conducted using a Thermo Scientific Delta V spectrometer. The carbon and nitrogen isotope ratios (${}^{13}C/{}^{12}C$ and ${}^{15}N/{}^{14}N$) were expressed in delta (δ) notation, defined as the parts per thousand (∞) relative to a standard material (VPDB). The analytical precision for the measurements was 0.2‰. $\delta^{15}N$ was used as a proxy for nitrogen fluxes, assimilation and allocation (Kalcsits et al., 2014) supplementary to leaf N content. Furthermore, water-use strategies were approached by $\delta^{13}C$, which was employed as a proxy for _iWUE (intrinsic water-use efficiency; Pérez-Harguindeguy et al., 2013) because it relies on the ratio of

intercellular to ambient CO_2 concentrations (C_i/C_a) and therefore is affected by transpiration at the stomata (g_s) and net assimilation $(A_N;$ Farquhar et al., 1989).

2.4 | Leaf traits and physiological measurements

During field sampling, we collected 10 completely developed mature leaves in each population (1 leaf per individual) and measured their areas in the field by digital image analysis using 'Easy Leaf Area' software (Easlon & Bloom, 2014). Dry weight (DW) was determined after drying the leaves at 60°C until constant weight was reached and leaf mass per area (LMA) was then calculated as the ratio between DW and leaf area (g DW/m²). LMA was selected as a crucial variable to account for leaf constructions costs as well as plant growth rate (low LMA indicates thin leaf blade or not very dense tissue, or both, which contribute to short leaf life spans and fast growth rates; Lambers & Poorter, 1992; Westoby et al., 2002; Wright et al., 2004), also being highly variable within functional groups and responsive to environmental conditions (Poorter et al., 2009).

Furthermore, the maximum quantum efficiency of PSII photochemistry (F_v/F_m) and the chlorophyll *a* fast kinetics were registered in 30 min dark-adapted, completely developed leaves (n = 25 per protocol) from randomly selected individuals of each population, using FMS-2 (Hansatech Instruments Ltd., UK) and FluorPen FP100 (Photo System Instruments, Czech Republic) portable pulse-modulated fluorimeters, respectively. For the chlorophyll *a* transient data, we used the pre-programmed OJIP test protocols of FluorPen FP100 and calculated derived parameters following Strasser et al. (2010). In particular, we analysed the informative energy transduction fluxes on a leaf cross-section basis (absorbed, ABS/CS, trapped, TR/CS, electron transport, ET/CS, and dissipated, DI/CS, energy fluxes) since significant differences in the density of reaction centres (RC/CS) among cytotypes were previously found (one-way analysis of variance, ANOVA: $F_{3,246} = 5.91$; p < 0.05). F_v/F_m and OJIP test-derived total performance index (PI_{Total}) were considered based on the broad view they offer about the overall photochemical activity and their sensitivity to stressful conditions, as those of the sampling time in summer.

2.5 | Edaphic divergence of polyploids

After obtaining a detailed description of the edaphic niche characteristics in the 12 sampled *D. broteri* populations, we estimated their phylogenetic relationships from a previously performed phylogram based on amplified fragment length polymorphism (AFLP) data (see Balao et al., 2010, 2011). Polytomies were resolved in random order using the *multi2di* function from the 'APE' package in R (Paradis et al., 2004). Using the 'ADEPHYLO' R package (Jombart, Balloux, et al., 2010), we then performed a phylogenetic principal component analysis (pPCA; Jombart, Pavoine, et al., 2010), which can detect

sets of soil traits presenting a similar phylogenetic signal and hence informs about the general influence of phylogeny on the measured variables. Through this method, we attempted to identify sources of inter-cytotype variation due to phylogenetic relatedness as well as local divergence (i.e. adaptation or phenotypic plasticity). On the one hand, axes with the most positive eigenvalues reflected positive phylogenetic autocorrelation, corresponding to early diverged traits (global structures sensu Jombart, Pavoine, et al., 2010). At the microevolutionary scale studied in the D. broteri complex, early divergence is tightly related to polyploid formation (Balao et al., 2010), although it is not completely possible to separate direct effects due to polyploidization from rapid selection promoting the polyploid establishment without experimenting with synthetic polyploids. On the other hand, axes with the most negative eigenvalues mirrored the most recently diverged traits (lower phylogenetic autocorrelation than null expected or local structures), related to post-polyploidization processes such as adaptation and phenotypic plasticity.

2.6 | Divergence and integration in the LES covariation network

For the analyses involving LES and its relationship with polyploidy, we used the previously introduced meaningful variables to describe ecological strategies from nutrient (LMA, leaf N, P, K and δ^{15} N) to water use (δ^{13} C), but also the photochemical behaviour (ABS/CS, TR/CS, ET/CS, DI/CS, F_v/F_m and Pl_{Total}). Thus, we carried out a pPCA, as previously described, using the 12 selected functional variables to investigate the effects of phylogeny and downstream selection on the LES. The influence of ploidy level on both sets of variables (functional and edaphic) was analysed using one-way ANOVAs in the two most explanatory axes from each pPCA. In case of significant results, post-hoc Tukey's HSD tests ($\alpha = 0.05$) were performed to identify inter-cytotype pairwise differences.

The overall covariation network in LES traits considering the influence of phylogeny was analysed using the 'corrr' package in R (Kuhn et al., 2020). We also guantified the magnitude of integration in the LES network, as a measure of coordination between functional traits, for each D. broteri cytotype separately based on the index of phenotypic integration (INT; Cheverud et al., 1989; Wagner, 1984), following the approach of Damián et al. (2020). We first constructed one correlation matrix per cytotype using Pearson's r and compared them by Mantel tests. We calculated INT values by accounting for the variance among eigenvalues (λ_i) of each correlation matrix and then corrected the values as $INT_c = (Var (\lambda_i) - (number of traits - 1)/$ number of individuals per cytotype). We transformed each INT, to percentage of the maximum integration, which equals the total number of traits in the correlation matrix. Finally, we obtained the 95% confidence intervals for each INT_c by the bootstrapping method (R = 10,000). High INT_c values were indicative of high variance among eigenvalues and therefore strong phenotypic integration (i.e. high trait coordination), whereas low INT_c values meant that phenotypic integration was weak in a particular cytotype (i.e. low

trait coordination). Less overlapping confidence intervals among cytotypes were considered as evidence of variation in phenotypic integration.

2.7 | LES and edaphic niche evolution: Divergence in nutrient uptake and use efficiency

Complementary to both pPCAs (i.e. soil and LES), we tested for phylogenetic signal on the most explanatory axes with global structures using the Blomberg's K (Blomberg et al., 2003) in the 'PHYTOOLS' R package (Revell, 2012). Given the cytotype relatedness pattern, the global structures can be interpreted as a consequence of niche/functional conservatism (following a Brownian motion model of gradual drift; BM) or divergence due to polyploidization (following an Ornstein-Uhlenbeck, OU, model of stabilizing selection with different optima; Butler & King, 2004). To investigate these alternatives, we compared the relative fit of several evolutionary models in case of significant phylogenetic signal: BM as well as an OU model with a single optimum (OU1) and an OU model with four optima (corresponding to one optimum per ploidy; OU4). Models were performed with the 'MVMORPH' R package (Clavel et al., 2015) and compared by the weights of their Akaike information criterion values (AIC). In case of pPCA axes with local structures, we checked them by fitting a null model of trait evolution (NP) through the simulation of a star-shaped phylogenetic tree.

Furthermore, to investigate the degree to which edaphic niche characteristics predicts LES variables (and vice versa; i.e. correlated evolution), we employed procrustean randomization tests (Peres-Neto & Jackson, 2001) through the functions *procrustes* and *protest* in the 'VEGAN' R package. We compared the pPCA scores obtained for each population in the two axes that explained more variation (PC1 and PC2 for the soil-pPCA; PC1 and PC11 for the LES-pPCA), using 9,999 permutations. A significant high correlation (p < 0.05) between PCs indicated interdependence of datasets and correlated evolution of characteristics.

Finally, we also investigated a wide variety of element contents in leaves. Macronutrient uptake and use efficiency in the four *D*. *broteri* cytotypes were determined, for each location, by accounting for the differences between leaves and soils in the concentration of N, K and P (in mg/g of dry matter). The influence of ploidy level on these latter variables was analysed using generalized linear models (one-way ANOVAs) with normal or gamma distribution of errors. Significant results were followed by post-hoc Tukey's HSD tests ($\alpha = 0.05$) to identify inter-cytotype pairwise differences. All statistical analyses were performed in R software ver. 4.0.0 (R Core Team, 2020).

3 | RESULTS

D. broteri populations showed a pattern of inter-cytotype differentiation in the edaphic space, as represented in the two most

explanatory pPCA axes (soil-PC1 and PC2). Both axes explained more than half of the total variance in the edaphic space (37.7% and 20.1%, respectively) and showed positive eigenvalues, so local structures (related to recently diverged traits) had little influence on the variation of soil niche characteristics. Moreover, variable contributions on these axes were generally low (always <0.5 in absolute values), revealing high levels of correlation between edaphic variables. Despite this result, the first axis (soil-PC1) clearly mirrored an alkalinity gradient and the second one (soil-PC2) a nutrient content gradient in soils. In this edaphic space, we noticed niche displacement and expansion of the polyploid cytotypes, being 4x, 6x and 12× niches clearly different and wider than the 2× one. Thus, diploids had the most restricted niche, which almost completely overlapped the tetraploid space (Figure 1B). Congruently, the two lower cytotypes (2× and 4×) did not present significant differences in any pPCA axis (Figure 1C,D) and they were associated with milder edaphic conditions. Their niches were characterized by clay, semiacid soils with high EC as well as elevated contents of essential macronutrients (N, P and Ca) and several important micronutrients (Cu and Mn; Figure 1A,B). Oppositely, the 6x cytotype was located in a significantly divergent position of soil-PC1 compared to the rest of ploidies ($F_{3,8} = 6.18$; p < 0.05; Figure 1C). In this axis, the niche space of hexaploids corresponded to silty and alkaline soils, with $pH \ge 7$ and high levels of total carbonates and active limestone. Lastly, the 12× cytotype appeared in an extreme position of soil-PC2, marginally different from the other cytotypes ($F_{3.8} = 2.37$; p = 0.07; Figure 1D). The niche space of dodecaploids in soil-PC2 was related to nutrient-poor soils (except for Fe), with reduced concentrations of all the essential macronutrients (N, P, K, Ca and Mg), organic matter and electrical conductivity.

D. broteri also demonstrated a notable variation of the LES traits, which was mainly explained by two pPCA axes accounting for 47.7% and 25.0% of the total inertia, respectively (Figure 2A). The first most explanatory axis had a positive eigenvalue but the second one had a negative eigenvalue, indicating that both global and local structures affect to LES covariation in D. broteri. Therefore, in the first axis (LES-PC1), we found the earliest diverged traits, which were those from the classical LES. LES-PC1 reflected intercytotype divergence, being the 12× significantly differentiated from the rest of cytotypes ($F_{3,8} = 16.78$; p < 0.05; Figure 2C). The dodecaploids were characterized in this axis by high levels of leaf macronutrients (N, P and K) and high $\mathsf{PI}_{\mathsf{Total}}$ as well as low LMA and δ^{13} C (Figure 2A,B). In this same axis, the other three ploidies overlapped and appeared in more central positions of the pPCA, with LES traits showed by the 2x cytotype encompassing both 4× and 6× (Figure 2B). Nonetheless, hexaploids were occupying the most opposite location in PC1 compared to dodecaploids. The second represented axis (LES-PC11) gathered recently diverged traits and was mainly characterized by photochemical parameters (TR/CS, ET/CS, DI/CS and F_v/F_m). The scores of LES-PC11 did not differ among cytotypes ($F_{3,8} = 0.19$; p > 0.05), being this axis mostly defined by intra-cytotype variation, specifically within the 4× (Figure 2D). Interestingly, the photosystems efficiency (F_v/F_m)



FIGURE 1 Phylogenetic principal component analysis (pPCA) obtained for the sampled populations of *Dianthus broteri*, using soil-related variables regarding nutrient composition and physicochemical properties. Only the two axes with greater inertia explained are represented. Panel (A) shows the distribution of the selected variables loading on the axes. Panel (B) presents the edaphic niches of the four cytotypes (ellipses encompass occurrence points) in the two main axes. Panels (C) and (D) are a breakdown of (B) representing each axis separately (soil-PC1 and PC2, respectively). These show the edaphic range of the ploidies as violin plots, where white circles correspond to the median, thick black bars to the interquartile range and thin bars to the 95% confidence interval. Different letters indicate cytotypes that are significantly different from each other (one-way ANOVA: p < 0.05, Tukey's HSD test: $\alpha = 0.05$). Letters with asterisks designate marginally significant differences (Tukey's HSD test: $0.05 < \alpha < 0.10$)

and performance (PI_{Total}) showed opposite patterns for all the ploidies considering their reverse contribution signs in both axes (Figure 2A).

Overall, the well-described covariation relationships for functional traits from the LES were maintained in *D. broteri*. A strong positive relationship was found between the essential macronutrients N and P (r = 0.72; p < 0.05), which were negatively related to δ^{13} C (Figure 3). This isotopic composition was, in turn, negatively related to δ^{15} N but positively to LMA. However, photochemical variables were separated from the classical LES traits in this covariation network but integrated between them (Figure 3). Thus, the total performance index (Pl_{Total}) presented tight positive correlations with the absorbed (ABS/CS; r = 0.62; p < 0.05) and dissipated fluxes (DI/CS; r = 0.90; p < 0.05), but the relationship was markedly negative with F_v/F_m (r = -0.85; p < 0.05). This latter parameter of PSII efficiency was more related to fluxes of primary photochemistry (TR/CS and ET/CS; Figure 3). Furthermore, the direction of fundamental, globally described, leaf trait-trait relationships (those between LMA, N and P; Wright et al., 2004) were maintained within cytotypes.

However, we found differences when comparing the coordination of functional traits among ploidies considering the complete network. The 2× cytotype had the highest integration of LES traits, which decreased with polyploidization, being $6\times$ and $12\times$ cytotypes



FIGURE 2 Phylogenetic principal component analysis (pPCA) obtained for the sampled populations of *Dianthus broteri*, using leaf economics spectrum (LES) and other functional variables. Only the two axes with greater inertia explained are represented. Panel (A) shows the distribution of the selected variables loading on the axes and panel (B) presents the functional space of the four cytotypes (ellipses encompass occurrence points) in the two main axes. Panels (C) and (D) are a breakdown of (B) representing each axis separately (LES-PC1 and PC11, respectively). These show the functional range of the ploidies as violin plots, where white circles correspond to the median, thick black bars to the interquartile range and thin bars to the 95% confidence interval. Different letters indicate cytotypes that are significantly different from each other (one-way ANOVA: p < 0.05, Tukey's HSD test: $\alpha = 0.05$)

those with the lowest INT_c and percentages of maximum integration (%INT_{max}; Table 1). Additionally, the Mantel tests revealed a greatly dissimilar structure of LES traits (lowest Pearson's *r*) between diploids and the other cytotypes (4×, 6× and 12×; Table S2). In contrast, these three cytotypes showed similar LES traits covariation matrices, although the Mantel tests were not significant (p > 0.05; Table S2).

In both soil and LES pPCAs, we only found significant phylogenetic signal ($K \ge 1, p < 0.05$) for the most explanatory axes (PC1s). The variation in these axes better fitted the OU4 model (i.e. the one considering a scenario with four optima, corresponding to the four cytotypes), showing AIC weights above 75 (virtually 100 for LES-PC1; Table 2). These results supported the conservatism of the edaphic niche and functional traits at population level (i.e. within ploidies), but the divergence among *D. broteri* cytotypes. Nevertheless, both sets of characteristics were not correlated according to the results of the procrustean tests. The two pairs of pPCA scores did not show a pattern of multivariate concerted evolution, based on their low and non-significant correlation (r = 0.36; p > 0.05).

In contrast to the above-mentioned divergence in soil nutrient contents among *D. broteri* cytotypes, their leaves showed a clearly distinct pattern, marked by significantly higher contents of essential macronutrients in the 12× cytotype (C, N, P and K; p < 0.05; Table S3) and a greater variation in the 2× cytotype (as indicated its overall higher standard errors; Table S3). Thus, nutrient acquisition and accumulation rates and efficiencies differed among ploidies, especially affecting 6× and 12×. With regard to the variables accounting for the difference in macronutrient concentrations between leaf



FIGURE 3 Correlation network of functional variables regarding nutrient, water and light use in the sampled populations of *Dianthus broteri*. Tightly related variables appear closer and lines connecting them are wider and have darker colours (red for positive relationships and blue for negative ones). Only correlations with Pearson's r > 0.3 are drawn

 TABLE 1
 Summary of the integration analysis in the leaf

 economics spectrum traits for each Dianthus broteri cytotype

Ploidy	N	INT _c	Lower Cl	Upper Cl	%INT _{max}
2×	64	3.04 ± 1.00	2.29	3.33	25.33
4×	66	2.95 ± 0.90	2.49	3.10	24.58
6×	64	2.39 ± 0.93	1.83	2.52	19.92
12×	81	2.70 ± 0.88	2.41	2.76	22.50

Note: Values represent mean \pm *SD*.

Abbreviations: %INT_{max}, percentage of the total maximum integration; CI, confidence interval; INTc, corrected phenotypic integration index; *N*, total number of sampled individuals.

TABLE 2Performance of models for the evolution of leaf
economic traits and soil niche properties in *Dianthus broteri*. Only
pPCA axes with significant phylogenetic signal are represented.
Models are ranked according to their Akaike information criterion
(AIC) weight

LES-PC1			Soil-PC1			
Model	AIC	Weight	Model	AIC	Weight	
OU4	39.2	99.1	OU4	48.3	77.9	
BM	49.3	0.6	BM	51.8	13.8	
OU1	51.3	0.2	OU1	53.8	5.1	
NP	55.1	0.0	NP	54.7	3.2	

and soil, dodecaploids presented again the overall most elevated values (Figure 4A–C). Thus, the 12× cytotype had significantly greater differences in N than the rest of ploidies (ANOVA: $F_{3,8} = 6.66$; p < 0.05; Figure 4A). Dodecaploids also exhibited marginally higher differences in K ($F_{3,8} = 2.23$; p = 0.08; Figure 4B) and P ($F_{3,8} = 2.39$; p = 0.07; Figure 4C) compared to 2× and 6× cytotypes, respectively. We also detected large differences in several micronutrients such as Na, which was significantly higher in the 6× cytotype than in the remaining ones ($F_{3,8} = 5.17$; p < 0.05; Table S3).



FIGURE 4 Violin plots representing differences in the main macronutrients between leaves from the four *Dianthus broteri* cytotypes and soils from their respective niches. Panel (A) corresponds to nitrogen, panel (B) to potassium and panel (C) to phosphorus. For each element, higher values represent more efficient acquisition rates from soil to leaves. In these plots, white circles depict the median, thick black bars correspond to the interquartile range and thin bars show the 95% confidence interval. Different letters indicate cytotypes that are significantly different from each other (one-way ANOVA: p < 0.05, Tukey's HSD test: $\alpha = 0.05$). Letters with asterisks designate marginally significant differences (Tukey's HSD test: $0.05 < \alpha < 0.10$)

4 | DISCUSSION

4.1 | Soil niche divergence and expansion associated with polyploidy

Most of the variation in the edaphic niche of *D. broteri* would be explained by processes associated with the short-term effects of polyploidization as suggested the *global structures* found in the two main soil-pPCA axes. The immediate phenotypic novelties derived from genome duplication, such as those found in *D. broteri* synthetic neotetraploids for several functional traits similar to the ones explored in this study (gas-exchange and photochemical variables; Domínguez-Delgado et al., 2021), would be the main drivers of the divergence in soil niche. However, an early post-polyploidization character displacement caused by competition for niche space (Beans, 2014) or niche filling cannot be ruled out. The highest cytotypes (6x and 12x) would have been displaced to stressful environmental edges with much poorer soils in terms of macronutrient content and potential availability than the lower cytotypes (López-Jurado et al., 2019). Furthermore, the divergence pattern was accompanied by an expansion of the edaphic niche in polyploids, which could be linked with amplified ecological tolerances (Molina-Henao & Hopkins, 2019; te Beest et al., 2012) and/or enhanced competitive abilities (Rey et al., 2017).

4.2 | Divergent positioning along the LES associated with polyploidy

Although soil variables are supposed to be the strongest environmental predictors of leaf traits (Maire et al., 2015), the results of the procrustean tests linking edaphic niche characteristics and LES variables suggested that they are not coevolving. Nevertheless, functional traits related to nutrient (N, P and K) and water economy (δ^{13} C) explained by LES-PC1 also showed an early divergence (global structure) associated with polyploidization. Leal-Bertioli et al. (2017) and Wei et al. (2020) also demonstrated large effects of polyploidy per se on leaf morphology and anatomy by finding analogous results in wild and induced polyploids. Moreover, Münzbergová and Haisel (2019) found a similar pattern for the photosynthetic pigment contents in autotetraploids.

In agreement with soil-PC1, each cytotype reached a different LES-PC1 optimum given the highest performance of OU4 models. Thus, divergent selection was the major force driving the evolution of D. broteri polyploids in functional traits and soil niche characteristics, which agreed with a scenario of inter-cytotypic competitive interactions (Laport et al., 2013; Rey et al., 2017). In contrast to its most restricted edaphic niche, the $2 \times$ cytotype had the overall greatest variation in leaf nutrient concentrations. This fact could be explained by larger pools of genetic variation in this cytotype than the other ploidies (Balao et al., 2010). Notwithstanding the higher genetic variation of the diploids, they showed reduced leaf nutrient contents, which were similar to those of $4 \times$ and $6 \times$ cytotypes. However, we would have expected the leaf nutrients to be increased in the polyploids based on the greater need for these in taxa with larger genomes (Leitch & Leitch, 2008) and the complex polygenic uptake mechanisms for essential macronutrients (N, P and K; Lea & Azevedo, 2006; Vance et al., 2003; Véry & Sentenac, 2003), which would be facilitated by post-polyploidization processes (Cheng et al., 2018). This finding could be explained by the phenotypic diploidization phenomenon, an evolutionary tendency of returning to diploid-like phenotype which has been recently demonstrated for photosynthetic characteristics of established D. broteri 4× individuals (Domínguez-Delgado et al., 2021).

Oppositely, the highest-order cytotype (12×) showed, as expected, an elevated leaf accumulation of K, P and N. On the basis of the low-P and N soils where the 12× ploidy is distributed, their elevated content in leaves would be useful for dealing with uncertainties in availability (Clarkson et al., 2005; Ostertag, 2010; Ruiz-Navarro et al., 2016). Moreover, dodecaploids would benefit from a higher K content because this element has a key role in numerous stress response mechanisms (e.g. drought, cold or salinity; Wang et al., 2013).

In the D. broteri complex, LES-PC1 reflected the acquisitiveconservative continuum (fast-slow, sensu Reich, 2014), which was in accordance with water-use strategies (through δ^{13} C) as Prieto et al. (2018) described in other Mediterranean species. Meanwhile, the highest ploidy (12×) occupied an extreme of the continuum, the second highest ploidy (6x) was located at the opposite extreme, and 2x and 4x filled more intermediate spaces. The 6x cytotype showed a nutrient and water-conservative or slow strategy (i.e. high LMA, low leaf nutrient accumulation plus high δ^{13} C), whereas the 12× cytotype presented a nutrient and water-acquisitive or rapid strategy (i.e. low LMA, high leaf nutrient accumulation plus low δ^{13} C). The 12x cytotype showed the lowest intrinsic water-use efficiency (WUE; estimated from leaf δ^{13} C), which would explain its elevated leaf nutrient concentrations since the low WUE is offset with high nutrient uptake efficiency through increased stomatal conductance (g.; Cramer et al., 2009). Nevertheless, the other highly droughtexposed cytotype (6x) would prioritize water conservation (causing high ¡WUE) through the alleged reduction of g. The functioning at lower internal CO₂ concentrations (C_i) would help to reduce water loss (Goud et al., 2019). Additionally, the higher LMA in hexaploids would allow a better adjustment of leaf water potential with small changes in water content (optimizing water uptake from dry soils; Niinemets, 2001) and, at the same time, the alleviation of extreme heat stress (Leigh et al., 2012) and the adaptation to high light intensities (Poorter et al., 2009). Congruently, high-LMA species have been extensively associated with semi-arid Mediterranean environments as the one inhabited by the 6x cytotype (e.g. Costa-Saura et al., 2016; de la Riva et al., 2016). However, increased LMA entails generally low photosynthetic rates (A_N) , since it would limit CO_2 diffusion through the mesophyll (Tomás et al., 2013), as well as N investment in photosynthetic proteins because of the higher cell wall mass fraction (Onoda et al., 2017).

Interestingly, the rapid or acquisitive strategy characterizing the 12× cytotype cannot be maintained indefinitely in a perennial plant (Onoda et al., 2017). Therefore, this functional strategy should be promoted in periods of opportunity, limiting the growing season to when water and nutrients become sufficiently available (Querejeta et al., 2018). These adaptations would enable *D. broteri* dodecaploids to cope with the remarkably seasonal Mediterranean climate under which they live, with highly variable precipitation events (Deitch et al., 2017) and long dry spells (Sánchez et al., 2011). Supporting this hypothesis, we have found stomatal closure and increased _iWUE during summer in the 12× cytotype (López-Jurado et al., 2019).

4.3 | The impact of polyploidization and subsequent local processes on phenotypic integration and the flexibility of photochemical responses

Despite the mentioned acquisitive-conservative adaptative strategies were defined by a common network of functional traits for all the cytotypes, a gradual uncoupling of these traits was found with increasing ploidy. Interestingly, the same pattern was previously detected in *D. broteri* using a multivariate morphological study of reproductive and vegetative traits (Balao et al., 2011). The enhanced allelic diversity in autopolyploids (Parisod et al., 2010) could lead to transgressive associations in the pleiotropic genes regulating LES trait covariation, then promoting novel extreme phenotypes (Vasseur et al., 2012). The strong selection on specific adaptive traits during initial phases after polyploid formation would have also contributed to the LES uncoupling in *D. broteri* polyploids. Thus, the higher phenotypic flexibility exhibited by polyploids may facilitated the rapid adaptation of *D. broteri* 4×, 6× and 12× cytotypes to overcome the initial minority cytotype exclusion (Husband et al., 2016; Oswald & Nuismer, 2011).

Nonetheless, divergent selection was not the only driver of functional variation in this complex. The local structures of LES-PC11 (mainly explained by photochemical parameters) suggested a relative role of local selection and/or plasticity, which had already been proposed for the evolution of this complex (Alonso et al., 2016; López-Jurado et al., 2020). In addition, LES-PC11 occupied a different and independent functional dimension from LES-PC1 as a likely effect of inherent physiological constraints, given that shade tolerance is independent of the tolerance to other abiotic stresses (Puglielli et al., 2021). However, although the four cytotypes clearly overlapped in the LES-PC11, polyploids (4x, 6x and 12x) showed higher variation compared to diploids, highlighting the effect of polyploidy on promoting phenotypic plasticity and local adaptation (Chen, 2007; Hahn et al., 2012). The high flexibility and independence of the photochemical behaviour of D. broteri polyploids would contribute to the colonization of heterogeneous light microenvironments at any soil-climate interaction (summarized in soilpPCA and López-Jurado et al., 2019). Interestingly, the highest-order cytotype (12×) showed the highest constitutive PI_{Total} together with the lowest F_v/F_m . This opposite pattern might depend on the contrasting performance of photosystems. While F_v/F_m only accounted for PSII efficiency, $\mathrm{PI}_{\mathrm{Total}}$ is calculated from both PSI and PSII yields. In this regard, PSII could be more prone to ploidy dosage effects due to its larger number of subunits and regulatory loci (Coate et al., 2011), which could cause misassembled PSII complexes, impairing photochemical performance in high polyploids (Baena-González & Aro, 2002; Hwang et al., 2008).

4.4 | Final remarks

Although singularities in the LES at the intraspecific level have been detected (Fajardo & Siefert, 2018), our straightforward results

suggest that polyploidy can be a powerful driving force overcoming limitations in LES universality across evolutionary scales and levels of organization. The results of this study, together with the rapid radiation of polyploid races within D. broteri (Balao et al., 2010), suggested an ongoing scenario of diversification similar to the one found in Allium (governed by a high ploidy diversity and ecological drivers; Han et al., 2020). We demonstrated that inter-cytotype trait divergence in D. broteri is mostly caused by short-term consequences of whole-genome duplication and captured by the LES (almost 50% of the total functional variance was explained by LES-PC1). Trait novelty in early generations of polyploid races would be then related to the higher adaptive and niche exploring capacity of these polyploid genomes to overcome fitness disadvantage compared to diploids (Baker et al., 2017). The dominance of polyploidization effects over subsequent evolution in D. broteri functional traits is likely explained by the relatively recent divergence of the cytotypes (Balao et al., 2010), as Čertner et al. (2019) proposed for Knautia serpentinicola. Nevertheless, we have also shown the influence of local processes after polyploidization events, such as adaptation and phenotypic plasticity, on the divergence of *D. broteri* polyploids through their photochemical responses under variations in environmental conditions. To date, there are no additional studies demonstrating these patterns together, especially dealing with several cytotypes and using a multi-resource-based LES, accounting for nutrient, water and light. Albeit this line of research clearly deserves further investigation, our work is the first one unambiguously reflecting the LES mediated by autopolyploidy at the microevolutionary scale. Finally, we generated new evidence for the inclusion of photochemical variables as an additional dimension of the LES to explain light acquisition and use in plants.

ACKNOWLEDGEMENTS

We thank the University of Seville Herbarium and Mass Spectrometry General Services for their collaboration and providing facilities and equipment. We are also grateful to Innoagral laboratories and CEBAS-CSIC Ionomics Service for their efficient work in soil and leaf analyses. Thanks to the two anonymous referees and editor for their constructive and helpful comments on previous versions of the manuscript. This study was supported by the Research Project PGC2018-098358-B-I00 from the Spanish MICINN, Áridos La Melera S.L. (FIUS project 2234/0724) and fellowships to J.L.-J. from the V and VI PPIT-US (Fifth and Sixth Research and Transfer Plans from the University of Seville).

CONFLICT OF INTEREST

The authors have no conflict of interest to declare.

AUTHORS' CONTRIBUTIONS

F.B. and E.M.-N. conceived the idea; J.L.-J., E.M.-N. and F.B. gathered the data; F.B. and J.L.-J. designed the analyses; J.L.-J. performed the analyses and wrote the first draft of the manuscript; all authors interpreted the results, provided corrections to manuscript drafts and discussed ideas within it.

PEER REVIEW

The peer review history for this article is available at https://publons. com/publon/10.1111/1365-2745.13823.

DATA AVAILABILITY STATEMENT

Data available from the Dryad Digital Repository https://doi. org/10.5061/dryad.nvx0k6dtp (López-Jurado et al., 2021a) and R scripts from Zenodo https://doi.org/10.5281/zenodo.5742426 (López-Jurado et al., 2021b).

ORCID

Javier López-Jurado https://orcid.org/0000-0002-6354-0800 Enrique Mateos-Naranjo https://orcid.org/0000-0001-6276-5664 Francisco Balao https://orcid.org/0000-0003-2104-3846

REFERENCES

- Alonso, C., Balao, F., Bazaga, P., & Pérez, R. (2016). Epigenetic contribution to successful polyploidizations: Variation in global cytosine methylation along an extensive ploidy series in *Dianthus broteri* (Caryophyllaceae). *New Phytologist*, 212(3), 571–576. https://doi. org/10.1111/nph.14138
- Baena-González, E., & Aro, E.-M. (2002). Biogenesis, assembly and turnover of photosystem II units. Philosophical Transactions of the Royal Society B: Biological Sciences, 357(1426), 1451–1460. https://doi. org/10.1098/rstb.2002.1141
- Baker, R. L., Yarkhunova, Y., Vidal, K., Ewers, B. E., & Weinig, C. (2017). Polyploidy and the relationship between leaf structure and function: Implications for correlated evolution of anatomy, morphology, and physiology in *Brassica. BMC Plant Biology*, 17, 3. https://doi. org/10.1186/s12870-016-0957-3
- Balao, F., Casimiro-Soriguer, R., Talavera, M., Herrera, J., & Talavera, S. (2009). Distribution and diversity of cytotypes in *Dianthus broteri* as evidenced by genome size variations. *Annals of Botany*, 104(5), 965–973. https://doi.org/10.1093/aob/mcp182
- Balao, F., Herrera, J., & Talavera, S. (2011). Phenotypic consequences of polyploidy and genome size at the microevolutionary scale: A multivariate morphological approach. New Phytologist, 192(1), 256–265. https://doi.org/10.1111/j.1469-8137.2011.03787.x
- Balao, F., Herrera, J., Talavera, S., & Dötterl, S. (2011). Spatial and temporal patterns of floral scent emission in *Dianthus inoxianus* and electroantennographic responses of its hawkmoth pollinator. *Phytochemistry*, 72(7), 601–609. https://doi.org/10.1016/j.phytochem.2011.02.001
- Balao, F., Valente, L. M., Vargas, P., Herrera, J., & Talavera, S. (2010). Radiative evolution of polyploid races of the Iberian carnation *Dianthus broteri* (Caryophyllaceae). *New Phytologist*, 187(2), 542– 551. https://doi.org/10.1111/j.1469-8137.2010.03280.x
- Beans, C. M. (2014). The case for character displacement in plants. *Ecology* and Evolution, 4(6), 862–875. https://doi.org/10.1002/ece3.978
- Blomberg, S. P., Garland Jr, T., & Ives, A. R. (2003). Testing for phylogenetic signal in comparative data: Behavioral traits are more labile. *Evolution*, 57(4), 717-745. https://doi.org/10.1111/j.0014-3820. 2003.tb00285.x
- Brouillette, L. C., Mason, C. M., Shirk, R. Y., & Donovan, L. A. (2014). Adaptive differentiation of traits related to resource use in a desert annual along a resource gradient. *New Phytologist*, 201(4), 1316– 1327. https://doi.org/10.1111/nph.12628
- Butler, M. A., & King, A. A. (2004). Phylogenetic comparative analysis: A modeling approach for adaptive evolution. *The American Naturalist*, 164(6), 683–695. https://doi.org/10.1086/426002
- Carvalho, B., Bastias, C. C., Escudero, A., Valladares, F., & Benavides, R. (2020). Intraspecific perspective of phenotypic coordination of

functional traits in Scots pine. *PLoS ONE*, 15(2), e0228539. https://doi.org/10.1371/journal.pone.0228539

- Čertner, M., Sudová, R., Weiser, M., Suda, J., & Kolář, F. (2019). Ploidyaltered phenotype interacts with local environment and may enhance polyploid establishment in *Knautia serpentinicola* (Caprifoliaceae). *New Phytologist*, 221, 1117–1127. https://doi. org/10.1111/nph.15426
- Chen, Z. J. (2007). Genetic and epigenetic mechanisms for gene expression and phenotypic variation in plant polyploids. Annual Review of Plant Biology, 58(1), 377–406. https://doi.org/10.1146/annurev.arplant.58.032806.103835
- Cheng, F., Wu, J., Cai, X., Liang, J., Freeling, M., & Wang, X. (2018). Gene retention, fractionation and subgenome differences in polyploid plants. *Nature Plants*, 4(5), 258–268. https://doi.org/10.1038/ s41477-018-0136-7
- Cheverud, J. M., Wagner, G. P., & Dow, M. M. (1989). Methods for the comparative analysis of variation patterns. *Systematic Zoology*, 38(3), 201–213. https://doi.org/10.2307/2992282
- Clarkson, B. R., Schipper, L. A., Moyersoen, B., & Silvester, W. B. (2005). Foliar ¹⁵N natural abundance indicates phosphorus limitation of bog species. *Oecologia*, 144(4), 550–557. https://doi.org/10.1007/ s00442-005-0033-4
- Clavel, J., Escarguel, G., & Merceron, G. (2015). mvMORPH: An R package for fitting multivariate evolutionary models to morphometric data. *Methods in Ecology and Evolution*, 6(11), 1311–1319. https:// doi.org/10.1111/2041-210X.12420
- Coate, J. E., Schlueter, J. A., Whaley, A. M., & Doyle, J. J. (2011). Comparative evolution of photosynthetic genes in response to polyploid and nonpolyploid duplication. *Plant Physiology*, 155(4), 2081–2095. https://doi.org/10.1104/pp.110.169599
- Costa-Saura, J. M., Martínez-Vilalta, J., Trabucco, A., Spano, D., & Mereu, S. (2016). Specific leaf area and hydraulic traits explain niche segregation along an aridity gradient in Mediterranean woody species. *Perspectives in Plant Ecology, Evolution and Systematics*, 21, 23–30. https://doi.org/10.1016/j.ppees.2016.05.001
- Cramer, M. D., Hawkins, H. J., & Verboom, G. A. (2009). The importance of nutritional regulation of plant water flux. *Oecologia*, 161(1), 15– 24. https://doi.org/10.1007/s00442-009-1364-3
- Damián, X., Ochoa-López, S., Gaxiola, A., Fornoni, J., Domínguez, C. A., & Boege, K. (2020). Natural selection acting on integrated phenotypes: Covariance among functional leaf traits increases plant fitness. New Phytologist, 225(1), 546–557. https://doi.org/10.1111/ nph.16116
- de la Riva, E. G., Olmo, M., Poorter, H., Ubera, J. L., & Villar, R. (2016). Leaf mass per area (LMA) and its relationship with leaf structure and anatomy in 34 mediterranean woody species along a water availability gradient. *PLoS ONE*, 11(2), e0148788. https://doi. org/10.1371/journal.pone.0148788
- Deitch, M. J., Sapundjieff, M. J., & Feirer, S. T. (2017). Characterizing precipitation variability and trends in the world's Mediterraneanclimate areas. *Water*, 9(4), 259. https://doi.org/10.3390/w9040259
- Díaz, S., Kattge, J., Cornelissen, J. H. C., Wright, I. J., Lavorel, S., Dray, S., Reu, B., Kleyer, M., Wirth, C., Colin Prentice, I., Garnier, E., Bönisch, G., Westoby, M., Poorter, H., Reich, P. B., Moles, A. T., Dickie, J., Gillison, A. N., Zanne, A. E., ... Gorné, L. D. (2016). The global spectrum of plant form and function. *Nature*, *529*, 167–171. https://doi. org/10.1038/nature16489
- Domínguez-Delgado, J. J., López-Jurado, J., Mateos-Naranjo, E., & Balao, F. (2021). Phenotypic diploidization in plant functional traits uncovered by synthetic neopolyploids in *Dianthus broteri*. *Journal of Experimental Botany*, 72, erab179. https://doi.org/10.1093/jxb/erab179
- Donovan, L. A., Maherali, H., Caruso, C. M., Huber, H., & de Kroon, H. (2011). The evolution of the worldwide leaf economics spectrum. *Trends in Ecology & Evolution*, 26(2), 88–95. https://doi. org/10.1016/j.tree.2010.11.011

- Easlon, H. M., & Bloom, A. J. (2014). Easy leaf area: Automated digital image analysis for rapid and accurate measurement of leaf area. Applications in Plant Sciences, 2(7), 1400033. https://doi. org/10.3732/apps.1400033
- Evans, J. R. (1989). Photosynthesis and nitrogen relationships in leaves of C₃ plants. *Oecologia*, 78, 9–19. https://doi.org/10.1007/BF003 77192
- Fajardo, A., & Siefert, A. (2018). Intraspecific trait variation and the leaf economics spectrum across resource gradients and levels of organization. *Ecology*, 99(5), 1024–1030. https://doi.org/10.1002/ ecy.2194
- Farquhar, G. D., Ehleringer, J. R., & Hubick, K. T. (1989). Carbon isotope discrimination and photosynthesis. Annual Review of Plant Biology, 40, 503–537. https://doi.org/10.1146/annurev.pp.40.060189.002443
- Fernandez-Hernandez, A., Mateos, R., Garcia-Mesa, J. A., Beltran, G., & Fernandez-Escobar, R. (2010). Determination of mineral elements in fresh olive fruits by flame atomic spectrometry. *Spanish Journal* of Agricultural Research, 8(4), 1183–1190. https://doi.org/10.5424/ sjar/2010084-1206
- Flagel, L. E., & Wendel, J. F. (2009). Gene duplication and evolutionary novelty in plants. *New Phytologist*, 183, 557–564. https://doi. org/10.1111/j.1469-8137.2009.02923.x
- Goud, E. M., Sparks, J. P., Fishbein, M., & Agrawal, A. A. (2019). Integrated metabolic strategy: A framework for predicting the evolution of carbon-water tradeoffs within plant clades. *Journal of Ecology*, 107(4), 1633–1644. https://doi.org/10.1111/1365-2745.13204
- Hahn, M. A., van Kleunen, M., & Müller-Schärer, H. (2012). Increased phenotypic plasticity to climate may have boosted the invasion success of polyploid *Centaurea stoebe*. *PLoS ONE*, 7(11), e50284. https://doi.org/10.1371/journal.pone.0050284
- Hallik, L., Niinemets, Ü., & Kull, O. (2012). Photosynthetic acclimation to light in woody and herbaceous species: A comparison of leaf structure, pigment content and chlorophyll fluorescence characteristics measured in the field. *Plant Biology*, 14, 88–99. https://doi. org/10.1111/j.1438-8677.2011.00472.x
- Hallik, L., Niinemets, Ü., & Wright, I. J. (2009). Are species shade and drought tolerance reflected in leaf-level structural and functional differentiation in Northern Hemisphere temperate woody flora? New Phytologist, 184, 257–274. https://doi. org/10.1111/j.1469-8137.2009.02918.x
- Han, T. S., Zheng, Q.-J., Onstein, R. E., Rojas-Andrés, B. M., Hauenschild, F., Muellner-Riehl, A. N., & Xing, Y.-W. (2020). Polyploidy promotes species diversification of *Allium* through ecological shifts. *New Phytologist*, 225(1), 571–583. https://doi.org/10.1111/nph.16098
- Husband, B. C., Baldwin, S. J., & Sabara, H. A. (2016). Direct vs. indirect effects of whole-genome duplication on prezygotic isolation in *Chamerion angustifolium*: Implications for rapid speciation. *American Journal of Botany*, 103(7), 1259–1271. https://doi.org/10.3732/ ajb.1600097
- Hwang, H. J., Nagarajan, A., McLain, A., & Burnap, R. L. (2008). Assembly and disassembly of the photosystem II manganese cluster reversibly alters the coupling of the reaction center with the lightharvesting phycobilisome. *Biochemistry*, 47(37), 9747–9755. https:// doi.org/10.1021/bi800568p
- Ji, W., LaZerte, S. E., Waterway, M. J., & Lechowicz, M. J. (2020). Functional ecology of congeneric variation in the leaf economics spectrum. New Phytologist, 225, 196–208. https://doi.org/10.1111/ nph.16109
- Jombart, T., Balloux, F., & Dray, S. (2010). adephylo: New tools for investigating the phylogenetic signal in biological traits. Bioinformatics, 26(15), 1907–1909. https://doi.org/10.1093/bioinformatics/ btq292
- Jombart, T., Pavoine, S., Devillard, S., & Pontier, D. (2010). Putting phylogeny into the analysis of biological traits: A methodological approach. *Journal of Theoretical Biology*, 264(3), 693–701. https://doi. org/10.1016/j.jtbi.2010.03.038

- Kalcsits, L. A., Buschhaus, H. A., & Guy, R. D. (2014). Nitrogen isotope discrimination as an integrated measure of nitrogen fluxes, assimilation and allocation in plants. *Physiologia Plantarum*, 151, 293–304. https://doi.org/10.1111/ppl.12167
- Kalisz, S., & Kramer, E. M. (2008). Variation and constraint in plant evolution and development. *Heredity*, 100(2), 171–177. https://doi. org/10.1038/sj.hdy.6800939
- Kuhn, M., Jackson, S., & Cimentada, J. (2020). corrr: Correlations in R. Retrieved from https://CRAN.R-project.org/package=corrr; https://github.com/tidymodels/corrr
- Lambers, H., & Poorter, H. (1992). Inherent variation in growth rate between higher plants: A search for physiological causes and ecological consequences. Advances in Ecological Research, 23, 187–261. https://doi.org/10.1016/S0065-2504(03)34004-8
- Lambers, H., Raven, J. A., Shaver, G. R., & Smith, S. E. (2008). Plant nutrient-acquisition strategies change with soil age. *Trends in Ecology & Evolution*, 23(2), 95–103. https://doi.org/10.1016/j. tree.2007.10.008
- Laport, R. G., Hatem, L., Minckley, R. L., & Ramsey, J. (2013). Ecological niche modeling implicates climatic adaptation, competitive exclusion, and niche conservatism among *Larrea tridentata* cytotypes in North American deserts. *Journal of the Torrey Botanical Society*, 140(3), 349–363.
- Lea, P. J., & Azevedo, R. A. (2006). Nitrogen use efficiency. 1. Uptake of nitrogen from the soil. Annals of Applied Biology, 149(3), 243–247. https://doi.org/10.1111/j.1744-7348.2006.00101.x
- Leal-Bertioli, S. C. M., Moretzsohn, M. C., Santos, S. P., Brasileiro, A. C. M., Guimarães, P. M., Bertioli, D. J., & Araujo, A. C. G. (2017). Phenotypic effects of allotetraploidization of wild Arachis and their implications for peanut domestication. American Journal of Botany, 104(3), 379–388. https://doi.org/10.3732/ajb.1600402
- Leigh, A., Sevanto, S., Ball, M. C., Close, J. D., Ellsworth, D. S., Knight, C. A., Nicotra, A. B., & Vogel, S. (2012). Do thick leaves avoid thermal damage in critically low wind speeds? *New Phytologist*, 194(2), 477– 487. https://doi.org/10.1111/j.1469-8137.2012.04058.x
- Leitch, A. R., & Leitch, I. J. (2008). Genomic plasticity and the diversity of polyploid plants. *Science*, 320(5875), 481–483. https://doi. org/10.1126/science.1153585
- López-Jurado, J., Balao, F., & Mateos-Naranjo, E. (2016). Deciphering the ecophysiological traits involved during water stress acclimation and recovery of the threatened wild carnation, *Dianthus inoxianus*. *Plant Physiology and Biochemistry*, 109, 397–405. https://doi. org/10.1016/j.plaphy.2016.10.023
- López-Jurado, J., Balao, F., & Mateos-Naranjo, E. (2020). Polyploidymediated divergent light-harvesting and photoprotection strategies under temperature stress in a Mediterranean carnation complex. Environmental and Experimental Botany, 171, 103956. https:// doi.org/10.1016/j.envexpbot.2019.103956
- López-Jurado, J., Mateos-Naranjo, E., & Balao, F. (2019). Niche divergence and limits to expansion in the high polyploid *Dianthus broteri* complex. *New Phytologist*, 222(2), 1076–1087. https://doi. org/10.1111/nph.15663
- López-Jurado, J., Mateos-Naranjo, E., & Balao, F. (2021a). Data from: Polyploidy promotes divergent evolution across the leaf economics spectrum and plant edaphic niche in the *Dianthus broteri* complex. *Dryad Digital Repository*, https://doi.org/10.5061/dryad. nvx0k6dtp
- López-Jurado, J., Mateos-Naranjo, E. & Balao, F. (2021b). Scripts from: Polyploidy promotes divergent evolution across the leaf economics spectrum and plant edaphic niche in the *Dianthus broteri* complex. *Zenodo Repository*, https://doi.org/10.5281/zenodo.5742426
- López-Jurado, J., Mateos-Naranjo, E., García-Castaño, J. L., & Balao, F. (2019). Conditions for translocation of a key threatened species, *Dianthus inoxianus* Gallego, in the southwestern Iberian Mediterranean forest. *Forest Ecology and Management*, 446, 1–9. https://doi.org/10.1016/j.foreco.2019.05.008

- Luo, W., Elser, J. J., Lü, X. T., Wang, Z., Bai, E., Yan, C., Wang, C., Li, M.-H., Zimmermann, N. E., Han, X., Xu, Z., Li, H., Wu, Y., & Jiang, Y. (2015). Plant nutrient do not covary with soil nutrients under changing climatic conditions. *Global Biogeochemical Cycles*, 29(September), 1298–1308. https://doi.org/10.1002/2015GB005089
- Luo, W., Sardans, J., Dijkstra, F. A., Peñuelas, J., Lü, X.-T., Wu, H., Li, M.-H., Bai, E., Wang, Z., Han, X., & Jiang, Y. (2016). Thresholds in decoupled soil-plant elements under changing climatic conditions. *Plant and Soil*, 409(1–2), 159–173. https://doi.org/10.1007/s1110 4-016-2955-5
- Lusk, C. H., Reich, P. B., Montgomery, R. A., Ackerly, D. D., & Cavender-Bares, J. (2008). Why are evergreen leaves so contrary about shade? *Trends in Ecology & Evolution*, 23(6), 299–303. https://doi. org/10.1016/j.tree.2008.02.006
- Maire, V., Wright, I. J., Prentice, I. C., Batjes, N. H., Bhaskar, R., van Bodegom, P. M., Cornwell, W. K., Ellsworth, D., Niinemets, Ü., Ordonez, A., Reich, P. B., & Santiago, L. S. (2015). Global effects of soil and climate on leaf photosynthetic traits and rates. *Global Ecology and Biogeography*, 24(6), 706–717. https://doi.org/10.1111/ geb.12296
- Martínez, L. M., Fernández-Ocaña, A., Rey, P. J., Salido, T., Amil-Ruiz, F., & Manzaneda, A. J. (2018). Variation in functional responses to water stress and differentiation between natural allopolyploid populations in the *Brachypodium distachyon* species complex. *Annals of Botany*, 121, 1369–1382. https://doi.org/10.1093/aob/mcy037
- Mason, C. M., Goolsby, E. W., Humphreys, D. P., & Donovan, L. A. (2016). Phylogenetic structural equation modelling reveals no need for an 'origin' of the leaf economics spectrum. *Ecology Letters*, 19, 54–61. https://doi.org/10.1111/ele.12542
- Messier, J., McGill, B. J., Enquist, B. J., & Lechowicz, M. J. (2017). Trait variation and integration across scales: Is the leaf economic spectrum present at local scales? *Ecography*, 40, 685–697. https://doi. org/10.1111/ecog.02006
- Molina-Henao, Y. F., & Hopkins, R. (2019). Autopolyploid lineage shows climatic niche expansion but not divergence in Arabidopsis arenosa. American Journal of Botany, 106(1), 61–70. https://doi.org/10.1002/ ajb2.1212
- Münzbergová, Z., & Haisel, D. (2019). Effects of polyploidization on the contents of photosynthetic pigments are largely populationspecific. *Photosynthesis Research*, 140(3), 289–299. https://doi. org/10.1007/s11120-018-0604-y
- Niinemets, Ü. (2001). Global-scale climatic controls of leaf dry mass per area, density, and thickness in trees and shrubs. *Ecology*, 82(2), 453– 469. https://doi.org/10.1890/0012-9658(2001)082[0453:GSCCO L]2.0.CO;2
- Niinemets, Ü. (2015). Is there a species spectrum within the world-wide leaf economics spectrum? Major variations in leaf functional traits in the Mediterranean sclerophyll *Quercus ilex*. *New Phytologist*, 205, 79–96.
- Niinemets, Ü., & Kull, K. (2005). Co-limitation of plant primary productivity by nitrogen and phosphorus in a species-rich wooded meadow on calcareous soils. *Acta Oecologica*, 28, 345–356. https://doi. org/10.1016/j.actao.2005.06.003
- Onoda, Y., Wright, I. J., Evans, J. R., Hikosaka, K., Kitajima, K., Niinemets, Ü., Poorter, H., Tosens, T., & Westoby, M. (2017). Physiological and structural tradeoffs underlying the leaf economics spectrum. *New Phytologist*, 214(4), 1447–1463. https://doi.org/10.1111/nph.14496
- Osborn, T. C., Chris Pires, J., Birchler, J. A., Auger, D. L., Jeffery Chen, Z., Lee, H.-S., Comai, L., Madlung, A., Doerge, R. W., Colot, V., & Martienssen, R. A. (2003). Understanding mechanisms of novel gene expression in polyploids. *Trends in Genetics*, 19(3), 141–147. https://doi.org/10.1016/S0168-9525(03)00015-5
- Ostertag, R. (2010). Foliar nitrogen and phosphorus accumulation responses after fertilization: An example from nutrientlimited Hawaiian forests. *Plant and Soil*, 334, 85–98. https://doi. org/10.1007/s11104-010-0281-x

- Oswald, B. P., & Nuismer, S. L. (2011). Neopolyploidy and diversification in *Heuchera grossulariifolia*. Evolution, 65(6), 1667–1679. https://doi. org/10.1111/j.1558-5646.2010.01208.x
- Paradis, E., Claude, J., & Strimmer, K. (2004). APE: Analyses of phylogenetics and evolution in R language. *Bioinformatics*, 20(2), 289–290. https://doi.org/10.1093/bioinformatics/btg412
- Parisod, C., Holderegger, R., & Brochmann, C. (2010). Evolutionary consequences of autopolyploidy. New Phytologist, 186(1), 5–17. https:// doi.org/10.1111/j.1469-8137.2009.03142.Keywords
- Peres-Neto, P. R., & Jackson, D. A. (2001). How well do multivariate data sets match? The advantages of a procrustean superimposition approach over the Mantel test. *Oecologia*, 129(2), 169–178. https:// doi.org/10.1007/s004420100720
- Pérez-Harguindeguy, N., Díaz, S., Garnier, E., Lavorel, S., Poorter, H., Jaureguiberry, P., Bret-Harte, M. S., Cornwell, W. K., Craine, J. M., Gurvich, D. E., Urcelay, C., Veneklaas, E. J., Reich, P. B., Poorter, L., Wright, I. J., Ray, P., Enrico, L., Pausas, J. G., de Vos, A. C., ... Cornelissen, J. H. C. (2013). New handbook for standardised measurement of plant functional traits worldwide. *Australian Journal of Botany*, *61*, 167–234. https://doi.org/10.1071/BT12225
- Pérez-Ramos, I. M., Roumet, C., Cruz, P., Blanchard, A., Autran, P., & Garnier, E. (2012). Evidence for a 'plant community economics spectrum' driven by nutrient and water limitations in a Mediterranean rangeland of southern France. *Journal of Ecology*, 100, 1315–1327. https://doi.org/10.1111/1365-2745.12000
- Poorter, H., Lambers, H., & Evans, J. R. (2014). Trait correlation networks: A whole-plant perspective on the recently criticized leaf economic spectrum. *New Phytologist*, 201(2), 378–382. https://doi. org/10.1111/nph.12547
- Poorter, H., Niinemets, Ü., Poorter, L., Wright, I. J., & Villar, R. (2009).
 Causes and consequences of variation in leaf mass per area (LMA):
 A meta-analysis. New Phytologist, 182(3), 565–588. https://doi. org/10.1111/j.1469-8137.2009.02830.x
- Prieto, I., Querejeta, J. I., Segrestin, J., Volaire, F., & Roumet, C. (2018). Leaf carbon and oxygen isotopes are coordinated with the leaf economics spectrum in Mediterranean rangeland species. *Functional Ecology*, 32(3), 612–625. https://doi.org/10.1111/1365-2435.13025
- Puglielli, G. (2019). Beyond the concept of winter-summer leaves of Mediterranean seasonal dimorphic species. *Frontiers in Plant Science*, 10, 696. https://doi.org/10.3389/fpls.2019.00696
- Puglielli, G., Hutchings, M. J., & Laanisto, L. (2021). The triangular space of abiotic stress tolerance in woody species: A unified tradeoff model. *New Phytologist*, 229(3), 1354–1362. https://doi.org/ 10.1111/nph.16952
- Puglielli, G., & Varone, L. (2018). Inherent variation of functional traits in winter and summer leaves of Mediterranean seasonal dimorphic species: Evidence of a 'within leaf cohort' spectrum. *AoB PLANTS*, 10, ply027. https://doi.org/10.1093/aobpla/ply027
- Querejeta, J. I., Prieto, I., Torres, P., Campoy, M., Alguacil, M. M., & Roldán, A. (2018). Water-spender strategy is linked to higher leaf nutrient concentrations across plant species colonizing a dry and nutrientpoor epiphytic habitat. *Environmental and Experimental Botany*, 153, 302–310. https://doi.org/10.1016/j.envexpbot.2018.06.007
- R Core Team. (2020). R: A language and environment for statistical computing. R Foundation for Statistical Computing. Retrieved from http:// www.R-project.org/
- Reich, P. B. (2014). The world-wide 'fast-slow' plant economics spectrum: A traits manifesto. *Journal of Ecology*, 102(2), 275–301. https://doi. org/10.1111/1365-2745.12211
- Reich, P. B., Wright, I. J., Cavender-Bares, J., Craine, J. M., Oleksyn, J., Westoby, M., & Walters, M. B. (2003). The evolution of plant functional variation: Traits, spectra, and strategies. *International Journal* of Plant Sciences, 164, S143–S164. https://doi.org/10.1086/374368
- Revell, L. J. (2012). phytools: An R package for phylogenetic comparative biology (and other things). *Methods in Ecology and Evolution*, 3(2), 217–223. https://doi.org/10.1111/j.2041-210X.2011.00169.x

- Rey, P. J., Manzaneda, A. J., & Alcántara, J. M. (2017). The interplay between aridity and competition determines colonization ability, exclusion and ecological segregation in the heteroploid *Brachypodium distachyon* species complex. *New Phytologist*, 215(1), 85–96. https:// doi.org/10.1111/nph.14574
- Rojas-Andrés, B. M., Padilla-García, N., de Pedro, M., López-González, N., Delgado, L., Albach, D. C., Castro, M., Castro, S., Loureiro, J., & Martínez-Ortega, M. M. (2020). Environmental differences are correlated with the distribution pattern of cytotypes in *Veronica* subsection *Pentasepalae* at a broad scale. *Annals of Botany*, 125(3), 471–484.
- Ruiz-Navarro, A., Barberá, G. G., Albaladejo, J., & Querejeta, J. I. (2016). Plant δ¹⁵N reflects the high landscape-scale heterogeneity of soil fertility and vegetation productivity in a Mediterranean semiarid ecosystem. New Phytologist, 212(4), 1030–1043. https://doi. org/10.1111/nph.14091
- Sánchez, E., Domínguez, M., Romera, R., de la Franca, N. L., Gaertner, M. A., Gallardo, C., & Castro, M. (2011). Regional modeling of dry spells over the Iberian Peninsula for present climate and climate change conditions. *Climatic Change*, 107(3), 625–634. https://doi. org/10.1007/s10584-011-0114-9
- Sardans, J., & Peñuelas, J. (2015). Potassium: A neglected nutrient in global change. Global Ecology and Biogeography, 24(3), 261–275. https://doi.org/10.1111/geb.12259
- Sartori, K., Vasseur, F., Violle, C., Baron, E., Gerard, M., Rowe, N., Ayala-Garay, O., Christophe, A., Jalón, L. G. D., Masclef, D., Harscouet, E., Granado, M. D. R., Chassagneux, A., Kazakou, E., & Vile, D. (2019). Leaf economics and slow-fast adaptation across the geographic range of *Arabidopsis thaliana*. *Scientific Reports*, *9*, 10758. https://doi.org/10.1038/s41598-019-46878-2
- Seehausen, O. (2004). Hybridization and adaptive radiation. Trends in Ecology & Evolution, 19(4), 198–207. https://doi.org/10.1016/j. tree.2004.01.003
- Smith, J. L., & Doran, J. W. (1997). Measurement and use of pH and electrical conductivity for soil quality analysis. In J. W. Doran & A. J. Jones (Eds.), *Methods for assessing soil quality* (Vol. 49, pp. 169–185). SSSA Special Publications. https://doi.org/10.2136/sssaspecpub49.c10
- Soltis, D. E., Visger, C. J., Marchant, D. B., & Soltis, P. S. (2016). Polyploidy: Pitfalls and paths to a paradigm. *American Journal of Botany*, 103(7), 1146–1166. https://doi.org/10.3732/ajb.1500501
- Strasser, R. J., Tsimilli-Michael, M., Qiang, S., & Goltsev, V. (2010). Simultaneous in vivo recording of prompt and delayed fluorescence and 820-nm reflection changes during drying and after rehydration of the resurrection plant *Haberlea rhodopensis*. *Biochimica et Biophysica Acta–Bioenergetics*, 1797(6–7), 1313–1326. https://doi. org/10.1016/j.bbabio.2010.03.008
- te Beest, M., Le Roux, J. J., Richardson, D. M., Brysting, A. K., Suda, J., Kubešová, M., & Pyšek, P. (2012). The more the better? The role of polyploidy in facilitating plant invasions. *Annals of Botany*, 109(1), 19–45. https://doi.org/10.1093/aob/mcr277
- Tomás, M., Flexas, J., Copolovici, L., Galmés, J., Hallik, L., Medrano, H., Ribas-Carbó, M., Tosens, T., Vislap, V., & Niinemets, Ü. (2013). Importance of leaf anatomy in determining mesophyll diffusion conductance to CO₂ across species: Quantitative limitations and scaling up by models. *Journal of Experimental Botany*, 64(8), 2269– 2281. https://doi.org/10.1093/jxb/ert086
- Usman, A. I., Seydou, H., Abubakar, A., & Bala, M. S. (2017). Validation of Atomic Absorption Spectroscopy (AAS) for trace elements analysis of environmental samples. *Research & Reviews: Journal of Physics*, 6(2), 8–13. Retrieved from www.stmjournals.com
- Vance, C. P., Uhde-Stone, C., & Allan, D. L. (2003). Phosphorus acquisition and use: Critical adaptations by plants for securing a

nonrenewable resource. *New Phytologist*, 157, 423–447. https://doi. org/10.1046/j.1469-8137.2003.00695.x

- Vasseur, F., Violle, C., Enquist, B. J., Granier, C., & Vile, D. (2012). A common genetic basis to the origin of the leaf economics spectrum and metabolic scaling allometry. *Ecology Letters*, 15(10), 1149–1157. https://doi.org/10.1111/j.1461-0248.2012.01839.x
- Véry, A.-A., & Sentenac, H. (2003). Molecular mechanisms and regulation of K⁺ transport in higher plants. *Annual Review of Plant Biology*, 54, 575–603. https://doi.org/10.1146/annurev.plant. 54.031902.134831
- Violle, C., Navas, M., Vile, D., Kazakou, E., Fortunel, C., Hummel, I., & Garnier, E. (2007). Let the concept of trait be functional! *Oikos*, *116*, 882–892. https://doi.org/10.1111/j.2007.0030-1299.15559.x
- Visger, C. J., Germain-Aubrey, C. C., Patel, M., Sessa, E. B., Soltis, P. S., & Soltis, D. E. (2016). Niche divergence between diploid and autotetraploid *Tolmiea*. American Journal of Botany, 103(8), 1396–1406. https://doi.org/10.3732/ajb.1600130
- Volaire, F., Gleason, S. M., & Delzon, S. (2020). What do you mean 'functional' in ecology? Patterns versus processes. *Ecology and Evolution*, 10(21), 11875–11885. https://doi.org/10.1002/ece3.6781
- Wagner, G. P. (1984). On the eigenvalue distribution of genetic and phenotypic dispersion matrices: Evidence for a nonrandom organization of quantitative character variation. *Journal of Mathematical Biology*, 21(1), 77–95. https://doi.org/10.1007/BF00275224
- Wang, M., Zheng, Q., Shen, Q., & Guo, S. (2013). The critical role of potassium in plant stress response. *International Journal of Molecular Sciences*, 14(4), 7370–7390. https://doi.org/10.3390/ijms14047370
- Wei, N., Cronn, R., Liston, A., & Ashman, T.-L. (2019). Functional trait divergence and trait plasticity confer polyploid advantage in heterogeneous environments. New Phytologist, 221, 2286–2297. https:// doi.org/10.1111/nph.15508
- Wei, N., Du, Z., Liston, A., & Ashman, T.-L. (2020). Genome duplication effects on functional traits and fitness are genetic context and species dependent: Studies of synthetic polyploid *Fragaria*. American Journal of Botany, 107(2), 262–272. https://doi.org/10.1002/ ajb2.1377
- Westoby, M., Falster, D. S., Moles, A. T., Vesk, P. A., & Wright, I. J. (2002). Plant ecological strategies: Some leading dimensions of variation between species. *Annual Review of Ecology and Systematics*, 33, 125–159. https://doi.org/10.1146/annurev.ecols ys.33.010802.150452
- Wright, I. J., Reich, P. B., Westoby, M., Ackerly, D. D., Baruch, Z., Bongers, F., Cavender-Bares, J., Chapin, T., Cornelissen, J. H. C., Diemer, M., Flexas, J., Garnier, E., Groom, P. K., Gulias, J., Hikosaka, K., Lamont, B. B., Lee, T., Lee, W., Lusk, C., ... Villar, R. (2004). The worldwide leaf economics spectrum. *Nature*, 428(6985), 821–827. https://doi. org/10.1038/nature02403

SUPPORTING INFORMATION

Additional supporting information may be found in the online version of the article at the publisher's website.

How to cite this article: López-Jurado, J., Mateos-Naranjo, E., & Balao, F. (2022). Polyploidy promotes divergent evolution across the leaf economics spectrum and plant edaphic niche in the *Dianthus broteri* complex. *Journal of Ecology*, 110, 605–618. https://doi.org/10.1111/1365-2745.13823