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# Combined drug triads for synergic neuroprotection in retinal degeneration

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#### ABSTRACT

This review focuses on retina degeneration occurring during glaucoma, age-related macular degeneration (AMD), diabetic retinopathy (DR), and retinitis pigmentosa (RP), and on the potential therapeutic use of triads of repositioned medicines, addressed to distinct but complementary targets, to prevent, delay or stop retina cell death. Although myriad pathogenic mechanisms have been implicated in these disorders, common signaling pathways leading to apoptotic cell death to all of them, and to all neurodegenerative diseases are (i) calcium dyshomeostasis/excitotoxicity: (ii) oxidative stress/mitochondrial dysfunction, and (iii) neuroinflammation/ P2X7 receptor activation. From a therapeutic point of view, it is relevant to consider the multitarget approach based on the use of combined medicines acting on complementary pathogenic mechanisms that has been highly successful in the treatment of chronic diseases such as cancer, AIDS, pain, hypertension, Parkinson's disease, cardiac failure, depression, or the epilepsies as the basic mechanisms of cell death do not differ between the different CNS degenerative diseases. We suggest the multi-target therapy approach could be more effective compared with single-drug treatments. Used at doses lower than standard, these triads may also be safer and more efficient. After the establishment of a proof-of-concept in animal models of retinal degeneration, potential successful preclinical trials of such combinations may eventually drive to test this concept in clinical trials in patients, first to evaluate the safety and efficacy of the drug combinations in humans and then their therapeutic advantages, if any, seeking the prevention and/or the delay of retina degeneration and blindness.

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*Abbreviations*: A2E, N-retinyl-N-retinylidene ethanolamine; AD, Alzheimer's disease; AGE, advanced glycation end product; ALS, amyotrophic lateral sclerosis; AMPA, α-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptor; AMD, age related macular degeneration; Ang, angiopoietin; BBB, brain-blood barrier; anti-Aβ, anti-amyloid beta; BzATP, 2,3-O-(4-benzoylbenzoyl)-ATP; CaBP, calcium-binding protein; CaMKII, Ca<sup>2+</sup>/calmodulin-dependent protein kinase II; CAT, combined antioxidant therapy; Cav, voltage-dependent calcium channels; CICR, Ca<sup>2+</sup>-induced Ca<sup>2+</sup> release; CREB, cyclic AMP response element binding protein; CT, clinical trial; CYP, cytochrome P450; DHA, docosahexaenoic acid; DM, diabetes mellitus; DR, diabetic retinopathy; DREAM, downstream regulatory antagonist modulator; EAAT2, excitatory amino acid transporter 2; EGF, epidermal growth factor; ER, endoplasmic reticulum; ERG, electroretinogram; FDA, US Food and Drug Administration; GCL, ganglion cell layer; GluR, glutamate receptor; HCMD, high calcium microdomain; HD, Huntington's disease; INL, inner nuclear layer; IOP, intraocular pressure; IP<sub>3</sub>, inositol trisphosphate; IP<sub>3</sub>R, inositol trisphosphate receptor; mCU, mitochondrial Ca<sup>2+</sup> uniporter; mPTP, mitochondrial permeability transition pore; mRGC, melanopsin-containing retinal ganglion cell; MS, multiple sclerosis; NFAT, nuclear factor of activated T-cells; NFL, nerve fiber layer; NMDA, N-metyl-Daspartate; NO, nitric oxide; NOS, nitric oxide synthase; oATP, oxidized ATP; ONL, outer nuclear layer; P2X7R, purinergic P2X7 receptor; PD, Parkinson's disease; PPADS, pyridoxal-phosphate-6-azophenyl-2,4-disulfonic acid; PUFA, polyunsaturated fatty acids; RBB, retinal-blood barrier; RP, retinitis pigmentosa; RGC, retinal ganglion cell; ROS, reactive oxygen species; RPE, retinal pigment epithelial cell; RyR, ryanodine receptor; S1P, sphingosine-1-phosphate; TLR, toll-like receptor; TRPC, transient receptor potential canonical; TRPV, transient receptor potential vanilloid; UPR, unfolded protein r

#### 1. Introduction

The eye retina is a brain tissue. As such, vessels, glia, and neurons are its main constituents. Photoreceptors, bipolar cells, horizontal cells, amacrine cells, and retinal ganglion cells (RGCs) are the neuronal types of the retina. Their disposition in the different retinal layers is schematically displayed in Fig. 1. Located in the outer nuclear layer (ONL) of the vertebrate retina, the photoreceptors rods and cones initiate the phototransduction process of converting light energy into electrical signals. The cell bodies of interneurons (horizontal, bipolar, and amacrine cells), predominantly located at the inner nuclear layer (INL), modify and relay the visual information from photoreceptors to the retinal ganglion cells (RGCs), at the ganglion cell layer (GCL). Their axons initially run in the nerve fiber layer (NFL) to form the optic nerve, conveying the visual signals to the brain to interpret them as visual images [1].

A complex light-sensitive network such as the retina, which is formed by multiple layers of interconnected neurons, exhibits high vulnerability upon its exposure to several types of stress injury. This is exemplified by degenerative diseases primarily originated in the retina as well as those secondary to neurodegenerative diseases affecting different brain



**Fig. 1.** Retinal cytoarchitecture. An overall view of the eye. Representation of a vertical section of a human ocular globe (top) showing the main retinal layers and cell types (bottom). RPE, retinal pigment epithelial cell; ONL, outer nuclear layer; OPL, outer plexiform layer; INL, inner nuclear layer; GCL, ganglion cell layer; R, rods; C, cones; B, bipolar cells; H, horizontal cells; A, amacrine cells; G, ganglion cells; M, Müller cells; Mi, microglial cells.

tissues. The following are the main eye diseases leading to retina degeneration, which will be discussed in this review: (i) glaucoma, a leading cause of blindness, is associated to augmented intraocular pressure (IOP), degeneration of RGCs, and optic nerve damage; (ii) agerelated macular degeneration (AMD), the main cause of irreversible vision loss in the elderly, secondary to dysfunction and loss of retinal pigment epithelial (RPE) cells and photoreceptors, with the stimulation of vascular angiogenesis, an increase in the formation of lipofuscin granules, an accumulation of advanced glycation end products, drusen formation, and breakdown of the blood retinal barrier and changes in pigmentation, altogether with a reduction in melanosomes, increased thickness of Bruch's membrane and mitochondrial DNA deletions; (iii) diabetic retinopathy (DR), a complication occurring in patients with poorly controlled diabetes, blindness is secondary to blood vessels growth on the retinal surface and apoptotic death of RGC, photoreceptors, horizontal or amacrine cells; and (iv) retinitis pigmentosa (RP), a group of inherited diseases leading to blindness, photoreceptor degeneration occurs, being usually the rods affected first, and then cones [2, 31.

Retinal damage secondary to neurodegenerative diseases has also been observed. This is the case of retinopathy in Alzheimer's disease [4] and Parkinson's disease (PD) [5]. In PD, the degeneration of photosensitive melanopsin-containing RGC has recently been associated to disorders of sleep and circadian rhythms [6]. A recent study concludes that, as happens to be the case in striatal dopaminergic neurons, the amacrine dopaminergic neurons also degenerate in the retina of PD patients, thus explaining the visual alterations above mentioned [7].

Myriad receptors, ion channels, intracellular signaling pathways, and mutated proteins have been implicated in neurodegenerative diseases. Several of these pathological hallmarks are also present in the pathogenesis of retinal degeneration.

The basic pathogenic pathways leading to cell death in either retinal degenerative diseases and neurodegerative diseases such as for instance, Alzheimer's disease (AD) or Parkinson's disease (PD), do not significantly differ. Therefore, the drug targets being investigated are similar in both types of diseases.

In this context, we thought of interest to review here three pathogenic features that are common to both neurodegenerative diseases and retina degeneration, that is, excitotoxicity linked to Ca<sup>2+</sup> dyshomeostasis, oxidative stress associated to mitochondrial dysfunction, and neuroinflammation mediated by the purinergic receptor P2X7 (P2X7R). Based on the therapeutic concept that various chronic diseases are best treated with combined drugs acting on different targets, the concept here raised suggests that novel multi-target therapeutic approaches with orally administered combined triads of repositioned medicines acting on the three above mentioned common pathogenic pathways may delay, slow, or even stop the progression of retina degeneration in glaucoma, DR, AMD and RP, versus single-target drug treatments. We will next review the implication of those three signaling pathways in the pathogenesis of retinal degeneration.

# 2. Calcium dyshomeostasis, excitotoxicity and retinal degeneration

To better understand the role of distorted  $Ca^{2+}$  dynamics in retinal degeneration we will first briefly review the mechanisms involved in the handling of  $Ca^{2+}$  by excitable cells and its implication in cell survival and death.

### 2.1. Calcium homeostasis and neuronal function

At each moment of cell activity, the homeostasis of  $Ca^{2+}$  is determined by fluxes between three compartments: the extracellular medium, the cytosol and the  $Ca^{2+}$ -storage organelles. At rest, the free  $Ca^{2+}$ concentrations are in the range of  $10^{-7}$  M in the cytosol and the mitochondrial matrix, and around  $10^{-3}$  M at the extracellular medium and at



**Fig. 2.** Intracellular calcium homeostasis. The cytosolic Ca<sup>2+</sup> concentration ([Ca<sup>2+</sup>]<sub>c</sub>) at each moment of cell activation is determined by Ca<sup>2+</sup> fluxes among different cellular compartments, including (i) Ca<sup>2+</sup> entry through plasmalemmal voltage-dependent calcium channels (Cav) that open during cell activation, (ii) Ca<sup>2+</sup> sequestration into and Ca<sup>2+</sup> release from the endoplasmic reticulum (ER) and mitochondria (M), and (iii) plasmalemmal Ca<sup>2+</sup> efflux transporters (PCET). The ER stores high concentrations of free Ca<sup>2+</sup> at rest, while it is present at low concentrations in M and cytosol. After cell activation, Ca<sup>2+</sup> enters into the cytosol through Cav or is released from the ER. The M take up vast amounts of Ca<sup>2+</sup> either from subplasmalemmal sites and/or from Ca<sup>2+</sup> released from the ER. http://smart.servier.com/.

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the endoplasmic reticulum (ER) store. Consequently, there are huge concentration gradients favoring  $Ca^{2+}$  diffusion to the cytosol. Thus, the increase of local cytosolic  $Ca^{2+}$  concentrations ( $[Ca^{2+}]_c$ ) is controlled by (i)  $Ca^{2+}$  entry through plasmalemmal calcium channels that open during cell activation, (ii)  $Ca^{2+}$  sequestration into and  $Ca^{2+}$  release from the ER and mitochondria, and (iii) plasmalemmal  $Ca^{2+}$  efflux transporters (Fig. 2) [8].

The entry of Ca<sup>2+</sup> into neurons mainly occurs through ionotropic glutamate receptor channels (i.e. N-methyl-D-aspartate, NMDAR;  $\alpha$ -amino-3-hydroxy-5-methylisoxazol-4-propionic acid, AMPAR) and voltage-activated calcium channels (Cav). Upon cell activation, Ca<sup>2+</sup> ions enter the cell through a 10<sup>4</sup> Ca<sup>2+</sup> gradient, giving rise to high [Ca<sup>2+</sup>]<sub>c</sub> microdomains (HCMDs) at specific sites [9,10]. The generation of HCMDs may also occur by ER Ca<sup>2+</sup> release through inositol trisphosphate (IP<sub>3</sub>) receptor channels or via ryanodine receptors (RyRs) through Ca<sup>2+</sup>-induced Ca<sup>2+</sup> release (CICR) [11].

Calcium transport by mitochondria has received much attention because its three well-defined roles: (i) the shaping of the  $[Ca^{2+}]_c$  transients; (ii) the regulation of mitochondrial respiration and ATP synthesis; and (iii) the activation of the programmed cell death [12].  $Ca^{2+}$  is taken up through the mitochondrial  $Ca^{2+}$  uniporter (mCU), a low-affinity/high-capacity system [13]; the driving force of the mitochondrial membrane potential (-150 to -180 mV) would promote the accumulation of  $Ca^{2+}$  into the mitochondrial matrix up to 5–6 orders of magnitude above the  $[Ca^{2+}]_c$  [14].  $Ca^{2+}$  exchanger and also through a Na<sup>+</sup>/Ca<sup>2+</sup> exchanger and also through a Na<sup>+</sup>-independent system [15]. Plasma membrane  $Ca^{2+}$  exit is due to joint operation of a high-affinity  $Ca^{2+}$ -ATPase and the Na<sup>+</sup>/Ca<sup>2+</sup>

exchange transporter [16].

Finally, cytosolic calcium-binding proteins (CaBPs) with low Ca<sup>2+</sup> affinity also contribute to the clearance of HCMDs. The brain is rich in CaBPs namely parvalbumin, calbindin-D28K, calmodulin, calcineurin, calretinin, and the S-100 family. Parvalbumin, calbindin-D28K, and calretinin are generally regarded as Ca<sup>2+</sup> buffering proteins rather than Ca<sup>2+</sup> regulatory proteins; hence, they play a vital role in neuronal Ca<sup>2+</sup> homeostasis [17,18]. The expression levels of CaBPs in specific neurons is conditioning the shaping of  $[Ca^{2+}]_c$  transients, as well as the Ca<sup>2+</sup>-dependent vulnerability of those neurons to different types of stressors. For instance, cortical neurons containing calretinin are resistant to calcium overload and excitotoxicity. Furthermore, in tissues from patients with either amyotrophic lateral sclerosis (ALS) or temporal lobe epilepsy, neurons containing calbindin or parvalbumin survive while those lacking them undergo degeneration [19,20].

This complex machinery for the regulation of  $Ca^{2+}$  homeostasis is required for multiple Ca2+-dependent functions namely neurotransmitter release [21], short and long-term synaptic plasticity [22] or expression of genes [23]. These functions are exerted by a variety of  $Ca^{2+}$  responsive elements, such as proteins involved in synaptic vesicle fusion with the plasmalemma during exocytosis (synaptotagmins),  $Ca^{2+}$ -dependent kinases and phosphatases ( $Ca^{2+}/CaM$  kinases and Ca<sup>2+</sup>-dependent phosphatase calcineurin), Ca<sup>2+</sup>-dependent signaling enzymes (adenylate cyclase and nitric oxide synthase), and Ca<sup>2+</sup>-dependent transcription factors (cyclic AMP response element binding protein (CREB), calcineurin β-controlled nuclear factor of activated T-cells (NFAT), and Ca<sup>2+</sup> binding downstream regulatory antagonist modulator (DREAM)). The diversity of these Ca<sup>2+</sup>-dependent elements provides a means for Ca<sup>2+</sup>-dependent regulation of neuronal function in a time scale ranging from microseconds (as in the case of vesicle fusion during neurotransmitter release), to seconds and minutes (as in the case of Ca<sup>2+</sup>-dependent protein phosphorylation and dephosphorylation), to days and years (as in the case of Ca<sup>2+</sup>-dependent changes in neuronal gene expression). These Ca<sup>2+</sup>-dependent processes lead to short- and long-term changes in neuronal excitability (by modulating ion channel expression and activity) and synaptic transmission (by modifying the synaptic machinery and facilitating the formation or disassembly of synaptic connections) [24].

## 2.2. Calcium, neuronal viability and neuronal death

It is intriguing that  $Ca^{2+}$  ions may promote cell survival (at lower cytosolic concentrations) or cell death (at higher cytosolic concentrations). This could be explained considering the extreme sensitivity of neurons to variations in  $Ca^{2+}$  signals. Thus, even subtle defects and abnormalities in the complex machinery that control the signals might lead to devastating consequences, as in the case of neurodegenerative diseases [25,26].

The role of  $Ca^{2+}$  as a death trigger was first suggested by Albrecht Fleckenstein in 1974. He proposed that excess Ca<sup>2+</sup> influx into cardiac myocytes could be the mechanism underlying the pathology of cardiac ischemia [27]. This concept was extended soon to other tissues showing that both, receptor overstimulation [28] and cytotoxic agents [29,30] induced lethal influx of Ca<sup>2+</sup> into cells. Additionally, excess Ca<sup>2+</sup> entry through L-type voltage-activated Ca<sup>2+</sup> channels (Cav) also elicited mitochondrial disruption and cell death [31]. It is however puzzling that a mild sustained  $[Ca^{2+}]_{c}$  elevation may have opposite neuroprotective effects; this is the case of the rescue of neurons from death by a mild elevation of the extracellular K<sup>+</sup> concentration [32–35]. Also, depolarization preconditioning produced neuroprotection [36]; in this direction are the observations that the  $K^{\!+}$  channel blocker tetraethylammonium [37] or blockers of the M-type K<sup>+</sup> current [38] also caused neuroprotection. The mechanism underlying these effects may be linked to cell depolarization, activation of Cav (particularly the L-subtype), and elevation of  $[Ca^{2+}]_c$  [39].

The formulation of the interesting hypothesis of the Ca<sup>2+</sup> set point



**Fig. 3.** Plausible pathway of cell death in  $Ca^{2+}$ -dependent excitotoxicity. High rate of  $Ca^{2+}$  influx through NMDA and AMPA receptors or voltage-activated  $Ca^{2+}$  channels (Cav) seems to be the main pathway leading to neuronal injury in excitotoxicity. Enhanced  $Ca^{2+}$  entry through the P2X7R pore also elicits  $Ca^{2+}$  overload. Elevated intracellular  $Ca^{2+}$  induces the activation of cellular enzymes with potential toxic effect, mitochondrial dysfunction, ROS formation and the subsequent death of postsynaptic neurons. Activation of P2X7Rs in glial cells are involved in the inflammasome-dependent cell death pathway. Extracellular ATP released from lesioned neurons or glia causes further activation of microglia leading to the release of pro-inflammatory cytokines, the propagation of gliosis, and cell death. EAAT2, excitatory amino acid transporter 2.

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indicates that a minimal  $[Ca^{2+}]_c$  elevation is required to maintain neuronal viability [40]. When the  $[Ca^{2+}]_c$  moves below or above this set point, the apoptotic cascade is rapidly activated, leading to neuronal death. This apoptotic effect may involve the transcriptional activation and synthesis of antiapoptotic factors; also, it could be that  $Ca^{2+}$  influx keeps the ER  $Ca^{2+}$  store filled, thereby preventing its depletion by apoptotic stimuli or ER stress responses [41].

### 2.3. Calcium and excitotoxicity

Pioneering experiments demonstrated the Ca<sup>2+</sup>-dependence of glutamate-elicited excitotoxicity [42]. Soon thereafter, this mechanism was linked to several diseases of the nervous system [43] and was considered to be the final common pathway leading to neuronal death in neurotrauma, stroke, epilepsy, and neurodegenerative diseases [44,45].

In the central nervous system, the basal extracellular concentration of glutamate is kept at around 0.6  $\mu$ M, as monitored by intracerebral microdialysis [46]. Its elevation to 2–5  $\mu$ M is sufficient to cause excessive glutamate receptor stimulation and neuronal death [47,48]. Physiologically, AMPA receptors mediate fast excitatory neurotransmission while NMDA receptors are responsible for the late component of excitatory neurotransmission [49], playing a key role in synaptic plasticity [50].

Ionotropic glutamate receptors are permeable to Na<sup>+</sup>, Ca<sup>2+</sup>, and K<sup>+</sup> to different degrees, while metabotropic receptors are coupled to G-proteins to control second messenger pathways. Changes in the tetramer subunits may change the Ca<sup>2+</sup> permeability of ionotropic receptors. For instance, the down regulation of glutamate receptor 2 (GluR2) results in enhanced Ca<sup>2+</sup> influx through AMPA receptors and enhanced death of vulnerable neurons upon brain ischemia [51]. Also, at postsynaptic levels, energy depletion makes neurons more vulnerable, leading to

NMDA receptor-mediated neuronal damage in the presence of normal levels of glutamate [52]. Thus, the excess  $Ca^{2+}$  influx through NMDA and AMPA receptors or Cav channels seems to be the main pathway leading to neuronal injury in excitotoxicity [42,53]. The plausible mechanism underlying neuronal death induced by excess  $Ca^{2+}$  is schematically presented in Fig. 3.

Excess  $Ca^{2+}$  entry causes a sustained elevation of the cytosolic  $Ca^{2+}$  concentration, which elicits the activation of protein kinase C, phospholipases, lipases, endonucleases, proteases, protein phosphatases, nitric oxide synthase, or xanthine oxidase. Furthermore, altered mitochondrial  $Ca^{2+}$  handling and overproduction of reactive oxygen species are two interacting mechanisms that also contribute to excitotoxic cell death [54,55].

#### 2.4. Calcium dyshomeostasis, excitotoxicity and retinal degeneration

As discussed above, excitotoxic neuronal death occurs as a consequence of excess activation by glutamate of NMDARs and AMPARs, as well as the activation of Cav channels (particularly of the L-subtype); this lead to augmented Ca<sup>2+</sup> entry, increased  $[Ca^{2+}]_c$ , mitochondrial Ca<sup>2+</sup> overload, and apoptosis [56,57]. Various experiments suggest the involvement of Ca<sup>2+</sup> overload in retinal damage. Thus, prolonged cytosolic and mitochondrial Ca<sup>2+</sup> overload through these three calcium channels contributes to retinal damage during ischemia [58,59]. This excess Ca<sup>2+</sup> influx activates the calpain-dependent apoptosis pathway and calcineurin activation in retinal pathologies such as glaucoma or retinitis pigmentosa [60–64].

A few experiments suggest the involvement of the L-subtype of Cav channels in excitotoxicity. So, the blocker nifedipine attenuated the slow excitatory amino acid-elicited neurotoxicity [65]. Additionally, on an in vivo rat model of retinal ischemia, pretreatment with nifedipine



**Fig. 4.** Link between oxidative stress, mitochondrial calcium handling, and cell death. C I-C IV: mitochondrial complexes I-IV; IP<sub>3</sub>R: inositol trisphosphate receptor; NOS: nitric oxide synthase; NO: nitric oxide; PCET: plasmalemmal Ca<sup>2+</sup> efflux transporters; ROS: reactive oxygen species; RYR: ryanodine receptor. Images modified from Servier Medical Art, licensed under a Creative Commons Attribution 3.0 Generic License. http://smart.servier.com/.

facilitated the recovery of the b-wave amplitude of the electroretinogram [66]. Also, Cav channel blockers as diltiazem or nivaldipine have shown neuroprotective effects in animal models of RP [67], although the effects of calcium channel blockers on photoreceptor rescue remain controversial. In murine models, cyclosporine A and NIM811, two inhibitors of the mitochondrial permeability transition pore (mPTP), blocked all Ca<sup>2+</sup>-induced apoptosis of rod photoreceptors [68]. Furthermore, the topical "wide-spectrum" Cav blocker flunarizine reduced IOP and protected the retina against ischemia-excitotoxicity [69,70]. Flunarizine also decreased Ca<sup>2+</sup> overload and lipofuscin formation in human RPE cells and maintained cellular vitality, showing its probable clinical relevance in AMD [71].

Brimonidine and other  $\alpha 2$  agonists can protect RGCs under disease conditions by preventing abnormal elevations of  $[Ca^{2+}]_c$  either in RGCs and/or in their presynaptic cells through the modulation of L-type Cav channel activity [72]. The blocking of Na<sup>+</sup> and Ca<sup>2+</sup> influx by beta-blockers as timolol and levobetaxolol is also neuroprotective, as shown in a rat model of ischemia/reperfusion [59,73].

RGCs are sensitive to damage elicited by excessive glutamate [74] due to activation of a  $Ca^{2+}$ -dependent caspase cascade [75]. The observation that intraocular glutamate levels were increased in glaucoma patients [76] raised the hypothesis that blockade of glutamate receptors could protect RGCs; this was shown to be true in a high IOP model in the rat, using the potent NMDAR antagonist MK-801 [77,78]. On the other hand, the reversible NMDAR antagonist memantine afforded robust neuroprotection against glutamate toxicity in RGCs [75, 79]. Furthermore, the peptide somatostatin has a neuroprotective role in the retina against glutamate-induced neurotoxicity and ischemia [80]. Some studies have shown that the activation of somatostatin receptors may protect RGCs by the suppression of T-type  $Ca^{2+}$  currents and the intracellular NO/cGMP/PKG signaling pathway [81]. Moreover, somatostin can regulate intracellular  $Ca^{2+}$  levels via L-type Cav channels [82].

Evidence also points to the involvement of transient receptor potential (TRP) calcium channels in retinal degeneration, first shown in experiments in *Drosophila* [83] and then in mammals, due to their relationship with  $Ca^{2+}$  homeostasis [84]. In glaucoma, the exposure of retinal ganglion cells to elevated pressure can induce an increase of intracellular  $Ca^{2+}$  accumulation due to activation of transient receptor potential vanilloid 1 (TRPV1) and 4 (TRPV4) channels, what leads to apoptosis of RGCs [85–87]. Thus, high extracellular levels of glutamate and the increased  $Ca^{2+}$  entry through NMDARs, AMPARs, Cav channels and TRPV4 channels influence the RGC death [88,89].

An activated unfolded protein response (UPR) and mitochondrial dysfunction characterize the pathology of autosomal dominant RP in rat models. The sustained UPR in the wild-type retina can promote retinal degeneration through increased intracellular  $Ca^{2+}$  and  $Ca^{2+}$ -mediated calpain activation [90,91]. Other studies suggest that  $Ca^{2+}$  accumulation activates photoreceptor death by caspase-dependent apoptosis. Thus, in degenerating photoreceptors of the rd1 model of RP, enhanced intracellular accumulation of  $Ca^{2+}$  was accompanied by augmented calpain activation and apoptosis; congruent with this, calcium channel blockers decreased calpain activation and apoptosis both in vitro and in rd1 mice, suggesting that elevated  $Ca^{2+}$  is required for calpain activation and calpain-induced cell-death [92]. In this context, another study showed that inhibition of calpain reduced photoreceptor death in the rd10 mouse model of RP [93].

Concerning DR, it is known that hyperglicemia is a major risk factor for endothelial dysfunction and vascular disease in diabetic patients. High glucose levels in diabetic patients can influence the influx of  $Ca^{2+}$ in endothelial cells, with an impact on  $Ca^{2+}$ -induced cell death. Some experiments have shown that high glucose levels can increase  $Ca^{2+}$ entry, as that induced by ATP in bovine aortic endothelial cells by enhancing the expression of transient receptor potential canonical 1 (TRPC1) channels [94]. In the macaque cell line RF/6 A of choroid-retinal endothelial cells cultured in hyperglycemic conditions,  $Ca^{2+}$  entry activates  $Ca^{2+}$ /calmodulin-dependent protein kinase II (CaMKII), which contributes to cell apoptosis by activating both Fas-dependent and mitochondria-dependent apoptosis pathways [95].

In summary, evidence supporting the view that neuronal (and mitochondrial)  $Ca^2$  overload lead to apoptotic cell death is overwhelming. In the case of the retina, evidence is accumulating proving that  $Ca^{2+}$  overload is also a pathogenic feature in various experimental models of retinal degeneration and hence the pharmacological mitigation of such  $Ca^{2+}$  overload could drive retinal protection.

# 3. Oxidative stress, mitochondrial dysfunction and neurodegeneration

Again, to better understand the implication of excess oxidative stress in retinal degeneration, we will briefly comment on its concept and link with distorted mitochondrial  $Ca^{2+}$  handling and its role in neuronal death. Then, these concepts will be extrapolated to retinal degeneration.

#### 3.1. Oxidative stress

The concept of oxidative stress has its origin in the pioneering observation that patients with hematological diseases had defective glutathione reductase [96]. Later on, it was demonstrated that in isolated heart mitochondria, the respiratory chain was capable of producing superoxide anion and indirectly hydrogen peroxide [97]. Afterwards, oxidative stress was defined as an imbalance between the biochemical reactions that generate reactive oxygen species (ROS) and those responsible for their removal.

In mammalian cells, ROS are generated in mitochondria, endoplasmic reticulum, cytosol, and the extracellular space. In mitochondria, the most active sites for ROS production are complex I and complex III of the oxidative phosphorylation chain [98]. Mitochondria are not only the major source of ROS; they are also particularly vulnerable to ROS-inflicted damage, particularly mitochondrial DNA. This damage has detrimental functional consequences with excess ROS production and decreased ATP synthesis; this results in bioenergetic failure thereby limiting the energy available for key cellular processes such as neurotransmitter release and repair after tissue injury. This explains that mitochondrial dysfunction contributes to the pathogenesis of neurodegenerative diseases [99] and optic neuropathies [100].

# 3.2. Link between oxidative stress, distorted mitochondrial calcium handling and neuronal death

Conditions that provoke oxidative/nitrosative stress results in Ca<sup>2+</sup> overload and neuronal death [101,102] (Fig. 4). ROS are generated as byproducts of the mitochondrial respiratory chain [103]. On the other hand, there is a positive correlation between the Ca<sup>2+</sup>-dependent metabolic rate and ROS production [104]. Furthermore, Ca<sup>2+</sup> stimulates nitric oxide (NO) by NO synthase (NOS) which blocks complex IV leading to ROS production in complex III [105]. Conversely, ROS regulate several Ca<sup>2+</sup> transporters and signaling proteins of the mitochondria-endoplasmic reticulum communication system [106]. ROS also modulate ryanodine receptors [107] and IP<sub>3</sub> receptors (IP<sub>3</sub>Rs) as well as plasmalemmal and ER Ca<sup>2+</sup> transporters [105] thus contributing to elevated [Ca<sup>2+</sup>]<sub>c</sub> during oxidative stress. Therefore, in neurodegenerative diseases mitochondrial dysfunction is associated to four interacting pathways namely, altered Ca<sup>2+</sup> homeostasis, excessive ROS production, induction of apoptosis, and neuronal death [101,102,108, 1091.

#### 3.3. Oxidative stress and retinal degeneration

Augmented oxidative stress has been implicated in various eye diseases leading to retinal degeneration. In glaucoma patients, NOS and nitrotyrosine are augmented in the trabecular meshwork, which is responsible for draining the aqueous humor in the anterior eye chamber [110]. RGCs are highly sensitive to excess oxidative stress linked to mitochondrial dysfunction in glaucoma [111]. Also, apoptotic RGC death is directly linked to high intraocular pressure; in its turn, death of RGCs increases ROS, leading to optic nerve retinopathy. The influence of mitochondrial misbalance and ROS in the development of the neurodegenerative process is crucial from the early stages of the disease [112–114].

Oxidative damage also contributes to both, the onset and progression of AMD [115,116] and accounts for the main pathological

manifestations, including the dysfunction of RPE and breakdown of the blood retinal barrier [117,118]. During ageing, oxidized polyunsaturated fatty acids (PUFAs) are not efficiently digested and are deposited in RPE cells in the form of lipofuscin, a chromophore involved in ROS formation. Furthermore, cones, which are highly sensitive to free radicals and are the predominant photoreceptors at the macula, have a high demand of oxygen and also contribute to ROS production. With age, the DNA repair system and antioxidant defences decrease, contributing to the increase of accumulated damage [115]. The high local oxygen exposure, together with lifestyle choices as smoking, sunlight exposure or diet leads to oxidative damage in AMD [116]. Many preclinical studies in animal models of AMD have shown a protective effect of antioxidant substances as resveratrol, carotenoids and omega-3 fatty acids (reviewed in [119]).

ROS also contribute to DR initiation and progression. Oxidative stress is a key factor as indicated by the observation that hyperglycemia induces mitochondrial overproduction of such stress [120,121]. With the antioxidant defences impaired in DR, ROS are increased first by the increase of oxidized glucose and mitochondrial overproduction of ROS, but also via other metabolic pathways namely the polyol pathway, the advanced glycation end product (AGE) pathway, and the protein kinase C (PKC) pathway, which have key repercussions in the pathogenesis of DR [120,121].

Oxidative stress has also a main role in inherited retinal degenerative processes; it is increased in the outer retina of RP animal models and in the ocular samples of RP patients. On the other hand, anti-oxidant interventions delay photoreceptor cell death in experimental RP [122]. Also, DHA has shown protection in murine models of light damage [123, 124]. After the rod cell death, oxygen consumption is reduced in RP retinas, resulting in high tissue levels of oxygen in the outer retina. Then, cones exposed to high levels of tissue oxygen begin to degenerate. As oxidative stress spreads, cones dye from the periphery to the central zone of the retina, decreasing the area of surviving cones and diminishing the remaining visual field, ending at the macula, which is the area of highest cone cell density [125]. Furthermore, the oxidative damage is directly involved in microglia activation and the neuroinflammatory process, and anti-oxidant treatments can suppress the activation of microglia [122]. In line with this is a study done in the rd10 mouse model of RP; antioxidants reduced inflammatory markers, suggesting that oxidative stress contributes to the induction of apoptosis [126].

In summary, oxidative stress is a relevant pathogenic pathway leading to neuronal cell death in neurodegenerative diseases; similarly, oxidative stress is implicated in retinal degeneration in the four eye diseases here analyzed. The pharmacological mitigation of oxidative stress rescues retinal cells from death in several cell and animal models of disease.

# 4. Neuroinflammation linked to P2X7 receptors and retinal degeneration

Neuroinflammation associated to glial activation is implicated in practically all neurodegenerative diseases and also in retinal degeneration. We will briefly review this general topic to better understand its introduction in the field of retinal damage.

### 4.1. The P2X7R as a gatekeeper of inflammation

The concept of purinergic signaling mediated by ATP and purinergic receptors arose about 50 years ago [127]. Purinergic receptors are classified as P1 and P2; they are activated by adenosine and ATP, respectively. P2Y receptors are metabotropic and P2X<sub>1-7</sub> receptors are ATP-gated ion channels permeable to Na<sup>+</sup>, Ca<sup>2+</sup>, and K<sup>+</sup>, and to larger molecules up to 1 kDa [128–131]. P2X7Rs activation triggers IL-1 $\beta$  release and is involved in macrophage communication and differentiation [132,133]; this led Francesco di Virgilio to formulate the proposal that P2X7Rs are acting as mediators of chronic inflammatory responses



**Fig. 5.** Implication of P2X7Rs in brain neuroinflammation. ATP released from dying neurons activate P2X7Rs, with intracellular Ca<sup>2+</sup> overload, inflammasome activation and cytolysis. The release of proinflammatory cytokines as TNF- $\alpha$  and IL-1 which promotes further microglia activation, gliosis and cell death. Original images were obtained and modified from Servier Medical Art, licensed under a Creative Commons Attribution 3.0 Generic License. http://smart.servier.com/.



**Fig. 6.** Cells expressing P2X7Rs in the mammalian retina. P2X7Rs are expressed in retinal pigment epithelial cells (RPE), retinal ganglion cells (G), Müller cells (M), microglia (Mi) and astrocytes (As), as well as in the synaptic ribbons of rods (R), cones (C), horizontal (H) and amacrine cells (A), shown in green. RPE, retinal pigment epithelial cell; OS, outer segments; ONL, outer nuclear layer; OPL, outer plexiform layer; INL, inner nuclear layer; GCL, ganglion cell layer; B, bipolar cells.

[133,134]. The concept is supported by the fact that P2X7Rs are expressed in immune and inflammatory cells such as dendritic cells, osteoclasts, mast cells, natural killer cells, and T and B lymphocytes [131].

The implication of P2X7Rs in brain neuroinflammation is supported by their expression in astrocytes and microglia, where they activate the inflammasome. The assembly of the NLRP3 inflammasome complex in microglia is elicited by P2X7R activation by the high ATP concentrations released from damaged neurons; as a result, the secretion of proinflammatory cytokines such as IL-1 $\beta$  takes place [135,136] (Fig. 5).

### 4.2. P2X7Rs, neuroinflammation and retinal degeneration

P2X7Rs are abundantly expressed in several types of retinal cells such as astrocytes, microglia, Müller cells, RPE, or pericyte-containing microvessels [137–139]. They are also expressed in neuronal type cells, including RGCs, horizontal and amacrine cells of the inner nuclear layer, the outer plexiform layer (synaptic sites of rods and cones), and the inner plexiform layer [139–141] (Fig. 6).

Through the physiological activation of ionotropic P2X7Rs, ATP contributes to fast excitatory transmission in the retina [142,143]. This is in line with the observation that P2X7R activation elicits changes in the a- and b-waves of the electroretinogram (ERG) as well as in the oscillatory potentials [139,144–146].

P2X7Rs also play a prominent role in retinal gliosis and retinal degeneration. through two mechanisms: excessive Ca<sup>2+</sup> entry through the P2X7R pore elicits (i)  $Ca^{2+}$  overload and cytolysis [147]; and (ii) activation of the inflammasome-dependent cell death pathway[148]. Excess ATP release from lesioned neurons or glia causes the further activation of microglia, leading to the release of pro-inflammatory cytokines IL-1 $\beta$  and TNF- $\alpha$  this promotes further microglia activation, the propagation of gliosis, with secondary cell death [149,150], and the progression of retinal degeneration [3,148] (Fig. 5). The activation of microglia and the implication of P2X7Rs has been shown to occur in models of retina damage induced by elevated IOP. In a rat model of chronic ocular hypertension, excessive ATP release was accompanied by RGC damage linked to P2X7Rs; by shifting purinergic balance and blocking P2X7Rs, inhibition of microglial activation and protection of RGCs was observed [150]. In another study, the pressure-induced damage of RGCs was prevented by P2X7R blockade, indicating once more the implication of this receptor in glaucoma [151] and the potential of P2X7R blockers as pharmacological protecting targets in the



**Fig. 7.** Number of clinical trials found in the FDA database ClinicalTrials.gov, done in patients with diabetic retinopathy (DR), age-related macular degeneration (AMD), glaucoma, and retinitis pigmentosa (RP). A, total number of clinical trials (CTs). B, CTs with small molecules. C, CTs with small molecules that target Ca<sup>2+</sup> dyshomeostasis/excitotoxicity, oxidative stress (OS), P2X7Rs activity-linked neuroinflammation (P2X7R) or other targets outside the scope of this review.

disease. Other studies have shown the activation of microglia, and hence, the presence of neuroinflammation in this disease [152]. In a mouse model of chronic glaucoma with early microglia activation, anti-inflammatory microglia improved optic nerve integrity and reduced retinal microglial activation [153]. Also, high IOP induced the upregulation of retinal GFAP and MHC-II, as well as microglia reactivity in mice retina [154] and reactive nonproliferative gliosis in a mouse model of glaucoma [155]. Glial toll-like receptors (TLRs) have also been implicated in stressed glaucomatous tissue [156]. Increased secretion of inflammatory proteins such as TNF- $\alpha$  from glaucomatous tissues and upregulation of their expression [152] is congruent with the observation that inhibition of reactive microglia with anti-inflammatory minocycline augmented RGC survival [157,158]. In this frame, the inhibition of signaling cascades such as NO synthase or TNF-a initiated in reactive microglia, are potential therapeutic strategies in glaucoma [158–160]. Minocycline has also shown protection against inflammation-induced photoreceptor cell death in murine models of inherited retinal degeneration [161] and light damage [162,163]. Furthermore, as in the human disease, immunostimulatory signaling can also be triggered by glial TLRs; this could also become another therapeutic target.

In AMD, calcium dyshomeostasis can be mediated through the activation of P2X7Rs, that could cause the dysfunction and apoptosis of RPE. Yang et al. (2011) showed that in human RPE cells both the nonselective endogenous agonist ATP and the selective P2X7R agonist 2,3-O-(4-benzoylbenzoyl)-ATP (BzATP) can increase intracellular Ca<sup>2+</sup> via extracellular Ca<sup>2+</sup> influx and induce apoptosis. This Ca<sup>2+</sup> increase was significantly inhibited by the P2X7R antagonist oxidized ATP (oATP) but not by the P2 receptor antagonist suramin, inhibited by BAPTA-AM and also by low extracellular Ca<sup>2+</sup> [164]. P2X7Rs are expressed in the RPE and different layers of the retina and are functional both in vitro and in vivo, thus mediating AMD-like defects [165]. P2X7Rs expressed in microglia and macrophages normally function as scavenger receptors; so, functional alterations in this process can predispose individuals to AMD [166]. Moreover, one of the main components of lipofuscin is the bisretinoid N-retinyl-N-retinylidene ethanolamine (A2E), which cannot be enzymatically degraded, and it is accumulated in aged RPE cells. On the other hand, microglial cells in culture can internalize A2E and change it into an activated pro-inflammatory phenotype [167].

Concerning DR, it is notable that elevated glucose levels increase the extracellular ATP concentration, which activates the P2X7 receptors (P2X7Rs) and evokes the release of proinflammatory cytokines IL-6, TNF- $\alpha$ , and CCL2 in primary microglia [168]. In DR, purinergic-elicited vasotoxicity influences the death of microvascular

pericytes and endothelial cells [169]. In a rat model of diabetes induced by streptozotocin, it was shown that P2X7R antagonists caused the reversion of increased vascular permeability, VEGF accumulation, and IL-6 expression [170]. In diabetic retinas, Müller cells show an increased expression of CD40 and P2X7Rs. CD40 ligation causes phospholipase C- $\gamma$ 1 (PLC- $\gamma$ 1) activation, and secretion of extracellular ATP by Müller cells, which binds to P2X7Rs leading to secretion of TNF- $\alpha$  and IL-1 $\beta$ . The CD40-ATP-P2X7R pathway is linked with the programmed cell death of bystander retinal endothelial cells, what likely contributes to degeneration of capillaries [171].

Inflammation also has a central role in RP. In fact, infiltration of the degenerating regions of the retina by microglia and blood-derived macrophages is present in rd1 and rd10 retinal degeneration mice as well as in post-mortem samples from RP patients [172–174]. Accordingly, P2X7Rs also seem to have a role in RP. In this sense, the intravitreal injection of the purinergic antagonist pyridoxal-phosphate-6-azophenyl-2,4-disulfonic acid (PPADS) protected against ATP-mediated apoptosis in rats with intravitreal injection of high concentrations of ATP. This antagonist also increased the photoreceptor survival in the rd1 mouse [175].

In summary, both in neurodegenerative diseases and in retinal degeneration neuroinflammation mediated by P2X7R activation is a major cause of neuronal death and retinal cell damage; thus, in some studies the pharmacological blockade of P2X7Rs rescue retinal cells from death in various models of eye diseases. This suggests that this receptor is a good target to develop antagonists with potential neuroprotective effects in eye diseases leading to retinal degeneration.

# 5. Pharmacological targets addressed in clinical trials in patients with eye diseases that lead to retinal degeneration

In a search of clinical trials (CTs) at the US Food and Drug Administration (FDA) database ClinicalTrials.gov, the total number of registered studies are as follows: DR 611, AMD 1346, glaucoma 1099, and RP 178 (Fig. 7A) [176]. Some studies are done with large molecules genetic approaches. Many others are done with small molecules: 350 in DR, 1380 in AMD, 1060 in glaucoma, and 20 in RP (Fig. 7B). The number of small molecules directed to targets linked to the three pathogenic pathways of retinal degeneration here discussed (calcium dyshomeostasis, oxidative stress, and neuroinflammation) are considerably smaller (Fig. 7C). Thus, in DR, AMD, and RP there are 13, 7 and 6 studies addressing  $Ca^{2+}$  dyshomeostasis respectively. Targets associated to oxidative stress are present in 11, 8 and 5 studies in DR, AMD and RP respectively. On the other hand, the number of CTs addressing

#### Table 1

Main pharmacological targets in clinical use or under evaluation in the treatment of glaucoma, AMD, DR and RP and example drugs.

| Disease  | Targets                         | Drug  |
|----------|---------------------------------|---|
| Glaucoma | $\beta$ -adrenergic blockers    | Timolol   |
|          | $\alpha$ 2-adrenergic agonists  | Brimonidine                                     |
|          | Cholinergic agonists            | Pilocarpine                                     |
|          | Prostaglandin analogs           | Latanoprost                                     |
|          | Carbonic anhydrase inhibitors   | Brinzolamide                                    |
|          | Rho-kinase inhibitors (under    | Riparsudol                                      |
|          | evaluation)                     |   |
|          | Adenosine receptor agonists     | Trabodenoson                                    |
|          | (under evaluation)              |   |
|          | Modified prostaglandin analogs  | Latanoprostende bunod                           |
|          | (under evaluation)              |   |
|          | Combined antioxidants (under    | $\alpha$ -tocoferol, Vitamins A, B, E,          |
|          | evaluation)                     | Ginkgo biloba                                   |
| AMD      | Anti-vascular endotelial growth | Antibodies: ranibixumab                         |
|          | factor (VEGF) therapy,          | Antibody fragments:                             |
|          | sometimes combined with         | brolucizumab                                    |
|          | corticosteroids (dexamethasone) | Fusion proteins: conbercept                     |
|          | Antibodies anti-Angiopoietin-2  | Faricimab                                       |
|          | (under study)                   | - 1 11  |
|          | Anti-Platelet derived growth    | Pegpleranib                                     |
|          | Antibody anti antigratina 1     | Cononaizumah                                    |
|          | phosphate (under study)         | Sonepcizuniab                                   |
| DR       | Corticosteoids                  | Triamcinolone                                   |
|          | Antioxidants, monotherapy or in | Combination of lutein,                          |
|          | combination (under study)       | $\alpha$ -tocoferol, $\beta$ -carotene, niacin, |
|          |                                 | ascorbic acid, zinc, selenium,                  |
|          |                                 | copper, and manganese                           |
| RP       | Dietary supplements with        | DHA, Vitamin A                                  |
|          | antioxidants (under study)      |   |

### Table 2

Examples of successful drug repositioning. The generic drug name is shown together with the original indication, the new indication that arose from repositioning programs, and the reference.

| Drug               | Original indication  | New Indication           | Reference |
|--------------------|----------------------|--------------------------|-----------|
| Amantadine         | Influenza            | Parkinson's disease      | [236]     |
| Aspirin            | Pain, fever,         | Coronary artery disease  | [237]     |
|                    | inflammation         |                          |           |
| Amphotericine<br>B | Fungal infections    | Leishmaniasis            | [238]     |
| Bromocriptine      | Parkinson's disease  | Diabetes mellitus        | [239]     |
| Bupropion          | Smoking cessation    | Depression               | [240]     |
| Celecoxib          | Osteoarthritis,      | Breast and colon cancer  | [241]     |
|                    | reumathoid arthritis |                          |           |
| Duloxetine         | Depression           | Urinary incontinence     | [242]     |
| Finasteride        | Prostatic            | Male androgenic alopecia | [243]     |
|                    | hypertrophy          |                          |           |
| Fluoxetine         | Depression           | Premenstrual dysphoria   | [244]     |
| Galantamine        | Recovery from curare | Alzheimer's disease      | [245]     |
|                    | in anesthesia        |                          |           |
| Gemcitabine        | Viral infections     | Cancer                   | [246]     |
| Methotrexate       | Cancer               | Reumathoid arthritis,    | [247]     |
|                    |                      | psoriasis                |           |
| Minoxidil          | Hypertension         | Male androgenic alopecia | [248]     |
| Paclitaxel         | Cancer               | Prevention of coronary   | [249]     |
|                    |                      | restenosis               |           |
| Raloxifene         | Prostate and breast  | Postmenopausal           | [250]     |
|                    | cancer               | osteoporosis             |           |
| Ropinirole         | Hypertension         | Parkinson's disease      | [251]     |
| Sildenafil         | Chest angina         | Erectile dysfunction     | [252]     |
| Thalidomide        | Hyperemesis          | Multiple myeloma         | [253]     |
|                    | gravidarum           |                          |           |
| Topiramate         | Epilepsy             | Binge eating disorder    | [254]     |
|                    |                      | associated with obesity  |           |
| Zidovudine         | Cancer               | AIDS                     | [255]     |

neuroinflammation linked to P2X7Rs are scarcely represented, i.e. 2 studies in each DR or AMD. In glaucoma, only 2 studies addressing oxidative stress or  $Ca^{2+}$  dyshomeostasis were found. Therefore, most

studies in DR (27), AMD (90), glaucoma (8), and RP (2) address other targets. We next comment on CTs and drug targets addressed in glaucoma, AMD, DR, and RP. Main targets and drugs in clinical use or under development are summarized in Table 1.

### 5.1. Clinical trials and drug targets in glaucoma

Topical drug options for the lowering of IOP in glaucoma include beta-adrenergic blockers (timolol), alpha-2 adrenergic agonists (brimonidine), cholinergic agonists (pilocarpine), prostaglandin analogs (latanoprost), or carbonic anhydrase inhibitors (brinzolamide); these drugs act either by increasing aqueous humor outflow and/or by decreasing its production. Several large, controlled, and randomized CTs have demonstrated the efficacy of reducing IOP to prevent glaucoma development from ocular hypertension [177] as well as disease progression in patients with established glaucoma [178]. One meta-analysis summarized 28 CTs of topical monotherapy for glaucoma [179] and a second one analyzed 41 CTs of fixed drug combinations for topical treatment of glaucoma [180]. More recently, novel candidates for treatment of primary open-angle glaucoma are being developed in clinical trials; some of them are rho kinase inhibitors (ripasudol, netarsudil, AMA0076, AR-12286, DE-104, SNJ-1656), adenosine receptor agonists (trabodenoson), and modified prostaglandin analogs (latanoprostene bunod, ONO-9054) [176,181,182].

Guidelines by the American Academy of Ophthalmology recommend the use of adrenergic beta-blockers and prostaglandin (PG) analogs as first-line treatment of high IOP. This is not always effective in attenuating disease progression in glaucoma. Thus, other drugs acting directly on the optic nerve and on the process of retinal degeneration can provide additional protection [183].

Adrenergic alpha-2 receptors are located both in ciliary processes and RGCs. These are the major cell types affected in glaucoma, and thus, the  $a^2$  agonist brimonidine seems to produce both lowering of IOP through reduction of aqueous humor formation and neuroprotection by acting on  $a^2$  receptors at RGCs [184]; in fact, one CT found less visual field loss with brimonidine compared with timolol [185]. But the effects of brimonidine also include the upregulation of brain-derived neurotrophic factor expression, inhibition of glutamate release, modulation of NMDA receptors, and regulation of cytosolic Ca<sup>2+</sup>, that probably accounts for its final effect. In this sense, targeting of NMDA receptors has been another strategy to treat glaucoma patients. However, the reversible NMDAR antagonist memantine failed to demonstrate a significant effect on disease progression (cited by [183]).

CTs with combined antioxidants (mainly polyphenol derivatives) reported some benefits in glaucoma. This was the case of mexidol®,  $\alpha$ -tocoferol, or combined vitamins A, B, and E (reviewed by [186]). *Ginkgo biloba*, that exhibits vasoregulatory, neuroprotective, and antioxidant effects can be found in up to 8 CTs, alone or in combination with antioxidants as  $\alpha$ -tocopherol [176].

In summary, most CTs and therapeutic strategies in glaucoma are directed towards the reduction of IOP and very few studies focus on direct pharmacological protection of the retina in patients with glaucoma.

#### 5.2. Clinical trials and drug targets in age-related macular degeneration

Most therapies in AMD focus on the blockade of the proliferation and survival of subretinal neovascular membranes [187]. This is achieved through the intravitreal administration of monoclonal antibodies against vascular endothelial growth factor (anti-VEGF). Three of them are in clinical use, namely ranibizumab, bevacizumab, and aflibercept. Intravitreal implants of the corticosteroids dexamethasone and triamcinolone are also used as adjunct treatments, combined with anti-VEGF.

These medications have shown to be safe and efficacious to reduce severe visual impairment. Despite this, many patients continue losing vision, impairing their abilities to read, drive and perform daily



**Fig. 8.** An in vitro experimental example and a scheme on the synergic effects of drug combinations. A, cell death elicited by oxidative stress of mitochondrial origin, by exposing SH-SY5Y cells to combined rotenone plus oligomycin (R/O). Cell viability (MTT reduction in %, ordinate) showed a 30% cell loss. Neither galantamine (G) nor melatonin (M) at low concentrations, elicited neuroprotection. However, the combination of both drugs exerted a significant neuroprotection. B, scheme showing the desired pharmacological activity and the toxic effect of high drug concentrations (left) of three drugs (d1, d2, d3) and the synergistic effects of activity, but not on toxicity, of combined drugs 1, 2 and 3 at lower concentrations (right). (A) Adapted from [259]. (B) Adapted from [261].

activities [188]. Furthermore, the therapeutic regimens are cumbersome as due to the relatively short half-life of anti-VEGF, repeated intravitreal injections have to be done monthly. This requires frequent visits to the hospital with concomitant loss of time and productivity [189]. Thus, new therapies are needed for neovascular AMD [190].

Various approaches that are being investigated mostly have the VEGF (and other secondary growth factors) as drug targets. The objective is related to the extension of the half-life of anti-VEGF so that the therapeutic regimen can be reduced to, for instance, quarterly intravitreal injections of anti-VEGF. This was the case of conbercept, a VEGF trap-like molecule that blocks all isoforms of VEGF-A, VEGF-B and VEGF-C [191]. Of interest is brolucizumab, a single-chain antibody fragment that inhibits VEGF-A; it displays similar efficacy than other anti-VEGF but injection frequency is also quarterly [192].

Other drugs acting at different but complementary targets to VEGF are also in clinical development. This is the case for the anti-platelet derived growth factor (anti-PDGF) pegpleranib, combined with anti-VEGF; however, results of a phase III CT gave disappointing outcomes [193]. Another target tested was angiopoietin 2 (Ang-2); faricimab is an antibody that binds to Ang-2 and VEGF-A. A phase II CT with 12- and 16-weeks regimens compared well with monthly treatment with ranibizumab, as far as efficacy was concerned [194]. Of note is an ingenious device to release ranibizumab by passive diffusion from a refilled device, surgically-implanted in the pars plana of the eye. A phase II CT demonstrated that patients with the so-called port delivery system (100 mg/ml) showed visual outcomes comparable to those of monthly 0.5 mg ranibizumab; 79.8% of the patients did not require a refill for 6 months [195].

An alternative target being explored is sphingosine-1-phosphate (S1P) that induces significant capillary formation, acting as a proangiogenic factor that directly interacts with other growth factors including epidermal growth factor (EGF) and VEGF receptors. Thus, a monoclonal antibody raised against S1P may be an effective therapy for AMD [196]. Furthermore, reverse transcriptase inhibitors that block the inflammasome and abort the inflammatory process, are also being explored [197]. Gene therapy based in the intravitreal injection of adenovirus carrying a gene expressing a VEGF inhibitory protein has shown limited efficacy so far [198]. Worth mentioning are some studies on inhibitors of tyrosine kinases such as valatanib and pazopanib that do inhibit VEGF receptor subtypes. However, CTs have given negative outcomes [199]. Finally, RNA interference to silence VEGF-A messenger RNA is also being investigated.

Concerning other therapeutic strategies, only a few CTs have assayed

molecules with antioxidant properties, some of them also with effect on cytosolic Ca<sup>2+</sup> modulation, as  $\alpha$ -lipoic acid, the antibiotic doxycycline, the immunosuppressant rapamycin or the vasodilator MC-1101, which has also an anti-inflammatory and antioxidant profile. Molecules with multiple mechanisms of action as metformin or brimonidine have also been assayed in CTs to test their neuroprotective potential.

In summary, all therapeutic strategies being applied to patients with AMD, and many other that are in clinical development, focus on the mitigation of the macular neovascularization, mostly by suppressing growth factors with specific monoclonal antibodies intravitreally applied, particularly anti-VEGF antibodies. Targets on retina cells themselves to afford retinal cell protection are scarcely being investigated.

#### 5.3. Clinical trials and drug targets in diabetic retinopathy

In trying to delay DR onset and to mitigate disease progression once diagnosed, ophthalmologists do give priority to the adequate control of diabetes mellitus (DM); therapy focus on control of blood glucose and lipids as well as blood pressure, with diet and specific medications [200]. Once diabetes is stabilized, DR treatment is based in standard therapies such as photocoagulation, vitrectomy, or intraocular injections of corticosteroids or anti-VEGF monoclonal antibodies. However, some patients are resistant to these treatments, and hence they suffer DR progression [112]; this has been associated mainly to oxidative stress [201]. Thus, complementary therapeutic approaches focusing on the rebalance of excess free radical production and/or restoration of antioxidant natural systems, have been and are being tested in CTs. Both specific single antioxidant compounds or various antioxidants given in combination are tested [112]; also, diet/nutraceutical supplementation is being studied [202].

A recent comprehensive review comments on 31 CTs done in patients with DR; those studies have been performed with single antioxidant compounds or with combined antioxidant therapy (CAT) [112]. In spite of the large heterogeneity of studies (dosing, use of single antioxidant or CAT, treatment duration), some positive outcomes emerged. For instance, a long-lasting study (5 years) evaluated the effect of CAT (lutein, alpha-tocopherol, niacin, beta-carotene, ascorbic acid, zinc, selenium, copper, and manganese) in 105 patients with DM2 and non-proliferative DR. Authors found a retardation of DR progression but no effect on visual acuity; the antioxidant plasma status was maintained [203]. Another CT with a complex CAT formula (18 antioxidant ingredients!) showed improved visual acuity but no changes in retinal

### Table 3

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Small molecules that have been, or are being tested individually in clinical trials, acting on one of the three common pathogenic pathways of neurodegenerative diseases and retinal degeneration: calcium overload/NMDARs-AMPARs/Cav channels/excitotoxicity, mitochondrial dysfunction/oxidative stress, and neuro-inflammation/microgliosis/P2X7Rs. AD: Alzheimer disease; ALS: amyotrophic lateral sclerosis; AMD: age-related macular degeneration; CT: clinical trial; GA: geographic atrophy; MS: multiple sclerosis; PD: Parkinson disease; RP: retinitis pigmentosa.

| Pathogenic pathway                              | Small molecule  | Mechanism of action  | References  |
|---|---|--|---|
| Calcium overload / NMDA-                        | Brimonidine   | Blocks NMDA receptors; decreases intracellular Ca <sup>2</sup> *.  | [270] Neuroprotective effects in RP patients, after topical administration  |
| AMPA receptors-Cav<br>channels/ excitotoxicity  | Cannabis (cannabidiol:<br>THC, 1:1)<br>Ceftriaxone<br>Curcumin  | Inhibits L-type calcium channels, TRPV1, TRPA1<br>(activates and desensitizes TRP)<br>Reduces glutamatergic hyperactivity.<br>Modulates HVGCC, CRAC, VGCC (Cav2.2, Cav 2.1).<br>Blocks Ca <sup>2+</sup> channels. Blocks Ca <sup>2+</sup> mobilization | administration<br>[271] Early phase I CT for RP. Well tolerated by most<br>patients<br>[176,272] Phase III CT in ALS<br>[273–278] Well tolerated. Poor bioavailability. Low<br>bioavailability, which can be increased with several<br>formulations, as well as safety, tolerability, and efficacy. |
|   | DHA   | Reduces endothelial calcium influx.  | Phase II CTs in AD. Phase I CTs for AMD, Phase II in<br>combination with resveratrol and quercetin<br>[216,279,280] Phase II CT in RP. No effects in slowing RP<br>progression were detected  |
|   | Empagliflozin   | Reduces calcium dysregulation; reduces Ca <sup>2+</sup> /<br>calmodulin-dependent kinase II  | [281] May prevent preclinical DR with metformin   |
|   | Everolimus<br>Homotaurine                                       | Improved endothelin-induced calcium flux<br>Modulates ER stress, Ca <sup>2</sup> * homeostasis   | [282,283] Phase II CTs in AMD<br>[284,285] In combination with vitamin E and citicoline<br>increases contrast sensitivity in glaucoma patients  |
|   | Isradipine<br>Lutein  | Blocks L-type Cav channels<br>Inhibits TRP   | [286,287] Did not show efficacy in Phase III CT in PD<br>[288] Lutein serum levels are associated with decreased<br>AMD progression   |
|   | Memantine   | Non-competitive blockade of NMDARs   | [289–293] Approved for the treatment of AD. Phase II CT<br>in PD. Phase III CTs did not prevent glaucoma progression  |
|   | Metformin   | Reduces calcium influx   | [294,295] Retrospective studies associate metformin<br>with decreased risk of AMD   |
|   | Mynocicline<br>Nilvadipine                                      | Chelates Ca <sup>2</sup> * decreases calcium influx<br>Blocks L-type Cav channels  | [296–298] Phase II CT in AMD and RP<br>[219,299] No benefits observed in Phase III CTs in mild to<br>moderate AD. Phase II CT shows that could retard<br>progression of visual field defect in RP   |
|   | Rapamycin   | Binds to calcium channels and FKBP52, affects intracellular calcium signaling  | [300] Phase II CT in AMD. Well tolerated<br>subconjunctival. No benefits observed in Phase I/II CTs<br>subconjunctival nor intravitreal   |
|   | Riluzole  | Blocks glutamatergic neurotransmission   | [301,302] Phase II CT in MS, delays progression. To see if<br>it also delays retinal damage   |
|   | Somatostatin  | Blocks L- and T-type $\mathrm{Ca}^{2*}$ channels, inhibits calcium influx  | [270] Neuroprotective effects in RP patients, after topical<br>administration. Phase II CT  |
|   | Talampanel<br>Ubiquinone, CoQ                                   | Non-competitive AMPA antagonist<br>Suppresses voltage-dependent calcium influx and MAPK<br>signaling   | [303,304] Failed in Phase II CTs in ALS patients<br>[305-308] Safe and well tolerated. Administered<br>topically improves RGC loos in AD patients. Good clinical<br>outcomes in phase II CT in DR   |
|   | Unoprostone   | Inhibits calcium influx. Activates large conductance $Ca^{2*}$ -activated K* (BK) channels   | [309,310] Well tolerated topically. Increased macular sensitivity in RP patients. Phase II CT for RP  |
| Mitochondrial dysfunction /<br>oxidative stress | 9-cis β Carotene  | Scavenger  | [203,311–313] Well tolerated orally. Poorly absorbed.<br>Phase II CT for RP   |
|   | Alpha lipoic acid<br>BN82451B                                   | Inhibits p38MAPK pathway. ROS scavenger<br>Several mechanisms proposed: socium channel blocking<br>potential (antiexcytotoxic), antioxidant, anti-<br>inflammatory (COX inhibitory potential), and<br>mitochondrial protective                         | [314,315] Well absorbed orally. Negative outcomes in GA<br>[316] Poorly tolerated. Phase I CT in Huntington disease   |
|   | Creatine<br>Curcumin  | Antioxidant. Stabilizes mitochondrial membranes. Also<br>stimulates glutamate uptake into synaptic vesicles<br>Regulates ROS and mitochondrial depolarization.<br>Decreases oxidative stress   | [317,318] Good safety profile and well tolerated. Phase II<br>CT for ALS<br>See above   |
|   | Doxycycline<br>Edaravone  | Scavenges ROS<br>Free radical scavenger  | [319–321] Phase II CT in DR. Well tolerated<br>[322,323] Approved by FDA for ALS. Phase II CT for<br>alcohol-induced brain injury. Phase II CT for acute<br>ischemic stroke patients.   |
|   | Ethyl eicosapentaenoic<br>acid (EPA)                            | Free radical scavenger   | [324] Phase II CTs for AD. Phase III for AMD (DHA/EPA)  |
|   | Finerenone  | Decreases oxidative stress, atenuates endothelial dysfunction  | [325] Observational study with DR patients  |
|   | Hydroxycloroquine   | Inhibits calcium channels  | [326,327] Preliminary promising results for EM, not for AD  |
|   | Metformin<br>Lutein<br>MitoQ<br>Minocycline<br>N-acetylcysteine | Inhibits Complex I. Redox regulator<br>Decreases oxidative stress.<br>Antioxidant that supports mitochondrial function<br>Radical scavenger<br>Antioxidant   | <ul> <li>[328] Phase I CT for MS. Phase I DR. Phase II CT for AMD</li> <li>[329] Phase II CT in RP</li> <li>[330] Phase II CT in MS and PD</li> <li>[331] Phase II CT in RP and AMD</li> <li>[332,333] Phase II CT in RP. Safe administered orally, but results are still inclonclusive</li> </ul>  |
|   | Pioglitazone<br>Pridopidine                                     | $\ensuremath{\text{PPAR}}\ensuremath{\gamma}$ agonist Sigma-1 receptor agonist. Enhances mitochondrial function  | [334] Phase III CT in patients with Friedreich's Ataxia<br>[335,336] Phase II CT in patients with Huntington's<br>disease. Well tolerated. Still not conclusive data.   |
|   |   |  | (communed on next page)   |

#### Table 3 (continued)

| Pathogenic pathway                                   | Small molecule              | Mechanism of action   | References  |
|--|-----------------------------|---|---|
|  | Rapamycin<br>Ruboxistaurine | Decreases oxidative stress Inhibits PKC $\beta$ and reduces oxidative stress  | See above<br>[337] Tested in CTs for indications related to diabetes,<br>including DB   |
|  | Sulodexide                  | Antioxidant. Preserves endothelial function   | [338,339] Prevents cognitive impairment in AD. Tested   |
|  | Ubiquinone,<br>Coenzyme Q10 | Quenching of free radicals  | In DR for the treatment of hard exulates<br>[340,341] Phase II CT in DR suggest therapeutic efficacy.<br>Tested in several brain disorders as AD, PD, MS, epilepsy,<br>depression. Well telerated, pacificible side effects |
|  | Urate                       | Antioxidant (given as inosine)  | [342] Good profile of safety and tolerability. Negative<br>outcomes   |
|  | Zeaxanthin                  | Decreases OS  | [343,344] Encouraging results in AMD and glaucoma after dietary supplementation   |
| Neuroinflammation /<br>microgliosis / P2X7 receptors | AZD3241                     | Inhibitor of myeloperoxidase  | [345] Tested in patients with PD and Multiple System<br>Atrophy, Safe and well tolerated  |
|  | AZD9056                     | Blocker of P2X7Rs   | [346] CT in patients with rheumatoid arthritis, with<br>negative outcomes but good tolerability.  |
|  | Celecoxib                   | COX inhibitor   | [347] Phase III CTs in Tested in AD. Initially no good<br>outcomes  |
|  | Curcumin                    | Decreases P2X7R expression and activation.<br>Represses NLRP3 inflammasome activation via P2X7<br>activation                        | [275] See above   |
|  | Exenatide                   | Agonist of glucagon-like peptide 1 (GLP-1)  | [348,349] Phase II CTs in PD and AD. Initial positive   |
|  | Glatiramer acetate          | Immunomodulator   | outcomes. Ongoing phase III CTs in PD<br>[350,351] Approved by FDA for MS. Phase III CTs in AMD<br>with no formal report of results   |
|  | Ibuprofen                   | COX inhibitor   | [352] Phase IV CTs in patients with AD  |
|  | JNJ-55308942                | Blocker of P2X7Rs   | [176] 2020. Phase I CTs; good safety profile  |
|  | JNJ-54175446                | Blocker of P2X7Rs   | [176] 2020. Phase II CTs; in depression; negative outcome but good safety profile   |
|  | Laquinimod                  | Down-regulates proinflammatory cytokines and up-<br>regulates anti-inflammatory citokines   | [353,354] Phase II CTs in MS. Generally, well tolerated.<br>Initial good results  |
|  | Metformin                   | Inhibits P2X7R  | See above   |
|  | Minocycline                 | Inhibition of ROS production  | See above   |
|  | Naloxone                    | Inhibits retinal microglial activation  | [355] No beneficious initial results in AD as monotherapy   |
|  | Naproxen                    | COX inhibitor   | [356] Phase III CTs in Tested in AD. Initially no good<br>outcomes. Initial disappointing outcomes AD   |
|  | Pioglitazone                | PPARy agonist   | [334] Phase IV CTs in DR. No results officially posted  |
|  | Saffron                     | Saffron reduces ATP-induced retinal cytotoxicity by<br>targeting P2X7 receptors. Anti-apoptotic, anti-<br>inflammatory, antioxidant | [357,358] Results suggest possible therapeutic potential<br>in retinal degenerative pathologies. Phase I CTs for AMD  |
|  | Valproic acid               | Inhibits histone deacetylase and autophagy  | [359] Heterogeneous clinical repots in RP. Not<br>beneficious for all genotypes.  |

thickness [204]. In contrast, the supplementation for 3 months with the CAT formula in patients suffering from DM1 or DM2 showed an improvement of visual acuity, contrast sensitivity, and foveal thickness [205].

Another interesting approach has been the association of CAT with anti-VEGF therapies. So, in a study on 55 DM2 patients with diabetic macular edema treated with CAT and ranibizumab for 3 years, authors found lower macular thickness in the group supplemented with antioxidants, compared with control patients [206]. An additional effective combination was fasudil (a rho kinase inhibitor) and anti-VEGF bevacizumab; intravitreal injection of both agents improved visual acuity in patients with diabetic macular edema [207].

Various antioxidants (flavonoids, coenzyme Q, N-acetylcysteine, vitamin C, calcium dobesilate) have been studied in CTs, with erratic controversial results (see [112,176]). Two recent CTs with antioxidant saffron-derived compounds (safranal, crocetin, and dimethylcrocetin) show promising early data for the treatment of diabetic maculopathy [208]. A recent CT has shown a synergistic effect of the co-administration of curcumin, homotaurine, and vitamin D3, decreasing soluble mediators of inflammation and retinal damage, so suggesting the potential benefits of a multi-target strategy [209]. A few molecules with effect on cytosolic Ca<sup>2+</sup> levels have been tested in CTs against AMD. This is the case of  $\alpha$ -lipoic acid, empagliflozin, rapamycin, or ubiquinone, all of them with other mechanisms of action [176].

In summary, CTs with single or combined antioxidant drugs in DR patients have provided controversial results. However, some of them are

promising, particularly those using combined antioxidants. Long-term studies should consider the efficacy and safety of antioxidants at different doses; this is hard to test with complex formulations containing several compounds. Thus, although there is a tendency to use antioxidants in complex combinations, "cleaner" CTs should be performed with single antioxidants or, at most, with 2–3 combined antioxidants.

#### 5.4. Clinical trials and drug targets in retinitis pigmentosa

An interesting research line focus on advanced therapies in RP; these include gene replacement, genome editing, and stem cells treatments [210–213]. As these treatments might take time in reaching the clinic, focus is being placed in pharmacological approaches to target pathways leading to cell death, in order to delay disease progression [214]. Dietary supplements with antioxidant properties such as vitamin A, do-cosahexaenoic acid (DHA), and lutein are the prescribed antioxidants in RP. Concerning vitamin A, early CTs reported a protective effect at high doses [215]. In another CT, combined DHA plus vitamin A improved photoreceptor survival [216]. A more recent CT showed a slowing in the progression of visual field loss in patients with X-linked RP, treated with oral daily supplement of DHA for 4 years [217]. Also, a CT in 34 RP patients treated with lutein supplementation for 6 months showed positive clinical outcomes in preserving the central visual field [218].

Long-term treatment with the blocker of L-type  $Ca^{2+}$  channels nilvadipine, improved average visual field sensitivity in RP patients. Authors of this study suggested that further improvement could be



**Fig. 9.** Scheme showing the proposed combination of three drugs (triads) acting specifically and selectively on each one of the three pathogenic targets that are common to DR, AMD, glaucoma, and RP. F1: drug blocking NMDA channels and reducing Ca<sup>2+</sup> overload and excitotoxicity; F2: antioxidant drug, reducing oxidative stress; F3: anti-inflammatory drug blocking P2X7Rs. Some images were obtained and modified from Servier Medical Art, licensed under a Creative Commons Attribution 3.0 Generic License. http://smart.servier.com/.

#### Table 4

Proposed triads of repositioned medicines that at low doses may exhibit synergistic retinal protective activity with low toxic effects in retinal degenerative diseases.

| Triad   | Excitotoxicity | Oxidative stress | Neuroinflammation |
|---------|----------------|------------------|-------------------|
| Triad 1 | Memantine      | Pioglitazone     | Mynocicline       |
| Triad 2 | Nivaldipine    | Edaravone        | Naloxone          |
| Triad 3 | Ceftriaxone    | MitoQ            | Valproic acid     |

achieved with the combination of nilvadipine and an antioxidant [219]. Several molecules with effects on cytosolic  $Ca^{2+}$  levels have been explored, as cannabidiol (as a plant extract) and also minocycline and ubiquinone, both of them with antioxidant properties too [176].

In summary, targeting oxidative stress with antioxidant supplementation has been the main focus in CTs for RP. Calcium dyshomeostasis has been explored in at least one study with a blocker of L-type Cav channels. Finally, in neuroinflammation some studies focused on chemokine receptor inhibitors; however, no CTs have yet been done with P2X7R blockers to mitigate the neuroinflammatory pathogenic pathway of RP.

### 6. Multi-target therapy approach to combat chronic diseases

Here, we will discuss the first basis for therapeutic combinations of drugs acting on different but complementary pathogenic signaling pathways in various chronic diseases. We will then extrapolate these concepts to the treatment of retinal degeneration with such drug combinations.

# 6.1. The "one-target-one-drug" paradigm versus the "multitarget-drug combination" paradigm

The biological definition of a key target in the pathogenic cascade of a given disease led medicinal chemists to strategically design a molecule that interacts with such target with certain degree of selectivity; in doing so, such molecule could become a potential therapeutic candidate for clinical trials, and eventually it may reach the status of a new medicine to treat such disease. Unquestionably, this "one target-one compound" paradigm has been highly successful in the past; for example, in betablockers for coronary artery disease, cyclosporine as immunosupressant in organ transplantation, or proton pump inhibitors in acid gastroduodenal diseases. During the second half of the XX century, hundreds of successful drug therapies have been introduced in the clinic following this research paradigm. More recently, genomic and proteomic studies have revealed new potential targets that gave rise to highly selective medicines essentially based on monoclonal antibodies for inflammatory and cancer diseases. Precisely, these and many other chronic diseases such as cancer [220], hypertension [221], asthma [222] and AIDS [223] are better controlled with combinations of drugs with different but complementary mechanisms of action associated to various targets of the disease.

This multi-target approach is grounded on various facts. One of them is that most cells have a sort of "back-up" systems leading to the same final outcome, namely, gene expression, receptor-triggered signaling pathways, protein synthesis, or protein degradation. Thus, proteins and intermediates involved in these back-up mechanisms may be completely different; hence, drugs targeting a given primary signaling pathway may have no effect over the back-up pathways, a phenomenon known as redundancy [224]. Additionally, cellular networks and signaling pathways are strongly buffered, thus preventing major output changes, in spite of the pronounced changes that their components may undergo [225,226].

So, a multi-target therapy approach can be more efficacious as it may mitigate the redundancy escape by interfering simultaneously with three target pathways; in so doing, this triad of medicines could sequentially act on the complex multiple pathogenic mechanisms giving rise to neuronal death in neurodegeneration [227,228]. Here we propose a novel therapeutic strategy to combat retina degeneration by acting on three common and interrelated pathogenic pathways, namely: (i) excitotoxicity linked to overactivation of NMDARs, AMPARs, and Cav channels with ensuing neuronal and mitochondrial Ca<sup>2+</sup> overload; (ii) mitochondrial dysfunction and oxidative stress with augmented ROS production and decreased ATP production; and (iii) neuroinflammation linked to P2X7Rs. In the search of three compound candidates in order to

#### Table 5

Pharmacokinetic characteristics of the repositioned medicines included in Table 4.

| Drug             | Oral<br>bioavailability<br>(%)                                  | % Bound in plasma                               | V<br>distribution<br>(L/Kg)                   | t <sub>1/2</sub>  | CYP450 effects   | BBB                       | RBB   | Comments  |
|------------------|---|---|---|---|--|---------------------------|---|---|
| Memantine        | 100 [371]   | 45 [372]  | 9–11 [372]                                    | 60–80 h [372]   | Low CYP<br>inhibitory<br>promiscuity<br>[373]  | +[373]                    | + rats [374]<br>+ rats and monkeys<br>experimental<br>glaucoma [375]            | Linear pharmacokinetics<br>over the therapeutic dose<br>range.<br>Combination with other<br>drugs as citicoline have<br>shown better<br>pharmacokinetic parameters<br>[376] |
| Pioglitazone     | 83 [377]<br>97 [378]  | > 99 [379]                                      | 0.63 + /<br>- 0.41<br>[379]<br>0.25 [378]     | 3–7 h, (16–24 h<br>active<br>metabolites)<br>[379]<br>9 h [377,378] | Inhibitor<br>CYP450 1A2,<br>2C9, 2C19. High<br>inhibitory<br>promiscuity<br>[379]<br>No effects [377]              | +[379,<br>380]            | + rats, diabetes<br>model, STZ induced<br>[381]                                 | 20% metabolized by CYP3A4<br>(possible interindividual<br>variability in PK, possible<br>inhibition by other drugs)<br>[378]  |
| Minocycline      | 95–100 [382]  | 76<br>[383,384]                                 | 67.5–115<br>[383,385]                         | 11.1–22.1 [383]   | Low CYP<br>inhibitory<br>promiscuity<br>[383]  | +[386]                    | + murine ischemia-<br>reperfusion, retinal<br>detachment<br>[387–389]           | Needs monitoring for > 6<br>months treatments<br>(hepatotoxicity, lupus<br>eritematosus, pigmentation)<br>[390]   |
| Nilvadipine      | 14–19 [391]   | 97.5–98.7<br>[392]                              | 94.7 [393]                                    | 15–20 [394]<br>11 + /- 2.3 h<br>[393]<br>9.8–18.2 [395]             | Inhibitor:<br>CYP4501A2,<br>2C9, 2C19,3A4<br>High CYP<br>inhibitory<br>promiscuity<br>[396]                        | -[396]                    | + rat retinal<br>ischemia [397]   | High first-pass metabolism<br>[394]<br>Substrate of CYP3A4 [396]<br>Therapeutic plasma levels<br>are reached after oral<br>administration [395]                             |
| Edaravone        | 60% [398]   | 92% [399]                                       | 18.5 [400]                                    | 4.5–10.9 h<br>[398,400]   | No [401]   | +[400,<br>402]            | + mouse model<br>normal tension<br>glaucoma [403]                               | Well tolerated [404]<br>Protects retina against<br>several insults<br>(isquemia/reperfusion,<br>steptozocin induced<br>diabetes, retinal<br>detachment).                    |
| Naloxone         | < 2% [405] but<br>significant<br>sublingual<br>absorption [406] | 45% [407]<br>50% [408]                          | 200 [407]                                     | 1–2 h [408]   | Low CYP<br>inhibitory<br>promiscuity<br>[407]  | +[407,<br>409]            | + mouse [409]   | Substrate CYP450 2D6, 3A4<br>[407]<br>High presystemic<br>metabolism [408]  |
| Ceftriaxone      | 8% (24% rectal<br>absorption with<br>enhancer) [410]            | 95% [411]                                       | 5.78–3.5 (iv,<br>im) [411]<br>8.5–10<br>[412] | 5.8–13.5 h (im)<br>[413]<br>6 h (iv) [412]                          | Inhibitor<br>CYP450 3A4.<br>High CYP<br>inhibitory<br>promiscuity<br>[413]   | -[413]<br>+(low)<br>[414] | Detectable in<br>vitreous after<br>intramuscular<br>administration [415<br>416] | Substrate of CYP450 3A4 [413]   |
| Mitoquinone      | 10% as methane-<br>sulfonate [417,<br>418]                      | Binds to<br>serum<br>albumine in<br>vitro [419] |   | 14 h rats [420]   | Few information<br>available [421]<br>CoQ10<br>undergoes<br>structural<br>changes when<br>exposed to CYPs<br>[422] | +[423]                    | + mouse<br>inherited<br>photoreceptor<br>degeneration<br>[424]                  | Accumulates several<br>hundredfolds into the<br>mitochondria.<br>Ongoing several CTs for<br>usefulness in MS, PD, AD,<br>Asthma, and others                                 |
| Valproic<br>acid | 90–100 [425]  | 82–90<br>[426]                                  | 0.1–0.4<br>[427]                              | 13–19 h [426]<br>6–8 h [427]<br>9–16 h [428]                        | Low CYP<br>inhibitory<br>promiscuity<br>[426]  | +[426]                    | + in vitro assays<br>[429,430]  | It is absorbed rapidly. Its<br>bioavailability is complete<br>independently of the preparation<br>used [427]  |

block simultaneously those common pathogenic pathways, we also propose the search of three medicines in clinical use through the shorter strategy of drug repositioning, that we comment next.

### 6.2. Drug repositioning

For years, the finding of second indications for medicines in clinical use has been fortuitous. For instance, in 1951 isoniazid and its isopropyl derivative iproniazid were developed for the treatment of tuberculosis. When their clinical use started, it was soon found that iproniazid (but not isoniazid) had mood-elevating effects in tuberculosis patients; this was associated to its ability to block the enzyme monoamine oxidase (MAO), that allowed the introduction of MAO inhibitors in psychiatry for the treatment of depressed patients [229]. A few other serendipitous discoveries of second indications of drugs opened the way for a more systematic search of new indications for old drugs, so called repositioning or repurposing approach, that started to gain interest at the beginning of this century [230].

The principle that establishes the basis of drug repositioning can be defined as multiple compound-targets; this refers to the phenomenon by which the majority of compounds and medicines bind to more than one biological target; in so doing, they exhibit pharmaco-physiological effects secondary to the initially desired one [231]. In addition, one single target can be relevant in more than one mechanism of disease. Two main tools are being followed in repositioning programs: (i) generation of connection hypotheses through literature-based search of related

cellular and pathogenic mechanisms; (ii) study of bioinformatic features, pharmacological properties and prediction of interactions via computational approaches [232]. These two complementary approaches offer valuable insights into the relationships between drugs, targets and diseases, which are key for successful repositioning. The increasing attention to this strategy during the last two decades is grounded on the following facts: (i) increasing high costs of drug discovery and development [233]; (ii) lower number of drugs approved by regulatory agencies each year [234]; (iii) several steps of the drug development pipeline can be skipped during drug repositioning; (iv) the candidate drug has a known biological effect in humans and comply with the requirements of safety and bioactivity. Thus, in most cases the evaluation of a suitability of a repositioned drug is directly initiated in phase II studies. This shortens the time span of development, reduces risks, and compensates for the economic pressure characteristic of pharma industry [235].

There is a long list of examples of repositioned drugs; Table 2 shows some critical and interesting examples. Overall, approximately 170 drugs entered the process of repositioning approach; the majority of them are in distinct phases of clinical trials and around 10% of them have been approved, which denotes an advantageous outcome in comparison to the standard drug development program.

#### 6.3. The pharmacological basis of drug combinations

As discussed above, the repositioning of approved medicines has gained new momentum for the rapid identification and clinical development of new therapeutic indications for diseases that lack effective treatments. This strategy has proven to be successful in the last decade. Nevertheless, a limitation of this approach has been the relatively low potency that some of the repositioned medicines exhibit on the new explored target. Thus, upon their oral administration, they may produce tolerated plasma concentrations that may be lower than those required for therapeutic efficacy. This unfavorable therapeutic/safety ratio may be overcome by the combination of two or more medicines with different but complementary mechanisms of action in disease pathogenesis; this combination is an alternative approach to increase the success rate of drug repositioning [256–258].

By targeting different pathogenic signaling pathways, the use of drugs in combinations may provide a synergistic therapeutic effect. This results in a reduction of the required effective drug concentration for each individual drug, with respect the concentration required to inhibit its specific target. Once identified in a given repositioning program, the selected drugs can be tested on in vitro and in vivo models of the disease being explored. For instance, a study found that at sub-threshold concentrations with no effect for single drugs, the association of melatonin and galantamine afforded a synergistic neuroprotection in neuronal cultures [259] (Fig. 8A). Also, in an in vivo model of stroke in mice, additive effects in reducing infarct volume with combined citicoline and nimodipine were found [260]. So, drug combinations may allow administration schedules at doses low enough to achieve the desired therapeutic effect at human plasma concentrations below their toxicity effects [261] (Fig. 8B). This has been particularly successful in cancer, were combined drugs exhibit superior efficacy and safety, compared with monotherapies [262].

Drug-drug interactions may also limit the combination of medicines. Although pharmacodynamic interactions may occur, pharmacokinetic (PK) interactions are the most common. Such is the case of the estrogen receptor modulator tamoxifen and the selective serotonin reuptake inhibitor paroxetine that is used to treat breast cancer in women with depression. Cytochrome P450 2D6 (CYP2D6) metabolizes tamoxifen to its active metabolite endoxifen; thus, in inhibiting CYP2D6, paroxetine decreases the metabolism of tamoxifen, thereby diminishing the endoxifen levels [263]. A latter study demonstrated enhanced risk of death from breast cancer in women who took paroxetine and tamoxifen, compared with women taking only tamoxifen [264]. Thus, care must be exerted in drug combinations, that should be avoided when one of them inhibits CYP3A4, as it is the case for ketoconazole, that leads to higher toxic levels of the other drugs [265]; or contrarily, the drug may induce CYP3A4, such as rifampicin, leading to lower plasma levels and lower efficacy of the other drugs [266].

In summary, there are three major advantages for combination therapies emerging from drug repositioning strategies: (i) the potential for synergistic effects that significantly reduces drug concentrations for each of the individual drug; this greatly augments the chances of their application in combination, which are otherwise insufficiently active as single agents; (ii) the reduction or delay of the development of drug resistance as a result of multiple targeting mechanisms of the drug combinations; and (iii) the partial inhibition of a small number of targets could be more therapeutically efficient than complete inhibition of a single target [267].

# 7. Combined drug triads for synergic neuroprotection in retinal degeneration

Clinical trials with drug combinations in neurodegenerative diseases are scarce. For instance, in AD, only 2% of phase II CTs and 6% of phase III CTs were done with drug combinations [268]. This is also true for eye diseases leading to retinal degeneration; here, drug combinations have been practically restricted to antioxidant compounds [3,269]. Table 3 collects the drugs (small molecules) and mechanisms of action on Ca<sup>2+</sup> dyshomeostasis/excitotoxicity, oxidative stress/mitochondrial dysfunction and neuroinflammation/P2X7Rs, addressed in CTs.

Multitarget single drugs or drug combinations addressed to different targets are the two approaches that look for a higher efficacy to combat retinal degeneration with lower side effects. Some examples illustrate the first approach. Thus, in DR, curcumin has shown to alter Ca<sup>2+</sup> mobilization, (i) by modulating or blocking L-type Cav channels [360, 361], (ii) by decreasing oxidative stress through regulation of ROS and mitochondrial depolarization [362-364] and through diminution of the expression and activation of P2X7Rs [365], and (iii) by amending  $Ca^{2+}$ dysregulation in microglia [366]. So, this is an illustrative example of a multi-target single molecule. Another example is the antidiabetic metformin [367], that has been explored in DR and AMD, showing decreased oxidative stress and inflammation by modulating P2X7R-mediated signaling pathway [368] and by decreasing Ca<sup>2+</sup> influx as well [367]. Of interest is also the reduction by saffron of ATP-induced retinal cytotoxicity, by altering the P2X7R-mediated Ca<sup>2+</sup> signaling in a model of DR [369]. Finally, in glaucoma taurine has been shown to modulate ER stress and  $Ca^{2+}$  homeostasis thereby protecting against P2X7R-mediated toxicity [370].

The second approach is related to the combination of three drugs (triads) acting specifically and selectively on each one of the three pathogenic targets that are common to the four eye diseases here discussed, namely, DR, AMD, glaucoma, and RP (Fig. 9). The proposal implies the systemic oral administration of the triad as to complement and reinforce the topical eye treatments currently used. As the triads are intended to be selected in the frame of a program of repositioned drugs in clinical use, their pharmacodynamic and pharmacokinetic properties are well known. The ideal properties of the triad medicines should therefore comprise: (i) known effective drug concentrations for their main activity (ED<sub>50</sub>); (ii) lipophilicity to cross cell membranes and so, to pass through the brain-blood barrier (BBB) and retinal-blood barrier (RBB); (iii) low first-pass hepatic extraction and poor metabolism by cytochrome P450 liver enzymes; (iv) lack of P450 enzyme induction or inhibition to prevent drug-drug interactions; (v) ample toxicity window with a large therapeutic ratio.

Looking at the drugs included in Table 3, that have been or are being tested in CTs, the following triads are proposed (Table 4). The PK characteristics of these 9 medicines are summarized in Table 5. These three triads are being suggested as examples of drugs that have optimal characteristics to cross the BBB, do not have high interactions with the

hepatic cytochrome P450 and a good bioavailability.

#### 8. Conclusions and perspectives

Blindness due to retinal degeneration is a major health problem. Poor quality of life for patients and their families and a huge burden to society and health systems are some of these health, social and economic problems. Here we focused on four eye diseases that are major causes of retinal degeneration namely, glaucoma, AMD, DR, and RP. Until geneand cell-based therapies evolve to successfully target the etiology and/or restore the cell functionality in neurodegenerative diseases, a pharmacological approach is, for now, our best chance to slow down the degenerative process. These diseases have multiple etiologies which are unknown in most cases. From a pathogenic point of view, they have myriad mechanisms and signaling pathways that are specific for each one. However, they also share some mechanisms that end up with degeneration of the different retinal cell types, ie, Ca<sup>2+</sup> overload (NMDAR and AMPAR mediated excitotoxicity), oxidative stress (distorted mitochondrial Ca<sup>2+</sup> handling with augmented free radical production by the respiratory chain), and neuroinflammation (overactivation of the inflammasome by P2X7R overstimulation by high ATP concentrations locally released by damaged tissues). Here we argue that redundancy of several pathogenic pathways can compensate one of them that has been blocked by a single drug. This could explain the failure of hundreds CTs with a single-test compound, that provided negative outcomes in both brain neurodegenerative diseases and the pathogenically related eye diseases leading to retinal degeneration. CTs with combined drugs have provided highly positive outcomes in several chronic diseases as cancer, AIDS, cardiac failure, asthma, the epilepsies, Parkinson's disease, or major depression. In spite of this vast evidence in favor of drugs combinations, scarce if any clinical trials are being done with drug combinations in both brain neurodegenerative diseases and retinal degeneration.

In this review, we provide strong arguments to ground the hypothesis that a triad of repositioned medicines could generate better outcomes in CTs with patients suffering from retinal degeneration. These three medicines showing complementary mechanisms of action on excitotoxicity, oxidative stress, and neuroinflammation may lead to better outcomes in clinical trials in patients suffering from glaucoma, AMD, DR, and RP. A major challenge to this strategy could be the potential PK interactions between the three medicines included in a given therapeutic triad. However, as the PK profiles of these medicines in medical use are well known, those interactions may be overcome.

In the middle of a high scientific controversy, the FDA just approved the anti-amyloid beta (anti-A $\beta$ ) monoclonal antibody aducanumab; the outcomes from two large phase III clinical trials were modest in one trial and negative in the other. The cost of such medication will be around 40.000 euros/year, with monthly administration of the antibody for an undetermined time. Nevertheless, the FDA has asked Biogen, the company that will manufacture the medicine, to perform a novel phase III clinical trial. This example illustrates, once more, the problem of addressing a treatment to combat the complexity of neurodegeneration processes with a single-target drug. For reasons of patent protection and competitive business, pharmaceutical companies rarely agree in performing a clinical trial with combinations of their proprietary drugs. This reluctancy may be overcome by academia, with programs of drug repositioning that have been quite successful in the last decades. We predict that a combined triad of old medicines that went out of patent protection, targeting the three pathogenic mechanisms here addressed, will provide better outcomes in clinical trials aimed at testing its potential to prevent and delay retinal degeneration and blindness in patients suffering from glaucoma, AMD, DR, and RP; this may be even more successful if triads are added as adjunct to standard current treatments of glaucoma, AMD, DR, and RP. To date, many scientific and economic efforts, have been made to slow or stop retinal degenerative processes. Here we propose a new approach that considers more than

one target. The whole way to the elucidation of the therapeutic possibilities and applicability of these triads is long but, considering the efforts made up to date and the results obtained so far, we consider it worth to try.

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## CRediT authorship contribution statement

AGG conceived the manuscript. AGG and VM drafted the manuscript. VM and AMG searched for bibliography. AGG, VM, PL, NC and AMG designed the figures. AGG and VM designed the tables. AGG, VM, AMG, PL and NC revised the final manuscript.

#### References

- H. Kolb, How the retina works, Am. Sci. 91 (2003) 28, https://doi.org/10.1511/ 2003.11.841.
- [2] R. Boia, N. Ruzafa, I.D. Aires, X. Pereiro, A.F. Ambrosio, E. Vecino, A.R. Santiago, Neuroprotective strategies for retinal ganglion cell degeneration: current status and challenges ahead, Int. J. Mol. Sci. 21 (2020), https://doi.org/10.3390/ iims21072262.
- [3] N. Cuenca, L. Fernandez-Sanchez, L. Campello, V. Maneu, P. De la Villa, P. Lax, I. Pinilla, Cellular responses following retinal injuries and therapeutic approaches for neurodegenerative diseases, Prog. Retin Eye Res. 43 (2014) 17–75, https:// doi.org/10.1016/j.preteyeres.2014.07.001.
- [4] E. Kirby, S. Bandelow, E. Hogervorst, Visual impairment in Alzheimer's disease: a critical review, J. Alzheimers Dis. 21 (2010) 15–34, https://doi.org/10.3233/ JAD-2010-080785.
- [5] I. Bodis-Wollner, Retinopathy in Parkinson disease, J. Neural Transm. 116 (2009) 1493–1501, https://doi.org/10.1007/s00702-009-0292-z.
- [6] I. Ortuno-Lizaran, G. Esquiva, T.G. Beach, G.E. Serrano, C.H. Adler, P. Lax, N. Cuenca, Degeneration of human photosensitive retinal ganglion cells may explain sleep and circadian rhythms disorders in Parkinson's disease, Acta Neuropathol. Commun. 6 (2018) 90, https://doi.org/10.1186/s40478-018-0596-
- [7] I. Ortuno-Lizaran, X. Sanchez-Saez, P. Lax, G.E. Serrano, T.G. Beach, C.H. Adler, N. Cuenca, Dopaminergic retinal cell loss and visual dysfunction in Parkinson disease, Ann. Neurol. 88 (2020) 893–906, https://doi.org/10.1002/ana.25897.
- [8] A.G. Garcia, A.M. Garcia-De-Diego, L. Gandia, R. Borges, J. Garcia-Sancho, Calcium signaling and exocytosis in adrenal chromaffin cells, Physiol. Rev. 86 (2006) 1093–1131, https://doi.org/10.1152/physrev.00039.2005.
- [9] E. Neher, Vesicle pools and Ca2+ microdomains: new tools for understanding their roles in neurotransmitter release, Neuron 20 (1998) 389–399, https://doi. org/10.1016/s0896-6273(00)80983-6.
- [10] S.M. Simon, R.R. Llinas, Compartmentalization of the submembrane calcium activity during calcium influx and its significance in transmitter release, Biophys. J. 48 (1985) 485–498, https://doi.org/10.1016/S0006-3495(85)83804-2.
- [11] M.T. Alonso, M.J. Barrero, P. Michelena, E. Carnicero, I. Cuchillo, A.G. Garcia, J. Garcia-Sancho, M. Montero, J. Alvarez, Ca2+-induced Ca2+ release in chromaffin cells seen from inside the ER with targeted aequorin, J. Cell Biol. 144 (1999) 241–254, https://doi.org/10.1083/jcb.144.2.241.
- M.R. Duchen, Mitochondria and calcium: from cell signalling to cell death, J. Physiol. 529 (Pt 1) (2000) 57–68, https://doi.org/10.1111/j.1469-7793.2000.00057.x.
- [13] T.E. Gunter, D.R. Pfeiffer, Mechanisms by which mitochondria transport calcium, Am. J. Physiol. 258 (1990) C755–C786, https://doi.org/10.1152/ aipcell.1990.258.5.C755.
- [14] P. Bernardi, Mitochondrial transport of cations: channels, exchangers, and permeability transition, Physiol. Rev. 79 (1999) 1127–1155, https://doi.org/ 10.1152/physrev.1999.79.4.1127.
- [15] T.E. Gunter, K.K. Gunter, S.S. Sheu, C.E. Gavin, Mitochondrial calcium transport: physiological and pathological relevance, Am. J. Physiol. 267 (1994) C313–C339, https://doi.org/10.1152/ajpcell.1994.267.2.C313.
- [16] C. Villalobos, L. Nunez, M. Montero, A.G. Garcia, M.T. Alonso, P. Chamero, J. Alvarez, J. Garcia-Sancho, Redistribution of Ca2+ among cytosol and organella during stimulation of bovine chromaffin cells, FASEB J. 16 (2002) 343–353, https://doi.org/10.1096/fj.01-0630com.
- [17] R.J.R. A, Calretinin and calbindin-D28k in rat brain: patterns of partial colocalization Title, (n.d.). (https://doi.org/10.1016/0306-4522(92)90525-7).

- [18] L. Winsky, D.M. Jacobowitz, Radioimmunoassay of calretinin in the rat brain, Neurochem. Int. 19 (1991) 517–522, https://doi.org/10.1016/0197-0186(91) 90070-T.
- [19] P. Ince, N. Stout, P. Shaw, J. Slade, W. Hunziker, C.W. Heizmann, K. G. Baimbridge, Parvalbumin and calbindin D-28k in the human motor system and in motor neuron disease, Neuropathol. Appl. Neurobiol. 19 (1993) 291–299, https://doi.org/10.1111/j.1365-2990.1993.tb00443.x.
- [20] R.S. Sloviter, Calcium-binding protein (calbindin-D28k) and parvalbumin immunocytochemistry: localization in the rat hippocampus with specific reference to the selective vulnerability of hippocampal neurons to seizure activity, J. Comp. Neurol. 280 (1989) 183–196, https://doi.org/10.1002/ cne.902800203.
- [21] B. Katz, R. Miledi, The effect of calcium on acetylcholine release from motor nerve terminals, Proc. R. Soc. L B Biol. Sci. 161 (1965) 496–503, https://doi.org/ 10.1098/rspb.1965.0017.
- [22] M.J. Berridge, P. Lipp, M.D. Bootman, The versatility and universality of calcium signalling, Nat. Rev. Mol. Cell Biol. 1 (2000) 11–21, https://doi.org/10.1038/ 35036035.
- [23] A. Ghosh, D.D. Ginty, H. Bading, M.E. Greenberg, Calcium regulation of gene expression in neuronal cells, J. Neurobiol. 25 (1994) 294–303, https://doi.org/ 10.1002/neu.480250309.
- [24] M.J. Berridge, M.D. Bootman, P. Lipp, Calcium-a life and death signal, Nature 395 (1998) 645–648, https://doi.org/10.1038/27094.
- [25] A.M.G. de Diego, S. Lorrio, J.M. Hernández-Guijo, L. Gandía, A.G. García, Multitarget drugs for stabilization of calcium cycling and neuroprotection in neurodegenerative diseases and stroke, in: Therapeutic Targets, John Wiley & Sons, Inc, Hoboken, NJ, USA, 2012, pp. 123–200, https://doi.org/10.1002/ 9781118185537.ch4.
- [26] E.C. Toescu, A. Verkhratsky, Role of calcium in normal aging and neurodegeneration, Aging Cell 6 (2007) 265, https://doi.org/10.1111/j.1474-9726.2007.00299.x.
- [27] A. Fleckenstein, J. Janke, H.J. Doring, O. Leder, Myocardial fiber necrosis due to intracellular Ca overload-a new principle in cardiac pathophysiology, Recent Adv. Stud. Card. Struct. Metab. 4 (1974) 563–580. (https://www.ncbi.nlm.nih. gov/pubmed/4468468).
- [28] J.P. Leonard, M.M. Salpeter, Agonist-induced myopathy at the neuromuscular junction is mediated by calcium, J. Cell Biol. 82 (1979) 811–819, https://doi.org/ 10.1083/jcb.82.3.811.
- [29] F.A. Schanne, A.B. Kane, E.E. Young, J.L. Farber, Calcium dependence of toxic cell death: a final common pathway, Science 206 (1979) 700–702, https://doi. org/10.1126/science.386513.
- [30] B.F. Trump, I.K. Berezesky, Calcium-mediated cell injury and cell death, FASEB J. 9 (1995) 219–228, https://doi.org/10.1096/fasebj.9.2.7781924.
- [31] M.F. Cano-Abad, M. Villarroya, A.G. Garcia, N.H. Gabilan, M.G. Lopez, Calcium entry through L-type calcium channels causes mitochondrial disruption and chromaffin cell death, J. Biol. Chem. 276 (2001) 39695–39704, https://doi.org/ 10.1074/jbc.M102334200.
- [32] J.L. Franklin, E.M. Johnson Jr., Suppression of programmed neuronal death by sustained elevation of cytoplasmic calcium, Trends Neurosci. 15 (1992) 501–508, https://doi.org/10.1016/0166-2236(92)90103-f.
- [33] V. Gallo, A. Kingsbury, R. Balazs, O.S. Jorgensen, The role of depolarization in the survival and differentiation of cerebellar granule cells in culture, J. Neurosci. 7 (1987) 2203–2213. (https://www.ncbi.nlm.nih.gov/pubmed/2886565).
   [34] T. Koike, D.P. Martin, E.M. Johnson Jr., Role of Ca2+ channels in the ability of
- [34] T. Koike, D.P. Martin, E.M. Johnson Jr., Role of Ca2+ channels in the ability of membrane depolarization to prevent neuronal death induced by trophic-factor deprivation: evidence that levels of internal Ca2+ determine nerve growth factor dependence of sympathetic ganglion cells, Proc. Natl. Acad. Sci. USA 86 (1989) 6421–6425, https://doi.org/10.1073/pnas.86.16.6421.
- [35] G.J. Thompson, C. Langlais, K. Cain, E.C. Conley, G.M. Cohen, Elevated extracellular [K+] inhibits death-receptor- and chemical-mediated apoptosis prior to caspase activation and cytochrome c release, Biochem. J. 357 (2001) 137–145, https://doi.org/10.1042/0264-6021:3570137.
- [36] C. Orozco, A.M. Garcia-de-Diego, E. Arias, J.M. Hernandez-Guijo, A.G. Garcia, M. Villarroya, M.G. Lopez, Depolarization preconditioning produces cytoprotection against veratridine-induced chromaffin cell death, Eur. J. Pharmacol. 553 (2006) 28–38, https://doi.org/10.1016/j.ejphar.2006.08.084.
- [37] S.P. Yu, L.M. Canzoniero, D.W. Choi, Ion homeostasis and apoptosis, Curr. Opin. Cell Biol. 13 (2001) 405–411, https://doi.org/10.1016/s0955-0674(00)00228-3.
- [38] S. Xia, P.A. Lampe, M. Deshmukh, A. Yang, B.S. Brown, S.M. Rothman, E. M. Johnson Jr., S.P. Yu, E.M. Johnson, S.P. Yu, Multiple channel interactions explain the protection of sympathetic neurons from apoptosis induced by nerve growth factor deprivation, J. Neurosci. 22 (2002) 114–122, https://doi.org/10.1523/JNEUROSCI.22.01-00114.2002.
- [39] F. Collins, J.D. Lile, The role of dihydropyridine-sensitive voltage-gated calcium channels in potassium-mediated neuronal survival, Brain Res. 502 (1989) 99–108, https://doi.org/10.1016/0006-8993(89)90465-4.
- [40] J.L. Franklin, E.M. Johnson Jr., Elevated intracellular calcium blocks programmed neuronal death, Ann. N. Y. Acad. Sci. 747 (1994) 195–204, https:// doi.org/10.1111/j.1749-6632.1994.tb44410.x.
- [41] S. Orrenius, B. Zhivotovsky, P. Nicotera, Regulation of cell death: the calciumapoptosis link, Nat. Rev. Mol. Cell Biol. 4 (2003) 552–565, https://doi.org/ 10.1038/nrm1150.
- [42] D.W. Choi, Ionic dependence of glutamate neurotoxicity, J. Neurosci. 7 (1987) 369–379. (https://www.ncbi.nlm.nih.gov/pubmed/2880938).
- [43] D.W. Choi, Glutamate neurotoxicity and diseases of the nervous system, Neuron 1 (1988) 623–634, https://doi.org/10.1016/0896-6273(88)90162-6.

- [44] J.T. Coyle, P. Puttfarcken, Oxidative stress, glutamate, and neurodegenerative disorders, Science 262 (1993) 689–695, https://doi.org/10.1126/ science.7901908.
- [45] S.A. Lipton, P.A. Rosenberg, Excitatory amino acids as a final common pathway for neurologic disorders, N. Engl. J. Med. 330 (1994) 613–622, https://doi.org/ 10.1056/NEJM199403033300907.
- [46] H. Benveniste, J. Drejer, A. Schousboe, N.H. Diemer, Elevation of the extracellular concentrations of glutamate and aspartate in rat hippocampus during transient cerebral ischemia monitored by intracerebral microdialysis, J. Neurochem. 43 (1984) 1369–1374, https://doi.org/10.1111/j.1471-4159.1984.tb05396.x.
- [47] B. Meldrum, J. Garthwaite, Excitatory amino acid neurotoxicity and neurodegenerative disease, Trends Pharmacol. Sci. 11 (1990) 379–387, https:// doi.org/10.1016/0165-6147(90)90184-a.
- [48] P.A. Rosenberg, S. Amin, M. Leitner, Glutamate uptake disguises neurotoxic potency of glutamate agonists in cerebral cortex in dissociated cell culture, J. Neurosci. 12 (1992) 56–61. (https://www.ncbi.nlm.nih.gov/pubmed/ 1345946).
- [49] G.L. Collingridge, R.A. Lester, Excitatory amino acid receptors in the vertebrate central nervous system, Pharmacol. Rev. 41 (1989) 143–210. (https://www.ncbi. nlm.nih.gov/pubmed/2558391).
- [50] R.A. Nicoll, R.C. Malenka, Expression mechanisms underlying NMDA receptordependent long-term potentiation, Ann. N. Y. Acad. Sci. 868 (1999) 515–525, https://doi.org/10.1111/j.1749-6632.1999.tb11320.x.
- [51] J.A. Gorter, J.J. Petrozzino, E.M. Aronica, D.M. Rosenbaum, T. Opitz, M. V. Bennett, J.A. Connor, R.S. Zukin, Global ischemia induces downregulation of Glur2 mRNA and increases AMPA receptor-mediated Ca2+ influx in hippocampal CA1 neurons of gerbil, J. Neurosci. 17 (1997) 6179–6188. (https://www.ncbi.nlm.nih.gov/pubmed/9236229).
- [52] A. Novelli, J.A. Reilly, P.G. Lysko, R.C. Henneberry, Glutamate becomes neurotoxic via the N-methyl-D-aspartate receptor when intracellular energy levels are reduced, Brain Res. 451 (1988) 205–212, https://doi.org/10.1016/0006-8993(88)90765-2.
- [53] P. Van Damme, M. Dewil, W. Robberecht, L. Van Den Bosch, Excitotoxicity and amyotrophic lateral sclerosis, Neurodegener. Dis. 2 (2005) 147–159, https://doi. org/10.1159/000089620.
- [54] J.A. Dykens, Isolated cerebral and cerebellar mitochondria produce free radicals when exposed to elevated CA2+ and Na+: implications for neurodegeneration, J. Neurochem. 63 (1994) 584–591, https://doi.org/10.1046/j.1471-4159.1994.63020584.x.
- [55] M. Urushitani, T. Nakamizo, R. Inoue, H. Sawada, T. Kihara, K. Honda, A. Akaike, S. Shimohama, N-methyl-D-aspartate receptor-mediated mitochondrial Ca(2+) overload in acute excitotoxic motor neuron death: a mechanism distinct from chronic neurotoxicity after Ca(2+) influx, J. Neurosci. Res. 63 (2001) 377–387, https://doi.org/10.1002/1097-4547(20010301)63:5<377::AID-JNR1032>3.0. CO;2-#.
- [56] D.W. Choi, Calcium-mediated neurotoxicity: relationship to specific channel types and role in ischemic damage, Trends Neurosci. 11 (1988) 465–469, https:// doi.org/10.1016/0166-2236(88)90200-7.
- [57] P. Van Damme, L. Van Den Bosch, E. Van Houtte, G. Callewaert, W. Robberecht, GluR2-dependent properties of AMPA receptors determine the selective vulnerability of motor neurons to excitotoxicity, J. Neurophysiol. 88 (2002) 1279–1287, https://doi.org/10.1152/jn.2002.88.3.1279.
- [58] A. Bringmann, O. Uckermann, T. Pannicke, I. Iandiev, A. Reichenbach, P. Wiedemann, Neuronal versus glial cell swelling in the ischaemic retina, Acta Ophthalmol. Scand. 83 (2005) 528–538, https://doi.org/10.1111/j.1600-0420.2005.00565.x.
- [59] N.N. Osborne, J.P. Wood, G. Chidlow, R. Casson, L. DeSantis, K.G. Schmidt, Effectiveness of levobetaxolol and timolol at blunting retinal ischaemia is related to their calcium and sodium blocking activities: relevance to glaucoma, Brain Res. Bull. 62 (2004) 525–528, https://doi.org/10.1016/S0361-9230(03)00070-4.
- [60] M. Azuma, T.R. Shearer, The role of calcium-activated protease calpain in experimental retinal pathology, Surv. Ophthalmol. 53 (2008) 150–163, https:// doi.org/10.1016/j.survophthal.2007.12.006.
- [61] F. Doonan, M. Donovan, T.G. Cotter, Activation of multiple pathways during photoreceptor apoptosis in the rd mouse, Investig. Ophthalmol. Vis. Sci. 46 (2005) 3530–3538, https://doi.org/10.1167/iovs.05-0248.
- [62] D.P. McKernan, M.B. Guerin, C.J. O'Brien, T.G. Cotter, A key role for calpains in retinal ganglion cell death, Investig. Ophthalmol. Vis. Sci. 48 (2007) 5420–5430, https://doi.org/10.1167/iovs.07-0287.
- [63] W. Huang, J.B. Fileta, A. Dobberfuhl, T. Filippopolous, Y. Guo, G. Kwon, C. L. Grosskreutz, Calcineurin cleavage is triggered by elevated intraocular pressure, and calcineurin inhibition blocks retinal ganglion cell death in experimental glaucoma, Proc. Natl. Acad. Sci. USA 102 (2005) 12242–12247, https://doi.org/10.1073/pnas.0505138102.
- [64] W. Huang, J. Fileta, I. Rawe, J. Qu, C.L. Grosskreutz, Calpain activation in experimental glaucoma, Investig. Ophthalmol. Vis. Sci. 51 (2010) 3049–3054, https://doi.org/10.1167/iovs.09-4364.
- [65] J.H. Weiss, D.M. Hartley, J. Koh, D.W. Choi, The calcium channel blocker nifedipine attenuates slow excitatory amino acid neurotoxicity, Science 247 (1990) 1474–1477, https://doi.org/10.1126/science.2157282.
- [66] C.E. Crosson, J.A. Willis, D.E. Potter, Effect of the calcium antagonist, nifedipine, on ischemic retinal dysfunction, J. Ocul. Pharmacol. 6 (1990) 293–299, https:// doi.org/10.1089/jop.1990.6.293.

- [67] M. Nakazawa, Effects of calcium ion, calpains, and calcium channel blockers on retinitis pigmentosa, J. Ophthalmol. 2011 (2011), 292040, https://doi.org/ 10.1155/2011/292040.
- [68] D.A. Fox, A.T. Poblenz, L. He, J.B. Harris, C.J. Medrano, Pharmacological strategies to block rod photoreceptor apoptosis caused by calcium overload: a mechanistic target-site approach to neuroprotection, Eur. J. Ophthalmol. 13 Suppl 3 (2003) S44–S56, https://doi.org/10.1177/112067210301303s08.
- [69] N.N. Osborne, J.P. Wood, A. Cupido, J. Melena, G. Chidlow, Topical flunarizine reduces IOP and protects the retina against ischemia-excitotoxicity, Investig. Ophthalmol. Vis. Sci. 43 (2002) 1456–1464. (https://www.ncbi.nlm.nih.gov/ pubmed/11980861).
- [70] K. Takahashi, T.T. Lam, D.P. Edward, E.R. Buchi, M.O. Tso, Protective effects of flunarizine on ischemic injury in the rat retina, Arch. Ophthalmol. 110 (1992) 862–870, https://doi.org/10.1001/archopht.1992.01080180134041.
- [71] L. Zhang, Y.N. Hui, Y.S. Wang, J.X. Ma, J.B. Wang, L.N. Ma, Calcium overload is associated with lipofuscin formation in human retinal pigment epithelial cells fed with photoreceptor outer segments, Eye 25 (2011) 519–527, https://doi.org/ 10.1038/eye.2011.7.
- [72] C.J. Dong, Y. Guo, L. Wheeler, W.A. Hare, Alpha2 adrenergic receptor-mediated modulation of cytosolic Ca++ signals at the inner plexiform layer of the rat retina, Investig. Ophthalmol. Vis. Sci. 48 (2007) 1410–1415, https://doi.org/ 10.1167/iovs.06-0890.
- [73] N.N. Osborne, C. Cazevieille, A.L. Carvalho, A.K. Larsen, L. DeSantis, In vivo and in vitro experiments show that betaxolol is a retinal neuroprotective agent, Brain Res. 751 (1997) 113–123, https://doi.org/10.1016/s0006-8993(96)01393-5.
- [74] D.R. Sisk, T. Kuwabara, Histologic changes in the inner retina of albino rats following intravitreal injection of monosodium L-glutamate, Graefes Arch. Clin. Exp. Ophthalmol. 223 (1985) 250–258, https://doi.org/10.1007/BF02153655.
- [75] C.K. Vorwerk, S.A. Lipton, D. Zurakowski, B.T. Hyman, B.A. Sabel, E.B. Dreyer, Chronic low-dose glutamate is toxic to retinal ganglion cells. Toxicity blocked by memantine, Investig. Ophthalmol. Vis. Sci. 37 (1996) 1618–1624. (https://www. ncbi.nlm.nih.gov/pubmed/8675405).
- [76] E.B. Dreyer, D. Zurakowski, R.A. Schumer, S.M. Podos, S.A. Lipton, Elevated glutamate levels in the vitreous body of humans and monkeys with glaucoma, Arch. Ophthalmol. 114 (1996) 299–305, https://doi.org/10.1001/ archopht.1996.01100130295012.
- [77] P. Chaudhary, F. Ahmed, S.C. Sharma, MK801-a neuroprotectant in rat hypertensive eyes, Brain Res. 792 (1998) 154–158, https://doi.org/10.1016/ s0006-8993(98)00212-1.
- [78] L. Guo, T.E. Salt, A. Maass, V. Luong, S.E. Moss, F.W. Fitzke, M.F. Cordeiro, Assessment of neuroprotective effects of glutamate modulation on glaucomarelated retinal ganglion cell apoptosis in vivo, Investig. Ophthalmol. Vis. Sci. 47 (2006) 626–633, https://doi.org/10.1167/iovs.05-0754.
- [79] W.A. Lagreze, R. Knorle, M. Bach, T.J. Feuerstein, Memantine is neuroprotective in a rat model of pressure-induced retinal ischemia, Investig. Ophthalmol. Vis. Sci. 39 (1998) 1063–1066. (https://www.ncbi.nlm.nih.gov/pubmed/9579489).
- [80] D. Cervia, G. Casini, P. Bagnoli, Physiology and pathology of somatostatin in the mammalian retina: a current view, Mol. Cell Endocrinol. 286 (2008) 112–122, https://doi.org/10.1016/j.mce.2007.12.009.
- [81] Q. Li, Y. Zhang, N. Wu, N. Yin, X.H. Sun, Z. Wang, Activation of somatostatin receptor 5 suppresses T-type Ca(2+) channels through NO/cGMP/PKG signaling pathway in rat retinal ganglion cells, Neurosci. Lett. 708 (2019), 134337, https:// doi.org/10.1016/j.neulet.2019.134337.
- [82] J. Johnson, M.L. Caravelli, N.C. Brecha, Somatostatin inhibits calcium influx into rat rod bipolar cell axonal terminals, Vis. Neurosci. 18 (2001) 101–108, https:// doi.org/10.1017/s0952523801181095.
- [83] J. Yoon, H.C. Ben-Ami, Y.S. Hong, S. Park, L.L. Strong, J. Bowman, C. Geng, K. Baek, B. Minke, W.L. Pak, Novel mechanism of massive photoreceptor degeneration caused by mutations in the trp gene of Drosophila, J. Neurosci. 20 (2000) 649–659. (https://www.ncbi.nlm.nih.gov/pubmed/10632594).
- [84] B. Minke, The TRP calcium channel and retinal degeneration, in: W. Baehr, K. Palczewski (Eds.), Photoreceptors Calcium. Advances in Experimental Medicine and Biology, Springer, Boston, MA., 2002, pp. 601–622, https://doi. org/10.1007/978-1-4615-0121-3\_34.
- [85] Z. Pan, H. Yang, P.S. Reinach, Transient receptor potential (TRP) gene superfamily encoding cation channels, Hum. Genom. 5 (2011) 108–116, https:// doi.org/10.1186/1479-7364-5-2-108.
- [86] D.A. Ryskamp, P. Witkovsky, P. Barabas, W. Huang, C. Koehler, N.P. Akimov, S. H. Lee, S. Chauhan, W. Xing, R.C. Renteria, W. Liedtke, D. Krizaj, The polymodal ion channel transient receptor potential vanilloid 4 modulates calcium flux, spiking rate, and apoptosis of mouse retinal ganglion cells, J. Neurosci. 31 (2011) 7089–7101, https://doi.org/10.1523/JNEUROSCI.0359-11.2011.
- [87] R.M. Sappington, T. Sidorova, D.J. Long, D.J. Calkins, TRPV1: contribution to retinal ganglion cell apoptosis and increased intracellular Ca2+ with exposure to hydrostatic pressure, Investig. Ophthalmol. Vis. Sci. 50 (2009) 717–728, https:// doi.org/10.1167/iovs.08-2321.
- [88] D.A. Ryskamp, A.M. Frye, T.T. Phuong, O. Yarishkin, A.O. Jo, Y. Xu, M. Lakk, A. Iuso, S.N. Redmon, B. Ambati, G. Hageman, G.D. Prestwich, K.Y. Torrejon, D. Krizaj, TRPV4 regulates calcium homeostasis, cytoskeletal remodeling, conventional outflow and intraocular pressure in the mammalian eye, Sci. Rep. 6 (2016) 30583, https://doi.org/10.1038/srep30583.
- [89] U. Wojda, E. Salinska, J. Kuznicki, Calcium ions in neuronal degeneration, IUBMB Life 60 (2008) 575–590, https://doi.org/10.1002/iub.91.
- [90] J. Kaur, S. Mencl, A. Sahaboglu, P. Farinelli, T. van Veen, E. Zrenner, P. Ekstrom, F. Paquet-Durand, B. Arango-Gonzalez, Calpain and PARP activation during

photoreceptor cell death in P23H and S334ter rhodopsin mutant rats, PLoS One 6 (2011), e22181, https://doi.org/10.1371/journal.pone.0022181.

- [91] V. Shinde, P. Kotla, C. Strang, M. Gorbatyuk, Unfolded protein response-induced dysregulation of calcium homeostasis promotes retinal degeneration in rat models of autosomal dominant retinitis pigmentosa, Cell Death Dis. 7 (2016), e2085, https://doi.org/10.1038/cddis.2015.325.
- [92] D. Sanges, A. Comitato, R. Tammaro, V. Marigo, Apoptosis in retinal degeneration involves cross-talk between apoptosis-inducing factor (AIF) and caspase-12 and is blocked by calpain inhibitors, Proc. Natl. Acad. Sci. USA 103 (2006) 17366–17371, https://doi.org/10.1073/pnas.0606276103.
- [93] N. Rodríguez-Muela, A.M. Hernández-Pinto, A. Serrano-Puebla, L. García-Ledo, S. H. Latorre, E.J. de la Rosa, P. Boya, Lysosomal membrane permeabilization and autophagy blockade contribute to photoreceptor cell death in a mouse model of retinitis pigmentosa, Cell Death Differ. 22 (2015) 476–487, https://doi.org/10.1038/cdd.2014.203.
- [94] N.B. Bishara, H. Ding, Glucose enhances expression of TRPC1 and calcium entry in endothelial cells, Am. J. Physiol. Hear. Circ. Physiol. 298 (2010) H171–H178, https://doi.org/10.1152/ajpheart.00699.2009.
- [95] J. Li, P. Wang, S. Yu, Z. Zheng, X. Xu, Calcium entry mediates hyperglycemiainduced apoptosis through Ca(2+)/calmodulin-dependent kinase II in retinal capillary endothelial cells, Mol. Vis. 18 (2012) 2371–2379. (https://www.ncbi. nlm.nih.gov/pubmed/23049237).
- [96] N.V.V. Paniker, S.K.K. Srivastava, E. Beutler, Glutathione metabolism of the red cells effect of glutathione reductase deficiency on the stimulation of hexose monophosphate shunt under oxidative stress, Biochim. Biophys. Acta Gen. Subj. 215 (1970) 456–460, https://doi.org/10.1016/0304-4165(70)90096-6.
- [97] G. Loschen, A. Azzi, On the formation of hydrogen peroxide and oxygen radicals in heart mitochondria, Recent Adv. Stud. Card. Struct. Metab. 7 (1975) 3–12. (https://www.ncbi.nlm.nih.gov/pubmed/179119).
- [98] E. Cadenas, K.J. Davies, Mitochondrial free radical generation, oxidative stress, and aging, Free Radic. Biol. Med. 29 (2000) 222–230, https://doi.org/10.1016/ s0891-5849(00)00317-8.
- [99] M.T. Lin, M.F. Beal, Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases, Nature 443 (2006) 787–795, https://doi.org/ 10.1038/nature05292.
- [100] P. Yu-Wai-Man, P.G. Griffiths, P.F. Chinnery, Mitochondrial optic neuropathiesdisease mechanisms and therapeutic strategies, Prog. Retin Eye Res. 30 (2011) 81–114, https://doi.org/10.1016/j.preteyeres.2010.11.002.
- [101] K.T. Kishida, E. Klann, Sources and targets of reactive oxygen species in synaptic plasticity and memory, Antioxid. Redox Signal 9 (2007) 233–244, https://doi. org/10.1089/ars.2007.9.ft-8.
- [102] M. Valko, D. Leibfritz, J. Moncol, M.T. Cronin, M. Mazur, J. Telser, Free radicals and antioxidants in normal physiological functions and human disease, Int. J. Biochem. Cell Biol. 39 (2007) 44–84, https://doi.org/10.1016/j. biocel.2006.07.001.
- [103] V. Adam-Vizi, Production of reactive oxygen species in brain mitochondria: contribution by electron transport chain and non-electron transport chain sources, Antioxid. Redox Signal 7 (2005) 1140–1149, https://doi.org/10.1089/ ars.2005.7.1140.
- [104] G. Zundorf, S. Kahlert, V.I. Bunik, G. Reiser, alpha-Ketoglutarate dehydrogenase contributes to production of reactive oxygen species in glutamate-stimulated hippocampal neurons in situ, Neuroscience 158 (2009) 610–616, https://doi.org/ 10.1016/j.neuroscience.2008.10.015.
- [105] R.F. Feissner, J. Skalska, W.E. Gaum, S.S. Sheu, Crosstalk signaling between mitochondrial Ca2+ and ROS, Front. Biosci. (Landmark Ed. ) 14 (2009) 1197–1218, https://doi.org/10.2741/3303.
  [106] G. Csordas, G. Hajnoczky, SR/ER-mitochondrial local communication: calcium
- [106] G. Csordas, G. Hajnoczky, SR/ER-mitochondrial local communication: calcium and ROS, Biochim. Biophys. Acta 2009 (1787) 1352–1362, https://doi.org/ 10.1016/j.bbabio.2009.06.004.
- [107] C. Hidalgo, Cross talk between Ca2+ and redox signalling cascades in muscle and neurons through the combined activation of ryanodine receptors/Ca2+ release channels, Philos. Trans. R. Soc. L B Biol. Sci. 360 (2005) 2237–2246, https://doi. org/10.1098/rstb.2005.1759.
- [108] D.G. Nicholls, Mitochondrial calcium function and dysfunction in the central nervous system, Biochim. Biophys. Acta 1787 (2009) 1416–1424, https://doi. org/10.1016/j.bbabio.2009.03.010.
- [109] G. Zundorf, G. Reiser, The phosphorylation status of extracellular-regulated kinase 1/2 in astrocytes and neurons from rat hippocampus determines the thrombin-induced calcium release and ROS generation, J. Neurochem. 119 (2011) 1194–1204, https://doi.org/10.1111/j.1471-4159.2011.07527.x.
- [110] R. Fernandez-Durango, A. Fernandez-Martinez, J. Garcia-Feijoo, A. Castillo, J. M. de la Casa, B. Garcia-Bueno, B.G. Perez-Nievas, A. Fernandez-Cruz, J.C. Leza, Expression of nitrotyrosine and oxidative consequences in the trabecular meshwork of patients with primary open-angle glaucoma, Investig. Ophthalmol. Vis. Sci. 49 (2008) 2506–2511, https://doi.org/10.1167/iovs.07-1363.
- [111] V. Chrysostomou, F. Rezania, I.A. Trounce, J.G. Crowston, Oxidative stress and mitochondrial dysfunction in glaucoma, Curr. Opin. Pharmacol. 13 (2013) 12–15, https://doi.org/10.1016/j.coph.2012.09.008.
- [112] J.J. Garcia-Medina, E. Rubio-Velazquez, E. Foulquie-Moreno, R.P. Casaroli-Marano, M.D. Pinazo-Duran, V. Zanon-Moreno, M. Del-Rio-Vellosillo, Update on the effects of antioxidants on diabetic retinopathy: in vitro experiments, animal studies and clinical trials, Antioxidants 9 (2020), https://doi.org/10.3390/ antiox9060561.
- [113] Q. Liu, W.K. Ju, J.G. Crowston, F. Xie, G. Perry, M.A. Smith, J.D. Lindsey, R. N. Weinreb, Oxidative stress is an early event in hydrostatic pressure induced

retinal ganglion cell damage, Investig. Ophthalmol. Vis. Sci. 48 (2007) 4580–4589, https://doi.org/10.1167/iovs.07-0170.

- [114] M.D. Pinazo-Duran, V. Zanon-Moreno, R. Gallego-Pinazo, J.J. Garcia-Medina, Oxidative stress and mitochondrial failure in the pathogenesis of glaucoma neurodegeneration, Prog. Brain Res. 220 (2015) 127–153, https://doi.org/ 10.1016/bs.pbr.2015.06.001.
- [115] J. Blasiak, G. Petrovski, Z. Vereb, A. Facsko, K. Kaarniranta, Oxidative stress, hypoxia, and autophagy in the neovascular processes of age-related macular degeneration, Biomed. Res. Int. 2014 (2014), 768026, https://doi.org/10.1155/ 2014/768026.
- [116] P.X. Shaw, T. Stiles, C. Douglas, D. Ho, W. Fan, H. Du, X. Xiao, Oxidative stress, innate immunity, and age-related macular degeneration, AIMS Mol. Sci. 3 (2016) 196–221, https://doi.org/10.3934/molsci.2016.2.196.
- [117] A. Tisi, M. Feligioni, M. Passacantando, M. Ciancaglini, R. Maccarone, The impact of oxidative stress on blood-retinal barrier physiology in age-related macular degeneration, Cells 10 (2021) 64, https://doi.org/10.3390/cells10010064.
- [118] E.B. Domènech, G. Marfany, The relevance of oxidative stress in the pathogenesis and therapy of retinal dystrophies, Antioxidants 9 (2020) 347, https://doi.org/ 10.3390/antiox9040347.
- [119] J. Dziedziak, K. Kasarełło, A. Cudnoch-Jędrzejewska, Dietary antioxidants in agerelated macular degeneration and glaucoma, Antioxidants 10 (2021) 1743, https://doi.org/10.3390/antiox10111743.
- [120] J.J. Garcia-Medina, V. Zanon-Moreno, M.D. Pinazo-Duran, E. Foulquie-Moreno, E. Foulquie-Moreno, R.P. Casaroli-Marano, M. Del-Rio-Vellosillo, Oxidative stress in diabetic retinopathy, in: V.R. Preedy (Ed.), Diabetes, 2020, pp. 49–57. https ://doi.org/10.1016/B978-0-12-815776-3.00005-X.
- [121] M.Y. Wu, G.T. Yiang, T.T. Lai, C.J. Li, The oxidative stress and mitochondrial dysfunction during the pathogenesis of diabetic retinopathy, Oxid. Med. Cell Longev. 2018 (2018), 3420187, https://doi.org/10.1155/2018/3420187.
- [122] Y. Murakami, Y. Nakabeppu, K.H. Sonoda, Oxidative stress and microglial response in retinitis pigmentosa, Int. J. Mol. Sci. 21 (2020), https://doi.org/ 10.3390/ijms21197170.
- [123] N. Taveau, A. Cubizolle, L. Guillou, N. Pinquier, E. Moine, D. Cia, V. Kalatzis, J. Vercauteren, T. Durand, C. Crauste, P. Brabet, Preclinical pharmacology of a lipophenol in a mouse model of light-induced retinopathy, Exp. Mol. Med. 52 (2020) 1090–1101, https://doi.org/10.1038/s12276-020-0460-7.
- [124] K. Ramchani-Ben Othman, C. Cercy, M. Amri, M. Doly, I. Ranchon-Cole, Dietary supplement enriched in antioxidants and omega-3 protects from progressive lightinduced retinal degeneration, PLoS One 10 (2015), e0128395, https://doi.org/ 10.1371/journal.pone.0128395.
- [125] P.A. Campochiaro, T.A. Mir, The mechanism of cone cell death in Retinitis Pigmentosa, Prog. Retin Eye Res. 62 (2018) 24–37, https://doi.org/10.1016/j. preteyeres.2017.08.004.
- [126] Y. Chen, M. Yang, Z.-J. Wang, (Z)-7,4'-Dimethoxy-6-hydroxy-aurone-4-Oβ-glucopyranoside mitigates retinal degeneration in Rd10 mouse model through inhibiting oxidative stress and inflammatory responses, Cutan. Ocul. Toxicol. 39 (2020) 36–42, https://doi.org/10.1080/15569527.2019.1685535.
- [127] G. Burnstock, Purinergic nerves, Pharmacol. Rev. 24 (1972) 509–581. (https:// www.ncbi.nlm.nih.gov/pubmed/4404211).
- [128] S. Cockcroft, B.D. Gomperts, ATP induces nucleotide permeability in rat mast cells, Nature 279 (1979) 541–542, https://doi.org/10.1038/279541a0.
- [129] T.H. Steinberg, S.C. Silverstein, Extracellular ATP4- promotes cation fluxes in the J774 mouse macrophage cell line, J. Biol. Chem. 262 (1987) 3118–3122. (https://www.ncbi.nlm.nih.gov/pubmed/2950094).
- [130] K.A. Jacobson, M.F. Jarvis, M. Williams, Purine and pyrimidine (P2) receptors as drug targets, J. Med. Chem. 45 (2002) 4057–4093, https://doi.org/10.1021/ jm020046y.
- [131] F. Calzaferri, C. Ruiz-Ruiz, A.M.G. de Diego, R. de Pascual, I. Mendez-Lopez, M. F. Cano-Abad, V. Maneu, C. de Los Rios, L. Gandia, A.G. Garcia, The purinergic P2X7 receptor as a potential drug target to combat neuroinflammation in neurodegenerative diseases, Med. Res. Rev. 40 (2020) 2427–2465, https://doi. org/10.1002/med.21710.
- [132] R. Laliberte, D. Perregaux, L. Svensson, C.J. Pazoles, C.A. Gabel, Tenidap modulates cytoplasmic pH and inhibits anion transport in vitro. II. Inhibition of IL-1 beta production from ATP-treated monocytes and macrophages, J. Immunol. 153 (1994) 2168–2179. (http://www.ncbi.nlm.nih.gov/pubmed/8051418).
- [133] S. Falzoni, M. Munerati, D. Ferrari, S. Spisani, S. Moretti, F. Di Virgilio, The purinergic P2Z receptor of human macrophage cells. Characterization and possible physiological role, J. Clin. Investig. 95 (1995) 1207–1216, https://doi. org/10.1172/JCI117770.
- [134] F.Di Virgilio, The P2Z purinoceptor: an intriguing role in immunity, inflammation and cell death, Immunol. Today 16 (1995) 524–528, https://doi.org/10.1016/ 0167-5699(95)80045-X.
- [135] D.J. DiSabato, N. Quan, J.P. Godbout, Neuroinflammation: the devil is in the details, J. Neurochem. 139 (2016) 136–153, https://doi.org/10.1111/jnc.13607.
- [136] K.V. Swanson, M. Deng, J.P.-Y. Ting, The NLRP3 inflammasome: molecular activation and regulation to therapeutics, Nat. Rev. Immunol. 19 (2019) 477–489, https://doi.org/10.1038/s41577-019-0165-0.
- [137] H. Kawamura, T. Sugiyama, D.M. Wu, M. Kobayashi, S. Yamanishi, K. Katsumura, D.G. Puro, ATP: a vasoactive signal in the pericyte-containing microvasculature of the rat retina, J. Physiol. 551 (2003) 787–799, https://doi.org/10.1113/ jphysiol.2003.047977.
- [138] A. Wurm, T. Pannicke, I. Iandiev, M. Francke, M. Hollborn, P. Wiedemann, A. Reichenbach, N.N. Osborne, A. Bringmann, Purinergic signaling involved in Muller cell function in the mammalian retina, Prog. Retin Eye Res. 30 (2011) 324–342, https://doi.org/10.1016/j.preteyeres.2011.06.001.

- [139] J. Sanderson, D.A. Dartt, V. Trinkaus-Randall, J. Pintor, M.M. Civan, N. A. Delamere, E.L. Fletcher, T.E. Salt, A. Grosche, C.H. Mitchell, Purines in the eye: recent evidence for the physiological and pathological role of purines in the RPE, retinal neurons, astrocytes, Muller cells, lens, trabecular meshwork, cornea and lacrimal gland, Exp. Eye Res. 127 (2014) 270–279, https://doi.org/10.1016/j. exer.2014.08.009.
- [140] T. Puthussery, E.L. Fletcher, Synaptic localization of P2X7 receptors in the rat retina, J. Comp. Neurol. 472 (2004) 13–23, https://doi.org/10.1002/cne.20045.
- [141] K.A. Vessey, E.L. Fletcher, Rod and cone pathway signalling is altered in the P2X7 receptor knock out mouse, PLoS One 7 (2012), e29990, https://doi.org/10.1371/ journal.pone.0029990.
- [142] G.D. Housley, A. Bringmann, A. Reichenbach, Purinergic signaling in special senses, Trends Neurosci. 32 (2009) 128–141, https://doi.org/10.1016/j. tins.2009.01.001.
- [143] M.M. Ward, T. Puthussery, K.A. Vessey, E.L. Fletcher, The Role of Purinergic Receptors in Retinal Function and Disease, 2010, pp. 385–391. https://doi.org/ 10.1007/978-1-4419-1399-9\_44.
- [144] T. Puthussery, P. Yee, A.J. Vingrys, E.L. Fletcher, Evidence for the involvement of purinergic P2X receptors in outer retinal processing, Eur. J. Neurosci. 24 (2006) 7–19, https://doi.org/10.1111/j.1460-9568.2006.04895.x.
- [145] S. Chavda, P.J. Luthert, T.E. Salt, P2X7R modulation of visually evoked synaptic responses in the retina, Purinergic Signal 12 (2016) 611–625, https://doi.org/ 10.1007/s11302-016-9522-7.
- [146] P. Kupenova, E. Popova, L. Vitanova, Purinergic modulation of frog electroretinographic responses: the role of the ionotropic receptor P2X7, Vis. Neurosci. 34 (2017), E015, https://doi.org/10.1017/S0952523817000128.
- [147] C.H. Mitchell, W. Lu, H. Hu, X. Zhang, D. Reigada, M. Zhang, The P2X(7) receptor in retinal ganglion cells: a neuronal model of pressure-induced damage and protection by a shifting purinergic balance, Purinergic Signal 4 (2008) 313–321, https://doi.org/10.1007/s11302-008-9125-z.
- [148] A. Reichenbach, A. Bringmann, Purinergic signaling in retinal degeneration and regeneration, Neuropharmacology 104 (2016) 194–211, https://doi.org/ 10.1016/j.neuropharm.2015.05.005.
- [149] M. Monif, G. Burnstock, D.A. Williams, Microglia: proliferation and activation driven by the P2X7 receptor, Int. J. Biochem. Cell Biol. 42 (2010) 1753–1756, https://doi.org/10.1016/j.biocel.2010.06.021.
- [150] L. Dong, Y. Hu, L. Zhou, X. Cheng, P2X7 receptor antagonist protects retinal ganglion cells by inhibiting microglial activation in a rat chronic ocular hypertension model, Mol. Med. Rep. 17 (2018) 2289–2296, https://doi.org/ 10.3892/mmr.2017.8137.
- [151] V. Resta, E. Novelli, G. Vozzi, C. Scarpa, M. Caleo, A. Ahluwalia, A. Solini, E. Santini, V. Parisi, F. Di Virgilio, L. Galli-Resta, Acute retinal ganglion cell injury caused by intraocular pressure spikes is mediated by endogenous extracellular ATP, Eur. J. Neurosci. 25 (2007) 2741–2754, https://doi.org/10.1111/j.1460-9568.2007.05528.x.
- [152] E.C. Johnson, J.C. Morrison, Friend or foe? Resolving the impact of glial responses in glaucoma, J. Glaucoma. 18 (2009) 341–353, https://doi.org/10.1097/ IJG.0b013e31818c6ef6.
- [153] A. Bosco, M.R. Steele, M.L. Vetter, Early microglia activation in a mouse model of chronic glaucoma, J. Comp. Neurol. 519 (2011) 599–620, https://doi.org/ 10.1002/cne.22516.
- [154] B.I. Gallego, J.J. Salazar, R. de Hoz, B. Rojas, A.I. Ramirez, M. Salinas-Navarro, A. Ortin-Martinez, F.J. Valiente-Soriano, M. Aviles-Trigueros, M.P. Villegas-Perez, M. Vidal-Sanz, A. Trivino, J.M. Ramirez, IOP induces upregulation of GFAP and MHC-II and microglia reactivity in mice retina contralateral to experimental glaucoma, J. Neuroinflamm. 9 (2012) 92, https://doi.org/10.1186/1742-2094-9-92
- [155] D.M. Inman, P.J. Horner, Reactive nonproliferative gliosis predominates in a chronic mouse model of glaucoma, Glia 55 (2007) 942–953, https://doi.org/ 10.1002/glia.20516.
- [156] C. Luo, X. Yang, A.D. Kain, D.W. Powell, M.H. Kuehn, G. Tezel, Glaucomatous tissue stress and the regulation of immune response through glial Toll-like receptor signaling, Investig. Ophthalmol. Vis. Sci. 51 (2010) 5697–5707, https:// doi.org/10.1167/iovs.10-5407.
- [157] A. Bosco, S.D. Crish, M.R. Steele, C.O. Romero, D.M. Inman, P.J. Horner, D. J. Calkins, M.L. Vetter, Early reduction of microglia activation by irradiation in a model of chronic glaucoma, PLoS One 7 (2012), e43602, https://doi.org/ 10.1371/journal.pone.0043602.
- [158] R. Seitz, A. Ohlmann, E.R. Tamm, The role of Muller glia and microglia in glaucoma, Cell Tissue Res. 353 (2013) 339–345, https://doi.org/10.1007/ s00441-013-1666-y.
- [159] A.H. Neufeld, Pharmacologic neuroprotection with an inhibitor of nitric oxide synthase for the treatment of glaucoma, Brain Res. Bull. 62 (2004) 455–459, https://doi.org/10.1016/j.brainresbull.2003.07.005.
- [160] M. Roh, Y. Zhang, Y. Murakami, A. Thanos, S.C. Lee, D.G. Vavvas, L.I. Benowitz, J.W. Miller, Etanercept, a widely used inhibitor of tumor necrosis factor-alpha (TNF-alpha), prevents retinal ganglion cell loss in a rat model of glaucoma, PLoS One 7 (2012), e40065, https://doi.org/10.1371/journal.pone.0040065.
- [161] E. Ozaki, C. Delaney, M. Campbell, S.L. Doyle, Minocycline suppresses diseaseassociated microglia (DAM) in a model of photoreceptor cell degeneration, Exp. Eye Res. 217 (2022), 108953, https://doi.org/10.1016/j.exer.2022.108953.
- [162] K. Dannhausen, C. Möhle, T. Langmann, Immunomodulation with minocycline rescues retinal degeneration in juvenile Neuronal Ceroid Lipofuscinosis (jNCL) mice highly susceptible to light damage, Dis. Model. Mech. (2018), https://doi. org/10.1242/dmm.033597.

- [163] R. Scholz, M. Sobotka, A. Caramoy, T. Stempfl, C. Moehle, T. Langmann, Minocycline counter-regulates pro-inflammatory microglia responses in the retina and protects from degeneration, J. Neuroinflamm. 12 (2015) 209, https://doi. org/10.1186/s12974-015-0431-4.
- [164] D. Yang, S.G. Elner, A.J. Clark, B.A. Hughes, H.R. Petty, V.M. Elner, Activation of P2X receptors induces apoptosis in human retinal pigment epithelium, Investig. Ophthalmol. Vis. Sci. 52 (2011) 1522–1530, https://doi.org/10.1167/iovs.10-6172.
- [165] D. Yang, Targeting the P2X7 receptor in age-related macular degeneration, Vis 1 (2017), https://doi.org/10.3390/vision1020011.
- [166] K.A. Vessey, B.J. Gu, A.I. Jobling, J.A. Phipps, U. Greferath, M.X. Tran, M. A. Dixon, P.N. Baird, R.H. Guymer, J.S. Wiley, E.L. Fletcher, Loss of function of P2X7 receptor scavenger activity in aging mice: a novel model for investigating the early pathogenesis of age-related macular degeneration, Am. J. Pathol. 187 (2017) 1670–1685, https://doi.org/10.1016/j.ajpath.2017.04.016.
- [167] M.P. Rozing, J.A. Durhuus, M. Krogh Nielsen, Y. Subhi, T.B. Kirkwood, R. G. Westendorp, T.L. Sorensen, Age-related macular degeneration: a two-level model hypothesis, Prog. Retin Eye Res. 76 (2020), 100825, https://doi.org/ 10.1016/j.preteyeres.2019.100825.
- [168] I. Akhtar-Schafer, L. Wang, T.U. Krohne, H. Xu, T. Langmann, Modulation of three key innate immune pathways for the most common retinal degenerative diseases, EMBO Mol. Med. 10 (2018), https://doi.org/10.15252/emmm.201708259.
- [169] T. Sugiyama, Role of P2X7 receptors in the development of diabetic retinopathy, World J. Diabetes 5 (2014) 141–145, https://doi.org/10.4239/wjd.v5.i2.141.
- [170] C. Clapp, N. Diaz-Lezama, E. Adan-Castro, G. Ramirez-Hernandez, B. Moreno-Carranza, A.C. Sarti, S. Falzoni, A. Solini, F. Di Virgilio, Pharmacological blockade of the P2X7 receptor reverses retinal damage in a rat model of type 1 diabetes, Acta Diabetol. 56 (2019) 1031–1036, https://doi.org/10.1007/s00592-019-01343-4.
- [171] C.S. Subauste, The CD40-ATP-P2X 7 receptor pathway: cell to cell cross-talk to promote inflammation and programmed cell death of endothelial cells, Front. Immunol. 10 (2019) 2958, https://doi.org/10.3389/fimmu.2019.02958.
- [172] N. Gupta, K.E. Brown, A.H. Milam, Activated microglia in human retinitis pigmentosa, late-onset retinal degeneration, and age-related macular degeneration, Exp. Eye Res. 76 (2003) 463–471, https://doi.org/10.1016/S0014-4835(02)00332-9.
- [173] C.J. Zeiss, E.A. Johnson, Proliferation of microglia, but not photoreceptors, in the outer nuclear layer of the rd-1 mouse, Investig. Opthalmol. Vis. Sci. 45 (2004) 971, https://doi.org/10.1167/iovs.03-0301.
- [174] L. Zhao, M.K. Zabel, X. Wang, W. Ma, P. Shah, R.N. Fariss, H. Qian, C. N. Parkhurst, W.-B. Gan, W.T. Wong, Microglial phagocytosis of living photoreceptors contributes to inherited retinal degeneration, EMBO Mol. Med. 7 (2015) 1179–1197, https://doi.org/10.15252/emmm.201505298.
- [175] T. Puthussery, E. Fletcher, Extracellular ATP induces retinal photoreceptor apoptosis through activation of purinoceptors in rodents, J. Comp. Neurol. 513 (2009) 430–440, https://doi.org/10.1002/cne.21964.
- [176] No Title, (n.d.). (https://clinicaltrials.gov/).
- [177] M.A. Kass, The ocular hypertension treatment study, J. Glaucoma. 3 (1994) 97–100, https://doi.org/10.1097/00061198-199400320-00001.
  [178] Chauhan, [Canadian glaucoma study:] 3. Impact of risk factors and intraocular
- [178] Chauhan, [Canadian glaucoma study:] 3. Impact of risk factors and intraocular pressure reduction on the rates of visual field change (Archives of Ophthalmology (2010), 128, 10, (1249-1255)), Arch. Ophthalmol. 128 (2010) 1633, https://doi. org/10.1001/archophthalmol.2010.304.
- [179] R. van der Valk, C.A.B. Webers, T. Lumley, F. Hendrikse, M.H. Prins, J.S.A. G. Schouten, A network meta-analysis combined direct and indirect comparisons between glaucoma drugs to rank effectiveness in lowering intraocular pressure, J. Clin. Epidemiol. 62 (2009) 1279–1283, https://doi.org/10.1016/j. iclineni.2008.04.012.
- [180] J.W. Cheng, S.W. Cheng, L. Di Gao, G.C. Lu, R.L. Wei, Intraocular pressurelowering effects of commonly used fixed-combination drugs with timolol: a systematic review and meta-analysis, PLoS One 7 (2012), https://doi.org/ 10.1371/journal.pone.0045079.
- [181] L.J. Lu, J.C. Tsai, J. Liu, Novel pharmacologic candidates for treatment of primary open-angle glaucoma, Yale J. Biol. Med. 90 (2017) 111–118.
- [182] N.A. Mehran, S. Sinha, R. Razeghinejad, New glaucoma medications: latanoprostene bunod, netarsudil, and fixed combination netarsudil-latanoprost, Eye 34 (2020) 72–88, https://doi.org/10.1038/s41433-019-0671-0.
- [183] S. He, D.L. Stankowska, D.Z. Ellis, R.R. Krishnamoorthy, T. Yorio, Targets of Neuroprotection in Glaucoma, (n.d.). (https://doi.org/10.1089/jop.2017.0041).
- [184] W.S. Lambert, L. Ruiz, S.D. Crish, L.A. Wheeler, D.J. Calkins, Brimonidine prevents axonal and somatic degeneration of retinal ganglion cell neurons, Mol. Neurodegener. 6 (2011) 4, https://doi.org/10.1186/1750-1326-6-4.
- [185] T. Krupin, J.M. Liebmann, D.S. Greenfield, R. Ritch, S. Gardiner, A randomized trial of brimonidine versus timolol in preserving visual function: results from the low-pressure glaucoma treatment study, Am. J. Ophthalmol. 151 (2011) 671–681, https://doi.org/10.1016/j.ajo.2010.09.026.
- [186] A. Giaquinta Aranda, A. Fernández Araque, R. Curbelo Rodriguez, A. Rojo Aragues, Glaucoma y antioxidantes: revisión sistemática, Rev. Mex. Oftalmol. 91 (2017) 112–121, https://doi.org/10.1016/j.mexoft.2016.03.007.
- [187] P.A. Campochiaro, Retinal and choroidal neovascularization, J. Cell. Physiol. 184 (2000) 301–310, https://doi.org/10.1002/1097-4652(200009)184:3<301::AID-JCP3>3.0.CO;2-H.
- [188] P.A. Keane, G. De Salvo, D.A. Sim, S. Goverdhan, R. Agrawal, A. Tufai, Clinical Ophthalmology Dovepress Strategies for improving early detection and diagnosis of neovascular age-related macular degeneration, 2015. (https://doi.org/10 .2147/OPTH.S59012).

- [189] K.L. Spooner, C.T. Mhlanga, T.H. Hong, G.K. Broadhead, A.A. Chang, The burden of neovascular age-related macular degeneration: a patient's perspective, Clin. Ophthalmol. (2018) 12–2483, https://doi.org/10.2147/OPTH.S185052.
- [190] V. Kniggendorf, J.L. Dreyfuss, C.V. Regatieri, Age-related macular degeneration: a review of current therapies and new treatments, Arq. Bras. Oftalmol. 83 (2020) 552–561, https://doi.org/10.5935/0004-2749.20200082.
- [191] K. Liu, Y. Song, G. Xu, J. Ye, Z. Wu, X. Liu, X. Dong, M. Zhang, Y. Xing, S. Zhu, X. Chen, Y. Shen, H. Huang, L. Yu, Z. Ke, P.J. Rosenfeld, P.K. Kaiser, G. Ying, X. Sun, X. Xu, R. Li, Q. Wu, X. Wang, F. Kuang, J. Lv, Z. Niu, Conbercept for treatment of neovascular age-related macular degeneration: results of the randomized phase 3 PHOENIX study, Am. J. Ophthalmol. 197 (2019) 156–167, https://doi.org/10.1016/j.ajo.2018.08.026.
- [192] P.U. Dugel, A. Koh, Y. Ogura, G.J. Jaffe, U. Schmidt-Erfurth, D.M. Brown, A. V. Gomes, J. Warburton, A. Weichselberger, F.G. Holz, HAWK and HARRIER: Phase 3, multicenter, randomized, double-masked trials of brolucizumab for neovascular age-related macular degeneration, Ophthalmology 127 (2020) 72–84, https://doi.org/10.1016/j.ophtha.2019.04.017.
- [193] E.N. Dunn, V.S. Sheth, S.M. Hariprasad, An overview of the fovista and rinucumab trials and the fate of anti-PDGF medications, Ophthalmic Surg. Lasers Imaging Retin 48 (2017) 100–104, https://doi.org/10.3928/23258160-20170130-02.
- [194] U. Chakravarthy, C. Bailey, D. Brown, P. Campochiaro, M. Chittum, K. Csaky, A. Tufail, P. Yates, P. Cech, M. Giraudon, P. Delmar, P. Szczesny, J. Sahni, A. Boulay, S. Nagel, S. Fürst-Recktenwald, D. Schwab, Phase I trial of anti-vascular endothelial growth factor/anti-angiopoietin 2 bispecific antibody RG7716 for neovascular age-related macular degeneration, Ophthalmol. Retin. 1 (2017) 474–485, https://doi.org/10.1016/j.oret.2017.03.003.
- [195] P.A. Campochiaro, D.M. Marcus, C.C. Awh, C. Regillo, A.P. Adamis, V. Bantseev, Y. Chiang, J.S. Ehrlich, S. Erickson, W.D. Hanley, J. Horvath, K.F. Maass, N. Singh, F. Tang, G. Barteselli, The port delivery system with ranibizumab for neovascular age-related macular degeneration: results from the randomized phase 2 ladder clinical trial, Ophthalmology 126 (2019) 1141–1154, https://doi.org/ 10.1016/j.ophtha.2019.03.036.
- [196] R.A. Sabbadini, Sphingosine-1-phosphate antibodies as potential agents in the treatment of cancer and age-related macular degeneration, Br. J. Pharmacol. 162 (2011) 1225–1238, https://doi.org/10.1111/j.1476-5381.2010.01118.x.
- [197] B.J. Fowler, B.D. Gelfand, Y. Kim, N. Kerur, V. Tarallo, Y. Hirano, S. Amarnath, D. H. Fowler, M. Radwan, M.T. Young, K. Pittman, P. Kubes, H.K. Agarwal, K. Parang, D.R. Hinton, A. Bastos-Carvalho, S. Li, T. Yasuma, T. Mizutani, R. Yasuma, C. Wright, J. Ambati, Nucleoside reverse transcriptase inhibitors possess intrinsic anti-inflammatory activity, Science 346 (2014) 1000–1003, https://doi.org/10.1126/science.1261754.
- [198] G.A. Rodrigues, E. Shalaev, T.K. Karami, J. Cunningham, N.K.H. Slater, H. M. Rivers, Pharmaceutical development of AAV-based gene therapy products for the eye, Pharm. Res. 36 (2019), https://doi.org/10.1007/s11095-018-2554-7.
- [199] K.G. Csaky, P.U. Dugel, A.J. Pierce, M.A. Fries, D.S. Kelly, R.P. Danis, J. I. Wurzelmann, C.-F. Xu, M. Hossain, T. Trivedi, Clinical evaluation of pazopanib eye drops versus ranibizumab intravitreal injections in subjects with neovascular age-related macular degeneration, Ophthalmology 122 (2015) 579–588, https:// doi.org/10.1016/j.ophtha.2014.09.036.
- [200] A.P. Cabrera, F. Monickaraj, S. Rangasamy, S. Hobbs, P. McGuire, A. Das, Do genomic factors play a role in diabetic retinopathy? J. Clin. Med. 9 (2020) 216, https://doi.org/10.3390/jcm9010216.
- [201] R.A. Kowluru, Diabetic retinopathy, metabolic memory and epigenetic modifications, Vis. Res. 139 (2017) 30–38, https://doi.org/10.1016/j. visres.2017.02.011.
- [202] R.R. Robles-Rivera, J.A. Castellanos-González, C. Olvera-Montaño, R.A. Flores-Martin, A.K. López-Contreras, D.E. Arevalo-Simental, E.G. Cardona-Muñoz, L. M. Roman-Pintos, A.D. Rodríguez-Carrizalez, Adjuvant therapies in diabetic retinopathy as an early approach to delay its progression: the importance of oxidative stress and inflammation, Oxid. Med. Cell. Longev. 2020 (2020) 1–23, https://doi.org/10.1155/2020/3096470.
- [203] J.J. Garcia-Medina, M.D. Pinazo-Duran, M. Garcia-Medina, V. Zanon-Moreno, S. Pons-Vazquez, A 5-year follow-up of antioxidant supplementation in type 2 diabetic retinopathy, Eur. J. Ophthalmol. 21 (2011) 637–643, https://doi.org/ 10.5301/EJO.2010.6212.
- [204] A.P. Chous, S.P. Richer, J.D. Gerson, R.A. Kowluru, The diabetes visual function supplement study (DiVFuSS), Br. J. Ophthalmol. 100 (2016) 227–234, https:// doi.org/10.1136/bjophthalmol-2014-306534.
- [205] B.-J. Hu, Y.-N. Hu, S. Lin, W.-J. Ma, X.-R. Li, Application of Lutein and Zeaxanthin in nonproliferative diabetic retinopathy, Int. J. Ophthalmol. 4 (2011) 303–306, https://doi.org/10.3980/j.issn.2222-3959.2011.03.19.
- [206] M. Lafuente, L. Ortín, M. Argente, J.L. Guindo, M.D. López-Bernal, F.J. López-Román, J.C. Domingo, J. Lajara, Three-year outcomes in a randomized singleblind controlled trial of intravitreal ranibizumab and oral supplementation with docosahexaenoic acid and antioxidants for diabetic macular edema, 2019. (htt ps://eudract.ema.europa.eu/).
- [207] H. Ahmadieh, R. Nourinia, A. Hafezi-Moghadam, H. Sabbaghi, S. Nakao, S. Zandi, M. Yaseri, Z. Tofighi, S. Akbarian, Intravitreal injection of a Rho-kinase inhibitor (fasudil) combined with bevacizumab versus bevacizumab monotherapy for diabetic macular oedema: a pilot randomised clinical trial, Br. J. Ophthalmol. 103 (2019) 922–927, https://doi.org/10.1136/bjophthalmol-2018-312244.
- [208] S. Sepahi, S.A. Mohajeri, S.M. Hosseini, E. Khodaverdi, N. Shoeibi, M. Namdari, S. A.S. Tabassi, Effects of crocin on diabetic maculopathy: a placebo-controlled randomized clinical trial, Am. J. Ophthalmol. 190 (2018) 89–98, https://doi.org/10.1016/j.ajo.2018.03.007.

- [209] M. Filippelli, G. Campagna, P. Vito, T. Zotti, L. Ventre, M. Rinaldi, S. Bartollino, R. dell'Omo, C. Costagliola, Anti-inflammatory effect of curcumin, homotaurine, and vitamin D3 on human vitreous in patients with diabetic retinopathy, Front. Neurol. 11 (2021), https://doi.org/10.3389/fneur.2020.592274.
- [210] M.F. Dias, K. Joo, J.A. Kemp, S.L. Fialho, A. da Silva Cunha, S.J. Woo, Y.J. Kwon, Molecular genetics and emerging therapies for retinitis pigmentosa: basic research and clinical perspectives, Prog. Retin. Eye Res. 63 (2018) 107–131, https://doi.org/10.1016/j.preteyeres.2017.10.004.
- [211] A.V. Garafalo, A.V. Cideciyan, E. Heon, R. Sheplock, A. Pearson, C. WeiYang Yu, A. Sumaroka, G.D. Aguirre, S.G. Jacobson, Progress in treating inherited retinal diseases: early subretinal gene therapy clinical trials and candidates for future initiatives, Prog. Retin Eye Res. 77 (2020), 100827, https://doi.org/10.1016/j. preteyeres.2019.100827.
- [212] M. Yanik, B. Müller, F. Song, J. Gall, F. Wagner, W. Wende, B. Lorenz, K. Stieger, In vivo genome editing as a potential treatment strategy for inherited retinal dystrophies, Prog. Retin. Eye Res. 56 (2017) 1–18, https://doi.org/10.1016/j. preteyeres.2016.09.001.
- [213] D. Terrell, J. Comander, Current stem-cell approaches for the treatment of inherited retinal degenerations, Semin. Ophthalmol. 34 (2019) 287–292, https:// doi.org/10.1080/08820538.2019.1620808.
- [214] F. Newton, R. Megaw, Mechanisms of photoreceptor death in retinitis pigmentosa, Genes 11 (2020), https://doi.org/10.3390/genes11101120.
- [215] E.L. Berson, B. Rosner, M.A. Sandberg, K.C. Hayes, B.W. Nicholson, C. Weigel-DiFranco, W. Willett, A randomized trial of vitamin A and vitamin E supplementation for retinitis pigmentosa, Arch. Ophthalmol. 111 (1993) 761–772, https://doi.org/10.1001/archopht.1993.01090060049022.
- [216] E.L. Berson, Clinical trial of docosahexaenoic acid in patients with retinitis pigmentosareceiving vitamin A treatment, Arch. Ophthalmol. 122 (2004) 1297, https://doi.org/10.1001/archopht.122.9.1297.
- [217] D.R. Hoffman, D.K. Hughbanks-Wheaton, R. Spencer, G.E. Fish, N.S. Pearson, Y.-Z. Wang, M. Klein, A. Takacs, K.G. Locke, D.G. Birch, Docosahexaenoic acid slows visual field progression in X-linked retinitis pigmentosa: ancillary outcomes of the DHAX trial, Investig. Opthalmol. Vis. Sci. 56 (2015) 6646, https://doi.org/ 10.1167/iovs.15-17786.
- [218] H. Bahrami, M. Melia, G. Dagnelie, Lutein supplementation in retinitis pigmentosa: PC-based vision assessment in a randomized double-masked placebocontrolled clinical trial [NCT00029289], BMC Ophthalmol. 6 (2006) 23, https:// doi.org/10.1186/1471-2415-6-23.
- [219] M. Nakazawa, Y. Suzuki, T. Ito, T. Metoki, T. Kudo, H. Ohguro, Long-term effects of Nilvadipine against progression of the central visual field defect in retinitis pigmentosa: an extended study, Biomed. Res. Int. 2013 (2013) 1–6, https://doi. org/10.1155/2013/585729.
- [220] R.W. Humphrey, L.M. Brockway-Lunardi, D.T. Bonk, K.M. Dohoney, J. H. Doroshow, S.J. Meech, M.J. Ratain, S.L. Topalian, D.M. Pardoll, Opportunities and challenges in the development of experimental drug combinations for cancer, JNCI J. Natl. Cancer Inst. 103 (2011) 1222–1226, https://doi.org/10.1093/jnci/ djr246.
- [221] G. Glass, Cardiovascular combinations, Nat. Rev. Drug Discov. 3 (2004) 731–732, https://doi.org/10.1038/nrd1501.
- [222] H.S. Nelson, Advair: Combination treatment with fluticasone propionate/ salmeterol in the treatment of asthma, J. Allergy Clin. Immunol. 107 (2001) 397–416, https://doi.org/10.1067/mai.2001.112939.
- [223] B.A. Larder, S.D. Kemp, P.R. Harrigan, Potential mechanism for sustained antiretroviral efficacy of AZT-3TC combination therapy, Science 269 (1995) 696–699, https://doi.org/10.1126/science.7542804.
- [224] J.L. Hartman IV, Principles for the buffering of genetic variation, Science 291 (2001) 1001–1004, https://doi.org/10.1126/science.291.5506.1001.
- [225] B. Papp, C. Pál, L.D. Hurst, Metabolic network analysis of the causes and evolution of enzyme dispensability in yeast, Nature 429 (2004) 661–664, https:// doi.org/10.1038/nature02636.
- [226] H. Kitano, A robustness-based approach to systems-oriented drug design, Nat. Rev. Drug Discov. 6 (2007) 202–210, https://doi.org/10.1038/nrd2195.
- [227] J.J. Palop, J. Chin, L. Mucke, A network dysfunction perspective on neurodegenerative diseases, Nature 443 (2006) 768–773, https://doi.org/ 10.1038/nature05289.
- [228] A.-L. Barabási, N. Gulbahce, J. Loscalzo, Network medicine: a network-based approach to human disease, Nat. Rev. Genet. 12 (2011) 56–68, https://doi.org/ 10.1038/nrg2918.
- [229] H. Weil-Malherbe, D.C. Wmhington, The biochemistry of the functional psychoses\*, 1967.
- [230] T.T. Ashburn, K.B. Thor, Drug repositioning: identifying and developing new uses for existing drugs, Nat. Rev. Drug Discov. 3 (2004) 673–683, https://doi.org/ 10.1038/nrd1468.
- [231] M.L. Bolognesi, Polypharmacology in a single drug: multitarget drugs, Curr. Med. Chem. 20 (2013) 1639–1645, https://doi.org/10.2174/0929867311320130004.
- [232] H. Li, H. Xiao, L. Lin, D. Jou, V. Kumari, J. Lin, C. Li, Drug design targeting protein–protein interactions (PPIs) using multiple ligand simultaneous docking (MLSD) and drug repositioning: discovery of raloxifene and bazedoxifene as novel inhibitors of IL-6/GP130 interface, J. Med. Chem. 57 (2014) 632–641, https:// doi.org/10.1021/jm401144z.
- [233] C.P. Adams, V.V. Brantner, Estimating the cost of new drug development: is it really \$802 million? Health Aff. 25 (2006) 420–428, https://doi.org/10.1377/ hlthaff.25.2.420.
- [234] A.L. Gassman, C.P. Nguyen, H.V. Joffe, FDA regulation of prescription drugs, N. Engl. J. Med. 376 (2017) 674–682, https://doi.org/10.1056/NEJMra1602972.

- [235] J.A. DiMasi, Z. Smith, K.A. Getz, Assessing the financial benefits of faster development times: the case of single-source versus multi-vendor outsourced biopharmaceutical manufacturing, Clin. Ther. 40 (2018) 963–972, https://doi. org/10.1016/j.clinthera.2018.04.011.
- [236] G. Hubsher, M. Haider, M.S. Okun, Amantadine: the journey from fighting flu to treating Parkinson disease, Neurology 78 (2012) 1096–1099, https://doi.org/ 10.1212/WNL.0b013e31824e8f0d.
- [237] C. Patrono, L.A. García Rodríguez, R. Landolfi, C. Baigent, Low-dose aspirin for the prevention of atherothrombosis, N. Engl. J. Med. 353 (2005) 2373–2383, https://doi.org/10.1056/NEJMra052717.
- [238] C. Bern, J. Adler-Moore, J. Berenguer, M. Boelaert, M. den Boer, R.N. Davidson, C. Figueras, L. Gradoni, D.A. Kafetzis, K. Ritmeijer, E. Rosenthal, C. Royce, R. Russo, S. Sundar, J. Alvar, Reviews of anti-infective agents: liposomal Amphotericin B for the treatment of visceral Leishmaniasis, Clin. Infect. Dis. 43 (2006) 917–924, https://doi.org/10.1086/507530.
- [239] R.I.G. Holt, A.H. Barnett, C.J. Bailey, Bromocriptine: old drug, new formulation and new indication, Diabetes Obes. Metab. 12 (2010) 1048–1057, https://doi. org/10.1111/j.1463-1326.2010.01304.x.
- [240] C. Lerman, R. Niaura, B.N. Collins, P. Wileyto, J. Audrain-McGovern, A. Pinto, L. Hawk, L.H. Epstein, Effect of bupropion on depression symptoms in a smoking cessation clinical trial, Psychol. Addict. Behav. 18 (2004) 362–366, https://doi. org/10.1037/0893-164X.18.4.362.
- [241] V. Jendrossek, Targeting apoptosis pathways by Celecoxib in cancer, Cancer Lett. 332 (2013) 313–324, https://doi.org/10.1016/j.canlet.2011.01.012.
- [242] D.D. Sweeney, M.B. Chancellor, Treatment of stress urinary incontinence with duloxetine hydrochloride, Rev. Urol. 7 (2005) 81–86. (http://www.ncbi.nlm.nih. gov/pubmed/16985814).
- [243] A. Rossi, C. Cantisani, M. Scarnò, A. Trucchia, M.C. Fortuna, S. Calvieri, Finasteride, 1 mg daily administration on male androgenetic alopecia in different age groups: 10-year follow-up, Dermatol. Ther. 24 (2011) 455–461, https://doi. org/10.1111/j.1529-8019.2011.01441.x.
- [244] M. Steiner, S. Steinberg, D. Stewart, D. Carter, C. Berger, R. Reid, D. Grover, D. Streiner, Fluoxetine in the treatment of premenstrual dysphoria, N. Engl. J. Med. 332 (1995) 1529–1534, https://doi.org/10.1056/NEJM199506083322301.
- [245] S. Lilienfeld, Galantamine a novel cholinergic drug with a unique dual mode of action for the treatment of patients with Alzheimer's disease, CNS Drug Rev. 8 (2006) 159–176, https://doi.org/10.1111/j.1527-3458.2002.tb00221.x.
- [246] R.S. King, Gemcitabine. New first-line therapy for pancreatic cancer. Cancer Pract. 4 (n.d.) 353–354. (http://www.ncbi.nlm.nih.gov/pubmed/9128490).
- [247] G.-R. Burmester, A.J. Kivitz, H. Kupper, U. Arulmani, S. Florentinus, S.L. Goss, S. S. Rathmann, R.M. Fleischmann, Efficacy and safety of ascending methotrexate dose in combination with adalimumab: the randomised CONCERTO trial, Ann. Rheum. Dis. 74 (2015) 1037–1044, https://doi.org/10.1136/annrheumdis-2013-204769.
- [248] E.A. Olsen, F.E. Dunlap, T. Funicella, J.A. Koperski, J.M. Swinehart, E.H. Tschen, R.J. Trancik, A randomized clinical trial of 5% topical minoxidil versus 2% topical minoxidil and placebo in the treatment of androgenetic alopecia in men, J. Am. Acad. Dermatol. 47 (2002) 377–385, https://doi.org/10.1067/mjd.2002.124088.
   [249] A. Gershlick, I. De Scheerder, B. Chevalier, A. Stephens-Lloyd, E. Camenzind,
- [249] A. Gershlick, I. De Scheerder, B. Chevalier, A. Stephens-Lloyd, E. Camenzind, C. Vrints, N. Reifart, L. Missault, J.-J. Goy, J.A. Brinker, A.E. Raizner, P. Urban, A. W. Heldman, Inhibition of restenosis with a paclitaxel-eluting, polymer-free coronary stent, Circulation 109 (2004) 487–493, https://doi.org/10.1161/01. CIR 0000109694 58299 A0
- [250] A. Cranney, J.D. Adachi, Benefit-risk assessment of raloxifene in postmenopausal osteoporosis, Drug Saf. 28 (2005) 721–730, https://doi.org/10.2165/00002018-200528080-00006.
- [251] K.D. Sethi, Ropinirole for the treatment of early Parkinson disease: a 12-month experience, Arch. Neurol. 55 (1998) 1211–1216, https://doi.org/10.1001/ archneur.55.9.1211.
- [252] H.A. Ghofrani, I.H. Osterloh, F. Grimminger, Sildenafil: from angina to erectile dysfunction to pulmonary hypertension and beyond, Nat. Rev. Drug Discov. 5 (2006) 689–702, https://doi.org/10.1038/nrd2030.
- [253] S. Lindner, J. Krönke, The molecular mechanism of thalidomide analogs in hematologic malignancies, J. Mol. Med. 94 (2016) 1327–1334, https://doi.org/ 10.1007/s00109-016-1450-z.
- [254] S.L. McElroy, I.M. Arnold, N.A. Shapira, P.E. Keck, N.R. Rosenthal, M.R. Karim, M. Kamin, J.I. Hudson, Topiramate in the treatment of binge eating disorder associated with obesity: a randomized, placebo-controlled trial, Am. J. Psychiatry 160 (2003) 255–261, https://doi.org/10.1176/appi.ajp.160.2.255.
- [255] S. Broder, The development of antiretroviral therapy and its impact on the HIV-1/ AIDS pandemic, Antivir. Res. 85 (2010) 1–18, https://doi.org/10.1016/j. antiviral.2009.10.002.
- [256] P.A. Ascierto, F.M. Marincola, Combination therapy: the next opportunity and challenge of medicine, J. Transl. Med. 9 (2011) 115, https://doi.org/10.1186/ 1479-5876-9-115.
- [257] P.C. Trippier, K. Jansen Labby, D.D. Hawker, J.J. Mataka, R.B. Silverman, Targetand mechanism-based therapeutics for neurodegenerative diseases: strength in numbers, J. Med. Chem. 56 (2013) 3121–3147, https://doi.org/10.1021/ im3015926.
- [258] J. Foucquier, M. Guedj, Analysis of drug combinations: current methodological landscape, Pharmacol. Res. Perspect. 3 (2015), e00149, https://doi.org/10.1002/ prp2.149.
- [259] A. Romero, J. Egea, A.G. García, M.G. López, Synergistic neuroprotective effect of combined low concentrations of galantamine and melatonin against oxidative stress in SH-SY5Y neuroblastoma cells, J. Pineal Res. 49 (2010) 141–148, https:// doi.org/10.1111/j.1600-079X.2010.00778.x.

- [260] M. Sobrado, M. López, F. Carceller, A. García, J. Roda, Combined nimodipine and citicoline reduce infarct size, attenuate apoptosis and increase bcl-2 expression after focal cerebral ischemia, Neuroscience 118 (2003) 107–113, https://doi.org/ 10.1016/S0306-4522(02)00912-0.
- [261] W. Sun, P.E. Sanderson, W. Zheng, Drug combination therapy increases successful drug repositioning, Drug Discov. Today 21 (2016) 1189–1195, https://doi.org/ 10.1016/j.drudis.2016.05.015.
- [262] M.J. Lee, A.S. Ye, A.K. Gardino, A.M. Heijink, P.K. Sorger, G. MacBeath, M. B. Yaffe, Sequential application of anticancer drugs enhances cell death by rewiring apoptotic signaling networks, Cell 149 (2012) 780–794, https://doi.org/ 10.1016/j.cell.2012.03.031.
- [263] Y. Jin, Z. Desta, V. Stearns, B. Ward, H. Ho, K.-H. Lee, T. Skaar, A.M. Storniolo, L. Li, A. Araba, R. Blanchard, A. Nguyen, L. Ullmer, J. Hayden, S. Lemler, R. M. Weinshilbourn, J.M. Rae, D.F. Hayes, D.A. Flockhart, CYP2D6 genotype, antidepressant use, and tamoxifen metabolism during adjuvant breast cancer treatment, JNCI J. Natl. Cancer Inst. 97 (2005) 30–39, https://doi.org/10.1093/ jnci/dji005.
- [264] C.M. Kelly, D.N. Juurlink, T. Gomes, M. Duong-Hua, K.I. Pritchard, P.C. Austin, L. F. Paszat, Selective serotonin reuptake inhibitors and breast cancer mortality in women receiving tamoxifen: a population based cohort study, c693–c693, BMJ 340 (2010), https://doi.org/10.1136/bmj.c693.
- [265] F.P. Guengerich, Inhibition of Cytochrome P450 enzymes by drugs-molecular basis and practical applications, Biomol. Ther. 30 (2022) 1–18, https://doi.org/ 10.4062/biomolther.2021.102.
- [266] B. Williamson, K.E. Dooley, Y. Zhang, D.J. Back, A. Owen, Induction of influx and efflux transporters and cytochrome P450 3A4 in primary human hepatocytes by Rifampin, Rifabutin, and Rifapentine, Antimicrob. Agents Chemother. 57 (2013) 6366–6369, https://doi.org/10.1128/AAC.01124-13.
- [267] P. Csermely, V. Agoston, S. Pongor, The efficiency of multi-target drugs: the network approach might help drug design, Trends Pharmacol. Sci. 26 (2005) 178–182, https://doi.org/10.1016/j.tips.2005.02.007.
- [268] Alzheimer's Drug Discovery Foundation, Alzheimer CTs Report, 2018.
- [269] V. Maneu, P. Lax, N. Cuenca, Current and future therapeutic strategies for the treatment of retinal neurodegenerative diseases, Neural Regen. Res. 17 (2022) 103, https://doi.org/10.4103/1673-5374.314305.
- [270] R. Simó, C. Hernández, M. Porta, F. Bandello, J. Grauslund, S.P. Harding, S. J. Aldington, C. Egan, U. Frydkjaer-Olsen, J. García-Arumí, J. Gibson, G.E. Lang, R. Lattanzio, P. Massin, E. Midena, B. Ponsati, L. Ribeiro, P. Scanlon, C. Lobo, M.Â. Costa, J. Cunha-Vaz, Effects of topically administered neuroprotective drugs in early stages of diabetic retinopathy: results of the EUROCONDOR clinical trial, Diabetes 68 (2019) 457–463, https://doi.org/10.2337/db18-0682.
- [271] T. Iuvone, G. Esposito, D. De Filippis, C. Scuderi, L. Steardo, Cannabidiol: a promising drug for neurodegenerative disorders? CNS Neurosci. Ther. 15 (2009) 65–75, https://doi.org/10.1111/j.1755-5949.2008.00065.x.
- [272] C.E. Dkowicz, S. Titus, M. Kearney, H. Yu, A. Sherman, D. Schoenfeld, D. Hayden, A. Shui, B. Brooks, R. Conwit, D. Felsenstein, D.J. Greenblatt, M. Keroack, J. T. Kissel, R. Miller, J. Rosenfeld, J.D. Rothstein, E. Simpson, N. Tolkoff-Rubin, L. Zinman, J.M. Shefner, Safety and efficacy of ceftriaxone for amyotrophic lateral sclerosis: a multi-stage, randomised, double-blind, placebo-controlled trial, Lancet Neurol. 13 (2014) 1083–1091, https://doi.org/10.1016/S1474-4422(14) 70222-4.
- [273] S.D. Voulgaropoulou, T.A.M.J. van Amelsvoort, J. Prickaerts, C. Vingerhoets, The effect of curcumin on cognition in Alzheimer's disease and healthy aging: a systematic review of pre-clinical and clinical studies, Brain Res. 2019 (1725), 146476, https://doi.org/10.1016/j.brainres.2019.146476.
- [274] M. Nebbioso, F. Franzone, A. Greco, M. Gharbiya, V. Bonfiglio, A. Polimeni, Recent advances and disputes about curcumin in retinal diseases, Clin. Ophthalmol. 15 (2021) 2553–2571. https://doi.org/10.2147/OPTH.S306706.
- [275] M. Mohseni, A. Sahebkar, G. Askari, T.P. Johnston, B. Alikiaii, M. Bagherniya, The clinical use of curcumin on neurological disorders: an updated systematic review of clinical trials, Phyther. Res. (2021), https://doi.org/10.1002/ptr.7273.
- [276] S. Hu, P. Maiti, Q. Ma, X. Zuo, M.R. Jones, G.M. Cole, S.A. Frautschy, Clinical development of curcumin in neurodegenerative disease, Expert Rev. Neurother. 15 (2015) 629–637, https://doi.org/10.1586/14737175.2015.1044981.
- [277] S. Pandaran Sudheeran, D. Jacob, J. Natinga Mulakal, G. Gopinathan Nair, A. Maliakel, B. Maliakel, R. Kuttan, K.I.M. Safety, Tolerance, and enhanced efficacy of a bioavailable formulation of curcumin with fenugreek dietary fiber on occupational stress, J. Clin. Psychopharmacol. 36 (2016) 236–243, https://doi. org/10.1097/JCP.000000000000508.
- [278] R. Jamwal, Bioavailable curcumin formulations: a review of pharmacokinetic studies in healthy volunteers, J. Integr. Med. 16 (2018) 367–374, https://doi.org/ 10.1016/j.joim.2018.07.001.
- [279] I.C. Arellanes, N. Choe, V. Solomon, X. He, B. Kavin, A.E. Martinez, N. Kono, D. P. Buennagel, N. Hazra, G. Kim, L.M. D'Orazio, C. McCleary, A. Sagare, B. V. Zlokovic, H.N. Hodis, W.J. Mack, H.C. Chui, M.G. Harrington, M.N. Braskie, L. S. Schneider, H.N. Yassine, Brain delivery of supplemental docosahexaenoic acid (DHA): a randomized placebo-controlled clinical trial, EbioMedicine 59 (2020), 102883, https://doi.org/10.1016/j.ebiom.2020.102883.
- [280] A. Sala-Vila, E.M. Arenaza-Urquijo, G. Sánchez-Benavides, M. Suárez-Calvet, M. Milà-Alomà, O. Grau-Rivera, J.M. González-de-Echávarri, M. Crous-Bou, C. Minguillón, K. Fauria, G. Operto, C. Falcón, G. Salvadó, R. Cacciaglia, S. Ingala, F. Barkhof, H. Schröder, N. Scarmeas, J.-D. Gispert, J.L. Molinuevo, A. Beteta, A. Brugulat-Serrat, A. Cañas, N. Carranza, C. Deulofeu, R. Dominguez, M. Emilio, L. Hernandez, G. Huesa, J. Huguet, I. Knezevic, P. Marne, T. Menchón, A. Polo, S. Pradas, M. Shekari, A. Soteras, M. Vilanova, N. Vilor-Tejedor, DHA intake relates to better cerebrovascular and neurodegeneration neuroimaging

phenotypes in middle-aged adults at increased genetic risk of Alzheimer disease, Am. J. Clin. Nutr. 113 (2021) 1627–1635, https://doi.org/10.1093/ajcn/nqab016.

- [281] M.C. Sabaner, R. Duman, M. Dogan, M. Akdogan, A. Vurmaz, E. Bozkurt, S. Beysel, Do SGLT2 inhibitors prevent preclinical diabetic retinopathy? A prospective pilot optical coherence tomography angiography study, J. Fr. Ophtalmol. 44 (2021) 1159–1167, https://doi.org/10.1016/j.jfo.2021.01.005.
- [282] US National Library of Medicine. CinicalTrials.gov, n.d. (https://www.clinicaltrials.gov/ct2/show/results/NCT00857259?term=NCT00857259&draw=2&ran k=1).
- [283] H.S. Sandhu, J. Lambert, Y. Xu, H.J. Kaplan, Systemic immunosuppression and risk of age-related macular degeneration, PLoS One 13 (2018), e0203492, https://doi.org/10.1371/journal.pone.0203492.
- [284] M. Jakaria, S. Azam, M.E. Haque, S.-H. Jo, M.S. Uddin, I.-S. Kim, D.-K. Choi, Taurine and its analogs in neurological disorders: focus on therapeutic potential and molecular mechanisms, Redox Biol. 24 (2019), 101223, https://doi.org/ 10.1016/j.redox.2019.101223.
- [285] P. Marino, G. Rossi, G. Campagna, D. Capobianco, C. Costagliola, Effects of Citicoline, Homotaurine, and Vitamin E on contrast sensitivity and visual-related quality of life in patients with primary open-angle glaucoma: a preliminary study, Molecules 25 (2020) 5614, https://doi.org/10.3390/molecules25235614.
- [286] Isradipine versus placebo in early Parkinson disease, Ann. Intern. Med. 172 (2020) 591–598. (https://doi.org/10.7326/M19–2534).
- [287] C.S. Venuto, L. Yang, M. Javidnia, D. Oakes, D. James Surmeier, T. Simuni, Isradipine plasma pharmacokinetics and exposure–response in early Parkinson's disease, Ann. Clin. Transl. Neurol. 8 (2021) 603–612, https://doi.org/10.1002/ acn3.51300.
- [288] M. Khalighi Sikaroudi, S. Saraf-Bank, Z.S. Clayton, S. Soltani, A positive effect of egg consumption on macular pigment and healthy vision: a systematic review and meta-analysis of clinical trials, J. Sci. Food Agric. 101 (2021) 4003–4009, https:// doi.org/10.1002/jsfa.11109.
- [289] K.A. Wesnes, D. Aarsland, C. Ballard, E. Londos, Memantine improves attention and episodic memory in Parkinson's disease dementia and dementia with Lewy bodies, Int. J. Geriatr. Psychiatry 30 (2015) 46–54, https://doi.org/10.1002/ gps.4109.
- [290] I. Leroi, R. Atkinson, R. Overshott, Memantine improves goal attainment and reduces caregiver burden in Parkinson's disease with dementia, Int. J. Geriatr. Psychiatry 29 (2014) 899–905, https://doi.org/10.1002/gps.4077.
- [291] P. Borghammer, M. Vafaee, K. Ostergaard, A. Rodell, C. Bailey, P. Cumming, Effect of memantine on CBF and CMRO 2 in patients with early Parkinson's disease, Acta Neurol. Scand. 117 (2008) 317–323, https://doi.org/10.1111/ j.1600-0404.2007.00943.x.
- [292] R.N. Weinreb, J.M. Liebmann, G.A. Cioffi, I. Goldberg, J.D. Brandt, C.A. Johnson, L.M. Zangwill, S. Schneider, H. Badger, M. Bejanian, Oral memantine for the treatment of glaucoma, Ophthalmology 125 (2018) 1874–1885, https://doi.org/ 10.1016/j.ophtha.2018.06.017.
- [293] L. Blanco-Silvente, D. Capellà, J. Garre-Olmo, J. Vilalta-Franch, X. Castells, Predictors of discontinuation, efficacy, and safety of memantine treatment for Alzheimer's disease: meta-analysis and meta-regression of 18 randomized clinical trials involving 5004 patients, BMC Geriatr. 18 (2018) 168, https://doi.org/ 10.1186/s12877-018-0857-5.
- [294] A.L. Boxer, D.S. Knopman, D.I. Kaufer, M. Grossman, C. Onyike, N. Graf-Radford, M. Mendez, D. Kerwin, A. Lerner, C.-K. Wu, M. Koestler, J. Shapira, K. Sullivan, K. Klepac, K. Lipowski, J. Ullah, S. Fields, J.H. Kramer, J. Merrilees, J. Neuhaus, M.M. Mesulam, B.L. Miller, Memantine in patients with frontotemporal lobar degeneration: a multicentre, randomised, double-blind, placebo-controlled trial, Lancet Neurol. 12 (2013) 149–156, https://doi.org/10.1016/S1474-4422(12) 70320-4.
- [295] D.F. Romdhoniyyah, S.P. Harding, C.P. Cheyne, N.A.V. Beare, Metformin, a potential role in age-related macular degeneration: a systematic review and metaanalysis, Ophthalmol. Ther. 10 (2021) 245–260, https://doi.org/10.1007/ s40123-021-00344-3.
- [296] R. Howard, O. Zubko, R. Bradley, E. Harper, L. Pank, J. O'Brien, C. Fox, N. Tabet, G. Livingston, P. Bentham, R. McShane, A. Burns, C. Ritchie, S. Reeves, S. Lovestone, C. Ballard, W. Noble, R. Nilforooshan, G. Wilcock, R. Gray, Minocycline at 2 different dosages vs placebo for patients with mild Alzheimer disease, JAMA Neurol. 77 (2020) 164, https://doi.org/10.1001/ jamaneurol.2019.3762.
- [297] P.H. Gordon, D.H. Moore, R.G. Miller, J.M. Florence, J.L. Verheijde, C. Doorish, J. F. Hilton, G.M. Spitalny, R.B. MacArthur, H. Mitsumoto, H.E. Neville, K. Boylan, T. Mozaffar, J.M. Belsh, J. Ravits, R.S. Bedlack, M.C. Graves, L.F. McCluskey, R. J. Barohn, R. Tandan, Efficacy of minocycline in patients with amyotrophic lateral sclerosis: a phase III randomised trial, Lancet Neurol. 6 (2007) 1045–1053, https://doi.org/10.1016/S1474-4422(07)70270-3.
- [298] H.-S. Kim, Y.-H. Suh, Minocycline and neurodegenerative diseases, Behav. Brain Res. 196 (2009) 168–179, https://doi.org/10.1016/j.bbr.2008.09.040.
- [299] B. Lawlor, R. Segurado, S. Kennelly, M.G.M. Olde Rikkert, R. Howard, F. Pasquier, A. Börjesson-Hanson, M. Tsolaki, U. Lucca, D.W. Molloy, R. Coen, M.W. Riepe, J. Kálmán, R.A. Kenny, F. Cregg, S. O'Dwyer, C. Walsh, J. Adams, R. Banzi, L. Breuilh, L. Daly, S. Hendrix, P. Aisen, S. Gaynor, A. Sheikhi, D.G. Taekema, F. R. Verhey, R. Nemni, F. Nobili, M. Franceschi, G. Frisoni, O. Zanetti, A. Konsta, O. Anastasios, S. Nenopoulou, F. Tsolaki-Tagaraki, M. Pakaski, O. Dereeper, V. de la Sayette, O. Sénéchal, I. Lavenu, A. Devendeville, G. Calais, F. Crawford, M. Mullan, Nilvadípine in mild to moderate Alzheimer disease: a randomised controlled trial, PLoS Med. 15 (2018), e1002660, https://doi.org/10.1371/ journal.pmed.1002660.

- [300] W.T. Wong, S. Dresner, F. Forooghian, T. Glaser, L. Doss, M. Zhou, D. Cunningham, K. Shimel, M. Harrington, K. Hammel, C.A. Cukras, F.L. Ferris, E. Y. Chew, Treatment of geographic atrophy with subconjunctival sirolimus: results of a Phase I/II clinical trial, Investig. Opthalmol. Vis. Sci. 54 (2013) 2941, https:// doi.org/10.1167/iovs.13-11650.
- [301] D.C. Matthews, X. Mao, K. Dowd, D. Tsakanikas, C.S. Jiang, C. Meuser, R. D. Andrews, A.S. Lukic, J. Lee, N. Hampilos, N. Shafian, M. Sano, P. David Mozley, H. Fillit, B.S. McEwen, D.C. Shungu, A.C. Pereira, Riluzole, a glutamate modulator, slows cerebral glucose metabolism decline in patients with Alzheimer's disease, Brain (2021), https://doi.org/10.1093/brain/awab222.
- [302] S. Vucic, M.C. Kiernan, P. Menon, W. Huynh, A. Rynders, K.S. Ho, R. Glanzman, M.T. Hotchkin, Study protocol of RESCUE-ALS: a Phase 2, randomised, doubleblind, placebo-controlled study in early symptomatic amyotrophic lateral sclerosis patients to assess bioenergetic catalysis with CNM-Au8 as a mechanism to slow disease progression, BMJ Open 11 (2021), e041479, https://doi.org/ 10.1136/bmjopen-2020-041479.
- [303] R.M. Pascuzzi, J. Shefner, A.S. Chappell, J.S. Bjerke, R. Tamura, V. Chaudhry, L. Clawson, L. Haas, J.D. Rothstein, A phase II trial of talampanel in subjects with amyotrophic lateral sclerosis, Amyotroph. Lateral Scler. 11 (2010) 266–271, https://doi.org/10.3109/17482960903307805.
- [304] H.J. Wobst, K.L. Mack, D.G. Brown, N.J. Brandon, J. Shorter, The clinical trial landscape in amyotrophic lateral sclerosis—past, present, and future, Med. Res. Rev. 40 (2020) 1352–1384, https://doi.org/10.1002/med.21661.
- [305] M.F. Beal, D. Oakes, I. Shoulson, C. Henchcliffe, W.R. Galpern, R. Haas, J. L. Juncos, J.G. Nutt, T.S. Voss, B. Ravina, C.M. Shults, K. Helles, V. Snively, M. F. Lew, B. Griebner, A. Watts, S. Gao, E. Pourcher, L. Bond, K. Kompoliti, P. Agarwal, C. Sia, M. Jog, L. Cole, M. Sultana, R. Kurlan, I. Richard, C. Deeley, C. H. Waters, A. Figueroa, A. Arkun, M. Brodsky, W.G. Ondo, C.B. Hunter, J. Jimenez-Shahed, A. Palao, J.M. Miyasaki, J. So, J. Tetrud, L. Reys, K. Smith, C. Singer, A. Blenke, D.S. Russell, C. Cotto, J.H. Friedman, M. Lannon, L. Zhang, E. Drasby, R. Kumar, T. Subramanian, D.S. Ford, D.A. Grimes, D. Cote, J. Conway, A.D. Siderowf, M.L. Evatt, B. Sommerfeld, A.N. Lieberman, M.S. Okun, R. L. Rodriguez, S. Merritt, C.L. Swartz, W.R.W. Martin, P. King, N. Stover, S. Guthrie, R.L. Watts, A. Ahmed, H.H. Fernandez, A. Winters, Z. Mari, T. M. Dawson, B. Dunlop, A.S. Feigin, B. Shannon, M.J. Nirenberg, M. Ogg, S. A. Ellias, C.-A. Thomas, K. Frei, I. Bodis-Wollner, S. Glazman, T. Mayer, R. A. Hauser, R. Pahwa, A. Langhammer, R. Ranawaya, L. Derwent, K.D. Sethi, B. Farrow, R. Prakash, I. Litvan, A. Robinson, A. Sahay, M. Gartner, V.K. Hinson, S. Markind, M. Pelikan, J.S. Perlmutter, J. Hartlein, E. Molho, S. Evans, C. H. Adler, A. Duffy, M. Lind, L. Elmer, K. Davis, J. Spears, S. Wilson, M.A. Leehey, N. Hermanowicz, S. Niswonger, H.A. Shill, S. Obradov, A. Rajput, M. Cowper, S. Lessig, D. Song, D. Fontaine, C. Zadikoff, K. Williams, K.A. Blindauer, J. Bergholte, C.S. Propsom, M.A. Stacy, J. Field, D. Mihaila, M. Chilton, E.Y. Uc, J. Sieren, D.K. Simon, L. Kraics, A. Silver, J.T. Boyd, R.W. Hamill, C. Ingvoldstad, J. Young, K. Thomas, S.K. Kostyk, J. Wojcieszek, R.F. Pfeiffer, M. Panisset, M. Beland, S.G. Reich, M. Cines, N. Zappala, J. Rivest, R. Zweig, L.P. Lumina, C. L. Hilliard, S. Grill, M. Kellermann, P. Tuite, S. Rolandelli, U.J. Kang, J. Young, J. Rao, M.M. Cook, L. Severt, K. Boyar, A randomized clinical trial of high-dosage coenzyme Q10 in early Parkinson disease, JAMA Neurol. 71 (2014) 543, https:// doi org/10/1001/jamaneurol/2014/131 [306] R.H. Karakahya, T.Ş. Özcan, Salvage of the retinal ganglion cells in transition phase in Alzheimer's disease with topical coenzyme Q10: is it possible? Graefe's
- Arch. Clin. Exp. Ophthalmol. 258 (2020) 411–418, https://doi.org/10.1007/ s00417-019-04544-3.
   [307] A. Kumar, A. Singh, A review on mitochondrial restorative mechanism of
- [307] A. Kunai, A. Singi, A review on informular resonative mechanism of antioxidants in Alzheimer's disease and other neurological conditions, Front. Pharmacol. 6 (2015), https://doi.org/10.3389/fphar.2015.00206.
- [308] J. Hernández-Ojeda, E.G. Cardona-Muñoz, L.M. Román-Pintos, R. Troyo-Sanromán, P.C. Ortiz-Lazareno, M.A. Cárdenas-Meza, S. Pascoe-González, A. G. Miranda-Díaz, The effect of ubiquinone in diabetic polyneuropathy: a randomized double-blind placebo-controlled study, J. Diabetes Complicat. 26 (2012) 352–358, https://doi.org/10.1016/j.jdiacomp.2012.04.004.
- [309] M. Akiyama, Y. Ikeda, N. Yoshida, S. Notomi, Y. Murakami, T. Hisatomi, H. Enaida, T. Ishibashi, Therapeutic efficacy of topical unoprostone isopropyl in retinitis pigmentosa, Acta Ophthalmol. 92 (2014) e229–e234, https://doi.org/ 10.1111/aos.12293.
- [310] C. Shiragami, M. Miyake, A. Fujiwara, Y. Morizane, A. Tsujikawa, A. Yamashita, F. Shiraga, Effect of topical isopropyl unoprostone on macular atrophy progression in eyes with exudative age-related macular degeneration, Medicine 96 (2017), e6422, https://doi.org/10.1097/MD.00000000006422.
- [311] Bohn, Carotenoids and markers of oxidative stress in human observational studies and intervention trials: implications for chronic diseases, Antioxidants 8 (2019) 179, https://doi.org/10.3390/antiox8060179.
- [312] W. Stahl, W. Schwarz, H. Sies, Human serum concentrations of all-trans β- and α-carotene but not 9-cis β-carotene increase upon ingestion of a natural isomer mixture obtained from Dunaliella salina (Betatene), J. Nutr. 123 (1993) 847–851, https://doi.org/10.1093/jn/123.5.847.
- [313] Y. Rotenstreich, D. Harats, A. Shaish, E. Pras, M. Belkin, Treatment of a retinal dystrophy, fundus albipunctatus, with oral 9-cis- -carotene, Br. J. Ophthalmol. 94 (2010) 616–621, https://doi.org/10.1136/bjo.2009.167049.
- [314] D. Kaur, T. Behl, A. Sehgal, S. Singh, N. Sharma, S. Chigurupati, A. Alhowail, A. Abdeen, S.F. Ibrahim, C. Vargas-De-La-Cruz, M. Sachdeva, S. Bhatia, A. Al-Harrasi, S. Bungau, Decrypting the potential role of α-lipoic acid in Alzheimer's disease, Life Sci. 284 (2021), 119899, https://doi.org/10.1016/j. lifs.2021.119899.

- [315] B.J. Kim, A. Hunter, A.J. Brucker, P. Hahn, K. Gehrs, A. Patel, A.O. Edwards, Y. Li, R.N. Khurana, I. Nissim, E. Daniel, J. Grunwald, G.-S. Ying, M. Pistilli, M. G. Maguire, J.L. Dunaief, Orally administered alpha lipoic acid as a treatment for geographic atrophy, Ophthalmol. Retin. 4 (2020) 889–898, https://doi.org/ 10.1016/j.oret.2020.03.019.
- [316] S. Bohlen, I. Paty, M. Volteau, I. Meyer, W. Rein, C. Kosinski, R. Reilmann, J07 What can we learn from a phase ii study with BN82451B in hd beyond safety and tolerability – clinical versus objective motor measures, in: Clinical Therapeutics, BMJ Publishing Group Ltd, 2018, https://doi.org/10.1136/jnnp-2018-EHDN.267.
- [317] S. Babu, E.A. Macklin, K.E. Jackson, E. Simpson, K. Mahoney, H. Yu, J. Walker, Z. Simmons, W.S. David, P.E. Barkhaus, L. Simionescu, M.M. Dimachkie, A. Pestronk, J.S. Salameh, M.D. Weiss, B.R. Brooks, D. Schoenfeld, J. Shefner, S. Aggarwal, M.E. Cudkowicz, N. Atassi, Selection design phase II trial of high dosages of tamoxifen and creatine in amyotrophic lateral sclerosis, Amyotroph. Lateral Scler. Front. Degener. 21 (2020) 15–23, https://doi.org/10.1080/ 21678421.2019.1672750.
- [318] C.J. Xu, W.E. Klunk, J.N. Kanfer, Q. Xiong, G. Miller, J.W. Pettegrew, Phosphocreatine-dependent glutamate uptake by synaptic vesicles, J. Biol. Chem. 271 (1996) 13435–13440, https://doi.org/10.1074/jbc.271.23.13435.
- [319] N.O. Mansour, M.A. Shama, R.H. Werida, The effect of doxycycline on neuronspecific enolase in patients with traumatic brain injury: a randomized controlled trial, 204062232110243, Ther. Adv. Chronic Dis. 12 (2021), https://doi.org/ 10.1177/20406223211024362.
- [320] D. Varges, H. Manthey, U. Heinemann, C. Ponto, M. Schmitz, W.J. Schulz-Schaeffer, A. Krasnianski, M. Breithaupt, F. Fincke, K. Kramer, T. Friede, I. Zerr, Doxycycline in early CJD: a double-blinded randomised phase II and observational study, J. Neurol. Neurosurg. Psychiatry 88 (2017) 119–125, https://doi.org/10.1136/jnnp-2016-313541.
- [321] I.U. Scott, G.R. Jackson, D.A. Quillen, M. Larsen, R. Klein, J. Liao, S. Holfort, I. C. Munch, T.W. Gardner, Effect of doxycycline vs placebo on retinal function and diabetic retinopathy progression in patients with severe nonproliferative or non–high-risk proliferative diabetic retinopathy, JAMA Ophthalmol. 132 (2014) 535, https://doi.org/10.1001/jamaophthalmol.2014.93.
- [322] A. Al-Chalabi, A. Chiò, C. Merrill, G. Oster, R. Bornheimer, W. Agnese, S. Apple, Clinical staging in amyotrophic lateral sclerosis: analysis of Edaravone Study 19, J. Neurol. Neurosurg. Psychiatry 92 (2021) 165–171, https://doi.org/10.1136/ jnnp-2020-323271.
- [323] J. Shefner, T. Heiman-Patterson, E.P. Pioro, M. Wiedau-Pazos, S. Liu, J. Zhang, W. Agnese, S. Apple, Long-term edaravone efficacy in amyotrophic lateral sclerosis: post-hoc analyses of Study 19 (MCI186–19), Muscle Nerve 61 (2020) 218–221, https://doi.org/10.1002/mus.26740.
- [324] J.J. Ferreira, A. Rosser, D. Craufurd, F. Squitieri, N. Mallard, B. Landwehrmeyer, Ethyl-eicosapentaenoic acid treatment in Huntington's disease: a placebocontrolled clinical trial, Mov. Disord. 30 (2015) 1426–1429, https://doi.org/ 10.1002/mds.26308.
- [325] Bayer, A Study That Uses Data From Routine Eye Examinations of Patients Participating in Studies FIDELIO-DKD and FIGARO-DKD to Explore Whether Finerenone Can Delay the Progression of a Diabetes Complication That Affects the Eyes (Diabetic Retinopathy,DR), n.d. (https://ichgcp.net/clinical-trials-registry/ NCT04477707).
- [326] M.W. Koch, S. Kaur, K. Sage, J. Kim, M. Levesque-Roy, G. Cerchiaro, V.W. Yong, G.R. Cutter, L.M. Metz, Hydroxychloroquine for primary progressive multiple sclerosis, Ann. Neurol. 90 (2021) 940–948, https://doi.org/10.1002/ana.26239.
- [327] W.A. Van Gool, H.C. Weinstein, P.K. Scheltens, G.J. Walstra, Effect of hydroxychloroquine on progression of dementia in early Alzheimer's disease: an 18-month randomised, double-blind, placebo-controlled study, Lancet 358 (2001) 455–460, https://doi.org/10.1016/S0140-6736(01)05623-9.
- [328] C. Rotermund, G. Machetanz, J.C. Fitzgerald, The therapeutic potential of metformin in neurodegenerative diseases, Front. Endocrinol. 9 (2018), https:// doi.org/10.3389/fendo.2018.00400.
- [329] E.Y. Chew, T.E. Clemons, E. Agrón, L.J. Launer, F. Grodstein, P.S. Bernstein, Effect of Omega-3 fatty acids, Lutein/Zeaxanthin, or other nutrient supplementation on cognitive function, JAMA 314 (2015) 791, https://doi.org/10.1001/ jama.2015.9677.
- [330] B.J. Snow, F.L. Rolfe, M.M. Lockhart, C.M. Frampton, J.D. O'Sullivan, V. Fung, R. A.J. Smith, M.P. Murphy, K.M. Taylor, Protect Study Group, A double-blind, placebo-controlled study to assess the mitochondria-targeted antioxidant MitoQ as a disease-modifying therapy in Parkinson's disease, Mov. Disord. 25 (2010) 1670–1674, https://doi.org/10.1002/mds.23148.
- [331] D. Blum, A. Chtarto, L. Tenenbaum, J. Brotchi, M. Levivier, Clinical potential of minocycline for neurodegenerative disorders, Neurobiol. Dis. 17 (2004) 359–366, https://doi.org/10.1016/j.nbd.2004.07.012.
- [332] G. Tardiolo, P. Bramanti, E. Mazzon, Overview on the effects of N-acetylcysteine in neurodegenerative diseases, Molecules 23 (2018) 3305, https://doi.org/ 10.3390/molecules23123305.
- [333] Š. Šalamon, B. Kramar, T.P. Marolt, B. Poljšak, I. Milisav, Medical and dietary uses of N-acetylcysteine, Antioxidants 8 (2019) 111, https://doi.org/10.3390/ antiox8050111.
- [334] Pioglitazone in early Parkinson's disease: a phase 2, multicentre, double-blind, randomised trial, Lancet Neurol. 14 (2015) 795–803. (https://doi. org/10.1016/S1474-4422(15)00144-1).
- [335] A randomized, double-blind, placebo-controlled trial of pridopidine in Huntington's disease, Mov. Disord. 28 (2013) 1407–1415. (https://doi.org/ 10.1002/mds.25362).

- [336] M. Jabłońska, K. Grzelakowska, B. Wiśniewski, E. Mazur, K. Leis, P. Gałązka, Pridopidine in the treatment of Huntington's disease, Rev. Neurosci. 31 (2020) 441–451, https://doi.org/10.1515/revneuro-2019-0085.
- [337] M.D. Davis, M.J. Sheetz, L.P. Aiello, R.C. Milton, R.P. Danis, X. Zhi, A. Girach, M. C. Jimenez, L. Vignati, Effect of Ruboxistaurin on the visual acuity decline associated with long-standing diabetic macular edema, Investig. Opthalmol. Vis. Sci. 50 (2009) 1, https://doi.org/10.1167/iovs.08-2473.
- [338] J. Lasierra-Cirujeda, M.J.A. Pascual-Salcedo, M.M.A. Pascual-Salcedo, Sulodexide and Alzheimer's disease: a preliminary prospective study, World J. Cardiovasc. Dis. 06 (2016) 54–71, https://doi.org/10.4236/wjcd.2016.62007.
- [339] J.H. Song, H.S. Chin, O.W. Kwon, S.J. Lim, H.K. Kim, Effect of sulodexide in patients with non-proliferative diabetic retinopathy: diabetic retinopathy sulodexide study (DRESS), Graefe's Arch. Clin. Exp. Ophthalmol. 253 (2015) 829–837, https://doi.org/10.1007/s00417-014-2746-8.
- [340] D. Hill, C. Compagnoni, M.F. Cordeiro, Investigational neuroprotective compounds in clinical trials for retinal disease, Expert Opin. Investig. Drugs 30 (2021) 571–577, https://doi.org/10.1080/13543784.2021.1896701.
- [341] N. Pradhan, C. Singh, A. Singh, Coenzyme Q10 a mitochondrial restorer for various brain disorders, Naunyn. Schmiede Arch. Pharmacol. 394 (2021) 2197–2222, https://doi.org/10.1007/s00210-021-02161-8.
- [342] M.A. Schwarzschild, A. Ascherio, M.F. Beal, M.E. Cudkowicz, G.C. Curhan, J. M. Hare, D.C. Hooper, K.D. Kieburtz, E.A. Macklin, D. Oakes, A. Rudolph, I. Shoulson, M.K. Tennis, A.J. Espay, M. Gartner, A. Hung, G. Bwala, R. Lenehan, E. Encarnacion, M. Ainslie, R. Castillo, D. Togasaki, G. Barles, J.H. Friedman, L. Niles, J.H. Carter, M. Murray, C.G. Goetz, J. Jaglin, A. Ahmed, D.S. Russell, C. Cotto, J.L. Goudreau, D. Russell, S.A. Parashos, P. Ede, M.H. Saint-Hilaire, C.-A. Thomas, R. James, M.A. Stacy, J. Johnson, L. Gauger, J. Antonelle de Marcaida, S. Thurlow, S.H. Isaacson, L. Carvajal, J. Rao, M. Cook, C. Hope-Porche, L. McClurg, D.L. Grasso, R. Logan, C. Orme, T. Ross, A.F.D. Brocht, R. Constantinescu, S. Sharma, C. Venuto, J. Weber, K. Eaton, Inosine to increase serum and cerebrospinal fluid urate in Parkinson disease, JAMA Neurol. 71 (2014) 141, https://doi.org/10.1001/jamaneurol.2013.5528.
- [343] N.K. Scripsema, D.-N. Hu, R.B. Rosen, Lutein, Zeaxanthin, and meso -Zeaxanthin in the clinical management of eye disease, J. Ophthalmol. 2015 (2015) 1–13, https://doi.org/10.1155/2015/865179.
- [344] J. Loughman, E. Loskutova, J.S. Butler, W.F. Siah, C. O'Brien, Macular pigment response to Lutein, Zeaxanthin, and meso-zeaxanthin supplementation in openangle glaucoma, Ophthalmol. Sci. 1 (2021), 100039, https://doi.org/10.1016/j. xops.2021.100039.
- [345] A. Jucaite, P. Svenningsson, J.O. Rinne, Z. Cselényi, K. Varnäs, P. Johnström, N. Amini, A. Kirjavainen, S. Helin, M. Minkwitz, A.R. Kugler, J.A. Posener, S. Budd, C. Halldin, A. Varrone, L. Farde, Effect of the myeloperoxidase inhibitor AZD3241 on microglia: a PET study in Parkinson's disease, Brain 138 (2015) 2687–2700, https://doi.org/10.1093/brain/awv184.
- [346] E.C. Keystone, M.M. Wang, M. Layton, S. Hollis, I.B. McInnes, Clinical evaluation of the efficacy of the P2X 7 purinergic receptor antagonist AZD9056 on the signs and symptoms of rheumatoid arthritis in patients with active disease despite treatment with methotrexate or sulphasalazine, Ann. Rheum. Dis. 71 (2012) 1630–1635, https://doi.org/10.1136/annrheumdis-2011-143578.
- [347] Naproxen and celecoxib do not prevent AD in early results from a randomized controlled trial, Neurology 68 (2007) 1800–1808. (https://doi.org/10.1212/01. wnl.0000260269.93245.d2).
- [348] D. Athauda, K. Maclagan, S.S. Skene, M. Bajwa-Joseph, D. Letchford, K. Chowdhury, S. Hibbert, N. Budnik, L. Zampedri, J. Dickson, Y. Li, I. Aviles-Olmos, T.T. Warner, P. Limousin, A.J. Lees, N.H. Greig, S. Tebbs, T. Foltynie, Exenatide once weekly versus placebo in Parkinson's disease: a randomised, double-blind, placebo-controlled trial, Lancet 390 (2017) 1664–1675, https:// doi.org/10.1016/S0140-6736(17)31585-4.
- [349] B. Zhou, J. Zissimopoulos, H. Nadeem, M.A. Crane, D. Goldman, J.A. Romley, Association between exenatide use and incidence of Alzheimer's disease, Alzheimer's Dement. Transl. Res. Clin. Interv. 7 (2021), https://doi.org/10.1002/ trc2.12139.
- [350] I.A. Zavalishin, E.I. Gusev, N.N. Iakhno, M.A. Ronkin, T.E. Shmidt, T.L. Demina, A.I. Kugoev, L.S. Adarcheva, A.S. Niiazbekova, M.N. Zakharova, A.B. Peresedova, G.G. Toropina, L.A. Klishevskaia, I.M. Maksimenko, M.V. Krotenkova, S. N. Konovalov, O.I. Rebrova, T.D. Zhuchenko, [Results of open post-registration clinical trials of copaxone in patients with multiple sclerosis], Zh. Nevrol. Psikhiatrii Im. S.S. Korsakova. Suppl. (2002) 59–64. (http://www.ncbi.nlm.nih. gov/pubmed/12418394).
- [351] A.N. Boyko, N.Y. Lashch, S.N. Sharanova, M.N. Zakharova, O. V. Trifonova, T.O. Simaniv, E. V. Lysogorskaya, O.E. Guryanov, S. V. Kotov, T.I. Iakushin, V.Y. Lizhdvoy, Y.A. Belova, F.A. Khabiro, N.N. Babichev, T.I. Khaibullin, E. V. Granatov, L.A. Averyanova, D. V. Sazonov, M.M. Odinak, Y. V. Trinitatsky, L.A. Tsukurov, A.I. Sergeeva, R.A. Ivanov, M.S. Shustova, [Comparative, placebo-controlled clinical study of efficacy and safety of glatiramer acetate 20 mg in patients with relapsing-remitting multiple sclerosis: results of the first year of the study], Zhurnal Nevrol. i Psikhiatrii Im. S.S. Korsakova. 116 (n.d.) 61–67. (https://doi.org/10.17116/jnevro201611610261–67).
- [352] A. Villarejo-Galende, M. González-Sánchez, V.A. Blanco-Palmero, S. Llamas-Velasco, J. Benito-León, Non-steroidal anti-inflammatory drugs as candidates for the prevention or treatment of Alzheimer's disease: do they still have a role? Curr. Alzheimer Res. 17 (2021) 1013–1022, https://doi.org/10.2174/ 1567205017666201127163018.
- [353] G. Comi, Y. Dadon, N. Sasson, J.R. Steinerman, V. Knappertz, T.L. Vollmer, A. Boyko, P. Vermersch, T. Ziemssen, X. Montalban, F.D. Lublin, M.A. Rocca, R. Volkinshtein, S. Rubinchick, N. Halevy, M. Filippi, CONCERTO: a randomized,

placebo-controlled trial of oral laquinimod in relapsing-remitting multiple sclerosis, 13524585211032804, Mult. Scler. (2021), https://doi.org/10.1177/13524585211032803.

- [354] G. Giovannoni, V. Knappertz, J.R. Steinerman, A.P. Tansy, T. Li, S. Krieger, A. Uccelli, B.M.J. Uitdehaag, X. Montalban, H.-P. Hartung, M. Pia Sormani, B.A. C. Cree, F. Lublin, F. Barkhof, A randomized, placebo-controlled, phase 2 trial of laquinimod in primary progressive multiple sclerosis, Neurology 95 (2020) e1027–e1040, https://doi.org/10.1212/WNL.000000000010284.
- [355] V.W. Henderson, E. Roberts, C. Wimer, E.L. Bardolph, H.C. Chui, A.R. Damasio, P. J. Eslinger, M.F. Folstein, L.S. Schneider, E.L. Teng, L.E. Tune, L.P. Weiner, P. J. Whitehouse, Multicenter trial of naloxone in alzheimer's disease, Ann. Neurol. 25 (1989) 404–406, https://doi.org/10.1002/ana.410250413.
- [356] L.A. Hershey, R.B. Lipton, Naproxen for presymptomatic Alzheimer disease, Neurology 92 (2019) 829–830, https://doi.org/10.1212/ WNL.000000000007233.
- [357] J. Fernández-Albarral, R. de Hoz, A. Ramírez, I. López-Cuenca, E. Salobrar-García, M. Pinazo-Durán, J. Ramírez, J. Salazar, Beneficial effects of saffron (Crocus sativus L.) in ocular pathologies, particularly neurodegenerative retinal diseases, Neural Regen. Res. 15 (2020) 1408, https://doi.org/10.4103/1673-5374.274325.
- [358] S. Sepahi, A. Ghorani-Azam, S.M. Hossieni, S.A. Mohajeri, E. Khodaverdi, Pharmacological effects of saffron and its constituents in ocular disorders from in vitro studies to clinical trials: a systematic review, Curr. Neuropharmacol. 19 (2021) 392–401, https://doi.org/10.2174/1570159X18666200507083346.
- [359] L. Todd, C. Zelinka, Valproic acid for a treatment of retinitis pigmentosa: reasons for optimism and caution, J. Neurosci. 37 (2017) 5215–5217, https://doi.org/ 10.1523/JNEUROSCI.0774-17.2017.
- [360] K. Liu, B. Gui, Y. Sun, N. Shi, Z. Gu, T. Zhang, X. Sun, Inhibition of L-type Ca2+ channels by curcumin requires a novel protein kinase-theta isoform in rat hippocampal neurons, Cell Calcium 53 (2013) 195–203, https://doi.org/ 10.1016/j.ceca.2012.11.014.
- [361] J. Tabeshpour, S. Banaeeyeh, F. Eisvand, T. Sathyapalan, M. Hashemzaei, A. Sahebkar, Effects of curcumin on ion channels and pumps: a review, iub.2054, IUBMB Life (2019), https://doi.org/10.1002/iub.2054.
- [362] A.C. Uğuz, A. Öz, M. Nazıroğlu, Curcumin inhibits apoptosis by regulating intracellular calcium release, reactive oxygen species and mitochondrial depolarization levels in SH-SY5Y neuronal cells, J. Recept. Signal Transduct. 36 (2016) 395–401, https://doi.org/10.3109/10799893.2015.1108337.
- [363] C. Muangnoi, U. Sharif, P. Ratnatilaka Na Bhuket, P. Rojsitthisak, L. Paraoan, Protective effects of curcumin ester prodrug, curcumin diethyl disuccinate against H2O2-induced oxidative stress in human retinal pigment epithelial cells: potential therapeutic avenues for age-related macular degeneration, Int. J. Mol. Sci. 20 (2019) 3367, https://doi.org/10.3390/ijms20133367.
- [364] Z. Ran, Y. Zhang, X. Wen, J. Ma, Curcumin inhibits high glucose-induced inflammatory injury in human retinal pigment epithelial cells through the ROS-PI3K/AKT/mTOR signaling pathway, Mol. Med. Rep. (2018), https://doi. org/10.3892/mmr.2018.9749.
- [365] W. Zhang, Y. Guo, W. Han, M. Yang, L. Wen, K. Wang, P. Jiang, Curcumin relieves depressive-like behaviors via inhibition of the NLRP3 inflammasome and kynurenine pathway in rats suffering from chronic unpredictable mild stress, Int. Immunopharmacol. 67 (2019) 138–144, https://doi.org/10.1016/j. intimp.2018.12.012.
- [366] Z. Wang, W. Ren, F. Zhao, Y. Han, C. Liu, K. Jia, Curcumin amends Ca2+ dysregulation in microglia by suppressing the activation of P2X7 receptor, Mol. Cell. Biochem. 465 (2020) 65–73, https://doi.org/10.1007/s11010-019-03668-8.
- [367] R.V. Sharma, R.C. Bhalla, Metformin attenuates agonist-stimulated calcium transients in vascular smooth muscle cells, Clin. Exp. Hypertens. 17 (1995) 913–929, https://doi.org/10.3109/10641969509033643.
- [368] G. Wang, S. Chen, Z. Shao, Y. Li, W. Wang, L. Mao, J. Li, X. Mei, Metformin alleviates hydrogen peroxide-induced inflammation and oxidative stress via inhibiting P2X7R signaling in spinal cord tissue cells neurons, Can. J. Physiol. Pharmacol. 99 (2021) 768–774, https://doi.org/10.1139/cjpp-2020-0373.
   [369] L. Corso, A. Cavallero, D. Baroni, P. Garbati, G. Prestipino, S. Bisti, M. Nobile,
- [369] L. Corso, A. Cavallero, D. Baroni, P. Garbati, G. Prestipino, S. Bisti, M. Nobile, C. Picco, Saffron reduces ATP-induced retinal cytotoxicity by targeting P2X7 receptors, Purinergic Signal 12 (2016) 161–174, https://doi.org/10.1007/ s11302-015-9490-3.
- [370] C.-C. Chao, P. Chan, C.-S. Kuo, C.-L. Gong, T.-H. Cheng, Z.-M. Liu, P.-C. Shen, C.-C. Huang, Y.-M. Leung, Protection of differentiated neuronal NG108-15 cells from P2X7 receptor-mediated toxicity by taurine, Pharmacol. Rep. 66 (2014) 576–584, https://doi.org/10.1016/j.pharep.2014.01.005.
- [371] EMA, Ebixa. Summary of product characteristics, n.d.
- [372] FDA, Namenda. Prescribing information, n.d. [373] DrugBank, Memantine, n.d.
- [374] W. Hare, E. WoldeMussie, R. Lai, H. Ton, G. Ruiz, B. Feldmann, M. Wijono, T. Chun, L. Wheeler, Efficacy and safety of memantine, an NMDA-type openchannel blocker, for reduction of retinal injury associated with experimental glaucoma in rat and monkey, Surv. Ophthalmol. 45 (2001) S284–S289, https:// doi.org/10.1016/S0039-6257(01)00200-4.
- [375] Y. Shinozaki, S. Akanuma, Y. Mori, Y. Kubo, K. Hosoya, Comprehensive evidence of carrier-mediated distribution of amantadine to the retina across the blood-retinal barrier in rats, Pharmaceutics 13 (2021) 1339, https://doi.org/ 10.3390/pharmaceutics13091339.
- [376] B.B. Sysuev, D.K. Salakhetdinov, In vivo study of pharmacokinetic parameters of a new combination drug based on citicoline and memantine, Res. Results Pharmacol. 7 (2021) 23–30, https://doi.org/10.3897/rrpharmacology.7.60380.
- [377] M. Hanefeld, Pharmacokinetics and clinical efficacy of pioglitazone, Int. J. Clin. Pract. Suppl. (2001) 19–25. (http://www.ncbi.nlm.nih.gov/pubmed/11594240).

- [378] D. Eckland, M. Danhof, Clinical pharmacokinetics of pioglitazone, Exp. Clin. Endocrinol. Diabetes 108 (2000) 234–242, https://doi.org/10.1055/s-2000-8525.
- [379] DrugBank, Pioglitazone, n.d.
- [380] C. Grommes, J.C. Karlo, A. Caprariello, D. Blankenship, A. DeChant, G. E. Landreth, The PPARγ agonist pioglitazone crosses the blood–brain barrier and reduces tumor growth in a human xenograft model, Cancer Chemother. Pharmacol. 71 (2013) 929–936, https://doi.org/10.1007/s00280-013-2084-2.
- [381] K. Mishra, M. Nath, N. Halder, T. Velpandian, Evaluation of the possibility of selective modulation of retinal glucose transporters in diabetic complications: an experimental study, Indian J. Pharmacol. 52 (2020) 495, https://doi.org/ 10.4103/ijp.JJP 403 17.
- [382] N. Garrido-Mesa, A. Zarzuelo, J. Gálvez, Minocycline: far beyond an antibiotic, Br. J. Pharmacol. 169 (2013) 337–352, https://doi.org/10.1111/bph.12139.
   [383] DrugBank, Minocycline, (n.d.).
- [384] J. Zhou, B.T. Tran, V.H. Tam, The complexity of minocycline serum protein
- binding, J. Antimicrob. Chemother. 72 (2017) 1632–1634, https://doi.org/ 10.1093/jac/dkx039.
- [385] S. Saivin, G. Houin, Clinical pharmacokinetics of doxycycline and minocycline, Clin. Pharmacokinet. 15 (1988) 355–366, https://doi.org/10.2165/00003088-198815060-00001.
- [386] S. Nagarakanti, E. Bishburg, Is minocycline an antiviral agent? A review of current literature, Basic Clin. Pharmacol. Toxicol. 118 (2016) 4–8, https://doi. org/10.1111/bcpt.12444.
- [387] S.F. Abcouwer, C. Lin, S. Shanmugam, A. Muthusamy, A.J. Barber, D.A. Antonetti, Minocycline prevents retinal inflammation and vascular permeability following ischemia-reperfusion injury, J. Neuroinflamm. 10 (2013) 913, https://doi.org/ 10.1186/1742-2094-10-149.
- [388] L. Yang, Minocycline inhibition of photoreceptor degeneration, Arch. Ophthalmol. 127 (2009) 1475, https://doi.org/10.1001/ archophthalmol 2009 288
- [389] Y.-I. Chen, Y.-J. Lee, D.A. Wilkie, C.-T. Lin, Evaluation of potential topical and systemic neuroprotective agents for ocular hypertension-induced retinal ischemia-reperfusion injury, Vet. Ophthalmol. 17 (2014) 432–442, https://doi. org/10.1111/vop.12105.
- [390] S. Nazarian, H. Åkhondi, Minocycline, n.d. (https://www.ncbi.nlm.nih.gov/ books/NBK554519/).
- [391] J. Rosenthal, Nilvadipine: profile of a new calcium antagonist. An overview, J. Cardiovasc. Pharmacol. 24 Suppl 2 (1994) S92–S107. (http://www.ncbi.nlm.ni h.gov/pubmed/7898101).
- [392] T. Niwa, Y. Tokuma, H. Noguchi, Plasma protein binding of nilvadipine, a new dihydropyridine calcium antagonist, in man and dog, Res. Commun. Chem. Pathol. Pharmacol. 55 (1987) 75–88. (http://www.ncbi.nlm.nih.gov/pubmed/ 3563108).
- [393] M. Terakawa, Y. Tokuma, A. Shishido, H. Noguchi, Pharmacokinetics of Nilvadipine in healthy volunteers, J. Clin. Pharmacol. 27 (1987) 111–117, https://doi.org/10.1002/j.1552-4604.1987.tb02170.x.
- [394] A. von Nieciecki, H.J. Huber, F. Stanislaus, Pharmacokinetics of nilvadipine, J. Cardiovasc. Pharmacol. 20 Suppl 6 (1992) S22–S29. (http://www.ncbi.nlm.ni h.gov/pubmed/1283185).
- [395] R.N. Brogden, D. McTavish, Nilvadipine, Drugs Aging 6 (1995) 150–171, https:// doi.org/10.2165/00002512-199506020-00007.
- [396] DrugBank, NIlvadipine, n.d.
- [397] A. Uemura, A. Mizota, Retinal concentration and protective effect against retinal ischemia of nilvadipine in rats, Eur. J. Ophthalmol. 18 (n.d.) 87–93. (https://doi. org/10.1177/112067210801800115).
- [398] H. Shimizu, Y. Nishimura, Y. Shiide, H. Matsuda, M. Akimoto, M. Matsuda, Y. Nakamaru, Y. Kato, K. Kondo, Evaluation of pharmacokinetics, safety, and drug-drug interactions of an oral suspension of edaravone in healthy adults, Clin. Pharmacol. Drug Dev. 10 (2021) 1174–1187, https://doi.org/10.1002/cpdd.925.
- [399] EMA, Radicava. Withdrawal assessment report, n.d.
- [400] Drugbank, Edaravone, n.d.
- [401] M.P. Cruz, Edaravone (Radicava): a novel neuroprotective agent for the treatment of amyotrophic lateral sclerosis, P T 43 (2018) 25–28. (http://www.ncbi.nlm.nih. gov/pubmed/29290672).
- [402] C. Fong, Improved Edaravone delivery to the brain and crossing the blood brain barrier: using quantum mechanics. [Research Report] Eigenenergy, Adelaide, Australia, 2019. ffhal-02292553v2f, n.d. (https://hal.archives-ouvertes.fr/hal-02292553/document).
- [403] K. Akaiwa, K. Namekata, Y. Azuchi, X. Guo, A. Kimura, C. Harada, Y. Mitamura, T. Harada, Edaravone suppresses retinal ganglion cell death in a mouse model of normal tension glaucoma, e2934–e2934, Cell Death Dis. 8 (2017), https://doi. org/10.1038/cddis.2017.341.
- [404] M. Kaste, S. Murayama, G.A. Ford, D.W.J. Dippel, M.R. Walters, T. Tatlisumak, Safety, tolerability and pharmacokinetics of MCI-186 in patients with acute

ischemic stroke: new formulation and dosing regimen, Cerebrovasc. Dis. 36 (2013) 196–204, https://doi.org/10.1159/000353680.

- [405] K. Smith, M. Hopp, G. Mundin, S. Bond, P. Bailey, J. Woodward, D. Bell, Low absolute bioavailability of oral naloxone in healthy subjects, Int. J. Clin. Pharmacol. Ther. 50 (2012) 360–367, https://doi.org/10.5414/CP201646.
- [406] D.M. Strickland, J.K. Burson, Sublingual absorption of naloxone in a large clinical population, J. Drug Metab. Toxicol. 09 (2018), https://doi.org/10.4172/2157-7609.1000240.
- [407] DrugBank, Naloxone, n.d.
- [408] D. Koyyalagunta, Opioid analgesics, in: Pain Management, Elsevier, 2007, pp. 939–964, https://doi.org/10.1016/B978-0-7216-0334-6.50117-5.
- [409] H. Chapy, P. André, X. Declèves, J.-M. Scherrmann, S. Cisternino, A polyspecific drug/proton antiporter mediates diphenhydramine and clonidine transport at the mouse blood-retinal barrier, Br. J. Pharmacol. 172 (2015) 4714–4725, https:// doi.org/10.1111/bph.13246.
- [410] B. Ba, K. Gaudin, A. Désiré, T. Phoeung, M.-H. Langlois, C.R. Behl, J. Unowsky, I. H. Patel, A.W. Malick, M. Gomes, N. White, T. Kauss, Ceftriaxone absorption enhancement for noninvasive administration as an alternative to injectable solutions, Antimicrob. Agents Chemother. 62 (2018), https://doi.org/10.1128/ AAC.01170-18.
- [411] N. Dailymed, Ceftriaxone, n.d. (https://dailymed.nlm.nih.gov/dailymed/drugInf o.cfm?setid=4d1ad77f-2c6b-4250-82e5-ab3574444e08).
- [412] I.H. Patel, S. Chen, M. Parsonnet, M.R. Hackman, M.A. Brooks, J. Konikoff, S. A. Kaplan, Pharmacokinetics of ceftriaxone in humans, Antimicrob. Agents Chemother. 20 (1981) 634–641, https://doi.org/10.1128/AAC.20.5.634.
- [413] DrugBank, Ceftriaxone, n.d.
- [414] R. Nau, H.W. Prange, P. Muth, G. Mahr, S. Menck, H. Kolenda, F. Sörgel, Passage of cefotaxime and ceftriaxone into cerebrospinal fluid of patients with uninflamed meninges, Antimicrob. Agents Chemother. 37 (1993) 1518–1524, https://doi. org/10.1128/AAC.37.7.1518.
- [415] L. Brockhaus, D. Goldblum, L. Eggenschwiler, S. Zimmerli, C. Marzolini, Revisiting systemic treatment of bacterial endophthalmitis: a review of intravitreal penetration of systemic antibiotics, Clin. Microbiol. Infect. 25 (2019) 1364–1369, https://doi.org/10.1016/j.cmi.2019.01.017.
- [416] M. Sharir, G. Triester, J. Kneer, E. Rubinstein, The intravitreal penetration of ceftriaxone in man following systemic administration, Investig. Ophthalmol. Vis. Sci. 30 (1989) 2179–2183. (http://www.ncbi.nlm.nih.gov/pubmed/2793358).
- [417] R.A.J. Smith, M.P. Murphy, Animal and human studies with the mitochondriatargeted antioxidant MitoQ, Ann. N. Y. Acad. Sci. 1201 (2010) 96–103, https:// doi.org/10.1111/j.1749-6632.2010.05627.x.
- [418] R.A. Zinovkin, A.A. Zamyatnin, Mitochondria-targeted drugs, Curr. Mol. Pharmacol. 12 (2019) 202–214, https://doi.org/10.2174/ 1874467212666181127151059.
- [419] H. Sies, L. Packer, Quinones and Quinone Enzymes, Elsevier Inc, 2004.
- [420] K.M. Taylor, R.A.J. Smith, Mitoquinone derivatives used as mitochondrially targeted antioxidants, n.d.
- [421] N.M. Zaki, Strategies for oral delivery and mitochondrial targeting of CoQ10, Drug Deliv. (2014) 1–14, https://doi.org/10.3109/10717544.2014.993747.
- [422] I. Bogeski, R. Gulaboski, R. Kappl, V. Mirceski, M. Stefova, J. Petreska, M. Hoth, Calcium binding and transport by coenzyme Q, J. Am. Chem. Soc. 133 (2011) 9293–9303, https://doi.org/10.1021/ja110190t.
- [423] J.M. Villalba, C. Parrado, M. Santos-Gonzalez, F.J. Alcain, Therapeutic use of coenzyme Q 10 and coenzyme Q 10 -related compounds and formulations, Expert Opin. Investig. Drugs 19 (2010) 535–554, https://doi.org/10.1517/ 13543781003727495.
- [424] D. Vlachantoni, A.N. Bramall, M.P. Murphy, R.W. Taylor, X. Shu, B. Tulloch, T. Van Veen, D.M. Turnbull, R.R. McInnes, A.F. Wright, Evidence of severe mitochondrial oxidative stress and a protective effect of low oxygen in mouse models of inherited photoreceptor degeneration, Hum. Mol. Genet. 20 (2011) 322–335, https://doi.org/10.1093/hmg/ddq467.
- [425] G. Zaccara, A. Messori, F. Moroni, Clinical pharmacokinetics of valproic acid– 1988, Clin. Pharmacokinet. 15 (1988) 367–389, https://doi.org/10.2165/ 00003088-198815060-00002.
- [426] DrugBank, Valproic acid, n.d.
- [427] R. Gugler, G.E. von Unruh, Clinical pharmacokinetics of valproic Acid1, Clin. Pharmacokinet. 5 (1980) 67–83, https://doi.org/10.2165/00003088-198005010-00002.
- [428] FDA, Depakene prescribing information, n.d.
- [429] Y. Kubo, S. Akanuma, K. Hosoya, Influx transport of cationic drug at the blood-retinal barrier: impact on the retinal delivery of neuroprotectants, Biol. Pharm. Bull. 40 (2017) 1139–1145, https://doi.org/10.1248/bpb.b17-00090.
- [430] Y. Kubo, E. Fukui, S.-I. Akanuma, M. Tachikawa, K.-I. Hosoya, Application of membrane permeability evaluated in in vitro analyses to estimate blood–retinal barrier permeability, J. Pharm. Sci. 101 (2012) 2596–2605, https://doi.org/ 10.1002/jps.23171.