



Article MIF rs755622 and IL6 rs1800795 Are Implied in Genetic Susceptibility to End-Stage Renal Disease (ESRD)

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Abstract: Chronic kidney disease (CKD) is characterized by an increased risk of kidney failure and end-stage renal disease (ESRD). Aging and comorbidities as cardiovascular diseases, metabolic disorders, infectious diseases, or tumors, might increase the risk of dialysis. In addition, genetic susceptibility factors might modulate kidney damage evolution. We have analyzed, in a group of ESRD patients and matched controls, a set of SNPs of genes (Klotho rs577912, rs564481, rs9536314; FGF23 rs7955866; IGF1 rs35767; TNFA rs1800629; IL6 rs1800795; MIF rs755622, rs1007888) chosen in relation to their possible involvement with renal disease and concomitant pathologies. Analysis of the raw data did indicate that IL6 rs180795 and MIF rs755622 SNPs might be markers of genetic susceptibility to ESRD. In particular, the C positive genotypes of MIF rs755622, (dominant model) seem to be an independent risk factor for ESDR patients (data adjusted for age, gender, and associated pathologies). Stratifying results according to age MIF rs755622 C positive genotype frequencies are increased in both the two age classes considered (<59 and ≥ 59 -year-old subjects). Analyses of data according to gender allowed us to observe that ESRD women shoved a significantly reduced frequency of genotypes bearing IL6 rs180795 C allele. In addition, MIF rs755622 might interact with diabetes or hypercholesterolemia in increasing susceptibility to ESRD. In conclusion, our data indicate that some polymorphisms involved in the regulation of both renal function and inflammatory response can influence the evolution of chronic kidney disease and suggest that the modulation of the activities of these and other genes should also be considered as therapeutic targets on to intervene with innovative therapies.

Keywords: MIF; IL6; SNP; CKD; ESRD

1. Introduction

Chronic kidney disease (CKD) is characterized by a progressive decrease of renal function with an increased risk of kidney failure and end-stage renal disease (ESRD), the disease stage where dialysis and transplantation are needed [1]. The National Kidney Foundation's Kidney Disease Outcomes Quality Initiative (K/DOQI) has defined the stage of chronic kidney disease (CKD) based on the value of glomerular filtration rate (GFR) [2]. According to this classification, a GFR \leq 59 mL/min/1.73 m² or less defines a moderate to severe decrease in renal function.

In recent years, many studies have been carried out aimed at identifying the molecules to be used as the most sensitive biomarkers of kidney function, many of which are proteins



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Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). produced by different segments of the nephrons, cytokines, receptors that might be implied in the kidney acute or chronic damage [3,4]. In addition, genes coding for these mediators are highly polymorphic and the effect of their interactions on the pathogeneses of different diseases might be influenced by the different individual genetic backgrounds [5].

Recently, the scenario of possible biomarkers in chronic renal failure has been enriched by new players such as Fibroblast Growth Factor-23 (FGF-23) and Klotho. The progressive deterioration of renal function is characterized by a progressive reduction of Klotho protein and an increase of the FGF-23 levels [6–8].

Klotho has anti-inflammatory and anti-fibrotic activity, preventing vascular calcifications. In humans, the circulating levels of Klotho decrease with age in serum, and polymorphisms of the *Klotho* gene related to premature aging have also been identified [9,10].

Circulating Klotho can be considered an early diagnostic marker of CKD. Already for estimated glomerular filtration rate (eGFR) values below 90 mL/min, there is a progressive reduction. Clinical and experimental studies have shown that a significant decrease in *Klotho* expression in the kidney has a direct correlation with the reduction of GFR-23 in CKD subjects [11]. High levels of FGF-23 and reduction of circulating Klotho levels in CKD correlate with a variety of clinical conditions such as left ventricular hypertrophy, vascular calcifications, and even an increased risk of mortality [12] in CKD patients. In this scenario, the SNPs of these genes can play a role.

In particular, rs577912 polymorphism of the Klotho gene sequence might affect the expression of the protein and therefore its serum levels. The CC genotype is associated with a 16-21% lower Klotho expression compared to the AA/AC genotype with an association with an increased risk of mortality at 1 year from the start of dialysis [13]. In addition, the SNP rs7955866 in exon 3 of FGF23 has been related to renal damage [14]. The minor allele of this SNP induces a missense mutation (T239M) with a conformational change in the beta-sheets of the protein. Human embryonic kidney cells transfected in vitro with the *FGF23-239M* plasmid release higher concentrations of FGF-23 [14]. On the other hand, Klotho polymorphisms involved in metabolic disorders and cardiovascular pathologies might play a role in favoring progression to ESRD. In particular, rs564481 and rs9536314 might play a role, being implied in metabolic diseases and potentially influencing the prognosis of CKD [15–18]. In particular, rs56481 might influence hypertension susceptibility and creatinine serum levels in different populations [15–17] whereas rs9536314 might influence the kidney failure susceptibility impinging on the kidney function decline due to aging. Actually, a pathophysiological modification associated with chronic kidney disease mimics that of elderly individuals, which underlies shared clinical characteristics such as an increased risk of infection and atherosclerosis [18,19]. Moreover, rs9536314 was the polymorphism most frequently involved in association studies on cardiovascular and non-cardiovascular mortality. This may be particularly relevant in patients with chronic kidney disease who have decreased Klotho levels in tissues and blood [20].

More recently, the interactions between the Klotho/FGF-23 system and the growth hormone (GH)/insulin-like growth factor-1 (IGF-1) axis have been analyzed. The GH/IGF-1 system appears relevant for glomerular and tubular function, as well as the synthesis of Klotho [21]. On the other hand, beneficial effects of IGF-1 in CKD are antagonized by Suppressor of Cytokine Signaling, due to CKD-associated chronic inflammation that might be responsible for GH insensitivity observed [22] and could be modulated by SNPs of *IGF1* gene [23,24]. Moreover, as reported in a recent meta-analysis [25], *IGF1 rs35767* increases the risk of renal disease.

Actually, the chronic inflammation state that characterizes CKD and ESRD induces an up-regulation of proinflammatory cytokines and the associated biomarkers (interleukin (IL)-1 β , IL-1 receptor antagonist, IL-6, tumor necrosis factor (TNF)- α , C-reactive protein, and fibrinogen) [26,27].

Proinflammatory markers are shown to be important in the development of atherosclerotic complications and inflammation. In addition, it has been associated with hypertension in both experimental animal models and human studies [28–32], however, due to the complexity of this relationship, the mechanisms between inflammation and vascular involvement are still unclear.

On the other hand, an aging kidney can be considered a paradigmatic model of immunosenescence characterized by defective immune responses and increased systemic inflammation. In the kidney, resident macrophages and fibroblasts are continuously exposed to environmental stimuli, and the effects of cellular reprogramming induced by local immune responses, which accumulate with age, might have a role in the increased susceptibility to kidney disease among elderly individuals [19,33] and premature aging [34]. Key proinflammatory cytokines have been analyzed by different research groups both as circulating markers and as components of the genetic background predisposing to CKD. Cytokines of the TNF family [35–37], *TNF* SNPs [38,39], IL-6 serum levels [40,41], and *IL6* gene SNPs [42–44], as well as macrophage inhibiting factor (MIF) and *MIF* gene variants [45–48], might play a role in favoring kidney damage and have been studied. In particular, the *rs1800629* and *rs1800795* polymorphisms of *TNFA* and *IL6* genes can selectively affect spontaneous production of inflammatory cytokines influencing kidney function decrease and consequent development and progression of chronic kidney disease.

Herein, data on some genetic variants of key cytokine, mediator, and receptors involved in the complex scenario of kidney chronic failure have been reported and their relevance as risk profile markers useful in the prediction of kidney disease development and progression have been evaluated.

2. Materials and Methods

2.1. Patients and Controls

In collaboration with the Department of Nephrology and Dialysis with C.R.R. for arterial hypertension, "Paolo Giaccone" University Hospital, a population of 93 patients with end-stage renal chronic failure (ESRD) who were routinely undergoing dialysis therapy was recruited. Two control populations, for a total of 213 individuals, were recruited, one of healthy young people and one of the people over 59 years old, at the Department of Medical Biotechnology of the University of Palermo and in collaboration with the U.O.C of Transfusion Medicine of the Paolo Giaccone" University Hospital. The cut-off for age between the two control population was chosen on the basis of the median age of ESRD patients. The \geq 59-year-old control population included both healthy subjects and people with hypercholesterolemia, type 2 diabetes, or hypertension without renal impairment (glomerular filtration rate, GFR \geq 90 mL/min). Persons with autoimmune or tumor pathologies or with GFR < 90 were excluded from the control group. The clinical and demographic characteristics of the populations studied are shown in Table 1.

Demographic	ES	RD	Controls		<59-Year-Old Controls		≥59-Year-Old Controls		p *
and Clinical Characteristics	N.	%	N.	%	N.	%	N.	%	-
Age, mean \pm SD	69.01	± 13.02	55.97	± 10.99	43.75	± 5.46	66.32	± 5.24	0.1118
>59 years old	71	76.3	67	31.45	-	-	67	31.45	-
Women	42	45.16	94	44.13	64	43.83	30	44.78	1.000
				Pathologies	in ESRD and	d Controls			
Hypertension	32	34.41	17	8.01	-	-	17	8.01	< 0.0001
Type II Diabetes	19	20.43	8	3.76	-	-	8	3.76	< 0.0001
Hypercholesterolemia	13	13.98	21	9.85	-	-	21	9.85	0.3242

Table 1. Demographic and clinical characteristics of 93 patients with end-stage renal disease (ESRD) and 213 control subjects.

* analyses of significant differences between ESRD patients and \geq 59-year-old controls.

Our study was performed in accordance with ethical standards of the Helsinki Declaration of the World Medical Association and Italian legislation, and was approved by the local institutional review board (Comitato Etico Palermo 1, protocol code CET1 04/2020, date of approval 22 April 2020). All participants gave their informed consent and the data has been encoded to ensure the protection of patient privacy and controls. The EDTA

peripheral blood samples used for molecular typing respectively were collected and stored at 70 $^{\circ}\mathrm{C}$ until their use.

2.2. SNP Genotyping

DNA were extracted using a salting-out protocol and DNA quality was verified by 260/280 nm absorbance ratio and gel electrophoresis as previously reported [49]. SNP typing was performed using an on-demand assay developed by KBioscience Ltd. (Middlesex, UK) and based on homogeneous FRET (Fluorescence Resonance Energy Transfer) analysis on the products of a specific allele PCR (Kaspar) [50,51]. SNPs are listed in Table 2. Information on these polymorphisms was acquired from dbSNP NCBI (https://www.ncbi.nlm.nih.gov/snp/, accessed on 18 January 2022).

Gene	SNP	Major Allele	Minor Allele	Chr Location
	rs577912	С	А	13:33036014
Klotho	rs564481	С	Т	13: 33060846
	rs9536314	Т	G	13:33054001
FGF23	rs7955866	G	А	12:4370383
IGF1	rs35767	С	Т	12:102481791
TNFA	rs1800629	G	А	6:31575254
IL6	rs1800795	G	С	7:22727026
	rs755622	G	С	22:23894205
IVIIF	rs1007888	Т	С	22:23898914

Table 2. Typed SNPs.

2.3. Statistical Analysis

Allele and genotype frequencies were evaluated by gene count. Pearson's test was applied to test the Hardy–Weinberg equilibrium. Power of calculation has been evaluated for all comparisons made using an online tool (https://sample-size.net/, accessed on 17 February 2022). Significant differences in genotype distributions between groups were calculated using chi-square or Fisher's exact test applied to the appropriated 2×2 or 3×2 contingency tables. Multiple logistic regression models were applied using overdominant (homozygous major allele and/or homozygous minor allele genotypes versus heterozygous genotype), codominant (heterozygous genotype versus homozygous major allele plus homozygous minor allele genotypes), dominant (homozygous plus heterozygous minor allele genotypes versus homozygous major allele genotype), and recessive (homozygous minor allele genotype versus homozygous major allele plus heterozygous genotypes) of heredity and adjusting the results for age, sex, and concomitance of existing pathologies. The choice of the most suitable model was based on the Bayesian Information Criterion (BIC) and the Akaike Information Verification Test (AIC). Odds ratio (OR) values and 95% confidence intervals (95% CI) were calculated and a p-value < 0.05 was considered statistically significant. Statistical analyzes were conducted using version 3.06 of Graph-PadInStat software (GraphPad, San Diego, CA, USA) and online statistical analysis tools (https://www.snpstats.net/start.htm, accessed on 15 November 2021) devoted to the evaluation of the association of SNPs with diseases in the presence of other biological, genetic, or clinical variances.

3. Results

In our study we analyzed two populations of ESRD patients and controls, assessed in relation to demographic and clinical characteristics. As reported above, SNPs were chosen in relation to their possible involvement with renal disease and concomitant pathologies. Table 3 shows the allele and genotype frequencies in the distribution of single nucleotide polymorphisms, between subjects with chronic renal failure on dialysis therapy and controls.

	() ID	Alleles/	Con	itrols	ES	RD	OR	1 1 1
GENE	SNP	Genotypes	Nr	Freq.	Nr	Freq.	(95% CI)	¹ <i>p</i> -Value
		С	360	0.85	162	0.87		0.450
		А	66	0.15	24	0.13	1.04	0.458
	rs577912	CC	150	0.7	71	0.76	1.24	
		CA	60	0.28	20	0.22	(0.75–2.05)	0.442
		AA	3	0.01	2	0.02		
		С	262	0.62	113	0.61		0.055
		Т	164	0.38	73	0.39	0.07	0.857
Klotho	rs564481	CC	82	0.38	34	0.37	0.97	
		СТ	98	0.46	45	0.48	(0.68–1.38)	0.927
		TT	33	0.15	14	0.15		
		Т	355	0.83	154	0.83		0.007
		G	71	0.17	32	0.17	0.07	0.907
	rs9536314	TT	153	0.72	63	0.68	0.96	
		TG	49	0.23	28	0.3	(0.61–1.52)	0.242
		GG	11	0.05	2	0.02		
		G	380	0.89	168	0.9		0 77 4
		А	46	0.11	18	0.1	1 10	0.774
FGF23	rs7955866	GG	175	0.82	77	0.83	1.13	
10120 10100000		GA	30	0.14	14	0.15	(0.64 - 2.01)	0.757
		AA	8	0.04	2	0.02		
		С	348	0.82	150	0.81		0.000
		Т	78	0.18	36	0.19	0.02	0.822
IGF1	rs35767	CC	146	0.69	64	0.69	0.93	
		СТ	56	0.26	22	0.24	(0.60–1.45)	0.673
		TT	11	0.05	7	0.08		
		G	388	0.91	176	0.95		0.145
		А	38	0.09	10	0.05	1 70	
TNFA	rs1800629	GG	178	0.84	85	0.91	1.72	
		GA	32	0.15	6	0.06	(0.64-5.54)	0.105
		AA	3	0.01	2	0.02		
		G	311	0.73	152	0.82		0.024
		С	115	0.27	34	0.18	0.61	
IL6	rs1800795	GG	118	0.55	65	0.70	(0.20, 0.02)	
		GC	75	0.35	22	0.24	(0.39 - 0.93)	0.059
		CC	20	0.09	6	0.06		
		G	301	0.71	108	0.58		0.0028
		С	125	0.29	78	0.42	1 83	
MIE	rs755622	GG	128	0.60	42	0.45	(1 12 2 00)	
		GC	45	0.21	24	0.26	(1.12–2.99)	0.042
		CC	40	0.19	27	0.29		
.,		Т	231	0.54	107	0.58		0.480
		С	195	0.46	79	0.42	1 1 /	
	rs1007888	TT	70	0.33	34	0.37	1.14	
		TC	91	0.43	39	0.42	(0.01-1.02)	0.778
		CC	52	0.24	20	0.22		

Table 3. SNP allelic and genotypic frequencies of 93 dialyzed patients (ESRD) and 213 control subjects.

¹ Evaluation of SNP alleles and genotypes, significant differences between ESRD and controls (crude analyses) was performed using the chi-square test or Fisher's exact test applied to 2×2 or 3×2 contingency tables.

The statistical analysis of the raw data did indicate that the *IL6 rs180795* and *MIF rs755622* SNPs crude frequencies of the patient group were significantly different with respect to the control group in particular *rs180795* C allele frequency, associated to reduced production of IL-6 is significantly reduced in ESRD patients whereas *rs755622* G positive genotypes is increased.

We then proceeded to apply logistic regression models, using codominant, dominant, and recessive models (chosen on the basis of BIC and AIC criteria) of inheritance and adjusting the results for biological variables (age, sex) and concomitant comorbidities (hypertension, diabetes, hypercholesterolemia).

The best results of multivariate analysis, applied to each of the nine selected SNPs (Table 4), allowed us to confirm that C positive genotypes of *MIF rs755622*, (dominant model) is an independent risk factor for ESDR patients (independent of age, gender, and associated pathologies). In addition, a rough significant influence of both *TNFA rs1800629* homozygous genotypes have been detected (complete model analyses for each SNP are reported in Supplementary Tables S1–S9).

Table 4. Evaluation of SNP associations with ESRD (adjusted by age, gender, hypertension, diabetes, and hypercholesterolemia).

Gene	SNP	Genetic Models	OR (95% CI)	<i>p</i> -Value
	rs577912	Dominant	0.61 (0.32-1.20)	0.15
Klotho	rs564481	Dominant	1.09 (0.60-1.98)	0.77
	rs9536314	Recessive	0.25 (0.05-1.29)	0.067
FGF23	rs7955866	Recessive	0.90 (0.16-4.98)	0.9
IGF1	rs35767	Recessive	1.27 (0.39-4.12)	0.69
TNFA	rs1800629	Overdominant	0.35 (0.12-1.06)	0.046
IL6	rs1800795	Dominant	0.76 (0.42-1.39)	0.37
MIF	rs755622 rs1007888	Dominant Dominant	3.37 (1.80–6.30) 0.87 (0.48–1.58)	<0.0001 0.65

Evaluation of SNP associations with ESRD was performed using the best genetic transmission model of minor allele according to the lowest values of Bayesian and Akaike information criteria. Significant OR, and 95% CI are reported in bold.

To evaluate the possible interactions between SNPs and biological (age and gender) and clinical variables (hypertension, diabetes, and hypercholesterolemia), genotype frequencies of ESRD patients and controls were stratified and compared according to the different variables. Complete results of these analyses are reported in Supplementary Tables S10–14.

Stratifying results according to age, *MIF* rs755622 *C* positive genotype frequencies are increased both in <59 and \geq 59-year-old patients than in controls (Table 5). No other SNP has been shown to play a role in ESRD predisposition in association with age cut-off (Supplementary Table S10).

Table 5. Significant SNP genotype frequencies (gen. freq.) in end-stage renal disease patients (ESRD) and controls (CTRL) stratified according to 59 years age cut off (adjusted by gender, diabetes, hypertension, and hypercholesterolemia).

Genes and SNP Alleles			Ι	Age < 59		$Age \ge 59$				
		CTRL	ESRD	OR	<i>n</i> -Value	CTRL	ESRD OR		u Valua	
		N (Gen	ı. Freq.)	(95% CI)	p luite	N (Ger	n. Freq.)	(95% CI)	<i>p</i> -value	
	G/G	82 (0.56)	6 (0.28)	0.29 (0.11–0.79)	0.021	46 (0.69)	36 (0.51)	0.47 (0.23–0.94)	0.038	
MIF rs755622	C/G	29 (0.20)	8 (0.36)	3.95 (1.19–13.17)	0.013	16 (0.24)	16 (0.22)	0.93 (0.73–2.44)	1.00	
	C/C	35 (0.24)	8 (0.36)	1.81 (0.70–4.68)	0.292	5 (0.07)	19 (0.27)	4.56 (1.69–12.97)	0.003	
	C/*	64 (0.44)	16 (0.72)	3.30 (1.40–10.89)	0.021	21 (0.31)	35 (0.49)	2.13 (1.06–4.27)	0.038	

Analyses of data according to gender allowed us to observe that ESRD women shoved a significantly reduced frequency of genotypes bearing the *IL6 rs180795 C* allele. No differences were observed for men (Table 6).

Table 6. Significant SNP genotype frequencies (gen. freq.) in end-stage renal disease patients (ESRD) and controls (CTRL) stratified according to Gender (adjusted by 59 years of age cut-off, diabetes, hypertension, and hypercholesterolemia).

Genes and SNP Alleles			FEMALE		MALE				
		CTRL ESRD		OR	<i>n</i> -Value	CTRL	ESRD	OR	u Valua
		N (Gen	ı. Freq.)	(95% CI)	p mae	N (Gen	. Freq.)	(95% CI)	<i>p</i> -value
	G/G	50 (0.53)	31 (0.74)	2.48 (1.12-5.51)	0.036	68 (0.57)	34 (0.67)	1.50 (0.76-2.98)	0.306
	C/G	34 (0.36)	9 (0.21)	0.63 (0.23-1.69)	0.111	41 (0.35)	13 (0.25)	0.65 (0.32-1.38)	0.284
IL6	C/C	10 (0.11)	2 (0.05)	0.49 (0.09–2.67)	0.342	10 (0.08)	4 (0.08)	0.92 (0.27-3.13)	1.00
rs1800795	C/*	44 (0.47)	11 (0.26)	0.40 (0.18–0.89)	0.036	51 (0.43)	17 (0.33)	0.67 (0.34–1.32)	0.306

No other SNP has been shown to play a role in ESRD predisposition in association with gender (Supplementary Table S11).

Next, we evaluated the genotype frequency distribution of the nine SNPs, in relation to the presence of comorbidities as covariances. The power calculation was analyzed for the statistical analyses performed, when data were stratified for hypertension diabetes and hypercholesterolemia power of calculation was weak, so the statistical significance of these data was not considered to be relevant. No differences in frequencies of the SNP genotypes in ESRD patients affected by hypertension with respect to no hypertensive ESDR and hypertensive control groups (see Supplementary Table S12).

On the other hand, 15 of the 19 ESDR Diabetic patients were positive for homozygous GG genotype of *MIF rs755622* with a significant increase of frequency with respect to nondiabetic patients (Supplementary Table S13). When genotype frequencies were analyzed according to the presence of the Hypercholesterolemia of 22 ESRD-affected patients, 9 were positive for C allele bearing genotype of *MIF rs755622* with an increase of genotype frequencies with respect to hypercholesterolemic controls (Supplementary Table S14).

All in all, our results indicate that *MIF rs755622* might influence the susceptibility to ESDR both as an independent risk factor and in presence of comorbidities.

4. Discussion

The last stage of chronic renal failure (ESRD) is the result of the progressive reduction of renal function and the onset of a progressively more severe chronic renal failure whose evolution is strongly influenced by the underlying disease, age, sex, and genetic background of the patients. If we consider the progressive aging trend of the population and the increase in the prevalence of the main diseases predisposing to chronic kidney failure (CKF) (diabetes, hypertension, and cardiovascular diseases), identifying predicting genetic markers that allow early identification of patients with an increased risk to develop severe CKF and reach the ESRD appears to be mandatory [52,53].

Analyses of alleles and genotypes crude frequencies of nine SNPs of genes codifying for key cytokine and receptors involved in kidney damage pathogenesis do not allow to identify associations with the *Klotho*, in spite of the above-mentioned data on the role of *rs577912* polymorphism as risk factors of mortality in dialyzed patients [13] and the role of *rs9536314* in the so-called longevity trait [17]. In a study performed in 2016, it was shown that the GG genotype is associated with reduced survival and poor prognosis in dialysis patients compared to the TT and TG genotypes [18] but we were unable to confirm this observation in our population. Similarly, *FGF23 rs7955866* and *IGF1 rs35767* were not found to be associated with ESRD. On the other hand, more recently, other *FGF23* SNPs have been associated with CKD [54]. So further studies are warranted on this point. Regarding IGF-1, this cytokine plays major a role in the main pathways in the progression of

metabolic traits, such as progression of T2DM complications, as CKD, and the development of cardiovascular disease. In particular, *rs*35767 is known to contribute to the development of diabetes in various populations [25].

Interleukin-6 (IL-6), another important proinflammatory cytokine has been shown to play a critical role in the development of atherosclerosis and atherosclerotic disease including kidney diseases [55,56], and it is well known that patients having CKD are at higher risk for accelerated atherosclerosis [33]. Our results show that frequencies of genotypes bearing the C allele of *IL6 rs1800795* are lower in patients than in controls allowing us to hypothesize that the presence of the C allele might be protective against progression to ESRD. This was not confirmed by adjusting the odds ratio for biological variances and clinical comorbidities (Table 4), whereas the protective association was confirmed stratifying data according to gender in women but not in men.

It has long been known that the *rs1800795G/C* variant located in the gene promoter (-174G/C) is associated with changes in transcriptional expression [57].

Even if IL-6 tracks with disability and age-related diseases, published data on the interaction among *rs1800795* and gender are not conclusive. Studies on aging frailty or longevity trait have demonstrated the possibility that genetically determined high production of IL-6 and women gender are risk factors for age-related disease [58,59]. In this view, the IL-6 low production associated with the *rs1800795* C allele might be protective against ESRD for CKD-affected women. On the other hand, the relatively small size of the patient sample does not allow to consider this observation conclusive.

In diabetes, the C allele is associated with a low risk of nephropathy [60,61]. On the other hand, the C allele was found to be associated with sepsis, high levels of circulating IL-6, and mortality increase in dialyzed subjects [62–64].

These apparently conflicting results are not easy to interpret. On the other hand, genetically determined low production of IL-6 might be a vantage in the contrasting progression of systemic diseases characterized by a chronic low rate of the inflammatory response [65], as CKD, but might be detrimental in acute inflammatory response characterizing sepsis or dialysis complications.

As reported in Table 4, after correction for confounding factors, a weak association of *TNFA rs1800629* genotypes has been detected. Cytokines of the TNF family and their receptor has been recently proposed as biomarkers of kidney disease. Actually, TNF signaling pathways play important roles in the progression of atherosclerotic and kidney disease [31–33] and in particular in the progression of diabetic nephropathy [66,67].

Studies on *TNFA* SNPs have suggested that *rs1800629*, associated with an increased *TNFA* gene transcription, might be a risk factor for multifactorial diseases [68], in particular, *rs1800629* might be a risk factor for CKD in particular in diabetic patients but data have been not conclusive [38,39]. Generally, TNF- α concentrations are found to be elevated in various cardiovascular conditions (e.g., advanced health failure, cardiomyopathy) and may also induce vascular inflammation, which may contribute to the pathogenesis of atherosclerosis [69].

Arterial hypertension is one of the main causes of kidney damage leading to end-stage renal disease (ESRD) [70,71], and cardiovascular disease is the leading cause of mortality in patients with renal disease [72].

In spite of this, our data do not allow us to identify the association of hypertension and SNP genotype studied as risk or protective factors in our group of ESRD patients.

Typing of *MIF* rs1007888, which was found associated with an increased risk of diabetes [48], does not indicate a role for this SNP in ESRD predisposition, whereas the C positive genotypes of rs755622, located at -173bp before the transcription initiation site of *MIF* gene and associated with a higher production of the cytokine [73], is more frequent in patients than in controls. This observation was also confirmed by adjusting the data for confounding factors. In addition, C positive genotypes of *MIF* rs755622 seem to be associated with an increased risk to develop an ESRD both in patients \geq 59 and <59 years old.

Aging is per se a risk factor for kidney diseases and in particular for CKD. However, it is well known that glomerular filtration rate (eGFR) declines with aging and this might impinge on the risk of CKD evolution to terminal stage and mortality. As recently reported by Delanaye et al. [74], for people older than 65 years (reference eGFR 75–89 mL/min per 1.73 m²), the risk was relevant only when eGFR had fallen below 45 mL/min. In this view, the possibility to refine the analyses combining age and significant SNP markers could be useful for the best management of CKD prevention and therapy. However, our data indicate that the *MIF rs755622* C allele is an age and gender independent risk factor for ESRD progression

Our results are in good agreement with data of other groups indicating that high production of MIF is correlated with an increased susceptibility for a large number of inflammatory and metabolic diseases including chronic kidney failure [75–79]. However, in renal pathologies, MIF seems to act according to a model of antagonistic pleiotropism, in fact, in different clinical situations and in different stages of renal pathology it can exert both harmful and protective effects [47] which can be likened to the story of Dr. Jekyll and Mr. Hyde. Indeed, MIF can be considered a potential biomarker of kidney damage. Elevated urinary and plasma MIF levels correlate with acute renal failure [80], chronic kidney disease [77], and the polycystic kidney [81].

On the other hand, in chronic kidney diseases (CKD), MIF limits the pro-fibrotic and proinflammatory activation of tubular cells by counteracting the arrest of the cell cycle and promoting its regeneration; In addition, in acute kidney damage (AKI), MIF also counteracts the apoptosis of tubular cells with a protective effect [43,82,83]. So it is possible that the net effect of the presence of *rs755622* on CKD predisposition and ESRD progression might be influenced by individual biological factors such as aging [84] or the presence of comorbidities. As it is well known, the presence of metabolic inflammation-based diseases, in particular diabetes and atherosclerosis, constitute a high-risk factor for CKD and progression towards ESRD [85].

As it is well known, diabetes is one of the major comorbidities associated with CKD. CKD can be a consequence of the alteration of the glucose status or we can have a kidney disease on which type II diabetes is superimposed. In diabetic nephropathy, the endothelial involvement and local induction of cytokines and chemokines induce immune cell recruitment accompanying and promoting tissue remodeling and nephron atrophy [86]. Moreover, CKD patients often have both quantitative and qualitative alterations in the lipid metabolism of lipoproteins. These lipid changes are present in all stages of the disease but differ in part on the basis of the deficit of renal function, the presence of a replacement treatment (dialysis and transplant), and the extent of proteinuria. The most frequently observed modification is the constant reduction in HDL, and hypercholesterolemia is a risk factor for chronic kidney disease [87,88].

The analysis of the association between the polymorphisms we typed and the risk of dialysis in subjects suffering from hypercholesterolemia or diabetes do not allow us to identify associations with polymorphisms known to be associated with changes in lipid and carbohydrate metabolism as *Klotho rs564481* [15,16], whereas *MIF rs755622* genetic polymorphism might play a role in ESRD progression in subjects affected by these pathologies. However, our data were obtained analyzing a relatively low number of subjects bearing diabetes or hypercholesterolemia, so a definitive conclusion cannot be drawn, even if some literature reports seem to support this view [78,89,90].

5. Conclusions

In conclusion, CKD might be considered a paradigm of the multifactorial diseases in which preexisting pathological conditions and genetic susceptibility factors might interplay with each other, resulting in different trajectories of damage evolution to the final stage as ESRD. In this view, we have analyzed the role of some genetic variants of key cytokines, mediators, and receptors involved in the complex scenario of kidney chronic failure and their interaction with comorbidities. Our data indicate that some polymorphisms, *IL6*

rs1800795 and *MIF rs755622*, involved in the regulation of both renal function and inflammatory response can influence the evolution of chronic kidney disease. In particular, *IL6 rs1800795G* genotype and *MIF rs755622C* alleles associated with increased production of the respective cytokines inducing increased inflammatory damages of kidney parenchyma might be useful as a marker for evaluation of ESRD evolution risk.

In addition, it is hypothesized that the modulation of the activities of these and other genes can also be considered therapeutic targets for innovative therapies. In this light, further studies will certainly be needed to investigate the role and interaction of these SNPs, in particular *MIF rs755622*, in the progression of end-stage renal disease.

Supplementary Materials: The following supporting information can be downloaded at: https://www.action.com/actionals //www.mdpi.com/article/10.3390/genes13020226/s1, Table S1. Evaluation of Klotho rs577912 associations with ESRD (adjusted by age, gender, hypertension, diabetes, and hypercholesterolemia) using different genetic transmission models of minor alleles; Table S2. Evaluation of Klotho rs564481 associations with ESRD (adjusted by age, gender, hypertension, diabetes, and hypercholesterolemia) using different genetic transmission models of minor alleles; Table S3. Evaluation of Klotho rs9536314 associations with ESRD (adjusted by age, gender, hypertension, diabetes, and hypercholesterolemia) using different genetic transmission models of minor alleles; Table S4. Evaluation of FGF23 rs7955866 associations with ESRD (adjusted by age, gender, hypertension, diabetes, and hypercholesterolemia) using different genetic transmission models of minor alleles; Table S5. Evaluation of IGF1 rs35767 associations with ESRD (adjusted by age, gender, hypertension, diabetes, and hypercholesterolemia) using different genetic transmission models of minor alleles; Table S6. Evaluation of TNFA rs1800629 associations with ESRD (adjusted by age, gender, hypertension, diabetes, and hypercholesterolemia) using different genetic transmission models of minor alleles; Table S7. Evaluation of IL6 rs1800795 associations with ESRD (adjusted by age, gender, hypertension, diabetes, and hypercholesterolemia) using different genetic transmission models of minor alleles; Table S8. Evaluation of MIF rs755622 associations with ESRD (adjusted by age, gender, hypertension, diabetes, and hypercholesterolemia) using different genetic transmission models of minor alleles; Table S9. Evaluation of MIF rs1007888 associations with ESRD (adjusted by age, gender, hypertension, diabetes, and hypercholesterolemia) using different genetic transmission models of minor alleles; Table S10: SNP frequencies in end-stage renal disease patients (ESRD) and controls (CTRL) stratified according to 59 years age cut off (adjusted by gender, diabetes, hypertension, and hypercholesterolemia); Table S11: SNP frequencies in end-stage renal disease patients (ESRD) and controls (CTRL) stratified according to gender (adjusted by 59 years age cut off, diabetes, hypertension, and hypercholesterolemia); Table S12: SNP frequencies in end-stage renal disease patients affected by hypertension (Hy ESRD) compared to ESRD patients with normal blood pressure (No Hy ESRD) and hypertensive controls (HyCTRL) (adjusted by 59 years age cut off, gender, diabetes, and hypercholesterolemia); Table S13: SNP frequencies in end-stage renal disease patients affected by diabetes mellitus (DM ESRD) compared to ESRD patients without diabetes (No T2DM ESRD) and diabetic controls (DM CTRL) (adjusted by 59 years age cut off, gender, hypertension, and hypercholesterolemia); Table S14: SNP frequencies in end-stage renal disease patients affected by hypercholesterolemia (HC ESRD) compared to ESRD patients without hypercholesterolemia (No HC ESRD) and hypercholesterolemic controls (HC CTRL) (adjusted by 59 years age cut off, gender, hypertension, and diabetes).

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Data Availability Statement: All data generated or analyzed during this study are stored in electronic archives that can be supplied on request.

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References

- 1. Zoccali, C.; Kramer, A.; Jager, K.J. Chronic kidney disease and end-stage renal disease—A review produced to contribute to the report 'the status of health in the European union: Towards a healthier Europe'. *NDT Plus* **2010**, *3*, 213–224. [CrossRef] [PubMed]
- 2. National Kidney Foundation. K/DOQI clinical practice guidelines for chronic kidney disease: Evaluation, classification and stratification. *Am. J. Kidney Dis.* **2002**, *39* (Suppl. S1), S1–S266.
- 3. Zhang, W.R.; Parikh, C.R. Biomarkers of acute and chronic kidney disease. Annu. Rev. Physiol. 2019, 81, 309–333. [CrossRef]
- 4. Balistreri, C.R.; Candore, G.; Accardi, G.; Bova, M.; Buffa, S.; Bulati, M.; Forte, G.I.; Listi, F.; Martorana, A.; Palmeri, M.; et al. Genetics of longevity. data from the studies on Sicilian centenarians. *Immun. Ageing* **2012**, *9*, 8. [CrossRef] [PubMed]
- Jiang, W.; Wang, X.; Geng, X.; Gu, Y.; Guo, M.; Ding, X.; Zhao, S. Novel predictive biomarkers for acute injury superimposed on chronic kidney disease. *Nefrologia* 2021, 41, 165–173. [CrossRef] [PubMed]
- Phan, P.; Saikia, B.B.; Sonnaila, S.; Agrawal, S.; Alraawi, Z.; Kumar, T.K.S.; Iyer, S. The Saga of Endocrine FGFs. *Cells* 2021, 10, 2418. [CrossRef]
- Dalton, G.D.; Xie, J.; An, S.W.; Huang, C.L. New insights into the mechanism of action of soluble klotho. *Front. Endocrinol.* 2017, 8, 323. [CrossRef]
- 8. Kuro-o, M. Overview of the FGF23-Klotho axis. Pediatr. Nephrol. 2010, 25, 583–590. [CrossRef]
- 9. Yamazaki, Y.; Imura, A.; Urakawa, I.; Shimada, T.; Murakami, J.; Aono, Y.; Hasegawa, H.; Yamashita, T.; Nakatani, K.; Saito, Y.; et al. Establishment of sandwich ELISA for soluble alpha-klotho measurement: Age-dependent change of soluble alpha-klotho levels in healthy subjects. *Biochem. Biophys. Res. Commun.* **2010**, *398*, 513–518. [CrossRef]
- 10. Arking, D.E.; Krebsova, A.; Macek, M., Sr.; Arking, A., Jr.; Mian, I.S.; Fried, L.; Hamosh, A.; Dey, S.; McIntosh, I.; Dietz, H.C. Association of human aging with a functional variant of klotho. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 856–861. [CrossRef]
- 11. Webster, A.C.; Nagler, E.V.; Morton, R.L.; Masson, P. Chronic kidney disease. Lancet 2017, 389, 1238–1252. [CrossRef]
- 12. Zou, D.; Wu, W.; He, Y.; Ma, S.; Gao, J. The role of klotho in chronic kidney disease. *BMC Nephrol.* **2018**, *19*, 285. [CrossRef] [PubMed]
- 13. Friedman, D.J.; Afkarian, M.; Tamez, H.; Bhan, I.; Isakova, T.; Wolf, M.; Ankers, E.; Ye, J.; Tonelli, M.; Zoccali, C.; et al. Klotho variants and chronic hemodialysis mortality. *J. Bone Miner. Res.* **2009**, *24*, 1847–1855. [CrossRef] [PubMed]
- Rendina, D.; Esposito, T.; Mossetti, G.; De Filippo, G.; Gianfrancesco, F.; Perfetti, A.; Magliocca, S.; Formisano, P.; Prié, D.; Strazzullo, P. A functional allelic variant of the FGF23 gene is associated with renal phosphate leak in calcium nephrolithiasis. *J. Clin. Endocrinol. Metab.* 2012, *97*, E840–E844. [CrossRef] [PubMed]
- 15. Xu, X.; Liang, X.; Hu, G.; Zhang, J.; Lei, H. Renal function and klotho gene polymorphisms among Uygur and Kazak populations in Xinjiang, China. *Med. Sci. Monit.* **2015**, *21*, 44–51. [CrossRef] [PubMed]
- 16. Akbari, H.; Asadikaram, G.; Aria, H.; Fooladi, S.; Vakili, S.; Masoumi, M. Association of Klotho gene polymorphism with hypertension and coronary artery disease in an Iranian population. *BMC Cardiovasc. Disord.* **2018**, *18*, 237. [CrossRef]
- 17. Revelas, M.; Thalamuthu, A.; Oldmeadow, C.; Evans, T.J.; Armstrong, N.J.; Kwok, J.B.; Brodaty, H.; Schofield, P.R.; Scott, R.J.; Sachdev, P.S.; et al. Review and meta-analysis of genetic polymorphisms associated with exceptional human longevity. *Mech. Ageing Dev.* **2018**, *175*, 24–34. [CrossRef]
- Marchelek-Myśliwiec, M.; Różański, J.; Ogrodowczyk, A.; Dutkiewicz, G.; Dołęgowska, B.; Sałata, D.; Budkowska, M.; Safranow, K.; Stępniewska, J.; Wiśniewska, M.; et al. The association of the *Klotho* polymorphism *rs9536314* with parameters of calcium-phosphate metabolism in patients on long-term hemodialysis. *Ren. Fail.* 2016, *38*, 6–80. [CrossRef]
- 19. Sato, Y.; Yanagita, M. Immunology of the ageing kidney. Nat. Rev. Nephrol. 2019, 15, 625–640. [CrossRef]
- 20. Lanzani, C.; Citterio, L.; Vezzoli, G. Klotho: A link between cardiovascular and non-cardiovascular mortality. *Clin. Kidney J.* **2020**, 13, 926–932. [CrossRef]
- Haffner, D.; Grund, A.; Leifheit-Nestler, M. Renal effects of growth hormone in health and in kidney disease. *Pediatr. Nephrol.* 2021, *36*, 2511–2530. [CrossRef] [PubMed]
- 22. Wiezel, D.; Assadi, M.H.; Landau, D.; Troib, A.; Kachko, L.; Rabkin, R.; Segev, Y. Impaired renal growth hormone JAK/STAT5 signaling in chronic kidney disease. *Nephrol. Dial. Transplant.* **2014**, *29*, 791–799. [CrossRef] [PubMed]
- Teumer, A.; Qi, Q.; Nethander, M.; Aschard, H.; Bandinelli, S.; Beekman, M.; Berndt, S.I.; Bidlingmaier, M.; Broer, L.; CHARGE Longevity Working Group; et al. Genomewide meta-analysis identifies loci associated with IGF-I and IGFBP-3 levels with impact on age-related traits. *Aging Cell* 2016, 15, 811–824. [CrossRef] [PubMed]
- 24. Honda, H.; Qureshi, A.R.; Heimburger, O.; Barany, P.; Wang, K.; Pecoits Filho, R.; Stenvinkel, P.; Lindholm, B. Serum albumin, C reactive protein, interleukin 6, and fetuin A as predictors of malnutrition, cardiovascular disease and mortality in patients with ESRD. *Am. J. Kidney Dis.* **2006**, *47*, 139–148. [CrossRef] [PubMed]
- 25. Tong, X.; He, J.; Liu, S.; Peng, S.; Yan, Z.; Zhang, Y.; Fan, H. Macrophage migration inhibitory factor-173G/C gene polymorphism increases the risk of renal disease: A meta-analysis. *Nephrology* **2015**, *20*, 68–76. [CrossRef] [PubMed]

- 26. Smykiewicz, P.; Segiet, A.; Keag, M.; Żera, T. Proinflammatory cytokines and ageing of the cardiovascular-renal system. *Mech. Ageing Dev.* **2018**, *175*, 35–45. [CrossRef]
- 27. Tripathi, G.; Borkar, M.; Akhter, A.; Sankhwar, S.N.; Sharma, R.K.; Agrawal, S. Association of proinflammatory cytokines with end stage renal disease. *Cytokine* 2010, *50*, 278–283. [CrossRef]
- Cottone, S.; Mulè, G.; Nardi, E.; Vadalà, A.; Lorito, M.C.; Guarneri, M.; Arsena, R.; Palermo, A.; Cerasola, G. C-reactive protein and intercellular adhesion molecule-1 are stronger predictors of oxidant stress than blood pressure in established hypertension. *J. Hypertens.* 2007, 25, 423–428. [CrossRef]
- Cottone, S.; Mulè, G.; Nardi, E.; Vadalà, A.; Guarneri, M.; Briolotta, C.; Arsena, R.; Palermo, A.; Riccobene, R.; Cerasola, G. Relation of C-reactive protein to oxidative stress and to endothelial activation in es-sential hypertension. *Am. J. Hypertens.* 2006, 19, 313–318. [CrossRef]
- Rodriguez-Iturbe, B.; Pons, H.; Johnson, R.J. Role of the immune system in hypertension. *Physiol. Rev.* 2017, 97, 1127–1164. [CrossRef]
- Biasucci, L.M.; CDC; AHA. CDC/AHA workshop on markers of inflammation and cardiovascular disease: Application to clinical and public health practice: Clinical use of inflammatory markers in patients with cardiovascular diseases: A background paper. *Circulation* 2004, 110, e560–e567. [CrossRef] [PubMed]
- 32. Cottone, S.; Lorito, M.C.; Riccobene, R.; Nardi, E.; Mulè, G.; Buscemi, S.; Geraci, C.; Guarneri, M.; Arsena, R.; Cerasola, G. Oxidative stress, inflammation and cardiovascular disease in chronic renal failure. *J. Nephrol.* **2008**, *21*, 175–179. [PubMed]
- 33. Niu, H.; Jiang, R.; Dong, S.; Xia, L.; Fang, H. Research on association between levels of serum adiponectin, Hs-CRP, and sICAM-1 and hypertensive cerebrovascular complications. *Biomed. Res. Int.* **2021**, 2021, 4455038. [CrossRef]
- Zoccali, C.; Vanholder, R.; Massy, Z.A.; Ortiz, A.; Sarafidis, P.; Dekker, F.W.; Fliser, D.; Fouque, D.; Heine, G.H.; Jager, K.J.; et al. European renal and cardiovascular medicine (EURECA-m) working group of the European Renal Association—European Dialysis Transplantation ASSOCIATION (ERA-EDTA). The systemic nature of IRC. *Nat. Rev. Nephrol.* 2017, 13, 344–358. [CrossRef]
- 35. Bae, E.; Cha, R.H.; Kim, Y.C.; An, J.N.; Kim, D.K.; Yoo, K.D.; Lee, S.M.; Kim, M.H.; Park, J.T.; Kang, S.W.; et al. Circulating TNF receptors predict cardiovascular disease in patients with chronic kidney disease. *Medicine* **2017**, *96*, e6666. [CrossRef]
- Gohda, T.; Niewczas, M.A.; Ficociello, L.H.; Walker, W.H.; Skupien, J.; Rosetti, F.; Cullere, X.; Johnson, A.C.; Crabtree, G.; Smiles, A.M.; et al. Circulating TNF receptors 1 and 2 predict stage 3 CKD in type 1 diabetes. J. Am. Soc. Nephrol. 2012, 23, 516–524. [CrossRef]
- Niewczas, M.A.; Gohda, T.; Skupien, J.; Smiles, A.M.; Walker, W.H.; Rosetti, F.; Cullere, X.; Eckfeldt, J.H.; Doria, A.; Mayadas, T.N.; et al. Circulating TNF receptors 1 and 2 predict ESRD in type 2 diabetes. J. Am. Soc. Nephrol. 2012, 23, 507–515. [CrossRef] [PubMed]
- Peng, Y.; Li, L.J. TNF-α-308G/A polymorphism associated with TNF-α protein expression in patients with diabetic nephropathy. *Int. J. Clin. Exp. Pathol.* 2015, *8*, 3127–3131.
- Fathy, S.A.; Mohamed, M.R.; Ali, M.A.M.; El-Helaly, A.E.; Alattar, A.T. Influence of IL6, IL-10, IFN-γ and TNF-α genetic variants on susceptibility to diabetic kidney disease in type 2 diabetes mellitus patients. *Biomarkers* 2019, 24, 43–55. [CrossRef] [PubMed]
- 40. Avci Çiçek, E.; Rota, S.; Dursun, B.; Kavalci, E. Evaluation of serum NGAL and hepcidin levels in chronic kidney disease patients. *Ren. Fail.* **2016**, *38*, 35–39. [CrossRef]
- 41. Amdur, R.L.; Feldman, H.I.; Gupta, J.; Yang, W.; Kanetsky, P.; Shlipak, M.; Rahman, M.; Lash, J.P.; Townsend, R.R.; Ojo, A.; et al. Inflammation and progression of CKD: The CRIC study. *Clin. J. Am. Soc. Nephrol.* **2016**, *11*, 1546–1556. [CrossRef] [PubMed]
- Hassan, M.O.; Duarte, R.; Dickens, C.; Dix-Peek, T.; Naidoo, S.; Vachiat, A.; Grinter, S.; Manga, P.; Naicker, S. Interleukin-6 gene polymorphisms and interleukin-6 levels are associated with atherosclerosis in CKD patients. *Clin. Nephrol.* 2020, *93*, 82–86. [CrossRef] [PubMed]
- 43. Rocha, S.; Valente, M.J.; Coimbra, S.; Catarino, C.; Rocha-Pereira, P.; Oliveira, J.G.; Madureira, J.; Fernandes, J.C.; do Sameiro-Faria, M.; Miranda, V.; et al. Interleukin 6 (rs1800795) and pentraxin 3 (rs2305619) polymorphisms-association with inflammation and all-cause mortality in end-stage-renal disease patients on dialysis. *Sci. Rep.* **2021**, *11*, 14768. [CrossRef] [PubMed]
- 44. Ururahy, M.A.; de Souza, K.S.; Oliveira, Y.M.; Loureiro, M.B.; da Silva, H.P.; Freire-Neto, F.P.; Bezerra, J.F.; Luchessi, A.D.; Doi, S.Q.; Hirata, R.D.; et al. Association of polymorphisms in IL6 gene promoter region with type 1 diabetes and increased albumin-to-creatinine ratio. *Diabetes Metab. Res. Rev.* 2015, *31*, 500–506. [CrossRef] [PubMed]
- 45. Calandra, T.; Roger, T. Macrophage migration inhibitory factor: A regulator of innate immunity. *Nat. Rev. Immunol.* **2003**, *3*, 791–800. [CrossRef] [PubMed]
- 46. Nishihira, J.; Koyama, Y.; Mizue, Y. Identification of macrophage migration inhibitory factor (MIF) in human vascular endothelial cells and its induction by lipopolysaccharide. *Cytokine* **1998**, *10*, 199–205. [CrossRef] [PubMed]
- 47. Boor, P. MIF in kidney diseases: A story of Dr, Jekyll and Mr, Hyde. Pathologe 2019, 40, 25–30. [CrossRef]
- Aslani, S.; Hossein-nezhad, A.; Maghbooli, Z.; Mirzaei, K.; Karimi, F. Genetic variation in macrophage migration inhibitory factor associated with gestational diabetes mellitus and metabolic syndrome. *Horm. Metab. Res.* 2011, 43, 557–561. [CrossRef]
- Scola, L.; Giarratana, R.M.; Pisano, C.; Ruvolo, G.; Marinello, V.; Lio, D.; Balistreri, C.R. Genotyping strategy of SMAD-3 rs3825977 gene variant for a differential management of ascending aorta aneurysm in women people: Gender oriented diagnostic tools. Meta Gene 2020, 25, 100706. [CrossRef]

- 50. Scola, L.; Giarratana, R.M.; Marinello, V.; Cancila, V.; Pisano, C.; Ruvolo, G.; Frati, G.; Lio, D.; Balistreri, C.R. Polymorphisms of pro-inflammatory IL6 and IL-1β cytokines in ascending aortic aneurysms as genetic modifiers and predictive and prognostic biomarkers. *Biomolecules* 2021, *11*, 943. [CrossRef]
- 51. Balistreri, C.R.; Ammoscato, C.L.; Scola, L.; Fragapane, T.; Giarratana, R.M.; Lio, D.; Piccione, M. Susceptibility to heart defects in down syndrome is associated with single nucleotide polymorphisms in HAS 21 interferon receptor cluster and VEGFA genes. *Genes* **2020**, *11*, 1428. [CrossRef] [PubMed]
- 52. Jha, V.; Garcia-Garcia, G.; Iseki, K.; Li, Z.; Naicker, S.; Plattner, B.; Saran, R.; Wang, A.Y.; Yang, C.W. Chronic kidney disease: Global dimension and perspectives. *Lancet* 2013, *382*, 260–272. [CrossRef]
- 53. Cortinovis, M.; Ruggenenti, P.; Remuzzi, G. Progression, remission and regression of chronic renal diseases. *Nephron* **2016**, *134*, 20–24. [CrossRef]
- Liang, Y.; Luo, S.; Schooling, C.M.; Au-Yeung, S.L. Genetically predicted fibroblast growth factor 23 and major cardiovascular diseases, their risk factors, kidney function, and longevity: A two-sample mendelian randomization study. *Front. Genet.* 2021, 12, 699455. [CrossRef]
- Noto, D.; Cottone, S.; Cefalù, A.; Vadalà, A.; Barbagallo, C.; Rizzo, M. Interleukin 6 plasma levels predict with high sensitivity and specificity coronary stenosis detected by coronary angiography. *Thromb. Haemost.* 2007, 98, 1362–1367.
- 56. Hartman, J.; Frishman, W.H. Inflammation and atherosclerosis: A review of the role of interleukin-6 in the development of atherosclerosis and the potential for targeted drug therapy. *Cardiol. Rev.* **2014**, *22*, 147–151. [CrossRef]
- Napolioni, V.; MacMurray, J. Infectious diseases, IL6-174G>C polymorphism, and human development. *Brain Behav. Immun.* 2016, *51*, 196–203. [CrossRef] [PubMed]
- Kuh, D.; Cooper, R.; Sattar, N.; Welsh, P.; Hardy, R.; Ben-Shlomo, Y. Systemic inflammation and cardio-renal organ damage biomarkers in middle age are associated with physical capability up to 9 years later. *Circulation* 2019, 139, 1988–1999. [CrossRef]
- Sebastiani, P.; Thyagarajan, B.; Sun, F.; Honig, L.S.; Schupf, N.; Cosentino, S.; Feitosa, M.F.; Wojczynski, M.; Newman, A.B.; Montano, M.; et al. Age and sex distributions of age-related biomarker values in healthy older adults from the long life family study. J. Am. Geriatr. Soc. 2016, 64, e189–e194. [CrossRef]
- 60. Chen, B.; Wu, M.; Zang, C.; Li, Y.; Xu, Z. Association Between IL6 Polymorphisms and Diabetic Nephropathy Risk: A Metaanalysis. *Am. J. Med. Sci.* 2019, 358, 363–373. [CrossRef]
- 61. Chang, W.T.; Huang, M.C.; Chung, H.F.; Chiu, Y.F.; Chen, P.S.; Chen, F.P.; Lee, C.Y.; Shin, S.J.; Hwang, S.J.; Huang, Y.F.; et al. Interleukin-6 gene polymorphisms correlate with the progression of nephropathy in Chinese patients with type 2 diabetes: A prospective cohort study. *Diabetes Res. Clin. Pract.* **2016**, *120*, 15–23. [CrossRef] [PubMed]
- Panayides, A.; Ioakeimidou, A.; Karamouzos, V.; Antonakos, N.; Koutelidakis, I.; Giannikopoulos, G.; Makaritsis, K.; Voloudakis, N.; Toutouzas, K.; Rovina, N.; et al. Hellenic Sepsis Study Group. -572 G/C single nucleotide polymorphism of interleukin-6 and sepsis predisposition in chronic renal disease. *Eur. J. Clin. Microbiol. Infect. Dis.* 2015, 34, 2439–2446. [CrossRef] [PubMed]
- 63. Verduijn, M.; Maréchal, C.; Coester, A.M.; Sampimon, D.E.; Boeschoten, E.W.; Dekker, F.W.; Goffin, E.; Krediet, R.T.; Devuyst, O. The -174G/C variant of IL6 as risk factor for mortality and technique failure in a large cohort of peritoneal dialysis patients. *Nephrol. Dial. Transplant.* **2012**, *27*, 3516–3523. [CrossRef] [PubMed]
- 64. Hasuike, Y.; Nonoguchi, H.; Ito, K.; Naka, M.; Kitamura, R.; Nanami, M.; Tokuyama, M.; Kida, A.; Otaki, Y.; Kuragano, T.; et al. Interleukin-6 is a predictor of mortality in stable hemodialysis patients. *Am. J. Nephrol.* **2009**, *30*, 389–398. [CrossRef]
- Di Bona, D.; Vasto, S.; Capurso, C.; Christiansen, L.; Deiana, L.; Franceschi, C.; Hurme, M.; Mocchegiani, E.; Rea, M.; Lio, D.; et al. Effect of interleukin-6 polymorphisms on human longevity: A systematic review and meta-analysis. *Ageing Res. Rev.* 2009, *8*, 36–42. [CrossRef]
- Huang, Y.S.; Fu, S.H.; Lu, K.C.; Chen, J.S.; Hsieh, H.Y.; Sytwu, H.K.; Wu, C.C. Inhibition of tumor necrosis factor signaling attenuates renal immune cell infiltration in experimental membranous nephropathy. *Oncotarget* 2017, *8*, 111631–111641. [CrossRef] [PubMed]
- 67. Mehaffey, E.; Majid, D.S.A. Tumor necrosis factor-α, kidney function, and hypertension. *Am. J. Physiol. Renal. Physiol.* **2017**, 313, F1005–F1008. [CrossRef]
- 68. Knisely, M.R.; Maserati, M.; Heinsberg, L.W.; Shah, L.L.; Li, H.; Zhu, Y.; Ma, Y.; Graves, L.Y.; Merriman, J.D.; Conley, Y.P. Symptom science: Advocating for inclusion of functional genetic polymorphisms. *Biol. Res. Nurs.* **2019**, *21*, 349–354. [CrossRef]
- Kalliolias, G.D.; Ivashkiv, L.B. TNF biology, pathogenic mechanisms and emerging therapeutic strategies. *Nat. Rev. Rheumatol.* 2016, 12, 49–62. [CrossRef]
- Coresh, J.; Wei, G.L.; McQuillan, G.; Brancati, F.L.; Levey, A.S.; Jones, C.; Klag, M.J. Prevalence of high blood pressure and elevated serum creatinine level in the United States: Findings from the third National Health and Nutrition Examination Survey (1988–1994). Arch. Intern. Med. 2001, 161, 1207–1216. [CrossRef]
- Buckalew, V.M., Jr.; Berg, R.L.; Wang, S.R.; Porush, J.G.; Rauch, S.; Schulman, G. Modification of Diet In Renal Disease Study Group. Prevalence of hypertension in 1795 subjects with chronic renal disease: The modification of diet in renal disease study baseline cohort. *Am. J. Kidney Dis.* 1996, 28, 811–821. [CrossRef]
- Collins, A.J.; Li, S.; Gilbertson, D.T.; Liu, J.; Chen, S.C.; Herzog, C.A. Chronic kidney disease and cardiovascular disease in the Medicare population. *Kidney Int.* 2003, 87, S24–S31. [CrossRef] [PubMed]

- 73. Bruchfeld, A.; Wendt, M.; Miller, E.J. Macrophage migration inhibitory factor in clinical kidney disease. *Front. Immunol.* **2016**, *7*, 8. [CrossRef] [PubMed]
- 74. Delanaye, P.; Jager, K.J.; Bökenkamp, A.; Christensson, A.; Dubourg, L.; Eriksen, B.O.; Gaillard, F.; Gambaro, G.; van der Giet, M.; Glassock, R.J.; et al. CKD: A call for an age-adapted definition. J. Am. Soc. Nephrol. 2019, 30, 1785–1805. [CrossRef] [PubMed]
- Mittal, R.D.; Manchanda, P.K. Association of interleukin (IL)-4 intron-3 and IL6-174 G/C gene polymorphism with susceptibility to end-stage renal disease. *Immunogenet* 2007, 59, 159–165. [CrossRef] [PubMed]
- 76. Bernhagen, J.; Krohn, R.; Lue, H.; Gregory, J.L.; Zernecke, A.; Koenen, R.R.; Dewor, M.; Georgiev, I.; Schober, A.; Leng, L.; et al. MIF is a noncognate ligand of CXC chemokine receptors in inflammatory and atherogenic cell recruitment. *Nat. Med.* 2007, 13, 587–596. [CrossRef]
- 77. Bruchfeld, A.; Carrero, J.J.; Qureshi, A.R.; Lindholm, B.; Barany, P.; Heimburger, O.; Hu, M.; Lin, X.; Stenvinkel, P.; Miller, E.J. Elevated serum macrophage migration inhibitory factor (MIF) concentrations in chronic kidney disease (CKD) are associated with markers of oxidative stress and endothelial activation. *Mol. Med.* **2009**, *15*, 70–75. [CrossRef]
- 78. Herder, C.; Klopp, N.; Baumert, J.; Müller, M.; Khuseyinova, N.; Meisinger, C.; Martin, S.; Illig, T.; Koenig, W.; Thorand, B. Effect of macrophage migration inhibitory factor (MIF) gene variants and MIF serum concentrations on the risk of type 2 diabetes: Results from the MONICA/KORA Augsburg case-cohort study. 1984–2002. *Diabetologia* 2008, *51*, 276–284. [CrossRef]
- 79. Du, G.L.; Luo, J.Y.; Wang, D.; Li, Y.H.; Fang, B.B.; Li, X.M.; Gao, X.M.; Yang, Y.N. *MIF* gene *rs755622* polymorphism positively associated with acute coronary syndrome in Chinese Han population: Case-control study. *Sci. Rep.* **2020**, *10*, 140. [CrossRef]
- Hong, M.Y.; Tseng, C.C.; Chuang, C.C.; Chen, C.L.; Lin, S.H.; Lin, C.F. Urinary macrophage migration inhibitory factor serves as a potential biomarker for acute kidney injury in patients with acute pyelonephritis. *Mediat. Inflamm.* 2012, 2012, 381358. [CrossRef]
- Chen, L.; Zhou, X.; Fan, L.X.; Yao, Y.; Swenson-Fields, K.I.; Gadjeva, M.; Wallace, D.P.; Peters, D.J.; Yu, A.; Grantham, J.J.; et al. Macrophage migration inhibitory factor promotes cyst growth in polycystic kidney disease. J. Clin. Investig. 2015, 125, 2399–2412. [CrossRef] [PubMed]
- 82. Shachar, I. An essential MIF-CD74 signaling axis in kidney tubular regeneration, with prospects for precision medicine and pharmacological augmentation. *Am. J. Physiol. Renal Physiol.* **2017**, *313*, F1084–F1086. [CrossRef] [PubMed]
- Ochi, A.; Chen, D.; Schulte, W.; Leng, L.; Moeckel, N.; Piecychna, M.; Averdunk, L.; Stoppe, C.; Bucala, R.; Moeckel, G. MIF-2/D-DT enhances proximal tubular cell regeneration through SLPI- and ATF4- dependent mechanisms. *Am. J. Physiol. Renal Physiol.* 2017, 313, F767–F780. [CrossRef] [PubMed]
- 84. Abdel-Rahman, E.M.; Okusa, M.D. Effects of aging on renal function and regenerative capacity. *Nephron. Clin. Pract.* 2014, 127, 15–20. [CrossRef] [PubMed]
- Abdelhafiz, A.H.; Brown, S.H.; Bello, A.; El Nahas, M. Chronic kidney disease in older people: Physiology, pathology or both? Nephron. Clin. Pract. 2010, 116, c19–c24. [CrossRef]
- Anders, H.J.; Huber, T.B.; Isermann, B.; Schiffer, M. CKD in diabetes: Diabetic kidney disease versus nondiabetic kidney disease. *Nat. Rev. Nephrol.* 2018, 14, 361–377. [CrossRef] [PubMed]
- 87. Kwan, B.C.; Kronenberg, F.; Beddhu, S.; Cheung, A.K. Lipoprotein metabolism and lipid management in chronic kidney disease. *J. Am. Soc. Nephrol.* **2007**, *18*, 1246–1261. [CrossRef]
- Emanuelsson, F.; Nordestgaard, B.G.; Benn, M. Familial hypercholesterolemia and risk of peripheral arterial disease and chronic kidney disease. J. Clin. Endocrinol. Metab. 2018, 103, 4491–4500. [CrossRef]
- 89. Kleemann, R.; Bucala, R. Macrophage migration inhibitory factor: Critical role in obesity, insulin resistance, and associated comorbidities. *Mediat. Inflamm.* **2010**, 2010, 610479. [CrossRef]
- 90. Illescas, O.; Gomez-Verjan, J.C.; García-Velázquez, L.; Govezensky, T.; Rodriguez-Sosa, M. Macrophage migration inhibitory factor-173 G/C polymorphism: A global meta-analysis across the disease spectrum. *Front. Genet.* **2018**, *9*, 55. [CrossRef]