Focus on Gonadotrophin Signalling

What have gonadotrophin overexpressing transgenic mice taught us about gonadal function?

Susana B Rulli¹ and Ilpo Huhtaniemi²

¹Institute of Biology and Experimental Medicine-CONICET, Vuelta de Obligado 2490, 1428 Buenos Aires, Argentina and ²Institute of Reproductive and Developmental Biology, Imperial College Faculty of Medicine, Hammersmith Campus, Du Cane Road, London W12 0NN, UK

Correspondence should be addressed to I Huhtaniemi; Email: ilpo.huhtaniemi@imperial.ac.uk

Abstract

The two gonadotrophins, follicle-stimulating hormone and luteinising hormone, are pivotal regulators of the development and maintenance of normal fertility by maintaining testicular and ovarian endocrine function and gametogenesis. Too low gonadotrophin secretion, i.e. hypogonadotrophic hypogonadism, is a common cause of infertility. But there are also physiological and pathophysiological conditions where gonadotrophin secretion and/or action are either transiently or chronically elevated, such as pregnancy, pituitary tumours, polycystic ovarian syndrome, activating gonadotrophin receptor mutations, perimenopause and menopause. These situations can be either the primary or secondary cause of infertility and gonadal pathologies in both sexes. Also the role of gonadotrophins as tumour promoters is possible. Recently, the possibility to combine information from genetically modified mice and human phenotypes in connection with mutations of gonadotrophin or gonadotrophin receptor genes has elucidated many less well known mechanisms involved in dysregulation of gonadotrophin function. Among the genetically modified mouse models, transgenic mice with gonadotrophin hypersecretion have been developed during the last few years. In this review, we describe the key findings on transgenic mouse models overexpressing gonadotrophins and present their possible implications in related human pathologies. In addition, we provide examples of genetic mouse models with secondary effects on gonadotrophin production and, consequently, on gonadal function.

Reproduction (2005) 130 283-291

Physiological actions of gonadotrophins

Gonadal function is regulated by complex physiological and molecular processes that are to a great extent under the control of the hypothalamic-pituitary-gonadal axis. The integrity of this regulatory cascade is crucial, since disturbances at any of its levels can produce reproductive defects. The pituitary gonadotrophins follicle-stimulating hormone (FSH) and luteinising hormone (LH) and their receptors (R) are the main regulators of gonadal function, and have been extensively studied (Simoni et al. 1997, Ascoli et al. 2002, Burger et al. 2004). LH and FSH, together with thyroid-stimulating hormone, are members of the pituitary glycoprotein hormone family. They are heterodimers composed of a common α -subunit and the hormone-specific β-subunit. LH and FSH exert their effects through binding to their cognate G-protein-coupled transmembrane receptors (GPCR; Simoni et al. 1997, Ascoli et al. 2002).

The decapeptide gonadotrophin-releasing hormone (GnRH) is released in a pulsatile fashion from specific hypothalamic nuclei into the hypothalamo-hypophyseal portal circulation, and it stimulates the synthesis and secretion of LH and FSH. Gonadal steroid and peptide (mainly inhibin) hormones exert negative and positive feedback effects on gonadotrophin synthesis and secretion, either directly at the pituitary level or indirectly via the hypothalamus, mainly by modulating GnRH secretion (Burger et al. 2004). The physiological actions of LH and FSH are well characterised and they are essential for folliculogenesis, ovulation and steroidogenesis in females, and for testicular growth, spermatogenesis and steroidogenesis in males. The placental analogue of LH, human chorionic gonadotrophin (hCG), is a fourth member of the glycoprotein hormone family. However, it interacts with the same LH/hCG receptor as LH, and functions as an LH agonist with a longer half-life and higher biopotency than its pituitary counterpart (Jameson & Hollenberg 1993).

The first experimental evidence suggesting that the gonadal function is regulated by pituitary-derived compound(s) was based on implantation of the anterior pituitary gland into immature male and female rodents, which induced precocious puberty, enlargement of the ovaries and superovulation (Smith & Engle 1927). A few years later, the gonadotrophic principle was proposed, due to the extraction from the pituitary gland of two different hormones that both stimulated the gonads, displaying biological properties of the follicle-stimulating and luteinising hormones respectively as we know them today (Fevold *et al.* 1931). With regard to hCG, it was demonstrated that the blood and urine of pregnant women presented with gonad-stimulating properties, originating from the placenta.

Subsequently, the isolation of increasingly purer gonadotrophin preparations from pituitary glands for experimental studies and from urinary extracts for clinical use has advanced our knowledge of gonadotrophin physiology, and improved the management of infertility. However, it is difficult to develop protocols for chronic administration of exogenous gonadotrophins that would mimic physiological effects of gonadotrophin secretion in the long term. Since these methods rely on pharmacological strategies to elevate gonadotrophin levels, findings on short- and long-term gonadotrophin treatments in experimental animals seem to be strongly age, dose, and time dependent (Risbridger et al. 1982, Scott et al. 1990). Moreover, formation of antibodies against heterologous proteins poses a confounding factor upon long-term experiments. In most experimental conditions, treatment of animals chronically with LH, hCG, or FSH results in decreased responses of their testicular and ovarian target cells through receptor down-regulation and desensitisation of signalling (Conti et al. 1976, Dufau et al. 1979).

The advent of recombinant DNA technology and transgenic animals made it possible to obtain new insight into the effects of chronically elevated gonadotrophin levels and the physiopathology of function of the hypothalamic—pituitary—gonadal axis. Consequently, mouse models exist now for many human reproductive abnormalities due to genetic alterations in gonadotrophin secretion or action (Themmen & Huhtaniemi 2000, Huhtaniemi & Themmen 2005, Themmen *et al.* 2005). Despite the major advancement in this field, many reproductive disturbances still remain idiopathic.

Elevated gonadotrophin levels and human diseases

There are different physiological and pathophysiological conditions where gonadotrophins can be either transiently or chronically elevated during the lifetime of humans. One such situation is pregnancy when hCG is produced in very high amounts during the first trimester to maintain the progesterone production of the corpus luteum of pregnancy, which prepares the uterus for implantation, and for embryonic and placental development (Jameson & Hollenberg 1993). hCG also stimulates foetal testicular

testosterone production which is needed for masculinisation of the male foetus.

Another physiological condition with highly elevated gonadotrophin levels occurs in women during ageing. Gonadotrophin secretion starts slowly increasing during the years preceding the menopause (premenopause), and increases 10- to 20-fold after the last menstrual bleeding, menopause. Thereafter, the female body is chronically exposed to high levels of gonadotrophins for decades. The postmenopausal increase in gonadotrophin levels is basically due to exhaustion of the ovarian follicle pool and consequent cessation of the negative feedback of ovarian hormones (oestrogen, progesterone, inhibin) at the hypothalamic-pituitary level. Despite the cessation of ovarian function, this prolonged exposure to gonadotrophins may have effects on postmenopausal women, especially in view of the ubiquitous extragonadal expression of LHR/hCGR (Filicori et al. 2005).

In terms of pathological conditions, surgical or chemical gonadectomy (Huirne & Lambalk 2001), primary gonadal failure with missing feedback regulation (Salbenblatt *et al.* 1985, Quigley 2002), hypothalamic dysfunction (Marshall *et al.* 2001), or gonadotroph adenomas (Roberts *et al.* 2005) are other known conditions that induce high gonadotrophin levels. On the other hand, expression of hCG occurs in trophoblastic diseases and testicular germ cell tumours (Cole & Butler 2002, Stenman *et al.* 2004). Finally, polycystic ovarian syndrome (PCOS) is a condition where chronic elevation of LH levels is a diagnostic hallmark (Franks & McCarthy 2004).

Many clinical and epidemiological studies have suggested that inappropriately elevated gonadotrophin action leads to infertility and gonadal pathologies in both sexes. For instance, elevated gonadotrophin levels are associated with ovarian hyperstimulation syndrome (Delvigne & Rozenberg 2002) and PCOS (Franks & McCarthy 2004). Exposure to elevated gonadotrophins after the menopause or infertility treatments is proposed to be a risk factor for developing ovarian tumours (Risch 1998, Konishi et al. 1999, Riman et al. 2004). In contrast, reduced risk for ovarian cancer is associated with multiple pregnancies, breast-feeding, oral contraceptive use, and oestrogen replacement therapy, all of which lead to lower levels and reduced exposure to gonadotrophins (Gnagy et al. 2000, La Vecchia 2001). Much less is known about possible pathological effects of gonadotrophin dysregulation in men, and on the basis of available evidence it seems that the female reproductive system is more vulnerable to gonadotrophin dysregulation than the male.

In men, the best examples of reproductive disturbances due to defective gonadotrophin action are the naturally occurring mutations of LHR and FSHR (Themmen & Huhtaniemi 2000, Huhtaniemi & Themmen 2005). The activating mutations permanently stimulate, in the absence of their cognate ligand, the receptor signalling pathways, evoking a condition of precocious, chronically elevated gonadotrophin action. In the case of LHR, a number of

activating mutations causing male-limited gonadotrophinindependent hypersecretion of testosterone and precocious puberty have been described (Themmen & Huhtaniemi 2000). Interestingly, one specific mutation (Asp⁵⁷⁸His) has been reported in association with Leydig cell adenomas (Liu et al. 1999). This observation supports the hypothesis that gonadotrophin action is involved in gonadal tumorigenesis. On the other hand, only one activating mutation of the FSHR gene has so far been identified in humans (Gromoll et al. 1996). This patient, a male who was hypophysectomised and treated with testosterone, displayed normal spermatogenesis and was, unexpectedly, fertile in spite of undetectable gonadotrophins.

Interestingly, FSHR mutations making the receptor responsive to hCG stimulation were recently identified by two different groups in association with familial gestational spontaneous ovarian hyperstimulation syndrome (Smits et al. 2003, Vasseur et al. 2003). These mutations display abnormal hypersensitivity to hCG activation through the structurally altered FSHR, which presented during pregnancy when the circulating levels of endogenous hCG are very high. This finding supports the concept that the FSH pathway has a pivotal role in the pathophysiology of ovarian hyperstimulation syndrome.

Although gonadotrophin actions are mainly directed to the regulation of gonadal function, the recently discovered extragonadal sites of gonadotrophin subunit and receptor expression suggest that extragonadal gonadotrophin effects may also exist (Filicori et al. 2005). This possibility extends the spectrum of gonadotrophin actions, especially in women, beyond reproductive age and suggests the possibility that the high postmenopausal gonadotrophin levels could have a physiological role. However, the topic of extragonadal gonadotrophin action remains controversial, not the least in light of the very recent findings on LHR knockout mice (Pakarainen et al. 2005).

Transgenic mouse models as a tool to study gonadal function

The generation of genetically modified animal models provides powerful means to understand the physiological role of gonadotrophins in reproductive function as well as the pathologies arising from their dysregulation. These techniques provide in vivo models to study the role of a particular hormone throughout the life of the animal, including the very early developmental stages. Increasing interest has emerged among researchers to generate animal models that mimic human pathologies (Burns & Matzuk 2002, Huhtaniemi et al. 2002, 2005). Gain-of-function models consisting of mice overexpressing a certain gene of interest may mimic the effects of hypersecretion syndromes and activating mutations in humans. Another alternative is the generation of mice bearing targeted point mutations that may be even closer phenocopies of human activating mutations. On the other hand, loss-of-function mutations in knockout mice are able to recreate hormone deficiency and resistance syndromes in humans (Kumar et al. 2005). We will present below key examples of transgenic mice overexpressing gonadotrophins and their possible implications in relation to human pathologies (Table 1).

Transgenic mice overexpressing FSH

In order to elucidate the biological role of FSH in gonadal growth, function and tumour development, Kumar et al. (1999) developed gain-of-function mutant bi-transgenic mice overexpressing the human glycoprotein hormone-α and human (h) FSHB-subunits under the ectopically expressing mouse metallothionein-1 promoter. With this approach, one line of mice with a low copy number of hFSHβ directed the expression of circulating hFSH at levels comparable to those in postmenopausal women. In this case, both males and females were fertile and did not present any abnormalities in other tissues. These studies showed that prolonged exposure to elevated FSH levels for more than one year did not directly cause ovarian tumorigenesis or other functional abnormalities.

A second line of these transgenic mice had a high copy number of the FSHβ transgene, expressing hFSH levels that by far exceeded those found in postmenopausal women (Kumar et al. 1999). Female transgenic mice were infertile and developed haemorrhagic and cystic ovaries. They had elevated testosterone, oestradiol and progesterone levels in serum, and developed enlarged and cystic kidneys. These mice died before 13 weeks of age due to urinary tract obstruction, but had no signs of ovarian tumours. This concept was further confirmed with the generation of mouse models with different genetic approaches that supported the influence of FSH as an important trophic modifier for gonadal tumorigenesis. Inhibindeficient mice that have increased FSH and activin levels developed multiple sex-cord stromal tumours of the granulosa and Sertoli cell lineage (Matzuk et al. 1992). It was demonstrated that gonadotrophins are required for tumour development in these mice, since double-mutant mice homozygous for the gonadotrophin-deficient hypogonadal (hpg) mutation in the inhibin null background did not develop tumours (Kumar et al. 1996). Moreover, inhibin and FSH null mice showed a significant delay in tumour development and a less aggressive phenotype compared with mice deficient only in inhibin (Kumar et al. 1999). A similar role for gonadotrophins as tumour promoters was demonstrated in transgenic mice expressing the SV40 T-antigen under the inhibin α -subunit promoter, since development of gonadal somatic cell tumours in these mice was dependent on gonadotrophin secretion (Kananen et al. 1997, Mikola et al. 2003). However, in these mice it was found that LH was the main tumourpromoting gonadotrophin.

Male hFSH transgenic mice were infertile, produced elevated levels of serum testosterone and presented with enlarged seminal vesicles. Testicular growth and spermatogenesis, however, appeared morphologically normal.

Table 1 Transgenic mouse models with gonadotrophin overexpression.

Construct	Phenotype	References
MT-1 promoter/human FSHαβ	Males infertile, normal spermatogenesis, reproductive tract obstruction; females infertile, haemorrhagic and cystic ovaries	Kumar <i>et al.</i> (1999)
Rat insulin II promoter/FSH $\alpha\beta \times hpg$	Selective FSH action independently of LH	Allan et al. (2004)
Rat androgen-binding protein promoter/ mutant FSH receptor × hpg	Constitutive signalling and cell-surface expression of FSH receptor; male specific; FSH-like Sertoli cell response	Allan et al. (2004)
Inhibin α knockout	High FSH and activin; sex-cord stromal tumours	Matzuk et al. (1992)
Inhibin α knockout \times hpg	Sex-cord stromal tumours abolished	Kumar et al. (1996)
Inhibin α knockout × FSH knockout	Sex-cord stromal tumours delayed	Kumar <i>et al.</i> (1999)
hCGβ promoter/hCGβ cluster	No phenotype	Strauss et al. (1994)
bLHα promoter/bLHβ-CTP	Males fertile; females polycystic ovaries, granulosa/theca cell	Risma et al. (1995),
	tumours, pituitary adenomas, mammary tumours	Mann et al. (2003)
Ubiquitin C promoter/hCGβ	Males fertile; females infertile, ovaries luteinised, prolactinomas, metastatic mammary adenocarcinomas	Rulli <i>et al.</i> (2002)
Ubiquitin C promoter/hCGαβ	Males infertile, foetal Leydig cell adenomas, urethral and kidney defects; females infertile, ovarian teratomas	Rulli <i>et al.</i> (2003), Huhtaniemi <i>et al.</i> (2005), Ahtiainen <i>et al.</i> (2005)
MT-1 promoter/hCGβ	Males and females infertile; cystic and haemorrhagic ovaries	Matzuk et al. (2003)
MT-1 promoter/hCGαβ	Males infertile, Leydig cell hyperplasia; females infertile, cystic and haemorrhagic ovaries, cystic kidneys	Matzuk et al. (2003)
hCG-LH receptor complex	Male and female gonadal defects; altered hormone levels	Meehan et al. (2005)
Estrogen α receptor knockout	Males infertile, seminiferous tubules disrupted; females infertile, high LH; cystic and haemorrhagic ovaries	Schomberg et al. (1999)
Estrogen $\alpha\beta$ receptor knockout	High LH; males infertile; females infertile, follicle transdiferentiation to seminiferous tubules	Couse et al. (1999b)
Aromatase knockout	High testosterone, LH and FSH; males progressively infertile; females infertile, disrupted folliculogenesis	Britt <i>et al.</i> (2001)

MT-1, metallothionein 1.

The infertility of these mice was probably due to an obstruction in the ejaculatory tract that prevented the access of epididymal sperm to semen (Kumar *et al.* 1999).

The results obtained from the FSH overexpressing female mice resemble known human reproductive pathologies, in which ovarian cyst formation and haemorrhage are often associated with ovarian cancer in postmenopausal women or in patients with ovarian hyperstimulation syndrome. In contrast to females, there is no direct evidence that inadvertently elevated FSH levels can affect male fertility in humans.

In this respect, in an attempt to study the specific role of FSH in gonadal function independently of LH activity, a series of genetic mouse models was created by combining the gonadotrophin-deficient background of *hpg* mice with transgenic mice expressing the heterodimeric FSH, or a mutant human FSH receptor containing a single amino acid substitution (Asp⁵⁶⁷Gly) equivalent to activating mutations in related glycoprotein hormone receptors (Allan *et al.* 2004). These findings revealed that full Sertoli cell proliferation could be accomplished by FSH activity without LH. There were no obvious gonadal phenotypes on the normal mouse background, suggesting that, in these models, the transgene expression did not exceed the physiological FSH response.

Transgenic mice overexpressing LH

To address the impact of chronically elevated LH action on the reproductive system, different animal models with chronic hypersecretion of LH or hCG have been developed. Transgenic LHBCTP mice first reported by Risma et al. (1995) expressed a fusion gene of the coding sequence of bovine (b) LHB subunit fused in-frame with the C-terminal peptide sequence of the human chorionic gonadotrophin-β subunit (CTP), driven by the pituitaryspecific bovine glycoprotein- α subunit promoter. This transgene was targeted to the pituitary gonadotroph cells, and it achieved physiological levels of LH in the circulation, ranging from a 5- to 10-fold increase in females, but no apparent LH elevation occurred in transgenic males. Female LHBCTP mice presented with precocious puberty (Risma et al. 1997), and suffered from accelerated depletion of primordial follicles in the ovary (Flaws et al. 1997). These mice are infertile primarily due to chronic anovulation (Risma et al. 1995), which can be reversed by administration of an LH surge (Mann et al. 1999). In addition to this, the hormone imbalance of the LHBCTP females produces defects in uterine receptivity and induces mid-gestation pregnancy failure (Mann et al. 1999). Since these mice develop enlarged ovaries containing multiple cysts producing increased levels of testosterone and oestradiol, chronic hypersecretion of LH in the LHBCTP mice was proposed as a useful model to recapitulate PCOS in humans (Risma et al. 1995, Mann et al. 2003).

The majority of human ovarian tumours are epithelial in origin, to a lesser extent they are of sex-cord stromal or germ cell origin, and granulosa cell tumours are very rare (Amsterdam & Selvaraj 1997). Gonadotrophin hypersecretion has long been implicated in ovarian tumour development, especially based on epidemiological studies (Risch

1998), and recently, these data have been supported by studies in experimental animal models (Risma et al. 1995, Kananen et al. 1997, Kumar et al. 1999). In this respect, the LHBCTP female mice occasionally develop granulosa cell tumours in old age (Risma et al. 1995). However, it was later demonstrated that the nature of ovarian tumour formation was directly dependent on the genetic background of the mouse strain used for breeding, and this feature, i.e. the need for a specific genetic constellation, may also contribute to the rarity of these tumours in women. For instance, in the CF-1 outbred strain, elevated LH causes granulosa cell tumours by 5 months of age. It was suggested that the susceptibility to granulosa cell tumours is an oligogenic trait controlled by three unlinked genes (Keri et al. 2000). Hybrid mice, instead, displayed cystic ovaries with a highly luteinised phenotype reminiscent of luteomas of pregnancy (Keri et al. 2000). Interestingly, the same ovarian phenotype was developed when LHBCTP (CF-1) mice were treated with repetitive hCG surges, indicating that the ovulatory LH/hCG surges are able to prevent granulosa cell tumours, independently of the mouse strain used (Owens et al. 2002).

Kero et al. (2000) demonstrated that chronically elevated LH in female LHBCTP mice causes adrenal hyperplasia, and induces LHR expression in the adrenal gland, together with stimulation of corticosterone production. This effect appears to be dependent on ovarian hyperfunction, probably due to its polycystic condition, since gonadectomy abolishes the adrenal disturbances. These mice have a phenotype reminiscent of Cushing's syndrome, and they may provide a useful model to study human adrenal hyperfunction associated with chronically elevated gonadotrophin secretion, such as occurs in postmenopausal women or in those with PCOS.

Transgenic mice overexpressing hCG

Many endocrine disorders are attributed to excessive secretion of the hormone beyond physiological levels. Consequently, we have recently developed a transgenic mouse model that secretes pharmacological levels of hCG, although compared with humans not exceeding those occurring in pregnancy (Rulli et al. 2002, 2003). By intentionally exaggerating hCG production, we were able to recognise novel phenotypes both in males and females that could not be revealed by models displaying more moderate elevation of hormone secretion.

We first generated a transgenic mouse model bearing the hCGB subunit under the human ubiquitin promoter $(hCG\beta +)$, in which hCG is moderately overproduced in a large number of tissues (Rulli et al. 2002, 2003). By association with the endogenously expressed common α -subunit in the pituitary gland, bioactive hCG α/β dimers were produced and secreted into the circulation, achieving around a 40-fold increase in LH/hCG bioactivity in female animals (Rulli et al. 2002), but only a 3- to 4-fold increase in males (Rulli et al. 2003). In this model, the availability of the endogenous α -subunit becomes rate limiting in the dimerisation process of hCG, since the hCGB subunit is expressed in excess. Dimerisation is obligatory for hormonal activity, as the individual hCG subunits are devoid of bioactivity (Narayan et al. 2002). Consequently, the sexual dimorphism observed in the secreted bioactive forms of hCG would be attributed to a differential sex steroid feedback regulation of common α subunit expression at the hypothalamic-pituitary level, as was observed elsewhere (Risma et al. 1995).

The female phenotype of the $hCG\beta + mice$ presented with precocious puberty, disrupted oestrous cycles and infertility due to ovarian and uterine defects (Rulli et al. 2002). The ovaries appeared with occasional haemorrhagic cysts and looked massively luteinised, resembling luteomas. The presence of luteinised cells filled with lipid droplets suggested active steroid synthesis as a consequence of direct hCG hyperstimulation. Accordingly, high levels of oestradiol, testosterone and progesterone were produced from early stages of sexual development. An interesting phenotype emerging from this model was the development of pituitary adenomas and malignant mammary gland tumours in older age, which were strictly dependent on ovarian function, since ovariectomy prevented both effects despite persistent high levels of hCG. Female hCG + mice were hyperprolactinaemic and showed lactotroph hyperplasia followed by development of prolactinomas, probably due to overexposure to oestrogens during peripuberty, followed by persistently elevated levels of androgens as a source of locally produced oestrogens. Due to its luteotrophic properties (Freeman et al. 2000), the high prolactin level produced by the hyperplastic/tumorigenic pituitary may help to maintain the luteinised ovary in an active state of progesterone overproduction. In addition, the role of prolactin in mammary gland development and tumorigenesis has been established (Clevenger et al. 2003). Consequently, the indirectly orchestrated high hCG production, along with increased oestrogen, progesterone and prolactin levels, thus brought about mammary gland proliferation and differentiation of the $hCG\beta + mice$. The response originally resembled the lactating state of the gland, but was subsequently followed by the appearance of adenocarcinoma with metastatic properties in older age. Due to its complex and multistep hormonal dysregulation, this is a good animal model for the understanding of the hormone-dependent pathogenesis of the pituitary and mammary gland tumours.

The $hCG\beta$ + males were fertile and showed only a mild phenotype, in agreement with their moderately increased levels of bioactive hCG. These mice had smaller testes in the face of full spermatogenesis and normal sperm quality, thus failing to demonstrate adverse effects of hCG on male fertility (Rulli et al. 2003).

Besides the $hCG\beta$ + mouse with rather mild hormonal aberration, our interest was to achieve a mouse model that maintains higher levels of hCG, in order to analyse the consequences of clearly exaggerated response to hCG. We therefore generated a double transgenic mouse model harbouring both the common α - and hCG β -subunit transgenes under the same ubiquitin promoter (hCG $\alpha\beta$ + mice), by crossbreeding the hCG β + mice with another transgenic mouse line overexpressing α -subunit under the same promoter. These double transgenic mice are able to co-express both subunits in excess in different tissues, and to produce efficiently the dimeric form of hCG, reaching as high as 2000-fold levels, in terms of biological activity in the circulation, in both males and females (Rulli *et al.* 2003, SB Rulli, P Ahtiainen, M Poutanen and I Huhtaniemi, unpublished observations).

The hCGα-subunit overexpressing mice were normal and fertile, confirming the absence of biological effects of free gonadotrophin subunits. In contrast to the single $hCG\beta + model$, the double $hCG\alpha\beta + males$ were infertile and their reproductive organs were severely altered (Rulli et al. 2003). The first series of studies, performed in adulthood, showed smaller testes, enlarged seminal vesicles and prostate, dilated vasa deferentia and urinary bladder, as well as kidney defects. Testicular steroidogenesis was enhanced despite a clear down-regulation of LH/hCGR expression. In agreement with previous studies, based on long-term LH/hCG treatments in vivo (Risbridger et al. 1982, Gaytan et al. 1994), these mice developed focal Leydig cell hyperplasia/hypertrophy, but failed to promote testicular tumours. This was intriguing, since in humans, a specific activating mutation of the LHR (Asp⁵⁷⁸His) is associated with Leydig cell adenomas (Liu et al. 1999, Richter-Unruh et al. 2002). Interestingly, recent studies in young hCG $\alpha\beta$ + mice demonstrated postnatally clear Leydig cell adenomas of foetal Leydig cell origin, but these tumours disappeared at puberty (Ahtiainen et al. 2005). Whether the activation of alternative intracellular signalling pathways of the LHR, besides cAMP (Liu et al. 1999), explain the early Leydig cell tumorigenesis remains to be elucidated. An abnormal function of the accessory sex organs and the lower urinary tract was also evident in these males. Functional obstruction of vasa deferentia due to overproduction of secretory fluids or impaired emptying of the glands is the likely cause for the male infertility observed in this model. Progressive degenerative changes in the seminiferous epithelium and epididymides are associated with obstruction through a backpressure effect, and most of such changes may be due to severe steroid imbalance, in concordance with the male phenotype of oestrogen receptor-α knockout mice with elevated androgen levels (Eddy et al. 1996).

Interestingly, our recent investigations demonstrated that $hCG\alpha\beta+$ female mice with overexpression of excessive levels of hCG developed ovarian tumours that are phenocopies of human teratomas (Huhtaniemi, *et al.* 2005, SB Rulli, P Ahtiainen, M Poutanen and I Huhtaniemi, unpublished observations). This novel finding strengthens the applicability of this model for studying human diseases, since no other model has previously shown any linkage between ovarian teratomas and gonadotrophin action. It is well

known that these tumours derive from parthenogenetically activated oocytes within the ovary (Mutter 1997, Ulbright 2004). The mechanism that triggers the initial steps of tumour formation in our model is currently under investigation.

More recently, another transgenic model for hCG has been reported, where either one or both subunits of hCG were overexpressed using the mouse metallothionein promoter (Matzuk et al. 2003). In this model, hCGβ overexpressing females were infertile and progressively developed cystic ovaries, whereas males were infertile despite no discernible phenotype. In contrast, transgenic male mice co-expressing hCGα- and hCGβ-subunits showed multiple reproductive defects resembling those found in our previous model, such as infertility, Levdig cell hyperplasia, increased testosterone, reduced testis size and enlarged seminal vesicles. Double-transgenic females were infertile, had elevated oestradiol levels, and developed cystic ovaries with thecal layer enlargement and stromal cell proliferation, and degenerating kidneys (Matzuk et al. 2003). No evidence for tumorigenesis in gonadal or extragonadal tissue was reported in this model.

A transgenic mouse model expressing a constitutively active yoked hormone–receptor complex was recently generated, where hCG was covalently linked to the N-terminus of the rat LH receptor in a fusion protein (Meehan et al. 2005). Males exhibited prepubertal increases in testosterone levels and seminal vesicle weights, and decreases in serum FSH, LH, testis weight, and the size of the seminiferous tubules. Females presented with precocious sexual development and progressive ovarian lesions, from enhanced follicular development to degenerating follicles and haemorrhagic cysts.

Taken together, these studies indicate that chronically elevated hCG leads to multiple gonadal and extragonadal defects in males and females, including ovarian and testicular tumours as the primary effect, whereas the alterations found in the pituitary and mammary glands are due to secondary effects of the aberrant gonadal function. The distinct phenotypes emerging from the existing mouse models overexpressing hCG/LH *in vivo* may be related to the level of hCG/LH production, age, characteristics of the transgene expression, and the genetic background. However, the phenotypic similarities among the different models further emphasise the role of gonadotrophin action in reproductive pathophysiology.

Genetically modified mouse models with secondarily altered gonadotrophin secretion

Gonadotrophin synthesis and secretion is modulated by different steroidal and non-steroidal ovarian factors. Consequently, genetic mouse models directed to those factors can induce secondary gonadotrophin imbalance and display disturbed reproductive phenotypes similar to those found in gonadotrophin overexpressing mice. One typical example is the knockout mouse model for oestrogen receptor α (α ERKO), which has elevated LH levels due to the loss of oestrogen-mediated feedback regulation of the hypothalamic-pituitary-gonadal axis, and shows some similarities with the phenotypes of gonadotrophin overexpressing models (Schomberg et al. 1999). It was demonstrated that high LH levels are responsible for the defective ovarian phenotype in αERKO mice, since increased LH and polycystic follicle development are prevented by GnRH antagonist treatment (Couse et al. 1999a).

αERKO male mice are infertile due to abnormal fluid reabsortion in the epididymis, leading to disrupted spermatogenesis and seminiferous tubule organisation through a backpressure effect (Eddy et al. 1996). These mice exhibit elevated serum testosterone levels, but LH and FSH levels are not significantly different from those of wild-type males. These results are in line with the male phenotype of hCG $\alpha\beta$ + mice (Rulli et al. 2003), in which elevated androgen levels may be responsible for the defective phenotype of the male reproductive system.

The aromatase knockout model (ArKO) is characterised by lack of oestrogen production, which induces testosterone, LH and FSH levels in circulation. Similarly to αERKO, the ArKO mice are infertile due to disrupted folliculogenesis and failure to ovulate, and develop haemorrhagic cystic follicles (Britt et al. 2001). The ovarian phenotype of the ArKO mouse was then attributed to the altered hormonal milieu, where the conversion of androgens to oestrogens is blocked in the presence of elevated gonadotrophins.

Concluding remarks

Application of new technologies for the development of experimental animal models together with the characterization of human phenotypes with genetic defects have provided a great deal of novel information on less well known mechanisms involved in pathological effects of gonadotrophin function in both sexes. The different transgenic mouse models with gonadotrophin overexpression available in the literature are increasing in number and reveal marked differences and similarities in their phenotypes (Table 1). As a consensus from most of the models, female fertility and ovarian physiology are particularly vulnerable to alterations of gonadotrophic action, mainly manifested by different levels of hormone profile disruption, and development of cystic and/or tumorigenic ovaries as the primary effect. In the most severe cases, secondary gonad-dependent phenotypes, such as urinary tract disturbances and extra-gonadal tumour development (mammary and pituitary gland tumours, adrenal lesions) are also observed. The male phenotypes in the different models appear much milder in terms of fertility and gonadal tumour development. However, discrete phenotypes of the male reproductive tract also occur.

Taken together, these 'lessons' from transgenic mice provide important evidence for the impact of gonadotrophin hypersecretion on the gonadal function and its relation with human pathologies. The novel findings clearly advance our knowledge beyond that obtained previously in experiments using various gondotrophin treatments or ablations. Since many aspects of reproductive pathophysiology are not completely understood, and new questions are constantly arising, additional studies providing more detailed information are needed in the future.

Acknowledgements

The authors declare that there is no conflict of interest that would prejudice the impartiality of this scientific work.

References

- Ahtiainen P, Rulli S, Pelliniemi LJ, Toppari J, Poutanen M & Huhtaniemi I 2005 Fetal but not adult Leydig cells are susceptible to adenoma formation in response to persistently high hCG level; a study on hCG overexpressing transgenic mice. Oncogene In Press.
- Allan CM, Garcia A, Spaliviero J, Zhang FP, Jimenez M, Huhtaniemi I & Handelsman DJ 2004 Complete Sertoli cell proliferation induced by follicle-stimulating hormone (FSH) independently of luteinizing hormone activity: evidence from genetic models of isolated FSH action. Endocrinology 145 1587-1593.
- Amsterdam A & Selvaraj N 1997 Control of differentiation, transformation, and apoptosis in granulosa cells by oncogenes, oncoviruses, and tumour suppressor genes. Endocrine Reviews 18 435-461.
- Ascoli M, Fanelli F & Segaloff DL 2002 The lutropin/choriogonadotropin receptor, a 2002 perspective. Endocrine Reviews 23 141–174.
- Britt KL, Drummond AE, Dyson M, Wreford NG, Jones ME, Simpson ER & Findlay JK 2001 The ovarian phenotype of the aromatase knockout (ArKO) mouse. Journal of Steroid Biochemistry and Molecular Biology 79 181-185.
- Burger LL, Haisenleder DJ, Dalkin AC & Marshall JC 2004 Regulation of gonadotropin subunit gene transcription. Journal of Molecular Endocrinology 33 559-584.
- Burns KH & Matzuk MM 2002 Minireview: genetic models for the study of gonadotropin actions. Endocrinology 143 2823-2835.
- Clevenger CV, Furth PA, Hankinson SE & Schuler LA 2003 The role of prolactin in mammary carcinoma. Endocrine Reviews 24 1-27.
- Cole LA & Butler S 2002 Detection of hCG in trophoblastic disease. The USA hCG reference service experience. Journal of Reproductive Medicine 47 433-444.
- Conti M, Harwood JP, Hsueh AJ, Dufau ML & Catt KJ 1976 Gonadotropin-induced loss of hormone receptors and desensitization of adenylate cyclase in the ovary. Journal of Biological Chemistry 251
- Couse JF, Bunch DO, Lindzey J, Schomberg DW & Korach KS 1999a Prevention of the polycystic ovarian phenotype and characterization of ovulatory capacity in the estrogen receptor-alpha knockout mouse. Endocrinology 140 5855-5865.
- Couse JF, Hewitt SC, Bunch DO, Sar M, Walker VR, Davis BJ & Korach KS 1999b Postnatal sex reversal of the ovaries in mice lacking estrogen receptors alpha and beta. Science 286 2328-2331.
- Delvigne A & Rozenberg S 2002 Epidemiology and prevention of ovarian hyperstimulation syndrome (OHSS): a review. Human Reproduction Update 8 559-577.
- Dufau ML, Cigorraga S, Baukal AJ, Sorrell S, Bator JM, Neubauer JF & Catt KJ 1979 Androgen biosynthesis in Leydig cells after testicular desensitization by luteinizing hormone-releasing hormone and human chorionic gonadotropin. Endocrinology 105 1314–1321.
- Eddy EM, Washburn TF, Bunch DO, Goulding EH, Gladen BC, Lubahn DB & Korach KS 1996 Targeted disruption of the estrogen receptor gene in male mice causes alteration of spermatogenesis and infertility. Endocrinology 137 4796-4805.
- Fevold SL, Hisaw FL & Leonard SL 1931 The gonad-stimulating and the luteinizing hormones of the anterior lobe of the hypophysis. American Journal of Physiology 97 291–301.

- Filicori M, Fazleabas AT, Huhtaniemi I, Licht P, Rao ChV, Tesarik J & Zygmunt M 2005 Novel concepts of human chorionic gonadotropin; reproductive system interactions and potentials in the management of infertility. Fertility and Sterility In Press.
- Flaws JA, Abbud R, Mann RJ, Nilson JH & Hirshfield AN 1997 Chronically elevated luteinizing hormone depletes primordial follicles in the mouse ovary. *Biology of Reproduction* **57** 1233–1237.
- **Franks S & McCarthy M** 2004 Genetics of ovarian disorders: polycystic ovary syndrome. *Reviews of Endocrine and Metabolic Disorders* **5** 69–76.
- Freeman ME, Kanyicska B, Lerant A & Nagy G 2000 Prolactin: structure, function, and regulation of secretion. *Physiology Reviews* **80** 1523–1631.
- Gaytan F, Pinilla L, Romero JL & Aguilar E 1994 Differential effects of the administration of human chorionic gonadotropin to postnatal rats. *Journal of Endocrinology* 142 527–534.
- **Gnagy S**, **Ming EE**, **Devesa SS**, **Hartge P & Whittemore AS** 2000 Declining ovarian cancer rates in US women in relation to parity and oral contraceptive use. *Epidemiology* **11** 102–105.
- **Gromoll J, Simoni M & Nieschlag E** 1996 An activating mutation of the follicle-stimulating hormone receptor autonomously sustains spermatogenesis in a hypophysectomized man. *Journal of Clinical Endocrinology and Metabolism* **81** 1367–1370.
- **Huhtaniemi IT & Themmen APN** 2005 Mutations in human gonadotropin and gonadotropin receptor genes. *Endocrine* In Press.
- Huhtaniemi I, Zhang FP, Kero J, Hamalainen T & Poutanen M 2002 Transgenic and knockout mouse models for the study of luteinizing hormone and luteinizing hormone receptor function. *Molecular and Cellular Endocrinology* 87 49–56.
- **Huhtaniemi I, Rulli S, Ahtiainen P & Poutanen M** 2005 Multiple sites of tumorigenesis in transgenic mice overproducing hCG. *Molecular and Cellular Endocrinology* **234** 117–126.
- Huirne JA & Lambalk CB 2001 Gonadotropin-releasing hormonereceptor antagonists. Lancet 358 1793–1803.
- **Jameson JL & Hollenberg AN** 1993 Regulation of chorionic gonadotropin gene expression. *Endocrine Reviews* **14** 203–221.
- Kananen K, Rilianawati, Paukku T, Markkula M, Rainio EM & Huhtanemi IT 1997 Suppression of gonadotropins inhibits gonadal tumorigenesis in mice transgenic for the mouse inhibin α-subunit promoter/simian virus 40 T antigen fusion gene. Endocrinology 138 3521–3531.
- Keri RA, Lozada KL, Abdul-Karim FW, Nadeau JN & Nilson JH 2000 Luteinizing hormone induction of ovarian tumors: oligogenic differences between mouse strains dictates tumor disposition. PNAS 97 383–387.
- Kero J, Poutanen M, Zhang FP, Rahman N, McNicol AM, Nilson JH, Keri RA & Huhtaniemi IT 2000 Elevated luteinizing hormone induces expression of its receptor and promotes steroidogenesis in the adrenal cortex. *Journal of Clinical Investigation* 105 633–641.
- Konishi I, Kuroda H & Mandai M 1999 Review: gonadotropins and development of ovarian cancer. *Oncology* **57** 45–48.
- Kumar TR, Wang Y & Matzuk MM 1996 Gonadotropins are essential modifier factors for gonadal tumor development in inhibin-deficient mice. *Endocrinology* 137 4210–4216.
- Kumar TR, Palapattu G, Wang P, Woodruff TK, Boime I, Byrne MC & Matzuk MM 1999 Transgenic models to study gonadotropin function: the role of follicle-stimulating hormone in gonadal growth and tumorigenesis. *Molecular Endocrinology* 13 851–886.
- **Kumar TR** 2005 What have we learned about gonadotropin function from gonadotropin subunit and receptor mice? *Reproduction* **130** 293–302.
- La Vecchia C 2001 Epidemiology of ovarian cancer: a summary review. European Journal of Cancer Prevention 10 125–129.
- **Liu G**, **Duranteau L**, **Carel JC**, **Monroe J**, **Doyle DA & Shenker A** 1999 Leydig-cell tumors caused by an activating mutation of the gene encoding the luteinizing hormone receptor. *New England Journal of Medicine* **341** 1731–1736.

- Mann RJ, Keri RA & Nilson JH 1999 Transgenic mice with chronically elevated luteinizing hormone are infertile due to anovulation, defects in uterine receptivity, and midgestation pregnancy failure. *Endocrinology* **140** 2592–2601.
- Mann RJ, Keri RA & Nilson JH 2003 Consequences of elevated luteinizing hormone on diverse physiological systems: use of the LHbetaCTP transgenic mouse as a model of ovarian hyperstimulation-induced pathophysiology. *Recent Progress in Hormone Research* **58** 343–375.
- Marshall JC, Eagleson CA & McCartney CR 2001 Hypothalamic dysfunction. *Molecular and Cellular Endocrinology* **183** 29–32.
- Matzuk MM, Finegold MJ, Su JG, Hsueh AJ & Bradley A 1992 Alpha-inhibin is a tumour-suppressor gene with gonadal specificity in mice. *Nature* **360** 313–319.
- Matzuk MM, DeMayo FJ, Hadsell LA & Kumar TR 2003 Overexpression of human chorionic gonadotropin causes multiple reproductive defects in transgenic mice. *Biology of Reproduction* 69 338–346.
- Meehan TP, Harmon BG, Overcast ME, Yu KK, Camper SA, Puett D & Narayan P 2005 Gonadal defects and hormonal alterations in transgenic mice expressing a single chain human chorionic gonadotropin–lutropin receptor complex. *Journal of Molecular Endocrinology* 34 489–503.
- Mikola M, Kero J, Nilson JH, Keri RA, Poutanen M & Huhtaniemi I 2003 High levels of luteinizing hormone analog stimulate gonadal and adrenal tumorigenesis in mice transgenic for the mouse inhibin-alpha-subunit promoter/Simian virus 40 T-antigen fusion gene. *Oncogene* 22 3269–3278.
- Mutter GL 1997 Role of imprinting in abnormal human development. Mutation Research 396 141–147.
- Narayan P, Gray J & Puett D 2002 Yoked complexes of human choriogonadotropin and the lutropin receptor: evidence that monomeric individual subunits are inactive. *Molecular Endocrinology* 16 2733–2745.
- Owens GE, Keri RA & Nilson JH 2002 Ovulatory surges of hCG prevent hormone-induced granulosa cell tumor formation leading to the identification of tumor-associated changes in the transcriptome. *Molecular Endocrinology* 16 1230–1242.
- **Quigley CA** 2002 Editorial: the postnatal gonadotropin and sex steroid surge insights from the androgen insensitivity syndrome. *Journal of Clinical Endocrinology and Metabolism* **87** 24–28.
- Pakarainen T, Zhang F-P, Poutanen M & Huhtaniemi I 2005 Fertility in LH receptor knockout mice after wild-type ovary transplantation demonstrates redundancy of extragonadal LH action. *Journal of Clinical Investigation* In Press.
- Richter-Unruh A, Wessels HT, Menken U, Bergmann M, Schmittmann-Ohters K, Schaper J, Tappeser S & Hauffa BP 2002 Male LH-independent sexual precocity in a 3.5-year-old boy caused by a somatic activating mutation of the LH receptor in a Leydig cell tumor. *Journal of Clinical Endocrinology and Metabolism* 87 1052–1056.
- Riman T, Nilsson S & Persson IR 2004 Review of epidemiological evidence for reproductive and hormonal factors in relation to the risk of epithelial ovarian malignancies. *Acta Obstetrica et Gynecologica Scandinavica* 83 783–795.
- Risbridger GP, Robertson DM & de Kretser DM 1982 The effects of chronic human chorionic gonadotropin treatment on Leydig cell function. *Endocrinology* 110 138–145.
- **Risch HA** 1998 Hormonal etiology of epithelial ovarian cancer, with a hypothesis concerning the role of androgens and progesterone. *Journal of the National Cancer Institute* **90** 1774–1786.
- Risma KA, Clay CM, Nett TM, Wagner T, Yun J & Nilson JH 1995 Targeted overexpression of luteinizing hormone in transgenic mice leads to infertility, polycystic ovaries, and ovarian tumors. *PNAS* **92** 1322–1326.
- Risma KA, Hirshfield AN & Nilson JH 1997 Elevated luteinizing hormone in prepubertal transgenic mice causes hyperandrogenemia, precocious puberty, and substantial ovarian pathology. *Endocrinology* 138 3540–3547.

- Roberts JE, Spandorfer S, Fasouliotis SJ, Lin K & Rosenwaks Z 2005 Spontaneous ovarian hyperstimulation caused by a follicle-stimulating hormone-secreting pituitary adenoma. Fertility and Sterility 83 208-210.
- Rulli SB, Kuorelahti Al, Karaer O, Pelliniemi L, Poutanen M & Huhtaniemi IT 2002 Reproductive disturbances, pituitary lactotrope adenomas, and mammary gland tumors in transgenic female mice producing high levels of human chorionic gonadotropin. Endocrinology 143 4084-4095.
- Rulli SB, Ahtiainen P, Makela S, Toppari J, Poutanen M & Huhtaniemi I 2003 Elevated steroidogenesis, defective reproductive organs, and infertility in transgenic male mice overexpressing human chorionic gonadotropin. Endocrinology 144 4980-4990.
- Salbenblatt JA, Bender BG, Puck MH, Robinson A, Faiman C & Winter JS 1985 Pituitary-gonadal function in Klinefelter syndrome before and during puberty. Pediatric Research 19 82-86.
- Schomberg DW, Couse JF, Mukherjee A, Lubahn DB, Sar M, Mayo KE & Korach KS 1999 Targeted disruption of the estrogen receptoralpha gene in female mice: characterization of ovarian responses and phenotype in the adult. Endocrinology 140 2733-2744.
- Scott IS, Charlton HM, Cox BS, Grocock CA, Sheffield JW & O'Shaughnessy PJ 1990 Effect of LH injections on testicular steroidogenesis, cholesterol sidechain cleavage P450 mRNA content and Leydig cell morphology in hypogonadal mice. Journal of Endocrinology 125 131-138.
- Simoni M, Gromoll J & Nieschlag E 1997 The follicle-stimulating hormone receptor: biochemistry, molecular biology, physiology, and pathophysiology. Endocrine Reviews 18 739-773.
- Smith PE & Engle ET 1927 Experimental evidence of the role of anterior pituitary in development and regulation of gonads. American Journal of Anatomy 40 159-161.

- Smits G, Olatunbosun O, Delbaere A, Pierson R, Vassart G & Costagliola \$ 2003 Ovarian hyperstimulation syndrome due to a mutation in the follicle-stimulating hormone receptor. New England Journal of Medicine 349 760-766.
- Stenman UH, Alfthan H & Hotakainen K 2004 Human chorionic gonadotropin in cancer. Clinical Biochemistry 37 549-561.
- Strauss BL, Pittman R, Pixley MR, Nilson JH & Boime I 1994 Expression of the beta subunit of chorionic gonadotropin in transgenic mice. Journal of Biological Chemistry 269 4968-4973.
- Themmen APN & Huhtaniemi IT 2000 Mutations of gonadotropins and gonadotropin receptors: elucidating the physiology and pathophysiology of pituitary-gonadal function. Endocrine Reviews 21 551-583
- Themmen APN 2005 An update of the pathophysiology of human gonadotrophin subunit and receptor mutations and polymorphisms. Reproduction **130** 263-274.
- Ulbright TM 2004 Gonadal teratomas: a review and speculation. Advances in Anatomy and Pathology 11 10-23.
- Vasseur C, Rodien P, Beau I, Desroches A, Gerard C, de Poncheville L, Chaplot S, Savagner F, Croue A, Mathieu E, Lahlou N, Descamps P & Misrahi M 2003 A chorionic gonadotropin-sensitive mutation in the follicle-stimulating hormone receptor as a cause of familial gestational spontaneous ovarian hyperstimulation syndrome. New England Journal of Medicine 349 753-759.

Received 9 May 2005 First decision 10 June 2005 Accepted 30 June 2005