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¹ Selectivity in the Photo-Fries Rearrangement of Some Aryl ² Benzoates in Green and Sustainable Media. Preparative and ³ Mechanistic Studies

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8 Supporting Information

ABSTRACT: Irradiation of a series of p-substituted aryl 9 benzoates under N2 atmosphere in homogeneous and micellar 10 media was investigated by means of steady-state condition and 11 of time-resolved spectroscopy. A notable selectivity in favor of 12 the 2-hydroxybenzophenone derivatives was observed in 13 micellar media. The benzophenone derivatives were the 14 main photoproduct. On the other hand, in homogeneous 15 media (cyclohexane, acetonitrile, and methanol) the observed 16 17 product distribution was entirely different, viz. substituted 2hydroxybenzophenones, p-substituted phenols, benzyl and 18 benzoic acid were found. The binding constants in the 19



20 surfactant were also measured and NOESY experiments showed that the aryl benzoates were located in the hydrophobic core of

- the micelle. Laser flash photolysis experiments led to the characterization of both *p*-substituted phenoxy radical and substituted
- 22 2-benzoylcyclohexadienone transients in homogeneous and micellar environment.

23 INTRODUCTION

24 Photochemical reactions grant access to a variety of scaffolds 25 difficult, if not impossible, to access through thermal chemistry. 26 However, the energetic advantage of populating the excited 27 states is often paid with a lack of selectivity in product formation. 28 For this reason, reactions involving intermediates such as radical 29 pairs or radical-ion pairs represent a significant challenge 30 regarding product distribution for the synthetic organic 31 photochemist. Zeolites, micelles, polyolefin films, cavitands, 32 dendrimers, etc., as a useful heterogeneous media, can be helpful 33 to direct the selectivity of photoproducts in photoinduced 34 reactions.¹

Surfactants are amphiphilic molecules and aggregation in 35 solution to form micelles can be achieved when their 36 37 concentration is 100 times higher than the critical micellar 38 concentration (cmc). Then, micelles solubilize efficiently 39 hydrophobic compounds in water, albeit micelles are not static 40 species showing a dynamic equilibrium.² Also, micelle can 41 concentrate guest molecules into relatively small effective 42 volumes promoting their re-encounter consequently.^{1c} Inside 43 the homophobic core of the micelles, significant cage effects are 44 observed when compared to homogeneous media, with 45 magnitudes impossible to explain considering the sole micro-46 viscosity in the constrained environment. The main reason for 47 this behavior is based on the hydrophobicity of the solutes where 48 inhibition of their diffusion into the aqueous phase is 49 noteworthy. Thus, the reaction intermediates show a high

lifetime in the restricted hydrophobic core of the micelle. For 50 example, geminate radical pairs that are produced photochemi-51 cally within the micellar core, have their rotational and 52 translational mobility constrained inside the micelles.² Indeed, 53 the mobility restricted within the hydrophobic cores of radicals, 54 radical cations, or other reactive intermediates limits unwanted 55 reactions (e.g., radical-radical self-quenching reactions) and the 56 access of adventitious reagents (e.g., water and oxygen) that 57 would cause their collapse in solution. Therefore, micellar 58 solution can induce a product distribution and a relative 59 chemical yield that can be significantly different when compared 60 with homogeneous conditions.^{1,2}

There have been many studies on the control of the reactivity 62 of radical species generated within the hydrophobic core of the 63 micelle, and some physical parameters (e.g., electrostatic, 64 polarity, hydrophobic interactions, viscosity, as well as hydro- 65 gen-bonding solvation) may be involved in determining their 66 reactivities.³ In addition, several studies directed to analyze and 67 quantify the reactivity, selectivity, and efficiency of micellar cage 68 on photochemical reactions in water have been carried out. 69

Among the vast number of photochemical transformations, a 70 particular example is represented by the photo-Fries rearrange-71 ment. Anderson and Reese⁴ have discovered the photoreaction 72 where a homolytic fragmentation of a carbon—heteroatom bond 73

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74 is involved, i.e., C–O, C–S, and C–N, of esters, thioesters and 75 amides, respectively.⁵ The photo-Fries rearrangement proceeds 76 via a well-established radical mechanism, mainly occurring 77 through the excited singlet state.^{1b,6} Typically, the photoinduced 78 Fries rearrangement reaction of (hetero)aryl benzoates in 79 homogeneous media affords *ortho-* and *para-*regioisomers as 80 well as the corresponding phenols (Scheme 1).^{7,1b}





The product distribution of the photo-Fries rearrangement 81 82 underlines the competition between in-cage radical recombina-83 tion versus out-of-cage diffusion, and consequently, aryl esters, 84 as model substrates, have been chosen to study heterogeneous environments.⁸ In these studies, SDS (sodium dodecyl sulfate) 85 86 was the preferred surfactant to be tested. In the literature, there 87 are examples of photo-Fries reactions, i.e., irradiation of 88 benzamides in SDS solution and irradiation of aqueous solutions 89 of acetanilides confined in cyclodextrin.^{8b,d} Recently, we have 90 studied the photo-Fries rearrangement of a variety of substituted 91 acetanilides in micellar solution showing the high selectivity of 92 the photoreaction in favor of the ortho-rearranged photo-93 products, viz. substituted 2-aminoacetophenones.⁹ The prep-94 aration of benzoyloxy benzophenone derivatives requires the use 95 of 2-hydroxybenzophenones as key compounds, demonstrating 96 biological activity and pharmaceutical properties (e.g., anti-97 inflammatory and estrogenic activity).¹⁰ Therefore, we carried 98 out the systematic study the photo-Fries rearrangement of a 99 series of para-substituted phenyl benzoates in surfactant 100 solutions with the aim of evaluating the selectivity toward the formation of 5-substituted 2-hydroxybenzophenones in a 101 102 constrained environment. Scheme 2 shows the structures of 103 the surfactants as well as the aryl benzoates employed in this systematic study. 104

In the present paper, we describe the results on the photo-106 Fries rearrangement of several *para*-substituted aryl benzoates in 107 homogeneous and micro-heterogeneous media. The binding 108 constants (K_b) and the location of the substrates within the 109 micelles are measured through spectroscopic methods. From a 110 preparative point of view, the use of anionic and neutral

Scheme 2. Structures of surfactants and aryl benzoates



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surfactant micelles shows a high selectivity in favor of the 111 formation of the 2-hydroxybenzophenone derivatives achieving 112 yields higher than 90%. From a mechanistic viewpoint, it 113 furnishes the characterization of both *para*-substituted phenoxy 114 radical and substituted 2-benzoylcyclohexadienone transients in 115 homogeneous and micellar environments. The in-cage and outof-cage rate constants (k_R and k_E , respectively) of *para*- 117 substituted phenoxy radicals are measured for the first time by 118 means of laser flash photolysis. 119

Steady-State Photolysis. *Photoirradiation of Aryl Ben*- 121 *zoates in Homogeneous Media.* Irradiation of aryl benzoates 122 (1–8) in cyclohexane, MeOH, and MeCN, which were chosen 123 as representative nonpolar, protic polar, and aprotic polar 124 solvents, with $\lambda_{exc} = 254$ nm under N₂ atmosphere provided the 125 expected photoproducts from the photo-Fries rearrangement, 126 viz. formation of 2-hydroxybenzophenone derivatives (1a–8a), 127 the corresponding phenols (1b–8b), and benzoic acid. The 128 photochemical reaction is depicted in Scheme 3. The formation 129 s3

Scheme 3. Photoinduced Fries rearrangement of aryl benzoates (1 - 8)



of benzoic acid was attributed to the oxidation of benzaldehyde, 130 which is the primary photoproduct formed, because of the 131 presence of residual molecular oxygen in the reaction mixture.¹¹ 132 In all of the performed reactions, the yield of benzoic acid was 133 between 5 and 15%. 134

When esters are consumed, the chemical yields collected in 135 Table 1 show that benzophenones 1a-8a are the main 136 t1 photoproducts in up to 94% yield. Furthermore, the product 137 distribution did not change significantly with the nature of the 138 solvent, and poor selectivity in favor of the benzophenone 139 derivatives was observed. 140

The quantum yields of consumption of the aryl benzoates 141 (ϕ_r) in polar and nonpolar solvents were measured (see Table 142 1). The ϕ_r values were found to be higher than 0.30, implying 143 that the photo-Fries rearrangement reaction occurred efficiently. 144 Moreover, in every solvent, a marked increase of the ϕ_r values 145 was observed, moving from esters bearing electron-donor to 146

Table 1. Irradiation of Aryl Benzoates in Homogeneous Solution: Yield of Photoproducts,^{*a*} Reaction Quantum Yield (ϕ_r) ,^{*b*} and Fluorescence Quantum Yield $(\phi_f)^c$

		photoproduct y	rield (%)		
aryl benzoates	solvent	benzophenones (a)	phenols (b)	$\phi_{ m r}$	$\phi_{ m f}$
1	cyclohexane	46	27	0.30	0.08
	MeCN	62	17	0.36	0.03
	MeOH	66	17	0.33	0.10
2	cyclohexane	55	28	0.37	0.002
	MeCN	69	19	0.71	0.013
	MeOH	73	23	0.44	0.002
3	cyclohexane	42	22	0.40	0.25
	MeCN	94	5	0.62	0.07
	MeOH	80	20	0.31	0.11
4	cyclohexane	59	27	0.50	0.001
	MeCN	95	5	0.35	0.025
	MeOH	75	25	0.63	0.002
5	cyclohexane	40 ^e	30	0.36	0.001
	MeCN	60 ^e	33	0.14	0.005
	MeOH	58 ^e	26	0.14	0.002
6	cyclohexane	41	23	0.59	0.012
	MeCN	73	18	0.32	0.035
	MeOH	85	14	0.31	0.033
7	cyclohexane	38	19	0.51	0.002
	MeCN	84	15	0.82	0.025
	MeOH	75	26	0.93	0.002
8	cyclohexane	31	34	0.02	NF ^d
	MeCN	60	40	0.07	NF
	MeOH	50	52	0.07	NF

^{*a*}Yield of photoproducts determined by ¹H NMR spectroscopy in the reaction mixture. Concentration of aryl benzoates: 5.0×10^{-3} M. ^{*b*}Actinometer: KI (0.6 M), KIO₃ (0.1 M) and Na₂B₂O₇.10H₂O (0.01 M) solution in water; ϕ (I₃⁻) = 0.74; λ_{exc} = 254 nm.¹² Error: ±0.01. ^{*c*}Actinometer: 4-chloroanisole acetonitrile solution under Ar atmosphere; ϕ_{f} = 0.019.¹³ Error: ±0.002. ^{*d*}NF: nonfluorescent substrate. ^{*e*}4-Hydroxybenzophenone is also formed: Cyclohexane 30%; MeCN 7%; MeOH 15%.

¹⁴⁷ esters substituted with electron-acceptor substituents. The only ¹⁴⁸ exception found was ester **8**. The ϕ_r values measured for ¹⁴⁹ compound **8** are lower than 0.10 in all the solvents studied. The ¹⁵⁰ aryl benzoates are all poorly fluorescent chromophores (see ¹⁵¹ Table 1) with the exception of benzoate **8** that was found to be ¹⁵² nonfluorescent. The spin coupling effect of the nitro group ¹⁵³ explains the peculiarity of ester **8**. ^{14,15} Indeed, intersystem ¹⁵⁴ crossing pathway competes with both the photo-Fries rearrange-¹⁵⁵ ment reaction and the fluorescence emission.

UV-vis and NMR spectroscopies were used to follow the 156 photochemical reaction, and aryl benzoates 1, 2, 3, and 7 have 157 158 been chosen as representative examples for such spectroscopic 159 studies. Figure 1a showed the time-resolved UV-vis absorption 160 spectrum of the photoreaction of *p*-methylphenyl benzoate (3) 161 in cyclohexane showing the growth of a new band located at 352 162 nm during the irradiation time. This band was assigned to the 163 $n_{,}\pi^{*}$ transition of the carbonyl group of benzophenone 3a¹⁴ and was also observed in MeOH and MeCN. However, no 164 165 significant solvent effect was observed in the maximum 166 wavelength of the n,π^* transition band upon change of the 167 solvent polarity. A similar solvent effect on the maximum 168 wavelength of the n,π^* band of the 2-hydroxybenzophenone 169 derivatives (1a, 2a, 4a-8a) was also observed (see Figure S1, 170 Supporting Information). On the other hand, a noticeable

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substituent effect on the $n_{\pi}\pi^*$ band of the substituted 2- 171 hydroxybenzophenones (1a-8a) was observed. In fact, a 172 hypsochromic effect ($\Delta \lambda = -73$ nm) ascribed to the $n_n \pi^*$ 173 band was measured by changing the substituent from the 174 electron donor MeO ($\lambda_{max} = 378$ nm) to the electron acceptor 175 NO₂ (λ_{max} = 305 nm). The course of the photoreaction of ester 1 176 is detailed in Figure 1b, where the benzoyl [1,3]-migration to 177 form benzophenone 1a is shown to be the primary process. Parts 178 c and d of Figure 1 show the relative formation of 179 benzophenones 2a and 7a (starting from the corresponding 180 esters 2 and 7), respectively, in nonpolar and polar solvents. The 181 relative rates of formation of 2a are similar in MeCN and 182 cyclohexane, while in MeOH the rate somewhat lower, implying 183 that in protic polar solvent radiative and nonradiative decay rates 184 compete with the photoreaction pathway. The rates of 185 formation of benzophenone 7a are quite similar in all of the 186 solvents tested. For the other esters studied, the relative rates of 187 formation of the corresponding benzophenone derivatives 188 showed a similar behavior (see Figure S2, Supporting 189 Information). 190

The ¹H NMR spectra of the photoreaction mixture (6 h ¹⁹¹ irradiation) of 4-cyanophenyl benzoate (7) in cyclohexane ¹⁹² under N_2 atmosphere were also recorded (see Figure S3, ¹⁹³ Supporting Information). As expected, the formation of the ¹⁹⁴ photoproducts benzophenone 7a and phenol 7b was confirmed ¹⁹⁵ through their diagnostic signals (see Figure S3, Supporting ¹⁹⁶ Information) along with the unreacted ester 7. The experiments ¹⁹⁷ demonstrated poor selectivity of the photoreaction in ¹⁹⁸ homogeneous media, and this trend was observed for all the ¹⁹⁹ esters (1–8) studied (see Table 1). ²⁰⁰

Photoirradiation of Aryl Benzoates in Micellar Media. ²⁰¹ Irradiation of aryl benzoates (1–8) in aqueous SDS (0.10 M) ²⁰² with $\lambda_{exc} = 254$ nm under air caused the selective generation of 2- ²⁰³ hydroxybenzophenone derivatives (1a–8a) in high yields along ²⁰⁴ with lower amounts of the corresponding phenols (1b–8b) (for ²⁰⁵ structures, refer to Scheme 3). In these experiments, ²⁰⁶ consumption up to 95% of aryl benzoates (1–8) was obtained. ²⁰⁷ The chemical yields of benzophenone derivatives (1a–8a) are ²⁰⁸ collected in Table 2. ²⁰⁹ t2

As is apparent from Table 2, SDS and Brij-P35 micellar 210 solutions promoted a high selectivity on the photo-Fries 211 rearrangement of the aryl benzoates 2-8, favoring the formation 212 of the corresponding substituted benzophenones (2a-8a) over 213 the para-substituted phenols. The observed selectivity was 214 attributed to the confinement of the aryl benzoates and the 215 radicals formed after the C-O bond cleavage within the 216 hydrophobic core provided by the micellar medium. Moreover, 217 the para-substituted phenols were formed in a minor extent or 218 not formed at all, evidencing the suppression of products arising 219 from cage escape. It is worth noticing that the irradiation of p- 220 nitrophenyl benzoate (8) gave 2-hydroxy-5-nitrobenzophenone 221 (8a) only in 44% yield (the consumption of the starting material 222 was 66% after 6 h of irradiation). No p-nitrophenol was detected 223 in the reaction mixture. Competitive deactivation of the singlet 224 state of *p*-nitrophenyl benzoate (8) through intersystem 225 crossing accounted for the observed chemical yield of 226 benzophenone **8a.**¹⁴ The competitive process populates the 227 triplet excited state, due to the spin-orbit coupling provided by 228 the nitro group, which is an unproductive excited state of 229 benzoate 8. 230

Quantum yields of consumption of the aryl benzoates (ϕ_r , see 231 Table 2) were measured in micellar media, viz. SDS and Brij- 232 P35, and were found to be of the same order of magnitude. 233



Figure 1. (a) Time-resolved UV–visible absorption spectrum of **3** in cyclohexane. Blue line: initial time; red line: 300 s. (b) Relative yield profile vs time of **1** in MeOH: ester **1** (circles); benzophenone **1a** (triangles); phenol **1b** (square). (c) Relative absorbance at 358 nm (A/A_{∞}) of formation of **2a** in MeOH (circles); MeCN (triangles); cyclohexane (square). (d) Relative absorbance at 333 nm (A/A_{∞}) of formation of **7a** in MeOH (circles); MeCN (triangles); cyclohexane (square).

However, the quantum yields measured in SDS solution were larger than in Brij-P35 solutions. This behavior can be attributed to an enhancement of the nonradiative pathway from the singlet state of the aryl benzoates in Brij-P35 solutions. The highend consumption quantum yields and low or no fluorescence emission from aryl benzoates (1-8) was consistent with a fast reaction from the singlet state. However, nonradiative and intersystem crossing pathways from the singlet state compete with the photo-Fries rearrangement.

UV-vis spectroscopy was used for following the photo-244 reaction and *p*-methoxyphenyl benzoate (1) was selected as a 245 representative aryl benzoate. Thus, the reaction of 1 in SDS 246 (0.10 M) was followed by UV-vis absorption spectroscopy, and 247 the UV-vis spectral change vs time is shown in Figure 2a. It is 248 apparent from the UV-vis spectra that the photo-Fries 249 rearrangement of compound 1 to form 2-hydroxy-5-methox-250 ybenzophenone (1a) is the primary process, according to the 251 appearance of the characteristic n,π^* band of the carbonyl group

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located at 372 nm. Similar behavior was observed for 252 compounds 2–8 (see Figure S1, Supporting Information). 253

The course of the photoreaction of ester **1** is depicted in 254 Figure 2b and clearly shows that the benzoyl [1;3]-migration to 255 form benzophenone **1a** is the main process. On the other hand, 256 we selected aryl benzoates 7 and **4** as examples of aryl benzoates 257 to show the relative formation of benzophenones **7a** and **4a**, 258 respectively, in cyclohexane and 0.10 M SDS and 0.01 M Brij-P35 solutions (see Figure 2c,d). The relative rates of formation 260 of **7a** in surfactant media (see Figure 2c) are slightly lower than 261 in cyclohexane and can be attributed to the radiative and 262 nonradiative decay rates that compete with the photoreaction 263 pathway. Similar behavior is observed for the rates of formation 264 of benzophenone **4a** (see Figure 2d) as well as with for the other 265 benzophenone derivatives (see Figure S2, Supporting Informa-266 tion). 267

The photoreaction of compound 1 in micellar environment $_{268}$ (SDS 0.10 M) was also followed by NMR spectroscopy. The 1 H $_{269}$

Table 2. Irradiation of Aryl Benzoates in Micellar Solution: Yield of 5-Substituted 2-Hydroxybenzophenones,^{*a*} Reaction Quantum Yield (ϕ_r) ,^{*b*} and Fluorescence Quantum Yield $(\phi_f)^c$ of Aryl Benzoates

	photoproduct yield (%)			$\phi_{ m f}$	
benzophenones	SDS	Brij-P35	SDS	Brij-P35	SDS
1a	94	95	0.13	0.12	0.026
2a	76	85	0.79	0.21	0.026
3a	70	92	0.55	0.41	0.024
4a	85	89	0.54	0.26	0.025
5a	90 ^e	92 ^e	0.50	0.21	0.029
6a	87	90	0.53	0.21	0.062
7a	80	78	0.45	0.10	0.024
8a	44	56	0.13	0.01	NF ^d

^{*a*}Yield of photoproducts determined by ¹H NMR spectroscopy in the reaction mixture. Concentration of aryl benzoates: 5.0×10^{-3} M. ^{*b*}Actinometer: KI (0.6 M), KIO₃ (0.1 M) and Na₂B₂O₇·10H₂O (0.01 M) solution in water; $\phi(I_3^-) = 0.74$; $\lambda_{exc} = 254$ nm.¹² Error: ± 0.01 . ^{*c*}Actinometer: 4-chloroanisole acetonitrile solution under Ar atmosphere; $\phi_f = 0.019$; ¹³ Error: ± 0.002 . ^{*d*}NF: nonfluorescent substrate. ^{*e*}4-Hydroxybenzophenone is also formed in 8–10%.

270 NMR spectra of the reaction mixture of benzoate 1 in SDS (0.10 271 M) irradiated during 6 h with λ_{exc} = 254 nm (see Figure S4, 272 Supporting Information) showed that benzophenone 1a was 273 formed in 94% yield along with *p*-methoxyphenol and benzoic 274 acid in yields lower than 5%. In the same spectra, the signals 275 belonging to the surfactant (SDS) were also observed. The 276 consumption of benzoate 1 was quantitative. Similar results were 277 obtained with all the aryl benzoates studied (2–8), and the 278 photoproduct yields are collected in Table 2.

Binding Constants (K_b) of Aryl Benzoates in Micellar 279 280 Media. When ionic and neutral surfactant solutions are used as 281 microreactors to perform photoreactions the knowledge of the 282 reactant's location in the micellar system is required. In order to 283 know the reactant's positioning within micelles, UV-vis and ¹H 284 NMR studies of guest molecules (aryl benzoates) in micellar 285 solution were conducted. UV-vis spectroscopy was used to 286 determine the binding constant (K_b) between micelles and aryl 287 benzoates, applying a methodology that has been reported 288 previously for aryl acetamide.⁹ Both bathochromic and hyper-289 chromic shifts of the lower energy absorption banrd of the aryl 290 benzoates were observed by addition of increasing amounts of 291 surfactant, demonstrating the binding of the substrates to the 292 micelle that took place within the hydrophobic core of the 293 micelle. Indeed, the binding of the aryl benzoates to the micelle 294 can be described as equilibrium, and $K_{\rm b}$ can be written according 295 to eq 1, where S represents the benzoates, Surf the surfactants, 296 and [S-Surf] the complex formed between benzoates and the 297 surfactants.

$$[S] + [Surf] \stackrel{K_{b}}{\rightleftharpoons} [S-Surf] \quad k_{b} = \frac{[S-Surf]}{[S][Surf]}$$
(1)

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Equation 2 was obtained after application of the Lambert– 300 Beer law on eq 1. A_0 and A are the absorbances at the maximum 301 wavelength in the absence and presence of surfactant, 302 respectively. The molar absorptivity of the complex and the 303 benzoates are dubbed accordingly $\varepsilon_{\rm C}$ and $\varepsilon_{\rm S}$. A linear 304 relationship is observed between $(A - A_0)^{-1}$ and the reciprocal 305 of the concentration of the surfactant in eq 3, which was 306 obtained after rearranging eq 2.

$$\frac{(A - A_0)}{A_0} = \frac{\varepsilon_C K_b[\text{Surf}]}{\varepsilon_S(1 + K_b[\text{Surf}])}$$
(2) 307

$$\frac{A_0}{(A-A_0)} = \frac{\varepsilon_{\rm S}}{\varepsilon_{\rm C}} + \frac{\varepsilon_{\rm S}}{\varepsilon_{\rm C} K_{\rm b}} \frac{1}{[{\rm Surf}]} = A_0 \frac{1}{\Delta A}$$
(3) 308

The experimental data obtained for some aryl benzoates and 309 SDS and Brij-P35 are shown in Figure 3, and the best linear 310 f3 regression curves are also included in the same figure. The plots 311 for the other systems are collected in Figure S5 (see the 312 Supporting Information). The $K_{\rm b}$ values for 1–8 calculated from 313 the ratio between the slope and the intercept of the regression 314 curve are shown in Table 3. From these data, it is apparent that 315 t3 the benzoates bind to the micelles, and the ones possessing 316 hydrophobic character, i.e., benzoates 1 and 4 in SDS micellar 317 solution, show high $K_{\rm h}$ values. Likewise, benzoates 1, 4, and 7 318 possess greater $K_{\rm b}$ compared to the other substrates in the case 319 of Brij-P35 micellar solution. Generally, the $K_{\rm b}$ values obtained 320 for benzoates 1-8 are typical of aromatic solutes as it pointed 321 out by Quina, Treiner, and co-workers.¹⁶ Estimation of $K_{\rm b}$ values 322 $\leq 100 \text{ M}^{-1}$ in SDS for weakly hydrophobic substrates such as 323 phenyl chloroformate have also been reported.¹⁷ Likewise, K_b 324 values have been reported for the binding of Brij-P35 surfactant 325 and a series of benzoyl chloride derivatives, and they were found 326 to be higher than those obtained for other surfactants such as 327 SDS and CTAC.¹⁸ Estimation of a minimum value around 190 328 M^{-1} for the binding constant ($K_{\rm b}$) of Brij-P35 surfactant was 329 reported and accounts for a more apolar environment when 330 pyrene was used as a micropolarity probe.¹⁸ 331

2D NMR spectroscopy has been recorded to confirm 332 qualitatively the location of the aryl benzoates within the 333 micelle. NOESY experiments have been often used to determine 334 the localization of substrates inside the micelle as well as to 335 determine the extent of coaggregation between two different 336 kinds of surfactants in water.¹⁹ Positive results are achieved 337 when cross-peaks between diagnostic signals of the substrate and 338 the surfactants, respectively, are observed in the corresponding 339 contour plots.⁹ Thus, the NOESY experiments were performed 340 in D₂O, and Figure 4 shows the 2D NMR spectrum for a 341 f4 solution of SDS (7 mM) in the presence of benzoate 1 (10 mM). 342 The inset red frames recognize the NOE (nuclear Overhauser 343 effect) between the signals of the surfactant SDS (bulk 344 hydrogens and α hydrogen) and the signals belonging to the 345 aromatic protons (H-2/H-6, H-3/H-5, H-10/H-14, H-12, and 346 H-12/H-13) of p-cyanophenyl benzoate 7. Also, Figure 6 shows 347 the labels of the protons of the surfactant SDS and the aryl 348 benzoate 7, respectively. Similar spectroscopic behavior was 349 observed for solutions of SDS and Brij-P35 in D2O in the 350 presence of aryl benzoates 1 and 3 (see Figures S6-S8, 351 Supporting Information). The cross-peaks of diagnostic signals 352 observed in the 2D NMR contour plots are in agreement with 353 and reinforce the UV-vis spectroscopic analyses. 354

However, we cannot estimate the location of the benzoates 355 with accuracy but we can suggest that the benzoates are located 356 inside the hydrophobic core of the micelle because the proton 357 nuclei of the aryl benzoates correlate nicely with the proton 358 nuclei of the surfactants as can be seen through the cross-peaks 359 of the contour plots. 360

Laser Flash Photolysis of Aryl Benzoates. Irradiation of $_{361}$ p-methoxyphenyl benzoate (1) in acetonitrile and cyclohexane $_{362}$ solutions with a laser pulse (266 nm) under nitrogen $_{363}$ atmosphere gives the transient absorption spectra shown in $_{364}$ Figure 5a,b. Four absorption bands with maximum wavelengths $_{365}$ fs



Figure 2. (a) UV–vis spectral change vs time of 1 in SDS 0.10 M in water. Blue line: initial time; red line: 300 s. (b) Relative yield profile vs time of 1 in MeOH: ester 1 (circles); benzophenone 1a (triangles); phenol 1b (square). (c) Relative absorbance at 375 nm (A/A_{∞}) of formation of 7a in cyclohexane (circles); 0.10 M SDS solution (triangles); 0.10 M Brij-P35 solution (square). (d) Relative absorbance at 355 nm (A/A_{∞}) of formation of 4a in cyclohexane (circles); 0.10 M SDS solution (triangles); 0.10 M Brij-P35 solution (square).

366 at 290, 340, 400, and 720 nm were observed in the transient 367 spectra. According to the data reported in the literature, we attributed the bands located at 290 and 400 nm to the p-368 methoxyphenoxyl radical, while those bands centered at 340 and 369 370 720 nm were attributed to the cyclohexadienone transient.²⁰ Two consecutive pathways from the singlet excited state of ester 371 are involved in the formation of both transients after the pulse 1 372 (266 nm): (i) homolytic fragmentation of the C–O bond of the 373 ester group affording p-methoxyphenoxy and benzoyloxy radical 374 375 species in the solvent cage (intermediates A and B, respectively, 376 in Scheme 4) and then (ii) coupling of both radical species to give the 4-methoxy-2-benzoylcyclohexadienone intermediate 377 (C in Scheme 4). Also, in Figure 5c,d are reported the transient 378 absorption spectra of compounds 6 and 2 in acetonitrile after the 379 laser pulse (266 nm). Two characteristic bands located around 380 340 and 400 nm are observed which were assigned to the 381 382 substituted 2-benzoylcyclohexadienone and substituted phe-383 noxy radical intermediates, respectively. Note that y similar 384 results are obtained for the other aryl benzoates studied (see 385 Figure S9, Supporting Information).

The decay traces of the transient signal assigned to 4-386 substituted phenoxyl radicals were also measured at 400 nm in 387 N₂-saturated cyclohexane and acetonitrile solutions (after a laser 388 pulse at 266 nm). The experiment was done with the aim of 389 determining both the rate constants of radical out-of-cage escape 390 $(k_{\rm E})$ and ortho coupling reaction $(k_{\rm R})$ (see Scheme 4). Some 391 selected examples of the decay traces are shown in Figure 6. The 392 f6 decay traces of 4-substituted phenoxyl radicals show biexpo- 393 nential decay fitting with r^2 values >0.998 independent of the 394 solvent used. Two half lifetime values were obtained from the 395 nonlinear fittings, $\tau_{\rm E}$ and $\tau_{\rm R}$: the short lifetime $(\tau_{\rm R})$ was assigned 396 to an in-cage coupling process, while the large lifetime $(au_{
m F})$ was $_{397}$ assigned to the phenoxyl radical out-of-cage process by 398 comparison with previously reported data regarding thiyl 399 radicals.^{20e,21} This biexponential behavior can be interpreted 400 considering the two competitive pathways the 4-substituted 401 phenoxyl radical can undergo, viz. out-of-cage escape and in- 402 cage coupling pathways (see Scheme 4). This behavior can be $_{403}$ described according to eq 4 where ArO[•] represents the 404

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Figure 3. Reciprocal plotting $((A_0/(A - A_0) \text{ vs concentration of surfactants})$ in water at room temperature: (a) SDS (circles: 2; squares: 4; triangles: 7) and (b) Brij-P35 (circles: 2; squares: 4; triangles: 1). In all of the linear fitting regressions: $r^2 > 0.99$.

Table 3. Binding Constant (<i>K</i> _b) in Wate	r of Surfactants (SDS
and Brij-P35) and Aryl Benzoates	

				$K_{\rm b}$ (N	I ⁻¹)			
aryl benzoates	1	2	3	4	5	6	7	8
SDS	1402	80	127	539	42	10	18	74
Brij-P35	1398	581	69	423	63	26	1394	181
NC 3	$2 \qquad 10$ $2 \qquad 0$ $6 \qquad 0$ $5 \qquad 0$	$ \begin{array}{c} 11 \\ 12 \\ 14 \\ 13 \end{array} $	\sim	Вı	ilk	~~	∠SO₃Na	
	•	H-10/H-14	H-3/	H-5 H-12	н н-11-н-13	I-2/H-6		
					-			-0.5
Bulk	· ·	4		•			• 1•	-1.0
β		6	·•	•		98 		-1.5
								-2.0
								-2.5 ^d
								-3.0
								-3.5
α		0	. •		o	۵		-4.0
	3.5 8.4 8.1	8.2 8.1	8.0 7.9	7.8 7.7	7.6 7.5	7.4 7.	3 7.2 7.1	4.5

Figure 4. 2D NOESY contour plot of a solution of SDS (7 mM) and 7 (10 mM) in D_2O at room temperature.

405 substituted phenoxy radical and PhCO[•] represents the benzoyl 406 radical.

$$-\frac{d[ArO^{\bullet}]}{dt} = k_{E}[ArO^{\bullet}] + k_{R}ArO^{\bullet}[Ph\dot{C}O]$$
(4)

⁴⁰⁸ The out-of-cage escape of the substituted phenoxy radical is a ⁴⁰⁹ unimolecular pathway. Therefore, the rate constant ($k_{\rm E}$) of this process can be calculated from the reciprocal of the lifetime, $k_{\rm E} = 410$ $1/\tau_{\rm E}$ (see Table 4). The in-cage coupling rate constants ($k_{\rm R}$) 411 t4 were obtained by plotting the reciprocal of the concentration of 412 the substituted phenoxy radicals against time, and excellent 413 linear correlations were observed (see Figure S11, Supporting 414 Information, for the linear correlations). Then, after application 415 of a linear regression fitting the in-cage coupling rate constants 416 $(k_{
m R})$ were obtained from the slopes, and these data are also 417 shown in Table 4. As can be seen in Table 4, it is apparent that 418 the out-of-cage rate constants $(k_{\rm E})$ for the unimolecular escape 419 process of 4-substituted phenoxyl radicals are quite similar in all 420 the solvents studied $((1.5-8.3) \times 10^5 \text{ s}^{-1})$ and somewhat 421 independent of the substituent. On the other hand, the 422 bimolecular in-cage coupling of 4-substituted phenoxyl and 423 benzoyl radicals was found to be a second-order rate constant 424 $(k_{\rm R})$ of $10^9 - 10^{10}~{
m M}^{-1} \cdot {
m s}^{-1}$ in N2-saturated solvents with no 425 significant substituent effect associated to it. 426

A similar spectroscopic analysis was carried out with aryl 427 benzoates in aqueous SDS (0.10 M) solutions after irradiation 428 with a pulse at 266 nm. Two characteristic bands (340–360 and 429 400 nm) are observed in the transient absorption spectra, and 4- 430 substituted phenoxyl radicals showed second-order kinetic 431 decay traces. The in-cage coupling rate constants (k_R) were 432 obtained by plotting the reciprocal of the concentration of the 4- 433 substituted phenoxyl radicals against time, and after applying a 434 linear regression fitting, the rate constants (k_R) were obtained 435 from the slopes (see Table 4). No significant substituent effect 436 on the rate constants (k_R) was observed when the solvent was a 437 micellar solution (SDS 0.10 M), which gave a trend similar to 438 that observed in homogeneous media. 439

Because phenyl benzoate (5) has no substituent in the *para* 440 position, two possible *o*- and *p*-benzoylcyclohexadienone 441 regioisomers can be proposed (see Scheme 5). Therefore, we 442 s5 have recorded the transient absorption spectra and transient 443 decay traces of compound 5 in different N₂-saturated solvents 444 with a laser pulse at 266 nm systematically. Figure 7a shows both 445 t7 the transient absorption spectra and decay trace recorded at 400 446 nm in MeCN. The band located at 400 nm in the transient 447 absorption spectra was assigned to the phenoxyl radical, while 448



Figure 5. Transient absorption spectra obtained after a laser pulse (100 μ s; λ_{exc} : 266 nm) of solutions (5.1 × 10⁻⁴ M) of (a) 1 in cyclohexane, (b) 1 in acetonitrile, (c) 6 in acetonitrile, and (d) 2 in acetonitrile under nitrogen atmosphere.

Scheme 4. Formation of Transient Species after the Laser Pulse (266 nm)



⁴⁴⁹ the large structured band located between 330 and 385 nm was ⁴⁵⁰ assigned to *o*-benzoylcyclohexadienone and *p*-benzoylcyclohex-⁴⁵¹ adienone transients (see Scheme 5).

⁴⁵² On the other hand, Figure 7b shows the decay trace of ⁴⁵³ phenoxyl radical measured at 400 nm in N₂-saturated ⁴⁵⁴ acetonitrile solution after the laser pulse (266 nm). Biexponen-⁴⁵⁵ tial decay was observed after a nonlinear fitting. This ⁴⁵⁶ biexponential behavior can be interpreted considering the ⁴⁵⁷ competitive pathways the phenoxyl radical can take viz. out-of-⁴⁵⁸ cage escape and *ortho* and *para* in-cage coupling pathways (see ⁴⁵⁹ Scheme 5). The rate constant (k_E) of the out-of-cage escape ⁴⁶⁰ pathway was calculated from the reciprocal of the lifetime, k_E = ⁴⁶¹ $1/\tau_E$. The rate constants thus obtained are shown in Table 5. In ⁴⁶² addition, the *ortho* and *para* in-cage coupling constants, k_{ortho} ⁴⁶³ and k_{para} , were obtained by plotting the reciprocal of the ⁴⁶⁴ concentration of the phenoxy radical against time, and two nice

t5

linear correlations were observed (see Figure S12, Supporting 465 Information, for the linear correlations). Then, after a linear 466 regression fitting was applied to the linear correlations, the in- 467 cage coupling constants were obtained from the slopes, and 468 these data are also shown in Table 5. No significant solvent effect 469 on the rate constants was observed. However, the in-cage *para*- 470 coupling pathway is rather lower than the *ortho*-coupling 471 pathway.

DISCUSSION

473

As it was described above, direct irradiation (254 nm) of aryl 474 benzoates in homogeneous media (cyclohexane, MeCN and 475 MeOH) as well as in micro heterogeneous media (SDS and Brij- 476 P35 aqueous solutions) under N_2 atmosphere took place 477 efficiently (see Figures 1 and 2). During the irradiation of aryl 478 benzoates, a noticeable selectivity in favor of the benzophenone 479



Figure 6. Decay traces of 4-substituted phenoxyl radical recorded at 400 nm after the laser pulse (λ_{exc} : 266 nm) of solutions (5.1 × 10⁻⁴ M) of (a) 4-methoxyphenyl benzoate (1) in cyclohexane; (b) 4-methoxyphenyl benzoate (1) in acetonitrile; (c) 4-*tert*-butylphenyl benzoate (4) in cyclohexane; (d) 4-phenoxyphenyl benzoate (2) in acetonitrile under N₂ atmosphere.

Table 4. Out-of-Cage (k_E) and in-Cage Coupling (k_R) Rate Constants of 4-Substituted Phenoxyl Radicals Measured by Laser Flash Photolysis (266 nm) in Different Solvents under N₂ Atmosphere^{*a*}

Rate constants								
R	MeCN		МеОН		Cyclohexane		SDS (0.10 M)	
R	$k_{\rm E} {\rm x10^{-5}} / {\rm s^{-1}}$	$k_{\rm R} { m x10^{-9}} / { m M^{-1}.s^{-1}}$	$k_{\rm E} {\rm x} 10^{-5} / {\rm s}^{-1}$	$k_{\rm R} { m x10^{-9}} / { m M^{-1}.s^{-1}}$	$k_{\rm E} { m x} 10^{-5} / { m s}^{-1}$	$k_{\rm R} { m x10^{-9}} / { m M^{-1}.s^{-1}}$	$k_{\rm R} { m x10^{-9}} /{ m M^{-1}.s^{-1}}$	
OMe (1)	2.2±0.1	5.5±0.1	2.8±0.1	15±1	4.5±0.1	8.9±0.1	3.9±0.1	
OPh (2)	8.3±0.2	14±1	1.0±0.2	24±1	2.2±0.2	46±1	25±1	
Me (3)	3.0±0.1	$1.4{\pm}0.1$	6.7±0.1	1.7±0.1	2.5±0.1	7.3±0.1	3.0±0.1	
t-Bu (4)	1.5±0.1	28±1	3.4±0.1	12±1	3.4±0.1	3.4±0.1	26±1	
Ph (6)	5.0±0.2	4.6±0.1	1.0±0.2	6.1±0.1	Ins	soluble	1.2±0.1	
CN (7)	4.8±0.2	9.5±0.1	2.3±0.2	9.9±0.1	1.3±0.2	36±1	17±1	

^{*a*}Concentration of aryl benzoates: 5.0×10^{-4} M.

 $_{480}$ derivatives was observed in micellar media. In this case, the $_{481}$ formation of the corresponding substituted phenols was lower $_{482}$ than 2% because of the confined hydrophobic core of the micelle $_{483}$ (compare the data shown in Tables 1 and 2). The reaction $_{484}$ mechanism depicted in Scheme 6 was sustained by the results

obtained under steady-state conditions and laser flash photolysis 485 experiments. When aryl benzoates are irradiated at 254 nm, the 486 population of the singlet state is achieved efficiently. This excited 487 state is the photoreactive state of the photoreaction as reported 488 in the literature.⁵ Two pathways are involved in the deactivation 489



Scheme 5. Formation of Transient Species after the Laser Pulse (266 nm) from Phenyl Benzoate

Figure 7. (a) Transient absorption spectra of phenyl benzoate (5) recorded in N₂-saturated acetonitrile solution $(5.1 \times 10^{-4} \text{ M})$ and (b) decay trace of phenoxyl radical recorded at 400 nm after the laser pulse $(100 \,\mu\text{s}; \lambda_{exc}: 266 \text{ nm})$ of N₂-saturated acetonitrile solution $(5.1 \times 10^{-4} \text{ M})$ of phenyl benzoate (5).

Table 5. Out-of-Cage $(k_{\rm E})$ and in-Cage Coupling $(k_{\rm ortho}$ and $k_{\rm para}$) Rate Constants of Phenoxyl Radicals Measured by Laser Flash Photolysis (266 nm) in Different Solvents under N₂ Atmosphere^{*a*}

	rate constants					
solvents	$k_{\rm E} \times 10^{-5}$ (s ⁻¹)	$\substack{k_{\mathrm{ortho}} \times 10^{-9} \\ (\mathrm{M}^{-1} \cdot \mathrm{s}^{-1})}$	$k_{\rm para} { m tme} { m x10}^{-9} \ ({ m M}^{-1} { m \cdot} { m s}^{-1}$			
cyclohexane	4.8 ± 0.1	8.7 ± 0.1	4.1 ± 0.1			
MeCN	4.8 ± 0.1	8.4 ± 0.1	3.2 ± 0.1			
MeOH	3.1 ± 0.1	2.7 ± 0.1	1.4 ± 0.1			
SDS (0.10 M)		5.5 ± 0.2	1.5 ± 0.2			
		4				

^{*a*}Concentration of phenyl benzoates: 5.0×10^{-4} M.

of the singlet state: (i) homolytic fragmentation of the C–O 490 bond (path a; Scheme 6) affording aryl phenoxy (ArO[•]) and 491 benzoyl (PhCO[•]) radicals that evolve to 5-substituted 2- 492 hydroxybenzophenone and 4-substituted phenol and (ii) 493 photophysical deactivation (k_d ; Scheme 3) of the singlet state 494 involving fluorescence emission and internal conversion path- 495 ways that give the aryl benzoates in their ground state. In the case 496 of ester **8**, intersystem crossing pathway must be considered as a 497 process involved in the physical deactivation pathway estimating 498 a ϕ_T value around 0.50.^{14,15} Irradiation of aryl benzoates in N₂- 499 saturated solutions with a laser pulse ($\lambda_{exc} = 266$ nm) gave the 500 transient absorption of 4-substituted phenoxyl radical and 5- 501 substituted 2-benzoylcyclohexadienone, and these transients 502 were formed immediately at 10 μ s after the incident light (see, 503

Scheme 6. Reaction Mechanism for the Irradiation of Aryl Benzoates with $\lambda_{exc} = 254$ nm





504 for example, Figure 5 for compounds 1, 2, and 6), demonstrating 505 that C–O homolytic fragmentation (path a in Scheme 6) and in-506 cage coupling pathway (path d in Scheme 6) occurred 507 efficiently.

The decay traces of the phenoxyl radical transients were 508 obtained at 400 nm in homogeneous media under inert 509 ⁵¹⁰ atmosphere, and biexponential decay was observed. Second-⁵¹¹ order kinetics (k_R) of 10^9-10^{10} M⁻¹·s⁻¹ were obtained (see 512 Table 4 and Figure 6) and were attributed to the in-cage coupling of the substituted phenoxy radical and benzoyl radical 513 (path d in Scheme 6) providing the substituted 2-benzoylcy-514 515 clohexadienone intermediates C. These last intermediates, i.e., 516 intermediates C, are long-lived species showing lifetime values s17 higher than 200 μ s. The [1;3]-hydrogen migration and 518 aromatization reaction pathway (path e in Scheme 6) were s19 estimated to occur with first-order kinetics $(k_{\rm H})$ lower than 5 × $_{520}$ 10³ s⁻¹, providing the substituted 2-hydroxybenzophenone derivatives as the main photoproducts (see Table 1). On the 521 other hand, first-order kinetics $(k_{\rm F})$ of $10^5 \, {\rm s}^{-1}$ were also obtained 522 (see Table 4) which were assigned to the escape of the phenoxy 523 524 radical from the solvent cage (path b in Scheme 6) that in turn 525 evolved to the 4-substituted phenol by abstraction of hydrogen 526 from the reaction solvent (path c in Scheme 6; SH: solvent). Second-order rate constants $(k_{\rm R})$ in the range of $10^9 - 10^{10}$ 527 $_{528}$ M⁻¹·s⁻¹ were obtained (see Table 4) from the fitting analysis of 529 the decay traces of the phenoxyl radicals measured at 400 nm in 530 micellar media (SDS and Brij-P35) under N2 atmosphere. These values describe the in-cage coupling within the core of the 531 micelle of the substituted phenoxy radical (ArO[•]) and benzoyl 532 radical $(PhC(O)^{\bullet})$ (see path d in Scheme 6). This pathway 533 afforded the substituted 2-benzoylcyclohexadienone intermedi-534

sist another the substituted 2 behavior performance intermedial sist ates C, also formed within the hydrophobic core of the micelle, sist that evolved through a [1;3]-hydrogen migration and size aromatization reactions to substituted 2-hydroxybenzophenone. sist Because intermediates C are also long-lived species in micellar sign media ($\tau > 200 \ \mu$ s), the rate constants $k_{\rm H}$ were estimated to be side lower than $5 \times 10^3 \ {\rm s}^{-1}$ as observed in homogeneous media.

It is worth mentioning that the only photoproducts detected in micellar media were the 2-hydroxybenzophenone derivatives $\mathbf{1a}-\mathbf{8a}$ formed with chemical yields up to 95% (see Table 2), and it he yields of the substituted phenols were lower than 5%. Therefore, we suggest that the escape of the phenoxy radical it from the hydrophobic core of the micelle (path b in Scheme 6) is it not a productive pathway. The decay traces of phenoxy radicals in micellar media did not show biexponential decay traces as is observed in homogeneous media but second-order kinetics, so which is in agreement with the in-cage coupling pathway (path d is Scheme 6). Additional comments about the irradiation of phenyl 552 benzoate (5) in homogeneous and micellar media are needed. 553 The results obtained under steady-state conditions and time-554 resolved spectroscopy on phenyl benzoate led us to advance the 555 different reaction pathways depicted in Scheme 7. Irradiation of 556 s7 phenyl benzoate populated the singlet excited state efficiently, 557 and competitive physical deactivation (k_d) and homolytic 558 fragmentation (path a in Scheme 7) pathways occurred. 559

After fragmentation phenoxy and benzoyl radical species were 560 formed in the solvent cage, escape of the radical species from the 561 solvent cage (path b) and hydrogen abstraction from the solvent 562 (path c) gave the conventional products. Laser flash photolysis 563 experiments provided rate constants $(k_{\rm E})$ in cyclohexane, 564 MeCN, and MeOH (see Table 5) following the decay trace of 565 the phenoxy radical at 400 nm. No escape of the radical species 566 was detected in micellar solution. Because phenyl benzoate has 567 no substituent in the para position, two possible o- and p- 568 benzoylcyclohexadienone regioisomers viz. intermediates C and 569 D were formed through the in-cage coupling of the radical 570 species (path d and path e in Scheme 7). Both intermediates C 571 and D were observed in the absorption transient spectra (see 572 Figure 7) with characteristic bands located in the range of 340- 573 380 nm. Besides, the $k_{\rm ortho}$ and $k_{\rm para}$ values on the order of 10⁹ 574 $M^{-1} \cdot s^{-1}$ belonging to in-cage coupling pathways (paths d and d' 575 in Scheme 7) implied that the coupling reaction occurred 576 efficiently in all the solvents studied. Then intermediates C and 577 D formed in the solvent cage evolved to the regioisomers 2- 578 hydroxybenzophenone and 4-hydroxybenzophenone through 579 the sequence [1,3]-hydrogen migration and aromatization 580 pathways (path e and path f in Scheme 7). Again, intermediates 581 C and D showed lifetimes higher than 200 μ s, and the rate 582 constant $k_{\rm H}$ was estimated to be lower than 10³ s⁻¹. These 583 photoproducts were formed with 30-60% yields together with 584 the corresponding phenol in homogeneous media (see Table 1), 585 while in micellar solution they were formed in up to 95% yield 586 and no phenol was detected in the micellar reaction mixture. 587

CONCLUSIONS

The photochemical reaction of aryl benzoates examined in this 589 paper takes place efficiently in homogeneous and micellar 590 media. High selectivity in the formation of the 5-substituted 2- 591 hydroxybenzophenone derivatives was observed in micellar 592 media, providing these photoproducts in yields up to 95% 593 without the formation of the corresponding phenols. Location of 594 the aryl benzoates with 2D NOESY NMR spectroscopy in the 595 shell or in the hydrophobic core of the micelle and measurement 596 of the binding constants (K_b) between the benzoates and the 597 surfactants account for the selective behavior observed where 598

588

599 diffusion of the radical species from the micelle is inhibited. On 600 the other hand, benzophenone derivatives, as the main 601 photoproducts, and the *para*-substituted phenols were formed 602 when the irradiations were carried out in homogeneous media 603 such as cyclohexane, MeCN, and MeOH, but no selectivity was 604 observed.

605 Laser flash photolysis led us to characterize two intermediates 606 viz. the substituted phenoxy radical and the 5-substituted 2-607 benzoylcyclohexadienone transients. These intermediates were 608 formed in the cage solvent within 10 μ s after the laser pulse. In 609 addition, the phenoxy radical escapes from the solvent cage with 610 first-order rate constants ($k_{\rm E}$) of 10⁵ s⁻¹ that in turn evolve to the 611 corresponding phenols by hydrogen abstraction from the 612 reaction solvent (see Schemes 6 and 7). The kinetic parameters $_{613}$ ($k_{\rm R}$) for in-cage coupling pathways of the radical species viz. 614 substituted phenoxy and benzoyl radicals were also measured in 615 all of the solvents studied, providing the corresponding 5-616 substituted 2-benzovlcvclohexadienone intermediates (inter-617 mediate C in Scheme 6 and intermediates C and D in Scheme 618 7). These species which are formed in the solvent cage evolved 619 to the regioisomeric 5-substituted 2-hydroxybenzophenone 620 derivatives through the sequence [1;3]-hydrogen migration 621 and aromatization pathways (path e in Scheme 6 and path f in 622 Scheme 7). Because these intermediates showed lifetimes higher 623 than 200 μ s, the rate constants $k_{\rm H}$ were estimated to be lower 624 than 10^3 s⁻¹ in all of the solvents studied.

Finally, the finding that the selectivity observed in the photo-Fries rearrangement of some aryl benzoates in green and sustainable micellar media gives 5-substituted-2-hydroxybenzophenone derivatives in yields up to 95% could be applied in the preparation of a new wide variety of substituted 2hydroxybenzophenoe derivatives.

631 **EXPERIMENTAL SECTION**

Materials and Equipment. Para-substituted phenols, benzoyl 632 633 chloride, pyridine, sodium dodecyl sulfonate, and Brij-P35 were 634 obtained from commercial sources. Spectroscopic grade solvents 635 were used as received. Pyridine was distilled and stored over KOH 636 pellets. Melting points were determined with a Fisher Jones apparatus 637 and are not corrected. ¹H and ¹³C NMR spectra were recorded in 638 CDCl₃ on a 300 MHz spectrometer; chemical shifts (δ) are reported in 639 part per million (ppm), relative to signal of tetramethylsilane, used as 640 internal standard. 2D NOESY spectra were recorded in D₂O on a 500 641 MHz spectrometer, using a NOESY-ph pulse sequence with a 600 ms 642 mixing time and a recovery delay of 1.5 s. 2K data points were collected 643 for 512 increments of 16 scans, using TPPI flquadrature detection; 644 chemical shifts (δ) are reported in part per million (ppm), relative to 645 the signal of trimethylsilylpropionic acid, used as internal standard. 646 Coupling constant (J) values are given in hertz. The measurements 647 were carried out using standard pulse sequences. GC analysis was 648 carried out on a Hewlett-Packard 5890 gas chromatograph using an 649 Ultra 2 capillary chromatographic column. The chromatograms were 650 recorded with the following program: initial temperature: 100 °C, 2 651 min; gradient rate: 10 °C.min⁻¹; final temperature: 250 °C, 10 min. The 652 UV-vis spectra were measured with a Shimadzu UV-1203 653 spectrophotometer using two-faced stoppered quartz cuvettes (1 mm × 1 mm) at 298 K. 654

655 Determination of the Binding Constants (K_b) of Phenyl 656 Benzoates in Micellar Media. Solutions of phenyl benzoates were 657 prepared in deionized water (Milli-Q), and their concentrations varied 658 between 5.5 × 10⁻⁵ M and 1.0 × 10⁻⁴ M. An aliquot (2 mL) of the 659 phenyl benzoate solution was placed in a fluorescence-stoppered quartz 660 cuvette provided with a stirring bar, and the UV–vis spectrum was 661 recorded. The initial absorbance value at the maximum absorption 662 wavelength (A_0) was read. Subsequently, aliquots of concentrated 663 surfactant solution (10 μL) were added. The UV–vis spectra were registered, recording for each solution the *A* value at the maximum 664 absorption wavelength. After each addition of surfactant, the solution 665 was stirred for 20 min before measuring the absorbance. With the values 666 of A_0 and *A* in hand, the values of $(A_0/(A - A_0))$ versus the reciprocal of 667 the concentration of the micellar surfactant were plotted, and the data 668 were fitted with a linear regression program. The K_b values were 669 obtained by calculating the ratio of the slope and the origin. 670

Laser Flash Photolysis. The laser pulse photolysis apparatus 671 consisted of a Flash lamp-pumped Q-switched SpitLight-100 Nd:YAG 672 laser from InnoLas used at the fourth harmonic of its fundamental 673 wavelength. The LP920-K monitor system (supplied by Edinburgh 674 Instruments), arranged in a cross-beam configuration, consisted of a 675 high-intensity 450 W ozone free Xe arc lamp (operating in pulsed 676 wave), a Czerny-Turner with triple grating turret monochromator, and 677 a five-stage dynode photomultiplier. The signals were captured by 678 means of a Tektronix TDS 3012C digital phosphor oscilloscope, and 679 the data were processed with the L900 software supplied by Edinburgh 680 Instruments. The solutions to be analyzed were placed in a fluorescence 681 cuvette (d = 10 mm).

Synthesis of Phenyl Benzoates 1-8. To a solution of the 683 substituted phenols (0.010 mol) in pyridine (10 mL) cooled in an ice 684 bath was added benzoyl chloride (0.012 mol) dropwise over 10 min 685 with stirring. Subsequently, the reaction mixture was kept under stirring 686 for 60 min. After total consumption of the starting material was 687 confirmed by TLC, the reaction mixture was extracted with 688 dichloromethane (10 mL) and washed with a solution of diluted HCl 689 (10 mL). The organic phase was then washed with water, dried on 690 Na₂SO₄, filtrated, and evaporated under pressure. The phenyl 691 benzoates were purified from the solid residue by recrystallization 692 using ethanol–water mixtures to give the corresponding phenyl 693 benzoates in excellent yields (>90%). The aryl benzoates 1-8 were 694 characterized by comparing the physical constant (mp) and 695 spectroscopic data (¹H NMR and ¹³C NMR) with the ones reported 696 in the literature.

Photoirradiation of Phenyl Benzoates in Homogeneous 698 Media. A stock solution of a given benzoate (1-8, 0.106 mmol in 200 699 mL cyclohexane) was placed in a stoppered Erlenmeyer quartz flask and 700 degassed with argon for 30 min. The flask was placed in a homemade 701 optical bench provided with the possibility to use four or eight lamps. 702 The solution was stirred during the entire irradiation. Irradiations with 703 λ_{exc} = 254 nm were carried out with four germicide lamps (Philips, each 704 of 20 W, purchased in Argentina). The reaction progress was monitored 705 by TLC [eluent: hexane-ethyl acetate (8:2 v/v); spots were visualized 706 with UV light (254 and 366 nm)] and by GC analysis (Ultra 2 capillary 707 column). When the conversion of the starting material was higher than 708 90%, the photolyzed solution was carefully evaporated to dryness under 709 reduced pressure. The yellowish solid residue obtained was purified by 710 silica gel column chromatography (eluent: hexane 100% followed by 711 hexane-ethyl acetate mixtures). From the eluted fractions, the 712 photoproducts were isolated and characterized by means of physical 713 and spectroscopic methods. 714

Photoirradiations of Phenyl Benzoates in Micellar Media. 715 Stock solutions of surfactants in deionized water (SDS 0.10 M and Brij-716 P35 0.05 M) were freshly prepared before each experiment. The aryl 717 benzoate (5 mg) was placed in a stoppered quartz cell provided with a 718 stirring bar (3 mL), and the surfactant stock solution (2 mL) was added. 719 Then the solution was vigorously stirred for 1 h and degassed with 720 argon for 20 min. The quartz cell was placed in a homemade optical 721 bench provided with two germicide lamps (each of 20 W). The progress 722 of the photoreaction was monitored by two different methods: (i) UV- 723 vis spectroscopy and GC analysis (Ultra 2 capillary column). The 724 conversion of the benzoates was kept below 20% to avoid secondary 725 reactions and the formation of byproducts. Previous to the injection 726 into the GC apparatus, the micellar solutions were treated as follows. 727 The photolyzed solutions were diluted with 2 mL of an aqueous 728 solution of NaCl and then extracted with ethyl acetate $(3 \times 2 \text{ mL})$ while 729 the system was carefully shaken to avoid the formation of emulsions. 730 The organic layer was separated, dried over Na₂SO₄, and evaporated to 731 dryness under vacuum. The yellowish solid residue was diluted in 732 dichloromethane (2.00 mL), and this solution was injected into the GC 733

734 for chromatographic analysis. The products were characterized by 735 comparison of physical constant (mp) and spectroscopic data (1 H 736 NMR and 13 C NMR) with those reported in the literature.

737 *p-Methoxyphenyl Benzoate (1).* White needles (2.24 g; 98%). Mp: 738 89–90 °C (lit.²² mp 87–88 °C). ¹H NMR (300 MHz, CDCl₃): δ 8.23 739 (d, J = 8.6 Hz, J = 1.4 Hz, 2H), 7.66 (dd, J = 8.6 Hz, J = 1.3 Hz, 2H), 7.53 740 (t, J = 7.8 Hz, J = 1.1 Hz, 1H), 7.17 (d, J = 9.4 Hz, 2H), 6.97 (d, J = 9.4 741 Hz, 2H), 3.85 (s, 3H). ¹³C{¹H} NMR (75 MHz, CDCl₃): δ 165.4, 742 157.2, 144.3, 133.4, 130.0, 129.6, 128.4, 122.3, 114.4, 55.5.

743 *p-Phenoxyphenyl Benzoate (2).* White plates (2.84 g; 98%). Mp: 744 100–101 °C.²³ ¹H NMR (300 MHz, CDCl₃): δ 8.24 (d, *J* = 7.1, 1.0 Hz, 745 2H), 7.67 (t, *J* = 7.7 Hz, 1H), 7.55 (t, *J* = 7.9 Hz, 2H), 7.39 (t, *J* = 8.4 Hz, 746 2H), 7.21 (d, *J* = 8.9 Hz, 2H), 7.18–7.04 (m, 5 H). ¹³C{¹H} NMR (75 747 MHz, CDCl₃): δ 165.2, 157.1, 146.2, 133.5, 130.1, 129.7, 129.4, 128.5, 748 123.5, 122.7, 119.6, 118.7.

749*p-Methylphenyl Benzoate* (3). White needles (2.02 g; 95%). Mp: 71750°C (lit.²⁴ mp 72 °C). ¹H NMR (300 MHz, CDCl₃): δ 8.24 (d, J = 8.2751Hz, J = 1.4 Hz, 2H), 7.65 (dd, J = 7.4 Hz, J = 1.2 Hz, 2H), 7.54 (t, J = 7.8752Hz, J = 1.2 Hz, 1H), 7.25 (d, J = 8.3 Hz, 2H), 7.13 (d, J = 8.5 Hz, 2H),7532.41 (s, 3H). ¹³C{¹H} NMR (75 MHz, CDCl₃): δ 165.3, 148.6, 135.4,754133.4, 130.1, 129.6, 129.9, 128.4, 121.3, 20.8.

755 *p-tert-Butylphenyl Benzoate* (**4**). White plates (2.44 g; 96%). Mp: 756 83–84 °C (lit.²⁵ mp 82–83 °C). ¹H NMR (300 MHz, CDCl₃): δ 8.23 757 (dd, *J* = 7.9, 1.6 Hz, 2H), 7.66 (t, *J* = 7.4 Hz, 2H), 7.55 (t, *J* = 8.7 Hz, 758 1H), 7.47 (d, *J* = 8.7 Hz, 2H), 7.17 (d, *J* = 8.7 Hz, 2H), 1.35 (s, 9H). 759 ¹³C{¹H} NMR (75 MHz, CDCl₃): δ 165.2, 148.6, 148.5, 133.4, 130.0, 760 129.6, 128.4, 126.3, 120.9, 34.4, 31.6.

761 *Phenyl Benzoate (5).* White solid (1.94 g; 98%). Mp: 69–70 °C 762 (lit.^{6a} mp 67–69 °C). ¹H NMR (300 MHz, CDCl₃): δ 8.25 (d, *J* = 7.4 763 Hz, 2H), 7.67 (t, *J* = 7.6 Hz, 1H), 7.55 (t, *J* = 8.1 Hz, 2H), 7.45 (t, *J* = 764 8.3, 7.8 Hz, 2H), 7.32 (t, *J* = 8.1, 7.2 Hz, 1H), 7.24 (d, *J* = 7.2 Hz, 2H). 765 $^{13}C{^{1}H}$ NMR (75 MHz, CDCl₃): δ 165.1, 150.9, 133.5, 130.1, 129.5, 766 129.4, 128.5, 125.8, 121.6.

p-Phenylphenyl Benzoate (6). White needles (2.52 g; 92%). Mp: 150–151 °C.²³ ¹H NMR (300 MHz, CDCl₃): δ 8.20 (d, *J* = 8.2, 1.2 Hz, 269 2H), 7.72–7.61 (m, 5H), 7.57 (t, *J* = 7.9 Hz, 2H), 7.49 (t, *J* = 7.8 Hz, 770 2H), 7.41 (d, *J* = 7.2 Hz, 1H), 7.33 (d, *J* = 8.6 Hz, 2H). ¹³C{¹H} NMR 771 (75 MHz, CDCl₃): δ 165.1, 150.3, 140.3, 138.9, 133.5, 130.1, 129.4, 772 128.7, 128.5, 128.1, 127.3, 127.0, 121.9.

p-*Cyanophenyl Benzoate* (7). White solid (2.01 g; 90%). Mp: 94– 774 95 °C (lit.²⁶ mp 91–92 °C). ¹H NMR (300 MHz, CDCl₃) δ: 8.22 (d, *J* 775 = 8.4 Hz, 2H), 7.74 (d, *J* = 7,64 Hz 2H), 7.70 (t, *J* = 7.0, 7.64 Hz, 2H) 776 7.56 (t, *J* = 7.6 Hz, 2H), 7.4 (d, *J* = 8.4 Hz, 2H). ¹³C{¹H} NMR (75 777 MHz, CDCl₃) δ: 164.2, 154.1, 134.0, 133.6, 130.1, 128.6, 128.5, 122.8, 778 118.2, 109.7.

p-*Nitrophenyl Benzoate* (8). Pale yellow needles (2.21 g; 91%). Mp: 780 144–145 °C (lit.²⁷ mp 142–144 °C). ¹H NMR (300 MHz, CDCl₃) δ: 781 8.34 (d, *J* = 9.1 Hz, 2H), 8.23 (dd, *J* = 7.6, 1.3 Hz, 2H), 7.71 (t, *J* = 7.5 782 Hz, 1H), 7.57 (t, *J* = 7.5 Hz, 2H), 7.45 (d, *J* = 9.1 Hz, 2H). ¹³C{¹H} 783 NMR (75 MHz, CDCl₃) δ: 164.1, 155.6, 145.3, 134.2, 130.2, 128.7, 784 128.4, 125.2, 122.5.

785 **2-Hydroxy-5-methoxybenzophenone** (1*a*). Pale yellow needles 786 (188 mg; 94%). Mp: 83–84 °C (lit.²⁸ mp 84 °C). ¹H NMR (300 MHz, 787 CDCl₃) δ : 11.58 (s, 3H); 7.69 (dd, *J* = 8.5, 1.6 Hz, 2H), 7.60 (t, *J* = 7.5 788 Hz, 1H), 7.51 (t, *J* = 7.8 Hz, 2H), 7.14 (d, *J* = 9.0, 2.9 Hz, 1H), 7.06 (d, *J* 789 = 3.1 Hz, 1H), 7.02 (d, *J* = 9.0 Hz, 1H), 3.70 (s, 3H). ¹³C{¹H} NMR 790 (75 MHz, CDCl₃) δ : 201.1, 157.7, 151.6, 138.0, 132.1, 130.3, 129.2, 791 128.5, 124.3, 119.4, 116.5, 56.1.

2-*Hydroxy-5-phenoxybenzophenone* (2*a*). Yellow needles (193 mg; 76%). Mp: 47–48 °C. ¹H NMR (300 MHz, CDCl₃) δ: 11.85 (s, 794 1H), 7.70 (d, *J* = 8.1 Hz, 2H), 7.59 (t, *J* = 7.4, 1H), 7.50 (t, *J* = 7.6 Hz, 795 2H), 7.34–7.26 (m, 4 H), 7.12 (d, *J* = 8.7 Hz, 1H), 7.07 (t, *J* = 7.4 Hz, 796 1H), 6.94 (d, *J* = 8.7 Hz, 2H). ¹³C{¹H} NMR (75 MHz, CDCl₃) δ: 797 200.9, 159.6, 158.6, 147.7, 137.5, 132.2, 129.8, 129.2, 128.9, 128.5, 798 123.8, 122.8, 119.7, 119.2, 117.3. Anal. Calcd for $C_{19}H_{14}O_3$: C, 78.61; 799 H, 4.86. Found: C, 78.57; H, 4.90.

800 2-Hydroxy-5-methylbenzophenone (**3a**). Pale yellow needles (130 801 mg; 70%). Mp: 83–84 °C (lit.²⁹ mp 84 °C). ¹H NMR (300 MHz, 802 CDCl₃) δ: 11.84 (s, 1H), 7.67 (dd, J = 8.4, 1.2 Hz, 2H), 7.60 (t, J = 7.4, 803 1H), 7.52 (t, J = 7.8 Hz, 2H), 7.36 (d, J = 2.8 Hz, 1 H), 7.32 (dd, J = 8.5, 2.3 Hz, 1H), 6.98 (d, J = 8.54 Hz, 1H), 2.28 (s, 3H).). ¹³C{¹H} NMR ⁸⁰⁴ (75 MHz, CDCl₃) δ : 201.7, 161.3, 138.2, 137.5, 133.3, 131.9, 129.2, ⁸⁰⁵ 128.5, 127.9, 118.9, 118.3, 20.6. ⁸⁰⁶

2-Hydroxy-5-t-butylbenzophenone (4a). White solid (189 mg; 807 85%). M: 67–68 °C (lit.²⁵ mp 67–68 °C). ¹H NMR (300 MHz, 808 CDCl₃) δ : 11.87 (s, 1 H), 7.72 (dd, *J* = 1.6, 8.1 Hz, 2H), 7.63 (t, *J* = 1.3, 809 7.4, 2H), 7.60 (t, *J* = 2.5, 8.5 Hz, 1H), 7.55 (t, *J* = 1.6, 7.2 Hz, 2 H), 7.05 810 (d, *J* = 8.3 Hz, 1H), 1.28 (s, 9H). ¹³C{¹H} NMR (75 MHz, CDCl₃) δ : 811 201.6, 161.0, 141.3, 138.1, 133.9, 131.9, 129.8, 129.3, 128.3, 118.4, 812 117.9, 34.1, 31.3.

2-Hydroxybenzophenone (**5***a*). Yellow needles (156 mg; 90%). 814 Mp: 37–38 °C (lit.^{6a} mp 37–38 °C). ¹H NMR (300 MHz, CDCl₃) δ : 815 12.05 (s, 1H), 7.68 (dd, *J* = 1.4, 8.4 Hz, 2H), 7.61–7.57 (m, 2H), 7.53–816 7.48 (m, 3H), 7.08 (dd, *J* = 1.1, 8.5 Hz, 1 H), 6.88 (dd, *J* = 1.1, 7.2 Hz, 817 1H). ¹³C{¹H} NMR (75 MHz, CDCl₃) δ : 201.7, 163.3, 138.0, 136.4, 818 133.7, 132.0, 129.3, 129.2, 128.5, 119.2, 118.8, 118.5, 118.3. 819

4-Hydroxybenzophenone. Yellow plates (14 mg; 8%). Mp: 133– 820 134 °C (lit.^{6a,30} mp 133–134 °C). ¹H NMR (300 MHz, CDCl₃) δ: 7.77 821 (d, *J* = 8.8 Hz, 2H), 7.75 (d, *J* = 8.4, 2H), 7.57 (t, *J* = 7.4, 1.3 Hz, 1H), 822 7.47 (t, *J* = 7.5 Hz, 2 H), 6.95 (d, *J* = 8.7 Hz, 2H). ¹³C{1H} NMR (75 823 MHz, CDCl₃) δ: 197.2, 161.1, 138.1, 133.3, 132.4, 130.0, 129.5, 128.4, 824 115.6. 825

2-Hydroxy-5-phenylbenzophenone (**6a**). White needles (209 mg; 826 87%). Mp: 91–92 °C (lit.³¹ mp 91–92 °C). ¹H NMR (300 MHz, 827 CDCl₃) δ: 12.06 (s, 1 H), 7.80 (dd, J = 2.4, 8.6 Hz, 1H), 7.77 (dd, J = 828 1.5, 8.5, 2H), 7.70–7.62 (m, 2H), 7.56 (t, J = 7.7, 1.7 Hz, 2H), 7.49 (dd, 829 J = 8.5, 1.4 Hz, 2 H), 7.44 (t, J = 7.5, 1.5 Hz, 2H), 7.35 (t, J = 7.2 Hz, 830 1H), 7.21 (d, J = 8.6 Hz, 1H). ¹³C{¹H} NMR (75 MHz, CDCl₃) δ: 831 201.7, 162.6, 139.8, 137.9, 135.1, 133.7, 132.1, 132.0, 131.8, 129.3, 832 128.9, 128.8, 128.5, 119.3, 118.9.

2-Hydroxy-5-cyanobenzophenone (**7a**). Pale yellow needles (156 s34 mg; 80%). Mp: 120–121 °C (lit.³² mp 120–121 °C). ¹H NMR (300 s35 MHz, CDCl₃) δ: 12.48 (s, 1H), 7.96 (d, J = 2.1, 1H), 7.74 (dd, J = 8.7, 836 2.1 Hz, 1H), 7.70–7.65 (m, 3H), 7.60–7.55 (m, 2H), 7.16 (d, J = 8.7 kz, 1H). ¹³C{¹H} NMR (75 MHz, CDCl₃) δ: 200.6, 166.4, 138.7, 838 138.4, 136.6, 133.1, 129.3, 129.0, 120.2, 119.4, 118.3, 102.6. 839

2-Hydroxy-5-nitrobenzophenone (**8a**). Intense yellow plates (94 840 mg; 44%). Mp.: 123–124 °C (lit.³³ mp 123–124 °C). ¹H NMR (300 841 MHz, CDCl₃) δ : 12.67 (s, 1H), 8.60 (d, *J* = 2.7 Hz, 1H), 8.40 (dd, *J* = 842 9.2, 2.7 Hz, 1H), 7.72 (d, *J* = 8.4, 1.3 Hz, 2H), 7.69 (t, *J* = 7.5, 1.3 Hz, 843 1H), 7.59 (t, *J* = 7.8, 1.8 Hz, 2H), 7.19 (d, *J* = 9.2 Hz, 1H). ¹³C{¹H} 844 NMR (75 MHz, CDCl₃) δ : 200.7, 168.1, 139.6, 136.5, 133.3, 131.1, 845 129.8, 129.4, 129.2, 119.7, 118.1.

ASSOCIATED CONTENT

S Supporting Information

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UV-vis absorption spectra under steady-state and time- 851 resolved spectroscopy. Relative absorption profiles. 852 Determination of the constants of binding $K_{\rm b.}$ 2D 853 NOESY NMR spectra in micellar media. Determination 854 of the rate constants $k_{\rm E}$ and $k_{\rm R}$ in homogeneous and 855 heterogeneous media. 1 H and 13 C spectra of aryl 856 benzoates and of 2-hydroxy-5-substituted benzophenones 857 (PDF) 858

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