

Evidence of positive selection towards Zebuine haplotypes in the BoLA region of Brangus cattle

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The Brangus breed was developed to combine the superior characteristics of both of its founder breeds, Angus and Brahman. It combines the high adaptability to tropical and subtropical environments, disease resistance, and overall hardiness of Zebu cattle with the reproductive potential and carcass quality of Angus. It is known that the major histocompatibility complex (MHC, also known as bovine leucocyte antigen: BoLA), located on chromosome 23, encodes several genes involved in the adaptive immune response and may be responsible for adaptation to harsh environments. The objective of this work was to evaluate whether the local breed ancestry percentages in the BoLA locus of a Brangus population diverged from the estimated genome-wide proportions and to identify signatures of positive selection in this genomic region. For this, 167 animals (100 Brangus, 45 Angus and 22 Brahman) were genotyped using a high-density single nucleotide polymorphism array. The local ancestry analysis showed that more than half of the haplotypes (55.0%) shared a Brahman origin. This value was significantly different from the global genome-wide proportion estimated by cluster analysis (34.7% Brahman), and the proportion expected by pedigree (37.5% Brahman). The analysis of selection signatures by genetic differentiation (F_{st}) and extended haplotype homozygosity-based methods (iHS and Rsb) revealed 10 and seven candidate regions, respectively. The analysis of the genes located within these candidate regions showed mainly genes involved in immune response-related pathway, while other genes and pathways were also observed (cell surface signalling pathways, membrane proteins and ion-binding proteins). Our results suggest that the BoLA region of Brangus cattle may have been enriched with Brahman haplotypes as a consequence of selection processes to promote adaptation to subtropical environments.

Keywords: Brangus, major histocompatibility complex, selection signatures, BoLA, ancestral haplotypes

Implications

Since Taurine breeds are poorly adapted to tropical environments, Brangus was developed as a composite breed to combine the adaptability, disease resistance, maternal instincts and overall hardiness of Brahman cattle with the fertility and meat quality traits of the Angus breed. Bearing this in mind, the results of this work demonstrate the enrichment of Zebuine haplotypes in the bovine leucocyte antigen (BoLA) region of Brangus cattle, probably as consequence of selection processes to promote adaptation to tropical and subtropical environments. This is consistent with the original objectives of the founder Brangus breeders and may be applied to select other Taurine—Zebuine composite breeds.

Introduction

The Brangus breed is a composite breed developed to combine the adaptability, disease resistance, maternal instincts and overall hardiness of Brahman cattle with the fertility and meat quality traits of the Angus breed. In most of the countries that currently breed Brangus cattle, the founder breed composition (FBC) has been fixed at 5/8 (62.5%) Angus and 3/8 Zebu (37.5%) (mainly Brahman). However, the Argentinian association took a different path producing animals with different fractions of Zebuine genes that go from 1/4 (25.0%) to 5/8 (62.5%). This larger fraction was the outcome of an intent to promote even better adaptation to warm and humid environments (Cantet, 2013). When considering a single locus, the fraction of animals with haplotypes from each breed is expected to be coincident to the average FBC. However, specific regions of the genome

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may present a different FBC as a result of direct (or indirect) selection or drift, and such could be the case of the major histocompatibility complex (MHC). The MHC region encodes several genes involved in the immune response in mammals. In cattle, this complex is known as BoLA and is located on chromosome 23 (BTA23: 7,013,913–28,998,760). Genetic variation in BoLA has been associated with resistance or susceptibility to several conditions such as mastitis, leucosis and parasitic infections (Takeshima and Aida, 2006). These conditions cause substantial losses for ranchers, including decreased production levels, discarded products and treatment costs, thus having a substantial economic impact on the livestock sector (Martinez et al., 2006; Biegelmeyer et al., 2015).

Many statistical approaches are available to detect selection signatures using single nucleotide polymorphism (SNP) data (Gautier and Vitalis, 2012), and they are generally classified into three categories: frequency-based methods (Tajima's D and Fay and Wu's H), population differentiationbased methods (F_{st}), and linkage disequilibrium (LD)-based methods (Rsb and iHS), as reviewed by Vitti et al. (2013). Each of these approaches allows the user to identify footprints of selection in terms of different time scale contexts (Vitti et al., 2013). Sabeti et al. (2002) identified genetic footprints of recent positive selection in humans by analysing long-range haplotypes with a measure called extended haplotype homozygosity (EHH). This measure is defined as the probability that two randomly chosen chromosomes carrying the core allele of interest are identical by descent from the core allele to a specific distance. Under a neutral evolution scenario, a new variant will require many generations to reach a high frequency in the population, and the surrounding LD will decay due to recombination events. However, when an allele rises rapidly in frequency due to strong selection, it tends to have high levels of haplotype homozygosity extending much further than expected under a neutral model. Voight et al. (2006) proposed a within population test based on the integral of the EHH decay, called integrated EHH (iHH). They defined the integrated haplotype score (iHS) as the log-ratio of iHH estimated at the ancestral and the derived focal alleles. The iHS scores are then standardized to have a mean of zero and variance of one. The limitation of this intrapopulation test is that it has low power when the selected allele is near fixation, because it will present an integral close to zero or no integral at all. To overcome this limitation, Tang et al. (2007) developed a similar procedure to compare EHH profiles between pairs of populations. This method, based on a measure called sitespecific EHH (EHHS), consists in estimating a weighted average of the EHH at both alleles of each SNP in each population. Then, the distribution of the standardized log-ratio of the integrated EHHS (iES) between pairs of populations, which is called Rsb, is used to detect signals of selection. The advantage of this method is that it calculates IHH for the entire population instead of partitioning it into ancestral and derived alleles, which eliminates the allele frequency constraint and makes it capable of detecting selection sweeps near fixation. Besides, the comparison with

another population in which the selective sweep may not have happened provides more power to the method. These EHH-based tests have been proved efficient in identifying the identification of footprints of recent selection in humans and other species (Voight et al., 2006; Tang et al., 2007). For instance, signatures of artificial selection have been identified in Holstein and Jersey dairy cattle by examining the decay of EHH against other cattle breeds (Oanbari et al., 2010; Kim et al., 2015). Other recent studies have identified genome-wide footprints of selection in tropical-adapted cattle populations (Gautier et al., 2009; Flori et al., 2012 and 2014; Bahbahani et al., 2015; Xu et al., 2015). The regions identified in these works included genes involved in the regulation of the immune system, male reproductive characteristics, and skin and hair structure. To the best of our knowledge, no such studies have been conducted in the Brangus breed until now. Considering the role of BoLA genes in immune response, specific regions of this complex are expected to present evidence of selection in the Brangus breed as a consequence of positive selection towards Zebuine variants in tropical and subtropical environments. Thus, the objectives of the present study were: (1) to evaluate whether the local ancestry percentages of the BoLA region diverged from the genome-wide estimated proportion; and (2) to identify signatures of positive selection in the BoLA region and to verify whether they were recent or ancestral through a population differentiation method (F_{st}) and two LD-based methods: iHS and Rsb.

Material and methods

Animal samples

A total of 167 bovine samples representing the two ancestral breeds (Angus (n=45), Brahman (n=22)) and the composite breed (Brangus (n=100)) were analyses. The animals were selected by the corresponding argentine breeders' associations (Asociación Argentina de Brangus, www. brangus.org.ar; Asociación Criadores Brahman Argentina, http://www.brahman.org.ar/) to represent the key animals of local populations according to breed pedigree data and expected progeny differences values. The Institutional Committee on Care and Use of Experimental Animals from the School of Veterinary Sciences of the National University of La Plata (Buenos Aires, Argentina) reviewed and approved all animal procedures.

DNA extraction, genotyping and quality control

Genomic DNA was extracted from venous blood, hair follicle and semen samples through organic extraction, standardized NaOH/HCl methods, and commercial kits (WizardTM Genomic DNA Purification Kit (Promega, Madison, WI, USA), DNeasy Blood and Tissue Kits (Qiagen, Hilden, Germany)). Then, DNA samples were genotyped in a GeneTitanTM platform (Affymetrix, Santa Clara, CA, USA) using the AxiomTM Bos 1 Genotyping Array r3 (Affymetrix), which contains 648 855 SNPs.

Raw data were processed using the Axiom[™] Analysis Suite software (Affymetrix) and SNPs were filtered by sample (≥97%) and SNP call rates (≥97%). Finally, processed data

were filtered by position to obtain the SNPs located in the BoLA region (BTA23: 7,013,913–28,998,760), which resulted in 5585 SNPs. SNP Annotation was performed using the Axiom_GW_Bos_SNP_1.na35.annot file (Affymetrix) and positions were assigned according to the bovine genome assembly UMD 3.1.

Genome-wide breed composition analysis

The global FBC of the Brangus samples was estimated from a subset of 4556 autosomal and evenly spaced SNPs from the Bos1 Array (available from the corresponding author upon request) through the clustering algorithm implemented in STRUCTURE 2.3.4 software (Pritchard *et al.*, 2000). Since the software assumes independence in SNP data, SNPs were selected to have an approximate distance of 0.5 Mb between each other, because this is the average LD block length for many cattle breeds. The analysis was performed considering two clusters (K = 2) according to the historical number of ancestral populations (Angus and Brahman). This test was executed considering an admixture model with independent allele frequencies, and a burn-in of 100 000 iterations followed by 1 000 000 Markov Monte Carlo iterations.

Local ancestry inference in the bovine leucocyte antigen region (ancestral haplotypes)

An interactive pipeline was developed using BioSmalltalk (Morales and Giovambattista, 2013) to automate the process of inference of local ancestry in the Brangus population (Supplementary Figure S1). Briefly, processed data were exported from the AxiomTM Analysis Suite software in three tabular-like files: a 'forward strand base call' report file and two complementary files (PED/MAP) for further analysis with bioinformatics tools. Then, reference alleles and family information columns were added to the PED/MAP, which was binarized using PLINK c1.07 (Purcell *et al.*, 2007). Haplotype inference and missing genotype imputation were conducted using Shapelt v2 (r644) software (Delaneau *et al.*, 2014) with a window size of 2 Mbp and effective population sizes of 100, 106 and 150 for Angus, Brangus and Brahman, respectively.

Our pipeline software was then used to transpose the Angus and Brahman haplotypes, to format Brangus genotypes and to extract SNP positions from the MAP file. The resulting data were used as input to infer percentages of local ancestry through the LAMP-LD software (Baran $et\ al.$, 2012). The convertLAMP-LD, which is compressed using SNP intervals, into a full haplotype-like format. Results were plotted by position using the ROASSAL package (Araya $et\ al.$, 2013) in the BioSmalltalk environment. Finally, a χ^2 statistical test was applied to evaluate whether the average local ancestry proportion diverged from the global composition estimated in the previous analysis.

Signatures of positive selection

A combination of genetic differentiation and linkage disequilibrium-based methods was used to identify signatures of positive selection across the BoLA region. The genetic differentiation of Branqus was evaluated by estimation of Weir and Cockerham's F_{st} against Angus and Brahman. The analysis was conducted using vcftools (Danecek et al., 2011) in windows of 3 kbp after filtering data by MAF \geqslant 0.05. The upper 1% of the distribution was arbitrarily chosen as threshold to determine candidate regions, which were defined as two or more significant windows separated by \leqslant 1 Mb. Additionally, two EHH-derived statistics, iHS (intrapopulation) and Rsb (interpopulation), were implemented to identify footprints of selection in the BoLA region. These statistics were calculated using the output files from the BioSmalltalk pipeline and the rehh package for R software (Gautier and Vitalis, 2012). To compute iHS, the Brangus ancestral allele was defined as the allele with the highest frequency estimated

$$f = w_{AA}f_{AA} + w_{Rh}f_{Rh}$$

where $w_{AA} = 0.625$ and $w_{Bh} = 0.375$ represent the average expected proportions of Angus and Brahman ancestries in the Brangus genome and f_{AA} and f_{Bh} represent the allele frequencies in Angus and Brahman (Hacia *et al.*, 1999; Gautier and Naves, 2011). The *iHS* value was calculated for SNPs with MAF \geqslant 0.05 within the Brangus population. Then, a two-tailed *Z*-test was applied to detect statistically significant SNPs under selection with unusually extended haplotypes of ancestral (positive *iHS* values) or derived alleles (negative *iHS* values) since the standardized *iHS* values were normally distributed (Supplementary Figure S2). *P*-values were derived as $-\log_{10}(1-2|\Phi(iHS)-0.5|)$, where $\Phi(iHS)$ represents the cumulative distribution function of the Gaussian density.

The *Rsb* statistic was calculated between Brangus and each of the two ancestral breeds. A *Z*-test was performed to detect statistically significant SNPs under selection (positive *Rsb* scores). One-sided *P*-values were derived as $-\log_{10}(1-\Phi(Rsb))$, where $\Phi(Rsb)$ represents the Gaussian cumulative distribution function. The significance threshold was set at $-\log_{10}(P\text{-value}) = 2$ for both *iHS* and *Rsb*. Candidate regions were defined as two or more significant SNPs separated by ≤ 1 Mb.

Identification of genes and pathways in candidate regions Genes in the candidate regions resulting from the F_{st} and LD-based methods were retrieved from Biomart ENSEMBL (http://www.ensembl.org/biomart) using the Ensembl Genes 87 — Cow (UMD 3.1) database and the Multiple Chromosomal Regions filter. The list of candidate genes obtained from Biomart was analysed on PANTHER Classification System (Huaiyu *et al.*, 2016) to group and classify them according to biological process and pathway representation.

Results and discussion

Genome-wide breed composition analysis

All the Brangus samples showed admixed composition in the genome-wide study conducted with STRUCTURE (Figure 1, Table 1). The average the Brahman fraction was $34.7\% \pm 9.0\%$ and the median value was 33.1%, ranging from 22.3% to 81.1% (Supplementary Figure S3). These results are

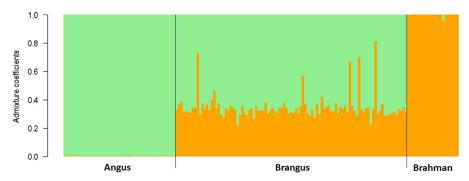


Figure 1 Brangus genome-wide composition analysis. The Brahman (orange) and Angus (green) genetic fractions are shown for each individual. Values were estimated from a subset of SNP data (5K panel) using STRUCTURE software (K = 2).

Table 1 Average Brangus cattle composition values estimated by STRUCTURE and LAMP-LD software

Method	Angus	Brahman
STRUCTURE (Genome-wide)	65.3	34.7
LAMP-LD (BoLA region)	45.0	55.0

consistent with those recently reported by Cantet (2013), who detected composition values closer to 2/3 Angus (66.6%) – 1/3 Brahman (33.3%) using a microsatellite panel. The fractions are concordant with the theoretical expectation of heterosis retention for a two-breed crossbreeding system that stabilizes at 2/3:1/3 (Dickerson, 1973; Table 1). Several conjectures about selection drift or other genetic mechanisms can be formulated based on these results, but they are beyond the scope of this article. This genome-wide value of breed composition will be considered from here on as neutral reference to specifically analyse the BoLA region on the following sections of this article.

Local ancestry inference in the bovine leucocyte antigen region of Brangus

Local ancestry in the BoLA region was estimated through the LAMP-LD software. More than half of the BoLA haplotypes were inferred to be shared with Brahman (0.55 \pm 0.04), which suggests a Zebuine origin for these haplotypes. This local value was significantly different from the genome-wide proportion obtained from STRUCTURE, which was of 34.7% (P< 0.01). Results are shown in Figure 2, where uneven breed origin is observed across the BoLA region. Two specific regions exhibited notable deviations (>2 SD) towards Brahman haplotypes at 10 to 20 Mbp and 25 to 29 Mbp, while the region at 20 to 25 Mbp showed a Brahman fraction more akin to the genome-wide proportion estimated with STRUCTURE. Considering the crucial role of the BoLA region in immune response, the increase in Brahman haplotypes may be reflecting adaptation to tropical and subtropical environments.

Identification of selection signatures in the bovine leucocyte antigen region

The first approach to detect evidence of selection in the BoLA region was performed via genetic differentiation (F_{st}) between

pairs of populations. When the analysis was conducted between the two parental breeds (Angus–Brahman), the interpopulation genetic differentiation accounted for 33% of the total genetic variance ($F_{st} = 0.33$). These results are concordant with previous works reporting a large divergence in haplotype and allele composition between Taurine and Zebuine breeds in the BoLA region (Mikko and Anderson, 1995); this difference enables the detection of selection events in the Taurine–Zebuine Brangus breed. The F_{st} estimations for the Brangus–Angus and Brangus–Brahman comparisons resulted in weighted values of 0.06 and 0.17, respectively. Seven candidate regions were detected in the former case and three regions were detected in the latter (upper 1%; Figure 3a and b: Table 2).

In addition, two LD-based methods, iHS and Rsb, were used to identify more recent selection signatures in the BoLA region. The results of these analyses are presented in Figure 4a, b and c and Table 3. The iHS analysis showed four candidate regions (R4, R5, R6, R7) with significantly different IHH values between the ancestral and derived alleles of the Brangus population ($-\log (P\text{-value}) > 2$) (Table 3). On the other hand, the Rsb analyses between Brangus and each of the two ancestral populations revealed six candidate regions. Of these six, four were detected against Angus (R1, R2, R3, R6) and four were detected against Brahman (R2, R4, R5, R6) (Table 3). Interestingly, region R6 showed significant values in the three tests (*iHS*, Rsb_{AA} , Rsb_{BH}), while three regions exhibited significant values for two tests: R2 (Rsb_{AA} , Rsb_{BH}), R4 (iHS, Rsb_{BH}) and R5 (iHS, Rsb_{BH}). In summary, seven candidate regions containing between 1 and 124 significant SNPs were identified in the BoLA region of the Brangus population through *iHS* and *Rsb*.

The differences in the number and distribution of candidate regions obtained in the F_{st} analysis and the EHH-based studies may be due to the nature of the analysis methods themselves, since population differentiation-based methods such as F_{st} are best suited to detect selection events that occurred further in past, while LD-based methods are more appropriate for recent events (Ferrer-Admetlla *et al.*, 2014).

Identification of genes and pathways in candidate regions The candidate regions obtained in the F_{st} analyses were large (five of them longer than 0.5 Mb), which led to a wide list of

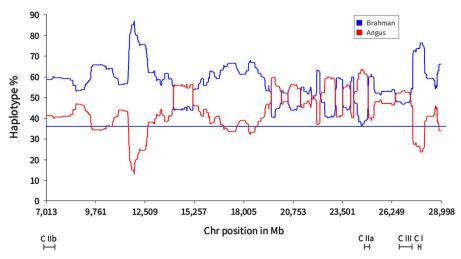


Figure 2 Local ancestry percentage (Angus = red, Brahman = blue) across the bovine leucocyte antigen region of the Brangus population derived from LAMP-LD software. The expected Brahman percentage (34.7%) is shown as a horizontal blue line.

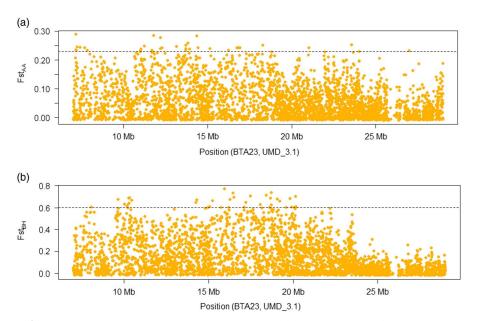


Figure 3 Manhattan plots of the pairwise bovine leucocyte antigen region F_{st} analyses. (a) Brangus v. Angus, (b) Brangus v. Brahman.

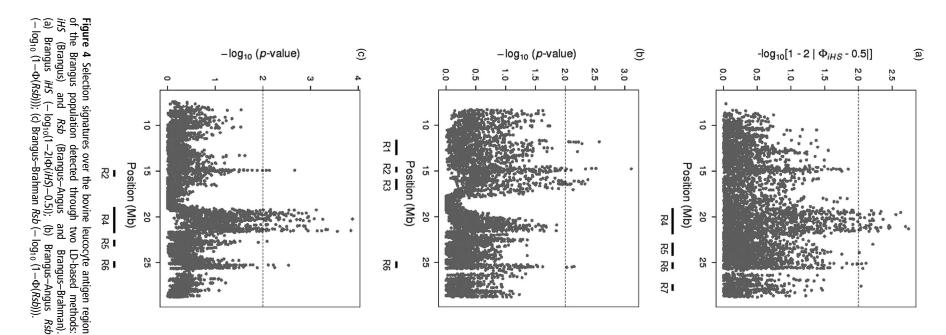
143 candidate genes involved in several different biological processes. In order to obtain a global view of the types of genes or pathways that were represented within the list, PANTHER Classification System was used (Supplementary Table S1). Most of those genes participate in a wide range of different cellular (59 genes), metabolic (57 genes), and localization processes (19 genes), while others participate in the response to stimulus (22 genes) and the immune system (13 genes). In addition, 18 were assigned to specific biological pathways: as angiogenesis, T-cell activation, Ras pathway, FGF signalling pathway and Gonadotropinreleasing hormone receptor pathway, all related to the immune system. For instance, induction of angiogenesis is mediated by macrophages, which produce pro-inflammatory cytokines, endothelial growth factors (VEGF, bFGF), and proteases (MMP9) that either directly stimulate growth of new vessels, degrade tissue matrix or stimulate other cells to

produce angiogenic factors. Regarding T-cell activation, the second pathway, is a crucial part of the cellular immune response; while Ras, Rac and Rho proteins play roles in regulating the functions of T and B lymphocytes and dendritic cells. Fibroblast growth factor receptors are expressed by some T-cells, and provide co-stimulation for them, and lastly, GnRH is also known to be involved in the modulation of immune system function.

Concerning the *EHH*-based analyses, region R6 was the most interesting region for being the only one to show evidence of recent selection in the three tests (iHS, Rsb_{AA} and Rsb_{BH}). It encompassed 0.55 Mb and included seven BoLA Class II genes and one BoLA Class II pseudogene. In addition, two other genes associated with immune response and fatty acid synthesis were also located in this region. This result may be a consequence of recent selection processes promoted by breeders, since several works have reported

 Table 2 Candidate regions for positive selection by F_{st} pairwise comparisons with the ancestral breeds (Brangus-Angus and Brangus-Brahman)

Candidate region	Comparison	Genomic interval (Kb)	Number of SNPs	Max F _{st}	Candidate genes
A1	Angus	7149 7842	9	0.289	HLA-DMA, BOLA-DOA, BRD2, COL11A2, RXRB, SLC39A7, HSD17B8, RING1, VPS52, RPS18, B3GALT4, WDR46, PFDN6, RGL2, TAPBP, DAXX, KIFC1, PHF1, CUTA, SYNGAP1, ZBTB9, BAK1, ITPR3, IP6K3, LEMD2, ZBTB22, bta-mir-2376
A2	Angus	10 998 15 132	20	0.285	PIM1, TMEM217, TBC1D22B, RNF8, CMTR1, MDGA1, ZFAND3, GLO1, DNAH8, GLP1R, SAYSD1, KCNK5, KCNK17, KCNK16, DAAM2, LRFN2, UNC5CL, TSPO2, APOBEC2, OARD1, NFYA, TREML1, TREM2, TREML2, U6, 7SK, bta-mir-2377, U6, SNORD112
A3	Angus	16 233 16 929	6	0.243	UBR2, PRPH2, GLTSCR1L, RPL7L1, PTCRA, CNPY3, GNMT, PEX6, PPP2R5D, KLHDC3, CUL7, MRPL2, KLC4, PTK7, SRF, CUL9, DNPH1, SLC22A7, CRIP3, ZNF318, TBCC, 5S_rRNA, U6
A4	Angus	18 267	1	0.252	SUPT3H
A5	Angus	21 024	1	0.242	
A6	Angus	23 547 23 550	2	0.253	
A7	Angus	26 970	1	0.233	NOTCH4
B1	Brahman	8067 8070	2	0.608	GRM4
B2	Brahman	9657 10 461	10	0.692	ARMC12, CLPS, LHFPL5, SRPK1, SLC26A8, MAPK14, MAPK13, BRPF3, PNPLA1, C6orf222, KCTD20, STK38, RPL27, U6
B3	Brahman	14 241 20 172	31	0.772	LRFN2, UNC5CL, TSPO2, APOBEC2, OARD1, NFYA, TREML1, TREM2, TREML2, TREM1, FOXP4, MDFI, TFEB, PGC, FRS3, USP49, MED20, BYSL, CCND3, TAF8, C6orf132, GUCA1A, GUCA1B, MRPS10, TRERF1, UBR2, PRPH2, GLTSCR1L, RPL7L1, PTCRA, CNPY3, GNMT, PEX6, PPP2R5D, KLHDC3, CUL7, MRPL2, KLC4, PTK7, SRF, CUL9, DNPH1, SLC22A7, CRIP3, ZNF318, DLK2, TJAP1, LRRC73, YIPF3, POLR1C, XPO5, POLH, GTPBP2, MAD2L1BP, RSPH9, MRPS18A, VEGFA, AARS2, MRPL14, TMEM63B, CDC5L, CAPN11, SLC29A1, HSP90AB1, SLC35B2, NFKBIE, RUNX2, CLIC5, ENPP4, ENPP5, RCAN2, CYP39A1, SLC25A27, TDRD6, PLA2G7, ANKRD66, MEP1A, ADGRF5, MGC137036, TBCC, U6, U6, 55_rRNA, U6, 55_rRNA, U6, U6



able 3 Candidate regions for positive selection in Brangus cattle by the intrapopulation iHS and interpopulation Rsb tests (Brangus—Angus and Brangus—Brahman)

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Region	Interval (Kb)	Peak SNP	Peak position	;HZ*	iHS* Rsb _{AA} * *	Rsb _{BH} ** Genes	Genes
R1	11 793-12 871	rs208391975	1,179,8813		2.57		ZFAND3, BTBD9, GL01, DNAH8
R 2	14724-14917	rs383325987	14,760,200		3.11	2.67	UNCSCL
22	15 951-17 043	rs472225883	15,951,447		2.38		TRERF1, UBR2, PRPH2, GLTSCR1L, RPL7L1, PTCRA, CNPY3, GNMT, PEX6, PPP2R5D, KLHDC3,
							CULZ, MRPLZ, KLC4, PTK7, SRF, CUL9, DNPH1, SLC22A7, CRIP3, ZNF318, DLK2, TJAP1, LRRC73, YIPF3, POLR1C, TBCC, 55 rRNA, U6
R4	19224-21 583	rs209240475	20,242,474	2.70		3.88	CLIC5, ENPP4, ENPP5, RCAN2, CYP39A1, SLC25A27, TDRD6, PLA2G7, ANKRD66, MEP1A,
							ADGRF5, ADGRF1, TNFRSF21, CD2AP, ADGRF2, ADGRF4, OPN5, PTCHD4, U6
83	22 683-23 907	rs110198940	23,006,324	2.21		2.09	TFAP2D, TFAPB, PKHD1
R6	25 081-25 639	rs382005618	25,292,973	2.11	2.15	2.54	ELOVIS, BOLA-DQA2, BOLA-DQB, BOLA-DQA5, BOLA-DRB3, BOLA-DQA1, LA-DRB,
							BOLA-DRB2, BOLA-DRA, BTNL2, LOC100851058
R7	27 572	rs380665096	I	2.04			DDX39B
* – logs (1	loα.。(12lΦ(iHS)0 5l)						

associations between BoLA Class II polymorphisms and resistance/susceptibility to infectious disease, including bovine leukaemia virus-induced lymphoma, mastitis and dermatophilosis. Moreover, BoLA has been suggested to influence other production traits such as milk yield, growth, reproduction and immune response (revised by Takeshima and Aida, 2006).

Regions R2, R4 and R5 exhibited significant values for two scores (Table 3). No genes were identified within R2, but interestingly, UNC5CL (UniProtKB - E1BGH1) was located close to the downstream end. UNC5CL is involved in the TNF-alpha/NF-kB signalling pathway, which plays a role in the regulation of immune cells. Region R4 encompassed 2.3 Mb and included TNFRSF21, which is involved in the TNF and ERK signalling pathways, and two genes with G-protein coupled receptor and transmembrane signalling receptor activities (ADGRF1 and ADGRF5). R5 spanned 1.22 Mb and contained three genes: TFAP2D, TFAP2B and PKHD1. The first two genes are transcription factors that modulate different processes during development and have no clear link with immunity, but the third, PKHD1, has been suggested to have a role in immunity since its expression is upregulated in activated T-cells (Hogan et al., 2004). The remaining regions (R1, R3 and R7) showed significant values for only one statistic index and included a number of additional genes related with the immune system (UBR2, CUL7) among others with several different functions.

The list of genes in candidate regions obtained with EHH-based analyses included 77 genes, which were then used to identify groups and biological pathways with PANTHER (Supplementary Table S2). There were 62 genes participating in the same processes: metabolic (25 genes) and cellular processes (23 genes), response to stimulus (21 genes) and immune system (17 genes). In the case of response to stimulus, cellular defense result in 12 genes (BoLA-DQA2, BoLA-DQA1, BTNL2, BoLA-DQB1, LOC101902890, BoLA-DRA, BoLA-DQB, LOC525599, LRRC73, CRIP3, BoLA-DRB3, BoLA-DQA5), while the immune system was represented by eight genes with antigen processing and presenting functions (BoLA-DQA, BoLA-DQA2, BoLA-DQB1, LOC101902890, BoLA-DRA, BoLA-DOB, BoLA-DRB3, BoLA-DQA5) and five to other immune response-related functions (ADGRF1, ADGRF5, ADGRF2, GPR115, TNFRSF21). In addition, T-cell activation pathway was the only with more than one gene detected (BoLA-DQA, BoLA-DQA5, BoLA-DQA2, BoLA-DRA). Other pathways were: EGF receptor (PPP2R5D), FGF (PPP2R5D), Gonadotropin-releasing hormone receptor (SRF), Interleukin (SRF), PDGF (SRF), Ras (SRF), Wnt (PPP2R5D) and p38 MAPK (SRF) pathways.

Class II genes, such as *BoLA-DRB3*, *BoLA-DQA1* and *BoLA-DQB*, play an important role in immune response mechanisms (peptide binding and antigen presentation). It is worth mentioning that although immune response pathways were represented in both of our analyses (genetic differentiation and LD), the selected regions included genes involved in different immune mechanisms and other biologically important processes. As previously mentioned, these differences may be caused by the specific concepts exploited by each

 $\log_{10} (1 - \Phi(Rsb))$

of these methods, with time scale likely having a large impact (Vitti *et al.*, 2013, Cadzow *et al.*, 2014).

Several works have reported selection signatures based on adaptation to tropical and subtropical environments in the last years (Gautier and Naves, 2011; Flori et al., 2012; Kim and Rothschild, 2014). The selected regions included candidate genes involved in reproductive development and reproductive functions, milk yield and composition, metabolism pathways. thermotolerance, and horn development. Furthermore, candidate genes associated with innate immune responses (Kim and Rothschild, 2014) and cellular signalization of immune response (Gautier and Naves, 2011; Iso-Touru et al., 2016) have been also reported. Our results provide additional evidence about the presence of selection signatures in genomic regions involved in the immune response in cattle. Besides, this study illustrates the relevance of using composite breeds with well-known parental populations to identify footprints of selection in livestock species (Khayatzadeh et al., 2016).

In conclusion, our results reveal an enrichment of Brahman haplotypes with respect to the average genome proportion in the BoLA region of the Brangus breed. Moreover, the segments under selection in the BoLA region included genes involved in immune response pathways. This may be a consequence of selection processes to promote adaptation to tropical and subtropical environments, which is consistent with the objectives of the foundation of the Brangus breed.

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Supplementary material

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