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Reduction of the Pectoral Spine and Girdle in Domesticated Channel Catfish is Likely Caused by Changes in Selection Pressure

Michael L. Fine

Virginia Commonwealth University, mfine@vcu.edu

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1 Reduction of the pectoral spine and girdle in domesticated Channel Catfish is likely caused by
2 changes in selection pressure

3

4 Michael L. Fine¹, Shweta Lahiri¹, Amanda D. H. Sullivan¹, Mark Mayo¹, Scott H. Newton² and
5 Edward N. Sismour²

6

7 ¹Department of Biology, Virginia Commonwealth University, Richmond, VA 23284-2012

8 ²Agricultural Research Station, Virginia State University, Petersburg, VA 23806

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10 Corresponding author: mfine@vcu.edu

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12

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21 Locked pectoral spines of the Channel Catfish *Ictalurus punctatus* more than double the fish's
22 width and complicate ingestion by gape-limited predators. The spine mates with the pectoral
23 girdle, a robust structure that anchors the spine. This study demonstrates that both spine and
24 girdle exhibit negative allometric growth and that pectoral spines and girdles are lighter in
25 domesticated than in wild Channel Catfish. This finding could be explained by changes in
26 selection pressure for spine growth during domestication or by an epigenetic effect in which
27 exposure to predators in wild fish stimulates pectoral growth. We tested the epigenetic
28 hypothesis by exposing domesticated Channel Catfish fingerlings to Largemouth Bass
29 *Micropterus salmoides* predators for 13 weeks. Spines and girdles grow isometrically in the
30 fingerlings, and regression analysis indicates no difference in proportional pectoral growth
31 between control and predator-exposed fish. Therefore a change in selection pressure likely
32 accounts for smaller pectoral growth in domesticated Channel Catfish. Decreasing spine growth
33 in older fish suggests anti-predator functions are most important in smaller fish. Additionally,
34 growth of the appendicular and axial skeleton is controlled differentially, and mechanical
35 properties of the spine and not just its length are an important component of this defensive
36 adaptation.

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43 Spines have been examined extensively as anti-predator adaptations in a number of aquatic
44 species including *Daphnia* (Tollrian and Dodson 1999) and the Three-Spined Stickleback
45 *Gasterosteus aculeatus* (Huntingford and Coyle 2007). A major adaptation of catfishes, one of
46 the most successful teleost groups, is a hypertrophied first pectoral spine that can be bound and
47 locked (Fine et al. 1997). Binding involves active muscular force that engages friction-locking
48 surfaces between the spine base and the pectoral girdle. Locking requires full abduction of the
49 spine, which traps a spine process within a recess in the coracoid preventing further movement
50 (abduction or adduction). In addition the spine can be rubbed against the cleithrum to produce
51 stridulatory sounds that can serve in distress, agonistic and courtship capacities (Fine and Ladich
52 2003; Kaatz et al. 2010; Parmentier et al. 2010). These specialized functions are mediated by
53 rearranged pectoral muscles (Diogo et al. 2001; Miano et al. 2013) and derived processes
54 (dorsal, anterior and ventral) on the spine base (Hubbs and Hibbard 1951) that mate with
55 complimentary structures on the pectoral girdle (Diogo et al. 2001; Fine et al. 1997). The
56 pectoral girdle (a fused cleithrum, coracoid and scapula) is a robust structure (Fine et al. 1997),
57 providing rigid support for the spine (Schaefer 1984). When locked in a fully-abducted position,
58 the stout pectoral spines of Channel Catfish, *Ictalurus punctatus* Rafinesque more than double
59 the fish's width (Sismour et al. 2013) and impede ingestion by gape-limited predators. Intact
60 Channel Catfish fingerlings are three times less likely to be eaten by Largemouth bass,
61 *Micropterus salmoides* Lacepede than comparably-sized individuals with clipped spines (Bosher
62 et al. 2006), and Largemouth bass consume fewer Channel Catfish than Goldfish, *Carassius*
63 *auratus* L. and Bluegills, *Lepomis macrochirus* Rafinesque in choice experiments, supporting
64 Forbes' dangerous prey hypothesis (Sismour et al. 2013).

65 Channel Catfish spines collected beneath eagle nests (Duvall 2007) exhibited subtle
66 morphological differences from those of domesticated stocks (Fine et al. 1997) calling into
67 question the effects of domestication on spine development. Domestication in Channel Catfishes
68 has selected for more rapid growth rate than in natural populations and a reduction of genetic
69 diversity although introduction of new fish has likely minimized inbreeding (Hallerman et al.
70 1986; Smitherman and Dunham 2003).

71 We compared pectoral spines of Channel Catfish from the James River, Virginia with
72 domesticated fish purchased from an Arkansas stock. Smaller spines found in domesticated fish
73 could result from changes in selection pressure over multiple generations. Alternately, exposure
74 to predators in wild fish could induce an epigenetic effect that turns on genes that induce spine
75 growth (phenotypic plasticity). We tested the epigenetic hypothesis by exposing domesticated
76 Channel Catfish fingerlings to Largemouth Bass behind a mesh barrier for several months.
77 Previous work with these individuals demonstrated that fingerlings exposed to Largemouth Bass
78 move and eat less than controls and grow more slowly (Fine et al. 2011). The current study
79 demonstrates that although control fingerlings were longer and heavier than experimental fish,
80 the proportional pectoral spine and girdle weight are similar in control fish and experimental fish
81 exposed to Largemouth Bass, indicating that predators did not induce increased growth of the
82 pectoral skeleton.

83

84 *Methods*

85 Wild Channel Catfish *I. punctatus* were caught by electroshocking (Virginia Department of
86 Game and Inland Fisheries and Virginia Commonwealth University VADGIF permit number
87 0444631 and IACUC AD20216) in the James River, Virginia. Domesticated fish were obtained

88 from the aquaculture facility of Virginia State University from stocks purchased from Arkansas.
89 Fish were weighed in grams and measured for total length in mm (TL). Frozen fish were thawed
90 and boiled briefly to clear the skeletons. After drying, pectoral spines were measured for length
91 with digital calipers, and spines and girdles were weighed in milligrams. Since spine tips often
92 break, the longer and heavier spine was used for analysis. Measurements were linearized by log-
93 log transformation, regressed against TL or weight, and regressions of domesticated and wild
94 fish were compared by analysis of covariance. Adjusted means for domesticated and wild fish
95 were calculated from regressions for 400 mm TL and 1,000 g Channel Catfish. Data were plotted
96 on a linear scale to illustrate growth rate.

97 Experimental protocols for non-consumptive effects of Largemouth Bass *Micropterus*
98 *salmoides* predators are provided in Fine et al. (2011) and will be summarized briefly. Juvenile
99 Channel Catfish were measured for total length TL in mm and weighed to 0.1 g. Largemouth
100 Bass (29 to 43 cm TL) were obtained by hook and line from a small impoundment at the Rice
101 Center of Virginia Commonwealth University (Charles City County, Virginia). Fish were
102 maintained in eight 300 L fiberglass tanks at 23 C under a 14:10 LD cycle. A black polyethylene
103 plastic mesh barrier (6.5 mm square openings) was erected across the center of each tank
104 separating them into halves, and a clay flowerpot was added to the catfish side for shelter. Ten
105 Channel Catfish were acclimated in the right half of each tank for one week, after which a
106 Largemouth Bass was added to four of the tanks. Largemouth bass are a generalized fish
107 predator that commonly consumes Channel Catfish in rivers and ponds (Sismour et al. 2013).
108 The other four tanks with no bass served as controls. There was no difference in mean size of
109 control and experimental Channel Catfish at the beginning of the experiment (Fine et al. 2011).
110 Largemouth bass were fed at least twice a week with a dead Channel Catfish from the same

111 stock as the experimental catfish. Catfish were fed to satiation multiple days per week using a
112 standard 32% protein floating-pellet catfish ration. The experiment was conducted for 13 weeks.

113 Based on data from wild and domesticated fish and our hypothesis that predators would
114 increase spine growth, we compared means from control and experimental tanks with a one-
115 tailed T-test. TL, fish weight, pectoral spine length and weight and girdle weight from the four
116 control and four experimental tanks were averaged so that each tank was treated as a unit (N = 4
117 per treatment), and $p < 0.05$ was considered significant. Because of differences in size of control
118 and experimental fish at the termination of the experiment (see results), tank means were not
119 sufficient to determine if relative spine and girdle growth are affected by predators. Growth was
120 therefore evaluated by linear regressions of spine length against fish TL and spine and girdle
121 weight against fish weight. Regressions were compared by analysis of covariance. Because of
122 possible tank effects, we first compared regressions across tanks within control and within
123 experimental treatments. Slopes and intercepts were not significant for spine length, spine weight
124 or girdle weight within either treatment. Since there were no tank effects, data from individual
125 tanks were combined, and relative growth for spines and girdles was evaluated and regressions
126 were compared between control and experimental tanks using individual fish.

127

128 *Results*

129 **PECTORAL COMPARISONS BETWEEN WILD AND DOMESTICATED CHANNEL** 130 **CATFISH**

131 Spine length, spine weight and girdle weight increased continuously with fish size although at a
132 decelerating rate in catfish that ranged from 87 to 562 mm TL (Fig. 1). Adjusted means for
133 domesticated and wild fish respectively were 40.4 and 49.9 mm for spine length, 0.60 and 1.04 g

134 for spine weight and 5.42 and 7.50 g for girdle weight. Wild catfish had longer and heavier
135 spines and heavier girdles than domesticated individuals. Slopes for spine lengths did not differ
136 between domesticated and wild fish ($F_{1,91}=1.23$, $p=0.2669$), but elevations were greater in wild
137 fish ($F_{1,92}=219.6$, $p<0.0001$). Slope for spine weight against fish weight were so much greater in
138 wild fish ($F_{1,91}=10.7$, $p=0.0015$) that intercepts could not be tested. Slopes for girdle weight were
139 similarly greater in wild fish ($F_{1,86}=8.10$, $p=0.0055$).

140

141 **PREDATOR EXPOSURE EXPERIMENT**

142 Mean TL ranged from 16.40 to 17.59 mm for control and 14.38 to 16.06 for experimental tanks
143 and weights from 37.24 to 45.35 and 21.20 to 31.65 respectively. Control Channel Catfish were
144 longer ($T_6=3.229$, $p=0.0179$) and heavier ($T_6=4.384$, $p=0.0046$) than experimental fish at the
145 termination of the experiment (Fig. 2a, b) indicating that a predator across the barrier retarded
146 growth (Fine et al. 2011). The control fish had longer ($T_6=2.420$, $p=0.0259$) and heavier
147 ($T_6=2.078$, $p=0.0415$) pectoral spines (Fig. 2c, d) as would be expected of larger individuals, but
148 the difference in girdle weight did not reach significance ($T_6=1.808$, $p=0.1206$). In this size
149 range spines and girdles of both control and experimental catfish grew linearly (Table 1; Fig 3).
150 The r^2 values for linear regressions of control fish ranged from 0.75 to 0.93 for spine length
151 against TL, spine weight against weight and girdle weight against weight except for control spine
152 length against TL with an r^2 of 0.46 (Table 1). Data points for control and experimental fish co-
153 scattered, and analysis of covariance indicated no significant difference between proportional
154 pectoral spine length, weight or girdle weight of control fish and fish exposed to Largemouth
155 Bass (Table 1, Fig 3). The largest individuals were all in control tanks, and these individuals
156 were likely responsible for slight though not significant differences in intercepts of spine and

157 girdle weight regressions. Similar experimental and control regressions indicate that exposure to
158 predators did not increase pectoral growth.

159 *Discussion*

160 The most parsimonious hypothesis to account for the decreased pectoral spine and girdle weight
161 we observed in domesticated fish is changes in selection pressure for this defensive adaptation
162 rather than an epigenetic effect caused by exposure to predators. Domestication of Channel
163 Catfish has involved selection for fast growth in ponds without fish predation, and the pectoral
164 apparatus was not considered when choosing breeding stock (Dunham and Smitherman 1982;
165 Hallerman et al. 1986; Smitherman and Dunham 1993). However, breeding for fast growth
166 could have inadvertently selected for fish with smaller pectoral spines and girdles. We note that
167 spines are not a concern in large commercial ventures since fish are rarely handled individually
168 until processing when they are stunned with a weak electric current before decapitation
169 (Marshall 2004). Additionally smaller spines in aquaculture fish are still an effective adaptation
170 and deter predation by Largemouth Bass (Bosher et al. 2006 Sismour et al. 2013).

171 Relaxation of selection pressure during domestication and inadvertent selection for
172 smaller spines are complimentary hypotheses to explain spine reduction although inadvertent
173 selection might be unlikely in the presence of fish predators. Unraveling the course of selection
174 on spine size will be complex since contemporary aquaculture stocks do not have a single
175 population of origin (Smitherman and Dunham 1983). Therefore, comparisons between cultured
176 stocks with a single wild population of origin would not be possible. Experimentation to confirm
177 either hypothesis could involve developing a cultivated stock from a wild population and
178 following spine development over several generations. Such an endeavor would require at least
179 a decade given that Channel Catfish in Virginia require 4-6 years to mature (Hubert 2000). It

180 should also be possible to compare spine size in populations of domesticated fish with different
181 growth rates.

182 Selection pressure on spine length has been demonstrated in populations of *Gasterosteus*
183 *aculeatus* under different predation regimes: populations dominated by fish predators have long
184 spines whereas populations with invertebrate predators have shorter ones (Huntingford and
185 Coyle 2007). Similarly, in Nine-Spined Sticklebacks, a common garden experiment
186 demonstrated that predator density and food availability did not affect body shape or armor,
187 indicating that anti-predator traits are constitutive rather than inducible (Välämäki et al. 2012).
188 The classic example of a predator-induced defense in fishes is an increase in body depth in
189 Crucian carp, *Carassius carassius* (Brönmark and Miner 1992). Similarly, the presence of
190 predators caused morphological changes in perch and roach (Eklöv and Jönsson 2007) and in
191 pumpkinseed sunfish (Januszkiewicz and Robinson 2007).

192 The 13-week experiment reported here resulted in significant growth differences but not in
193 changes in pectoral development when normalized to fish size. Our results do not rule out the
194 possibility of an epigenetic effect on smaller or wild individuals. The negative allometry of
195 pectoral girdle growth suggests that the axial and appendicular skeletons are controlled by
196 different genetic mechanisms. One would expect axial and appendicular skeletons to grow
197 proportionately in most fishes in order to provide normal control of fine movement. Our finding
198 therefore supports the importance of the fused pectoral girdle (Schaefer 1984) as a major
199 component of the anti-predator adaptation provided by the pectoral spines. Furthermore, linear
200 pectoral spine growth in young but not older fish emphasizes the importance of the adaptation in
201 small fish that face increased predation risk. We caution that growth rate in domesticated and
202 wild fish differ.

203 The pectoral spine of Channel Catfish is an enlarged flattened fin ray that tapers toward the
204 tip (Fine et al. 1997). The enlarged horizontal profile will increase resistance in the horizontal
205 plane, which opposes forces caused by passage through a predator's mouth or underwater
206 obstructions. Spine and girdle growth is isometric in small juveniles used in this study, and the
207 negative allometry in spine length and weight is due to decreasing growth in larger individuals
208 who would be less vulnerable to predation. Additionally breakage of spine tips occurs commonly
209 in both domestic and wild individuals. Decreasing girdle weight supports a growth effect and not
210 just breakage. The increase in spine weight would be determined by linear dimensions,
211 particularly near the wider spine base, and spine weight per millimeter of spine length increases
212 exponentially with TL (Duvall 2007). Wild Channel Catfish have wider spines (Duvall 2007),
213 which increases the moment of inertia (a greater cross sectional area further from the midline)
214 and therefore breakage resistance of the structure. The larger mass of the pectoral girdle in wild
215 fish is striking since it is a major component of the fish's girth. Finally, the increased spine mass
216 suggests that in addition to spine length (Tollrian and Dodson 1999; Huntingford and Coyle
217 2007) other dimensions that contribute to the material properties of a defensive spine are
218 important to its function.

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225

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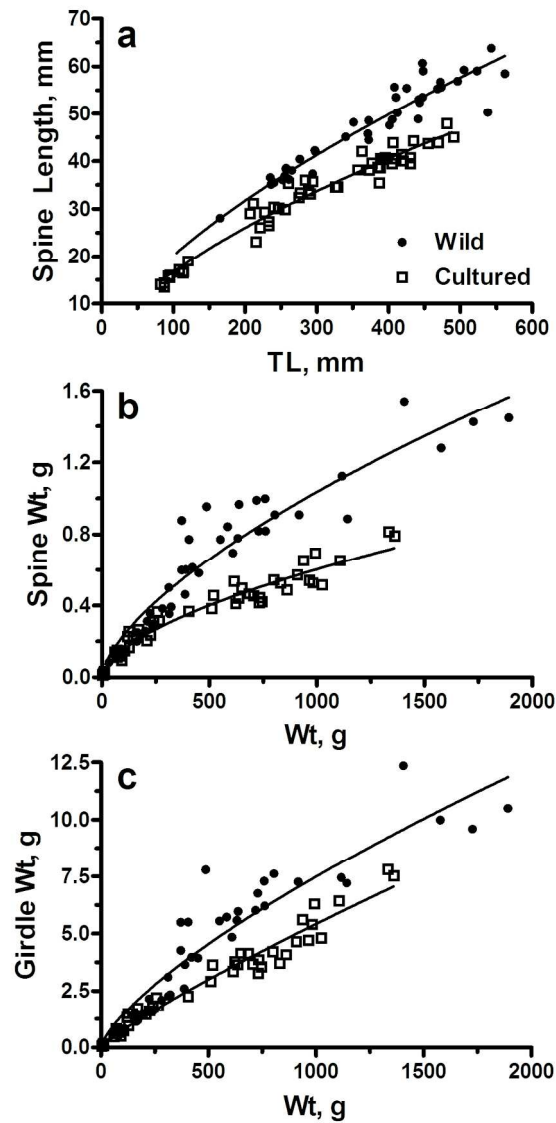
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Figure Legends

Figure 1. Relationship of spine length to total length (a), spine weight to fish weight (b) and pectoral girdle weight to fish weight (c) for wild and domesticated catfish. Equations for James River fish: $\text{Log Spine L} = 0.001654 + 0.6518 \text{ Log TL}$, $r^2 = 0.92$; $\text{Log Spine Wt} = -1.928 + 0.6480 \text{ Log Wt}$, $r^2 = 0.89$; $\text{Log Girdle Wt} = -1.284 + 0.7197 \text{ Log Wt}$, $r^2 = 0.87$. Equations for domesticated fish: $\text{Log Spine L} = -0.0561 + 0.639 \text{ Log TL}$, $r^2 = 0.96$; $\text{Log Spine Wt} = -1.984 + 0.5885 \text{ Log Wt}$, $r^2 = 0.96$; $\text{Log Girdle Wt} = -1.833 + 0.8558 \text{ Log Wt}$, $r^2 = 0.96$.

Figure 2. Mean \pm SE total length, fish weight, spine length, spine weight, spine length for control and experimental Channel Catfish. * $p < 0.05$, ** $p < 0.01$.

Figure 3. Relationship of spine length to total length (a), spine weight to total weight (b) and girdle weight to total weight (c) in control and experimental Channel Catfish.



Relationship of spine length to total length (A), spine weight to fish weight (B) and pectoral girdle weight to fish weight (C) for wild and domesticated catfish. Equations for James River fish: $\text{Log Spine L} = 0.001654 + 0.6518 \text{ Log TL}$, $r^2 = 0.92$; $\text{Log Spine Wt} = -1.928 + 0.6480 \text{ Log Wt}$, $r^2 = 0.89$; $\text{Log Girdle Wt} = -1.284 + 0.7197 \text{ Log Wt}$, $r^2 = 0.87$. Equations for domesticated fish: $\text{Log Spine L} = -0.0561 + 0.639 \text{ Log TL}$, $r^2 = 0.96$; $\text{Log Spine Wt} = -1.984 + 0.5885 \text{ Log Wt}$, $r^2 = 0.96$; $\text{Log Girdle Wt} = -1.833 + 0.8558 \text{ Log Wt}$, $r^2 = 0.96$.
120x223mm (300 x 300 DPI)

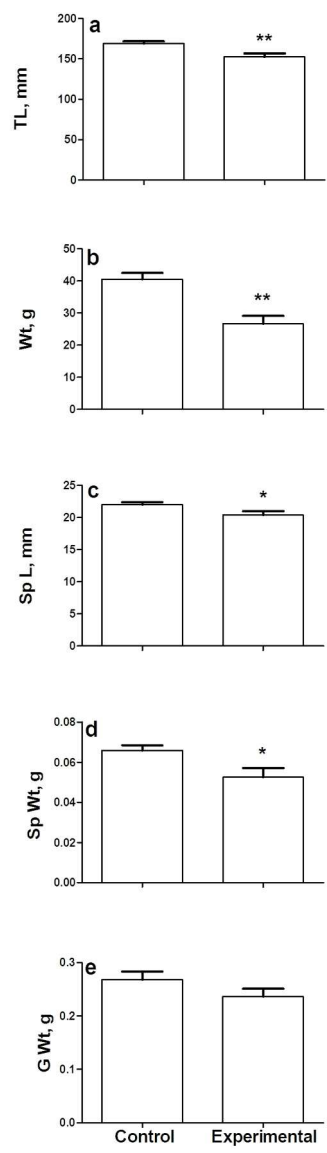


Figure 2. Mean ± SE total length, fish weight, spine length, spine weight, spine length for control and experimental Channel catfish. * p < 0.05, **p < 0.01. 72x216mm (300 x 300 DPI)

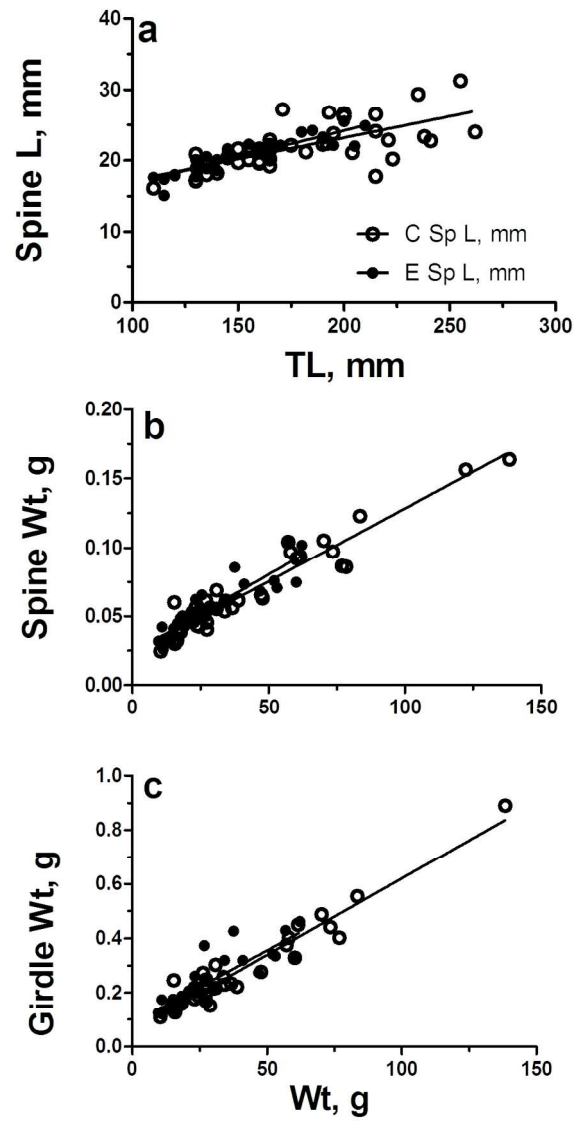


Figure 3. Relationship of spine length to total length (a), spine weight to total weight (b) and girdle weight to total weight (c) in control and experimental Channel catfish.
113x201mm (300 x 300 DPI)

Table 1 Regression equation and coefficient of determination for spine length against total length, spine weight against fish weight and girdle weight against fish weight, analysis of covariance for slopes and intercepts and adjusted means calculated for 160 mm TL and 50 g weight Channel catfish

Regression Equations	Slope			Intercept		Adjusted Mean
	r^2	F	p	F	p	
Ctrl Spine L=0.0601 TL+11.18	0.46	$F_{1,67}=0.97$	0.32	$F_{1,68}=0.68$	0.41	20.80 mm
Exp Spine L=0.0752 TL+9.168	0.79					21.20 mm
Ctrl Spine Wt=0.0010 Wt+0.022	0.92	$F_{1,67}=0.48$	0.4	$F_{1,68}=2.37$	0.13	0.072 g
Exp Spine Wt=0.0011 Wt+0.023	0.83					0.078 g
Ctrl Girdle Wt= 0.0056 Wt+0.058	0.93	$F_{1,65}=0.14$	0.7	$F_{1,66}=3.57$	0.06	0.338 g
Exp Girdle Wt=0.0054 Wt+0.086	0.75					0.356 g