

Regular Article

Highlighted Paper selected by Editor-in-Chief

Development of Scaled-Up Synthetic Method for Retinoid X Receptor Agonist NEt-3IB Contributing to Sustainable Development Goals

Yuta Takamura, Ken-ichi Morishita, Shota Kikuzawa, Masaki Watanabe, and Hiroki Kakuta*

Division of Pharmaceutical Sciences, Graduate School of Medicine, Dentistry and Pharmaceutical Sciences, Okayama University; 1-1-1 Tsushimanaka, Kita-ku, Okayama 700-8530, Japan.

Received October 22, 2021; accepted November 14, 2021

Small-molecular drugs, which are generally inexpensive compared with biopharmaceuticals and can often be taken orally, may contribute to the Sustainable Development Goals (SDGs) adopted by the United Nations. We previously reported the retinoid X receptor (RXR) agonist 4-(ethyl(3-isobutoxy-4-isopropylphenyl)amino)benzoic acid (NEt-3IB, **1**) as a small-molecular drug candidate to replace biopharmaceuticals for the treatment of inflammatory bowel disease. The previous synthetic method to **1** required a large amount of organic solvent and extensive purification. In line with the SDGs, we aimed to develop an environmentally friendly, inexpensive method for the large-scale synthesis of **1**. The developed method requires only a hydrophobic ether and EtOH as reaction and extraction solvents. The product was purified by recrystallization twice to afford 99% pure **1** at 100 mmol scale in about 30% yield. The optimized process showed a 35-fold improvement of the E-factor (an index of environmental impact) compared to the original method. This work, which changes the solvent used to environmentally preferable ones based on the existing synthetic method for **1**, illustrates how synthetic methods for small-molecular drugs can be adapted and improved to contribute to the SDGs.

Key words Sustainable Development Goal; process chemistry; green sustainable chemistry; retinoid X receptor; inflammatory bowel disease

Introduction

In 2015, the United Nations (UN) Summit listed 17 Sustainable Development Goals (SDGs) to be achieved in the 15 years to 2030.¹⁾ One of them, “Ensure healthy lives and promote well-being for all at all ages (SDG 3)” was established partly due to the disparity in drug supply between developed and developing countries.²⁾ In recent years, various biopharmaceuticals have been developed to treat intractable diseases, such as inflixivab and adalimumab for the treatment of inflammatory bowel disease (IBD). However, the efficacy of biopharmaceuticals can be compromised by the emergence of antibodies, and they are generally expensive to produce. From this point of view, small-molecular drugs are again becoming attractive.

Small-molecular drugs are generally relatively inexpensive and can often be taken orally, but on the other hand, they have disadvantages such as a greater likelihood of off-target effects compared to biopharmaceuticals. Current small-molecular drugs used to treat IBD include 5-aminosalicylic acid and its derivatives, steroidal anti-inflammatory drugs, the Janus kinase (JAK) inhibitor tofacitinib, and the immunosuppressant azathioprine, but these have varying efficacy for mucosal healing and remission,³⁾ and there is still a need for alternatives.

We focused on retinoid X receptor (RXR) as a target to treat IBDs. The RXR agonist bexarotene (Targretin[®], Fig. 1) has been used to treat cutaneous T-cell lymphoma (CTCL), and has been the subject of drug repositioning studies for various inflammatory diseases, including IBD.⁴⁻⁷⁾ However, bexarotene causes side effects such as hypothyroidism.^{8,9)} Therefore, we aimed to create RXR ligands with reduced side effects. We

previously found that 4-(ethyl(3-isobutoxy-4-isopropylphenyl)amino)benzoic acid¹⁰⁾ (NEt-3IB, **1**, Fig. 1) maintains the ability to activate RXR transcription without causing the above side effects.¹¹⁾ Positron emission tomography (PET) imaging revealed that **1** accumulates in the liver after administration and is rapidly transferred to the lower gastrointestinal tract.¹²⁾ A therapeutic effect of **1** accompanied with mucosal healing was confirmed in IBD model mice.¹³⁾

To obtain sufficient amounts of **1** for further evaluation, we required a scaled-up, low-cost, green synthetic process. In particular, the key objective was to minimize the use of organic solvents in line with the UN's SDGs, such as “Ensure availability and sustainable management of water and sanitation for all (SDG 6).” However, our original synthesis of **1** uses various organic solvents and requires multiple purifications by column chromatography¹⁰⁾ (Chart 1, Medicinal Chemistry method). Therefore, in this study, we focused on hydrophobic ether solvents that are widely used in process chemistry, not only as reaction media, but also as extraction solvents. For process chemistry, cyclopentylmethyl ester (CPME), 2-methyltetrahydrofuran (2-MeTHF), 4-methyl-

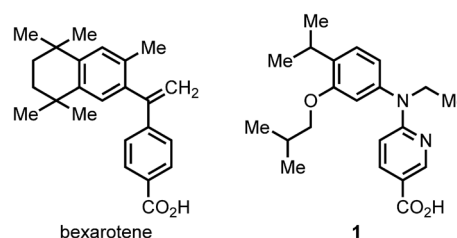
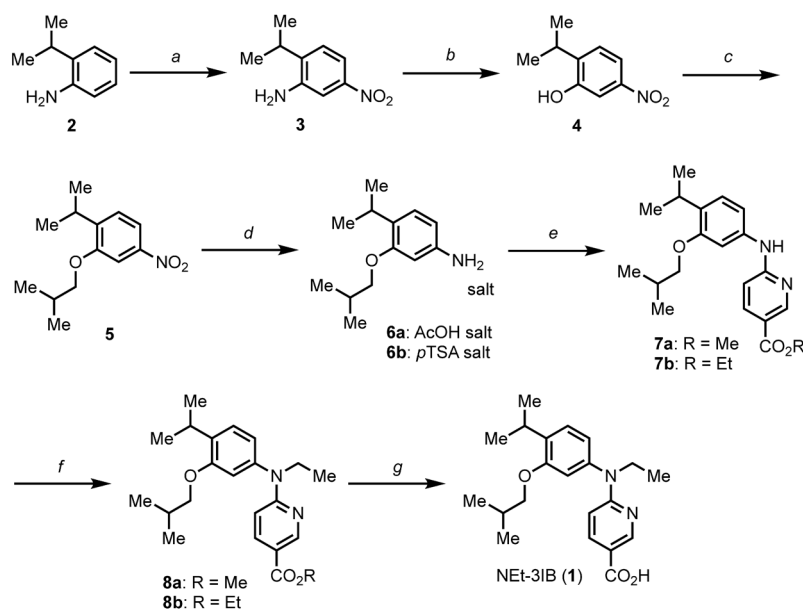


Fig. 1. Chemical Structures of RXR Agonists Bexarotene and NEt-3IB (**1**)

This paper is dedicated to Dr. Koichi Shudo, who passed away on 7 July 2021.

* To whom correspondence should be addressed. e-mail: kakuta-h@okayama-u.ac.jp



step	Medicinal Chemistry method ^{ref. 10}					Process method (This work)				
	Reagents	Solvent	Run time	Purification	Yield	Reagents	Solvent	Run time	Purification	Yield
a	HNO ₃ / H ₂ SO ₄ = 2/5 (v/v)	H ₂ SO ₄	<30 min	extracted with EtOAc	81%	KNO ₃	H ₂ SO ₄	1 h	-	-
b	1) NaNO ₂ 2) H ₂ SO ₄ / H ₂ O = 4/3 (v/v)	60% H ₂ SO ₄	<30 min	extracted with EtOAc, column chromat ography	75%	NaNO ₂ , H ₂ O	15% H ₂ SO ₄ CPME	1 h	washed with H ₂ O	-
c	<i>i</i> BuBr, K ₂ CO ₃ , KI	dry DMF	2 h	extracted with EtOAc, column chromat ography	71%	<i>i</i> BuBr, NaOH, TBAB	CPME H ₂ O	8 h	washed with H ₂ O	-
d	H ₂ , Pd/C	AcOH	6 h	-	q.y.	1) H ₂ , Pd/C 2) <i>p</i> TSA	EtOH	24 h	-	-
e	1) 6-chloro- nicotinic acid 2) H ₂ SO ₄	1) AcOH 2) dry MeOH	1) 4 h 2) o/n	extracted with EtOAc column chromat ography	62% for 2 steps	ethyl 6-chloro-n icotinate	CPME	28 h	washed with H ₂ O recrystall ized from EtOH, 70% EtOH	49% for 5 steps
f	EtI, NaH	dry DMF	o/n	extracted with EtOAc	-	EtI, KOH, TBAB	CPME	14 h	-	-
g	NaOH	MeOH H ₂ O	1 h	extracted with EtOAc, recrystal lized from CH ₂ Cl ₂ / <i>n</i> -hexane	41% for 2 steps	KOH	EtOH H ₂ O	1 h	recrystall ized from 70% EtOH	66% for 2 steps
<ul style="list-style-type: none"> • separation 6 times • column chromatography 3 times • recrystallization 1 time • total yield 11% (<100 mg) 					<ul style="list-style-type: none"> • separation 3 times • column chromatography 0 time • recrystallization 2 times • total yield 32% (>10 g) 					

Chart 1. The Medicinal Chemistry Method and the Newly Developed Process Method to **1**

tetrahydropyran (MTHP) were considered as candidates.^{14–17} These solvents can be azeotropically dehydrated and reused after reactions, so that organic solvent waste is minimized. In this study, we aimed to establish a new process synthesis of **1** using CPME as an organic solvent, since it is inexpensive and readily available. The finally developed method required only CPME and EtOH. The product was obtained in a total of

7 steps, and purified by recrystallization twice, affording 99% pure **1** at 100mmol scale in at least 30% overall yield.

Results and Discussion

Examination of Conditions for Each Reaction Step

Step a: Nitration

The first step is the nitration of 2-isopropylaniline (**2**). In the

Medicinal Chemistry method, nitroaniline **3** was synthesized using mixed acid ($\text{HNO}_3/\text{H}_2\text{SO}_4 = 2/5$).¹⁰ However, in increasing the reaction scale, the use of a highly reactive mixed acid may be dangerous, and nitric acid is volatile, so that it is difficult to calculate the number of equivalents precisely. Therefore, KNO_3 was added as a nitrating reagent in this step. The solubility of **2** was 3 mmol in 600 μL of *conc.* H_2SO_4 , and so this was used as the maximum concentration for examination in *Step b*. It was considered that the nitroaniline **3** can be obtained as a single product by adding 1 eq of KNO_3 to a *conc.* H_2SO_4 solution of **2**.

Step b: Sandmeyer reaction

In the Medicinal Chemistry method, after confirming the completion of *Step a*, neutralization, extraction with EtOAc, and evaporation under reduced pressure were performed, and the residue was dissolved in *dil.* H_2SO_4 again to carry out *Step b*.¹⁰ In order to prevent solvent wastage due to the neutralization and extraction, we considered that *Step b* should be performed continuously by adding the *conc.* H_2SO_4 solution of nitroaniline **3** to ice to obtain *dil.* H_2SO_4 solution. In the Medicinal Chemistry method, nitroaniline **3** was suspended in *dil.* H_2SO_4 , and NaNO_2 aq was added dropwise to form the diazonium salt, and then the solution was then added to heated *dil.* H_2SO_4 (about 50%) to obtain nitrophenol **4**.¹⁰ To avoid the increase in the volume of the reaction solution, NaNO_2 was added as a powder to the cooled *dil.* H_2SO_4 solution of nitrophenol **4**. In addition, we considered that dropping the diazonium salt solution into heated *dil.* H_2SO_4 should be simplified by directly heating the diazonium salt; this would also reduce the required amount of solvent. The important point here is the concentration of *dil.* H_2SO_4 during heating. A high concentration of *dil.* H_2SO_4 might lead to sulfonation as a side reaction,¹⁸ so we examined various concentrations up to 20% (Table 1, entries 1–4). In this experiment, the reaction mixture

was diluted to about 50 μM , and the reaction progress was monitored by HPLC. While 15% *dil.* H_2SO_4 showed the highest area%, the yield was only 43% (Table 1, entry 3). Decreasing the concentration of substrate **3** did not lead to a significant improvement (Table 1, entry 5). As a side reaction of hydroxylation by means of the Sandmeyer reaction, dimer formation between the produced phenol and unreacted aniline has been reported.¹⁸ Heating the *dil.* H_2SO_4 after adding an organic solvent has been reported to prevent such dimerization and to increase the product yield.¹⁸ Since CPME works as the organic solvent in the Sandmeyer reaction,¹⁹ we examined CPME in the present reaction. When CPME was added to the *dil.* H_2SO_4 and the mixture was directly refluxed, the area% reached 72% (Table 1, entry 6). A small further improvement was obtained (73%) by decreasing the concentration of substrate **3**, and finally we judged that the conditions of entry 7 were optimal for this reaction. On the other hand, the conditions of Entry 6 may also be noteworthy from the viewpoint of the amount of waste.

Step c: O-alkylation

In the Medicinal Chemistry method, *O*-isobutoxylated substrate **5** was obtained in a yield of 71% by stirring for 2 h using dry DMF as a solvent, K_2CO_3 as a base, potassium iodide (KI) as a catalyst and isobutyl bromide as a reaction reagent.¹⁰ In this method, nitrophenol **4** was not completely eliminated, and purification by column chromatography was required.¹⁰ First, we tried CPME as a solvent, but the reaction did not reach completion (Table 2, entry 2). This reaction between phenoxide ion and isobutyl bromide is considered to proceed *via* an $\text{S}_{\text{N}}2$ mechanism. NaOH , a stronger base than K_2CO_3 , was used (Table 2, entry 3). However, under the conditions of entry 2, NaOH did not sufficiently dissolve in CPME, and the reaction did not proceed. Next, in order to dissolve NaOH , the same amount of H_2O as CPME was added. Under this condition, KI was not added, because it is expected to dissolve in H_2O . When the reaction was carried out at 70 °C for 24 h, a part of the starting material **4** still remained (Table 2, entry 4). The starting material **4** is considered to exist as a phenoxide ion in the H_2O layer under basic conditions. On the other hand, isobutyl bromide is expected to exist in the CPME layer. Therefore, we focused on tetrabutylammonium bromide (TBAB), a phase-transfer catalyst that catalyzes the reaction at the interface between the organic layer and the H_2O layer.²⁰ Addition of a catalytic amount of TBAB significantly promoted the reaction and the reaction progress reached 56% after 24 h (Table 2, entry 5). As described above, CPME is known to azeotrope with H_2O at 83 °C.¹⁷ The boiling point of isobutyl bromide is about 90 °C, and it was expected that

Table 1. Comparison of Conditions for Steps a and b

Entry	Conc. of H_2SO_4 aq (v/v%)	Conc. of 3 (mol/L)	Solvent	Area% of 4 ^{a)}
1	5	0.25	5% H_2SO_4 aq	7%
2	10	0.25	10% H_2SO_4 aq	27%
3	15	0.25	15% H_2SO_4 aq	43%
4	20	0.25	20% H_2SO_4 aq	18%
5	15	0.125	15% H_2SO_4 aq	46%
6	15	0.125	15% H_2SO_4 aq/CPME	72%
7	15	0.084	15% H_2SO_4 aq/CPME	73%

a) Area% was detected by HPLC.

Table 2. Comparison of Conditions for Step c

Entry	Base	Catalyst	Solvent	Internal temp.	Result ^{a, b)}
1	K_2CO_3 (1.2 equiv.)	KI (0.1 equiv.)	Dry DMF	70 °C	Medicinal Chemistry method
2	K_2CO_3 (1.2 equiv.)	KI (0.1 equiv.)	CPME	70 °C	No reaction
3	NaOH (1.2 equiv.)	KI (0.1 equiv.)	CPME	70 °C	No reaction
4	NaOH (1.2 equiv.)	—	CPME/ H_2O	70 °C	17% progress at 24 h
5	NaOH (1.2 equiv.)	TBAB (0.1 equiv.)	CPME/ H_2O	70 °C	56% progress at 24 h
6	NaOH (1.2 equiv.)	TBAB (0.1 equiv.)	CPME/ H_2O	Reflux	70% progress at 24 h
7	NaOH (2.4 equiv.)	TBAB (0.1 equiv.)	CPME/ H_2O	Reflux	Complete at 10 h
8	KOH (2.4 equiv.)	TBAB (0.1 equiv.)	CPME/ H_2O	Reflux	Complete at 8 h

a) Area% was determined by HPLC. b) Reaction progress was confirmed every 2 h in all cases.

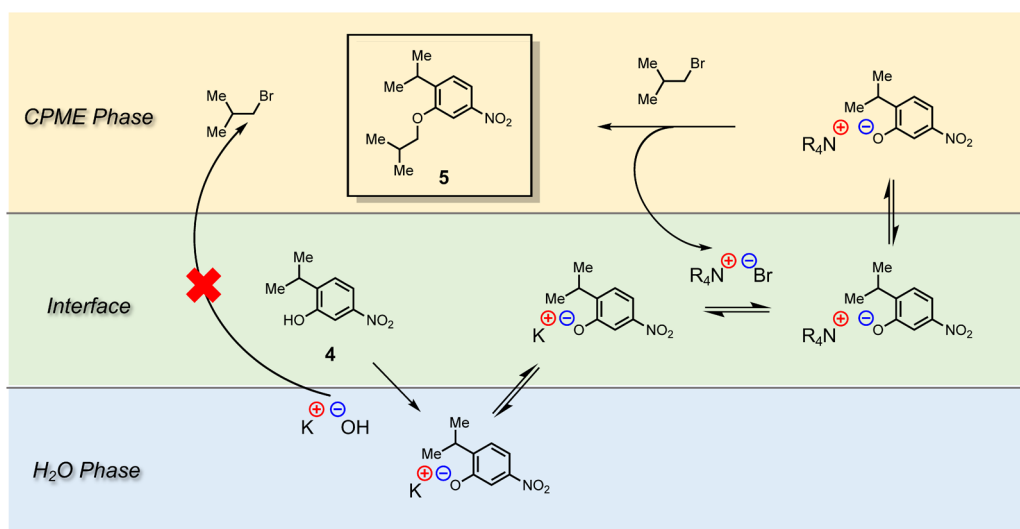


Fig. 2. Expected Mechanism of Step c

reflux could be achieved by using CPME and H₂O as solvents. When the reaction mixture obtained under the conditions of entry 5 was refluxed, the reaction progress reached 70% after 24 h (Table 2, entry 6). In order to completely eliminate the starting material **4**, the amount of base was increased, and in this case the reaction was completed in 10 h (Table 2, entry 7). Since KOH is used as a base in Step f (*N*-alkylation), we also examined the use of KOH in this reaction (Table 2, entry 8). It is considered important to reduce the number of types of reagents used in process synthesis. The reaction was completed in 8 h under these conditions. The starting material **4** is considered to exist as a potassium salt in the H₂O layer. The presence of the phase-transfer catalyst TBAB allows the potassium salt to become a quaternary amine salt, promoting the reaction with isobutyl bromide in the CPME layer. Furthermore, this was expected to prevent the loss of isobutyl bromide due to S_N2 reaction with KOH^{21,22} (Fig. 2). Overall, the conditions of entry 8 were judged as optimal for Step c.

Step d: Reduction

In the Medicinal Chemistry method, Pd-catalytic reduction using acetic acid as a solvent was performed. Then, since the subsequent coupling reaction with 6-chloronicotinic acid is carried out under acidic conditions, the aniline **6** obtained by Pd-catalytic reduction was isolated as an acetate salt.¹⁰ Since Pd-catalytic reduction is potentially dangerous in process chemistry when the reaction is scaled up, this reduction was performed after sufficient degassing to remove oxygen.²³ Reduction conditions using CPME as a solvent at normal pressure were examined, but without success. Therefore, in anticipation of promoting the reaction, we used EtOH, which is a protonic solvent. As a result, the starting material **5** disappeared and the target aniline **6** was obtained. Then, the solvent was changed from EtOH to CPME for the subsequent coupling reaction. In addition, *p*-toluenesulfonic acid (*p*TSA) salt **6b** was selected because it was expected to be more soluble in CPME than the acetate salt **6a** used in the Medicinal Chemistry method.

Step e: Coupling reaction

In the Medicinal Chemistry method, aminoacetate salt **6a** was dissolved in acetic acid, and coupled with 6-chloronicotinic acid, then the solvent was replaced with MeOH and

methyl ester **7a** was obtained by esterification with H₂SO₄.¹⁰ In order to simplify this reaction, we aimed to obtain the ethyl ester **7b** in one step by coupling amino *p*TSA salt **6b** and ethyl 6-chloronicotinate in CPME. If H₂O is present, hydrolysis may occur. It was confirmed that hydrolysis could be prevented by dehydration in a Dean–Stark apparatus before the reaction, and the ethyl ester **7b** was obtained as a single product.

Step f and Step g: *N*-alkylation and hydrolysis

In the Medicinal Chemistry method, *N*-alkylation was performed by using EtI and NaH as a base, and the final compound **1** was synthesized by subsequent hydrolysis using MeOH and NaOHaq.¹⁰ In scaling up this reaction, NaH was considered unsuitable due to safety concerns, and different bases were examined. With CPME as a solvent, *N*-alkylation and hydrolysis of the ester might proceed simultaneously in the presence of H₂O. Therefore, before the reaction, the CPME solution of ethyl ester **7b** was sufficiently dehydrated using a Dean–Stark apparatus. As a base, KOH (used in Step c) was first examined (Table 3, entries 2, 3). To enable subsequent hydrolysis of the ester in one pot, a large excess of base was added. *N*-Alkylation proceeded at 60 °C, but not at room temperature. However, even at 60 °C, the reaction was not complete after 12 h, and hydrolysis continued over 24 h. This result suggested that extension of the reaction time may cause hydrolysis by H₂O, which was thought to be produced by abstracting the proton of the secondary amine with a hydroxide ion. Therefore, we attempted to promote the *N*-alkylation rather than hydrolysis by adding TBAB (used in Step c). Under these conditions, the starting material **7b** disappeared and the *N*-Et ethyl ester **8b** was obtained in 6 h (Table 3, entry 4). When NaOH was employed, which was also used in Step c, hydrolysis proceeded at the same time (Table 3, entry 5). Since NaOH is highly deliquescent, it was considered that KOH, which is less deliquescent, would be more suitable as a base.^{24,25} At the interface between solid KOH and CPME, the reaction was promoted by TBAB, and the desired *N*-Et ethyl ester **8b** was obtained^{21,22} (Fig. 3). Based on these results, the conditions of entry 4 were judged as the most suitable for Step f.

In the final step of hydrolysis, CPME was evaporated under reduced pressure, and EtOH and H₂O were added to a flask containing *N*-Et ethyl ester **8b** and KOH. The mixture was

Table 3. Comparison of Conditions for Step f

Entry	Base	Catalyst	Solvent	Internal temp.	Result ^{a, b)}
1	NaH (1.2 equiv.)	—	DMF	r.t.	Medicinal Chemistry method
2	KOH (4.0 equiv.)	—	CPME	r.t.	Almost no reaction (approx.1%)
3	KOH (4.0 equiv.)	—	CPME	60 °C	30% progress at 12h Hydrolysis progressed over 24h
4	KOH (4.0 equiv.)	TBAB (0.1 equiv.)	CPME	60 °C	100% progress at 6h
5	NaOH (4.0 equiv.)	TBAB (0.1 equiv.)	CPME	60 °C	50% progress at 12h Hydrolysis progressed over 24h

a) Area% was determined by HPLC. b) Reaction progress was confirmed every 2h in all cases. r.t.: room temperature.

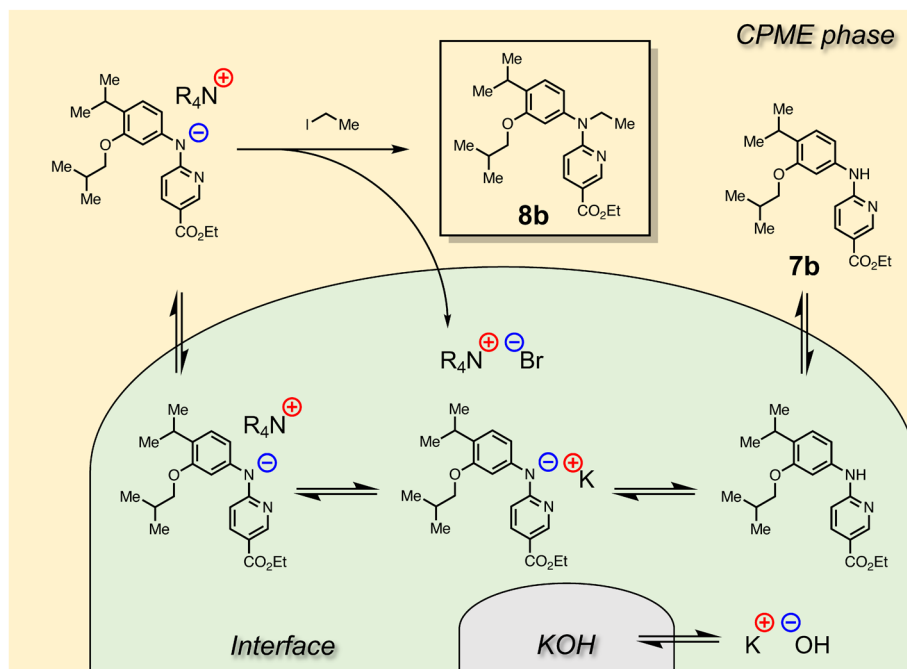


Fig. 3. Expected Mechanism of Step f

refluxed to obtain the target compound **1**, which was purified by recrystallization.

Synthesis on 100 mmol Scale

Continuous synthesis of **1** was performed using 100 mmol (13.5 g) of the starting material **2**.

[Steps a and b] To a solution of **2** (13.5 g, 100 mmol) in *conc.* H₂SO₄ (180 mL), KNO₃ (10.1 g, 100 mmol) was added to afford nitroaniline **3** with 99% purity (Supplementary Fig. S1). The obtained *conc.* H₂SO₄ solution of nitroaniline **3** was poured onto ice (1080 mL), and NaNO₂ (7.25 g, 105 mmol) was added to the resulting solution on an ice bath to prepare the diazonium salt. CPME (400 mL) was added to the flask, and Sandmeyer reaction was initiated by heating. Since unreacted **3** and inorganic substances were present in the H₂O layer, washing the CPME layer with H₂O gave nitrophenol **4** with a purity of 65% (Supplementary Fig. S2).

[Step c] Compound **4** obtained in the previous step was dissolved in CPME (100 mL) and H₂O (100 mL). To the mixture were added KOH (13.5 g, 240 mmol), TBAB (3.22 g, 10 mmol) and isobutyl bromide (20.6 g, 150 mmol). After refluxing for 8h the disappearance of nitrophenol **4** was confirmed (Fig. 4A). Since all reagents other than the desired product **5** were present in the H₂O layer, the CPME layer was washed with H₂O to afford **5**. The combined aqueous layer afforded the

main product with purity of 67% (Supplementary Fig. S3).

[Step d] After evaporation under reduced pressure, the solvent was replaced with EtOH (200 mL), and Pd-catalyzed reduction using H₂ gas was performed. The disappearance of the starting material **5** was confirmed after reaction for 24h. Then, EtOH was evaporated under reduced pressure and a CPME solution (100 mL) of aniline **6** was prepared. The concentration of aniline **6** was measured by HPLC, and an equimolar amount of *p*TSA was added. The mixture was stirred at room temperature to obtain **6b** as pale brown needles with a purity of 85% (Supplementary Fig. S4).

[Step e] To the CPME solution of **6b** was added ethyl 6-chloronicotinate (11.3 g, 60.7 mmol). The solution was refluxed for 21h, during which time **6b** disappeared (Fig. 4B, 4C). After the reaction, the CPME solution was washed with H₂O to remove *p*TSA. The ethyl ester **7b** was obtained as a black solid with a purity of 68% (Supplementary Figs. S5B, S5D). Residual ethyl 6-chloronicotinate was also observed. Recrystallization from EtOH gave **7b** (17.5 g, 49.1 mmol) as a pale brown solid with a purity of 92% (Supplementary Fig. S5). It should be noted that it was difficult to achieve decolorization at the final step.

[Steps f and g] After dissolving ethyl ester **7b** (17.5 g, 49.1 mmol) in CPME (200 mL), the solution was dehydrated in

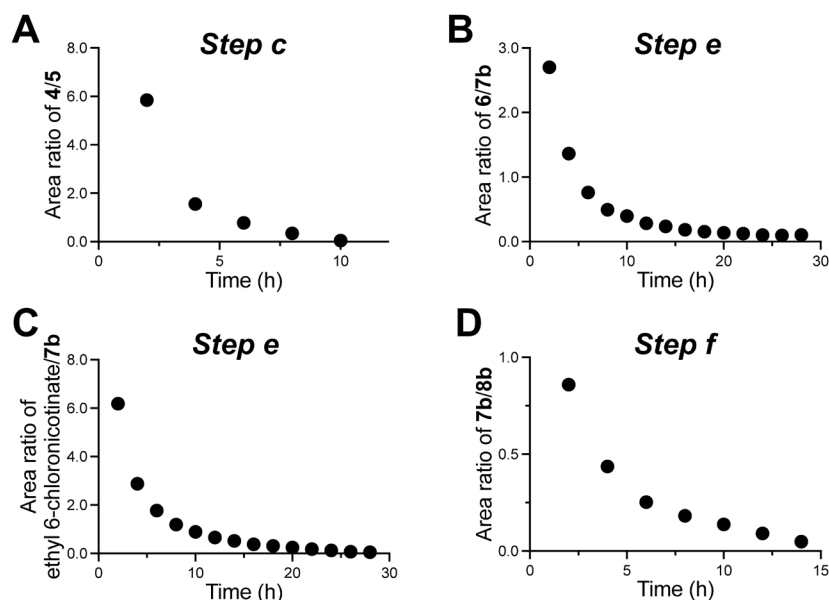


Fig. 4. Reaction Progress of (A) Step c, (B, C) Step e, and (D) Step f

The vertical axis shows the area ratio of each starting material/product determined by HPLC.

a Dean–Stark apparatus. Then, to the resulting solution were added KOH (11.0 g, 196 mmol), TBAB (1.58 g, 4.9 mmol) and EtI (9.17 g, 58.8 mmol) under an Ar atmosphere, and the mixture was heated at 60 °C for 14 h. Compound **7b** disappeared and the *N*-Et ethyl ester **8b** (Fig. 4D, Supplementary Fig. S6) was formed. CPME was removed under reduced pressure and EtOH and H₂O (100 mL each) were added. The mixture was heated for 1 h to afford the final compound **1** as a pale brown solid, which was recrystallized from 70% EtOH to afford pale brown needles with a purity of 99%, in a total yield of 32% over 7 steps (Supplementary Fig. S7).

CPME was recovered and reused as much as possible during the 100 mmol-scale synthesis. At this scale, the amounts of waste CPME and EtOH were about 50 mL and 650 mL, respectively.

Measurement of the Amount of Waste and Calculation of the E-Factor We were interested in evaluating how much more environmentally friendly the new process is, compared to the Medicinal Chemistry method. In multi-step synthesis, the E-factor is widely used as an index of environmental impact.^{26,27} This is defined with the weight (kg) of industrial waste (solvent, silica gel, etc.) produced to obtain 1 kg of the target product. In this calculation, water may or may not be included as waste. As there is a concern that the amount of waste would be increased excessively by including water, E-factors are calculated excluding waste water in many cases. For the Medicinal Chemistry method and the process method established above, we compared the effects of both methods on the environment by calculating the E-factor. In both methods, 20 mmol (2.7 g) of starting material **2** was used, and the amount of waste solvent in each step and the amount of silica gel (when column chromatography was carried out) were all calculated. The E-factor excluding water was 4669 for the Medicinal Chemistry method and 124 for the process method (Supplementary Figs. S8–S11, Table S2). Thus, the new process method showed an improvement of about 35 times compared to the Medicinal Chemistry method, and is therefore a more environmentally friendly synthetic method for **1**. In

general, the E-factor in pharmaceutical syntheses is between 25 and 200,^{26,27} so it can be said that the new process method is acceptable. When water was included in the waste, the E-factors of the two methods were 7559 and 390, respectively (Supplementary Table S3).

Process Method Using Other Hydrophobic Ether Solvents

In addition to CPME, 2-MeTHF and MTHP have been used as hydrophobic ether solvents for process chemistry. Thus, we evaluated whether the synthesis of **1** was possible using these solvents, starting from 20 mmol (2.7 g) of **2**. Compound **1** was obtained in total yields of 7 and 50%, respectively (Supplementary Figs. S12, S13, Table S4). The low total yield for 2-MeTHF is considered to be due to the low yield in Step b (Sandmeyer reaction). The main product in Step b in 2-MeTHF was isolated and analyzed by ¹H-NMR. The NMR chart revealed that 1-isopropyl-4-nitrobenzene (**3'**) was produced *via* the introduction of a proton into the diazonium site (Supplementary Fig. S14).²⁸ The HPLC retention time of **3'** was consistent with that of the by-product in Step b of the 100 mmol scale synthesis using CPME. Although the safety of the solvent has not been evaluated yet,¹⁴ MTHP gave the highest yield and could be of interest. Nevertheless, CPME is inexpensive and easily available, and may be advantageous for use in process chemistry.

Conclusion

In this study, we aimed to establish a scaled-up process synthesis for RXR agonist NEt-3IB (**1**), a drug candidate for IBD, that would be consistent with SDGs. The only organic solvents used for reactions and extraction are a hydrophobic ether (CPME) and EtOH, and the ether can be azeotropically dehydrated and reused. Simple purification by recrystallization twice from EtOH afforded 99% pure **1** in a yield of over 30%. When CPME was used, the amounts of waste CPME and EtOH were about 50 and 650 mL, respectively, from a 100 mmol scale reaction, and the E-factor was at least 35 times lower than that of the Medicinal Chemistry method. This work provides a scaled-up green process for synthesiz-

ing **1**, and we believe this work, which changes the solvent used to environmentally preferable ones based on the existing synthetic method for **1**, also offers a useful illustration of how synthetic methods for small-molecular drugs can be adapted and improved to contribute to the SDGs.

Experimental

General The reagents used in this study were purchased from the indicated sources: 2-isopropylaniline (**2**) (Sigma-Aldrich, MO, U.S.A.), KNO₃ (Wako, Osaka, Japan), conc.H₂SO₄ (Wako), NaNO₂ (Kanto Kagaku, Tokyo, Japan), urea (TCI, Tokyo, Japan), CPME (Wako), 1-bromo-2-methylpropane (Sigma-Aldrich), KOH (Wako), NaOH (Sigma-Aldrich), TBAB (TCI), N₂ gas (Chugoku Air Water, Japan), H₂ gas (Chugoku Air Water), EtOH (Wako), *p*-toluenesulfonic acid monohydrate (TCI), ethyl 6-chloronicotinate (TCI), ethyl iodide (Wako), conc.HCl (Wako), 2-MeTHF (TCI), and MTHP (TCI).

The progress of each reaction was monitored by TLC on a 0.2 mm TLC plates (Merck, aluminium-backed, silica gel 60 F245) and spots were located under UV light. Silica gel 60 (particle size 0.035–0.070 mm) was used for flash column chromatography. The progress of each reaction was also monitored by HPLC. The HPLC system used was a Shimadzu liquid chromatographic system (Japan) consisting of a LC-20AT pump, SPD-20A detector, and CTO-10AS column oven. Data were processed using Labsolutions software. The reaction mixture (20 μL) was injected onto an Inertsil ODS-3 column (4.6 mm i.d. × 100 mm, 5 μm, GL Sciences, Tokyo, Japan) fitted with a guard column of Intersil ODS-3 (4.0 mm i.d. × 10 mm, 3 μm, GL Sciences) at 40 °C; the mobile phase was MeOH/H₂O = 85/15 + 0.1% formic acid. The flow rate was 0.7 mL/min. A photodiode array (PDA) detector was used to monitor absorbance at 260 nm.

¹H-NMR (600 MHz) and ¹³C-NMR (150 MHz) spectra were recorded on a Varian NMR System PS600 at room temperature. Deuterated chloroform (CDCl₃) was used as the solvent for all routine NMR measurements. Chemical shifts are reported in ppm relative to the respective deuterated solvent peak, δ 7.26 ppm for ¹H-NMR and δ 77.2 ppm for ¹³C-NMR, and coupling constants are given in Hz. FAB-MS spectra (low- and high-resolution mass spectra) were measured on a JEOL JMS-700 mass spectrometer.

Synthesis

Examining the Conditions for Each Step

Examining the Conditions of Step a and Step b (Table 1)

Compound **2** was added to conc.H₂SO₄ (0.6 mL) in a 100 mL round-bottomed flask on ice and stirred to dissolve the solid. The solution was cooled below 5 °C, then KNO₃ (1.0 mol equiv.) was added. The reaction mixture was stirred at 5 °C for 30 min, then poured onto ice and washed with H₂O. NaNO₂ (1.05 mol equiv.) was slowly added at below 5 °C. The presence of excess nitrite ion was checked by using potassium iodide starch paper. To the reaction mixture were added urea (0.125 mol equiv.) and CPME as indicated (+/–). The reaction mixture was refluxed for 1 h, then diluted to about 50 μM. The reaction progress was monitored by HPLC. The reaction conditions are listed in Supplementary Table S1.

Examining the Conditions of Step c (Table 2)

To a solution of **3** (10 mmol) in solvent (CPME 20 mL or CPME/H₂O = 10 mL/10 mL) in a 100 mL round-bottomed flask were added base (K₂CO₃ or NaOH or KOH, 12 mmol

or 24 mmol), and catalyst (KI or TBAB, 1.0 mmol). The reaction mixture was heated at 60 °C for 30 min. Then 1-bromo-2-methylpropane (*i*BuBr, 15 mmol) was added, and the mixture was heated to 70 °C (internal temperature) or refluxed. The reaction mixture was diluted to about 50 μM and the reaction progress was monitored by HPLC.

Examining the Conditions of Step f (Table 3)

To a solution of **7b** (10 mmol) in CPME (20 mL) in a 100 mL round-bottomed flask were added base (NaOH or KOH, 40 mmol) and catalyst (TBAB 1.0 mmol or not), and the mixture was stirred at 60 °C for 30 min. Then EtI (12 mmol) was added and stirring was continued at 60 °C (internal temperature). The reaction mixture was diluted to about 50 μM and the reaction progress was monitored by HPLC.

100 mmol-Scale Synthesis

[Step a] Conc.H₂SO₄ (180 mL) was poured to a 1-L 3-neck flask equipped with a mechanical stirrer and a thermometer, and cooled to below 5 °C. To the flask, 2-isopropylaniline (**2**, 13.5 g, 100 mmol) was added at 1 mL/min with stirring (250 rpm). After the addition, the precipitated solid was dissolved by increasing the temperature to r.t. After cooling the reaction mixture to below 5 °C again, powdered KNO₃ (10.1 g, 100 mmol) was added slowly (1 g/min) with stirring (200 rpm). The progress of the reaction was monitored by HPLC (the total reaction time was 1 h). The retention times of **2** and **3** were 2.45 and 2.64 min, respectively (Supplementary Fig. S1).

[Step b] The reaction mixture was poured onto ice (300 mL) in a 500-mL beaker. The solution was transferred into a 1-L 3-neck flask and the beaker was washed with H₂O (820 mL). The concentration of H₂SO₄ at this stage was 15% (180 mL conc.H₂SO₄/1080 mL H₂O). The solution was cooled to below 5 °C, and powdered NaNO₂ (7.25 g, 105 mmol) was added slowly (200 mg/30 s) to the solution with stirring (200 rpm). The presence of excess nitrite ion was checked by the use of potassium iodide starch paper. To the reaction mixture were added urea (750 mg, 12.5 mmol) and CPME (400 mL with stirring (250 rpm) at 80 °C (internal temperature). After confirming completion of the reaction by HPLC (the total reaction time was 1 h), the reaction mixture was cooled to room temperature. The retention time of **4** was 3.25 min (Supplementary Fig. S2). The reaction mixture was poured into a separating funnel and the water layer was removed. The organic layer was washed with H₂O (200 mL × 3) and evaporated under reduced pressure.

[Step c] To make the reaction solution basic (pH 11), H₂O (100 mL) and KOH (13.5 g, 240 mmol) were added to **3** in a 1-L 3-neck flask. To the solution were added CPME (100 mL) and TBAB (3.22 g, 10 mmol), and the mixture was stirred (200 rpm) at 60 °C (internal temperature) for 2 h. Then, 1-bromo-2-methylpropane (20.6 g, 150 mmol) was added, and stirring was continued (200 rpm) at 80 °C (internal temperature). The progress of the reaction was monitored by HPLC (the total reaction time was 8 h). The retention time of **5** was 8.29 min (Supplementary Fig. S3). After the reaction was finished, the solution was cooled and then poured into a separating funnel. The organic layer was separated and washed with H₂O (100 mL × 3). The CPME phase was evaporated under reduced pressure.

[Step d] After adding EtOH (200 mL) to **5** in a 1-L Erlenmeyer flask, the air in the flask was replaced with nitrogen gas. Then, Pd/C (1.19 g, 5% (w/w) of **5**) was added to the flask, which was purged again with nitrogen. After replacement of N₂ with H₂ from a balloon, the solution was stirred at room

temperature under an H₂ atmosphere. The progress of the reaction was monitored by HPLC (the total reaction time was 24 h). The retention time of **6** was 1.81 min (Supplementary Fig. S4). After confirming completion of the reaction, the H₂ atmosphere was replaced with nitrogen gas. The reaction mixture was filtered through *Celite* and washed with EtOH (50 mL). The filtrate was evaporated under reduced pressure to remove EtOH. The residue was dissolved in CPME (100 mL) and the concentration of the product was determined by HPLC (14.2 g, 68.6 mmol). Then, *p*-toluenesulfonic acid (13.0 g, 68.6 mmol) was added and the mixture was stirred for 1 h at room temperature, affording **6b** as precipitated pale brown needles.

[Step e] A 1-L 3-neck flask was equipped with a mechanical stirrer, a thermometer, and a Dean–Stark apparatus with a Dimroth condenser. Before adding reagents, the CPME solution of **6b** was heated to remove water using the Dean–Stark apparatus. The boiling temperature was initially 85 °C, rising to 105 °C (internal temperature) after the removal of water. Then, ethyl 6-chloronicotinate (12.7 g, 68.6 mmol) was added to the CPME solution of **6b** and the reaction mixture was stirred (200 rpm) under reflux. The progress of the reaction was monitored by HPLC (the total reaction time was 28 h from the boiling). The retention time of **7b** was 9.29 min (Supplementary Fig. S5). After confirming completion of the reaction by HPLC, the reaction mixture was poured into a separating funnel and washed with H₂O (100 mL × 3). The CPME solution was concentrated under reduced pressure. The residue was recrystallized from EtOH (40 mL) and the precipitated solid was collected. Recrystallization was performed one more time from 70% EtOH_{aq} (20 mL) and the precipitated solids were combined to yield **7b** as a pale white solid (17.5 g, 49.1 mmol). ¹H-NMR (400 MHz, CDCl₃): δ = 8.81 (1H, dd, *J* = 2.3, 0.7 Hz), 8.05 (1H, dd, *J* = 8.8, 2.3 Hz), 7.31 (1H, s), 7.18 (1H, d, *J* = 7.8 Hz), 6.86–6.81 (3H, m), 4.35 (2H, q, *J* = 7.2 Hz), 3.71 (2H, d, *J* = 6.6 Hz), 3.33 (1H, seq, *J* = 7.4 Hz), 2.13 (1H, seq, *J* = 6.6 Hz), 1.38 (3H, t, *J* = 7.2 Hz), 1.23 (7H, d, *J* = 7.4 Hz), 1.06 (6H, d, *J* = 6.6 Hz). ¹³C-NMR (150 MHz, CDCl₃): δ = 165.73, 159.16, 157.27, 150.93, 139.32, 137.24, 134.15, 126.82, 117.18, 114.32, 106.79, 106.13, 74.52, 60.91, 28.66, 26.98, 22.82, 19.61, 14.59. FAB-MS *m/z*: 357 [M + H]⁺. HRMS (FAB+) *m/z*: [M + H]⁺ Calcd for C₂₁H₂₈N₂O₃, 357.2657. Found, 357.2128.

[Step f] Solid **7b** from the previous step was dissolved in CPME (200 mL) and the solution was added to a 1-L 3-neck flask equipped with a mechanical stirrer, a thermometer, and a Dean–Stark apparatus with a Dimroth condenser. After removal of water by the Dean–Stark apparatus, KOH (11.0 g, 196.0 mmol) and TBAB (1.58 g, 4.9 mmol) were added. The mixture was stirred (200 rpm) at 50 °C (internal temperature) under an Ar atmosphere for 1 h, then EtI (9.17 g, 58.8 mmol) was added, and stirring was continued at 60 °C (internal temperature) under an Ar atmosphere. The progress of this reaction was monitored by HPLC (the total reaction time was 14 h). The retention time of **8b** was 19.72 min (Supplementary Fig. S6). The reaction mixture was evaporated under reduced pressure.

[Step g] EtOH (50 mL) and H₂O (50 mL) were added to **8a** in a 1-L 3-neck flask equipped with a mechanical stirrer, a thermometer and a Dimroth condenser, and the mixture was stirred (200 rpm) at 80 °C (internal temperature). The progress

of reaction was monitored by HPLC (the total reaction time was 1 h). The retention time of **1** was 9.40 min (Supplementary Fig. S7). The reaction mixture was transferred to a 500-mL eggplant flask, and the EtOH was evaporated under reduced pressure. To acidify the solution to pH 4, *conc.*HCl (10 mL) was added and precipitated solid was filtered with the aid of water (50 mL) using a suction funnel. The solid was dried and recrystallized overnight from 70% EtOH_{aq} (750 mL) with stirring at r.t. The collected crystals were dried overnight under reduced pressure at 40 °C to give 11.5 g of NEt-3IB (**1**) as pale brown needles (32.3 mmol, 99% purity by HPLC). ¹H-NMR (600 MHz, CDCl₃): δ = 8.93 (1H, d, *J* = 2.2 Hz), 7.84 (1H, dd, *J* = 9.1, 2.2 Hz), 7.26 (1H, d, *J* = 8.0 Hz), 6.77 (1H, dd, *J* = 8.0, 2.0 Hz), 6.64 (1H, d, *J* = 2.0 Hz), 6.25 (1H, d, *J* = 9.1 Hz), 4.05 (2H, q, *J* = 7.0 Hz), 3.68 (2H, d, *J* = 6.5 Hz), 3.36 (1H, seq, *J* = 6.5 Hz), 2.12 (1H, seq, *J* = 6.7 Hz), 1.26 (6H, d, *J* = 6.5 Hz), 1.25 (3H, t, *J* = 7.0 Hz), 1.06 (6H, d, *J* = 6.7 Hz). ¹³C-NMR (150 MHz, CDCl₃): δ = 171.59, 160.87, 157.54, 152.10, 141.85, 137.75, 136.30, 127.17, 119.61, 113.37, 110.60, 107.58, 74.34, 45.50, 28.45, 26.87, 22.55, 19.38, 13.09. mp 189.6–191.0 °C. FAB-MS *m/z*: 357 [M + H]⁺. HRMS (FAB+) *m/z*: [M + H]⁺ Calcd for C₂₁H₂₈N₂O₃, 357.2666. Found, 357.2178.

Anal. Calcd for C₂₁H₂₈N₂O₃: C, 70.76; H, 7.92; N, 7.86. Found: C, 71.02; H, 8.09; N, 7.88.

20mmol-Scale Synthesis

Compound **2** (2.70 g, 20 mmol) was used as a starting material.

Medicinal Chemistry Method

All synthetic steps were performed according to reference S1. The final compound **1** was obtained in a yield of 0.91 g (2.56 mmol) (13%).

Process Method (using CPME)

All synthesis were performed as described above. The final compound **1** was obtained in a yield of 1.95 g (5.46 mmol) (27%).

Comparison of E-Factors for the Medicinal Chemistry and Process Methods

All the weights of waste materials, including waste solvent, either including or excluding water, were calculated. The E-factors were evaluated according to the following equation.

$$\text{E-factor} = \text{weight of waste (g)} / \text{weight of product (g)}$$

Process Method (using 2-MeTHF)

All synthetic procedures were performed as described above, except that 2-MeTHF was used instead of CPME. The final compound **1** was obtained in a yield of 0.51 g (1.43 mmol) (7%). mp 191.5–192.7 °C.

Process Method (using MTHP)

All synthetic procedures were performed as described above, except that MTHP was used instead of CPME. The final compound **1** was obtained in a yield of 3.56 g (9.98 mmol) (50%). mp 180.1–181.3 °C.

Acknowledgments The authors are grateful to the Division of Instrumental Analysis, Okayama University for the NMR and MS measurements.

This work was partially supported by Grants from Okayama University Science, Technology, and Innovation Creation Fellowship (to Y.T.).

Author Contributions Y.T., K.M. and H.K. conceived

and designed the project. Y.T., S.K., and M.W. examined each reaction condition. Y.T. and K.M. performed 100mmol scale synthesis. Y.T. performed 20mmol scale synthesis and calculated E-factors. The manuscript was written by Y.T. and H.K.

Conflict of Interest This study received funding from AIBIOS K.K. The founder was not involved in the study design, collection, analysis, interpretation of data, the writing of this article or the decision to submit it for publication. All authors declare no other competing interests.

Supplementary Materials This article contains supplementary materials.

References

- 1) "The Sustainable Development Agenda.": <<https://www.un.org/sustainabledevelopment/development-agenda/>>, cited 8 February, 2021.
- 2) Nabukalu J. B., Asamani J. A., Nabyonga-Orem J., *Int. J. Health Policy Manag.*, **9**, 297–308 (2020).
- 3) Seyedian S. S., Nokhostin F., Malamir M. D., *J. Med. Life*, **12**, 113–122 (2019).
- 4) Janakiram N. B., Mohammed A., Qian L., Choi C. I., Steele V. E., Rao C. V., *Neoplasia*, **14**, 159–168 (2012).
- 5) Mukherjee R., Davies P. J. A., Crombie D. L., Bischoff E. D., Cesario R. M., Jow L., Hamann L. G., Boehm M. F., Mondon C. E., Nadzan A. M., Paterniti J. R. Jr., Heyman R. A., *Nature* (London), **386**, 407–410 (1997).
- 6) Mariani M. M., Malm T., Lamb R., Jay T. R., Neilson L., Casali B., Medarametla L., Landreth G. E., *Sci. Rep.*, **7**, 42270 (2017).
- 7) McFarland K., Spalding T. A., Hubbard D., Ma J. N., Olsson R., Burstein E. S., *ACS Chem. Neurosci.*, **4**, 1430–1438 (2013).
- 8) Sherman S. I., *Clin. Lymphoma*, **3**, 249–252 (2003).
- 9) Standeven A. M., Escobar M., Beard R. L., Yuan Y. D., Chandraratna R. A., *Biochem. Pharmacol.*, **54**, 517–524 (1997).
- 10) Takamatsu K., Takano A., Yakushiji N., Morohashi K., Morishita K., Matsuura N., Makishima M., Tai A., Sasaki K., Kakuta H., *ChemMedChem*, **3**, 780–787 (2008).
- 11) Kakuta H., Ohsawa F., Yamada S., Makishima M., Tai A., Yasui H., Yoshikawa Y., *Biol. Pharm. Bull.*, **35**, 629–633 (2012).
- 12) Kobayashi T., Furusawa Y., Yamada S., Akehi M., Takenaka F., Sasaki T., Akahoshi A., Hanada T., Matsuura E., Hirano H., Tai A., Kakuta H., *ACS Med. Chem. Lett.*, **6**, 334–338 (2015).
- 13) Matsumoto R., Takahashi D., Watanabe M., Nakatani S., Takamura Y., Kurosaki Y., Kakuta H., Hase K., *Front. Pharmacol.*, **12**, 715752 (2021).
- 14) Prat D., Pardigon O., Flemming H. W., Letestu S., Ducandas V., Isnard P., Guntrum E., Senac T., Ruisseau S., Cruciani P., Hosek P., *Org. Process Res. Dev.*, **17**, 1517–1525 (2013).
- 15) Antonucci V., Coleman J., Ferry J. B., Johnson N., Mathe M., Scott J. P., Xu J., *Org. Process Res. Dev.*, **15**, 939–941 (2011).
- 16) Kobayashi S., Tamura T., Yoshimoto S., Kawakami T., Masuyama A., *Chem. Asian J.*, **14**, 3921–3937 (2019).
- 17) Azzena U., Carraro M., Pisano L., Monticelli S., Bartolotta R., Pace V., *ChemSusChem*, **12**, 40–70 (2019).
- 18) Yu Z., Ye X., Xu Q., Xie X., Dong H., Su W., *Org. Process Res. Dev.*, **21**, 1644–1652 (2017).
- 19) Taniguchi T., Imoto M., Takeda M., Nakai T., Mihara M., Iwai T., Ito T., Mizuno T., Nomoto A., Ogawa A., *Heteroatom Chem.*, **26**, 411–416 (2015).
- 20) Johnson C. R., Ansari M. I., Coop A., *ACS Omega*, **3**, 10886–10890 (2018).
- 21) Makosza M., Ludwikow M., *Rocz. Chem.*, **39**, 1223 (1965).
- 22) Kitamura M., Shirakawa S., Maruoka K., *Angew. Chem. Int. Ed.*, **44**, 1549–1551 (2005).
- 23) Gallagher W. P., Marlatt M., Livingston R., Kiau S., Muslehiddinoglu J., *Org. Process Res. Dev.*, **16**, 1665–1668 (2012).
- 24) Scapino L., Zondag H. A., Bael J. V., Diriken J., Rindt C. C. M., *Renew. Sustain. Energy Rev.*, **76**, 1314–1331 (2017).
- 25) Konings R. J. M., Cordfunke E. H. P., *J. Chem. Thermodyn.*, **20**, 103–108 (1988).
- 26) Tobiszewski M., Marć M., Gałuszka A., Namieśnik J., *Molecules*, **20**, 10928–10946 (2015).
- 27) Sheldon R. A., *Green Chem.*, **9**, 1273–1283 (2007).
- 28) Sharma Y., Nikam A. V., Kulkarni A. A., *Org. Process Res. Dev.*, **23**, 170–176 (2019).