

Effect thresholds for the earthworm *Eisenia fetida*: Toxicity comparison between conventional and biodegradable microplastics

Ding, Weili; Li, Zhen; Qi, Ruimin; Jones, David; Liu, Qiuyun; Liu, Qin; Yan, Changrong

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1 Effect thresholds for the earthworm *Eisenia fetida*: Toxicity
2 comparison between conventional and biodegradable microplastics

3 Weili Ding^{a,b,&}, Zhen Li^{a,b,&}, Ruimin Qi^{a,b,c,&}, Davey L. Jones^{c,d}, Qiuyun Liu^e, Qin Liu^{a,b}, Changrong
4 Yan^{a,b,*}

5 ^a *Institute of Environment and Sustainable Development in Agriculture, Chinese Academy of Agricultural*
6 *Sciences, Beijing 100081, P.R. China*

7 ^b *Key Laboratory of Prevention and Control of Residual Pollution in Agricultural Film, Ministry of Agriculture*
8 *and Rural Affairs, Beijing 100081, P.R. China*

9 ^c *School of Natural Sciences, Bangor University, Bangor, Gwynedd, LL57 2UW, UK*

10 ^d *SoilsWest, UWA School of Agriculture and Environment, The University of Western Australia, Perth, WA 6009,*
11 *Australia*

12 ^e *The Biocomposites Centre, Bangor University, Bangor, Gwynedd, LL57 2UW, UK*

13

14 [&] *These authors contributed equally to this work and should be considered as co-first authors. E-mail address:*
15 *dingweili@caas.cn (Weili Ding), lizhen@caas.cn (Zhen Li), qiruimin529@163.com (Ruimin Qi),*

16 ^{*} *Corresponding author. E-mail address: yanchangrong@caas.cn (Changrong Yan). Tel/Fax: +86-10-82106018*

17

18 **HIGHLIGHTS**

- 19 • Response of earthworms exposed to PE, PLA and PPC microplastics was studied.
20 • Avoidance, survival, biomass and reproduction of earthworms were tested.
21 • Earthworms clearly avoided microplastic concentrations > 40 g kg⁻¹.
22 • Number of cocoons during reproduction was significantly reduced at 53 g kg⁻¹.
23 • PLA and PPC microplastics showed no less toxicity compared to PE.

24

25 **ABSTRACT**

26 Biodegradable plastics have been developed to eliminate the progressive accumulation and
27 ever-growing threat posed by conventional fossil fuel-derived plastics. The impact of these bioplastics,

28 particularly in an agricultural context (e.g. biopolymer mulch films), however, remains poorly
29 understood. In this study, we compared the biotoxicity of biodegradable (polylactic acid, PLA;
30 polypropylene carbonate, PPC) and non-degradable (polyethylene, PE) microplastics using a series of
31 standardized bioassays using the earthworm *Eisenia fetida*. The responses studied included: avoidance
32 behavior, mortality, biomass, and reproduction responses. We incubated earthworms in artificial soils
33 amended with different concentrations of microplastic (0, 0.125, 1.25, 12.5, 125, 250, and 500 g kg⁻¹)
34 under laboratory conditions. This wide range allowed linear regression modeling and estimation of
35 microplastic effect thresholds. Our results showed that microplastic concentration rather than plastic
36 type was more important in regulating earthworm responses to soil contamination. The critical
37 threshold for microplastic contamination was 40 g kg⁻¹, after which earthworms exhibit microplastic
38 avoidance behavior. A significant reduction (EC₁₀) in number of cocoons and juvenile earthworms
39 occurred at a concentration of 53 g kg⁻¹ and 97 g kg⁻¹, respectively; while no significant effect was
40 found for survival of earthworm until levels of 500 g kg⁻¹. Overall, the two biodegradable materials
41 (PLA and PPC), appeared to be no more biofriendly than PE. Based on reported levels of plastic
42 contamination in soil of up to 67 g kg⁻¹, we conclude that microplastics are now starting to pose a
43 threat to earthworm population. To better evaluate the risk posed by biodegradable and nondegradable
44 plastics, further mechanistic studies on how microplastics affect earthworm behaviour and the
45 potential long-term impacts of this on soil functioning are required.

46 **Keywords:** Plastic mulching film, Microplastic safety, Earthworm response, Biototoxicity, Effect
47 threshold

48

49 **1. Introduction**

50 Due to the action of heat, UV irradiation, mechanical forces and microbial degradation, large
51 plastic debris in soil progressively deteriorates, leading to its fragmentation and the formation of
52 microplastics (Li et al., 2019; Steinmetz et al 2016; Ammala et al., 2011; Laycock et al., 2017). Plastic
53 debris less than 5 mm in size (i.e. microplastics) are generally thought to be more harmful when they
54 enter the environment in comparison to macroplastics (Steinmetz et al., 2016; Rillig et al., 2017). A
55 growing number of studies have reported ingestion of microplastics by different organisms, causing
56 inflammation and damage to tissues and organs, and which also leads to further transport and
57 accumulation in the food chain when these organisms are consumed (Lwanga et al., 2017). In addition,
58 microplastics have the ability to bind xenobiotics and undergo long distance migration which further

59 adds to their hazardous effect and the spread of pollution (Qi et al., 2020; Qian et al., 2020). In recent
60 years, microplastics have been increasingly recognized as one of the most important environmental
61 pollutants that threaten organismal health and the sustainability of ecosystem food webs (Gall and
62 Thompson, 2015; Horton et al., 2017; Hurley and Nizzetto, 2018). A recent literature survey on the
63 behaviour and fate of microplastics in the environment indicated that most studies have focused on
64 aquatic ecosystems, especially oceans (71% of the total) or on sediments from aquatic environments,
65 or beaches and sludges (24% of the total) (Qi et al., 2020). There is therefore a paucity of knowledge
66 regarding microplastic pollution in agricultural soils and terrestrial ecosystems (Bakir et al., 2016; Qi
67 et al., 2020)

68 Plastic mulching has been widely adopted in many regions of the world to promote agricultural
69 production due to its proven ability to improve water and nutrient use efficiency and suppress weed
70 growth (Kader et al., 2017; Li et al., 2020; Tang et al., 2020). However, incomplete removal of plastic
71 mulch films from soil at the end of the growing season has led to a progressive accumulation of
72 macroplastic fragments in soil. This has been reported to cause deterioration in soil health by
73 negatively affecting the soil's water holding capacity, damaging soil structure, slowing nutrient
74 cycling and adversely affecting soil organisms (Liu et al., 2014; Steinmetz et al., 2016; Yan et al.,
75 2014; Li et al., 2020). To overcome the problem of soil contamination by conventional film fragments,
76 the agricultural industry is rapidly adopting the use of biodegradable mulch films which are designed
77 to degrade in soil within 18 to 24 months (Feng, et al., 2019; Rodrigues et al., 2021). Typical materials
78 used in biodegradable mulch films include biobased polymers such as polylactic acid (PLA), and
79 chemo-synthetic polymers for instance polypropylene carbonate (PPC). Despite the growing market
80 for biodegradable mulch films, and many studies looking at improving their tensile strength and
81 functional properties (temperature and moisture conservation) (Rodrigues et al., 2021; Deng et
82 al., 2019), it is still unclear whether biodegradable mulch films and their constituents are truly
83 environmentally benign (Qi et al., 2018). This is of particular importance given the large amount of
84 microplastic particles that may be produced by biodegradable mulch films in a concentrated time
85 period before ultimate degradation (Qi et al., 2021; Sintim et al., 2019; Steinmetz et al., 2016;
86 Ammala et al., 2011; Laycock et al., 2017). There is therefore a critical need to investigate and
87 compare the effect of biodegradable and nondegradable plastics on agroecosystem health.

88 Due to their presence in upper trophic levels in soil food webs, earthworms (e.g. *Eisenia fetida*
89 and *Lumbricus terrestris*) are often used as bioindicators for assessing critical thresholds for pollutant

90 loading in soil (Calisi et al., 2013). As earthworms are also central to the delivery of a wide range of
91 soil-based ecosystem services, these thresholds can also be used to predict when a loss of soil
92 functioning will occur (Pérès et al., 2011). Studying the response of earthworms to non-degradable
93 and biodegradable microplastics therefore represents an important measure to evaluate how these
94 contaminants affect soil quality (Spurgeon et al., 2003; Zhang et al., 2018). Many studies have
95 indicated that non-degradable plastic can adversely affect earthworm fitness by causing intestinal
96 damage (*E. andrei*) under the exposure conditions of 125 mg kg⁻¹, producing an immune stress
97 response (Rodriguez-Seijo et al., 2017), and reducing the growth rate of earthworms (*Lumbricus*
98 *terrestris*) at high exposure levels (> 280 g kg⁻¹) in soil litter (Huerta Lwanga et al., 2016). Cao et al.
99 (2017) also highlighted that polystyrene particles (58 µm) at a loading rate of 10-20 g kg⁻¹
100 significantly inhibited the growth and increased the mortality of *E. fetida*, while Jiang et al. (2020)
101 also reported that exposure to polystyrene microplastics damaged the intestinal cells and DNA of *E.*
102 *fetida*. However, there are few studies on the effect of degradable plastic particles on earthworms, and
103 none have compared biodegradable materials with non-biodegradable materials.

104 The objectives of this study were therefore to: (1) ascertain the acute and chronic effect of
105 microplastics on *Eisenia fetida*; (2) determine the effect thresholds of microplastics to different
106 toxicity endpoint traits; and (3) determine the toxicity of biodegradable and nondegradable
107 microplastic particles.

108

109 **2. Materials and Methods**

110 *2.1. Artificial soil media*

111 An artificial soil was used to exclude the possibility that the soil may contain plastic particles,
112 earthworms and their cocoons and is an internationally accredited method for evaluating the
113 biotoxicity of pollutants (OECD, 2006). The artificial soil was prepared according to ISO 11268-1 and
114 ISO 11268-2 (ISO, 2012). Peat was bought from the Beijing Guangda Hengyi Technology Co., Ltd.,
115 China, and kaolinite clay and quartz sand were bought from the Shanghai Macklin Biochemical
116 Technology Co., Ltd., China. The different constituents were separately air-dried at room temperature,
117 and then mixed in a ratio (w/w) of 1:2:7. Subsequently, deionized water and calcium carbonate were
118 added to adjust the water content and pH value of the soil. After thorough mixing, the soil was stored
119 at room temperature for 48 h to equilibrate. The pH value was determined using standard electrodes
120 (Mettler Toledo, Switzerland) using a soil: distilled water ratio of 1:2.5 (w/w). Soil water holding

121 capacity was measured according to ISO11268-1 Annex C (ISO11268-1, 2012). The pH of the
122 artificial soil was 6.5 ± 0.5 and the final water content was 30% (i.e. 50% of the maximum water
123 holding capacity).

124

125 2.2. Earthworm cultivation

126 Adult earthworms of the species *Eisenia fetida* were purchased from Dilongli Group (Tianjin,
127 China) and incubated for several generations in the laboratory. Before the experiment, the worms
128 were incubated for a week in the artificial soil to adapt to the experimental conditions during which
129 time they were regularly fed with cow dung. Subsequently, they were transferred into artificial soils
130 without cow dung for 24 h to clean up the intestines before use in experiments. Adult worms at age of
131 2-3 months, with wet mass of 0.4 ± 0.05 g, and a clitellum that represents their maturity were chosen
132 for the subsequent incubation experiments. This is the growth period when most earthworms become
133 mature and are ready to produce offspring (Guo et al., 1981). A population of ten earthworms was
134 assigned to each mesocosm in the biotoxicity assays according to ISO 11268-1 and ISO 11268-2 (ISO,
135 2012).

136

137 2.3. Microplastics

138 Three types of microplastic particles were purchased from Zoomlion Plasticizing Ltd. (Changsha,
139 China). The properties of the three plastics, namely polyethylene (PE), polylactic acid (PLA), and
140 polypropylene carbonate (PPC) are shown in Table 1. A gradient concentration of microplastics (PE,
141 PLA or PPC) were used for the biotoxicity assays, that begins from an under environmentally relevant
142 exposure to 50% soil dry weight (0, 0.125, 1.25, 12.5, 125, 250 and 500 g kg⁻¹). This doses were
143 chosen to provide a sufficient microplastic range for the linear regression modeling and calculation of
144 effect thresholds.

145

146 2.4. Mesocosm design

147 The mesocosms consisted of polypropylene plastic boxes with dimensions 19 cm × 12.5 cm × 10
148 cm (length × width × height) (Fig. S1). Aeration holes ($n = 15$) were placed along the two longer sides
149 while a further 4 holes were placed in the top cap (Fig. S1). The mesocosms were filled with 450 g
150 (dry weight) of artificial soil. The mesocosms were placed in a RDN-800D-4 climate chamber
151 (Ningbo Southeast Instrument Co. Ltd., China) with temperature of 20°C, light intensity of 400~800

152 lux, a 12 h photoperiod and relative humidity of 70% (ISO11268-2, 2011). The moisture content of
153 the soil was maintained at 30% by periodically weighing the mesocosms and replacing any water
154 which had been lost by evaporation.

155

156 *2.5. Earthworm avoidance in response to microplastic exposure*

157 Microplastics (PE, PLA or PPC) were added to soil at six different concentrations in the
158 avoidance test: 0.125, 1.25, 12.5, 125, 250 and 500 g kg⁻¹ dry soil. In this experiment, a split
159 mesocosm approach was used whereby artificial soil was placed in one half of the container (450 g)
160 and plastic-contaminated soil (450 g) placed in the other half. A baffle plate was initially used to
161 separate the two compartments. At the start of the experiment, the baffle plate was removed and ten
162 earthworms placed on the soil surface at the boundary of the two compartments. Fresh cow dung (5 g)
163 was placed on the soil surface in the center of each compartment. Each treatment had four
164 independent replicates. After incubation for 48 h in the climate-controlled chambers, the numbers of
165 worms on each side of the test container were recorded alongside the mass of cow dung remaining.

166

167 *2.6. Earthworm biomass, reproduction and mortality in response to microplastic exposure*

168 Bio-toxicity assays were performed according to the international standard procedures ISO
169 11268-1 and ISO 11268-2 (ISO, 2012). Boric acid was used as a reference substance to validate the
170 condition of laboratory testing. As expected, the survival rate of earthworms to H₃BO₃, and the mean
171 50% lethal concentration (LD₅₀) to H₃BO₃ (Supplementary materials A) were highly consistent to the
172 reference values presented in ISO 11268 (ISO, 2012).

173 The incubation conditions for toxic bioassays of PE, PLA and PPC microplastics were identical
174 to those used in the H₃BO₃ test. In detail, ten earthworms of uniform age and weight were incubated
175 in replicate mesocosms ($n = 3$) containing artificial soil (450 g) and various concentrations of either
176 PE, PLA or PPC (0, 125, 250, and 500 g kg⁻¹). The mortality of earthworms was assessed by
177 recording the percentage of dead individuals after either 7 or 14 d.

178 In a parallel experiment, the earthworms were initially washed, dried with paper towels and
179 weighed (± 0.0001 g) before being placed in the mesocosms. The earthworms were recovered at day 7,
180 14, 21 and 28 and reweighed. Mechanical handling and the time out of soil was kept to a minimum.
181 After 28 d, we removed the adult worms and then counted the number of earthworm cocoons
182 according to ISO 11268-2 (ISO, 2012). The cocoons were then returned back to the original plastic

183 container so that the offspring number and biomass could be recorded at day 56.

184

185 2.7. Data analysis

186 All statistical analysis was undertaken in the R platform (R-Core-Team, 2019) and lme4 package
187 (Bates et al., 2014). The cut-off for statistical significance was considered to be $p < 0.001$. Linear
188 regression analyses were conducted using SPSS v22.0 (IBM Inc, Armonk, NY).

189 For the earthworm avoidance assays we recorded the number of live earthworms (Fig. S2) and
190 calculated the rate of avoidance (R) as follows:

$$191 \quad R = (N_c - N_p) / N_t \quad \text{(Eqn. 1)}$$

192 where N_c , N_p and N_t were the number of earthworms in the control compartment, in the
193 plastic-amended compartment and in total, respectively (Martinez Morcillo et al., 2013). This gives a
194 proportion-like variable ranging from -1 to 1, and $(R+1)/2$ values ranging from 0 to 1. We used the
195 transformation arcsine square root of $(R+1)/2$ to normalize the data (denoted hereafter as TR). Then,
196 we analyzed this dataset in two ways. Firstly, we included all the data. We considered the combination
197 of plastic material and a concentration as a treatment and the contrast as a separate treatment.
198 Therefore, there were nineteen treatments in total. We built a linear model taking the transformed rate
199 of avoidance (TR) as a response variable and treatment as an independent variable. For the second
200 approach, we excluded the contrast. We built another linear model taking TR as the response variable
201 and type of plastic and concentration as independent variables.

202 For the biomass, reproduction and mortality assays, we excluded the contrast. We built linear
203 mixed models by taking the logarithmically transformed biomass (or reproduction or mortality) as
204 response variables, material, concentration and days as independent variables and experiment box as a
205 random effect. The calculation was implemented with the stats package on the R platform (R Core
206 Team 2019).

207 In addition, the effect concentrations of EC_{10} and EC_{50} for behavior and development of
208 earthworms were calculated by solving the linear regression models for 10% and 50% effect doses
209 compared to C_0 control (0 g kg^{-1}).

210

211 3. Results

212 3.1. Earthworm avoidance test

213 Statistical analysis revealed that rates of earthworm avoidance sharply increased with PE, PLA,

214 and PPC microplastic concentration. However, interestingly, the avoidance behavior of earthworms
215 was relatively less sensitive to PLA in comparison to PE and PPC (Fig. S3). As shown in Figure 1, the
216 avoidance behavior of earthworms to PLA started at a concentration of 50 g kg⁻¹, behind that of PE
217 and PPC. Overall, however, that avoidance behavior of earthworms was not shown to be significantly
218 affected by the different types of plastic ($p = 0.894$), but was highly responsive to soil microplastic
219 concentration ($p < 0.001$). The interaction between plastic type and concentration was not significant.
220 In addition, the residual amounts of cow dung remaining on the soil surface on the plastic
221 contaminated side of the mesocosm after 48 h increased with increasing microplastic concentration
222 (Fig. S4). This reflected the avoidance behavior of the earthworms in the plastic contaminated
223 compartment. The disappearance of cow dung also showed no significant difference between the three
224 types of plastic tested. The changes in the abundance of cow dung on the surface of the soil at each of
225 the six concentration levels (i.e. 0.125, 1.25, 12.5, 125, 250, and 500 g kg⁻¹) and microplastics (i.e. PE,
226 PLA, and PPC) is shown in Figure S5.

227

228 *3.2. Mortality of earthworms exposed to microplastics*

229 In the unamended soil (control) and 125 g kg⁻¹ PE treatment, no earthworm mortality was
230 recorded. In contrast, earthworm mortality was recorded in all other treatments. It is worth mentioning
231 that while PE microplastics showed a more moderate effect than PLA and PPC at relatively low
232 concentrations, it caused more severe mortality of earthworms compared to PLA and PPC at high
233 concentrations. When the microplastic increased from 125 to 500 g kg⁻¹, the mortality of worms
234 increased from 0% to 12.5% under the PE treatment, 2.5% to 5% under the PLA, but stabilized at 6%
235 for PPC. Exposure time ($p = 0.264$) and the interaction of the type and concentration of plastic ($p =$
236 0.075) had no significant influence on earthworm mortality rate. Regardless of time and concentration,
237 the type of plastic (PE, PLA, and PPC) had no significant influence on earthworm death rate ($p =$
238 0.256) (Fig. 2).

239

240 *3.3. Earthworm biomass and reproduction changes in response to microplastic exposure*

241 Earthworm biomass significantly increased with exposure time in the PE, PLA, and PPC
242 microplastic treatments ($p < 0.001$, Fig. 3); although the increase in biomass generally decreased with
243 microplastic concentration. Specifically, earthworm growth rate in all the PE treatments and higher
244 concentrations (250 and 500 g kg⁻¹) of the PPC treatment were lower in comparison to the control

245 treatment. In contrast, at all PLA doses and the 125 g kg⁻¹ of PPC treatment, earthworm growth rates
246 were higher than in the control treatment. Overall, the type and concentration of microplastic and their
247 interaction had no significant influence on earthworm biomass ($p > 0.1$).

248 At harvest on day 28, the number of earthworm cocoons in soil was found to decrease with
249 increasing microplastic concentration. The amount of cocoons were similar between the control
250 treatment (C₀) and at lower levels of plastic contamination (PE, PLA, and PPC at 125 g kg⁻¹). This
251 contrasts with the lower cocoon counts recorded in the PE, PLA, and PPC treatments at doses of 250
252 and 500 g kg⁻¹ (Fig. 4 and 5). Again, compared to the biodegradable microplastics (PLA and PPC), PE
253 displayed less severe impact at lower doses, but more severe damage at higher contamination levels.

254 After returning the cocoons to the soil and incubation for a further 28 d, juvenile earthworms
255 were collected and their number and biomass recorded. The results showed that the number of
256 offspring in the PE₁₂₅ treatment was the highest, followed by that in the C₀, PLA₁₂₅ and PPC₁₂₅
257 treatments, while the lowest numbers were reported in the PE₅₀₀ and PLA₅₀₀ treatments (Fig. 6).
258 Regardless of plastic type (PE, PLA, and PPC), the number of offspring decreased as microplastic
259 concentration increased ($p < 0.001$).

260 The total biomass of all the juvenile earthworms in each test container was determined on day 56.
261 The total biomass of offspring in the PE treatment showed no difference to the control at PE₁₂₅ (10.2
262 g), but was lowest in the PE₅₀₀ treatment (7.4 g), decreasing significantly by 27%. In comparison, the
263 total biomass of offspring only decreased by 7% for PLA from 9.10 g in PLA₁₂₅ to 8.5 g in PLA₅₀₀,
264 and from 15% for PPC from 10.7 g to 9.02 g (Table S6).

265

266 3.4. RDA analysis and biotoxicity thresholds

267 RDA was used to explain the biomass and reproduction of earthworms (response variables) using
268 concentration and plastic type (explanatory variables) after incubation (Fig. 7, $p = 0.002$). We found
269 that the biomass and reproduction of earthworms were negatively correlated with microplastic
270 concentration, but not significantly with the type of plastic material.

271 Therefore, we calculated the EC₁₀ and EC₅₀ effect concentrations of the microplastics using
272 generalized linear regression models (Fig. 8), which instead of distinguishing between the types of
273 microplastics reflected them all as a whole. This was used to determine EC₁₀ and EC₅₀ values for the
274 effect of microplastic on the survival, development, behavior and reproduction of earthworms. Our
275 results showed that plastic avoidance is a very sensitive response of earthworms to soil microplastic

276 contamination, with EC₁₀ and EC₅₀ values of 40 g kg⁻¹ and 207 g kg⁻¹, respectively. Reproduction of
277 earthworms were also significantly affected by microplastic exposure, as with number of cocoons and
278 juvenile earthworms sharply reduced by 10% at 53 g kg⁻¹, 97 g kg⁻¹, and 50% at 347 g kg⁻¹, 500 g kg⁻¹,
279 respectively (Table 2). However, microplastics caused no significant effect on survival of earthworms
280 until they were present at extremely high concentrations (500 g kg⁻¹).
281

282 **4. Discussion**

283 *4.1. Acute response of earthworms upon exposure to microplastics*

284 Earthworms are commonly used as model organisms to assess the potential toxicity of soil
285 contaminants (ISO11268, 2012; Rombke et al., 2007). In our artificial mesocosms we showed that
286 earthworms exhibited clear avoidance behavior when the concentration of microplastics in soil
287 reached 40 g kg⁻¹. This supports the previous study of Huerta Lwanga et al. (2017) who reported that
288 earthworms migrated to deeper soil layers when polyethylene concentrations in soil litter layers
289 reached up to 70 g kg⁻¹. A key finding from this study was that all microplastic types induced
290 avoidance behavior, irrespective of chemical formulation, suggesting that the avoidance behavior of
291 earthworms was mainly related to the physical properties of the microplastic or its chemical properties.
292 Some previous studies have reported that high concentrations of microplastic can adversely affect soil
293 structure (e.g. soil bulk density, water holding capacity, and soil aggregates), unfavorable for
294 earthworm movement and soil ingestion (de Souza Machado et al., 2018). In addition, microplastics
295 have been shown to cause burns and lesions on the surface of earthworms (Baeza et al., 2020), leading
296 to avoidance behavior. It is worth mentioning that the avoidance behavior of earthworms to PLA was
297 always relatively lower than to PE and PPC (Fig. S3). A possible reason might be that PLA is a
298 biopolymer material obtained by polymerization of lactic acid that might represent a supply of
299 available carbon for the earthworms at relatively low concentrations. However, the mechanistic basis
300 and factors influencing earthworm avoidance behavior needs to be investigated further.

301 In our experiment, exposure time (7 or 14 days) had no significant influence on the mortality of
302 earthworms. It is possible that the earthworms had adapted to the presence of microplastics, especially
303 low levels, after incubation for one week. Mortality was, however, significantly higher in the high PE
304 treatment (250 g kg⁻¹) in comparison to the control treatment. This result is similar to that reported by
305 Huerta Lwanga et al. (2016), however, it should be noted that these concentrations represent extreme
306 addition rates to soil which are typically only seen in waste contaminated urban soils (He et al., 2019;

307 Yang et al., 2018). It is also likely that these urban soils would contain a range of other
308 co-contaminants (e.g. metals, PAHs) which may also compound the effect of the plastics (Browne et
309 al., 2013; Gomiero et al., 2018). It is worth mentioning that, PE, PLA, and PPC had a different
310 influence on the death rate of worms, with PE being particularly toxic at high concentrations, while
311 less harmful than PPC and PLA at lower contamination levels. The underlying reason may be related
312 to selective uptake of different materials and degradable degrees of microplastics in the earthworm
313 intestine (Zhang et a., 2018). Biodegradable microplastics such as PLA and PPC might also be
314 ingested by earthworms at higher proportions due to their higher degradability and thus greater
315 associated biofilm and microbial load (Zhang et a., 2018). The greater biofilm and nutritional content
316 may also reduce their toxicity. In contrast, PE might accumulate more in the earthworm
317 intestines/typhlosole inducing blockages (Chen et al., 2020; Huerta Lwanga et al., 2016). Huerta
318 Lwanga et al. (2018) indicated that low-density polyethylene (LDPE) microplastics could be degraded
319 by bacteria isolated from the *Lumbricus terrestris* gut, however, this breakdown process is expected to
320 be very slow relative to the transit time through the gut. Zhang et al. (2018) also reported that
321 earthworms did not ingest PE, but foraged partial field-weathering biodegradable microplastics with
322 smaller particle sizes for food. Currently, there is no consistent evidence on the adverse effect of
323 nondegradable and biodegradable microplastics on earthworm intestines. In spite of this, the
324 discussion above indicates the potential risk of microplastic particles to the survival of earthworms.

325

326 4.2. Chronic response of earthworms to microplastics

327 The growth rate and biomass of earthworms decreased with the increasing concentration of PE in
328 this study, a finding also demonstrated by Huerta Lwanga et al. (2016), who indicated that the growth
329 rate of *L. terrestris* decreased with a higher percentage of PE microplastics (280 g kg⁻¹, 450 g kg⁻¹, and
330 600 g kg⁻¹, size < 150 µm) in the soil litter layer. A similar dose-dependent decrease of growth rate
331 was also reported by Redondo-Hasselerharm et al. (2018) in a study of freshwater benthic
332 macroinvertebrates with polystyrene microplastics ranging from 0 g kg⁻¹ to 400 g kg⁻¹ in sediment. In
333 addition, the number and mass of microplastics inside the body of *G. pulex* showed a positive
334 relationship to sediment exposure (Redondo-Hasselerharm et al., 2018). The adverse effects of
335 microplastics would be mainly caused by the significant accumulation of microplastics in the gut and
336 stomach of organisms, which can damage their immune systems and affect their feeding behavior and
337 development (Eltemsah et al., 2019; Liu et al., 2019; Rist et al., 2017; Rodriguez-Seijo et al., 2017;

338 Yin et al., 2019). Moreover, it is important to stress that biomass is not strictly a reliable indicator on
339 the growth of earthworms, as it may also include the weight of microplastics that have not been
340 egested.

341 For the eggs and reproduction (including offspring number and biomass) of earthworms, there
342 was no obvious distinction between the low concentration treatment (125 g kg⁻¹ of PE, PLA, and PPC)
343 and the control (C0) which decreased with increasing concentration level. Kwak and An (2021)
344 exposed earthworms to two different sizes of polyethylene microplastic for 21 days, and concluded
345 that microplastics affected coelomocyte viability and caused damage to male reproductive organs,
346 while having negligible effects on female reproductive organs, which may affect the reproduction of
347 earthworms.

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349 *4.3. Effect thresholds of microplastics on development and behavior of earthworms*

350 Nanoplastics generated from ingested microplastics can be introduced into soils through cast
351 excretion and these may pose an additional risk to soil organism and environment (Rillig, 2012;
352 Rodriguez-Seijo et al., 2017). Our results showed clear avoidance of earthworms to microplastic
353 exposure > 40 g kg⁻¹, which is the most sensitive response of earthworms observed in our study,
354 suggesting an instinct capability for self-preservation. In addition, microplastics also caused a
355 significant inhibition on the reproduction of earthworms as the number of cocoons sharply decreased
356 by 10% at concentrations of 53 g kg⁻¹. On the basis of reported levels of plastic contamination in
357 terrestrial soils as high as 67 g kg⁻¹ (Fuller et al., 2016), it is possible that microplastics are already
358 starting to pose a threat to earthworm populations. However, no significant effect of microplastics was
359 observed on the survival of earthworms, which may suggest some autoregulation and physiological
360 protective effects within the adult earthworm population. Further studies should seek to gain a deeper
361 mechanistic understanding of the effects of different microplastics on earthworms. In our study,
362 microplastic concentration was the dominant factor affecting earthworm biomass and reproduction,
363 while material type had a much lesser effect. PLA and PPC, as two biodegradable materials, were no
364 more benign than PE. The biosafety of biodegradable plastic film remains to be verified (Qi et al.,
365 2018; Sintim et al., 2019; Zhang et al., 2018). Additionally, this study mainly compared the effects of
366 three types of microplastics (PE, PLA, and PPC) on earthworm behavior and survival, and did not
367 explore their ecotoxicological mode of action. The ingestion of microplastics in earthworm bodies and
368 its effect on the pathological tissue of earthworms should therefore be further studied.

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5. Conclusions

Here we evaluated the response of *Eisenia fetida* in soils amended with different concentrations and types of microplastic. We found that the biomass and reproduction of the earthworms was negatively affected at microplastic concentrations greater than 40 g kg⁻¹. With microplastic concentrations as high as 67 g kg⁻¹ being reported in terrestrial environments, this suggests that microplastics may already be adversely affecting native earthworm populations and thus negatively impacting on soil functioning. Concentration proved to be the dominant factor affecting earthworm biomass and reproduction, rather than type of plastic material. The two biodegradable microplastics (PLA and PPC) did not appear to be more environmentally benign than PE. To improve our understanding of microplastic behavior in agricultural soil, further work is needed to identify the production rate of microplastics from biodegradable and nondegradable films, and their distribution in the natural environment. Additionally, further studies are needed to gain a better mechanistic understanding of how biodegradable microplastics affect earthworms and the potential long-term impacts of these effects on soil functioning. Together, these will allow a more holistic evaluation of the safety of biodegradable plastic use in agriculture.

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