

### Marine Ecology Progress Series

<b>Manuscript:</b>	MEPS-2021-06-001/R1 RESUBMISSION
<b>Title:</b>	Functional thermal limits are determined by rate of warming during simulated marine heatwaves
<b>Authors(s):</b>	Rebecca De Leij (Corresponding Author), Laura Grange (Co-author), Lloyd Peck (Co-author)
<b>Keywords:</b>	Climate change, Echinoderm , Extreme warming events, Polar, Segmented regression, Sub-lethal limits, Thermal tolerance
<b>Type:</b>	Research Article

1 Article to Marine Ecology Press Series

2

3 **Functional thermal limits are determined by rate of**  
4 **warming during simulated marine heatwaves**

5

6 **Rebecca De Leij<sup>1,2</sup>, Laura J. Grange<sup>2</sup>, Lloyd S. Peck<sup>3</sup>**

7 <sup>1</sup>University of Southampton, Waterfront Campus, European Way,

8 Southampton, SO14 3ZH

9 <sup>2</sup>School of Ocean Sciences, Bangor University, Bangor, Gwynedd, North

10 Wales, LL57 2DG

11 <sup>3</sup>British Antarctic Survey, High Cross, Madingley Rd, Cambridge, CB3 0ET

12 Correspondence: Email: [rid1n17@soton.ac.uk](mailto:rid1n17@soton.ac.uk); Phone: +44(0)7544553603

13 Running head: Functional thermal limits during marine heatwaves

1 **ABSTRACT**

2 Marine heatwaves (MHWs) are increasing in both intensity and frequency  
3 against a backdrop of gradual warming associated with climate change. In the  
4 context of MHWs, animals are likely to experience sub-lethal, rather than lethal  
5 effects, defining long-term limits to survival and/or impacting individual and  
6 population fitness. This study investigated how functional sub-lethal limits track  
7 critical thresholds and how this relationship changes with warming rate. To this  
8 end we monitored basic functioning, specifically the ability to right, feed and  
9 assimilate energy, as well as oxygen consumption rate in the common Antarctic  
10 sea urchin, *Sterechinus neumayeri*. Water temperature in experimental  
11 systems was increased at rates of 1°C day<sup>-1</sup>, 0.5°C day<sup>-1</sup> and 0.3°C day<sup>-1</sup>, in  
12 line with the characteristics of MHW events previously experienced at the site  
13 where the study urchins were collected on the Antarctica Peninsula.  
14 Functioning was assessed during the simulation of MHWs and sub-lethal limits  
15 determined when the rate of functional degradation changed as temperature  
16 increased. Results suggest that thermal sensitivity varies between the key  
17 biological functions measured, with the ability to right having the highest  
18 thermal threshold. Arguably, the most interesting result was that functions  
19 deteriorated at lower temperatures when warming was more rapid (1°C day<sup>-1</sup>),  
20 contrary to lethal critical thresholds, which were reached at lower temperatures  
21 when warming was slower (0.3°C day<sup>-1</sup>). MHWs and their impacts extend far  
22 beyond Antarctica and in this context, our analyses indicate that the onset rate  
23 of MHWs is critical in determining an organism's ability to tolerate short-term  
24 elevated temperatures.

1 Key words: Extreme warming events, sub-lethal limits, thermal tolerance,  
2 climate change, polar, segmented regression, echinoderm

### 3 **1. INTRODUCTION**

4 Historical temperature records have now detected positive temperature trends  
5 for the majority of the Earth's surface (Myrvoll-Nilsen et al. 2019), with the  
6 oceans being key to the regulation and capture of much of the excess heat  
7 present in the atmosphere (Marshall et al. 2015). As a result, marine  
8 environments are changing both physically and biochemically (Bopp et al.  
9 2013). Included in these changes is the occurrence of marine heat waves  
10 (MHWs), which are increasing in duration, magnitude and frequency, with  
11 alarming ecological consequences (Garrabou et al. 2009, Rubio-Portillo et al.  
12 2016, Oliver et al. 2018).

13 Physiological flexibility of species is crucial to survival during MHW events  
14 (Peck 2011) and species at low latitudes may be able to acclimate and adapt  
15 across generations to altered environments (Donelson et al. 2012, Salinas &  
16 Munch 2012, Clark et al. 2019a). As a result, predicting effects of MHWs on  
17 lower latitude species may need to consider shifting thermal ranges as these  
18 species adapt to climate change. It is unlikely that the same will apply to  
19 Antarctic species, since many are physiologically limited by their capacity to  
20 acclimate and adapt to new temperatures because of their long generation  
21 times and delayed reproductive maturity (Peck et al. 2014, Peck 2018). For  
22 example, several invertebrate species such as the Antarctic scallop  
23 *Adamussium colbecki*, the limpet *Nacella concinna*, and the bivalves, *Laternula*  
24 *elliptica* and *Adacnarca nitens*, take 4 – 7 years to mature. The Antarctic  
25 bivalve, *Aequiyoldia eightsi*, starts reproducing at around 12 years (Peck &

1 Bullough 1993) and the brachiopod *Liothyrella uva*, can take up to 18 years  
2 before brooding young (Peck 2005, 2018, Oliver et al. 2019).

3 Predicting species and ecosystem responses to MHWs is challenging, owed to  
4 the past infrequency and variability of each event (Oliver et al. 2018). However,  
5 if we can track the functional deterioration of organisms when temperatures  
6 exceed their typical thermal range, this can inform our understanding of the  
7 relationships between the sub-lethal and lethal limits likely to be encountered  
8 during MHW events.

9 For organisms with slow growth and development and long generation times,  
10 like many of those found in Antarctica, thermal stress caused by MHWs is likely  
11 to trigger other mechanisms for survival such as biochemical and cellular stress  
12 responses (e.g. Clark & Peck 2009, Payton et al. 2016). Biochemical and  
13 genetic mechanisms, including a range of chaperone proteins, provide a short-  
14 term buffer that allow functioning to continue temporarily at temperatures  
15 outside an organism's thermal niche (Deschaseaux et al. 2010, Clark et al.  
16 2019b). Once animals are no longer able to maintain basic functions by these  
17 mechanisms, the sub-lethal limit to survival is reached.

18 Data on the functional thermal limits of species and MHW characteristics (i.e.  
19 rate, magnitude and duration) at which these thresholds are reached are rare,  
20 especially in fluctuating environments (Janecki et al. 2010, Peck et al. 2014,  
21 Ardor Bellucci & Smith 2019). Little is known about functional deterioration as  
22 a species approaches its critical thermal limit, and in the context of MHWs,  
23 animals are likely to experience temperatures that cause sub-lethal, rather than  
24 lethal effects, defining long-term limits to survival and/or inhibiting population  
25 health (Pörtner et al. 2007).

1 This study aims to understand how functional (sub-lethal) limits track critical  
2 (lethal) limits and how this relationship changes with warming rate during a  
3 simulated MHW. To this purpose, we monitored the ability to right, feed and  
4 assimilate energy as well as oxygen consumption rate, in the common Antarctic  
5 sea urchin, *Sterechinus neumayeri*.

## 6 **2. MATERIALS AND METHODS**

### 7 2.1 Sample site and animal collections

8 *Sterechinus neumayeri* were sampled from South Cove, Rothera Point  
9 (67°34'09.1"S 68°07'52.7"W), from sites near the British Antarctic Survey's  
10 Rothera Research Station on the Western Antarctic Peninsula (WAP) during  
11 December 2019 (Figure S1). 120 adult urchins (test diameter range, 28 mm –  
12 49 mm) were SCUBA-diver collected at depths of 10-20 m and returned to the  
13 Rothera aquarium facility within two hours of collection.

14 *Sterechinus neumayeri* is one of the most common and locally abundant  
15 members of the Antarctic marine shallow benthos, forming a significant  
16 component of the benthic community (Brockington 2001, Pierrat et al. 2012),  
17 with reported densities up to 600 m<sup>2</sup> (Barnes & Brockington 2003). It is a major  
18 scavenger of dead organisms and in iceberg scours on the shallow Antarctic  
19 seabed (Dunlop et al. 2014), and is a significant grazer and bioturbator of  
20 sediments (Lenihan et al. 2018). Because of this *S. neumayeri* is an important  
21 carbon transformer in Antarctic shallow seas. Further to this, due to its  
22 abundance and ease of maintenance in laboratory culture systems, *S.*  
23 *neumayeri* has been the subject of extensive study of its embryonic and larval  
24 development, which is highly extended, and up to in excess of 100 days (Bosch  
25 et al. 1987). It has also been the subject of studies of the effects of temperature

1 on embryonic and larval development (Stanwell-Smith & Peck 1998), the  
2 impact of ocean acidification on reproduction (Suckling et al. 2014) and energy  
3 budgets (Morley et al. 2016). Furthermore, it has been shown that there are  
4 long-term cycles in its reproduction (De Leij et al. 2021). These factors all make  
5 *S. neumayeri* one of the most important members of the Antarctic shallow  
6 benthic ecosystem and key to investigating responses to MHWs.

## 7 2.2 Experimental set-up and warming system

8 A decade of temperature data (1997-2017) from Ryder Bay on the WAP  
9 (sourced from the Rothera Time-Series (RaTS) environmental monitoring  
10 programme (Clarke et al. 2008, Venables et al. 2013)) was used in the R  
11 package “heatwaveR” (Schlegel & Smit 2018), to detect past warming events  
12 (Figure 1) (see details of warming event analysis methodology and  
13 characteristics summary in the Supplementary Materials, Text S1, Table S1,  
14 Figure S2). Studying the characteristics of these past warming events, including  
15 onset rate and magnitude, allowed us to set realistic warming rates for the  
16 experimental systems.

17 Urchins were held in flow-through aquaria (170 L) at ambient temperatures  
18 typical for December and January (-1.5°C to +0.5°C) for six weeks on a  
19 continuous light regime. During this time, animals were not fed to allow any  
20 ingested food to be processed and the production of faeces to cease. The  
21 cessation of faeces production is an indicator that metabolic rates had reached  
22 a “standard” level at the start of the experiment. Previous research suggests  
23 that these urchins are able to sustain and experience natural periods of  
24 starvation for up six months during winter (Brockington 2001), and hence six

1 weeks without feeding was unlikely to be detrimental to the physiological  
2 metrics measured in this study. Previous studies of oxygen consumption in  
3 Antarctic marine invertebrates has demonstrated that standard levels are  
4 reached in less, and often significantly less, than this time in the brachiopod  
5 *Liothyrella uva* and the limpet *Nacella concinna* (Peck 1989), in the amphipod  
6 *Waldeckia obesa* (Chapelle et al. 1994), in the isopod *Glyptonotus antarcticus*  
7 (Robertson et al. 2001), and in the sea star *Odontaster Validus* (Peck et al.  
8 2008).

9 After urchins were maintained in the flow-through aquarium (170 L) at ambient  
10 temperatures, 30 urchins were distributed to four main aquarium tanks to  
11 represent each warming treatment as well as the ambient control treatment.  
12 Urchins were distributed at random. Replication within each of these treatments  
13 was achieved by floating five separate 6-litre tanks, each containing six urchins  
14 in each main aquarium tank (170 L). Each main aquarium tank functioned as a  
15 temperature bath (Figure S3; 30 urchins per treatment, 5 replicates per  
16 treatment where data from urchins in the same replicate floating tank were  
17 pooled). Temperature treatments were not replicated due to space restrictions.  
18 The same treatment conditions (i.e., temperature) was translated to all replicate  
19 urchins, and as such, temperature was closely monitored to note and control  
20 variability (Figure S4).

21 The water in each floating tank was aerated using air stones and refreshed by  
22 50% water change every other day. Water changes not only ensured that  
23 overall water quality was maintained, but also meant any metabolic products,  
24 especially potentially toxic nitrogenous chemical species, were maintained at  
25 very low levels. Tank water samples were periodically analysed for pH (ranging



1 7.5 - 8.0), NO<sub>2</sub> (ranging 0.05 mg l<sup>-1</sup> – 0.1 mg l<sup>-1</sup>), NO<sub>3</sub> (ranging 0.5 mg l<sup>-1</sup> - 1.0  
2 mg l<sup>-1</sup>) and NH<sub>4</sub> (stable at 0.1 mg l<sup>-1</sup>) to ensure good water quality. Throughout  
3 the experiment, concentrations of the aforementioned compounds remained  
4 within the ranges stated.

5 Urchins within each replicate tank were separated by aquaria egg crates and  
6 fine mesh partitions to ensure individuals were isolated and any faeces  
7 produced was retained within compartments (Figure S3). During warming trials  
8 experimental temperatures in the aquaria water baths were raised by 1°C,  
9 0.5°C or 0.3°C each evening, depending on treatment. Temperatures in the  
10 floating tanks increased more gradually than the water baths, allowing urchins  
11 to adjust slowly to each new temperature. Temperatures were checked every  
12 30 minutes after each temperature change to ensure required temperatures  
13 were achieved and kept constant. Initially, temperatures fluctuated by up to ±  
14 0.3°C before stabilising after 1-2 hrs. Temperatures were subsequently  
15 monitored throughout the following day and held within ± 0.1°C of the target  
16 experimental temperature (Figure S4). For ambient controls, urchins were held  
17 in the aquarium with the set-up and light conditions identical to the warming  
18 treatment conditions. Temperatures were maintained at those experienced in  
19 Ryder Bay which naturally fluctuated between 0.9 °C and 1.9°C.

### 20 2.3 Feeding trials

21 Urchins were fed pre-portioned amounts of food every 48 hrs. Previous studies  
22 fed *S. neumayeri* high protein diets, such as fish fillets, *Polachius virens*  
23 (Suckling et al. 2014, Morley et al. 2016). In the current study, urchins were fed  
24 the foot of the common Antarctic limpet, *Nacella concinna*, which has a  
25 comparable protein content to that of *P. virens* muscle. Based on feeding

1 protocols in Morley et al. (2016b), urchins were fed ~4% of their mean body  
2 mass every three weeks, but this was spread across 48 hr feeding increments  
3 in order to keep feeding activity constant and reduce the variability in daily  
4 metabolic activity.

5 Limpets were chosen as a food source since nutrient content could be  
6 controlled and pre-portioned. A more representative diet would be a varied one  
7 with algal biofilm, animal tissues and/ or detritus (McClintock 1994). However,  
8 administering a varied diet would make it difficult to assess the amount of food  
9 consumed per urchin at the same time as standardising the nutritional content.

10 There is evidence that diet, especially protein levels, can affect development  
11 and gonad growth (Liu et al. 2007, Zupo et al. 2019) as well as ingestion and  
12 assimilation rates in sea urchins (Azad et al. 2011). As such, by feeding a diet  
13 of limpets it is possible that body condition may be altered and the ability to  
14 tolerate stress may be improved as a result.

15 Feeding was initiated two days before the beginning of the experiment to start  
16 the digestion process. Each urchin was allowed to feed for 48 hrs before any  
17 remaining food was removed and refreshed. After 48 hrs, each urchin was  
18 recorded as feeding or not feeding. Infrequently, urchins may have only partially  
19 consumed the food piece, which was recorded.

## 20 2.4 Faecal collection

21 Faecal production began four days into the experiment, 6-days after feeding  
22 was initiated. The presence of faeces was recorded for all urchins every 48 hrs.  
23 To measure faecal production, faeces were collected every 48 hrs by pipette  
24 and transferred to falcon tubes from 10 urchins per treatment, where at least

1 one sample was taken from each replicate tank within the treatment. The same  
2 urchins were targeted for faecal collection to minimise subconscious  
3 preferences towards urchins producing more faeces. This was not always  
4 possible since sometimes urchins did not produce any faeces or else  $CT_{max}$   
5 was reached, and these urchins were removed. In these cases, a different  
6 urchin was chosen at random to sample from. For all other urchins, any  
7 remaining faecal matter was removed.

8 Collected faecal matter was centrifuged and the supernatant seawater  
9 decanted. Faeces were then rinsed with RO (Reverse Osmosis purified) water  
10 by agitating and centrifuging to remove any seawater salt. Washed faeces were  
11 pipetted into pre-ashed and pre-weighed foil boats and dried at 60°C for 24 hrs.  
12 Dry foil boats and faeces were placed in a desiccator to cool and then weighed  
13 ( $\pm 1$  mg). Dry faeces were subsequently ignited in a muffle furnace at 475°C for  
14 6 hrs. Foil boats and ashed faeces were cooled in a desiccator and weighed ( $\pm$   
15 1 mg). Dry mass (DM) and Ash-Free Dry Mass (AFDM) (i.e., organic content)  
16 were obtained by subtraction.

## 17 2.5 Respirometry

18 Oxygen consumption was recorded for 10 urchins per treatment, sampling two  
19 individuals from each replicate tank within each treatment. Oxygen  
20 consumption was recorded for the same urchins for every 2°C rise in  
21 temperature from ambient in each treatment. Methods for measuring oxygen  
22 consumption followed those described by Suckling et al., (2015), using 200 -  
23 250 ml volume chambers. For each urchin, live wet mass ( $\pm 0.01$  g) was  
24 recorded where  $O_2$  consumption was measured. AFDM was determined from  
25 live wet mass vs AFDM regressions determined from a subsample of urchins

1 (n = 40) collected from the same site. To obtain the ash mass of urchins,  
2 individuals were weighed live before freezing in liquid nitrogen and storing at –  
3 40°C. Frozen urchins were then placed in pre-ashed and pre-weighed ceramic  
4 crucibles and dried at 60°C until constant mass was obtained ( $\pm 0.01$  g). Once  
5 dried, urchins were ignited in a muffle furnace at 475°C for 6 hrs and  
6 subsequently weighed to obtain ash mass after cooling in a desiccator ( $\pm 1$  mg).

## 7 2.6 Righting

8 The time taken for urchins to right themselves was recorded for 10 urchins per  
9 treatment, sampling two urchins from each replicate tank within each treatment.  
10 The time taken to right was recorded for the same urchins every 2°C rise in  
11 temperature from ambient in each treatment. Ten individuals were removed  
12 from their experimental tanks and placed in individual containers. These  
13 containers were previously filled and floated in water already at the  
14 experimental target temperature. Urchins were immediately inverted following  
15 transfer from experimental tanks to the floating containers and timed until the  
16 individual was fully upright. Urchins could not reach the sides of containers to  
17 aid in righting. Once righted, urchins were returned to their experimental tanks.

## 18 2.7 Critical temperature limits ( $CT_{max}$ )

19 The critical thermal limit ( $CT_{max}$ ) was recorded for all experimental urchins in  
20 the warming treatments, where the limit was defined as the point at which the  
21 individual was unable to right itself within 12 hrs, had stopped eating and  
22 producing faeces. When an urchin began to show signs of reaching the  $CT_{max}$   
23 (not feeding or producing faeces), they were inverted in the tank and left for 12

1 hrs. If the urchin had not righted itself after this period, they were removed and  
2 weighed suspended in water to obtain live wet volumes ( $\pm 0.01$  mL).

### 3 2.8 Statistical Analysis

4 Where multiple urchins were sampled within the same floating tank,  
5 measurements of feeding, faecal production, righting, and oxygen consumption  
6 were pooled so that  $n = 5$ , and the standard errors were calculated from these  
7 five replicate tanks.

8 To determine differences in functional responses between treatments, a one-  
9 way repeat measures analysis of variance (ANOVA) was carried out in R (v.  
10 4.0.5). This analysis was considered appropriate for this experiment due to the  
11 related and non-independent groups at each temperature timepoint. For this  
12 analysis, treatment group variances were compared when treatments reached  
13 the same temperature increments. For ambient controls, temperature  
14 timepoints were aligned with measurements taken at similar dates to treatment  
15 sampling. Variances were compared between groups and within timepoints for  
16 righting and oxygen consumption rates and the resultant p-value was adjusted  
17 using the Bonferroni correction method. Significant differences ( $p < 0.05$ ) were  
18 followed up with a paired t-test and again, p-values were adjusted using the  
19 Bonferroni correction method. Data were initially log transformed to ensure  
20 assumptions of normal distribution were met.

21 Segmented linear regression models were fitted in the R package 'segmented'  
22 (Muggeo 2008) to identify breakpoints in the linear relationships between  
23 functional process and temperature. Breakpoints were identified where the  
24 gradient of the relationship changed (McWhorter et al. 2018). The change in

1 gradient was used to define the functional threshold of the process measured.  
2 It was especially important to use a method such as segmented regression to  
3 identify breakpoints in process rates. Segmented regressions were used to  
4 model these relationships not necessarily for the purpose of fitting the simplest  
5 model, but rather to identify any change in the regressions gradient which then  
6 indicated that the functions response to temperature increase had changed. In  
7 some cases, a linear regression would be sufficient to explain the relationship,  
8 however a linear model could mask the subtle change in the rate of degradation  
9 experienced when a species hits a thermal threshold. Alternatives would be to  
10 fit curves and identify changes in slope (e.g. Pörtner et al. 2006), but curves  
11 were not appropriate here. A Davies test was also conducted to determine  
12 significant ( $p < 0.05$ ) differences in the gradients of the segmented slopes.

13 Size effects on functional response were explored through scatter plots. Where  
14 relationships were observed, the effect of size (as test diameter) and  
15 temperature on the functional response, was assessed with a linear mixed  
16 effects model using the package 'lme4' and the function 'lmer' in R (v. 4.0.5).  
17 Test diameter and temperature were added as interacting fixed terms and  
18 replicate tank ID was added as a random effect. Prior to any modelling, function  
19 responses were transformed to achieve normality in the distribution.

## 20 **3. RESULTS**

### 21 3.1 Feeding and faecal egestion

22 On average,  $80\% \pm 19\%$  of animals fed in ambient conditions for the duration  
23 of the experiment. For the first four days of the experiment, in treatments where  
24  $T \uparrow 1^\circ\text{C day}^{-1}$ , the proportion of animals feeding exceeded all other treatments  
25 ( $97\% \pm 4\%$ ), including ambient conditions ( $87\% \pm 10\%$ ). Fifty percent of animals

1 stopped feeding in treatments when temperatures exceeded 7.2°C, 8.2°C, and  
2 9.2°C, where T↑ by 1°C, 0.5°C and 0.3°C day<sup>-1</sup>, respectively (Figure 1).

3 A breakpoint (where the slope of the regression changed) for the % individuals  
4 feeding was identified at 4.0°C and 6.2°C in treatments where T↑ 1°C day<sup>-1</sup>  
5 and 0.5°C day<sup>-1</sup>, respectively (Table 1). However, changes in the segmented  
6 slope gradients were not significantly different from linear regressions for these  
7 two treatments (Davies p-value = 0.329 and 0.301, respectively). A breakpoint  
8 for the % feeding in T↑ 0.3°C day<sup>-1</sup> was identified at 8.2°C (Table 1), from which  
9 point the % individuals feeding declined rapidly and the relationship between  
10 temperature and the proportion of individuals feeding became significant (p  
11 <0.001). The mean temperature breakpoint for the function of % feeding was  
12 6.1°C ± 1.2°C, averaged across all treatments.

13 The percentage of animals producing faeces tracked the proportion of animals  
14 feeding after the first four days (Figure 1). Following each breakpoint, the  
15 relationship between temperature and % individuals producing faeces became  
16 significant (Table 1). For the fastest rate of warming where T↑ 1°C day<sup>-1</sup>, a  
17 breakpoint was identified at 5.2°C, above which the % individuals producing  
18 faeces rapidly declined from 100% to 10.3% within 6 days. Where T↑ 0.3°C  
19 day<sup>-1</sup> and 0.5°C day<sup>-1</sup>, the regression breakpoint for faecal production was  
20 8.3°C and 4.5°C respectively (Table 1). The mean temperature breakpoint for  
21 the function of % producing faeces was 6.0°C ± 2.0°C, averaged across all  
22 treatments.

23 The mean mass of faeces produced in treatments where T↑ 0.3°C day<sup>-1</sup>, was  
24 significantly greater than the faecal mass produced in ambient control

1 conditions and treatments where  $T \uparrow 1^\circ \text{C day}^{-1}$ , until temperatures exceeded  
2  $2.1^\circ \text{C}$  ( $t_{(4)} = 8.74$ ,  $p = 0.006$  and  $t_{(4)} = 5.02$ ,  $p = 0.044$ , respectively). Where  $T \uparrow$   
3  $0.5^\circ \text{C day}^{-1}$ , the mass of faeces produced was significantly greater than  
4 treatments where  $T \uparrow 1^\circ \text{C day}^{-1}$ , until temperatures exceeded  $2.1^\circ \text{C}$  ( $t_{(4)} = 5.31$ ,  
5  $p = 0.036$ ). Despite this observation, no additional food was consumed in these  
6 treatments. There was no significant difference between the treatments or  
7 control as temperatures increased beyond  $2.1^\circ \text{C}$ .

8 Breakpoints in regressions were identified at  $5.0^\circ \text{C}$  and  $3.1^\circ \text{C}$  for treatments  
9 where  $T \uparrow 0.5^\circ \text{C day}^{-1}$  and  $0.3^\circ \text{C day}^{-1}$ , respectively (Table 1). The breakpoints  
10 for these regressions marked a reduction in the gradient of the 2<sup>nd</sup> slope,  
11 whereby faeces produced  $\text{day}^{-1} \text{ mgAFDM}^{-1}$  as a function of temperature  
12 decreased at a slower rate as temperatures increased. The mean temperature  
13 breakpoint for faeces produced was  $4.1^\circ \text{C} \pm 0.95^\circ \text{C}$ , averaged across the  
14 slowest ( $T \uparrow 0.3^\circ \text{C day}^{-1}$ ) and intermediate ( $T \uparrow 0.5^\circ \text{C day}^{-1}$ ) rates of warming.

### 15 3.2 Righting

16 In treatments where  $T \uparrow 1.0^\circ \text{C day}^{-1}$ , time taken to right became significantly  
17 longer than ambient controls when temperatures reached  $9.2^\circ \text{C}$  ( $t_{(4)} = 6.06$ ,  $p <$   
18  $0.022$ ). For treatments where  $T \uparrow 0.3^\circ \text{C day}^{-1}$ , time taken to right only became  
19 significantly longer than ambient controls just before  $\text{CT}_{\text{max}}$  was reached, when  
20 temperatures reached  $11.2^\circ \text{C}$  ( $t_{(4)} = 6.04$ ,  $p < 0.023$ ). For treatments where  $T \uparrow$   
21  $0.5^\circ \text{C day}^{-1}$ , time taken to right never exceeded ambient controls significantly,  
22 however mean righting times were consistently higher than control conditions  
23 throughout the warming period.



1 A breakpoint in the linear regression was identified at 8.7°C in treatments where  
2 temperature was raised at 0.3°C day<sup>-1</sup> (Table 1). The relationship between  
3 temperature and the time taken to right became significant above this  
4 breakpoint temperature ( $p < 0.001$ ). For the other treatments righting time  
5 increased linearly without a breakpoint in the regression.

6 The interactive effect of urchin size and temperature on the time taken to right  
7 was significant ( $t_{(204)} = 2.11$ ,  $p = 0.034$ ), where larger urchins took longer to right  
8 at higher temperatures (Figure S5, Table S3).

### 9 3.3 Oxygen consumption

10 Oxygen consumption rates were significantly higher in heatwave treatments  
11 compared to ambient controls when temperatures reached 7.2°C for all  
12 treatments. However, oxygen consumption rates were significantly higher than  
13 ambient controls from lower temperatures of 3.2°C in treatments where  $T \uparrow$   
14 0.3°C day<sup>-1</sup> ( $t_{(4)} = 5.62$ ,  $p = 0.030$ ) and 5.2°C in treatments where  $T \uparrow$  1.0°C  
15 day<sup>-1</sup> ( $t_{(4)} = 4.98$ ,  $p = 0.045$ ). Overall, there was a positive linear trend between  
16 oxygen consumption and temperature for all treatments. However, where  $T \uparrow$   
17 1°C day<sup>-1</sup>, a drop in O<sub>2</sub> consumption occurred at 9.2°C, and where  $T \uparrow$  0.3°C  
18 day<sup>-1</sup>, a drop occurred just before the CT<sub>max</sub> at 11.2°C.

19 O<sub>2</sub> consumption increased at a faster rate per increase in temperature where  
20 warming rates were fastest at 1°C day<sup>-1</sup> (slope gradient = 1.50) and increased  
21 at the slowest rate when warming rates were slowest at 0.3°C day<sup>-1</sup> (slope  
22 gradient = 0.96) (Table 1). No breakpoint was identified in any treatment.

### 23 3.4 CT<sub>max</sub>

1 The  $CT_{max}$  for urchins in treatments where  $T \uparrow 0.3^{\circ}\text{C day}^{-1}$ ,  $T \uparrow 0.5^{\circ}\text{C day}^{-1}$  and  
2  $T \uparrow 1^{\circ}\text{C day}^{-1}$  ranged from  $10.6^{\circ}\text{C} - 13.8^{\circ}\text{C}$ ,  $11.2^{\circ}\text{C} - 13.7^{\circ}\text{C}$ , and  $12.2^{\circ}\text{C} -$   
3  $14.2^{\circ}\text{C}$ , respectively. The effect of warming rate on the  $CT_{max}$  was significant  
4 ( $F_{(2, 12)} = 7.29$ ,  $p = 0.008$ ), with post-hoc analysis identifying that for treatments  
5 where temperature increased at the fastest rate ( $T \uparrow 1^{\circ}\text{C day}^{-1}$ ), the  $CT_{max}$  was  
6 significantly higher compared to treatments where temperature increased at a  
7 slower rate ( $T \uparrow 0.3^{\circ}\text{C day}^{-1}$ ) ( $t_{(8)} = -6.02$ ,  $p = 0.001$ ).

8 Across all functions where breakpoints were identified, the slowest rate of  
9 warming ( $T \uparrow 0.3^{\circ}\text{C day}^{-1}$ ) had a mean temperature breakpoint of  $8.3^{\circ}\text{C} \pm 1.3^{\circ}\text{C}$ .  
10 In comparison, the mean temperature breakpoint was  $5.4^{\circ}\text{C} \pm 0.5^{\circ}\text{C}$ , and  $4.6^{\circ}\text{C}$   
11  $\pm 0.6^{\circ}\text{C}$  for intermediate ( $T \uparrow 0.5^{\circ}\text{C day}^{-1}$ ) and fast ( $T \uparrow 1^{\circ}\text{C day}^{-1}$ ) warming  
12 rates, respectively.

#### 13 **4. DISCUSSION**

14 MHWs are predicted to increase in frequency, intensity, and duration in the  
15 coming decades. Deterioration of basic animal functioning, critical for long-term  
16 survival, will likely be a more frequent consequence of the short-term warming  
17 (i.e., weeks-months) caused by MHWs, rather than mortality. However, little is  
18 known about functional impacts, especially thresholds and how these limits  
19 deteriorate with respect to  $CT_{max}$ . By understanding how key biological  
20 functions are affected by short term temperature elevations and different  
21 warming rates, we can better understand how extreme climate events, typified  
22 by short-term warming, may impact individuals and populations, and hence  
23 communities.

1 In this study, we investigated the effect of warming rates typical of those  
2 expected during Antarctic MHW events on the functioning of the Antarctic sea  
3 urchin, *S. neumayeri*. Functional thresholds were identified using segmented  
4 regressions, where a breakpoint indicated a gradient change in the response  
5 trend with temperature. The identification of regression breakpoints, or slope  
6 changes has been used previously to define ecological thresholds, and is  
7 considered a more flexible and realistic approach when interpreting complex,  
8 often non-linear, ecological relationships (Piepho & Ogutu 2003, Ferrarini 2011,  
9 Morley et al. 2014).

10 Several studies have shown that faster warming rates result in higher  $CT_{max}$  in  
11 terrestrial (e.g. Terblanche et al. 2007, Allen et al. 2016) and marine (Peck et  
12 al. 2009) species. These observations, along with the  $CT_{max}$  data in this study,  
13 follow the failure rate model proposed by Kingsolver & Umbanhowar (2018),  
14 who showed that critical limits are reached at lower temperatures when  
15 warming accumulates over extended periods. However, our results for  
16 functional thermal limits follow the opposite trend to the  $CT_{max}$ , where functions  
17 are impacted negatively at lower temperatures when warming is rapid. Overall,  
18 in this study higher functional thresholds were reached when temperatures  
19 were raised slowly (thresholds averaging  $8.3^{\circ}C \pm 1.3^{\circ}C$ ). At the faster warming  
20 rates functional thresholds were lower ( $5.4^{\circ}C \pm 0.5^{\circ}C$  or  $4.6^{\circ}C \pm 0.6^{\circ}C$ ). There  
21 was even evidence that some functions declined linearly, with significant  
22 deterioration from temperatures  $+2.8^{\circ}C$  above ambient when warmed at the  
23 fastest rate. Thus, short-term exposure to more extreme temperatures has  
24 more impact on functioning than longer, chronic exposure to more slowly  
25 elevated temperatures.

1 Although metabolic acclimation is unlikely over such short time periods  
2 (apparent from the oxygen consumption data here, and also previous research  
3 on long-term acclimation of *S. neumayeri* (Peck et al. 2014, Suckling et al.  
4 2015)), short-term acclimation for some functions might be possible after an  
5 initial shock response when temperatures are increased slowly. In our study,  
6 the shock response did not appear to subside at faster rates of warming, and  
7 instead mean functional thresholds were lower as warming rate increased.  
8 These results suggest that functional and lethal limits are likely driven and  
9 determined by different mechanisms. Previous studies have shown that lethal  
10 limits are likely set by one or both of physiological processes or cellular and  
11 biochemical mechanisms. At very rapid rates of warming, such as  $1^{\circ}\text{C h}^{-1}$  or  
12  $1^{\circ}\text{C day}^{-1}$ , physiological mechanisms such as nervous and circulatory failure  
13 appear to be the limiting factors (Young et al. 2006, Pörtner et al. 2007, Bilyk &  
14 DeVries 2011). At slower rates of warming ( $1^{\circ}\text{C 3 days}^{-1}$  to  $1^{\circ}\text{C month}^{-1}$ ) cellular  
15 and biochemical mechanisms such as accumulation of toxic products, e.g.  
16 protein carbonyls, enzyme tolerances or insufficiency of chaperone protein  
17 capacity appear to be limiting (Peck et al. 2009, Clark et al. 2017, 2018).  
18 Recently the factors setting thermal limits and responses to warming have been  
19 shown to be highly species specific (Clark et al. 2021, Collins et al. 2021).

20 Our results also indicate that thermal sensitivity varies among key biological  
21 functions. For example, the function of righting in urchins was similar between  
22 treatments and ambient control conditions until temperatures reached  $9.2^{\circ}\text{C}$  for  
23 the fastest rates of warming, and the highest breakpoint of  $8.7^{\circ}\text{C}$  was identified  
24 in the slowest rates of warming. However, lower thresholds were identified for  
25 the other functions related to digestion such as % feeding or producing faeces.

1 Variation between functional thresholds could be related to function complexity,  
2 where a function involving multiple processes would be more likely to fail  
3 (Pörtner et al. 2007, Stevens et al. 2010, Peck 2011). Another explanation could  
4 be related to the extent to which functions limit survival and fitness, where an  
5 organism's energy reserves allow for short periods of negative energy balance.  
6 In Antarctic marine species such periods of negative energy balance can be  
7 very long, extending to months or even years of low food supply or starvation,  
8 because of the extreme environmental seasonality and the very low metabolic  
9 energy use characteristic of this fauna (Brockington et al. 2001, Harper & Peck  
10 2003, Obermüller et al. 2010). However, being able to right provides immediate  
11 protection from predation, equivalent to mechanisms such as the ability to stay  
12 attached to the substratum in limpets (Morley et al. 2012b) or reburying in  
13 infaunal clams when disturbed and removed from the sediment by, for example,  
14 iceberg scour (Peck et al. 2004). Finally, where a function has a higher  
15 metabolic energy demand, it is more likely to be limited by food availability and  
16 energy delivery capacity (van der Meer 2006, Morley et al. 2012a, Peck 2018).

17 The breakpoints identified for the mass of faeces produced might not indicate  
18 a functional threshold. Instead, the initial high faecal production in the slowest  
19 and intermediate warming rates is likely a result of the initial increase in  
20 temperature causing food to move faster through the urchin, as also seen in  
21 the Antarctic plunderfish *Harpagifer antarcticus* (Boyce et al. 2000). This  
22 elevation in faecal production was only observed when temperatures increased  
23 initially, after which faecal production reduced to rates similar to ambient control  
24 conditions. This effect was not observed in treatments with the fastest rates of  
25 warming since these slight increases in temperature of 1°C – 2°C were likely

1 not maintained long enough for gut passage rate to increase. Therefore, our  
2 results indicate that the breakpoints for faecal production may not have any  
3 direct implications on functionality and instead give evidence for the relationship  
4 between temperature and gut evacuation rate (GER).

5 In thermally stressed environments, animals usually increase their oxygen  
6 uptake in order to meet increasing demands of functional processes (Gillooly  
7 et al. 2001). However, when oxygen uptake is increased, yet functioning  
8 deteriorates, it is hypothesised that this indicates a threshold where uptake,  
9 transport, and delivery of oxygen can no longer meet the animal's functional  
10 demands. This theory has been termed the oxygen and capacity limited thermal  
11 tolerance hypothesis (OCLTT) (Pörtner et al. 2017). This theory focuses on the  
12 limitations set by the animal's physiology. However, as temperature increases  
13 the concentration of oxygen diminishes, further reducing the availability of  
14 oxygen to the animal and potentially amplifying the effects of OCLTT. Reducing  
15 the concentration of oxygen in the water can limit functioning (Peck et al. 2007,  
16 Pörtner et al. 2007) and as such, the functional thresholds identified in this study  
17 may not only indicate thermal limits but may also be influenced by the reduced  
18 oxygen content as temperatures increased. If oxygen concentration was  
19 controlled and elevated throughout warming, the functional thresholds identified  
20 would likely be higher (Pörtner et al. 2006). However, warmer oceans will be  
21 accompanied by lower oxygen concentrations (Oschlies et al. 2018, Spicer et  
22 al. 2019) and as such the functional thresholds determined in this study will be  
23 more representative of a natural system than if oxygen were controlled.

24 Food availability and quality can also be a significant factor in determining  
25 functional scope (Welch et al. 1998, Lemoine & Burkepille 2012, Cheng et al.

1 2018), whereby the nutritional status and condition of the animal could affect  
2 energy delivery capacity similarly to OCLTT. For example, feeding and  
3 digestive capacity limited the thermal tolerance of juvenile spiny lobsters,  
4 *Sagmariasus verreauxi* (Fitzgibbon et al. 2017) and digestive capacity and food  
5 intake of individuals at high temperatures related to depressed mitochondrial  
6 respiratory capacity in brown trout *Salmo trutta* (Salin et al. 2016). The capacity  
7 to assimilate energy would also play a role in determining energy delivery to  
8 tissues and is determined by physiological processes including consumption  
9 rate, absorption of food and GER (Boyce et al. 2000, Angilletta 2001). Hence,  
10 assimilation itself is energetically demanding and may limit functional thermal  
11 thresholds (Sandersfeld et al. 2015, Salin et al. 2016).

12 Thus, OCLTT may be a possible mechanism for determining functional limits  
13 observed in our experiments. However, there is no empirical support in our data  
14 for this theory. In both experiments and in natural MHWs, other factors are likely  
15 to be important, and obtaining sufficient energy from food may be important for  
16 successful functioning. Impacts on animal condition from warming may be  
17 especially important in highly seasonal polar environments where warming in  
18 winter, when food supplies are scarce, would increase energy use with little or  
19 no opportunity to mitigate the cost (Peck 2018). Species such as *S. neumayeri*  
20 that have been shown to spend periods in winter up to seven months without  
21 feeding (Brockington 2001) may be particularly vulnerable to such impacts.

22 Our experiment included a period of six weeks without feeding to allow  
23 metabolic activity to stabilise and be comparable between individuals.  
24 However, a caveat to this initial standardisation of condition could influence the  
25 urchin's physiological response to the warming in treatments. Nutritional status

1 has been shown to affect the reproductive state of *S. neumayeri*, with a  
2 reduction in gonad index and maturation of gametes following six weeks without  
3 food, comparative to animals foraging naturally in the environment (De Leij  
4 2021). Functional capacity has also been affected in other invertebrates under  
5 low food coupled with environmental stress, for example the blue mussel  
6 *Mytilus edulis* had a reduced ability to repair shells when high CO<sub>2</sub> was coupled  
7 with low food (Melzner et al. 2011) and the green sea urchin *Strongylo-*  
8 *centrotus droebachiensis*, exhibited severe metabolic acidosis when exposed  
9 to elevated CO<sub>2</sub> with empty digestive tracts (Stumpp et al. 2012). Hence, we  
10 might consider that the elevated temperatures coupled with the suboptimal  
11 nutritional status at the start of the experiment, may have impacted the thermal  
12 limits of certain functions. This would likely have resulted from a mismatch  
13 between a limited energy supply and stores, and an increased energy demand  
14 of the animal. However, the data in this study shows a reduction in the number  
15 of urchins feeding as temperatures increase, suggesting that food was not the  
16 limiting factor when this species approached its functional thermal limits.

17 From our analysis of the RaTS environmental data, previous MHW events  
18 reached maximum temperatures of  $2.3^{\circ}\text{C} \pm 0.36^{\circ}\text{C}$ , with onset rates of  $0.3^{\circ}\text{C}$   
19  $\text{day}^{-1}$ . Days at heatwave status have extended up to 95 days, and cumulative  
20 intensities (a combination of temperature intensity and heatwave duration) have  
21 reached maxima of  $54^{\circ}\text{C} \times \text{day}$  (Figure S2). Mean climate temperatures are  
22 predicted to shift by  $+2^{\circ}\text{C}$  by 2100, and with that, climate extremes such as  
23 MHWs will increase in magnitude relative to this (IPCC 2014, 2019). Our results  
24 suggest that functions such as feeding and faecal egestion are likely to be  
25 affected by MHW events occurring in 2100, if not before, and this will include



1 increased metabolic demands with consequent impacts on annual energy  
2 budgets.

3 For a long-lived (>40 year (Brey et al. 1995)) and slow to mature (8-9 years  
4 (Peck 2018)) species such as *S. neumayeri*, there will be less scope for  
5 phenotypic and genotypic adaptations to a warming climate as might be  
6 possible for short-lived and rapidly maturing species (Peck 2011, Donelson et  
7 al. 2012, Salinas & Munch 2012). However, there may still be opportunity for *S.*  
8 *neumayeri* to adapt to a warmer world. Within 80 years (2020 - 2100), eight  
9 generations of *S. neumayeri* will have succeeded the present population, and  
10 in the year 2100, the 5<sup>th</sup>, 6<sup>th</sup> and 7<sup>th</sup> generation could be present and  
11 reproducing in populations around Antarctica. If we consider the evidence of *S.*  
12 *neumayeri*'s capacity to acclimate, it may be possible for this species to  
13 acclimate and adapt successfully to function in a +2°C warmer world (Morley et  
14 al. 2016). It is still uncertain, however, how this species will respond to acute  
15 warming, like that experienced during MHWs, in this warmer climate. The data  
16 in this study cannot predict the implications of acclimation and adaptation on  
17 the subsequent tolerance to MHWs for *S. neumayeri*. Instead, the data provides  
18 insight into the effect of onset rate of acute warming, the thermal vulnerability  
19 of key biological functions, and the difference between critical thermal limits and  
20 functional thermal limits. Thus, according to our data we could see reduced  
21 energy availability for *S. neumayeri* from changes in feeding and food  
22 processing rates during MHWs in warmer oceans, which would very likely  
23 reduce survival in marginal environments.

1 Following the results from this study, it would be important to explore recovery  
2 following MHW events. Our data indicate reduced functioning as temperatures  
3 are raised across all rates of warming. However, the ability and rate of *S.*  
4 *neumayeri* to resume 'normal' functioning if returned to ambient temperatures  
5 is uncertain. It has been shown that the marine snail, *Littorina littorea*, loses  
6 motility under thermal stress, however if temperatures are lowered again, this  
7 function returns (Hamby 1975). To resume a single function may not indicate  
8 full recovery, and our study shows that different biological functions have  
9 varying thermal tolerances. As such, performance of all functions, including  
10 metabolic activity, would need to return to baseline levels for an animal to  
11 recover completely (Walter et al. 2013). Developing our understanding of  
12 recovery following acute warming and even the effects of repeat MHW events,  
13 could better predict the long-term implications of MHWs for this species.

14 It is important to note that the functional and critical limits measured in this study  
15 are likely an example of a 'best case scenario'. Experiments such as these can  
16 only predict the isolated effects of one variable. However, the additional  
17 energetic costs associated with physical factors such as salinity change and  
18 biological factors including varying food quality and quantity, species  
19 interactions, diseases and scavenging for food, need to be included before we  
20 can obtain dependable predictions for 'real world' scenarios that give  
21 information relevant to the variable conditions experienced across a species  
22 distribution range. What is limiting at the range margins for a species will differ  
23 from core areas (Kolzenburg et al. 2021).

24 Our data highlight that the deterioration of functioning when temperatures are  
25 raised, especially during MHWs, has implications for long term survival, and

1 physiological functions. Therefore, functioning should be considered when  
2 determining organism thermal limits, rather than traditional critical thermal  
3 limits. Our findings show that fitness cannot be determined from a single  
4 function and instead functions vary in thermal sensitivity. A whole organism  
5 approach to functional fitness is therefore necessary, considering functional  
6 complexity, importance, and energetic demand. Our results suggest that  
7 contrary to the relationship between critical thermal limits and onset rate,  
8 functional degradation occurs at lower temperatures when exposed to rapid  
9 warming ( $1^{\circ}\text{C day}^{-1}$ ). Therefore, when investigating the impact of MHWs on  
10 organisms and populations, it is important to consider the key features of the  
11 heatwave event, including the onset rate, exposure duration, and how these  
12 characteristics act together to determine functional thresholds.

### 13 **ACKNOWLEDGMENTS**

14 This work was supported by the Natural Environmental Research Council [grant  
15 number NE/L002531/1]. We would like to acknowledge the Collaborative  
16 Antarctic Science Scheme (CASS) for providing the funds to support the travel  
17 and carry out this work at Rothera Research Station. We would like to thank  
18 the British Antarctic Survey Rothera Marine team for the support provided  
19 during the experimental period of this work, in particular, the laboratory  
20 manager, Aurelia Reichardt for her technical guidance and assistance in the  
21 aquarium, and Dr. Simon Morley for his expertise and assistance during the  
22 Antarctic season.

### 23 **AUTHOR CONTRIBUTIONS**

1 L.S.P and R.D conceived and designed the study. R.D carried out the  
2 practical work and data processing. R.D, L.J.G and L.S.P analysed the data,  
3 drafted the manuscript and approved its publication.

#### 4 **COMPETING INTERESTS**

5 The authors declare no competing interests.

#### 6 **LITERATURE CITED**

- 7 Allen JL, Chown SL, Janion-Scheepers C, Clusella-Trullas S (2016)  
8 Interactions between rates of temperature change and acclimation affect  
9 latitudinal patterns of warming tolerance. *Conserv Physiol* 4:1–14.
- 10 Ardor Bellucci LM, Smith NF (2019) Crawling and righting behavior of the  
11 subtropical sea star *Echinaster (Othilia) graminicola*: effects of elevated  
12 temperature. *Mar Biol* 166:1–9.
- 13 Azad AK, Pearce CM, McKinley RS (2011) Effects of diet and temperature on  
14 ingestion, absorption, assimilation, gonad yield, and gonad quality of the  
15 purple sea urchin (*Strongylocentrotus purpuratus*). *Aquaculture* 317:187–  
16 196.
- 17 Barnes DKA, Brockington S (2003) Zoobenthic biodiversity, biomass and  
18 abundance at Adelaide Island, Antarctica. *Mar Ecol Prog Ser* 249:145–  
19 155.
- 20 Bilyk KT, DeVries AL (2011) Heat tolerance and its plasticity in Antarctic  
21 fishes. *Comp Biochem Physiol - A Mol Integr Physiol* 158:382–390.
- 22 Bopp L, Resplandy L, Orr JC, Doney SC, Dunne JP, Gehlen M, Halloran P,  
23 Heinze C, Ilyina T, Séférian R, Tjiputra J, Vichi M (2013) Multiple  
24 stressors of ocean ecosystems in the 21st century: Projections with  
25 CMIP5 models. *Biogeosciences* 10:6225–6245.
- 26 Bosch I, Beauchamp KA, Steele ME, Pearse JS (1987) Development,  
27 metamorphosis, and seasonal abundance of embryos and larvae of the  
28 Antarctic sea urchin *Sterechinus neumayeri*. *Biol Bull* 173:126–135.

- 1 Boyce SJ, Murray AWA, Peck LS (2000) Digestion rate, gut passage time and  
2 absorption efficiency in the Antarctic spiny plunderfish. *J Fish Biol*  
3 57:908–929.
- 4 Brey T, Pearse J, Basch L, McClintock J, Slattery M (1995) Growth and  
5 production of *Sterechinus neumayeri* (Echinoidea: Echinodermata) in  
6 McMurdo Sound, Antarctica. *Mar Biol* 124:279–292.
- 7 Brockington S (2001) The seasonal ecology and physiology of *Sterechinus*  
8 *neumayeri* (Echinodermata; Echinoidea) at Adelaide Island, Antarctica.  
9 PhD thesis The Open University.
- 10 Brockington S, Clarke A, Chapman ALG (2001) Seasonality of feeding and  
11 nutritional status during the austral winter in the Antarctic sea urchin  
12 *Sterechinus neumayeri*. *Mar Biol* 139:127–138.
- 13 Chappelle G, Peck LS, Clarke A (1994) Effects of feeding and starvation on the  
14 metabolic rate of the necrophagous Antarctic amphipod *Waldeckia obesa*  
15 (Chevreux, 1905). *J Exp Mar Bio Ecol* 183:63–76.
- 16 Clark MS, Peck LS (2009) Triggers of the HSP70 stress response:  
17 environmental responses and laboratory manipulation in an Antarctic  
18 marine invertebrate (*Nacella concinna*). *Cell Stress Chaperones* 14:649–  
19 660.
- 20 Clark MS, Peck LS, Thyrring J (2021) Resilience in Greenland intertidal  
21 *Mytilus*: The hidden stress defense. *Sci Total Environ* 767:144366.
- 22 Clark MS, Sommer U, Sihra JK, Thorne MAS, Morley SA, King M, Viant MR,  
23 Peck LS (2017) Biodiversity in marine invertebrate responses to acute  
24 warming revealed by a comparative multi-omics approach. *Glob Chang*  
25 *Biol* 23:318–330.
- 26 Clark MS, Suckling CC, Cavallo A, Mackenzie CL, Thorne MAS, Davies AJ,  
27 Peck LS (2019a) Molecular mechanisms underpinning transgenerational  
28 plasticity in the green sea urchin *Psammechinus miliaris*. *Sci Rep* 9:1–12.
- 29 Clark MS, Thorne MAS, King M, Hipperson H, Hoffman JI, Peck LS (2018)  
30 Life in the intertidal: Cellular responses, methylation and epigenetics.  
31 *Funct Ecol* 32:1982–1994.

- 1 Clark MS, Villota Nieva L, Hoffman JI, Davies AJ, Trivedi UH, Turner F,  
2 Ashton G V, Peck LS (2019b) Lack of long-term acclimation in Antarctic  
3 encrusting species suggests vulnerability to warming. *Nat Commun* 10:1–  
4 10.
- 5 Clarke A, Meredith MP, Wallace MI, Brandon MA, Thomas DN (2008)  
6 Seasonal and interannual variability in temperature, chlorophyll and  
7 macronutrients in northern Marguerite Bay, Antarctica. *Deep Res Part II*  
8 *Top Stud Oceanogr* 55:1988–2006.
- 9 Collins M, Peck LS, Clark MS (2021) Large within, and between, species  
10 differences in marine cellular responses: Unpredictability in a changing  
11 environment. *Sci Total Environ* 794:148594.
- 12 De Leij R (2021) Functional response of the Antarctic sea urchin, *Sterechinus*  
13 *neumayeri*, to environmental change and extreme events in the context of  
14 a warming climate (In Press). PhD Thesis, University of Southampton
- 15 De Leij R, Peck LS, Grange LJ (2021) Multiyear trend in reproduction  
16 underpins interannual variation in gametogenic development of an  
17 Antarctic urchin. *Sci Rep* 11:1–13.
- 18 Deschaseaux ESM, Taylor AM, Maher WA, Davis AR (2010) Cellular  
19 responses of encapsulated gastropod embryos to multiple stressors  
20 associated with climate change. *J Exp Mar Bio Ecol* 383:130–136.
- 21 Donelson JM, Munday PL, McCormick MI, Pitcher CR (2012) Rapid  
22 transgenerational acclimation of a tropical reef fish to climate change. *Nat*  
23 *Clim Chang* 2:30–32.
- 24 Dunlop KM, Barnes DKA, Bailey DM (2014) Variation of scavenger richness  
25 and abundance between sites of high and low iceberg scour frequency in  
26 Ryder Bay, West Antarctic Peninsula. *Polar Biol* 37:1741–1754.
- 27 Ferrarini A (2011) Detecting ecological breakpoints: a new tool for piecewise  
28 regression. *Comput Ecol Softw* 1:121–124.
- 29 Fitzgibbon QP, Simon CJ, Smith GG, Carter CG, Battaglione SC (2017)  
30 Temperature dependent growth , feeding , nutritional condition and  
31 aerobic metabolism of juvenile spiny lobster, *Sagmariasus verreauxi*.

1           Comp Biochem Physiol Part A 207:13–20.

2   Garrabou J, Coma R, Bensoussan N, Bally M, Chevaldonné P, Cigliano M,  
3           Diaz D, Harmelin JG, Gambi MC, Kersting DK, Ledoux JB, Lejeusne C,  
4           Linares C, Marschal C, Pérez T, Ribes M, Romano JC, Serrano E,  
5           Teixido N, Torrents O, Zabala M, Zuberer F, Cerrano C (2009) Mass  
6           mortality in Northwestern Mediterranean rocky benthic communities:  
7           Effects of the 2003 heat wave. *Glob Chang Biol* 15:1090–1103.

8   Gillooly JF, Brown JH, West GB, Savage VM, Charnov EL (2001) Effects of  
9           size and temperature on metabolic rate. *Science* (80- ) 293:2248–2251.

10   Hamby RJAY (1975) Heat Effects on a Marine Snail. *Biol Bull* 149:331–347.

11   Harper EM, Peck LS (2003) Predatory behaviour and metabolic costs in the  
12           Antarctic muricid gastropod *Trophon longstaffi*. *Polar Biol* 26:208–217.

13   IPCC (2014) Climate Change 2014: Synthesis Report. Contribution of  
14           Working Groups I, II and III to the Fifth Assessment Report of the  
15           Intergovernmental Panel on Climate Change. Core Writing Team  
16           Pachauri RK and Meyer LA (ed) IPCC, Geneva, Switzerland, 151pp.

17   IPCC (2019) Summary for Policymakers. In: Climate Change and Land: an  
18           IPCC special report on climate change, desertification, land degradation,  
19           sustainable land management, food security, and greenhouse gas fluxes  
20           in terrestrial ecosystems. Shukla PR, Skea J, Calvo Buendia E, Masson-  
21           Delmotte V, Pörtner HO, Roberts DC, Zhai P, Slade R, Connors S, van  
22           Diemen R, Ferrat M, Haughey E, Luz S, Neogi S, Pathak M, Petzold J,  
23           Portugal Pereira J, Vyas P, Huntley E, Kissick K, Malley J (ed) In press.

24   Janecki T, Kidawa A, Potocka M (2010) The effects of temperature and  
25           salinity on vital biological functions of the Antarctic crustacean *Serolis*  
26           *polita*. *Polar Biol* 33:1013–1020.

27   Kingsolver JG, Umbanhowar J (2018) The analysis and interpretation of  
28           critical temperatures. *J Exp Biol* 221.

29   Kolzenburg R, D’Amore F, McCoy SJ, Ragazzola F (2021) Marginal  
30           populations show physiological adaptations and resilience to future  
31           climatic changes across a North Atlantic distribution. *Environ Exp Bot*

1 188:104522.

2 Lenihan HS, Peterson CH, Miller RJ, Kayal M, Potoski M (2018) Biotic  
3 disturbance mitigates effects of multiple stressors in a marine benthic  
4 community. *Ecosphere* 9.

5 Liu H, Kelly MS, Cook EJ, Black K, Orr H, Zhu JX, Dong SL (2007) The effect  
6 of diet type on growth and fatty-acid composition of sea urchin larvae, I.  
7 *Paracentrotus lividus* (Lamarck, 1816) (Echinodermata). *Aquaculture*  
8 264:247–262.

9 Marshall J, Scott JR, Armour KC, Campin JM, Kelley M, Romanou A (2015)  
10 The ocean's role in the transient response of climate to abrupt  
11 greenhouse gas forcing. *Clim Dyn* 44:2287–2299.

12 McClintock J (1994) Trophic biology of Antarctic shallow-water echinoderms.  
13 *Mar Ecol Prog Ser* 111:191–202.

14 van der Meer J (2006) An introduction to Dynamic Energy Budget (DEB)  
15 models with special emphasis on parameter estimation. *J Sea Res*  
16 56:85–102.

17 Melzner F, Stange P, Trubenbach K, Thomsen J, Casties I, Panknin U, Gorb  
18 SN, Gutowska MA (2011) Food Supply and Seawater pCO<sub>2</sub> Impact  
19 Calcification and Internal Shell Dissolution in the Blue Mussel *Mytilus*  
20 *edulis*. *PLoS One* 6:e24223.

21 Morley SA, Lai CH, Clarke A, Tan KS, Thorne MAS, Peck LS (2014) Limpet  
22 feeding rate and the consistency of physiological response to  
23 temperature. *J Comp Physiol B Biochem Syst Environ Physiol* 184:563–  
24 570.

25 Morley SA, Martin SM, Bates AE, Clark MS, Ericson J, Lamare M, Peck LS  
26 (2012a) Spatial and temporal variation in the heat tolerance limits of two  
27 abundant Southern Ocean invertebrates. *Mar Ecol Prog Ser* 450:81–92.

28 Morley SA, Martin SM, Day RW, Ericson J, Lai CH, Lamare M, Tan KS,  
29 Thorne MAS, Peck LS (2012b) Thermal Reaction Norms and the Scale of  
30 Temperature Variation: Latitudinal Vulnerability of Intertidal Nacellid  
31 Limpets to Climate Change. *PLoS One* 7:7–10.



- 1 Morley SA, Suckling CC, Clark MS, Cross EL, Peck LS (2016) Long-term  
2 effects of altered pH and temperature on the feeding energetics of the  
3 Antarctic sea urchin, *Sterechinus neumayeri*. *Biodiversity* 17:34–45.
- 4 Muggeo VMR (2008) Segmented: An R Package to Fit Regression Models  
5 with Broken-Line Relationships. *R News* 3:343–4.
- 6 Myrvoll-Nilsen E, Fredriksen HB, Sørbye SH, Rypdal M (2019) Warming  
7 Trends and Long-Range Dependent Climate Variability Since Year 1900:  
8 A Bayesian Approach. *Front Earth Sci* 7:1–8.
- 9 Obermüller BE, Morley SA, Barnes DKA, Peck LS (2010) Seasonal  
10 physiology and ecology of Antarctic marine benthic predators and  
11 scavengers. *Mar Ecol Prog Ser* 415:109–126.
- 12 Oliver ECJ, Burrows MT, Donat MG, Sen Gupta A, Alexander L V., Perkins-  
13 Kirkpatrick SE, Benthuyesen JA, Hobday AJ, Holbrook NJ, Moore PJ,  
14 Thomsen MS, Wernberg T, Smale DA (2019) Projected Marine  
15 Heatwaves in the 21st Century and the Potential for Ecological Impact.  
16 *Front Mar Sci* 6:1–12.
- 17 Oliver ECJ, Donat MG, Burrows MT, Moore PJ, Smale DA, Alexander L V.,  
18 Benthuyesen JA, Feng M, Sen Gupta A, Hobday AJ, Holbrook NJ,  
19 Perkins-Kirkpatrick SE, Scannell HA, Straub SC, Wernberg T (2018)  
20 Longer and more frequent marine heatwaves over the past century. *Nat*  
21 *Commun* 9:1–12.
- 22 Oschlies A, Brandt P, Stramma L, Schmidtko S (2018) Drivers and  
23 mechanisms of ocean deoxygenation. *Nat Geosci* 11:467–473.
- 24 Payton SL, Johnson PD, Jenny MJ (2016) Comparative physiological,  
25 biochemical and molecular thermal stress response profiles for two  
26 unionid freshwater mussel species. *J Exp Biol* 219:3562–3574.
- 27 Peck LS (2018) Antarctic Marine Biodiversity: Adaptations, Environments and  
28 Responses to Change. *Oceanogr Mar Biol An Annu Rev* 56:105–236.
- 29 Peck LS (2011) Organisms and responses to environmental change. *Mar*  
30 *Genomics* 4:237–243.
- 31 Peck LS (2005) Prospects for survival in the Southern Ocean: Vulnerability of

- 1 benthic species to temperature change. *Antarct Sci* 17:497–507.
- 2 Peck LS (1989) Temperature and basal metabolism in two Antarctic marine  
3 herbivores. *J Exp Mar Bio Ecol* 127:1–12.
- 4 Peck LS, Bullough LW (1993) Growth and population structure in the infaunal  
5 bivalve *Yoldia eightsi* in relation to iceberg activity at Signy Island,  
6 Antarctica. *Mar Biol* 117:235–241.
- 7 Peck LS, Clark MS, Morley SA, Massey A, Rossetti H (2009) Animal  
8 temperature limits and ecological relevance: Effects of size, activity and  
9 rates of change. *Funct Ecol* 23:248–256.
- 10 Peck LS, Morley SA, Pörtner H-O, Clark MS (2007) Thermal limits of  
11 burrowing capacity are linked to oxygen availability and size in the  
12 Antarctic clam *Laternula elliptica*. *Oecologia* 154:479–484.
- 13 Peck LS, Morley SA, Richard J, Clark MS (2014) Acclimation and thermal  
14 tolerance in Antarctic marine ectotherms. *J Exp Biol* 217:16–22.
- 15 Peck LS, Webb KE, Bailey DM (2004) Extreme sensitivity of biological  
16 function to temperature. *Funct Ecol* 18:625–630.
- 17 Peck LS, Webb KE, Miller A, Clark MS, Hill T (2008) Temperature limits to  
18 activity, feeding and metabolism in the Antarctic starfish *Odontaster*  
19 *validus*. *Mar Ecol Prog Ser* 358:181–189.
- 20 Piepho HP, Ogutu JO (2003) Inference for the break point in segmented  
21 regression with application to longitudinal data. *Biometrical J* 45:591–601.
- 22 Pierrat B, Saucède T, Laffont R, De Ridder C, Festeau A, David B (2012)  
23 Large-scale distribution analysis of Antarctic echinoids using ecological  
24 niche modelling. *Mar Ecol Prog Ser* 463:215–230.
- 25 Pörtner H-O, Bock C, Mark FC (2017) Oxygen- & capacity-limited thermal  
26 tolerance: Bridging ecology & physiology. *J Exp Biol* 220:2685–2696.
- 27 Pörtner H-O, Peck LS, Hirse T (2006) Hyperoxia alleviates thermal stress in  
28 the Antarctic bivalve, *Laternula elliptica*: Evidence for oxygen limited  
29 thermal tolerance. *Polar Biol* 29:688–693.
- 30 Pörtner H-O, Peck LS, Somero G (2007) Thermal limits and adaptation in

- 1 marine Antarctic ectotherms: An integrative view. *Philos Trans R Soc B*  
2 *Biol Sci* 362:2233–2258.
- 3 Robertson R, El-Haj AJ, Clarke A, Peck LS, Taylor E (2001) The effects of  
4 temperature on metabolic rate and protein synthesis following a meal in  
5 the isopod *Glyptonotus antarcticus* Eights (1852). *Polar Biol* 24:677–686.
- 6 Rubio-Portillo E, Izquierdo-Muñoz A, Gago JF, Rosselló-Mora R, Antón J,  
7 Ramos-Esplá AA (2016) Effects of the 2015 heat wave on benthic  
8 invertebrates in the Tabarca Marine Protected Area (southeast Spain).  
9 *Mar Environ Res* 122:135–142.
- 10 Salinas S, Munch SB (2012) Thermal legacies: Transgenerational effects of  
11 temperature on growth in a vertebrate. *Ecol Lett* 15:159–163.
- 12 Schlegel RW, Smit AJ (2018) HeatwaveR: A central algorithm for the  
13 detection of heatwaves and cold-spells. *J Open Source Softw* 3:821.
- 14 Spicer JI, Morley SA, Bozinovic F (2019) Physiological diversity, biodiversity  
15 patterns and global climate change: Testing key hypotheses involving  
16 temperature and oxygen. *Philos Trans R Soc B Biol Sci* 374:8–11.
- 17 Stanwell-Smith D, Peck LS (1998) Temperature and embryonic development  
18 in relation to spawning and field occurrence of larvae of three Antarctic  
19 echinoderms. *Biol Bull* 194:44–52.
- 20 Stevens MM, Jackson S, Bester SA, Terblanche JS, Chown SL (2010)  
21 Oxygen limitation and thermal tolerance in two terrestrial arthropod  
22 species. *J Exp Biol* 213:2209–2218.
- 23 Suckling CC, Clark MS, Beveridge C, Brunner L, Hughes AD, Harper EM,  
24 Cook EJ, Davies AJ, Peck LS (2014) Experimental influence of pH on the  
25 early life-stages of sea urchins II: Increasing parental exposure times  
26 gives rise to different responses. *Invertebr Reprod Dev* 58:161–175.
- 27 Suckling CC, Clark MS, Richard J, Morley SA, Thorne MAS, Harper EM, Peck  
28 LS (2015) Adult acclimation to combined temperature and pH stressors  
29 significantly enhances reproductive outcomes compared to short-term  
30 exposures. *J Anim Ecol* 84:773–784.
- 31 Terblanche JS, Deere JA, Clusella-Trullas S, Janion C, Chown SL (2007)

1 Critical thermal limits depend on methodological context. Proc R Soc B  
 2 Biol Sci 274:2935–2942.

3 Venables HJ, Clarke A, Meredith MP (2013) Wintertime controls on summer  
 4 stratification and productivity at the western Antarctic Peninsula. Limnol  
 5 Oceanogr 58:1035–1047.

6 Walter J, Jentsch A, Beierkuhnlein C, Kreyling J (2013) Ecological stress  
 7 memory and cross stress tolerance in plants in the face of climate  
 8 extremes. Environ Exp Bot 94:3–8.

9 Young JS, Peck LS, Matheson T (2006) The effects of temperature on  
 10 walking and righting in temperate and Antarctic crustaceans. Polar Biol  
 11 29:978–987.

12 Zupo V, Glaviano F, Paolucci M, Ruocco N, Polese G, Di Cosmo A, Costantini  
 13 M, Mutalipassi M (2019) Roe enhancement of *Paracentrotus lividus*:  
 14 Nutritional effects of fresh and formulated diets. Aquac Nutr 25:26–38.

15

16 **TABLES**

17 Table 1: Summary statistics for linear regression relationships between the  
 18 measured functions of *Sterechinus neumayeri* and temperature.  $\beta$  indicates the  
 19 slope of the linear regression lines before the breakpoint (Slope\_1) and after  
 20 the breakpoint (Slope\_2);  $SE_a$  indicates standard error for the intercept and  
 21 slopes; df = degrees of freedom; bold p-values indicate significant relationships  
 22 ( $p < 0.05$ ) between temperature and the variable measured and bold Davies p-  
 23 values represent a significant change ( $p < 0.05$ ) in the gradient of the slope of  
 24 segmented regressions. Values in the column BP indicate the localisation of  
 25 the breakpoint or else NA indicates a single linear regression;  $SE_b$  (standard  
 26 error) and  $R^2$  refers to the goodness of fit for the entire model.

Function	$\beta$	$SE_a$	p-value	BP	$SE_b$	$R^2$	Davies p-value
----------	---------	--------	---------	----	--------	-------	----------------

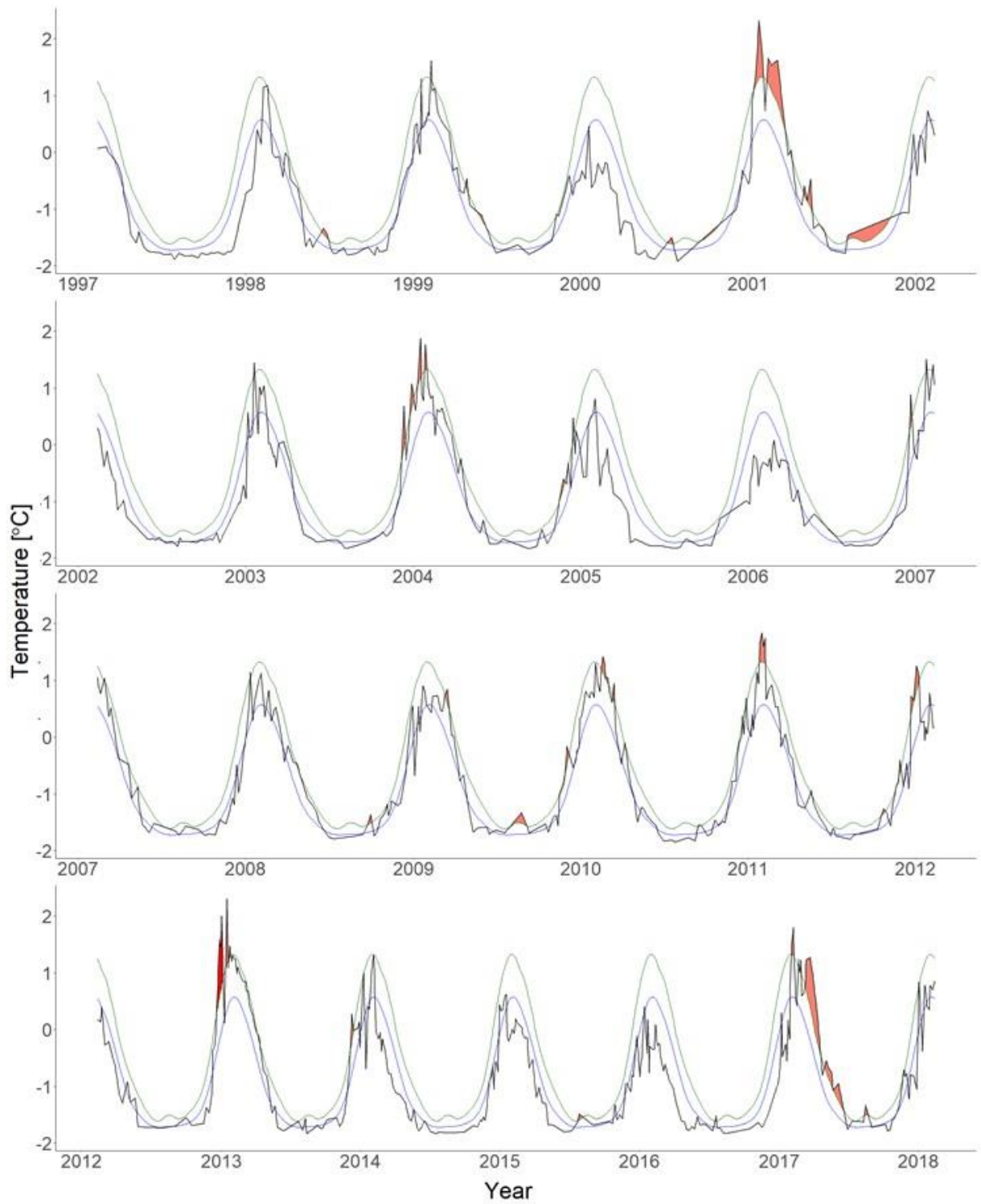
Individuals feeding, 1°C day <sup>-1</sup> (Intercept) Slope_1 Slope_2	89.0 3.45 -12.9	25.4 10.5 2.35	df=3 0.039 0.764 <b>0.012</b>	4.0	14.9	0.894	0.329
Individuals feeding, 0.5°C day <sup>-1</sup> (Intercept) Slope_1 Slope_2	110.3 -6.34 -11.5	12.7 3.14 1.05	df=7 <0.001 0.083 <b>&lt;0.001</b>	6.2	6.78	0.964	0.301
Individuals feeding, 0.3°C day <sup>-1</sup> (Intercept) Slope_1 Slope_2	95.3 -2.73 -20.3	7.53 1.38 2.92	df=12 <0.001 0.071 <b>&lt;0.001</b>	8.2	8.48	0.922	<b>0.001</b>
Individuals producing faeces, 1°C day <sup>-1</sup> (Intercept) Slope_1 Slope_2	-29.0 24.1 -13.3	23.1 9.54 2.13	df=3 0.298 0.085 <b>0.008</b>	5.2	13.5	0.881	<b>0.019</b>
Individuals producing faeces, 0.5°C day <sup>-1</sup> (Intercept) Slope_1 Slope_2	34.0 13.3 -10.3	28.6 8.54 8.68	df=7 0.274 0.162 <b>&lt;0.001</b>	4.5	12.1	0.844	<b>0.039</b>
Individuals producing faeces, 0.3°C day <sup>-1</sup> (Intercept) Slope_1 Slope_2	77.9 -0.306 -18.6	11.1 2.02 4.29	df=12 <0.001 0.882 <b>&lt;0.001</b>	8.3	12.5	0.762	<b>0.006</b>
Faeces produced, 1°C day <sup>-1</sup> (Intercept) Slope_1	0.645 -0.040	0.137 0.027	df=14 <0.001 0.165	NA	0.216	0.071	0.858
Faeces produced, 0.5°C day <sup>-1</sup> (Intercept) Slope_1 Slope_2	1.52 -0.23 -0.06	0.214 0.072 0.025	df=31 <b>&lt;0.001</b> <b>0.007</b> <b>0.016</b>	4.9	1.11	0.664	<b>0.043</b>
Faeces produced, 0.3°C day <sup>-1</sup> (Intercept) Slope_1 Slope_2	3.54 -0.718 -0.051	0.509 0.202 0.020	df=34 <b>&lt;0.001</b> <b>0.001</b> <b>0.012</b>	3.3	0.294	0.729	<b>&lt;0.001</b>
Time taken to right, 1°C day <sup>-1</sup> (Intercept) Slope_1 <sup>1</sup>	-8.60 6.83	9.04 1.35	df=26 0.350 <0.001	NA	23.3	0.476	NA
Time taken to right, 0.5°C day <sup>-1</sup> (Intercept) Slope_1 <sup>1</sup>	8.88 2.61	5.03 0.731	df=26 0.089 <b>0.001</b>	NA	13.1	0.302	NA
Time taken to right, 0.3°C day <sup>-1</sup> (Intercept) Slope_1 Slope_2	14.6 0.384 55.7	20.1 3.66 13.8	df=25 0.237 0.459 <b>&lt;0.001</b>	8.7	0.556	0.588	<b>&lt;0.001</b>
Oxygen consumption, 1°C day <sup>-1</sup> (Intercept) Slope_1 <sup>1</sup>	1.64 1.50	1.76 0.248	df=28 0.358 <b>&lt;0.001</b>	NA	4.64	0.551	NA
Oxygen consumption, 0.5°C day <sup>-1</sup> (Intercept) Slope_1 <sup>1</sup>	4.29 0.611	1.10 0.134	df=33 <0.001 <0.001	NA	3.17	0.368	NA

Oxygen consumption, 0.3°C day <sup>-1</sup> (Intercept) Slope_1 <sup>1</sup>	3.30 0.957	1.36 0.185	df=28 <b>0.022</b> <b>&lt;0.001</b>	NA	3.49	0.471	NA
--	---------------	---------------	---	----	------	-------	----

---

<sup>1</sup> Reporting only a single slope (Slope\_1) indicates that no breakpoint was detected in the regression and statistics for a single linear regression model is reported for the data instead.

1 **FIGURES**

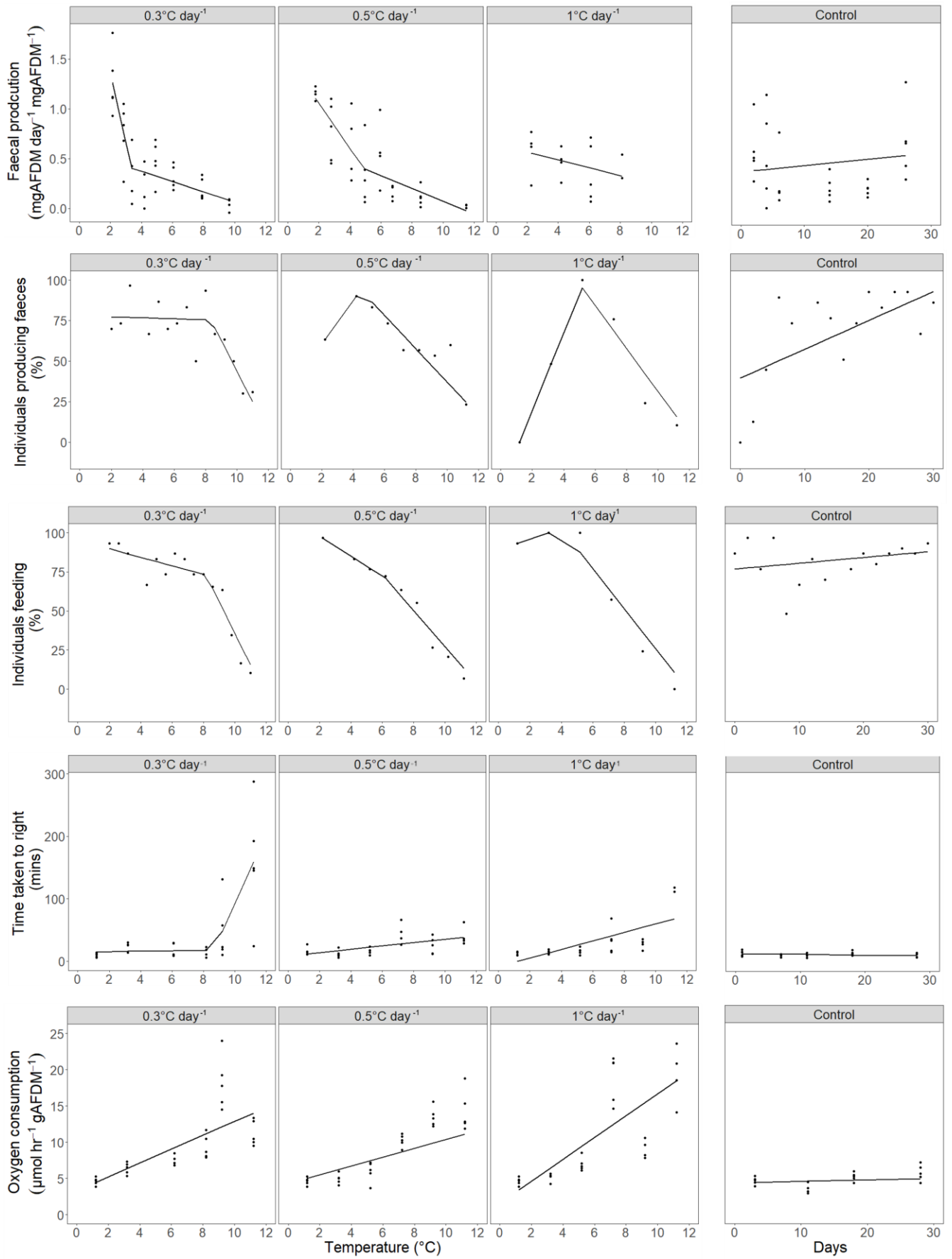


2

3 Figure 1: Times-series of temperatures (°C) experienced in Ryder Bay,  
4 Antarctica, at depths of 15 m, represented by the black lines. The data are split  
5 into panels to cover the entire span of the time-series, where the x-axis

1 represents time in years. Blue lines represent the seasonal climatology of the  
2 region based on the full time-series of daily temperatures (1997 – 2018). Green  
3 lines represent the seasonally varying threshold for a marine heatwave (90<sup>th</sup>  
4 percentile). Temperatures exceeding the threshold for  $\geq 5$  days are highlighted  
5 in red and indicate the occurrence of a marine heatwave.





1 Figure 2: *Sterechnus neumayeri*. Biological functions measured in *Sterechnus*

1 *neumayeri* in experimental conditions where temperatures were increased daily  
2 by 0.3°C, 0.5°C and 1°C. Functions in warming conditions are plotted against  
3 increasing temperature and ambient control treatments are plotted against the  
4 number of days in the experiment. Data points represent the pooled data within  
5 replicate floating tanks (n=5). Regressions are either segmented where  
6 appropriate for treatment conditions or linear for controls and treatment data  
7 where breakpoints were not identified.