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1 **Gastrointestinal diseases and their impact on drug solubility: Celiac disease**

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18 **Abstract**

19 The aim of this study was to develop an *in vitro* tool for predicting drug solubility and  
20 dissolution in intestinal fluids of patients with Celiac disease (CED). Biorelevant media for  
21 patients with CED were developed based on published information and a Design of Experiment  
22 (DoE) approach. The CED biorelevant media were characterised according to their surface  
23 tension, osmolality, dynamic viscosity and buffer capacity. By performing solubility studies of  
24 six drugs with different physicochemical properties in CED media, we aimed to identify drugs  
25 at high risk of altered luminal solubility in CED patients. Identified differences in CED patients  
26 compared to healthy subjects were related to a higher concentration of bile salts, lecithin and  
27 cholesterol and included as factors in the DoE resulting in 8 CED biorelevant media.  
28 Differences in media properties were observed for the surface tension between biorelevant  
29 media based on CED patients and healthy subjects. In terms of solubility, only a minimal effect  
30 of CED on the solubility of the hydrophilic neutral compound azathioprine was observed. For  
31 neutral moderately lipophilic compounds (budesonide, celecoxib) a higher surfactant  
32 concentration resulted in most cases in a higher drug solubility, while it was specific to each  
33 drug whether this was mainly driven by bile salts or lecithin. In comparison, drug solubilisation  
34 of ionisable compounds with moderate to high lipophilicity was less impacted by CED  
35 differences. The developed biorelevant CED media serve as *in vitro* tool to identify the main  
36 media factors impacting on drug solubility.

37 **Keywords**

38 Gastrointestinal diseases; Celiac disease; Biorelevant media; Physicochemical properties;  
39 Solubility

## 40 **1. Introduction**

41 Celiac disease (CED) is a chronic auto-inflammatory disease induced by an intolerance to  
42 dietary gluten, a storage protein of wheat, rye, barley and oats. Approximately 1% of the  
43 population is affected by CED and its aetiology is a combination of genetic predisposition and  
44 environmental factors (e.g., breastfeeding, time of gluten introduction and the microbiota)  
45 (Koehler et al., 2014). CED mainly affects the small intestine resulting in gastrointestinal (GI)  
46 symptoms such as bloating, diarrhoea, malabsorptive symptoms and weight loss. Additionally,  
47 CED patients can present extra-intestinal symptoms such as dermatitis herpetiformis, anaemia  
48 or osteoporosis (Leffler et al., 2015). The diagnosis involves serological testing for  
49 autoantibodies (anti-tTG, anti-EMA) and an endoscopic biopsy (Turner et al., 2015).  
50 Depending on the damage to the small intestine, the disease can be classified in different  
51 disease grades based on histological findings such as crypt hyperplasia, the constitution of the  
52 villi and the intra-epithelial lymphocytes in the jejunum and duodenum (Oberhuber et al.,  
53 1999). For the treatment of CED, patients need to adhere to a gluten-free diet, the only known  
54 effective treatment to date, since the reintroduction of dietary gluten results in a relapse of the  
55 disease (Gottlieb et al., 2015). More treatment options are expected to emerge in the near future,  
56 since several new active pharmaceutical ingredients have reached clinical phases of drug  
57 development in recent years (Gottlieb et al., 2015).

58 Patient convenience dictates that oral administration is the preferred route of drug  
59 administration for most drugs. Consequently, patients with CED are likely to be treated with  
60 orally administered drug products for concomitant conditions or extra-intestinal manifestations  
61 of CED. Since oral drug administration is, apart from drug and formulation properties,  
62 dependent on gastrointestinal physiology, pathophysiological changes in CED could affect  
63 drug safety and efficacy. GI diseases can affect various processes involved in oral drug delivery  
64 e.g., drug release from the formulation, drug dissolution, permeation through the GI membrane

65 and gut or hepatic metabolism (Effinger et al., 2019). Altered drug absorption in CED patients  
66 compared to healthy subjects has previously been attributed to a reduced small intestinal  
67 surface area, a different intestinal CYP enzyme abundance, a higher jejunal permeability and  
68 differences in gastric emptying (Tran et al., 2013).

69 So far, there is only a small number of drugs for which drug product performance has been  
70 investigated in CED patients and these studies included only a small number of patients (Tran  
71 et al., 2013). Due to the high costs of clinical trials, it is expected that in the future investigations  
72 in CED patients will remain rare.

73 For poorly soluble drugs, drug absorption can be limited by the dissolution rate or the solubility  
74 of the drug in gastrointestinal fluids (Amidon et al., 1995). If this is the case, *in vitro* release  
75 and dissolution testing can be used as surrogate for a drug's *in vivo* performance (Amidon et  
76 al., 1995). To simulate closely the conditions present in the GI tract, biorelevant media were  
77 developed mimicking the composition of the gastrointestinal fluids of healthy subjects (Galia  
78 et al., 1998; Jantratid et al., 2008; Markopoulos et al., 2015; Vertzoni et al., 2010; Vertzoni et  
79 al., 2005). The composition of the gastrointestinal fluids can be altered in patients with GI  
80 disease and therefore, *in vitro* dissolution and solubility studies with biorelevant media adapted  
81 to pathophysiological conditions could result in better predictions of drug product performance  
82 in patient populations (Effinger et al., 2019).

83 This study aims to identify drugs at risk of altered solubility in GI fluids of CED patients.  
84 Biorelevant media for patients with CED representative of the small intestinal fluid in the fasted  
85 and fed state were developed. Information from literature was collected to identify differences  
86 in the composition of luminal contents of patients with CED compared to healthy subjects.  
87 Biorelevant media for CED patients were developed based on biorelevant media for healthy  
88 subjects and a Design of Experiment (DoE) approach by integrating the identified differences

89 as factors with two levels. Subsequently, the CED biorelevant media were characterised in  
90 terms of surface tension, osmolality, buffer capacity and dynamic viscosity. Additionally, the  
91 solubility of six compounds with different physicochemical properties (including azathioprine,  
92 budesonide, celecoxib, dipyridamole, loperamide and sulfasalazine), in the developed  
93 biorelevant media based on CED patients and healthy subjects was determined.

## 94 **2. Materials**

95 Acetic acid High Performance Liquid Chromatography (HPLC) grade, chloroform, sodium  
96 oleate, budesonide, phosphoric acid and sodium hydroxide were purchased from Sigma-  
97 Aldrich Company Ltd., Dorset, England. Sulfasalazine, loperamide hydrochloride,  
98 dipyridamole, celecoxib, azathioprine, methanol HPLC grade and acetonitrile HPLC grade  
99 were purchased from VWR International Ltd, Lutterworth, UK. Sodium chloride,  
100 trifluoroacetic acid (TFA), potassium dihydrogen orthophosphate, dimethyl sulfoxide and  
101 maleic acid were used from Fisher Scientific UK Ltd., Loughborough, England. Other  
102 chemicals used included sodium taurocholate (Prodotti Chimici Alimentari S.P.A., Basaluzzo,  
103 Italy), egg lecithin–Lipoid EPCS (Lipoid GmbH, Ludwigshafen, Germany), glyceryl  
104 monooleate–Rylo Mg 19 (Danisco, Brabrand, Denmark) and cholesterol (95%, Acros  
105 Organics, Geel, Belgium). Water was ultra-pure (Milli-Q) laboratory grade.

## 106 **3. Methods**

### 107 3.1. Media development

#### 108 3.1.1. GI physiological differences in CED compared to healthy subjects

109 To identify differences in the composition of GI fluids of untreated CED patients compared to  
110 healthy subjects, a literature search was performed. Since to date the GI fluids of CED patients  
111 have not been directly characterised, studies investigating parameters that most likely impact  
112 on GI fluids were considered.

113 The bile flow and biliary lipid output has previously been measured in untreated CED patients  
114 and healthy subjects during a constant infusion of a liquid formula diet using a duodenal  
115 intubation technique (Vuoristo and Miettinen, 1985). Biliary lipid outputs such as cholesterol,  
116 bile acids and phospholipids could be estimated in comparison to the dilution of a marker  
117 (polyethylene glycol 4000). The bile flow was with  $232\pm 29$  mL/h (mean  $\pm$  SD) significantly  
118 higher in CED patients compared to  $132\pm 24$  mL/h in healthy subjects (Student's t-test,  $p<0.05$ ).  
119 The biliary cholesterol output normalised to the body weight was significantly increased in  
120 CED patients ( $0.82\pm 0.10$  vs  $0.43\pm 0.06$  mg/kg\*h,  $p<0.02$ ). Similarly, the biliary output of  
121 phospholipids was also highly increased in CED patients compared to healthy subjects  
122 ( $0.26\pm 0.05$  vs  $0.08\pm 0.02$  mg/kg\*h,  $p<0.02$ ). Additionally, a higher bile acid output was  
123 observed in CED patients ( $9.28\pm 1.65$  vs  $4.64\pm 0.45$  mg/kg\*h). In accordance, it was observed  
124 that the bile salt pool is three times higher in CED patients compared to healthy subjects, which  
125 could be related to a very effective ileal reabsorption of bile acids or a sluggish contraction of  
126 the gall bladder (Low-Beer et al., 1973). Since our study was based on untreated CED patients  
127 (not adhering to a gluten-free diet), dietary differences between CED patients and healthy  
128 subjects were not considered.

### 129 3.1.2. Development of CED media with Design of Experiment

130 The development of biorelevant media for CED patients followed a DoE approach and CED  
131 biorelevant media representative of the small intestinal fluid in the fasted and fed state were  
132 developed. Biorelevant media previously developed based on healthy subjects were used as the  
133 basis for CED biorelevant media and included Fasted-State Simulated Intestinal Fluid-Version  
134 2 (FaSSIF-V2) and Fed-State Simulated Intestinal Fluid-Version 2 (FeSSIF-V2) (Jantratid et  
135 al., 2008). According to the identified differences described in Section 3.1.1, biorelevant media  
136 based on healthy subjects were modified by including the differences as factors in the  
137 experimental design. For both prandial states, the integrated factors in the experimental design

138 were the concentration of bile salts, lecithin and cholesterol. Since the biliary secretion is the  
139 main source of bile salts, lecithin and cholesterol present in the intestinal fluids, a direct  
140 correlation between biliary output and intestinal concentration was assumed. Since the three  
141 parameters were not directly measured in the GI fluids, an indirect percental approach was  
142 followed to determine the level of the corresponding factor according to

$$143 \quad x_{CED-BM} = \frac{y_{CED}}{y_H} * x_{H-BM} \quad (1)$$

144 where  $x_{CED-BM}$  is the high level of the factor in CED media,  $y_{CED}$  and  $y_H$  are the median of  
145 the corresponding biliary output observed in CED patients and healthy subjects, respectively  
146 and  $x_{H-BM}$  is the level of the factor in biorelevant media based on healthy subjects.

147 The three factors were integrated with two levels in the experimental design, a low and a high  
148 level. The low level was based on the concentration in biorelevant media based on healthy  
149 subjects (Table 1) and the high level corresponded to the median percentage of the respective  
150 concentration in the healthy medium. For cholesterol, the low level concentration was based  
151 on the median concentration of cholesterol observed in human intestinal fluid as observed by  
152 Riethorst et al. (2016) [fasted state: 0.08 mM, fed state: 0.57 mM], since cholesterol is not a  
153 component of FaSSIF-V2 and FeSSIF-V2.

154 The DoE was performed with Statgraphics Centurion 18 (Statpoint Technologies Inc., VA, US)  
155 with a full factorial design for CED intestinal biorelevant media for the fasted and fed state.  
156 An overview of the DoE is given in Figure 1. Biorelevant media were prepared as previously  
157 described with an additional step of adding cholesterol (Jantratid et al., 2008). The cholesterol  
158 solution (50 mg/mL in chloroform) was mixed with a lecithin solution (100 mg/ml in  
159 dichloromethane) using a magnetic stirrer, before being added to the bile salt/buffer mixture  
160 and driven off using a rotary evaporator Büchi Rotovapor R-114 (Büchi Labortechnik, Flawil,  
161 Switzerland) according to the published protocol. The osmolality of CED media was set to the



162 value in the corresponding biorelevant medium based on healthy subjects by adjusting the  
163 concentration of sodium chloride.

### 164 3.1.3. Media characterisation

165 Surface tension, osmolality, dynamic viscosity and buffer capacity of biorelevant media  
166 previously developed based on healthy subjects and newly developed for CED patients were  
167 measured in triplicate. The results are reported as mean with standard deviation.

#### 168 3.1.3.1. Surface tension

169 Surface tension measurements were performed at room temperature with a ring tensiometer  
170 (Sigma 700 Force tensiometer, Attension, UK) using approximately 10 mL of each medium,  
171 placed in a glass vessel with a diameter of 46 mm. A platinum Du Noüy ring was lowered  
172 below the meniscus of the medium. Subsequently, by pushing and pulling the ring through the  
173 surface of the medium, the force exerted by the meniscus was measured and related to the  
174 surface tension of the medium (Butt et al., 2004).

#### 175 3.1.3.2. Osmolality

176 Osmolality was determined with an Advanced Instruments Inc. micro-osmometer Model 3300  
177 (Norwood, MA, US). Therefore, the freezing-point depression of a 20 µl sample was measured  
178 with a high-precision thermistor following the supercooling and induced crystallisation of the  
179 sample.

#### 180 3.1.3.3. Dynamic viscosity

181 The dynamic viscosity at 37°C was measured with a Bohlin Rheometer C-VOR (Malvern  
182 instruments, UK). Therefore, a cone-plate measuring system, including a rotating upper cone  
183 (4°, 40mm) and a fixed lower plate with the medium contained between them, was used. The  
184 shear rate was measured while twenty different shear stresses, logarithmically distributed in  
185 the range of 0.05 to 0.15 Pa, were exerted on the sample of the medium. The ratio of shear  
186 stress to shear rate corresponds to the dynamic viscosity.

187 3.1.3.4. Buffer capacity

188 Buffer capacity was determined using a potentiometric titration method. Therefore, small  
189 volumes of 0.5 M hydrochloric acid were added to 10 mL of sample until a change of one pH  
190 unit was recorded by a Mettler Toledo SevenCompact S220 pH meter (Schwerzenbach,  
191 Switzerland). Equation (2) was used to calculate the buffer capacity ( $\beta$ ) according to

192 
$$\beta = \left( \frac{0.5M * V_{acid}}{\Delta pH} \right) * \frac{1000}{V_s} \quad (2)$$

193 where  $V_{acid}$  is the volume of the acid added,  $V_s$  is the volume of the sample and  $\Delta pH$   
194 corresponds to the change in pH (Rabbie et al., 2015).

195 3.2. Compound selection

196 For the solubility studies, low soluble compounds belonging to Biopharmaceutics  
197 Classification System (BCS) class II (low solubility, high permeability) or IV (low solubility,  
198 low permeability) were selected as shown in Table 1. Additionally, the selected drugs varied  
199 in their ionization properties (pKa) and lipophilicity (logP). Drugs with indication for  
200 gastrointestinal diseases were preferred.

201 **Table 1:** Properties and indication of selected compounds for solubility studies.

Drug	pKa (acid/base)	logP	BCS class	Intrinsic aqueous solubility [mg/mL]	Indication
<b>Azathioprine</b>	7.9 (acid) (Mitra and Narurkar, 1987)	0.1 (Hansch et al., 1995)	IV (Lindenberg et al., 2004)	0.171 (Llinas et al., 2008)	Immunosuppressive
<b>Budesonide</b>	12.0 (acid) (Corey and Fossel, 2016)	2.6 (Bharate et al., 2016)	II (Bhatt et al., 2014)	0.028 (Ali et al., 2010)	Locally acting corticosteroid in IBD
<b>Celecoxib</b>	11.1 (acid) (G.D. Searle LLC Division of Pfizer Inc, 2019)	3.5 (G.D. Searle LLC Division of Pfizer Inc, 2019)	II (Paulson et al., 2001)	0.003 - 0.007 (Paulson et al., 2001)	Nonsteroidal anti-inflammatory drug
<b>Dipyridamole</b>	6.4 (base) (Pedersen, 1979)	2.2 (Betageri and Dipali, 1993)	II (Zaki et al., 2010)	0.003 (Hopfinger et al., 2009)	Platelet aggregation inhibitor
<b>Loperamide</b>	8.6 (base) (Manallack, 2007)	5.5 (Dickson et al., 2017)	II (Zaki et al., 2010)		Anti-diarrheal agent
<b>Sulfasalazine</b>	2.3, 7.9 (acid) (Shalaeva et al., 2008)	2.9 (Graham and Pile, 2015)	II/IV (Lindenberg et al., 2004)	$0.29 \times 10^{-3}$ (Llinas et al., 2008)	Anti-inflammatory agent in IBD

202

### 203 3.3. Solubility studies

204 The shake-flask method was used to determine the solubility of the investigated compounds  
 205 (Baka et al., 2008). Therefore, an excess amount of drug was added to 5 mL of the respective  
 206 medium in a glass tube, which was then placed in a shaking water bath (Grant instruments,  
 207 UK) and maintained at 37°C and 200 strokes/min for 24 h. Subsequently, GF/D membrane  
 208 filters with a pore size of 2.7 µm (Whatman® Puradisc, diameter 13 mm) were used to filter the  
 209 sample followed by quantitative analysis with HPLC/UV. The solubility studies were

210 performed in triplicate in CED disease media and healthy media and average solubility  
211 differences between CED media and healthy media were expressed as a % Relative effect on  
212 solubility  $(((S_{CED}-S_{Healthy})/ S_{Healthy}) \times 100)$ . A higher drug solubility in CED media compared to  
213 healthy media is indicated by a positive value, whereas the opposite is indicated for negative  
214 values. HPLC analysis was performed with an Agilent Technologies 1200 series HPLC system  
215 (Santa Clara, CA) including a binary pump (G1212A), an autosampler (G1329A), a  
216 thermostatted column compartment (G1316A) and a diode array detector (G1315D). The  
217 methods used for the HPLC-UV analysis of the six drugs were modifications of previously  
218 published methods (presented in Gastrointestinal diseases and their impact on drug solubility:  
219 Crohn's disease) (Effinger et al., 2020).

#### 220 3.4. Statistical analysis

221 Differences between media properties and drug solubility in biorelevant media based on CED  
222 patients compared to healthy subjects were identified with the software XLSTAT (Addinsoft,  
223 France) using one-way analysis of variance (ANOVA) with a post-hoc Tukey's test and a  
224 significance level of  $p \leq 0.05$ .

225 A multifactorial ANOVA performed in Statgraphics Centurion 18 (Statpoint Technologies  
226 Inc., VA, US) was used to estimate the effects of the three categorical variables (bile salts,  
227 lecithin, cholesterol) and two-factor interactions in the DoE on the solubility of each of the six  
228 investigated compounds. Factors were considered statistically significant if the p-value was  
229 less than 0.05, indicating an effect on drug solubility at the 95.00% confidence level.

## 230 **4. Results and discussion**

### 231 4.1. Media characterisation

232 The surface tension of intestinal CED biorelevant media is shown in Figure 2 and was in the  
233 range of 45.5 to 51.6 mN/m and of 26.6 to 35.7 mN/m for the fasted and fed state, respectively.

234 In the fasted state, the surface tension of all media with low bile salt concentration was higher  
235 compared to the healthy medium ( $p<0.05$ ). This finding is consistent with another study, where  
236 a higher surface tension was observed for reduced bile salt concentrations in fasted state  
237 simulating fluids without cholesterol (Xie et al., 2014). Additionally, media with at the same  
238 time high bile salt and lecithin concentrations possessed a significantly higher surface tension  
239 compared to the healthy medium but a lower surface tension compared to all CED media with  
240 low bile salt concentrations ( $p<0.05$ ). In the fed state, the surface tension of all CED media  
241 with low lecithin concentrations, except for the medium with at the same time low bile salt and  
242 cholesterol concentrations, was significantly decreased ( $p<0.05$ ).

243 The osmolality of biorelevant media based on CED patients and healthy subjects was not  
244 significantly different.

245 The measured dynamic viscosities of CED biorelevant media at a shear stress of 0.06 Pa, 0.08  
246 Pa and 0.15 Pa are presented in Figure 3. All healthy and CED media showed shear thinning  
247 behaviour. The viscosity of CED biorelevant media at an applied shear stress of 0.15 Pa was  
248 in the range of 3.26 to 3.56 mPas, at 0.08 Pa in the range of 3.70 to 4.56 mPas and at 0.06 Pa  
249 in the range of 4.28 to 6.42 mPas, respectively. No significant differences between biorelevant  
250 media based on CED patients and healthy subjects were observed considering all three different  
251 shear stresses ( $p<0.05$ ).

252 The buffer capacity was not significantly different in fasted and fed state intestinal media based  
253 on healthy subjects compared to CED patients, since the same buffer composition was used  
254 and no changes of the media pH were applied (data not shown).

#### 255 4.2. Solubility of drugs in CED biorelevant media

256 Considering the final pH value of the medium after 24 h, the pH was within  $6.5 \pm 0.1$  and  $5.8$   
257  $\pm 0.1$  in all cases except for the sulfasalazine studies in fasted (final medium pH:  $6.2 \pm 0.1$ ) and  
258 fed state (final medium pH  $5.7 \pm 0.1$ ) intestinal media.

##### 259 4.2.1. Neutral drugs

260 The results of the solubility studies with neutral compounds in CED fasted and fed state  
261 intestinal media are illustrated in Figure 4.

262 For azathioprine, the solubility in the fasted state was not significantly different in CED media  
263 compared to healthy media. In the fed state, the solubility of azathioprine was significantly  
264 higher in CED biorelevant media with high concentrations of bile salts but the relative increase  
265 was for all media below 15%.

266 For budesonide, the solubility in all fasted state CED biorelevant media was significantly  
267 higher compared to the healthy medium ( $p < 0.05$ ), whereby the solubility of budesonide was  
268 highest in CED media with high bile salt concentrations. The positive effect of bile salts is in  
269 accordance with a previous study showing that an increase of the concentration of bile salts in  
270 a fixed 4:1 ratio of bile salts to lecithin resulted in an increase in budesonide solubility  
271 (Soderlind et al., 2010). Additionally, the positive effect of cholesterol on budesonide  
272 solubilisation indicates a drug-cholesterol interaction or a positive solubilisation effect of more  
273 complex vesicles (sodium taurocholate-lecithin-cholesterol) as previously reported for  
274 fenofibrate (Khoshakhlagh et al., 2015).

275 In the fed state, the solubility of budesonide in the CED media with at the same time low  
276 concentrations of bile salts and lecithin was significantly decreased compared to the healthy  
277 medium ( $p<0.05$ ), indicating a competition for solubilisation between cholesterol and  
278 budesonide possibly due to the similarity of their chemical structure. In contrast, a significantly  
279 higher solubility was observed in CED media with high concentrations of bile salts and lecithin  
280 and CED media with either a high concentration of bile salts or lecithin and a low concentration  
281 of cholesterol ( $p<0.05$ ), indicating a positive effect of higher surfactant concentration and a  
282 negative effect of cholesterol on budesonide solubility.

283 For celecoxib, the solubility in fasted state CED media with a high concentration of lecithin  
284 and a low concentration of cholesterol was significantly higher compared to the healthy  
285 medium. In contrast, in all other CED fasted state media, the solubility of celecoxib was  
286 significantly lower ( $p<0.05$ ). The positive effect of lecithin on celecoxib solubility is in  
287 accordance with previous results revealing a higher solubility of celecoxib in FaSSIF (higher  
288 concentration of lecithin) compared to FaSSIF-V2 (Shono et al., 2009).

289 In the fed state, the solubility of celecoxib was significantly higher in CED media with at the  
290 same time high concentrations of bile salts and lecithin ( $p<0.05$ ), suggesting a positive effect  
291 of luminal surfactants on celecoxib solubility.

#### 292 4.2.2. Weak acid

293 The results of the solubility studies in CED fasted and fed state intestinal media with  
294 compounds possessing different ionisation properties are presented in Figure 5.

295 For the weak acid sulfasalazine, the solubility in fasted state CED media with at the same time  
296 high concentrations of lecithin and low concentrations of cholesterol is significantly lower  
297 compared to the healthy medium ( $p<0.05$ ). In fed state intestinal media, the solubility of  
298 sulfasalazine was significantly higher in CED media with high bile salt concentrations and in

299 the medium with a low concentration of bile salts and lecithin and a high concentration of  
300 cholesterol.

#### 301 4.2.3. Weak bases

302 For the weak base dipyridamole, the solubility was significantly higher in fasted state CED  
303 media with high bile salt concentrations and to a lower extent also in the medium with a high  
304 concentration of lecithin and low concentrations of bile salts and cholesterol ( $p<0.05$ ). The  
305 positive effect of bile salts on the solubility of dipyridamole is most likely the result of  
306 electrostatic interactions of the weak base with sodium taurocholate. In the fed state, the  
307 solubility of dipyridamole in the CED medium with a high concentration of lecithin and low  
308 concentrations of bile salts and cholesterol was significantly lower compared to the  
309 corresponding healthy medium ( $p<0.05$ ).

310 For loperamide hydrochloride, the solubility in the fasted state CED media with high  
311 concentrations of lecithin and cholesterol and a low concentration of bile salts was significantly  
312 lower compared to the corresponding healthy medium ( $p<0.05$ ). This is possibly due to less  
313 bile salts being available for drug solubilisation due to the need for lecithin and cholesterol  
314 solubilisation. In the fed state, the solubility of loperamide hydrochloride was not significantly  
315 different in CED media compared to the corresponding healthy medium ( $p<0.05$ ).

#### 316 4.3. Multifactorial statistical analysis of solubility in CED media

317 For CED fasted state intestinal media, the significant effects and two-factor interactions  
318 affecting the drug solubility of the six investigated drugs are presented in Table 2.

319 For azathioprine and budesonide, only the bile salt concentration had a positive impact on their  
320 solubility. For celecoxib, the highest positive effect on solubility was observed for the lecithin  
321 concentration, followed by a negative effect of cholesterol. Additionally, all two-factor  
322 interactions were significant for the solubility of celecoxib but less influential in comparison



323 to both main effects. For dipyridamole, the highest positive impact on its solubility was  
 324 observed for bile salts. Other significant effects for dipyridamole were a positive effect of  
 325 lecithin, a negative effect of cholesterol and the interaction between bile salts and cholesterol  
 326 was significant. Considering loperamide, bile salts showed a positive and cholesterol a negative  
 327 impact on solubility, respectively. For sulfasalazine solubility, a positive effect of cholesterol  
 328 was observed, followed by a significant interaction of bile salts and cholesterol and a positive  
 329 effect of the bile salt concentration.

330

331 **Table 2:** Significant effects and two-factor interactions in CED fasted state intestinal media.

<b>Main effects/ interactions</b>	<b>AZA</b>	<b>BUD</b>	<b>CEL</b>	<b>DIP</b>	<b>LOP</b>	<b>SSZ</b>
<b>BS</b>	+	+		+	+	+
<b>Lec</b>			+	+		
<b>Chol</b>			-	-	-	+
<b>BS/Lec</b>			-			
<b>BS/Chol</b>			+	+		+
<b>Lec/Chol</b>			-			

332 +: positive effect, -: negative effect, BS: bile salts, Lec: lecithin, Chol: cholesterol, AZA:  
 333 azathioprine, BUD: budesonide, CEL: celecoxib, DIP: dipyridamole, LOP: loperamide, SSZ:  
 334 sulfasalazine

335 For CED fed state intestinal media, the significant effects and two-factor interactions with an  
 336 impact on the drug solubility of all six drugs are shown in Table 3.

337 For azathioprine, the bile salt concentration had the highest positive impact on solubility,  
 338 followed by a positive impact of cholesterol. Considering budesonide solubility, all three main  
 339 effects were significant with the highest positive impact of bile salts, followed by a positive  
 340 impact of lecithin and a negative impact of cholesterol. The two-factor interactions bile

341 salts/cholesterol and lecithin/cholesterol were also significant but less influential compared to  
 342 the main effects. For celecoxib, the lecithin concentration had the highest positive impact on  
 343 its solubility, followed by a positive effect of the bile salt concentration. For dipyridamole, bile  
 344 salts and cholesterol had a positive impact on solubility. Additionally, the interaction of bile  
 345 salts and cholesterol was significant. Considering loperamide solubility, a negative impact of  
 346 cholesterol was observed and a smaller positive effect of the lecithin concentration. For  
 347 sulfasalazine, only the bile salt concentration had a positive impact on its solubility.

348

349 **Table 3:** Significant effects and two-factor interactions in CED fed state intestinal media.

<b>Main effects/ interactions</b>	<b>AZA</b>	<b>BUD</b>	<b>CEL</b>	<b>DIP</b>	<b>LOP</b>	<b>SSZ</b>
<b>BS</b>	+	+	+	+		+
<b>Lec</b>		+	+		+	
<b>Chol</b>	+	-		+	-	
<b>BS/Lec</b>						
<b>BS/Chol</b>		-		+		
<b>Lec/Chol</b>		-				

350 +: positive effect, -: negative effect, BS: bile salts, Lec: lecithin, Chol: cholesterol, AZA:  
 351 azathioprine, BUD: budesonide, CEL: celecoxib, DIP: dipyridamole, LOP: loperamide, SSZ:  
 352 sulfasalazine

353

#### 354 4.4. Drugs at risk of altered solubility in luminal fluids of CED patients

355 For hydrophilic compounds, only small differences in drug solubility are expected between  
 356 intestinal fluids of CED patients and healthy subjects as shown by the low impact of CED  
 357 alterations on azathioprine solubility.

358 A higher impact of CED on drug solubility is expected for neutral compounds with moderate  
359 to high lipophilicity. For these drugs, a higher luminal surfactant concentration (bile salts,  
360 lecithin) is expected to result in a higher solubility. It seems to be specific to each drug whether  
361 this increase in solubility is mainly driven by bile salts as in the case of budesonide or lecithin  
362 as in the case of celecoxib.

363 A lower risk of altered intestinal solubility in CED is expected for ionisable compounds with  
364 moderate to high lipophilicity since drug solubilisation was less impacted by CED changes  
365 integrated in the DoE compared to neutral lipophilic compounds.

366 The investigation of solubility differences for six compounds in simulated gastrointestinal  
367 fluids representing CED patients compared to healthy subjects provided an initial  
368 biopharmaceutics risk assessment in CED patients. To reach broader conclusions a bigger  
369 database including additional compounds is needed.

370 The present study considered differences in CED patients in terms of luminal concentrations  
371 of bile salts, lecithin and cholesterol. More studies are needed to characterise the luminal fluid  
372 composition of CED patients to investigate additional differences (e.g., luminal pH since a  
373 higher jejunal surface pH has been reported, luminal protein concentrations that are potentially  
374 increased by protein leakage through the intestinal membrane), which could not be adequately  
375 explored in this study (Kitis et al., 1982).

## 376 **5. Conclusion**

377 In the current study, biorelevant media developed to be representative of the small intestinal  
378 fluids in fasted and fed state of CED patients showed differences in media properties and drug  
379 solubilisation compared to biorelevant media developed based on healthy subjects. In terms of  
380 media properties, some CED media showed a higher surface tension in the fasted state  
381 compared to biorelevant media based on healthy subjects, whereas a lower surface tension was

382 observed in some CED media in the fed state. Differences in drug solubility in CED media  
383 compared to biorelevant media based on healthy subjects were mainly observed for moderately  
384 lipophilic compounds with a higher surfactant concentration (bile salts, lecithin) resulting in  
385 most cases in a higher drug solubility. The driving factor behind the increase in drug solubility  
386 (higher bile salt or lecithin concentration) seemed to be specific to each drug. Further solubility  
387 studies with additional compounds would increase the database for biopharmaceutics risk  
388 assessment in CED patients and additional studies investigating the composition of luminal  
389 contents in CED patients are needed.

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## 395 **7. Declaration of interest**

396 None.

397

398 **8. References**

- 399 Ali, H.S.M., York, P., Blagden, N., Soltanpour, S., Acree, W.E., Jouyban, A., 2010.  
400 Solubility of Budesonide, Hydrocortisone, and Prednisolone in Ethanol + Water Mixtures at  
401 298.2 K. *Journal of Chemical & Engineering Data* 55, 578-582.
- 402 Amidon, G.L., Lennernas, H., Shah, V.P., Crison, J.R., 1995. A theoretical basis for a  
403 biopharmaceutic drug classification: the correlation of in vitro drug product dissolution and in  
404 vivo bioavailability. *Pharm Res* 12, 413-420.
- 405 Baka, E., Comer, J.E., Takacs-Novak, K., 2008. Study of equilibrium solubility measurement  
406 by saturation shake-flask method using hydrochlorothiazide as model compound. *J Pharm*  
407 *Biomed Anal* 46, 335-341.
- 408 Betageri, G.V., Dipali, S.R., 1993. Partitioning and thermodynamics of dipyridamole in the  
409 n-octanol/buffer and liposome systems. *J Pharm Pharmacol* 45, 931-933.
- 410 Bharate, S.S., Kumar, V., Vishwakarma, R.A., 2016. Determining Partition Coefficient (Log  
411 P), Distribution Coefficient (Log D) and Ionization Constant (pKa) in Early Drug Discovery.  
412 *Comb Chem High Throughput Screen* 19, 461-469.
- 413 Bhatt, H., Naik, B., Dharamsi, A., 2014. Solubility Enhancement of Budesonide and  
414 Statistical Optimization of Coating Variables for Targeted Drug Delivery. *J Pharm (Cairo)*  
415 2014, 262194.
- 416 Butt, H., Graf, K., Kappl, M., 2004. Liquid Surfaces, in: Butt, H., Graf, K., Kappl, M. (Eds.),  
417 *Physics and Chemistry of Interfaces*. WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim,  
418 Germany, pp. 4-25.
- 419 Corey, E.J., Fossel, E.T., 2016. Transdermal formulations of fluticasone (US 2016/0081915).  
420 Google Patents.
- 421 Dickson, C.J., Hornak, V., Pearlstein, R.A., Duca, J.S., 2017. Structure-Kinetic Relationships  
422 of Passive Membrane Permeation from Multiscale Modeling. *J Am Chem Soc* 139, 442-452.

423 Effinger, A., O'Driscoll, C.M., McAllister, M., Fotaki, N., 2019. Impact of gastrointestinal  
424 disease states on oral drug absorption - implications for formulation design - a PEARRL  
425 review. *J Pharm Pharmacol* 71, 674-698.

426 Effinger, A., O'Driscoll, C.M., McAllister, M., Fotaki, N., 2020. Gastrointestinal diseases and  
427 their impact on drug solubility. Part I. Crohn's disease. *Eur J Pharm Sci* (in press).

428 G.D. Searle LLC Division of Pfizer Inc, 2019. CELEBREX- celecoxib capsule prescribing  
429 information, New York, NY, US. Available from:  
430 <http://labeling.pfizer.com/ShowLabeling.aspx?id=793> [accessed 09.06.2019].

431 Galia, E., Nicolaidis, E., Horter, D., Lobenberg, R., Reppas, C., Dressman, J.B., 1998.  
432 Evaluation of various dissolution media for predicting in vivo performance of class I and II  
433 drugs. *Pharm Res* 15, 698-705.

434 Gottlieb, K., Dawson, J., Hussain, F., Murray, J.A., 2015. Development of drugs for celiac  
435 disease: review of endpoints for Phase 2 and 3 trials. *Gastroenterol Rep (Oxf)* 3, 91-102.

436 Graham, G.G., Pile, K.D., 2015. Sulfasalazine and Related Drugs, in: Parnham, M. (Ed.),  
437 Compendium of Inflammatory Diseases. Springer, Basel, Switzerland, pp. 1-5.

438 Hansch, C., Leo, A., Hoekman, D., 1995. Exploring QSAR: Hydrophobic, Electronic, and  
439 Steric Constants. American Chemical Society, Washington, DC, US.

440 Hopfinger, A.J., Esposito, E.X., Llinas, A., Glen, R.C., Goodman, J.M., 2009. Findings of the  
441 challenge to predict aqueous solubility. *J Chem Inf Model* 49, 1-5.

442 Jantratid, E., Janssen, N., Reppas, C., Dressman, J.B., 2008. Dissolution media simulating  
443 conditions in the proximal human gastrointestinal tract: an update. *Pharm Res* 25, 1663-1676.

444 Khoshakhlagh, P., Johnson, R., Langguth, P., Nawroth, T., Schmueser, L., Hellmann, N.,  
445 Decker, H., Szekely, N.K., 2015. Fasted-State Simulated Intestinal Fluid "FaSSIF-C", a  
446 Cholesterol Containing Intestinal Model Medium for In Vitro Drug Delivery Development. *J*  
447 *Pharm Sci* 104, 2213-2224.

448 Kitis, G., Lucas, M.L., Bishop, H., Sargent, A., Schneider, R.E., Blair, J.A., Allan, R.N.,  
449 1982. Altered jejunal surface pH in coeliac disease: its effect on propranolol and folic acid  
450 absorption. *Clin Sci (Lond)* 63, 373-380.

451 Koehler, P., Wieser, H., Konitzer, K., 2014. Chapter 1 Celiac Disease—A Complex Disorder,  
452 in: Koehler, P., Wieser, H., Konitzer, K. (Eds.), *Celiac Disease and Gluten*. Academic Press,  
453 London, UK, pp. 1-96.

454 Leffler, D.A., Green, P.H., Fasano, A., 2015. Extraintestinal manifestations of coeliac  
455 disease. *Nat Rev Gastroenterol Hepatol* 12, 561-571.

456 Lindenberg, M., Kopp, S., Dressman, J.B., 2004. Classification of orally administered drugs  
457 on the World Health Organization Model list of Essential Medicines according to the  
458 biopharmaceutics classification system. *Eur J Pharm Biopharm* 58, 265-278.

459 Llinas, A., Glen, R.C., Goodman, J.M., 2008. Solubility challenge: can you predict  
460 solubilities of 32 molecules using a database of 100 reliable measurements? *J Chem Inf*  
461 *Model* 48, 1289-1303.

462 Low-Bear, T.S., Heaton, K.W., Pomare, E.W., Read, A.E., 1973. The effect of coeliac  
463 disease upon bile salts. *Gut* 14, 204.

464 Manallack, D.T., 2007. *The pK(a) Distribution of Drugs: Application to Drug Discovery*.  
465 *Perspect Medicin Chem* 1, 25-38.

466 Markopoulos, C., Andreas, C.J., Vertzoni, M., Dressman, J., Reppas, C., 2015. In-vitro  
467 simulation of luminal conditions for evaluation of performance of oral drug products:  
468 Choosing the appropriate test media. *Eur J Pharm Biopharm* 93, 173-182.

469 Mitra, A.K., Narurkar, M.M., 1987. Kinetics of azathioprine degradation in aqueous solution.  
470 *Int J Pharm* 35, 165-171.

471 Oberhuber, G., Granditsch, G., Vogelsang, H., 1999. The histopathology of coeliac disease:  
472 time for a standardized report scheme for pathologists. *Eur J Gastroenterol Hepatol* 11, 1185-  
473 1194.

474 Paulson, S.K., Vaughn, M.B., Jessen, S.M., Lawal, Y., Gresk, C.J., Yan, B., Maziasz, T.J.,  
475 Cook, C.S., Karim, A., 2001. Pharmacokinetics of celecoxib after oral administration in dogs  
476 and humans: effect of food and site of absorption. *J Pharmacol Exp Ther* 297, 638-645.

477 Pedersen, A.K., 1979. Specific determination of dipyridamole in serum by high-performance  
478 liquid chromatography. *J Chromatogr* 162, 98-103.

479 Rabbie, S.C., Flanagan, T., Martin, P.D., Basit, A.W., 2015. Inter-subject variability in  
480 intestinal drug solubility. *Int J Pharm* 485, 229-234.

481 Riethorst, D., Mols, R., Duchateau, G., Tack, J., Brouwers, J., Augustijns, P., 2016.  
482 Characterization of Human Duodenal Fluids in Fasted and Fed State Conditions. *J Pharm Sci*  
483 105, 673-681.

484 Shalaeva, M., Kenseth, J., Lombardo, F., Bastin, A., 2008. Measurement of dissociation  
485 constants (pKa values) of organic compounds by multiplexed capillary electrophoresis using  
486 aqueous and cosolvent buffers. *J Pharm Sci* 97, 2581-2606.

487 Shono, Y., Jantratid, E., Janssen, N., Kesisoglou, F., Mao, Y., Vertzoni, M., Reppas, C.,  
488 Dressman, J.B., 2009. Prediction of food effects on the absorption of celecoxib based on  
489 biorelevant dissolution testing coupled with physiologically based pharmacokinetic  
490 modeling. *Eur J Pharm Biopharm* 73, 107-114.

491 Soderlind, E., Karlsson, E., Carlsson, A., Kong, R., Lenz, A., Lindborg, S., Sheng, J.J., 2010.  
492 Simulating fasted human intestinal fluids: understanding the roles of lecithin and bile acids.  
493 *Mol Pharm* 7, 1498-1507.

494 Tran, T.H., Smith, C., Mangione, R.A., 2013. Drug absorption in celiac disease. *Am J Health*  
495 *Syst Pharm* 70, 2199-2206.



496 Turner, G.D., Dunne, M.R., Ryan, A.W., 2015. Celiac Disease: Background and Historical  
497 Context, in: Ryan, A.W. (Ed.), Celiac Disease: Methods and Protocols. Springer, New York,  
498 NY, US, pp. 3-14.

499 Vertzoni, M., Diakidou, A., Chatziliias, M., Soderlind, E., Abrahamsson, B., Dressman, J.B.,  
500 Reppas, C., 2010. Biorelevant media to simulate fluids in the ascending colon of humans and  
501 their usefulness in predicting intracolonic drug solubility. *Pharm Res* 27, 2187-2196.

502 Vertzoni, M., Dressman, J., Butler, J., Hempenstall, J., Reppas, C., 2005. Simulation of  
503 fasting gastric conditions and its importance for the in vivo dissolution of lipophilic  
504 compounds. *Eur J Pharm Biopharm* 60, 413-417.

505 Vuoristo, M., Miettinen, T.A., 1985. Increased Biliary Lipid Secretion in Celiac Disease.  
506 *Gastroenterology* 88, 134-142.

507 Xie, X., Cardot, J.M., Garrait, G., They, V., El-Hajji, M., Beyssac, E., 2014. Micelle  
508 dynamic simulation and physicochemical characterization of biorelevant media to reflect  
509 gastrointestinal environment in fasted and fed states. *Eur J Pharm Biopharm* 88, 565-573.

510 Zaki, N.M., Artursson, P., Bergstrom, C.A., 2010. A modified physiological BCS for  
511 prediction of intestinal absorption in drug discovery. *Mol Pharm* 7, 1478-1487.

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513

514 **Figure Legends**

515 Figure 1: Design of Experiments for the development of Celiac disease intestinal biorelevant  
516 media (\*value observed in human intestinal fluids (Riethorst et al., 2016)).

517 Figure 2: Surface tension (blue, left y-axis) and osmolality (rose, right y-axis) of Celiac  
518 disease biorelevant media according to the Design of Experiment (green: high level, red: low  
519 level, white: healthy) and healthy media (H).

520 Figure 3: Dynamic viscosity of biorelevant media based on Celiac disease patients and the  
521 corresponding biorelevant media based on healthy subjects (H) at different shear stress (0.06  
522 Pa: blue, 0.08 Pa: red, 0.15 Pa: black) according to the Design of Experiments (green: high  
523 level, red: low level, white: healthy).

524 Figure 4: % Relative effect (RE) on the solubility of neutral (at pH 5.8-6.5) investigated  
525 drugs in Celiac disease intestinal biorelevant media compared to the corresponding media  
526 based on healthy subjects according to Design of Experiments (red: low concentration of  
527 cholesterol, blue: high concentration of cholesterol, grey point: medium based on healthy  
528 subjects).

529 Figure 5: % Relative effect on the solubility of weak acids and bases in Celiac disease  
530 intestinal biorelevant media compared to the corresponding media based on healthy subjects  
531 according to Design of Experiments (red: low concentration of cholesterol, blue: high  
532 concentration of cholesterol, grey point: medium based on healthy subjects).

533

534

535

536

	<b>Celiac disease</b>			
<b>Prandial state</b>	<b>Fasted state</b>		<b>Fed state</b>	
<b>Compartment</b>	<b>intestine</b>		<b>intestine</b>	
<b>Level</b>	<b>low</b>	<b>high</b>	<b>low</b>	<b>high</b>
<b>Bile salts [mM]</b>	3.0	5.1	10.0	17.0
<b>Lecithin [mM]</b>	0.2	0.6	2.0	6.0
<b>Cholesterol [mM]</b>	0.08*	0.16	0.57*	1.14

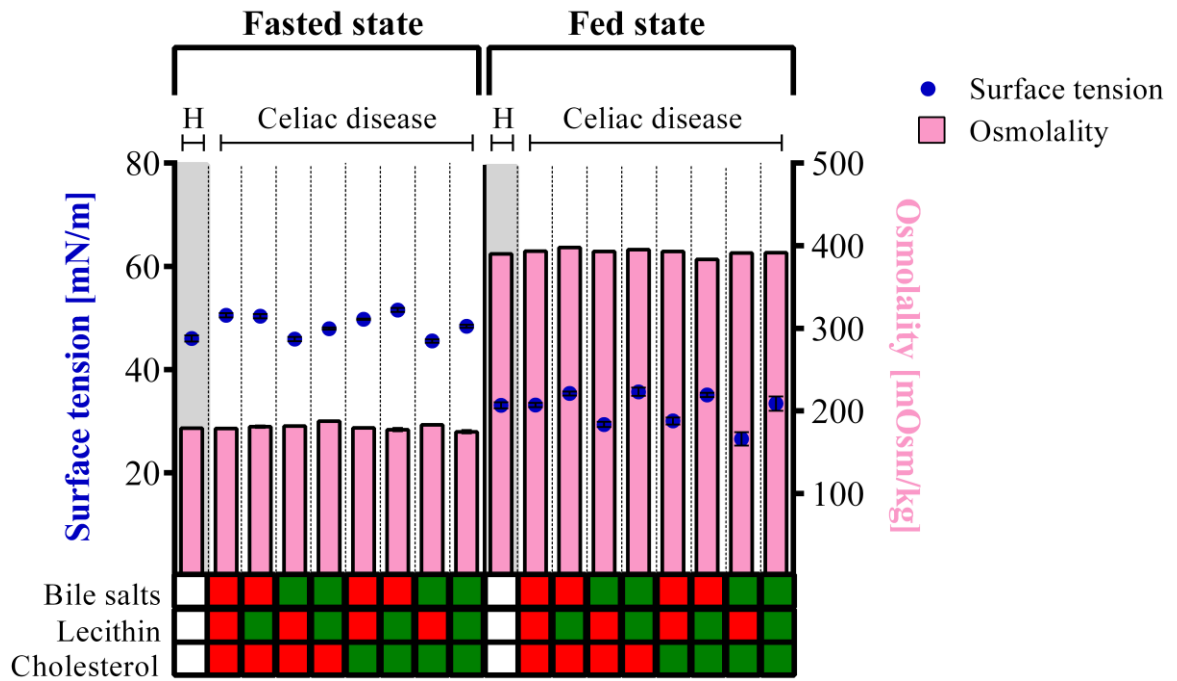
	increase
	value in healthy biorelevant media

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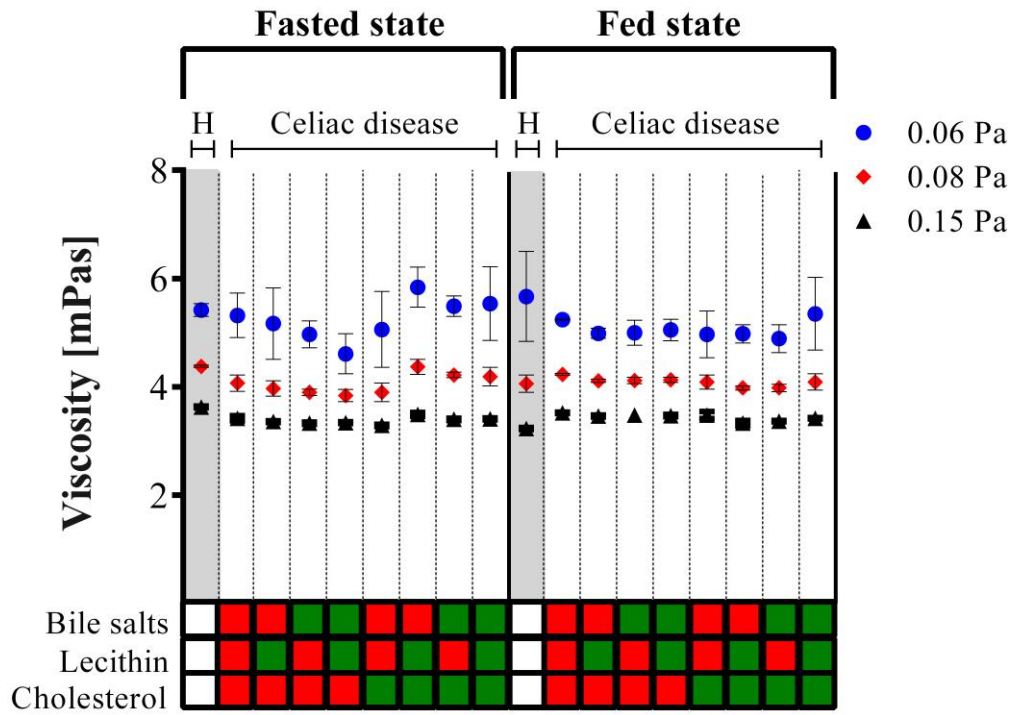


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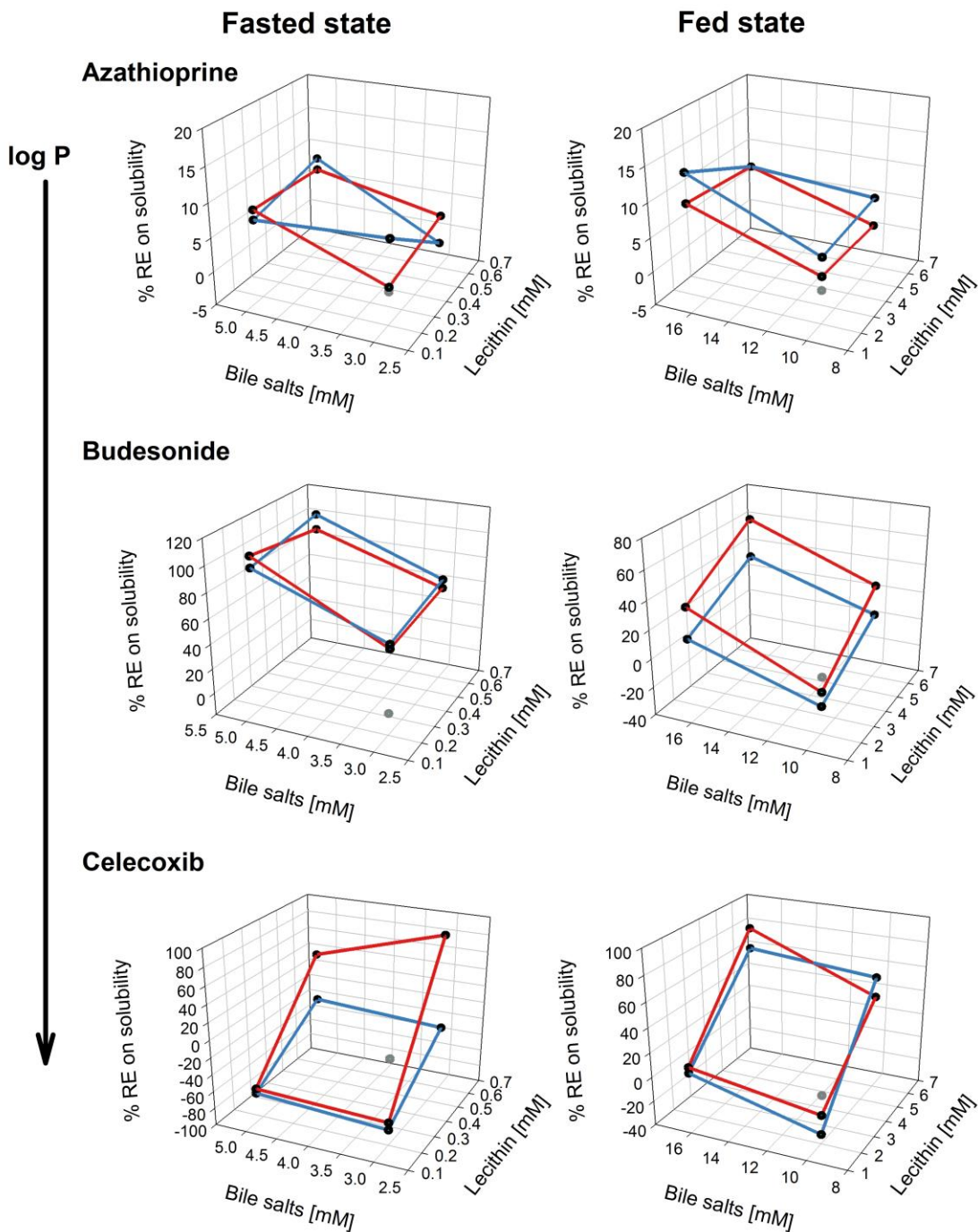
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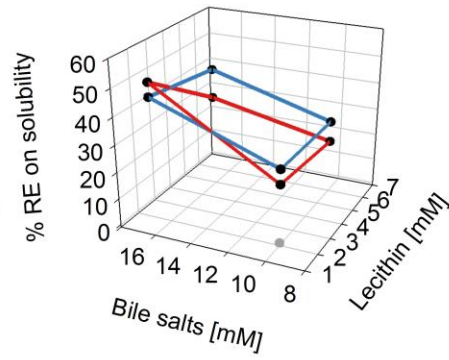
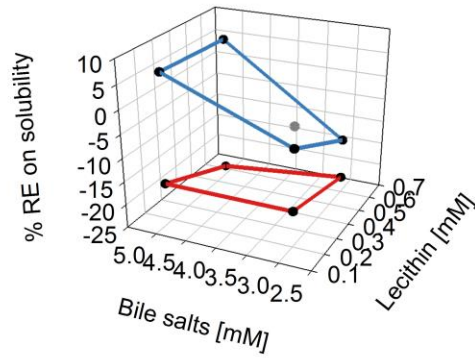


**Fasted state**

**Fed state**

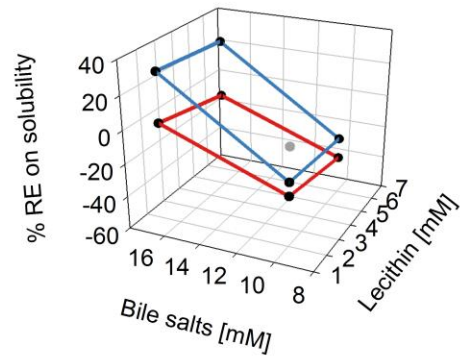
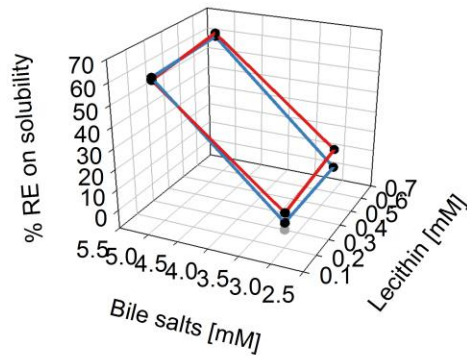
**Weak acid**

**Sulfasalazine**  
pKa 2.2



**Weak bases**

**Dipyridamole**  
pKa 6.4



**Loperamide-HCl**  
pKa 8.6

