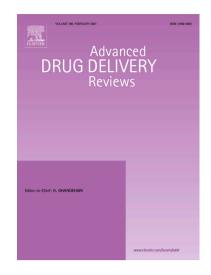
Electrohydrodynamic Atomisation Driven Design and Engineering of Opportunistic Particulate Systems For Applications in Drug Delivery, Therapeutics and Pharmaceutics

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1 Abstract

2 Electrohydrodynamic atomisation (EHDA) technologies have evolved significantly over the 3 past decade; branching into several established and emerging healthcare remits through timely 4 advances in the engineering sciences and tailored conceptual process designs. More specifically for pharmaceutical and drug delivery spheres, electrospraying (ES) has presented 5 itself as a high value technique enabling a plethora of different particulate structures. However, 6 7 when coupled with novel formulations (e.g. co-flows) and innovative device aspects (e.g., 8 materials and dimensions), core characteristics of particulates are manipulated and engineered 9 specifically to deliver an application driven need, which is currently lacking, ranging from 10 imaging and targeted delivery to controlled release and sensing. This demonstrates the holistic 11 nature of these emerging technologies; which is often overlooked. Parametric driven control 12 during particle engineering via the ES method yields opportunistic properties when compared to conventional methods, albeit at ambient conditions (e.g., temperature and pressure), making 13 14 this extremely valuable for sensitive biologics and molecules of interest. Furthermore, several processing (e.g., flow rate, applied voltage and working distance) and solution (e.g., polymer 15 16 concentration, electrical conductivity and surface tension) parameters impact ES modes and 17 greatly influence the production of resulting particles. The formation of a steady cone-jet and 18 subsequent atomisation during ES fabricates particles demonstrating monodispersity (or near 19 monodispersed), narrow particle size distributions and smooth or textured morphologies; all of 20 which are successfully incorporated in a one-step process. By following a controlled ES 21 regime, tailored particles with various intricate structures (hollow microspheres, nanocups, 22 Janus and cell-mimicking nanoparticles) can also be engineered through process head 23 modifications central to the ES technique (single-needle spraying, coaxial, multi-needle and 24 needleless approaches). Thus, intricate formulation design, set-up and combinatorial 25 engineering of the EHDA process delivers particulate structures with a multitude of 26 applications in tissue engineering, theranostics, bioresponsive systems as well as drug dosage 27 forms for specific delivery to diseased or target tissues. This advanced technology has great 28 potential to be implemented commercially, particularly on the industrial scale for several unmet 29 pharmaceutical and medical challenges and needs. This review focuses on key seminal 30 developments, ending with future perspectives addressing obstacles that need to be addressed 31 for future advancement.

Key words: electrohydrodynamic atomisation (EHDA), electrospraying, coaxial, particle
 engineering, drug delivery systems, core-shell micro/nano particles, dosage design, targeting

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1 1. Introduction

2

3 Particulate systems hosting active pharmaceutical ingredients (API), biologics and functional 4 chemical entities are dynamic and opportunistic platforms to meet key attributes and 5 requirements for specific drug delivery and therapeutic applications (anti-cancer [1], 6 theranostics [2] and tissue engineering [3]). Enhancing fundamental characteristics of particles 7 can be an intricate challenge. However, when overcome, optimum properties are achievable 8 using formulation or emerging engineering technologies; some of which have been adapted to 9 the therapy and drug delivery remits over the last decade. These provide noteworthy improvements (e.g., toxicity and consequential side effects) over conventional drug delivery 10 11 systems. Engineering methods have been adapted, manipulated and modified to fabricate an 12 array of particulate systems by exploiting the morphology and surface chemistry of resulting 13 structures. Successful engineering of tailored particles enables opportunistic pathways for 14 therapeutic and diagnostic agents to enhance efficacy at specific target tissue and sites of 15 interest. Fine-tuning morphological characteristics of particles during an engineering process 16 or set of processes, yields greater control over particle properties. For example; particle size, 17 size distribution, shape, rigidity and topography are known to influence therapeutic delivery 18 and therefore drug delivery strategies [4]. The particle size for example, determines the surface 19 area of particles which interact with biological milieu and can significantly impact dosing and 20 release characteristics. These, in turn, can affect blood circulation times [5], toxicity, particle 21 stability and drug loading [6].

22

23 It is therefore crucial to consider both particle surface chemistry and the nature of the intended 24 biological environment (at site of action). The interplay between these governs the degree of 25 interactions, surface charges, adhesive forces, chemical composition and functionality. For 26 instance, extensive studies have shown that zeta potential is dependent on the surface charge of particles [7], facilitates nanoparticle stability and impacts initial adsorption, biodistribution 27 28 and blood clearance levels [8]. These variable chemistries are therefore essential to determine 29 biocompatibility and toxicity of particle interactions with cellular tissues [9]. Physiochemical 30 properties of particulate drug delivery systems provides an approach to address evolving 31 therapeutic needs where such properties can be pivotal. Applications where cellular uptake 32 poses a challenge, several parameters (i.e. particle size, charge and morphology) can influence 33 how active molecules enter tissues effectively. If the drug delivery system undergoes surface 34 modification, it is imperative that biocompatibility for targeted delivery is not compromised

[10]. Where drugs are encapsulated in matrix type devices, the particle matrix material plays a key role in drug delivery. A controlled release mechanism is often accomplished with the preferred material of choice (e.g., polymer type and length) and offers a known degradation profile with minimal toxicity to surrounding cells [11]. The degradation behaviour is also known to impact cellular uptake for site-specific delivery [12].

6 Particle engineering includes a host of fabrication techniques and these can deploy either top-7 down or bottom-up methodologies. Top-down approaches involve size reduction of larger 8 particles and bulk material to the nanometre or micrometer scale [13]. This can be achieved 9 using techniques such as milling, homogenisation and atomisation [14]. In contrast, bottom-up 10 approaches generate nano-scaled architectures through assembly of individual molecules. This 11 is usually accomplished by imparting knowledge of supramolecular chemistry, where 12 interactions such as Van der Waals, hydrogen bonding and hydrophobic forces lead to 13 molecular self-assembly progressing to larger structures [15]. Examples of engineering 14 processes deploying such approaches include microfluidics, electrohydrodynamic atomisation 15 (EHDA) [16], hot-melt extrusion and co-precipitation methods [17].

16 A plethora of technologies have been used to engineer particles. One of these well-established 17 techniques is spray drying where liquid droplets are atomised by controlled evaporation, 18 eventually forming microspheres. A dry powder is formed consisting of API and other 19 excipients [18]. Spray dried particles can exhibit different diameters (10 - 100µm) [19] and 20 morphologies (crystalline, skin-forming and agglomerate) [20]. Similarly, freeze drying is another solvent-evaporation approach where frozen material containing drug components 21 22 undergoes sublimation to produce dried powder formulations. Microparticles can also be 23 generated from other engineering methods including nanoprecipitation, microfluidics [21] and 24 supercritical fluid processing, or self-emulsifying techniques such as hot-melt extrusion [22].

25 The aforementioned advanced particle engineering methods have been utilised for decades, 26 however, are accompanied with several drawbacks and bottlenecks. Engineering processes 27 comprising multiple steps need to be managed effectively and are known to cause reduced 28 production rates, loss of material and are therefore time-consuming. In this regard, EHDA is 29 an efficient technique which can be used to prepare several particulate drug delivery systems 30 with various sizes, compositions and morphologies at ambient conditions in a single-step [23]. This technique has been extensively studied due to its applicability for the preparation of 31 32 diverse drug delivery systems for different pharmaceutical and biomedical engineering

1 applications [24]. EHDA generates atomised liquid droplets via the deployment of a strong 2 electric field to produce structures at the micrometre and nanometer scales [25]. In addition to 3 one-step engineering, it also offers structure versatility, efficiency in producing particles with 4 desired morphology, narrow size distribution, high drug encapsulation and structure driven 5 control on drug release kinetics [26]. Therefore, an alternative route of engineering is emerging 6 which circumvents limitations associated with established methods for dosage development 7 (i.e. low encapsulation efficiencies, poor drug release kinetics, disruptive engineering 8 conditions and prolonged manufacturing time).

9 This review focuses on the EHDA process mainly for particle engineering. The fundamental 10 principles of EHDA technologies are summarised; elaborating on the types of particulate 11 structures which can be formed. Appropriate applications of this technique are discussed with 12 regards to facilitating pharmaceutical and therapeutic strategies with the aim to elucidate the 13 benefits such technologies bring to the fore for addressing challenges during dosage or particle 14 production in the pharmaceutical industry

15 2. Principles of electrohydrodynamic atomisation technologies

16

17 2.1 The electrohydrodynamic atomisation process

Recent advances (from theory to application) in particulate technology have meant techniques 18 19 such as EHDA have been increasingly utilised to engineer structures with desired properties. 20 According to early studies, the phenomena underpinning EHDA stems from the English 21 physicist Sir William Gilbert, who in the 1600s observed interactions between electrostatic and 22 magnetic forces. Gilbert's pioneering work revealed that a piece of charged amber was drawn 23 up into a conical shape, (what would eventually become known as the Taylor cone/Gilbert-24 Taylor cone) when exposed to water. The electric field employed by the charged amber resulted 25 in a fine aerosol of charged water droplets from the tip of the cone, thereby distinguishing the behavioural forces between the two materials [27]. In 1882, Lord Rayleigh also observed these 26 27 interactions and calculated a charge threshold limit which, when reached, can potentially hinder 28 the capability to engineer particles. This is known as the 'Rayleigh criterion limit' where 29 Rayleigh demonstrated that an isolated liquid droplet became unstable and disintegrated into a 30 jet of multiple drops when the critical threshold was exceeded compared to the stabilising effect 31 of the solution surface tension [28]. Further analysis into the realm of EHDA technology was 32 continued by John Zeleny, who worked on the physical behaviour of fluids and their 33 mathematical modelling during the electrohydrodynamic process [29]. Taylors theory emerged

in 1964, which subsequently led to the characterisation of the value at which a cone formed at
a liquid surface under the influence of an electrostatic force, and hence confirmed the Rayleigh
limit [30]. The formation of a droplet shape now known as the Taylor cone is used as a basis
to fabricate highly monodisperse (or near monodisperse) particles under stable processing with
structures exhibiting near uniformity and increased likelihood of reproducibility in the medical
industry.

7

8 The fundamental principal of EHDA is built on the atomisation of formulations by electrical 9 forces when exposed to a high voltage [31]. This results in the formation of fibers or particles 10 at the tip of a nozzle which are deposited on a grounded collection plate [32]. The EHDA set-11 up entails a simplistic framework comprising of an apparatus which includes a syringe pump 12 connected to a syringe filled with the formulation, a stainless-steel needle (also referred to as nozzle or spinneret), collection plate and a high voltage generator (Figure 1). During the EHDA 13 14 process, the desired formulation is allowed to flow through a capillary needle at a controlled rate. The ultimate exposure of a strong electrical field to the spheroid droplet results in a build-15 16 up of charge on the liquid surface [33]. Increasing the voltage further subsequently leads to the 17 stable cone-jet mode, thereafter, breaking up into finer particulate or fibrous microstructures 18 and nanostructures depending on a number of parameters [30].

19

20 2.2 Types of electrohydrodynamic atomisation technologies

EHDA is an umbrella term which encompasses a range of different structure engineering processes. Fiber (electrospinning) and particle (electrospraying (ES)) fabrication techniques make use of identical base technology and the latter is addressed in detail further in this review. The interplay of various parameters merits the atomisation method to be utilised for different therapeutic approaches.

26

27 2.2.1 Electrospraying

ES is a versatile jetting technique offering liquid atomisation of formulations consisting of a polymer and API when exposed to a high voltage. Electrostatic forces enable the formation of a Taylor cone and when these cohesive forces of surface tension are finally overcome, it results in uniform sized microparticles or nanoparticles (Figure 2) [14]. ES is a distinctive method in the family of EHDA technologies as particles fabricated by this technique exhibit selfdispersing behaviour due to the Coulomb repulsion. Therefore, the agglomeration and coalescence of particles are reduced, thus inhibiting the potential hazard of nozzle clogging in

the apparatus [34]. Major parameters which influence the morphology, size and encapsulation
capacities of the structures formed can be assigned to the flow rate and applied voltage,
developing various modes of atomisation [33].

4

5 In comparison to traditional systems, ES provides copious benefits for pharmaceutical applications owing to its ability to fabricate uniformly sized particles, high encapsulation 6 7 efficiencies and the potential to incorporate several bioactive compounds within polymer 8 matrices. As modern technology emphasizes the use of pharmacokinetic and pharmacodynamic profiles, the unique characteristic of particle size distribution is highly 9 acclaimed in the therapeutic industry as it can control the performance of drug delivery systems 10 and in turn minimise variations between batches [35]. Applications of the ES method include 11 12 the fabrication of particulate delivery systems for anti-cancer therapeutics [36], antiinflammatory drugs [37], antibiotics [38], proteins including gene and vaccine delivery [39] as 13 14 well as a wide variety of bioengineering treatments [40], all of which will be discussed in comprehensive detail below. 15

16

17 2.2.2. Electrospinning

Electrospinning is an electrohydrodynamic variant of ES using the same experimental set-up 18 19 but rather polymeric formulations with a higher viscosity than those employed in ES, resulting 20 in the production of fibers instead of particles [33]. One of the earliest appearances of the 21 electrospinning mechanism in literature is accredited to Morton in his patent submitted in the 22 1900s, [41] which consequently propagated a number of further studies on the electrospinning technique [42-44]. This emerging technique can create microfibres and nanofibers by the 23 24 utilisation of electrostatic forces exerted on a solution [45]. The fibrous structures are produced when an electrostatically charged jet containing the polymeric solution elongates at the needle 25 26 exit by an unstable whipping motion, resulting in a deposition of fibers on the grounded collection plate [46]. Electrospun fibers possess attractive characteristics as they are simple, 27 28 cost effective and versatile with respect to the range of polymers which can be incorporated 29 within the solution. They have therefore gained curiosity over the past few years in various 30 fields including tissue engineering [47], drug encapsulation [48], wound management devices 31 [49] and gene therapy [50].

32

1 2.2.3 Microbubbling

2 Research over the last decade has led to the exponential development of EHDA processing 3 techniques in the pharmaceutical industry. Coaxial EHDA, an extension of EHDA consists of 4 an outer and inner flow from a coaxial needle thereby forming a coaxial electrified jetting 5 system. The coaxial EHDA method has been extensively used for encapsulation systems, from which techniques such as microbubbling have stemmed from. Microbubbling has been 6 7 established by Farook et al in 2007, where the novel preparation of gas filled microparticles 8 (or bubbles) was initially reported. In this, microbubbles were prepared using a glycerol 9 medium which flowed through capillary tubes and formed a stable Taylor cone. Simultaneously, a secondary inner needle incorporated the gaseous phase which ultimately 10 11 produced monodisperse microbubbles that were less than 10 µm in size [51]. The fabrication 12 of microbubbles is of great interest in medical therapeutics [52] for applications such as 13 diagnostic imaging [53], drug delivery vehicles [54], and biomedical engineering [55].

14 Li et al for example, employed a co-axial electrospray method for the generation of 15 microbubbles to be used as ultrasound contrast agents for medical imaging. In this study, the 16 microbubble shell was comprised of a mixture consisting of polyethylene glycol (PEG) and 17 glycerol. The addition of PEG 400 significantly reduced the viscosity and microbubble size, 18 hence improving microbubble stability. The ability to fine-tune the formulation and operating 19 parameters deems the co-axial jetting system favourable as fabricated microbubbles can exhibit 20 an array of sizes and shell thicknesses. Thus, the microbubbling process as reported in this 21 study was found to be an exceptional technique, finding applications in imaging where 22 microbubbles play an efficient role as drug carriers [56]

23

24 2.2.4 Printing

25 Alternatively, printing has also been developed using EHDA technology which can provide 26 medical pathways to bioengineering [57], making use of 3D EHDA printed structures and more 27 recently near field electrospinning. Printing is a novel approach which involves the use of 28 various biomaterials to tailor topographies or coarse structures using desired quantities of 29 materials, which, when deposited, result in fabricated three-dimensional structural scaffolds 30 [58]. Near field electrospinning is an emerging printing technique which provides exceptional 31 control by allowing customisable adjustment of the voltage potential where the employment of 32 an electric field drives the ink and eventually prints micropatterns and nanopatterns on 33 functionalised topographical structures [59]. Printing techniques can provide applications in 34 tissue regeneration [60], targeted drug delivery [61] and biomedical engineering [62].

3. Principles of electrohydrodynamic atomisation technologies with respect to particle engineering

3

Significant strides have been made to accomplish engineered particles for different drug delivery platforms. To target and achieve specific therapeutic approaches, the influence of different solution and processing parameters can be adjusted, all of which impact the electric field and variations in resulting jetting modes. The characteristic versatility of the EHDA technique can therefore be employed as a useful tool for the deposition of an array of monodisperse particles.

10

11 3.1 Solution parameters

12 Several fluidic properties affect the characteristics of different atomised structures that are used for various engineering platforms. The electrohydrodynamic model, specifically ES, is reliant 13 14 on sufficient electrical conductivity which can impact the formation of a Taylor cone and the 15 particle production efficiency [63]. During the atomisation step, droplets exposed to an electric field result in an accumulation of charge on the surface [26]. This involves the interaction of 16 electrostatic forces between charged particles and the grounded collection plate, where low 17 18 conductivity is preferential for the generation of monodisperse electrosprayed particles [33]. 19 For this reason, the electrical conductivity of the solution should be considered as a key 20 parameter when optimising the ES method and has been previously demonstrated by scaling 21 laws [64]. The dielectric constant or relative permittivity determines how easily the atomised 22 liquid can be polarised due to the application of an electric field. It has been studied that a 23 solution with electrical conductivity as well as relative permittivity culminates into steady jet 24 formation upon the addition of an electric field [65].

25

Surface tension is another crucial parameter which facilitates the ES method in achieving a hydrostatic equilibrium and subsequently a stable Taylor cone. A key requirement for the EHDA process is the atomisation of the droplet from the needle tip and hence the surface tension of the solution is important [66]. As electrostatic coulomb forces increase, the effect of surface tension decreases. Solutions with high surface tension can result in jet instability whereas low surface tension formulations in conjugation with low viscosity, form optimised particles in a controlled manner [67].

33

1 Another formulation variable for the generation of successful ES is the visco-elastic properties 2 of the solution [66]. Viscosity is a critical factor that distinguishes the ES and electrospinning 3 methods. Solutions exhibiting low viscosity have been used ideally for the formation of stable 4 cone jetting which results in particles with uniform size (or near uniform) and smooth surface 5 morphology [68]. Increasing the viscosity further subsequently leads to the generation of continuous electrospun fibers which, after a certain threshold is reached, results in poor jet 6 7 emission due to blockage at the needle orifice [69]. For this reason, it is of paramount 8 importance to identify if either particles or fibers are desired so that an optimum viscosity can 9 be established during solution preparation.

10

The density of a solution is also considered as a vital physical property which determines the Taylor cone jet diameter and can be defined as the relative measurement by the ratio of mass to volume. As elaborated, viscosity plays a crucial role in the jet stability, however, this parameter is greatly dependent on the density of the formulation [64]. During ES, an increase in density has been associated with larger particle diameter sizes due to the influence of gravity on the conical droplet shape [46].

17

18 3.2 Processing parameters

19 Processing parameters such as the flow rate, voltage, working distance (needle exit to collector) 20 and the needle diameter can also alter the functioning process of the EHDA set-up and 21 determine the ability of the EHDA mode to produce spherical nanoparticles. Flow rate is the 22 rate at which the liquid solution travels through the capillary needle and therefore a crucial 23 parameter for optimising particulate structures. The variation in flow rate can impact the 24 particle size, particle size distribution and morphology. Generally, a lower flow rate ($\leq 1 \text{ mL/h}$) has been suggested for the ES mode of EHDA to achieve spherical monodisperse nanoparticles 25 26 with narrow size distributions [66]. Coaxial EHDA, a further sophisticated EHDA variant, is a 27 branch of ES in which two or more liquid solutions are passed through two or more 28 concentrically located capillary needles. The flow rate of core and shell liquids must be 29 optimised, where a suitable ratio between these should be established to achieve a successful 30 coaxial ES system [68].

31

The applied voltage acts as a driving force for atomisation of the liquid and significantly impacts the various modes of ES, resulting in a number of conical sprays from initial dripping to stabilised Taylor cone formation and partially defines the overall outcome of the ES process

[70, 71]. The increase in applied voltage has shown to decrease the particle size of atomised formulations. However, after a certain threshold, the particles begin to morph into elongated or beaded fibers due to the presence of Coulomb forces and an increase in charge imposed on the droplet at the solution interface [16]. On the contrary, conflicting studies suggest that an increase in the applied voltage consequently results in an increased particle diameter size. For this reason, it is imperative that the applied voltage is chosen depending on the individual formulation and fine-tuned to accommodate the desired particle characteristics.

8

9 It is postulated that the working distance can considerably influence the capability of the EHDA process to engineer optimal particles. Typically, a shorter working distance results in a stronger 10 11 electric field and therefore the fabrication of monodisperse particles [66]. As the collection 12 distance increases, a higher voltage is required to provide a substantial electric field, otherwise, this may result in the failure of controlled atomised structures and potential material loss [72]. 13 14 Too small a working distance equates to the inability of complete solvent evaporation, eventually forming a wet particulate system and possible particle aggregation on the grounded 15 16 collection plate [73]. Therefore, an ideal working distance must be utilised during the ES mode, such optimisation could be achieved with adequate jetting maps [74]. The optimisation of the 17 18 working distance in combination with jetting map experimentations can be used to predict the 19 distance at which spheroid particles are formed. This is a useful technique, proving great 20 flexibility during particle engineering in pharmaceutical manufacturing.

21

22 As well as the types of needles employed (Figure 1a), the diameter of the needle orifice, often 23 expressed in gauge, also plays an instrumental role in the ES set-up and affects the properties 24 of resultant structures [75]. The inner diameter of the nozzle also impacts the size and size 25 distribution of fabricated droplets as it is heavily reliant on the development of the Taylor cone 26 domain by means of its apex. Particles obtained using smaller needle gauges (small nozzle 27 diameters) produce fine monodisperse particles and a stable ES mode [68]. In contrast, the 28 utilisation of large needle gauges (large needle orifice diameters) demonstrates unstable 29 sputtering, thereby generating particulates which display polydisperse behaviour [73].

30

31 3.3 Local environment engineering parameters

32 In addition to the significance of processing parameters and properties of materials used in ES 33 for the effective production of particle engineering, the ambient environment is also of critical 34 importance. For example, an increase in temperature results in accelerated evaporation of the

solvent medium, a decrease in polymer viscosity and a reduced particle size [70]. Additionally,
a high humidity level influences the solidification process of ES and impacts the morphology
of droplets due to condensation which could potentially lead to the diffusion of water into the
particulate system as well as crystallisation [76]. For a more controllable atmosphere, the ES
set-up can be placed in an isolated chamber where the cocooning nature of the chamber
prevents contamination and allows the adjustability of ambient conditions [66].

7

8 3.4 Other considerations

Various modes of the ES phenomena can be obtained by optimising the parameters discussed 9 10 earlier, including modes categorised into either dripping or jetting [77] (Figure 1b). This is 11 based on the geometrical shape of the liquid droplet at the needle exit during the employment 12 of an electric field and eventually the deformation of the droplet interface. There are several 13 modes, these comprise of initial dripping, micro-dripping, spindle and multi-spindle modes, 14 where the liquid solution forms fragments of the formulation. In jetting, however, a continuous 15 jet is produced out of the capillary needle exit at a distance from the spinneret and includes 16 cone-jet (Taylor cone), precession, oscillating-jet or multi-jet variations.

17 The choice of polymer and solvent employed during ES is crucial in determining the 18 characteristics of fabricated particulates, including the weight, porosity and surface properties 19 of the developed structures [78]. Various polymer properties should be considered (i.e. 20 molecular weight, concentration, viscosity, surface tension, immunogenicity and 21 biocompatibility) when streamlining the ES process and optimising solutions.

22

The solvent used during ES can also ultimately impact the particle morphology and the size of 23 24 resultant structures. Highly volatile solvents positively correlate with greater evaporation rates 25 and coincide with increased particle diameter [46]. Solvent properties must also include high 26 miscibility with a large range of solutes [68] and high electrical conductivity, thus reducing the 27 amount of applied voltage necessitated for the ES process [46]. The surface tension of the 28 solvent is another essential criterion for determining the type of solvent material which should 29 be employed during the atomisation process. In principle, solvents with high electrical 30 conductivity and low surface tension are preferable to fabricate monodisperse particles [79].

31

The aforementioned processing and material parameters provide a detailed insight into the intricacies involved in EHDA and specifically ES technologies with respect to particle engineering. The broad selection of parameters indicates the ability to fabricate diverse

particles with multifunctional properties, which, in particular is an overarching challenge
 during drug delivery and related therapeutic (e.g. theranostic) applications.

3

4 4. Engineering various types of particulate structures using the 5 electrospraying set-up. 6

- 7 Optimisation of the experimental arrangement, specifically the needles employed to fabricate 8 particles have the potential to control a diverse range of functional structures that can be 9 generated from ES (Table 1).
- 10

11 4.1 Pre-atomised formulations

Before the introduction of ES, various orthodox approaches can be utilised to prepareparticulate systems including nanoprecipitation, emulsification and solid dispersion.

14

Nanoprecipitation, also known as agitated solvent displacement, involves the dissolution of the 15 16 active in an aqueous polymer solution prior to the addition of the anti-solvent medium (containing distilled water or an aqueous buffer) [80]. Rapid desolvation of the polymeric 17 18 material results in a precipitated polymer and drug entrapment in the polymer matrix. The 19 nanoparticles which are ultimately formed can be explained by the Marangoni effect where 20 interfacial turbulences between the two phases results in the fabrication of nanoparticles [81]. 21 Instability in this phenomenon can cause coalescence and broad size distributions of polymeric 22 nanoparticles [82].

23

24 Emulsion-based techniques have been widely employed for hydrophobic molecules, improving 25 their release profiles for targeted drug delivery [83]. Emulsification methods can be further 26 classified into single-emulsion (w/o, o/w and o/o) or double emulsion (w/o/w and o/w/o) 27 disperse systems, where varying the proportions of oil and water molecules significantly 28 impacts the emulsion blend and therefore the hydrophilic and lipophilic characteristics. For 29 these methods, an aqueous solution of hydrophilic entities is thoroughly mixed with the 30 appropriate polymeric formulation, forming microdroplets which are subsequently dried by 31 solvent evaporation, eventually producing nanoparticles or microparticles [66]. The use of 32 organic solvents is however accompanied with possible drug denaturation (due to loss of drug 33 during emulsification or washing steps) leading to low drug encapsulation efficiencies [84].

34

Solid dispersion is a specific process referring to the dispersion of the drug at solid state into a polymeric solution [33]. The successful formation of particulate systems is achieved by establishing an optimum drug to polymer ratio as well as desired intermolecular interactions between the drug and the polymeric carrier matrix [85]. Although considered an advantageous method utilised to improve the bioavailability of poorly water-soluble drugs, polymers used in solid dispersions tend to absorb moisture, triggering a number of complications including phase separation, drug hydrolysis and crystal growth [86].

8

9 Due to the limitations associated with these particulate systems alone, it can be gathered that 10 by combining these particulate formulations with ES can potentially avoid drawbacks related 11 to conventional methods. Therefore, the introduction of ES in formulations can act as a 12 progressive step following final solution preparation to circumvent any limitations and achieve 13 highly monodisperse electrosprayed nanostructures.

14

15 4.2 Single-needle electrospraying

Single-needle ES is the most common and simple atomisation technique which employs a single spraying needle and can produce several types of particulate systems [87]. Upon mixing the polymer with an active agent (in a suitable solvent), a homogenous polymeric formulation is created, which, if complementary, forms a stable ES process and ideally a perfect Taylor cone, depending on the parameters discussed earlier (section 3). Single-needle ES shows great promise in various applications such as gene therapy [39], chemotherapeutic drug delivery [1] and targeted drug delivery [88].

23

24 4.3 Coaxial electrospraying

In addition to conventional Single-needle electrosprayed particles, further developments in the 25 26 engineering field have meant that other assembled ES systems have been utilised to yield 27 exceptionally functional particulate structures including various spinneret arrangements 28 (Figure 1a). These include coaxial and triaxial ES systems (commonly three needles aligned in 29 a co-axial configuration) as well as an array of different needle arrangements. In this review, 30 triaxial ES is similar in arrangement to coaxial tri-needle unless stated otherwise. These 31 complex ES techniques are used to prepare particulate systems with unique morphologies thus 32 further expanding the opportunities to fabricate intricate particulate systems.

33

1 The coaxial EHDA mode provides an efficient technique which can be utilised to produce 2 electrified liquid jets where possible resultant structures include core-shell, capsule and solid-3 shell solid-core formations. To assemble a coaxial ES arrangement, two concentric needles are 4 employed in conjunction, in which an outer liquid encapsulates the inner formulation. This results in microparticulate or nanoparticulate systems characteristically possessing particles 5 6 with near monodispersity [33, 89]. Capsulated structures are usually comprised of an aqueous 7 core solution encased in a larger organic shell solution, in which both formulations are 8 individually extruded through a bi-component needle system. The shell is usually made up of 9 the polymeric solution while the drug is encapsulated within the polymer, where both flowing 10 streams exit the dual needle configuration in an orifice. Various parameters can impact the experimental coaxial EHDA technique, in particular flow rates of both the core and shell 11 12 formulations which ultimately determine morphological and surface characteristics of the resultant capsule system [66]. In addition, miscibility of the core and shell solution is a key 13 14 characteristic which plays a vital factor in determining morphology of fabricated particles [90]. 15

16 The successful application of coaxial EHDA has been reported by Lee *et al* for the fabrication of a monodisperse polymeric coated core-shell structure in which nearly 100% drug 17 18 encapsulation efficiency was reported when coating Budesonide with poly(lactic-co-glycolic acid) (PLGA) [91]. Coaxial ES has been increasingly studied in recent years due to its 19 20 attractive characteristics including its ability to protect sensitive actives (as core materials) via 21 shell formation, improving drug encapsulation efficiencies, generating monodisperse particles, 22 enhancing drug stability, and achieving controlled drug release patterns [79]. The applicability 23 of this technique indicates the potential implementation of ES in various therapeutic or active 24 delivery fields such as tissue engineering scaffolds, [92] anti-cancer theranostics, [93] coatings and biomedical imaging [51]. 25

26

Multi-layered particles have also been fabricated by the ES mode where the transition to a coaxial set up entails a seamless arrangement of three coaxial needles [94]. A novel device containing this experimental set up was reported by Ahmad *et al*, where multi-layered bubbles, gas encapsulated threads and nanocapsules were generated and hence highlighted the numerous prospects of tri-needle coaxial spinnerets for drug delivery devices [95].

1 4.4 Other novel configurations of processing needles

Advancement in the realm of EHDA and specifically ES technology has allowed further
modification of more sophisticated needle variations which can be developed to form a plethora
of different microstructured or nanostructured particles.

5

6 Aligned needles have been designed with novel non-concentric spinnerets and angled tips at 7 the needle outlet to form drug encapsulated Janus particles (particles that display 8 multifunctional surfaces). Zhang et al, for instance, found that the utilisation of an aligned non-9 concentric angular nozzle resulted in minimal contact at the cone area (30°), whilst a nozzle angle of 60° produced stable co-jetting atomisation with continuous flow and eventual break-10 11 up at the needle apex. This led to the formation of Janus particles with desired morphological 12 properties, where the model dye (Sudan Red G) and the drug (Indomethacin) were both 13 successfully encapsulated into the Janus particulate system. The resultant particles displayed 14 independent release patterns, hence demonstrating the potential for tailored combinatorial drug 15 release [96]. In comparison to aligned needles, multiple needles can also be prepared 16 possessing angular heads and a large variation of tips which can be modified to optimise drug 17 encapsulation and release profiles, thereby indicating potential use for drug delivery 18 applications.

19

20 In contrast to the utilisation of needles in the EHDA set-up, needleless approaches have also 21 been investigated as a more cost-effective method to fabricate functional particulate systems. 22 This type of process is reliant on jet formation from open liquid surfaces with the use of external 23 forces [25]. In one study a two-layered formulation system was subjected to an electric field 24 and additionally a magnetic field by the employment of a magnet or coil, resulting in solidified 25 nanofibers [97]. A needleless electrospray apparatus was also prepared by Wang et al in which 26 a spiral tower was employed to atomise polyvinylpyrrolidone (PVP)/ Fe₃O₄ ferrofluid. The 27 addition of an external magnetic field in combination with an electric field formed multiple 28 cone-jets. This ultimately resulted in a magnetic film composed of the PVP polymer in which 29 Fe₃O₄ nanoparticles were homogenously distributed [98]. Alternatively, needleless devices can 30 also include the utilisation of orifices. Bocanegra et al for example, drilled orifices in dielectric 31 materials with hydrophobic properties. The hydrophobic characteristics of the dielectric 32 materials provided an optimal Taylor cone-jet, successfully resulting in multi-electrosprays in 33 up to 37 holes which displayed size distributions comparable to single needle ES [99]. 34 Needleless approaches are therefore highly effective as they exhibit an extremely simplistic process without the need of a feed unit and reduce complications associated with conventional
 tips (nozzle clogging within the ES set-up).

3

4 During the ES process, the application of an electrostatically charged jet at the nozzle exit 5 results in the deposition of particles on a grounded substrate positioned under the tip of the 6 capillary needle exit which serves as a collector [16]. Collecting substrates can be made up of 7 a variety of materials including silicon, glass, aluminium and copper, where a conductive 8 substrate is preferential as it limits the deposition of particles to the charged area [33]. The cone 9 apex can break up into charged droplets, depending on the attraction of the formulation 10 contained within the jet to the substrate collector [26].

11

12 It can therefore be concluded that various particulate structures can be engineered by fine-13 tuning the components of the ES set-up. Nozzle geometry as well as the use of multi-needle 14 spinneret arrangements have a significant effect on the subsequent particle characteristics 15 which can be modified for optimal drug delivery, targeting or imaging aspects. Substrate 16 adjustments can also alter resultant particulates. Modifying these components can ultimately 17 generate complex microparticles or nanoparticles for targeting specific anatomical sites, all of 18 which will be discussed further in this review (section 8).

19

20 5. Engineered particulate structures21

22 Manipulation of the EHDA system, (specifically for ES) to deposit microparticulate and 23 nanoparticulate materials can potentially fabricate an array of specialised structures (Figure 3). 24 The unique conformation of such particles including intricate features distinguish them from ordinary matrix based particulate structures. This is due to significant control over their shape, 25 26 surface morphology, composition and overall particle chemistry. Technological advancement 27 in a number of novel pharmaceutical applications has meant that the synthesis of desired 28 nanoparticles of certain shapes and sizes are significant to complement drug delivery devices 29 and enhance the particle engineering process [100]. The ES technique has been previously 30 demonstrated as possessing the capability to produce smooth spherical particulates [101], 31 [102], [103]. However, by the exploitation of various parameters and the adjustment of the 32 spraying nozzles employed, a variety of decorated particulate structures can be fabricated [104]. These particulate systems can potentially exhibit several morphologies [105] including 33

- donut shapes [106], nanocups [107], hollow microspheres [108] as well as cell-mimicking 1
- 2 particles [109] as demonstrated in Table 1.
- 3

4 5.1 Active polymer composite systems

ES is an advanced emerging technique for the preparation of microparticles or nanoparticles 5 6 (including microspheres or nanospheres and microcapsules or nanocapsules) [110]. Some 7 delivery systems only comprise two materials once the base solvent has evaporated: the 8 polymeric matrix and API. The polymeric matrix acts as a carrier in which the active is 9 dispersed or embedded. Ibuprofen has been embedded into zein microstructures using ES. 10 Here, Li et al coated an epoxy resin around a traditional 20G needle tip for engineering. 11 Conventional ES employing a standard 20G stainless-steel needle decreased the productivity 12 of the process due to clogging. In comparison, implementing an epoxy coated spraying head 13 prevented any clogging at the needle tip and ultimately generated microparticles exhibiting 14 homogenous structures and a narrow size distribution. In-vitro dissolution tests of 15 electrosprayed ibuprofen microstructures demonstrated a sustained release profile with a 16 smaller burst release when compared to traditional ES (stainless-steel needle). Active-polymer 17 loaded structures (into a single matrix material) can therefore be successfully developed to expand the applications of ES to novel drug delivery research [111]. Furthermore, ES paclitaxel 18 19 as a polymeric particle system for the potential treatment of malignant glioma has been shown. 20 In this study, Xie *et al* used paclitaxel and various polymer types (PCL and PLGA) as well as 21 different polymer ratios to fabricate microparticles containing the active and a polymeric 22 material. Cell cycling studies indicated promising results including an 80% encapsulation 23 efficiency for all samples and a sustained release profile over a period of 30 days, therefore 24 demonstrating ES as an efficient technique to fabricate an array of drug delivery systems based 25 on a drug-polymer composite system [36].

26

27 5.2 Active-multi excipient composite systems

28 Composite structures comprising multiple excipients and the API can also be prepared via ES. 29 These then transfer functional properties of excipients into nano or microparticulate systems.

- 30 For instance, timolol maleate was encapsulated within various polymers and solidified chitosan
- 31 was employed as a permeation enhancer, which when atomised using ES, resulted in the
- 32 development of stable ocular lens coating. The flexibility to modify the polymeric formulations
- 33
- allowed controllable release of timolol maleate for improved permeation through the cornea
- 34 and highlighted the advantage of ES for polymeric based dosages [112].

In addition, a study by Liu *et al* utilised the coaxial ES method, where an epoxy coated concentric spray was used for the fabrication of core-shell microparticles. In this, nanocomposites were formed from a PVP matrix and acyclovir distributed within the inner core, while the outer core consisted of sucralose and the organic compound sodium dodecyl sulfate which acted as a transmembrane enhancer. *In vitro* dissolution studies indicated that the solid dispersion composites rapidly released the active, thereby enhancing water solubility of acyclovir as well as increased permeation across porcine sublingual mucosa [113].

8

9 5.3 Multi-layered structures

ES as a technique for preparing multi-layered structures for drug delivery-based systems is a complex yet effective approach. This advanced technique can improve the size and morphology of polymeric matrix carriers, including the formation of either liquid or solid layers. Multilayered structures can be prepared using the coaxial ES mode consisting of a triple-needle device. Labbaf *et al* successfully demonstrated the fabrication of spherical multi-layered particles by the employment of three liquid streams (containing the polymers PLGA, polymethylsilsesquioxane (PMSQ) and polycaprolactone (PCL)), which flowed

17 simultaneously and resulted in a cone-jet at the needle orifice. A near monodisperse particulate 18 system was achieved by the addition of three layers all combined in a single-step process. The 19 prepared tri-layered system was found to have applications in sustained and prolonged drug 20 release [114]. In addition to this, microencapsulation techniques have also been incorporated 21 in the ES method to develop multi-shell capsules as shown by Kim *et al.* In this study, three 22 immiscible flowing liquids (ethylene glycol, 4-hydroxybutyl acrylate and olive oil) when 23 atomised using the ES set-up, resulted in multi-shell encapsulation [115]. Multi-layered 24 encapsulated structures can provide valuable characteristics including homogenous monodisperse size distributions, tailored drug release patterns and a protective core in hostile 25 26 environments (extreme pH levels, enzymatic degradation and temperature) for sensitive 27 therapeutic agents.

28

29 5.4 Live-cell entrapment based structures

Living cells existing abundantly in nature can be entrapped within an ES set-up (more commonly referred to as bio-ES) and represents a revolutionary technique in the field of drug delivery research. The EHDA method requires ambient conditions of temperature, humidity and pressure, and is therefore a beneficial process for sensitive biomaterials and living cells [40]. This has been explored by Ma *et al* where pancreatic islet cells were isolated from

1 Sprague-Dawley rats and encased within core-shell hydrogel microcapsules by the coaxial ES 2 mode. A simplistic atomisation step allowed islet cell entrapment within the core region of the 3 microcapsules whilst masking the cells, resulting in immuno-protective properties and 4 improved encapsulation [116]. Bio-ES can be employed for the fabrication of potential carriers 5 such as biological materials including bacteria and viruses. In one study, adenovirus was 6 encapsulated within alginate beads by the ES process. ES was shown to be an effective 7 approach due to tuneable characteristics in which alginate concentrations, flow rate and voltage 8 could be adjusted to optimise the fabrication of cross-linked adenovirus alginate beads [117]. 9 As a result, adenovirus exhibited release in a controlled manner from the alginate carrier over 10 a period of seven days to target cancerous cells. This study emphasised the role ES plays in 11 improving protection and delivery for virion related release and could therefore have potential 12 applications for advanced technologies in vaccine development [118].

13

14 5.5 Cell shaped particles

15 The unique approach of using polymeric particles that mimic cells can be utilised in biological 16 research to reduce intracellular toxicity and cell variations.

17

The optimisation of several characteristics of solutions involved in the ES process (e.g., solvent 18 19 evaporation rate, density and surface tension) can lead to the production of different particulate 20 structures such as cell shaped particles. For instance, one study found a positive correlation 21 between particle morphology and physical properties of the solution (e.g., solvent evaporation 22 and polymer diffusion) [119]. In situations where the solvent rapidly evaporated from the 23 droplet surface at the needle apex of the ES set-up, Lee *et al* found that the generated specialised 24 structures possessed a biconcave discoidal shape comparable to that of the human red blood cell. Uniform red blood cell shaped particles were also fabricated during ES using the 25 26 biopolymer chitosan [109]. Ju et al developed a stable process by monitoring solvent 27 evaporation during ES, where they identified that the concave-like morphology of particulates 28 was obtained due to solvent diffusion which occurred prior to the deposition of electrosprayed 29 particles on the grounded substrate [109].

30

31 5.6 Spindle shaped particles

32 In addition to conventional spherical particle shapes, other shape of particulate structures can

be fabricated *via* ES. ES can generate particles with novel structural configurations including
 anisotropic particles (particles that possess unique shapes with directional interactions) (e.g.

spindle-like structures) and non-anisotropic morphological particles (e.g. rod shaped particles).
Spindle-shaped particles have been fabricated in the field of nutraceuticals by a study
conducted by Khoshakhlagh *et al.* In this, researchers investigated the atomisation of an
emulsion formed between D-limonene, Alyssum homolocarpum seed gum and Tween 20,
which, when electrosprayed, under high electrical conductivity resulted in spindle-like
morphologies and ultimately nanocapsules. These heterogenous structures demonstrated high
encapsulation efficiencies (73.4%) and improved storage stability in harsh conditions [120].

9 5.7 Rod shaped particles

Other non-anisotropic morphological structures including rod, disc and toroidal shaped 10 particulate systems can be prepared using ES. Biodegradable microparticles were fabricated by 11 12 Bhaskar et al employing a side-by-side ES configuration. The prepared particles were generated by the co-jetting atomisation technique using PLGA polymers which resulted in 13 14 distinct shapes including rods, discs and spheres. In particular, the formation of a Taylor cone 15 at the needle apex resulted in rapid evaporation (solidification of the travelling jet) and 16 consequently the fabrication of rod-shaped particles. These novel particles can be utilised for 17 several multifunctional applications in drug delivery and medical imaging [121].

18

19 5.8 Donut shaped particles

Donut-like structures are another type of particulate system demonstrating unique morphology 20 which can be engineered by the ES method. Xie et al used the EHDA system to prepare 21 22 paclitaxel loaded polymeric microparticles for local and sustained delivery. Different solvents 23 were utilised all of which when atomised, resulted in a variety of morphologies including donut 24 shaped particles, spheres and corrugated particles. These particles inhibited the proliferation of 25 C6 glioma cells, thereby reducing the malignant tumor growth rate. The resultant 26 electrosprayed drug delivery devices demonstrated sizes ranging from 100 nm-10 µm and an encapsulation efficiency of $\sim 80\%$ hence finding applications in sustained delivery of 27 28 anticancer therapeutics [36]. Single and coaxial ES were also employed for the preparation of 29 Eudragit L100-5 polymeric nanoparticles loaded with prednisolone. When electrosprayed, toroidal donut-like structures were fabricated possessing a narrow size distribution. Dissolution 30 studies revealed a site-specific release of prednisolone for the targeted treatment of 31 32 inflammatory bowel disease and colorectal cancer [106].

33

1 5.9 Nanocups

Titania nanocups were engineered by the employment of titania isopropoxide (TIP) with 2 3 polyvinyl acetate (PVAc) [107]. The combination of these materials when electrosprayed 4 resulted in the formation of nanoparticles, where a depression formed in the solid structures 5 generated a particulate system that displayed definitive cup-like morphology. The unique structures had a size distribution ranging from 200-500nm. Additionally, the model protein 6 7 bovine serum albumin (BSA) was successfully loaded into the nanocups, where loading and 8 release profiles of the nanocups demonstrated sustained release. Electrosprayed titania 9 nanocups displayed higher protein adsorption and a gradual release compared to Titania nanoparticles and can therefore be considered as a novel system for protein drug delivery [107]. 10 11 Similarly, Park et al also synthesised titanium oxide nanocups via ES using titanium 12 tetraisopropoxide and the synthetic polymer polymethylmethacrylate (PMMA) as prospective 13 drug delivery devices. The appearance of the particles demonstrated cup-like profiles where 14 cavities in the structures were clearly observed, these were potentially generated during ES 15 where solvent evaporation caused phase separation [122].

16

17 5.10 Hollow microspheres

ES can also be utilised to generate different types of hollow polymeric micropsheres. For 18 19 example, Chang et al prepared PMSQ hollow polymeric microspheres which encapsulated the core liquid perfluorohexane (PFH). The close monitoring of certain parameters included in the 20 21 ES set-up resulted in different particle properties including variations in particle diameter, shell 22 thickness and particle uniformity. This demonstrated the capability of EHDA to successfully 23 obtain a versatile range of hollow microspheres [108]. These types of particulate systems can 24 be utilised in microencapsulation techniques for the protective coating of various molecules, to 25 control their release as well as shielding them from extreme environments. Thus, these systems 26 can be employed as controlled drug delivery systems for various applications in the medical 27 and biological industry [123].

28

In a study by Zhou *et al*, coaxial ES was employed using a formulation consisting of PCL/chloroform (5 wt. %) (shell) and PEG/chloroform (15 wt.%) (core). In this, researchers reported the use of various solutions which when accompanied with the coaxial ES atomisation method, fabricated hollow particulate systems. Further experimentation using ethanol as a collecting substrate formed single-hole hollow microspheres due to solvent evaporation from the needle apex and eventual desolvation of the PCL carrier, consequently leading to hollow-

like structures. The research study highlighted the importance of hollow microparticle based
 systems for an array of applications. For example, hollow microspheres could function as tumor
 cell mimicking phantoms, designed to replicate characteristics of biological material and can
 therefore be identified as crucial structures for anti-cancer therapeutics [124].

5

6 5.11 Porous microcarriers

In addition, other structural conformations which can be fabricated by ES include particles 7 8 which display a well-connected network of pores on the solidified surface [125]. Wu et al 9 investigated a novel ES technique, where they studied the influence of applying various 10 atmospheric pressures in contrast to the conventional ES method which employs ambient 11 pressure conditions. By tuning the ES parameters discussed earlier (section 3), solvent 12 evaporation and subsequently the solidification process, resulted in PCL polymeric particles 13 which exhibited porous geometry on the surface. Researchers deduced that the porous structure 14 of the particles was a result of phase separation at the spinneret which contained an 15 electrostatically charged jet. It was concluded that adjusting the atmospheric pressure utilised 16 during ES can be seen as a vital parameter and is particularly useful for sensitive drug delivery 17 methods [126]. In addition, porous particulate systems can also be fabricated for other applications, for example, ES was utilised as a versatile technique for the preparation of 18 19 inhalable porous microspheres. In this study, an anti-inflammatory drug (oridonin) was loaded 20 into polymer coated microspheres which resulted in porous-like structures during ES. The 21 porous structures demonstrated high anti-cancer effects (attributed to high lung deposition of 22 oridonin-loaded electrosprayed porous microspheres) due to their geometry. The delivery 23 system demonstrated high efficiency of inhalable drugs deep into the lungs and the subsequent 24 rapid release of them to the surroundings. Inhalable porous microcarriers could therefore be 25 utilised for applications in targeted drug delivery for the treatment of lung carcinomas [37].

- 26
- 27 5.12 Janus particles

Janus particles are types of unique microparticles or nanoparticles which contain anisotropic structures where the surface is exposed to the environment. Due to the surface chemistry of these particles, Janus shaped structures have an array of functional characteristics which can be applied to a diverse range of particle engineering fields. Rahmani *et al* designed nanoparticles with controlled characteristics using electrohydrodynamic co-jetting. Fabricated Janus particles exhibited a mean diameter of 105.7 nm and contained additional functional groups on the particle surface for further modification. Hence, bicompartmental nanoparticles

1 like Janus structures can be successfully applied as drug delivery carriers as well as diagnostic 2 imaging agents [127]. Others have gone on to further strengthen the claims of utilising Janus 3 particles for use as imaging agents in biomedical applications. Uniformly sized 4 compartmentalised Janus particles containing both rose Bengal dye and the anti-cancer drug 5 carmofur in separate compartments were prepared using side by side ES. The particles demonstrated a narrow size distribution, and the great potential these particulate systems 6 7 exhibit since their surfaces can be modified to include several sections, which are tailored for 8 segmented targeting for a number of drug delivery vehicles and photodynamic therapy [128].

9

10 5.13 Other unique morphologies

11

12 5.13.1 Strawberry shaped particles

A novel method for the preparation of particulates was also reported in which silver 13 14 nanoparticle-doped SiO₂ microspheres were generated using the EHDA technique [129]. 15 Scanning electron microscopy images revealed peculiar morphologies of particles which were 16 strawberry-like and mirrored aggregates of achenes found on the surface of strawberries. The engineered particles demonstrated efficacious inhibitory effects against the Escherichia coli 17 bacterium due to the silver nanoparticles embedded within the microspheres. This resulted in 18 19 increased antibacterial activity of the fabricated structures, these structures could therefore be 20 utilised as antibacterial materials for biomedical applications.

21

22 5.13.2 Fibril shaped particles

Particles resembling fibril-like shapes can also be prepared using the ES method as shown by Khanum *et al.* In this study, an array of solvents and a thiophene derivative (7,9-di(thiophen-2-yl)-8H-cyclopenta[a]acenaphthylen-8-one) (DTCPA) were used in the ES process to generate spike-spheres and spike morphologies which imitated fibril particulate structures. The study brought to light how the ES process can be potentially employed for the novel fabrication of organic molecules, and thus could be applied to photoactive materials for biomedical applications [130].

30

31 5.13.3 Yolk-shell particles

32 Recently a novel EHDA system consisting of a tri-needle coaxial device was developed. By

33 the use of ES, Zhang *et al* fabricated a multicomponent particulate system consisting of

34 particles that displayed unique yolk-shell like morphology. These were comprised of magnetic

1 nanoparticles, silicone oil and a polymeric core. Multiple model probes when co-encapsulated 2 into these particulate systems demonstrated an array of release profiles which could be fine-3 tuned via the employment of an external auxiliary magnetic field. Owing to their 4 compartmentalised structure, the engineered magnetic polymeric yolk-shell particles could be 5 implemented for multiple drug loading as well as applications in dual imaging modality [131]. 6 The delicate interplay of several parameters involved in the ES technique allows the precise 7 control of the size, size distribution, chemical compositions and morphological characteristics 8 of the particulate system. Effects of parameters such as material modification, nozzle design, 9 types of substrates used, voltage, flow rate and working conditions (e.g., temperature, humidity 10 and pressure) all contribute to the several types of architectural particulate structures that can be developed. It can therefore be concluded that ES provides an exceptional platform to 11 12 fabricate an array of elegant structures all of which have been thoroughly discussed above as well as their applications in a plethora of drug delivery fields. Hence, ES is elucidated as a 13 14 prospective facile technique for the future development of opportunistic particulate systems for addressing significant challenges faced by the pharmaceutical engineering industry. 15

16 **6. Active agent selection**

17

Microparticles and nanoparticles are attractive drug delivery systems that can be employed for 18 19 a wide range of applications. They offer interesting advantages including a high surface area, 20 ability to encapsulate large amounts of active molecules, biodegradability and capability of 21 achieving controlled release [132]. The use of ES to produce microparticulate and 22 nanoparticulate structures provides a plethora of advantages as it is a one-step, versatile system 23 which operates under ambient conditions. This allows an increased number of actives and 24 biomaterials (e.g. proteins) [133] and other APIs which are sensitive to elevated temperatures 25 or shear stress to be processed and encapsulated within a system [134]. EHDA and specifically 26 ES, have a significant impact on API entrapment, solubility and release which will be 27 discussed.

28

29 6.1 Choice of active pharmaceutical ingredient and other molecules

The aqueous solubility of APIs is critical for drug efficacy, specifically when the drug is to be orally administered. Low solubility results in inconsistent absorption, decreased bioavailability and slower onset, thus limiting their use in pharmaceutical applications. A handful of methods have been developed over the years to improve solubility through modification at either the

1 molecular, colloidal or particulate level. ES works by modification on a particulate level and 2 more specifically via nanonisation and amorphisation. Drugs with a low solubility are often 3 processed into microparticle or nanoparticle formulations to enhance their low bioavailability. 4 A substantial number of APIs are poorly water soluble, however through use of ES in 5 combination with a formulation consisting of a stable emulsion (a hydrophilic polymer, 6 surfactant and water-insoluble API), it is possible to produce microparticles with improved solubility [68]. Moreover, ES is an interesting technique with great capability of enhancing 7 8 the solubility of poorly water-soluble actives via amorphisation. Through the use of electrical 9 forces, formulation containing the API is atomised upon which the solvent rapidly evaporates, 10 and the droplets almost immediately solidify. APIs are left in an amorphous form, partially due 11 to the rapid solvent evaporation. When in an amorphous form, APIs have increased free energy 12 as well as a larger surface area due to the submicron particles, thus improving the solubility of poorly soluble molecules [66]. For example, Bohr et al loaded the poorly soluble drug 13 14 Celecoxib into PLGA microspheres using ES. Resultant microspheres were near-monodisperse 15 (1-5 µm) and displayed a smooth morphology. Differential scanning calorimetry confirmed 16 that the Celecoxib was entrapped in an amorphous form in electrosprayed PLGA microspheres. Celecoxib loaded microspheres showed a bi-phasic release pattern (initial burst release 17 18 followed by sustained release). This research demonstrates the potential use of ES in entrapping 19 poorly soluble drugs into particulate drug delivery systems and as a result improving their 20 solubility [135].

21

22 6.2 Stability of biomolecules and larger entities

Therapeutic proteins for pharmaceutical applications are problematic to process due to their specific properties and functions. Often methods (e.g. primary emulsion) of manufacturing and processing proteins result in protein denaturation and aggregation as they deploy harsh conditions. In contrast, ES encapsulates actives under ambient temperature and pressure making it a favourable method for processing sensitive drugs or biomolecules (proteins, peptides and cells).

29

For example, Coaxial ES was used in several studies to encapsulate different types of proteins into biodegradable polymeric microcapsules under ambient conditions, overcoming the limitations associated with the primary emulsion technique [136], [133] and [137]. ES was used by Yu Fuki *et al* to electrospray a sodium alginate aqueous solution directly into a 0.5 wt % chitosan aqueous solution resulting in the fabrication of polyelectrolyte complex

1 microcapsules. The produced microcapsules exhibited a narrow size distribution and controlled 2 diameters ranging from 80-230 µm. The research group were successful in encapsulating 3 protein, dextran and a polymeric microsphere individually within the polyelectrolyte complex 4 with encapsulation efficiencies >99%. Yeast was also encapsulated in the polyelectrolyte 5 microparticles via ES, where it was found that the encapsulated yeast preserved its activity. 6 This shows the possible advantage of using ES for the encapsulation of physiologically active 7 substrates as their activity is not hindered during the process [138]. Enzymes are other sensitive 8 biomolecules that can be encapsulated using ES in order to improve their stability and 9 bioavailability. For example, Fung et al successfully electrosprayed Coenzyme Q10 loaded 10 Copovidone microparticles (3–5 µm) to improve Coenzyme Q10 solubility and bioavailability. 11 *In-vitro* studies showed a reduction in both the crystallinity and particle size which could 12 improve bioavailability. Dissolution studies and in-vivo oral bioavailability were assessed 13 using a murine model, where the electrosprayed particles demonstrated improved dissolution 14 properties and oral bioavailability respectively when compared to both the raw materials and 15 physical mixture [139].

16

17 6.3 Encapsulation of small and volatile/sensitive actives

Nanoencapsulation is the process by which a bioactive compound is loaded or entrapped into a carrier matrix or material in the nano range [140]. ES allows for increased encapsulation efficiency owing to the absence of an external medium which permits migration or dissolution of water-soluble molecules [137]. Thus, ES is a useful technique in producing a drug loaded particulate based delivery systems with high payloads.

23

24 Loading of APIs into mesoporous materials (such as silica) has been a favourable technique to 25 improve drug solubility, permeability and bioavailability [141]. However, conventional 26 loading techniques that are used to load drugs into a mesoporous matrix usually lead to low 27 entrapment efficiency. Recently, ES was explored as a loading technique to incorporate poorly 28 water-soluble drugs into a mesoporous silica matrix. Sayed *et al* utilised ES to encapsulate 29 KAZ3; a novel poorly water soluble chalcone with anticancer properties, into mesoporous 30 materials (SBA-15 and MCM-41). This loading method was compared with the conventional 31 common solvent impregnation loading process. It was found that through using the ES 32 technique, KAZ3 was successfully loaded within the pores of the silica particles with a high 33 encapsulation efficiency in an amorphous form. Solvent impregnated formulations indicated a 34 lower encapsulation efficiency and partial crystallinity of the loaded drug. In addition, drug

dissolution studies demonstrated a 30-fold improvement in drug dissolution for the ES loaded particles when compared to the pure drug. Moreover, *ex-vivo* studies showed formulations to have increased the permeability across murine intestine in comparison to the solvent impregnated formulations or pure drug. This research, for the first time, highlights the ability of ES to produce loaded mesoporous silica particles with high payloads, an improved dissolution profile and enhanced permeability across the intestine [1].

7

8 6.4 Drug loading

Limited drug delivery efficiency occurs as a result of poor encapsulation, however, ES has 9 10 been used as a prospective technique to increase encapsulation efficiency whilst protecting 11 drugs from degradation [63]. Often, polymeric and lipid nanoparticles are used as a means of 12 transport as well as encapsulating APIs and proteins, largely because they possess the capacity 13 to protect the active from environmental conditions which often results in degradation [132]. 14 In a study by Zamani et al, BSA was used as a model protein and loaded into PLGA 15 nanoparticles via two different ES methods; co-axial ES and emulsion ES. It was found that 16 the solvent, PLGA molecular weight, and PLGA concentration influenced the particle 17 diameter, producing particles between 3.0-5.5 µm. The electrosprayed particles displayed a core-shell structure, where the encapsulation efficiency of structures fabricated via co-axial ES 18 19 was found to be significantly higher (69.2% and 71.8%) than those electrosprayed using 20 emulsion ES (53.5% and 46.7%) for both high and low molecular weight PLGA respectively. 21 In the initial 24 hours, a small burst release of 8-12% was displayed in emulsion electrosprayed 22 particles, in comparison, those which were co-axially electrosprayed had a core-shell structure 23 which demonstrated a release of 24-27%. This study highlights the capability of the ES process 24 to fabricate protein particles which display both a uniform size distribution and high 25 encapsulation efficiency making them advantageous for use in the drug delivery remit [142].

- 26
- 27 6.5 Release kinetics

ES has shown to control the shape, size and morphology of fabricated particles, all of which heavily impact the rate of degradation, drug diffusion and therapeutic effect of actives [143]. Formulation scientists have employed various strategies to control the active release and dissolution in order to improve their solubility, to sustain their release or to target the site of action. These strategies include the use of amorphous solid dispersions [144], inclusion into mesoporous silica [1], nanosizing [145], and ES [146]. Occasionally, a targeted regiospecific release is required, for example, when site specific administration is necessary in the

1 gastrointestinal tract. Coaxial ES is advantageous in the processing of such formulations as it 2 fabricates monodisperse micro sized or nano sized amorphous solid dispersion particles. These 3 particles can be subsequently coated with a responsive polymer, ultimately dissolving and 4 releasing the encapsulated active at the required site. For example, Smeets *et al*, utilised coaxial 5 ES to fabricate core-shell microparticles in which an amorphous solid dispersion of hydroxypropyl methylcellulose (HPMC) or PVP based matrix was used. The antiviral drug 6 7 Darunavir was coated using a gastro-resistant polymeric coating (co-polymer poly(methacrylic 8 acid-co-methyl methacrylate)) which is insoluble at a low pH of the stomach but dissolves at 9 higher pHs of the intestine. Thus, the polymeric coating prevented the premature release of the drug in the stomach, protecting it from degradation and ensuring delivery of the active to the 10 11 site of absorption (intestine). This research demonstrates the applicability of coaxial ES in the 12 fabrication of core-shell particles as a delayed release dosage form [144].

13

14 6.6 *In-vitro* and *In-vivo* performance of actives

15 Aforementioned, through careful adjustment of process and solution parameters, numerous particulate systems with different structures can be obtained [40]. Hao et al, employed coaxial 16 17 ES for the fabrication of aspirin loaded nanoparticles with an enteric coating for sustained 18 release. A pH responsive polymer (Eudragit L100-55) was used as an outer coating while a 19 sustained release polymer (Eudragit RS) was chosen for the inner core. Through altering the 20 flow rate of both the inner and outer solutions, a maximum loading capacity of 23.66% was 21 observed for the nanoparticles and an entrapment efficiency close to 100% was achieved. In-22 vitro release was carried out via a method previously used by Hosny et al. [147]. In-vitro release 23 studies demonstrated both pH sensitive and sustained release which occurred with less than 5% 24 of the aspirin being released in the gastric simulated fluid. A change in the pH to 6.8 resulted 25 in a substantially increased release rate. Sustained release was observed in the simulated 26 intestinal fluid where more than 95% of the aspirin was released from the nanoparticles over 27 120 hours (5 days). This study demonstrated how coaxial ES can be exploited to produce core-28 shell nanoparticles with an enteric coating to provide sustained release whilst also protecting 29 the stomachs delicate lining from the side effects of aspirin [148]. A site-specific delivery 30 system consisting of gambogic acid nanoparticles was produced using ES. A poly(D, L-Lactic 31 Acid (PDLLA) matrix was used and gambogic acid was successfully encapsulated. In-vivo 32 liver distribution studies were carried out alongside pharmacokinetic profiles and anti-tumor 33 efficacy tests for hepatocellular carcinoma. Results highlighted the advantages of using ES to 34 fabricate gambogic acid particles to attain substantial antitumor efficacy as well as overcoming

limitations associated with gambogic acid such as poor water solubility and side effects due to
 its pharmacological toxicity [149].

3

Through the above-mentioned studies, ES can be demonstrated as a versatile technique which can be used to process several APIs and sensitive biomolecules. Thus, the ES process enables particle sizing and structure enhancement to improve solubility of APIs, therefore improving bioavailability as well as obtaining desirable drug release profiles. In addition to this, ES can be used to encapsulate sensitive and toxic molecules to protect them from the biological environment and minimise adverse drug reactions respectively.

10

11 7. Dosage design

12

13 Dosage design plays a key role in developmental and engineering processes for active 14 particulate systems. This entails the consideration of several physiochemical and biological 15 characteristics of the drug as well as their ultimate targeted application when fabricating drug delivery systems. Establishing the required dosage design during ES involves the careful 16 17 monitoring of relevant parameters (e.g., polymer matrix concentration, solution type, flow rate 18 and applied voltage) which ultimately define the physiochemical characteristics of the resultant 19 particulates. ES can serve as a prospective technique where the interplay between parameters can fabricate both nanostructures and microstructures with improved biopharmaceutical 20 21 performance, ultimately spanning several targeted therapies. These can include an arsenal of 22 different systems such as matrix, multi-layer encapsulation and bio-responsive strategies.

23

24 7.1 Matrix systems

Matrix drug delivery particulate systems generally implies delivery devices in which the API 25 26 is homogenously dispersed either molecularly or in the solid state into a polymeric carrier. 27 Formulating drugs as a matrix type dosage form can aid in attaining different release profiles 28 and is particularly useful for drugs with short half-lives which are eliminated from the 29 bloodstream at a quicker rate. For instance, a study by Cavalli et al investigated the use of the 30 ES technique to fabricate spherical poly(amidoamine) (PAA) PAA-cholesterol nanoparticles 31 by carefully fine-tuning the applied voltage and flow rate parameters. In addition, tamoxifen, 32 an anti-cancer therapeutic for the treatment of breast cancer was molecularly dispersed within 33 the PAA-cholesterol conjugate matrix. The nanoparticles exhibited a drug loading efficiency

of 40% and a prolonged drug release profile. The study highlighted the effectiveness of ES as a credible technique for fabricating matrix-based system nanoparticles. More specifically, the work was conducted in a single-step without additional excipients and therefore concluded as a cost-effective method which can be implemented in the commercial industry [150].

5

7

6 7.2 Layered particulate systems

Dosage forms designed with layered particles can provide a vast array of advantages; a 8 9 protective shell to sensitive bioagents embedded within the core and encapsulating drugs within 10 a layered system to enable a variety of release kinetics. The coaxial ES method can be utilised 11 for the production of core-shell particulate structures in the nanometre or micrometre size range 12 [151]. These innovative structures play an important role in designing dosage forms with 13 required attributes for the pharmaceutical industry. Core-shell particles can aid in formulating 14 dosage forms for several purposes including; protecting actives from degradation (stomach 15 acidity), an array of release profiles for example, immediate release [152], targeted release [153], and sustained release [154] or co-delivery of different actives [155]. For instance, a fast 16 17 onset dosage form can be designed by ensuring an immediate release of the active by coating 18 it with a thin layer of fast-dissolving polymeric shell [152]. In contrast, a sustained dosage form 19 can be obtained using a slowly degrading solid lipid shell [154]. Moreover, a delayed onset 20 dosage form with a targeted therapeutic action can be attained by coating the core structure with a responsive polymer that only dissolves at the required site of action under the response 21 22 of physiological stimuli [144]. Layered particulate structures are also promising in combining 23 different actives in a single dosage form, thus capable of achieving delivery of different types 24 of actives simultaneously [155].

25

26 Yu et al electrosprayed helicid loaded nanoparticles in order to improve its poor aqueous 27 solubility, bioavailability and onset of action. A core-shell structure was fabricated consisting 28 of the biopolymer shellac at the core enveloped by a PVP-helicid shell, ultimately resulting in 29 solid composite nanoparticles. In-vitro studies demonstrated a faster dissolution of helicid from 30 the core-shell particles due to the presence of the active which exhibited amorphous characteristics within the PVP layer. Moreover, the presence of a thin solid nanocoating on the 31 32 particles enabled immediate release of helicid (one minute) compared to raw helicid where 33 only $11.4 \pm 4.2\%$ of the active dissolved into the dissolution medium (after a period of 30 34 minutes). The results of the study indicated that the core-shell matrix composites displayed

characteristics of rapidly dissolving dosage forms and could therefore alleviate symptoms at a
 faster rate when processed using ES, specifically employing a dual concentric spray nozzle
 [152].

4

5 A novel EHDA technique was proposed by Labbaf *et al* but rather consisting of a four-needle 6 device in comparison to the conventional single conductive needle utilised during ES. A four-7 layered particulate system was developed simultaneously, in which each layer was comprised 8 of different organic polymers, namely: PLGA, PCL, PMSQ and PEG. When electrosprayed, 9 particles displayed a smooth spherical morphology, a mean particle size of 620 ± 150 nm and 10 polydispersity index of 26%. To determine if four layered structures had potential significance in the pharmaceutical remit, dye release studies were conducted, revealing 11 12 different release rates due to a variation in thickness of each polymeric layer. It was elucidated from the study that modern therapeutics could benefit from a multilayer delivery system as a 13 14 controlled release dosage form could be generated by adjusting the ES processing parameters 15 [101].

16

Similarly, multi-layered structures were generated using a triaxial electrospray system by Kim 17 18 et al. In this study, triple layered capsulated shells were prepared with the use of acetonitrile 19 and trifluoroethanol solutions containing biocompatible polymers (PLGA and PDLLA) in a 20 single-step process. The assembled system consisted of a triaxial nozzle where the generation of a stable Taylor cone at the needle apex resulted in multi-shell capsules. The anti-cancer 21 22 drugs Doxorubicin and Paclitaxcel were loaded in the innermost and intermediate shell 23 respectively, with a high entrapment efficiency (>80%). *In-vitro* release profiles for both drugs 24 from fabricated capsules demonstrated zero-order release kinetics with minimal burst release. 25 The retention of both anti-tumor drugs within the shell of the capsulated structures could be 26 manipulated by varying the shell thickness and mean diameter. This was achieved by fine-27 tuning the flow rate parameter of the ES set-up as well as the concentration of the polymeric 28 formulation. Thus, it was gathered from the study that ES is an amenable process where 29 tailoring the solution (polymer concentration) and process (flow rate) parameters can result in 30 multi-layered structures for applications in controlled drug release [156].

31

32 7.3 Targeted systems

Different active agents can be utilised during pharmaceutical engineering to generate dosage
 forms with a plethora of therapeutic effects. For an efficient dosage form design, other

pharmaceutical excipients (e.g., penetration enhancers, targeting ligands or solubility enhancers) can be incorporated within the API to achieve desired pharmacological characteristics. ES can serve as a viable technique to fabricate particulate systems by refining the operating parameters of the ES mode and hence load actives in conjugation with different types of excipients in a single-step. Thus, ES is capable of improving the *in-vivo* performance of drug moieties involved, for example, improving their onset of action [134], cellular uptake [157], bioavailability [152] and targeting ability [158].

8

Hyaluronic acid-ceramide and solubility enhancer Soluplus[®] were electrosprayed to form 9 nanocomposites for tumor targeted drug delivery. Lee *et al* fabricated nanostructures where the 10 11 anti-cancer stibenoid resveratrol was enveloped within the nanoparticulate system and when 12 atomised demonstrated a narrow size distribution (230 nm). Drug entrapment studies calculated a high entrapment efficiency where 80% of the active resveratrol was successfully entrapped 13 14 within the nanocomposite. Here, hyaluronic acid-ceramide was employed as a targeting moiety to interact with the CD44 receptor, a cell-surface glycoprotein upregulated in breast cancer. In-15 16 *vivo* studies revealed that the electrosprayed resveratrol-Soluplus[®] nanocomposites displayed a higher cellular uptake and cancer targeting ability when compared to the raw resveratrol-17 18 Soluplus[®] nanocomposite. From this it was confirmed that the tumor targeting drug delivery 19 system benefitted from utilising the ES process as atomised composites demonstrated 20 decreased *in-vivo* clearance, sustained drug release patterns and extended blood circulation 21 times of resveratrol, all which can be deemed favourable for tumor targeted drug therapy [158].

22

23 Grafting targeting ligands (e.g functional groups or monoclonal antibodies) on the surface of 24 engineered particulate delivery systems is a unique approach to target specific tissues or cells 25 where the therapeutic action of an active is required [159]. Grafting ligands on preformed 26 nanoparticles usually requires a chemical conjugation reaction which is time consuming, 27 expensive and sometimes leads to loss of the loaded active. However, ES is a unique procedure 28 that can attach ligands on drug loaded particles under ambient conditions *via* a single-step, 29 thereby designing a dosage form with an improved therapeutic action and a reduced toxicity 30 profile [160]. These particles are capable of selectively binding to structures (antigen or 31 receptors) that are upregulated only in diseased tissues, hence enhancing cellular uptake and 32 protecting healthy tissues from the toxicity of actives.

33

1 Aptamers can also be utilised to promote and augment the targeting ability of drugs by binding 2 to specific target molecules. Docetaxel, also known as Taxotere is a clinically established 3 chemotherapeutic drug used for several treatments including cancers of the lungs, stomach and 4 ovaries. It does however come with complications owing to its poor aqueous solubility, low 5 bioavailability and increased toxicity, thus limiting its use in anti-cancer therapeutics [161]. ES 6 was employed by Ghasammi et al to potentially overcome drawbacks associated with 7 Docetaxel and improve its physiochemical properties. Docetaxel loaded nanoparticles were 8 prepared using Ecoflex and PEG 6000 at a flow rate of 1mL/h and were subsequently attached 9 to a HER-2-specific aptamer (for site-specific delivery), eventually yielding aptamer targeted 10 Ecoflex nanoparticles. During analysis, the electrosprayed delivery system displayed higher 11 cellular uptake and anti-cancer efficacy in comparison to non-targeted Taxotere loaded 12 nanoparticles. ES was therefore concluded as an efficient technique for preparing targeted drug delivery systems, in particular, those involving aptamer-based technology [157]. 13

14

15 7.4 Systems hosting toxic agents

ES has also shown its potential for the delivery of cytotoxic agents where employing the novel 16 17 technique can fabricate particulate systems with reduced toxicity, therefore establishing 18 suitable drug safety profiles for clinical applications. Cytotoxic drugs exert unselective toxicity 19 against both cancerous cells as well as healthy living tissue [40]. Moreover, during ES, solvents 20 play a fundamental role in the formulation process, however toxic solvents can damage 21 biomolecules (genes, enzymes and proteins). ES can be fine-tuned to produce particulate-based 22 dosage systems where issues associated with toxic drugs and solvents can be overcome whilst 23 maintaining therapeutic effects.

24

Cisplatin is a standard anticancer drug used in chemotherapeutic treatments for several 25 26 malignant tumors. However, due to its significant toxicity it can trigger a number of lethal side 27 effects including kidney dysfunction [162] and thus, its therapeutic use in a number of 28 applications is hindered. ES can be employed as a potential technique to fabricate delivery 29 systems for cytotoxic agents such as cisplatin, thereby diminishing clinical limitations 30 associated with cytotoxic drugs. Parhizkar et al, for example, utilised single-needle ES for the 31 atomisation of PLGA polymer and cisplatin into spherical particles exhibiting smooth surface 32 morphology and a high encapsulation efficiency (>70%). Through the use of ES, cisplatin was 33 encapsulated within a polymeric matrix, thus protecting it from possible degradation within the 34 biological environment. In addition to this, particles produced via ES which are in the nano

1 range often have prolonged circulation times. Initial sharp release is undesirable for cytotoxic 2 drugs as this often does not reduce side effects related to premature delivery. Drug release 3 kinetic studies revealed that nanoparticles with reduced sizes demonstrated a biphasic release 4 profile; initial burst release followed by sustained release. This is beneficial for cytotoxic drugs 5 as dosage forms must be designed to reduce the risk of adverse events which could affect patient safety. Moreover, nanoparticles produced using a 10 wt% formulation displayed a 6 7 release of 14% after 4 hours whereas a formulation of 5 wt% released 45% of the loaded 8 cisplatin. Using ES, where the flow rate and applied voltage could be manipulated, resulted in 9 tailored controlled release of drug particles. ES can therefore be considered as a potential 10 approach to reduce toxicity of anti-cancer drugs as well as improving encapsulation capacities 11 and hence can be applied to an array of chemotherapeutic treatments [102].

12

13 Organic solvents are essential in most techniques that aim to encapsulate actives or 14 biomolecules into particulate systems. However, some of these biological entities (e.g., cells, nucleotides, enzymes) are sensitive to solvents and denature or degrade upon contact with 15 16 them. It is therefore crucial to develop a delivery system where toxic organic solvents may be 17 used, however, are encased within a protective layer for safe drug delivery systems. Co-axial 18 ES is a prospective technique to protect these biomolecules or cells from degradation by 19 solvents during the encapsulation process via the use of concentric needles. Esfahani et al for 20 example fabricated core-shell microcapsules consisting of PLGA and 21 chloroform/dimethylformamide by co-jetting ES technology, resulting in the successful 22 encapsulation of live cells. Although the solvents displayed high toxicity against viable cells, 23 the coaxial system provided an effective approach to shield biomolecules from the solvent 24 formulation, mitigating toxicity issues and consequently providing a safe environment for 25 encapsulating live cells. The development of such microcapsules is an innovative technology 26 that can be useful in different pharmaceutical applications such as regenerative medicine, tissue 27 engineering and cell-based drug delivery platforms [163].

28

29 7.5 Stimuli-responsive systems

Stimuli responsive drug delivery systems allow a site-specific release and/or targeting action.
These types of systems can release their contents upon the response of different derived stimuli
including external (ultrasound, light, electrical and magnetic fields) or internal (temperature,
pH, ionization and enzymes) [164] stimuli. Engineering of stimuli responsive drug delivery
devices using ES can enable dosage forms with high bioavailability, exceptional targeting

ability and on-demand release profiles, thus these delivery systems can be utilised as an
evolutionary approach for several clinical applications. ES can be employed to design dosages
with tailored drug release by enveloping the active as a core within microcapsules nanocapsules
where the shell acts a responsive polymer and dissolves only in the presence of a specific
trigger.

6

7 For example, a coaxial device was developed by Cao et al which contained a formulation 8 consisting of Silk fibroin protein (outer needle), polyvinyl alcohol (PVA) polymer (inner 9 needle) and the anti-cancer drug Doxorubicin (1% w/w) encapsulated within the PVA core. 10 When infused into a coaxial system under relevant processing parameters, the formulation 11 generated a variety of smooth core-shell structures by modifying the PVA concentration (0.3-12 0.5 wt%) and applied voltage (12-20kV) of the ES process. The electrosprayed core-shell particles demonstrated encapsulation efficiencies over 90%, whilst release profiles indicated 13 14 an initial burst release followed by sustained release of Doxorubicin. Low intensity focused ultrasound was employed as an external trigger for tailored drug release, where groups treated 15 16 with the stimulation technique displayed twice the cell apoptosis rate in comparison to the 17 control group (no stimulus). The utilisation of compatible polymers to coaxially ES core-shell 18 structures was concluded as a suitable method, particularly modifying the PVA/silk-fibroin 19 ratio and the addition of ultrasound for tumor related therapy, which resulted in stimuli-20 responsive structures displaying double the amount of apoptotic activity in human breast cancer 21 cell lines [153]. Atomized stimuli responsive core-shell delivery systems can therefore be a 22 successful approach to enhance the cytotoxicity of anticancer drugs thereby reducing their 23 adverse effects.

24

25 7.6 Bio-responsive systems

Bio-responsive systems or environmentally responsive systems are a promising stimuli 26 27 responsive delivery system that release their contents upon change in the biological 28 surrounding or in certain cellular micro-environment. These intelligent drug delivery systems 29 deliver their cargo upon the effect of a biological stimulus (e.g., enzymes, redox, antibodies, 30 hormones and pH), triggering a diverse range of responses which can find applications for 31 clinical and biomedical routes. The construction of these drug delivery devices in combination 32 with ES can enable dosage forms which can generate specific targeting and therefore elicit 33 efficacious therapeutic doses when optimising engineered drug delivery platforms, particularly 34 those displaying a stimuli-responsive nature.

2 ES can be utilized to engineer responsive drug delivery systems by dispersing the drug in a 3 polymeric nanoparticle where the polymer can dissolve only under biological stimuli. For 4 example, Wu et al designed a novel concept to fabricate bioresponsive controlled particulates 5 using an elastin-like polypeptide polymeric solution and doxorubicin, co-dissolved in a mixture 6 of solvents with the use of single-needle ES. The developed nanoparticles were spherical in 7 shape and displayed smooth morphology with particle diameters within the range of 150-570 8 nm, influenced by the processing (applied voltage and flow rate) and solution (polymer 9 concentration and molecular weight) parameters. Doxorubicin loaded particles (20 w/w%) 10 demonstrated pH responsive behaviour where drug release followed pH-dependant solubility 11 of the bioresponsive polymer (elastin-like polypeptide), impacting drug release profiles due to 12 the nature of the polymer [165]. Polymeric carriers that trigger specific stimuli for enhanced 13 control of drug release profiles could therefore provide a suitable platform for stimuliresponsive particulate systems when employing ES in conjugation with bioresponsive 14 15 materials.

16

1

17 7.7 Systems hosting biopharmaceuticals

Biological entities such as ligands, nucleotides or antibody-drug conjugates can provide an exceptional platform for biopharmaceutical applications. As ES is a simplistic single-step procedure under ambient conditions, it can combine the convectional solution consisting of the polymer and solvent but with additional therapeutic agents, biological entities and targeting ligands, simultaneously. Thereby generating dosage designs which can deliver drugs and biological molecules with high specificity to diseased cells.

24

25 Wu et al engineered a transferrin grafted lipolex nanoparticulate system loaded with 26 oligodeoxynucleotide using coaxial ES. Lipoplex molecules are classified as synthetic carriers 27 of API molecules or genetic material. They are therefore beneficial owing to their 28 encapsulation properties within the hydrophobic bilayers or in the hydrophilic core of the 29 particulate system. After co-axial atomisation, fabricated nanoparticles displayed a mean 30 particle size of 190 ± 39 nm as well as an encapsulation capacity of $90 \pm 6\%$ (measured using gel electrophoresis). In addition, the glycoprotein transferrin was conjugated to lipoplex 31 32 nanoparticles to improve nanoparticle targeting and cellular uptake. Transferrin-conjugated 33 lipoplex nanoparticles demonstrated efficient delivery to human leukaemia cells. As a result, 34 this downregulated the bcl-2 protein expression by $57 \pm 3\%$, resulting in effective treatment of

leukaemia cancer cells and thereby decreasing their multidrug resistance [166]. This study
 clearly shows the immense potential of ES in encapsulating molecules such as nucleotides,
 siRNA and plasmid DNA into suitable particulate systems. Thus, designing a delivery system
 that can successfully play a role in treating genetic based diseases such as sickle cell anaemia,
 crohn's disease and cancer.

6

7 The employment of biological material which can undergo further modification such as 8 conjugation with other actives or targeting ligands is a valuable approach in biopharmaceutical 9 applications. Furthermore, this concept can be expanded to include dosage designs utilising 10 drug-conjugates which can be successfully translated into gene drug delivery or tissue 11 engineering applications.

12

13 7.8 Theranostic systems

Theranostic technology is an advanced approach in the field of pharmaceutical drug delivery 14 15 and research referring to an amalgamation between the use of therapeutic agents and diagnostic 16 applications [167]. It can include the incorporation of targeting moieties in conjugation with 17 nanomedicines, hence leading to the development of a smart dosage system. Rasekh et al, for 18 example developed an ES system incorporating a concentric design for the fabrication of a 19 potential theranostic agent to be used in combined imaging and therapy. The co-flow system 20 included a formulation consisting of superparamagnetic iron oxide nanoparticles (SPIONs), 21 PEG polymer, genistein (model drug) and a fluorescent dye, which were all encapsulated 22 within the triglyceride derivate, tristearin. Although SPIONS display exceptional applications 23 as contrast agents for magnetic resonance imaging, the characteristics of these nanoparticles 24 tend to form aggregates. To overcome these limitations, the combination of materials in the formulation when coaxially sprayed, resulted in encapsulated SPIONS (diameter 0.65-1.2µm). 25 26 The release profiles of composite tristearin-SPIONs (single-needle electrospray) and 27 encapsulated SPIONs (co-axial electrospray) were investigated, where encapsulated genistein 28 exhibited a triphasic release profile, useful for controlled drug release where drugs may be 29 required for a longer duration. The co-axial technique was found to be a facilitative approach 30 as atomised microparticles could reduce aggregates formed on SPIONS by adjusting processing parameters and encapsulating these SPIONs with active components for magnetic 31 32 resonance imaging related multimodular theranostic applications [2].

1 7.9 Sensor systems

2 ES has also been utilised to spray biologically active substances to include dosage designs for 3 gene expression, for the detection of chromosomal aberrations in cancer therapy or to identify 4 specific protein biomarkers [168]. Here, ES can be employed to create microarrays for the 5 detection of several biomolecules including DNA and proteins and in turn reduce limitations associated with conventional techniques including poor flow rates. For example, Morozov et 6 7 al electrosprayed solutions of biological molecules (proteins, DNA and dyes) through an array 8 of voids presented in a dielectric mask, ultimately depositing multiple dots of biological 9 material onto various grounded substrates. Fabricated microdots demonstrated a uniform 10 deposition on the collecting substrate, displaying a mean size of 2-6 µm for resultant structures. 11 ES was found to be a promising technique where the shifting of the di-electric mask generated 12 a multi-component matrix and in turn enhanced the rate of deposition. In addition, biological 13 molecules maintained their functional activity, where ultimately the atomisation process could 14 be employed for gene profiling as well as clinical diagnostic devices for applications in 15 diseased states [169].

16

In summary, intelligently prepared dosage designs focus on efficient drug uptake, release 17 18 mechanisms, biocompatibility of involved materials as well as the incorporation of API which 19 display complementary characteristics. By tailoring the solution and processing parameters, 20 the ES technique can engineer particulate delivery systems with uniform particle dimensions, 21 higher encapsulation efficiencies, increased cellular uptake and improved drug targeting, thus 22 fabricating exceptional dosage forms with highly efficacious therapeutic value. ES displays 23 clear cut advantages over existing platforms at ambient environments with all processing 24 assembled into a single-step, eliminating complicated pre-preparation during the 25 manufacturing process. ES therefore offers an extraordinary approach to address the emerging 26 requirements of the pharmaceutical industry by developing smart dosage designs for multi-27 functional applications in theranostics, targeted therapy and stimulus-based approaches.

28

29 8. Applications of electrosprayed particulate systems

30

Over the years, research has been heavily focused on utilising ES for both pharmaceutical and biomedical applications. ES has been used to prepare different types of drug delivery systems such as nanoemulsions [170], nanoparticles [171] and liposomes [172], using different materials including polymers, lipids and inorganic materials [66]. Nanoparticles fabricated *via*

ES have the ability to encapsulate large amounts of drug and act as specific drug carriers owing to their active surface absorption, binding or complexation, where the size of the nanoparticles produced is critical in the therapeutic remit [173]. The chosen route of administration (ROA) heavily influences therapeutic efficacy, and each ROA presents unique advantages and drawbacks, thus often resulting in the need for a specific delivery vehicle. Electrosprayed engineered particles have been used in different drug delivery systems including; oral, transdermal, parenteral, ocular, pulmonary, buccal and topical (Table 2, Figure 4).

8

9 8.1 Oral drug delivery systems

Oral drug delivery is a favourable ROA owing to its ease of administration (increased patient 10 11 compliance), and the potential of having a solid dosage form with an extended shelf life. 12 However, the oral ROA presents a number of barriers such as degradation of drugs due to the 13 acidic nature of the stomach. In addition to this, oral drug delivery is also limited due to the 14 poor solubility and bioavailability of a number of drugs, low systemic availability and reduced 15 efficacy [174] [175]. ES has been used to process a number of drugs, the incorporation of a polymeric carrier has shown to improve drug release characteristics via the encapsulation of 16 17 drugs, thus leading to different release patterns including sustained release [173]. Through using ES for the coating or encapsulation of molecules, improved bioavailability [176] and 18 19 solubility of poorly water-soluble drugs [177] can be achieved. PLGA nanoparticles have been 20 used as a carrier for Naproxen; Yang Cao *et al*, used coaxial ES to fabricate nanoparticles of 21 PVP/PLGA and PCL/PLGA with a distinct core-shell structure in which PLGA was used for 22 the outer shell each time. A single-step process was employed in which both rhodamine B 23 (hydrophilic) and Naproxen (hydrophobic) were encapsulated. A high encapsulation efficiency 24 was observed; greater than 85% Rhodamine B encapsulated in the inner PCL core, whilst naproxen was encapsulated in the outer PLGA shell. Different release patterns were observed 25 26 which can be attributed to dual drug encapsulation and the distinctive core-shell (drug and 27 polymeric matrix interaction) structure of the fabricated nanoparticles. In addition, release 28 kinetics were investigated and demonstrated a dual drug release profile. Through the use of ES 29 and modification of the polymeric matrix, it was possible to achieve the desired drug release 30 patterns, highlighting the use of coaxial ES for co-delivery of actives [178].

31

In another study by Mehmood *et al*, novel nanospherules were fabricated *via* ES to improve the aqueous solubility and oral bioavailability of the poorly soluble drug fenofibrate. PVP and Labrafil M 2125 (non-ionic surfactant) were used as carriers for fenofibrate. The nanospherules

were less than 200 nm in size and fenofibrate was entrapped in an amorphous state. An improved solubility of fenofibrate ($32.5\% \mu g/mL$) and a high dissolution rate of 85% within 10 minutes was observed. In addition, oral bioavailability was seen to be 2.5-fold better in comparison to the free drug, thus highlighting the possibility of using ES for nanospherule production for the improvement of solubility and oral bioavailability of the drug. This study proved ES to be a promising technique for the fabrication of drug delivery systems for the oral administration of poorly water-soluble drugs [179].

8

9 A study by Mai et al., 2017 used ES to fabricate curcumin loaded poly-lactic acid (PLA) microcapsules. Curcumin is a therapeutic molecule and has anti-septic, analgesic and anti-10 inflammatory properties, however, it possesses poor water solubility which, in turn, causes a 11 12 decrease in bioavailability, thus limiting its applications. In addition, it is unstable in a number of physical and chemical surroundings and therefore electrosprayed PLA microspheres can be 13 14 used to encapsulate therapeutic agents such as curcumin whilst providing high encapsulation efficiency and sustained release profiles. Atomized PLGA microcapsules were found to be 15 16 monodispersed and spherical with a diameter in the range of 3.8 µm to 4.4 µm. Microcapsules with a 15% loading of curcumin showed a release of 67.6% after 24 hours and followed a 17 18 Ritger-Peppas model which indicated sustained release following an initial burst release at 12 19 hours. It could be concluded that spherical microcapsules fabricated via ES have a broad range 20 of applications specifically for oral drug delivery [103].

21

22 8.2 Systemic drug delivery systems

Parenteral administration offers many advantages over other routes such as oral administration 23 as it provides enhanced bioavailability, reliable dosing and also avoids first pass metabolism 24 [180]. However, disadvantages of the parenteral route include the requirement of sterile 25 26 conditions, patient compliance as well as water soluble API for a feasible ROA [181]. ES has 27 been used to fabricate different types of nanoparticles which have been used for various 28 applications such as cancer targeting [36] and vaccine delivery [182]. ES is advantageous for 29 the processing of proteins compared to other techniques such as emulsions due to the use of 30 decreased shear forces and reduced contact times with solvents [39]. Protein antigens have 31 been encapsulated in polymeric microparticles using emulsion techniques which often results 32 in non-neutralising antibodies. Coaxial ES was used in a study by Gallovic et al, to produce a microparticulate anthrax vaccine in which the recombinant protective antigen and the adjuvant 33 34 resiguimod were encapsulated in either the same or separate acetalated dextran microparticles.

1 This study highlighted the advantage of using ES for the encapsulation of proteins, with 2 arguably the most significant advantage being the ability of the protein to maintain its 3 bioactivity following microparticulate fabrication. In addition to this, research by this group 4 was the first to show that through using specially engineered delivery vehicles in which the 5 vaccine was encapsulated, a greater degree of protection was demonstrated [183].

6

7 ES has been used to process a number of anticancer drugs such as paclitaxel, etoposide and 8 cisplatin. Zhang et al used coaxial ES for the production of Paclitaxel and etoposide loaded 9 PLGA microspheres. It was found that the atomized microspheres exhibited a core-shell structure, a high entrapment efficiency of 85.8% and a size range of 1-4 µm. Controlled release 10 of both drugs was achieved as follows; after 24 hours, 17% of paclitaxel was released, whereas 11 12 22% of etoposide was released, following this, after 120 hours 31% of paclitaxel was released compared to 33% for etoposide. The paclitaxel and etoposide loaded electrosprayed 13 14 microspheres demonstrated an improved cytotoxic effect on saos-2 osteosarcoma cells in 15 comparison to the pure drugs individually, thus highlighting the application of electrosprayed 16 microspheres for combinatorial drug therapy in the medical remit [155].

17

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19 8.3 Transdermal drug delivery systems

Transdermal drug delivery is an alternative ROA compared to oral and parenteral 20 21 administration. This method overcomes the challenges associated with the aforementioned 22 ROA, such as poor absorption and degradation of drugs occurring in the gastrointestinal tract 23 or the liver, whilst providing a sustained release of drugs with a single application. It should be 24 noted, only a selective number of medications can be delivered via the transdermal route in 25 therapeutic amounts, however microneedles can be employed to improve transdermal delivery 26 [184-186]. Microneedles work by creating micropunctures in the skin through which drugs 27 and nutrients can be transported into deeper layers within the skin. In addition, the use of 28 microneedles is convenient, inexpensive, non-invasive, painless, and can be self-administrated 29 thus improving patient compliance [187]. The ES process has the ability to fabricate controlled 30 particle coatings for stainless-steel microneedles. This improves drug delivery, specifically for 31 sensitive biomolecules such as peptides and proteins which remain stable during the ES process 32 but cannot be delivered orally. The ES method is favourable over others such as dip coating 33 due to the fact that it allows a controlled coating for only the microneedle tips as opposed to 34 the whole microneedle substrate, as is the case with dip coating [188]. Serdar Tort *et al*,

produced loaded (drug and insulin) nanoparticle coatings for microneedles *via* ES, insulin loaded nanoparticles with a size in the range of 522 ± 261 nm were produced. Following this, the effectiveness of the electrosprayed insulin coated microneedles was tested on diabetic rats, where the study observed a significant decline in fluctuating blood glucose levels when compared to subcutaneous injections [189].

6

7 8.4 Pulmonary drug delivery systems

Pulmonary drug delivery systems are often utilised for both systemic and local drug delivery 8 9 applications. Through use of inhaled drug delivery, the drug is able to directly target the site of 10 action, thus allowing the use of a reduced dose whilst maintaining an increased drug 11 concentration at the site of action and minimized systemic side effects [190]. Often, the use of 12 particulate pulmonary drug delivery systems is limited by the size polydispersity of the particles. In order to provide successful pulmonary drug delivery, particle size and uniformity 13 14 is critical, with an aerodynamic diameter in the range of 1-5 µm considered suitable for drug delivery devices. By controlling particle size, it is possible to regulate drug absorption and 15 16 bioavailability. Through the use of ES, monodispersed particles with a narrow particle size 17 distribution can be fabricated [191]. ES allows tight control over both particle size and 18 polydispersity, thus making it desirable for the fabrication of inhalable pharmaceuticals [192]. 19

20 ES has been used for a number of pulmonary applications in which targeted drug delivery via 21 inhalation is required, an example of these is asthma or cystic fibrosis. When in the cone-jet 22 mode, the ES technique gives rise to the production of droplets which are a few micrometers 23 in diameter with a very narrow size distribution. Droplet size can be altered *via* strict control 24 over the process parameters, where the ability to control droplet size is key in maximising distal lung deposition and thus the electrospray must be appropriate to target drug inhalation. 25 26 BatellePharma developed novel electrospray inhalers which were able to nebulise fine drug 27 loaded electrosprayed droplets which reached the distal regions of the lung alveoli and 28 bronchioles [68, 193]. Youliang hong et al, successfully loaded PLGA with Rifampicin, 29 forming microparticles via a modified ES technique. Particles produced had a mean diameter, 30 with a range of 2-5 µm, making it suitable for pulmonary drug delivery applications [194]. 31 Yaqoubi et al, employed single-needle ES to produce a dry powder inhalation formulation of 32 montelukast and budesonide, individually and in conjugation for pulmonary drug delivery. This formulation was processed using ES, in which montelukast behaved as both an API and a 33 34 carrier for budesonide in the excipient free formulation, producing an inhalable dry powder

formulation for the treatment of asthma. Research demonstrated that the therapeutic efficiency in controlling asthmatic episodes was significantly enhanced when budesonide was inhaled in combination with orally administered montelukast. This method of administration was found to be as effective as doubling the dose of budesonide would. It would therefore be reasonable to produce a formulation consisting of both montelukast and budesonide for dry powder inhalation *via* ES [190].

7

8 8.5 Nasal drug delivery systems

The nasal pathway has been a favourable route of administration for a number of years due to 9 10 the effect of a significant amount of drugs being more prominent when delivered via this 11 pathway. The use of nanotechnology for nasal drug delivery has shown to have a number of 12 advantages including targeted drug delivery, increased bioavailability and decreased toxicity 13 [195]. ES has been used to process asthmatic drugs, Midhun *et al*, utilised ES for the production 14 of Budesonide loaded PCL nanobeads. The effectiveness of Budesonide is restricted due to its 15 rapid elimination, thus encapsulation into a biodegradable polymer to obtain sustained release is necessary. Particles with a size of 116.1 ± 19 nm and a drug encapsulation efficiency of 75 16 17 \pm 2.4 % were achieved using ES under optimised conditions. Controlled drug release was obtained *in-vitro* at pH 7.4 and 5.6 [196]. Through the employment of ES, high drug 18 19 encapsulation efficiencies and sustained release profiles were demonstrated, thereby forming 20 efficient nanocarriers for the treatment of inflammatory disorders.

21

22 8.6 Ocular drug delivery systems

Ocular delivery is an advancing area within the drug delivery remit, largely due to the 23 24 challenges associated with delivery to the eye irrespective of the ease of access [197]. The eye 25 possesses several anatomical and physiological barriers which protect it from foreign objects. 26 As a result, it is vital that a delivery system is produced to overcome these barriers and allow targeting to specific ophthalmic tissues for the control and treatment of ocular diseases [134]. 27 28 Conventional drug delivery methods such as eye-drops are disadvantageous as a significant 29 amount of the drug (>90%) is lost upon administration via tear production and drainage 30 mechanisms [198]. In addition to this, drug bioavailability is poor due to reduced residence 31 time in the eye. ES overcomes these barriers when used as a coating method for contact lenses. Mehta et al, encapsulated timolol maleate into three different polymeric matrixes; PVP, 32 33 poly(Nisopropylacrylamide) (PNIPAM) and PVP:PNIPAM (50:50%w/w) from which both fibrous and particulate samples were generated. Over 52% of the fabricated structures for all 34

formulations were less than 200 nm in diameter. *In-vitro* release studies found a biphasic release for all formulations where the PNIPAM and timolol maleate coating released 89.8% of the drug after a period of 24 hours. *In-vitro* studies suggested through the combination of a fast-dissolving polymer (PVP) and a sustained dissolving polymer (PNIPAM), a polymeric coating which offers controlled prolonged release could be achieved *via* ES [199].

6

7 8.7 Other routes of administration

Electrosprayed particles for other ROAs such as buccal, topical and brain offer a number of 8 9 possibilities [200, 201]. Buccal mucosa is a promising route for systemic drug delivery, owing 10 to its convenience, accessibility and high vasculature. ES has been used to process mucoadhesive biopolymers for applications in buccal drug delivery. ES is a favourable method 11 12 for the fabrication of drug loaded multi-layered membrane mucoadhesive patches [202]. The 13 skin acts as a mechanical barrier, preventing the penetration of several drug substances, 14 however, this aside, the skin is an advantageous site for drug delivery. Often, products which 15 are formulated for topical use are categorised into two groups; those which are used for local 16 drug delivery or systemic drug delivery systems [203]. Najme Hazeri et al used ES for sericin 17 (an antibacterial agent) and a resultant nano-powder was formed with a mean particle size of 18 25 nm. The sericin nanoparticles were tested for their moisture absorption, where a moisture 19 regain of 56.2% was reported, thus indicating the potential use of these nanoparticulates in 20 topical applications such as moisturisers and sun creams [200]. Nie *et al* used co-axial ES to 21 produce paclitaxel and suramin loaded core-shell microspheres for the treatment of brain 22 tumors. Both drugs possessed varying hydrophilic characteristics and were encapsulated into Poly(L-lactide) PLLA (core) and PLGA (shell) microspheres. Furthermore, two different types 23 24 of microspheres $(10 - 20 \,\mu\text{m})$ were produced with swapped distributions as follows; sample A consisted of paclitaxel encapsulated in the core and suramin in the shell whereas sample B 25 26 consisted of suramin encapsulated in the core and paclitaxel in the shell. In-vitro studies were 27 carried out and sample B demonstrated enhanced apoptotic activity in comparison to sample A 28 and drug controls. In addition, sample A and B were superior in inducing apoptosis in 29 comparison to single drugs, therefore highlighting the advantages of using dual drug delivery 30 to inhibit U87 cell growth. Similarly, in-vivo studies demonstrated tumor inhibition of U87 31 glioma in nude mice, in which samples A and B showed substantial decrease in the number of 32 tumor cells, again highlighting the advantages of using ES to fabricate dual drug delivery 33 systems [204].

9. Emerging applications utilising electrohydrodynamic atomisation

Further advancements in biomedical and pharmaceutical research areas have resulted in the
need for more sophisticated therapies, such as regenerative medicine bioactive cargo carriers,
scaffolds for tissue engineering, implantables, coatings and sensing devices.

6

2

7 9.1 Tissue Engineering

8 ES is beneficial when used for tissue engineering applications in the regenerative medicine 9 remit [205]. Commonly, scaffolds have been utilised to support; bone cell growth and tissue 10 regeneration, as well as promoting bone-forming cells through natural proteins and growth 11 factors. Often porous surface topographies are fabricated to mimic the extracellular matrix 12 which in turn induces restoration of damaged bone tissues [10]. Bio-ES is a promising 13 technique in tissue engineering for cell delivery in scaffolds owing to the low current used 14 making it safe to process cells. Previously a number of cells have been processed via bio-ES 15 including bone marrow derived mesenchymal stem cells [206], embryonic stem cells [207] and 16 cardiac cells [208]. In recent years, artificial 3D constructs have been explored as a means of 17 direct delivery of mesenchymal cells for tissue engineering. Limitations with these constructs have arisen due to the lack of cell infiltration, reduced cell functioning and minimal diffusion 18 19 of essential nutrients and oxygen through the scaffold. For feasible tissue engineering, a 20 uniform occupation of cells is critical [209]. ES and electrospinning were used in combination 21 to overcome some of these limitations, for example, Braghirolli et al combined poly(lactide-22 co-glycolide) fibers produced via electrospinning with the bio-ES of a suspension of 23 mesenchymal stem cells, as a means of directly integrating cells into fibers. The scaffolds were 24 cultivated and following this, cells remained viable. Confocal imaging confirmed cell 25 adaptation and spreading between fibers and scanning electron microscopy assessed the morphology of the scaffolds, which indicated the presence of a significant number of cells 26 27 within the scaffold structure. Through combining bio-ES and electrospinning, uniform cellular 28 distribution was promoted across the 3D structures. The data attained highlights the possibility 29 of combining electrospinning and bio-ES for the successful fabrication of 3D cell-integrated 30 scaffolds which increase the development of tissues and is therefore favourable for use in 31 regenerative medicine [210].

32

One of the main challenges in tissue engineering is the localised delivery of growth factorswhere the administration of these molecules can result in potential inflammation and ectopic

1 bone formation in soft tissues. ES as a means of producing delivery systems has shown great promise in engineering microparticles to deliver biomolecules (e.g., proteins, enzymes and 2 3 growth factors) to bones whilst maintaining their bioactivity throughout the manufacturing 4 process [211]. Bock et al utilized the ES technique to encapsulate growth factors (vascular 5 endothelial growth factor and bone morphogenetic protein 7 (BMP-7)) into microparticles of 6 PLGA, PEG and trehalose composite. Here, the growth factor loaded PLGA particles were 7 fabricated and combined with a protein stability enhancer (PEG and trehalose) in a single-step. 8 Electrosprayed particles loaded with BMP-7 were cultured with preosteoblasts, where 9 substantial cell differentiation into osteoblasts was seen up to 3 weeks in culture. This research highlighted the successful delivery of active growth factors specific to bone tissue engineering 10 through the use of electrosprayed microparticles. Moving forward it would be useful to assess 11 the way in which microparticulate systems, specifically those consisting of sensitive molecules 12 such as growth factors are analysed [212]. 13

14

Through continuously evolving research in the tissue engineering remit, products have been 15 developed to enhance the regenerative medicine sector with advances in scaffold production 16 for bone, tendon and ligament repair via EHDA. By using bio-ES in combination with 17 18 electrospinning, there is an opportunity to fabricate complex living 3D architectures which have potential uses in regenerative medicine. Bio-ES is an evolving technique which allows 19 20 the encapsulation of cells directly into scaffolds as well as having potential uses in the 21 advancement of organs-on-chip technologies. Currently, the ES process works on a small scale 22 which is not sufficient for commercial use, however through employing a number of needles, 23 the ES process is becoming closer to being used on a much larger scale [40]. In another study 24 conducted by Shokraei et al, nanocomposites for use in cardiac tissue engineering were fabricated using electrospinning (polyurethane fibers) and ES (multiwall carbon nanotubes) 25 26 simultaneously. Carbon nanotubes were electrosprayed onto a rotating collector and 27 polyurethane fibers were electrospun at the same time from the opposite side of the collector. 28 The resulting scaffolds demonstrated cytocompatibility for cardio myoblasts, inducing cardio myoblast attachment and proliferation on the novel conductive nanocomposite patches. This 29 30 study emphasised the potential use of electrosprayed scaffolds in cardiac tissue engineering 31 moving forward [3].

1 9.2 Implant device coatings

Aforementioned, the ES process is a favourable technique for the fabrication of polymeric 2 3 nanoparticles as drug carriers. Implant failure often occurs as a result of infections following 4 implantation. With the ever-evolving area of implants, it is vital to propose an approach to 5 prevent surgical site infections. Here, Tsiapla et al used ES to produce biodegradable nanoparticles as drug carriers for the treatment of orthopaedic infections. Titanium metal 6 7 implants were coated with vancomycin loaded PCL nanoparticles using ES. Drug release was 8 investigated, and a bi-phasic release was observed with an initial burst release of 57.9% 9 followed by sustained release over a 61-day period. The initial burst release from the nanoparticles is desirable for treating the onset of infections specifically following surgery. A 10 11 subsequent sustained release pattern is also important, as often the administration of high doses 12 of antimicrobial agents is required in the infected area over a period of 6-8 weeks. Emerging 13 research following this study could focus on exploiting these electrosprayed nanoparticles for 14 use in cytotoxicity studies as well as researching their interactions with different cell types such 15 as mesenchymal stem cells or bone marrow stromal cells [213].

16

ES has been used for the fabrication of polymer coatings on implant surfaces, thus allowing 17 18 tight control over the surface texture. Guo et al., produced a number of surface micro-19 topographies using polyhedral oligosilsesquioxane thermoplastic polyurethane as a coating for 20 stainless steel coronary stents. Upon altering the electric field over a range of voltages, surface 21 coatings of three different morphologies; smooth, roughened and fibrous were achieved at 1.5 22 kV, 1.6 – 1.7 kV and 1.8 kV, respectively. The control over process parameters in ES is 23 advantageous as it allows the manipulation of coating topographies which in turn regulates the 24 integration of implant devices with tissue in the surrounding environment [214]. Due to the 25 adherence between the charged droplets and the conductive metallic stent (collector), coatings 26 produced via ES for metallic stents have proved to be superior in robustness and uniformity in 27 comparison to those formed *via* conventional methods [215]. Drug eluting stents are used for 28 the treatment of acute symptoms caused by coronary artery disease. By using ES to produce 29 dual action stent coatings, it is possible to enhance the therapeutic potential by providing more 30 personalised treatment approaches. This can be achieved through tight control over drug 31 loading and release kinetics due to ES deposition. McKittrick et al developed a novel coronary 32 stent coating for the controlled release of sirolimus from accelerate essentially combining an 33 anti-proliferative agent and a bioactive polymer coating which promotes re-endothelialisation 34 to form a dual action coating. The coating was deposited via ES which allowed strict control

over coating thickness, roughness, drug loading and release kinetics providing a significant therapeutic potential. The study demonstrated an improvement in the attachment of primary porcine endothelial cells to the surface. Further advancements in this area would benefit from investigating the effect of the ES process on drug loading and release kinetics on the performance of drug eluting stents [216].

6

7 9.3 Sensing Devices

Novel devices have also incorporated sensors for drug delivery systems, where the addition of 8 9 ES has found to be an attractive approach. Ruecha et al developed a solution consisting of 10 graphene, PVP and polyaniline (PANI), which, when atomised using ES, produced 11 nanocomposites onto a grounded substrate consisting of a paper-based biosensor and a rotating 12 drum. Fabricated nanocomposites displayed a mean particle size of 160 ±1.02 nm. The 13 modification of the biosensor by the ES technique also led to nanoparticulates exhibiting high 14 conductivity and a large surface area, subsequently improving electrochemical sensitivity for 15 cholesterol detection. Thus, ES was found to provide a diverse platform for improving the 16 detection of sensing devices for biologics, applicable in medical diagnostic therapies [217].

17

18 9.4 Eco-friendly and green technologies

The timely strategic expansion of the pharmaceutical industry and more specifically the 19 advancement of such emerging technologies indicates the potential to address unmet clinical 20 21 needs is promising. Furthermore, beyond the scope of this review is the adaptation of such 22 technologies in more eco-friendly and 'green' channels. Employing organic solvents in 23 engineering processes can be potentially harmful and consequently raise issues of cell 24 toxicology and environmental safety [40]. As ES methods have shown extended applications 25 in tissue engineering, biomedical therapeutics and theranostic systems it is of critical 26 importance to utilise non-toxic solvents when fabricating various particulate structures [218]. 27 'Green ES' is therefore a prospective technique which, if adopted in the pharmaceutical 28 industry, could potentially improve the environmental profile of such processes.

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1 Conclusion and future perspectives

2 The successful control and subsequent targeted drug release of therapeutic moieties is of 3 overarching importance with regards to the pharmaceutical industry. Currently, conventional 4 particle fabrication methodologies although well established, are accompanied with several shortfalls including poor drug loading, inefficient encapsulation capacities, unstable 5 therapeutic compounds as well as undesirable drug release kinetics. The ES method can 6 7 therefore be adopted as a facilitative approach for particle engineering to address these 8 limitations. These include upscaling manufacture of desired drug delivery systems, enabling 9 batch-batch reproducibility, improving stability as well as cost efficiency, all of which can be 10 implemented into the industry to yield highly efficacious drug delivery systems. The interplay 11 of processing parameters involved in the ES technique can have an immense impact on the 12 fabricated particulates which can exhibit highly functional structures, thus providing 13 tremendous scope in the field of pharmaceutics and drug delivery. Nevertheless, bottle necks 14 for the ES method can be assigned to its poor throughput due to low flow rates, particularly in 15 the Taylor cone-jet mode owing to its stable geometry. As a result, rendering a particulate 16 system with long production times and therefore insufficient to be utilised during scale-up. To 17 further refine the electrospray method, multiple cone-jet systems have been considered as a feasible approach to tackle these issues where multi-tip emitters can be employed each 18 19 possessing separate capillaries (which individually display the steady cone-jet mode). These 20 can generate monodisperse particles with a controllable spray plume, thus allowing effective 21 deposition of particles on the grounded substrate. This in turn increases the yield of resultant 22 particles for continuous production in the commercial industry. In addition, "Green ES" has 23 been used to further enhance the safety of the ES technique, by limiting the use of organic 24 solvents and their associated toxicity with biological entities (genes, peptides and enzymes) 25 and alternatively using aqueous solvents which are non-toxic in nature.

26

27 Furthermore, the employment of the ES technique in the pharmaceutical remit to fabricate 28 nanoparticles has shown great potential. This competent method can improve drug efficacy at 29 the site of interest demonstrating higher loading capacities and payload release in addition to 30 minimal side effects and toxicity. ES also provides a versatile technique to enhance the 31 solubility and bioavailability when encapsulating therapeutic compounds into drug carriers. 32 Following particle engineering, extensive in-depth analysis on the relationship between 33 generated particulate systems and biological structures is of paramount importance. This 34 involves the consideration of permeability, pharmacokinetics and physiochemical stability

which are critical factors in the development of desired particles for therapy, diagnosis and targeted drug delivery. Further research in the realm of EHDA should focus on diagnosing and monitoring the therapeutic response of efficacious delivery systems, particularly in animal models and clinical trials, making it a robust method for industrial implementation and hence translating particle engineering from small scale manufacture into commercial clinical practice.

7 On a final note, the industrialisation of EHDA technologies hinges on several aspects ranging 8 from industrial perspectives (e.g., business models, regulatory matters, production scale, multi-9 mode testing and product validation) to the actual implementation within the clinic. It is well known that the pharma remit is resilient to change and therefore adopting new technologies 10 11 must be of significantly higher value than those currently deployed. The current state of the 12 art has shown EHDA technologies to provide multiple benefits (process, material and space) 13 when compared to established protocols. In fact, the versatility of this method makes it promising for numerous ROA and target sites. At present several SMEs and indeed large 14 corporate pharma have shown a luke-warm interest indicating the need to scale-up of such 15 systems with emphasis on control, management, utilisation and product reproducibility on a 16 larger scale. This is the first major challenge and quite possibly the biggest challenge. The next 17 18 few stages then move towards *in-vivo* models and then potentially small-scale clinical trials.

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1 **References**

- 2 [1] E. Sayed, C. Karavasili, K. Ruparelia, R. Haj-Ahmad, G. Charalambopoulou, T. Steriotis,
- 3 D. Giasafaki, P. Cox, N. Singh, L.N. Giassafaki, A. Mpenekou, C.K. Markopoulou, I.S.
- 4 Vizirianakis, M. Chang, D.G. Fatouros, Z. Ahmad, Electrosprayed mesoporous particles for
- 5 improved aqueous solubility of a poorly water soluble anticancer agent: in vitro and ex vivo
- 6 evaluation, J. Controlled Release 278 (2018) 142-155.
- 7 [2] M. Rasekh, Z. Ahmad, R. Cross, J. Hernández-Gil, J.D.E.T. Wilton-Ely, P.W. Miller,
- 8 Facile Preparation of Drug-Loaded Tristearin Encapsulated Superparamagnetic Iron Oxide
- 9 Nanoparticles Using Coaxial Electrospray Processing, Mol. Pharm. 14 (2017) 2010-2023.
- 10 [3] N. Shokraei, S. Asadpour, S. Shokraei, M. Nasrollahzadeh Sabet, R. Faridi-Majidi, H.
- 11 Ghanbari, Development of electrically conductive hybrid nanofibers based on CNT-
- 12 polyurethane nanocomposite for cardiac tissue engineering, Microsc. Res. Tech. 82 (2019)
- 13 1316-1325.
- 14 [4] J.A. Finbloom, F. Sousa, M.M. Stevens, T.A. Desai, Engineering the drug carrier
- biointerface to overcome biological barriers to drug delivery, Adv. Drug Deliv. Rev. 167(2020) 89-108.
- 17 [5] M. Cooley, A. Sarode, M. Hoore, D.A. Fedosov, S. Mitragotri, A. Sen Gupta, Influence of
- 18 particle size and shape on their margination and wall-adhesion: implications in drug delivery
- 19 vehicle design across nano-to-micro scale, Nanoscale 10 (2018) 15350-15364.
- 20 [6] D. Chenthamara, S. Subramaniam, S.G. Ramakrishnan, S. Krishnaswamy, M.M. Essa,
- 21 F.H. Lin, M.W. Qoronfleh, Therapeutic efficacy of nanoparticles and routes of
- administration, Biomater. Res. 23 (2019) 20-019-0166-x. eCollection 2019.
- [7] M.K. Rasmussen, J.N. Pedersen, R. Marie, Size and surface charge characterization of
 nanoparticles with a salt gradient, Nature Communications 11 (2020) 2337.
- 25 [8] P. Aggarwal, J.B. Hall, C.B. McLeland, M.A. Dobrovolskaia, S.E. McNeil, Nanoparticle
- 26 interaction with plasma proteins as it relates to particle biodistribution, biocompatibility and
- therapeutic efficacy, Adv. Drug Deliv. Rev. 61 (2009) 428-437.
- [9] A. Albanese, P.S. Tang, W.C. Chan, The effect of nanoparticle size, shape, and surface
 chemistry on biological systems, Annu. Rev. Biomed. Eng. 14 (2012) 1-16.
- 30 [10] P. Jayaraman, C. Gandhimathi, J.R. Venugopal, D.L. Becker, S. Ramakrishna, D.K.
- 31 Srinivasan, Controlled release of drugs in electrosprayed nanoparticles for bone tissue
- 32 engineering, Adv. Drug Deliv. Rev. 94 (2015) 77-95.
- [11] Y. Wang, J.D. Byrne, M.E. Napier, J.M. DeSimone, Engineering nanomedicines using
 stimuli-responsive biomaterials, Adv. Drug Deliv. Rev. 64 (2012) 1021-1030.
- 35 [12] W. De Jong H., P.J.A. Borm, Drug delivery and nanoparticles:applications and hazards, 36 International journal of nanomedicine 3 (2008) 133-149
- 36 International journal of nanomedicine 3 (2008) 133-149.

- 1 [13] H. Chan, P.C.L. Kwok, Production methods for nanodrug particles using the bottom-up 2 approach, Adv. Drug Deliv. Rev. 63 (2011) 406-416.
- 3 [14] C. Yurteri, R. Hartman, J. Marijnissen, Producing Pharmaceutical Particles via
- Electrospray-ing with an Emphasis on Nano and Nano Structured Particles-A Review,
 KONA Powder and Particle Journal No 28 (2010).
- [15] P. Iqbal, J. Preece, P. Mendes, Nanotechnology: The "Top-Down" and "Bottom-Up"
 Approaches, in: Anonymous 2012, .
- 8 [16] R. Sverdlov Arzi, A. Sosnik, Electrohydrodynamic atomization and spray-drying for the
 9 production of pure drug nanocrystals and co-crystals, Adv. Drug Deliv. Rev. 131 (2018) 79100.
- 11 [17] H. de Waard, H.W. Frijlink, W.L.J. Hinrichs, Bottom-up preparation techniques for 12 nanocrystals of lipophilic drugs, Pharm. Res. 28 (2011) 1220-1223.
- 13 [18] G. O'Connor, L.E. Gleeson, A. Fagan-Murphy, S.A. Cryan, M.P. O'Sullivan, J. Keane,

14 Sharpening nature's tools for efficient tuberculosis control: A review of the potential role and

15 development of host-directed therapies and strategies for targeted respiratory delivery, Adv.

- 16 Drug Deliv. Rev. 102 (2016) 33-54.
- 17 [19] E.M. Both, R.M. Boom, M.A.I. Schutyser, Particle morphology and powder properties
- during spray drying of maltodextrin and whey protein mixtures, Powder Technol 363 (2020)
- 19 519-524.
- [20] D.E. Walton, The morphology of spray-dried particles a qualitative view, Drying
 Technol 18 (2000) 1943-1986.
- 22 [21] N.T. Nguyen, S.A. Shaegh, N. Kashaninejad, D.T. Phan, Design, fabrication and
- 23 characterization of drug delivery systems based on lab-on-a-chip technology, Adv. Drug
- 24 Deliv. Rev. 65 (2013) 1403-1419.
- 25 [22] P. Khadka, J. Ro, H. Kim, I. Kim, J.T. Kim, H. Kim, J.M. Cho, G. Yun, J. Lee,
- Pharmaceutical particle technologies: An approach to improve drug solubility, dissolution
 and bioavailability, Asian Journal of Pharmaceutical Sciences 9 (2014) 304-316.
- 28 [23] Y. Gao, M. Chang, Z. Ahmad, J. Li, Magnetic-Responsive Microparticles with
- 29 Customized Porosity for Drug Delivery, RSC Adv. 6 (2016).
- 30 [24] E.S. Thian, Z. Ahmad, J. Huang, M. Edirisinghe, S. Jayasinghe, D. Ireland, R.A. Brooks,
- 31 N. Rushton, W. Bonfield, S. Best, Bioactivity of Nanoapatite Produced by
- 32 Electrohydrodynamic Atomization, Journal of Bionanoscience 1 (2007) 60-63.
- 33 [25] P. Mehta, A. Zaman, A. Smith, M. Rasekh, R. Haj-Ahmad, M.S. Arshad, S. van der
- 34 Merwe, M.-. Chang, Z. Ahmad, Broad Scale and Structure Fabrication of Healthcare
- 35 Materials for Drug and Emerging Therapies via Electrohydrodynamic Techniques, Adv.
- 36 Therap. 2 (2019) 1800024.

- 1 [26] M. Enayati, Z. Ahmad, E. Stride, M. Edirisinghe, One-step electrohydrodynamic
- production of drug-loaded micro- and nanoparticles, Journal of the Royal Society, Interface 7
 (2010) 667-675.
- 4 [27] W. Gilbert, in: Anonymous De Magnete, Magneticisque Corporibus, et de Magno
- 5 Magnete Tellure, London, 1628, .
- [28] L. Rayleigh, On The Instability Of Jets, Proceedings of the London Mathematical
 Society s1-10 (1878) 4-13.
- 8 [29] J. Zeleny, Instability of Electrified Liquid Surfaces, Phys. Rev. 10 (1917) 1-6.
- 9 [30] G.I. Taylor, Disintegration of water drops in an electric field, Proceedings of the Royal
- 10 Society of London. Series A. Mathematical and Physical Sciences 280 (1964) 383-397.
- 11 [31] P. Mehta, R. Haj Ahmad, M. Rasekh, S. Arshad, A. Smith, S. Merwe, X. Li, M. Chang,
- 12 Z. Ahmad, Pharmaceutical and biomaterial engineering via electrohydrodynamic atomization 13 technologies, Drug Discov, Teday 22 (2016)
- 13 technologies, Drug Discov. Today 22 (2016).
- 14 [32] A. Bohr, J.P. Boetker, T. Rades, J. Rantanen, M. Yang, Application of spray-drying and
- 15 electrospraying/electospinning for poorly water-soluble drugs: a particle engineering
- 16 approach, Curr. Pharm. Des. 20 (2014) 325-348.
- [33] N. Bock, T.R. Dargaville, M.A. Woodruff, Electrospraying of polymers with therapeutic
 molecules: State of the art, Progress in Polymer Science 37 (2012) 1510-1551.
- [34] H. Brandenberger, D. Nüssli, V. Piëch, F. Widmer, Monodisperse particle production: A
 method to prevent drop coalescence using electrostatic forces, J. Electrostatics 45 (1999) 227238.
- [35] J. Chen, B. Guo, Z. Guo, L. Li, J. Jiang, Y. Zhan, J. Wu, C. Zhang, Association of serum
 gastric inhibitory polypeptide and pancreatic polypeptide levels with prolonged esophageal
- 23 gastric initiotory porypeptide and pancreatic porypeptide levels with prolonged esophageal
 24 acid exposure time in refractory gastroesophageal reflux disease, Medicine (Baltimore) 98
- 25 (2019). doi:10.1097/MD.000000000015965.
- [36] J. Xie, J.C.M. Marijnissen, C. Wang, Microparticles developed by electrohydrodynamic
 atomization for the local delivery of anticancer drug to treat C6 glioma in vitro, Biomaterials
 27 (2006) 3321-3332.
- 29 [37] L. Zhu, M. Li, X. Liu, Y. Jin, Drug-Loaded PLGA Electrospraying Porous Microspheres
- 30 for the Local Therapy of Primary Lung Cancer via Pulmonary Delivery, ACS Omega 2
- 31 (2017) 2273-2279.
- 32 [38] Y.H. Hsu, D.W. Chen, M.J. Li, Y.H. Yu, Y.C. Chou, S.J. Liu, Sustained Delivery of
- 33 Analgesic and Antimicrobial Agents to Knee Joint by Direct Injections of Electrosprayed
- 34 Multipharmaceutical-Loaded Nano/Microparticles, Polymers (Basel) 10 (2018) 890. doi:
- 35 10.3390/polym10080890.

- 1 [39] R.T. Steipel, M.D. Gallovic, C.J. Batty, E.M. Bachelder, K.M. Ainslie, Electrospray for
- 2 generation of drug delivery and vaccine particles applied in vitro and in vivo, Materials
- 3 Science and Engineering: C 105 (2019) 110070.
- 4 [40] J. Wang, J.A. Jansen, F. Yang, Electrospraying: Possibilities and Challenges of
- Engineering Carriers for Biomedical Applications—A Mini Review, Frontiers in Chemistry 7
 (2019) 258.
- 7 [41] W. Morton J., Method of dispersing fluids, (1902).
- 8 [42] J. Doshi, D.H. Reneker, Electrospinning process and applications of electrospun fibers,
- 9 Journal of Electrostatics 35 (1995) 151-160.
- 10 [43] J.F. Cooley, Apparatus for electrically dispersing fluids. (1902).
- 11 [44] A. Formhals, Process and apparatus for preparing artificial threads. (1934).
- 12 [45] Z. Ahmad, M.A. Vargas-Reus, R. Bakhshi, F. Ryan, G.G. Ren, F. Oktar, R.P. Allaker,
- 13 Chapter five Antimicrobial Properties of Electrically Formed Elastomeric Polyurethane-
- 14 Copper Oxide Nanocomposites for Medical and Dental Applications, Meth. Enzymol. 509
- 15 (2012) 87-99.
- [46] S. Chakraborty, I.C. Liao, A. Adler, K.W. Leong, Electrohydrodynamics: A facile
 technique to fabricate drug delivery systems, Adv. Drug Deliv. Rev. 61 (2009) 1043-1054.
- [47] S.Y. Chew, R. Mi, A. Hoke, K.W. Leong, Aligned Protein-Polymer Composite Fibers
 Enhance Nerve Regeneration: A Potential Tissue-Engineering Platform, Adv. Funct. Mater.
- 20 17 (2007) 1288-1296.
- [48] J. Zeng, X. Xu, X. Chen, Q. Liang, X. Bian, L. Yang, X. Jing, Biodegradable
 electrospun fibers for drug delivery, J. Controlled Release 92 (2003) 227-231.
- 23 [49] M.S. Khil, D.I. Cha, H.Y. Kim, I.S. Kim, N. Bhattarai, Electrospun nanofibrous
- polyurethane membrane as wound dressing, J. Biomed. Mater. Res. B. Appl. Biomater. 67
 (2003) 675-679.
- 26 [50] Y. Park, E. Kang, O. Kwon, T. Hwang, H. Park, J.M. Lee, J.H. Kim, C. Yun, Ionically
- crosslinked Ad/chitosan nanocomplexes processed by electrospinning for targeted cancer
 gene therapy, J. Controlled Release 148 (2010) 75-82.
- 29 [51] U. Farook, H.B. Zhang, M.J. Edirisinghe, E. Stride, N. Saffari, Preparation of
- microbubble suspensions by co-axial electrohydrodynamic atomization, Med. Eng. Phys. 29
 (2007) 749-754.
- 32 [52] Z. Ekemen, Z. Ahmad, M. Edirisinghe, E. Stride, Forming of Protein Bubbles and
- 33 Porous Films Using Co-Axial Electrohydrodynamic Flow Processing, Macromol. Mater.
- 34 Eng. 296 (2011) 8-13.
- 35 [53] S.L. Poliachik, W.L. Chandler, P.D. Mourad, M.R. Bailey, S. Bloch, R.O. Cleveland, P.
- 36 Kaczkowski, G. Keilman, T. Porter, L.A. Crum, Effect of high-intensity focused ultrasound

- 1 on whole blood with and without microbubble contrast agent, Ultrasound Med. Biol. 25
- 2 (1999) 991-998.
- 3 [54] E.C. Unger, E. Hersh, M. Vannan, T.O. Matsunaga, T. McCreery, Local drug and gene 4 delivery through microbubbles, Prog. Cardiovasc. Dis. 44 (2001) 45-54.
- 5 [55] Z. Ekemen, H. Chang, Z. Ahmad, C. Bayram, Z. Rong, E.B. Denkbas, E. Stride, P.
- 6 Vadgama, M. Edirisinghe, Fabrication of biomaterials via controlled protein bubble
- 7 generation and manipulation, Biomacromolecules 12 (2011) 4291-4300.
- 8 [56] Y. Li, J. Li, H. Zhang, Y. Su, Microbubble Suspensions Prepared via
- 9 Electrohydrodynamic Jetting Process, 2008 International Conference on BioMedical
- 10 Engineering and Informatics 2 (2008) 445-449.
- 11 [57] Z. Ahmad, E.S. Thian, J. Huang, M.J. Edirisinghe, S.M. Best, S.N. Jayasinghe, W.
- 12 Bonfield, R.A. Brooks, N. Rushton, Deposition of nano-hydroxyapatite particles utilising
- 13 direct and transitional electrohydrodynamic processes, J. Mater. Sci. Mater. Med. 19 (2008)
- 14 3093-3104.
- 15 [58] A.J. Halliday, S.E. Moulton, G.G. Wallace, M.J. Cook, Novel methods of antiepileptic
- 16 drug delivery -- polymer-based implants, Adv. Drug Deliv. Rev. 64 (2012) 953-964.
- 17 [59] P.P. Mehta, V.S. Pawar, 22 Electrospun nanofiber scaffolds: Technology and
- 18 applications, in: Inamuddin, A.M. Asiri and A. Mohammad (Eds.), Applications of
- 19 Nanocomposite Materials in Drug Delivery, Woodhead Publishing, 2018, pp. 509-573.
- 20 [60] S. Saghazadeh, C. Rinoldi, M. Schot, S.S. Kashaf, F. Sharifi, E. Jalilian, K. Nuutila, G.
- 21 Giatsidis, P. Mostafalu, H. Derakhshandeh, K. Yue, W. Swieszkowski, A. Memic, A.
- 22 Tamayol, A. Khademhosseini, Drug delivery systems and materials for wound healing
- 23 applications, Adv. Drug Deliv. Rev. 127 (2018) 138-166.
- 24 [61] P. Rattanakit, S.E. Moulton, K.S. Santiago, S. Liawruangrath, G.G. Wallace, Extrusion
- printed polymer structures: a facile and versatile approach to tailored drug delivery platforms,
 Int. J. Pharm. 422 (2012) 254-263.
- 27 [62] P. Fattahi, J.T. Dover, J.L. Brown, 3D Near-Field Electrospinning of Biomaterial
- 28 Microfibers with Potential for Blended Microfiber-Cell-Loaded Gel Composite Structures,
- Adv. Healthc. Mater. 6 (2017) 10.1002/adhm.201700456. doi: 10.1002/adhm.201700456.
 Empt 2017 Jun 20
- 30 Epub 2017 Jun 29.
- 31 [63] J. Xie, J. Jiang, P. Davoodi, M.P. Srinivasan, C. Wang, Electrohydrodynamic
- 32 atomization: A two-decade effort to produce and process micro-/nanoparticulate materials,
- 33 Chemical Engineering Science 125 (2015) 32-57.
- [64] A.M. Gañán-Calvo, J. Dávila, A. Barrero, Current and droplet size in the electrospraying
 of liquids. Scaling laws, J. Aerosol Sci. 28 (1997) 249-275.
- 36 [65] W. Wei, Z. Gu, S. Wang, Y. Zhang, K. Lei, K. Kase, Numerical simulation of the cone-
- 37 jet formation and current generation in electrostatic spray Modeling as regards space
- 38 charged droplet effect, J Micromech Microengineering 23 (2012) 015004.

- 1 [66] D.N. Nguyen, C. Clasen, G. Van den Mooter, Pharmaceutical Applications of
- 2 Electrospraying, J. Pharm. Sci. 105 (2016) 2601-2620.
- [67] S. Zhang, C. Campagne, F. Salaün, Influence of Solvent Selection in the Electrospraying
 Process of Polycaprolactone, Applied Sciences 9 (2019) 402.
- 5 [68] S.K. Boda, X. Li, J. Xie, Electrospraying an enabling technology for pharmaceutical and 6 biomedical applications: A review, J. Aerosol Sci. 125 (2018) 164-181.
- 7 [69] J.M. Deitzel, J. Kleinmeyer, D. Harris, N.C. Beck Tan, The effect of processing
- 8 variables on the morphology of electrospun nanofibers and textiles, Polymer 42 (2001) 2619 272.
- [70] N. Bhardwaj, S.C. Kundu, Electrospinning: A fascinating fiber fabrication technique,
 Biotechnol. Adv. 28 (2010) 325-347.
- [71] T.J. Sill, H.A. von Recum, Electrospinning: Applications in drug delivery and tissue
 engineering, Biomaterials 29 (2008) 1989-2006.
- 14 [72] B. Niu, P. Shao, Y. Luo, P. Sun, Recent advances of electrosprayed particles as
- 15 encapsulation systems of bioactives for food application, Food Hydrocoll. 99 (2020) 105376.
- 16 [73] N. Arya, S. Chakraborty, N. Dube, D.S. Katti, Electrospraying: a facile technique for
- synthesis of chitosan-based micro/nanospheres for drug delivery applications, J. Biomed.
 Mater, Res. B. Appl. Biomater, 88 (2009) 17-31.
- [74] Q. Wang, Z. Wang, S. Yang, B. Li, H. Xu, K. Yu, J. Wang, Experimental study on
 electrohydrodynamic atomization (EHDA) in stable cone-jet with middle viscous and low
 conductive liquid, Exp. Therm. Fluid Sci. 121 (2021) 110260.
- 22 [75] V. Guarino, W.K. Khodir, L. Ambrosio, Biodegradable microparticles and nanoparticles
- 23 by electrospraying techniques, J. Appl. Biomater. Funct. Mater. 10 (2012) 191-196.
- [76] M. Ikeuchi, R. Tane, K. Ikuta, Electrospray deposition and direct patterning of polylactic
 acid nanofibrous microcapsules for tissue engineering, Biomed. Microdevices 14 (2012) 3543.
- [77] A. Jaworek, A. Krupa, CLASSIFICATION OF THE MODES OF EHD SPRAYING, J.
 Aerosol Sci. 30 (1999) 873-893.
- [78] A. Frenot, I.S. Chronakis, Polymer nanofibers assembled by electrospinning, Current
 Opinion in Colloid & Interface Science 8 (2003) 64-75.
- 31 [79] A. Pawar, S. Thakkar, M. Misra, A bird's eye view of nanoparticles prepared by
- electrospraying: advancements in drug delivery field, Journal of Controlled Release 286
 (2018) 179-200.
- 34 [80] H. Fessi, F. Puisieux, J.P. Devissaguet, N. Ammoury, S. Benita, Nanocapsule formation
- by interfacial polymer deposition following solvent displacement, Int. J. Pharm. 55 (1989)
 R1-R4.

- 1 [81] S. Salatin, J. Barar, M. Barzegar-Jalali, K. Adibkia, F. Kiafar, M. Jelvehgari,
- 2 Development of a nanoprecipitation method for the entrapment of a very water soluble drug
- 3 into Eudragit RL nanoparticles, Res. Pharm. Sci. 12 (2017) 1-14.
- 4 [82] M. Chidambaram, K. Krishnasamy, Modifications to the conventional nanoprecipitation
- technique: an approach to fabricate narrow sized polymeric nanoparticles, Adv. Pharm. Bull.
 4 (2014) 205-208.
- 7 [83] P. Paximada, Y. Echegoyen, A.A. Koutinas, I.G. Mandala, J.M. Lagaron, Encapsulation
- 8 of hydrophilic and lipophilized catechin into nanoparticles through emulsion electrospraying,
- 9 Food Hydrocoll. 64 (2017) 123-132.
- 10 [84] N. Bock, M.A. Woodruff, D.W. Hutmacher, T.R. Dargaville, Electrospraying, a
- 11 Reproducible Method for Production of Polymeric Microspheres for Biomedical
- 12 Applications, Polymers 3 (2011).
- 13 [85] L.S. Usmanova, M.A. Ziganshin, I.T. Rakipov, N.M. Lyadov, A.E. Klimovitskii, T.A.
- 14 Mukhametzyanov, A.V. Gerasimov, Microspherical Particles of Solid Dispersion of
- 15 Polyvinylpyrrolidone K29-32 for Inhalation Administration, Biomed. Res. Int. 2018 (2018)
- 16 2412156.
- 17 [86] D. Allawadi, N. Sharma, S. Singh, S. Arora, Solid Disperisons: A Review on Drug
- delivery system and Solubility enhancement, International Journal of Pharmaceutical
 Sciences and Research 4 (2013) 2094-2105.
- 20 [87] Y. Gao, Y. Bai, D. Zhao, M. Chang, Z. Ahmad, J. Li, Tuning Microparticle Porosity
- during Single Needle Electrospraying Synthesis via a Non-Solvent-Based Physicochemical
 Approach, Polymers 7 (2015).
- 23 [88] B. Almería, W. Deng, T.M. Fahmy, A. Gomez, Controlling the morphology of
- electrospray-generated PLGA microparticles for drug delivery, J. Colloid Interface Sci. 343
 (2010) 125-133.
- 26 [89] R. Bakhshi, Z. Ahmad, M. Soric, E. Stride, M. Edirisinghe, Nanoparticle delivery
- systems formed using electrically sprayed co-flowing excipients and active agent, J. Biomed.
 Nanotechnol 7 (2011) 782-793.
- [90] Y. Zhou, Y. Wan, Z. Ye, Z. He, Q. Liu, Y. Shi, How Hepatitis B virus causes cirrhosis
 and liver cancer, Medical Hypotheses 108 (2017) 52-53.
- 31 [91] Y. Lee, F. Mei, M. Bai, S. Zhao, D. Chen, Release profile characteristics of
- biodegradable-polymer-coated drug particles fabricated by dual-capillary electrospray, J.
 Controlled Release 145 (2010) 58-65.
- 34 [92] Z. Ekemen, Z. Ahmad, E. Stride, D. Kaplan, M. Edirisinghe, Electrohydrodynamic
- bubbling: an alternative route to fabricate porous structures of silk fibroin based materials,
- 36 Biomacromolecules 14 (2013) 1412-1422.
- [93] L. Guo, Q. Zhao, M. Wang, Triggered release of anticancer drug and theranostics from
 microspherical vehicles made by coaxial electrospray, (2016).

- 1 [94] Z. Yao, C. Zhang, Z. Ahmad, Y. Peng, M. Chang, Microparticle Formation via Tri-
- needle Coaxial Electrospray at Stable Jetting Modes, Ind Eng Chem Res 59 (2020) 1442314432.
- [95] Z. Ahmad, H.B. Zhang, U. Farook, M. Edirisinghe, E. Stride, P. Colombo, Generation of
 multilayered structures for biomedical applications using a novel tri-needle coaxial device
 and electrohydrodynamic flow, J. R. Soc. Interface 5 (2008) 1255-1261.
- 7 [96] C. Zhang, M. Chang, Y. Li, Y. Qi, J. Wu, Z. Ahmad, J. Li, Janus particle synthesis via
- 8 aligned non-concentric angular nozzles and electrohydrodynamic co-flow for tunable drug
- 9 release, RSC Adv. 6 (2016) 77174-77178.
- [97] A.L. Yarin, E. Zussman, Upward needleless electrospinning of multiple nanofibers,
 Polymer 45 (2004) 2977-2980.
- [98] H. Wang, Z. Chen, B. Liu, L. Ren, L. Zhuang, K. Sun, L. Jiang, Needleless electrospray
 of magnetic film from magnetization-induced cone array, Mater. Manuf. Process. (2017) 1-6.
- 14 [99] R. Bocanegra, D. Galán, M. Márquez, I.G. Loscertales, A. Barrero, Multiple
- 15 electrosprays emitted from an array of holes, J. Aerosol Sci. 36 (2005) 1387-1399.
- 16 [100] Z. Ahmad, M. Nangrejo, M. Edirisinghe, E. Stride, P. Colombo, H.B. Zhang,
- 17 Engineering a material for biomedical applications with electric field assisted processing,
- 18 Applied Physics A 97 (2009) 31-37.
- 19 [101] S. Labbaf, H. Ghanbar, E. Stride, M. Edirisinghe, Preparation of multilayered
- 20 polymeric structures using a novel four-needle coaxial electrohydrodynamic device,
 21 Macromol. Rapid Commun. 35 (2014) 618-623.
- 22 [102] M. Parhizkar, P.J.T. Reardon, J.C. Knowles, R.J. Browning, E. Stride, P.R. Barbara,
- 23 A.H. Harker, M. Edirisinghe, Electrohydrodynamic encapsulation of cisplatin in poly (lactic-
- 24 co-glycolic acid) nanoparticles for controlled drug delivery, Nanomedicine: Nanotechnology,
- 25 Biology and Medicine 12 (2016) 1919-1929.
- 26 [103] Z. Mai, J. Chen, T. He, Y. Hu, X. Dong, H. Zhang, W. Huang, F. Ko, W. Zhou,
- Electrospray biodegradable microcapsules loaded with curcumin for drug delivery systems
 with high bioactivity, RSC Adv. 7 (2017) 1724-1734.
- [104] M. Enayati, Z. Ahmad, E. Stride, M. Edirisinghe, Preparation of polymeric carriers for
 drug delivery with different shape and size using an electric jet, Curr. Pharm. Biotechnol. 10
 (2009) 600-608.
- [105] Y. Gao, D. Zhao, M. Chang, Z. Ahmad, X. Li, H. Suo, J. Li, Morphology control of
 electrosprayed core–shell particles via collection media variation, Mater Lett 146 (2015) 59 64.
- 35 [106] T. Shams, U.E. Illangakoon, M. Parhizkar, A.H. Harker, S. Edirisinghe, M. Orlu, M.
- 36 Edirisinghe, Electrosprayed microparticles for intestinal delivery of prednisolone, J. R. Soc.
- 37 Interface 15 (2018) 20180491. doi: 10.1098/rsif.2018.0491.

- 1 [107] A.S. Kranthi Kiran, K. Madhumathi, T.S. Sampath Kumar, Electrosprayed titania
- nanocups for protein delivery, Colloid and Interface Science Communications 12 (2016) 17 20.
- 4 [108] M.W. Chang, E. Stride, M. Edirisinghe, Controlling the thickness of hollow polymeric
- 5 microspheres prepared by electrohydrodynamic atomization, J. R. Soc. Interface 7 Suppl 4 6 (2010) S451-60.
- [109] X. Ju, X. Wang, Z. Liu, R. Xie, W. Wang, L. Chu, Red-blood-cell-shaped chitosan
 microparticles prepared by electrospraying, Particulogy 30 (2016).
- 9 [110] R. Haj-Ahmad, M. Rasekh, K. Nazari, E.V. Onaiwu, B. Yousef, S. Morgan, D. Evans,
- 10 M.W. Chang, J. Hall, C. Samwell, Z. Ahmad, Stable increased formulation atomization using
- 11 a multi-tip nozzle device, Drug Deliv. Transl. Res. 8 (2018) 1815-1827.
- [111] W. Li, D. Yu, K. Chen, G. Wang, G.R. Williams, Smooth preparation of ibuprofen/zein
 microcomposites using an epoxy-coated electrospraying head, Mater Lett 93 (2013) 125-128.
- 14 [112] P. Mehta, A.A. Al-Kinani, M.S. Arshad, N. Singh, S.M. van der Merwe, M. Chang,
- 15 R.G. Alany, Z. Ahmad, Engineering and Development of Chitosan-Based Nanocoatings for
- 16 Ocular Contact Lenses, Journal of Pharmaceutical Sciences 108 (2019) 1540-1551.
- 17 [113] Z.P. Liu, L. Cui, D.G. Yu, Z.X. Zhao, L. Chen, Electrosprayed core-shell solid
- dispersions of acyclovir fabricated using an epoxy-coated concentric spray head, Int. J.
 Nanomedicine 9 (2014) 1967-1977.
- 20 [114] S. Labbaf, S. Deb, G. Cama, E. Stride, M. Edirisinghe, Preparation of
- 21 multicompartment sub-micron particles using a triple-needle electrohydrodynamic device, J.
- 22 Colloid Interface Sci. 409 (2013) 245-254.
- [115] W. Kim, S.S. Kim, Multishell encapsulation using a triple coaxial electrospray system,
 Anal. Chem. 82 (2010) 4644-4647.
- 25 [116] M. Ma, A. Chiu, G. Sahay, J.C. Doloff, N. Dholakia, R. Thakrar, J. Cohen, A. Vegas,
- 26 D. Chen, K.M. Bratlie, T. Dang, R.L. York, J. Hollister-Lock, G.C. Weir, D.G. Anderson,
- 27 Core-shell hydrogel microcapsules for improved islets encapsulation, Adv. Healthc. Mater. 2
- 28 (2013) 667-672.
- 29 [117] H. Park, P.H. Kim, T. Hwang, O.J. Kwon, T.J. Park, S.W. Choi, C.O. Yun, J.H. Kim,
- Fabrication of cross-linked alginate beads using electrospraying for adenovirus delivery, Int.
 J. Pharm. 427 (2012) 417-425.
- 32 [118] S. Zafar, M.S. Arshad, S. Fatima, A. Ali, A. Zaman, E. Sayed, M.W. Chang, Z. Ahmad,
- 33 COVID-19: Current Developments and Further Opportunities in Drug Delivery and
- 34 Therapeutics, Pharmaceutics 12 (2020) 945. doi: 10.3390/pharmaceutics12100945.
- 35 [119] H. Lee, S. An, S. Kim, B. Jeon, M. Kim, I.S. Kim, Readily Functionalizable and
- 36 Stabilizable Polymeric Particles with Controlled Size and Morphology by Electrospray,
- 37 Scientific Reports 8 (2018) 15725.

- 1 [120] K. Khoshakhlagh, M. Mohebbi, A. Koocheki, A. Allafchian, Encapsulation of D-
- 2 limonene in Alyssum homolocarpum seed gum nanocapsules by emulsion electrospraying:
- 3 Morphology characterization and stability assessment, Bioactive Carbohydrates and Dietary
- 4 Fibre 16 (2018) 43-52.
- 5 [121] S. Bhaskar, K.M. Pollock, M. Yoshida, J. Lahann, Towards designer microparticles:
 6 simultaneous control of anisotropy, shape, and size, Small 6 (2010) 404-411.
- [122] S. Park, J.B. Kim, S. Lee, J.H. Bang, Y.S. Kim, Synthesis of titanium oxide nanocups
 by electrospraying, Mater Lett 69 (2012) 34-36.
- 9 [123] H. Abdelakder, D.I.D.S.A. Hussain, N. Abdullah, Review on micro-encapsulation with 10 Chitosan for pharmaceuticals applications, MOJ Current Research & Reviews 1 (2018).
- 11 [124] F.L. Zhou, A. Chirazi, J.E. Gough, P.L. Hubbard Cristinacce, G.J.M. Parker, Hollow
- 12 Polycaprolactone Microspheres with/without a Single Surface Hole by Co-Electrospraying,
- 13 Langmuir 33 (2017) 13262-13271.
- 14 [125] M. Nangrejo, E. Bernardo, P. Colombo, U. Farook, Z. Ahmad, E. Stride, M.
- Edirisinghe, Electrohydrodynamic forming of porous ceramic capsules from a preceramicpolymer, Mater Lett 63 (2009) 483-485.
- [126] Y. Wu, S.J. Kennedy, R.L. Clark, Polymeric particle formation through electrospraying
 at low atmospheric pressure, J. Biomed. Mater. Res. B. Appl. Biomater. 90 (2009) 381-387.
- 19 [127] S. Rahmani, C.H. Villa, A.F. Dishman, M.E. Grabowski, D.C. Pan, H. Durmaz, A.C.
- 20 Misra, L. Colón-Meléndez, M.J. Solomon, V.R. Muzykantov, J. Lahann, Long-circulating
- Janus nanoparticles made by electrohydrodynamic co-jetting for systemic drug delivery
 applications, J. Drug Target. 23 (2015) 750-758.
- 23 [128] B. Sanchez-Vazquez, A.J.R. Amaral, D. Yu, G. Pasparakis, G.R. Williams,
- Electrosprayed Janus Particles for Combined Photo-Chemotherapy, AAPS PharmSciTech 18
 (2017) 1460-1468.
- 26 [129] Z. Ma, H. Ji, D. Tan, G. Dong, Y. Teng, J. Zhou, M. Guan, J. Qiu, M. Zhang, Large-
- 27 scale preparation of strawberry-like, AgNP-doped SiO2 microspheres using the
- 28 electrospraying method, Nanotechnology 22 (2011) 305307-4484/22/30/305307. Epub 2011
 29 Jul 1.
- 30 [130] K. Khanum, S. S. P. Ramamurthy, Design and Morphology Control of Thiophene 21 Derivative through Electrogramming Using Various Solventa, BSC, Adv. 5 (2015)
- 31 Derivative through Electrospraying Using Various Solvents, RSC Adv. 5 (2015).
- 32 [131] C. Zhang, Z.C. Yao, Q. Ding, J.J. Choi, Z. Ahmad, M.W. Chang, J.S. Li, Tri-Needle
- 33 Coaxial Electrospray Engineering of Magnetic Polymer Yolk-Shell Particles Possessing
- 34 Dual-Imaging Modality, Multiagent Compartments, and Trigger Release Potential, ACS
- 35 Appl. Mater. Interfaces 9 (2017) 21485-21495.
- 36 [132] M. Eltayeb, E. Stride, M. Edirisinghe, Electrosprayed core-shell polymer-lipid
- an nanoparticles for active component delivery, Nanotechnology 24 (2013) 465604-
- 38 4484/24/46/465604. Epub 2013 Oct 28.

- [133] J. Xie, W.J. Ng, L.Y. Lee, C. Wang, Encapsulation of protein drugs in biodegradable
 microparticles by co-axial electrospray, J. Colloid Interface Sci. 317 (2008) 469-476.
- 3 [134] P. Mehta, R. Haj-Ahmad, A. Al-Kinani, S. Arshad, M. Chang, R. Alany, Z. Ahmad,
- 4 Approaches in topical ocular drug delivery and developments in the use of contact lenses as
- 5 drug-delivery devices, Therapeutic Delivery 8 (2017) 521-541.
- 6 [135] A. Bohr, J. Kristensen, E. Stride, M. Dyas, M. Edirisinghe, Preparation of microspheres
- containing low solubility drug compound by electrohydrodynamic spraying, Int. J. Pharm.
 412 (2011) 59-67.
- 9 [136] E. Bell, M. Zeles-Hahn, C. Lengsfeld, Understanding EHDA and Protein Stability,
 2007.
- 11 [137] A. Sosnik, K.P. Seremeta, Advantages and challenges of the spray-drying technology
- 12 for the production of pure drug particles and drug-loaded polymeric carriers, Adv. Colloid
- 13 Interface Sci. 223 (2015) 40-54.
- 14 [138] Y. Fukui, T. Maruyama, Y. Iwamatsu, A. Fujii, T. Tanaka, Y. Ohmukai, H.
- 15 Matsuyama, Preparation of monodispersed polyelectrolyte microcapsules with high
- 16 encapsulation efficiency by an electrospray technique, Colloids Surf. Physicochem. Eng.
- 17 Aspects 370 (2010) 28-34.
- [139] W.Y. Fung, M.T. Liong, K.H. Yuen, Preparation, in-vitro and in-vivo characterisation
 of CoQ10 microparticles: electrospraying-enhanced bioavailability, J. Pharm. Pharmacol. 68
 (2016) 159-169.
- 21 [140] A. Alehosseini, B. Ghorani, M. Sarabi Jamab, N. Tucker, Principles of Electrospraying:
- A New Approach in Protection of Bioactive Compounds in Foods, Crit. Rev. Food Sci. Nutr.
- 23 58 (2018) 2346-2363.
- 24 [141] Y. Alyassin, E.G. Sayed, P. Mehta, K. Ruparelia, M.S. Arshad, M. Rasekh, J.
- 25 Shepherd, I. Kucuk, P.B. Wilson, N. Singh, M. Chang, D.G. Fatouros, Z. Ahmad,
- 26 Application of mesoporous silica nanoparticles as drug delivery carriers for chemotherapeutic
- 27 agents, Drug Discov. Today 25 (2020) 1513-1520.
- 28 [142] M. Zamani, M.P. Prabhakaran, E.S. Thian, S. Ramakrishna, Protein encapsulated core-
- shell structured particles prepared by coaxial electrospraying: Investigation on material and
 processing variables, Int. J. Pharm. 473 (2014) 134-143.
- 31 [143] M. Jafari-Nodoushan, J. Barzin, H. Mobedi, Size and morphology controlling of PLGA
- microparticles produced by electro hydrodynamic atomization, Polym. Adv. Technol. 26
 (2015) 502-513.
- [144] A. Smeets, R. Koekoekx, W. Ruelens, M. Smet, C. Clasen, G. Van den Mooter, Gastro resistant encapsulation of amorphous solid dispersions containing darunavir by coaxial
 electrospraying, Int. J. Pharm. 574 (2020) 118885.
- [145] W. Dai, L.C. Dong, Y. Song, Nanosizing of a drug/carrageenan complex to increase
 solubility and dissolution rate, Int. J. Pharm. 342 (2007) 201-207.

- 1 [146] L. Xiaoyan, Z. Zheng, D. Yu, X. Liu, Y. Qu, H. Li, Electrosprayed sperical
- 2 ethylcellulose nanoparticles for an improved sustained-release profile of anticancer drug, 3 Cellulose 24 (2017).
- 4 [147] E.A. Hosny, Formulation and comparative evaluation of bioadhesive containing
- 5 diclofenac sodium and commercial enteric coated tablets in-vitro and in dogs. Int. J. Pharm. 6 133 (1996) 149-153.
- 7 [148] S. Hao, B. Wang, Y. Wang, Y. Xu, Enteric-coated sustained-release nanoparticles by
- 8 coaxial electrospray: Preparation, characterization, and in vitro evaluation, Journal of
- 9 Nanoparticle Research 16 (2014).
- 10 [149] D. Yin, Y. Yang, H. Cai, F. Wang, D. Peng, L. He, Gambogic acid-loaded
- 11 electrosprayed particles for site-specific treatment of hepatocellular carcinoma, Mol. Pharm. 12 11 (2014) 4107-4117.
- 13 [150] R. Cavalli, A. Bisazza, R. Bussano, M. Trotta, A. Civra, D. Lembo, E. Ranucci, P.
- 14 Ferruti, Poly(amidoamine)-Cholesterol Conjugate Nanoparticles Obtained by Electrospraying
- as Novel Tamoxifen Delivery System, J. Drug Deliv. 2011 (2011) 587604. 15
- 16 [151] Y. Gao, D. Zhao, M. Chang, Z. Ahmad, J. Li, Optimising the shell thickness-to-radius
- 17 ratio for the fabrication of oil-encapsulated polymeric microspheres, Chem. Eng. J. 284 18 (2016) 963-971.
- 19 [152] D. Yu, X. Zheng, Y. Yang, X. Li, G.R. Williams, M. Zhao, Immediate release of
- 20 helicid from nanoparticles produced by modified coaxial electrospraying, Appl. Surf. Sci.
- 21 473 (2019) 148-155.
- 22 [153] Y. Cao, F. Liu, Y. Chen, T. Yu, D. Lou, Y. Guo, P. Li, Z. Wang, H. Ran, Drug release
- from core-shell PVA/silk fibroin nanoparticles fabricated by one-step electrospraving, 23
- Scientific Reports 7 (2017) 11913. 24
- 25 [154] S. Han, P. Dwivedi, F.A. Mangrio, M. Dwivedi, R. Khatik, D.E. Cohn, T. Si, R.X. Xu,
- 26 Sustained release paclitaxel-loaded core-shell-structured solid lipid microparticles for
- 27 intraperitoneal chemotherapy of ovarian cancer, Artif. Cells Nanomed Biotechnol. 47 (2019) 957-967. 28
- 29 [155] M. Zhang, Y. Tang, Z. Zhu, H. Zhao, J. Yao, D. Sun, Paclitaxel and etoposide-loaded
- Poly (lactic-co-glycolic acid) microspheres fabricated by coaxial electrospraying for dual 30
- 31 drug delivery, J. Biomater. Sci. Polym. Ed. 29 (2018) 1949-1963.
- 32 [156] W. Kim, S.S. Kim, Synthesis of biodegradable triple-layered capsules using a triaxial 33 electrospray method, Polymer 52 (2011) 3325-3336.
- 34 [157] E. Ghassami, J. Varshosaz, A. Jahanian-Najafabadi, M. Minaiyan, P. Rajabi, E. Hayati,
- 35 Pharmacokinetics and in vitro/in vivo antitumor efficacy of aptamer-targeted Ecoflex(®)
- nanoparticles for docetaxel delivery in ovarian cancer, Int. J. Nanomedicine 13 (2018) 493-36 504.
- 37

- 1 [158] S.Y. Lee, J.J. Lee, J.H. Park, J.Y. Lee, S.H. Ko, J.S. Shim, J. Lee, M.Y. Heo, D.D.
- 2 Kim, H.J. Cho, Electrosprayed nanocomposites based on hyaluronic acid derivative and
- 3 Soluplus for tumor-targeted drug delivery, Colloids Surf. B Biointerfaces 145 (2016) 267-274.
- 4
- 5 [159] Y.B. Patil, U.S. Toti, A. Khdair, L. Ma, J. Panyam, Single-step surface
- 6 functionalization of polymeric nanoparticles for targeted drug delivery, Biomaterials 30 7 (2009) 859-866.
- 8 [160] J. Kim, Y. Yamagata, B. Kim, S. Takeuchi, H. Toshiro, SU-8 based Cantilever
- Biosensor with the Electrospray Deposited Protein Film, The proceedings of the JSME 9 10 annual meeting 2003.5 (2003) 21-22.
- 11 [161] M.F. Sohail, M. Rehman, H.S. Sarwar, S. Naveed, O. Salman, N.I. Bukhari, I. Hussain,
- 12 T.J. Webster, G. Shahnaz, Advancements in the oral delivery of Docetaxel: challenges,
- 13 current state-of-the-art and future trends, Int. J. Nanomedicine 13 (2018) 3145-3161.
- 14 [162] V. Volarevic, B. Djokovic, M.G. Jankovic, C.R. Harrell, C. Fellabaum, V. Djonov, N.
- 15 Arsenijevic, Molecular mechanisms of cisplatin-induced nephrotoxicity: a balance on the
- 16 knife edge between renoprotection and tumor toxicity, J. Biomed. Sci. 26 (2019) 25-019-
- 17 0518-9.
- 18 [163] R. Esfahani, H. Jun, S. Rahmani, A. Miller, J. Lahann, Microencapsulation of Live
- 19 Cells in Synthetic Polymer Capsules, ACS Omega 2 (2017) 2839-2847.
- [164] A. Raza, T. Rasheed, F. Nabeel, U. Hayat, M. Bilal, H.M.N. Iqbal, Endogenous and 20
- 21 Exogenous Stimuli-Responsive Drug Delivery Systems for Programmed Site-Specific
- 22 Release, Molecules 24 (2019) 1117. doi: 10.3390/molecules24061117.
- 23 [165] Y. Wu, J.A. MacKay, J.R. McDaniel, A. Chilkoti, R.L. Clark, Fabrication of elastin-
- 24 like polypeptide nanoparticles for drug delivery by electrospraying, Biomacromolecules 10
- (2009) 19-24. 25
- 26 [166] Y. Wu, B. Yu, A. Jackson, W. Zha, L.J. Lee, B.E. Wyslouzil, Coaxial
- 27 Electrohydrodynamic Spraying: A Novel One-Step Technique To Prepare
- 28 Oligodeoxynucleotide Encapsulated Lipoplex Nanoparticles, Mol. Pharmaceutics 6 (2009)
- 29 1371-1379.
- 30 [167] S. Jeelani, R.C. Reddy, T. Maheswaran, G.S. Asokan, A. Dany, B. Anand,
- 31 Theranostics: A treasured tailor for tomorrow, J. Pharm. Bioallied Sci. 6 (2014) S6-8.
- 32 [168] M.B. Miller, Y.W. Tang, Basic concepts of microarrays and potential applications in 33 clinical microbiology, Clin. Microbiol. Rev. 22 (2009) 611-633.
- 34 [169] V.N. Morozov, T.Y. Morozova, Electrospray Deposition as a Method for Mass
- 35 Fabrication of Mono- and Multicomponent Microarrays of Biological and Biologically 36 Active Substances, Anal. Chem. 71 (1999) 3110-3117.
- 37 [170] B.N. Dubey, M. Howarth, P. Paximada, Electrosprayed particles derived from nano-38 emulsions as carriers of fish oil, (2018).

- 1 [171] H. Valo, L. Peltonen, S. Vehviläinen, M. Karjalainen, R. Kostiainen, T. Laaksonen, J.
- 2 Hirvonen, Electrospray Encapsulation of Hydrophilic and Hydrophobic Drugs in Poly(L-
- 3 lactic acid) Nanoparticles, Small 5 (2009) 1791-1798.
- 4 [172] M.A. Collier, E.M. Bachelder, K.M. Ainslie, Electrosprayed Myocet-like Liposomes:
- 5 An Alternative to Traditional Liposome Production, Pharm. Res. 34 (2017) 419-426.
- 6 [173] R. Sridhar, S. Ramakrishna, Electrosprayed nanoparticles for drug delivery and 7 pharmaceutical applications, Biomatter 3 (2013) e24281.
- 8 [174] Y.N. Gavhane, A.V. Yadav, Loss of orally administered drugs in GI tract, Saudi
- 9 pharmaceutical journal : SPJ : the official publication of the Saudi Pharmaceutical Society 20
 10 (2012) 331-344.
- 11 [175] B. Homayun, X. Lin, H.J. Choi, Challenges and Recent Progress in Oral Drug Delivery
- 12 Systems for Biopharmaceuticals, Pharmaceutics 11 (2019) 129. doi:
- 13 10.3390/pharmaceutics11030129.
- 14 [176] H. Jayan, M. Maria Leena, S.K. Sivakama Sundari, J.A. Moses, C.
- 15 Anandharamakrishnan, Improvement of bioavailability for resveratrol through encapsulation
- 16 in zein using electrospraying technique, Journal of Functional Foods 57 (2019) 417-424.
- 17 [177] R. Sun, C. Shen, S. Shafique, O. Mustapha, T. Hussain, I.U. Khan, Y. Mehmood, K.
- 18 Anwer, Y. Shahzad, A.M. Yousaf, Electrosprayed Polymeric Nanospheres for Enhanced
- 19 Solubility, Dissolution Rate, Oral Bioavailability and Antihyperlipidemic Activity of
- 20 Bezafibrate, International journal of nanomedicine 15 (2020) 705-715.
- [178] Y. Cao, B. Wang, Y. Wang, D. Lou, Dual Drug Release from Core–Shell Nanoparticles
 with Distinct Release Profiles, J. Pharm. Sci. 103 (2014) 3205-3216.
- 23 [179] A.M. Yousaf, O. Mustapha, D.W. Kim, D.S. Kim, K.S. Kim, S.G. Jin, C.S. Yong, Y.S.
- 24 Youn, Y. Oh, J.O. Kim, H. Choi, Novel electrosprayed nanospherules for enhanced aqueous
- solubility and oral bioavailability of poorly water-soluble fenofibrate, International journal of
 nanomedicine 11 (2016) 213-221.
- 27 [180] J. Zhang, Z. Xie, N. Zhang, J. Zhong, Chapter 13 Nanosuspension drug delivery
- 28 system: preparation, characterization, postproduction processing, dosage form, and
- 29 application, in: E. Andronescu and A.M. Grumezescu (Eds.), Nanostructures for Drug
- 30 Delivery, Elsevier, 2017, pp. 413-443.
- 31 [181] J.D. Spahn, S.J. Szefler, Chapter 16 Pharmacology of the Lung and Drug Therapy, in:
- 32 L.M. Taussig and L.I. Landau (Eds.), Pediatric Respiratory Medicine (Second Edition),
- 33 Mosby, Philadelphia, 2008, pp. 219-233.
- 34 [182] M.A. Collier, R.D. Junkins, M.D. Gallovic, B.M. Johnson, M.M. Johnson, A.N.
- 35 Macintyre, G.D. Sempowski, E.M. Bachelder, J.P.-. Ting, K.M. Ainslie, Acetalated Dextran
- 36 Microparticles for Codelivery of STING and TLR7/8 Agonists, Molecular pharmaceutics 15
- 37 (2018) 4933-4946.

- 1 [183] M.D. Gallovic, K.L. Schully, M.G. Bell, M.A. Elberson, J.R. Palmer, C.A. Darko, E.M.
- 2 Bachelder, B.E. Wyslouzil, A.M. Keane-Myers, K.M. Ainslie, Acetalated Dextran
- 3 Microparticulate Vaccine Formulated via Coaxial Electrospray Preserves Toxin
- 4 Neutralization and Enhances Murine Survival Following Inhalational Bacillus Anthracis
- 5 Exposure, Adv. Healthc. Mater. 5 (2016) 2617-2627.
- [184] K. Ita, Transdermal Delivery of Drugs with Microneedles-Potential and Challenges,
 Pharmaceutics 7 (2015) 90-105.
- 8 [185] G. Shingade, Review on: Recent Trend on Transdermal Drug Delivery System, JDDT 2
 9 (2012).
- 10 [186] R. Ali, P. Mehta, P. Kyriaki Monou, M.S. Arshad, E. Panteris, M. Rasekh, N. Singh, O.
- 11 Qutachi, P. Wilson, D. Tzetzis, M. Chang, D.G. Fatouros, Z. Ahmad,
- 12 Electrospinning/electrospraying coatings for metal microneedles: A design of experiments
- 13 (DOE) and quality by design (QbD) approach, European Journal of Pharmaceutics and
- 14 Biopharmaceutics 156 (2020) 20-39.
- 15 [187] X. He, J. Sun, J. Zhuang, H. Xu, Y. Liu, D. Wu, Microneedle System for Transdermal
- 16 Drug and Vaccine Delivery: Devices, Safety, and Prospects, Dose-response : a publication of
- 17 International Hormesis Society 17 (2019) 1559325819878585-1559325819878585.
- 18 [188] R. Haj-Ahmad, H. Khan, M.S. Arshad, M. Rasekh, A. Hussain, S. Walsh, X. Li, M.W.
- 19 Chang, Z. Ahmad, Microneedle Coating Techniques for Transdermal Drug Delivery,
- 20 Pharmaceutics 7 (2015) 486-502.
- 21 [189] S. Tort, N.B. Mutlu Agardan, D. Han, A.J. Steckl, In vitro and in vivo evaluation of
- 22 microneedles coated with electrosprayed micro/nanoparticles for medical skin treatments, J. 23 Microoneansul 37 (2020) 517 527
- 23 Microencapsul. 37 (2020) 517-527.
- 24 [190] S. Yaqoubi, K. Adibkia, A. Nokhodchi, S. Emami, A.A. Alizadeh, H. Hamishehkar, M.
- 25 Barzegar-Jalali, Co-electrospraying technology as a novel approach for dry powder inhalation
- formulation of montelukast and budesonide for pulmonary co-delivery, Int. J. Pharm. 591
 (2020) 119970.
- 28 [191] M. Zamani, M.P. Prabhakaran, S. Ramakrishna, Advances in drug delivery via
- electrospun and electrosprayed nanomaterials, International journal of nanomedicine 8 (2013)
 2997-3017.
- [192] A. Sosnik, Production of Drug-Loaded Polymeric Nanoparticles by Electrospraying
 Technology, Journal of Biomedical Nanotechnology 10 (2015).
- [193] A. Gomez, The electrospray and its application to targeted drug inhalation, Respir.
 Care 47 (2002) 1419-31; discussion 1431-3.
- 35 [194] Y. Hong, Y. Li, Y. Yin, D. Li, G. Zou, Electrohydrodynamic atomization of quasi-
- monodisperse drug-loaded spherical/wrinkled microparticles, J. Aerosol Sci. 39 (2008) 525 536.

- 1 [195] A.H. Bahmanpour, M. Ghaffari, S. Ashraf, M. Mozafari, 23 - Nanotechnology for
- 2 pulmonary and nasal drug delivery, in: M. Mozafari (Ed.), Nanoengineered Biomaterials for 3 Advanced Drug Delivery, Elsevier, 2020, pp. 561-579.
- 4 [196] B.T. Midhun, K.T. Shalumon, K. Manzoor, R. Jayakumar, S.V. Nair, M. Deepthy,
- 5 Preparation of budesonide-loaded polycaprolactone nanobeads by electrospraving for
- 6 controlled drug release, J. Biomater. Sci. Polym. Ed. 22 (2011) 2431-2444.
- 7 [197] P. Mehta, A. Al-Kinani, O. Qutachi, M.S. Arshad, A. Algahtani, M. Chang, W.M.
- 8 Amoaku, R.G. Alany, Z. Ahmad, Assessing the ex vivo permeation behaviour of
- 9 functionalised contact lens coatings engineered using an electrohydrodynamic technique,
- 10 Journal of Physics: Materials 2 (2018) 014002.
- 11 [198] P. Mehta, L. Justo, S. Walsh, M.S. Arshad, C.G. Wilson, C.K. O'Sullivan, S.M.
- 12 Moghimi, I.S. Vizirianakis, K. Avgoustakis, D.G. Fatouros, Z. Ahmad, New platforms for
- 13 multi-functional ocular lenses: engineering double-sided functionalized nano-coatings, J.
- 14 Drug Target. 23 (2015) 305-310.
- 15 [199] P. Mehta, A.A. Al-Kinani, R. Haj-Ahmad, M.S. Arshad, M. Chang, R.G. Alany, Z.
- Ahmad, Electrically atomised formulations of timolol maleate for direct and on-demand 16
- 17 ocular lens coatings, European Journal of Pharmaceutics and Biopharmaceutics 119 (2017)
- 18 170-184.
- 19 [200] N. Hazeri, H. Tavanai, A.R. Moradi, Production and properties of electrosprayed
- 20 sericin nanopowder, Science and technology of advanced materials 13 (2012) 035010-21 035010.
- 22 [201] P. Toman, C. Lien, Z. Ahmad, S. Dietrich, J.R. Smith, Q. An, É Molnár, G.J.
- 23 Pilkington, D.C. Górecki, J. Tsibouklis, E. Barbu, Nanoparticles of alkylglyceryl-dextran-
- 24 graft-poly(lactic acid) for drug delivery to the brain: Preparation and in vitro investigation,
- 25 Acta Biomaterialia 23 (2015) 250-262.
- 26 [202] A. Rohani Shirvan, A. Bashari, N. Hemmatinejad, New insight into the fabrication of
- 27 smart mucoadhesive buccal patches as a novel controlled-drug delivery system. European 28 Polymer Journal 119 (2019) 541-550.
- 29 [203] S. Madhusudhan, D. Gowda, N.V. Gupta, A. R. An overview on topical drug delivery
- 30 system - Updated review, International Journal of Research in Pharmaceutical Sciences 11
- (2020) 368-385. 31
- 32 [204] H. Nie, Y. Fu, C. Wang, Paclitaxel and suramin-loaded core/shell microspheres in the 33 treatment of brain tumors, Biomaterials 31 (2010) 8732-8740.
- 34 [205] N. Maurmann, L.E. Sperling, P. Pranke, Electrospun and Electrosprayed Scaffolds for 35 Tissue Engineering, Adv. Exp. Med. Biol. 1078 (2018) 79-100.
- 36 [206] S. Sahoo, W.C. Lee, J.C. Goh, S.L. Toh, Bio-electrospraying: A potentially safe
- 37 technique for delivering progenitor cells, Biotechnol. Bioeng. 106 (2010) 690-698.

- 1 [207] A. Abeyewickreme, A. Kwok, J.R. McEwan, S.N. Jayasinghe, Bio-electrospraying
- embryonic stem cells: interrogating cellular viability and pluripotency, Integr. Biol. 1 (2009)
 260-266.
- 4 [208] K. Ng, P. Joly, S.N. Jayasinghe, B. Vernay, R. Knight, S.P. Barry, J. McComick, D.
- 5 Latchman, A. Stephanou, Bio-electrospraying primary cardiac cells: In vitro tissue creation
- 6 and functional study, Biotechnology Journal 6 (2011) 86-95.
- 7 [209] Z. McCrea, Y. Arnanthigo, S. Cryan, S. O'Dea, A Novel Methodology for Bio-
- 8 electrospraying Mesenchymal Stem Cells that Maintains Differentiation, Immunomodulatory
- 9 and Pro-reparative Functions, Journal of Medical and Biological Engineering 38 (2017) 1-17.
- 10 [210] D.I. Braghirolli, F. Zamboni, G.A.X. Acasigua, P. Pranke, Association of
- 11 electrospinning with electrospraying: a strategy to produce 3D scaffolds with incorporated
- 12 stem cells for use in tissue engineering, International journal of nanomedicine 10 (2015)
- 13 5159-5169.
- 14 [211] E. San Thian, Z. Ahmad, J. Huang, M.J. Edirisinghe, S.N. Jayasinghe, D.C. Ireland,
- 15 R.A. Brooks, N. Rushton, W. Bonfield, S.M. Best, The role of electrosprayed apatite
- 16 nanocrystals in guiding osteoblast behaviour, Biomaterials 29 (2008) 1833-1843.
- 17 [212] N. Bock, T.R. Dargaville, G.T.S. Kirby, D.W. Hutmacher, M.A. Woodruff, Growth
- 18 Factor-Loaded Microparticles for Tissue Engineering: The Discrepancies of In Vitro
- 19 Characterization Assays, Tissue engineering.Part C, Methods 22 (2016) 142-154.
- 20 [213] A.R. Tsiapla, V. Bakola, V. Karagkiozaki, E. Pavlidou, S. Logothetidis, Biodegradable
- Electrosprayed NPs as Drug Carriers for Optimal Treatment of Orthopaedic Infections,
 Materials Today: Proceedings 19 (2019) 110-116.
- 23 [214] Q. Guo, J. Mather, P. Yang, M. Boden, P. Mather, Fabrication of Polymeric Coatings
- 24 with Controlled Microtopographies Using an Electrospraying Technique, PLOS ONE 10
- 25 (2015) e0129960.
- 26 [215] M. Zamani, M.P. Prabhakaran, J. Varshosaz, P.S. Mhaisalkar, S. Ramakrishna,
- 27 Electrosprayed Montelukast/poly (lactic-co-glycolic acid) particle based coating: A new
- therapeutic approach towards the prevention of in-stent restenosis, Acta Biomaterialia 42
- 29 (2016) 316-328.
- 30 [216] C.M. McKittrick, M.J. Cardona, R.A. Black, C. McCormick, Development of a
- Bioactive Polymeric Drug Eluting Coronary Stent Coating Using Electrospraying, Ann.
 Biomed. Eng. 48 (2020) 271-281.
- 33 [217] N. Ruecha, R. Rangkupan, N. Rodthongkum, O. Chailapakul, Novel paper-based
- 34 cholesterol biosensor using graphene/polyvinylpyrrolidone/polyaniline nanocomposite,
- 35 Biosens. Bioelectron. 52 (2014) 13-19.
- 36 [218] S. Agarwal, A. Greiner, On the way to clean and safe electrospinning—green
- electrospinning: emulsion and suspension electrospinning, Polym. Adv. Technol. 22 (2011)
 372-378.

	Journal Pre-proofs
1	
2	List of tables:
3	Table 1: Summary of the types of particulate structures engineered using the electrospraying
4	process (*ID – inner diameter, OD – outer diameter)
5	
6	Table 2: Summary of the applications of EHDA technologies for drug delivery, therapeutics
7	and pharmaceutics
8	
9	List of figures:
10	Figure 1: Schematic diagram of the EHDA process
11	a) Spinneret needle designs: a1) Single-needle elecctrospraying a2) Coaxial, a3) Multi-
12	tip emitter, a4) triple-needle coaxial [95] a5) angular nozzle arrangements [96] a5i) θ
13	= 30°, a5ii) θ = 60°, a6) needleless approach using an external magnetic field [98] a7)
14	needleless approach using multiple orifices [99].
15	b) Jetting modes: b1) no flow, b2) initial dripping, b3) microdripping, b4) oscillating jet,
16	b5) unstable jetting, b6) stable jetting (Taylor-cone), b7) unstable multi-jet, b8) stable
17	multi-jet.
18	
19	Figure 2: Schematic diagram of forces acting in the liquid cone at the tip of the capillary
20	needle
21	
22	Figure 3: Various types of particulate structures engineered using the electrospraying
23	process:
24	
25	a) 6% PCL matrix active-polymer microspheres loaded with paclitaxel [36] b)Titanium oxide
26	nanocups produced via electrospraying nitromethane solution of 8 wt% PMMA and titanium
27	tetraisopropoxide [122] c) Triple layered PLGA–PCL–PMSQ nanoparticles [114] d) Islet cells
28	encapsulated in core-shell hydrogel microcapsules using a concentric nozzle and coaxial jetting
29	[116] e) 5 w/v % PCL in DCM polymeric porous carrier particles at an externally applied
30	pressure of -150 mmHg [126] f) PVP-RBC (5%w/v PVP in ethanol, 0.1% w/v Rose Bengal
31	and 0.86% w/v Carmofur) Janus particles via side-by-side electrospraying [128] g) Spindle
32	particles obtained from electrospraying D-limonene (20%) with Alyssum homolocarpum seed
33	gum (0.75%) and Tween 20 (0.1%) [120] h) microrods generated from co-jetting of a 3.4%

Journal Pre-proofs

1 w/w solution (in 95:5 v/v chloroform: dimethylformamide (DMF)) of each polymer (PLGA 85:15 and PLGA 50:50) and triethylamine (3.6 vol% of solvent) [121] i) Red-blood-cell-shaped 2 3 chitosan microparticles with evaporable ethanol (20 vol%) and diffusible DMSO (30 vol%) [109] j) Fibril spike-like structures from electrosprayed DTCPA (1.5 wt%) in Chlorobenzene 4 5 [130] k) 3% Ethylene/vinyl acetate copolymer (EVAC) toroidal-like donut shaped 6 microparticles loaded with anti-cancer drug paclitaxel [36] I) Disc shaped biphasic particles 7 fabricated from co-jetting of 1.3% w/w solution (in 95:5 v/v chloroform: dimethylformamide 8 (DMF)) of each polymer (PLGA 85:15 and PLGA 50:50) [121] m) 0.5g Silver nanoparticle-9 doped SiO₂ strawberry-like microspheres [129] n) Adenovirus encapsulating cross linked alginate (0.5% wt) beads [117] o) PCL/chloroform (5 wt. %) + PEG/chloroform (15 wt.%) 10 hollow microspheres with single surface hole collected in an ethanol bath substrate by coaxial 11 12 spraying [124] **p**) 2%ibuprofen/12%zein microparticles coated with an epoxy resin needle [111] **q**) Magnetic volk shell particles prepared by coaxial electrospraving using 20% w/v PCL 13 14 in glacial acetic acid with magnetic nanoparticles and model probes. Nile blue/PCL (outer shell), silicone oil/Sudan Red G (central layer) and Acridine yellow/PCL (inner layer) [131] r) 15 16 Four layered structure prepared with a four-needle coaxial device, electrospraying various dyes 17 in the polymeric layers of PCL, PEG, PMSQ and PLGA [101].

18

19 Figure 4: Applications of EHDA engineered particles as established and emerging 20 technologies

21

22 a) Paclitaxel and suramin loaded core-shell microspheres for glioma treatment [204] b) 23 Montelukast and budesonide microparticles for treatment of asthma [190] c) Cisplatin 24 encapsulated nanoparticles as chemotherapeutic agents [102] d) Prednisolone loaded toroidal microstructures for treatment of inflammatory bowel disease and colorectal cancer [106] e) 25 26 Gambogic acid loaded nanoparticles to treat hepatocellular carcinoma [149] **f**) Sericin 27 nanoparticles with antibacterial properties [200] g) Uniform nanoparticle coatings for 28 microneedles [189] h) Nanospherules for improved oral bioavailability of fenofibrate [179] i) 29 Timolol maleate contact lens coating for glaucoma treatment [199] 30

31

	NOZZLE GEOMETRY	NOZZLE SIZE (MM)	VOLTAGE (kV)	MATRIX /CARRIER	ACTIVE /INNER MATERIAL	PARTICLE SIZE (μm)	APPLICATIONS	KEY DETAILS	REFEREENCE
Active-polymer composite microspheres	Single-needle	0.91	Nozzle- 8.8 Ring-7.1	PCL	Paclitaxel	17	Anti-cancer drug delivery	Formulation filled with 6% PCL matrix solid microparticles and loaded with Paclitaxel. Using a flow rate of 3mL/h fabricated matrix solid structures.	[36]
Active-polymer composite structure	Single-needle	ID- 0.60mm OD-0.91mm	20	-	Ibuprofen and Zein	1.78±0.31	Controlled release drug delivery platforms	Formulation consisted of 2% ibuprofen and 12% zein (w/v) microparticles in 85% ethanol. Epoxy resin was rapidly coated around the needle tip. Particles were collected onto an aluminium foil substrate at a distance of 15 cm.	[111]
structures	Coaxial (triple needle device)	Inner- OD (0.5mm) ID (0.2mm) Central- OD (1.5mm) ID (1.0mm) Outer- OD (2.6mm) ID (2.0mm)	6.9-7.9	PLGA (outer) PCL (central) PMSQ (inner)	-	0.2(±0.008)- 0.32 (±0.08)	Used for a variety of controlled release drug delivery systems	Spherical multi-layered nanoparticles were generated using a combination of polymers: 5wt% PLGA in DMC, 6wt% PCL in DCM and 12wt% EtOH. Varying flow rates were used 300- (outer)50(central)-5(inner) μL/h, which when atomised were collected at a working distance of 15cm.	[114]
Red blood cell shaped	Single-needle	Not specified	5-6	0	Chitosan	Various sizes but as small as 5 (similar to red blood cells)	Autofluorescence imaging for biomedical applications	Particles mimicking red blood cells were fabricated containing chitosan, evaporable ethanol (20vol%) and diffusible DMSO (30%vol%).Collecting distance was kept at 5cm as well as a flow rate of 120 µl/h.	[109]
Spindle shaped	Single-needle	Not specified	15	Alyssum homolocarpu m seed gum	D-limonene Tween 20	Various sizes below 0.5	Nutraceutical stability	Spindle-like particles were obtained by emulsion Electrospraying. D- limonene (20%) with Alyssum homolocarpum seed gum (0.75%) and Tween 20 (0.1%). A flow rate of 0.05ml/h- 0.1ml/h was utilised, and particles were collected at working	[120]

								distance of 15cm.	
Rod shaped	Dual capillary needle system	Not specified	6 ±-0.1	PLGA DMF Triethylamin e	-	Mean lengths of 18.37 ±6.17	Drug delivery and cell targeting	Microrods were generated using a 3.4% w/w solution (in 95:5 v/v chloroform: dimethylformamide (DMF)) of each polymer (PLGA 85:15 and PLGA 50:50). A flow rate of 0.45ml/h and a collection distance of 28-33cm, yielded rod-shaped microparticulates.	[121]
Fibril shaped	Single-needle	Not specified	15	Chlorobenze ne.	(7, 9-di (thiophen-2- yl)-8H- cyclopenta[a]acenaphthyl en-8-one) (DTCPA)	6-8	Photoactive materials for biomedical imaging	Fibril shaped structures from DTCPA (1.5wt%) in Chlorobenzene. Flow rate of 1.5 ml/hr is used as well as an aluminium plate with a collection distance of 12cm from the nozzle.	[130]
Donut shaped	Single-needle	0.34mm	Nozzle- 8.8 Ring-7.1	Ethylene/vin yl acetate copolymer (EVAC)	Paclitaxel	Around 10 in size	Anti-cancer therapeutics	A formulation consisting of 3% EVAC and loaded with paclitaxel fabricated toroidal-like donut shapes at a flow rate of 3ml/h.	[36]
Strawberry shaped	Single-needle	Not specified	15	SiO2	Ag-NPs	0.95	Antibacterial materials for biomedical applications	0.5g Silver nanoparticle(Ag-NPs)- doped SiO2 strawberry shaped microspheres were fabricated at a flow rate of 0.2ml/h, where particles were collected at a distance of 20cm. Ag- NPs were doped inside the SiO2 microspheres as well as embedded on the surface, therefore appearing strawberry-like.	[129]
Hollow microspheres	Co- electrosprayin g	Not specified	9	PCL/Chlorof orm (shell)	PEG/ Chloroform (core)	Below 15	Tumour cell mimicking phantoms for anti- cancer drug	PCL/chloroform (5 wt. %) and PEG/chloroform (15 wt.%) were co- electrosprayed and collected in an ethanol bath substrate using a flow	[124]

							delivery	rate of 1.0/3.0ml/h (core/shell) and. collected at a distance of 20cm.	
Nanocups	Single-needle	Not specified	12	Polymethy lmethacryl ate (PMMA) Titanium tetraisopro poxide	Nitromethane	Diameter of around 0.5	Applications in photocatalysis and drug delivery systems	Titanium oxide nanocups were produced by electrospraying nitromethane, 8% PMMA and titanium tetraisopropoxide . A flow rate of 2ml/h was used as well as a collection distance of 10cm.	[122]
Porous microcarriers	Single-needle	Not specified	12	PCL	-	3.5±0.4	Biomolecules, drug encapsulation and inhalable drug delivery platforms	5 w/v % PCL in DCM when electrosprayed fabricated porous microcarrier particles using an externally applied pressure of -150 mmHg. A flow rate of 0.5ml/h was applied, and particles were collected at a working distance of 30cm on an aluminium disc (additional 0.5kV).	[126]
Four layered structure	Coaxial (four needle)	-Needle 1 outermost (OD 4mm, ID 3.2mm) -Needle 2 second outermost (OD 2.6mm, ID 2.0mm) -Needle 3 second innermost (OD 1.5mm, ID 1.0mm) -Needle 4- innermost (OD 0.5mm, ID 0.2mm)	9-12	PCL PEG PMSQ PLGA	Various dyes (Evans blue, Pyronin B, Pinacyanol chloride, Hematoxylin)	0.62 ± 0.15	Multi-layered delivery system as a controlled release dosage form for combinatorial drug targeting	A four layered particulate system was developed : DCM:PEG 90:10 DCM:PLGA 95:5 DCM:PCL 97:3 EtOH:PMSQ 88:12. Additionaly a flow rate of 50-50-25- 10 (µl min-1) and a working distance of 10cm was utilised.	[101]
Janus	Side-by-side	Two 0.6mm	17	PVP	Rose Bengal	$0.607 \pm$	Photochemo-	PVP-RBC particles were fabricated	[128]

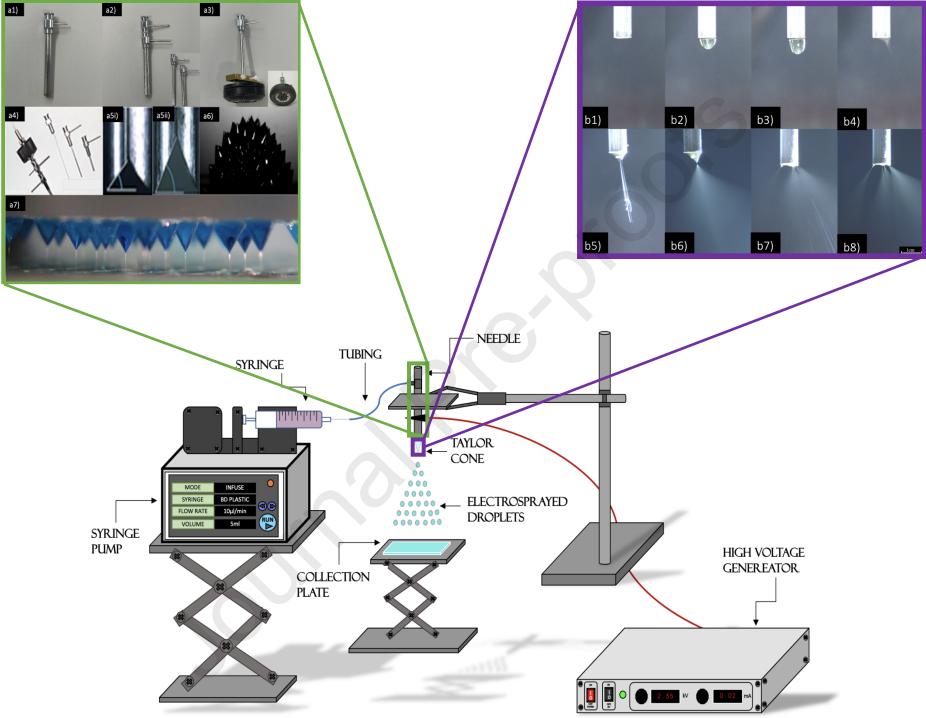
	electrosprayin g	ID spinnerets			Carmofur	0.191	therapy and imaging agents in biomedical applications	<i>via</i> side-by-side electrospraying using 5%w/v PVP in ethanol, 0.1% w/v Rose Bengal and 0.86% w/v Carmofur. Janus particles contained Rose Bengal in one compartment whilst Carmofour was in the other. Flow rate was at 0.5ml/h where particles were collected at a working distance of 20cm.	
Yolk-shell particles	Coaxial Electrosprayin g (triple needle device)	Inner needle- OD (0.50 mm) ID (0.31mm). Central needle- OD (1.60 mm) ID (1.07 mm) Outer needle- OD (2.85 mm) ID (2.26 mm)	10	PCL Silicone oil	Multiple probes	13.5 ± 1.74	Theranostic applications requiring multi- drug release	Magnetic yolk shell particles were prepared using 20% w/v PCL in glacial acetic acid with magnetic nanoparticles and model probes. Nile blue/PCL (outer shell), silicone oil/Sudan Red G (central layer) and Acridine yellow/PCL (inner layer). Flow rates varied for each layer: 10ml/h (outer shell), 2.5ml/h (central layer) and 1.6ml/h (inner layer).	[131]
Disc shaped	Dual capillary needle system	Not specified	6 ±0.1	PLGA DMF		3.41± 0.72	Biomedical imaging and drug delivery platforms	Disc shaped particles were fabricated using a 1.3% w/w solution (in 95:5 v/v chloroform: dimethylformamide (DMF)) of polymers (PLGA 85:15 and PLGA 50:50). A flow rate of 0.15ml/h was used, and discoid- biphasic particles were collected at a working distance of 28-33 cm.	[121]
<u>Ta</u>	<u>ble 1</u>		0						

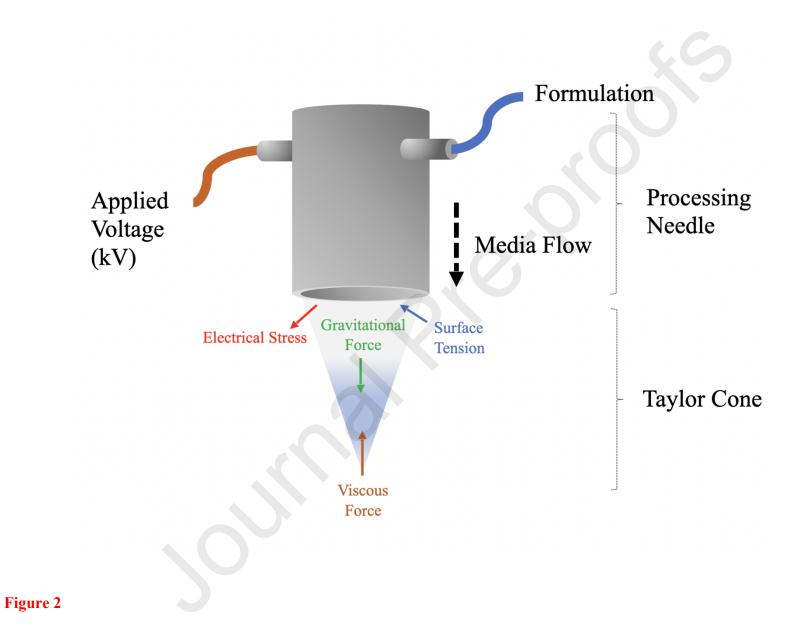
APPLICATIONS	EXPERIMENTAL DETAILS	STRUCTURE AND SIZE	COMMENTS	REFERENCE
Brain Delivery of paclitaxel and suramin microspheres for the treatment of brain tumors	Co-axial ES to produce microspheres with PLLA core and PLGA shell as a dual drug delivery system	Core-shell microspheres 10 – 20 μm	Electrosprayed microspheres were superior in inducing apoptosis compared to single drugs. <i>In-vivo</i> studies showed tumor inhibition of U87 glioma.	[204]
Ocular ES as a coating for contact lenses in the treatment of glaucoma	ES used to encapsulate timolol maleate into polymeric matrixes; PVP, PNIPAM and PVP:PNIPAM (50:50%w/w)	Fibrous and particulate structures of dimensions <200 nm were formed.	<i>In-vitro</i> release studies found a biphasic release for all formulations where the PNIPAM and timolol maleate coating released 89.8% of the drug after a period of 24 hours	[199]
<u>Nasal</u> Budesonide for the treatment of asthma	Single needle ES for the loading of Budesonide into PCL	Nanobeads with a Size of 116.1 ± 19 nm	Through ES drug encapsulation efficiency of 75 ± 2.4 % was achieved. Controlled drug release was obtained <i>in-vitro</i> at pH 7.4 and 5.6	[196]
Oral ES to improve the aqueous solubility and oral bioavailability of fenofibrate	Single needle ES to fabricate novel nanospherules. PVP and Labrafil M 2125 were used as a carrier for fenofibrate.	Nanospherules <200 nm in size and fenofibrate was in an amorphous state	2.5-fold improvement in oral bioavailability. Solubility of 32.5% μg/mL and a dissolution rate of 85% within 10 minutes was observed	[179]
Pulmonary ES to fabricate dry powder inhalation of montelukast and budesonide for the treatment of asthma	An excipient free formulation was used in single needle ES; montelukast behaved as both an API and a carrier for budesonide	Smooth spherical particles 1 – 5 μm suitable for respirable particles	Therapeutic efficiency in controlling asthmatic episodes was significantly enhanced when budesonide was inhaled in combination with orally administered montelukast thus highlighting the advantage of producing a formulation consisting of both montelukast and budesonide for dry powder inhalation	[190]

<u>Topical</u> Sericin with antibacterial properties for moisturisers and sun creams.	Single-needle ES (needle diameter 0.6 mm) Applied voltage of 15kV was used whilst	Uniform spheroidal nanoparticles Mean size < 80nm	Decreased concentration and feed rate with increased collection distance caused reduced particle size. Absorbance increased by 6x for sericin	[200]
<u>Transdermal</u> ES loaded (dye or insulin) nanoparticle coatings for microneedles	Concentration, flow rate and collection distance were varied Single needle ES used to fabricate drug-loaded nanoparticles as uniform coatings for microneedles	Spherical particles produced with a particle size of: Dye: 515 nm Insulin: 522nm	nanoparticles compared to sericin sponge. Microneedles with the optimised coating demonstrated a >70% transfer into porcine skins. <i>In-vivo</i> studies of coated microneedles on diabetic rats demonstrated a decrease in blood glucose levels fluctuations, compared to subcutaneous injections	[189]
Parenteral ES of cisplatin; anticancer drug used in chemotherapeutic treatments	Single needle ES for producing cisplatin encapsulated PLGA nanoparticles. Effect of applied voltage, flow rate and concentration of cisplatin on particle size was assessed.	Spherical particles with smooth surface morphology in the 550nm range	When using a flow rate of 5μ l/min, applied voltage of 16kV, decreased PLGA concentration (2%w/w) and increased cisplatin concentration (0.2%w/w) particle diameter was reduced from 1.2µm to 550nm.	[102].
<u>Liver</u> A site-specific delivery system of gambogic acid for the treatment of hepatocellular carcinoma	Single needle ES was used to encapsulate gambogic acid into a PDLLA matrix. Particles were collected using ultrapure water and residual solvent was removed <i>via</i> vacuum drying. Preparation parameters were varied thus producing different sized particles <i>In-vivo</i> liver studies were carried out	Optimal nanoparticles were spherical and were 185.6 nm in size	Upon increasing particle size from the nanoscale to the microscale, gambogic acid release rate sharply decreased. 2 weeks after administration, hepatocellular carcinoma mice treated with the particles demonstrated lower degree of tumor invasion and cell lesions as well as recovered liver function.	[149].

Colon	Coaxial ES was used to fabricate	Toroidal donut-like	Dissolution studies revealed site-specific	[106]
ES of prednisolone for	Eudragit L100-55 microparticles	structures	release of prednisolone for the targeted	
-			treatment of inflammatory bowel disease	
11	01			
disease and colon	needles; 6	were 1.7 µm and 0.6		
cancer		μm respectively		

<u>Table 2</u>





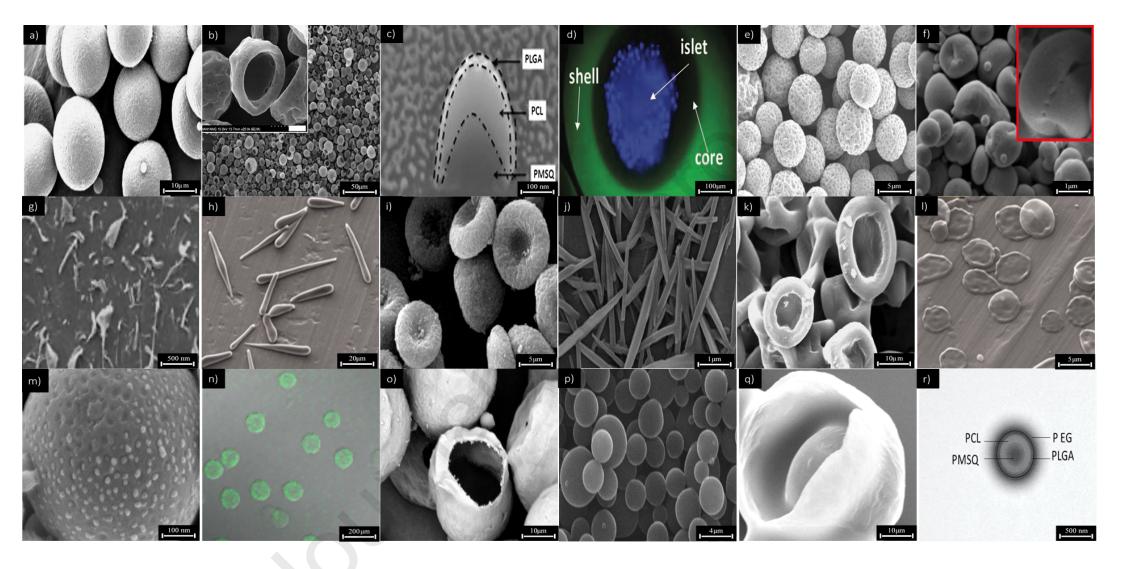
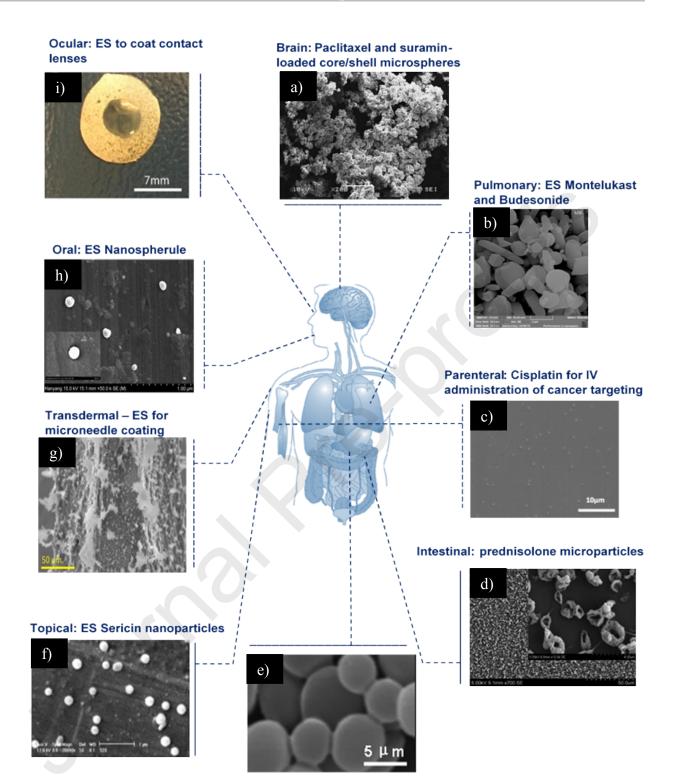


Figure 3

Journal Pre-proofs



Liver: Gabogic acid loaded particles for Hepatocellular Carcinoma

Figure 4

